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**Role of Chemical Dynamics Simulations in Mass Spectrometry Studies of
Collision-Induced Dissociation and Collisions of Biological Ions with Organic Surfaces**

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Abstract

In this article, a perspective is given of chemical dynamics simulations of collisions of biological ions with surfaces and of collision-induced dissociation (CID) of ions. The simulations provide an atomic-level understanding of the collisions and, overall, are in quite good agreement with experiment. An integral component of ion/surface collisions is energy transfer to the internal degrees of freedom of both the ion and the surface. The simulations reveal how this energy transfer depends on the collision energy, incident angle, biological ion, and surface. With energy transfer to the ion's vibration fragmentation may occur, i.e. surface-induced dissociation (SID), and the simulations discovered a new fragmentation mechanism, called shattering, for which the ion fragments as it collides with the surface. The simulations also provide insight into the atomistic dynamics of soft-landing and reactive-landing of ions on surfaces. The CID simulations compared activation by multiple "soft" collisions, resulting in random excitation, versus high energy single collisions and non-random excitation. These two activation methods may result in different fragment ions. Simulations provide fragmentation products in agreement with experiments, and hence can provide additional information regarding the reaction mechanisms taking place in experiment. Such studies paved the way on using simulations as an independent and predictive tool in increasing fundamental understanding of CID and related processes.

Keywords: Surface Induced Dissociation; Collision Induced Dissociation; Chemical Dynamics Simulations; Theoretical Modeling

I. Introduction

The study of biological ions fragmentation is an important component of mass spectrometry.¹⁻¹⁰ In this perspective article, collisions of these ions with surfaces¹⁻⁶ and their gas-phase collision-induced dissociation (CID)⁷⁻¹⁰ are considered. Ions with a broad range of mass and charge may be prepared by electrospray ionization (ESI).¹¹ The resulting fragmentation products, in CID or with a surface (surface-induced dissociation, SID^{12,13}), can provide additional information regarding the biological molecules under analysis. This review will highlight important advances that can now be obtained from simulations of such processes.

In SID, a specific ion is selected, given a fixed translational energy, and collided with a surface. When sufficient translational energy is imparted to the ion, it may fragment and provide information regarding the ion's dissociation energetics and mechanisms, as well as a fingerprint of the ion's primary structure. In addition to SID, the ion may be deposited on the surface by physisorption, soft-landing (SL),^{14,15} or chemisorption, reactive landing (RL),¹⁶ these are hyperthermal processes in contrast to thermal physisorption and chemisorption. When a molecule sticks to a surface, it can bind with either chemical interactions (chemisorption) or physical interactions (physisorption). Chemisorption involves the formation of a chemical bond between the adsorbate and surface.¹⁷ Physisorption has weaker interactions, involving dispersion and electrostatic forces. For SL the ion desorbs intact from the surface¹⁵ and is physisorption, while for RL the ion is chemically bound to the surface and is chemisorption. SL and RL interface nicely with traditional surface science experiments of non-thermal gas-surface collisions leading to physisorption and chemisorption.¹⁸⁻²³ The chemisorption probability has been investigated versus both the projectile's collision energy^{18,19} and vibrational energy.¹⁹ Physisorption has been studied experimentally for both translationally²⁰⁻²² and vibrationally²³ excited projectiles, and in chemical dynamics simulations versus translational energy.^{24,25}

SL and RL have important applications including purification of compounds from complex mixtures,²⁶⁻²⁸ preparation of protein or peptide microarrays,²⁹ development of biocompatible substrates and biosensors,^{30,31} deposition of mass-selected cluster ions,³²⁻³⁴ and preparation of novel synthetic materials,^{35,36} including nanomaterials.^{37,38} Understanding the molecular basis of SL and RL is thus crucial to further advances of such techniques.

In CID, ions collide with an inert gas, such as N_2 or a noble gas (Ar, Ne, Xe), and as in SID provide a fragmentation pattern giving information regarding the primary structure of the ion. One important application of CID is in characterization of polypeptides, whose different types of fragments have a specific nomenclature.³⁹ In peptide CID, several studies were performed to understand fragmentation products and related mechanisms.^{9,40-45} In CID there is a large number of different instruments (details can be found in dedicated literature⁴⁶) with specific characteristics which can influence product distribution. In particular, differences in the ion formation method, as well as the way the ion is controlled in the instrument and activated by collision with the inert gas may largely affect the types and the relative abundance of products. Obtaining direct experimental insight into the fragmentation mechanism⁴⁷⁻⁵² is possible by studying the reactants and fragments via Infra-Red Multiple Photon Dissociation (IRMPD) spectroscopy.⁵³ Such fundamental studies on CID fragmentation mechanisms were also done on other biological molecules, like nucleic acids⁵⁴⁻⁵⁸ and carbohydrates.⁵⁹⁻⁶⁵ Applications of CID are found in many fields like proteomics,⁶⁶⁻⁶⁸ metabolomics,⁶⁹ forensic sciences,⁷⁰ and analysis and detection of doping substances.⁷¹ Understanding the molecular mechanisms of CID is a key step to predict fragmentations independently from experiments. Chemical dynamics simulations can help at this aim, since it is possible (as we will detail in the present article) to use them to let the ions fragment by modeling the CID conditions. These simulations are able to predict fragmentation products as well as corresponding reaction mechanisms: they can now be used not only to understand experimental spectra but also to predict fragmentation mechanisms of new structures, like e.g. different isomers with the same chemical formula.

Chemical dynamics simulations of SID, SL, RL, and CID are reviewed in the following. An important component of the simulations is to compare with experiment to confirm the accuracy of the simulations and assist in interpreting experimental spectra. The simulations provide atomistic detail concerning the dynamics of SID, SL, RL, and CID processes, which is difficult to obtain from typical experiments.

II. Collisions of Biological Ions with Organic Surfaces

In recent work,^{72,73} a review and perspective were published which address chemical dynamics simulations of collisions of biological ions with surfaces. A motivating factor for these simulations was the finding that chemical dynamics simulations⁷⁴⁻⁷⁷ for $SiMe_3^+$ and $Cr^+(CO)_6$ give

collisional energy transfer probabilities in agreement with experiment.⁷⁸⁻⁸⁰ Here, theoretical methods for the chemical dynamics simulations, the simulation results, and the impact of the simulations on experiments are discussed. In the following, four aspects of the dynamics associated with collision of biological ions with organic surfaces are discussed. The first is the efficiency of energy transfer from the collision's translational energy to the ion's internal degrees of freedom and to the surface, upon collision with the surface. These are important for modeling SID^{12,13} of the ion. The second addresses fundamental dynamics for SID fragmentation. For the third, soft-landing (SL)^{14,15} of the ion is considered, where the ion physisorbs intact on the surface without fragmenting or reacting. The fourth is reactive-landing (RL),¹⁶ collisions in which the ion reacts with the surface and chemisorbs. SL and RL are processes for preparing materials with a broad range of technological applications. However, before these four features of biological ion + organic surface collisions are discussed, the methodology for the chemical dynamics simulations is first outlined.

A. Simulation methodology

Chemical dynamics simulations involve calculating ensembles of classical trajectories to represent experiments. Two important components of the simulations are the potential energy function and the method for calculating the classical trajectories. Each is described below.

A.1. Models for potential energy function

The potential energy function for the ion/surface is represented as

$$V_{total} = V_{ion} + V_{surface} + V_{ion-surface} \quad (1)$$

where V_{ion} is the biological ion's intramolecular potential, $V_{surface}$ is the surface's potential, and $V_{ion-surface}$ is the intermolecular potential between the ion and surface. Two models have been used for V_{ion} . The first model uses the AMBER molecular mechanics (MM) force field,⁸¹ which allows the ion to vibrate and absorb energy, but to not fragment. The second model obtains the ion's potential directly from a quantum mechanical (QM) electronic structure theory such as AM1 or MP2,^{82,83} resulting in a simulation referred to as QM+MM direct dynamics.⁸⁴ With this model, fragmentation of the ion may be studied.

Accurate MM potentials were used for $V_{surface}$. The MM potential developed by Mar and Klein⁸⁵ for isolated alkyl thiolate self-assembled monolayer (H-SAM) surfaces was modified⁷⁶ to

model hyper-thermal collisions with peptide-H⁺ ions. The potential used for diamond is a MM force field fit to the diamond phonon spectrum.⁸⁶ For perfluorinated alkyl thiolate self-assembled monolayer (F-SAM) surfaces, a MM potential was developed from *ab initio* electronic structure calculations for perfluoroalkanes.⁸⁷

To determine accurate energy transfer probabilities from the simulations to compare with experiments, high quality intermolecular potentials between biological ions and surfaces are needed.⁸⁸ MM potentials such as AMBER,⁸¹ CHARMM,⁸⁹ and OPLS⁹⁰ “roughly” approximate the long-range, attractive two-body potentials between the atoms of the ions and surfaces, but are grossly in error for the short-range repulsions.^{91,92} These latter terms are important for the transfer of the ion’s collision energy to the ion and surface internal degrees of freedom.^{88,93,94}

Biological ion + surface collisions are at high energy and collisional energy transfer to the ion and surface internal degrees of freedom is determined by the ion + surface repulsive potential. For collisions of peptide-H⁺ ions interacting with the H-SAM and diamond {111} hydrocarbon surfaces, these potentials were derived from *ab initio* calculations.⁹⁵ The peptide-H⁺/surface intermolecular potential is modeled by a sum of two-body potentials between the atoms of the ion and surface. The two-body potential is given by the Buckingham repulsion and an additional repulsive term; i.e.

$$V_{XY} = A_{XY} \exp(-B_{XY}r_{ij}) + C_{XY}/r_{ij}^n \quad (2)$$

where X corresponds to C or H atoms of the diamond or H-SAM surface, Y corresponds to H, C, O, or N atoms of peptide-H⁺, and r_{ij} is the distance between the atoms. To determine parameters for the above two-body potentials, *ab initio* potential energy curves were calculated between CH₄, as a model for the C and H atoms of the diamond {111} and H-SAM surfaces, and CH₄, NH₃, NH₄⁺, H₂CO, and H₂O as models for the different types of atoms and functional groups comprising protonated polyglycine and polyalanine peptide ions. The *ab initio* calculations were carried out at the MP2 level of theory with the frozen-core approximation and 6-311+G(2df,2pd) basis set. The molecules were held fixed in their optimized geometries and intermolecular potential energy curves for different orientations of the CH₄/CH₄, CH₄/NH₃, CH₄/NH₄⁺, CH₄/H₂CO, and CH₄/H₂O systems were calculated. The curves for the different orientations of a particular system were then

fit simultaneously by a sum of the two-body functions in Eq. 2, using nonlinear least squares, to obtain the potential energy parameters.

The above approach was used to obtain potential energy curves for polyglycine and polyalanine peptide ions colliding with a F-SAM surface.^{92,96} CF₄ was used as a model for the surface and CH₄, NH₃, NH₄⁺, and HCOOH molecules were used to model the atoms and functional groups of the peptide-H⁺ ions. In the initial fitting,⁹⁶ repulsive potential energy functions were determined as in Eq. (2). Later,⁹² attractive terms were included by calculating potential energy curves at the MP2/aug-cc-pVTZ level of theory and fitting these curves by the atom-surface two-body potential

$$V_{XY} = A_{XY} \exp(-B_{XY}r_{ij}) + C_{XY}/r_{ij}^n + D_{XY}/r_{ij}^m \quad (3)$$

where X corresponds to C or F atoms of F-SAM surface, Y corresponds to H, C, O, or N atoms of the peptide, and r_{ij} is the distance between the atoms. The fitting was accomplished with a combined genetic/nonlinear least squares algorithm, with $C_{XY} \leq 0$, for an attractive term, $D_{XY} \geq 0$, for an additional repulsive term, and $m - n \geq 3$. The formamide molecule, HCONH₂, was not included in deriving this peptide-H⁺/surface intermolecular potential and it was tested by using it to calculate potential energy curves for CF₄ interacting with the -NH₂ end of formamide and comparing these curves with those calculated with MP2/aug-cc-pVTZ theory. As shown in Figure 1, the curves are in quite good agreement for different orientations of CF₄ interacting with the -NH₂ group, illustrating the transferability of the derived potential energy parameters.

The potential energy function between a biological ion and surface is substantially more complex if ion + surface reactions are treated in the simulations. A QM/MM model⁸⁴ was used for these simulations. A QM electronic structure theory is used for the ion, as described above, as well as part of the surface, so that reactions between the two are possible. This entails selecting a region of the surface large enough to represent the ion/surface collision and sufficiently deep to include all surface atoms participating in the ion/surface reaction. The MM model is used to include interactions between the ion and other atoms of the surface, and there is a QM/MM interface between the QM and MM surface atoms. The QM surface region may be modified to represent reactive functional groups on the top of the surface.

A.2. Method for the chemical dynamics simulations

There are two principal components for performing the chemical dynamics simulations; one is choosing proper initial conditions for the trajectories to represent experiment and the second is the ensuing numerical integration of the classical equations of motion for the trajectories. A sufficiently large ensemble of trajectories, with random initial conditions, are calculated to obtain statistically meaningful results to compare with experiment.

The procedure for choosing initial conditions for the trajectories has been described in detail.⁹⁵ A classical molecular dynamics (MD) simulation is performed for the surface so that its vibrational energy is in accord with the surface temperature T_s , usually 300 K. The initial internal energy for the biological ion may be for a specific vibration/rotation state or sampled from a Boltzmann distribution of these states at temperature T_{ion} , usually 300 K, both selected by quasiclassical sampling.⁹⁷ The ion is randomly rotated about its Euler angles, and the incident collision energy E_i and angle θ_i are set. The center-of-mass of the ion is then randomly aimed within the central unit-cell of the surface and numerical integration of the trajectory commences.

B. Peptide-H⁺ + surface energy transfer efficiencies

Upon collision with the surface, there is transfer of some of the translational energy E_i of the ion to the surface vibrations E_{surf} and the ion's internal degrees of freedom E_{int} ;

$$E_i = E_f + \Delta E_{surf} + \Delta E_{int} \quad (4)$$

where E_f is the ion's final translational energy after collision with the surface. Numerous experiments have been performed to study energy transfer to internal degrees of freedom of biological ions in their collisions with surfaces.⁹⁸⁻¹⁰² The simulations also provide the transfer to E_{surf} and the amount remaining in E_f . Percentages of E_i transferred to ΔE_{int} , ΔE_{surf} , E_f are summarized in Table 1 for simulations that have been performed for peptide-H⁺ + surface collisions.¹⁰³⁻¹¹² Uncertainties of the percentages range from 1-2%. In the following, these energy transfer efficiencies are considered as a function of E_i and θ_i , the surface, peptide-H⁺ and its size, the V_{ion} intramolecular potential, and dynamics of the ion-surface collision.

B.1. Collision energy

Laskin and Futrell^{100,101} experimentally studied energy transfer in ala₂-H⁺ + F-SAM collisions for $E_i = 4.5 - 22.5$ eV and in des-Arg¹-bradykinin collisions with H-SAM, F-SAM, and diamond {111} surfaces for $E_i = 15 - 100$ eV. They found that the percentage energy transfer to

ΔE_{int} was independent of E_i . The percentages are 21% for $\text{ala}_2\text{-H}^+$ + F-SAM and 10.1, 20.5, and 19.2% for des-Arg¹-bradykinin colliding with the H-SAM, F-SAM, and diamond surfaces, respectively. The incident angle was normal to the surface for these experiments; i.e. $\theta_i = 0^\circ$. As shown in Table 1, the simulations also give a percentage energy transfer to ΔE_{int} independent of E_i . For the $\theta_i = 0^\circ$ simulations, the average percentage energy transfer to ΔE_{int} is 12% for $\text{gly}_8\text{-H}^+$ + H-SAM, 19% for $\text{ala}_2\text{-H}^+$ + F-SAM, and 21% for $\text{gly}_2\text{-H}^+$ + diamond. These percentages agree with those measured by Laskin and Futrell.¹⁰⁰⁻¹⁰²

For the $\theta_i = 45^\circ$ simulations, the average percentage transfer to ΔE_{int} is 12% for $\text{gly}_8\text{-H}^+$ + H-SAM, 23% for $\text{ala}_2\text{-H}^+$ + F-SAM, 15% for $\text{gly}_2\text{-H}^+$ + diamond, 18% for $\text{ala}_2\text{-H}^+$ + diamond, and 16% for $\text{gly}_3\text{-H}^+$ + diamond (the $E_i = 10$ eV datum is not included for this last percentage).

For each of the systems studied in Table 1, the percentage remaining in translational energy decreases and the percentage transferred to the surface increases with increasing collision energy E_i . This is consistent with the time-scales for peptide- H^+ translation and surface vibration becoming more commensurate with increases in E_i . From another perspective, the surface appears “softer” with increasing E_i and the collision becomes more inelastic.¹⁰⁵

B.2. V_{ion} intramolecular potential

Energy transfer simulations were performed with three different models for the peptide- H^+ intramolecular potential V_{ion} ; i.e. AMBER MM force field, and AM1 semi-empirical and MP2 *ab initio* electronic structure theories.^{103,104,106,107} As shown in Table 2, within statistical uncertainties, the same average energy transfer probabilities are found with AMBER, AM1, and MP2 for gly-H^+ + diamond collisions and AMBER and AM1 for $\text{gly}_2\text{-H}^+$ + diamond collisions. The energy transfer distributions are also in agreement for the different V_{ion} , as illustrated in Figure 2 for $\text{gly}_2\text{-H}^+$ + diamond collisions using AMBER and AM1. This agreement suggests the collisional energy transfer is only influenced by the peptide ion’s structure and forces about the ion’s potential energy minimum. Details of the peptide ion’s intramolecular potential are apparently unimportant for energy transfer.

B.3. Incident angle

As shown in Table 1 for the $\text{gly}_8\text{-H}^+$ + H-SAM and $\text{ala}_2\text{-H}^+$ + F-SAM collisions, energy transfer to peptide- H^+ is the same for θ_i of 0° and 45° , showing that for the SAM surfaces energy transfer to ΔE_{int} is not dependent on θ_i . The insensitivity in the percentage transfer to ΔE_{int} with θ_i may result from the high corrugation and substantial roughness of the SAM surfaces. In contrast

to this insensitivity for transfer to ΔE_{int} , the transfers to ΔE_{surf} and E_f are affected by θ_i , with transfer to the former decreasing and the latter increasing as θ_i is increased from 0° to 45° .

In contrast to the result for the SAM surfaces, for the diamond {111}, surface energy transfer to ΔE_{int} depends on θ_i as shown in Table 1. For diamond, there is a decrease in the transfer to ΔE_{int} upon changing θ_i from 0 to 45° ; that is, a factor of 0.66 decrease for gly-H⁺ collisions at 70 eV with MP2,¹⁰⁷ 0.63 decrease for gly₂-H⁺ at 70 eV with AM1 and AMBER,^{104,105} and 0.58 decrease for gly₈-H⁺ collisions at 100 eV with AM1.¹⁰⁹ This decrease in energy transfer is approximately the same as that expected for $\cos^2\theta_i$ scaling, which equals 0.5. However, for energy transfer to the surface and remaining in translation, the dependence on θ_i is the same as that above for the SAM surfaces.

The simulations for collisions of gly₈-H⁺ with the diamond surface at $E_i = 100$ eV and θ_i of 0° and 45° , in Table 1, may be compared with experiments for des-Arg¹-bradykinin colliding with the diamond surface.¹⁰¹ For gly₈-H⁺ the average percentage transfer to ΔE_{int} is 45% for $\theta_i = 0^\circ$ and 26% for $\theta_i = 45^\circ$. In contrast, in experiments for des-Arg¹-bradykinin + diamond, with $\theta_i = 0^\circ$, the transfer to ΔE_{int} is 19.2%.¹⁰¹ Such different values for energy transfer to ΔE_{int} for gly₈-H⁺ and des-Arg¹-bradykinin, both octapeptides, is unexpected. This difference may arise from the perfectly flat diamond {111} surface used for the simulations as compared with the experiment's "rough" diamond surface.^{101,112} The experimental surface is grown by "merging" different nucleation sites and is inherently rough. In addition, some of the carbon on the surface is graphitic and its fraction uncertain.¹¹³ Furthermore, the experimental diamond surface may have a termination that is different than the H-atom terminated surface used for the simulation. Thus, there are significant differences between the simulation and experimental surfaces. For normal collisions investigated experimentally, a perfectly flat diamond {111} surface may transfer substantially more energy to peptide-H⁺ ΔE_{int} than does a rough and irregular partially graphitic surface. However, a counter finding, from experiments,¹¹² is that three diamond surfaces of different roughness give the same energy transfer efficiency. As discussed above, energy transfer to ΔE_{int} is not strongly affected by θ_i for collisions of peptide-H⁺ ions with SAM surfaces as a result of their corrugation and roughness.

The simulations for gly₈-H⁺ + diamond with $\theta_i = 45^\circ$ may be representative of collisions with a rough "diamond-like" surface. For this incident angle, the percentage energy-transfer to ΔE_{int} is 26% compared to the experimental value of 19.2%. In addition, the experimental and simulation

distributions of ΔE_{int} energy transfer are qualitatively consistent. Though the percentage of energy-transfer to ΔE_{int} , for des-Arg¹-bradykinin is unaffected by varying the roughness of the “diamond-like” surface,¹¹² the smoothest experimental surface still has a 32 Å roughness.

B.4. Peptide-H⁺ size

As shown in Table 1, for peptide-H⁺ + H-SAM collisions, the average percentage transfer to ΔE_{int} is weakly dependent on the peptide-H⁺ size. For $\theta_i = 45^\circ$ collisions, the percentage transfer to ΔE_{int} is 7 to 8% for gly₃-H⁺ and 12% for gly₈-H⁺. This result is consistent with experiments with the F-SAM, where the percentage energy transfer to ΔE_{int} for ala₂-H⁺ and protonated bradykinin collisions are the same within statistical uncertainties.¹⁰⁰⁻¹⁰²

For the simulations of gly_n-H⁺ collisions with diamond, the energy transfer efficiency is also not strongly dependent on peptide size. As given above, the average energy transfer percentage to ΔE_{int} is 15% for gly₂-H⁺, 18% for ala₂-H⁺, and 16% for gly₃-H⁺. From Table 1, diamond and $\theta_i = 45^\circ$ simulations were performed at single E_i values for gly-H⁺, gly₅-H⁺, and gly₈-H⁺, and their energy transfer percentages are 11, 23, and 26%, respectively. The percentage increases from 15% for gly₂-H⁺ to 26% for gly₈-H⁺. A statistical model which assumes the energy transfer percentage is proportional to the number of vibrational modes of the peptide ion would predict a much larger increase. As discussed below, analyses of the energy transfer dynamics suggest torsional modes of peptide-H⁺ are efficiently excited by peptide-H⁺ + surface collisions.

B.5. Surface properties

As summarized in Table 1, the nature of the surface is very important for the energy transfer efficiencies. A comparison may be made between collisions of gly₈-H⁺ with H-SAM and diamond surfaces at 100 eV. For the H-SAM, the percentage transfers to ΔE_{int} , ΔE_{surf} , and E_f are nearly independent of θ_i , with values of 13, 77, and 10% for $\theta_i = 0^\circ$, respectively. In contrast, for diamond these percentages are strongly dependent on θ_i . For $\theta_i = 0^\circ$ they are 45, 26, and 29%, with ΔE_{int} and ΔE_{surf} receiving much more and much less energy, respectively. Similarly, a comparison may be made between ala₂-H⁺ collisions with the F-SAM and diamond surfaces. For the F-SAM surface, energy transfer to ΔE_{int} is not strongly dependent on E_i and θ_i . For $E_i = 22.5$ eV and $\theta_i = 45^\circ$, the percentage transfers to ΔE_{int} , ΔE_{surf} , and E_f are 23, 61, and 16%. In contrast, for the diamond surface with $E_i = 30$ eV and $\theta_i = 45^\circ$, these percentages are 18, 12, and 70%, with energy transfer to ΔE_{int} similar for the two surfaces, while the F-SAM receives much more energy than diamond. To summarize these comparisons, energy transfer to ΔE_{int} is similar for collisions with the F-SAM

and diamond surfaces, with less energy transfer to ΔE_{int} for collision with the H-SAM; findings consistent with experiment. The simulations show that the H-SAM and F-SAM surfaces receive more energy than does diamond, though this has not been determined experimentally.

B.6. Energy-transfer pathways and collision lifetime

Peptide ions have a hierarchy of vibrational modes ranging from high-frequency stretches to low-frequency torsions. To assist in understanding the peptide- H^+ + surface energy transfer dynamics, it is useful to know which peptide modes are most efficiently excited by collisional activation. In a gas-phase simulation of exciting unfolded gly₄- H^+ by collisions with Ar atoms at $E_i = 4.3$ eV, the role of different types of modes on the energy transfer was investigated by increasing individual mode frequencies to the high-frequency limit, so that the modes are constrained and do not accept energy.¹¹⁴ With none of the vibrations constrained, 58% of E_i is adsorbed by the peptide's internal degrees of freedom. When all of the modes are constrained, except the torsions, this is lowered to 49%, which indicates the torsions absorb ~84% of the internal energy transferred to the peptide.

As shown in Table 1, with none of the folded gly₃- H^+ modes constrained, 18, 9, and 73% of $E_i = 30$ eV is transferred to E_{int} , E_{surf} , and E_f , respectively, for collision with the diamond surface. With all of the modes of gly₃- H^+ , except the torsions, constrained as described above, these respective values become 14, 16, and 70%.⁸⁶ Thus, as found in the simulation of gas-phase Ar + gly₄ energy transfer, ~80% of the internal energy transferred to the peptide goes to torsions for gly₃- H^+ + diamond collisions.

In MM simulations of folded gly₃- H^+ colliding with the diamond {111} surface,⁹⁵ average lifetimes for the collisions were determined by calculating the average change in the gly₃- H^+ internal energy versus time for the simulations' ensembles of trajectories. Results of these analyses are shown in Figure 3 for the different E_i . The average collision lifetime decreases from ~ 1000 fs to ~ 220 fs, a factor of ~ 4.5, as the collision energy is increased from 10 to 110 eV. A simple model would assume that the gas-surface collision lifetime to be proportional to the collision velocity, i.e. proportional to $(E_i)^{1/2}$, and yielding a factor of 3.3. The simulation's decrease in the average collision lifetime is somewhat greater than this model's prediction.

B.7. Collision dynamics and energy transfer models

Model studies have been performed to investigate properties of peptide- H^+ + surface energy transfer dynamics.^{106,115} Effects of the peptide- H^+ vibrational temperature T_{vib} and surface

temperature T_{surf} on the energy transfer efficiency were investigated in gly₂-H⁺ + diamond {111} simulations by varying both T_{vib} and T_{surf} between 300 and 2000 K.¹⁰⁶ To a good approximation, the changes in T_{vib} and T_{surf} had a negligible effect on the transfer of E_i to the surface and gly₂-H⁺ internal degrees of freedom.

Effects of peptide orientation and surface impact site on the efficiencies of energy transfer were studied in gly₂-H⁺ + diamond {111} simulations.¹⁰⁶ The diamond surface has both H- and C-sites, and energy transfer to ΔE_{int} , ΔE_{surf} , and E_f was statistically the same for impact with the two sites. In contrast, for $\theta_i = 0^\circ$ collisions, the orientation of gly₂-H⁺ has a significant effect on energy transfer. Initially, the peptide ion is not rotationally excited and the ion's orientation for the collision was determined by defining a vector from the N-atom of the protonated amino group to the hydroxyl O-atom of the carboxylic group. The angle between this vector and the vector normal to the surface determined the initial orientation angle Ψ_i of the peptide ion. When gly₂-H⁺ collides with its backbone vertical to the surface plane with $\Psi_i = 90^\circ$ and, thus, a C- or N-terminus approach, the internal energy change is a maximum. When the ion collides horizontally, the opposite occurs. In addition, for vertical collisions more energy is transferred to the peptide ion if the C-terminus first strikes the surface instead of the N-terminus. For non-perpendicular collisions, with $\theta_i = 45^\circ$, energy transfer efficiency is less sensitive to the peptide orientation. The ion orientation becomes more important as E_i is increased. Energy transfer to ΔE_{int} versus the orientation angle Ψ_i is well fit by the quadratic expression $\Delta E_{int} = a(\Psi_i - \Psi_o)^2 + b$, where $\Psi_o = \sim 90^\circ$. For highly folded, globular peptides, there may not be a strong orientation effect for energy transfer.

As shown by the energy transfer percentages in Table 1, a good model for peptide-H⁺ + surface collisions versus E_i is that energy transfer to ΔE_{int} is independent of E_i , while transfers to ΔE_{surf} and E_f are E_i dependent. In analyses for gly₂-H⁺ and ala₂-H⁺ + F-SAM⁹⁹ and gly₈-H⁺ + H-SAM¹⁰¹ collisions, the percentage energy transfer to the surface was fit by a model based on the adiabaticity parameter for $T \rightarrow V$ energy transfer.¹¹⁶ It was found that a more physically realistic fit was obtained if the adiabaticity parameter was related to the collision energy rather than the velocity. The model for the percentage energy transfer to the surface is then given by

$$P_{surf}(E_i) = \langle \Delta E_{surf} \rangle / E_i = P_o \exp(-b/E_i) \quad (5)$$

where P_o (unit less) and b (in eV) are fitting parameters. In more complete analyses¹¹⁰ of the previous simulations¹⁰⁸ of $\text{gly}_2\text{-H}^+$ collisions with the F-SAM, P_o and b equal 0.85 and 11.83 eV at 0° and 0.72 and 6.73 eV at 45° . For the $\text{gly}_8\text{-H}^+$ + H-SAM simulations, P_o and b are 0.78 and 1.87 eV for 0° and 0.74 and 4.23 eV for 45° .

C. Fragmentation mechanisms

QM+MM simulations identify two limiting mechanisms for SID fragmentation of peptide- H^+ ions.⁹⁴ For one, peptide- H^+ “shatters” as it collides with the surface, forming multiple fragments which scatter off the surface and may undergo secondary dissociations. This process is termed shattering and is practically defined within simulations as a fragmentation event that occurs faster than the vibrational period of the broken bond (this value depends on the bond). For the second mechanism, the molecule is vibrationally excited by its collision with the surface and then fragments after bouncing off the surface. If intramolecular vibrational energy redistribution (IVR)¹¹⁷ is complete for peptide- H^+ before it dissociates, its unimolecular kinetics will be in accord with Rice-Ramsperger-Kassel-Marcus (RRKM) theory.¹¹⁸ Between the limits of shattering and RRKM fragmentation, a range of different unimolecular dynamics is expected depending on the extent of IVR.

Shattering was observed in the $\text{gly}_n\text{-H}^+$ + diamond {111} QM+MM simulations and the results are summarized in Table 3. The percentage of fragmentation which is shattering increases with increasing collision energy, illustrated by the $\text{gly}_2\text{-H}^+$ simulations with AM1. The shattering percentage is only 8% at 30 eV, but 70% at 100 eV. Shattering occurs via multiple fragmentation channels, with as many as 304 unique products for the $\text{gly}_8\text{-H}^+$ simulations at $E_i = 100$ eV and $\theta_i = 0^\circ$. As shown in Table 3, the shattering dynamics for gly-H^+ at $E_i = 100$ eV and $\theta_i = 45^\circ$ with the AM1 and MP2 QM models are nearly identical. This is an important finding which highlights that qualitatively similar results are obtained using the much less computationally expensive semi empirical methods.

Shattering was observed in Fourier transform ion cyclotron resonance mass spectrometry (FT-ICR MS) experiments of des-Arg1- and des-Arg⁹-bradykinin SID on a F-SAM surface.^{119,120} At low collision energies of 15-25 eV both ions fragment via a small number of pathways, but increasing the collision energy above 30 eV resulted in many more dissociation pathways. The same type of fragmentation dynamics was found in the simulations. Experimental time-resolved fragmentation of the ions indicated both slow and fast fragmentation, with the latter being time-

independent and the former accurately modeled by RRKM theory.¹¹⁸ The kinetics of shattering fragmentation were described using a “sudden death” approximation. Sudden death is a simplified kinetic model that assumes the fragmentation rate constant is a step function of the internal energy.

Fragmentation mechanisms and energetics were analyzed for the QM+MM simulations of gly_n-H⁺ ions colliding with diamond.^{103,104,107,109} For gly-H⁺ simulations at $E_i = 70$ eV and $\theta_i = 45^\circ$, AM1 gave three important pathways forming NH₂CH₂⁺ + C(OH)₂, NH₃ + CH₂COOH⁺, and NHCH₂ + H₂ + COOH⁺.¹⁰³ The first is a low energy pathway, expected from energetic considerations, while the latter is a high energy pathway, occurring predominantly by shattering. For the MP2 simulations at this E_i and θ_i ,¹⁰⁷ there were two important pathways NH₂CH₂⁺ + C(OH)₂ and NH₃ + CH₂COOH⁺. The MP2 simulations were also performed for $\theta_i = 0^\circ$ and there were the following five important pathways listed in descending importance, with the percentage shattering in parentheses; i.e. NH₃ + CH₂COOH⁺ (84), NH₂CH₂⁺ + C(OH)₂ (41), NH₃ + CO + CH₂OH⁺ (88), H₂ + NH₂CHCOOH⁺ (100), and NH₂CH₂⁺ + CO + H₂O (53). The first two were also the important pathways at $\theta_i = 45^\circ$. The 5th is the lowest energy pathway and expected to be the dominant pathway on energetic grounds.¹⁰³ Only for pathways 2 and 5 is the vast majority of the fragmentation not shattering. From energy resolved CID experiments for gly-H⁺ at low collision energies by Klassen and Kebarle,¹²¹ it was concluded that NH₂CH₂⁺ + CO + H₂O was the most likely dissociation pathway. This pathway was not observed in the AM1 simulation and, though it was observed in the MP2 simulation, it ranked 5th amongst the most important pathways. For later energy resolved CID experiments, Armentrout et al.¹²² found that the lowest energy pathway was formation of NH₂CH₂⁺(H₂O) + CO followed by formation of NH₂CH₂⁺ + CO + H₂O. These experiments extended to collision energies of 5.0 eV.

The differences between the gly-H⁺ + diamond SID simulations and the gly-H⁺ CID experiments are not surprising. The CID experiments focused on low energy collisions and determining the fragmentation threshold, which is less than 2.0 eV. The low energy CID may be fit by RRKM theory.¹²² In contrast, the SID simulations are at 70 eV and the fragmentation is dominated by shattering. It is possible that none of the fragmentation is statistical and in accord with RRKM theory.

For the gly-H⁺ and gly₂-H⁺ + diamond simulations,^{103,104,107} trajectories were analyzed to identify which backbone bond rupture initiated the fragmentation. The specific a- and x-type ions formed by the gly-H⁺ and gly₂-H⁺ fragmentations, and the following gly₈-H⁺ fragmentation, are

given in the research articles and not listed here. For the AM1 gly-H⁺ simulations,¹⁰³ the predominant initial backbone rupture was the ⁺H₃NCH₂---COOH bond, with a small amount of ⁺H₃N---CH₂ bond rupture forming NH₃ + CH₂COOH⁺. For the MP2 gly-H⁺ simulations,¹⁰⁷ the former initial bond rupture remains dominant, but ⁺H₃N---CH₂ bond rupture is more important than for the AM1 simulations.

At $E_i = 30$ eV, for the gly₂-H⁺ + diamond simulations,¹⁰⁴ ⁺H₃NCH₂---CONHCH₂COOH bond rupture is the only important initial bond dissociation pathway. At higher E_i , this remains the most important initial backbone bond cleavage, but the initial cleavage now occurs at all of the backbone bonds except ⁺H₃NCH₂CONH---CH₂COOH. From the simulations, at $E_i \leq 70$ eV, NH₂CH₂⁺ and its isomer NH₃CH⁺ are the principal fragment ions, consistent with collision energy resolved gly₂-H⁺ CID experiments.¹²¹ However, from the experiments it was proposed that the initial backbone cleavage for formation of these ions is the peptide bond, while the dominant initial cleavage from the simulations is the ⁺H₃NCH₂---CONHCH₂COOH bond.

Initial bond rupture dissociation sites were not determined for the gly₈-H⁺ simulations, but backbone cleavage patterns were determined.¹⁰⁹ Most of the fragmentations are shattering, with most occurring by two or three backbone cleavages. An important component of these simulations was analyses of product fragment rearrangements, a problem for peptide sequencing.¹²³⁻¹²⁵ Two rearrangement mechanisms were observed. One involves an initial association of the termini of a fragment, forming a cyclic structure. This intermediate then dissociates via a different backbone cleavage site, altering the backbone sequence. For the second, fragments recombine while they are near the surface.

Shattering was observed in QM/MM simulations of gly₂-H⁺ colliding and reacting with a F-SAM surface, with reactive -C(O)Cl and -C(H)O head groups.¹²⁶ However, the shattering percentage was not determined, given the difficulty in deconvoluting shattering and reactive trajectory events. There was no shattering in a simulation of gly₈-H⁺ collisions with a “soft” H-SAM surface.¹¹⁰ However, there was non-shattering fragmentation promoted by proton transfer.¹²⁷ The most important fragmentation pathways were cleavages of CH₂---CO and peptide bonds.

The mobile proton model is based on the concept that collisional excitation populates a thermodynamically less favorable conformation inducing proton transfer to less basic sites. As a consequence, the covalent bond adjacent to the final site is weakened and often breaks. Studying proton motion was the focus of Barnes’ and co-workers’¹²⁸ simulation of gly₈-H⁺+ F-SAM SID.

By carefully tracking motions of protons within the peptide and correlating proton motion to subsequent bond cleavage events, information was gained regarding the number of proton transfers required prior to fragmentation as well as the most probable sites for proton migration. Proton migration was considered relevant to a backbone cleavage event if the proton moved to or from either of the heavy atoms involved in the bond or a heavy atom connected to those involved. It was found that at collision energies of both 30 and 110 eV, nearly 50% of the backbone cleavage events occurred after a single proton hop. It was also observed that there was a striking even-odd nature regarding which hops were efficient. This feature was due to competition between two or more protonation sites. This may be illustrated through a simple example of two states, denoted A and B, which could be competing for a proton (the generalization to more sites is straightforward). Simulations showed that the first proton transfer frequently resulted in fragmentation, i.e. $A \rightarrow \mathbf{B}$ where the bold B denotes that fragmentation occurred from that protonation state. However, if insufficient energy was present in the relevant covalent bond, protonation state A could recover the proton, and if it was the initial, stable protonation site, the peptide ion was unlikely to fragment. Hence, the sequence $A \rightarrow B \rightarrow A \rightarrow \mathbf{B}$ leading to fragmentation becomes likely and justifies the even-odd fragmentation efficiency observed.

It is also noteworthy that qualitatively similar hop efficiencies are seen at 30 and 110 eV, suggesting the fragmentation mechanism is collision energy independent. The lag time between proton hop and fragmentation was also modeled using a simple kinetic scheme. It was found that 90% of the trajectories fragment within 1.5 ps of a relevant proton hop at 110 eV. To reach the same fragmentation fraction, following a relevant proton hop at 30 eV, 2.5 ps is required. This illustrates that as long as sufficient energy is transferred, the mechanism is collision energy independent but the reaction rate is dependent of the available energy.

In related work,¹²⁹ dynamics that take place following initial fragmentation events was examined. It was seen that fragmentation products were able to form complexes rather than simply drifting apart, a finding similar to that discussed above for $\text{gly}_8\text{-H}^+$ fragmentation.¹⁰⁹ These complexes typically were stabilized via hydrogen bonding between the charged and neutral fragment species. The presence of these complexes opened the possibility for secondary reactions leading to products that are only possible through complex formation. These non-covalent complexes have long lifetimes and specifying an average lifetime in units of time is misleading. Simulations necessarily examine a relatively short time window and the initial fragmentation event

may occur either early or late within the simulation. Hence, a lifetime fraction was defined relative to the total time the complex exists within the simulation. It was found that the average lifetime fraction was 83% at 30 eV and 55% at 110 eV. Some secondary fragmentation products resulted from combination of traditional intramolecular mobile proton motion along with intermolecular proton transfer between charged and neutral fragments. Formation of such complexes may play an important role in the overall reaction mechanisms of SID and CID.

D. Soft landing

In simulations of $\text{ala}_2\text{-H}^+$ colliding with a FSAM consisting of $\text{CF}_3\text{-(CF}_2)_7\text{-S-}$ chains, SL was studied.^{111,130} The $V_{ion\text{-}surface}$ potential used for the simulation included both accurate long-range attractive and short-range repulsive terms. In a separate molecular dynamics (MD) simulation of $\text{ala}_2\text{-H}^+$ desorbing from this surface,¹³¹ the $\text{ala}_2\text{-H}^+$ /F-SAM binding energy was found to be ~ 13 kcal/mol. When $\text{ala}_2\text{-H}^+$ is thermally equilibrated on the surface, its shape and size changes, becoming less compact by cleavage of a hydrogen bond between the -OH and -NH-groups. Of particular interest with respect to SL, is the percentage of trajectories which remained trapped on/in the F-SAM when the $\text{ala}_2\text{-H}^+$ + FSAM trajectories were terminated at 10 ps.¹¹¹ The percentage of trajectories which remain trapped decreases with increasing E_i . At 70 eV there are no trapped trajectories. For $\theta_i = 0^\circ$ collisions, 84, 65, 41, and 29% of the trajectories remained trapped at E_i of 5, 13.5, 22.5, and 30 eV, respectively. The percentage of the trajectories which are trapped decreases upon changing θ_i from 0° to 45° . This decrease is small at $E_i = 5$ eV, 84 to 69%, but large at $E_i = 22.5$ eV, 41 to 6%.

Mechanisms for trapping (SL) of $\text{ala}_2\text{-H}^+$ on the F-SAM include physisorption on top and penetration of the F-SAM.¹³⁰ Five different mechanisms were identified for $E_i = 5$ and 13.5 eV, and $\theta_i = 0^\circ$, for which trapping is important: (i) first physisorption, followed by penetration, and then physisorption when the trajectory is terminated (i.e., phys-pen-phys); the remaining four mechanisms are (ii) pen-phys-pen; (iii) pen-phys; (iv) phys-pen; and (v) only phys. For the 84% of the trajectories trapped for the 5 eV simulations, 40% first penetrate and 44% first physisorb. The pen-phys-pen, pen-phys, phys-pen, and phys mechanisms have similar probabilities. At 13.5 eV, 65% of the trajectories are trapped, with 47% first penetration and only 18% first physisorption. The pen-phys mechanism dominates and the phys-pen mechanism, important at 5 eV, is unimportant. A significant fraction of the 11 heavy atoms of $\text{ala}_2\text{-H}^+$ penetrate the F-SAM, when $\text{ala}_2\text{-H}^+$ is trapped. Most of the penetration occurred near the top of the $\text{CF}_3\text{-(CF}_2)_7\text{-S-}$ chains.

For the 30 eV simulation, penetration is only as deep as the second layer. For the $E_i = 70$ eV simulation, there is deep penetration to the 5th layer, but as discussed above there is no trapping.

Barnes and co-workers¹³² conducted simulations of Ac-ala₇-lys-H⁺ and Ac-lys-ala₇-H⁺ soft landing on an F-SAM surface using the same interaction potential as Hase and co-workers.⁹² While Ac-ala₇-lys and Ac-lys-ala₇ have similar composition, their secondary structures are dramatically different. In particular, Ac-ala₇-lys is the shortest stable alpha-helical polyalanine chain, whereas Ac-lys-ala₇ has a random coil or globular structure. The close similarity in sequence allows for an analysis of the effect of secondary structure on the SL efficiency. Trajectories were classified in a similar way as for the ala₂-H⁺ simulations,^{111,130} however, an additional “intermediate” class was included. Intermediate trajectories may eventually soft land or escape the surface, but their ultimate fate cannot be determined within the time frame of the simulation. The population within this class was minimal.

It was found that the soft-landing efficiency was nearly identical between Ac-ala₇-lys-H⁺ and Ac-lys-ala₇-H⁺, meaning secondary structure had little effect. A fast conformational change upon interaction with the surface was observed for both species. This change was quantified by defining a time dependent conformational entropy via an approach inspired by the work of Baxa et al.¹³³ The time dependent probability distribution for the Ramachandran angles of each peptide was calculated, which allowed for the entropy to be determined according to the statistical definition, namely $S(t) = -k_B \sum_i P_i(t) \ln(P_i(t))$, where $P_i(t)$ is the time dependent probability of having a Ramachandran angle within in the i^{th} bin (see Ref. 132 for greater detail). Although both secondary structures underwent fast conformational changes, the alpha-helical peptide produced more entropy throughout the course of the simulations. At the end of the simulations, both species approached a similar “globular” structure.

No significant penetration into the surface was observed at the collision energies considered, and hence the mechanism for soft landing was through physisorption to the surface. In addition, a “tethering” mechanism was observed. In this mechanism one portion of the peptide was strongly attracted and close to the surface - the tethering point, while another portion drifted quite far away before being tugged back and ultimately remaining close to the surface. In other words, without the strong attractive interaction at the tethering point, the peptide would not SL to the surface. This type of attachment mechanism is likely more common for larger peptides, explaining why it was not seen for the ala₂-H⁺ work.

E. Reactive landing

Reactive landing (RL) has been studied via simulations for several model systems.^{126,134,135} In order to include reactivity between the surface and the peptide, it was necessary to make some modifications to the general approach outlined above. Namely, it was a requirement to include some atoms of the surface along with all of the atoms of the peptide in the QM calculation. In these simulations, it is typical for the propyl-tip of the center chain to be included in the QM region via the linking atom approach. The first study involving this approach studied $\text{gly}_2\text{-H}^+$ colliding with a chemically modified F-SAM using RM1 semi-empirical theory. Two different chemical modifications were considered in which an F atom on the terminal C is replaced by either a $-\text{C}(\text{O})\text{Cl}$ or a $-\text{C}(\text{O})\text{H}$ moiety. These modifications were chosen to examine the effect of the “leaving group” (either Cl or H) on reactivity. A later study also considered the H-SAM system. Reactivity was significantly higher for the $-\text{C}(\text{O})\text{Cl}$ modification than for $-\text{C}(\text{O})\text{H}$.

Simulations showed that reactivity took place through a 4-centered transition state, which is not common. The TS involves forming two new bonds while simultaneously breaking two old bonds. In particular, the bonds that break involve an X-H bond on the peptide and a C-Y bond on the surface where Y is a “leaving group.” This is consistent with the observation that the $-\text{C}(\text{O})\text{Cl}$ moiety was more reactive, as Cl is a better leaving group. For both studies, good qualitative agreement was found with Laskin and coworkers experimental work.¹³⁶

Recently, Barnes and co-workers studied RL for the cyclic peptide $c(-\text{GGKG}-)$ for both neutral and +1 charged states.¹³⁵ This study represents the most direct comparison to work of Laskin and co-workers.¹³⁶ The work found that overall reactivity increased with collision energy, which would disagree with experiment. However, the experiment is not measuring total reactivity, but rather reactions in which the peptide reacted and remained intact. When examining this type of reaction within the simulations, good qualitative agreement was found with experiment. As in prior work,¹¹⁰ RL occurs through a 4-centered transition state.

Differences were seen between the neutral and charged species. In particular, the binding site on the peptide is different and RL is more likely for the neutral species than for the charged species. Given the size and cyclic nature of this system, the excess proton was placed on the lysine side chain. The basic nature of this amino acid served to deactivate RL at lysine, which accounts for the reactivity difference between the two species. This work also examined the effect of

including more chains in the QM region. Specifically, rather than including just the center chain, some simulations included it as well as the 8 nearest neighbor chains. For the charged species, nearly all of the RL events occur on the center chain, while for the neutral species roughly half occur on the surrounding chains. It is also striking that for the charged species, the covalent linking site is almost exclusively on the peptide backbone N's and O's, while for the neutral it is most likely to occur on the lysine side chain's N.

Lastly, we note that the simulation of the neutral species provides physical insight for the behavior of a large, charged species that happens to have a point of contact with the surface far from the charged site.

III. Collision-Induced Dissociation

Extensive simulations have been performed to study the CID of organic and biological ions. The simulation results have not been previously compared together and summarized. This is done here in the context of describing the role of chemical dynamics simulations for CID experiments.

A. Energy transfer to vibration and rotation

In CID, a crucial aspect is the energy that is transferred upon collision between the ion and the inert gas, which generally is a rare gas (Ar, Ne, Xe ...) or molecular nitrogen (N₂). During the collision, the ion is internally activated and can dissociate. Just as in SID, the probability of dissociation and the fragmentation mechanism are related to (at least) three aspects: (i) the amount of energy that is transferred after each collision; (ii) the partitioning between vibrational and rotational activation; (iii) the localization of the energy after the collision in a particular internal mode and the way this energy flows through the other modes. This last aspect will be discussed in section III.B, here we will first focus on what information simulations can provide regarding the energy transfer process.

Using chemical dynamics simulations, it is possible to directly model the collision between an ion and the inert gas. In particular, single collisions are studied and the collision energy is fixed as an input parameter. This collision energy, in the center-of-mass frame, can be related to the collision energy in the laboratory frame used in triple-quadrupole mass spectrometer:¹³⁷

$$E_{com} = \frac{m_{gas}}{m_{ion} + m_{gas}} E_{lab} \quad (6)$$

where E_{com} and E_{lab} are the energies in the center-of-mass and in the laboratory frame, respectively, and m_{ion} and m_{gas} are the masses of the ion and the inert gas, respectively. In this way, it is possible to make a direct correspondence between collisional simulations and MS/MS experiments done as a function of the collision energy. The collision energy is experimentally often reported in the laboratory frame and can be easily converted into center-of-mass frame via Eq. 6.

Part of the collision energy is transferred to the ion which is rovibrationally activated. The ion-gas interaction is generally described using an analytical function, similarly to SID (see section II.A.1). Often the analytical expression in Eq. 2 is used where A_{XY} , B_{XY} and C_{XY} are obtained by fitting interaction energies calculated *ab initio* and n can be either fixed or a parameter of the fit. Generally, the best fits are obtained for positive values for A_{XY} , B_{XY} and C_{XY} , which leads to a purely repulsive potential. The attractive part of the interaction is often disregarded since it is much smaller than the collision energy and to model the energy transfer it is very important to correctly reproduce the repulsive short-range potential. More details on this aspect can be found in Ref. 88.

When studying relatively large molecules, like peptides, a building block approach is used:⁸⁸ the interaction of the inert gas with CH₄, NH₃ and other elements is obtained from high level calculations (e.g. QCISD(T)). Generally, the interaction is set for each hybridization or chemical environment: for C there are values for sp³ and sp², for H the values are function of the group to which it is attached etc ... Using this approach, and fixing $n = 9$, Meroueh and Hase have developed and reported the interaction potential between Ar and building blocks of polyglycines.⁸⁸ Using the same functional form and approach, parameters for Ar interacting with aromatic rings, the sulfur atom, and some alkaline and alkaline-earth metals (Li⁺, Ca²⁺, and Sr²⁺) have been obtained.¹³⁸⁻¹⁴¹ The same functional form was also used to study N₂ colliding with protonated urea.¹⁴²

Other analytical expressions are used in the literature as for SID. For example, de Sainte Claire et al.¹⁴³ used Eq. 3 with $n = 6$ and $m = 12$ in studying the collision of Ar with Al clusters, and Rodriguez-Fernandez et al.¹⁴⁴ used the same Eq. 3 where n and m were also fitted to study the collision of Ar with [Li(Uracil)]⁺ cluster. Knyazev and Stein¹⁴⁵ have used the analytical expression of Eq. 3 with $n = 9$ and $n = 6$ and parameters obtained by high level *ab initio* calculations. In this way, they successfully studied the reactivity of n-Butylbenzene in CID.

More recently, for CID of a doubly protonated tripeptide, $[\text{TIK}+2\text{H}]^{2+}$, with N_2 , a full semi-empirical Hamiltonian (RM1) was used.¹⁴⁶ Semi-empirical methods can provide correct interactions for N_2 , but they are incapable of treating rare gas atoms. The choice of a full semi-empirical Hamiltonian (like RM1) or an analytical expression depends essentially on the availability of empirical (molecular mechanics, MM) parameters. Results of RM1 and MM for the simple case of N_2 colliding with CH_3OH show that both methods provide short-range interaction energies within 10-30 kcal/mol with respect to accurate QCISD(T)/6-31++G** calculations (see Figure 4).

In Table 4 we summarize results obtained in recent years for different systems using Eq. 2 or semi-empirical Hamiltonians to treat the ion-gas interaction. The average % of the transferred energy is reported. Values are often significantly smaller than 50% and they decrease as the collision energy increases. In fact, with increasing collision energy, the relative velocity also increases and thus the ion-neutral interaction time. While the average energy transfer is relatively small, the distribution is generally broad. As a result, some trajectories get a large portion of the collision energy. These trajectories are mostly reactive trajectories. An example of the distribution for the total energy transferred for reactive and non-reactive trajectories is shown in Figure 5.

Another important aspect which can be studied in such simulations is the partitioning between collisionally acquired rotational and vibrational energy. This information is important when studying the statistical reactivity of activated ions using simulation results. In fact, in RRKM unimolecular dissociation statistical theory, the rate constant is function of both vibrational and rotational internal energy.¹¹⁸ Rotational energy is often disregarded in practical RRKM calculations since in many cases the rotational activation is low and its effect on the rate constant is important only for higher values and/or when the rotational constants (and thus shapes) of reactant and transition state are very different. Note that when analyzing guided ion beam mass spectrometry experiments with statistical theory, the rotational energy has been taken into account, as reported by Armentrout and co-workers.^{147,148} These studies pointed out that a phase space theory approach, in which rotational and orbital angular momentum are explicitly conserved and allowed to interchange, is successful in reproducing the data, even if they also show that the assumptions on angular momentum have no effect on thermodynamic information.¹⁴⁹⁻¹⁵⁵

In general, it is not evident how the partitioning between rotational and vibrational energy results after collisional activation. Simulations can provide information on those aspects. In studies

on Al clusters and small peptides, it was shown that the rotational excitation is higher in case of planar shapes while it is smaller for globular ones.^{114,143} Later, simulations of collisions between protonated urea, uracil and Li-uracil have shown that rotational energy excitation is important.^{142,144,156} A summary of the percentage of rotational energy transfer for different systems we have studied in recent years is reported in Table 4. We should note in particular that rotational excitation is more important for small molecules and it decreases as the collision energy increases. This dependence is shown in more details in the study of collisions of protonated uracil with Ar.¹⁵⁶ For low collision energies, the rotational excitation is more important while, with increasing the energy, the vibrational excitation becomes dominant (see Figure 2 of Ref. 156). Using data from simulations at different collision energies, it is possible to fit the following analytical phenomenological expression to extrapolate the energy transfer at different values of E_{com} :

$$\langle \Delta E \rangle = a_1 e^{-b_1/\sqrt{E_{com}}} + a_2 \left[\operatorname{csch} \left(\frac{b_2}{\sqrt{E_{com}}} \right) \right]^2 \quad (7)$$

where a_1 , a_2 , b_1 and b_2 are adjustable. This expression was originally developed to study the energy transfer to solid surfaces¹⁵⁷ but it can be applied also to CID, since it is an extension of the original Mahan impulsive collision model.⁹³ Nogueira et al. have thoroughly tested this expression in large-scale simulations of SID with different projectiles.¹⁵⁸

Once the energy is transferred to the molecule after collision, this energy is available to promote fragmentation. As for SID, two main classes of fragmentation mechanisms can be defined: (i) shattering mechanisms; (ii) energy flow through the modes corresponding (in the limit of full IVR) to a statistical energy distribution and associated unimolecular decomposition. In this case one expects a RRKM behavior.

We now show how collisional dynamics can unravel the presence of shattering mechanisms and how they are important in understanding CID fragmentation.

B. Shattering fragmentation

As previously discussed, it is possible that a shattering fragmentation occurs (see section II.C). For peptides, trajectories reacting in less than 40 fs can be defined as shattering.¹⁵⁹ In fact, if the bond has no time to vibrate there is also no time for the energy to flow from this mode to other vibrational modes. This mechanism was initially observed in SID simulations but also in the

collision between a molecular ion and the inert neutral gases. This is a typical example of non-RRKM dynamics due to collisional activation, which is localized in space: the portion of the molecule where the projectile hits the ion can quickly react. Concerning small molecules, shattering dissociations and non-RRKM dynamics have been observed in experiments¹⁶⁰ and simulations¹⁶¹ of $\text{CH}_3\text{SH}^+ + \text{Ar}$ CID, experiments¹⁶² and simulations¹⁶³ of $\text{CH}_3\text{SCH}_3^+ + \text{Ar}$ CID, and simulations¹⁶⁴⁻¹⁶⁶ of $\text{Cr}^+(\text{CO})_6 + \text{Xe}$, $\text{H}_2\text{CO}^+ + \text{Ne}$ and $\text{UreaH}^+ + \text{Ar}$ CID. Moving to larger molecules, the first direct dynamics study of CID was performed by Meroueh et al.¹⁰³ where they compared CID with SID fragmentation of protonated glycine. CID simulation with Ar at 13 eV of collision energy and for zero impact parameter did not report shattering, while SID did.

In our recent studies, some shattering fragmentations were observed as reported in Table 4 as a function of the system and the collision energy. The fraction of shattering fragmentation increases with the collision energy (as expected) and that it is quite high for large molecules, like penta- and octa-glycines.¹⁶⁷ This high percentage is due partially to the reaction of the molecular tails, which are more exposed to the collision and thus they react due to a localized collision. Further, for such big systems, complete IVR is relatively slow and in the given simulation time it is less probable to observe trajectories in which the energy has time to redistribute and induce a fragmentation. In some cases, the first fragmentation occurs with a shattering mechanism, as for the NH_3 loss in amine-substituted diglycine, which forms the linear b_2^+ ion that then cyclizes into the well-known oxazolone structure, as observed for diglycine (see Figure 6).¹⁶⁸ A full pathway leading to an observed fragment can be composed of two steps: a first (shattering) in which a part of the molecule dissociates and a second (rearrangement as in the example of Figure 6) which forms the final structure. Of course, mechanisms can be more complex, since mixing of shattering and non-shattering can be involved, and, for example, isomerization can occur before fragmentation (as often is the case for peptides).

Shattering can be an important dynamical mechanism which cannot be accounted for using statistical theories. Thus, studies of the potential energy surface of different dissociation pathways can underestimate the probability of a pathway where shattering occurs.¹⁴⁰ On the other hand, limitation in simulation time-length is a source of overestimation of shattering fragmentations. To better include events at longer time, a statistical treatment is also necessary, as will be discussed in following sections.

C. Comparison with experiments

Chemical dynamics simulations provide the relative abundance of fragmentation products as a function of collision energy. This data provides a means to directly compare with experimental results.

Our first example is the fragmentation of protonated urea, for which collisional dynamics simulations, done at MP2 level of theory, were compared with experiments.¹⁶⁶ Experiments were performed on a QqTOF instrument which operates at low pressure close to the single collision limit. In this case, the fragmentation pattern was simple, consisting of two pathways: (1) neutral ammonia loss and formation of the CONH_2^+ ion (m/z 44); (2) formation of NH_4^+ ions (m/z 18) and neutral CONH. Pathway (2) is energetically favorable, but in experiments and simulations we obtained both pathways. From simulations it was possible, in particular, to explain why the high-energy pathway was observed: it is obtained by the cleavage of C-N bond followed by NH_3 loss, and this happens via a shattering mechanism. By hitting the molecule on that bond, if the leaving NH_3 group escapes with a sufficiently high translational energy it has no time to pick a proton from the NH_2 group forming NH_4^+ and neutral NHCO. This mechanism, elucidated for the first time in the prototypical case of protonated urea, was then obtained for other systems, in particular in protonated peptides in which the protonated N-terminal group can lead to NH_3 loss before any other internal rearrangement.¹⁶⁸

The same approach was used to examine fragmentation of protonated peptides. In the case of amino-substituted protonated di- and tri-glycines, we coupled our simulations with CID experiments and ion spectroscopy (IRMPD) used to characterize the structure of the precursor ion and of some products.¹⁶⁸ Simulations and experiments obtained the same product structures, and the mechanisms suggested from experiments were observed in chemical dynamics simulations. In the case of peptide fragmentation, the experimental literature agrees with the “mobile proton model”,¹²⁷ described in section II.C. From simulations, it was possible to observe the characteristic features of the mobile proton model in two steps: (i) we observed some proton transfers by populating high-energy tautomers; (ii) we used these tautomers as initial structures in further simulations, obtaining new fragmentation products. One key aspect is that the initial protonation site is more important in determining the final products than the conformation of each tautomer. This was observed not only in peptides but also in uracil.¹⁴¹ A proton transfer (PT) is a process that is generally slow with respect to the simulation time scale. If one PT is obtained, then a second reaction (or even a second PT) has a very low probability to be observed in simulations, mainly

due to the limitation in simulation time. Thus, putting together simulations done using different tautomers as initial structures, it is possible to reconstruct the full fragmentation scheme and understand the origin of the observed products.¹⁶⁹ Of course, by increasing the simulation time, it is also possible to obtain several proton transfers and related fragmentation.

One characteristic gas phase reactivity of peptides is that, when the linear *b* ion is formed by C-N bond cleavage, then there are two possible nucleophilic attacks leading to two different cyclic structures: (i) the 5-member ring oxazolone and (ii) the 6-member ring diketopiperazine, which is energetically more stable (but not observed in many cases).^{45,49,170-172} Simulations of polyglycines and N-Formylalanylamine provide as product the oxazolone in agreement with experiments.¹⁶⁷⁻¹⁶⁹ More recently the negative ion of deprotonated di-proline was studied, and the diketopiperazine *b*₂ ion was obtained from simulations, in agreement with IRMPD experiments probing the molecular structure of such a product.¹⁷³

Another successful performance of collision dynamics in reproducing CID spectrum and products of biologically relevant molecule is the study of protonated testosterone.¹⁷⁴ First, the fragment peak distribution obtained from simulations is very similar to what is obtained experimentally (see Figure 7).¹⁷⁵ Second, from simulations it was possible to determine the mechanisms leading to the different pathways, which provided computational conformation of some of them.¹⁷⁵⁻¹⁷⁷ Finally, the simulations find the same geometries obtained experimentally by IRMPD spectroscopy, thus strengthening our confidence in the other structures (and associated mechanisms) for which no experimental details are available. This study gives us confidence in using collisional simulations to study other similar systems in the future, with possible applications to metabolomics or characterization of doping substances.

As clearly shown from the study of testosterone, but also true in other systems, the simulations reproduce fairly well the experimentally obtained fragments, but often the intensity of the peaks is very different. The reasons are briefly summarized here:

- 1) The *ab initio* computational method employed is, by necessity, approximate in nature, since fully correlated methods (and the completed basis set limit) are computationally intractable for use in these direct dynamics simulations. In practice, it is possible to use DFT or MP2 with relatively small basis set for small molecules and semi-empirical Hamiltonians for larger systems. Comparisons between DFT and semi-empirical Hamiltonians were done in some cases, as we will discuss in section III.E.

- 2) The limitation in simulation time-scales is another source of discrepancy. Direct dynamics trajectories are limited to picoseconds, while, in experiments, fragmentations occur up to milliseconds. As already mentioned, the shattering fragmentations can be overestimated and thus the peaks which are not formed via shattering can have a much lower occurrence in simulations. To include longer time-scale fragmentation events, RRKM theory was also included in some studies when the transition states (TSs) connecting the reactant with the products were located.^{140,156} Recently, Martinez-Nunez has developed an automatic algorithm to find minima and TS from simulations using high-temperature trajectories to sample the configuration space,¹⁷⁸⁻¹⁸⁰ which was applied to the case of CID of protonated uracil.¹⁵⁶ Coupling it directly with the collisional dynamics is a clear possibility and a recent first study was reported for cysteine-sulfate fragmentation.¹⁸¹ However, this method cannot be used if a pathway is not observed in simulations and the usual approaches to determine reaction pathways and TSs must be used, which can be very problematic for large and flexible systems.
- 3) Collisional simulations are done in the single collision limit, while in experiments multiple collisions occur often. Many experiments are done in ion traps in which the activation mode is different and the collision energy is not clearly defined. This will be discussed in next section.
- 4) Finally, experimentally, the intensity of fragments depends on many details, including the mode of ionization and source conditions, ion transmission and detection, mode of collisional excitation, and time between ion activation and analysis. This phenomenon can be seen in the simple example of fragmentation of the simple protonated uracil molecule,^{141,182-184} where the reaction mechanisms were elucidated by simulations. The aim of modeling CID is to provide a global framework to understand the fragmentation mechanisms and to predict the possible products, not to model one (or each) specific mass spectrometer.

D. Single vs multiple collisions

CID is not at all limited to triple-quadrupole instruments and to the single collision limit. Ion traps are largely employed in mass spectrometry, in particular for large molecules. One key difference is that in ion traps, the fragmentation results from multiple low energy collision events. Figure 8 gives a graphical representation of the two activation modes for the model case of a

fragmenting Morse oscillator. In the case of ion trap like activation, we assume that after each collision the energy is totally randomized between the modes before the next collision occurs. This corresponds to random activation of the different modes (i.e. statistical distribution of the excess energy). It is modeled by selecting initial conditions for the ion from a microcanonical ensemble at the given total energy, E . As discussed below, for a large molecule a temperature T may be associated with E and this becomes a thermal activation method.¹⁴⁶ It was recently applied to fragmentation of a doubly charged tripeptide $[\text{TIK}+2\text{H}]^{2+}$ ^{146,185} and diproline anion ($[\text{Pro}_2 - \text{H}]^-$).^{173,186} Fragments were obtained and compared with collisional simulations in terms of their nature and the corresponding mechanisms. For example, we observed that side chain fragments are more abundant in single collision simulations than in internal energy activation simulations.

By using statistical mechanics, the (vibrational) internal energy of a set of classical harmonic oscillators can be related to the (vibrational) temperature via the simple relation¹⁸⁷

$$E = s k_B T \quad (8)$$

in which s is the number of normal modes, k_B the Boltzmann constant, and T the temperature. This expression holds for fragmentation simulations presented here since the equations of motions are propagated using classical physics, and it is valid when quantum nuclear effects can be disregarded.

If the unimolecular dynamics of the randomly excited ion energy E is ergodic, its decay will be exponential and a rate constant, $k(E)$, may be determined from exponential decay. This is a classical unimolecular rate constant, and does not include quantum effects such as tunneling and zero-point energy constraints. As shown previously,¹⁸⁸ if $s \approx (s - 1)$ for the unimolecular reactant's s vibrational modes and the unimolecular dissociation energy E_o is much less than the reactant's energy E (i.e. $E_o/E \ll 1$), the classical microcanonical RRKM unimolecular rate constant $k(E)$ at E becomes identical to the classical canonical transition state theory (TST) rate constant $k(T)$ at T with $E = s k_B T$,¹⁸⁷ where k_B is the Boltzmann constant. Thus, constant E simulations may be used to determine rate constants for constant T . This relation between E and T has also been discussed with respect to product energies in unimolecular dissociation.¹¹⁸

By performing simulations at different E , the rate constant $k(T)$ may be obtained at different T . Verifying Arrhenius behavior,

$$k(T) = Ae^{-\frac{E_a}{k_B T}} \quad (9)$$

it is possible to obtain the pre-exponential factor and activation energy for the total unimolecular dissociation. Rate constants for individual pathways, $k_i(T)$, can be obtained from $k(T)$, for which the individual $k_i(T)$ equals $k(T)$ multiplied by the number of dissociations for the i^{th} pathway divided by the total number of dissociations $p_i(T)$; that is, $k_i(T) = p_i(T) k(T)$. Thus, pre-exponential factors and activation energies may be determined for individual pathways. The activation energy E_a equals the classical potential energy barrier for the unimolecular dissociation.¹⁴⁶

The constraints $s \approx (s - 1)$ and $E_o/E \ll 1$ apply to $[\text{TIK}+2\text{H}]^{2+}$ and $[\text{Pro}_2 - \text{H}]^-$ dissociation and the above approach was used to determine pre-exponential factors and activation energies for some of their dissociation pathways.^{146,186} This analysis was possible for pathways that had statistically meaningful values of $p_i(T)$. Within statistical uncertainties, the resulting activation energies were in agreement with barrier heights of transition states located as saddle points.^{146,186} This opens the possibility of using simulations to directly locate energy thresholds and, as we discuss in section IV, use them to extrapolate results to longer times. Of course, statistically meaningful values of $k_i(T)$ will be obtained mainly for the important pathways, which will have lower activation energies and faster kinetics. To better describe slower and rare events, with this direct approach, a larger number of longer trajectories will be necessary. For large biological ions, the constraint $s \approx (s - 1)$ will always apply. Also, given the large value for s , it is expected that the constraint $E_o/E \ll 1$ will also be applicable, but it should always be tested.

E. Methods performances

The accuracy and reliability of the simulations can be established either by comparing results with experimental data or with other calculations. In previous sections, we have discussed how simulation results compare with experiments and which factors must be taken into account for a correct comparison. In particular, we should point out that simulations are limited in time-length such that fast processes are necessarily overestimated: this means that often secondary fragmentations are not sampled.

Another possibility is to compare different theoretical methods (i.e. dynamics done using different quantum chemistry methods to describe the reactivity). This is particularly important when using semi-empirical Hamiltonians. When the size of the system allows, as for uracil and N-Formylalanylamine, we have compared AM1 and PM3 results with density functional theory

(DFT) results using the B3LYP functional.^{141,169} Results show that semi-empirical Hamiltonians provide globally the same products. In general, fewer DFT-trajectories are performed and have a reduced simulation time with respect to those done with semi-empirical Hamiltonians and thus results are not fully converged, statistically speaking. Finally, in the case of TIK we compared AM1 and RM1 results, finding the same fragmentation patterns.¹⁴⁶

Concluding, the choice of semi-empirical method is based on the one hand on the practical implementation of the coupling between VENUS and the semi-empirical package, and on the other hand on the performances of the method. RM1 and PM6-D are suggested as good first choice to perform fragmentation simulations. In the future, a systematic assessment of semi-empirical methods (including also the tight-binding DFT¹⁸⁹) will surely be useful to provide a clear guide to choose the best method for the given system.

IV. Future Directions

There are a number of future directions for the research described in this work. It should be possible to use the information provided by experiments regarding the time dependence of ion formation to predict the ion yield at longer times. For small biological ions, it may be possible to use quantum dynamics to simulate their fragmentation. It would also be of interest to develop software for including a zero-point energy (ZPE) constraint in the classical dynamics simulations of fragmentation.¹⁸⁹⁻¹⁹² Proton transfer along a peptide ion's backbone is important for fragmentation, and rate constants may be determined from the simulations. Finally, there are a number of important simulations of biological molecules colliding with surfaces and of their CID to be performed, which are extensions of the previous studies, but require enhancements of the computational methodology.

A. Extrapolating the Fragmentation Dynamics to Long Times

One interesting future direction will be to use rate constants obtained from the simulations to extrapolate results to longer times. In particular, as shown for primary fragmentations,^{146,193} it is possible to follow the formation in time of different products and fit their abundance with a kinetic model. For unimolecular irreversible fragmentations, a simple model would consist of an initial ion fragmenting into n primary products which can eventually further fragment into m secondary products. The number of primary products and subsequent branching reactions will depend on specific rate constants. Each fragmentation step is characterized by a rate constant

which can be obtained as shown in section III.D directly from simulations, avoiding the procedure of locating the TSs. Then, the full kinetic systems can be solved analytically or numerically. By extrapolating the ratio of different ions to longer times, it may be possible to obtain a time-independent theoretical mass spectrum which will reflect theoretical ion ratios on time-scales closer to the experimental ones. This may overcome the problem arising from limitation in simulation time. In fact, one limitation of simulations is that short time-scale products are normally overestimated. If at least some low abundance products are obtained, then their rate constants can be fitted from trajectories and their abundance at longer time-scales estimated. Of course, the quality of the estimation will depend on the abundance of such rare events, and likely more trajectories with longer simulation times will be necessary. Furthermore, by fitting Arrhenius-like or RRKM expressions with rate constants obtained from simulations, it will be in principle possible to use such information in more specific models of fragmentation of ions as those specifically conceived by Knyazev and Stein for double-octopole guided beam, triple-quadrupole and quadrupole-hexapole-quadrupole tandem mass spectrometers.^{147,194}

B. Quantum Dynamics

The chemical dynamics simulations described above for SID and CID use classical mechanics for the atomistic dynamics. For some particular conditions, nuclear quantum effects may be important. One may be concerned with tunneling for proton transfer reactions and zero-point energy effects for ion fragmentations. In recent years, different methods have been developed for using classical trajectories to account for quantum effects.^{190-192,195-197} Wave-packet methods, even with efficient approaches like multi-configuration time-dependent Hartree,¹⁹⁸ are too computationally demanding for application to SID and CID systems, and this is expected to be the case for the near future. However, methods like Centroid Molecular Dynamics,¹⁹⁹ Ring Polymer Molecular Dynamics,²⁰⁰ and Quantum Thermal Bath²⁰¹ may be applicable. This will require implementations and accurate studies to evaluate the feasibility of their applications. A first application of Quantum Thermal Bath was reported by Spezia and Hammak for the model reaction $\text{CH}_4 \rightarrow \text{CH}_3 + \text{H}$.²⁰² These methods are defined for canonical ensembles and will need some reformulation and/or *ad hoc* modification to study collisions. The use of semi-classical methods, which may be defined for microcanonical ensembles, is tempting. While rigorous methods like Herman-Kluck²⁰³ are computationally intractable for CID and SID, more approximate methods, like frozen Gaussians²⁰⁴ or generalized Gaussian wave-packet²⁰⁵ dynamics, are probably

affordable. Future studies, incorporating quantum dynamics will be surely useful, not only for SID and CID, but in general for understanding gas phase ion reaction dynamics.

C. ZPE Constraint for Fragmentation

In classical chemical dynamics simulations, a fragmentation transition state (TS) may be crossed without ZPE in the vibrational modes orthogonal to the reaction coordinate, which results in lower threshold for fragmentation than that given by quantum dynamics. As a result, the classical fragmentation rate constant is larger than the quantum value. A ZPE constraint has been proposed for providing a correction to the classical dynamics,¹⁹¹ resulting in the quantum threshold and a rate constant closer to the quantum value. For a fragmentation without a saddle point, as for ${}^+\text{NH}_3\text{CH}_2\text{-COOH} \rightarrow \text{NH}_3\text{CH}_2^+ + \text{COOH}$, if each fragment has an internal energy which is less than the ZPE the velocities of the atoms are reversed and the trajectory returned to the reactant region of phase space. In this manner, the classical simulations have the quantum threshold. This procedure is continued until fragments are formed with ZPE and the reactant's lifetime is the total trajectory time for forming fragments with ZPE.

The above algorithm needs to be extended so that it may be applied to a TS at a saddle point, for which the requirement would be ZPE in the vibration modes orthogonal to the TS's reaction coordinate. The algorithm and software need to be developed for implementing this procedure into the classical chemical dynamics simulations. For the above bond dissociation reaction, without a saddle point, a sufficient ZPE constraint is ZPE in each fragment.

D. Determining $k(T)$ for Proton Transfer

An important component of the dynamics of peptide ion fragmentation is proton transfer along the its backbone. Knowledge of the rate constants $k(T)$, for these transfers, would be very helpful in modeling the ion's fragmentation. The same procedure, as used to study peptide- H^+ fragmentation versus temperature,^{146,186} may be used to determine $k(T)$ for proton transfer. The ion may be thermally excited with H^+ attached to different sites, e.g. NH_2 , COOH , and the C-N peptide bond, and $k(T)$ determined for proton transfer. If multiple transfers occur, $k(T)$ may be determined for each individual transfer. Arrhenius parameters, A and E_a , may be determined from the $k(T)$. E_a is the classical activation energy without ZPE, but it should be straightforward to develop models for including a ZPE correction.

E. Future CID and SID Simulations

Finally, after the series of successful studies on CID of organic and small biological molecules as amino-acids, peptides or sugars with less than 100 atoms, as described above, an important step will be to simulate larger systems, like extended polypeptides, sugars, DNA etc ... Towards this end, the electronic structure description must be greatly simplified to speed-up the calculations. Even semi-empirical Hamiltonians will be too computationally demanding for an appropriate statistical sampling. Analytical force fields, which describe the fragmentation dynamics, should be developed in order to study larger molecules and make the CID simulations applicable to proteomics, glycomics, and metabolomics. Data bases for the force fields could be constructed by direct dynamics simulations of the pathways for smaller molecules. A possibility to develop analytical potential for chemical reactivity will be using “machine learning” techniques.²⁰⁶⁻²⁰⁸

An interesting future extension of the simulation method will concern the reactivity of complexes with transition metals.^{209,210} In this case, a careful description of the potential energy surface is needed, with the likelihood of spin transitions also considered. The possibility of using modern DFT methods or further developments of electronic structure theory methods is appealing. At the present time, such simulations are limited by the computing time and by the difficulty of obtaining a reasonable description of the potential energy surface with methods not too computationally demanding. Studies in this direction are surely welcome.

For simulations of collisions of biological ions with surfaces, there is a need for additional studies of soft-landing and reactive-landing for comparison with experiment. To simulate collisions of large biological ions with surface, analytic potentials are needed and a strategy for their construction follow the above scheme described for the CID simulations. There is a considerable interest in simulating collisions of peptide dimers composed by units with different charge states. If there is prompt SID fragmentation of the dimer, there may not be extensive proton transfer between the different peptides with a considerable effect on final products. The SID fragmentation may be compared with random thermal excitation fragmentation and it would be instructive to understand how the two eventually differ.

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Table 1. Energy Transfer Dynamics for Peptide-H⁺ Collisions with Organic Surfaces^a

| Peptide-H ⁺ | Surface | E_i (eV) | θ_i (°) | ΔE_{int} (%) | ΔE_{surf} (%) | ΔE_f (%) | ref. |
|---|---------|----------------------------------|-------------------|-------------------------|--------------------------|---------------------|------------------|
| gly ₃ -H ⁺ folded | H-SAM | 30 | 45 | 7 | 63 | 30 | 95 |
| gly ₃ -H ⁺ extended | H-SAM | 30 | 45 | 8 | 54 | 38 | 95 |
| gly ₈ -H ⁺ H-SAM | | 5 | 0,45 | 12,10 | 57,38 | 30,52 | 111 ^b |
| | | 10 | 0,45 | 11,11 | 66,48 | 23,40 | |
| | | 30 | 0,45 | 11,13 | 74,62 | 16,26 | |
| | | 50 | 0,45 | 11,13 | 76,67 | 14,20 | |
| | | 70 | 0,45 | 13,11 | 76,71 | 13,16 | |
| | | 90 | 0,45 | 12,13 | 77,72 | 11,14 | |
| | | 100 | 0,45 | 12,13 | 76,73 | 12,13 | |
| ala ₂ -H ⁺ | F-SAM | 110 | 0,45 | 13,13 | 77,74 | 10,12 | 112 |
| | | 5 | 0,45 | 19,22 | 73,62 | 8,16 | |
| | | 13.5 | 0 | 18 | 71 | 11 | |
| | | 22.5 | 0,45 | 20,23 | 76,61 | 4,16 | |
| | | 30 | 0 | 19 | 78 | 3 | |
| gly-H ⁺ | diamond | 70 | 0,45 | 17,11 | 47,39 | 36,50 | 108 ^c |
| | | gly ₂ -H ⁺ | 5 | 45 | 16 | 0 | |
| gly ₂ -H ⁺ | diamond | 10 | 45 | 17 | 4 | 79 | 106 |
| | | 30 | 0,45 | 24,16 | 27,12 | 49,72 | (d) |
| | | 50 | 0 | 21 | 37 | 42 | 105 |
| | | 70 | 0,45 | 20,15 | 40,25 | 40,60 | (d) |
| | | 100 ^c | 0,45 | 17,13 | 48,34 | 35,53 | (d) |
| ala ₂ -H ⁺ | diamond | 5 | 45 | 19 | 1 | 80 | 105 |
| | | 10 | 45 | 19 | 5 | 76 | |
| | | 30 | 45 | 18 | 12 | 70 | |
| | | 70 | 45 | 21 | 21 | 58 | |
| | | 100 ^c | 45 | 15 | 31 | 54 | |
| gly ₃ -H ⁺ folded | diamond | 10 | 45 | 8 | 2 | 89 | 95 |
| | | 30 | 45 | 18 | 9 | 73 | |
| | | 70 | 45 | 17 | 21 | 60 | |
| | | 110 | 45 | 14 | 29 | 57 | |
| gly ₅ -H ⁺ folded | diamond | 30 | 45 | 23 | 5 | 72 | 95 |
| gly ₈ -H ⁺ | diamond | 100 | 0,45 | 45,26 | 26,12 | 29,62 | 110 ^f |

a. The simulations are performed with the AMBER molecular mechanics (MM) force field for the peptide ion's intramolecular potential, unless otherwise noted.

b. The simulations were performed with RM1 for the peptide ion's intramolecular potential.

c. The simulations were performed with MP2 for the peptide ion's intramolecular potential.

d. 0° results are AM1 from ref. 136 and 45° results are AMBER from ref. 105.

e. The 45° result is for $E_i = 110$ eV.

f. The simulations were performed with AM1 for the peptide ion's intramolecular potential.

Table 2. Peptide-H⁺ Intramolecular Potential and Energy Transfer Efficiencies

| Potential | E_i (eV) | ΔE_{int} (%) | ΔE_{surf} (%) | E_f (%) | Ref. |
|---|---------------|-------------------------|--------------------------|--------------|------|
| gly-H ⁺ + diamond, $\theta_i = 45^\circ$ | | | | | |
| AMBER | 70 | 11 | 37 | 52 | 104 |
| AM1 | 70 | 12 | 38 | 50 | 104 |
| MP2 | 70 | 11 | 39 | 50 | 108 |
| gly ₂ -H ⁺ + diamond, $\theta_i = 0^\circ$ | | | | | |
| AMBER | 35 | 27 | 29 | 44 | 107 |
| AM1 | 30 | 24 | 27 | 49 | 105 |
| AMBER | 70 | 22 | 41 | 37 | 107 |
| AM1 | 70 | 20 | 40 | 40 | 105 |
| gly ₂ -H ⁺ + diamond, $\theta_i = 45^\circ$ | | | | | |
| AMBER | 70 | 15 | 25 | 60 | 106 |
| AM1 | 70 | 12 | 27 | 61 | 105 |

Table 3. Percentage of Shattering SID in QM+MM Direct Dynamics Simulations with a Diamond {111} Surface

| Peptide-H ⁺ | QM ^a | E_i (eV) | θ_i (°) | Shattering ^b (%) | Channels ^c | Ref. |
|----------------------------------|-----------------|---------------|-------------------|--------------------------------|-----------------------|------|
| gly-H ⁺ | AM1 | 70 | 45 | 23 | 18 | 103 |
| gly-H ⁺ | MP2 | 70 | 0 | 57 | 96 | 107 |
| gly ₂ -H ⁺ | AM1 | 30 | 45 | 22 | 14 | 104 |
| | | 50 | 0 | 8 | 6 | |
| | | 70 | 0 | 13 | 23 | |
| | | 100 | 0 | 44 | 44 | |
| gly ₈ -H ⁺ | AM1 | 100 | 0 | 71 | 59 | 109 |
| | | 100 | 45 | 78 | 304 | |
| | | | | | - ^d | |

- a. Quantum mechanical method for the direct dynamics.
b. Percentage of the fragmentations which are shattering.
c. Number of different fragmentation channels.
d. Not determined.

Table 4. Summary of results obtained in collisional dynamics of different systems, as a function of collision energy (E_{col}), percentage of transferred energy (E_{transf}), of rotational activation (E_{rot}) and shattering mechanisms.

| Collision System | $E_{\text{col}}^{\text{a}}$ | $E_{\text{transf}}^{\text{b}}$ (%) | $E_{\text{rot}}^{\text{c}}$ (%) | Shattering ^d (%) | Reference |
|---|-----------------------------|---------------------------------------|------------------------------------|--------------------------------|-----------|
| UreaH ⁺ + Ar | 4.40 | 33 | 22 | 12 | 166 |
| | 5.66 | 32 | 20 | 33 | 166 |
| | 6.29 | 35 | 21 | 41 | 166 |
| UreaH ⁺ + N ₂ | 6.29 | 19 | 13 | 14 | 142 |
| CaUrea ²⁺ + Ar | 9.11 | 29 | 13 | - | 139 |
| | 13.00 | 20 | 8 | - | 139 |
| CaForm ²⁺ + Ar | 7.80 | 14 | 11 | - | 140 |
| | 9.97 | 10 | 8 | - | 140 |
| | 12.14 | 9 | 6 | - | 140 |
| SrForm ²⁺ + Ar | 7.80 | 9 | 6 | - | 140 |
| | 9.97 | 8 | 5 | - | 140 |
| | 12.14 | 7 | 4 | - | 140 |
| H ⁺ -Gly ₂ NH ₂ + Ar | 15.18 | 8 | 3 | - | 168 |
| H ⁺ -Gly ₃ NH ₂ + Ar | 15.18 | 12 | 0.7 | - | 168 |
| H ⁺ -Gly ₅ NH ₂ + Ar | 15.18 | 51 | 3 | 51 | 167 |
| H ⁺ -Gly ₅ OH + Ar | 15.18 | 14 | 0.6 | 50 | 167 |
| H ⁺ -Gly ₈ NH ₂ + Ar | 15.18 | 22 | 0.7 | 55 | 167 |
| H ⁺ -Gly ₈ OH + Ar | 15.18 | 48 | 1 | 81 | 167 |
| Pro ₂ ⁻ + Ar | 13.01 | 20 | - | - | 173 |
| [TIK+2H] ²⁺ + N ₂ | 10.8 | 23 | | 50.8 | 185 |
| | 12.9 | 18 | | 57.8 | 185 |
| | 30 | 3.5 | | 91.6 | 185 |
| [TLK+2H] ²⁺ + N ₂ | 12.9 | 11 | 1 | 42 | 185 |
| TestosteroneH ⁺ + Ar | 30 | - | - | 22 ^c | 174 |

a. Collision energy in eV in the center-of-mass frame.

b. Percentage of total energy transfer (average values calculated over non-reactive trajectories).

c. Percentage of the energy transfer to the polyatomic ion that is rotational energy (average values calculated over non-reactive trajectories).

d. Percentage of the ion fragmentation (over the reactive trajectories) that is shattering.

e. Shattering % for the most important fragments for which absolute number of trajectories are reported and global % can be obtained. Details for shattering vs fragment are reported in the original publication, Ref. 174.

Figure Captions

Figure 1. Comparison between intermolecular potential energy curves for CF_4 interacting with the $-\text{NH}_2$ group of HCONH_2 for different orientations; MP2-aug-ccpVTZ (solid circles) and fitted analytic potential (solid lines). (Reprinted with permissions from ref. 92. Copyright 2014 American Chemical Society).

Figure 2. Distributions of energy transfer to ΔE_{int} , ΔE_{surf} , and E_f for $\text{gly}_2\text{-H}^+$ + diamond {111} collisions at $E_i = 70$ eV and $\theta_i = 45^\circ$. Simulations for the AMBER (----) and AM1 (—) models for the $\text{gly}_2\text{-H}^+$ intramolecular potential are compared. (Reprinted with the permissions from ref. 105. Copyright 2003 Elsevier B.V.).

Figure 3. Ensemble averages of the internal energy of folded $\text{gly}_3\text{-H}^+$ versus time for $E_i = 10$ (upper left), 30 (upper right), 70 (lower left), and 110 eV (lower right) collisions with diamond {111}.

Figure 4. $\text{N}_2\text{-CH}_3\text{OH}$ intermolecular potential energy curves (different orientations are shown) as obtained from RM1, QCSID(T) and analytical potential (MM) of Eq.1.

Figure 5. Energy transfer distribution as obtained in collisional dynamics of $[\text{TIK}+2\text{H}]^{2+}$ with N_2 using a collision energy of 9.76 eV in the center-of-mass reference frame. Reactive and non-reactive trajectories are shown in red and black, respectively.

Figure 6. Example of reaction of protonated aminated-di-glycine forming first a linear b^+ ion (m/z 115) via a shattering mechanism followed by a rearrangement consisting in cyclization leading to oxazolone ion.

Figure 7. Mass spectra of protonated testosterone as obtained by CID simulations (left) and experiments (right). Figures are adapted from Ref. 174 and Ref. 175 for theory and simulations respectively. Reproduced by permission of the Elsevier B.V. (left panel) and Springer Nature (right panel).

Figure 8. Schematic picture of single collision (panel A) vs multiple collision (panel B) activation of a model Morse oscillator leading to dissociation.

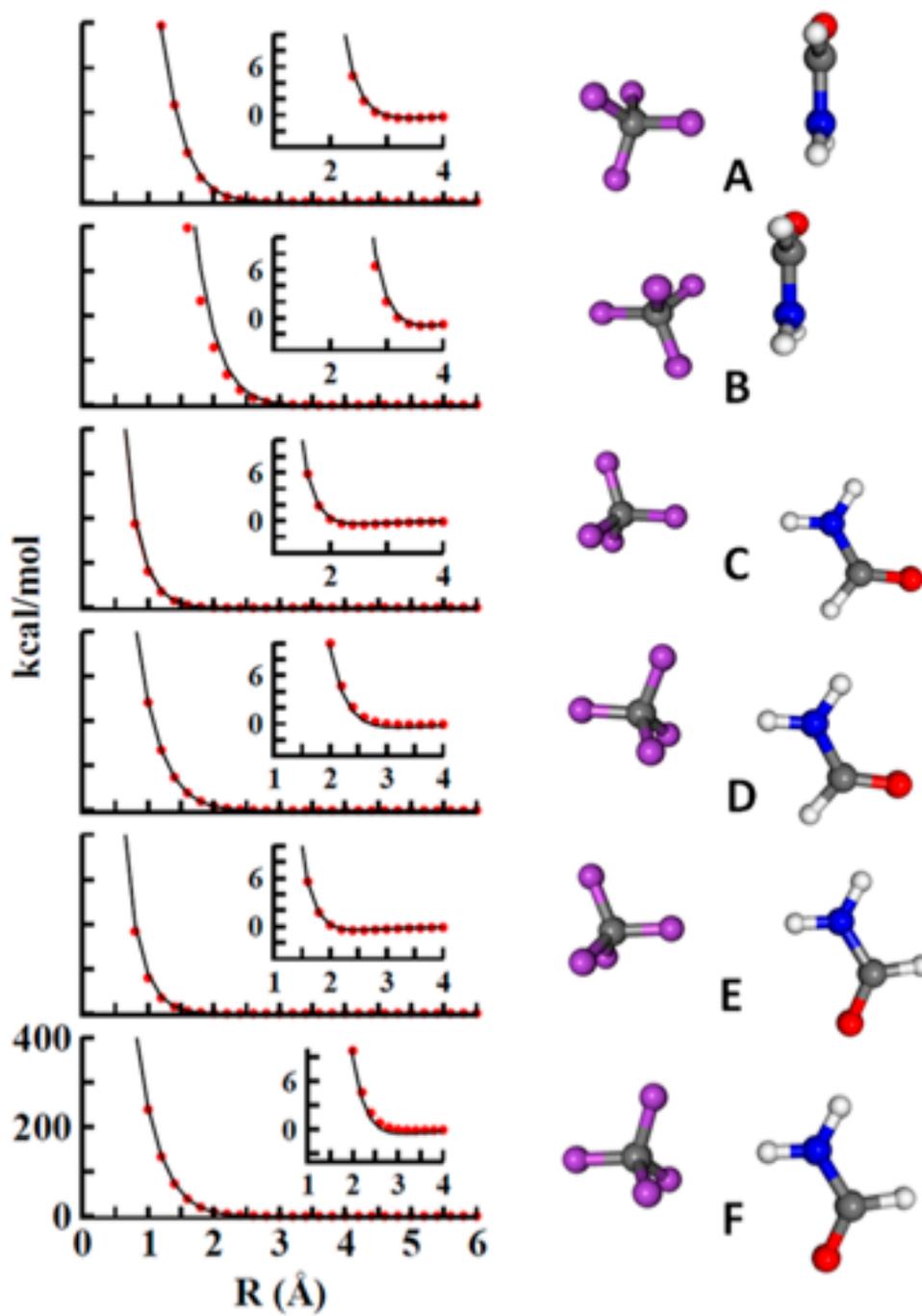


Figure 1.

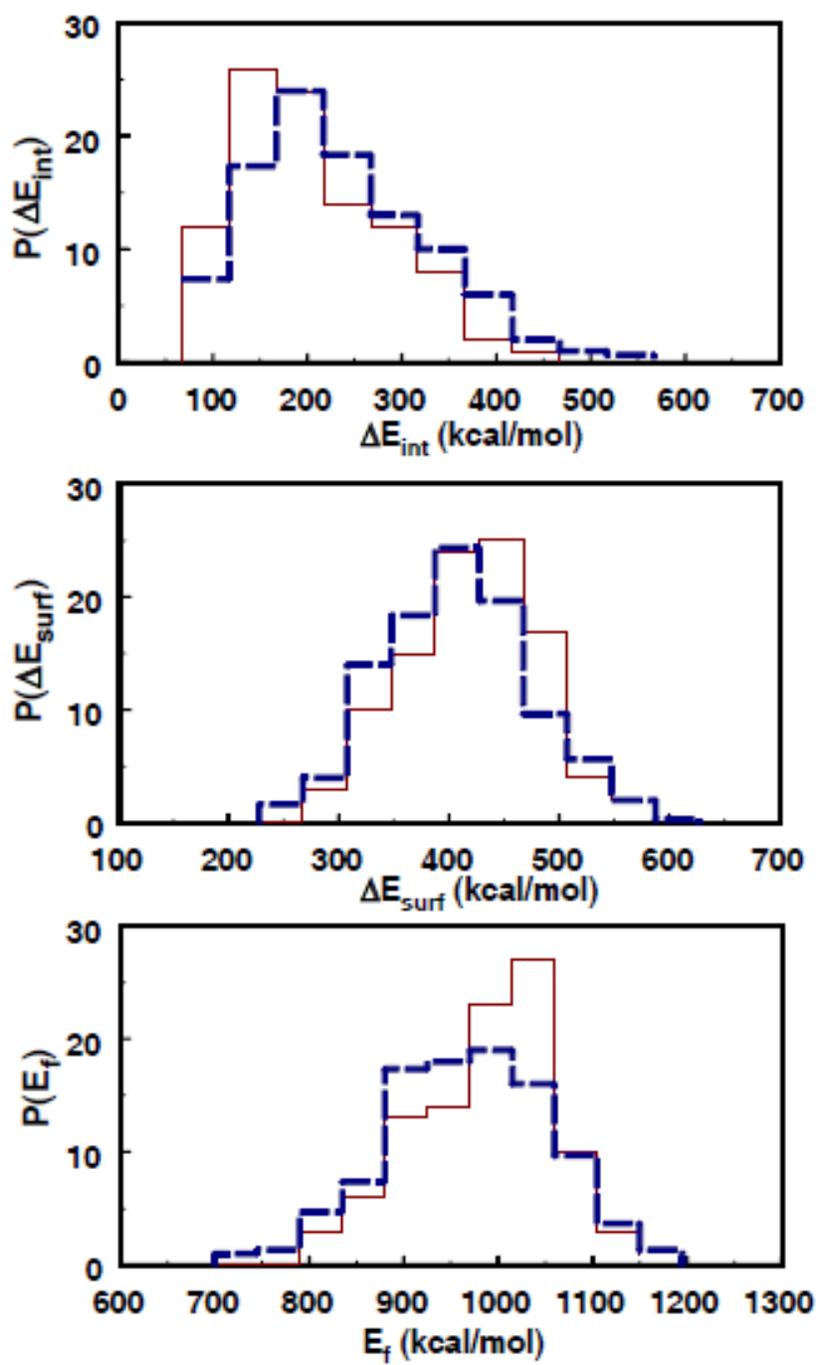


Figure 2.

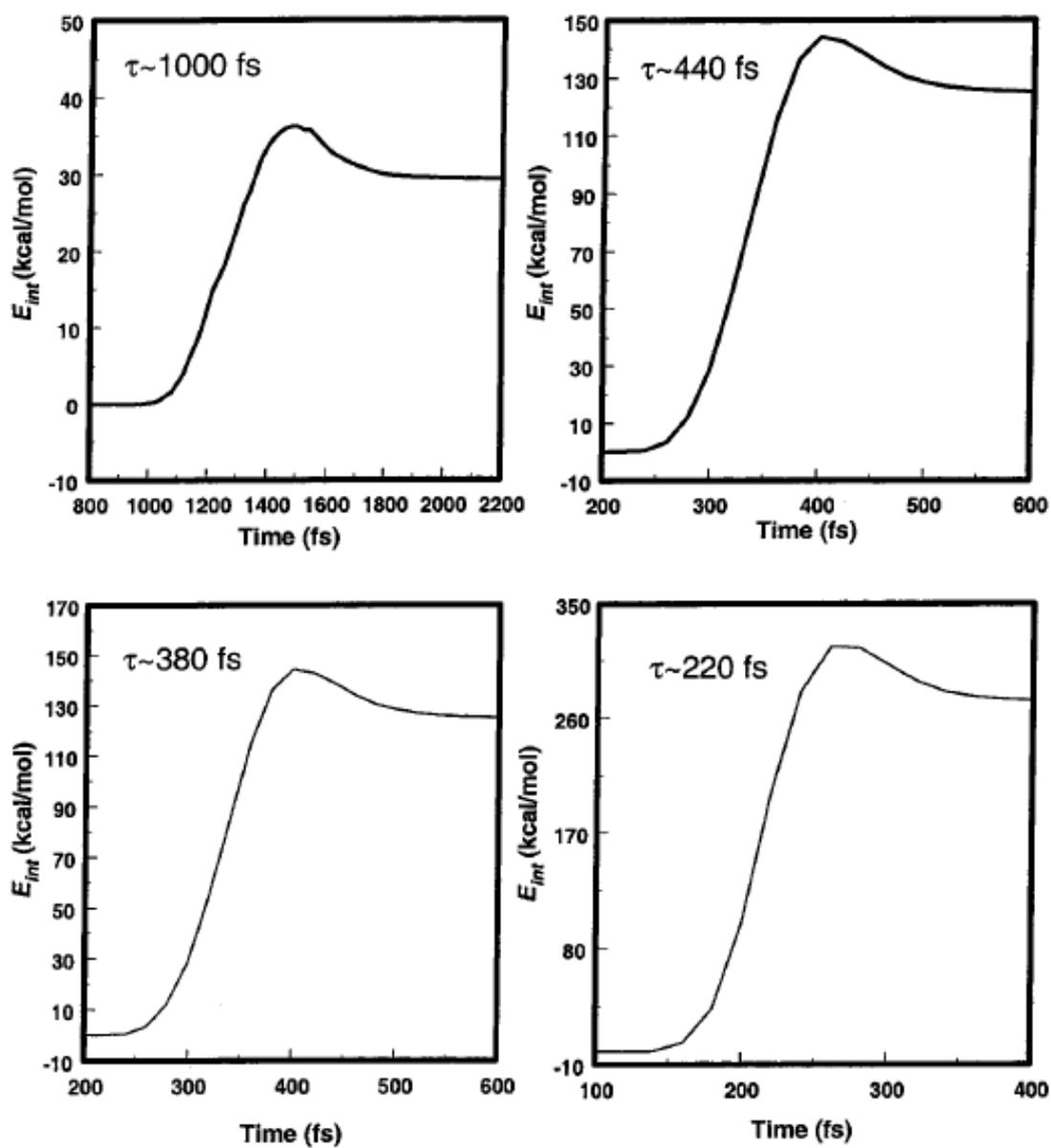


Figure 3.

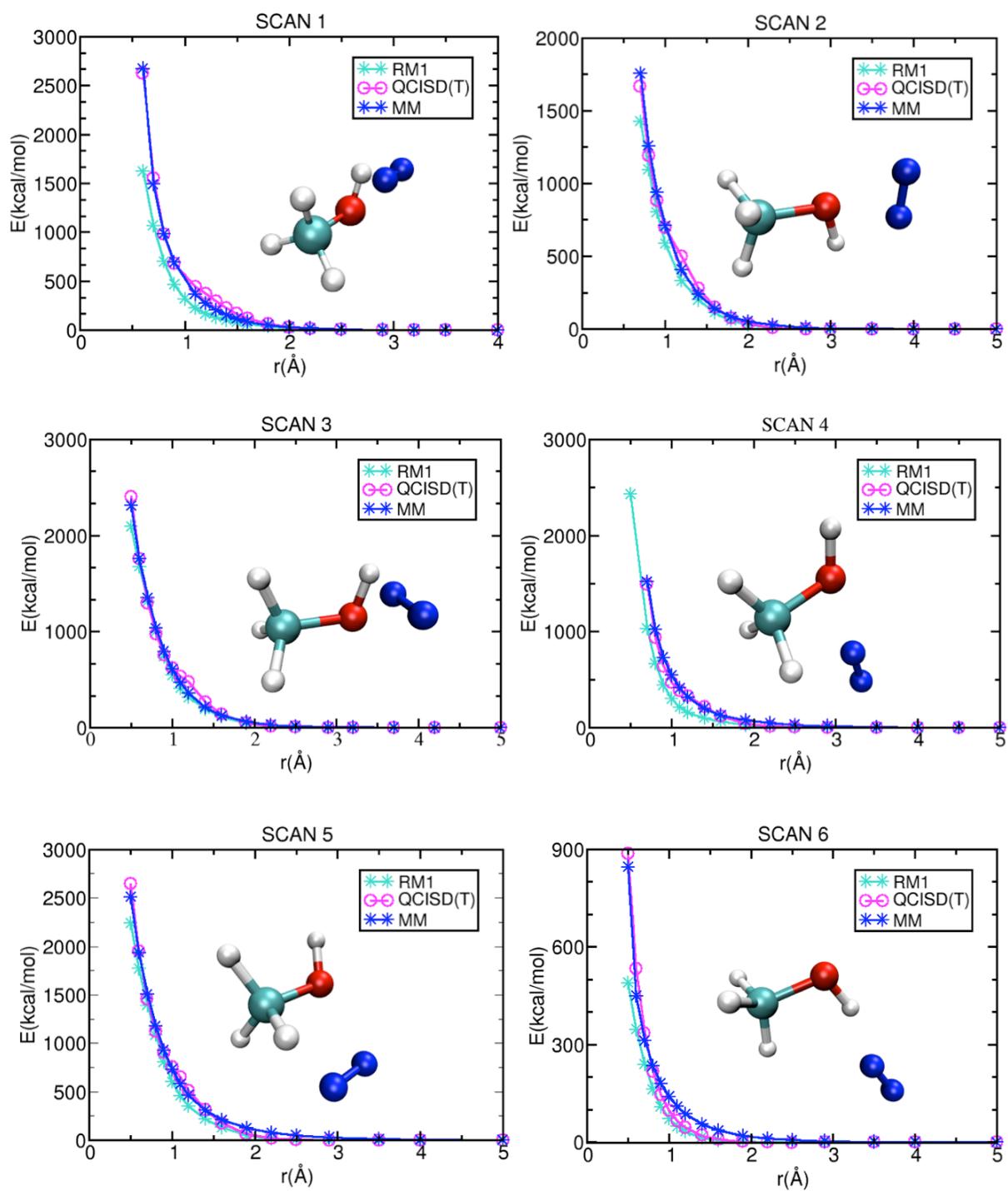


Figure 4.

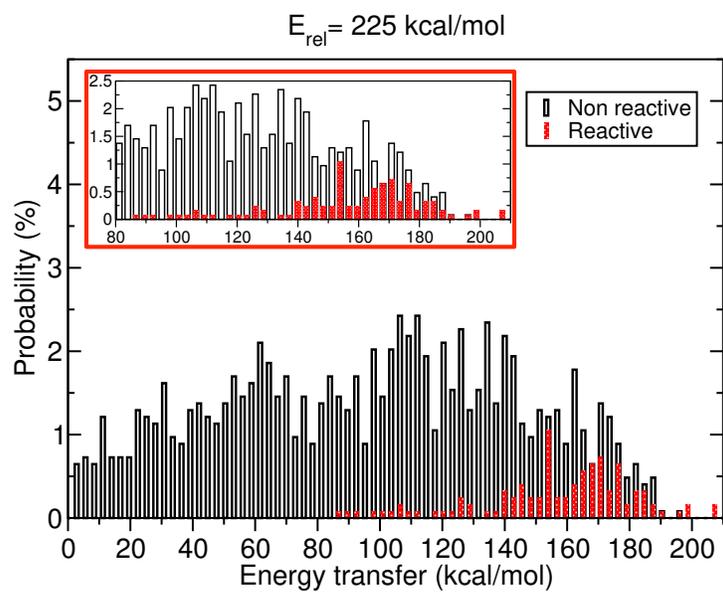


Figure 5.

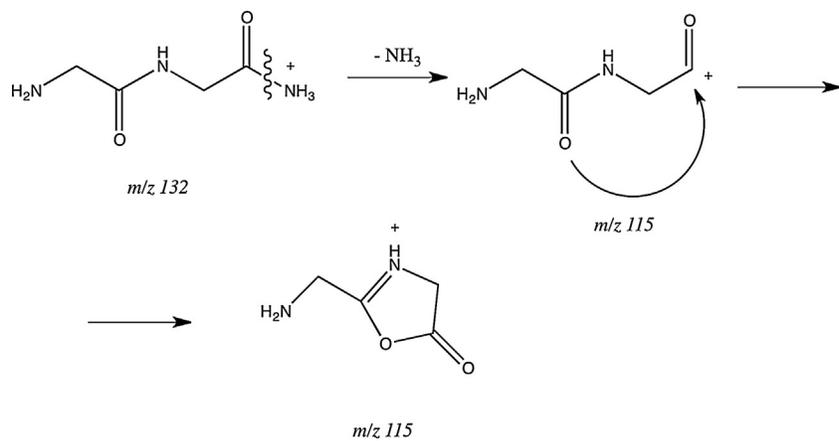


Figure 6.

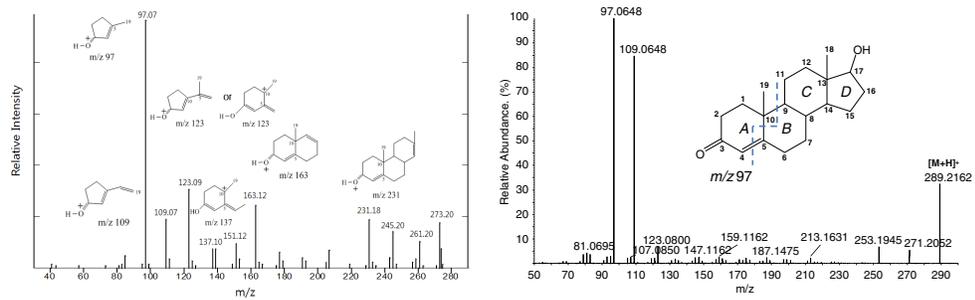


Figure 7.

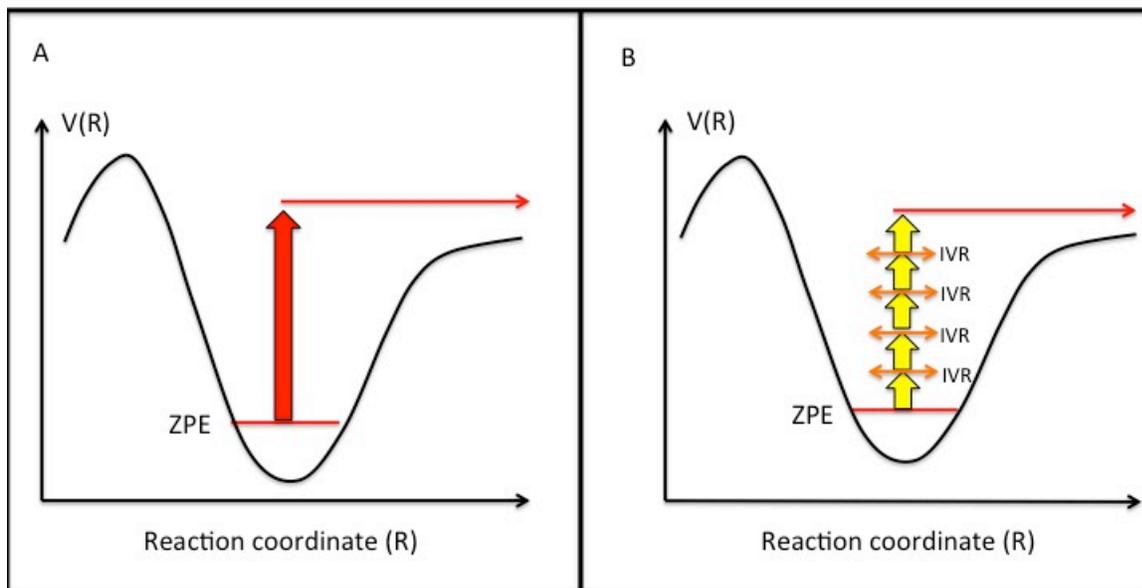
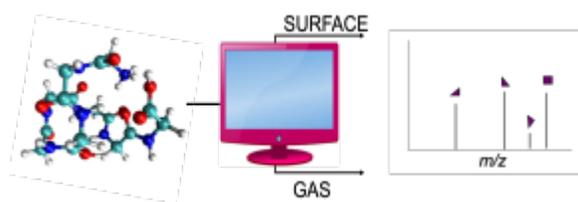


Figure 8.

For Table of Contents Use Only

Role of Chemical Dynamics Simulations in Mass Spectrometry Studies of Collision-Induced Dissociation and Collisions of Biological Ions with Organic Surfaces

Ana Martin Somer, Veronica Macaluso, George L. Barnes, Li Yang, Subha Pratihar, Kihyung Song, William L. Hase, and Riccardo Spezia



Starting from an initial structure of a biological ion, computer simulations can provide fragmentation products as in CID and SID experiments