

Resistance to third-generation cephalosporins in Escherichia coli in the French community: The times they are a-changin'?

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Audrey Jeanvoine, Hélène Bouxom, Joel Leroy, Houssein Gbaguidi-Haore, Xavier Bertrand, et al.. Resistance to third-generation cephalosporins in Escherichia coli in the French community: The times they are a-changin'?. International Journal of Antimicrobial Agents, 2020, pp.105909. 10.1016/j.ijantimicag.2020.105909. hal-02541479

HAL Id: hal-02541479 https://hal.science/hal-02541479v1

Submitted on 22 Aug 2022

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Resistance to third-generation cephalosporins in Escherichia coli in the French

2	community, The Times They Are A-Changin'?
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24	Running title: Resistance of <i>E. coli</i> to third-generation cephalosporins in outpatients

25 **Abstract**

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26	Introduction. Since the early 2000s, E. coli resistance to third-generation cephalosporins
27	(3GCs) has been increasing in all European countries, mainly due to the spread of extended

spectrum β -lactamases (ESBL). We present here a retrospective study that combines

resistance of E. coli to 3GCs and quinolones and the antibiotic use in the community in a

region of Northeastern France.

31 Materials and Methods. Since 2012, we conducted an observational surveillance of

32 antimicrobial resistance and antibiotic use in the community: data of antibiotic resistance in

E. coli isolates were collected from 11 private laboratories, and consumption data from the

three main healthcare insurances.

35 **Results.** A significant decrease of resistance prevalence to 3GCs (from 5.6 to 3.9%, p <

0.001), nalidixic acid (16.7 to 14.8%, p = 0.004) and ciprofloxacin (10.9 to 8.1%, p < 0.001)

was reported between 2015 and 2017. Although the total antibiotic consumption did not

vary significantly between 2012 and 2017, we observed a decrease of the consumption of

3GCs (-32%, p < 0.001) and fluoroquinolones (-25%, p < 0.001).

40 **Conclusion.** We report here a decrease of the prevalence of *E. coli* isolates resistant to 3GCs

and quinolones in outpatients in a context of significant decreasing consumptions of these

two antibiotic classes.

44 **Keywords:** *E. coli*; third-generation cephalosporins; quinolones; resistance; outpatients.

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1. Introduction

High rates of antimicrobial resistance in *Escherichia coli* are an important global public health concern [1]. Indeed, *E. coli* constitutes a major cause of both community-onset and hospital-acquired infections [2]. Since the early 2000s, *E. coli* resistance to third-generation cephalosporins (3GCs) has been increasing in hospital and community settings of all European countries, mainly due to the spread of extended spectrum β -lactamases (ESBL) [1,3,4]. In 2017, EARS-net report showed that the rate of resistance of *E. coli* to 3GCs reached 14.9 % in bacteremias in hospital setting and upward trends were observed in almost all European countries in the last decade [1]. Besides these hospital data, several surveillances have also reported an increasing trend of resistance to 3GCs in *E. coli* from the community [5,6]. This spread combines dissemination of epidemic clones such as ST131 - C1/*H*30-R and C2/*H*30-Rx clades, as well as horizontal transfers of plasmid-borne bla_{ESBL} [7]. The possibility that ESBL rates reach that of penicillinase within this species is clearly a matter of major concern.

ESBL-producing *E. coli* isolates are often resistant to other antibiotics such as fluoroquinolones and aminoglycosides, further complicating the management of patients infected with ESBL-producing *Enterobacteriaceae* [6,7]. Indeed, combined resistance to 3GCs, fluoroquinolones, and aminoglycosides was 6.3 % in 2017 in the EU, but did not significantly change during the period 2014–2017 [1]. Moreover, up to 70 % of ESBL-producing isolates were resistant to fluoroquinolones in community-onset infections [6]. The spread of ESBL-producing *E. coli* within the community is mainly driven by antimicrobial exposure, notably previous use of fluoroquinolones and oral 3GCs, which have a high ecological impact [8]. In that context, we implemented in our region an antimicrobial resistance stewardship program that monitored the antibiotic use in outpatients.

We present here a retrospective study that combines the resistance of *E. coli* to 3GCs and quinolones and the antibiotic use in the community in a region of Northeastern France.

2. Materials and methods

Since 2012, we conducted an observational surveillance (the OSCAR network) of antimicrobial resistance and antibiotic use in the community in Franche-Comté region, a Northeastern area of France with 1.2 million inhabitants.

3GCs and quinolones (nalidixic acid and ciprofloxacin) resistance data were collected during six years (2012-2017) from eleven private laboratories. All resistance data concerning nursing homes or clinics were excluded. For one patient, one isolate per susceptibility profile and per year was considered.

Antimicrobial consumption data were provided by the three main healthcare insurances of Franche-Comté and their beneficiaries represented 88% of the total population. These antibiotic reimbursement data for ambulatory care, based on antibiotics dispensed in community pharmacies, were available since 2012. The consumption data were expressed in defined daily dose (DDD) per 1000 beneficiaries per day for each antibiotic for systemic use (ATC group J01): tetracyclines (J01A), ß-lactams (J01C and J01D), sulfamide and trimethoprim (J01E), macrolides (J01F), aminoglycosides (J01G), fluoroquinolones (J01M) and associations and other antibiotics (J01R+J01X) [9].

We tested the variation in the prevalence of *E. coli* isolates resistant to 3GCs, nalidixic acid, and ciprofloxacin, using the χ^2 test. The trend of consumption of each antibiotic class over the period 2012 to 2017 was tested by using a negative binomial model for which the DDD were considered as counting data. The relative change, also calculated here, consists of the variation of consumption of each antibiotic class between the two years 2012 and 2017.

3. Results

3.1 Antimicrobial resistance in E. coli

A total of 34,767 *E. coli* isolates have been tested for their susceptibility between 2012 and 2017. Trend in resistance prevalence of *E. coli* to 3GCs, nalidixic acid, and ciprofloxacin, are shown in Figure 1a. Resistance rate for 3CGs was 3.9% in 2012, reached 5.9% in 2015 and significantly decreased between 2015 and 2017, from 5.9 to 4.2% (p < 0.001). Although the evolution of the prevalence of resistance to nalidixic acid differed from that to 3GCs and ciprofloxacin between 2012 and 2015, a significant decrease of the prevalence of resistance to quinolones was also reported between 2015 and 2017, from 16.7 to 14.8% for nalidixic acid (p = 0.004) and from 10.9 to 8.1% for ciprofloxacin (p < 0.001). Among *E. coli* isolates resistant to 3GCs collected between 2012 and 2017, the prevalence of nalidixic acid and ciprofloxacin resistance decreased in parallel between 2015 and 2017, from 73.8 to 64.2%, and from 66.8 to 52.9%, respectively (Figure 1b).

3.2 Antibiotics consumption

Although the total antibiotic consumption did not vary significantly between 2012 and 2017, the use of penicillins (J01C), and that of the broad-spectrum penicillins (J01CA) such as amoxicillin (J01CA04), increased significantly (p < 0.001) over the study period (Table 1). Except for tetracyclines (J01A) and associations and other antibiotics (J01R+J01X), we found a significant downward trend of consumption for the others studied group of antibiotics (Table 1). We notably observed a decrease of the 3GCs (J01DD, -32.1%, p < 0.001) and fluoroquinolones consumption (J01M, -25.5%, p < 0.001) over the study period (Table 1, Figure 1a).

4. Discussion

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After several years of constant rise, we report here a significant decrease of the prevalence of E. coli resistant to 3GCs and quinolones in outpatients from Franche-Comté between 2015 and 2017. Regarding the resistance to 3GCs, our results confirm those collected by another French surveillance network (Medqual) which nevertheless reported a lower decrease between 2015 and 2017 (from 4.5% in 2015 to 4% in 2017 versus 5.9% to 4.2% in Franche-Comté) [10]. These trends observed in the community coincided with those recorded in EARS-net database, the national survey of *E. coli* isolated from blood culture in French hospitals (11% in 2015 to 10.2% in 2017) [1]. Although this national downward trend was not significant, the decrease reported in Franche-Comté could be more general in France. Concerning fluoroquinolones, the decrease of resistance reported here was also highlighted by EARS-net data in France (from 17.7% in 2015 to 15% in 2017) [1]. The same trend was described by the Medqual network in community-onset infections for nalidixic acid. However, the decrease of the resistance rate of ciprofloxacin in *E. coli* was less pronounced, from 10.1% in 2015 to 9.6% in 2017 versus 10.9% to 8.1% in Franche-Comté [10]. According to other studies, we described here high resistance rates of quinolones in isolates resistant to 3GCs [6,11].

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Interestingly, the decrease of the prevalence of *E. coli* resistant to 3GCs in community-onset infections reported here occurred in a context of a significant decrease of the consumption of 3GCs and fluoroquinolones, despite a significant increase of penicillins consumption. Many studies have described the high-ecological impact of 3GCs and fluoroquinolones. For instance, recent phylogenic studies using whole genome sequencing

identified the most recent common ancestor of successful epidemic clones, thus strongly suggesting that fluoroquinolones had played an important role in the global expansion of the major pathogens *Escherichia coli* ST131 H30-Rx, methicillin-resistant *Staphylococcus aureus* EMRSA-15, *Pseudomonas aeruginosa* ST235 in the mid-1980s, and that of *Clostridium difficile* 027 in the early 1990s [12–15].

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Although this is not statistically demonstrated, the decrease of the use of fluoroquinolones and 3GCs, which began in 2013 in our region, presumably played a major role in the trend of resistance to 3GCs among community E. coli isolates. However, the reducing consumption of antibiotics is not immediately followed by the decrease of antibiotic resistance. Some studies reported that once the resistance to an antimicrobial is well established in a population of E. coli, a reduction in the use of that antimicrobial is unlikely to influence to resistance rates in the short-to-medium term [2,16]. This could be explained by two effects. The first one may be due to the prolonged carriage of multidrugresistant E. coli even in the absence of antimicrobial selection pressure, described for E. coli ST131 [17]. Hence, it may take several years to achieve a sufficient patient turnover and to observe the effect of antibiotic use reduction [16]. The second one is the nonlinear relationship between the antibiotic use and resistance; other parameters could be involved such as dissemination of resistant strains by cross-transmission [18]. A recent study has calculated antibiotic consumption thresholds below which the resistance rates are independent from the consumption. For instance, minimum thresholds were estimated for selected antibiotics and the rates of ESBL-producing or cefepime-resistant E. coli [18].

This study has several limitations. First, our data did not establish a significant relationship between the decrease of antibiotic resistance rate and their consumption. Then, we missed the genotyping data that could have allowed the understanding of the decrease of resistance reported here. However, we could speculate that the decrease in prevalence of the major multidrug-resistant clone *E. coli* ST131 H30-Rx participated, at least in part, to the decrease of the resistance in the species. OSCAR network surveillance must be continued to confirm the trend in resistance prevalence and antibiotics consumption reported here.

179	We report here a decrease of the prevalence of <i>E. coli</i> isolates resistant to 3GCs and to
180	quinolones in a context of decreasing consumptions of these two antibiotic classes. The
181	stewardship programs can efficiently decrease the antimicrobial resistance, making the
182	progression to multi-resistance not inevitable. However, actions improving antimicrobial
183	prescription should be continuous because of their delayed effect.
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185	Acknowledgments
186	We thank the private laboratory group CBM25, Centre de Biologie médicale, who enabled
187	the use of its antibiotic resistance data.
188	
189	Declarations
190	Funding. This study received a partial grant from the Regional Health Agency (ARS
191	Bourgogne-Franche-Comté).
192	Competing Interests: None
193	Ethical Approval: Not required
194	
195	Author contributions. Conceived and designed the experiments: XB, CBS, and JL. Performed
196	the experiments: CBS, HB, and HGH. Analyzed the data: CBS, HB, AJ, and HGH. Wrote the
197	paper: AJ, CBS, and XB.
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5. Conclusion

200 **References**

- 201 [1] European centre for disease prevention and control. Surveillance of antimicrobial
- resistance in Europe 2017, https://www.ecdc.europa.eu/en/publications-
- 203 data/surveillance-antimicrobial-resistance-europe-2017; 2017 [accessed 19 October
- 204 2019].
- 205 [2] Stapleton PJ, Lundon DJ, McWade R, Scanlon N, Hannan MM, O'Kelly F, et al. Antibiotic
- resistance patterns of *Escherichia coli* urinary isolates and comparison with antibiotic
- 207 consumption data over 10 years, 2005–2014. Ir J Med Sci 2017; 186:733–41.
- 208 https://doi.org/10.1007/s11845-016-1538-z.
- 209 [3] Cantón R, Novais A, Valverde A, Machado E, Peixe L, Baquero F, et al. Prevalence and
- spread of extended-spectrum β-lactamase-producing *Enterobacteriaceae* in Europe.
- 211 Clin Microbiol Infect 2008; 14:144–53. https://doi.org/10.1111/j.1469-
- 212 0691.2007.01850.x.
- 213 [4] Hawser SP, Bouchillon SK, Hoban DJ, Badal RE, Canton R, Baquero F. Incidence and
- antimicrobial susceptibility of Escherichia coli and Klebsiella pneumoniae with
- extended-spectrum-lactamases in community- and hospital-associated intra-abdominal
- infections in Europe: results of the 2008 study for monitoring antimicrobial resistance
- trends (SMART). Antimicrob Agents Chemother 2010; 54:3043–6.
- 218 https://doi.org/10.1128/AAC.00265-10.
- 219 [5] Boix-Palop L, Xercavins M, Badía C, Obradors M, Riera M, Freixas N, et al. Emerging
- 220 extended-spectrum β-lactamase-producing Klebsiella pneumoniae causing community-
- onset urinary tract infections: a case—control—control study. Int J Antimicrob Agents
- 222 2017; 50:197–202. https://doi.org/10.1016/j.ijantimicag.2017.03.009.

- 223 [6] Chervet D, Lortholary O, Zahar JR, Dufougeray A, Pilmis B, Partouche H. Antimicrobial 224 resistance in community-acquired urinary tract infections in Paris in 2015. Med Mal
- 225 Infect 2018; 48:188–92. https://doi.org/10.1016/j.medmal.2017.09.013.
- 226 [7] Stoesser N, Sheppard AE, Pankhurst L, De Maio N, Moore CE, Sebra R, et al.
- 227 Evolutionary history of the global emergence of the *Escherichia coli* epidemic clone
- 228 ST131. MBio 2016; 7:e02162. https://doi.org/10.1128/mBio.02162-15.
- [8] Paterson DL. "Collateral Damage" from cephalosporin or quinolone antibiotic therapy.
- 230 Clin Infect Dis 2004; 38:S341–5. https://doi.org/10.1086/382690.
- 231 [9] World Health Organization. Anatomical Therapeutic Chemical (ATC) classification index
- with defined daily doses (DDDs),
- 233 http://apps.who.int/medicinedocs/en/d/Js2202e/5.html, Norway; 2001 [accessed 29
- 234 October 2019].
- 235 [10] ONERBA. 2017 Rapport d'activité : Annual report,
- http://onerba.org/publications/rapports-onerba/; 2018 [accessed 29 Ocotober 2019].
- 237 [11] Azargun R, Sadeghi MR, Soroush Barhaghi MH, Samadi Kafil H, Yeganeh F, Ahangar
- Oskouee M, et al. The prevalence of plasmid-mediated quinolone resistance and ESBL-
- 239 production in *Enterobacteriaceae* isolated from urinary tract infections. Infect Drug
- 240 Resist 2018; 11:1007–14. https://doi.org/10.2147/IDR.S160720.
- [12] Ben Zakour NL, Alsheikh Hussain AS, Ashcroft MM, Khanh Nhu NT, Roberts LW, Stanton
- 242 Cook M, et al. Sequential acquisition of virulence and fluoroquinolone resistance has
- shaped the evolution of *Escherichia coli* ST131. MBio 2016; 7:e00347-16.
- 244 https://doi.org/10.1128/mBio.00347-16.

245 [13] He M, Miyajima F, Roberts P, Ellison L, Pickard DJ, Martin MJ, et al. Emergence and 246 global spread of epidemic healthcare-associated Clostridium difficile. Nat Genet 2013; 45:109–13. https://doi.org/10.1038/ng.2478. 247 248 [14] Holden MT, Hsu LY, Kurt K, Weinert LA, Mather AE, Harris SR, et al. A genomic portrait 249 of the emergence, evolution, and global spread of a methicillin-resistant 250 Staphylococcus aureus pandemic. Genome Res 2013; 23:653-64. 251 https://doi.org/10.1101/gr.147710.112. 252 [15] Treepong P, Kos VN, Guyeux C, Blanc DS, Bertrand X, Valot B, et al. Global emergence of the widespread Pseudomonas aeruginosa ST235 clone. Clin Microbiol Infect 2018; 253 24:258–66. https://doi.org/10.1016/j.cmi.2017.06.018. 254 255 [16] Muller A, Romand K, Suplisson D, Nattero JF, Leroy J, Bertrand X. Impact of a long-term 256 antibiotic stewardship program targeting fluoroquinolones in a French local hospital. Med Mal Infect 2019; 49:442-6. https://doi.org/10.1016/j.medmal.2018.10.006. 257 [17] Overdevest I, Haverkate M, Veenemans J, Hendriks Y, Verhulst C, Mulders A, et al. 258 Prolonged colonisation with Escherichia coli O25:ST131 versus other extended-259 260 spectrum beta-lactamase-producing E. coli in a long-term care facility with high 261 endemic level of rectal colonisation, the Netherlands, 2013 to 2014. Eurosurveillance 2016; 21. https://doi.org/10.2807/1560-7917.ES.2016.21.42.30376. 262 263 [18] López-Lozano JM, Lawes T, Nebot C, Beyaert A, Bertrand X, Hocquet D, et al. A nonlinear time-series analysis approach to identify thresholds in associations between 264 population antibiotic use and rates of resistance. Nat Microbiol 2019; 4:1160-72. 265 https://doi.org/10.1038/s41564-019-0410-0. 266

Figure captions

Figure 1. Resistance prevalence of *E. coli* isolated from community-onset infections to third-generation cephalosporins and quinolones between 2012 and 2017 in Franche-Comté (Northeastern France). **(a)** Trend in resistance rate of *E. coli* isolated from community-onset infections to third-generation cephalosporins (3GCs), nalidixic acid (NA) and ciprofloxacin (CIP) associated to 3GCs and quinolones consumption between 2012 and 2017 in the Northeastern France. Prevalence of resistance to 3GCs, NA and CIP are represented by dark grey, light grey and black lines, respectively. The prevalence of *E. coli* resistant to these antibiotics decreased between 2015 and 2017 (χ^2 test). Consumptions of 3GCs and fluoroquinolones (FQs) are represented by dark grey and light grey bars, respectively. **(b)** Trend in resistance rate to nalidix acid (NA) and ciprofloxacin (CIP) of *E. coli* isolates resistant to third-generation cephalosporins (3GCs-R) between 2012 and 2017 in the Northeastern France. The bars represent the number of *E. coli* isolates 3GCs-R between 2012 and 2017. Light grey and dark grey lines represent the percentage of 3GCs-R *E. coli* isolates resistant to NA and CIP, respectively.

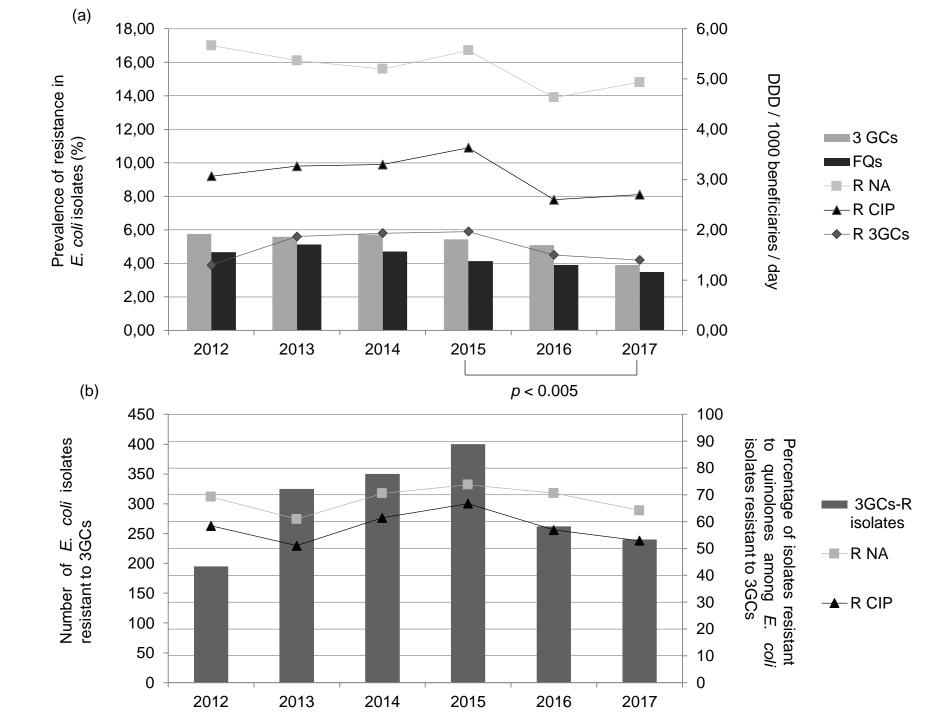


Table 1: Consumption of antibiotics for the treatment of community-onset infections in the Northeastern France between 2012 and 2017.

Antibiotic class	Antibiotic consumption (DDD / 1000 beneficiaries / day)						Relative change ¹	Trend² (p)
	2012	2013	2014	2015	2016	2017	(%)	
J01A - tetracyclines	2.57	2.93	2.64	2.47	2.61	2.52	- 1.8	0.208
J01C - β-lactams-penicillins	14.93	18.53	18.07	18.19	19.28	18.47	23.7	0.020*
J01CA - broad spectrum penicillins	7.92	10.38	10.25	10.68	11.69	11.68	47.5	< 0.001*
J01CR - penicillins associations	6.40	7.52	7.18	7.04	7.19	6.39	- 0,20	0.720
J01CR02 - amoxicillin-clavulanic acid	NA	NA	NA	7.03	7.18	6.38	NC	NC
J01D - other β-lactams	2.73	2.77	2.64	2.42	2.24	1.75	- 35.9	0.001*
J01DB - first-generation cephalosporins	0.22	0.035	0.16	0.015	0.015	0.007	- 96.8	< 0.001*
JO1DC - second-generation cephalosporins	0.59	0.87	0.56	0.59	0.52	0.43	- 26.8	0.011*
J01DD - third-generation cephalosporins	1.92	1.86	1.91	1.81	1.70	1.30	- 32.1	0.001*
intravenous 3GCs	0.12	0.15	0.14	0.15	0.14	0.13	13.6	0.327
per os 3GCs	1.80	1.71	1.77	1.66	1.56	1.17	- 35.1	0.001*
J01E- sulfamide and trimethoprim	0.67	0.81	0.77	0.37	0.36	0.35	- 47.3	0.001*
J01F-macrolides	3.47	3.87	3.41	3.49	3.46	3.08	- 11.4	0.027*
J01FG01-pristinamycin	NA	NA	NA	0.98	0.90	0.90	NC	NC
J01G- aminoglycosides	0.04	0.042	0.035	0.033	0.031	0.025	- 36.4	< 0.001*
J01M- fluoroquinolones	1.56	1.71	1.57	1.38	1.30	1.16	- 25.5	< 0.001*
intravenous fluoroquinolones	0.00051	0.0007	0.00027	0.00049	0.00013	0.00052	- 3.0	0.438
per os fluoroquinolones	1.56	1.71	1.57	1.38	1.30	1.16	- 25.5	< 0.001*
J01R+J01X - Associations and other antibiotics	0.52	1.05	0.98	1.0	0.96	0.87	67.7	0.290
Total	26.91	31.70	30.10	29.35	30.24	28.24	4.9	0.905

NA: not available, NC: not calculated, *: significant trend (p < 0.05)

¹Relative change consists of the variation of consumption of each antibiotic class between the two years 2012 and 2017.

²The trend of consumption of each antibiotic class was determined over the period 2012 to 2017 by using a negative binomial model