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High contribution of hydrocarbon transformation during the removal of polycyclic aromatic hydrocarbons from soils, humin and clay by thermal treatment at 100–200 °C

Hanzhong Jia¹ · Jinbo Liu¹ · Kecheng Zhu¹ · Pin Gao² · Eric Lichtfouse³

Abstract

Polycyclic aromatic hydrocarbons (PAHs) are major pollutants in air, soils and sediments. PAH-polluted soils can be cleaned rapidly by thermal treatment. PAH volatilization is considered as the main process explaining PAH removal at low temperature, yet other processes may occur. Particularly, we hypothesize that thermal transformation can also explain PAH removal, where transformation refers to both degradation and formation of bound PAHs. We thus studied the removal of spiked benzo[a]pyrene at 0.5 mg/g in bauxite soil, fluvo-aquic soil, chernozem soil, montmorillonite, humin, and quartz sand as control, from 100 to 200 °C. We measured concentrations of benzo[a]pyrene in the volatilized fraction and solid residues by high-performance liquid chromatography. We identified transformation products by gas chromatography–mass spectrometry. Results show that the contribution of thermal transformation to the removal of benzo[a]pyrene increased from 24.7 to 58.4 wt% for bauxite soil, from 4.4 to 38.2 wt% for fluvo-aquic soil, and from 11.5 to 35.9 wt% for chernozem soil, with temperature increasing from 100 to 200 °C. Transformation such as oxidation occurred in all samples except in benzo[a]pyrene-spiked quartz sands. Transformation of benzo[a]pyrene was thus partly explained by the presence of clay minerals, as evidenced for the montmorillonite assay where transformation contributed 74.6 wt% to the total removal of benzo[a]pyrene at 200 °C. Overall, our findings demonstrate a major overlooked contribution of transformation to PAH removal at low temperature.

Keywords Thermal treatment \cdot Polycyclic aromatic hydrocarbon \cdot Benzo[a]pyrene \cdot Transformation \cdot Volatilization \cdot Clay minerals

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Introduction

Polycyclic aromatic hydrocarbons (PAHs) are a class of priority organic pollutants according to the US Environmental Protection Agency. PAHs induce teratogenic, carcinogenic and mutagenic effects in living organisms (Juhasz and Naidu 2000; Li et al. 2008; Samanta et al. 2002). About 90% of environmental PAHs ultimately end up into soils and sediments, notably due to their excellent hydrophobic property (Eriksson et al. 2000; Field et al. 1992). After entering soils and sediments, PAHs are hardly degraded due to their chemical stability, oxidation resistance and environmental persistence (Jafarabadi et al. 2018; Lichtfouse et al. 1997; Zhang et al. 2015). Therefore, rapid and effective methods are needed to remove PAHs from contaminated soils (Henner et al. 1997).

Remediation techniques for PAH-contaminated soils include chemical oxidation (Si-Hyun et al. 2009), washing

(Trellu et al. 2016), thermal treatment (Bucala et al. 1994; Li et al. 2018) and bioremediation (Gan et al. 2009; Harvey et al. 2002; Wilson and Jones 1993). Thermal treatment is used for fast and efficient remediation (Merino and Bucalá 2007). Two types of thermal treatment are distinguished according to the process involved: desorption at low temperatures of 100-300 °C and degradation at high temperatures of 300-600 °C (Falciglia et al. 2011). High-temperature treatments remove more organic compounds, but require more energy and strongly alter soil physicochemical properties. Alternatively, low-temperature treatment causes minimal damage to soils, requires lower energy, yet still removes organic pollutants efficiently (Falciglia et al. 2011). As a consequence, low-temperature treatment is widely applied to remediate contaminated soils (Gilot et al. 1997; Falciglia et al. 2011; Merino and Bucalá 2007).

The main mechanisms proposed to explain removal of organic pollutants from soils by low-temperature treatment are volatilization and desorption (Falciglia et al. 2011; Gilot et al. 1997; Merino and Bucalá 2007; Talley et al. 2004). Nevertheless, recent investigations suggest that catalyzed oxidation might occur during low-temperature treatment of contaminated soils (Wang et al. 2015; Zhang et al. 2019). For example, Zhang et al. (2019) found that oxidation was a major process during thermal treatment of catechol-contaminated soils. Jia et al. (2019) further reported that minerals can induce oxidation of PAHs even under natural conditions. These observations suggest that oxidative transformation might contribute to PAH removal at low temperature. However, there is actually little quantitative knowledge on the contribution of PAH transformation versus volatilization during thermal treatment of PAH-contaminated soils at low temperature. Therefore, we investigated the transformation and volatilization of benzo[a]pyrene, a carcinogenic PAH, from benzo[a]pyrene-spiked soils to explore the underlying removal mechanisms.

Materials and methods

Chemicals

Benzo[a]pyrene, of 98% purity, HPLC-grade methanol, was purchased from J&K Chemical Ltd., Beijing, China. Dichloromethane and acetone of analytical grade were from Sinopharm Chemical Reagent Co., Shanghai, China. 100 mesh acid-purified quartz sands were from Tianjin Kemiou Chemical Reagent Co., Tianjin, China. SWy-3 montmorillonite was from the Clay Minerals Society, West Lafayette, USA. Humin was extracted from peat according to the method of the International Humic Substances Society (IHSS) (Saab and Martin-Neto 2008).

Soil sample collection and benzo[a]pyrene-spiked sample preparation

Three cultivated soils including bauxite soil, fluvo-aquic soil and chernozem soil were sampled from the 0–20 cm soil surface layer in Shaanxi, Henan and Heilongjiang provinces in China, respectively. Soils were air-dried for 24 h at 25 °C and sieved through 100-mesh prior to analysis.

Benzo[a]pyrene-spiked soil samples were prepared with previously described methods (Jia et al. 2018; Zhao et al. 2017). Samples included three soils: bauxite soil, fluvo-aquic soil and chernozem soil, and pure components: montmorillonite, humic acid and quartz sands. Typically, 1 g of sample was mixed with 1 mL of acetone containing 500 mg/L of benzo[a]pyrene. The sample-benzo[a]pyrene suspensions were mixed for 0.5 h and then placed in the dark at 25 °C until acetone was completely evaporated in about 5 min. The final concentration of benzo[a]pyrene in samples was determined after extracting by a mixture of acetone and dichloromethane (1/1 v/v), and the concentration values were in the range of 0.497–0.502 mg/g. The prepared samples were stored at -4 °C before use.

Low-temperature treatment

Samples contaminated with benzo[a]pyrene were heated using an experimental apparatus (Fig. 1), consisting of an oil bath heater from DragonLAB, MS-H-Pro⁺, China, a two-necked round-bottom flask, two porous gas washing bottles for capturing volatilized-benzo[a]pyrene by a mixture of acetone and dichloromethane (1/1 v/v), and a roundbottomed flask filled with water to absorb the exhaust gas. As the oil bath temperature reached 100, 125, 150, 175 or 200 °C, the two-necked round-bottom flask was put into the oil bath and kept for 60 min. After that, soil samples were collected and cooled down at 25 °C and then stored in the sealed polyethylene bags prior to analysis. The solution in porous gas washing bottles was collected and dried to 5 mL by a rotary evaporator from DragonLAB, RE100-Pro, China, and then analyzed using a U3000 high-performance liquid chromatograph (HPLC) from Thermo Scientific, Germering, Germany. Blanks of non-spiked samples and control of benzo[a]pyrene-spiked quartz sands were thermally treated in parallel as mentioned above. Each treatment was repeated three times. The removal (% Re) including transformed- and volatilized-benzo[a]pyrene are calculated as follows

$$\% \text{Re} = \frac{M_0 - M_f}{M_0} \times 100 = \% \text{Re}_{\text{transformed}} + \% \text{Re}_{\text{volatilized}}$$

where %Re corresponds to the total removal of benzo[a] pyrene after thermal treatment, M_0 and M_f correspond,

Fig. 1 Experimental apparatus for the thermal treatment of benzo[a]pyrene-spiked sand, montmorillonite, humin and soils. A, two-necked roundbottom flask; B and C, porous gas washing bottles; D, roundbottomed flask filled with water. The samples in A was collected and extracted after thermal treatment, and then, the residual benzo[a]pyrene was measured. The volatilized-benzo[a]pyrene was adsorbed by solvents in both **B** and **C** and then mixed for measurement



respectively, to the initial and final extractable amounts of benzo[a]pyrene in the samples, $\[mathcal{Re}_{transformed}\]$ and $\[mathcal{Re}_{volatilized}\]$ correspond, respectively, to the removal of benzo[a]pyrene by transformation and volatilization.

Chemical analysis

Organic carbon (OC), clay and metal contents, pH and cation exchange capacity (CEC) of the collected soil samples were determined with methods previously reported by Droge and Goss (2013). Briefly, the soil OC was measured by a total organic carbon (TOC) analyzer from Elementar vario series, Hesse-Darmstadt, Germany. To analyze metals, soil samples first were fully microwave-digested in aqua regia solutions (Janssen et al. 1997). The resulting digested solutions were measured using inductively coupled plasma atomic emission spectrometer (ICP-AES) from Thermo Scientific iCAP6300, Waltham, USA. For measuring pH, 5 g of soil was mixed with 25 mL of deionized water (1/5 m/v soil to water ratio) for 1 h, and then, the supernatant pH was measured. Cation exchange capacity (CEC) was determined by adding 0.2 M NH₄Cl into the supernatant for substituting exchangeable cations of K⁺, Na⁺, Ca²⁺, Mg²⁺, and Al³⁺ and then analyzed by ICP-AES. Clays in particle-size fractions below 2 mm of the target soils were determined semi-quantitatively by X-ray diffraction analysis (XRD, D8-ADVANCE). Soil textures were analyzed using a hydrometer method (Wen et al. 2002).

After thermal treatment, 50 mg of each sample was sacrificed and extracted by 1 mL of a mixture of acetone and dichloromethane (1/1 v/v) for three cycles. Extracts were collected and mixed and then filtrated using syringes with 0.45 μ m membrane filters. Benzo[a]pyrene in the filtrate was quantified by HPLC equipped with a Cosmosil C18 column (25 cm \times 4.6 mm, 5 μ m particle size). Pure methanol was used as mobile phase in isocratic mode with a flow rate of 1.0 mL/min. The sample injection volume was 10 μ L with an auto-sampler. The oven temperature was kept at 30 °C.

Thermal transformation products were identified with an Agilent 7890B–5977B gas chromatograph–mass spectrometer (GC–MS) by scanning from m/z 40 to m/z 500. A Rtx-5MS capillary column of 30 m×0.25 mm internal diameter (i.d.) and 0.25 μ m film thickness was used, and the He flow rate was 1.0 mL/min. The temperature program for separation started at 80 °C and held for 2 min, set at 10 °C/min to 290 °C and held for 10 min. The inlet temperature was 200 °C, and the detector temperature was 200 °C.

Results and discussion

Amount of removal of benzo[a]pyrene

We studied the transformation and volatilization of benzo[a] pyrene in the benzo[a]pyrene-spiked samples after 60 min at 100–200 °C. Samples included three soils: bauxite soil, fluvo-aquic soil and chernozem soil, and pure components: montmorillonite, humin, and quartz sands. Uncontaminated samples were also run under the same conditions as blanks.

Our results show that the removal of benzo[a]pyrene (%*Re*) in all samples increased with temperature (Fig. 2). For example, the concentration of benzo[a]pyrene in the bauxite soil decreased from 0.501 to 0.362 mg/g at 100 °C for 60 min. At 200 °C, the concentration of benzo[a]pyrene reduced to 0.169 mg/g, resulting in a removal 0.193 mg of benzo[a]pyrene. Similar results were also obtained for fluvo-aquic and chernozem soils, with reduction of



Benzo[a]pyrene removal (mg/g) in heated soils, clay, humin and sand

Fig. 2 Removal of benzo[a]pyrene in bauxite soil, fluvo-aquic soil, chernozem soil, montmorillonite, humin, and quartz sands during thermal treatment from 100 to 200 °C. Gray bars show the initial concentration of spiked benzo[a]pyrene. Blue bars depict the residual concentration of benzo[a]pyrene after treatment. Note the increasing

removal with temperature, the highest removal for pure quartz sands, the high removal for montmorillonite clay and the low removal for humin, a major soil organic component. Those findings reveal the strong interactions of soil components with PAHs, either promoting removal for clays, or inhibiting removal for humin

benzo[a]pyrene of 0.194 mg and 0.155 mg, respectively. These results are consistent with previous reports (Falciglia et al. 2011; Talley et al. 2004).

We observed the highest removal of benzo[a]pyrene in pure quartz sands, reaching 92.2% at 200 °C (Fig. 2). This finding suggests that soil components such as organic matter and minerals display strong interactions with PAHs during thermal treatment. These interactions either promote or inhibit PAH removal. For instance, montmorillonite promotes benzo[a]pyrene removal, with a high decrease of concentration to 0.111 mg/g at 200 °C, in agreement with our previous report on the impact of clays (Jia et al. 2016). On the contrary, results show that humin, a major organic component, inhibits benzo[a]pyrene removal, with a concentration almost unchanged (0.494 mg/g) versus initial concentration (0.503 mg/g), at 100 °C for 60 min. This protecting effect is also observed in fluvo-aquic and chernozem soils.

Overall, our results demonstrate the strong interactions of benzo[a]pyrene with soil components, either promoting removal by clays or inhibiting removal by organic matter. The next section deciphers removal mechanisms, in particular the respective contribution of PAH transformation versus PAH volatilization.

Mechanisms of removal of benzo[a]pyrene

Volatilization

Here, we assume that the removal of benzo[a]pyrene during thermal treatment is explained either by PAH volatilization or PAH transformation, in which transformedbenzo[a]pyrene includes thermal-degraded, e.g., oxidized, and soil-bound benzo[a]pyrene. Results show that proportion of volatilized-benzo[a]pyrene in all samples increased with temperature from 100 to 200 °C (Fig. 3, orange bars). For instance, using cultivated soils, the volatilizedbenzo[a]pyrene accounted, respectively, for 3.1–7.8 wt% in the bauxite soil, 3.1–7.9 wt% in the fluvo-aquic soils, and 3.9-9.9 wt% in the chernozem soil. Similarly, volatilization accounted ti 1.4-3.2 wt% for montmorillonite and 0.6–1.4 wt% for humin, in agreement with a previous report (Karimi-Lotfabad et al. 1996). By contrast, using pure quartz sands, volatilization accounted up to 97.3 wt% of removal. These findings reveal that the interaction of soil components with benzo[a]pyrene strongly reduce volatilization.



Benzo[a]pyrene removal (mg/g) in heated soils, clay, humin and sand

Fig. 3 Contribution of volatilized-benzo[a]pyrene (orange) and transformed-benzo[a]pyrene (green) to its total removal amount (grey) in bauxite soil, fluvo-aquic soil, chernozem soil, montmorillonite, humin and quartz sands during thermal treatment from 100 to 200 °C. Note the high contribution of transformation versus volatilization, except

in quartz sand. This finding reveals the major contribution of soil components, in particular clay and organic matter, to the transformation of benzo[a]pyrene. Transformation refers both to degraded, e.g., oxidized benzo[a]pyrene, and benzo[a]pyrene sequestrated into soil organic matter

Transformation

The concentration of transformed-benzo[a]pyrene increased with temperature and, more importantly, accounted for a much higher proportion of removal (Fig. 3, green bars) compared to volatilization. For example, transformation accounted for 24.7-58.4 wt% in the bauxite soil, 4.4-38.2 wt% in the fluvo-aquic soil, and 11.5-35.9 wt% in the chernozem soil. By contrast, transformation was very low in pure quartz sands. These findings reveal again the strong interactions of benzo[a]pyrene with two soil components, clays and organic matter. Indeed, results for the montmorillonite clay display a 42.7-74.6 wt% contribution of transformation. The clay influence is also supported by the clay content of our soil samples, of 26 wt% for the bauxite soil, 18.2 wt% for the fluvo-aquic soil and 19.3 wt% for the chernozem soil. These observations are supported by our previous studies showing that benzo[a] pyrene interacts with the clay surface via cation- π complexation, then promotes electron transfer followed by oxidation (Jia and Wang 2013; Jia et al. 2016). Alternatively, transformation of benzo[a]pyrene may be triggered by oxidative degradation in the presence of manganese and iron oxide in soil during heating (Umeh et al. 2018), as suggested by the presence of Mn and Fe in our soil samples (Table S1). Overall, minerals such as clays are likely to play a major role in benzo[a]pyrene transformation.

Concerning the role of organic matter, results for humin show up to 43.9 wt% of transformed-benzo[a]pyrene at 200 °C (Fig. 3). This observation is supported by the formation of humin-bound hydrophobic compounds such *n*-alkanes (Lichtfouse et al. 1998a, b) and benzo[a]pyrene (Zeng et al. 2018). PAHs strongly adsorb onto organic matters (Celis et al. 2006), thus enhancing PAH retention in soils and inhibiting the mineral-mediated transformation (Jia et al. 2015).

Overall, our findings show that transformation accounts for a major proportion of benzo[a]pyrene removal in soils, clay and humin, versus volatilization, during thermal treatment at 100-200 °C. Transformation of benzo[a]pyrene is explained by two possible processes, sequestration into soil organic matter as humin-bound compounds and oxidative degradation, the quantitative contribution of which is unknown. We further gained insights on the oxidation process by studying degradation products in the next section.

Mechanisms of thermal degradation of benzo[a] pyrene

chromatography–mass spectrometry (GC–MS). For quartz sands as control assay, benzo[a]pyrene is a major compound (Fig. 4a). This finding is consistent with our results showing that almost all of the spiked benzo[a]pyrene, about 97.3 wt%, in quartz sands was volatilized during thermal treatment at

Further insights on the contribution of thermal degradation to the transformation of benzo[a]pyrene was obtained by gas



Fig. 4 Total ion current and mass spectra of products of thermal treatment of benzo[a]pyrene-spiked samples, for quartz sand (a) and the bauxite soil (b), obtained by gas chromatography-mass spectrometry (GC-MS)

200 °C (see previous section). In contrast, several degradation products were identified in benzo[a]pyrene-spiked soils, i.e., in the bauxite soil, such as benzo[a]pyrene-1,6-dione, benzo[a] pyrene-6,12-dione, formic acid phenyl ester, acetic acid phenyl ester, and phenol (Fig. 4b). Noteworthy, some of those compounds were also observed in the Fenton oxidation of benzo[a] pyrene (Lee and Hosomi 2001). Our findings thus confirm the occurrence of oxidative transformation of benzo[a]pyrene during thermal treatment at 100–200 °C.

Thermal degradation may be induced by electron transfer between benzo[a]pyrene molecules and clay minerals (Jia et al. 2016). This assumption is strengthened by findings showing that benzo[a]pyrene can be adsorbed on clays by complexation of exchangeable cations on mineral surfaces (Celis et al. 2006). Direct electron transfer from benzo[a]pyrene molecules to the active sites of soil minerals may thus occur, producing the electron-deficient intermediates. In turn, these unstable intermediates react with dioxygen to form oxygenated compounds (Zhao et al. 2017). Oxidation is also supported by the reported production of reactive oxygen species (ROS) during thermal treatment (Zhang et al. 2019). ROS may thus react with adsorbed benzo[a]pyrene to generate oxidative cleavage products. As a result, oxidation is likely to contribute to the removal of benzo[a]pyrene by thermal treatment.

Conclusion

We studied the mechanisms for benzo[a]pyrene removal in soils during thermal treatment at 100–200 °C. Results showed that heating temperature and soil components had significant impacts on the volatilization and transformation of benzo[a]pyrene. The contribution of thermal transformation to the removal of benzo[a]pyrene increased with temperature. Clay minerals such as montmorillonite promote the oxidation of benzo[a]pyrene, producing oxygenatedbenzo[a]pyrene and other aromatic compounds. Transformation, rather than volatilization, accounts for a major part of benzo[a]pyrene removal in soils at 100–200 °C.

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References

- Bucala V, Saito H, Howard JB, Peters WA (1994) Thermal treatment of fuel oil-contaminated soils under rapid heating conditions. Environ Sci Technol 28:1801–1807. https://doi.org/10.1021/es000 60a008
- Celis R, Jonge HD, Jonge LWD, Real M, Cornejo J (2006) The role of mineral and organic components in phenanthrene and

dibenzofuran sorption by soil. Eur J Soil Sci 57:308–319. https://doi.org/10.1111/j.1365-2389.2005.00740.x

- Droge STJ, Goss KU (2013) Sorption of organic cations to phyllosilicate clay minerals: CEC-normalization, salt dependency, and the role of electrostatic and hydrophobic effects. Environ Sci Technol 47:14224–14232. https://doi.org/10.1021/es403187w
- Eriksson M, Dalhammar G, Borg-Karlson AK (2000) Biological degradation of selected hydrocarbons in an old PAH/creosote contaminated soil from a gas work site. Appl Microbiol Biotechnol 53:619–626. https://doi.org/10.1007/s002530051667
- Falciglia PP, Giustra MG, Vagliasindi FGA (2011) Low-temperature thermal desorption of diesel polluted soil: influence of temperature and soil texture on contaminant removal kinetics. J Hazard Mater 185:392–400. https://doi.org/10.1016/j.jhazm at.2010.09.046
- Field JA, De Jong E, Feijoo Costa G, de Bont JA (1992) Biodegradation of polycyclic aromatic hydrocarbons by new isolates of white rot fungi. Appl Environ Microbiol 58:2219–2226. https ://doi.org/10.1002/yea.320080813
- Gan S, Lau EV, Ng HK (2009) Remediation of soils contaminated with polycyclic aromatic hydrocarbons (PAHs). J Hazard Mater 172:532–549. https://doi.org/10.1016/j.jhazmat.2009.07.118
- Gilot P, Howard JB, Peters WA (1997) Evaporation phenomena during thermal decontamination of soils. Environ Sci Technol 31:461–466. https://doi.org/10.1021/es960293p
- Harvey P, Campanella B, Castro P, Harms H, Lichtfouse E, Schaeffner A, Smrcek S, Werck-Reichhart D (2002) Phytoremediation of polyaromatic hydrocarbons, anilines and phenols. Environ Sci Pollut Res 9:29–47. https://doi.org/10.1007/BF02987315
- Henner P, Schiavon M, Morel JL, Lichtfouse E (1997) Polycyclic aromatic hydrocarbon (PAH) occurrence and remediation methods. Analusis Mag 25:M56–M59
- Jafarabadi AR, Bakhtiari AR, Aliabadian M, Laetitia H, Toosi AS, Yap CK (2018) First report of bioaccumulation and bioconcentration of aliphatic hydrocarbons (AHs) and persistent organic pollutants (PAHs, PCBs and PCNs) and their effects on alcyonacea and scleractinian corals and their endosymbiotic algae from the Persian Gulf, Iran: inter and intra-species differences. Sci Total Environ 627:141–157. https://doi.org/10.1016/j.scito tenv.2018.01.185
- Janssen RP, Peijnenburg WJ, Posthuma L, Van Den Hoop MA (1997) Equilibrium partitioning of heavy metals in Dutch field soils. I. Relationship between metal partition coefficients and soil characteristics. Environ Toxicol Chem 16:2470–2478. https://doi. org/10.1002/etc.5620161206
- Jia H, Wang C (2013) Comparative studies on montmorillonite-supported zero-valent iron nanoparticles produced by different methods: reactivity and stability. Environ Technol 34:25–33. https:// doi.org/10.1080/09593330.2012.679698
- Jia H, Chen H, Nulaji G, Li X, Wang C (2015) Effect of low-molecular-weight organic acids on photo-degradation of phenanthrene catalyzed by Fe(III)-smectite under visible light. Chemosphere 138:266–271. https://doi.org/10.1016/j.chemosphere.2015.05.076
- Jia H, Nulaji G, Gao H, Wang F, Zhu Y, Wang C (2016) Formation and stabilization of environmentally persistent free radicals induced by the interaction of anthracene with Fe(III)-modified clays. Environ Sci Technol 50:6310–6319. https://doi.org/10.1021/acs.est.6b005 27
- Jia H, Zhao S, Shi Y, Zhu L, Wang C, Sharma VK (2018) Transformation of polycyclic aromatic hydrocarbons and formation of environmentally persistent free radicals on modified montmorillonite: the role of surface metal ions and polycyclic aromatic hydrocarbon molecular properties. Environ Sci Technol 52:5725–5733. https://doi.org/10.1021/acs.est.8b00425
- Jia H, Zhao S, Shi Y, Fan X, Wang T (2019) Formation of environmentally persistent free radicals during the transformation of

anthracene in different soils: roles of soil characteristics and ambient conditions. J Hazard Mater 362:214–223. https://doi. org/10.1016/j.jhazmat.2018.08.056

- Juhasz AL, Naidu R (2000) Bioremediation of high molecular weight polycyclic aromatic hydrocarbons: a review of the microbial degradation of benzo[a]pyrene. Int Biodeterior Biodegrad 45:57–88. https://doi.org/10.1016/s0964-8305(00)00052-4
- Karimi-Lotfabad S, Pickard MA, Gray MR (1996) Reactions of polynuclear aromatic hydrocarbons on soil. Environ Sci Technol 30:1145–1151. https://doi.org/10.1021/es950365x
- Lee BD, Hosomi M (2001) Fenton oxidation of ethanol-washed distillation-concentrated benzo(A)pyrene: reaction product identification and biodegradability. Water Res 35:2314–2319. https://doi. org/10.1016/S0043-1354(00)00485-1
- Li X, Li P, Lin X, Zhang C, Li Q, Gong Z (2008) Biodegradation of aged polycyclic aromatic hydrocarbons (PAHs) by microbial consortia in soil and slurry phases. J Hazard Mater 150:21–26. https://doi.org/10.1016/j.jhazmat.2009.07.044
- Li DC, Xu WF, Mu Y, Yu HQ, Crittenden JC (2018) Remediation of petroleum-contaminated soil and simultaneous recovery of oil by fast pyrolysis. Environ Sci Technol 52:5330–5338. https://doi. org/10.1021/acs.est.7b03899
- Lichtfouse E, Budzinski H, Garrigues P, Eglinton T (1997) Ancient polycyclic aromatic hydrocarbons in modern soils: ¹³C, ¹⁴C and biomarker evidence. Org Geochem 26:353–359. https://doi. org/10.1016/S0146-6380(97)00009-0
- Lichtfouse E, Leblond C, Da Silva M, Behar F (1998a) Occurrence of biomarkers and straight-chain biopolymers in humin: implication for the origin of soil organic matter. Naturwissenschaften 85:497–501. https://doi.org/10.1007/s001140050538
- Lichtfouse E, Wehrung P, Albrecht P (1998b) Plant wax n-alkanes trapped in soil humin by non-covalent bonds. Naturwissenschaften 85:449–452. https://doi.org/10.1007/s001140050529
- Merino J, Bucalá V (2007) Effect of temperature on the release of hexadecane from soil by thermal treatment. J Hazard Mater 143:455– 461. https://doi.org/10.1016/j.jhazmat.2006.09.050
- Saab SC, Martin-Neto L (2008) Characterization by electron paramagnetic resonance of organic matter in whole soil (Gleysoil) and organic-mineral fractions. J Braz Chem Soc 19:413–417. https:// doi.org/10.1590/S0103-50532008000300007
- Samanta SK, Singh OV, Jain RK (2002) Polycyclic aromatic hydrocarbons: environmental pollution and bioremediation. Trends Biotechnol 20:243–248. https://doi.org/10.1016/S0167 -7799(02)01943-1
- Si-Hyun D, Jeong-Hwan J, Young-Hoon J, Hong-Kyun H, Sung-Ho K (2009) Application of a peroxymonosulfate/cobalt (PMS/Co(II)) system to treat diesel-contaminated soil. Chemosphere 77:1127– 1131. https://doi.org/10.1016/j.chemosphere.2009.08.061
- Talley JW, Ghosh U, Furey JS, Tucker SG, Luthy RG (2004) Thermal program desorption mass spectrometry of PAHs from mineral

and organic surfaces. Environ Eng Sci 21:647–660. https://doi. org/10.1089/ees.2004.21.647

- Trellu C, Mousset E, Pechaud Y, Huguenot D, van Hullebusch ED, Esposito G, Oturan MA (2016) Removal of hydrophobic organic pollutants from soil washing/flushing solutions: a critical review. J Hazard Mater 306:149–174. https://doi.org/10.1016/j.jhazm at.2015.12.008
- Umeh AC, Duan L, Naidu R (2018) Time-dependent remobilization of nonextractable benzo [a] pyrene residues in contrasting soils: effects of aging, spiked concentration, and soil properties. Environ Sci Technol 52:12295–12305. https://doi.org/10.1021/acs. est.8b03008
- Wang Z, Zang L, Fan X, Jia H, Li L, Deng W, Wang C (2015) Defectmediated of Cu@TiO₂ core-shell nanoparticles with oxygen vacancies for photocatalytic degradation 2,4-DCP under visible light irradiation. Appl Surf Sci 358:479–484. https://doi. org/10.1016/j.apsusc.2015.08.051
- Wen B, Aydin A, Duzgoren-Aydin NS (2002) A comparative study of particle size analyses by sieve-hydrometer and laser diffraction methods. Geotech Test J 25:34–442. https://doi.org/10.1520/ GTJ11289J
- Wilson SC, Jones KC (1993) Bioremediation of soil contaminated with polynuclear aromatic hydrocarbons (PAHs): a review. Environ Pollut 81:229–249. https://doi.org/10.1016/0269-7491(93)90206 -4
- Zeng J, Zhu Q, Wu Y (2018) Oxidation of benzo[a]pyrene by laccase in soil enhances bound residue formation and reduces disturbance to soil bacterial community composition. Environ Pollut 242:462– 469. https://doi.org/10.1016/j.envpol.2018.06.075
- Zhang L, Cao Y, Colella NS, Liang Y, BréDas J-L, Houk KN, Briseno AL (2015) Unconventional, chemically stable, and soluble twodimensional angular polycyclic aromatic hydrocarbons: from molecular design to device applications. Acc Chem Res 48:500– 509. https://doi.org/10.1021/ar500278w
- Zhang Y, Guo X, Si X, Yang R, Zhou J, Quan X (2019) Environmentally persistent free radical generation on contaminated soil and their potential biotoxicity to luminous bacteria. Sci Total Environ 687:348–354. https://doi.org/10.1016/j.scitotenv.2019.06.137
- Zhao S, Jia H, Nulaji G, Gao H, Wang F, Wang C (2017) Photolysis of polycyclic aromatic hydrocarbons (PAHs) on Fe³⁺-montmorillonite surface under visible light: degradation kinetics, mechanism, and toxicity assessments. Chemosphere 184:1346–1354. https://doi.org/10.1016/j.chemospher e.2017.06.106

SUPPLEMENTARY MATERIAL

High contribution of hydrocarbon transformation during the removal of polycyclic aromatic hydrocarbons from soils, humin and clay by thermal treatment at 100-200 $^{\circ}$ C

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Table S1

Physicochemical properties of bauxite soil, fluvo-aquic soil, chernozem soil,

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Soil			Sample		
characteristics	Bauxite	Fluvo-aquic soil	Chernozem	Montmorillonite	Humin
	Shannxi	Henan	Heilongjiang		
Location	34°11′N,	35°00′N,	45°40′N,		
	113°01′E	113°41′E	126°37′E		
рН	7.90	8.07	6.27	6.54	7.00
OC [*] , wt%	0.96	1.03	2.07		7.89
Clay, wt%	26.01	18.18	19.33	100.00	
CEC**, cmol/kg	22.37	16.01	28.59		
Total Fe, g/kg	17.13	20.76	25.56	12.80	3.60
Total Mn, g/kg	0.37	0.46	0.63	0.09	0.07
Total Cu, g/kg	/	/	/	/	/
Total Zn, g/kg	0.06	0.07	0.13	0.09	0.03
Total Pb, g/kg	/	/	0.09	/	0.02
Total Mg, g/kg	5.72	7.98	5.20	7.40	0.30
Total Ca, g/kg	90.9	141.3	23.2	24.5	7.1

montmorillonite, and humin.

* OC: organic carbon.

**CEC: cation exchange capacity.