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1	SARS-CoV-2-like viruses from captive Guangdong pangolins generate circular RNAs
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3	
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11	
12	Viruses closely related to SARS-CoV-2, the virus responsible for the Covid-19
13	pandemic, have previously been identified in several Sunda pangolins (Manis javanica)
14	seized in Guangdong and Guangxi provinces of China ¹⁻⁴ . The virus extracted from
15	Guangdong pangolins is particularly intriguing as the amino-acid sequence of the
16	receptor binding domain of the spike protein is nearly identical to that of the human
17	SARS-CoV-2 virus ^{2,3} . This characteristic suggests it is capable of binding to the human
18	ACE2 receptor and may therefore be able to mediate infection of human cells.
19	Here, we assembled SARS-CoV-2-like genomes from the Sequence Read Archive
20	(SRA) data available in May 2020 for ten Guangdong pangolins. As previously
21	described ⁵ the genome assemblies were of poor quality, having high levels of missing
22	data and with possible DNA contamination from human, mouse and tiger. We found 10
23	circular RNAs (circRNAs) of 278-776 nucleotides (nt) containing SARS-CoV-2-like

- sequences in three pangolin lung samples. This is the first time this class of molecule is
 reported to be generated from an RNA-genome virus.

27	Next-generation sequencing (NGS) runs previously published for 16 pangolin samples
28	were analysed (Table 1) ¹⁻³ . A large number of SARS-CoV-2-like reads (\geq 184,870) were
29	found in SRA datasets generated from lungs and intestines of Guangxi pangolins, whereas a
30	limited number of reads (≤1,078) were found in SRA datasets generated from lungs of
31	Guangdong pangolins. These differences suggest higher RNA degradation in Guangdong
32	samples and explain why it was not possible to assemble a full virus genome from these data.
33	Assuming the same virus infected Guangdong pangolins ^{3,4} , we pooled together the 4688
34	SARS-CoV-2-like reads found in the ten Guangdong datasets to assemble a consensus
35	sequence. The resulting genome remained partial, with 15 fragments and 4% missing data
36	(mean coverage 16.6X). Two recent studies ^{3,4} had filled genome gaps by sequencing multiple
37	PCR products. However, these two published genomes differ at 15 nucleotide positions
38	(Supplementary Information) indicating the quality of Guangdong pangolin virus sequence
39	still needs to be improved ⁵ .
40	In three SRA datasets generated from lung samples of three Guangdong pangolins
41	(Table 1), we discovered several circRNAs by mapping NGS reads to the reference genome
42	(GISAID accession number EPI_ISL_410721 ³) using Geneious Prime® 2020.0.3 and the
43	"low sensitivity" option (maximum mismatch: 10%). For the GD/P5L dataset ³ (Table 1),
44	most SARS-CoV-2-like reads (566 of all 600 reads) mapped to a small portion of the ORF1ab
45	gene (Figure 1A). However, the 5'-and 3'-flanking regions of the contig were found to
46	contain chimeric reads, in which the extremities could not be aligned to the reference genome.
47	A preliminary consensus sequence of 748 nt was extracted and then used as a reference to
48	extend the contig by mapping GD/P5L reads (three iterations in Geneious Prime® 2020.0.3).

49 The final contig contained 635 reads, and the consensus sequence of 862 nt was then 50 extracted for further analysis. As shown in **Figure 1B**, its flanking regions (highlighted in 51 yellow) are parts of an original circRNA of 733-nt long, herein referred to as circRNA-52 GDP5L-1: the 5'-flanking region is identical to the 3'-end of circRNA-GDP5L-1 (positions 53 685-733), whereas the 3'-flanking region is identical to the 5'-end of circRNA-GDP5L-1 54 (positions 1-80). Such repetitions of identical sequences are consistent with a circular nature 55 of the original RNA molecule. In support of this finding, the high number of reads found for 56 *circRNA-GDP5L-1* (635 reads) suggests the molecule was still circular before reverse transcription (see protocol used for library construction³) as multiple cDNA copies of a single 57 58 circRNA can be generated through rolling circle amplification⁶. The structure of *circRNA*-59 GDP5L-1 is characterized by the presence of a 33-nt motif (positions 1-33, shown in red in 60 Figure 1B) which cannot be aligned to the reference genome, but which is the full reverse 61 complement of the sequence located 20-nt downstream (underlined sequence in Figure 1B). Based on computer modeling tools⁷, we can predict *circRNA-GDP5L-1* to form a highly 62 63 stable secondary structure (Supplementary Information). We found nine other highly 64 structured circRNAs showing similar features: two further in the GD/P5L dataset, two in the GD/P7L dataset, and five in the GD/P8L dataset. Their full sequences, characteristics and 65 66 secondary structures are detailed in Supplementary Information. The length of these 67 circRNAs varies between 278 and 776 nt. Only a single circRNA contained a viral sequence 68 without additional motif. The nine other circRNAs are composed of a viral genomic sequence 69 followed by the full reverse complement of a viral motif (length between 5 and 96 nt) found a 70 few nucleotides downstream (between 14 and 62 nt). Notably, a substantial proportion of 71 these genomic sequences are protein-coding-fragments (see details in Supplementary 72 Information).

CircRNAs form a large class of RNA molecules that are widespread in eukaryotes⁸. 73 74 They play important functions in gene regulation as they can have interactions with RNAbinding proteins, serve as translation templates, and act as microRNA sponges or 75 76 transcriptional regulatory factors⁸⁻¹⁰. Recent studies have reported that DNA-virus genomes are also capable of generating circRNAs^{11,12}. To our knowledge, these molecules have not 77 78 previously been described as by-products of RNA-virus genomes. It is therefore important to 79 know whether these circRNAs are natural or not, since some circRNAs have been described as possible artifacts of the reverse transcription process¹³. In addition, an intriguing issue is 80 81 that the three SRA datasets in which circRNAs were found are the same three datasets 82 containing human and mouse DNA (**Table 1**)⁵. The comparison between NGS runs 83 performed on RNAs extracts from pangolin lungs (Table 1) clearly show much more SARS-84 CoV-2-like reads for Guangxi datasets than for Guangdong datasets (≥184,870 versus 85 \leq 1,078). Two hypotheses can be formulated to explain these differences: Guangdong samples 86 contain lower concentrations of SARS-CoV-2-like virus; or RNA molecules extracted from 87 Guangdong samples were much more degraded due to poor sample preservation. In support of 88 the second hypothesis, it has been found that transcriptome degradation can result in circRNA enrichment as linear RNAs decay more rapidly than circRNAs¹⁴, suggesting that the 10 89 90 circRNAs discovered in the three Guangdong pangolin samples are true by-products of 91 SARS-CoV-2 like RNA-genomes. Another argument for the natural origin of these circRNAs is provided by the alignment of their 5'-end with homologous sequences extracted from 92 93 SARS-CoV-2 like genomes (Supplementary Information). In the case of *circRNA-GDP5L-1*, 94 the two additional base-pairing motifs involved in the secondary structure (highlighted in 95 green in Figure 3C) are found to be highly conserved among SARS-CoV-2-like viruses, 96 whereas the genomic fragment is a part of a protein-coding gene. In this context, the fact that 97 canonical base-pairing motifs have been restored through compensatory mutations in the virus

98 genome from Guangxi pangolins (position 2: G=>A and position 19: C=>T in Figure 3C) is 99 particularly relevant, as it suggests strong natural selection pressure for maintaining a RNA-100 stem-loop (hairpin) structure. This hypothesis is also corroborated by the high conservation of 101 canonical base-pairing motifs in all other circRNAs and by the detection of similar 102 compensatory mutations in three other circRNAs (circRNA-GDP7L-1, circRNA-GDP7L-2, 103 and *circRNA-GDP8L-3*; Supplementary Information), suggesting that the hairpin is an 104 important structure that may be directly involved in the circularization process. 105 But what can the role of viral circRNAs be during host infection? It has been 106 demonstrated that viral infection triggers the degradation of host-produced circRNAs which 107 in turn activates the innate immune response¹⁵. This mechanism suggests viral circRNAs 108 could have an important role, either to delay the host's immune response, or to initiate viral 109 proliferation in host cells. Our discovery of circRNAs generated from pangolin SARS-CoV-2-110 like virus opens up new perspectives for understanding virus-host interactions and has 111 possible implications in a wider range of RNA-virus infections including Covid-19.

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113 **References**

114 1. Liu, P. et al. Viral Metagenomics Revealed Sendai Virus and Coronavirus Infection of

115 Malayan Pangolins (*Manis javanica*). Viruses **11**, (2019).

116 2. Lam, T.T. *et al.* Identifying SARS-CoV-2 related coronaviruses in Malayan pangolins.

117 *Nature* doi: 10.1038/s41586-020-2169-0. (2020).

- 3. Xiao, K. *et al.* Isolation of SARS-CoV-2-related coronavirus from Malayan pangolins. *Nature* (2020).
- 120 4. Liu, P. et al. Are pangolins the intermediate host of the 2019 novel coronavirus (SARS-
- 121 CoV-2)? *PLoS Pathog.* **16**, (2020).

- 122 5. Hassanin, A. The SARS-CoV-2-like virus found in captive pangolins from Guangdong
- should be better sequenced. (submitted; doi: https://doi.org/10.1101/2020.05.07.077016).
- 124 6. Szabo, L., Salzman, J. Detecting circular RNAs: bioinformatic and experimental
- 125 challenges. *Nat Rev Genet.* **17**, (2016).
- 126 7. Gruber, A.R. et al. The ViennaRNA web services. *Methods Mol Biol.* 1269, (2015).
- 127 8. Memczak, S. et al. Nature. Circular RNAs are a large class of animal RNAs with
- 128 regulatory potency. **495**, (2013).
- 129 9. Chen, L.L. *et al.* The expanding regulatory mechanisms and cellular functions of circular
- 130 RNAs. Nat Rev Mol Cell Biol. Online ahead of print (2020).
- 131 10. Xie, R. *et al.* Circular RNAs are a large class of animal RNAs with regulatory potency.
- 132 *Nature* **495**, (2020).
- 133 11. Ungerleider, N. et al. The Epstein Barr virus circRNAome. PLoS Pathog. 14, (2018).
- 12. Toptan, T. *et al.* Circular DNA tumor viruses make circular RNAs. *Proc Natl Acad Sci U S A* 115, (2018).
- 136 13. Chen I, et al. Biogenesis, identification, and function of exonic circular RNAs. *Wiley*
- 137 *Interdiscip Rev RNA.* **6**, (2015).
- 138 14. Alhasan, A.A. *et al.* Circular RNA enrichment in platelets is a signature of transcriptome
 139 degradation. *Blood* 127, (2016).
- 140 15. Liu, C.X. et al. Structure and Degradation of Circular RNAs Regulate PKR Activation in
- 141 Innate Immunity. *Cell* **177**, (2019).
- 142
- 143

144 Figure 1. Identification and characterization of *circRNA-GDP5L-1*

145 (A) The reads of the *GD/P5L* SRA dataset were mapped to the SARS-CoV-2-like genome

sequenced from Guangdong pangolins (GISAID accession number EPI ISL 4107214) using

147 Geneious Prime® 2020.0.3. The preliminary consensus sequence was then used as a reference

148 for three mapping iterations in order to extend the contig.

149 (B) The final consensus sequence of 862 nucleotides was constructed from a contig of 635

reads. The *circRNA-GDP5L-1* is 733-nt long (red and black nucleotides not highlighted in

151 yellow). The 5'- and 3'-flanking regions, highlighted in yellow, support the circular nature of

the RNA molecule, as they are perfectly identical to the 3'- and 5'-ends of the *circRNA*-

153 *GDP5L-1* sequence, respectively. The 33-nt motif shown in red cannot be aligned to the

154 reference genome but is the perfect reverse complement of the underlined motif, located 20-

155 nucleotides downstream.

156 (C) The alignment of the genomic sequences located between the two complementary motifs

157 shows the existence of additional base-pairing motifs (highlighted in green) which are highly

158 conserved among SARS-CoV-2-like genomes, i.e., GX/P (same sequence found in five virus

159 genomes from Guangxi pangolins; GenBank accession numbers: MTO40333-6 and

160 MT072864), SARS-CoV-2 (NC_045512, human reference genome), RaGT13 (MN996532,

virus genome from the bat species *Rhinolophus affinis*), VZC45 (MG772933) and VZXC21

162 (MG772934), two virus genomes from the bat species *Rhinolophus sinicus*.



A. GD/P5L reads mapped to the reference genome (hCov-19/pangolin/Guangdong)

B. CircRNA-GDP5L-1 sequence

C. Alignment of the genomic sequences located between the two complementary motifs



 Table 1. Analyses of SRA data available for SARS-CoV-2-like viruses detected in captive pangolins from Guangdong (GD) and Guangxi (GX).

Illumina sequencing run			Viral genome assembly			Reads mapped to mitochondrial genomes			
Code - Tissue	NCBI SRA	Reads	Reads	МС	MD	pangolin NC_026781	human NC_012920	mouse NC_005089	
GD/P7L ¹ - Lung 07	SRR10168378	38,091,846	285*	1.4X	57%	98,226	28	0	1,634
GD/P8L ¹ - Lung 08	SRR10168377	32,829,850	1,078*	5.3X	19%	7,727	1,333	0	183
GD/P9L ¹ - Lung 09	SRR10168376	36,135,230	36	0.2X	88%	13,770	47	3,447	0
GD/P11L ¹ - Lung 11	SRR10168375	44,440,374	10	0.2X	99%	807,747	24	1,394	3
GD/P2S ² – Scale P2	SRR11093265	2,633	2,604	6.5X	29%	0	0	0	0
GD/P1L ³ - Lung Mj1	SRR11119766	46,4866,240	10	0.1X	97%	1,952,841	0	0	0
GD/P2L ³ - Lung Mj2	SRR11119765	424,322,500	0	NA	NA	492,015	0	0	0
GD/P3L ³ - Lung Mj3	SRR11119763	393,522,404	8	0.1X	97%	319,594	0	112	0
GD/P4L ³ - Lung Mj4	SRR11119762	547,302,862	58	0.3X	83%	281,251	0	0	0
GD/P5L ³ - Lung Mj5	SRR11119759	214,534,718	600*	3.0X	92%	333,399	9,637	0	20
GX/P1E ² - Intestine	SRR11093266	470,344	450,433	510.5X	1%	25	0	0	0
GX/P2V ² - Vero cells	SRR11093271	4,550,437°	57,356	289.8X	0%	4	0	0	0
GX/P3B ² - Blood	SRR11093270	19,607,536	1,794	7.3X	16%	52,875	1,174	17	9
GX/P4L ² - Lung	SRR11093269	520,254	184,870	2002.1X	3%	7	0	0	0
GX/P5E ² - Intestine	SRR11093268	416,659	401,378	3958.2X	1%	8	0	0	0
GX/P5L ² - Lung	SRR11093267	340,661	274,982	2749.3X	2%	59	0	0	0

*: datasets in which circRNA reads were detected (see main text for details). Abbreviations: MC: mean coverage; MD: missing data.

Supplementary information 1

<u>Nucleotide differences between the four SARS-CoV-2-like genomes sequenced from</u> <u>Guangdong pangolins.</u>

Apomorphy lists associated to the following parsimonious tree reconstructed in PAUP* version 4.0b10 (Swofford, 2003).

\		GDP1L/2019²
	\	GenBank MT121216 ⁴
 6	/5	GISAID EPI_ISL_4107214 ³
/		GenBank MT084071

Branch	Characte	er CI	Change
node_6> MT084071	5836	0.500	A> G
	5838	0.500	А> Т
	5851	0.500	A> C
	8870	1.000	G ==> A
	8875	1.000	T ==> A
	8878	1.000	T ==> C
	8883	1.000	A ==> C
	8887	1.000	C ==> G
	8889	1.000	T ==> C
	8891	1.000	C ==> G
	8892	1.000	A ==> G
	8893	1.000	T ==> C
	8894	1.000	T ==> C
	8895	1.000	T ==> G
	8898	1.000	T ==> G
	8900	1.000	C ==> G
	10216	1.000	A> G
	11865	1.000	A ==> T
	25027	1.000	T ==> A
	25028	1.000	T ==> A
	25535	1.000	A> G
	29420	1.000	С ==> Т
	29421	1.000	T ==> C
	29423	1.000	С> Т
	29427	1.000	A> T
node_6> node_5	4057	1.000	С> Т
	4168	1.000	G ==> T
	4171	1.000	С ==> Т
	4173	1.000	A ==> C
	4174	1.000	A ==> C
	4175	1.000	C ==> G

	4176	1.000	T ==> A
	4177	1.000	G ==> A
	4178	1.000	T ==> A
	4181	1.000	T ==> C
	4182	1.000	С> Т
	4183	1.000	С> Т
	6143	1.000	A ==> G
	6920	1.000	G> Т
	7240	1 000	А> Т
	8948	1 000	A> C
	15390	1 000	A> G
	22542	1 000	A> C
	25051	1.000	С> Т
	25051	1.000	
	25536	1.000	и ==> с с ==> т
	25538	1.000	$C \implies 1$ $C \implies \lambda$
	25530	1.000	G ==> Д С ==> Т
	25559	1.000	$C \rightarrow C$
	29422	1.000	
	29424	1.000	$C \rightarrow C$
	29423	1.000	C> G
	29420	1.000	A> G
	29800	1.000	$C \rightarrow T$
	29801	1.000	C> G
rada 5 > EDI ISI 4107214	29802	1.000	G> 1
10de_3> EP1_13L_410/214	3837	0.300	C ==> G
	9042	1.000	A ==> G
	9089	1.000	A ==> C
	23090	1.000	A> T
	23508	1.000	A == > T
	24233	1.000	C ==> T
$node_5 -> M1121216$	2193	1.000	A> G
	2730	1.000	A ==> G
	4993	0.500	C> T
	5836	0.500	A> G
	5838	0.500	A> T
	3831	0.500	A> C
	18094	1.000	C> T
	23433	1.000	A> C
	26536	1.000	G> T
node_6> GDP1L/2019	4967	1.000	G ==> T
	4993	0.500	С> Т
	5837	0.500	C> G
	5840	1.000	G ==> T
	8226	1.000	С> Т
	8253	1.000	G> T -
	19388	1.000	T ==> C
	23294	1.000	С> Т
	23395	1.000	С> Т
	29799	1.000	A> C

Supplementary Information 2

<u>Sequences and characteristics of the 10 circRNAs detected in SRA datasets generated</u> <u>from Guangdong pangolin samples in which SARS-Cov-2-like virus were identified.</u>

As shown in **Figure 1**, the reads were mapped to the SARS-CoV-2-like genome sequenced from Guangdong pangolins (GISAID accession number EPI_ISL_410721³). All contigs with chimeric reads at the 5'-and 3'-flanking regions were considered as possible circRNAs, and further studied.

The characteristics of the 10 circRNAs here detected are described by the following data:

- (1) the viral sequence identical to the reference genome is written in black;
- (2) 5'- and 3'-flanking sequences are highlighted in yellow;
- (3) the additional motif is shown in red;
- (4) its reverse complement in the viral genomic sequence is underlined; and
- (5) the viral sequence between these two complementary motifs is generally

characterized by additional base-pairing motifs, which are highlighted in green.

The evolution of these additional base-pairing motifs was analyzed by aligning the following SARS-CoV-2 genomes: GX/P (representing the five full genomes sequenced from Guangxi pangolins MTO40333-6 + MT072864); SARS-CoV-2 (NC_045512, human reference genome); RaGT13 (genome sequenced from the bat species *Rhinolophus affinis*); and VZC45 (MG772933) VZXC21(MG772934), two genomes isolated from the bat species *Rhinolophus sinicus*.

For each sequence, the centroid secondary structure was predicted using ViennaRNA web services⁷. In this structure, the location of the two full complementary motifs is indicated by a red arrow, whereas the location of additional base-pairing motifs is indicated by a green arrow.

Characteristics of the 10 circRNAs identified in this study in three lung samples from	n
Guangdong pangolins (length in nucleotides)	

circRNA name	VGF length	SARS gene(s)	Genomic position*	Additional motif length	Distance (nt) from its RC
circRNA-GDP5L-1	700	ORF1ab	625-1323	33	20
circRNA-GDP5L-2	268	ORF8	28003-28270	10	14
circRNA-GDP5L-3	412	N + ORF10 + 3' end	29424-29890	96	19
circRNA-GDP7L-1	302	ORF1ab	13629-13930	35	32
circRNA-GDP7L-2	329	ORF1ab	18694-19022	5	62
circRNA-GDP8L-1	610	ORF1ab	12120-12729	NA	NA
circRNA-GDP8L-2	512	ORF1ab	16230-16741	14	18
circRNA-GDP8L-3	681	ORF1ab	18694-19374	10	62
circRNA-GDP8L-4	758	ORF1ab	20677-21434	18	17
circRNA-GDP8L-5	555	Ν	28581-29135	5	22

Abbreviations: VGF=viral genomic fragment; RC= reverse complement in the VGF. *reference genome = hCoV-19/pangolin/Guangdong/EPI_ISL_410721³. °partial RC in the VGF.

>circRNA-GD/P5L-1 [733 nt; contig of 635 reads]

Alignment of genomic sequences located between the two complementary motifs

0	0	1
GD/P	A <mark>GTTCTT</mark> C	CTTCGC <mark>AAGAAC</mark> G
GX/P	- <mark>A</mark> -	CT <mark>T</mark> -
SARS-CoV-2	G <mark></mark> -	T <mark></mark> -
RaGT13	G <mark></mark> -	T <mark></mark> -
VZC45		T <mark></mark> -
VZXC21		T <mark></mark> -

The base-pairing motifs (the reverse complement being underlined) are fully conserved in other SARS-CoV-2-like genomes. In GX/P genomes, the canonical base-pairing motifs have been restored through compensatory mutations (G => A and C => T).

Centroid secondary structure (minimum free energy=256.20 kcal/mol)



>circRNA-GDP5L-2 [278 nt; contig of 17 reads]

Alignment of genomic sequences located between the two complementary motifs

GD/P	AA.	AG	T	CAGCAC	C	\mathbf{TTT}
GX/P		-A	_	-T	-	A
SARS-CoV-2		–A			_	
RaGT13		–A			_	
VZC45		–A	_		_	
VZXC21		–A			_	

The base-pairing motifs (the reverse complement being underlined) are partially conserved (3/4 nucleotides) in other SARS-CoV-2-like genomes. In GX/P genomes, different base-pairing motifs were found in the same region.

Centroid secondary structure (minimum free energy=49.20 kcal/mol)



>circRNA-GDP5L-3 [508 nt; contig of 17 reads]

Alignment of genomic sequences located between the two complementary motifs

GD/P	TTTGGA	TGATTTC	TCCAAA
GX/P			
SARS-CoV-2			
RaGT13		C	
VZC45		A	
VZXC21		A	

The base-pairing motifs (the reverse complement being underlined) are fully conserved in other SARS-CoV-2-like genomes.

Centroid secondary structure (minimum free energy=230.30 kcal/mol)



>circRNA-GDP7L-1 [337 nt; contig of 8 reads]

Alignment of genomic sequences located between the two complementary motifs

GD/P	C <mark>ATAGTT</mark> A	AGAGACACACTTTC	rC <mark>T</mark>	<mark>AACTAT</mark> C
GX/P		T		
SARS-CoV-2	Т <mark>G</mark>			C-
RaGT13	Т <mark>G</mark>			C-
VZC45	- <mark>G</mark>	GAA-G-	<mark>C</mark>	C-
VZXC21	- <mark>G</mark>	GAA-G-	C	C-

The base-pairing motifs (the reverse complement being underlined) are fully or partially conserved (6/7 nucleotides) in other SARS-CoV-2-like genomes. In pangolin viral genomes, the canonical base-pairing motifs have been restored through compensatory mutations (G=>A and C=>T).

Centroid secondary structure (minimum free energy=119.00 kcal/mol)



>circRNA-GDP7L-2 [332 nt; contig of 6 reads]

Alignment of genomic sequences located between the two complementary motifs

8	8 · · · · · · · · · · · · · · · · · · ·	
GD/P	TGCTGGCATCACTCAATTGGGTTCTTTCATGATTGATGTT	CAGCA
GX/P	<mark>T</mark> GAT	A
SARS-CoV-2	<mark>T</mark> TTATGT	A
RaGT13	<mark>T</mark> TTATGT	A
VZC45		
VZXC21		

The base-pairing motifs (the reverse complement being underlined) are fully or partially conserved (4/5 nucleotides) in other SARS-CoV-2-like genomes. In the Guangdong pangolin viral genome, the canonical base-pairing motifs have been restored through compensatory mutations (T=>C and A=>G).

Centroid secondary structure (minimum free energy=97.20 kcal/mol)



>circRNA-GDP8L-1 [610 nt; contig of 89 reads]

Alignment of genomic sequences in the 5'-part of the circRNA

GD/P	AC	TGCT	CA	GGAAGCTTA	TGAGCA	GCT	GT
GX/P		A	1	<u>A</u>		AG	
SARS-CoV-2			1	<u>A</u>		<mark>-</mark>	
RaGT13	_	C	1	<u>A</u>			
VZC45			1	AG		<mark>-</mark>	
VZXC21		AC	1	AG			

The base-pairing motifs (the reverse complement being underlined) are fully or partially conserved (at least 5/7 nucleotides) in other SARS-CoV-2-like genomes. In the VZXC21 genome, different base-pairing motifs were found in the same region.

Centroid secondary structure (minimum free energy=165.30 kcal/mol)



>circRNA-GDP8L-2 [526 nt; contig of 55 reads]

Alignment of genomic sequences located between the two complementary motifs

GD/P	TGT	TCTTTGTAATTC	ACA
GX/P		CC	
SARS-CoV-2		C	
RaGT13		C	
VZC45		GT-AC	
VZXC21		АТ-А	

The base-pairing motifs (the reverse complement being underlined) are fully conserved in other SARS-CoV-2-like genomes.

Centroid secondary structure (minimum free energy=137.80 kcal/mol)



>circRNA-GDP8L-3 [691 nt; contig of 85 reads]

Alignment of genomic sequences located between the two complementary motifs

GD/P	TGCTG	GCATCACTCACATGATTGATGTT	CAG	CA
GX/P	T		A	
SARS-CoV-2	T	TTT	A	
RaGT13	T	TTT	A	
VZC45	T	-AC		
VZXC21	T	-AC		

The base-pairing motifs (the reverse complement being underlined) are fully or partially conserved (4/5 nucleotides) in other SARS-CoV-2-like genomes. In the Guangdong pangolin viral genome, the canonical base-pairing motifs have been restored through compensatory mutations (T=>C and A=>G).

Centroid secondary structure (minimum free energy=166.30 kcal/mol)



>circRNA-GDP8L-4 [776 nt; contig of 40 reads]

Alignment of genomic sequences located between the two complementary motifs

GD/P	СТТ	ΓT.	ACAAAATGC	AAAG
GX/P				-G
SARS-CoV-2				
RaGT13	<mark></mark> 0	2-		
VZC45	0	2-	- T	-G
VZXC21			- T	-G

The base-pairing motifs (the reverse complement being underlined) are fully or partially conserved (at least 3/4 nucleotides) in other SARS-CoV-2-like genomes.

Centroid secondary structure (minimum free energy=164.30 kcal/mol)



>circRNA-GDP8L-5 [560 nt; contig of 92 reads]

Alignment of genomic sequences located between the two complementary motifs

GD/P	AGC	TGGACTTCCCTATGGT	GCT
GX/P			
SARS-CoV-2			
RaGT13			
VZC45			
VZXC21			

The sequence was found to be fully conserved in all SARS-CoV-2-like genomes.

Centroid secondary structure (minimum free energy=135.80 kcal/mol)

