



HAL
open science

Lipopolythiourea Transfecting Agents: Lysine Thiourea Derivatives

Jeanne Leblond, Nathalie Mignet, Celine Largeau, Johanne Seguin, Daniel Scherman, Jean Herscovici

► **To cite this version:**

Jeanne Leblond, Nathalie Mignet, Celine Largeau, Johanne Seguin, Daniel Scherman, et al.. Lipopolythiourea Transfecting Agents: Lysine Thiourea Derivatives. *Bioconjugate Chemistry*, 2007, <10.1021/bc7001924>. <hal-02512550>

HAL Id: hal-02512550

<https://hal.science/hal-02512550v1>

Submitted on 23 Mar 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



HAL Authorization

Lipopolythiourea transfecting agents: lysine thiourea derivatives.

Jeanne Leblond, Nathalie Mignet, Céline Largeau, Johanne Seguin, Daniel Scherman and Jean Herscovici**

Inserm, U640, Paris, F-75006 France; CNRS, UMR8151, Paris, F-75006 France; Ecole Nationale Supérieure de Chimie de Paris, Paris, F-75005; Unité de Pharmacologie Chimique et Génétique, France; Université Paris-Descartes, Faculté de Pharmacie, Paris, F-75270 France; Chimie Moléculaire de Paris Centre, CNRS, FR 2769, Paris, F-75005.

* Nathalie.Mignet@univ-paris5.fr; jean-herscovici@enscp.fr

TITLE RUNNING HEAD: Lipopolythioureas: new noncationic vector for gene transfer

ABSTRACT: Synthetic vectors represent an alternative to recombinant viruses for gene transfer. We have recently explored the transfection potential of a class of noncationic lipids bearing thiourea moieties as DNA associating headgroups. The encouraging results obtained with lipopolythioureas derived from serinol prompted us to investigate further this family of vectors. In the present study, we considered the transfection properties of a series of derivatives based on a different thiourea polar headgroup bearing a lysine scaffold. The synthesis of these compounds could be readily achieved in 3 steps with good yields. We found that these lipopolythioureas (LPT) might be considered as alternative systems for gene transfection, since their activity reached the same magnitude range as cationic vectors in the presence of serum. LPT with 14-carbon length chains appeared to be more efficient as transfecting agent than the ones with shorter chains. Toxicity studies proved that the hydration film

method led to particles well tolerated both by the cells *in vitro* and by the mice *in vivo*. The ability to induce gene expression *in vivo* was demonstrated by intratumoral injection. Finally, biodistribution studies showed that the quantity recovered in the blood circulation, 2h after systemic injection, was improved as compared to cationic lipids.

INTRODUCTION

Gene delivery requires, in most cases, a vector to protect DNA from enzymatic degradation and to allow DNA cellular uptake. Synthetic (1) and viral vectors (2) are currently being developed as gene delivery systems. Among artificial vectors, cationic lipids (3) and polymers (4) are particularly attractive as they can be easily prepared and have been extensively characterized (5). However, their cationic charges lead to nonspecific interaction with anionic serum components, strongly decreasing their transfection efficiency (6, 7) and inducing unspecific delivery *in vivo* (8). Numerous studies have been undertaken to improve serum resistance by hiding the cationic charges (9, 10, 11). Alternative delivery systems have been introduced, using nonelectrostatic interaction with DNA. For instance, Aoyama et al. succeeded to trap DNA in virus-like saccharidic clusters (12), and Barthelemy et al. developed nucleoside-based amphiphiles associating plasmids (13). However, with the exception of artificial glycovirus (14) these systems required a cationic charge to show some transfection activity *in vitro* (15). Strictly neutral systems, like liposomes (16) or spherulites (17), have been worked out for gene delivery, but, to the best of our knowledge, they have not shown any transfection properties.

Recent studies from our laboratory (18, 19) demonstrated the feasibility of using bis-thiourea as a noncationic lipid headgroup for gene transfer both *in vitro* and *in vivo* (20). Thiourea moieties are strong hydrogen bond donors and are able to interact with a wide range of derivatives including carbohydrates, carboxylates (21), nitrate (22) and phosphate groups (23). This last acid derivative is known to lead to the strongest interactions (24). These properties, together with their structural potential variety, make them good candidates to develop alternative vectors for gene transfer based on non-electrostatic interaction. In our growing interest in noncationic delivery systems based on thiourea interacting

groups, we developed a new family of lipopolythioureas (LPT) lipids designed around a 2,6-bis(thiocarbamoyl-amino)hexanoic acid scaffold. The new transfecting agents, described here, are readily available from lysine. In this paper, we describe the synthesis of these compounds, and present their transfection efficiency data, cytotoxicity studies and *in vivo* behavior.

EXPERIMENTAL PROCEDURES

Materials and methods

All solvents were purchased from Carlo Erba-SDS (Peypin France). Dichloromethane was distilled from P₂O₅. DMF was dried over 3 Å molecular sieves and pyridine over KOH. All chemicals were purchased from Sigma-Aldrich-Fluka or Lancaster. Tosyl chloride (petroleum ether) and cholesteryl chloroformate (acetone) were recrystallized, diglycolic anhydride was dehydrated (P₂O₅) prior to use. Other solvents and products were used without further purification. Reactions were monitored by thin-layer chromatography using Merck precoated 60 F254 silica gel plates. Column chromatography was performed over SDS (Peypin France) 35-70 m silicagel according to the method of Still, Khan, and Mitra (25) or using small column flash chromatography (SFC) according to the following procedure: a plastic syringe was filled with silicagel (product/silicagel 1/5) and connected to a vacuum pump. The column was equilibrated with heptane then the sample, dissolved in a minimum of dichloromethane, was added to the top. The column was eluted by ten fractions (volume equal to the silicagel volume) of a heptane/ethyl acetate mixture. For each fraction the amount of ethyl acetate was increased (10% to 100 % (v/v) heptane in ethyl acetate). Inverse phase SPE was performed on Supelco Superclean C8 cartridge using H₂O/CH₃CN/CH₃OH mixture. ¹H and ¹³C NMR spectra were recorded on a BRUKER Avance DRX-300 spectrometer at 300.13 MHz for the proton and 75.47 MHz for the carbon. NMR spectra were processed using Xwinnmr (Bruker) or SwaN-MR (26). MS were carried out on a Shimadzu 2010A LC-MS on ESI mode. High resolution MS and elemental analysis were carried out by the “Service Central de Micro-analyse du CNRS”, Vernaison France. Dipalmytoylphosphatidylcholine (DPPC), Dioleoylphosphatidylethanolamine (DOPE) and phosphatidylethanolamine-lissamine rhodamine

were purchased from Aventi Polar Lipids (Alabaster Alabama). Polyethyleneimine (PEI) was purchased from Sigma-aldrich-Fluka. The lipopolyamine 2-{3-[Bis-(3-amino-propyl)-amino]-propylamino}-N-ditetradecylcarbonyl methyl-acetamide (RPR209120) was previously prepared in our laboratory, the synthesis is described in the supplementary information of reference (9) Plasmid pVax1 was provided by Invitrogen.

Synthesis of lipolythioureas.

(S)-tert-butyl [1-(didecylcarbonyl)-6-tert-butoxycarbonylamino-hexyl]aminoformate (3a)

N-hydroxysuccinimide (0.093 g, 0.81 mmol) and *N,N'*-dicyclohexylcarbodiimide (DCC) (0.42 g, 2.03 mmol) were added to a solution of acid **2** (0.28 g, 0.81 mmol) in dry THF (30 mL). The solution was stirred for one hour before the addition of didecylamine **1a** (0.20 g, 0.68 mmol). The stirring was followed during 17 h then the reaction mixture was diluted with dichloromethane and washed twice with water. The organic layer was dried (MgSO₄) and evaporated under reduced pressure. The resulting oil was purified first by SFC then by flash chromatography (heptane/ethyl acetate 9/1) to yield **3a** as an oil (0.32 g, 63%). ¹H NMR (CDCl₃) δ (ppm) 0.88 (t, 6H, *J* = 6.0 Hz, CH₃), 1.26 (m, 30H, -CH₂-), 1.43 (s, 9H, (CH₃)₃), 1.45 (m, 2H, -CH₂-), 1.55 (m, 4H, -CH₂-), 1.80 (m, 2H, CH₂), 3.11 (m, 2H, -CH₂N), 3.25 (m, 2H, -CH₂N), 3.47 (m, 2H, -CH₂N), 4.52 (m, 1H, CH), 4.54 (m, 1H, NH), 5.35 (d, 1H, *J* = 7.5 Hz, NH). ¹³C NMR (CDCl₃) δ (ppm) 14.11 (s, CH₃), 22.73 (m, -CH₂-), 27.05 (s, -CH₂-), 27.67 (s, -CH₂-), 28.45 (s, (CH₃)₃), 29.62 (m, -CH₂-), 31.95 (s, -CH₂-), 33.09 (s, CH₂), 40.51 (s, CH₂N), 46.10 (s, CH₂N), 47.86 (s, CH₂N), 49.92 (s, CH), 79.18 (s, C(CH₃)₃), 79.80 (s, C(CH₃)₃), 155.94 (s, OC=O), 156.31 (s, OC=O), 171.52 (s, NC=O).

(S)-tert-butyl [1-(didodecylcarbonyl)-6-tert-butoxycarbonylamino-hexyl]aminoformate (3b)

N-hydroxysuccinimide (0.24 g, 2.05 mmol) and DCC (1.06 g, 5.1 mmol) were added to a solution of acid **2** (0.71 g, 2.05 mmol) in dry THF (60 mL). The solution was stirred for two hours before addition of didodecylamine **1b** (0.61 g, 1.72 mmol). The stirring was followed during 20h then the reaction mixture was diluted with dichloromethane and washed twice with water. The organic layer was dried (MgSO₄) and evaporated under reduced pressure. The resulting oil was purified by flash

chromatography (heptane/ethyl acetate 95/5) to yield **3b** as an oil (0.65 g, 46 %). ¹H NMR (CDCl₃) δ (ppm) 0.84 (t, 6H, *J* = 6.0 Hz, CH₃), 1.22 (m, 38H, -CH₂-), 1.39 (s, 9H, (CH₃)₃), 1.49 (m, 2H, CH₂), 1.54 (m, 6H, -CH₂-), 3.06 (m, 2H, -CH₂N), 3.22 (m, 2H, -CH₂N), 3.45 (m, 2H, -CH₂N), 4.49 (m, 1H, CH), 4.68 (m, 1H, NH), 5.36 (d, 1H, *J* = 8.7 Hz, NH). ¹³C NMR (CDCl₃) δ (ppm) 14.09 (s, CH₃), 22.65 (m, -CH₂-), 26.99 (s, -CH₂-), 27.63 (s, -CH₂-), 28.42 (s, (CH₃)₃), 29.38 (s, -CH₂-), 29.62 (m, -CH₂-), 31.93 (s, -CH₂-), 33.81 (s, CH₂), 40.43 (s, -CH₂N), 46.07 (s, -CH₂N), 47.83 (s, -CH₂N), 49.89 (s, CH), 78.95 (s, C(CH₃)₃), 79.39 (s, C(CH₃)₃), 155.56 (s, OC=O), 156.03 (s, OC=O), 171.92 (s, C=O). MS (ESI, *m/z*) 682 (M+H)⁺, 704 (M+Na)⁺.

(S)-tert-butyl [1-(ditetradecylcarbamoyl)-6-tert-butoxycarbonylamino-hexyl]aminofornate (**3c**)

N-hydroxysuccinimide (0.17 g, 1.44 mmol) and DCC (0.74 g, 3.60 mmol) were added to a solution of acid **2** in dry THF (50 mL). The solution was stirred for one hour before the addition of ditetradecylamine **1c** (0.50 g, 1.21 mmol). The stirring was continued for 20 h then the reaction mixture was diluted with dichloromethane and washed twice with water. The organic layer was dried (MgSO₄) and evaporated under reduced pressure. The resulting oil was purified by flash chromatography (heptane/ethyl acetate 95/5) to yield **3c** as an oil (0.73 g, 67%). ¹H NMR (CDCl₃) δ (ppm) 0.88 (t, 6H, *J* = 6.0 Hz, CH₃), 1.26 (m, 30H, -CH₂-), 1.44 (s, 9H, (CH₃)₃), 1.45 (m, 2H, CH₂), 1.58 (m, 6H, -CH₂-), 3.11 (m, 2H, -CH₂N), 3.25 (m, 2H, CH₂N), 3.49 (m, 2H, -CH₂N), 4.52 (m, 1H, CH), 4.54 (m, 1H, NH), 5.39 (d, 1H, *J* = 7.5 Hz, NH). ¹³C NMR (CDCl₃) δ (ppm) 14.17 (s, CH₃), 22.76 (m, -CH₂-), 27.05 (s, -CH₂-), 27.67 (s, -CH₂-), 28.45 (s, (CH₃)₃), 29.62 (m, -CH₂-), 31.94 (s, -CH₂-), 33.74 (s, CH₂), 40.40 (s, CH₂N), 46.10 (s, CH₂N), 47.85 (s, CH₂N), 49.90 (s, CH), 79.38 (s, C(CH₃)₃), 155.94 (m, OC=O), 171.85 (s, C=O). MS (ESI, *m/z*) 760 (M+Na)⁺.

(S)-2,7-diamino-*N,N*-didecyl-heptanamide (**4a**)

A solution of amide **3a** (1.31 g, 2.1 mmol) in dichloromethane (2 mL) was treated with a TFA/CH₂Cl₂ mixture (3:1, 8 mL). The reaction mixture was stirred for 24h then was treated with solid NaHCO₃, filtered and washed with water then 2N NaOH was added to the aqueous phase until pH 9. The water layer was extracted three times with dichloromethane. The organic layers were dried (MgSO₄) and

evaporated under reduced pressure to yield **4a** (0.9 g, 100 %) as a yellow oil, which was used without further purification. ¹H NMR (CDCl₃) δ (ppm) 0.87 (t, 6H, *J* = 6.0 Hz, CH₃), 1.26 (m, 30H, -CH₂-), 1.55 (m, 6H, -CH₂-), 1.65 (m, 2H, CH₂), 2.03 (m, 4H, NH), 2.72 (m, 2H, CH₂NH₂), 3.11 (m, 2H, -CH₂N), 3.24 (m, 2H, CH₂N), 3.52 (m, 2H, CH), 5.40 (m, 1H, NH). δ (ppm) 14.13 (s, CH₃), 22.74 (s, -CH₂-), 23.22 (s, -CH₂-), 27.08 (s, -CH₂-), 27.78 (s, -CH₂-), 29.62 (m, -CH₂-), 31.94 (s, -CH₂-), 33.06 (s, CH₂), 41.74 (s, CH₂NH₂), 46.28 (s, CH₂N), 47.55 (s, CH₂N), 51.17 (s, CH), 173.05 (s, C=O).

(S)-2,7-diamino-*N,N*-didodecyl-heptanamide (**4b**)

A solution of amide **3b** (0.65 g, 0.95 mmol) in dichloromethane (2 mL) was treated with a TFA/CH₂Cl₂ mixture (3:1, 5 ml). The reaction mixture was stirred for 3h then it was treated with solid NaHCO₃, filtered and washed with water. NaOH 2N was added to the aqueous phase until pH 9. The water layer was extracted three times with dichloromethane. The organic layers were dried (MgSO₄) and evaporated under reduced pressure to yield **4b** (0.5 g, 100 %) as a yellow oil, which was used without further purification. ¹H NMR (CDCl₃) δ (ppm) 0.86 (t, 6H, *J* = 6.0 Hz, CH₃), 1.23 (m, 38H, -CH₂-), 1.49 (m, 6H, -CH₂-), 2.94 (m, 2H, CH₂NH₂), 3.11 (m, 4H, CH₂N), 3.80 (m, 2H, CH), 6.24 (m, 4H, NH). ¹³C NMR (CDCl₃) δ (ppm) 14.09 (s, CH₃), 21.89 (s, -CH₂-), 22.71 (s, -CH₂-), 27.12 (s, -CH₂-), 27.51 (s, -CH₂-), 29.70 (m, -CH₂-), 31.96 (s, -CH₂-), 33.94 (s, CH₂), 39.16 (s, CH₂NH₂), 46.50 (s, CH₂N), 47.72 (s, CH₂N), 50.73 (s, CH), 173.10 (s, C=O). MS (ESI, *m/z*) 483 (M+H)⁺, 262 ((M+2H+2Na)/2)⁺.

(S)-2,7-diamino-*N,N*-ditetradecyl-heptanamide (**4c**)

A solution of amide **3c** (0.73 g, 1 mmol) in a TFA/CH₂Cl₂ mixture (4:1, 5 ml) was stirred for 3 h. Then the reaction mixture was treated with solid NaHCO₃, filtered, and washed with water. 2N NaOH was added to the aqueous phase until pH 9. The water layer was extracted three times with dichloromethane. The organic layers were dried (MgSO₄) and evaporated under reduced pressure to yield **4c** (0.36 g, 68%) as a yellow oil, which was used without further purification. ¹H NMR (CDCl₃) δ (ppm) 0.86 (t, 6H, *J* = 6.0 Hz, CH₃), 1.23 (m, 30H, -CH₂-), 1.47 (m, 6H, -CH₂-), 1.88 (m, 2H, CH₂), 2.81 (m, 2H, CH₂NH₂), 3.10 (m, 2H, CH₂N), 3.24 (m, 2H, CH₂N), 3.47 (m, 2H, CH), 5.40 (m, 1H, NH).

^{13}C NMR (CDCl_3) δ (ppm) 14.12 (s, CH_3), 22.72 (m, $-\text{CH}_2-$), 27.10 (s, $-\text{CH}_2-$), 27.70 (s, $-\text{CH}_2-$), 29.70 (m, $-\text{CH}_2-$), 31.97 (s, $-\text{CH}_2-$), 34.00 (s, $-\text{CH}_2-$), 40.91 (s, CH_2NH_2), 46.33 (s, CH_2N), 47.58 (s, CH_2N), 51.48 (s, CH), 171.85 (s, $\text{C}=\text{O}$). MS (ESI, m/z) 539 ($\text{M}+\text{H}$) $^+$.

(S)-2,7-bis[(2,2-dimethyl-1,3-dioxolan-4-yl)methylthiocarbamoylamino]-*N,N*-didecyl-heptanamide
(6a)

A solution of amine **4a** (0.21 g, 0.5 mmol), and *N,N*-diisopropylethylamine (0.87 mL, 5 mmol) in dichloromethane (5 mL) was stirred for 10 mn before addition of isothiocyanate **5** (0.22 g, 1.3 mmol). The reaction mixture was stirred overnight and evaporated under reduced pressure. Purification of the crude by SFC afforded **5a** (0.32 mg, 83 %) as a colorless oil. ^1H NMR (CDCl_3) δ (ppm) 0.86 (t, 6H, $J = 6.0$ Hz, CH_3), 1.24 (m, 32H, $-\text{CH}_2-$), 1.31 (s, 6H, $(\text{CH}_3)_2$), 1.33 (s, 6H, $(\text{CH}_3)_2$), 1.43 (m, 4H, $-\text{CH}_2-$), 1.66 (m, 2H, $-\text{CH}_2-$), 3.20 (m, 4H, CH_2NCO), 3.45 (m, 2H, CH_2NCS) 3.68 (m, 4H, CH_2NCS), 4.01 (m, 4H, CH_2O), 4.27 (m, 4H, CHO), 5.55 (m, 1H, CH), 6.85 (m, 2H, NH), 7.85 (m, 2H, NH). ^{13}C NMR (CDCl_3) δ (ppm) 13.98 (s, CH_3), 22.72 (s, $-\text{CH}_2-$), 23.40 (s, $-\text{CH}_2-$), 25.30 (s, $(\text{CH}_3)_2$), 27.05 (s, $(\text{CH}_3)_2$), 27.71 (s, $-\text{CH}_2-$), 28.76 (s, $-\text{CH}_2-$), 29.38 (s, $-\text{CH}_2-$), 29.66 (m, $-\text{CH}_2-$), 31.95 (s, $-\text{CH}_2-$), 33.70 (s, CH_2), 44.58 (s, CH_2NCS), 46.78 (m, CH_2NCS , CH_2N), 48.70 (s, CH_2N), 53.77 (s, CH), 68.67 (s, CH_2O), 75.05 (s, CHO), 109.50 (s, $\text{CO}_2(\text{CH}_3)_2$), 172.91 (s, $\text{C}=\text{O}$), 183.26 (s, $\text{C}=\text{S}$). Anal. calcd. for $\text{C}_{40}\text{H}_{77}\text{N}_5\text{O}_5\text{S}_2$: C, 62.22; H, 10.05; N, 9.07; S, 6.30. Found C, 62.17; H, 10.17; N, 8.94; S, 7.07. MS (ESI, m/z) = 794 ($\text{M}+\text{Na}$) $^+$, 770 ($\text{M}-\text{H}$) $^-$, 808 ($\text{M}+\text{Cl}$) $^+$.

(S)-2,7-bis[(2,2-dimethyl-1,3-dioxolan-4-yl)methylthiocarbamoylamino]-*N,N*-didodecyl-heptanamide
(6b)

A solution of amine **4b** (0.24 g, 0.5 mmol), and *N,N*-diisopropylethylamine (0.44 mL, 2.5 mmol) in dichloromethane (2 mL) was stirred for 10 minutes before the addition of isothiocyanate **5** (0.095 g, 0.55 mmol). The reaction mixture was stirred overnight and evaporated under reduced pressure. Purification of the crude by SFC afforded **6b** (0.23 g, 55 %) as a colorless oil. ^1H NMR (CDCl_3) δ (ppm) 0.87 (t, 6H, $J = 6.0$ Hz, CH_3), 1.25 (m, 40H, $-\text{CH}_2-$), 1.32 (s, 6H, $(\text{CH}_3)_2$), 1.34 (s, 6H, $(\text{CH}_3)_2$), 1.44 (m, 4H, $-\text{CH}_2-$), 1.68 (m, 2H, $-\text{CH}_2-$), 3.21 (m, 4H, CH_2NCO), 3.47 (m, 2H, CH_2NCS) 3.70 (m, 4H,

CH₂NCS), 4.05 (m, 4H, CH₂O), 4.29 (m, 4H, CHO), 5.59 (m, 1H, CH), (m, 2H, NH), 7.86 (m, 2H, NH). ¹³C NMR (CDCl₃) δ (ppm) 14.13 (s, CH₃), 22.73 (s, -CH₂-), 25.27 (s, (CH₃)₂), 26.87 (s, (CH₃)₂), 27.71 (s, -CH₂-), 28.71 (s, -CH₂-), 29.07 (s, -CH₂-), 29.68 (m, -CH₂-), 31.98 (s, -CH₂-), 33.68 (s, CH₂), 44.58 (s, CH₂NCS), 46.76 (m, CH₂NCO, CH₂NCS), 48.63 (s, CH₂NCO), 53.77 (s, CH), 66.63 (s, CH₂O), 75.01 (s, CHO), 109.62 (s, CO₂(CH₃)₂), 172.76 (s, C=O), 183.23 (s, C=S). Anal. calcd. for C₄₄H₈₅N₅O₅S₂ : C, 63.80; H, 10.34; N, 8.45; S, 7.74. Found C, 63.94; H, 10.51; N, 7.80; S, 7.46. MS (ESI, *m/z*) = 850 (M+Na)⁺, 828 (M+Cl)⁻.

(S)-2,7-bis[(2,2-dimethyl-1,3-dioxolan-4-yl)methylthiocarbamoylamino]-*N,N*-ditetradecyl-heptanamide (**6c**).

A solution of amine **4c** (0.12 g, 0.22 mmol), and *N,N*-diisopropylethylamine (0.3 mL, 2.2 mmol) in dichloromethane (1.5 mL) was stirred for 10 mn before the addition of isothiocyanate **5** (0.1 g, 0.55 mmol). The reaction mixture was stirred overnight and evaporated under reduced pressure. Purification of the crude by SFC afforded **6c** (75 mg, 38 %) as a colorless oil. ¹H NMR (CDCl₃) δ (ppm) 0.87 (t, 6H, *J* = 6.0 Hz, CH₃), 1.25 (m, 44H, -CH₂-), 1.33 (s, 6H, (CH₃)₂), 1.37 (s, 6H, (CH₃)₂), 1.44 (m, 4H, -CH₂-), 1.67 (m, 2H, CH₂), 3.20 (m, 4H, CH₂NCO), 3.45 (m, 2H, CH₂NCS), 3.69 (m, 4H, CH₂NCS), 4.05 (m, 4H, CH₂O), 4.29 (m, 4H, CHO), 5.59 (m, 1H, CH), 6.85 (m, 2H, NH), 7.80 (m, 2H, NH). ¹³C NMR (CDCl₃) δ (ppm) 14.15 (s, CH₃), 22.74 (m, -CH₂-), 25.26 (s, (CH₃)₂), 27.06 (s, (CH₃)₂), 27.15 (s, -CH₂-), 28.69 (s, -CH₂-), 29.07 (s, -CH₂-), 29.68 (m, -CH₂-), 31.97 (s, -CH₂-), 33.69 (s, CH₂), 44.54 (s, CH₂NCS), 46.74 (m, CH₂NCO, CH₂NCS), 46.63 (s, CH₂NCO), 53.74 (s, CH), 66.61 (s, CH₂O), 75.01 (s, CHO), 109.60 (s, CO₂(CH₃)₂), 172.79 (s, C=O), 183.19 (s, C=S). MS (ESI, *m/z*) 907 (M+Na)⁺, 931 (M+HCOO)⁻. Anal. calcd. for C₄₈H₉₃N₅O₅S₂ : C, 65.19; H, 10.60; N, 7.92; S, 7.25. Found C, 65.18; H, 10.66; N, 7.21; S, 6.58. HR-ESMS calcd. for C₄₈H₉₃N₅O₅NaS₂ : 906.6516. Found 906.6498.

Liposome preparation.

Ethanol injection method: Lipopolythiourea and DPPC were dissolved in ethanol and were added dropwise to 10 volumes of water under vigorous agitation. The mixture was stirred overnight and then

evaporated under reduced pressure at room temperature to obtain a fairly concentrated solution of liposomes.

As an example, compound **6c** (1.77 mg, 2 μ mol) and DPPC (1.47 mg, 2 μ mol) were dissolved in 300 μ L ethanol. This solution was added dropwise into 3 mL of stirred filtered water. The mixture was stirred overnight then evaporated under reduced pressure at RT to obtain a clear suspension of **6c** / DPPC (0.380 mL, 10.5 mM total lipids).

Film hydration method: Lipopolythiourea and DPPC were dissolved in chloroform. The solvent was evaporated slowly under vacuum to obtain a thin film on the glass. Water was then added to hydrate the lipids and obtain a fairly concentrated solution of liposomes.

As an example, compound **6c** (4.0 mg, 4.5 μ mol) and DPPC (3.2 mg, 4.3 μ mol) were dissolved in chloroform (500 μ L). The solvent was evaporated slowly under vacuum to obtain a thin film on the glass. Water (430 μ L) was then added to hydrate the lipids and obtain a solution of liposomes (20 mM total lipids).

Size measurement. Particle diameter was determined by dynamic light scattering on a Zeta Sizer NanoSeries Malvern (Malvern Instruments, France). Concentration of the lipids = 0.1 mM in H₂O, mean value of 3 runs are given (1 min equilibration, 3 min run).

Preparation of plasmid DNA. pVax2 is a derivative of the commercial plasmid pVax1 (InVitrogen). pVax1 was digested with the restriction enzymes HincII and BamHI to excise the promoter. The plasmid was then blunted with the Klenow fragment and dephosphorylated with alkaline phosphatase. pCMVbeta (Clontech) was digested with EcoR1 and BamHI to excise the CMV promoter. The CMV promoter was blunted with Klenow enzyme and ligated into the blunted pVax1 to give pVax2. The plasmid pXL3031 was digested with EcoRI and BamHI, and then treated with the Klenow fragment to produce a blunted fragment containing the luciferase cDNA. This fragment was ligated into pVax2 after EcoRV digestion and phosphatase alkaline dephosphorylation to give pVax2-Luc.

Preparation of LPT/DNA complexes. Plasmid DNA (pVax2-luc, 100 μ L, 0.02 g/L in H₂O) prepared using the procedure above, was added dropwise with constant vortexing to various amounts of LPT/DPPC liposomes (in 100 μ L H₂O) at RT. TU/P indicates the ratio in nmol of thiourea function (2 per lipid) versus DNA phosphates. Cationic lipoplexes were formed in the same way from RPR209120/DOPE liposomes (1/1, hydration film method, 4 nmol in 100 μ L NaCl 150 mM) mixed with plasmid DNA at ratio N/P = 8 (0.5 μ g in 100 μ L NaCl 150 mM). PEI/DNA polyplexes were prepared from PEI (1 μ g in 100 μ L glucose 5%) mixed with plasmid DNA (0.5 μ g in 100 μ L glucose 5%) at a N/P ratio = 15.

Gel retardation experiments. Complexes were prepared as described above with liposomes made by ethanolic injection (20 μ L) and 5 μ L of bromophenol blue were added. The mixture was loaded on a 0.8 % agarose gel in TBE buffer (1M Tris, 0.9M boric acid, 0.01 M EDTA) at 80V/cm. DNA was revealed with ethidium bromide and visualized under UV light.

DNA protection. 20 μ L of culture medium (MEM with 10% or 50 % murine fresh serum) were added on 20 μ L of LPT/DPPC/DNA complexes (0.1 g/L DNA, prepared by ethanolic injection) or of cationic lipoplexes made of RPR209120 (0.1 g/L DNA). Samples were incubated at 37° C. After 6h, 10 μ L of samples were frozen at -20°C. After 24h, all the samples were replaced at RT. SDS 2% (5 μ L), EDTA (2 μ L, 0.5M) and bromophenol blue (3 μ L) were added on each sample (10 μ L). The mixture was loaded on agarose gel 1% containing 0.05% SDS at 80 V/cm. After 24h washing in water, DNA was revealed with ethidium bromide and visualized under UV light.

Transfection method. B16 murine cells were grown into DMEM supplemented with L-glutamin (29.2 mg/mL), penicillin (50 units/mL), streptomycin (50 units/mL) and 10% fetal bovine serum. One day before transfection, cells were treated by trypsin and deposited into 24-wells plates (45000 cells/well) and incubated 24h at 37° C. 100 μ L of LPT/DNA complexes (prepared by ethanolic injection), 100 μ L of RPR209120/DOPE/DNA (prepared as above, ratio N/P = 8) and PEI/DNA polyplexes (prepared as above, ratio N/P = 15), all formulations corresponding to 0.5 μ g DNA, were loaded on each well and the

plates were incubated at 37° C for 48h. Then the cells were washed twice with PBS and treated with 200µL of a passive lysis buffer (Promega). After 15 min, the cells were centrifuged for 5 min at 12000 r/min. Supernatant (10 µL) and iodoacetamide (10 µL) were added to a 96-well plate which was incubated at 37° C for 1h. Protein quantification was performed with the BCA protein assay KIT (PIERCE) and reported to BSA taken as a reference curve. Luciferase activity was quantified using a commercial kit Luciferase assay system (PROMEGA). On 10 µL of the lysed cells, 50 µL of the luciferase substrate was injected via an injector and the absorbance was read immediately at 563 nm on a Wallac Victor2 1420 Multilabel Counter Perkin Elmer.

Cytotoxicity on B16 murine cells. Murine B16 melanoma cells were grown in DMEM containing 2 mM L-glutamine, 10% fetal bovine serum, 100 U/mL penicillin, and 100 µg/mL streptomycin (37 °C, 5% CO₂). Exponentially growing B16 cells were plated onto 96-well plates at 5000 cells per well in 100 µL of culture medium. Twenty-four hours after plating, 100 µL of medium containing the compound of interest (final concentrations ranging from 1.56 µg/mL to 100 µg/mL, in 2-fold dilutions) was added to the wells (in triplicate) containing the cells and incubated for 48 h at 37 °C and 5% CO₂. After the 48 h exposure period to the test compounds, cell viability was assayed using the MTT test (27) and absorbance was read at 562 nm in a microplate reader (BioKinetics Reader, EL340). Appropriate controls with DMEM only and MTT were run to subtract background absorbance. Results are presented as percent of controls cells. The concentration of compound that inhibited cell viability by 50% (inhibitory concentration for 50% of cells, or IC₅₀) was determined using the GraphPad Prism software. Results are presented as means ± SEM of 6 independent experiments each run in triplicate.

In vivo acute toxicity test. 6c/DPPC particles (0.75 µmol, 200 µL) were injected into Swiss mice tail vein (3 mice for the sample, 3 mice control). Animals were observed daily over 16 days about their weight, behavior pattern, skin and fur modifications and somatomotor activity (28). The dose was considered as toxic if the weight was lower than 80% of the controlled mice.

Intratumoral injection. **6c**/DPPC/DNA (40 μ L, 10 μ g DNA, 40 TU/P, TU/P = mol thiourea functions/mol DNA phosphate group) and RPR209120/DOPE/DNA (prepared as above, ratio N/P = 6) were injected in 3LL tumors of 3 mice C57BL/6 bearing 3LL tumors. Mice were sacrificed 24h after injection, then tumors were removed, weighed, crushed in 1mL lysis buffer, and centrifuged during 10 minutes (12000 rpm, 4 $^{\circ}$ C). The supernatant (50 μ L) was loaded on a 96-well plates and luciferase activity was quantified using a commercial kit (Protein Assay). The absorbance was read at 563 nm on a Wallac Victor2 1420 Multilabel Counter Perkin Elmer.

Biodistribution study.

Experiments were conducted following NIH recommendation for animal experimentation. Rhodamine-labeled **6c**/DPPC/DNA complexes (200 μ L, 11 μ g DNA, 40 TU/P) were injected in tail vein of five mice C57BL/6 bearing 3LL tumors. Blood was collected by cardiac puncture after 2h, and mice were euthanized at the end of the sample taking. In the distribution study, liver, lungs, spleen, and tumors were removed, weighed, and homogenized in pH 7.4 phosphate-buffered saline (PBS, 5mL/g tissue). Rhodamine-labeled lipids were extracted as previously described (29) from 100 μ L of blood or tissue homogenates with 3 mL of CHCl₃-CH₃OH (1:1, v/v) by vigorous mixing during 30 min for blood and 40 min for tissue homogenates followed by centrifugation (3000 rpm, 10 min). The fluorescence intensity corresponding to rhodamine was assayed on the supernatant with a fluorospectrophotometer (Victor, Perkin Elmer_Wallac multiplate reader, λ_{exc} 530 nm, λ_{em} 570 nm). The amount of complexes in the blood or tissue homogenates was evaluated with a calibration curve of the initial **6c**/DPPC/DNA complexes containing 0.5 % LPT-rhodamine and expressed as the remaining percentage of injected dose.

RESULTS

Chemistry.

The lipopolythioureas were prepared as shown in Scheme 1. The synthesis started from boc lysine **2** that reacted with dialkylamine **1a**, **1b** and **1c** to yield dialkylamide **3a-c** (45 to 67 %). Boc removal

using TFA in dichloromethane led to diamines **4a-c** in good yields (68, 100, 100% respectively). Treatment of the diamine with isothiocyanate **5** afforded the protected thioureas (**6a** 83%, **6b** 55%, **6c** 38 %). Attempts to remove the acetal group failed, giving only the starting alkylamine **1**. This result suggests a rearrangement leading to a cyclic thiourea under protic catalysis, since such cyclic compounds have been described in literature (30). However, despite our efforts we were unable to identify such a compound.

Scheme 1

Formulation study and DNA association.

The three compounds prepared **6a**, **6b** and **6c** exhibit the same thiourea headgroup and differentiate themselves by their lipidic length, varying from 10 to 12 to 14 carbons, respectively. First, we tried to evaluate if their hydrophilic/lipophilic balance would allow forming homogeneous particles. This was achieved thanks to the adjunction of dipalmytoyl-phosphatidylcholine (DPPC) as a ratio 1 to 1 with the polythiourea lipid. Particle diameter is given in Table 1. The diameter was slightly higher for **6c** as compared to **6b** and **6a**. Also, the suspension formed was more homogeneous with compound **6c**, as shown by the polydispersity index. This might suggest a better interaction of compound **6c** with DPPC due to its longer chain length.

LPT	Chain length	Formulation (ethanolic injection)	Diameter (nm)	PDI	TU/P for DNA association	LPT/DNA Diameter (nm)	PDI
6a	10 C	6a /DPPC 1/1	76.5±11	0.337	40	96±2	0.253
6b	12 C	6b /DPPC 1/1	102±8	0.329	20	122±6	0.165
6c	14 C	6c /DPPC 1/1	141±50	0.239	10	164±3	0.115

Table 1. Formulation and DNA association with lipopolythiourea **6a**, **6b** and **6c**. Diameter is given as a mean of 3 values. PDI: polydispersity index. TU/P for DNA association is the number of thiourea/DNA phosphate ratio needed to retain DNA on the wells in agarose gel. The diameters of LPT/DNA complexes are given at the ratio needed for the association.

Addition of DNA to these particles induced a slight increase in particle diameter. LPT/DNA interaction was evidenced by this size increase along with gel electrophoresis. We relate in Table 1 the ratio between the thiourea and the phosphate functions (TU/P) needed to fully retain DNA on the wells of an agarose gel. As can be seen, all LPT were able to complex the plasmid. Moreover, the longer the lipid chain, the less amount of LPT was necessary to obtain a full interaction with DNA, reaching a minimum for **6c** (10 TU/P). This could be linked to the suspension homogeneity obtained when DPPC was mixed with lipid **6c**.

Lipoplex sensitivity to serum.

The complexes formed between LPT and DNA have been submitted to a treatment in fresh mice serum at 37° C for 6h and 24h, and then loaded on a denaturing agarose gel. As observed on Figure 1, free plasmid DNA was partially degraded after 6h, and fully degraded after 24h under these conditions, whereas complexed DNA was still intact. As regard to the technique sensitivity, no difference between the three lipids could be detected. These results confirm that the interaction occurring between LPT and DNA is strong enough to protect plasmid DNA from degradation in the presence of serum.

Figure 1

Transfection efficiency in vitro.

Compounds **6a**, **6b** and **6c** were further evaluated for their transfection activity. Since the aim of these nonelectrostatic systems is to improve the in vivo gene delivery, we carried out in vitro studies as preliminary tests for in vivo experiments. Thus, we performed the experiments in the presence of serum with particles of nanometric size. Two different TU/P ratios were evaluated for the three different formulations and compared to the cationic lipid (RPR209120) (31) and the cationic polymer PEI (32) (Figure 2) mixed with DNA.

Figure 2

As can be seen on Figure 3, transfection efficiency was dependent on the LPT used. Hence, higher transfection activity was obtained when increasing the lipid length from 10 to 14 carbons, from **6a** to **6c** respectively. Moreover, increasing the amount of LPT, from TU/P= 20 to 40, also led to improved transfection. However, for higher LPT amount we observed a decrease of the protein amount, suggesting some cytotoxicity.

Cytotoxicity.

In order to reduce the cytotoxicity, we have studied the effect of different formulation protocols. Particles mentioned in the previous experiments were formulated via an ethanolic injection protocol (33). We also prepared LPT particles via the film hydration method (34). After preparation, we submitted the cells to a MTT test and compared the IC₅₀ of the two different formulations. As can be seen on Figure 4, the cytotoxicity IC₅₀ obtained for compound **6a** formulated via an injection method was 15μM, while no toxicity could be measured when the same compound was formulated via the hydration method. This significant reduction of toxicity might be due to the reduction of the number of thiourea presented at the surface of the liposome since multilamellar vesicles (MLV) are formed by the hydration method, whereas unilamellar vesicles (SUV) are obtained by ethanolic injection (35). The *in vitro* efficiency of these complexes was also evaluated and, showed slightly the same level of transfection as compared to the previous one (9900 RLU/μg protein for a ratio TU/P=20). This good tolerance prompted us to use the hydration method for the following *in vivo* studies

Figure 4

According to the *in vitro* cytotoxicity results, we evaluated the *in vivo* toxicity of the least toxic and most efficient formulation: **6c**/DPPC, prepared by the hydration film method. The formulation was

injected once at $t = 0$, then the mice weight, as well as different toxicity signs such as locomotion, behaviour, changes in skin and fur, were followed over 2 weeks. The formulations were considered as nontoxic if the weight loss was lower than 20% as compared to the controlled mice. The formulation **6c**/DPPC prepared via the hydration method displayed an excellent tolerance, since the somatomotor activity, the behavior pattern and skin and fur characteristics were unchanged as assayed over a period of 16 days. The weight also remained stable from 98.6% at day 2, to 97.6% at day 5 to 98.65% of the controls at day 16.

***In vivo* properties.**

The absence of toxicity for the formulation **6c**/DPPC prompted us to investigate further the *in vivo* behavior of these particles.

First, we evaluated the transfection activity of the formulation **6c**/DPPC after intratumoral injection, and compared it (36) to the lipoplexes composed of the lipopolyamine RPR209120. We did not observe a significant difference between the two formulations; both were able to induce a luciferase activity 24 h post-injection.

Biodistribution of **6c**/DPPC was evaluated by introduction of a fluorescent lipid marker (37) and compared to a cationic formulation of the lipopolyamine RPR209120/DOPE prepared via an identical hydration film procedure. 2h after injection, different organs were removed and assessed for their fluorescence content. As shown on Figure 5, a significative gain of a factor 4 to 5 ($p < 0.01$) was obtained in the blood circulation particles using the neutral **6c**/DPPC formulation as compared to the cationic one. Despite this higher circulation time in the blood, no significant difference was found in tumor localization. Furthermore, an important difference was observed in the lung ($p < 0.01$). Nonspecific accumulation of the particles in the lung, which has often been reported for cationic particles (38), did not occur in the case of the LPT formulation. A similar amount was found in the liver for the two formulations, which was expected as elimination by Kupffer cells is mostly related to the particle size (39).

Figure 5

DISCUSSION

The above data show that the described lysine derived polythiourea mixed with DPPC are able to associate to plasmid DNA, to protect it from degradation in the presence of serum and to induce gene expression in cultured cells. Transfection efficiency was shown to be chain length dependent, the lipid **6c** bearing the longer chain exhibiting the highest transfection activity. Some results have shown that a shorter chain was more appropriate for optimized transfection (40). We had also observed that with a lipopolythiourea bearing a serinol headgroup (20). Figure 6 represents the comparison of the in vitro results mediated by the two lipopolythiourea family. It shows that, indeed, the transfection level is reversed between the two LPT family, the shorter chain is preferable for the serinol thiourea lipids, shown on figure 7, while the longer chain is preferable for the lysine thiourea lipids. That could be related to the improved hydrophilic/lipophilic balance in the lipid. In this work, the hydrophobic nature of the LPT required the use of a colipid, and obviously the colipid effect is of great importance as was previously reported (41). Mixing LPT with DPPC (bearing 16 carbons in the alkyl chain) might be more appropriate for the LPT **6c**, inducing improved lipid packing as the chain length of the LPT increases from 10 to 14 carbons. Indeed, suspension polydispersity decreased from the formulation **6a**/DPPC to **6c**/DPPC. The shorter chain LPT might create disorder in the lipid bilayer eventually leading to unstable particles (42). Alternatively, we can hypothesize that the bulky and unsymmetrical LPT headgroup might also interfere with the lipid packing and that a stronger hydrophobic interaction between LPT and the colipid might counterbalance the headgroup interference.

Figure 6

Despite its structure, this lysine-based uncharged headgroup allowed for efficient interaction with DNA at a LPT/DNA ratio similar to the one previously obtained for LPT bearing a serinol headgroup (TU/P = 15). This ratio was the lowest for the formulation **6c**/DPPC, as compared to **6b** and **6a**, which is probably related to the more homogeneous structure formed between lipid **6c** and DPPC.

The most efficient transfection obtained with lipid **6c** was higher than the previously reported serinol LPT at the ratio used (20) (Figure 6) and comparable to that of cationic formulations in the presence of serum. This is of particular interest considering that LPT formulations exhibit no positive charges and might interact less favorably with the anionic cell membranes. The absence of cationic charges was also confirmed by the *in vivo* improved kinetic properties of LPT/DNA complexes, which avoided nonspecific accumulation in the lung and blood circulation. However, this “stealth advantage” did not enhance passive accumulation in our tumor model in opposite to the proposed enhanced permeability and retention effect (43). We have already observed this lack of passive accumulation on 3LL tumors after chemical passivation of the cationic charges of lipoplexes (9). This points the use of a targeting ligand at the surface of these DNA/lipid particles in order to deliver DNA at a proper site, especially as this strategy has already been validated in the case of lipopolythiourea (19).

Figure 7

In conclusion, we described here a new noncationic transfecting agents bearing a polythiourea moiety. From this and previous results, we can deduce that a wide range of geometry and terminus allowed LPT/DNA interaction. With the present compound **6c**, this association was found to be strong enough to protect plasmid in the presence of serum and to obtain *in vitro* efficient cell transfection, comparative to that obtained with cationic lipids. Moreover, we could find a nontoxic window for *in vivo* use of these complexes. Intratumoral injection of LPT/DNA complexes showed that *in vivo* transfection ability and biodistribution were favorably improved as compared to cationic complexes. Finally we have found in our previous studies that the size of the hydrophobic anchor seemed to be one of the factor controlling

the physico-chemical and the transfecting properties of the LPTU. Structural comparison between **6a-6c** and **1a-1c** (Figure 7) shows two main differences between them. First the lack of a linker, second the presence of a 1,5-diamine instead of a 1,3-diamine for the headgroups. This suggests that a larger diamino chain would lead to better phosphate/thiourea interactions. This hypothesis is supported by the study of Bianco et al. (44) that demonstrate strong thiourea/phosphate interaction for thiourea-linked glycooligomers (Figure 7, Compound **VI**). In these molecules the two thioureas are in a 1,4 relationships favoring their participation in bidentate hydrogen-bond interactions with oxoanions. Compounds **III**, **IV** and **V** (Figure 7) possess also a 1,5 relationship between the two thioureas (19). However their lack of transfecting properties is probably due to their hydrophobicity. In addition, the presence of the acetal ring adds a constraint that could diminished the strength of hydrogen bonds. New models have been synthesized and are now under investigation to verify these hypotheses.

ACKNOWLEDGMENT: This work was financially supported by INSERM, CNRS, MNERT, the Région Ile de France (SESAME, CPER). J.L. thanks the MNERT for a doctoral fellowship.

REFERENCES

-
- (1) Miller, A. D. (1998) Cationic liposomes for gene therapy. *Angew. Chem. Int. Ed.* 37, 1768-1785.
- (2) Verma, I. M., Weitzman, M. D. (2005) Gene therapy: twenty-first century medicine. *Annu. Rev. Biochem.* 74, 711-738.
- (3) Simberg, D., Weisman, S., Talmon, Y., Barenholz, Y. (2004) DOTAP (and other cationic lipids): chemistry, biophysics, and transfection. *Crit. Rev. Ther. Drug Carrier Syst.* 21, 257-317.
- (4) Pack, D. W., Hoffman, A. S., Pun, S., Stayton, P. S. (2005) Design and development of polymers for gene delivery. *Nature Rev. Drug Discovery* 4, 581-593.
- (5) Ewert, K., Slack, N. L., Ahmad, A., Evans, H. M., Lin, A. J., Samuel, C. E., Safinya, C. R. (2004) Cationic lipid-DNA complexes for gene therapy: understanding the relationship between complex structure and gene delivery pathways at the molecular level. *Curr. Med. Chem.* 11, 133-49.
- (6) Tandia, B. M., Vandenbranden, M., Wattiez, R., Lakhdar, Z., Ruyschaert, J. M., Elouahabi, A. (2003) Identification of human plasma proteins that bind to cationic lipid/DNA complex and analysis of their effects on transfection efficiency: implications for intravenous gene transfer. *Mol. Ther.* 8, 264-273.
- (7) Escriou, V., Ciolina, C., Lacroix, F., Byk, G., Scherman, D., Wils, P. (1998) Cationic lipid-mediated gene transfer: effect of serum on cellular uptake and intracellular fate of lipopolyamine/DNA complexes. *Biochim. Biophys. Acta* 1368, 276-288
- (8) Eliyahu, H., Serval, N., Domb, A., Barenholz, Y. (2002) Lipoplex-induced hemagglutination: potential involvement in intravenous gene delivery. *Gene Ther.* 9, 850-858.

(9) Thompson, B., Mignet, N., Hofland, H., Lamons, D., Seguin, J., Nicolazzi, C., de la Figuera, N., Kuen, R. L., Meng, X. Y., Scherman, D., Bessodes, M. (2005) Neutral postgrafted colloidal particles for gene delivery. *Bioconjugate Chem.* *16*, 608-14.

(10) Mignet, N., Gregoriadis, G. (2006) *Liposome technology 3rd ed.* London, vol 2, ch.16, *in press*.

(11) Faneca, H., Simoes, S., Pedroso de Lima, M. C., (2004) Association of albumin or protamine to lipoplexes: enhancement of transfection and resistance to serum. *J. Med. Chem.* *6*, 681-692.

(12) Aoyama, Y., Kanamori, T., Nakai, T., Sasaki, T., Horiuchi, S., Sando, S., Niidome, T. (2003) Artificial viruses and their application to gene delivery. Size-controlled gene coating with glycocluster nanoparticles. *J. Am. Chem. Soc.* *125*, 3455-7.

(13) Arignon, J., Prata, C. A., Grinstaff, M. W., Barthelemy, P. (2005) Nucleic acid complexing glycosyl nucleoside-based amphiphiles. *Bioconjug. Chem.* *16*, 864-872.

(14) Nakai, T.; Kanamori, T.; Sando, S and Aoyama, Y. (2003) Remarkably Size-Regulated Cell Invasion by Artificial Viruses. Saccharide-Dependent Self-Aggregation of Glycoviruses and Its Consequences in Glycoviral Gene Delivery. *J. Am. Chem. Soc.* *125*, 8465-8475.

(15) Chabaud, P., Camblo, M., Payet, D., Serin, G., Moreau, L., Barthelemy, P., Grinstaff, M. W. (2006) Cationic nucleoside lipids for gene delivery. *Bioconjug. Chem.* *17*, 466-472.

(16) Bailey, A., Sullivan, S. M. (2000) Efficient encapsulation of DNA plasmids in small neutral liposomes induced by ethanol and calcium, *Biochim. Biophys. Acta*, *1468*, 239-252.

(17) Roux, D., Chenevier, D., Pott, T., Navailles, L., Regev, O., Mondain Monval, O. (2004) Conception and realization of a non-viral and non-cationic DNA vector, *Curr. Med. Chem.*, *11*, 2, 169-177.

-
- (18) Tranchant, I., Mignet, N., Crozat, E., Leblond, J., Girard, C., Scherman, D., Herscovici, J. (2004) DNA complexing lipopolythiourea. *Bioconjug Chem.* *15*, 1342-8.
- (19) Leblond, J., Mignet, N., Leseurre, L., Largeau, C., Bessodes, M., Scherman, D., Herscovici, J., (2006) Design, Synthesis, and Evaluation of Enhanced DNA Binding New Lipopolythioureas *Bioconjug. Chem.* *17*, 1200-1208.
- (20) Leblond, J., Mignet, N., Largeau, C., Spanedda, M-V., Seguin, J., Scherman, D., Herscovici, J., (2007) Lipopolythioureas: a new non-cationic system for gene transfer. *Bioconjug Chem.* *18*, 484-493.
- (21) Benito, J.M., Gomez-Marcia, M., Jimenez Blanco, J.L., Ortiz Mellet, C., Garcia Fernandez, J.M. (2001) Carbohydrate-based receptors with multiple thiourea binding sites. Multipoint hydrogen bond recognition of dicarboxylates and monosaccharides. *J. Org. Chem.* *66*, 1366-72.
- (22) Takemoto Y. (2005) Recognition and activation by ureas and thioureas: stereoselective reactions using ureas and thioureas as hydrogen-bonding donors. *Org. Biomol. Chem.* *3*, 4299-306.
- (23) Gómez, D.E., Fabbriizzi, L., Licchelli, M., Monzani, E. (2005) Urea vs. thiourea in anion recognition. *Org. Biomol. Chem.* *3*, 1495-500.
- (24) Bühlmann, P., Nishizawa, S., Xiao, K.P., Umezawa, Y. (1997) Strong hydrogen-bond mediated complexation of $H_2PO_4^-$ by neutral bis-thiourea hosts. *Tetrahedron* *53*, 1647-1654.
- (25) Still, W. C., Kahn, M., Mitra, A. (1978) *J. Org. Chem.* *43*, 2923-2926.
- (26) Balacco, G. (1994) SwaN-MR: A Complete and Expansible NMR Software for the Macintosh. *J. Chem. Inf. Comput. Sci.* *34*, 1235-124.
- (27) Scudiero, D. A., Shoemaker, R. H., Paull, K. D., Monks, A., Tierney, S., Nofziger, T. H., Currens, M. J., Seniff, D. Boyd, M. R. (1988) Evaluation of a soluble tetrazolium/formazan assay for

cell growth and drug sensitivity in culture using human and other tumor cell lines. *Cancer Res.* 48, 4827-4833.

(28) Guidance Document on the Recognition Assessment and Use of Clinical Signs as Human Endpoints for Experimental Animals Used in Safety Evaluation. OECD Environmental Health and Safety Monograph. Series on Testing and Assessment No. 19 (2000).

(29) Nicolazzi C, Mignet N, de la Figuera N, Cadet M, Ibad RT, Seguin J, Scherman D, Bessodes M. (2003) Anionic polyethyleneglycol lipids added to cationic lipoplexes increase their plasmatic circulation time *J. Controlled Release* 88, 429-443.

(30) Mac Kay, A. F., Tarlton, E. J., Petri, S. I., Steyermark, P. R., Mosley, M. A. (1958) Amino Acids. V. 1,3-di-(ω -carboxyalkyl)-thioureas and their chemistry, *Can. J. Chem.* 80, 1510-1517.

(31) Byk, G., Dubertret, C., Escriou, V., Jaslin, G., Rangara, R., Pitard, B., Crouzet, J., Wils, P., Schwartz, B., Scherman, D. (1998) Synthesis, activity, and structure-activity relationship studies of novel cationic lipids for DNA transfer. *J. Med. Chem.* 41, 224-235.

(32) Boussif, O., Lezoualc'h, F., Zanta, M. A., Mergny, M. D., Scherman, D., Demeneix, B., Behr, J. P. (1995) A versatile vector for gene and oligonucleotide transfer into cells in culture and in vivo: polyethylenimine. *Proc. Natl. Acad. Sci. U.S.A.* 92, 7297-301.

(33) Campbell, M. J. (1995) Lipofection reagents prepared by a simple ethanol injection technique. *Biotechniques* 18, 1027-1032.

(34) Bangham, A. D., Standish, M. M., Watkins, J. C. (1965) Diffusion of univalent ions across the lamellae of swollen phospholipids. *J. Mol. Biol.* 13, 238-252.

(35) Delattre, J., Couvreur, P., Puisieux, F., Philippot, J.R., Schuber, F. (1993) Les liposomes, Aspects technologiques, biologiques et pharmacologiques. *Ed. Inserm* 54-62.

-
- (36) Tranchant I., Thompson B., Nicolazzi C., Mignet N., Scherman D. (2004) Physicochemical optimisation of plasmid delivery by cationic lipids. *J Gene Med.* S24-35.
- (37) Nicolazzi, C., Mignet, N., de la Figuera, N., Cadet, M., Ibad, R. T., Seguin, J., Scherman, D., Bessodes, M. (2003) Anionic polyethyleneglycol lipids added to cationic lipoplexes increase their plasmatic circulation time. *J. Controlled Release* 88, 429-443.
- (38) Simberg, D., Weisman, S., Talmon, Y., Faerman, A., Shoshani, T., Barenholz, Y. (2003) The role of organ vascularization and lipoplex-serum initial contact in intravenous murine lipofection. *J. Biol. Chem.* 278, 39858-65.
- (39) Takakura, Y., Mahato, R. I., Hashida, M. (1998) Extravasation of macromolecules. *Adv. Drug Deliv. Rev.* 34, 93-108.
- (40) Gilot, D., Miramon, M. L., Benvegna, T., Ferrieres, V., Loreal, O., Guguen-Guillouzo, C., Plusquellec, D., Loyer, P. (2002) Cationic lipids derived from glycine betaine promote efficient and nontoxic gene transfection in cultured hepatocytes. *J. Gene Med.* 4, 415-27.
- (41) Ferrari, E., Rusalov, D., Enas, J., Wheeler, C. (2002) Synergy between cationic lipid and co-lipid determines the macroscopic structure and transfection activity of lipoplexes. *Nucl. Acids Res.* 30, 1808-1816.
- (42) Sjibren O., Janout, V., DiGiorgio, A., Young, M. C., Regen, S. L. (2000) Detection of Nonideal Mixing of Phospholipids in Fluid Bilayers *J. Am. Chem. Soc.*, 122, 1200 -1204.
- (43) Gabizon, A., Shmeeda H, Zalipsky S. (2006) Pros and cons of the liposome platform in cancer drug targeting. *J. Controlled Release.* 16, 175-183.
- (44) Blanco, J. L., Bootello, P., Benito, J. M., Mellet, O., Fernandez, G. Urea-, thiourea-, and guanidine- as phosphate binders in water. (2006) *J Org Chem.* 71, 5136-43.

Scheme title

Scheme 1: Preparation of lipopolythiourea (i) NHS, DCC, THF, RT. (ii) TFA, CH₂Cl₂, RT. (iii) DIPEA, **5**, CH₂Cl₂, RT.

Scheme 1

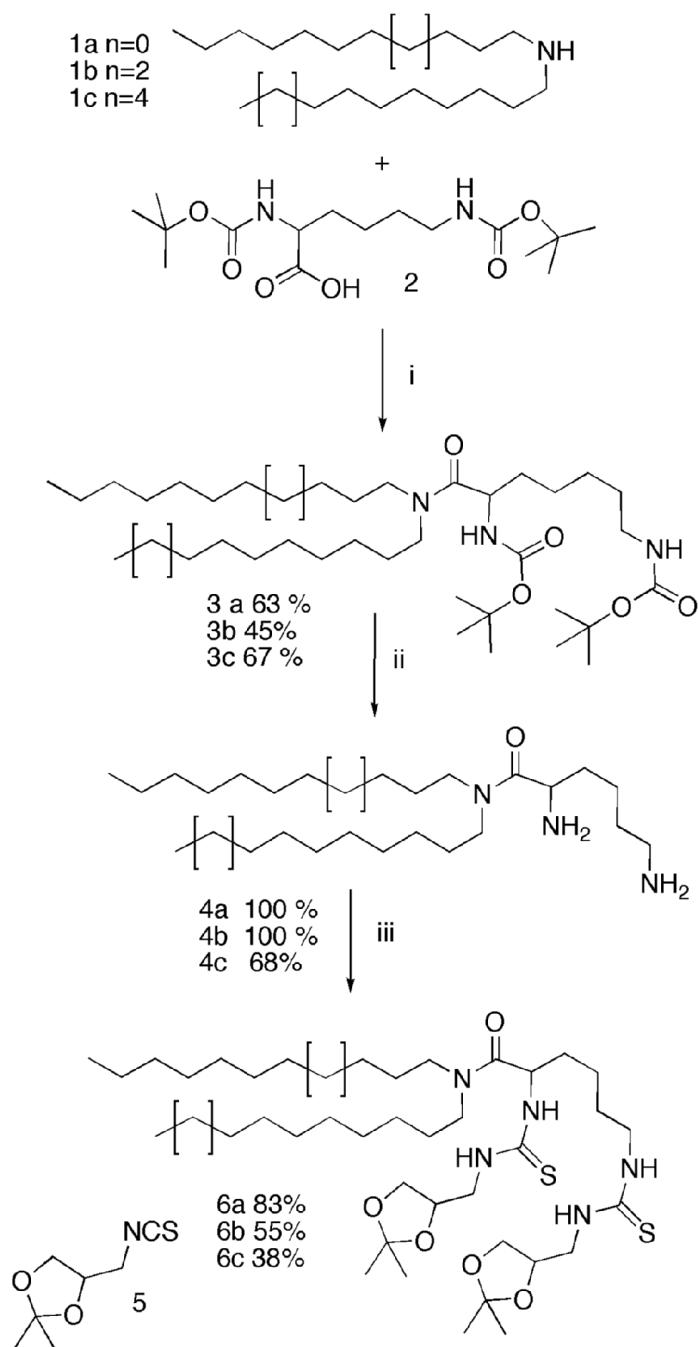


Figure Captions

Figure 1.

Complexes LPT/DNA (20 LPT/P, prepared by ethanolic injection) as well as the cationic lipid RPR209120 were incubated for 6 and 24h in 10% serum at 37°C, then treated with SDS 2% and EDTA 0.5M and loaded on an 1% agarose gel containing in TBE (1x) 0.05% of SDS (80V/cm). Complexes were revealed by ethidium bromide staining.

Figure 2.

2-{3-[Bis-(3-amino-propyl)-amino]-propylamino}-N-ditetradecylcarbamoyl methyl-acetamide (RPR209120) and Polyethyleneimine (PEI) used as reference transfecting agents.

Figure 3.

Hydrodynamic diameter (lozenge symbols, right scale) and gene transfection (bars, left scale), measured by luciferase activity of LPT/DNA complexes at 20 and 40 TU/P ratios after 48 h incubation on B16 cells in presence of serum. LPT/DNA complexes were prepared by ethanolic injection at 20 and 40 TU/P in H₂O. RPR209120/DOPE/DNA lipoplexes was prepared N/P ratio = 8 in NaCl 150 mM. PEI/DNA polyplexes were prepared at N/P = 15 in glucose 5%. The complexes were then added to the cells in DMEM + 10% FBS at 37°C. Results are expressed as the mean of an experiment performed in triplicate errors bars indicating standard error of the mean.

Figure 4.

IC₅₀ of **6c**/DPPC formulated by (*left*) ethanolic injection (IC₅₀ = 14.96μM) and (*right*) hydration method, determined by a MTT test *in vitro*.

Figure 5.

Biodistribution of rhodamine-labeled particles 2h after intravenous injection in mice bearing 3LL subcutaneous tumors. RPR209120/DOPE were prepared in NaCl 150 mM and **6c**/DPPC were prepared

by hydration film method. Results are expressed as the percentage of the injected dose. (** $p < 0.01$ by the Mann-Whitney U - test, significant in blood and lung, not in the tumor and RES)

Figure 6.

Comparison of gene transfection levels of lysine derivatives LPT (white bars) and previously published LPT (20) (grey bars) according to their ratio TU/P and chain length. Lysine LPT structures 10C, 12C, 14C correspond to **6a**, **6b**, **6c** respectively. Previously published LPT structures 10C, 12C, 14C correspond to compounds Ic, Ib and Ia represented on figure 7. Luciferase activity of LPT/DNA complexes at 20 and 40 TU/P ratios was measured after 48 h incubation on B16 cells in presence of serum. RPR209120/DOPE/DNA lipoplexes were prepared N/P ratio = 6 in NaCl 150 mM. Results are expressed as the mean of an experiment performed in triplicate errors bars indicating standard error of the mean.

Figure 7.

Previously published lipopolythiourea and thiourea compounds: I-II (20), III-V (19), VI (44).

Figure 1

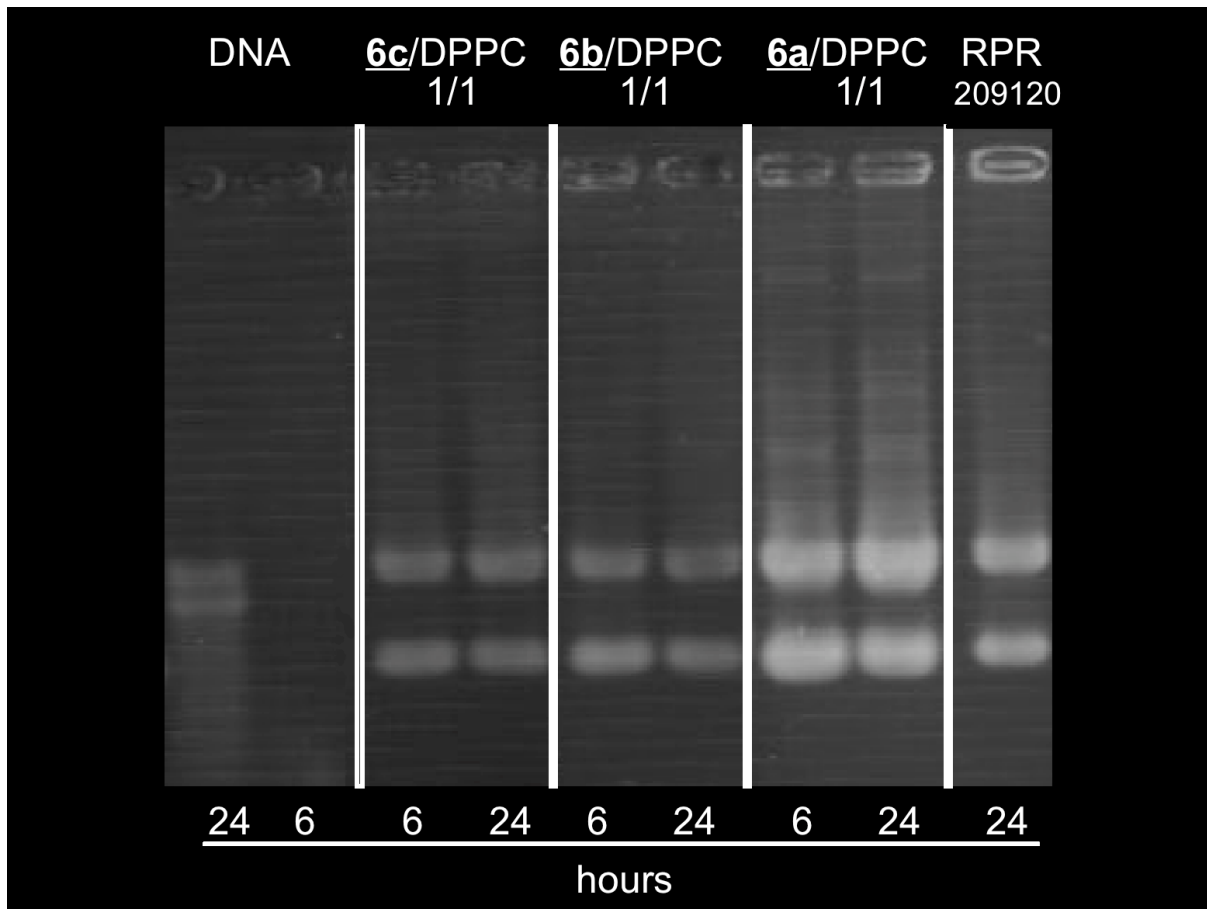


Figure 2

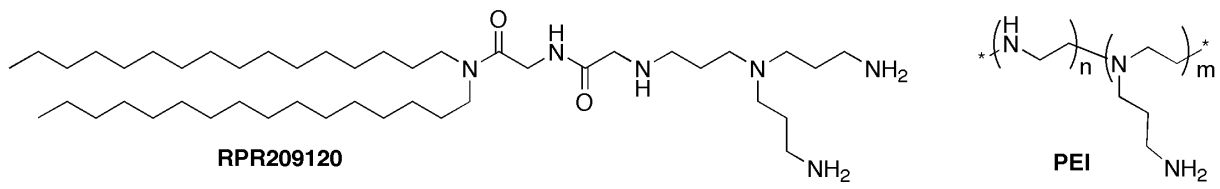


Figure 3

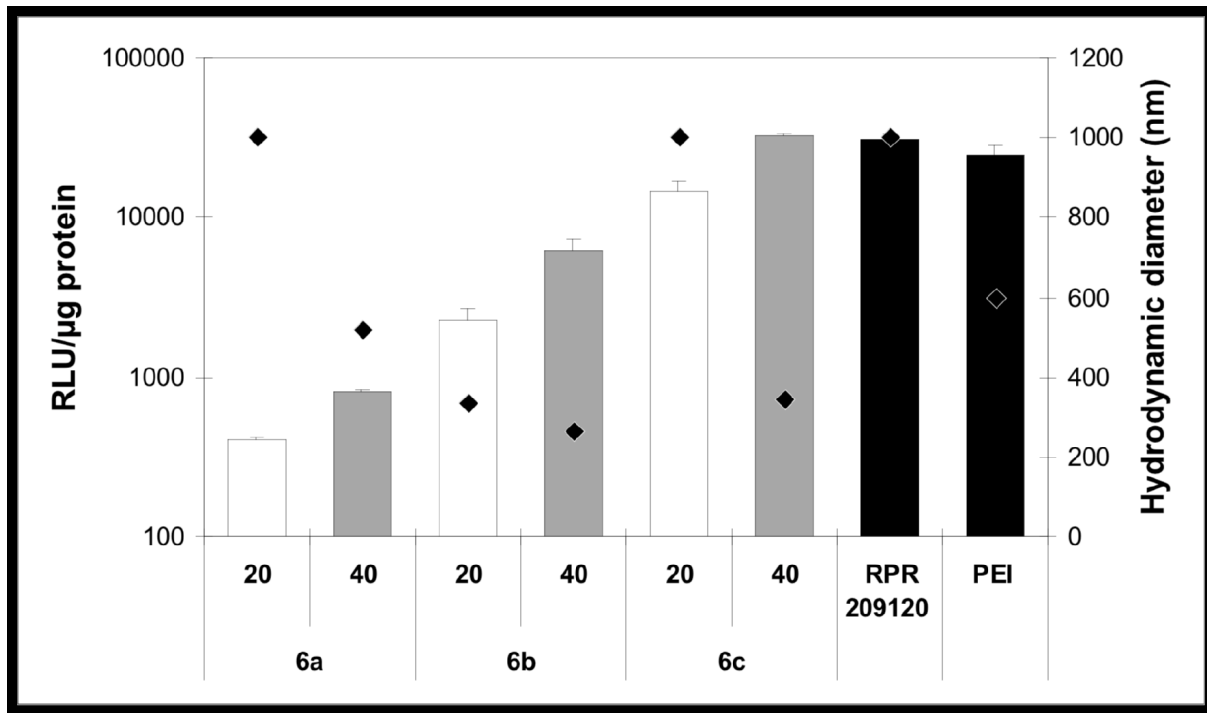


Figure 4

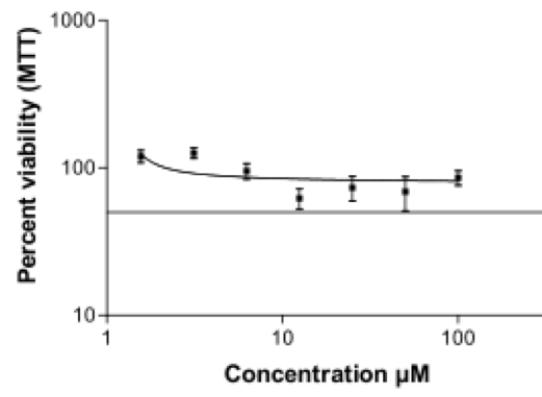
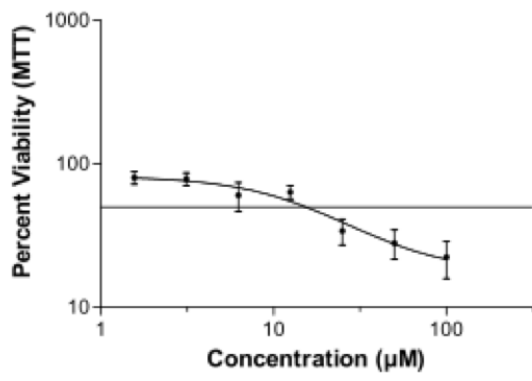


Figure 5

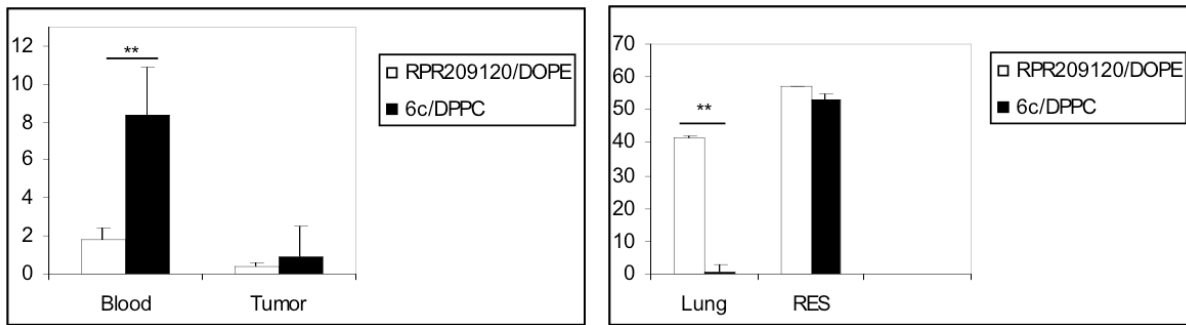


Figure 6

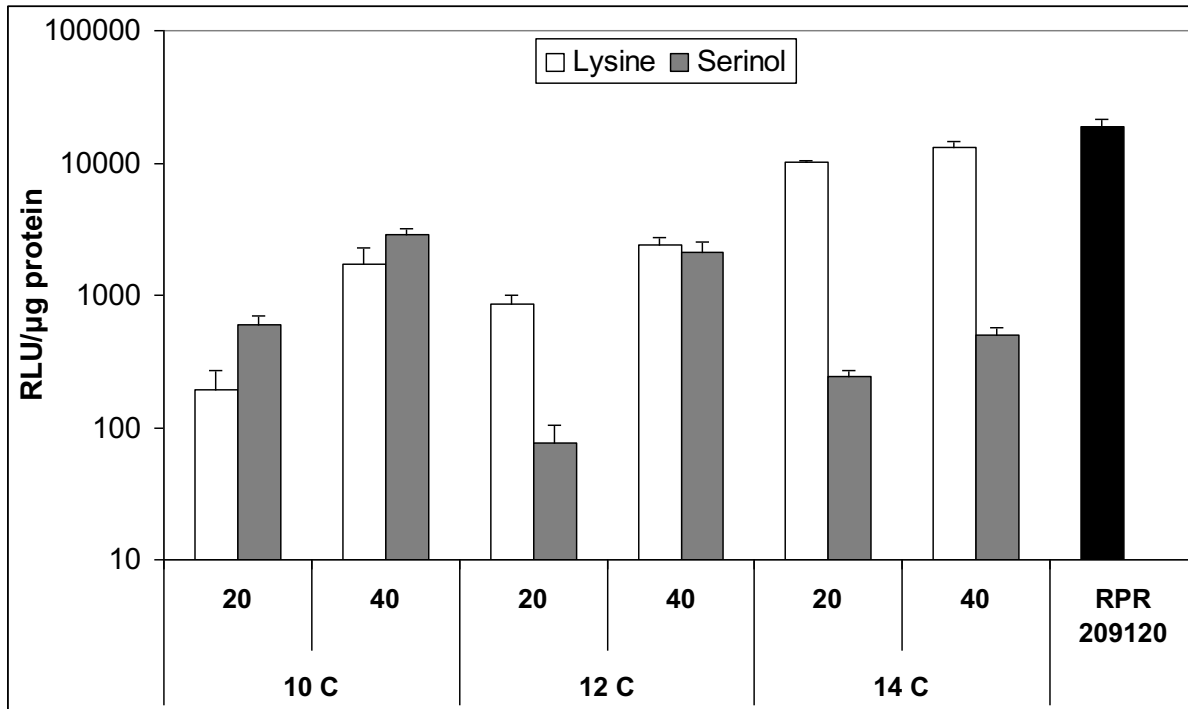
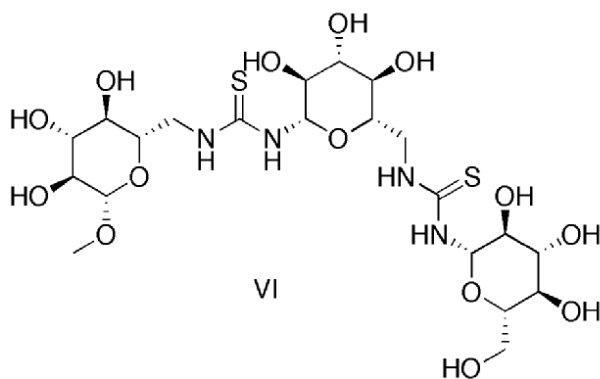
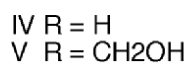
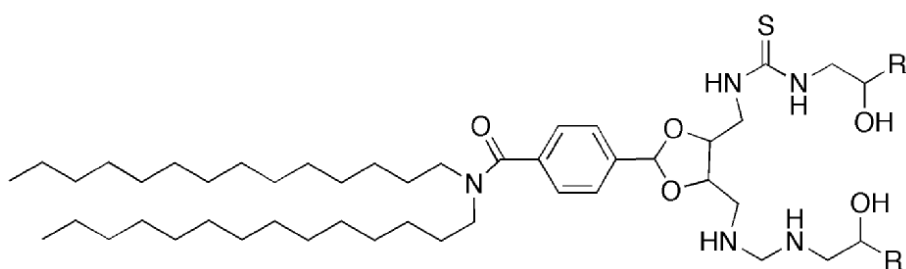
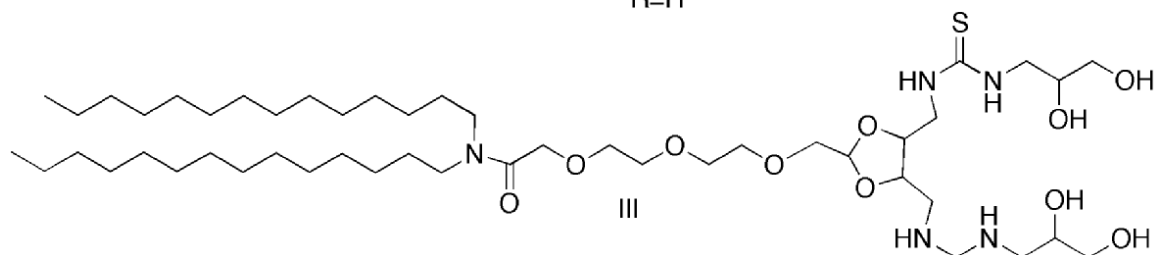
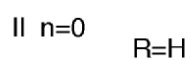
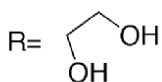
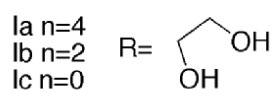
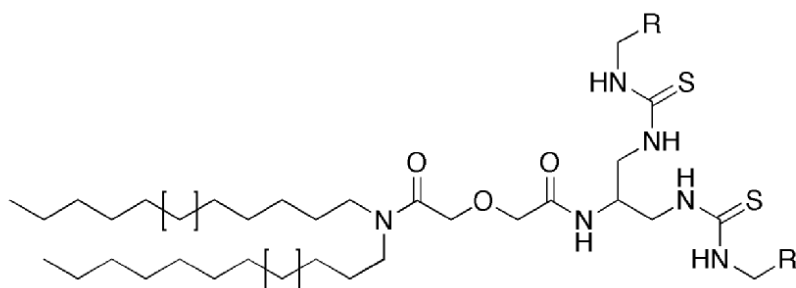


Figure 7



VI