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Interspecific hybridisation and interaction with cultivars affect the genetic variation of

Ulmus minor and U. glabra in Flanders

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Abstract

 Interspecific hybridisation and gene flow from cultivated plants may have profound effects on the evolution of wild species. Considering the cultural history and past use of *U. minor* and *U. glabra* trees in Flanders (northern Belgium), we investigated the extent of human impact on the genetic variation of the remaining, supposedly indigenous elm populations. We therefore examined the rate of interspecific hybridisation, which is expected to be higher under human influence, the occurrence of clones within and among locations, the presence of cultivars and their possible offspring. Based on results produced using 385 Amplified Fragment Length Polymorphic (AFLP) markers, 46% of the 106 investigated Flemish elms appeared to be F1 hybrids or backcrosses to one of the parent species, while no F2 hybrids (F1 x F1 progeny) were found. Clonality was mainly found among *U. minor* and hybrids, which are more likely to form root suckers or sprouts as opposed to *U. glabra*. The majority of the studied locations (76% of the locations with multiple samples) showed evidence of clonal reproduction. Several, sometimes distant, locations shared a multilocus lineage. We also found indications of gene flow from cultivated elms into native species. It is conceivable that reproductive material has been moved around extensively, obscuring the natural genetic structure of the elm populations. The results help guide the Flemish elm genetic resources conservation program.

- 17 Key words: Ulmus minor, Ulmus glabra, hybridisation, elm cultivars, clonal reproduction, human-
- 18 mediated disturbance

Introduction

Ulmus minor Mill. (Field elm) and U. glabra Huds. (Wych elm) are native deciduous trees in western Europe. Especially *U. minor* and *U. x hollandica* (i.e. the hybrid between *U. minor* and *U. glabra*) have been widely planted over the ages, in the cities as well as in the countryside, on roadsides and along property borders, in hedges, near manors and farmsteads, as standards as well as coppice. Planting stock was sometimes obtained from relatively distant provenances (Heybroek et al. 2009). In a forest in The Haque (the Netherlands), for example, elms from Flanders (northern Belgium) and Utrecht (the Netherlands) were planted in the 16th century (van Dissel 1912). Richens (1983) speculated that even prehistoric tribes carried live elms on their wanderings. But typically local material from surrounding forests, seedlings or root suckers, would have been used for plantings. Elm timber was greatly appreciated for many purposes, e.g. coppice produced firewood and pliable twigs. Consequently, elms have a long history of cultivation. Local cultivars were known since the 17th century and probably earlier. In the Mediterranean, but also in areas in western Europe where the Romans introduced their viticulture, elms were raised to act as vine supports, while the elm leaves served as an excellent fodder for cows and sheep (Forster and Heffner 1954; Fuentes-Utrilla et al. 2004). For this purpose, they brought a special cultivar, the 'Atinia' (Gil et al. 2004). More recent cultivars are often non-European elms or hybrids between European and non-European elms, such as U. wallichiana Planch. (Himalayan elm), U. japonica (Rehder) Sarg. (or U. davidiana var. japonica (Rheder) Nakai; Japanese elm) and *U. pumila* L. (Siberian elm).

In addition to habitat loss and fragmentation due to changes in land use, the Dutch Elm Disease (DED) severely reduced the number of elm trees starting from around 1910. DED is caused by the non-native fungi, *Ophiostoma ulmi* (Buisman) Nannf. and *O. novo-ulmi* Brasier, which are spread by bark beetles of *Scolytus* Geoffroy (Coleoptera, Scolitidae; Brasier 2001). The main vectors in western Europe are *S. scolytus* and *S. multistriatus* (Webber 2004). Considering the severe threats European elms are under, national actions were taken to conserve the remaining germplasm. In turn, this has led to the initiative within the European Forest Genetic Resources (EUFORGEN) cooperative program to realise a conservation plan on a European level (Collin 2002). Furthermore, an EU project on the "Co-ordination for conservation, characterisation, collection and utilisation of genetic resources of European elms" ran from 1997 till 2001 (RESGEN CT96-78). In Flanders, our study area, new clones of native elms are still being added to the *ex situ* collections, initiated within the RESGEN project. Still, limited knowledge is available on the level of genetic variation residing in elm populations and to what extent this was influenced by human activities.

U. minor and *U. glabra* co-occur in Flanders, but are also able to hybridise with each other (*U.* x hollandica Mill.), making their taxonomy extremely difficult (Goodall-Copestake et al. 2005). Although hybridisation between *U. minor* and *U. glabra* is a natural process, it can also be human-mediated, through habitat modification, species introduction or translocation. Human activities can therefore encourage hybridisation (Ellstrand and Schierenbeck 2000; Rhymer and Simberloff 1996). For example, *U. pumila* was introduced in Spain and Italy. Its hybrid with the native *U. minor* is now common in both countries (Brunet et al. 2013; Cogolludo-Agustin et al. 2000). Natural interspecific

hybridisation as well as gene flow from cultivated elms to their native relatives are processes with significant evolutionary consequences. A potential concern is outbreeding depression when coadapted gene-complexes are broken up, causing a decrease in fitness (Eucalyptus spp.: Potts et al. 2003; Edmands 2007; Laikre et al. 2010; Lynch 1991). Also, asymmetric gene flow could occur when one species is less abundant than the other, with females of the rare species receiving a high amount of heterospecific pollen, causing a higher risk of mate recognition errors (Mayr 1963; Wirtz 1999). Moreover, first generation backcrossing will more likely happen with the more common species, leading to asymmetric introgression (Rieseberg 1997). For instance, species abundance influenced the direction of introgression in oak populations (Lepais et al. 2009) as well as between two Morus spp. (Burgess et al. 2005). Interspecific hybridisation may, however, also result in the addition of genetic variability that facilitates habitat expansion (Arnold 1997). Conversely, the commercial release of cultivated trees (or cultivars) can result in homogenisation of the genetic composition and structure of the wild populations through introgression. These genetic risks are of particular concern when the planted trees represent a narrow gene pool spread on a wide scale and the related wild species is highly threatened by habitat reduction or diseases (e.g. Populus spp.: Santos-del-Blanco et al. 2013; Vanden-Broeck et al. 2012; Smulders et al. 2008; Malus spp.: Kron and Husband 2009; Coffea arabica: Aerts et al. 2013; Prunus avium: Mariette et al. 2010).

Considering the cultural history and past use of *U. minor* and *U. glabra* trees in Flanders, we investigated if the genetic variation of the remaining, supposedly indigenous elm populations is influenced by human activities. The answer to this research question could help guide the Flemish elm genetic resources conservation program. We therefore examined 1) the rate of interspecific hybridisation, which is expected to be elevated due to human activities, 2) the occurrence of ramets (i.e. genetically identical individuals originating vegetatively from a single ancestor) within and among locations and 3) the presence of cultivars and their offspring in Flemish indigenous elm populations.

Material and methods

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- Samples and DNA extraction
- 27 Three sets of samples were used in this study. Between 2007 and 2009 leaves of adult *Ulmus minor*,
- 28 *U. glabra* trees and their hybrids were collected in Flanders in northern Belgium for the first set (Table
- 29 1, Fig. 1). The leaves were collected in a gene bank containing ramets of the original trees (i.e. ortets)
- 30 except for the samples in the location coded BESP (Table 1). The indigenous ortets were at least 3 m
- 31 apart from each other to diminish the risk of sampling ramets. The census number of elm trees was
- 32 lower than 30, except at location BERI (n > 50) (Table 1). We limited the number of samples per
- 33 location to address the research questions on the regional scale (i.e. Flanders). The species of the
- 34 elm trees was identified morphologically according to Lambinon et al. (1998). This set of samples
- 35 ('dataset 1') entails the focus of our study and is therefore used in every analysis.
- 36 The second set ('dataset 2') comprises samples of *Ulmus minor* and *U. glabra*, which are used as
- 37 references for these three pure species in the hybrid detection analyses (Table 1). They were
- 38 obtained from the ex situ collection of the RESGEN CT96-78 project (indicated with 1 to 6 in

- 1 superscript in Table 1). They are originally from Belgium (location BETO), France, Italy, Germany and
- 2 Greece, and were collected mainly in forests and screened genetically by Goodall-Copestake et al.
- 3 (2005). Also, a few samples from six localities in France were added (locations FRAU, FRCB, FRLR,
- 4 FRLV, FRSP and FRBL), as well as one sample of *U. procera* Salisb. (English or Atinian elm)
- 5 collected in Kallstadt, Germany.
- 6 The third set of samples ('dataset 3') contains cultivars which were mainly collected in the
- 7 Netherlands, except for 'Klemmer', which was provided by Irstea (France) and originated from
- 8 Wakehurst Place (UK) (Table 2). The Dutch locations of elm cultivars are well known by Heybroek et
- 9 al. (2009); several recent cultivars in this study were even selected and released by H. Heybroek.
- 10 Some of the cultivars are hybrids with *U. wallichiana*, *U. japonica* or *U. pumila*. The cultivars were
- included in clone and parentage analyses to investigate their influence on supposedly indigenous
- individuals of *U. minor*, *U. glabra* and their interspecific hybrids. Well-known cultivars 'C. Buisman' (*U.*
- 13 minor), 'Sarniensis' (U. minor) and 'Horizontalis' (U. glabra) were also used as reference samples
- representing pure *U. minor* and *U. glabra* in hybrid detection analyses.
- 15 The leaves were stored on silica gel. Total DNA was extracted from ground leaf samples, partly with
- 16 QuickPick™ SML Plant DNA purification kit in combination with the PickPen 8-M magnectic tool or the
- 17 MagRo 8-M robotic workstation (Isogen Life Science) on 5 mg of dried leaf tissue, and partly with
- 18 DNeasy Plant Mini Kit (Qiagen) on 20 mg of dried leaf tissue. The integrity of the DNA was assessed
- 19 on 1.5% agarose gels, and the purity and concentration with the ND-1000 Nano-Drop
- 20 spectrophotometer (NanoDrop Technologies).
- 21 AFLP analysis
- 22 Amplified Fragment Length Polymorphism (AFLP) fingerprints were generated according to Vos et al.
- 23 (1995), but with restriction and ligation conducted in one single step. Initially, 21 primer combinations
- 24 (EcoRI/Msel) were tested on 16 samples (14 individuals and 2 replicates). The following two primer
- 25 combinations were selected for the selective amplifications: EcoRI-AGC(ned)/Msel-CTG and EcoRI-
- 26 ACC(ned)/Msel-CTG. We used the RawGeno v 2.0 R CRAN package (Arrigo et al. 2009) for
- automated scoring, which resulted in 385 polymorphic loci.
- 28 There were seven samples with poor quality AFLP electropherograms for one or both primer
- 29 combinations. To avoid series of missing values, we discarded these samples from further analysis,
- 30 leaving a total of 177 samples of which 34 (19%) were randomly replicated to assess reproducibility.
- 31 Mean typing error reached 4% (following Bonin et al. 2004) and mean Dice similarity between pairs of
- 32 replicates was 0.95. The polymorphism information content (PIC) of each locus was calculated
- 33 (Anderson et al. 1993).
- 34 Data analysis
- 35 Linkage disequilibrium
- The hybrid detection analysis and the sibship analysis were executed under the assumption of unlinked markers (see below). We therefore assessed linkage disequilibrium (LD) among AFLP loci

by performing logistic regressions between pairs of loci (e.g. Poncet et al. 2010). To correct for multiple testing, we used the false discovery rate approach (FDR: Benjamini and Hochberg 1995) with R package brainwaver (Achard 2012). The maximum FDR was set at 5%. Computations were done with the statistical software platform R version 2.15 (R Development Core Team 2010). Another method to detect linkage disequilibrium by Haubold et al. (1998) was used, implemented in LIAN 3.6 (Haubold and Hudson 2000). The standardized index of association (ISA) was calculated, based on 10,000 permutations. Values for dataset 1, with 309 polymorphic markers, were produced, as well as for three subsets of dataset 1: *U. glabra* samples (199 polymorphic markers), *U. minor* samples (183 polymorphic markers) and their hybrids (232 polymorphic markers). This subdivision was based on the results of the hybrid detection analyses. It is expected that U. minor and the hybrids will show higher LD because of their ability to reproduce asexually. The subsets of dataset 1 were reanalysed with duplicate ramets excluded, resulting in 197 polymorphic markers for the *U. glabra* samples, 172 for *U. minor* and 222 for hybrid samples. Recent hybridization between taxa derived from two gene pools with different allele frequencies, should exhibit strong LD. Still, associations among loosely linked markers decay rapidly with subsequent generations of hybrids, resulting in a reduced level of LD (Goodman et al. 1999).

Hybrid identification

 In order to investigate the actual extent of interspecific hybridisation among Flemish elms, we used the Bayesian method implemented in BAPS v 5.4 (Corander et al. 2008). An admixture analysis based on pre-defined groups was performed using 100 iterations. We restricted the analyses to samples identified as U. minor, U. glabra and their hybrids of all three datasets (i.e. 164 samples of 182). The samples screened genetically by Goodall-Copestake et al. (2005), were used as reference samples for U. minor and U. glabra (i.e. of the second dataset and the sample of location BETO from dataset 1). Also the well-known cultivars 'C. Buisman' (U. minor), 'Sarniensis' (U. minor) and 'Horizontalis' (U. glabra) were treated as reference samples for both pure species. Individuals with admixture values (q_i) \geq 0.90 for either of the two groups were assigned to purebreds. In other cases they were considered to be admixed (Vähä and Primmer 2006).

Another Bayesian approach, NewHybrids v1.1b3 (Anderson and Thompson 2002), was used on the same dataset. The program was run with the default parameters for the six genotype class frequencies (pure U. minor, pure U. glabra, F1 hybrids and F2 progeny (of F1 hybrid parents) and backcrossed F1 hybrids), uniform priors and two runs with a burn-in phase of 10,000 steps and 100,000 MCMC iterations. The same reference samples as in BAPS were used to represent the pure species. The samples used as priors were specified as sampled separately from the rest of the mixture of interest, except the sample of BETO located in Flanders. Individuals were considered to belong to one of the predefined classes if $P \ge 0.90$ for that genotypic class (Vähä and Primmer 2006).

It must be noted that the model underlying NewHybrids relies on the assumption that the genetic markers in the dataset are unlinked (Anderson 2008), which also applies for the BAPS analysis. Although we found significant LD among loci of the Flemish samples (see results), this concerned only a very small number of loci.

Clone assignment

Ramets (or clone mates) belonging to the same clonal lineage bear the same multilocus genotype or. if somatic mutations have occurred, the same multilocus lineage (MLL). To identify MLLs among the samples, we used GenoType (Meirmans and Van Tienderen 2004) on the three datasets combined. First, the discriminative power of the AFLP markers was assessed by plotting the number of loci versus genotypic richness (G:N; i.e. the ratio of the number of MLLs and number of samples) for an increasing number of loci. For the calculations of genotypic richness we first used a set of two highly informative markers successively adding sets of 16 markers with decreasing information content, resulting in calculations based on two, 18, 34, 50, 66, etc. markers, respectively. For each set of markers, the number of MLLs was obtained using several thresholds, ranging from 0.90 to 0.99 Dice similarity, to account for differences in actual thresholds between different sets of loci. When genotypic richness reaches a plateau, the markers are considered to possess enough resolution to reliably assign clonal lineages (Arnaud-Haond et al. 2005). The Dice similarity index was used as a distance measure between all pairs of individuals. As scoring errors and mutations can cause a genetic distance between members of the same MLL to be higher than zero, a threshold needs to be chosen to account for these differences, which are indicated by a multimodal frequency distribution of genetic distances (Arnaud-Haond et al. 2005; Douhovnikoff and Dodd 2003; Meirmans and Van Tienderen 2004). Consequently, the frequency distribution of the distances and the mean Dice similarity of the pairs of replicated samples were used to define an appropriate threshold.

Whether members of a MLL were the result of sexual reproduction, was evaluated through the clonal population structure test implemented in GENODIVE v2.0b17 (Meirmans and Van Tienderen 2004). Here, the observed clonal diversity is compared with the diversity achieved by randomising alleles over individuals (1,000 permutations). Clonal diversity is expected to be lower under clonal reproduction than under sexual reproduction (Gómez and Carvalho 2000). This test includes the threshold used for assigning MLLs. Nei's corrected diversity index (Nei 1978) was used as test statistic and the alleles were randomised over all individuals of the Flemish region (i.e. dataset 1). The same calculations were performed for the Flemish *U. minor*, *U. glaba* and hybrid trees separately. It should be noted that similar genotypes could also be the result of selfing, biparental inbreeding and population genetic structure (Halkett et al. 2005).

Parentage and sibship analysis

To evaluate past or present influence of elm cultivars on the putative indigenous elm populations of Flanders, we conducted a sibship and parentage analysis with Colony v2.0.1.9 (Jones and Wang 2009). In this program, a maximum likelihood method is used. Offspring are clustered into paternal and maternal families using a simulated annealing approach to maximise the group likelihood value. Then, candidate parents are assigned to the clusters at a 95% confidence level. If no candidate parents seem available, the program reconstructs parental genotypes. It can also deal with scoring errors. We excluded monomorphic loci, resulting in a set of 355 polymorphic markers. The offspring dataset contained unique ramets of dataset 1, while the cultivars of dataset 3 were set as the potential parents, considering their release dates are often quite old (18th – early 19th century) and/or their

- 1 widespread use, especially of 'Belgica', 'Major', 'Klemmer' and 'Vegeta'. The more recent cultivars,
- 2 mostly parented by non-native elms, were included as 'negative controls' in the candidate parents set.
- 3 We found some linkage among markers (see results). This could lower the precision of the analysis,
- 4 but is probably compensated by the high number of informative markers (Wang and Santure 2009).
- 5 We did not consider tree size or any other measure of tree age in the analysis. Because many trees in
- 6 our study are part of old hedges, coppice trees or root suckers of trees that succumbed to DED, age
- 7 estimates are highly unreliable. The results from this parentage and sibship analysis will therefore
- 8 only highlight close relations, rather than exact parent-offspring relationships.

Results

- 10 Marker information and independence
- 11 The AFLP loci are only moderately polymorphic (Table 3), with a comparable mean PIC for *U. minor*,
- 12 U. glabra and their hybrids (0.17-0.22). However, when considering loci with a band presence
- between 5% and 95%, a minimum of 145 loci still remained with a substantially higher mean PIC
- 14 between 0.26 and 0.30.
- 15 In dataset 1, 3.7% of the pairwise logistic regressions were significant. Although I^S_A was low (0.0174)
- it appeared significant (P < 0.0001). The proportion of significant logistic regressions decreased when
- only *U. minor* (0.6%) or hybrids (0.4%) were considered, while ISA slightly increased (0.0182 and
- 18 0.0175 with P <0.0001, respectively). Retaining only unique ramets further rendered insignificant
- 19 logistic regressions for *U. minor* and only one significant pairwise comparison (0.004%) for hybrids.
- The I_A^S value stayed the same for *U. minor*, but decreased to 0.0111 (P < 0.0001) for the hybrid
- 21 group. For the set of Flemish *U. glabra* samples, no significant logistic regressions were detected,
- while I^S_A stayed significant with even lower values (0.0064 and 0.0049 (P < 0.0001), with and without
- 23 duplicate ramets, respectively).
- 24 Hybrid detection
- 25 Out of 106 trees sampled in Flanders, only 28 were assigned as pure *U. glabra* and 29 as pure *U.*
- 26 minor with $q_i = 1$ using BAPS (Fig. 2). Three out of eight samples that were identified as hybrids in the
- 27 field, appeared to be pure species (one *U. glabra* and two *U. minor*). So, 49 (46%) of the Flemish
- 28 samples seemed genetically to be hybrids between these two species, of which 38 (78%) were
- 29 identified morphologically as a pure species. Most of the samples were assigned with high
- 30 probabilities using NewHybrids (80% with P ≥ 0.90). Eight Flemish samples showed probabilities
- 31 between 0.80 and 0.90. Only 12 Flemish samples had maximum probabilities below 0.80 with seven
- 32 below 0.70. These individuals with P < 0.90 were all classified as hybrids using BAPS, except for two
- 33 of location BERI and one sample of BEGE. Their maximum probabilities (0.78 < P < 0.89) given by
- 34 NewHybrids also suggested them to be purebreds. Furthermore, two reference samples of *U. minor* in
- 35 dataset 2, one of FRGS and one of GELD, were classified by NewHybrids as backcrosses to U.
- 36 minor, however with probabilities of 0.52 and 0.55, respectively. BAPS did not reclassify these
- 37 reference samples. No F2 hybrids were found, based on the NewHybrids results. As for the cultivars,
- 38 the analyses identified 'Groeneveld' and 'Vegeta' as (multiple generation) backcrosses to *U. minor*

- 1 and 'Dampieri' as a cultivar of *U. minor*. Furthermore, both replicates starting from leaf tissue of
- 2 'Major' appeared to be pure *U. minor*, but the cultivar is claimed to be a hybrid. The other cultivars
- 3 involving solely *U. minor* and/or *U. glabra* and the remaining samples of dataset 2 were assigned to
- 4 their predetermined group.
- 5 For all the following analyses, samples were assigned to the species class given by NewHybrids and
- 6 BAPS results.
- 7 Clonality
- 8 The resolution of the markers appeared high enough to reliably detect clones. The relationship
- 9 between number of markers and genotypic richness is asymptotic, reaching a plateau at
- 10 approximately 96 markers (Fig. 3). The frequency distribution of pairwise Dice similarity values was
- 11 more or less bimodal, with a small peak towards lower distances. Fig. 4 shows this small peak.
- 12 Although the valley between the first and second peak is considered a probable threshold (Meirmans
- 13 and Van Tienderen 2004), which is at a Dice similarity of 0.90, this seemed questionable. At this level
- of similarity, three French samples of two locations separated by more than 100 km, appeared to be
- 15 clones. Although this is not impossible through human mediation, this seems a less likely scenario.
- We therefore resided to select a more conservative threshold of 0.94 Dice similarity, which is slightly
- lower than the mean Dice similarity of 0.95 calculated for the duplicate samples to account for somatic
- mutations and which coincides with the first small drop in frequency in Fig. 4. In addition, three ramets
- of 'Lobel' and two of 'Plantyn', each collected at a different locality, were assigned to the correct MLL
- at this threshold.
- 21 Fig. 2 indicates the identified MLLs with multiple ramets for the Flemish locations. Clones were
- 22 detected at 12 of the 16 Flemish sampling locations and at four French locations, where more than
- one tree was sampled. They were mostly found among *U. minor* and hybrids, mainly with higher
- 24 admixture levels for *U. minor* genes, but also a few individuals of *U. glabra* appeared to be ramets.
- 25 Two MLLs with each two ramets of *U. glabra* were detected at location BERI (MLL '9' and '10'; Fig. 2).
- 26 The *U. glabra* ramets of MLL '9' are neighbouring trees.
- 27 Certain MLLs are present on more than one location (Euclidean distances between locations): MLL
- 28 'A' at locations BEDI, BEME2 and BESC (16 to 39 km), MLL 'B' at BEDM2 and BEHO (4.5 km), MLL
- 29 'C' at BEEG and BERI (70 km), and MLL 'E' at BEMA and BESC (4.5 km) (Fig. 1). Interestingly, one
- 30 individual of BEGE and all samples except one of BEME1 belong to the same MLL as the 'Klemmer'
- 31 cultivar.
- 32 Clonal diversity was significantly lower than diversity under random mating for dataset 1, as well as
- 33 for each separate species in Flanders (P < 0.001).
- 34 Also in France, some neighbouring *U. minor* trees at FRAU, FRLV, FRCB and FRLR, respectively,
- 35 had a MLL in common. Furthermore, a common MLL was found at locations FRLV and FRAU with a
- 36 distance of 16 km between them.

1 Parentage and sibship analysis

2 After retaining only unique ramets, dataset 1 contained 64 Flemish samples defined as offspring in 3 the analysis. As expected, no recent cultivars were inferred as parents of our Flemish samples. On 4 the other hand, 'Klemmer', 'Belgica' and 'Major' came up several times as potential parents with high 5 probabilities. These cultivars are at least 130 years old and known to have been widely planted in 6 Flanders. 'Klemmer' was inferred as a parent of a sample of BEGE and of BEME1, where also ramets 7 of 'Klemmer' were found (see the clonality results). 'Major' was found to be a potential parent of 8 individuals at the locations BEOO, BESP, BEEN and BEDI. Additionally, the BEDI location contained 9 another individual, possibly parented by 'Belgica'. This individual is a ramet of MLL 'A', which was 10 also present in BEME2 and BESC. Furthermore, MLL 'E' found in BEMA and BESC has 'Belgica' as a 11 potential parent. In four (50%) of these parent-offspring combinations, the admixture proportions of 12 the parents are different from those of the offspring. This suggests that a possible clonal relationship 13 as opposed to a parent-offspring relationship is less likely.

According to the results of Colony, many of the samples were found to be full-sibs or half-sibs (Fig. S1 in Supplementary Material). Only one of 569 pairwise full-sib relationships between individuals (of BEOO and BEEN) had a probability of 0.848, while other probabilities equalled one. Also, 1 of a total of 89 of inferred pairwise half-sib relationships had a probability of 0.943 and the remaining combinations received the maximum probability value. Location BERI holds many full-sibs of which a few were identified as backcrosses, while the majority seemed to be pure U. glabra. Also, a few peculiar full-sib and half-sib combinations could be detected between samples of distant locations, such as BERI, BEGE, BEEG and BEMA. Here, the full-sib relationships are often between potential pure Wych elms and hybrids (F1 or backcrosses to U. glabra). The majority of the full-sib combinations between pure *U. glabra* and hybrids, including backcrosses, can possibly be attributed by the uncertainty of the species assignment of several samples using NewHybrids. In fact, 90% of the 234 full-sib relationships between different species classes, involved samples that were classified as F1 hybrids or backcrosses to probabilities below 0.90, of which 50% was below 0.70. Taking admixture proportions of these individuals given by BAPS into account, did not change these unlikely combinations. The remaining full-sibs are mainly within particular locations (e.g. BEBR: U. minor) or between locations quite near to each other (e.g. BERI and BETO: *U. glabra*).

As stated before, because we were not able to estimate the age of the Flemish elms, they could just as well be a mixture of parents and offspring instead of pure offspring, potentially causing full-sib associations between individuals of different admixture classes. In addition, the cultivars assigned as potential parents, may be close relatives but not the parents of the alleged offspring. The results of the sibship analysis are therefore only indicative of family relations.

Discussion

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- 36 Occurrence of interspecific hybrids
- 37 Our results indicate that the formation of first-generation hybrids (F1) between *U. minor* and *U. glabra*
- 38 and backcrosses in either direction is common in Flanders. This suggests that no strong pre- or post-

fertilisation barriers between these two species exist. Former morphological studies already suggested that in Flanders and the Netherlands, the full spectrum of the *U. minor-U. glabra* complex is present (e.g. Touw 1963). Still, the hybrids do not seem to be recent, since the level of LD appears to be low and advanced generation hybrids are predominant among hybrids (Fig. 2). Because of the high number of hybrids, hybridisation could have been facilitated by human-mediated habitat disturbance (Ellstrand and Schierenbeck 2000; Rhymer and Simberloff 1996). Hybrid clone mates and full-sibs were often found among different locations, indicating human influence through movement of elm planting material and the establishment of cultivars, often hybrids, near natural elm populations (see below).

It seems that F1 elm hybrids are not sterile, judging by the backcrosses that were found. However, F2 offspring of F1 hybrids were not identified among the sampled trees. A possible reason for failing to detect F2 hybrids, could be an insufficient discriminative power of the AFLP markers, partly because of their dominant character. Still, assignment probabilities to the F2 genotype class never exceeded 0.15. A lack of F2 generations might suggest selection against offspring containing homozygous loci from either species. A loss in fitness might not arise until the second or later hybrid generations because of the increasing severity of homozygous incompatibilities according to the (Bateson-)Dobzhansky-Muller model (Turelli and Orr 2000) or because of outbreeding depression (Edmands 2007; Lynch 1991). Zalapa et al. (2009) also did not find second-generation hybrids between U. pumila and U. rubra. They suggested pollen-stigma incompatibilities between F1 hybrid parents as a possible cause, since F1 hybrids would contain S-alleles from both parents and thus be less likely to crossbreed successfully with another F1 hybrid. On the other hand, Brunet et al. (2013) did find F2 hybrids of *U. pumila* and *U. minor*, which they attributed to the relatively self-compatible nature of *U.* minor, described in the paper of Mittempergher and La Porta (1991). Based on many controlled selfings and crosses between several species and hybrids, including *U.minor* and *U.glabra*, Heybroek (1993) detected, however, a high degree of self-incompatibility. Over a period of ten years, producing 24,149 seedlings out of 2,378 pollination bags, 770 crosses and 196 selfings, he found that cross pollination gave 23 times more viable seeds and seedlings than selfing.

Out of 87 samples taken throughout Italy and in some locations in France, Brunet et al. (2013) identified 13 trees (15%) as F2 with a posterior probability of 0.90 or greater using NewHybrids, or 35% of the total number of hybrids (37). Bearing in mind these relatively high percentages and our more intensive sampling (i.e. on average the same amount of trees was sampled within a smaller region), missing an F2 hybrid seems less plausible, unless they are extremely rare.

Natural and artificial clonal propagation

Evidence of clonality was mostly found among *U. minor* and hybrids with a higher representation of *U. minor* genes. Although clonal reproduction seemed mainly to occur within locations, several ramets of the same MLL were found at different locations in Flanders, with distances of 4.5 km to more than 60 km. This suggests translocation of elm planting material, preferably root suckers, from one location to the other. Ramets of a genotype in BEHO (*U. minor*), of MLL 'C' in BERI and BEEG (*U. x hollandica*), and of MLL '3' in BEBR (*U. minor*) were even detected in the Netherlands using microsatellites (J.

- 1 Buiteveld, unpublished data). More pronounced examples of clonal spread through human activity
- were found for *Populus alba* in Sardinia (Brundu et al. 2008) and in Malta (Fussi et al. 2012).
- 3 Furthermore, many elms in Flanders were found in old hedges (of *U. minor;* Table 1), indicating past
- 4 human activity. One of the MLLs identified among the investigated elms appeared to be the cultivar
- 5 'Klemmer', confirming human involvement in elm establishment in Flanders. 'Klemmer' originates from
- 6 Belgium (Ypres) (Elwes et al. 1906). It dates back from 1877 or maybe 1789. It was always rare in the
- 7 Netherlands and common in Belgium and the north of France in the beginning of the 20th century.
- 8 However, it is now considered to be rare in general (Heybroek et al. 2009).
- 9 As mentioned before, it is difficult to infer an exact threshold to determine which of the samples
- 10 belong to the same MLL. It seemed highly unlikely that the inferred clone mates resulted from random
- mating (P < 0.001). Even though we cannot entirely rule out the occurrence of selfing, the elm species
- 12 in question are basically deemed self-incompatible (Heybroek 1993; Nielsen and Kjaer 2010;
- 13 Townsend 1975). Nearly identical genotypes can also be produced due to extensive inbreeding.
- Nevertheless, we also found such similar genotypes among distant locations and mainly among U.
- minor and hybrids which can easily be propagated vegetatively using root suckers. We are therefore
- 16 quite confident about the identified MLLs and the threshold used.
- 17 A few samples of *U. glabra*, of BERI, were detected as ramets of the same MLL. *U. glabra* is not
- 18 known to regenerate through sprouting, except on trunks of young trees, nor through root suckering.
- 19 The two closest clones in BERI could therefore be remnants of an old coppice tree, although no
- 20 apparent evidence pointing towards this hypothesis was found in the field. Planting, on the other
- 21 hand, is very plausible as cultivation of the species in Flanders has occurred since the 17th century
- 22 (Maes 2006). This could also explain the presence of related genotypes of *U. glabra* in BERI, BEGE
- 23 and BEEG.
- 24 Although it is feasible that, within locations, the clones among *U. minor* and its hybrids with *U. glabra*
- 25 were the products of natural clonal reproduction, they could just as well be planted. Even the ortet
- trees could have originated from another location, most probably from the same region. It is therefore
- 27 difficult to assess human interference on this level.
- 28 Influence of cultivars
- 29 Besides the samples with the same MLL as 'Klemmer' found in BEME1 and BEGE, this cultivar could
- 30 also be the parent of at least two other trees growing in the same locations. Moreover, 'Belgica' and
- 31 'Major' were found as potential parents of several sampled trees. Consequently, the three cultivars
- 32 have influenced the natural elm populations, either through planting and possibly clonal reproduction,
- and through hybridisation. Like 'Klemmer', cultivars 'Major' and 'Belgica' are old. 'Major' is probably
- 34 the oldest and dates from around 1600 or even before that (Heybroek et al. 2009). Its origin is
- 35 uncertain, but seemed to be brought to England in 1689 by Willem III from the Netherlands. It was
- planted frequently in the Netherlands until 'Belgica' became more popular mid-19th century. The latter
- was thought to originate from Belgium in the 18th century, in the Abbey of Dunes with nurseries in
- Furnes and Bruges (Elwes et al. 1906). It was extensively planted starting from 1850. Almost all elm

plantings in Belgium and the Netherlands consisted of this cultivar until 1928. In addition, for a long time, 'Belgica' was used as rootstock. Consequently, their age and frequent use in the past make the three cultivars feasible parents of some of the sampled elms. Due to the Flemish origin of at least 'Klemmer' and 'Belgica', with potentially close relatives still present in Flanders, family connections other than parent-offspring relationships are quite possible and should be taken into consideration. An alternative explanation for the assignment of cultivars as parents could be that some elms are in fact ramets of the cultivars in question, suggesting that more somatic mutations have occurred than accounted for by the current threshold for clone assignment. Nonetheless, half of these parent-offspring combinations displayed different admixture levels between parent and offspring, whereas the opposite would be expected among clones.

11 Consequences for conservation

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- 12 Our results suggest high human impact on elm populations obscuring their natural genetic structure.
- 13 A high number of (advanced generation) hybrids were detected, probably facilitated by human
- 14 activities such as translocation of trees. This was supported by the ramets found among locations. In
- addition, cultivar 'Klemmer' was identified among the investigated elms, as well as several individuals
- showing potential introgression from cultivars 'Major', 'Belgica' and 'Klemmer'.

LD among loci was low but significant and can partly be attributed to clonal reproduction (de Meeûs and Balloux 2004; Tibayrenc et al. 1991). The potential problem of mate or pollen limitation that comes with clonal reproduction could lead to a decrease in fertility or an increase in self-compatibility (Honnay and Jacquemyn 2008; Vallejo-Marín et al. 2010), of which examples can be found in domesticated crop plants (McKey et al. 2010). Linkage was still present when potential clone mates were excluded, which could be the result of a bottleneck in the elm populations (Flint-Garcia et al. 2003) caused by DED and habitat loss. In addition, the investigated elms seemed often highly related, which might suggest non-random mating. Non-random mating is likely to occur in small, fragmented woodland populations such as the Flemish elm populations. This can cause genetic drift and reduced genetic variation (Jump and Peñuelas 2006). Given the fact that biparental inbreeding is likely (e.g. Nielsen and Kjaer 2010), population differentiation might increase as a result of genetic drift and continuing losses of genotypes due to disease and additional loss of habitat. In addition, the spread of cultivars has probably influenced the remaining genetic resources of U. minor and U. glabra, potentially further altering the genetic variation. Steps taken to conserve the remaining elms in a field gene bank are therefore no wasted effort. The genetic markers and methodology used in this study will help in distinguishing clones, pure species, hybrids and cultivars among elms in the ex situ collection.

Since sexual reproduction appears to be rare in some locations in Flanders, especially in *U. minor*, energy put into the formation of F1 hybrids might be considered as wasted reproductive effort of the pure species, because F1 hybrids do not seem to succeed in creating F2 hybrids. The elm hybrids occupy, however, a great part of the ecological niches of both pure elm species in Flanders and therefore partly fulfil the ecological role of the parent taxa. Although the hybrids appear to be mainly

- 1 human induced, the focus should be on conserving their ecological function. In turn, this will help
- 2 maintain the structure of plant communities also containing (at one point) pure elms (Stronen and
- 3 Paquet 2013). Moreover, because of the abundance of clones and highly related individuals, the
- 4 residual gene pool, especially of the pure species, appears small and could become even smaller as
- 5 mentioned above. Introgression can be a source of genetic variation in the elm populations and may
- 6 promote adaptation (Grant and Grant 1994). Consequently, conserving the overall genetic variation is
- 7 warranted. Nonetheless, further research is required to determine whether elm hybrids in Flanders
- 8 have a competitive advantage over pure elm species, especially in regard to DED.

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Supplementary Material

- 16 Fig. S1 Best maximum likelihood sibship assignment plot of the sibship structure for the unique
- 17 ramets of Flemish multilocus lineages of the *U. minor-U. glabra* complex obtained with Colony
- 18 v2.0.1.9. Above diagonal (black dots): full-sibs; below diagonal (grey dots): half-sibs. On both axes,
- 19 the location code of each individual is given

20 Data Archiving Statement

21 AFLP data: data will be deposited at Dryad

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Figures

- **Fig. 1** Map depicting the locations of the elm samples of Flanders (northern Belgium). The codes are given in Table 1.
- **Fig. 2** Posterior probabilities of genotype frequency classes given by NewHybrids (a) and admixture proportions given by BAPS (b) for Flemish elm samples and cultivars of *U. glabra*, *U. minor* or *U. x hollandica*. The first half of the results is given above, the second half below. Samples are indicated with their location code (Table 1) or cultivar name (Table 2). Ramets of the same multilocus lineage (MLL) are indicated with the same number before the location code or a capital letter in case of a MLL found among locations. UM: *U. minor*, UG: *U. glabra*; F1: first generation hybrid; F2: offspring of F1 hybrids; BC to UM/UG: backcross to *U. minor/U. glabra*.
- **Fig. 3** Plot of number of loci versus genotypic richness (G:N) based consecutively on two highly polymorphic markers and sets of multiples of 16 markers with decreasing information content. For each set of loci, a range of G:N values (vertical lines), which were calculated using a different thresholds (0.90 to 0.99 Dice similarity). The line goes through mean G:N values.
- Fig. 4 Frequency distribution of pairwise Dice similarity index values with a maximum of 0.80.

Tables

Table 1 List of sampled locations of *Ulmus minor*, *U. glabra*, their hybrids and *U. procera*. N: number of sampled individuals; N AFLP: number of samples successfully analysed with AFLP; Nc: estimate of total number of individuals present on a location; Type LE: type of landscape element; UM: *U. minor*, UG: *U. glabra*; UH: *U. x hollandica*; UPR: *U. procera*; NA: not available.

Dataset 1: Flemish samples (with species identification based on morphology)										
Location	Country	City	Species (number of trees)	Lon	Lat	Ν	N AFLP	Nc	Type LE	
BEBR	Belgium	Brugge (Sint-Pieters)	UM	3.1728	51.2387	8	7	20	Old hedge	
BEDI	Belgium	Dilbeek	UG	4.2530	50.8536	2	2	10	Forest	
BEDM1	Belgium	Diksmuide	UM	NA	NA	2	2	NA	NA	
BEDM2	Belgium	Diksmuide	UM	2.9040	51.0095	4	4	5 - 10	Old hedge	
BEEG	Belgium	Heverlee	UG	4.6662	50.8541	8	6	20 - 30	Forest	
BEEN	Belgium	Oudenaarde (Ename)	UM	3.6593	50.8616	3	3	20 - 30	Wooded banks	
BEGE	Belgium	Geraardsbergen	UG (7), UH (1)	3.9290	50.7700	8	8	20	Forest	
BEHE	Belgium	Heusden	UM	3.8159	51.0099	2	2	10 - 20	Old hedge	
BEHO	Belgium	Houthulst	UM	2.8720	50.9770	4	4	10 - 20	Old hedge	
BELE	Belgium	Lemberge	UM	3.7693	50.9788	5	5	10 - 20	Old hedge	
BEMA	Belgium	Maarkedal	UM (1), UG (5), UH (1)	3.6520	50.8247	7	7	10 - 15	Wooded bank	
BEME1	Belgium	Merelbeke	UM (5), UH (1)	3.7198	50.9519	6	6	10	Forest	
BEME2	Belgium	Merelbeke	UH	3.7508	50.9739	3	3	10	Forest	
BEOO	Belgium	Oosterzele	UM	3.8065	50.9350	1	1	10	Forest edge	
BERI	Belgium	Riemst	UM (4), UG (29), UH (3)	5.6857	50.8058	38	36	50 - 100	Forest	
BESC	Belgium	Schorisse	UM (1), UG (4), UH (2)	3.7065	50.8028	7	7	20 - 30	Forest, wooded bank	
BESP	Belgium	Sint-Pieters-Kapelle (Herne)	NA	3.9795	50.6930	2	2	NA	NA	
BETO ^a	Belgium	Tongeren	UG	5.4908	50.8035	1	1	NA	Private park	
Dataset 2: reference samples										
Location	Country	City	Species	Lon	Lat	Ν	N AFLP			
FRAM	France	Amplier	UM	2.4010	50.1352	1	1			
FRAR ^b	France	Argentan	UM	-0.0187	48.7402	1	1			
FRAU	France	Aunay	UM	0.6307	49.0205	2	2			

$FRBB^{\mathtt{b}}$	France	Bourg-Blanc	UM	-4.5017	48.5005	1	1
FRBL	France	Blismes	UM	3.8202	47.1315	1	1
FRCB	France	La Chapelle-Bâton	UM	0.3297	46.4746	3	3
FRCM ^b	France	Saint-Martin-de-Ré	UM	-1.3593	46.2027	1	1
FRCU⁵	France	Cucq	UM	1.6207	50.4742	1	1
FRGO ^b	France	Godewaersvelde	UM	2.6380	50.7898	1	1
FRGS⁵	France	Grande-Synthe	UM	2.2897	51.0087	1	1
$FRIL^{\mathtt{b}}$	France	Illkirch-Graffenstaden	UM	7.7185	48.5243	1	1
FRLR	France	Le Rheu	UM	1.7954	48.1011	2	2
FRLV	France	Le Vey	UM	0.4701	48.9175	5	5
$FRLW^{b}$	France	La-Wantzenau	UM	7.8222	48.6575	1	1
FRMA ^b	France	Magnicourt-en-Comte	UM	2.4877	50.4018	1	1
FRME ^b	France	Meteren	UM	2.6880	50.7383	2	2
$FRMQ^b$	France	Mecquignies	UM	3.7890	50.2738	1	1
FROS⁵	France	Ostwald	UM	7.7058	48.5403	1	1
FRSP	France	Saint-Pé-de-Bigorre	UG	0.1552	43.0737	3	2
FRST ^b	France	Strasbourg	UM	7.7537	48.5845	1	1
GEGO1 ^c	Germany	Göttingen	UG	9.1500	51.3333	3	2
GEGO2 ^c	Germany	Göttingen	UM	9.9557	51.6502	1	1
GEKA	Germany	Pfalz	UPR	NA	NA	1	1
GELD ^c	Germany	Lüchow-Dannenberg	UM	10.8833	53.2667	3	3
$GRIR^{\mathtt{d}}$	Greece	Iraklion	UM	24.8062	35.3878	1	1
GRTH ^d	Greece	Thessaloniki	UM	23.7340	40.7523	1	1
ITBO ^e	Italy	Bolzano	UG	11.1167	46.6167	1	1
ITCA ^e	Italy	Catanzaro	UM	16.7500	38.8833	1	1
ITFV ^e	Italy	Fiume Veneto	UM	12.6833	45.9167	1	1
ITLA ^e	Italy	Latina	UM	13.0000	41.4667	1	1
ITMO ^e	Italy	Monfalcone	UM	13.5333	45.8000	1	1
ITNI ^e	Italy	Nimis	UM	13.2500	46.2167	1	1
ITSE ^e	Italy	Sesto Al Reghena	UM	12.7833	45.8667	1	1
ITTA ^e	Italy	Tamai	UM	12.5667	45.9333	2	2

Samples from the RESGEN collection provided by the following institutes:

^a Research Institute for Nature and Forest (INBO), Belgium

^b Institut national de recherche en sciences et technologies pour l'environnement et l'agriculture (Irstea), France

^c Niedersächsische Forstliche Versuchsanstalt (NFV), Germany

^d National Agricultural Research Foundation (NAGREF), Greece

^e Consiglio Nazionale delle Ricerche (CNR), Italy

Table 2 List of sampled elm cultivars included in the study (i.e. dataset 3).

Cultivars	Parents/ species				
Lobel (3 ramets)	clone 202 (<i>U. glabra</i> 'Exoniensis' × <i>U. wallichiana</i>) x				
	clone 336 ('Bea Schwarz' a, selfed)				
Clusius	clone 202 (<i>U. glabra</i> 'Exoniensis' × <i>U. wallichiana</i>) x				
	clone 336 ('Bea Schwarz' a, selfed)				
Sapporo Autumn Gold	U. pumila x U. japonica				
73P	U. pumila (mother tree of 'Sapporo Autumn Gold') x				
	? (open pollinated)				
2P	U. japonica				
Klemmer	U. x hollandica				
	or <i>U. minor</i>				
Dodoens	Selfed seedling of clone 202 (<i>U. glabra</i> 'Exoniensis'				
	× U. wallichiana)				
Groeneveld	clone 49 (U . glabra or U . \times hollandica) \times clone 1 (U .				
	minor)				
Commelin	U. x hollandica 'Vegeta' x clone 1 (U. minor)				
Plantyn (2 ramets)	Clone 202 (<i>U. glabra</i> 'Exoniensis' × <i>U. wallichiana</i>)				
	x clone 302 (<i>U. minor</i> '1' × <i>U. minor</i> '28')				
Christine Buisman	U. minor				
Vegeta	U. x hollandica				
Major	U. x hollandica				
Belgica	U. x hollandica				
Horizontalis	U. glabra				
Dampieri	U. x hollandica				
	or <i>U. minor</i>				
Den Haag	U. pumila x U. x hollandica 'Belgica'				
Columella	Probably selfed seedling of Plantyn				
Sarniensis	U. minor				

^a 'Bea Schwarz' is an *U. minor* or *U.* x hollandica.

Table 3 Information on the variation of the AFLP loci. N: number of samples; NPL: number of polymorphic loci; BF: mean band frequency; PIC: mean polymorphic information content; NPL $_{0.05-0.95}$: number of polymorphic loci with a band frequency between 0.05 and 0.95; PIC $_{0.05-0.95}$: mean polymorphic information content of loci with a band frequency between 0.05 and 0.95.

Primer combinations	N ^a	NPL	BF	PIC	NPL _{0.05-0.95}	PIC _{0.05-0.95}
			(st. dev.)	(st. dev.)		(st. dev.)
EcoRI-AGC/Msel-CTG	174	169	0.35 (0.35)	0.21 (0.17)	110	0.30 (0.14)
EcoRI-ACC/Msel-CTG	174	216	0.26 (0.35)	0.15 (0.15)	100	0.27 (0.14)
Species group	N ^a	NPL	BF	PIC	NPL _{0.05-0.95}	PIC _{0.05-0.95}
			(st. dev.)	(st. dev.)		(st. dev.)
Ulmus minor	77	314	0.31 (0.36)	0.17 (0.14)	187	0.26 (0.13)
Ulmus glabra	34	222	0.35 (0.35)	0.22 (0.15)	163	0.27 (0.14)
Hybrids <i>U. minor – U.</i> glabra	53	249	0.35 (0.36)	0.20 (0.16)	145	0.30 (0.13)
remaining samples	10	248	0.39 (0.27)	0.33 (0.12)	248	0.33 (0.12)

^a Excl. duplicate ramets of Lobel and Plantyn.







