



HAL
open science

Seroprevalence of hepatitis E virus among blood donors on Corsica, France, 2017

Lisandru Capai, Nathanaël Hoze, Jacques Chiaroni, Sylvie Gross, Rachid Djoudi, Rémi Charrel, Jacques Izopet, Frédéric Bosseur, Stephane Priet, Simon Cauchemez, et al.

► **To cite this version:**

Lisandru Capai, Nathanaël Hoze, Jacques Chiaroni, Sylvie Gross, Rachid Djoudi, et al.. Seroprevalence of hepatitis E virus among blood donors on Corsica, France, 2017. *Eurosurveillance*, 2020, 25 (5), pp.1-9. 10.2807/1560-7917.ES.2020.25.5.1900336 . hal-02493954

HAL Id: hal-02493954

<https://hal.science/hal-02493954>

Submitted on 2 Mar 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

Seroprevalence of hepatitis E virus among blood donors on Corsica, France, 2017

Lisandru Capai¹, Nathanaël Hozé², Jacques Chiaroni³, Sylvie Gross⁴, Rachid Djoudi⁴, Rémi Charrel⁵, Jacques Izopet^{6,7}, Frédéric Bosseur⁸, Stéphane Priet⁵, Simon Cauchemez², Xavier de Lamballerie⁵, Alessandra Falchi⁴, Pierre Gallian^{3,4,5}

1. EA 7310, Laboratoire de Virologie, Université de Corse, Corte, France
2. Mathematical Modelling of Infectious Diseases Unit, Institut Pasteur, UMR2000, CNRS, Paris, France
3. Etablissement Français du Sang Provence Alpes Côte d'Azur et Corse, Marseille, France
4. Etablissement Français du Sang, 93210, La Plaine-Saint-Denis, France
5. Unité des Virus Émergents (UVE): Aix Marseille Univ, IRD 190, INSERM 1207, IHU Méditerranée Infection, Marseille, France
6. Laboratoire de Virologie, Institut Fédératif de Biologie, Centre Hospitalier et Universitaire, Toulouse, France
7. Institut National de la Santé et de la Recherche Médicale Unité 1043, Université Toulouse III, Toulouse, France
8. Sciences Pour l'Environnement – UMR CNRS 6134 Université de Corse, Corte, France

Correspondence: Lisandru Capai (capai_l@univ-corse.fr)

Citation style for this article:

Capai Lisandru, Hozé Nathanaël, Chiaroni Jacques, Gross Sylvie, Djoudi Rachid, Charrel Rémi, Izopet Jacques, Bosseur Frédéric, Priet Stéphane, Cauchemez Simon, de Lamballerie Xavier, Falchi Alessandra, Gallian Pierre. Seroprevalence of hepatitis E virus among blood donors on Corsica, France, 2017. *Euro Surveill.* 2020;25(5):pii=1900336. <https://doi.org/10.2807/1560-7917.ES.2020.25.5.1900336>

Article submitted on 28 May 2019 / accepted on 04 Nov 2019 / published on 06 Feb 2020

Background: Hepatitis E virus (HEV) is an emerging zoonotic pathogen and an important cause of acute viral hepatitis in European countries. Corsica Island has been previously identified as a hyperendemic area for HEV. **Aim:** Our aim was to characterise the prevalence and titres of IgG antibodies to HEV among blood donors on Corsica and establish a model of the annual force of infection. **Methods:** Between September 2017 and January 2018, 2,705 blood donations were tested for anti-HEV IgG using the Wantai HEV IgG enzyme immunoassay. **Results:** The overall seroprevalence was 56.1%. In multivariate analysis, seroprevalence was higher in men than in women (60.0% vs 52.2%; $p < 0.01$), increased with age and was significantly higher among donors born on Corsica (60.6% vs 53.2%; $p < 0.01$). No significant difference was observed between the five districts of the island. IgG anti-HEV titres were mostly low (70% of positive donors had titres < 3 IU/mL). In Corsican natives, increasing seroprevalence by age could be explained by models capturing a loss of immunity (annual probability of infection: 4.5%; duration of immunity: 55 years) or by age-specific probabilities of infection (3.8% for children, 1.3% for adults). **Conclusion:** We confirmed the high HEV seroprevalence on Corsica and identified three aspects that should be further explored: (i) the epidemiology in those younger than 18 years, (ii) common sources of contamination, in particular drinking water, that may explain the wide exposure of the population, and (iii) the actual protection afforded by the low IgG titres observed and the potential susceptibility to secondary HEV infection.

Introduction

Hepatitis E virus (HEV) is a virus with a single-stranded positive-sense RNA of ca 7.5 kb and belongs to the

family *Hepeviridae* (genus *Orthohepevirus*) [1]. HEV was originally described as a non-enveloped virus [2,3] but a quasi-enveloped form has been observed in blood [4,5]. HEV strains infecting humans are currently classified in five genotypes (HEV-1 to HEV-4 and HEV-7) [6] but belong to a single serotype.

HEV-1 and 2 infections have been reported in humans, with person-to-person transmission occurring via the faecal-oral route. These strains are responsible for both epidemics and sporadic cases in low and middle income countries. HEV-3 and 4 are zoonotic pathogens infecting humans and domestic (e.g. pigs) and wild animal species (e.g. boars, deer) which constitute the animal reservoir [7]. The main route of infection for genotypes 3 and 4 is the consumption of contaminated food or water or the contact with infected animals [1,8]. Genotypes 3 and 4 are responsible for sporadic autochthonous cases in high-income countries.

HEV infection can lead to acute hepatitis but can also, in a considerable proportion of cases, be asymptomatic (or at least present in a way that does not call medical attention) [9-11]. Severe presentations (fulminant hepatitis) have been reported when infection occurs during pregnancy [12,13]. In low and middle income countries, chronic forms of infection have been reported in immunocompromised patients [14-17] or those with pre-existing liver diseases [18] infected by HEV genotype 3 [19,20] or genotype 4 [17,21].

Interpretation and comparison of the seroprevalence studies remain difficult because sensitivity and specificity rates of the different anti-HEV IgG assays vary widely [22,23]. The Wantai assay is frequently used in European countries [24] and has specificity and

TABLE 1

Univariate analysis of biological and epidemiological factors associated with IgG antibodies to hepatitis E virus, Corsica, 2017 (n = 2,705)

Parameters	Variables	n	Anti-HEV IgG-positive		Variable	p value	OR (95% CI)
			n	%			
Sex	Women	1,347	703	52.19	Men vs women	0.000024	1.37 (1.18–1.60)
	Men	1,358	815	60.01			
Age group (years)	18–27	605	278	45.95	One age group vs all other age groups	<0.00001 0.0008 NS <0.0001 0.0003	0.59 (0.49–0.70) 0.72 (0.59–0.88) NS 1.74 (1.44–2.12) 1.44 (1.17, 1.77)
	28–37	492	244	49.59			
	38–47	575	323	56.17			
	48–57	566	377	66.61			
	58–70	467	296	63.38			
Place of residence (district)	Ajaccio	1,047	611	58.36	One district vs all other districts	0.023 NS NS NS NS	1.18 (1.01–1.37) NS NS NS NS
	Bastia	560	304	54.29			
	Calvi	258	140	54.26			
	Corte	419	223	53.22			
	Sartène	421	240	57.01			
Native of Corsica	Yes	1,068	647	60.58	Native vs non-native	<0.0001	1.35 (1.16–1.58)
	No	1,637	871	53.21			
ABO blood group	A	1,019	569	55.84	One BG vs all other BG	NS NS NS NS	NS NS NS NS
	B	257	136	52.92			
	AB	105	65	61.90			
	O	1,324	748	56.50			
Rhesus D	Positive	2,250	1,261	56.04	Rh+ vs Rh–	NS	NS
	Negative	455	257	56.48			
Kell	Positive	238	135	56.72	Kell+ vs Kell–	NS	NS
	Negative	2,467	1,383	56.06			

BG: blood group; CI: confidence interval; HEV: hepatitis E virus; NS: non-significant value; Rh: Rhesus factor.

sensitivity values for HEV IgG in the order of magnitude of, respectively, 93.2% and 97.8% [25–27]. This test is based on a recombinant antigen corresponding to ORF2, which encodes the structural protein of the icosahedral capsid [28]. Epidemiological studies in Europe, conducted with the Wantai serological test, showed great geographical heterogeneity of anti-HEV carriers with seroprevalences ranging from 5–6% in Scotland [29,30] to 52% in south-western France [31]. Important variability was observed even within individual countries [32–34]. A nationwide seroprevalence study conducted in France reported a 22% anti-HEV IgG seroprevalence in the French blood donor population tested in 2011 and 2012 [32]. In this study, seropositivity was associated with male sex, increasing age, and consumption of oysters, pork meat and raw pork liver sausages. Drinking bottled water was associated with lower anti-HEV IgG seropositivity rate. A national report on the surveillance of HEV in France has shown an increase in the number of autochthonous cases detected between 2002 and 2016 (from nine to 2,292) [35]. However, this increase could be due to awareness among physicians and in the general population, resulting in increased testing [35].

With a population of 338,000 inhabitants in 2018 [36], Corsica is a large French island (8,680 km²) located in the Mediterranean Sea in the south-east of mainland France. The aim of the study was to investigate HEV seroprevalence in volunteer blood donors from Corsica, an area considered as a zone of high prevalence [32] and to assess the level of immunity in the population together with a fine-scale analysis of prevalence in Corsican districts.

Methods

Studied population

Blood donors accepted for donation according to the national requirements and living on Corsica were included in the study from September 2017 to January 2018. Only five of 2,710 eligible donors (0.18%) did not consent that their samples might be used for epidemiological studies and were excluded; accordingly the population studied provided a good picture of the population of blood donors in Corsica. French (including Corsican) blood donors are healthy volunteers 18 to 70 years-old. This population, when compared with the general population, presents well-known representativeness biases: children and young adults (<18-years-old) are not represented, the elderly population

TABLE 2

Multivariate analysis of biological and epidemiological factors associated with IgG antibodies to hepatitis E virus, Corsica, 2017 (n = 2,705)

Parameters	Variable	p value	AOR (95% CI)
Sex	Men vs women	0.002	1.28 (1.12–1.46)
Age group (years)	18–27	<0.00001	0.60 (0.49–0.73)
	28–37	0.018	0.74 (0.60–0.91)
	48–57	0.00021	1.68 (1.29–1.94)
	58–70	0.03	1.32 (1.07–1.63)
Native of Corsica	Native vs non-native	<0.0000001	1.62 (1.41–1.86)

AOR: adjusted odds ratio; CI: confidence interval.

Age of reference: 38–47 years. The table includes only risk factors associated with p value < 0.05 in the multivariate analysis.

(>65-years-old) is underestimated and blood donors are healthy. Such biases have never been associated in the age class of recruitment with a significant distortion of sero-epidemiological results of infectious diseases in comparison with the general population, as long as the analysis does not imply agents of chronic diseases disqualifying for blood donation. Information about the distribution of the studied population and the general population of Corsica by age group and sex is available in Supplementary Figure S1.

Only data recorded in the database of the French Public Transfusion service (Établissement Français du Sang: EFS) (age, sex, place of residence and birth) were used in the study after anonymisation. There was no specific epidemiological questionnaire for this study, we used the regular blood donation questionnaire that is identical in all French territories (this questionnaire assesses donor health, travel history as well as risk factors for the donor and the recipient such as blood borne infections and medications). Blood donors born in or outside Corsica were assigned to the 'native' or 'non-native' group, respectively.

Laboratory methods

Anti-HEV IgG detection

Plasma samples were tested for the presence of anti-HEV IgG (EFS Provence Alpes Cotes d'Azur and Corse, Marseille) using a single lot reagent of the Wantai HEV IgG enzyme immunoassay kit (Wantai Biologic Pharmacy Enterprise, Beijing, PRC). The assay is based on a recombinant antigen encoded by a structural region of ORF-2 derived from a Chinese isolate of HEV-1 [28,37] and its analytical and clinical performances have been evaluated recently [38]. For each sample, we calculated the ratio sample OD/cut-off OD. A result was considered positive if the sample ratio was ≥ 1 . The limit of detection of the Wantai IgG anti-HEV assay is 0.25 IU/mL and was determined using World Health Organization (WHO) standards [39].

Anti-HEV IgG quantification

Anti-HEV IgG titres were estimated as previously described [40] using dilution series (ranging from 0.25 to 5.0 IU/mL) from the WHO Anti-HEV IgG reference material, standard 95/584 (supplied by the National Institute for Biological Standards and Control, South Mimms, United Kingdom (UK)). For samples with titres >5.0 IU/mL, final titre was determined after testing dilutions of these samples until the signal was in the linearity part of the titration curve (0.25–5.0 IU/mL)

Blood grouping

ABO, Rhesus and Kell blood groups were determined using a fully automated microplate haemagglutination procedure [41] according to routine EFS procedures.

Statistical analysis

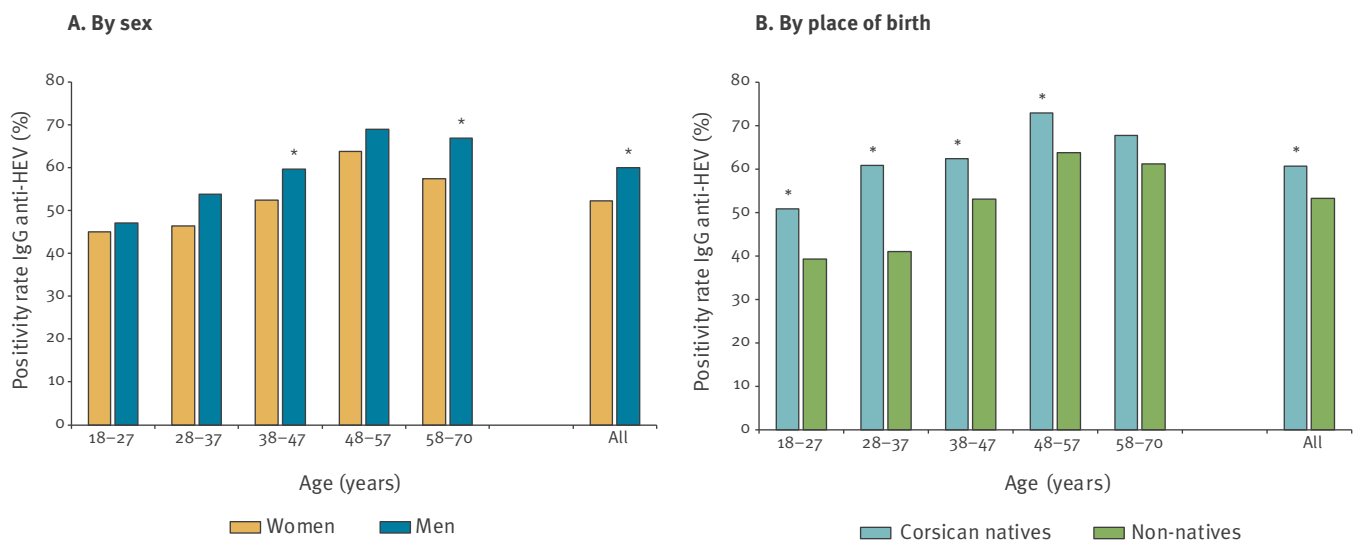
Association of the presence of anti-HEV IgG with biological factors (blood groups) and epidemiological data (sex, age, place of residence and place of birth) was analysed and tested using Fisher's exact test. All variables with a p value < 0.2 were included in the multivariate analyses using an unconditional logistic regression model. Statistical significance was set at a p value < 0.05. Multivariate analysis was performed using an unconditional logistic regression model. Results were considered statistically significant when the p value was lower than 0.05. All statistical analyses were performed using R software version 3.6.1 (R Foundation, Vienna, Austria).

Models of the annual force of infection

The analysis of seroprevalence stratified by age can provide insight about the history of circulation of HEV on Corsica, the risk factors associated with exposure as well as patterns of long-term antibody decay following HEV infection. Indeed, serocatalytic models are now commonly used to reconstruct from such data trends in the force of infection λ_a , defined as the rate of infection among susceptible individuals of age a [42]. For example, if the force of infection is constant with age ($\lambda_a = \lambda$) and if there is no decay in immunity, these models predict that the annual probability of infection for a susceptible individual is

FIGURE 1

Distribution of anti-HEV IgG seroprevalence by age group, Corsica, 2017 (n = 2,705)



The asterisk indicates a statistically significant difference (p value < 0.05) between variables (sex in panel A and place of birth in panel B), for a given age group.

$1 - \exp(-\lambda)$ and the seroprevalence in individuals of age a is $P_a = 1 - \exp(-\lambda \times a)$. By fitting model predictions to data, it is possible to estimate the force of infection.

Here, we considered three competing serocatalytic models to explain the observed age-stratified seroprevalence. Firstly, we considered the simple model described above where the force of infection is age-invariant ($\lambda_a = \lambda$) and there is no decay in immunity. Secondly, we assumed that the force of infection is different for children (λ_c ; <18-years-old) and for adults (λ_A), still assuming no decay in immunity. In this second model, the seroprevalence in individuals of age a is:

Finally, we assumed that the force of infection is age-invariant but immunity decays at a rate ρ :

In the third model, men and women can have a different risk of infection but are assumed to have the same duration of immunity.

The contribution to the likelihood of an individual of age a with seropositivity status s ($s=1$ if seropositive; otherwise $s=0$) is $(P_a)^s (1 - P_a)^{1-s}$. Parameters were estimated in a Bayesian framework using a Markov Chain Monte Carlo (MCMC) method implemented in R using the Rstan package [43]. Four independent chains of 5,000 iterations each were simulated, with the first 2,500 iterations corresponding to the burn-in. We chose flat priors for the force of infection λ and the seroreversion rate ρ .

Means and 95% credible intervals (CrI) of the posterior distributions were calculated. We report the annual

probability of infection of a susceptible individual using the formula $p = 1 - e^{-\lambda}$. The seroreversion rate was converted into a duration of immunity using the expression: $d = 1/\rho$.

Goodness of fit of the different models was assessed using the deviance information criterion (DIC), a measure of the deviance in the likelihood commonly used in parameter estimations in a Bayesian setting. The smaller the DIC of a model, the better the model fit [44]. A DIC difference > 4 corresponds to a substantial improvement of fit.

Ethical statement

The study received approval from the medical and scientific direction of the French Public Transfusion service EFS and from the ad hoc ethics committee (Comité de Protection des Personnes #2016-A01000-51). All French blood donors are voluntary and non-remunerated. Blood donors were informed that samples might be used for epidemiological studies. Donors who did not consent were not included.

Results

Anti-HEV IgG seroprevalence analyses

Study population

A total of 2,705 blood donors (18–70-years-old) living on Corsica for at least 6 months participated in this study, including 1,347 women (49.80%) with a mean age of 42 years and 1,358 men (50.20%) with a mean age of 43 years (Table 1).

TABLE 3

Parameter estimates for serocatalytic models of hepatitis E virus seroprevalence, Corsica, 2017

Models	Mean annual probability of infection in native susceptible men (95% CrI)	Mean annual probability of infection in native susceptible women (95% CrI)	Mean duration of immunity (range)
Age-independent	2.6% (2.3–2.8)	2.5% (2.3–2.8)	Indefinite
Age-dependent <18 years	3.9% (3.2–4.6)	3.7% (3.1–4.4)	Indefinite
Age-dependent >18 years	1.4% (0.89–1.9)	1.3% (0.8–1.9)	
Seroreversion	4.6% (3.6–6.2)	4.4% (3.4–5.8)	55 years (32–93)

CrI: credible interval.

Three models are considered. Age-independent: the force of infection is constant; age-dependent: the force of infection varies with age; seroreversion: the model includes decay of immunity.

Seroprevalence

The overall seroprevalence for anti-HEV IgG antibodies in this population was 56.1% (95% confidence interval (CI): 54.2–58.0). The adjusted estimate of seroprevalence in the general population of Corsica in the same age group was 55.8% (95% CI: 53.9–57.7; see Supplementary Table S1).

Sex

In both univariate and multivariate analysis (Table 1 and 2), the seroprevalence was significantly higher in male (60.0%) than in female (52.2%) donors (p value < 0.001). Significance was lost when the analysis was restricted to native individuals.

Age

Among the 18–57-years-old study participants, the seroprevalence increased with age for both men and women (Figure 1A and Supplementary Figure S2A). The overall increase in seroprevalence was estimated at ca 0.62% and ca 0.72% per year in women and men, respectively (Supplementary Figure S2A). A rough estimate of the increase in seroprevalence in those too young to give blood was calculated between the ages of 1 year (as a proxy for the loss of maternal antibodies and for potential exposure to food-borne contamination) and 23 years (which corresponds to the median age of the 18–27-year age group): the mean yearly seroprevalence increase was 2.05% and 2.14% in women and men, respectively.

Corsican natives vs non-natives

The seroprevalence was higher in Corsican native individuals (60.6%) than in those born elsewhere (53.2%) in both univariate and multivariate analyses (Table 1 and 2). This difference was observed in all age groups (Figure 1B and Supplementary Figure S3). The increase in seroprevalence in the 18–57 years age group was estimated at ca 0.68% and ca 0.86% per year in native and non-native donors, respectively (Supplementary Figure S3). In those too young to give blood (1–23 years-old, see above), seroprevalence was estimated to increase annually by 2.31% and 1.78% in native and non-native individuals, respectively.

In the subpopulation of 18–57-year-old blood donors born on Corsica, the increase in seroprevalence was estimated at ca 0.60% and ca 0.76% per year in women and men, respectively (Supplementary Figure S2B). In those aged 1–23 years, the corresponding annual increase was 2.35% and 2.26% in women and men individuals, respectively.

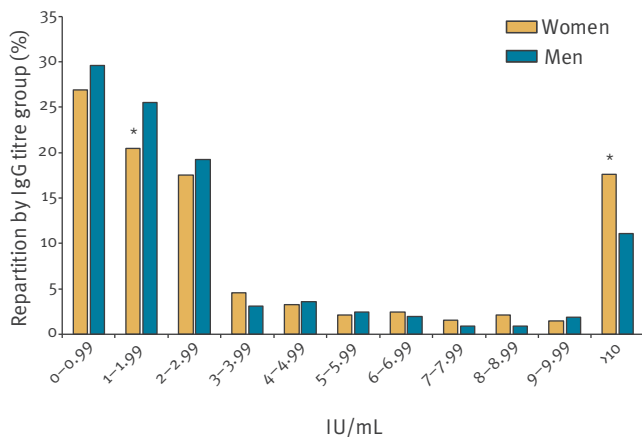
Models of the annual force of infection

This increase in seroprevalence per year of age is expected to underestimate the annual probability of infection in susceptible individuals (i.e. the force of infection) because it does not account for the fact that only a proportion of an age group is susceptible for infection. For example, if the force of infection is 2% but only 50% of the age group is susceptible, the increase in the seroprevalence (1%) will substantially underestimate the force of infection (2%). The fact that the seroprevalence increases at a lower rate in adults than in children may therefore be partly explained by higher levels of susceptibility in children. We therefore used serocatalytic models to derive, from these data, estimates of the force of infection.

Restricting the analysis to native individuals, we found that the model that assumed a constant age-independent force of infection and no loss of immunity over time could not reproduce the large seroprevalence observed in the younger age group nor the plateau in the seroprevalence of older men (Supplementary Figure S4A and B). To integrate these features, we tested two alternative models ('age-dependent' force of infection vs 'seroreversion'). Both models fit the data equally well. In the model where the force of infection was age-dependent (Supplementary Figure S4C and D), we found that the force of infection in children had to be 2.9 (95% CrI: 1.6–4.9) times higher than that in adults to explain the data (Table 3). Alternatively, a model assuming a long-term decay of immunity (average duration of immunity: 55 years; 95% CrI: 32–93) also provided a satisfying fit to the data as a loss of immunity in adults can explain the plateau in the age profile of seroprevalence (Supplementary Figure S4E and F). The estimated force of infection in that model was around 4% for men and women (Table 3).

FIGURE 2

Distribution of anti-HEV IgG titre (IU/mL) estimated using the World Health Organization standard 95/584, by sex, Corsica, 2017 (n = 2,705)



This asterisk indicates statistically significant difference, for a given titre, between sexes ($p < 0.05$).

Anti-HEV IgG geographical distribution

Seroprevalence of anti-HEV IgG was analysed in the five Corsican administrative districts. Positive anti-HEV IgG rates were high, ranging from 53.22% in the Corte district to 58.26% in the Ajaccio district (Table 1 and Supplementary Figure S5)

Anti-HEV IgG quantification

All positive anti-HEV IgG samples ($n = 1,518$) were quantified. Titres ranged from 0.25 to 153 IU/mL, mean and median titres were estimated at 2.91 IU/mL and 0.46 IU/mL, respectively. Around 70% of positive blood samples had an anti-HEV IgG titre < 3 IU/mL and 85.90% had a titre < 10 IU/mL (Figure 2). Among the positive donors, women had a higher mean antibody titre (6.35 IU/mL) than men (4.17 IU/mL) ($p < 0.05$). Among women, 17.64% of positive samples had a titre > 10 IU/ml compared with 11.04% among men ($p < 0.001$). No difference was observed between titre and age groups (Supplementary Figure S6).

Blood grouping

Among the 2,705 blood donors, the distribution of ABO blood groups was: O: 48.9% ($n = 1,324$), A: 37.7% ($n = 1,019$), B: 9.5% ($n = 257$) and AB: 3.9% ($n = 105$). Anti HEV IgG rates (Table 1) for ABO blood groups ranged from 52.92% (B) to 61.90% (AB). The prevalence values of Rhesus-positive (Rh D+ and Kell-positive (KEL+) phenotypes were 56.04% and 56.72%, respectively. No significant difference was observed when comparing the distribution of the anti-HEV IgG positive donations by blood groups or by combinations of them.

Discussion

In 2011 and 2012, a nationwide survey (using the Wantai assay) identified geographical heterogeneity in the distribution of the anti-HEV IgG serological status

among volunteer blood donors in mainland France, and Corsica Island was among the areas with the highest seroprevalence recorded (62%) [32]. The ficatellu, a local pork liver sausage, has been formally identified as a source of HEV food-borne contamination [45], but the actual impact of ficatellu consumption on HEV epidemiology is unknown and other potential sources of contamination remain to be explored. Here, we completed the abovementioned study by performing a specific analysis of a new and larger sample of Corsican blood donors.

Between September 2017 and January 2018 (a period without important touristic activity), we enrolled 2,705 blood donors residing on Corsica among whom 1,518 (56.1%) tested positive for anti-HEV IgG. This confirmed the high endemicity of HEV on Corsica. Other high prevalence areas have been identified in Europe using the same anti-HEV IgG assay, namely central Italy (Abruzzo region, 49%) [46] and south-western France (Midi-Pyrénées Region, 52.5%) [31].

The analysis of the raw data indicated that HEV seroprevalence is higher in men, a finding that was previously described in univariate but not multivariate analysis [32], and also higher in individuals born on Corsica. It was previously proposed that such differences may be explained by sociological factors (specific occupation, hunting, etc.), but there is to date no evidence-based explanation that takes into account the general high prevalence in both men and women and also the difference between the sexes. Association between human leukocyte antigen (HLA) or blood group antigens and infectious diseases has been documented in the literature [47-50] but has, to our best knowledge, never been investigated for HEV infection. Here, we did not identify such a link between ABO, Rhesus and Kell blood groups and the presence of anti-HEV antibodies.

Antibody titres are mostly low, with 77.2%, 81.6% and 85.9% of donors having anti-HEV IgG titres below 5, 7 and 10 IU/mL, respectively. The seroprevalence increases with age, except in those older than ca 60 years. In our models, the latter can be explained either by a force of infection that varies with age (but this is not supported to date by epidemiological or sociological information) or by a loss of specific antibodies over time. Previous studies have shown that antibody levels decrease over time and in some subjects, anti-HEV IgG can disappear after a follow-up ranging between 1 and 22 years [51]. A recent study among blood donors from central Italy using the Wantai assay has reported an HEV seroconversion rate of 2.1 per 100 person-years [52], in the same order of magnitude (1.3-4.6/100 person-years) as those estimated with our models.

Notably, the seroprevalence in the youngest age group of blood donors (18-27 years-old) was more than 10% higher in those born on Corsica (ca 51%) than in those born outside (ca 39%). This is reflected in a force of

infection of 4.3% (95% CrI: 3.5–5.6) per year in natives vs 2.8% (95% CrI: 2.5–3.2) per year in non-natives in the ‘seroreversion model’. However, in the absence of detailed epidemiological data for individuals younger than 18 years, the exact shape of the seroconversion curve in children and teenagers remains to be established. Clearly, identifying determinants of exposure in the Corsican population younger than 18 years is essential to understand the epidemiology of HEV locally. A comparison of the HEV seroprevalence measured here (56.1%) with archived sera from blood donors collected in the year 2000 in both departments of Corsica (53.3%; n=90) (data not shown) suggests that exposure to HEV has been stable in the Corsican population for at least two decades. In addition, we did not identify a significant variation in prevalence by administrative district of residence. Overall, the epidemiological information seems to converge towards a potential common, ubiquitous exposure to HEV infection for individuals living on Corsica.

The animal reservoir (mainly pigs and boars) is consensually regarded as a major source of HEV infection in humans [53,54]. However, the relative importance of direct (i.e. related to pig and boar meat consumption) or indirect routes of contamination remains unclear. Indirect contamination may be related to hand- or fomite-borne virus transmission, but also to contaminated drinking water. Previous information from the French nationwide study identified drinking bottled water as a protective factor against infection [32], and in the rural Auvergne region (central mainland France), water from the public network was identified as the common source of infection for a cluster of seven human cases, with HEV RNA detected in a private well that accidentally contaminated the public water network [55]. Moreover, a recent study in Sweden detected HEV genotype 3 strains in tap water and raw water before treatment [56]. Therefore, as more research is implemented to identify ubiquitous sources of HEV exposure on Corsica, the potential role of drinking water in the spread of HEV infection should be investigated.

Conclusion

Our study confirms that Corsica is a high endemic area for HEV infection, with homogeneous exposure in the different geographical districts. Seroprevalence increases with age until 60 years and is higher in men than in women. Our study identified three priority fields for further investigations on Corsica. Firstly, the epidemiology in the younger age group (under 18 years) is essentially unknown in the absence of biological data and should be explored further. Secondly, common sources of contamination, in particular drinking water, deserve further studies because HEV can be found in faeces and wastewater [57] and Corsica is a region where infection of pigs and boars is frequent [58,59]. The relationship between the animal reservoir, wastewater and the potential contamination of the public water network may be worth exploring. Thirdly, the high proportion of donors with low anti-HEV antibody

titre raises questions about the protection afforded by IgG antibodies and about susceptibility to secondary HEV infection.

Conflict of interest

None declared.

Authors’ contributions

Lisandru Capai, Alessandra Falchi, Pierre Gallian and Xavier de Lamballerie wrote the manuscript. Lisandru Capai, Frédéric Bosseur and Stephane Priet performed serological and/or statistical analyses. Nathanael Hozé and Simon Cauchemez performed modelling studies of the infection. Sylvie Gross, Remi Charrel and Jacques Chiaroni analysed the data. Pierre Gallian, Xavier de Lamballerie, Rachid Djoudi, and Jacques Izopet drafted the manuscript. All authors participated in review of the manuscript.

References

1. Kamar N, Bendall R, Legrand-Abravanel F, Xia NS, Ijaz S, Izopet J, et al. Hepatitis E. *Lancet*. 2012;379(9835):2477-88. [https://doi.org/10.1016/S0140-6736\(11\)61849-7](https://doi.org/10.1016/S0140-6736(11)61849-7) PMID: 22549046
2. Bradley DW. Enterically-transmitted non-A, non-B hepatitis. *Br Med Bull*. 1990;46(2):442-61. <https://doi.org/10.1093/oxfordjournals.bmb.a072409> PMID: 2116213
3. Emerson SU, Purcell RH. Hepatitis E virus. *Rev Med Virol*. 2003;13(3):145-54. <https://doi.org/10.1002/rmv.384> PMID: 12740830
4. Takahashi M, Tanaka T, Takahashi H, Hoshino Y, Nagashima S, Jirintai, et al. Hepatitis E Virus (HEV) strains in serum samples can replicate efficiently in cultured cells despite the coexistence of HEV antibodies: characterization of HEV virions in blood circulation. *J Clin Microbiol*. 2010;48(4):1112-25. <https://doi.org/10.1128/JCM.02002-09> PMID: 20107086
5. Chapuy-Regaud S, Dubois M, Plisson-Chastang C, Bonnefois T, Lhomme S, Bertrand-Michel J, et al. Characterization of the lipid envelope of exosome encapsulated HEV particles protected from the immune response. *Biochimie*. 2017;141:70-9. <https://doi.org/10.1016/j.biochi.2017.05.003> PMID: 28483690
6. Doceul V, Bagdassarian E, Demange A, Pavo N. Zoonotic hepatitis E virus: classification, animal reservoirs and transmission routes. *Viruses*. 2016;8(10):E270. <https://doi.org/10.3390/v8100270> PMID: 27706110
7. Pavo N, Meng XJ, Renou C. Zoonotic hepatitis E: animal reservoirs and emerging risks. *Vet Res*. 2010;41(6):46. <https://doi.org/10.1051/vetres/20100018> PMID: 20359452
8. Dalton HR, Bendall R, Ijaz S, Banks M. Hepatitis E: an emerging infection in developed countries. *Lancet Infect Dis*. 2008;8(11):698-709. [https://doi.org/10.1016/S1473-3099\(08\)70255-X](https://doi.org/10.1016/S1473-3099(08)70255-X) PMID: 18992406
9. Aggarwal R. Clinical presentation of hepatitis E. *Virus Res*. 2011;161(1):15-22. <https://doi.org/10.1016/j.virusres.2011.03.017> PMID: 21458513
10. Guillois Y, Abravanel F, Miura T, Pavo N, Vaillant V, Lhomme S, et al. High proportion of asymptomatic infections in an outbreak of hepatitis E associated with a spit-roasted piglet, France, 2013. *Clin Infect Dis*. 2016;62(3):351-7. <https://doi.org/10.1093/cid/civ862> PMID: 26429341
11. Said B, Ijaz S, Kafatos G, Booth L, Thomas HL, Walsh A, et al. Hepatitis E outbreak on cruise ship. *Emerg Infect Dis*. 2009;15(11):1738-44. <https://doi.org/10.3201/eid1511.091094> PMID: 19891860
12. Kumar A, Beniwal M, Kar P, Sharma JB, Murthy NS. Hepatitis E in pregnancy. *Int J Gynaecol Obstet*. 2004;85(3):240-4. <https://doi.org/10.1016/j.ijgo.2003.11.018> PMID: 15145258
13. Jilani N, Das BC, Husain SA, Baweja UK, Chattopadhyaya D, Gupta RK, et al. Hepatitis E virus infection and fulminant hepatic failure during pregnancy. *J Gastroenterol Hepatol*. 2007;22(5):676-82. <https://doi.org/10.1111/j.1440-1746.2007.04913.x> PMID: 17444855
14. de Niet A, Zaaier HL, ten Berge I, Weegink CJ, Reesink HW, Beuers U. Chronic hepatitis E after solid organ transplantation. *Neth J Med*. 2012;70(6):261-6. PMID: 22859417

15. Gauss A, Wenzel JJ, Flechtenmacher C, Navid MH, Eisenbach C, Jilg W, et al. Chronic hepatitis E virus infection in a patient with leukemia and elevated transaminases: a case report. *J Med Case Reports*. 2012;6(1):334. <https://doi.org/10.1186/1752-1947-6-334> PMID: 23031738
16. Debing Y, Moradpour D, Neyts J, Gouttenoire J. Update on hepatitis E virology: Implications for clinical practice. *J Hepatol*. 2016;65(1):200-12. <https://doi.org/10.1016/j.jhep.2016.02.045> PMID: 26966047
17. Behrendt P, Steinmann E, Manns MP, Wedemeyer H. The impact of hepatitis E in the liver transplant setting. *J Hepatol*. 2014;61(6):1418-29. <https://doi.org/10.1016/j.jhep.2014.08.047> PMID: 2519557
18. Crossan CL, Simpson KJ, Craig DG, Bellamy C, Davidson J, Dalton HR, et al. Hepatitis E virus in patients with acute severe liver injury. *World J Hepatol*. 2014;6(6):426-34. <https://doi.org/10.4254/wjh.v6.i6.426> PMID: 25018853
19. Hoofnagle JH, Nelson KE, Purcell RH. Hepatitis E. *N Engl J Med*. 2012;367(13):1237-44. <https://doi.org/10.1056/NEJMra1204512> PMID: 23013075
20. Te HS, Drobeniuc J, Kamili S, Dong C, Hart J, Sharapov UM. Hepatitis E virus infection in a liver transplant recipient in the United States: a case report. *Transplant Proc*. 2013;45(2):810-3. <https://doi.org/10.1016/j.transproceed.2012.08.020> PMID: 23498824
21. Geng Y, Zhang H, Huang W, J Harrison T, Geng K, Li Z, et al. Persistent hepatitis e virus genotype 4 infection in a child with acute lymphoblastic leukemia. *Hepat Mon*. 2014;14(1):e15618. PMID: 24596581
22. Shrestha AC, Flower RL, Seed CR, Stramer SL, Faddy HM. A comparative study of assay performance of commercial hepatitis E virus enzyme-linked immunosorbent assay kits in Australian blood donor samples. *J Blood Transfus*. 2016;2016:9647675. <https://doi.org/10.1155/2016/9647675> PMID: 27891290
23. Park HK, Jeong SH, Kim JW, Woo BH, Lee DH, Kim HY, et al. Seroprevalence of anti-hepatitis E virus (HEV) in a Korean population: comparison of two commercial anti-HEV assays. *BMC Infect Dis*. 2012;12(1):142. <https://doi.org/10.1186/1471-2334-12-142> PMID: 22726615
24. Hartl J, Otto B, Madden RG, Webb G, Woolson KL, Kriston L, et al. Hepatitis E seroprevalence in Europe: a meta-analysis. *Viruses*. 2016;8(8):E211. <https://doi.org/10.3390/v8080211> PMID: 27509518
25. Yan Q, Du H, Wang Y. Comparison of two diagnostic reagents to detect anti-hepatitis E virus IgG antibodies. *Chin J Zoonoses*. 2008;24:1087-9.
26. Abravanel F, Chapuy-Regaud S, Lhomme S, Miedougé M, Peron JM, Alric L, et al. Performance of anti-HEV assays for diagnosing acute hepatitis E in immunocompromised patients. *J Clin Virol*. 2013;58(4):624-8. <https://doi.org/10.1016/j.jcv.2013.10.003> PMID: 24183927
27. Wantai HEV-IgG ELISA. Beijing: Wantai Biological Pharmacy Enterprise Co, Ltd. Available from: http://www.ystwt.cn/IFU/HEV/HEV-IgG_CE.pdf
28. Zhang J, Ge SX, Huang GY, Li SW, He ZQ, Wang YB, et al. Evaluation of antibody-based and nucleic acid-based assays for diagnosis of hepatitis E virus infection in a rhesus monkey model. *J Med Virol*. 2003;71(4):518-26. <https://doi.org/10.1002/jmv.10523> PMID: 14556264
29. Cleland A, Smith L, Crossan C, Blatchford O, Dalton HR, Scobie L, et al. Hepatitis E virus in Scottish blood donors. *Vox Sang*. 2013;105(4):283-9. <https://doi.org/10.1111/vox.12056> PMID: 23763589
30. Thom K, Gilhooly P, McGowan K, Malloy K, Jarvis LM, Crossan C, et al. Hepatitis E virus (HEV) in Scotland: evidence of recent increase in viral circulation in humans. *Euro Surveill*. 2018;23(12):17-00174. <https://doi.org/10.2807/1560-7917.ES.2018.23.12.17-00174> PMID: 29589577
31. Mansuy JM, Bendall R, Legrand-Abravanel F, Sauné K, Miédouge M, Ellis V, et al. Hepatitis E virus antibodies in blood donors, France. *Emerg Infect Dis*. 2011;17(12):2309-12. <https://doi.org/10.3201/eid1712.110371> PMID: 22172156
32. Mansuy JM, Gallian P, Dimeglio C, Saune K, Arnaud C, Pelletier B, et al. A nationwide survey of hepatitis E viral infection in French blood donors. *Hepatology*. 2016;63(4):1145-54. <https://doi.org/10.1002/hep.28436> PMID: 27008201
33. Marcantonio C, Pezzotti P, Bruni R, Taliani G, Chionne P, Madonna E, et al. Incidence of Hepatitis E virus infection among blood donors in a high endemic area of central Italy. *J Viral Hepat*. 2018;jvh.13049. <https://doi.org/10.1111/jvh.13049>
34. Slot E, Hogema BM, Riezebos-Brilman A, Kok TM, Molier M, Zaaier HL. Silent hepatitis E virus infection in Dutch blood donors, 2011 to 2012. *Euro Surveill*. 2013;18(31):20550. <https://doi.org/10.2807/1560-7917.ES2013.18.31.20550> PMID: 23929229
35. Couturier E, Abravanel F, Figoni J, Van Cauteren D, Septfons A, Lhomme S, et al. Surveillance de l'hépatite E en France, 2002-2016. [Hepatitis E surveillance in France, 2002-2016]. *Bulletin Épidémiologique Hebdomadaire*. 2018;28:566-74. French. Available from: <https://www.santepubliquefrance.fr/maladies-et-traumatismes/hepatites-virales/hepatite-e/documents/article/surveillance-de-l-hepatite-e-en-france-2002-2016>
36. Institut national de la statistique et des études Économiques (INSEE). Estimations de population par sexe et âge au 1er janvier 2019. [Estimate of the population by sex and age on 1 January 2019]. Montrouge: INSEE; 2019. French. Available from: <https://www.insee.fr/fr/statistiques/2012692>
37. Li X, Kamili S, Krawczynski K. Quantitative detection of hepatitis E virus RNA and dynamics of viral replication in experimental infection. *J Viral Hepat*. 2006;13(12):835-9. <https://doi.org/10.1111/j.1365-2893.2006.00754.x> PMID: 17109683
38. Avellon A, Morago L, Garcia-Galera del Carmen M, Munoz M, Echevarría JM. Comparative sensitivity of commercial tests for hepatitis E genotype 3 virus antibody detection. *J Med Virol*. 2015;87(11):1934-9. <https://doi.org/10.1002/jmv.24251> PMID: 25959136
39. Abravanel F, Chapuy-Regaud S, Lhomme S, Miedougé M, Peron JM, Alric L, et al. Performance of anti-HEV assays for diagnosing acute hepatitis E in immunocompromised patients. *J Clin Virol*. 2013;58(4):624-8. <https://doi.org/10.1016/j.jcv.2013.10.003> PMID: 24183927
40. Abravanel F, Lhomme S, Chapuy-Regaud S, Mansuy JM, Muscari F, Sallusto F, et al. Hepatitis E virus reinfections in solid-organ-transplant recipients can evolve into chronic infections. *J Infect Dis*. 2014;209(12):1900-6. <https://doi.org/10.1093/infdis/jiu032> PMID: 24436450
41. Rookard LE, Edmondson O, Greenwell P. ABO reverse grouping: effect of varying concentrations of the enzyme bromelain. *Br J Biomed Sci*. 2009;66(2):93-7. <https://doi.org/10.1080/09674845.2009.11730251> PMID: 19637650
42. Hens N, Aerts M, Faes C, Shkedy Z, Lejeune O, Van Damme P, et al. Seventy-five years of estimating the force of infection from current status data. *Epidemiol Infect*. 2010;138(6):802-12. <https://doi.org/10.1017/S0950268809990781> PMID: 19765352
43. RStan: the R interface to Stan. R package version, 2(1). Vienna: Stan Development Team; 2019. Available from: <https://cran.r-project.org/web/packages/rstan/vignettes/rstan.html>
44. Spiegelhalter DJ, Best NG, Carlin BR, van der Linde A. Bayesian measures of model complexity and fit. *J R Stat Soc B*. 2002;64(4):583-639. <https://doi.org/10.1111/1467-9868.00353>
45. Colson P, Borentain P, Queyriaux B, Kaba M, Moal V, Gallian P, et al. Pig liver sausage as a source of hepatitis E virus transmission to humans. *J Infect Dis*. 2010;202(6):825-34. <https://doi.org/10.1086/655898> PMID: 20695796
46. Lucarelli C, Spada E, Taliani G, Chionne P, Madonna E, Marcantonio C, et al. High prevalence of anti-hepatitis E virus antibodies among blood donors in central Italy, February to March 2014. *Euro Surveill*. 2016;21(30):30299. <https://doi.org/10.2807/1560-7917.ES.2016.21.30.30299> PMID: 27494608
47. Heggelund JE, Varrot A, Imbert A, Krengel U. Histo-blood group antigens as mediators of infections. *Curr Opin Struct Biol*. 2017;44:190-200. <https://doi.org/10.1016/j.sbi.2017.04.001> PMID: 28544984
48. Politis C, Parara M, Kremastinou J, Hasapoulou E, Iniotaki A, Siorenta A, et al. Associations of ABO, D, and Lewis blood groups and HLA Class I and Class II alleles with West Nile virus Lineage 2 disease outcome in Greece, 2010 to 2013. *Transfusion*. 2016;56(8):2115-21. <https://doi.org/10.1111/trf.13667> PMID: 27245377
49. Kaidarova Z, Bravo MD, Kamel HT, Custer BS, Busch MP, Lanteri MC. Blood group A and D negativity are associated with symptomatic West Nile virus infection. *Transfusion*. 2016;56(7):1699-706. <https://doi.org/10.1111/trf.13622> PMID: 27189860
50. Nordgren J, Svensson L. Genetic susceptibility to human norovirus infection: an update. *Viruses*. 2019;11(3):E226. <https://doi.org/10.3390/v11030226> PMID: 30845670
51. Servant-Delmas A, Abravanel F, Lefrère JJ, Lionnet F, Hamon C, Izopet J, et al. New insights into the natural history of hepatitis E virus infection through a longitudinal study of multitransfused immunocompetent patients in France. *J Viral Hepat*. 2016;23(7):569-75. <https://doi.org/10.1111/jvh.12531> PMID: 26990022
52. Marcantonio C, Pezzotti P, Bruni R, Taliani G, Chionne P, Madonna E, et al. Incidence of hepatitis E virus infection among blood donors in a high endemic area of Central Italy. *J Viral Hepat*. 2019;26(4):506-12. PMID: 30548124

53. Thiry D, Mauroy A, Pavio N, Purdy MA, Rose N, Thiry E, et al. Hepatitis E virus and related viruses in animals. *Transbound Emerg Dis.* 2017;64(1):37-52. <https://doi.org/10.1111/tbed.12351> PMID: 25919649
54. Spahr C, Knauf-Witzens T, Vahlenkamp T, Ulrich RG, Johne R. Hepatitis E virus and related viruses in wild, domestic and zoo animals: A review. *Zoonoses Public Health.* 2018;65(1):11-29. <https://doi.org/10.1111/zph.12405> PMID: 28944602
55. Couturier E, Abravanel F, Figoni J, Van Cauteren D, Septfons A, Lhomme S, et al. Surveillance de l'hépatite E en France, 2002-2016. [Hepatitis E surveillance in France, 2002-2016]. *Bull Epidémiol Hebd.* 2018;(28):566-74. French. Available from: http://invs.santepubliquefrance.fr/beh/2018/28/2018_28_1.html
56. Wang H, Kjellberg I, Sikora P, Rydberg H, Lindh M, Bergstedt O, et al. Hepatitis E virus genotype 3 strains and a plethora of other viruses detected in raw and still in tap water. *Water Res.* 2020;168:115141. <https://doi.org/10.1016/j.watres.2019.115141> PMID: 31590036
57. Bisseux M, Colombet J, Mirand A, Roque-Afonso AM, Abravanel F, Izopet J, et al. Monitoring human enteric viruses in wastewater and relevance to infections encountered in the clinical setting: a one-year experiment in central France, 2014 to 2015. *Euro Surveill.* 2018;23(7):17-00237. <https://doi.org/10.2807/1560-7917.ES.2018.23.7.17-00237> PMID: 29471623
58. Capai L, Maestrini O, Casabianca F, Villechenaud N, Masse S, Bosseur F, et al. Drastic decline of hepatitis E virus detection in domestic pigs after the age of 6 months, Corsica, France. *Transbound Emerg Dis.* 2019;66(6):2462-73. <https://doi.org/10.1111/tbed.13304> PMID: 31336031
59. Jori F, Laval M, Maestrini O, Casabianca F, Charrier F, Pavio N. Assessment of domestic pigs, wild boars and feral hybrid pigs as reservoirs of hepatitis E virus in Corsica, France. *Viruses.* 2016;8(8):E236. <https://doi.org/10.3390/v8080236> PMID: 27556478

License, supplementary material and copyright

This is an open-access article distributed under the terms of the Creative Commons Attribution (CC BY 4.0) Licence. You may share and adapt the material, but must give appropriate credit to the source, provide a link to the licence and indicate if changes were made.

Any supplementary material referenced in the article can be found in the online version.

This article is copyright of the authors or their affiliated institutions, 2020.