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β -Cyclodextrin PAMAM dendrimer: how to overcome the tumbling process for getting fully available host cavities

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Keywords: β -cyclodextrin • Click Chemistry • PAMAM • dendrimer • tumbling process

Abstract: The synthesis of a G0 polyamidoamide (PAMAM) dendrimer end-capped with four β -cyclodextrin (β -CD) units, named as **Tetra-CD** was performed by using copper(I)-catalyzed alkyne-azide cycloaddition (CuAAC). This new platform presents a considerable high water-solubility in comparison to native β -CD. Tumbling process involving half of the CD cavities in D₂O was evidenced by ¹H NMR spectroscopy. Interaction of adamantane with **Tetra-CD** has been fully investigated by 1D/2D NMR techniques and mass spectrometry besides a joint study on new β -CD monomers (named as **A** and **B**) and on a dimer (**Di-CD**) and a trimer (**Tri-CD**). The obtained results demonstrate that the tumbling effect can be shifted in the presence of an appropriate hydrophobic guest resulting in the full host-ability of the CD units in water for that guest.

Introduction

Cyclodextrins (CDs) are a series of natural cyclic oligosaccharides comprising 6, 7 or 8 D-(+)-glucopyranose units linked by α -1,4-linkages, named as α -, β - or γ -CD, respectively,^[1] represented as a truncated cones with a hydrophilic exterior and a hydrophobic inner cavity. Their water-solubility is not correlated with their size but with the ability of their hydroxyl groups to form hydrogen bonds with water. It can be improved by eventual chemical modifications on these OH groups situated on the edges of the conical cylinder. Thanks to its size, hydrophobicity and hydrogen bonding of its inner cavity, β -CD can accommodate diverse guest molecules. Its low cost, water-solubility and biocompatible properties^[2] make it a molecule of choice for the construction of supramolecular systems.^[3] The interaction of CD with non-water-soluble molecules operates *via* a dynamic equilibrium process of host-guest complexation, following which the guest molecules continuously associate and dissociate from the CD cavity. Due to its inner cavity diameter of around 6.5 Å, β -CD has been found to be particularly attractive for preparation of drugs delivery systems.^[4]

Chemical modification of native β -CDs have been performed *via* reactions on the primary or secondary hydroxyl groups,^[5] leading to decorated β -CDs, formation of β -CD polymers,^[6] or generation of CD nanosponges.^[7] Among the used synthetic techniques, the copper(I)-catalysed alkyne-azide cycloaddition (CuAAC) “click” reaction^[8] has emerged as one of the most valuable synthetic

tools in the development of a wide variety of CD derivatives for biological applications in various fields, due to its simplicity, efficiency, and robustness properties.^[9] CuAAC is used for cycloaddition of the mono azido cyclodextrin (**β -CD-N₃**), obtained from the mono-(O-6)-tosylated β -CD,^[10] on the appropriate alkyne.^[11] In that way, a hydrophobic moiety can be grafted on one primary hydroxyl function of β -CD for obtaining CD monomers^[12] or more complex assemblies^[13] with modulated inclusion and transport properties. Besides, the use of hydrophilic dendritic structures has been proved to be a suitable strategy for obtaining highly soluble active ingredients in water.^[14]

In the present work, we used this strategy for designing a new platform, named as **Tetra-CD**, based on a central G0 PAMAM dendrimer (**IV**) substituted by 4 β -CD units *via* spacers formed by CuAAC. The presence of 4 peripheral β -CD units confers to this compound a high potential of guest accommodation. The PAMAM core was chosen because it could contribute to great solubility in water for **Tetra-CD**, without being a competitive encapsulating site contrarily to what was observed with higher generations of PAMAM.^[15] We used 2D ROESY NMR experiments to fully characterize the pristine platform and its inclusion complexes formed with adamantane sodium carboxylate (**AdCOONa**) and the poorly water-soluble adamantane carboxylic acid (**AdCOOH**). Adamantane was selected as a model guest molecule because it presents a high association constant with β -CD in water, of around 5.10⁴ M⁻¹ at room temperature.^[16] Besides, diverse molecules that are too voluminous to fit the β -CD cavity can be functionalized by an adamantyl moiety, offering them the possibility to be connected to inclusion complexes.^[17] Interaction of adamantyl unit with **Tetra-CD** was discussed in light of results obtained with previously reported trimer and dimer,^[18] named here as **Tri-CD** and **Di-CD**, respectively, as well as with new mono-CDs called **A** and **B**, used as assessment systems. The CD substituents of **A** and **B** are of similar nature to that of the spacers used for **Tri-CD** and **Di-CD**, and for **Tetra-CD**, respectively (see **scheme 1**). In that way, comparisons were made independently of the nature of the spacer.

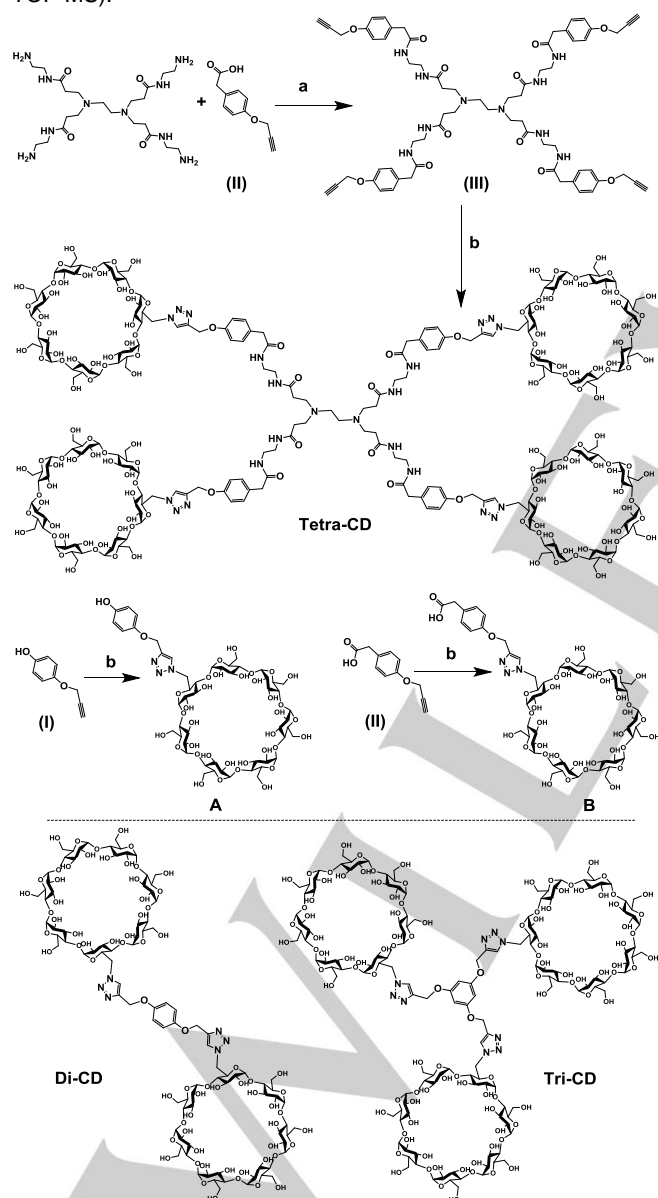
Results and Discussion

Synthesis and characterization of β CD-derivatives

The synthetic routes used for the synthesis of the β -CD derivatives are summarized in **Scheme 1**. For the construction of

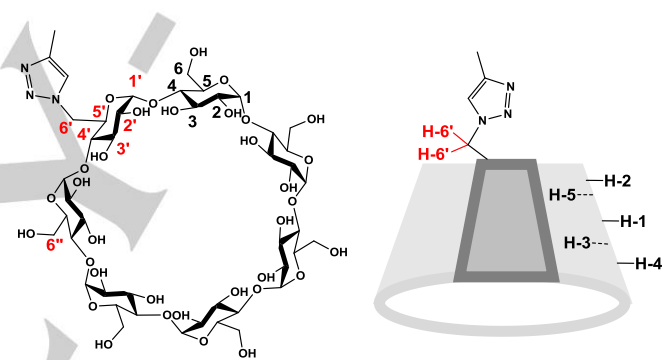
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Tetra-CD, the tetra-alkyne precursor (**III**) was prepared via amide bond formation between the amino groups of PAMAM G0 and carboxylic functions of **II**, using a mixture of EDC-HCl/HOBt as coupling reagents.^[19] The target molecules **Tetra-CD**, **A** and **B** were obtained using CuAAC in mild conditions with Cu(I) as catalyst. Catalytic Cu(I) species can be generated from Cu(0) as the source,^[20] but also *in situ* from a mixture of CuSO₄ and sodium ascorbate.^[21] This last method has been preferred because it excludes the formation of any by-products such as diacetylene and may be even applied in aqueous systems in presence of oxygen. Each compound was isolated in high yield (90 to 98%) after purification by size exclusion chromatography (SEC). Elemental analyses confirmed the high degree of purity for the obtained compounds, except for an extra-amount of water, namely about 6 water molecules per β -CD unit for **Tetra-CD**, and up to 10 for **A** and **B**. All compounds were characterized by electrospray ionisation time-of-flight mass spectrometry (ESI-TOF-MS).



Scheme 1. Top: synthetic routes to the target molecules. a. EDC-HCl/HOBt/DMF, b. β -CD-N₃, CuSO₄: sodium ascorbate 1:3 / DMSO:H₂O 5:2; Bottom: structures of Di-CD and Tri-CD.

The measured masses of the synthesized compounds were found to be in excellent agreement with the calculated ones (see “Experimental Section” part and spectra in ESI). The characterization of the new β -CD derivatives by NMR techniques (¹H, ¹³C, HMQC, COSY, NOESY) were carried out in detail in DMSO-*d*₆ (see text pp. SI-6,7 and corresponding figures). It resulted that a singularity was detected for some protons of the functionalized glucopyranose sub-unit of the β -CD cavity (see **Scheme 2**): protons H-1', H-5' and H-6' appeared at lower field than their respective correspondents H-1, H-5 and H-6 of non-functionalized sub-units (**Figure 1** top). This deshielding effect was particularly marked for H-6' protons. There was also a strong impact of the substitution on one -CH₂-OH fragment of the primary hydroxyl rim, since the signals of protons named as H-6" (two diastereotopic protons resonating at around 3.15 and 2.92 ppm) and OH-6" (around 4.30 ppm) appeared to be significantly up-shielded in comparison to their respective analogues H-6 and OH-6. Those protons were attributed to a glucopyranose sub-unit contiguous to the substituted one, their up-shielding might result from interaction with the vicinuous branch of the molecule. These observations were common to the 3 new compounds for which the CD cavities behave similarly.



Scheme 2. Representations of the CD cavity with atoms numbering (H-3 and H-5 are inner protons, H-2 and H-4 are outer protons).

Behaviour of β -CD derivatives in water

The new molecules were then investigated in water, which is the medium of choice for many applications or studies of CD derivatives.^[4,22] It is worth to notice that the isolated compounds **A** and **B** present good solubilities in water, of 37.5 mg/mL and 97.4 mg/mL, respectively, in comparison to 18.3 mg/mL for β -CD. Moreover, **Tetra-CD** exhibits remarkable high solubility in water (> 2580 mg/mL), which is of about 140 fold higher than that of β -CD.

The ¹H NMR spectrum of the compound **Tetra-CD** in D₂O showed a drastic change in the aromatic region, as compared to that obtained from DMSO-*d*₆ solution (**Figure 1**): the integration of the protons signal at around 8 ppm, the region where the triazole protons were identified in DMSO, matched only with 2 out of the 20 protons resonating in the 6.5 to 8.2 ppm region. This resulted from the previously observed tumbling process for CD tetramers and dimers in water,^[23-25] which affected the triazole protons that now appeared as a multiplet at around 8 ppm (assigned to H-triazole of branches wearing a non-reversed CD cavity) and also as a singlet centred on 7.04 ppm (assigned to the so-called

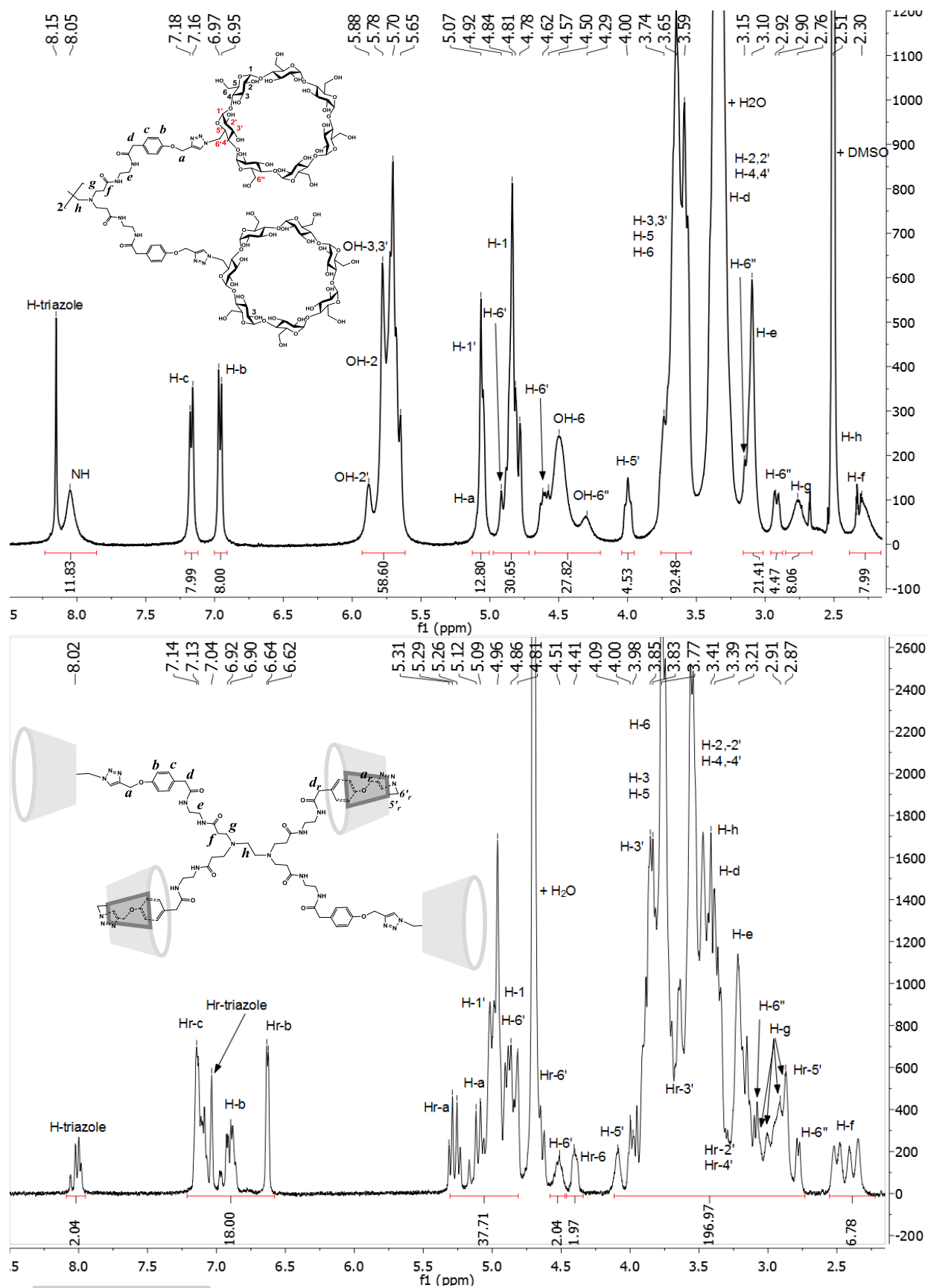
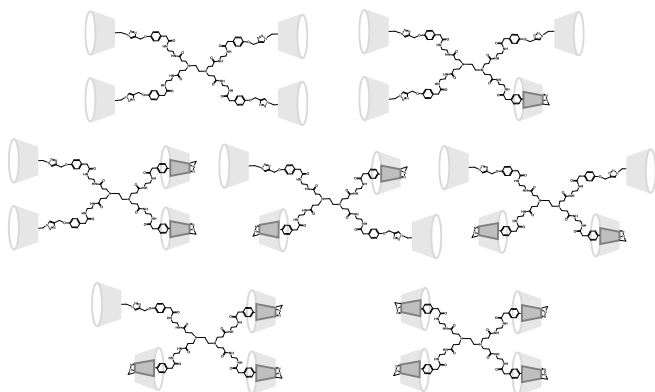


Figure 1. ¹H NMR spectrum of compound **Tetra-CD**. Top: in DMSO-*d*₆ with schematic representation of the molecule giving the atoms numbering for CD cavity. Bottom: in D₂O, with the representation of one of the 7 possible forms for compound **Tetra-CD** in water ("r" stands for the reversed form).

Hr-triazole of branches wearing a reversed CD cavity). The complexity of the signal at around 8 ppm was explained by the possibility of 7 conformations for compound **Tetra-CD** in water as sketched below.



Scheme 3. The 7 possible forms for **Tetra-CD** with 0 to 4 reversed cavities

The phenyl protons were also disturbed since they did not appear anymore as two doublets but as i) a first massif between 7.05 and 7.2 ppm originating from H-c and Hr-c aromatic protons of branches wearing a non-reversed and a reversed CD cavity, respectively, ii) a second massif between 6.8 and 7.0 ppm due to the H-b aromatic protons of branches wearing a non-reversed CD, and iii) a doublet ($J = 7.3$ Hz) at 6.63 ppm due to the Hr-b aromatic protons of branches wearing a reversed CD cavity. Integration of the signals indicated that, on average, half of the CD cavities underwent tumbling. Besides these changes observed in the aromatic region when going from DMSO to water, the rest of the spectrum was also affected by the tumbling effect. Assignments of the protons signals were made on the basis of the analysis previously performed in DMSO- d_6 (see SI), with the help of 1D and 2D NMR experiments in D_2O . Firstly, the 2D COSY spectrum (see **Figure SI-25**) gave the confirmation for chemical shifts values of aromatic protons and allowed also to assign some other CD proton signals. For example, the non-equivalent diastereotopic protons H-6' of the modified glucopyranose ring of β -CD appeared in water as two sets of two signals, one set originating from the non-inverted cavity (H-6' centred at around 4.9 and 4.6 ppm, respectively), the other from the reversed form (Hr-6' centred at around 4.7 and partially hidden by the water peak, and 4.4 ppm, respectively). The tumbling of the CD cavity resulted in a blending effect for those protons, as it was the case for the Hr-5' protons, detected at around 2.9 ppm. Secondly, the HSQC and HMBC spectra (see **Figures SI-26** and **-27**) allowed us to identify most of the protons and carbon signals and also to raise some ambiguities. For example, the H-a protons appeared as two multiplets centred at around 5.3 and 5.1 ppm, respectively, correlated with two carbon signals at 62.4 and 61.0 ppm, respectively. The comparison with the data for C-a/H-a in DMSO- d_6 led us to assign the 5.1/61.0 and 5.3/62.4 ppm correlations to H-a/C-a and Hr-a/Cr-a, respectively. The C-d signal was identified at around 41.7 ppm through HMBC with aromatic H-c protons, thus H-d protons were identified at around 3.4 ppm. Thirdly, 1D TOCSY experiments (see **Figures SI-28** to **-30**) gave rise to the chemical shifts values of the H-f and H-g protons of the branches and to those of the protons of the substituted glucopyranose units

for reversed and non-reversed CD cavities, as well as to the precise value of the chemical shift of Hr-c, found to be 7.15 ppm. Lastly, 2D ROESY experiment (see **Figure SI-31**) highlighted specific interactions of Hr-a, Hr-b, Hr-c and Hr-triazole protons with the inner protons of the reversed CD cavity. This fine analysis work led us to propose detailed assignments of the resonance domains of the CD protons (see **Figure 1** and **Figure SI-32**). The ^{13}C NMR spectrum in D_2O of **Tetra-CD** evidenced also some changes in comparison to the situation in DMSO- d_6 since the carbon signals are splitted in water as the result of tumbling. This is particularly apparent for the phenyl and triazolyl carbon signals, which are particularly sensitive to CD tumbling effect (**Figure SI-33**).

The situation was simpler for compounds **A** and **B** for whose no tumbling was detected in D_2O . Their respective 1H and ^{13}C NMR spectra in D_2O (see **Figures SI-34** and **35**) exhibited the same features as in DMSO- d_6 . Nevertheless, their 2D NOESY spectra (see **Figure SI-36**) revealed correlations between phenyl protons and inner protons of CD cavities, indicating interaction between the extremity of the arm of molecule and the CD cavity, as sketched in **Figure SI-36**, leading to supramolecular assemblies.^[26,27]

Interaction with adamantane carboxylic acid and its sodium salt

According to literature, head-to-tail assemblies similar to those observed with compounds **A** and **B** can be altered by the addition of **AdCOOH** that helps in releasing the CD cavity.^[26] Besides, the tumbling observed for **Tetra-CD** could result in an inclusion ability of an average of two molecules of guest per **Tetra-CD** platform, thus limited accessibility to the CD cavities,^[24,25,28] which is of major importance for considering its potential applications in aqueous medium. Due to the high affinity of adamantane for β -CD,^[16,29] the adamantyl unit (**Ad**) could be convenient to overcome the tumbling of the CD cavity of **Tetra-CD** in water.

To clarify this point, we have examined changes in 1H NMR spectra recorded in D_2O with increasing amounts of **AdCOONa** relative to **Tetra-CD**. The results were numbered using the molar fraction $X_{CD} = [\beta\text{-CD}] / ([\beta\text{-CD}] + [\text{AdCOONa}])$, for simplicity reasons (see **Figure 2**). For a 0.9 X_{CD} molar fraction, the spectrum shape was similar to that obtained for pure **Tetra-CD** in D_2O , unless a slight increase in integration for H-triazole protons (centred on 8 ppm) relatively to the total 20 protons resonating from 6.5 to 8.2 ppm. As X_{CD} decreased, the spectrum shape became simpler with the decrease of the multiplicity for the triazole and H-b protons, concomitant with the progressive disappearance of the Hr-b signal. As soon as X_{CD} reached 0.5, the signal of triazole proton looked as a singlet at 7.96 ppm, and that of phenyl protons as two doublets ($J = 8.8$ Hz) at 7.1 and 6.9 ppm, respectively, in a similar way to what was observed for pure **Tetra-CD** in DMSO- d_6 . This was interpreted as the total disappearance of reversed form of cyclodextrin.

Analyses of those mixtures by 2D ROESY NMR spectroscopy revealed also drastic changes upon addition of **AdCOONa**. The reference ROESY spectrum of pristine **Tetra-CD** in D_2O , given in **Figure SI-37**, displayed interactions of Hr-triazole, Hr-a, Hr-b, and Hr-c, with inner protons of reversed CD cavity. Conversely, when X_{CD} was set at 0.1, *i.e.* in the presence of a large excess of **AdCOONa**, only cross-peaks between the protons of CD cavity (H-3,3'; H-5,5') and those of **AdCOONa** (H- α ; H- β ; H- γ) were observed, which indicates that the CD cavities hosted only

AdCOONa molecules (see **Figure SI-38**). The absence of NOE effect involving triazole and aromatic protons suggests that no CD cavity remained in reversed conformation. Intermediate cases for which X_{CD} ranged from 0.8 to 0.4 are shown in **Figures SI-39** to **42**. For $X_{CD} = 0.8$, the internal protons of the CD cavities displayed NOE with triazole, aromatic and Hr-a protons, as well as with adamantyl protons: the interaction with inner CD protons occurred for both inverted and non-inverted CD rings. For a molar fraction of 0.6, inclusion occurrence was more pronounced, with intense correlation peaks in the 1.5-2 ppm region, in comparison to what was observed in the aromatic region. Correlation with Hr-a protons was no more seen. As X_{CD} reached 0.5, correlations involving inner CD protons were exclusively seen with adamantyl H- α , H- β , and H- γ protons; the same observation was made for $X_{CD} = 0.4$ (**Figure SI-40**). The use of **AdCOOH** as the guest led to same observations (see **Figure SI-43**). The apparent diffusion coefficients of adamantane carboxylate and adamantane carboxylic acid in the examined mixtures were evaluated through ^1H DOSY NMR experiments. The results, gathered in **Table 1**, indicated that as long as adamantyl unit was in default with respect to cyclodextrin, *i.e.* until a X_{CD} value of 0.5, its diffusion coefficient (D) value was low and rather constant, due to its inclusion into the CD cavity. Upon decreasing X_{CD} , a progressive increase in D value was observed, due to increasing amounts of free adamantane species in solution, until a maximum corresponding to these pure compounds. To summarize, beyond an equimolar ratio of adamantyl unit and cyclodextrin, no tumbling was detected by NMR spectroscopy and an increase in apparent value of D was measured. This could be indicative of formation of a [1:4] [**Tetra-CD:Ad**] complex.

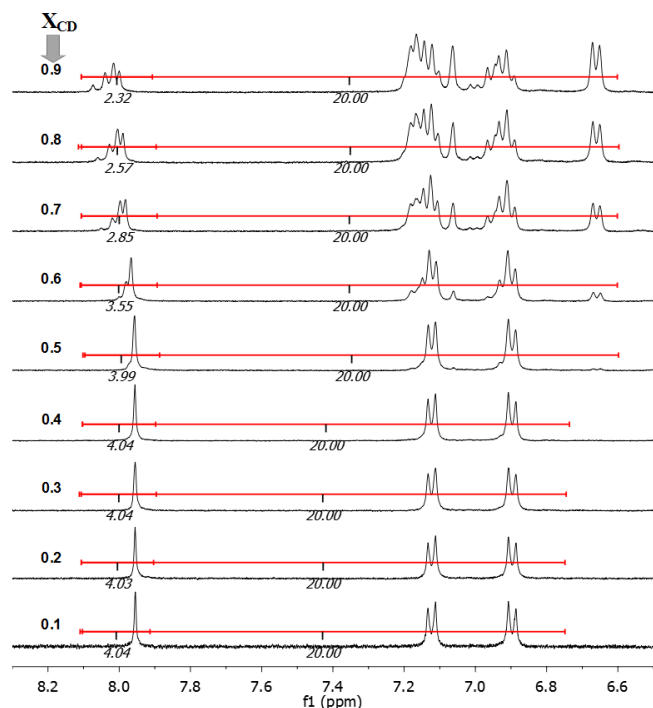


Figure 2. Evolution from top to bottom of ^1H NMR spectra in the 6.5-8.3 ppm region of mixtures of **Tetra-CD** and **AdCOONa** in D_2O with decreasing molar fraction, expressed with respect to the concentration in β -CD cavities ($X_{CD} = [\beta\text{-CD}]/([\beta\text{-CD}]+[\text{AdCOONa}])$). Italic: protons integration over the red segments domains. Total concentration $[\beta\text{-CD}]+[\text{AdCOONa}] = 8 \text{ mmol}\cdot\text{L}^{-1}$.

For evaluating the complex stoichiometry with adamantane carboxylate salt, Job's plot analyses were performed on compounds **A** and **Tetra-CD**. The cases of the so-called **Di-CD** and **Tri-CD** were also examined. These last molecules were chosen as comparison compounds, because the structure of their spacers was similar to that of compound **A**, and they constituted intermediate cases in between **A** and **Tetra-CD**: **Di-CD** is a dimer for which it was found that the tumbling process occurred for 90% of molecules in water,^[24] whereas **Tri-CD** underwent no reversion process for steric reasons as demonstrated by the presence of ^1H NMR singlets in D_2O at 8.15 and 6.42 ppm, originating from 3 triazolyl and 3 phenyl protons, respectively.^[18] **Figure SI-44** shows the continuous-variation plots of the ^1H NMR chemical shift change for the four compounds. For each case, a maximum value was obtained at $X_{CD} = 0.5$, which indicated the formation of a [1:1] complex, as expressed with regards to CD cavities. It followed that compounds **A** and **Tri-CD** lead in water, to [**A:AdCOONa**] and [**Tri-CD:AdCOONa**] complexes which stoichiometries are [1:1] and [1:3], respectively, as logically awaited, and [**Di-CD:AdCOONa**] and [**Tetra-CD:AdCOONa**] complexes exhibited respective stoichiometries of [1:2] and [1:4]. This set of analyses, based on NMR experiments, led us to the assumption that, even in presence of reversed CD cavities in the starting water solutions, it was possible to make them available for inclusion phenomena, thanks to shifting of the tumbling process by using the strongly affine host adamantyl unit.

In order to confirm the possibility for those two molecules to involve all CD cavities in inclusion phenomena, we have examined their isolated inclusion complexes (see Experimental part) by mass spectrometry (MS) with electrospray ionization (ESI) from $\text{MeOH}/\text{H}_2\text{O}$ solutions. Compounds **A** and **B** led to complexes which weights were compatible with MS-ESI detection in positive mode. The molecular peak for **A** was detected at $m/z = 1308.4$ (55%, $[\text{MH}]^+$), along with its sodium adduct at $m/z = 1330.4$ (100%, $[\text{M}+\text{Na}]^+$) (see **Figure SI-45**). The mass spectrum revealed also the signal of the [1:1] inclusion complex at $m/z = 1510.5$ (73%, $[\text{M}+\text{AdCOOH}+\text{Na}]^+$), and others corresponding to multi-charged species resulting to higher inclusion compounds, based on 2 up to 3 molecules **A** supposed to be included in cascade. Similar species were observed with complex formed from **B**, *i.e.* molecules **B** itself, [1:1] complex of [**B:AdCOOH**], and related molecules included in cascade (see **Figure SI-46**). Note for **B** complex analysis the 100% abundance for the peak at $m/z = 1552.6$ corresponding to the $[\text{M}+\text{AdCOOH}+\text{Na}]^+$ complex. Note also that, except for pristine **A** and **B** molecules, peaks of significant abundance always correspond to species for whose the CD cavity is hosting an adamantane molecule. Thus, as previously reported in literature,^[27] the formation of supramolecular assemblies due to head-to-tail interactions is not a major inconvenience for getting [1:1] complexes for compounds **A** and **B**. We also examined inclusion complex formed from **Di-CD** in the same conditions. MS-ESI analysis in the positive mode (see **Figure SI-47**) afforded the molecular peak for **Di-CD** at $m/z = 1275.9$ $[\text{M}+2\text{Na}]^{2+}$. The presence of [1:2] complex was detected by zooming in the m/z 1400-1500 zone as multicharged species forming adducts with sodium or potassium cations. Interestingly, MS-ESI in the negative mode revealed to be an efficient techniques for detecting more obviously this [1:2] complex as showed in **Figure SI-48**. The peak at $m/z = 1432$ (92% abundant) was identified as the di-charged $[\text{M}-2\text{H}+2\text{AdCOOH}]^{2-}$ complex whereas that at $m/z = 954.3$ (10% abundant) was related to the

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tri-charged complex. [Tetra-CD:AdCOOH] revealed also suitable for MS-ESI analysis from water solution in the negative mode: the peak at $m/z = 1640.6$ (12% abundant) was attributed to the [1:4] complex appearing as tetra-charged species $[M-4H+4AdCOOH]^{4-}$ [see Figure SI-49]. These last results suggest the possibility for the CD cavities to be fully available for hosting adamantane derivatives, despite their initial reversed conformation in water.

Table 1. Diffusion coefficients in D₂O of AdCOONa and AdCOOH, pure or in mixture with Tetra-CD, measured by ¹H DOSY NMR technique. The composition of the mixtures is given by X_{CD}. Total concentration $[\beta\text{-CD}]+[\text{Ad}] = 8 \text{ mM L}^{-1}$

X _{CD}	0.8	0.6	0.5	0.4	0.1	0
D _{AdCOONa} (10 ⁻¹⁰ m ² /s)	1.0	1.0	1.3	2.4	3.8	4.2
D _{AdCOOH} (10 ⁻¹⁰ m ² /s)	1.2	1.2	1.8	1.9	2.9	5.8

Conclusion

There are several reports on β -CD-based platforms used as drug carriers, particularly those obtained from grafting high molecular-weight polymers on primary/secondary face of the β -CD.^[30] One drawback of this strategy is that the polymer chains may compete with other guests and occupy the inner space of the β -CD cavity,^[31] limiting the host-guest interaction that causes a lower drug load. Therefore, we synthesized a highly water-soluble PAMAM β -CD derivative, Tetra-CD, initially thought to eliminate such host-host competition. We demonstrated that, despite the occurrence of tumbling in water, it was possible to form [1:4] inclusion complexes with adamantyl species in that medium. It results that the tumbling effect reported in literature^[23-25] can be efficiently shifted in the presence of an appropriate hydrophobic guest, resulting in the full host-ability of the CD units in water. This strategy is an encouraging alternative for the modification of PAMAM with carbohydrates^[32] which can help in mitigating the toxicity already known for low PAMAM generations.^[33] This tetramer combines thus high solubility, multiple functions surface of PAMAM dendrimer, and inclusion properties of β -CD, making it promising for drug loading purpose.^[34] Works in that aim are in progress.

Experimental Section

General: 4-(Propargyloxy)phenol (I),^[35] 2-[4-(propargyloxy)phenyl]acetic acid (II),^[36] 6-O-monotosyl- β -cyclodextrin (β -CD-OTs)^[37], 6-O-monoazido- β -cyclodextrin (β -CD-N₃)^[11] and the two platforms Di-CD and Tri-CD, were synthesized according to literature.^[18] All starting materials were purchased from Aldrich, Acros, Fluka and Strem. 1-Ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride (EDC-HCl), *N*-hydroxybenzotriazole (HOBt), PAMAM G0 dendrimer, copper sulfate (CuSO₄) and solvents were used as received. Sodium-ascorbate was freshly recrystallized from a water:ethanol mixture. Distilled water was used in all experiments. Reactions were performed under argon atmosphere and monitored by analytical TLC on pre-coated silica gel 60 F254 plates (Aldrich) with detection carried out under UV light. Purifications by column chromatography were performed on silica gel 60-200 μm VWR CHEMICALS. Size Exclusion Chromatography (SEC) was performed using water as the eluent with resins Bio-Gel® P-6 medium

(BIO-RAD Laboratory) and Sephadex G-15 (Aldrich). ¹H, ¹³C NMR and 2D HMQC, COSY, NOESY, ROESY, TOCSY experiments were obtained at 298 K on Bruker spectrometers (AV 400 MHz, Bruker Avance 500 spectrometer equipped with a 5 mm triple resonance inverse Z-gradient probe (TBI ¹H, ³¹P, BB) and Bruker NEO 600 spectrometer equipped with a 5 mm triple resonance inverse Z-gradient probe (TBI ¹H, ³¹P, BB)). All spectra were recorded at 25°C in the indicated deuterated solvents. Chemical shifts are reported in ppm (δ) and coupling constants (J) are reported in Hz. Multiplicities are reported by using the following abbreviations: s = singlet, d = doublet, t = triplet, br = broad, and m = multiplet. Protons and carbons signal assignments are indicated on the spectra reported in SI. Mass spectrometry was performed by the Mass Spectrometry Core Facility of ICT; ESI-TOF experiments have been made on a Q-TOF Premier spectrometer (Waters in positive and negative ionisation modes, using direct introduction. HR-ESI-TOF experiments have been made on a Q-TOF Premier (Waters) and on a Xevo G2 QTOF (Waters).

Synthesis

PAMAM G0-alkyne (III): According to a slightly modified general protocol,^[19] a mixture of compound II (515 mg, 2.7 mmol), EDC-HCl (727 mg, 3.79 mmol), and HOBt (366 mg, 2.7 mmol) in DMF (14 mL) was stirred for 4h at room temperature. Then, a solution of PAMAM G0 (250 mg, 0.483 mmol) in DMF (2 mL) was added dropwise. The resulting mixture was kept under stirring for 3 days at 35°C. After evaporation of the solvent under vacuum, the resulting solid was recrystallized in methanol, affording PAMAM G0-alkyne as a white powder (571 mg, 0.473 mmol, yield 98%). ¹H NMR (400 MHz, DMSO-*d*₆, δ ppm): 8.03 (br, 4H, NH), 7.96 (br, 4H, NH), 7.16 (d, $J = 8.7$ Hz, 8H, H-c), 6.90 (d, $J = 8.7$ Hz, 8H, H-b), 4.75 (d, $J = 2.4$ Hz, 8H, H-a), 3.53 (tr, $J = 2.4$ Hz, 4H, C≡H), 3.33 (8H, H-d overlapped with H₂O), 3.08 (br, 16H, H-e), 2.65 (m, 8H, H-g), 2.45 (br, 4H, H-h), 2.19 (m, 8H, H-f); ¹³C {¹H} NMR (101 MHz, DMSO-*d*₆, δ ppm): 171.95, 171.02, 156.26, 130.41, 129.49, 115.03, 79.84, 78.54, 55.80, 51.25, 50.04, 41.92, 38.98, 38.73, 33.60. ESI-TOF-MS $m/z = 1205.5$ (calcd. 1205.6 for [MH]⁺); 1227.5 (calcd. 1227.6 for [MNa]⁺). Elemental analysis: calculated for C₆₆H₈₀N₁₀O₁₂ + 1 H₂O: C: 64.80; H: 6.76; N: 11.45; O: 17.00; found C: 64.63; H: 6.77; N: 11.72; O: 16.22.

Compound Tetra-CD: PAMAM-G0 alkyne (130 mg, 0.107 mmol) and β -CD-N₃ (550 mg, 0.474 mmol) were dissolved in DMSO (5 mL) previously degassed by argon bubbling for 15 min. A solution of CuSO₄ (8 mg, 0.047 mmol) in 0.5 mL of H₂O, followed by a solution of sodium ascorbate (28 mg, 0.142 mmol) in H₂O (0.5 mL) added dropwise over 5 min. The mixture immediately turned yellow, and then was heated at 80 °C under vigorous stirring for 48h under argon atmosphere. After cooling down to room temperature, the mixture was precipitated in an excess of acetone (130 mL), giving a yellow powder recovered by filtration on a glass filter. The powder was dissolved in water and purified by SEC (Bio-Gel® P-6 medium). Evaporation of the solvent gave a pale yellow powder (579 mg, 0.099 mmol, yield 92.5%). ¹H NMR (400 MHz, DMSO-*d*₆, δ ppm): 8.15 (s, 4H, H triazole), 8.05 (s, 8H, NH), 7.17 (d, $J = 8.1$ Hz, 8H, H-c), 6.96 (d, $J = 8.1$ Hz, 8H, H-b), 5.88 (s, 4H, OH-2'), 5.78 (s, 24H, OH-2), 5.74-5.60 (m, 28H, OH-3,3'), 5.18-5.00 (m, 12H, H-1'; H-a), 4.95-4.75 (m, 28H, H-6'; H-1), 4.67-4.20 (m, 28H, H-6', OH-6, OH-6'), 4.00 (m, 4H, H-5'), 3.83-3.50 (m, 92H, H-6, H-3,3', H-5), 3.50-3.20 (m, 56 H, H4,4', H-2,2' overlapped with H₂O), 3.20-3.05 (m, 20H, H-6"; H-e), 2.91 (m, 4H, H-6"), 2.76 (s, 8H, H-g), 2.55-2.46 (m, 12H, H-d, H-h overlapped with DMSO), 2.30 (m, 8H, H-f); ¹³C {¹H} NMR (101 MHz, DMSO-*d*₆, δ ppm): 171.93, 171.14, 157.22, 143.00, 130.48, 128.98, 125.94, 114.86, 102.38, 102.04, 101.74, 83.43, 81.99, 73.50, 72.86, 70.64, 61.39, 60.21, 51.54, 51.52, 50.76, 41.92, 38.74, 33.43. ESI-TOF-MS $m/z = 1963.8$ (calcd. 1964.0 for [MHN₃]³⁺); 1478.6 (calcd. 1478.8 for [MHN₃]⁴⁺). Elemental analysis calculated for C₂₃₄H₃₅₆N₂₂O₁₄₈ + 25H₂O: C: 44.64; H: 6.50; N: 4.89; O: 43.96; found C: 43.65; H: 6.31; N: 4.80; O: 43.30.

Compound A: Compound I (70.2 mg, 0.47 mmol) and β -CD-N₃ (500 mg, 0.43 mmol) was dissolved in DMSO (5 mL) previously degassed by argon bubbling for 15 min. To a solution of compound I (70.2 mg, 0.47 mmol) and

β -CD-N₃ (500 mg, 0.43 mmol) dissolved in 5 mL of DMSO degassed by argon bubbling for 15 min, solutions of CuSO₄ (15 mg, 0.094 mmol) in 1 mL of H₂O, then sodium ascorbate (56 mg, 0.282 mmol) in 1 mL of H₂O, were added dropwise. The mixture turned rapidly yellow and was heated at 50 °C with vigorous stirring for 24 h. After cooling down to room temperature, the mixture was precipitated in an excess of acetone (80 mL). A green powder was recovered by filtration and then purified by SEC (Sephadex-G15). Evaporation of the solvent gave a pale yellow powder (507 mg, 0.39 mmol, yield 90%). ¹H NMR (400 MHz, DMSO-*d*₆, δ ppm): 8.94 (s, 1H, H-phenol), 8.12 (s, 1H, H-triazole), 6.85 (d, *J* = 9.0 Hz, 2H, H-b), 6.68 (d, *J* = 9.0 Hz, 2H, H-c), 5.90-5.60 (m, 14H, OH-2'-2'; OH-3,3'), 5.04 (s, 1H, H-1'), 4.97 (s, 2H, H-a), 4.99-4.75 (m, 7H, H-6'; H-1), 4.65-4.55 (m, 1H, H-6'), 4.55-4.40 (m, 5H, OH-6), 4.28 (m, 1H, OH-6'), 4.00 (m, 1H, H-5'), 3.75-3.50 (m, 23H, H-6; H-3,3'; H-5), 3.48-3.20 (m, 14H, H-4,4', H-2,2' overlapped with H₂O), 3.11 (m, 1H, H-6''), 2.93 (m, 1H, H-6''); ¹³C {¹H} NMR (101 MHz, DMSO-*d*₆, δ ppm): 152.08, 151.33, 143.35, 125.68, 116.21, 116.14, 102.67, 102.49, 102.45, 102.36, 101.72, 83.90, 82.52, 81.97, 81.82, 81.45, 73.67, 73.45, 73.10, 72.95, 72.82, 72.50, 72.21, 70.42, 60.39, 60.30, 61.98, 60.61, 60.42, 59.44, 50.79. HR-ESI-TOF-MS *m/z* = 1308.4429 [MH]⁺, 1330.4265 [MNa]⁺. Elemental analysis: calculated for C₅₁H₇₇N₃O₃₆ + 9 H₂O: C: 41.66; H: 6.51; N: 2.86; O: 48.97; found C: 41.55; H: 6.50; N: 2.49; O: 43.21.

Compound B: The same procedure for the obtainment of compound **A** was used, starting from Compound **II** (90 mg, 0.47 mmol) and **β -CD-N₃** (500 mg, 0.43 mmol). The obtained powder was purified by SEC (Bio-Gel® P-6 medium). Evaporation of the solvent gave a pale yellow powder (569 mg, 42 mmol, yield 98%). ¹H NMR (400 MHz, DMSO-*d*₆, δ ppm): 12.03 (s, 1H, COOH), 8.16 (s, 1H, H triazole), 7.18 (d, *J* = 8.0 Hz, 2H, Hc), 6.98 (d, *J* = 8.0 Hz, 2H, Hb), 5.88 (d, *J* = 8.0 Hz, 1H, OH-2'), 5.78 (s, 6H, OH-2), 5.75-5.55 (m, 7H, OH-3,3'), 5.08 (s, 2H, Ha), 5.05 (s, 1H, H-1'), 5.00-4.70 (m, 7H, H-6'; H-1), 4.61 (m, 1H, H-6'), 4.55-4.40 (m, 5H, OH-6), 4.29 (s, 1H, OH-6''), 4.00 (m, 1H, H-5'), 3.80-3.45 (m, 23H, H-6; H-3,3'; H-5), 3.45-3.25 (m, 14H, H-4,4', H-2,2' overlapped with H₂O), 3.25-3.10 (m, 1H, H-6''), 2.91 (m, 1H, H-6''), 2.55 (s, 2H, Hd overlapped with H₂O); ¹³C {¹H} NMR (101 MHz, DMSO-*d*₆, δ ppm): 173.45, 157.35, 143.02, 130.85, 127.75, 125.92, 114.90, 102.66, 102.35, 101.73, 83.90, 82.51, 81.98, 81.82, 81.47, 73.66, 73.45, 73.29, 73.10, 72.87, 72.81, 72.50, 72.20, 70.44, 61.37, 60.29, 59.44, 50.82, ~40.00 (overlap with DMSO). HR-ESI-TOF-MS *m/z* 1350.4503 [MH]⁺, 1372.4316 [MNa]⁺. Elemental analysis calculated for C₅₃H₇₉N₃O₃₇ + 10 H₂O: C: 41.60; H: 6.52; N: 2.75; O: 43.14; found C: 41.62; H: 5.90; N: 2.30; O: 43.85.

Solubility tests in water

The method reported by Jozwiakowski and Connors was used for the determination of the solubility of all platforms.^[38] Appropriate amounts of solid were placed in vials with 1 mL of H₂O. The samples were stirred in a constant temperature bath at 25.00 ± 0.01 °C for 48 hrs. The supernatants were then separated from the solid phase at 25 °C and filtered with Milli-Q membrane filters (0.45 μ m pore size) upon injection from 3 mL disposable plastic syringes and the solutions were analysed for dissolved solid. Supernatant of each sample was placed in at least three different vials. The samples were dried by freeze drying for 2 days and weighted to within ± 0.1 mg.

Formation of inclusion complexes.

Inclusion complexes with **AdCOOH** were prepared according to the freeze-drying method:^[39] 300 μ L of a 0.185 mmol.L⁻¹ solution of **AdCOOH** in MeOH was added to 3 mL of an aqueous solution of **β -CD** compounds (0.0185 mmol.L⁻¹ in CD cavity) under vigorous stirring at 40 °C for 24h. The resulting solution was freeze-dried to give inclusion complexes.

Job's plot method.

Two stock solutions, (**Sol1**) of **β -CD** compound 8 mmol.L⁻¹ in **β -CD** cavities, and (**Sol2**) of **AdCOONa**^[40] 8 mmol.L⁻¹, were prepared in D₂O. A series of nine samples in NMR tubes containing both **β -CD** compound and **AdCOONa** with a total concentration (**[AdCOONa]**+**[β -CD]**) fixed at 8 mmol.L⁻¹. This was accomplished by introducing 50; 100; 150; ... 450 μ L

of **Sol1** in the 1st; 2nd; 3rd; ...; 9th NMR tube, and then 450; 400; 350; ...; 50 μ L of **Sol2** in the 1st; 2nd; 3rd; ...; 9th tube. Thus, solutions with constant volume at varying **β -CD** molar fractions ($X_{CD} = \frac{[\beta\text{-CD}]}{([\beta\text{-CD}] + [\text{AdCOONa}])}$) in a complete range (0.1 < *r* < 0.9) were obtained. The tubes were submitted to NMR analysis, with D₂O as the internal standard. The continuous variation of the ¹H NMR chemical-shift change " $\Delta\delta \times X_{CD}$ " ($\Delta\delta$ taken for the adamantyl H- γ , see **Figure SI-38**), was plotted against " X_{CD} ".^[26]

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A G0 PAMAM dendrimer bearing four β -cyclodextrin cavities is synthesized through click chemistry. From results of ^1H NMR studies and mass spectrometry analyses, we demonstrate that the complexation strength of adamantyl species with cyclodextrin is favourable to the full availability of the cavities in water, despite of a spontaneous tumbling process in that medium for the pristine platform.