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1 **An optical in-situ tool for visualizing and understanding wetting dynamics in membrane distillation**

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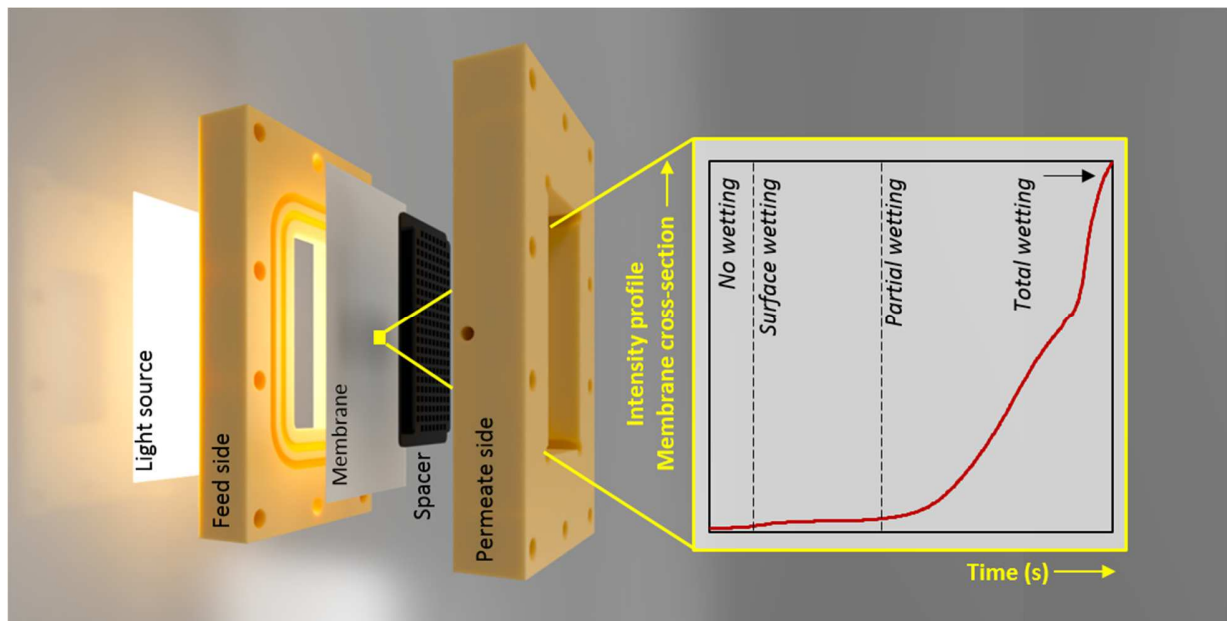
5

6 **Abstract**

7 This paper describes the development of a new optical tool to detect in-situ wetting in membrane
8 distillation. The principle of the detection tool is based on the phenomenon of light transmission. A
9 dedicated experimental setup was developed. The proof of concept of this optical tool is validated using
10 in parallel the Detection of Dissolved Tracer Intrusion (DDTI) method that was previously developed, and
11 which is based on the detection of remaining salt traces in wetted pores by scanning electron microscopy
12 and x-ray dispersion spectroscopy. The in-situ tool was used in standard operating conditions (feed
13 temperature 40°C, laminar flow and vacuum pressure 6 kPa) for vacuum membrane distillation with
14 deionized water, a synthetic saline solution, and seawaters. Controlled wetting was then induced by
15 adding a surfactant in the feed after 30 min of operation and wetting dynamics could be effectively
16 visualized. Additionally, the scalability (macro and mesoscales) of this tool was verified and wetting was
17 visualized at different locations on the membrane surface and compared with the wetting observations
18 made at a global scale. A definition of pore wetting is also proposed.

19 **Keywords:** wetting; membrane distillation; optical tool; in-situ detection

1 Graphical abstract



2

3 Highlights

1. In-situ wetting detection tool developed and tested
2. Link between wetting and light transmission established
3. Proportion of liquid intrusion in membrane thickness evaluated
4. Dynamics of pore wetting can be visualized
5. Wetting identified at different scales in-situ

9

1 **1. Introduction**

2 Wetting is an important issue in membrane distillation. Indeed, the risk of wetting occurrence is today
3 one of the major limitations for the industrial development of membrane distillation in various areas,
4 even if the potentiality of MD is very high for process intensification, during the removals of volatile
5 compounds from aqueous mixtures, brines, over-concentration of RO retentates, and better use of
6 energy by coupling with renewable or lost energy sources [1–5]. A recently published literature review
7 focusing on wetting in MD provides all the information about the state of knowledge about wetting
8 mechanisms, causes and effects [6]. It points out that wetting is a crucial challenge in MD and that
9 further assessments are needed.

10 Membrane distillation exploits the natural or induced hydrophobicity of the polymer used as
11 membranes. As a rule of thumb, porous polymers with low surface energy are selected for MD as they
12 are hydrophobic and ideally do not interact with the feed liquid. However, under adverse conditions, the
13 liquid enters the membrane pores, and the membrane might get “wet”.

14 To be able to make good choices of membranes and operating conditions with the criterion of avoiding
15 wetting, it is indeed necessary to gain knowledge about wetting. To achieve this goal some pertinent
16 tools are required to characterize and to evaluate this phenomenon. It is also very important to define
17 properly and to distinguish the concepts of wettability (risk of wetting occurrence or wetting potential)
18 and wetting (real occurrence of liquid intrusion in pores during MD operation) [7]. Indeed, wetting in
19 membrane distillation is very often understood as the ability of a liquid to maintain contact with a solid
20 surface, resulting from intermolecular interactions when the two are brought together. It is thus more a
21 definition of wettability (or wetting potential) than of wetting itself. Therefore, to better understand and
22 describe this phenomenon in membrane distillation, we have proposed to consider wetting at the pore
23 level and as a localized phenomenon: pore wetting indicates the displacement of the liquid-vapor (L/V)
24 interface inside the membrane pores [7]. Thus, the following definition of pore wetting is considered in
25 this paper:

26 ***“Pore wetting can be defined as the phenomenon where a shift in equilibrium of the liquid/vapor***
27 ***interface promotes liquid intrusion into the hydrophobic pore, i.e., a displacement of the liquid/vapor***
28 ***interface inside the pore, due to the various forces acting upon it, via the interaction of fluid (feed or***
29 ***permeate) and the membrane material under a given operating condition”.***

1 This definition is valid for all membrane distillation configurations.

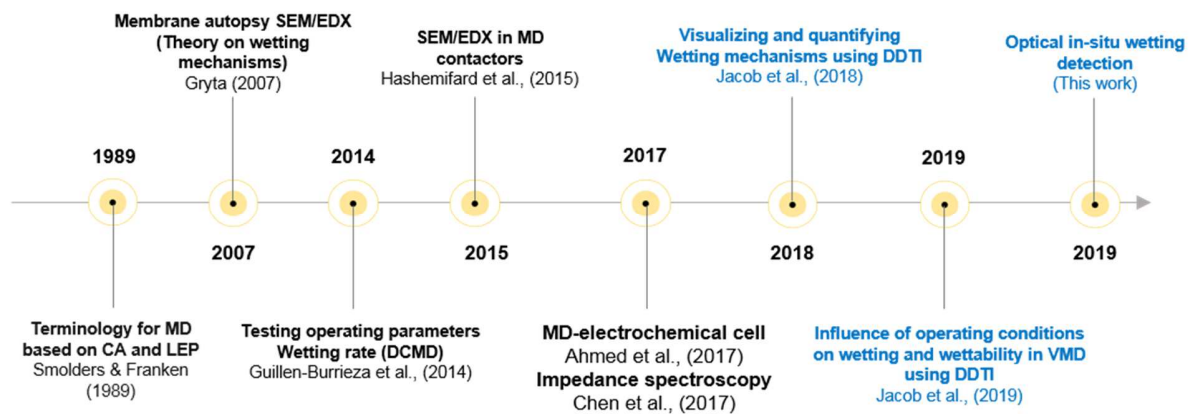
2 Today the assessment of wettability is routinely carried out based on such as Liquid Entry Pressure (LEP)

3 and contact angle (CA), according to the terminology definitions that were given by Smolders et al in

4 1989 [8]. Apart from the fact that these criteria provide information on wettability and not on wetting,

5 these methods are ex-situ measurements from the MD module and require sampling of membranes,

6 that are thus to be destroyed.



7

8 **Figure 1: Literature on wetting and wettability in membrane distillation and membrane contactors**

9 **that lead to the current work [6–13]**

10 In 2007, Gryta developed new approach towards wetting by classifying wetting into various its stage

11 (wetting mechanisms) after observing an autopsy of a wetted membrane by SEM/EDX [9] (Figure 1).

12 Other tools like AFM [14], FTIR [15,16], have also been proposed to characterize wettability and its

13 variation after membrane use, due to fouling or due to membrane aging. All these tools/methods

14 devoted to exploring wettability are *ex-situ*. They can give information on wettability potential on a local

15 scale, however, these tools are unable to give any information on the movement of the liquid-vapor

16 interface leading to wetting. Recently in one of our previous studies, we have proposed a new method

17 called the Detection of Dissolved Tracer Intrusion (DDTI) method [7], that is based on the SEM-EDX

18 detection of traces left by the saline intrusion with the movement of the L/V interface in the membrane

19 pores after the operation. The DDTI method allows to detect wetting mechanisms but it is yet still an *ex-*

20 *situ* and destructive technique.

1 On the other hand, significant efforts have been made to develop and adapt non-invasive techniques for
2 classical pressure based membrane processes however they are mainly devoted towards characterizing
3 fouling or scaling [17]. Most of these techniques are not adaptable to membrane distillation for wetting
4 detection. Presently, measuring conductivity [18,19] in the permeate is the most used in-situ wetting
5 detection technique, but wetting can then only be detected when a significant number of pores are
6 completely wetted, which can be too late. Some other technics that are being considered are volumetric
7 methods [20,21], measurement of electric current using a conductive layer on the membrane
8 compartment [11] or impedance across the membrane [12]. All these methods provide only global
9 information. There is an acute need to develop a technique that can combine the advantages of both in-
10 situ and ex-situ techniques and detect localized wetting on the membrane surface during operation.

11 A comparative assessment of the advantages and disadvantages of the presented methods/techniques
12 for identifying wetting in membrane distillation is presented in **Table 1**.

1 **Table 1: Comparison between in-situ and ex-situ methods for detecting wetting/wettability**

Method	Principle	Advantage	Limitation	Information obtained	Ref.
Contact angle	Surface energy	<ul style="list-style-type: none"> • Simplicity • Gives immediate information about hydrophobicity of the material and surface tension of sample liquid • Extensively studied 	<ul style="list-style-type: none"> • Higher risk/impact of impurities • Results depend on the consistency of the operator • Not representative only relative to the measured location • Characterizes wettability but not wetting itself 	Wettability	[22]
LEP	Provoked intrusion of liquid by filtration	<ul style="list-style-type: none"> • Simplicity • Standardized method • Extensively studied by various authors 	<ul style="list-style-type: none"> • Only gives information on the largest pore • Varies with temperature and feed used 	Wettability: Total liquid intrusion	[23] [24]
Detection of Dissolved Tracer Intrusion method (DTTI)	Operation with saline solution + detection of saline traces with SEM/EDX	<ul style="list-style-type: none"> • Local information on wetting • Wetting mechanisms can be identified • User-friendly operation • Fast results • Applicability in several MD configurations 	<ul style="list-style-type: none"> • Ex-situ • Destructive analysis • Preparation of samples can result in artefacts 	At local scale, all wetting mechanisms can be identified + 2 wetting indicators	[7,13]
Conductivity measurement	Conductivity and mass balance	<ul style="list-style-type: none"> • In-situ • Simple to setup and retrofit into existing pilots • Cheap 	<ul style="list-style-type: none"> • No information on the liquid vapor interface and surface wetting 	Wetting: partial / complete wetting at global scale	[18,19] [23].
Volumetric	Mass balance	<ul style="list-style-type: none"> • In situ • Needs no specialized equipment • Wetting can be studied at operating conditions 	<ul style="list-style-type: none"> • Sensitive • Needs pre-calibrated curve to interpret wetting • Membrane movement inside the cell may affect the reading • Need skilled operator 	Wetting: surface / partial wetting at global scale	[21] [20]

Method	Principle	Advantage	Limitation	Information obtained	Ref.
MD - electrochemical cell	Measuring electric current	<ul style="list-style-type: none"> • In situ • Potential expanded to large scale plants 	<ul style="list-style-type: none"> • Fabricating and maintaining electrically conductive layer • Mass transfer resistance due to the conductive layer 	Wetting: partial pore wetting	[11]
Electrochemical impedance spectroscopy	Impedance	<ul style="list-style-type: none"> • In situ • Potential expanded to large scale plants 	<ul style="list-style-type: none"> • Maybe be only useful in DCMD where 2 interfaces are in contact to the membrane surface 	Wetting: partial pore wetting and total wetting at global scale	[12]
Thermal conductivity detector	Thermal conductivity	<ul style="list-style-type: none"> • In situ • Used in membrane contactors 	<ul style="list-style-type: none"> • Maybe be only useful in SGMD and VMD applications • Emerging method in membrane contactor; proof of concept for MD still to be developed 	Wetting: partial pore wetting	[25–27]

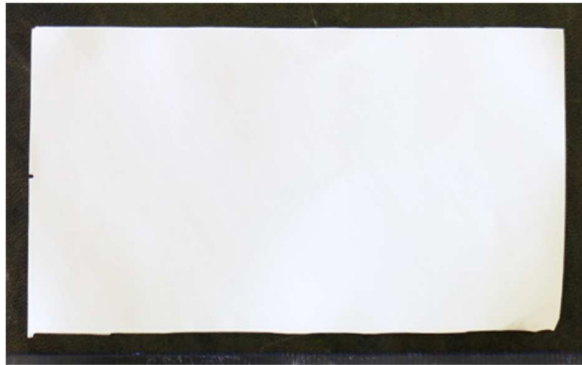
1 The objective of this paper is to develop a completely novel approach aiming to visualize wetting in-situ
2 by developing an optical tool based on light transmission. The principle of this method is based on
3 detection of the modification of the refractive index when the L/V interface penetrates inside the
4 membrane pores. The issue is to develop a piece of equipment that can detect this modification and
5 demonstrate that the obtained signal can be related to wetting occurrence. The primary objective of this
6 study is to provide the proof of concept of light transmission to detect localized in-situ wetting in
7 vacuum membrane distillation. Later, we aim to use this tool to study wetting in different feed solutions
8 for a given commercially available membrane and finally an attempt is made to compare and distinguish
9 wetting dynamics at the global scale and to compare it with wetting dynamics at local scale at different
10 locations on the membrane surface under the same operating conditions.

11 **2. Principle of light transmission in hydrophobic membrane**

12 For any material, when light falls on it, part of this light is absorbed, transmitted, reflected and scattered.
13 The amount of light scattered versus the amount transmitted determines the transparency of the
14 material. A membrane is a porous matrix of fibers with air inside and surrounding it, implying a
15 membrane with 75 % porosity (ϵ) is in fact 25 % material (membrane fibers) and the rest is air. Under
16 ambient conditions (standard temperature and pressure, (STP)) when light falls on a sheet of membrane
17 (Refractive index (N_D) 1.48 [28]) together with air (N_D 1), the majority of the light rays gets scattered or
18 reflected and only a part of the light gets transmitted through the membrane. This results in a
19 membrane which is opaque white due to light scattering. Light scattering depends on a variety of
20 variables (E.g. intrinsic properties, pore size, uniformity, etc.) including the difference in refractive index
21 between the membrane material and the medium (air) inside/surrounding it. Therefore, if the medium
22 (air) inside the membrane can be replaced by another medium (e.g. water N_D 1.33) with a similar
23 refractive index (N_D) as the one of the membrane material, the overall membrane can seem to be
24 translucent and ideally approaching transparent due to this refractive index matching. A described
25 example of this refractive index matching is presented in **Figure 2**. **Figure 2 (a)** visualizes a PVDF
26 membrane, where all the light falling on it is scattered resulting in the membrane to look like an opaque
27 white sheet of plastic similar to a sheet of paper. However, when water was forced into the same PVDF
28 membrane, refractive index matching occurred as the membrane pores were filled with water with a
29 closer refractive index to the membrane one. This results in a membrane translucent with a greater light

1 transmission (**Figure 2 (b)**). The idea is to exploit this phenomenon in membrane distillation to determine
 2 the degree of wetting at a localized scale during operation.

(a) Non-Wetted



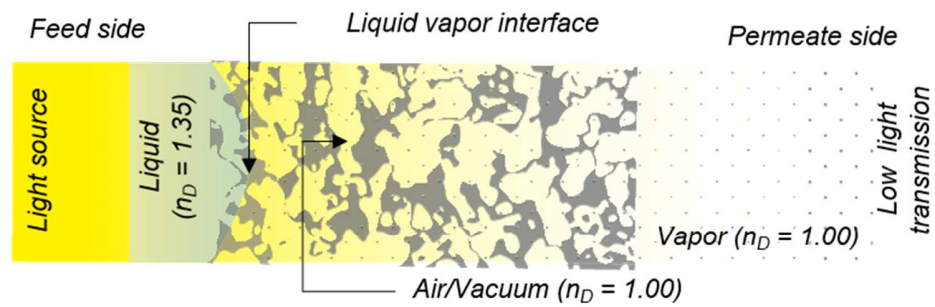
Opaque PVDF membrane

(b) Totally wet

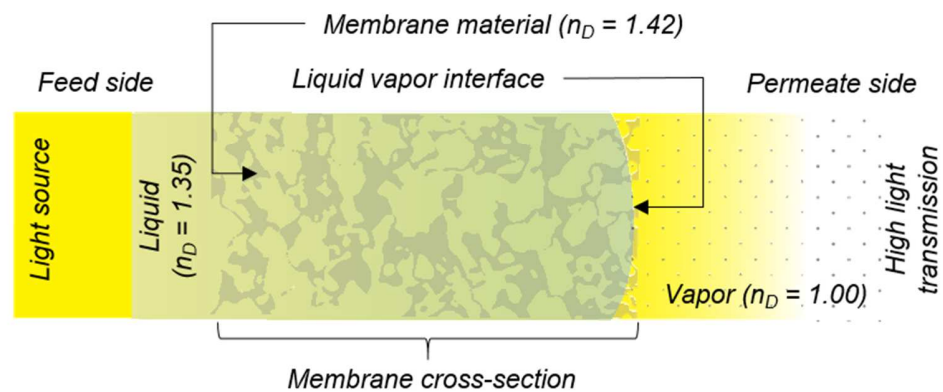


Translucent PVDF membrane

(c)



(d)



3
 4 **Figure 2: Light transmission through the same PVDF membrane (a) under non-wetted conditions (b)**
 5 **under totally wetted conditions using water at macro scale. Graphical representation of light**

1 **transmission through the membrane cross-section at microscale (c) where no wetting had occurred (d)**
2 **where total wetting has occurred (to be viewed in color)**

3 To better understand the theory of light transmission in the membrane pores, a graphical representation
4 of a membrane cross-section subjected to vacuum membrane distillation and observed at the microscale
5 is presented in **Figure 2**. The membrane cross-section was illuminated with a light source from the feed
6 side. Here 2 cases are presented. In the case of **Figure 2 (c)**, the liquid-vapor interface is located on the
7 feed side of the membrane. Here the light transmission from the feed side to the permeate side is low
8 due to the miss matching refractive indexes between liquid, membrane and partial air/vacuum inside the
9 membrane cross-section. However, when the interface moves inside the membrane and total wetting
10 occurs on the membrane cross-section (see **Figure 2 (d)**), light transmission increases due to the
11 matching refractive indexes of liquid and membrane material. Thus, light transmission increases, and
12 light scattering reduces significantly. Indeed, the light that was being initially reflected under no wetting
13 condition is now being transmitted by refraction causing the membrane to be ideally transparent while
14 being totally wet. Therefore, by using a camera/or a light sensor, a change in pixel intensity or light
15 intensity can be identified while keeping all the other parameters constant. Exploitation of this
16 phenomenon will result in identifying the movement of the liquid-vapor interface under a steady-state
17 operation of membrane distillation.

18

19 **3. Material and methods**

20 The first step in this study was to design and manufacture an experimental set-up based on both a
21 dedicated module which allowed both to perform VMD with good control of operating conditions and to
22 that allowed light transmission and on optical detection device.

23 In a second step, it was necessary to define a protocol allowing to detect wetting. For that purpose, it
24 was decided to provoke wetting by adding a surfactant during MD experiments, as surfactants (SDS and
25 Triton X) have been previously reported to induce wetting in membrane distillation [12,29,30]. Our
26 objective is thus to check which is the answer to the optical wetting sensor when the surfactant is
27 injected. It appeared interesting to check at the same time the answer given by wettability indicators and
28 by the DDTI method, for comparison.

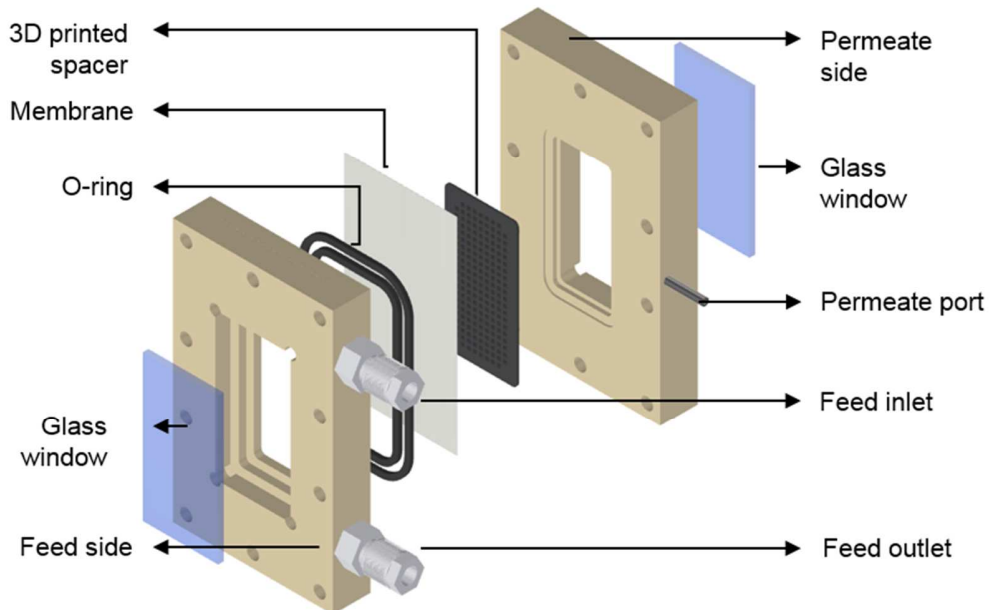
1 **3.1 The experimental setup: module + optical detection device**

2 The overall MD pilot plant was similar to classical vacuum membrane distillation setup described in our
3 previous paper [7,13]

4 **3.1.1 Special module for light transmission**

5 As presented in **Figure 3 (a)**, the module was designed and fabricated to have viewing windows on both
6 the feed and permeate sides of the membrane module to observe the active membrane surface during
7 operation. The rectangular flat sheet (FS) membrane module was made of two 20 mm C-PVC blocks
8 secured by two 10 mm thick stainless-steel plates. The windows are made in borosilicate glass that could
9 withstand high temperatures with 90 % light transmission for 350 – 2000 nm wavelength.

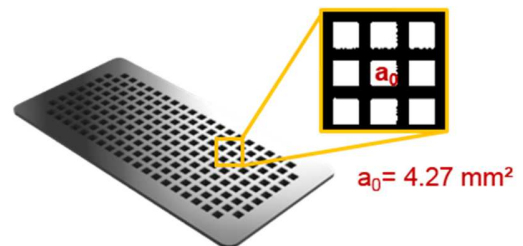
(a) Membrane cell



(b) Feed side internal volume



(c) Permeate spacer



1 **Figure 3: (a) Special module design (b) feed side internal geometry (c) 3-D printed spacer (to be**
2 **viewed in color)**

3 The dimensions of the internal feed side channel were 79 x 41 x 12 mm (length x width x depth) with an
4 edge radius of 7.5 mm. The 3-D printed spacer was only placed on the permeate side of the membrane
5 module to stabilize the membrane during operation. The volume of the internal feed side membrane cell
6 is presented in **Figure 3 (b)**. The feed inlet and outlet ports moved to the sides to have the active
7 membrane surface in view and to reduce the impact of hydrostatic pressure at the feed inlet. The
8 enclosed membrane had an active surface area of $3.24 \times 10^{-3} \text{ m}^2$ and was secured by two O-rings on the
9 feed side.

10 **3.1.2 Membrane and its characterization**

11 A Durapore membrane ($r_{\text{avg}} 0.22 \mu\text{m}$, PVDF) was used in this study. This membrane has been previously
12 characterized by several authors [31,32]. The virgin membrane was $117.2 \pm 0.9 \mu\text{m}$ thick (δ) with an
13 average porosity 0.75 (ϵ). The contact angle (CA) and LEP_w of the virgin membrane were $124 \pm 2.8^\circ$ and
14 $2.04 \pm 0.86 \text{ bar}$ respectively. The Knudsen permeability coefficient (K_M) was experimentally determined
15 in this membrane cell using temperature variation method [31] in laminar flow ($2.75\text{E-}06 \pm 1.1\text{E-}07$
16 $\text{s}\cdot\text{mol}^{1/2}\cdot\text{m}^{-1}\cdot\text{kg}^{-1/2}$) at the reference temperature of 20°C.

17 **3.1.3 Design of the membrane support**

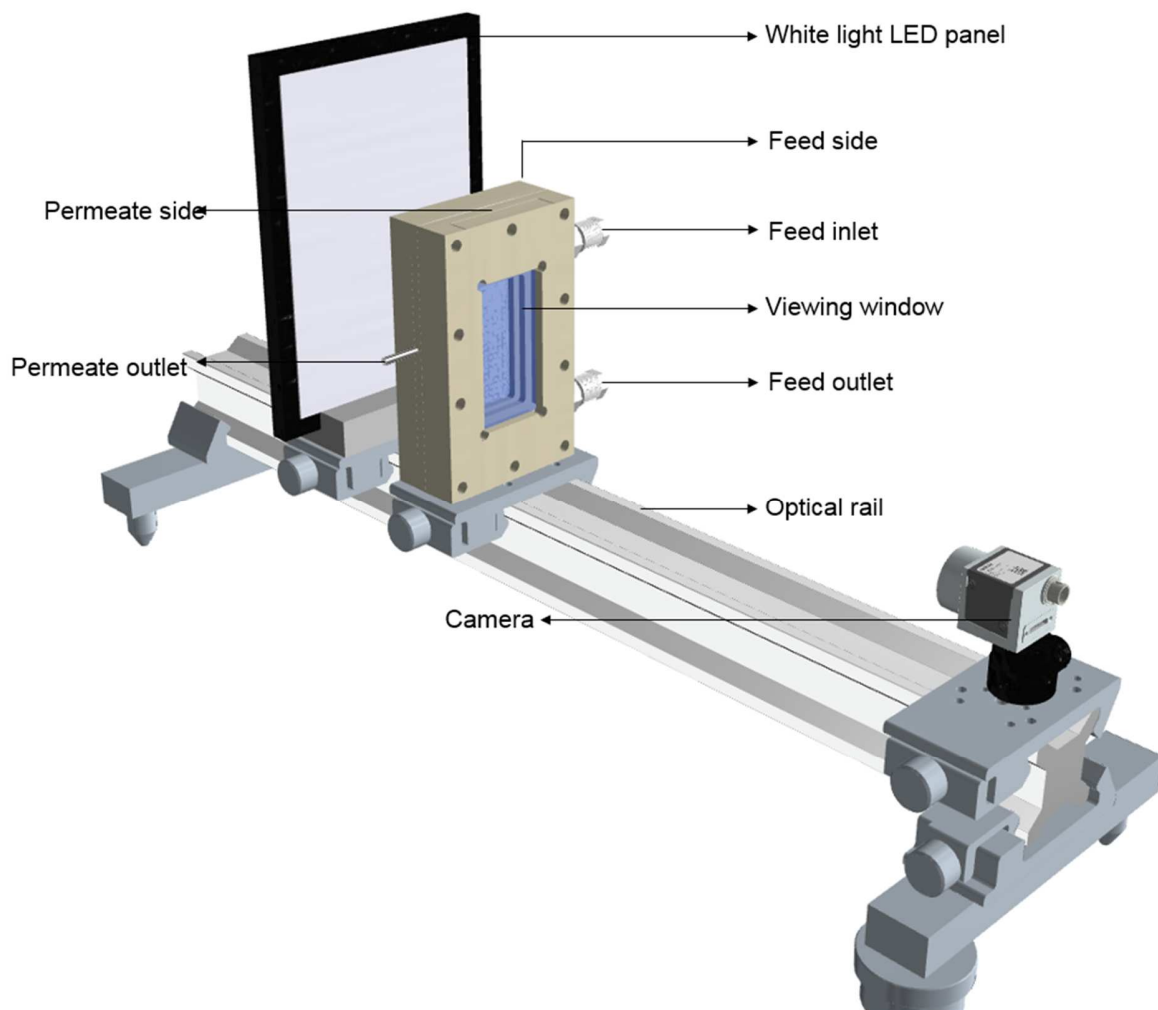
18 The advent and accessibility to manufacture custom parts using 3D printing has led to several
19 researchers developing and using custom support designs for membranes processes like RO[33,34] and
20 MD[35]. A 3-D printed support was introduced on the permeate side to support the membrane during
21 operation. The support had 185 openings with an open area (a_{oi}) of $4.27 \text{ mm}^2/\text{opening}$ (see **Figure 3 (c)**)
22 determined by image processing. This results in the visualization of only $7.94 \times 10^{-4} \text{ m}^2$ or $\sim 27.4 \%$ of the
23 active membrane surface. All wetting visualizations in this study are limited to these 185 openings or
24 $7.94 \times 10^{-4} \text{ m}^2$ or $\sim 27.4 \%$ of the active membrane surface (A).

25 **3.1.4 Optical wetting detection device**

26 The optical system consists of a LED panel, membrane cell, and camera (Basler ace acA1300-200uc with a
27 16mm lens) (see **Figure 4**). The whole setup was fixed and aligned on an optical rail for stability using
28 optomechanical clamps. The white light LED panel (Metaphase Technologies) was used as the light

1 source which emitted a uniform intensity. The feed side of the membrane surface faced the light source
2 while the permeate side faced the camera. The camera was focused and calibrated to view the
3 membrane surface through the spacer. Each image was 1200 x 600 pixels in RGB. The overall system has
4 a resolution of 13 pixels/mm or $\sim 77 \mu\text{m}/\text{pixel}$. The optical setup was designed and manufactured in-
5 house.

6 During operation, the whole system was placed inside the VMD pilot plant with the feed and permeate
7 ports of the membrane module connected to their respective pumps and setups. The camera recorded
8 images each second for the entire duration of the experiment. The viewing window on the permeate
9 side was heated to avoid vapor condensation on its surface. The whole system was covered with a matte
10 black paper and black fabric to avoid external light interference and signal noise.



1 **Figure 4: Optical system for localized wetting visualization (to be viewed in color)**

2 **3.2 Protocol of wetting experiments**

3 Each experiment is performed under standardized conditions and is based on 2 successive steps: MD
4 operations are first performed with a given feed solution for the first 30 minutes and then the feed
5 solution is injected with a surfactant under the same operating conditions, with the objective to provoke
6 wetting. The answers of both the MD system (permeate flux, J) and of the optical system are measured
7 on line in the same time.

8 In this study, laboratory grade Triton X-100 (Sigma Aldrich, France) was used as a wetting inducing agent
9 at 12.5 mg/L for all feed solution samples. The optimal concentration for wetting detection was
10 determined after several tests.

11 The temperature on the feed side temperature (T_f) was fixed at $40\pm 1^\circ\text{C}$ with a feed flow rate (F) $\sim Re \leq$
12 400, while the vacuum pressure was held at 6kPa. The feed solution (cf. 1.1.1) was pre-heated to the
13 desired temperature before circulating on the membrane surface. The vacuum membrane distillation
14 set-up was operated for 1 hour per experiment. For each experiment, the feed solution was circulated in
15 the membrane cell for 30 min. After that, the surfactant was injected into the feed tank to reach a final
16 concentration of 12.5 mg/L. Data for temperature (feed inlet and outlet, permeate), vacuum pressure
17 and conductivities (feed side and permeate flask) were collected and logged during the experimental
18 run. Whereas, images of the membrane surface illuminated from the feed side were taken using a
19 camera on the permeate side at 1 Hz for 1 hour.

20 **1.1.1. 3.2.1 Characterization of feed solution without and with surfactant**

21 4 different feed solutions were studied: de-ionized water (as a control/blank), a synthetic saline solution
22 and two seawaters. The synthetic saline solution was prepared to achieve NaCl concentration (C_f) of 35
23 g/L using crystalline NaCl salt (VWR, France) and deionized water. Seawater were collected at 2 locations
24 in the Mediterranean Sea (Location $42^\circ 50' 48.0''\text{N}$ $2^\circ 56' 33.4''\text{E}$ and $43^\circ 40' 42.1''\text{N}$ $7^\circ 13' 51.4''\text{E}$). The
25 samples were analyzed for conductivity, turbidity, pH, UV absorbance at 254 nm. All analytical
26 measurements were performed at least 3 times and the average value is reported in the following table.

1 **Table 2: Feed solution properties**

Parameters	Unit	Blank	Synthetic Saline solution	Seawater 1 SW1	Seawater 2 SW2
Conductivity	mS/cm	5×10^{-5}	51	43.8	51.1
pH	-	6.8	7.0	7.8	7.9
Turbidity	NTU	0.06	0.01	0.34	0.09
UV Absorbance	-	0	0	0.06	0.012
UV ₂₅₄ nm					
Refractive index	-	1.33241	1.33849	1.33754	1.33900
Surface tension (γ)	mN/m	72.0	73.5	73.9	72.8

Note: All measurements are reported for 25°C

2 Additionally, surface tension and refractive indexes of the feed solution, and the contact angle of the
 3 feed solution on the membrane surface were also analyzed and the protocols are described
 4 subsequently (cf. 3.2.2 and 3.2.3). **Table 2** presents the properties of pure water, saline solution and the
 5 seawaters termed as SW1 and SW2 respectively. Both seawaters were pre-filtered using a 0.45 μ m filter
 6 to remove sand and other large particles that may interfere with the measurements. Considering salinity
 7 (using conductivity measurements), SW1 was less saline than the synthetic saline solution and SW2.

8 Here it should also be noted that SW1 was collected from the inlet of a fish farm in a lake adjoining the
 9 Mediterranean Sea. This resulted in freshwater or surface water intrusion in SW1 that reduced salinity. It
 10 should be noted that SW1 contained higher organics matter as reported by Monnot et. al [36] studying
 11 seawater samples from the same location. This is not the case with SW2 where the salinity (measured by
 12 conductivity) is similar to the saline solution prepared at 35 g/L NaCl in the lab. Whereas considering pH,
 13 water and saline solutions had a pH close to neutral whereas both seawater samples were slightly basic.
 14 The surface tension and refractive indexes of all the three solutions were close to the blank solution.

15 **3.2.2 Surface tension and contact angle of the feed solutions**

16 The surface tension of the feed solution was measured using the pendant drop method and later the
 17 contact angle of these feed solutions with and without surfactants on the membrane surface were also
 18 measured using Drop Shape Analyzer – DSA25 (Kruss) at 25°C. The results were post-processed in

1 accompany software (ADVANCE). Tests were repeated over 10 times with average, and standard
2 deviation reported.

3 **3.2.3 Refractive index**

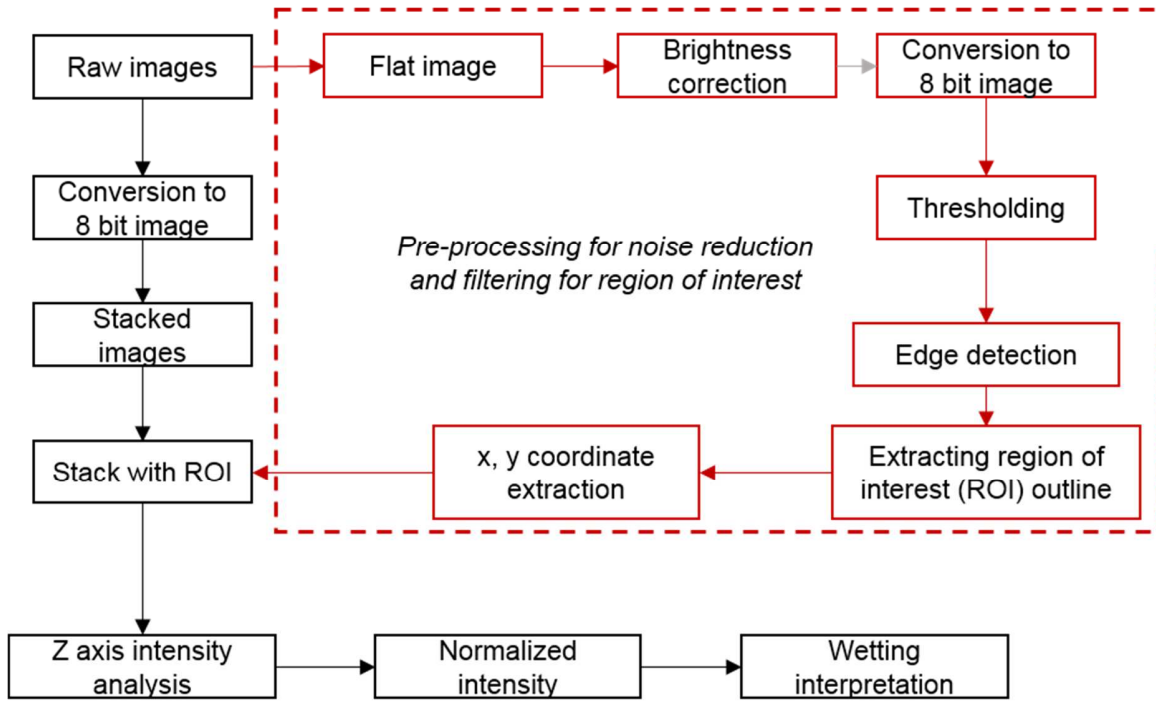
4 The refractive index (n_D) of these solutions were measured at 25°C using a refractometer (RM50
5 LiquiPhysics, Mettler Toledo). The samples were analyzed over three times with average, and standard
6 deviation reported.

7 **3.3 Wettability and wetting indicators**

8 Two classical wettability indicators (Liquid entry pressure (LEP_w) and contact angle (CA)) and one wetting
9 indicator from DDTI method previously developed were used in this study. LEP_w and CA characterization
10 were conducted on virgin membranes using feed solutions before and after adding surfactants. Whereas,
11 pore wetting ratio (ω_p), which is defined as the ratio between the depth of liquid intrusion (detected by a
12 tracer) to the total membrane cross-section [7], was used in validating the proof of concept and for
13 interpreting localized wetting visualization into different wetting mechanisms. The acquired images were
14 post-processed in FIJI[37] with the following algorithm (see **Figure 5**).

15 **3.4 Treatment of the images obtained from the optical device**

16 From each experiment 3600 images of 1200 x 600 pixel were collected which visualized 27.4 % of the
17 active membrane surface ($7.94 \times 10^{-4} \text{ m}^2$) through the spacer and the glass window. The images were
18 taken from the permeate side of the membrane surface with the continuous light source located on the
19 feed side. As an RGB camera is used, the images are converted in 8-bit grayscale.



1

2 **Figure 5: Flowchart for image processing algorithm for in-situ wetting detection**

3 A reference image is used to determine the regions of interest (ROI), which correspond to opening
 4 surface induced by the membrane support (i.e. the lighted parts in the images). This image corresponds
 5 to a steady state operating condition with no wetting. A threshold is applied, followed by a particle
 6 analysis [37], which will generate the ROIs from the reference image. The multi measure method from
 7 the ROI manager allow the area S_i and the position (x_i, y_i) to be determined for each ROI. Each ROI was
 8 labeled from 1-185. An image sequence is then imported corresponding to the experiment on which the
 9 ROIs are used in order to determine the mean light intensity $I_{ROI}(x_i, y_i)$ (mean gray value), in each ROI.
 10 The intensity was finally normalized for interpreting wetting mechanisms. No further data treatments
 11 were conducted as the noise in the acquired data were very low (0.8 – 1.7 gray values).

12 The averaged intensity values for the ROIs were calculated using Eq. 3.1 and presented over time.

$$I_{avg}(t) = \frac{\sum_{i=1}^M S_i \cdot I_{ROI}(x_i, y_i)(t)}{\sum_{i=1}^M S_i} \quad \text{Eq. 3.1}$$

1 Where I_{avg} is the overall average intensity of all region of interests (I_{ROI}) observed over time (t).
2 Whereas, $I_{ROI}(x_i, y_i)$ is the average intensity at one ROI at location x_i and y_i with an area S_i observed over
3 time (t). M is the number of ROI's and is here equal to 185.

4 Additionally, the average intensities were normalized to facilitate the wetting propagation through the
5 membrane cross-section using Eq. 3.2

$$N = \frac{(I_{observed} - I_{initial})}{(I_{max} - I_{initial})} \quad \text{Eq. 3.2}$$

6 Where N is the normalized light intensity, $I_{observed}$ is the intensity of one image at time (t) and I_{max} is
7 the intensity of an image captured when total wetting had occurred and finally, $I_{initial}$ was from the
8 reference image where no wetting had occurred after steady-state operation was achieved.

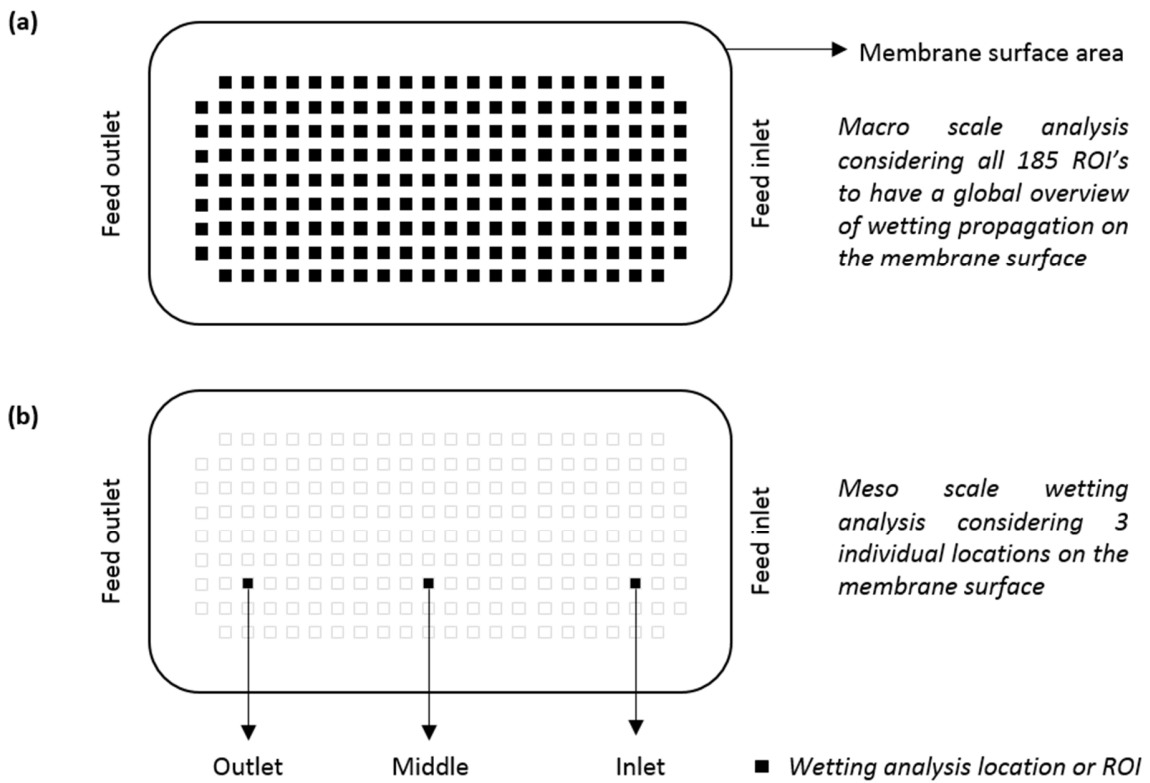
9 Consider an arbitrary ROI with S_i as the area on the membrane surface with the coordinates x_i, y_i at
10 steady state of operation, the liquid vapor interface on the membrane surface and the corresponding
11 light intensity is constant with a low gray value ($I_{initial}$) due to light scattering on the membrane surface.
12 However, when wetting starts to occur the liquid enters the membrane pores in Z axis. As the liquid and
13 the membrane have similar refractive indexes the amount of light transmitted through the membrane is
14 now higher as the relative distance between the light source and the sensor is reduced i.e. at a given x_i, y_i
15 coordinate on the membrane surface the camera is picking up a higher light intensity ($I_{observed}$). Finally,
16 when all the liquid has passed through the membrane there is no more increase in light transmission
17 thereby determining the highest intensity value (I_{max}). Therefore, by using the above treatment and
18 equations change in light intensity at a known location ($I_{ROI}(x_i, y_i)$) over time maybe used to
19 interpreted into movement of the L/V interface inside the membrane cross-section i.e. Z axis.

20 3.5 Wetting visualization at different scales

21 Using the above image treatment, we can choose and analyze wetting dynamics (or propagation) at a
22 global or a local scale as presented in **Figure 6**. The evaluations of these scales are detailed in the
23 following paragraphs.

1 **3.5.1 Macro scale**

2 If the choice is made to evaluate wetting progression considering all the 185 ROI's as seen in **Figure 6 (a)**,
3 wetting analysis using Eq. 3.1 and Eq. 3.2 would be considered at the macro scale or the global scale of
4 the membrane surface. Wetting analysis at this scale would result in an average increase in light intensity
5 of all ROI's in time during the entire duration of the experiment. It should be noted here that the results
6 sections from 4.1 and 4.2 utilizes wetting propagation considering all the ROIs.



7
8 **Figure 6: Wetting analysis with highlighted regions of interest (ROI) (a) at macro scale or global scale**
9 **(b) at meso scale or local scale**

10 **3.5.2 Meso scale**

11 At present, the finest tools that have been developed or used for determining in-situ wetting are at the
12 scale of the membrane. For example, conductivity measurement and recently developed impedance-
13 based technique. The information that can be derived from conductivity measurements is that some
14 pores have been compromised and total wetting has occurred. Wetting is considered at the global scale

1 of the active membrane surface area and not at a local scale. However, wetting may not occur at all
2 locations simultaneously as it is a localized phenomenon. If such a local analysis is deemed necessary, we
3 can choose to apply the same image treatment algorithm and equations on an individual ROI (4.2 mm² or
4 at the mesoscale) and have greater detail on wetting at this scale. Therefore, to illustrate the difference
5 in wetting propagation at different scales, 3 ROIs were selected on the membrane surface where; one is
6 close to the membrane surface close to the inlet, one at the middle of the membrane and one close to
7 the outlet of the membrane surface as illustrated in **Figure 6 (b)**. The difference in wetting propagation
8 at these 3 locations will be compared to the wetting propagation at the macro scale to illustrate the
9 effectiveness of this optical tool better.

10 **4. Results**

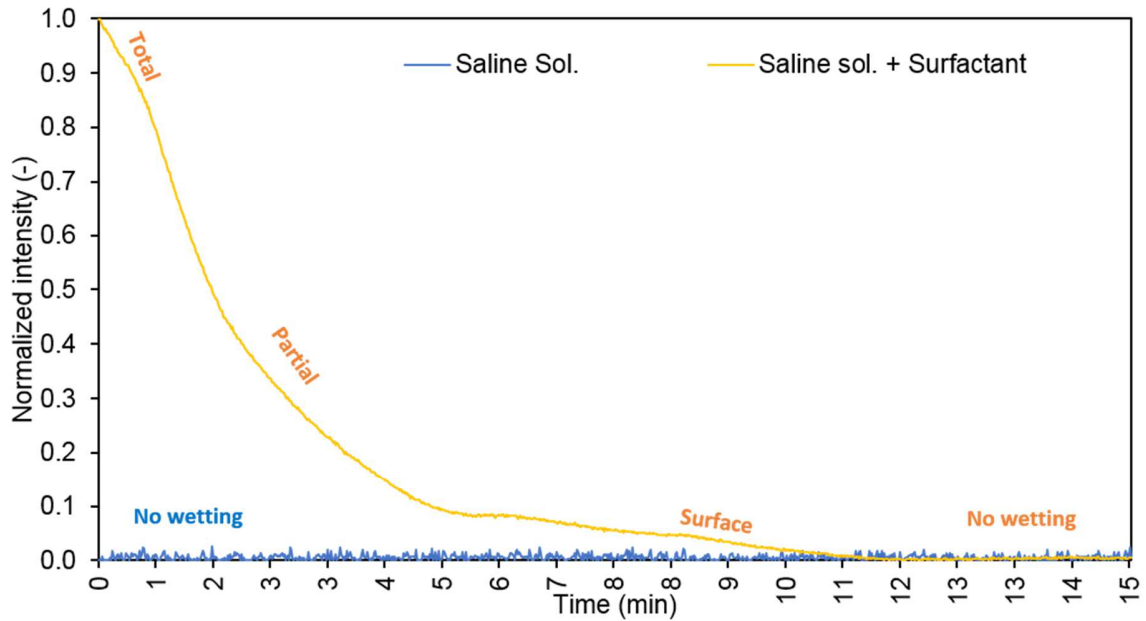
11 **4.1 Proof of concept of the optical in situ method for wetting detection**

12 The proof of concept of wetting detection using light transmission consisted of provoking wetting on a
13 membrane and observed the drying/de-wetting of the membrane pores using the optical device. These
14 observations can later be validated using pore wetting ratio from the DDTI method previously
15 developed. These observations could then be associated with the wetting mechanisms with the idea to
16 observe the wetting kinetics.

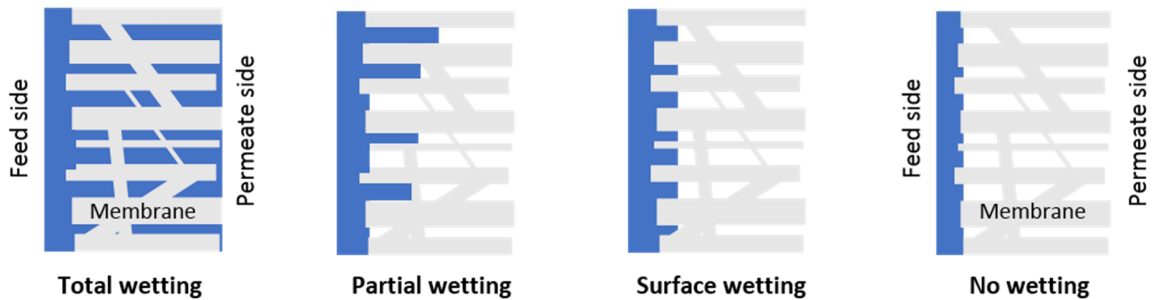
17 Experiments were conducted by operating the VMD pilot with:

- 18 • **Case 1:** A membrane submerged in saline solution (C_f 35 g/L NaCl) with 500 ppm of surfactant for
19 a week to ensure total wetting. Then the membrane was installed inside the membrane module.
20 The change in light intensity was observed through the membrane cross-section during the
21 drying of the membrane for 1 hour. During this time no feed was injected into the membrane
22 cell (as membrane was already wet and would have let water pass through membrane feed side
23 to the permeate side), but a vacuum of 6 kPa was still applied, and all other operating conditions
24 were maintained the same.
- 25 • **Case 2:** A reference membrane operated with the synthetic saline solution (C_f 35 g/L NaCl) only,
26 for which no wetting is expected (according to previous results) and observing the change in light
27 intensity for 1 hour through the membrane cross-section during VMD operation.

(a) Drying wetted membrane at macro scale



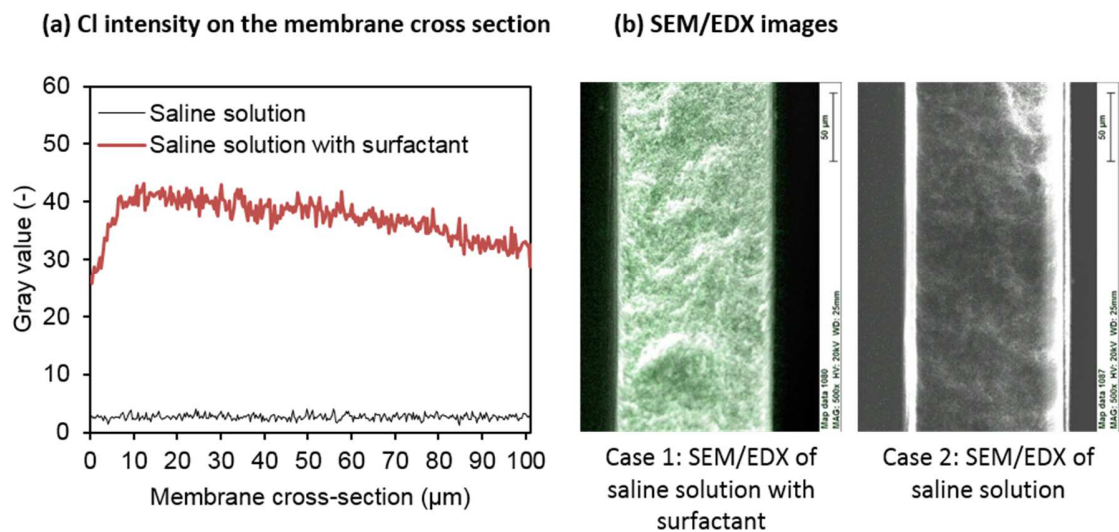
(b) Corresponding wetting mechanisms at pore scale through the membrane cross-section



1
2 **Figure 7: In-situ visualization and graphical representation of wetting mechanism (to be viewed in**
3 **color)**
4 The resulting normalized intensity profiles of these 2 cases are presented in **Figure 7(a)** for 15 min only.
5 Indeed, the membrane subjected to surfactant (case 1) had a higher intensity at the beginning, indicative
6 of total wetting, and as the membrane progressively dried with the water evaporation inside the
7 membrane pores, total wetting probably transitioned to partial wetting, and then to surface wetting .
8 We can suggest that the boundary of transitions between the different wetting mechanisms could
9 correspond to the changes in the curve slope but this assumption needs to be further investigated.

1 Finally, when all the water had evaporated, and the membrane dried entirely, the intensity reduced to its
2 lowest value indicative of no wetting. At the same time for case 2 where no surfactant was introduced,
3 no wetting was observed during the total experimental duration. These observations are at macroscale
4 but are representative of the transiting wetting mechanisms observed at the pore scale as illustrated in
5 **Figure 7 (b)** (for case 1).

6 To validate the above hypothesis, scanning electron microscopy with X-ray dispersion spectroscopy were
7 conducted on samples acquired from Case 1 and Case 2. Detailed protocols of sample preparation and
8 SEM/EDX analyses can be found elsewhere [7]. An average intensity profiles for Chlorine across the
9 membranes subjected to both cases are presented in **Figure 8(a)** whereas **Figure 8(b)** presents the
10 SEM/EDX micrographs of these membranes. Indeed, high Chlorine traces across the membrane sample
11 subjected to surfactant (case 1) allow validating that total wetting was prevalent in this sample. Whereas
12 for the membrane subjected to the saline solution only (Case 2) no traces of Chlorine on the membrane
13 cross section was found, indicating no wetting had occurred.



14
15 **Figure 8: (a) Chlorine intensity on the membrane cross-section (b) SEM/EDX micrographs for case 1**
16 **and case 2 (to be viewed in color)**

17 These preliminary results allow validating the proof of concept that an increase in light intensity at
18 steady state operation can be linked to a wetting propagation inside the membranes. In the following

1 section, the membrane was subjected to various feed solutions under steady-state operation, to better
 2 illustrate and understand wetting mechanisms.

3 **4.2 Influence of surfactant on feed properties, wettability parameters and MD permeate flux**

4 **Table 3** presents the properties of the different solutions used in this study before and after addition of
 5 the surfactant in the feed solution where deionized water was used as a control/blank. All
 6 measurements were conducted at 25°C and the seawaters were pre-filtered with a 0.45 µm membrane
 7 before characterization.

8 **Table 3: Feed properties and wettability parameters before and after surfactant injection**

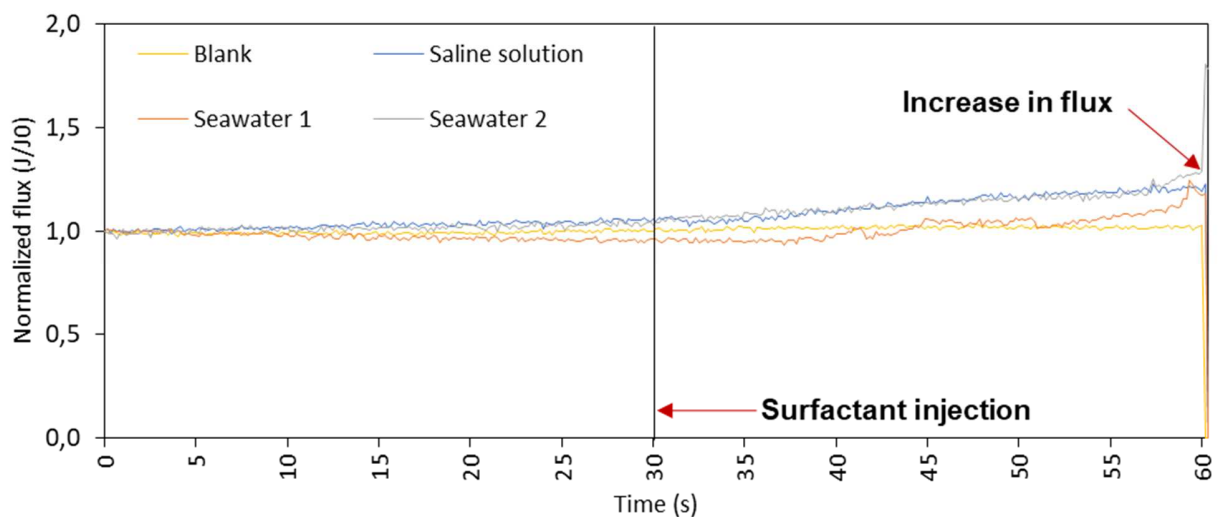
Parameters	Unit	Blank	Without surfactant			With surfactant		
			Saline	Seawater	Seawater	Saline	Seawater	Seawater
			solution	1	2	solution	1	2
Conductivity	mS/cm	5 x 10 ⁻⁵	51	43.8	51.1	51.5	41.1	49.5
pH	-	6.8	7	7.8	7.9	7.2	7.9	8.0
Turbidity	NTU	0.06	0.01	0.34	0.09	0.30	0.42	1.16
Absorbance UV254nm	-	0.000	0.000	0.060	0.012	0.049	0.093	0.048
Refractive index	-	1.33241	1.33849	1.33754	1.33900	1.33851	1.33719	1.33830
Surface tension	mN/m	72.0	73.5	73.9	72.8	62.3	64.1	57.1
Contact angle	°	123.7	125.9	125.3	123.7	122.1	123.6	119.3
LEP	bar	2.03	1.97	1.43	1.50	1.73	1.33	0.87

9 *Note: All measurements are reported for 25°C*

10 Considering the different feed properties, an addition of the surfactant generally increased the pH to
 11 slightly basic for all samples even though the turbidities of the samples were still clear. Similarly, it could
 12 be seen that all samples with an addition of surfactant had an overall increase in refractive index
 13 compared to the blank solution. However, after addition of the surfactant, there was a slight reduction in
 14 the refractive indexes in the samples. The principal changes in the feed that can be high-lighted is a 15 –

1 22 % reduction in the surface tension of the feed solution after surfactant addition, implying the wetting
2 potential of the feed solutions had increased.

3 Concerning the wettability parameters of the feed solutions with the studied membranes, both contact
4 angle and liquid entry pressure were measured. Only a 2-4 % reduction in contact angle was observed,
5 however, LEP presented a 7 to 42 % reduction. It can be seen for LEP that wetting potential of the feed
6 solution after surfactant addition had significantly increased. It worthy to mention that these
7 measurements were taken at standard temperature and pressures. Elevated temperature feed solution
8 with surfactant would have a significant reduction in surface tension thereby increasing the risk of
9 wetting under the operating conditions [29].



10

11 **Figure 9: Flux response for the feed solutions (to be seen in color)**

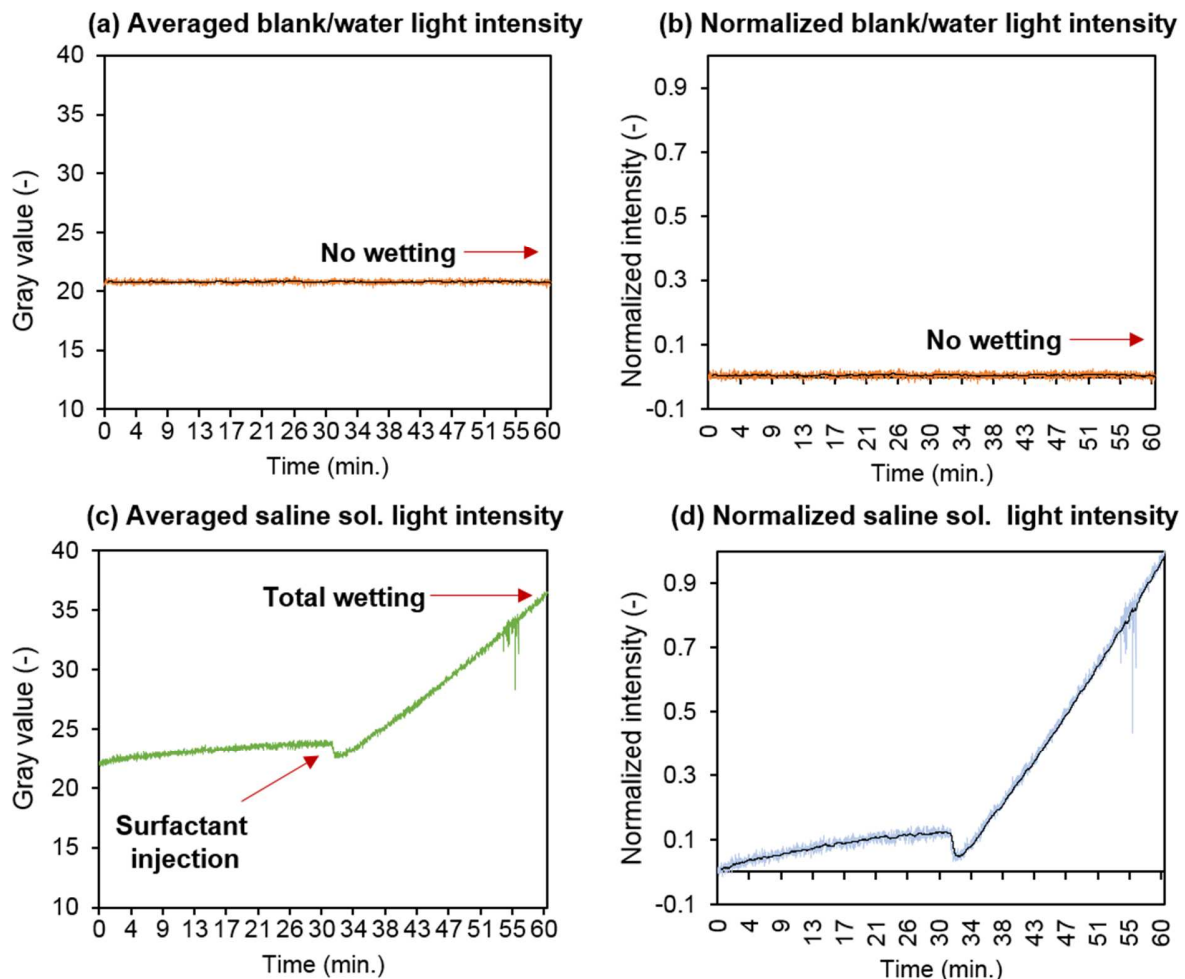
12 **Figure 9** summarizes the normalized flux response in the studied samples under fixed operating
13 conditions. Depending on the samples the permeate flux varied between 2.5 and 3.5 Kg/m².h and
14 generally could be considered stable during the first 30 minutes of operation. However, after addition of
15 the surfactant, a linear trend in flux is observed. This may be attributed to the inward movement of the
16 liquid-vapor interface thereby decreasing the vapor diffusion path and temporarily increasing the flux.
17 Similar observation of an increase in flux leading to wetting, later on, was also observed previously [18].
18 After 55 min of operation, the vapor flux started to be unstable, and feed solution could be seen to pass

1 through the membrane in its liquid state confirming total wetting had occurred at some location or
2 locations on the membrane surface during operation.

3

4 4.3 Visualizing induced wetting with different feed solutions

5 In this section, the different previously described feed solutions (cf. 3.5) were used to visualize wetting
6 progression under same steady-state operating conditions for 1 hour while using deionized water as
7 blank. Wetting was provoked after 30 min by injecting a surfactant (Triton x) into the feed solution to
8 reach a final concentration of 12.5 ppm.



9

1 **Figure 10: Averaged and normalized light intensities during experimental run (a, b) blank/water and (c,**
2 **d) saline solution with surfactant injection after 30 min**

3 **4.3.1 Blank and saline solution**

4 **Figure 10** presents the light transmission with (a) water (blank) and (c) saline solutions for the entire
5 duration of the experiment. In the case of blank, the intensity did not change during the entire
6 operation, and this state could attribute to the steady operating conditions where no wetting occurs. As
7 the feed contained neither salts nor the surfactant; the feed conductivity stayed stable $3.37 \pm 0.07 \mu\text{s/cm}$.
8 The normalized graph on the right side (**Figure 10 (b)**) also presents the same information. During the
9 total duration of the experiment, no feed liquid had appeared on the permeate side of the membrane
10 surface.

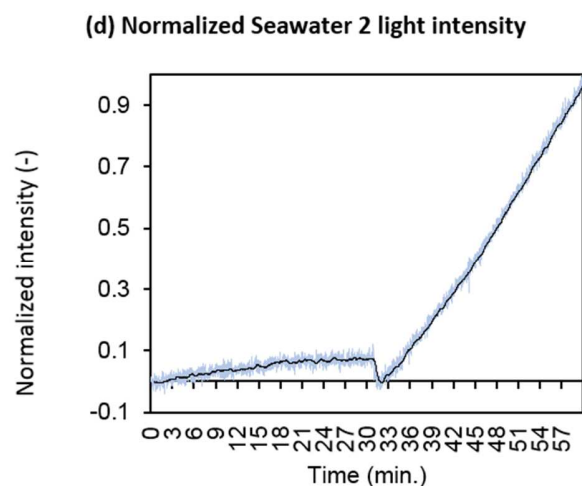
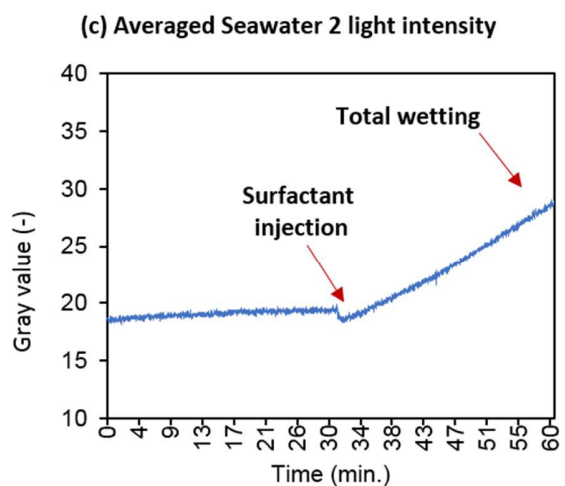
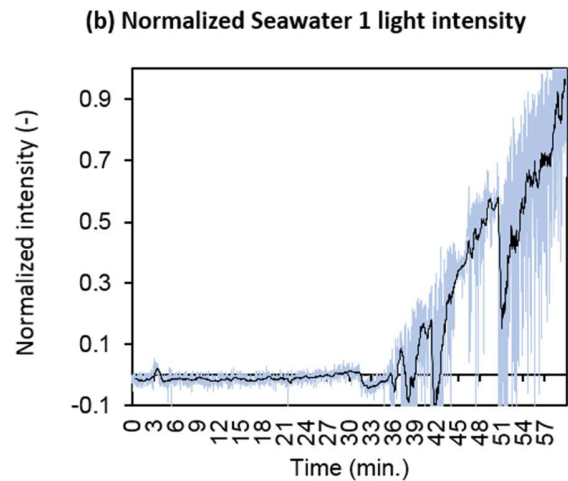
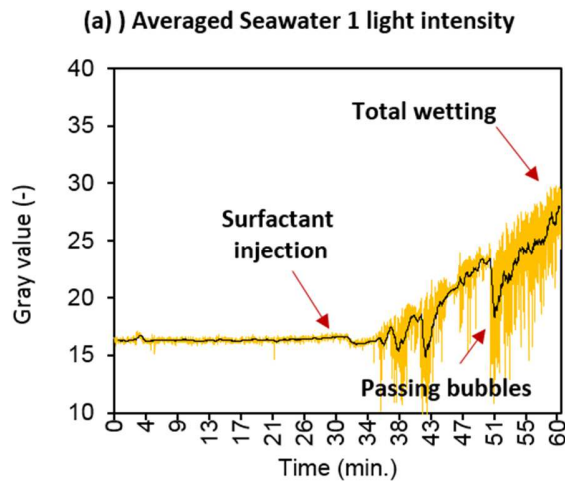
11 However, considering the saline solution (see **Figure 10 (c)**), the light transmission slightly increased as
12 the experiment continued. This increase in light intensity can be suggested as the inward movement of
13 the L/V interface. However, just after the addition of the surfactant at the $\frac{1}{2}$ hour mark, a sudden drop in
14 light intensity can be observed with a significant increase in light transmission soon afterwards. By the
15 end of the experiment, liquid water appeared on the permeate side of the membrane module validating
16 total wetting had occurred on the membrane surface. Considering the normalized data (**Figure 10 (d)**),
17 surfactant addition into feed solution introduces these molecules at the membrane's liquid vapor
18 interface resulting in a rapid reduction in light intensity. The surfactant interacts with the membrane
19 surface by reducing its hydrophobicity, as well as a reduced membrane contact angle, leading to wetting.
20 Details of progressive nature of surfactant induced wetting can be found elsewhere [29].

21 **4.3.2 Seawaters**

22 As the principal application of membrane distillation is for desalination, the application of wetting
23 detection using the optical technique was also tested for real seawaters. Considering light intensities
24 (see **Figure 11**), it can be noted that both samples had lower light transmission than observed for either
25 blank or saline solution. This may be due to additional solutes dissolved in seawater. For the first $\frac{1}{2}$ hour
26 SW1 performed more like the blank with no increase in wetting intensities. However, after the surfactant
27 injection, the intensity dropped like for saline solution but to a lesser extent and then it started to
28 increase. Here the signal was noisy so a running average filter ($n=30$) was introduced to better visualize
29 the trend in data. The significant noise in the signal was introduced by the surfactant interacting with this

1 seawater sample and generating small bubbles in the circulating feed samples. As these bubbles passed
2 in front of the visualized membrane surface, they created momentary dark regions resulting in the noisy
3 signal. However, a general trend in the increase in light signal could be still observed.

4 Considering SW2, this sample had a similar intensity profile as the saline solution. Here it can be noted
5 that there was a slight increase in intensity before the surfactant injection, implying an inward
6 movement in the L/V interface. However, after the surfactant injection, wetting progressed in the same
7 manner as the saline solution. As the surfactant accumulated on the membrane surface there again was
8 a reduction in the intensity and later wetting propagation occurred until a state of total wetting was
9 observed. Total wetting was confirmed by the accumulation of the liquid feed in the permeate side of
10 the membrane.

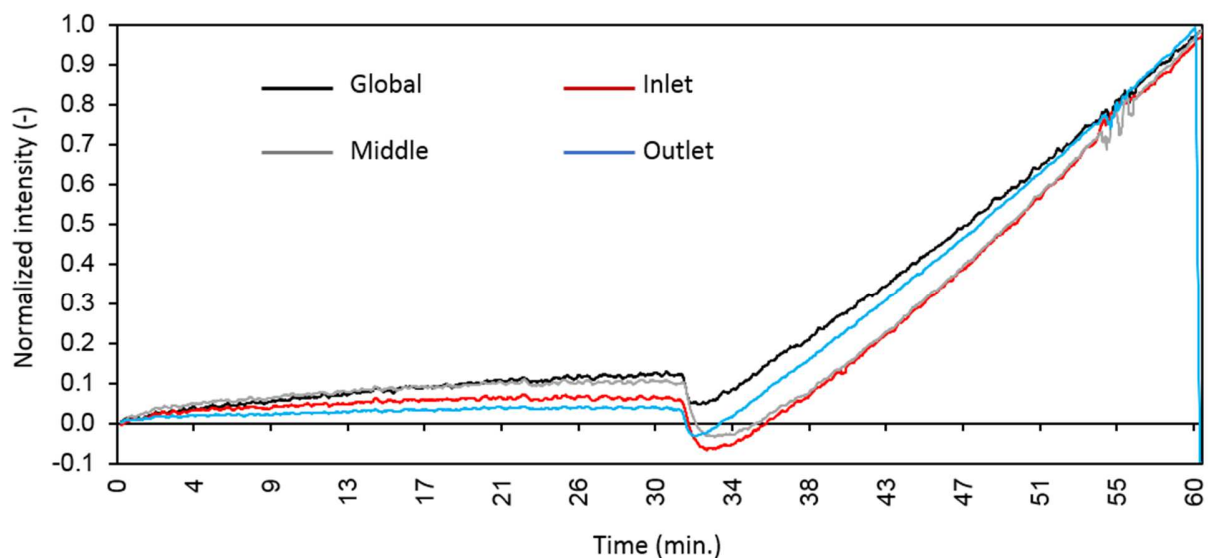


11

1 **Figure 11: Averaged and normalized light intensities during experimental run (a and b) seawater 1 (c**
2 **and d) seawater 2 with surfactant injection after 30 min**

3 **4.4 Difference in wetting propagation at macro and meso scale at different locations**

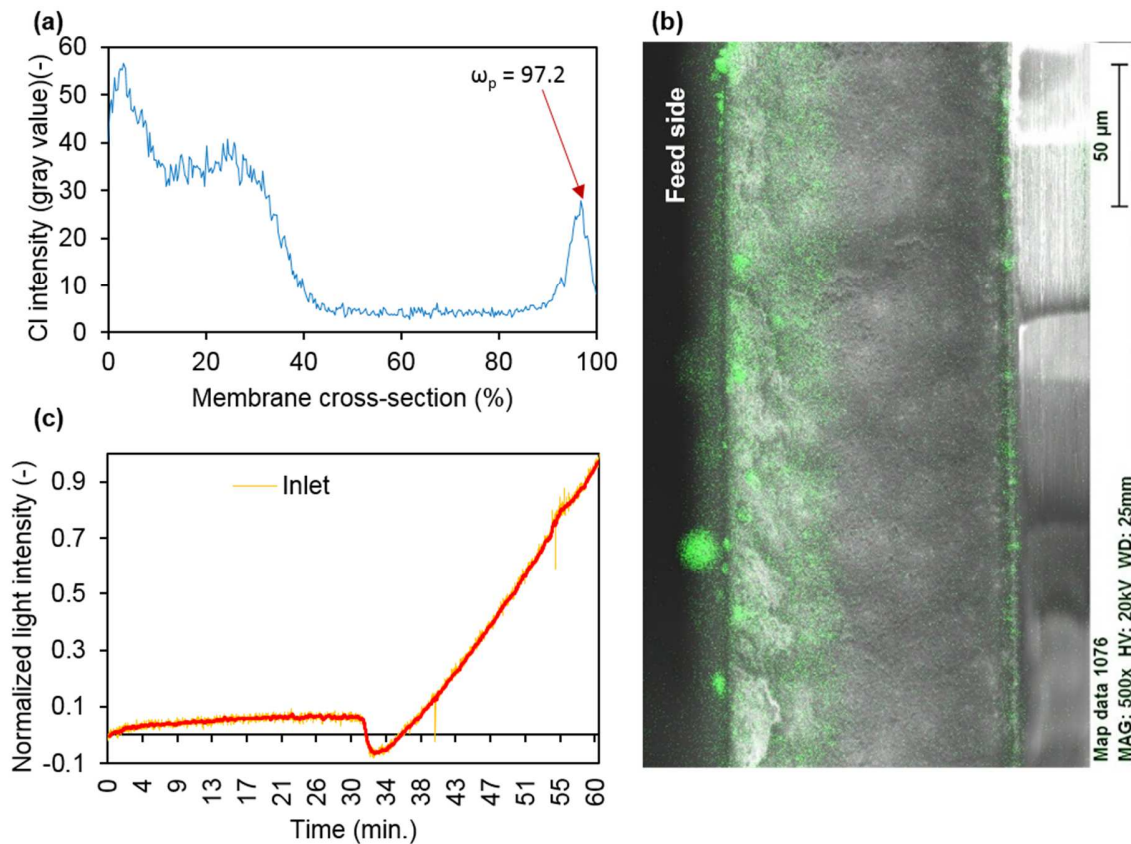
4 Using this optical tool, we can further differentiate wetting occurrences at a more local scale for example
5 at meso scale (0.01 to 30 mm) and compare it with the scale of the membrane. **Figure 12** presents the 4
6 curves of normalized intensity for the membrane surface at the inlet, middle, outlet and the global
7 intensity for the membrane subjected to the saline solution with the surfactant added at 30 min. The
8 data presented in **Figure 12** is a moving average filter (n=15) to reduce signal noise. Here it can be
9 observed that the global curve match closely to the data sampled at the middle until the surfactant is
10 added. However, wetting propagation at the feed outlet surpasses wetting propagation observed at the
11 inlet and middle of the membrane after surfactant addition. This may be due to the membrane's internal
12 morphology being different at each location or the fluid flow in the membrane feed chamber creating
13 slightly higher localized hydrostatic pressure resulting in faster wetting on the feed outlet. Wetting
14 induction at the feed outlet was faster than that for the inlet and middle even though the overall curves
15 mimic the same linear trend as the global scale. This difference in normalized light intensity is indicative
16 of the change in wetting rate experienced by the same membrane at different locations under the same
17 operating conditions. This level of detail in-situ localized wetting progression was not possible with the
18 current state of the art.



19

1 **Figure 12: Comparing wetting progression on the membrane surface at global and local scales: inlet,**
2 **center and outlet – (To be seen in color)**

3 Additionally, after the experiment, the membrane was sampled at the inlet location, and Cl⁻ was profiled
4 by SEM-EDX across the membrane cross-section using pore wetting ratio (ω_p) as presented in **Figure 13**
5 (a and b). Here the progressive change in light intensity can be interpreted by the Cl traces left by the
6 feed solution. The chloride deposits on the feed side membrane cross-section are indicative of an inward
7 movement of the L/V interface leading to surface wetting. The same surface wetting mechanism can also
8 be seen with a slight increase in light intensity from 0- 29 min (see **Figure 13(c)**). However, after
9 surfactant injection, an increase in light intensity could be observed as the liquid vapor interface moved
10 inside the membrane cross-section. At the end of the experiment, the highest intensity (indicated by
11 Normalized data = 1) was observed, indicative of a total wetting and this can be confirmed by the Cl
12 deposits on the permeate side of the membrane cross-section ((see **Figure 13(b)**) Pore wetting ratio also
13 confirms that the total wetted state of the membrane as Cl⁻ peaks can be seen at ω_p of 97.2 % of the
14 membrane cross-section.



1
 2 **Figure 13: (a) Chlorine Intensity profile of membrane cross-section (b) SEM/EDX micrograph of the**
 3 **membrane cross-section (c) Normalized in-situ light transmission of at the the same location with**
 4 **saline solution (to be seen in color)**

5 Wetting progression at the inlet (meso scale) can be validated by the SEM/EDX micrographs at the pore
 6 scale. Similar observations can be made all over the membrane surface and used to identify the
 7 propagation of wetting mechanisms at different locations. This would enable us to better understand the
 8 influence of feed solutions, operating conditions, and process hydrodynamics at a local scale during
 9 operation.

10 The scale and resolution of the wetting observation will depend on the resolution of the camera/sensor
 11 setup and area of the membrane visualized. Wetting may not be visualized where the membrane surface
 12 is covered for example with the spacer. Wetting propagation may be evaluated on the membrane by
 13 precisely determining and correlating SEM/EDX and light at different stages of wetting.

1 **5. Conclusions and perspectives**

2 In this study, a non-invasive in-situ optical tool was developed and tested for visualizing wetting
3 propagation in membrane distillation. The proof of concept that in-situ wetting can be detected using
4 light transmission was provided and validated using DDTI method. The tool was tested with saline
5 solutions and seawaters with controlled wetting induced by the surfactant. The optical tool was able to
6 provide information on wetting mechanisms in the studied feed solutions. The developed tool is
7 adaptable and can visualize wetting at both global and local scales. Earlier this level of scalability could
8 be only possible using ex-situ techniques like SEM/EDX.

9 The developed tool and pore wetting definition is simple and intuitive. The tool can be easily adapted to
10 all membrane distillation configurations for example; DCMD, SGMD, AGMD, and VMD with suitable
11 adaptation the membrane module. With this optical tool, the effects of operating parameters
12 (temperatures, flow rates, local hydrodynamic conditions, feed salinities, time and membrane, etc.) on
13 in-situ wetting can be evaluated. Additional to wetting detection techniques, there is a growing interest
14 in membrane regeneration after wetting. In this study, the proof of concept was based on the
15 observation of de-wetting with the optical method, which also appears as a promising tool to elaborate
16 de-wetting strategies.

17 The location of the liquid-vapor interface has been proven to be dynamic and dependent on the process
18 operating parameters and membrane material interactions which has shown to be vary significantly. The
19 developed optical tool can visualize and better interpret pore wetting in membrane distillation. **Table 4**
20 summarizes the current advantages of this optical wetting detection tool and further R&D challenges
21 that need to be addressed to make this tool even better.

22 **Table 4: Advantages and challenges of the developed optical tool**

Advantages of the optical tool	Challenges to be addressed in future studies
<ul style="list-style-type: none">• In-situ tool which can evaluate and visualize wetting propagation and mechanisms at the global and local scales	<ul style="list-style-type: none">• Wetting visualization is limited by the optics of the camera used therefore wetting can be visualized but not at the scale of the pores (i.e., microscale)

Advantages of the optical tool	Challenges to be addressed in future studies
<ul style="list-style-type: none"> • Can be adapted to the main membrane distillation configurations • Non-invasive • Non-destructive • Easy to setup and can be coupled with existing membrane cells with adaptations to visualize the membrane surface 	<ul style="list-style-type: none"> • Opaque particles / or translucent feed solutions and scaling/fouling may alter visualization of the wetting mechanism • Utilization limited to steady state as water properties, and operating parameters may influence light transmission • Thin, durable spacers designed to make more of the membrane surface available for visualization, ideally approaching complete surface visualization. • The optical method needs to be adapted to HF membrane configuration

1

2

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3 technologique pour analyser, remédier ou réduire les risques environnementaux (DS0102) 2014 for
4 funding the projet WETMEM (ANR-14-CE04-0008).

5 They also acknowledge Manon Montaner, from INSA-TBI for her contribution to the design and
6 manufacturing of the experimental module.

7

8 Nomenclature

Acronym	Description	Unit
A	Membrane area	m ²
AGMD	Airgap membrane distillation	
a _{oi}	Open area of the spacer/opening	mm ²
C	Salt concentration	g/L
CA	Contact angle	°
D.I. water	De-ionized water	
DCMD	Direct contact membrane distillation	
DDTI	Detection of Dissolved Tracer Intrusion	
E.g.	Example	
EDX	Energy-dispersive X-ray spectroscopy	
EIC	Electrochemical impedance spectroscopy	
F	Flow rate	m/s or L/h
FS	Flat sheet	
HF	Hollow fiber	
i.e.	That is	
I _{avg}	Overall average intensity	
I _{initial}	Intensity of a reference image where no wetting had occurred	
I _{max}	Intensity of an image captured when total wetting had occurred	
I _{observed}	Intensity of one image at time (t)	
I _{ROI} (X _i ,Y _i)	Mean light intensity in one ROI	Gray value
J	Flux	Kg/m ² .h
K _M	Membrane permeability in VMD at 20°C	s.mol ^{1/2} .m ⁻¹ .kg ^{-1/2}
L/V	Liquid vapor	
LED	Light-emitting diode	

LEP _w	Liquid entry pressure with D.I. water	bar
MD	Membrane distillation	
N	Normalized light intensity	
ND	Refractive index	
PVDF	Polyvinylidene fluoride	
r _{avg}	Average pore radius of the membrane	μm
Re	Reynolds number	
RGB	Red green blue	
RO	Reverse osmosis	
ROI	Region of interest	
SDS	Sodium dodecyl sulfate	
SEM	Scanning electron microscopy	
SFE	Surface free energy	mN/m
SGMD	Sweeping gas membrane distillation	
S _i	Area of one ROI with x _i and y _i as coordinates	
ST	Surface tension	mN/m
STP	Standard pressure and temperature	
T	Temperature	°C or K
t	Time	
TDS	Total dissolved solids	mg/L
T _f	Feed temperature	°C
UV	Ultra violet	
VMD	Vacuum membrane distillation	
w.r.t	With respect to	
x _i	Location of one ROI in x direction	
y _i	Location of one ROI in y direction	
δ	Membrane thickness	μm or m
ε	Porosity	
Υ	Surface tension	mN/m
ω _p	Pore wetting ratio	

6. References

- [1] E. Drioli, A. Ali, F. Macedonio, Membrane distillation: Recent developments and perspectives, *State-of-the-Art Rev. Desalination*. 356 (2015) 56–84. doi:10.1016/j.desal.2014.10.028.
- [2] A. Deshmukh, C. Boo, V. Karanikola, S. Lin, A.P. Straub, T. Tong, D.M. Warsinger, M. Elimelech, Membrane Distillation at the Water-Energy Nexus: Limits, Opportunities, and Challenges, *Energy Environ. Sci.* (2018).
- [3] J.-P. Mericq, S. Laborie, C. Cabassud, Vacuum membrane distillation of seawater reverse osmosis brines, *Water Res.* 44 (2010) 5260–5273. doi:10.1016/j.watres.2010.06.052.
- [4] J.-P. Mericq, S. Laborie, C. Cabassud, Evaluation of systems coupling vacuum membrane distillation and solar energy for seawater desalination, *Chem. Eng. J.* 166 (2011) 596–606. doi:10.1016/j.cej.2010.11.030.
- [5] A. Kujawska, J.K. Kujawski, M. Bryjak, M. Cichosz, W. Kujawski, Removal of volatile organic compounds from aqueous solutions applying thermally driven membrane processes. 2. Air gap membrane distillation, *J. Membr. Sci.* 499 (2016) 245–256. doi:10.1016/j.memsci.2015.10.047.
- [6] E. Guillen-Burrieza, M.O. Mavukkandy, M.R. Bilad, H.A. Arafat, Understanding wetting phenomena in membrane distillation and how operational parameters can affect it, *J. Membr. Sci.* 515 (2016) 163–174. doi:10.1016/j.memsci.2016.05.051.
- [7] P. Jacob, S. Laborie, C. Cabassud, Visualizing and evaluating wetting in membrane distillation: New methodology and indicators based on Detection of Dissolved Tracer Intrusion (DDTI), *Desalination*. 443 (2018) 307–322. doi:10.1016/j.desal.2018.06.006.
- [8] K. Smolders, A.C.M. Franken, Terminology for Membrane Distillation, *Desalination*. 72 (1989) 249–262. doi:10.1016/0011-9164(89)80010-4.
- [9] M. Gryta, Influence of polypropylene membrane surface porosity on the performance of membrane distillation process, *J. Membr. Sci.* 287 (2007) 67–78.
- [10] S.A. Hashemifard, T. Matsuura, A.F. Ismail, M. Rezaei Dasht Arzhandi, D. Rana, G. Bakeri, Characterization of partial pore wetting in hollow fiber gas absorption membrane contactors: An EDX analysis approach, *Chem. Eng. J.* 281 (2015) 970–980. doi:10.1016/j.cej.2015.07.036.
- [11] F.E. Ahmed, B.S. Lalia, R. Hashaikeh, Membrane-based detection of wetting phenomenon in direct contact membrane distillation, *J. Membr. Sci.* (2017). doi:10.1016/j.memsci.2017.04.035.
- [12] Y. Chen, Z. Wang, G.K. Jennings, S. Lin, Probing Pore Wetting in Membrane Distillation Using Impedance: Early Detection and Mechanism of Surfactant-Induced Wetting, *Environ. Sci. Technol. Lett.* (2017). doi:10.1021/acs.estlett.7b00372.
- [13] P. Jacob, T. Zhang, S. Laborie, C. Cabassud, Influence of operating conditions on wetting and wettability in membrane distillation using Detection of Dissolved Tracer Intrusion (DDTI), *Desalination*. 468 (2019) 114086. doi:10.1016/j.desal.2019.114086.
- [14] E. Guillen-Burrieza, R. Thomas, B. Mansoor, D. Johnson, N. Hilal, H. Arafat, Effect of dry-out on the fouling of PVDF and PTFE membranes under conditions simulating intermittent seawater membrane distillation (SWMD), *J. Membr. Sci.* 438 (2013) 126–139.

- [15] M. Gryta, The study of performance of polyethylene chlorinetrifluoroethylene membranes used for brine desalination by membrane distillation, *Desalination*. 398 (2016) 52–63. doi:10.1016/j.desal.2016.07.021.
- [16] M. Gryta, J. Grzechulska-Damszel, A. Markowska, K. Karakulski, The influence of polypropylene degradation on the membrane wettability during membrane distillation, *J. Membr. Sci.* 326 (2009) 493–502. doi:10.1016/j.memsci.2008.10.022.
- [17] V. Chen, H. Li, A.G. Fane, Non-invasive observation of synthetic membrane processes – a review of methods, *J. Membr. Sci.* 241 (2004) 23–44. doi:10.1016/j.memsci.2004.04.029.
- [18] M. Gryta, Long-term performance of membrane distillation process, *J. Membr. Sci.* 265 (2005) 153–159.
- [19] M. Gryta, M. Barancewicz, Influence of morphology of PVDF capillary membranes on the performance of direct contact membrane distillation, *J. Membr. Sci.* 358 (2010) 158–167. doi:10.1016/j.memsci.2010.04.044.
- [20] K.W. Lawson, D.R. Lloyd, Membrane distillation. I. Module design and performance evaluation using vacuum membrane distillation, *J. Membr. Sci.* 120 (1996) 111–121.
- [21] L. Peña, J.M.O. de Zárata, J.I. Mengual, Steady states in membrane distillation: Influence of membrane wetting, *J. Chem. Soc. Faraday Trans.* 89 (1993) 4333–4338.
- [22] L.D. Nghiem, F. Hildinger, F.I. Hai, T. Cath, Treatment of saline aqueous solutions using direct contact membrane distillation, *Desalination Water Treat.* 32 (2011) 234–241.
- [23] E. Guillen-Burrieza, A. Ruiz-Aguirre, G. Zaragoza, H.A. Arafat, Membrane fouling and cleaning in long term plant-scale membrane distillation operations, *J. Membr. Sci.* 468 (2014) 360–372.
- [24] R.B. Saffarini, B. Mansoor, R. Thomas, H.A. Arafat, Effect of temperature-dependent microstructure evolution on pore wetting in PTFE membranes under membrane distillation conditions, *J. Membr. Sci.* 429 (2013) 282–294. doi:10.1016/j.memsci.2012.11.049.
- [25] J. Hou, M.Y. Zulkifli, M. Mohammad, Y. Zhang, A. Razmjou, V. Chen, Biocatalytic gas-liquid membrane contactors for CO₂ hydration with immobilized carbonic anhydrase, *J. Membr. Sci.* 520 (2016) 303–313.
- [26] A. McLeod, B. Jefferson, E.J. McAdam, Toward gas-phase controlled mass transfer in micro-porous membrane contactors for recovery and concentration of dissolved methane in the gas phase, *J. Membr. Sci.* 510 (2016) 466–471.
- [27] S.-H. Yeon, K.-S. Lee, B. Sea, Y.-I. Park, K.-H. Lee, Application of pilot-scale membrane contactor hybrid system for removal of carbon dioxide from flue gas, *J. Membr. Sci.* 257 (2005) 156–160.
- [28] C. Duan, W.-N. Mei, W.-G. Yin, J. Liu, J. Hardy, M. Bai, S. Ducharme, Theoretical study on the optical properties of polyvinylidene fluoride crystal, *J. Phys. Condens. Matter.* 15 (2003) 3805.
- [29] Z. Wang, Y. Chen, X. Sun, R. Duddu, S. Lin, Mechanism of pore wetting in membrane distillation with alcohol vs. surfactant, *J. Membr. Sci.* 559 (2018) 183–195. doi:10.1016/j.memsci.2018.04.045.
- [30] L. Han, Y.Z. Tan, T. Netke, A.G. Fane, J.W. Chew, Understanding oily wastewater treatment via membrane distillation, *J. Membr. Sci.* 539 (2017) 284–294. doi:10.1016/j.memsci.2017.06.012.

- [31] T.D. Dao, J.-P. Mericq, S. Laborie, C. Cabassud, A new method for permeability measurement of hydrophobic membranes in Vacuum Membrane Distillation process, *Water Res.* 47 (2013) 2096–2104. doi:10.1016/j.watres.2013.01.040.
- [32] Y.Z. Tan, L. Han, W.H. Chow, A.G. Fane, J.W. Chew, Influence of module orientation and geometry in the membrane distillation of oily seawater, *Desalination.* 423 (2017) 111–123. doi:10.1016/j.desal.2017.09.019.
- [33] A. Siddiqui, N. Farhat, S.S. Bucs, R.V. Linares, C. Picioreanu, J.C. Kruithof, M.C. van Loosdrecht, J. Kidwell, J.S. Vrouwenvelder, Development and characterization of 3D-printed feed spacers for spiral wound membrane systems, *Water Res.* 91 (2016) 55–67.
- [34] N. Sreedhar, N. Thomas, O. Al-Ketan, R. Rowshan, H. Hernandez, R.K. Abu Al-Rub, H.A. Arafat, 3D printed feed spacers based on triply periodic minimal surfaces for flux enhancement and biofouling mitigation in RO and UF, *Desalination.* 425 (2018) 12–21. doi:10.1016/j.desal.2017.10.010.
- [35] N. Thomas, N. Sreedhar, O. Al-Ketan, R. Rowshan, R.K. Abu Al-Rub, H. Arafat, 3D printed triply periodic minimal surfaces as spacers for enhanced heat and mass transfer in membrane distillation, *Desalination.* 443 (2018) 256–271. doi:10.1016/j.desal.2018.06.009.
- [36] M. Monnot, H.T.K. Nguyễn, S. Laborie, C. Cabassud, Seawater reverse osmosis desalination plant at community-scale: Role of an innovative pretreatment on process performances and intensification, *EPIC2015.* 113 (2017) 42–55. doi:10.1016/j.cep.2016.09.020.
- [37] J. Schindelin, I. Arganda-Carreras, E. Frise, V. Kaynig, M. Longair, T. Pietzsch, S. Preibisch, C. Rueden, S. Saalfeld, B. Schmid, Fiji: an open-source platform for biological-image analysis, *Nat. Methods.* 9 (2012) 676.