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Pattern of latrine use by domestic cats on dairy farms and the implications for *Toxoplasma gondii* transmission

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Abstract

Toxoplasma gondii is the parasite responsible for toxoplasmosis, a highly prevalent zoonosis that affects humans and warm-blooded animals. Faeces of infected cats can contain millions of *T. gondii* oocysts, which remain infectious in the environment for months. Sites repeatedly used by cats for defecation ('latrines') are recognised as hotspots of *T. gondii* soil contamination, but this contamination varies from one latrine to another. To understand this spatial heterogeneity, camera traps were deployed in 39 cat latrines on three dairy farms with high-density cat populations and programmed to record visits during sixteen 10-day sessions, rotating between three farms over a period of a year. Generalized Linear Mixed Models were used to test the effects of cat sexual maturity, latrine location and season on the number of cat faeces deposited and on the number of cats defecating per latrine, as determined from the analysis of 41,282 video recordings. Sexually immature cats defecated 6.60-fold (95% CI = [2.87–15.25]) more often in latrines located close to a feeding site than in other latrines. This pattern was also observed for mature males (odds ratio [OR] = 9.42, 95% CI = [3.29–26.91]), especially during winter, but not for mature females (OR = 1.77, 95% CI = [0.80–3.94]). The number of defecating cats was also 2.67-fold (95% CI = [1.66 – 4.30], $P < 0.001$) higher in latrines located close to a feeding point than in those located far from it, regardless of cat category and season. Visits by intermediate *T. gondii* hosts (micromammals, birds and others) were also recorded. Out of the 39 latrines, 30 (76.92%) were visited by at least one intermediate host during the study period, and some latrines were highly frequented (up to 8.74 visits/day on average). These results provide evidence that the location of food resources in dairy farms influences the latrine use pattern by cats. Highly frequented latrines can be of high risk of *T. gondii* infection for definitive and intermediate hosts.

Keywords: *Felis silvestris catus*; video trap; cat faeces; toxoplasmosis; intermediate hosts.

1. Introduction

Wild and domestic carnivores are often infected by gastrointestinal zoonotic parasites (Polley 2005; Di Cerbo et al., 2008; Baneth et al., 2016), so their faeces are an important source of environmental contamination with parasitic eggs or (oo)cysts (Robertson and Thompson, 2002). The distribution of infected carnivore faeces largely determines the probability of contact between parasites and their hosts (Traversa et al. 2014). It may also be an important determinant of the zoonotic disease risk for humans especially when faeces are concentrated in human-use environment (Knapp et al., 2018, Bastien et al., 2018). As a consequence, identifying the spatiotemporal pattern of faeces deposition by carnivores constitutes a major issue in understanding the transmission dynamics of zoonotic parasites (Afonso et al., 2008; Milkovic et al., 2009; Raoul et al., 2015).

The faeces deposition pattern by carnivores is usually a non-random process. Several behavioural constraints could exert concurrently, such as the use of faeces for chemical communication (e.g. Rodgers et al., 2015), space use pattern and movements, themselves depending on ecological factors such as microhabitat and/or prey distribution (e.g. Guislain et al., 2007), selection of specific soil substrate to defecate (e.g. Soler et al., 2009), and/or repeated use of shared defecation sites by one or more individuals (e.g. Jordan et al., 2007). Repeated defecation at the same locations, called “latrines”, is observed in several wild carnivore species (see Macdonald, 1985; Rodgers et al., 2015; Buesching et al., 2016) as well as in the domestic cat (*Felis silvestris catus*) when living in social groups (Bradshaw et al., 2012). This particular pattern of faeces deposition may result in the accumulation of free parasite stages in spatially restricted areas, as demonstrated for the high occurrence of *Baylisascaris procyonis* eggs in raccoons (*Procyon lotor*) latrines (Page et al., 1998, Page et al. 2009) or *Toxoplasma gondii* oocysts in domestic cat latrines (Afonso et al., 2008; Simon et al., 2017).

Toxoplasma gondii is a protozoan parasite that can cause serious health issues for humans (Hill and Dubey, 2002) and other warm-blooded intermediate hosts like livestock (Tenter et al., 2000)

and wildlife (Conrad et al., 2005). Felids are the only definitive hosts. They can shed millions of oocysts after ingestion of any of the three infective stages of *T. gondii*. Their transmission pathway is through tachyzoites and bradyzoites contained in preys mainly, through sporozoites to a lesser extent (Dubey, 2006, 2010). After sporulation, oocysts can survive and remain infective for months in soil (Lélu et al. 2012) and water (Lindsay and Dubey, 2009). They represent a potential source of *T. gondii* infection in animals (Van Wormer et al., 2013), as well as in humans through the consumption of contaminated food (fruits and vegetables) and water, or through contact with contaminated soil (Muñoz-Zanzi et al., 2010; Boyer et al., 2011). Among definitive hosts, the domestic cat plays a key-role in the epidemiology of the parasite (Tenter et al., 2000; Dabritz and Conrad, 2010). This is especially true in locations where food resources are deliberately or accidentally provided from human activities. In this case, cats live at high local densities around spatially aggregated resources, such as in livestock farms, around rubbish dumps, industrial and hospital sites, or in certain public areas (Bradshaw et al., 2012). Dairy farms were especially identify as an important source of contamination in rural areas (Lehmann et al., 2003; Richomme et al., 2010; Afonso et al., 2013; Gotteland et al., 2014a). Farm owners use to leave milk available for cats to retain them on the site. This contributes to maintain farm cat populations in densities up to 50 cats/km² (Liberg et al., 2000) with high *T. gondii* seroprevalence and infection rates (Simon et al., 2018).

Two studies conducted on free-ranging cats living in high-density groups reported high levels of soil contamination by the parasite in cat latrines compared to other sites: from 9% to 13% and from 39% to 69% of latrine soil sample contained *T. gondii* DNA in an urban area (Afonso et al., 2008) and in dairy farms (Simon et al., 2017) respectively. However, these studies showed that not all latrines were systematically contaminated, suggesting heterogeneity in the spatial deposition of contaminated faeces. The determinants of such heterogeneity are important to understand, especially if latrines are located in areas where animals or humans can be in contact with the soil. On dairy farms, latrines close to a cat feeding area were more often contaminated

than those distant from a feeding area, suggesting their differential use by cats (Simon et al., 2017). However, authors did not find a significant effect of the number of detected faeces per latrine on the occurrence of *T. gondii* DNA in soil samples. They rather suggested a preferential use of latrines close to feeding areas by juvenile cats, known to be the main contributors to oocyst excretion in the environment (Dubey et al., 1995; Weigel et al., 1995; Gauss et al., 2003).

Juvenile cats have small home range centred on the main food resources (Bradshaw et al., 2012; Turner and Bateson, 2013) and are therefore more likely to use defecation sites close to a feeding area. On the contrary, sexually mature males and females living on farms have a larger home range, averaging 2 ha to 430 ha (Germain et al., 2008; Ferreira et al., 2011), which can vary seasonally in response to behavioural and environmental factors. Sexually mature males are known to extend their home range during the mating season (from January to June; Izawa et al., 1982) whereas time spent on farm by sexually mature females may depend on nursing kittens. In addition, adult cats spend more time on farm and move at shorter distances from buildings when precipitations are high and temperatures are low (Goszczynski et al., 2009; Horn et al., 2011). All these factors may influence the space use pattern by cats and their probability of using latrines, but their effect has never been tested.

In this study, based on a camera traps survey and a generalized linear mixed model analysis, we tested the effect of feeding area proximity, season and cat sexual maturity on cat latrine use, in order to ultimately understand why latrine location alter the *T. gondii* contamination of soil. We hypothesized that sexually immature individuals preferentially use latrines located near feeding sites, and that sexually mature cats mainly defecate in farm latrines during cold and wet months, with potential differences between males and females. Camera traps were also used to record latrines visits by *T. gondii* intermediate hosts (mammals and birds) to complete the identification of circumstances leading to hotspots of *T. gondii* transmission in dairy farms.

2. Material and methods

2.1. Study sites

The study was conducted on three dairy farms (A, B and C) in a rural area of north-eastern France (49°27'3.49"N, 4°47'0.7"E to 49°27'41.26"N, 5°3'3.35"E; Figure 1). In these farms, cats live in medium to large groups (seven to 24 individuals per group; Table 1). Incidence of *Toxoplasma gondii* in the three cat populations ranged from 0.88 to 0.96 seroconversion/cat/year (Simon et al., 2018) and 50% to 66% of the soil samples previously collected were contaminated with oocysts (Simon et al., 2017). The region has a temperate continental climate, with cold wet winters (average monthly temperature ranges from -1°C to 11°C and average monthly precipitation from 54 to 71 mm between November and March) and mild summers (average monthly temperature ranges from 4.4°C to 23.5°C between April and October; <https://fr.climate-data.org>). Cattle were permanently at pasture from April to October/November and remained inside farm buildings during winter. The three studied farms were situated at a distance of 9.67 km to 19.81 km apart and were located on the periphery of a village surrounded by fields of crops and pastures. As previously described by Simon et al. (2017), the study sites corresponded to the farm buildings plus a 20-m surrounding buffer zone, encompassing the core activity area of the cat population living there (Barrat, 1997; Gotteland et al., 2014b). The resulting delimited study area was 0.88 ha in farm A, 1.14 ha in farm B, and 1.58 ha in farm C (Figure 1).

2.2. Cat population monitoring

The cat population on each farm was intensively monitored during one year. Information on cat abundance, cat births and mortality was recorded monthly by direct observation and from information collected from farm owners. As a part of a capture-mark-recapture procedure (Simon et al., 2018), cats were sexed and photographed to create a portfolio used for individual identification based on coat colour pattern. Cats from the same farm with a similar coat colour were also fitted with plastic collars of different colours to be differentiated, as previously made in another study based on camera-traps (Elizondo and Loss, 2016). The age of cats was also estimated based on direct observations at very young age, information collected from farmers and teeth development (see Courchamp et al., 2000).

2.3. Latrine identification

The three studied farms were systematically searched for cat latrines during visits performed before each session of camera trap deployment (see below). Latrine identification was performed by the same person at each visit to avoid inter-individual detection bias. Following Simon et al. (2017), a latrine was defined as a site with at least two faeces of different ages (indicating reuse over time) within one metre of each other. Latrines may also show conspicuous signs of recent cat activity such as presence of footprints and scarification/scratches and/or ploughed substratum. Two latrine sites were considered as different entities when they were separated by at least five metres.

2.4. Camera trap survey

The latrines were surveyed by 10 to 13 infrared motion-triggered camera traps (an average of 12 per farm) deployed from August 2014 to July 2015 during sessions of 10 consecutive days per farm, rotating from one farm to another. Ten Bushnell Trophy Cam HD-119437 cameras and three Bushnell Trophy Cam HD-119537 cameras were used. Both models feature an 8-megapixel digital camera and an infrared flash that is considered less likely to disturb animals than visible white-light flashes (Glen et al., 2013; Trollet et al., 2014, but see Meek et al., 2016). The movement detection range was 15 m for the ten Bushnell-119437 cameras and 24 m for the three Bushnell-119537 cameras. The trigger speed was set to 1s and the minimal trigger interval to 5s. Depending on the size of a latrine, one to four cameras were deployed to survey the entire surface area used by cats to defecate. The cameras were positioned so as to optimise the detection of cats using the latrine (height and orientation of cameras; Rovero et al., 2010; Glen et al., 2013). However, camera placement was constrained by the availability of appropriate supports on a farm for mounting the camera trap, as this required a stable, solid support unlikely to be removed during the survey session and that did not interfere with the farmer's activities. All the cameras were programmed to record 40s of motion-triggered video footage continuously 24 hours a day. The date and time were displayed for each video capture. The duration of the

after-trigger recording time was determined based on a preliminary study performed in June and July 2014 on farm C. Forty seconds of duration was chosen as a good compromise between the time required to record a complete cat defecation sequence (positioning, elimination of faecal matter and, possibly, covering of faeces) and the time required to watch the video footage in order to analyse it. A total of sixteen 10-day recording sessions (five to six sessions per farm) were performed during the one-year study period, theoretically corresponding to a total of 160 recording days. However, due to organizational and unforeseen material issues (loss of battery power, camera failure, change in the field of view or camera disturbance by cats and farmers, *etc.*), some sessions were shorter or longer than 10 days (see Table 1).

2.5. Recorded information

The recorded video footage was watched and analysed by three readers, each analysing one farm. Doubtful events were discussed among readers to avoid divergent interpretations. The information collected from a farm's video footage was entered in a Microsoft Excel database and included the camera site, date, time, session, farm and information regarding the cat and defecation event (see below). The data in the three databases was then verified and homogenised by one person.

2.5.1. Identification of cat and defecation events

Cat defecation events observed at latrines were recorded in the database. An event was entered as 'defecation' when a cat was observed in a typical defecating position (rounded back with the anus close to the ground, with or without a visible elimination of faecal matter). A defecating cat observed in video footage was individually identified based on the colour pattern of its coat and/or coloured collar. Its individual characteristics (sex and age class) at the time of the video footage were also noted in the database. When cat identification was not possible, due to partial observation or to poor readability of the video, the individual was recorded as 'NI' (Not Identified). In this case, a sex and/or an age-class were assigned insofar as possible (see section 2.6).

2.5.2. *Latrine visits by intermediate hosts*

Intermediate hosts observed in the video footage were categorised in three groups: micromammals (small rodents and shrews), birds and others. A visit from an intermediate host was defined as an observation of one individual of a given species on successive footage within a single 15-min time interval (Kukielka et al., 2013). If several individuals were recorded in the same footage, one visit per individual was recorded. All observations of individuals in the defined latrine area or in its proximity (approximately 5 m around it) were recorded in the database. Visit rates, corresponding to the number of visits per day/latrine/session, were estimated separately for each group of intermediate hosts.

2.6. Definition of variables

For the analysis of latrine use by cats, the explanatory variables were the cat's category (*Cat*) according to its sex and sexual maturity, the season (*Season*) and the location of the latrine in relation to the nearest feeding site (*Distance*). Three cat categories were distinguished: 'sexually immature' (male or female), 'sexually mature male' and 'sexually mature female'. The sexual maturity of females was determined according to age and season: females born in spring and summer (from March to August) were considered mature from the beginning of the breeding period the following year (February), provided they had reached 6 months of age, whereas females born in autumn (after August) were considered mature from 6 months of age (Turner and Bateson, 2013). Based on average values given in published literature, males were considered mature when they reached 10 months of age (Say et al., 1999; Turner and Bateson, 2013). A latrine was considered near a feeding point when it was at a distance of less than 18.5 m, which corresponds to the estimated average maximum radius of cat activity areas around feeding points in the three studied farms during the study period (Simon et al., 2017). A feeding point corresponds to a site where farm owners feed cats daily (generally with milk, food remains or commercially available cat food).

To determine seasonal variation in latrine use, two ecological seasons were determined on the basis of changes in average monthly precipitation and temperature (data obtained from www.meteofrance.com): 1st April to 30th September ('summer') and 1st October to 31st March ('winter'). This season determination is also relevant from an epidemiological point of view, as cats on the three farms had a higher infection risk from October to March than from April to September (Simon et al., 2018).

2.7. Statistical analyses

The intensity of the latrine use by cats was assessed by two indicators: the number of defecations made by cats of a given category per latrine per session, and the total number of cats of a given category that defecated at least one time in a given latrine per session. Both were analysed in relation to the three fixed factors: *Cat*, *Season* and *Distance*. Generalized Linear Mixed Models (GLMMs) with a Poisson distribution and a logarithm link were used as is usual in analysing count data (Zuur et al., 2009). To take into account the duration of each session and the number of available cats in each farm, both the number of surveillance hours per latrine and per session and the total number of individuals per category and per farm (varying among sessions due to births and mortality) were considered as offsets (Table 1). The *Farm* and *Latrine-Session* (i.e. the 10-day survey session of a specific latrine) variables were considered as random effects to take into account the likely dependence of latrines within a given farm and the dependence of cat defecation events within a given latrine. The *Latrine* variable was also considered as a random effect to account for temporal autocorrelations that can result from the repeated survey of a latrine among sessions. These three nested effects were tested using the likelihood ratio test (LRT) on the complete model including all fixed effects and their 2 by 2 interactions.

After selecting random effects, all models were ranked based on Akaike's information criterion for a small sample size corrected for overdispersion (QAICc). The model with the lowest QAICc was considered the best fit to explain the data. A difference of > 2 in the QAICc between two models was used to consider them not equivalent (Burnham and Anderson, 2002). If the

difference in QAICc between two models was < 2 , only the most parsimonious model (*i.e.* the model with the smallest number of parameters) was retained. From the best model retained, Likelihood Ratio Tests (LRT) were performed to assess the significance of variables. Adjusted odds ratios were estimated with 95% confidence intervals for each comparison of classes, and Wald tests were used to determine the significance of differences between classes of each variable. As variance values were often higher than means, we checked whether over-dispersion was still present in the residuals of the selected models by calculating “sum of squared Pearson residuals/degree of freedom” ratios (Zuur et al., 2009). Residuals were visually examined for normality, homogeneity and independence.

All the analyses were performed with R 3.1.3 statistical software (R Core Team, 2014), with a threshold P-value of 0.05. Models were adjusted using the ‘lme4’ package.

3. Results

3.1. Pattern of latrine use by cats

3.1.1. Collected data

The number of cats per farm varied from six to 14 adults and zero to 13 juveniles, depending on farm and session (Table 1). It increased from spring to early autumn, with the succession of litters, but the high mortality of kittens during their first weeks rapidly reduced the population size.

A total of 39 latrines were surveyed during the study (three to nine per session per farm; Table 1). A total of 895 camera-days were recorded on latrines, resulting in 41,282 motion-triggered video recordings. However, of 28% of them were excluded from the database due to camera malfunction (*e.g.* loss of battery power, camera failure) or modifications to the field of view (often due to camera disturbance linked to farming activities). In total, 29,698 videos were analysed, leading to the characterization of 486 defecation events: 99 on farm A, 254 on farm B, and 133 on farm C. The individual identification of cats failed for 20 identification events, but 10 of these cats could be classified in one of the three categories. Defecation occurred in 66 out of the 93 recorded “latrine-sessions” (70.97%), but the frequency of defecation events was highly heterogeneous

among farms, survey sessions and latrines. Depending on the farm, cats were not observed defecating in five to 13 ‘latrine-sessions’. In contrast, 16 latrines (4 on farm A, 8 on farm B, and 4 on farm C) received 10–38 defecations during a given survey session.

3.1.2. Number of defecation events in latrines

During a 10-days survey session, the average number of defecations per latrine per session was 3.96 (standard deviation (sd) = 6.30) on farm A, 10.58 (sd = 13.33) on farm B, and 3.04 (sd = 3.79) on farm C. Only the nested effects of *Latrine* and *Latrine-Session* had significant random effects (Likelihood ratio test: $\chi^2 = 23.16$, df = 1, $P < 0.001$ and $\chi^2 = 349.20$, df = 1, $P < 0.001$ respectively). The most relevant explanatory model of the number of cat defecation events in latrines included the fixed effects of *Cat*, *Season* and *Distance* (Table 2 and Table 3). Interactions between *Cat* and *Season* and between *Cat* and *Distance* were also retained whereas the interaction between *Season* and *Distance* was not significant (Likelihood ratio test: $\chi^2 = 0.30$, df = 1, $P = 0.58$).

As expected, a higher number of defecation events from immature cats was observed in latrines located close to a feeding site than in latrines far from a feeding site (OR = 6.60, 95% CI = [2.87 – 15.25], $P < 0.001$; Figure 2). Sexually mature males also defecated more often in latrines close to a feeding site than in latrines far from a feeding site (OR = 9.42, 95% CI=[3.29 – 26.91], $P < 0.001$). However, seasonal variations were observed in mature males behaviour as the number of their faeces deposited in latrines, whatever the location, was higher in winter than in summer (OR = 2.19, 95% CI = [1.14 – 4.18], $P = 0.018$). Concerning sexually mature females, no difference was observed between their use of latrines close to a feeding site or far from a feeding site in either season (OR = 1.77, 95% CI = [0.80 – 3.94], $P = 0.16$ and OR = 1.04, 95% CI = [0.54 – 2.03], $P = 0.90$ respectively).

In summer, the number of faeces deposited in latrines located close to a feeding site was significantly higher from sexually immature individuals than from mature females (OR = 1.64, 95% CI = [1.20 – 2.25], $P=0.002$) and from mature males (OR = 2.51, 95% CI = [1.81 – 3.48], P

< 0.001; Figure 2). Mature females also defecated more often in these latrines than mature males (OR = 1.53, 95% CI = [1.07 – 2.18], P = 0.019). In latrines located far from a feeding site, mature females defecated more often than mature males and immature cats (OR = 7.92, 95% CI = [3.11 – 20.21], P < 0.001 and OR = 2.26, 95% CI = [1.16 – 4.41], P = 0.016 respectively) and immature cats defecated more often than mature males (OR = 3.52, 95% CI = [1.33 – 9.30], P = 0.011). In winter, no difference in the number of faeces deposited in latrines close to a feeding point was observed between immature cats and mature males or females (OR = 0.88, 95% CI = [0.63 – 1.24], P = 0.48 and OR = 1.32, 95% CI = [0.87 – 1.99], P = 0.18 respectively) neither between mature males and mature females (OR = 1.49, 95% CI = [1.00 – 2.23], P = 0.051; Figure 2). In latrines located far from a feeding site, mature females defecated more often than mature males and immature cats (OR = 3.57, 95% CI = [1.36 – 9.40], P = 0.01 and OR = 2.83, 95% CI = [1.34– 5.97], P = 0.006 respectively) but no difference was observed between immature cats and mature males (OR = 1.26, 95% CI = [0.46–3.43], P = 0.65).

3.3.3. Number of defecating cats

During a video trap session, one to eight cats used the same latrine to defecate. The average number of cats using a latrine was 2.47. The nested effects of *Latrine* and *Latrine-Session* had significant random effects (Likelihood ratio test: $\chi^2 = 8.17$, df = 1, P = 0.004 and $\chi^2 = 7.15$, df = 1, P = 0.007 respectively) whereas the effect of *Farm* was not retained in the model (Likelihood ratio test: $\chi^2 = 0.08$, df = 1, p = 0.77). The most parsimonious model included only the fixed variable *Distance*. The estimated parameters of this model (Table 3) indicated that the latrines located close to a feeding point were used by 2.67-fold more individuals than those located far from a feeding point, regardless of cat category and season (95% CI = [1.66 – 4.30], P < 0.001; Figure 3). The other variables considered in other models for which QAICc < 2 had no significant effects on the number of defecating cats (see Table 2).

3.2. Latrine visits by intermediate hosts

In total, the video footage for all farms revealed 74 visits to latrines from micromammals (17 rats *Rattus sp.*, 9 mice *Muridae*, four shrews *Soricidae*, one vole *Cricetidae*, and 43 unidentified), 1068 from birds (pigeons, corvids and other passerines) and 94 from other species groups (54 red foxes *Vulpes vulpes*, 38 European hedgehogs *Erinaceus europaeus*, one marten *Martes sp.*, one European hare *Lepus europaeus*). Visits from these intermediate hosts occurred in 30 of the 39 detected latrines corresponding to 76.92% of all the latrines: 64.28% on farm A, 84.61% on farm B and 83.33% on farm C. The visit rate of latrines was heterogeneous between species, farms and latrines (Figure 4). Visit rates were higher for birds (0.02 – 874 visits/day per latrine, with an average visit rate of 1.83 ± 2.28) than for micromammals (0.04 – 1.35 visits/day per latrine, average = 0.38 ± 0.44) and other species groups (0.05 – 1.17 visits/day, average = 0.27 ± 0.28).

4. Discussion

Considering that a cat defecates once a day (Liberg, 1982), and assuming that they defecate only in farm latrines, the expected number of defecation events during the survey period according to the number of cats per farm would be 579 on farm A, 1184 on farm B and 965 on farm C. Compared to these predictions, only 17.10% of defecation events on farm A, 21.45% on farm B, and 13.78% on farm C were observed. Several reasons may explain this result: i) some events may not have been captured due to failure of the cameras before the end of a session or have been mistakenly noted as urination rather than defecation as it was sometime difficult to discern the two behaviours on video; ii) some latrines located in areas inaccessible to humans may have been missed despite our efforts to examine all possible locations during the prospection sessions; iii) cats may also defecate outside latrines in their core activity area, as the presence of isolated faeces has been previously reported on farms (Kitts-Morgan et al., 2015; Simon et al., 2017), or outside the core activity area at a distance from the farm. Ishida and Shimizu (1998) and Forin-Wiart et al. (2014) reported in this regard that only 23.5% and 13.5% of faeces excreted by adult cats after ingestion of marked bait were found in the core activity area, suggesting that a significant proportion of the faeces were deposited at a distance away.

In this study, it was observed that latrines located close to a feeding point were used by almost three-fold more individuals than those located far from a feeding point and are used more intensively by immature cats and mature males than mature females. Both the use of those latrines by several cats and the multiple faecal deposits by same individuals could explain the high occurrence of *T. gondii* DNA previously found in soil samples (Simon et al., 2017). Considering that the probability of a cat becoming infected before age of six months may reach 73% on some farms (Simon et al., 2018) and that the *T. gondii* oocysts shedding period lasts up to 20 days (Dubey, 2010), the intense and repeated use of latrines by juveniles certainly contributes to a large part of the soil contamination in latrines located around food resources in dairy farms. The contamination of *T. gondii* in the most used latrines could also be accentuated by the high survival of oocysts in the environment (Lélu et al., 2012).

In contrast to juveniles, the high number of faeces from sexually mature males observed in the latrines close to a feeding site was less expected. In our study, we considered males to reach sexual maturity at 10 months but it has been shown that some males may delay reproduction until around the age of three years (Say et al., 1999) and that non-reproductive males spend more time around the resource centre than reproductive ones (Macdonald et al., 2000). Delay in access to reproduction may contribute to the contamination of latrines close to feeding areas by some males, especially from October to March when their use of latrines increases and when both cat seroconversion rates and oocyst shedding are high (Hermann et al., 2010, Schares et al., 2016; Simon et al., 2018).

The lower number of defecations from mature males in summer than in winter may result from two behavioural factors that decrease the time spent by cats on farms in summer: larger home ranges and movements during dry and warm months than during other months on one hand (Groszynski *et al.*, 2009; Horn *et al.*, 2011) and temporary dispersal of reproductive males from January to June on the other (Bradshaw *et al.*, 2012). On the opposite, in winter, cats spend most time in and around farm buildings (Horn et al., 2011). Furthermore, in this season, they may be

384 more dependent on human-provided food because of the low availability of prey (rodents and
385 birds; Krauze-Gryz et al., 2017). This may explain the absence of significant difference in the use
386 of latrines located close to a feeding site between the three cat categories in winter, whereas
387 juveniles use significantly more often these latrines than mature males and females in summer.
388 Despite mature females can also increase their movements outside farms during summer, their
389 smaller home ranges (ratio of 3.5:1 male:female; Liberg et al., 2000) and the presence of kittens
390 on farms may constrain them to spend time near farm buildings in summer. That may explain why
391 no seasonal effect was found on the use of latrines by mature females.

392 During the study, farm cats were also observed spending time at latrines for other activities than
393 defecating, such as playing, hunting, feeding and resting. It suggests latrines could be a risk for cat
394 infection by accidental ingestion of *T. gondii* oocysts. This may especially be the case for kittens,
395 which spend a lot of time scratching the ground and so can ingest soil or faeces (the ingestion of
396 cat faeces by a kitten was observed once in our video footage). Latrines may also pose a risk of
397 exposure of intermediate domestic and wild hosts (including cat preys) to *T. gondii* oocysts.

398 Intermediate wild hosts were seen on and around 77 % of the latrines during the study period.
399 Visit rates were highly heterogeneous between micromammals, birds and other species.

400 Compared to other species groups, a high number of bird visits was recorded on the three farms.
401 Most of them were small passerines characterised by a gregarious behaviour (del Hoyo et al.,
402 1992), resulting in a high number of individuals (up to 100) visiting a site at the same time. The
403 cameras position, optimized to detect cats movements rather than smaller animal movements, as
404 well as the characteristics of latrine sites, making them more attractive to certain intermediate
405 hosts than to others (for example exposed cereal storage areas that are used as latrines by cats are
406 also an attractive source of food for birds and rodents) may also explain this heterogeneity in the
407 visit rate of between species. The visits of wild intermediate hosts on latrines confirm that farms
408 may be areas where *T. gondii* circulates between wild and domestic environments (Gilot-Fromont
409 et al., 2012).

This study provides evidence that the location of food supplies provided by humans impacts the pattern of latrine use by cats on farms. Latrines close to a feeding area were : i) used by the highest number of individuals, ii) mainly used by juvenile cats, and iii) intensively used by mature males during autumn and winter that are periods of maximal risk of shedding *T. gondii* oocysts. These latrines could thus be high-risk sites for domestic animal and human *T. gondii* infection on farms especially because cat feeding sites are often located near the milking parlour (personal observation) which is daily frequented by livestock and farmers. In addition, cats usually defecate in areas with loose substratum, such as turn soil, gravel, sand or hay (Bradshaw et al., 2012) resulting in latrines location in areas where frequent contacts with animals and humans can occur (e.g. livestock cubicles with loose bedding material like straw, livestock food storage areas, sand pits, wood storage areas with wood chips, etc.). Previous studies showed that the infection of cattle is common in the region under study, and that the prevalence in cattle herd is associated to the presence of cats (Gilot-Fromont et al. 2009). Although cattle infection is not a major animal health issue, the risk of people to be infected from cattle meat may not be neglected (Tenter et al. 2000). Moreover, oocysts are a major source of direct human infection (VanWormer et al. 2013), thus the risk for people working or visiting farms is at concern. Feeding cats at distance from areas frequented by livestock and humans should modulate the *T. gondii* transmission dynamics in farm and reduce the risk of livestock and human infection. Future investigations are needed to better determine how the positioning of feeding points, as well as the management of areas potentially used as latrines, may influence the risk of infection.

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References

- Afonso, E., Germain, E., Poulle, M.-L., Ruetten, S., Devillard, S., Say, L., Villena, I., Aubert, D., Gilot-Fromont, E., 2013. Environmental determinants of spatial and temporal variations in the transmission of *Toxoplasma gondii* in its definitive hosts. *Int. J. Parasitol.* 2, 278–285.
- Afonso, E., Lemoine, M., Poulle, M.-L., Ravat, M.-C., Romand, S., Thulliez, P., Villena, I., Aubert, D., Rabilloud, M., Riche, B., Gilot-Fromont, E., 2008. Spatial distribution of soil contamination by *Toxoplasma gondii* in relation to cat defecation behavior in an urban area. *Int. J. Parasitol.* 38, 1017–1023.
- Baneth, G., Thamsborg S. M., Otranto, D., Guillot, J., Blaga, R., Deplazes, P., Solano-Gallego, L., 2016. Major parasitic zoonoses associated with dogs and cats in Europe. *J. Comp. Path.* 155, S54–S74.
- Barrat, D. G., 1997. Home range size, habitat utilization and movement patterns of suburban and farm cats *Felis catus*. *Ecography* 20, 271–280.
- Bastien, M., Vaniscotte, A., Combes, B., Umhang, G., Germain, E., Gouley, V., Pierlet, A., Quintaine, T., Forin-Wiart, M.-A., Villena, I., Aubert, D., Boue, F., Poulle, M.-L., 2018. High density of fox and cat faeces in kitchen gardens and resulting rodent exposure to *Echinococcus multilocularis* and *Toxoplasma gondii*. *Folia Parasitol.* 65, 002.
- Boyer, K., Hill, D., Mui, E., Wroblewski, K., Karrison, T., Dubey, J. P., Sautter, M., Noble, A. G., Withers, S., Swisher, C., Heydemann, P., Hosten, T., Babiarz, J., Lee, D., Meier, P., McLeod, R., et al., 2011. Unrecognized ingestion of *Toxoplasma gondii* oocysts leads to congenital toxoplasmosis and causes epidemics in North America. *Clin. Infect. Dis.* 53, 1081–1089.

460 Bradshaw, J. W. S., Casey, R. A., Brown, S. L., 2012. The behaviour of the domestic cat. Second
 461 Edition. CABI, 261 pp.

462 Buesching, C. D., Newman, C., Service, K, Macdonald, D. W., Riordan, P., 2016. Latrine
 463 marking patterns of badgers (*Meles meles*) with respect to population density and range
 464 size. *Ecosphere* 7(5): e01328.

465 Burnham, K. P., Anderson, D. R., 2002. Model selection and multimodel inference: a
 466 practical information-theoretic approach. Springer-Verlag New York, Inc. 496 pp.

467 Conrad, P. A., Miller, M. A., Kreuder, C., James, E. R., Mazet, J., Dabritz, H., Jessup, D. A.,
 468 Gulland, F., Grigg, M. E., 2005. Transmission of *Toxoplasma*: Clues from the study of sea
 469 otters as sentinels of *Toxoplasma gondii* flow into the marine environment. *Int. J. Parasitol.* 35,
 470 1155–1168.

471 Courchamp, F., Say, L., Pontier, D., 2000. Transmission of Feline Immunodeficiency Virus in a
 472 population of cats (*Felis catus*). *Wildl. Res.* 27, 603–611.

473 Dabritz, H. A., Conrad, P. A., 2010. Cats and *Toxoplasma*: implications for public health.
 474 *Zoonoses Public Health* 57, 34–52.

475 del Hoyo, J., Elliott, A., Sargatal, J., 1992. Handbook of the birds of the world, vol. 1. Barcelona:
 476 Lynx Edicions. 696 pp

477 Di Cerbo, A. R., Manfredi, M. T., Bregoli, M., Ferro Milone, N., Cova, M., 2008. Wild carnivores
 478 as source of zoonotic helminths in north-eastern Italy. *Helminthologia* 45, 13–19.

479 Dubey, J. P., 2006. Comparative infectivity of oocysts and bradyzoites of *Toxoplasma gondii* for
 480 intermediate (mice) and definitive (cats) hosts. *Vet. Parasitol.* 140, 69–75

481 Dubey, J. P., 2010. Toxoplasmosis of Animals and Man. CRC Press, Boca, Raton. Second edition,
 482 313 pp.

483 Dubey, J. P., Weigel, R. M., Siegel, A. M., Thulliez, P., Kitron, U. D., Mitchell, M. A., Mannelli,
 484 A., Mateus-Pinilla, N. E., Shen, S. K., Kwok, O. C. H., Todd, K. S., 1995. Sources and

reservoirs of *Toxoplasma gondii* infection on 47 swine farms in Illinois. J. Parasitol. 81, 723–729.

Elizondo, E. C., Loss, S. R., 2016. Using trail cameras to estimate free-ranging domestic cat abundance in urban areas. Wildl. Biol. 22, 246–252.

Ferreira, J. P., Leitão, I., Santos-Reis, M., Revilla, E., 2011. Human-related factors regulate the spatial ecology of domestic cats in sensitive areas for conservation. PLoS ONE 6(10), e25970. doi:10.1371/journal.pone.0025970

Forin-Wiart, M.-A., Gotteland, C., Gilot-Fromont, E., Poulle, M.-L., 2014. Assessing the homogeneity of individuals scat detection probability using the bait-marking method on a monitored free-ranging carnivore population. Eur. J. Wild. Res. 60, 665–672.

Gauss, C. B., Almeria, S., Ortuno, A., Garcia, F., Dubey, J. P., 2003. Seroprevalence of *Toxoplasma gondii* antibodies in domestic cats from Barcelona, Spain. J. Parasitol. 89, 1067–1068.

Germain, E., Benhamou, S., Poulle, M.-L., 2008. Spatio-temporal sharing between the European wildcat, the domestic cat and their hybrids. J. Zool. Lond. 276, 195–203.

Gilot-Fromont, E., Aubert, D., Belkilani, S., Hermitte, P., Gibout, R., Geers, R., Villena, I., 2009. Landscape, herd management and within-herd seroprevalence of *Toxoplasma gondii* in beef cattle herds from Champagne-Ardenne, France. Vet. Par. 161, 36–40.

Gilot-Fromont, E., Lélou, M., Dardé, M.-L., Richomme, C., Aubert, D., Afonso, E., Mercier, A., Gotteland, C., Villena, I., 2012. The life cycle of *Toxoplasma gondii* in the natural environment. In: Toxoplasmosis - Recent Advances, O Djurkovic-Djakovic (ed), In Tech, Rijeka, ISBN 980-953-307-298-7, pp. 1–36.

Glen, A. S., Cockburn, S., Nichols, M., Ekanayake, J., Warburton, B., 2013. Optimising camera traps for monitoring small mammals. PLoS ONE 8, e67940. doi:10.1371/journal.pone.0067940

Goszczynski, J., Krauze, D., Gryz, J., 2009. Activity and exploration range of house cats in rural areas of central Poland. Folia Zool. 58, 363–371.

511 Gotteland, C., Gilot-Fromont, E., Aubert, D., Poulle, M.-L., Dupuis, E., Dardé, M.-L., Forin-
 512 Wiart, M.-A., Rabilloud, M., Riche, B., Villena, I., 2014b. Spatial distribution of soil
 513 contamination by *Toxoplasma gondii* in a rural area: influence of cats and land use. *Vet.*
 514 *Parasitol.* 205, 629–637.

515 Gotteland, C., McFerrin, B. M., Zhao, X., Gilot-Fromont, E., Lélou, M., 2014a. Agricultural
 516 landscape and spatial distribution of *Toxoplasma gondii* in rural environment: an agent-based
 517 model. *Int. J. Health. Geogr.* 13–45.

518 Guislain, M.-H., Raoul, F., Poulle, M.-L., Giraudoux, P., 2007. Fox faeces and vole distribution
 519 on a local range: Ecological data in a parasitological perspective for *Echinococcus*
 520 *multilocularis*. *Parasite* 14, 299–308.

521 Herrmann, D. C., Pantchev, N., Vrhovec, M. G., Barutzki, D., Wilking, H., Frohlich, A., Luder,
 522 C. G. K., Conraths, F. J., Schares, G., 2010. Atypical *Toxoplasma gondii* genotypes identified
 523 in oocysts shed by cats in Germany. *Int. J. Parasitol.* 40, 285–292.

524 Hill, D., Dubey, J. P., 2002. *Toxoplasma gondii*: transmission, diagnosis and prevention. *Clin.*
 525 *Microbiol. Infect. Dis.* 8, 634–640.

526 Horn, J. A., Mateus-Pinilla, N., Warner, R. E., Heske, E. J., 2011. Home range, habitat use, and
 527 activity patterns of free-roaming domestic cats. *J. Wildl. Manage.* 75, 1177–1185.

528 Ishida, Y., Shimizu, M., 1998. Influence of social rank on defecating behaviors in feral cats. *J.*
 529 *Ethol.* 16, 15–21.

530 Izawa, M., Dot, T., Ono, Y., 1982. Grouping patterns of feral cats (*Felis catus*) living on a small
 531 island in Japan. *Jap. J. Ecol.* 32, 373–382.

532 Jordan, N. R., Cherry, M. I., Manser, M. B., 2007. Latrine distribution and patterns of use by
 533 wild meerkats: implication for territory and mate defence. *Anim. Behav.* 73, 613–622.

534 Kitts-Morgan, S. E., Caires, K. C., Bohannon, L. A., Parsons, E. I., Hilburn, K. A., 2015. Free-
 535 ranging farm cats: home range size and predation on a livestock unit in northwest Georgia.
 536 *PLoS ONE* 10, e0120513.doi: 10.1371/journal.pone.0120513

537 Knapp, J., Giraudoux, P., Combes, B., Umhang, G., Boué, F., Said-Ali, Z., Aknouche, S., Garcia,
 538 C., Vacheyrou, M., Laboissière, A., Raton, V., Comte, S., Favier, S., Demerson, J.-M., Caillot,
 539 C., Millon, L., Raoul, F., 2018. Rural and urban distribution of wild and domestic carnivore
 540 stools in the context of *Echinococcus multilocularis* environmental exposure. Int. J. Parasitol.
 541 48, 937–946.

542 Krauze-Gryz, D., Żmihorski, M., Gryz, J. 2017. Annual variation in prey composition of domestic
 543 cats in rural and urban environment. Urban Ecosyst. 20(4):945–952.

544 Kukielka, E., Baranosa, J. A., Cowie, C. E., Drewe, J. A., Gortazar, C., Cotarelo, I., Vicente, J.,
 545 2013. Spatial and temporal interactions between livestock and wildlife in South Central Spain
 546 assessed by camera traps. Prev. Vet. Med. 112, 213–221.

547 Lehmann, T., Graham, D. H., Dahl, E., Sreekumar, C., Launer, F., Corn, J. L., Gamble, H. R.,
 548 Dubey, J. P., 2003. Transmission dynamics of *Toxoplasma gondii* on a pig farm. Infect. Genet.
 549 Evol. 3, 135–141.

550 Lélou, M., Villena, I., Dardé, M.-L., Aubert, D., Geers, R., Dupuis, E., Marnef, F., Poulle, M.-
 551 L., Gotteland, C., Dumètre, A., Gilot-Fromont, E., 2012. Quantitative estimation of the
 552 viability of *Toxoplasma gondii* oocysts in soil. Appl. Environ. Microbiol. 78, 5127–5132.

553 Liberg, O., 1982. Correction factors for important prey categories in the diet of domestic cats.
 554 Acta Theriol. 27, 115–122.

555 Liberg, O., Sandell, M., Pontier, D., Natoli, E., 2000. Density, spatial organisation and
 556 reproductive tactics in the domestic cat and other felids. In: Turner DC, Bateson P (eds)
 557 The domestic cat: the biology of its behaviour. 2nd edn. Cambridge University Press,
 558 Cambridge, United-Kingdom, pp 119-147

559 Lindsay, D.S., Dubey, J.P., 2009. Long-term survival of *Toxoplasma gondii* sporulated oocysts
 560 in seawater. J. Parasitol. 95, 1019–1020.

561 Macdonald, D. W., 1980. Patterns of scent marking with urine and faeces amongst carnivore
 562 communities. Symp. Zool. Soc. London 45, 107–139.

563 Macdonald, D. W. 1985. The carnivores: order Carnivora. In: Brown, R., Macdonald, D.W.
 564 (Eds), Social odours in mammals. UK, Clarendon Press, Oxford, pp. 619–722
 565 Macdonald, D. W., Yamaguchi, N., Kerby, G., 2000. Group-living in the domestic cat: its
 566 sociobiology and epidemiology. In: Turner, D.C. & Bateson, P.P.G. (Eds.), The domestic
 567 cat: The biology of its behaviour (2nd ed.).Cambridge University Press, Cambridge, pp.
 568 95–115.
 569 Meek, P., Ballard, G., Fleming, P., Falzon, G., 2016. Are we getting the full picture? Animal
 570 responses to camera traps and implications for predator studies. Ecol. Evol. 6, 3216–3225.
 571 Milkovic, M., Carbajo, A. E., Rubel, D., 2009. Spatial distribution of canine faeces in Buenos
 572 Aires suburbs: implications for public health. Area 41(3), 310–318.
 573 Muñoz-Zanzi, C. A., Fry, P., Lesina, B., Hill, D., 2010. *Toxoplasma gondii* oocyst-specific
 574 antibodies and source of infection. Emerg. Infect. Dis. 16(10), 1591–1593.
 575 Page, L. K, Robert K., Kazacos, K. R., 1998. Raccoon latrine structure and its potential role in
 576 transmission of *Baylisascaris procyonis* to vertebrates. Am. Midl. Nat. 140 (1), 180-185.
 577 Page, L. K., Anchor, C., Luy, E., Kron, S., Larson, G., Madsen, L., Kellner, K., Smyser, T. J.
 578 2009. Backyard raccoon latrines and risk for *Baylisascaris procyonis* transmission to humans.
 579 Emerg. Infect. Dis. 15, 1530– 1531.
 580 Polley, L. 2005. Navigating parasite webs and parasite flow: Emerging and re-emerging
 581 parasitic zoonoses of wildlife origin. Int. J. Parasitol. 35, 1279–1294.
 582 R Core Team, 2014. R: A language and environment for statistical computing. R Foundation for
 583 Statistical Computing, Vienna, Austria. URL <http://www.R-project.org/>
 584 Raoul, F., Hegglin, D., Giraudoux, P., 2015. Trophic ecology, behaviour and host population
 585 dynamics in *Echinococcus multilocularis* transmission. Vet. Parasitol. 213, 162–171.
 586 Richomme, C., Afonso, E., Tolon, V., Ducrot, C., Halos, L., Alliot, A., Perret, C., Thomas, M.,
 587 Boireau, P., Gilot-Fromont, E., 2010. Seroprevalence and factors associated with *Toxoplasma*

588 *gondii* infection in wild boar (*Sus scrofa*) in a Mediterranean island. Epidemiol. Infect. 138,
589 1257–1266.

590 Robertson, I. D., Thompson, R. C., 2002. Enteric parasite zoonoses of domesticated dogs and cats.
591 Microbes Infect. 4, 867–873.

592 Rodgers, T. W., Giacalone, J., Heske, E. J., Pawlikowski, N. C., Schooley, R. L., 2015.
593 Communal latrines act as potentially important communication centers in ocelots *Leopardus*
594 *pardalis*. Mammalian Biology 80, 380–384.

595 Rovero, F., Tobler, M., Sanderson, J., 2010. Camera trapping for inventorying terrestrial
596 vertebrates. Man. Field Rec. Tech. Protoc. Taxa Biodivers. Invent. Monit. Belg. Natl. Focal
597 Point Glob. Taxon. Initiat. 100–128.

598 Say, L., Pontier, D., Natoli, E., 1999. High variation in multiple paternity of domestic cats (*Felis*
599 *catus* L.) in relation to environmental conditions. Proc. R. Soc. Lond. B 266, 2071–2074.

600 Schares, G., Ziller, M., Herrmann, D. C., Globokar, M. V., Pantchev, N., Conraths, F. J., 2016.
601 Seasonality in the proportions of domestic cats shedding *Toxoplasma gondii* or *Hammondia*
602 *hammondi* oocysts is associated with climatic factors. Int. J. Parasitol. 46, 263–273.

603 Simon, J. A., Kurdzielewicz, S., Jeanniot, E., Dupuis, E., Marnef, F., Aubert, D., Villena, I.,
604 Poulle M.-L., 2017. Spatial distribution of soil contaminated with *Toxoplasma gondii* in
605 relation to the distribution and use of domestic cat defecation sites on dairy farms. Int. J.
606 Parasitol. 47, 357–367.

607 Simon, J. A., Pradel, R., Villena, I., Aubert, D., Poulle, M.-L., 2018. A multi-event capture-
608 recapture analysis of seroconversion dynamic to *Toxoplasma gondii* in farm cats. Parasites &
609 Vectors 11: 339.

610 Soler, L., Lucherini, M., Manfredi, C., Ciuccio, M., Casanave, E. B., 2009. Characteristics of
611 defecation sites of the Geoffroy's cat *Leopardus geoffroyi*. Mastozool. Neotrop. 16, 485–
612 489.

613 Tenter, A. M., Heckeroth, A. R., Weiss, L. M., 2000. *Toxoplasma gondii*: from animals to
614 humans. Int. J. Parasitol. 30, 1217–1258.

615 Traversa, D., Frangipane di Regalbono, A., Di Cesare A., La Torre, F., Drake, J., Pietrobelli M.,
616 2014. Environmental contamination by canine geohelminths. Parasites & Vectors 7: 67.

617 Trollet, F., Huyen, M.-C., Vermeulen, C., Hambuckers, A., 2014. Use of camera traps for
618 wildlife studies. A review. Biotechnol. Agron. Société Environ. 18, 446–454.

619 Turner, D.C., Bateson, P., 2013. The domestic cat. The biology of its behaviour. Third Edition,
620 Cambridge Univ. Press, 279 pp.

621 VanWormer, E., Fritz, H., Shapiro, K., Mazet, J. A. K., Conrad, P. A., 2013. Molecules to
622 modeling: *Toxoplasma gondii* oocysts at the human-animal-environment interface. Comp.
623 Immunol. Microbiol. Infect. Dis. 36, 217–231.

624 Weigel, R. M., Dubey, J. P., Siegel, A. M., Kitron, U. D., Mannelli, A., Mitchell, M. A., Mateus-
625 Pinilla, N. E., Thulliez, P., Shen, S. K., Kwok, O. C. H., Todd, K. S., 1995. Risk-factors for
626 transmission of *Toxoplasma gondii* on swine farms in Illinois. J. Parasitol. 81, 736–741.

627 Zuur, A. F., Ieno, E. N, Walker, N. J., Saveliev, A. A., Smith, G. M., 2009. Mixed effects models
628 and extensions in ecology with R. Springer-Verlag New York, 574 pp.

629

Figure 1: Maps of the three studied farms (A, B and C) showing the demarcation of the study area on each farm, the farm layout (buildings and land cover) and the locations of latrines (black boxes) and feeding sites. Sites marked 's' indicate the location of a feeding site in summer; 'w' indicates the location of a feeding site in winter. Absence of 's' or 'w' indicates that the location of the feeding site remained the same throughout the year.

Figure 2: Number of cat defecation events per latrine and per session predicted by the best model. The number of cat defecations varied according to cat category, latrine location relatively to a feeding point (close: ≤ 18.5 m or far: > 18.5 m) and season. MF: sexually mature female, MM: sexually mature male, I: sexually immature cat. Summer: April–September, Winter: October–March.

Figure 3: Number of defecating cats per latrine predicted by the best model. The number of cats varied according to latrine location relative to a feeding point (close: ≤ 18.5 m or far: > 18.5 m).

Figure 4: Average latrine visit rates (number of visits/day/latrine) of micromammals (top), birds (middle) and other species (bottom) by latrine and by farm during the study period. The bars represent standard errors when latrines were visited during several sessions.

Table 1: Experimental design of the camera trap survey, composition of the cat population by session and number of recorded defecation events of individuals classified in one of the three categories: sexually immature cat, sexually mature male, sexually mature female.

Farm	Session	Season	No. days	No. cameras	No. detected latrines	No. mature ♂	No. mature ♀	No. immatures	No. defecations from mature ♂	No. defecations from mature ♀	No. defecations from immatures
A	S04	S	10	10	5	6	3	6	4	2	5
	S08	W	12	12	3	5	3	1	14	3	1
	S11	W	11	12	5	5	3	0	11	5	0
	S15	S	15	13	6	4	3	0	15	17	0
	S20	S	13	13	6	3	3	5	0	3	19
B	S3	S	10	13	4	6	3	13	1	12	22
	S7	W	12	12	5	6	4	12	8	8	29
	S12	W	10	12	4	6	6	12	25	15	25
	S16	S	10	13	4	6	6	11	11	13	30
	S19	S	10	12	7	7	7	9	7	8	34
C	S01	S	9	12	7	5	3	7	1	3	9
	S05	S	12	13	7	5	3	8	3	7	18
	S09	W	10	13	7	5	3	8	4	0	11
	S13	W	13	13	9	5	4	4	12	8	12
	S17	S	14	12	8	8	3	2	15	7	3
	S21	S	10	12	6	7	2	4	5	4	7

S: Summer (April–September), W: Winter (October–March), ♀: female, ♂: male.

Table 2: Model selection for the number of cat defecation events per latrine (a) and for the number of cats defecating in latrines (b). Only the first models showing the lowest Akaike's information criterion value corrected for a small sample size and overdispersion (QAICc) are presented. Values in bold correspond to the most parsimonious model. Δ_i : difference between the QAICc and the lowest QAICc value. Cat: sexually immature cat, sexually mature male or sexually mature female, Distance: location of latrine relative to the nearest feeding point ('close to' or 'far from'), Season: summer (April–September) or winter (October–March).

Model	Residual deviance	No. estimated parameters	QAICc	Δ_i	Model weight
(a) Number of defecations					
<i>Cat+Distance+Season+Cat*Distance+Cat*Season</i>	943.8	11	440.9	0.00	0.49
<i>Cat+Distance+Season+Cat*Distance+Cat*Season+Distance*Season</i>	943.5	12	442.9	2.00	0.17
<i>Cat+Distance+Cat*Distance</i>	964.5	8	443.5	2.65	0.13
<i>Cat+Distance+Season+Cat*Season</i>	962.4	9	444.7	3.86	0.07
<i>Cat+Distance+Season+Cat*Distance</i>	964.3	9	445.6	4.72	0.05
(b) Number of defecating cats					
<i>Distance</i>	549.4	4	299.4	0.00	0.20
<i>Distance+Season+Distance*Season</i>	542.3	6	299.8	0.46	0.16
<i>Distance+Season</i>	546.4	5	299.9	0.54	0.15
<i>Cat+Distance</i>	543.7	6	300.5	1.19	0.11
<i>Cat+Distance+Season+Distance*Season</i>	536.8	8	301.2	1.80	0.08
<i>Cat+Distance+Season</i>	540.9	7	301.2	1.84	0.08
<i>Cat+Distance+Cat*Distance</i>	537.9	8	301.7	2.38	3.06

Table 3: Parameters of the final models retained to explain the number of faeces deposited per latrine (a) and the number of cats defecating in latrines (b). For each variable modality the table indicates the parameter estimate and its standard error (SE). Values in bold correspond to P-values ≤ 0.05 . The modalities of each variable were: for Cat: sexually mature males, sexually mature females or sexually immature cats, for Distance: location of latrine relative to the nearest feeding point ('close to' or 'far from'), for Season: summer (April–September) or winter (October–March). The effects of Cat in the upper part of the table stands for the effect of Cat for the reference category, *i.e.*, individuals far from a feeding site and during summer. Because two interactions were selected (Cat*Distance and Cat*Season), the effect of Cat was not similar according to the distance and to the season. The Cat*Distance parameters give the effect of Cat in individuals living close to a feeding site, and the Cat*Season parameters stand for the effect of Cat in winter.

Variable	Description	Parameter estimate (SE)	P-value of the Wald test	Odds ratio	95% confidence interval
(a) Number of defecations					
<i>Cat</i>	Mature male (reference)				
	Mature female	2.07 (0.48)	<0.001	7.93	3.11–20.24
	Immature	1.26 (0.50)	0.011	3.52	1.33–9.31
<i>Season</i>	Summer (reference)				
	Winter	0.78 (0.34)	0.019	2.17	1.14–4.16
<i>Distance</i>	Far from a feeding site (reference)				
	Close to a feeding site	2.24 (0.54)	<0.001	9.42	3.29–26.93
<i>Cat*Distance</i>	Mature male*Close from a feeding site (reference)				
	Mature female*Close to a feeding site	-1.65 (0.48)	<0.001	0.19	0.07–0.49
	Immature*Close to a feeding site	-0.34 (0.50)	0.489	0.71	0.26–1.89
<i>Cat*Season</i>	Mature male*Winter (reference)				
	Mature female*Winter	-0.81 (0.27)	0.002	0.44	0.26–0.75
	Immature*Winter	-1.03 (0.24)	<0.001	0.36	0.22–0.57

(b) Number of defecating cats

<i>Distance</i>	Far from a feeding point (reference)				
	Close to a feeding point	0.98 (0.24)	< 0.001	2.67	1.66–4.30







