

# Insights from Microbial Transition State Theory on Monod's Affinity Constant

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#### 9 Abstract

Microbial transition state theory (MTS) offers a theoretically explicit mathematical model for substrate limited microbial growth. By considering a first order approximation of the MTS equation one recovers the well-known Monod's expression for growth, which was regarded as a purely empirical function. The harvest volume of a cell as defined in MTS theory can then be related to the affinity concept, giving a new physical interpretation to it, and a new way to determine its value. Consequences of such a relationship are discussed.

16

17 Since the success of Monod's expression (Equation 1) to model substrate-limited microbial growth<sup>1</sup>, 18 many expressions have been proposed<sup>2</sup>, accounting for a range of phenomena including substrate inhibition and population density effects <sup>3,4</sup>. All of these expressions rely on empirical rules, 19 differently to enzymology for which analogues of Monod and Haldane expressions have been 20 mathematically derived <sup>5</sup>). Microbial transition state theory <sup>6</sup> recently introduced a new expression 21 22 for microbial growth based on the statistics of molecules distribution in medium inspired from 23 chemical transition state theory. In this communication we explore the physical meaning of the affinity concept through the lens of MTS theory and particularly show how it may provide a novel 24 25 interpretation of Monod's growth function.

26 Equation 1

27

$$\mu_{max} \frac{s}{K_s + s}$$

Equation 1 represents the Monod growth function, where  $\mu_{max} [1/day]$  is the maximal growth rate, 28 s [g/L] represents the substrate concentration of the medium and  $K_s [g/L]$  is known as the 29 "affinity constant". Earlier works on kinetics <sup>7,8</sup> show the differences in reported literature values for 30 31 the affinity constant for the same species: these differences are explained by culture history, quality 32 of the experimental data, and posterior data analysis. However little to no consensus can be found in the literature on its interpretation. Furthermore in a review of theoretical derivations of the Monod 33 growth function <sup>9</sup> the author concludes that no clear interpretation may be given to the affinity 34 35 constant. A revision of the affinity concept in Microbiology was made by Button <sup>10</sup>, where fourteen different expressions for affinity are documented. The concept is largely influenced by the Michaelis-36 37 Menten model for enzyme kinetics interpretation of affinity from receptor and ligand binding sites, since Monod's expression for growth is mathematically equivalent to the Michaelis-Menten 38 39 expression. As stated by Monod himself, Monod's growth function is purely empirical, while Michaelis-Menten expression has a rigorous theoretical justification<sup>5</sup>, thus one might wonder if the 40 concept of affinity for representing cell growth has a solid conceptual ground. 41

42 MTS theory relates the growth rate to the amount of energy available to perform cellular work. The central idea of bioenergetics is that the energy consuming anabolism can only be thermodynamically 43 44 feasible if it is coupled with an energy yielding catabolism. The overall reaction resulting from the coupling is known as metabolism <sup>11</sup>. The formulation and complexity of both catabolism and 45 46 anabolism vary greatly depending on the objective the modeller has in mind. On the one hand, when 47 describing the metabolic pathways within a specific microbial species, the formulation takes into 48 account ATP formation and intra cellular intermediates and quickly becomes a very complex web, e.g.<sup>12</sup>. On the other hand if one is interested in observing the general metabolism of a culture at a 49 macroscopic level then the situation simplifies to just a couple of reactions <sup>11</sup>. We will focus on the 50 51 latter.

Let us consider a first reaction representing catabolism (Equation 2), a second reaction representing anabolism (Equation 3) then a linear combination of the two creates metabolism (Equation 4): by completing  $\lambda$  times the catabolism the energy requirements of the global metabolic reaction are fulfilled <sup>13</sup> (its negative free enthalpy constitutes the driving force for growth).

56 Equation 2

57 
$$E_d + aE_a \rightarrow bP$$
;  $\Delta_r G_{cat} < 0 \quad \langle \cdot \lambda > 0 \rangle$ 

58 Equation 3

59

 $dP \rightarrow B_x + c E_a$ ;  $\Delta_r G_{an} > 0$ 

60 Equation 4

61 
$$\lambda E_d + (\lambda a - c)E_a \rightarrow B_x + (\lambda b - d)P$$
;  $\Delta_r G_{met} = \lambda \Delta_r G_{cat} + \Delta_r G_{an} < 0$ 

where  $E_d$ ,  $E_a$ , and P stand for electron donor, electron acceptor, and products, respectively.  $B_x$ represents an equivalent biomass unit, for instance  $B_x = CH_{1.8}O_{0.5}N_{0.2}$  is a generic composition of one C-mole of biomass <sup>14</sup>. a, b, c, d are stoichiometric coefficients. Finally  $\Delta_r G$  represents the Gibbs free energy variation for each reaction.

The reader should notice that  $\lambda$  is the inverse of the yield as usually expressed  $(y_{x/s})$  in microbiology as shown in the equation 5.

68 Equation 5

69

$$y_{x/s} = \frac{1}{y_{s/x}} = \frac{1}{\lambda}$$

70  $y_{x/s}$  represents how many moles of biomass are formed per mole of substrate consumed, 71 conversely  $y_{s/x} = \lambda$  represents how many moles of substrate are being consumed per mole of 72 biomass formed. The methods reviewed by Kleerebezem et al. <sup>11</sup> allow computing  $\lambda$  from mass 73 balanced reactions with examples coming from a variety of biological process.

MTS theory demonstrates on a theoretically explicit ground a growth rate expression ( $\mu$ ) of a culture of bacteria limited by an electron donor in perfectly mixed conditions <sup>6</sup>. More precisely, if we denote by *s* the concentration of the limiting electron donor and *x* the concentration of the species then these two concentrations are dynamically related by: 78 Equation 6

79 
$$\dot{x} = \mu(s)x = \mu_{max} \exp\left(\frac{-\lambda}{V_h s}\right)x$$

80 where  $V_h$ , known as the harvest volume, represents the volume to which each microbe has access in 81 order to harvest the substrate *s* during the time between two cell divisions. It is worth pointing out 82 that the harvest volume is an average characteristic.

83



84

Figure 1. Example of plots of equations 1 and 6, with the values chosen such that  $K_s = \frac{\lambda}{V_h}$ . The measurement of the harvest volume from growth experiments can be obtained in an analogous fashion to the determination of the affinity constant: by noting s\* the value of substrate concentration at which the growth rate is  $e^{-1}\mu_{max}$  (represented by the black diamond) one obtains  $V_h$  by the formula  $V_h = \frac{1}{y_{x/s} s^*}$ , similarly to the  $K_s$  value identified as the concentration for which the specific growth rate  $\mu$  is equal to  $\frac{\mu_{max}}{2}$  in the Monod expression, (represented by the black square).

90 If one considers a first order approximation of the exponential function near zero (see 91 supplementary material) then one recovers Monod's expression of growth:

92 Equation 7

93 
$$\mu_{max}e^{-\frac{\lambda}{V_h \cdot s}} \approx \mu_{max}\frac{s}{s + \frac{\lambda}{V_h}} = \mu_{max}\frac{s}{s + \frac{1}{y_{x/s}V_h}}$$

The approximation holds true for high substrate concentrations. More precisely, it can be shown that the two curves differ by less than 10% for  $s \ge 1.92 K_s$ , (see supplementary material). In Figure 1 the graphical comparison of both growth functions can be seen for a given set of parameters. The MTS growth function is approximated very well by the Monod growth function, which is reassuring from a practical point of view: in a re-examination of the kinetics of *Escherichia coli* <sup>15</sup> different empirical substrate limiting expressions- all of them with a Monod-like shape- were compared and no difference was found in the identifiability of their parameters.

Note also that in equation 7,  $\frac{1}{\gamma_{x/s} V_h}$  replaces the  $K_s$  parameter of Monod's expression. In that sense 101 102 the affinity constant can be interpreted as a decreasing function of the harvest volume of the cell and its yield per mole of substrate. On one hand, associating low K<sub>s</sub> values to large harvest volumes is 103 104 well in line with our understanding of the affinity concept, since a cell that can harvest substrate 105 molecules in a more extended region should be less substrate limited. On the other hand, the fact 106 that a low  $K_s$  value could be due to a higher conversion yield of substrate to biomass sheds a new 107 light on the affinity concept. The order of magnitude of  $V_h$  can be seen from Table 1 for some 108 literature references for E. Coli ML 30. In the cases where no yield was reported the energy dissipation method <sup>11</sup> can be used as illustrated in table 1 and supplementary material. For 109 computing the yield a unique biomass formula was used ( $CH_{1,8}O_{0,5}N_{0,2}$ ). However, for each case, 110 111 the biomass composition could be different and, consequently, the yield, thus contributing to the explanation of the observed variability of K<sub>s</sub>. 112

$K_{\rm c}$ reported $[\mu q/L]$	λ (gS/gX)	$V_{h}\left[L/aX\right]$	$V_{\rm h}$ [ $\mu m^3$ /cell]	Radius [um]
for E. Coli ML 30	*Estimated by Energy		(cell dry weight:	of a sphere
	dissipation Method		$2.8 \cdot 10^{-13} \text{ gr/cell}$	of volume
	(Supplementary Material)		(Ref : <sup>16</sup> BNID:	$V_h$ .
	**Measured during		103904 Neidhart et	
	experiment			
			al. )	

33 (Ref: <sup>17</sup> )	$\lambda^* = 1.89$	$4.91 \cdot 10^4$	$1.6 \cdot 10^{7}$	156
33 (Ref: <sup>17</sup> )	$\lambda^* = 1.88$	$4.85 \cdot 10^4$	$1.6 \cdot 10^{7}$	156
53 (Ref: <sup>15</sup> )	$\lambda^* = 1.88$	$3.02 \cdot 10^4$	9.94 · 10 <sup>6</sup>	133
72 (Ref: <sup>15</sup> )	$\lambda^* = 1.88$	$2.22 \cdot 10^4$	7.32 · 10 <sup>6</sup>	120
76 (Ref: <sup>18</sup> )	$\lambda^{**} = 2.22$	$2.92 \cdot 10^4$	8.19 · 10 <sup>6</sup>	125
90 (Ref: <sup>18</sup> )	$\lambda^{**} = 2.22$	$2.47 \cdot 10^4$	6.91 · 10 <sup>6</sup>	118
100 (Ref: <sup>18</sup> )	$\lambda^{**} = 2.22$	$2.22 \cdot 10^4$	$6.22 \cdot 10^{6}$	114
132 (Ref: <sup>18</sup> )	$\lambda^{**} = 2.22$	$1.68 \cdot 10^4$	$4.71\cdot 10^6$	104
125 (Ref: <sup>18</sup> )	$\lambda^{**} = 2.22$	$1.77 \cdot 10^4$	$4.98\cdot 10^6$	105

113

**114** Table 1. Literature values (Table 2<sup>8</sup>) of  $K_s$  and calculation of  $V_h = \frac{\lambda}{K_s}$ , for different chemostat experiments using hexoses

115 as substrates.

116

117 On a more conceptual ground, the MTS approach proposes a way to revisit our current perception of 118 the "affinity-concept" of a microbial culture for a given substrate. It offers an alternative view of the 119 microbial affinity notion than its enzymatic analogue related to Michaelis-Menten theory. It unravels 120 a contribution that is related to the yield (mole of biomass formed per mole of substrate consumed) 121 from another that represents the capacity of the microbial culture to explore a fraction of its 122 surroundings in order to harvest substrate ( $V_h$  term). To this extent, it allows to compute the affinity 123 constant from the knowledge of the yield and the harvest volume, which is a completely new 124 approach to determining this constant.

This analysis thus plants a seed towards a more physically grounded view of affinity than earlier proposals made from attempts to theoretically derive Monod's equation<sup>9</sup>. The physical interpretation of the affinity concept raises new opportunities to analyse and experimentally challenge the meaning of the  $V_h$  parameter. Particularly interesting would be to assess to which extent  $V_h$  constitutes an intrinsic trait of the microbial culture, or if extrinsic attributes associated to the culture conditions

130	(such as agitation,	, viscosity or ionic force	e) could also significantl	y influence its value.	Such questions
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131 remain open and obviously await further studies.

#### 132 Conflict of Interest

133 The authors declare no conflict of interest.

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#### 182 Author Contributions

- 183 P.U. wrote the main manuscript text. T.B and E.D. conceived the harvest volume calculations. A.R.
- and P.U. derived the mathematical expressions. J.H. supervised the work. All authors participated in
- 185 the online meetings and corrected the manuscript.

# Supplementary Material

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### Approximation

Consider the first order approximation of the exponential function:

$$\exp(x) = 1 + x + o(x) \tag{1}$$

and the first order approximation:

$$\frac{1}{1+x} = 1 - x + o(x) \tag{2}$$

Then rewrite MTS expression as follows:

$$\mu_{max} exp\left(\frac{-\lambda}{V_h s}\right) \approx \mu_{max}\left(1 - \frac{\lambda}{V_h s}\right) \tag{3}$$

$$\approx \mu_{max} \frac{1}{1 + \frac{\lambda}{V_h s}} \tag{4}$$

$$=\mu_{max}\frac{s}{s+\frac{\lambda}{V_h}}\tag{5}$$

### Comparison of both expressions

The substrate limitation range can be studied through the ratio of both growth functions, shown in expressions (6) and (7), respectively.

$$\mu_{max} \frac{s}{s + \frac{\lambda}{V_h}} \tag{6}$$

$$\mu_{max} exp\left(\frac{-\lambda}{V_h s}\right) \tag{7}$$

Noting  $K_s := \frac{\lambda}{V_h}$  One then considers the ratio:

$$R(s) = \frac{\exp\left(\frac{-K_s}{s}\right)}{\frac{s}{s+K_s}} \tag{8}$$

Note that ratio (8) does not depend on  $\mu_{max}$ . It can be shown that  $R(s) \in (0, 1)$ , by using the well known inequality  $\exp(x) < \frac{1}{1-x}$  for x < 1. Which is valid since the term inside the exponential is negative:

$$R(s) \le \frac{1}{1 - \frac{-K_s}{s}} \frac{K_s + s}{s} = 1$$
(9)

The change of variables  $u = \frac{s}{K_s}$  is used to analyse the expression R(s), which gives equation (10).

$$F(u) = \exp\left(-\frac{1}{u}\right)\frac{u+1}{u} \tag{10}$$

$$F'(u) = \exp\left(-\frac{1}{u}\right)\frac{1}{u^2}\frac{u}{u+1} + \exp\left(-\frac{1}{u}\right)\frac{1}{(u+1)^2} > 0$$
(11)

Since F is monotonic, one gets that there exists a unique  $u^*$  such that  $\exp\left(-\frac{1}{u^*}\right)\frac{u^*+1}{u^*}=0.9$ , implying a unique  $s^*:=u^*K_s$ , such that  $R(s^*)=0.9$ . From the former it can be seen that for each  $K_s$  MTS expression approximates to 90 % of the Monod expression whenever  $s \geq u^*K_s$ .

The curve  $s \mapsto R(s)$  is shown in figure 1, for different  $K_s$  values of table 1 of the manuscript. One can see that  $u^* \approx 1.92$  therefore  $s \geq 1.92K_s$  then  $R(s) \geq 0.9$ .



Figure 1: R(s) curve for several  $K_s$  values obtained for E. Coli ML 30. The asterisk \* represents  $K_s$  values evaluated in expression R(s), note that  $R(K_s) \approx 0.74$ . The plus sign + represents the point where the R(s) = 0.9.