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The plastidial *Arabidopsis thaliana* NFU1 protein binds and delivers [4Fe-4S] clusters to specific client proteins

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ABSTRACT

Proteins incorporating iron-sulfur (Fe-S) cofactors are required for a plethora of metabolic processes. Their maturation depends on three Fe-S cluster assembly machineries in plants, located in the cytosol, mitochondria and chloroplasts. After *de novo* formation on scaffold proteins, transfer proteins load Fe-S clusters onto client proteins. Among plastidial representatives of these transfer proteins, NFU2 and NFU3 are required for the maturation of the [4Fe-4S] clusters present in photosystem I subunits, acting upstream of the high-chlorophyll fluorescence 101 (HCF101) protein. NFU2 is also required for the maturation of the [2Fe-2S] dihydroxyacid dehydratase, important for branched-chain amino acid synthesis. Here, we report that recombinant *Arabidopsis thaliana* NFU1 assembles one [4Fe-4S] cluster per homodimer. Performing co-immunoprecipitation experiments and assessing physical interactions of NFU1 with many [4Fe-4S]-containing plastidial proteins in binary yeast two-hybrid assays, we also gained insights into the specificity of NFU1 for the maturation of chloroplastic Fe-S proteins. Using bimolecular fluorescence complementation and *in vitro* Fe-S cluster transfer experiments, we confirmed interactions with two proteins involved in isoprenoid and thiamine biosynthesis, 1-hydroxy-2-methyl-2-(E)-butenyl 4-diphosphate synthase (ISPG) and 4-amino-5-hydroxymethyl-2-methylpyrimidine phosphate synthase (THIC), respectively. An additional interaction detected with the scaffold protein SUFD enabled us to build a model in which NFU1 receives its Fe-S cluster from the $SUFBC_2D$ scaffold complex and serves in the maturation of specific [4Fe-

4S] client proteins. The identification of the NFU1 partner proteins reported here more clearly defines the role of NFU1 in Fe-S client protein maturation in Arabidopsis chloroplasts among other SUF components.

Iron and sulfur are essential elements for all organisms, in particular as building blocks of iron-sulfur (Fe-S) proteins that are themselves essential for several key molecular processes. In plants, Fe-S proteins participate to photosynthesis and respiration, being present in the electron transfer chains found in chloroplasts and mitochondria, but also for sulfur and nitrogen assimilation, or chlorophyll and vitamin metabolisms to cite a few examples (1, 2). The most represented Fe-S cluster forms are the [2Fe-2S] and [4Fe-4S] clusters with some proteins such as the glutamate synthase (GOGAT) incorporating [3Fe-4S] clusters (2, 3). The maturation of Fe-S proteins is not a spontaneous process and relies on dedicated machineries that exist in all kingdoms although with some variations in the type and number of machineries present and in the molecular actors implicated (4). In plants, three machineries exist: the iron-sulfur cluster (ISC) machinery is found in mitochondria; the cytosolic iron-sulfur protein assembly (CIA) machinery provides Fe-S clusters for the maturation of both cytosolic and nuclear Fe-S proteins and is dependent on the ISC machinery; and the sulfur mobilization (SUF) machinery works independently and is involved in the maturation of plastidial proteins (3, 4). Regardless the machinery, the Fe-S cluster biogenesis can be divided into several steps. In early steps, a Fe-S cluster is built *de novo* on so-called scaffold proteins, necessitating a multiprotein assembly complex for the mobilisation, reduction and assembly of iron and sulfur atoms. The preformed Fe-S cluster will be conveyed to a set of transfer proteins, eventually with the help of chaperones. After possible conversion and exchange among these Fe-S cluster transfer proteins/complexes, the Fe-S cluster is delivered to final targets (4).

In the current model of the mitochondrial ISC machinery, a [2Fe-2S] cluster is assembled on ISU/ISCU scaffold proteins prior to its transfer to a glutaredoxin (GRX) referred to as Grx5 in yeast. Whereas these steps should be sufficient for the maturation of [2Fe-2S] proteins (5), the maturation of [4Fe-4S] proteins also requires a conversion from two [2Fe-2S] clusters to a [4Fe-4S] cluster by reductive coupling, probably occurring in the course of the interaction between GRX5 and ISCA1/2 heterodimer (Brancaccio et al., 2014). The BOLA1 and IBA57 maturation factors may be involved at this step. Further late acting Fe-S cluster transfer proteins, NFU1, IND1/INDH (when present) and BOLA3, participate to the maturation of some [4Fe-4S] proteins but not all (7–9).

In the current model of the chloroplastic SUF machinery, the cysteine desulfurase NFS2, assisted by SUFE proteins, provides the required sulfur atoms for the *de novo* synthesis of a Fe-S cluster onto the $SUFBC_2D$ scaffold complex. However, how the system is supplied with iron atoms and electrons is less clear. Accordingly, all these genes are essential as confirmed by the embryo-lethality of the corresponding *Arabidopsis thaliana* loss-of-function mutants (10–16). Then, only scarce information are available in the following stages albeit proteins belonging to the same family as the mitochondrial counterparts are present *i.e.*, GRXS14/16, BOLA1/4, IBA57.2, SUFA1, NFU1/2/3 and HCF101 (2, 3, 17–19). A first relevant functional information is that, except IBA57.2, each gene/protein complements the yeast mutants for the corresponding mitochondrial orthologs (20, 21). The physiological analysis of *hcf101*, *nfu2* and *nfu3* knock-down or knock-out *A. thaliana* mutants, which exhibit dwarf phenotypes, pointed to their involvement in the maturation of the [4Fe-4S] clusters found in photosystem I (PSI), with NFU2 and NFU3

acting directly upstream of HCF101 (22–25). HCF101 is also required for the maturation of the [4Fe-4S] cluster in the ferredoxin-thioredoxin reductase (FTR) (26, 27). In accordance with its capacity to bind both [2Fe-2S] and [4Fe-4S] clusters, NFU2 is required for the maturation of the [2Fe-2S] cluster present in the dihydroxyacid dehydratase (DHAD), an enzyme implicated in the synthesis of branched-chain amino acids (23, 28, 29). A valid biochemical characterization of NFU3 was hampered so far by the impossibility to express sufficient amounts of soluble, non-aggregated recombinant protein for a proper spectroscopic characterization, but it was suggested to bind both [3Fe-4S] and [4Fe-4S] clusters (22). On the other hand, no firm conclusion has been gained from the study of individual *grxS14*, *grxS16* and *sufa1* Arabidopsis mutants except that these proteins are dispensable, at least in the growth conditions tested (17, 23, 30, 31).

Unlike *nfu2* and *nfu3* mutants, an Arabidopsis *nfu1* mutant has no phenotype when grown under standard conditions (23). This result suggests that NFU1 has either an accessory or very specific function or a functional redundancy with other Fe-S transfer protein(s), rendering difficult the determination of its role *in planta*. Hence, to tackle the role of NFU1 in Arabidopsis, we have characterized the biochemical and spectroscopic properties of the recombinant protein, before identifying its plastidial partner proteins using co-immunoprecipitation (co-IP) experiments and a binary yeast-two hybrid (Y2H) screen with other SUF components and many candidate [4Fe-4S]-containing proteins. The *in planta* interaction was validated for a few proteins using bimolecular fluorescence complementation (BiFC) assays before assessing the capacity of NFU1 to transfer its Fe-S cluster to two selected client proteins, the 1-hydroxy-2-methyl-2-(E)-butenyl 4-diphosphate synthase (ISPG/GcpE/HDS) and the 4-amino-2-methyl-5-hydroxymethylpyrimidine phosphate synthase (THIC). The identification of these NFU1 partners allowed us to position NFU1 in the nebula of SUF components and Fe-S client proteins.

RESULTS

AtNFU1 binds a [4Fe-4S] cluster into a homodimer

The plastidial NFU1/2/3 isoforms have two NFU domains repeated in tandem but only the N-terminal domain possesses the conserved CXXC motif participating in Fe-S cluster ligation. They are all able to restore the growth defect of the yeast mutant for the mitochondrial Nfu1 isoform (21). This suggested that they share some common properties such as the capacity to bind a [4Fe-4S] cluster but this has not yet been explored for any plant NFU1 isoforms. Moreover, it is important to assess whether NFU1 can bind other types of Fe-S clusters as observed for *A. thaliana* NFU2 (29).

The mature form of Arabidopsis NFU1 was expressed in *Escherichia coli* as an untagged recombinant protein. Unlike NFU2, which contains [2Fe-2S] clusters as purified (25, 29), no color typical of the presence of Fe-S cluster was visible in the lysed cell extract and purified NFU1 was devoid of Fe-S clusters, based on a UV-visible absorption spectrum showing only a single absorption peak at 280 nm. To evaluate the ability of NFU1 to bind Fe-S clusters, *in vitro* enzymatic Fe-S cluster reconstitution experiments were performed under anaerobic conditions in the presence of *E. coli* cysteine desulfurase (EcIscS). The UV-visible absorption and CD spectra of reconstituted NFU1 are shown in Fig. 1A. The absorption spectrum, comprising broad shoulders centred at ~400 nm and ~320 nm, is characteristic of a $[4Fe\text{-}4S]^{2+}$ cluster and is very similar to that reported for the reconstituted [4Fe-4S] cluster-bound form of NFU2, see Fig. S1 (29). Resonance Raman provides a more definitive assessment of cluster type, based on the observed Fe-S stretching modes (32), and

the spectrum obtained with 457.9 nm laser excitation is uniquely indicative of a [4Fe-4S]$^{2+}$ cluster (Fig. 1B). Interestingly, the UV-visible CD and resonance Raman spectra of the [4Fe-4S] centers in NFU1 and NFU2 are quite distinct (Fig. S1). For example, the dominant symmetric breathing mode of the [4Fe-4S] core shifts from 336 cm^{-1} in NFU1 to 344 cm^{-1} in NFU2. The structural origins of such differences may be important for explaining why only NFU2 is capable of binding both [2Fe-2S] and [4Fe-4S] clusters at the subunit interface. X-ray crystal structures will be required for meaningful interpretation.

Quantification of both iron and acid-labile sulfur atoms bound to the protein showed that the sample contained 1.54 ± 0.29 Fe and 1.68 ± 0.11 acid-labile S per NFU1 monomer. Hence analytical data indicate that reconstituted samples contain ~80% of [4Fe-4S] cluster-loaded NFU1. This is in accord with the UV-visible absorption values of ε_{400} = 6.2 mM^{-1}cm^{-1} based on NFU1 monomer. This translates to ε_{400} = 12.4 mM^{-1}cm^{-1} based on NFU1 dimer and approximately 80% of NFU1 with a subunit bridging [4Fe-4S] cluster, based on typical values of ε_{400} = 15 ± 2 mM^{-1}cm^{-1} for one [4Fe-4S]$^{2+}$ cluster. Moreover, analytical gel filtration experiments have been performed using both apo- and holo-NFU1 (Fig. 1C). Both forms eluted as a single peak corresponding to an estimated molecular mass of 30 kDa for apo-NFU1 and 25 kDa for holo-NFU1, respectively (Fig. 1B). From the theoretical molecular mass of a mature NFU1 (17 kDa), we concluded that both protein forms exist as homodimers. Taken together, these results indicated that the Fe-S cluster is not required for dimerization and that a reconstituted NFU1 homodimer contained ~80% [4Fe-4S] cluster.

AtNFU1 physically interacts with SUFD, SUFA1 and various [4Fe-4S]-containing proteins

So far, there is no known NFU1 partners in plants and only negative results have been obtained using binary Y2H, either with HCF101 or with two putative targets, APS reductase 1 (APR1) and the PSI subunit, PsaC (23, 29). As a first approach to determine what are the SUF partners and client Fe-S proteins of NFU1, co-IP experiments using anti-GFP antibodies have been achieved from 2 week-old transgenic lines expressing either a NFU1-GFP fusion (*ProNFU1::gNFU1-GFP* construct) or a GFP alone fused downstream of the chloroplastic targeting peptide (CTP) of the NFU3 protein (*Pro35S::CTP$_{NFU3}$-GFP* construct) for a specific targeting of GFP into the stroma of chloroplasts. After verifying that the constructs allow the specific expression of both fusion proteins in chloroplasts (Fig. S2), four replicates have been performed using distinct transgenic lines. The table 1 lists the 49 proteins present in at least 3 replicates originating from the co-IP performed with the NFU1-GFP expressing lines and absent in the 4 replicates performed with the control lines. It also comprises 5 additional proteins (labelled with a star) representing Fe-S proteins or subunits associated to Fe-S proteins found using a slightly less stringent cut-off, *i.e*, also present in at least 3 replicates but found in one replicate of control experiments. Using the latter filter, a total of 89 additional proteins were retrieved (Table S1). Among these 138 proteins, we identified 6 known Fe-S proteins. HCF101 and GRXS16 are SUF components. Recovering HCF101 was surprising, because no interaction was previously detected by Y2H and BiFC (Touraine et al., 2019). It may be that it has been pulled down as part of a complex formed with other Fe-S proteins. Concerning GRXS16, the question of a direct physical interaction remains asked. Although no interaction was observed between NFU1 and GRXS16 by binary Y2H (see below), the NFU2 paralog has proven to be able to efficiently transfer a [2Fe-2S] cluster to GRXS16 (29). A role of GRXS16 in Fe-S cluster biogenesis has yet to be validated *in planta*, but *in vitro* the

recombinant protein incorporates either a [2Fe-2S] or a [4Fe-4S] cluster (30). Four putative Fe-S client proteins have been isolated. The 5'adenylyl-phosphosulfate reductase 2 (APR2), one of the three APR isoforms in Arabidopsis, catalyzes the second step of sulfate assimilation and is thus implicated in cysteine and methionine synthesis. The quinolinate synthase SUFE3 is involved in NAD synthesis (12). The glutamate synthase 2 (GLU2) participates to nitrogen assimilation by catalysing the conversion of glutamine into glutamate at the expense of ferredoxin. The last Fe-S protein firmly identified is THIC, an enzyme involved in the synthesis of vitamin B1 (thiamine) (33). APR2, SUFE3 and THIC are known to bind [4Fe-4S] clusters whereas GLU2 binds a [3Fe-4S] cluster (2). Another likely candidate partner is the isopropyl malate isomerase (IIL1/IPMI), an Fe-S enzyme catalyzing the isomerization between 2-isopropylmalate and 3-isopropylmalate and thus implicated in the synthesis of leucine and glucosinolates (34). It forms a protein complex involving a large subunit and a small subunit, and the three isoforms corresponding to the small subunits have been retrieved from these experiments. Despite being found in the four co-IP replicates, the large subunit, bearing the Fe-S cluster, was not retained here because it was also found in two of the control experiments. Then, what is noticeable among the set of the 45 other identified proteins is the presence a set of proteins performing redox reactions. It includes four sulfurtransferases (STR9, 10, 12, 14), enzymes containing a rhodanese domain with a conserved cysteine involved in trans-persulfidation reactions (35). It also comprises several thioredoxin (TRX)-like proteins, TRX z, TRX-lilium2 also referred to as the atypical Cys-His rich thioredoxin 2 and a novel putative TRX superfamily member (At5g65840). Having reactive cysteines, it may be that these proteins formed covalent bonds with the reactive cysteines of an apo-NFU1. In line with such a possible interaction, we have recently observed that mitochondrial TRXs o have the ability to reduce an intramolecular disulfide formed in the NFU domain of mitochondrial NFU5 (36). Whether this is true also for chloroplastic NFU1/TRX couples remains to be investigated and the physiological relevance assessed. TRX z is known to be part of the plastid-encoded RNA polymerase (PEP) complex, likely explaining why fructokinase-like 2 and plastid transcriptionally active 5 and 17 proteins have been retrieved too (37). To conclude on that approach, there are quite a few other proteins that have never been biochemically characterized and for which there is no attributed function. Consequently, it is difficult to extrapolate whether they could be or not novel Fe-S containing proteins or involved in the maturation process. For instance, several chloroplastic chaperones are present in the expanded list and it may be that they are required for Fe-S cluster exchange as documented for the assembly step of the mitochondrial ISC machinery. Although usually powerful, it is worth pointing that the co-IP approach may not be the best for supposedly weak or transient interactions or interactions that would rely on the presence of an oxygen-labile Fe-S cluster.

Hence, as a second more systematic approach, we sought to analyze the capacity of NFU1 to interact with the presumed scaffold and transfer proteins of the SUF machinery, *i.e.* SUFB, SUFC, SUFD, SUFA1, BOLA1, BOLA4, IBA57.2, GRXS14, GRXS16, NUF2, NFU3 and HCF101 using binary Y2H assays. An interaction was only observed with SUFD and SUFA1 (Fig. 2). Then, a large set of [4Fe-4S]-containing proteins involved in various metabolic pathways were tested. This included nitrite reductase (NIR) and sulfite reductase (SIR), two siroheme-containing proteins involved in nitrogen and sulfur assimilation. This also included several radical-SAM enzymes *i.e.*, THIC, the chloroplastic lipoate synthase (cLIP), which participates to the formation of lipoic acid and should incorporate two [4Fe-4S]

clusters by analogy to the bacterial and mitochondrial orthologs, and the tRNA modifying enzyme, MIAB. Additional targets are the glutamine phosphoribosyl pyrophosphate amidotransferase 2 (ASE2) implicated in the *de novo* synthesis of purine, the ISPG/GcpE/HDS and 1-hydroxy-2-methyl-2-(E)-butenyl 4-diphosphate reductase (ISPH/HDR) participating in the synthesis of isoprenoids, the β-carotene isomerase DWARF27.1 notably required for strigolactone synthesis and its two homologues (DWARF27.2/3), the IIL1/IPMI large subunit described above and the 7-hydroxymethyl chlorophyll *a* reductase (HCAR), a protein involved in the chlorophyll metabolism and binding two [4Fe-4S] clusters. Among these 13 candidate partners, interactions have been detected with NIR, cLIP, THIC, HCAR, DWARF27.1 and ISPG when NFU1 was fused to the GAL4 binding domain (Fig. 3). In these experiments, we could not firmly confirm the existence of an interaction with the large subunit of IPMI because the GAL4 auto-activation caused by this protein is strong and concluding on the growth difference in the presence of NFU1 remains questionable. Looking at the strength of the interactions, the strongest ones were observed with THIC and DWARF27.1, then with ISPG, HCAR and cLIP and finally NIR, the latter being visible only in the absence of 3-amino triazol (3-AT).

To challenge the relevance of these interactions in plant cells, we additionally performed BiFC assays in Arabidopsis protoplasts. We have first validated NFU1 homodimerisation as revealed by the BiFC signal in Arabidopsis chloroplasts in cells co-expressing NFU1-YFP fusions (*i.e.* NFU1 fused at the N-terminus of the N-terminal (AA_{1-155}) or C-terminal regions ($AA_{156-239}$) of YFP (Figs. 4A, S3 & S4). We next tested combinations of NFU1 with five selected partners isolated by one or the other above-described approach *i.e.*, SUFA1, ISPG, THIC, SUFE3 and cLIP (Figs. 4B, S3 & S4). Transfections with combinations of NFU1 fused to the C-terminal region of the YFP protein (NFU1-C) and of the partners fused to the N-terminal region of the YFP (protein-N) revealed positive BiFC in all cases, exclusively in the chloroplasts. These experiments revealed that NFU1 cooperates in a close environment with all the proteins tested in this plant reporter system.

NFU1 transfers its [4Fe-4S] cluster to apo-ISPG and -THIC in vitro

Based on the identified interactions, we tested whether we could obtain evidence for a Fe-S cluster transfer *in vitro* using two client proteins, ISPG and THIC, as their Fe-S cluster content was characterized previously (38, 39). Moreover, both proteins have a single [4Fe-4S] cluster per monomer, unlike HCAR and cLIP, and they are formed by a single domain and do not rely on additional subunits, unlike IPMI and SUFE3.

Hence, we produced the respective His-tagged recombinant proteins. After an aerobic purification, ISPG and THIC were both a mixture of apo- and holo-forms. By treating the proteins with an excess of EDTA and TCEP, we could obtain stable apo-proteins. Thus, we first assessed the ability of both apo-forms to incorporate [4Fe-4S] clusters by performing *in vitro* IscS-mediated Fe-S cluster reconstitution experiments (Fig. S5). Both reconstituted proteins had broad shoulders at ~320 nm and ~400 nm characteristic of a $[4Fe\text{-}4S]^{2+}$ center. This was quantitatively confirmed by analytical measurements of both iron and acid-labile sulfur atoms bound to the proteins as ISPG incorporated 3.99 ± 0.14 iron and 3.91 ± 0.11 labile sulfur per monomer whereas THIC contained 3.81 ± 0.43 iron and 3.63 ± 0.63 labile sulfur per monomer after reconstitution.

In a second step, *in vitro* Fe-S cluster transfer experiments were performed using a 2-fold molar excess of untagged holo-NFU1 (reconstituted as before) as compared to apo-acceptors. After 1h incubation, donor and acceptor proteins

were separated on a nickel affinity chromatography column. The unbound fractions contained NFU1 and eventually tiny amounts of the acceptor proteins (Figs. 5 and 6). The UV-visible absorption spectra indicated that the absorption bands typical of the Fe-S cluster in NFU1 were considerably diminished, though not totally, due to the 2-fold excess of NFU1 [4Fe-4S] clusters. On the other hand, the imidazole-eluted fractions, which contained ISPG or THIC, exhibited a brown-red colour. The UV-visible absorption spectra showed broad bands centred around 320 nm and 400 nm (Figs. 5 and 6) similar to the ones observed in the spectra of reconstituted proteins (Fig. S5). To assess transfer efficiency, we titrated the Fe and acid-labile S contents into acceptor proteins (Table S4) and compared these values to the theoretical amounts expected based on the presence of a [4Fe-4S] cluster per monomer of ISPG and THIC. After the 1 hour incubation time, we recovered ~50% and ~75% of holo-ISPG and holo-THIC, respectively. Altogether, these results pointed to an intact [4Fe-4S] cluster transfer from NFU1 to ISPG or THIC *in vitro*.

DISCUSSION

The maturation of all chloroplastic Fe-S proteins depends on the SUF machinery. The late-acting Fe-S cluster transfer proteins, including NFU1/2/3 and HCF101, are in principle in direct contact with client proteins. Unlike NFU1, some of the NFU2, NFU3 and HCF101 targets have been unveiled (11, 22–24, 40). The fact that an *A. thaliana nfu1* mutant has no macroscopic phenotype when grown under standard conditions prevented to delineate NFU1 function(s) and associated partners using a physiological approach (Touraine et al., 2019).

In this work, we purified the Arabidopsis NFU1 protein to investigate which type of Fe-S cluster(s) is bound to the protein *in vitro*. Previous studies showed that the large diversity in the domain organization of NFU proteins is associated to differences in their biochemical properties. The *Helicobacter pylori* Nfu, formed by a single NFU domain, and the Arabidopsis NFU2 formed by two NFU domains (only the N-terminal one containing the ligating cysteine residues) were shown to bind either one [2Fe-2S] cluster as purified from *E. coli* cells or one [4Fe-4S] cluster upon *in vitro* reconstitution (29, 41). The *Escherichia coli* and *Azotobacter vinelandii* NfuA that contain two domains, a C-terminal NFU domain fused to an N-terminal ISCA-type domain lacking the three conserved cysteine residues, were reported to bind only [4Fe-4S] clusters into homodimers (42, 43). In mitochondrial NFUs, the NFU domain is coupled to a domain of unknown function at its N-terminus. The human NFU1 was shown to incorporate a [4Fe-4S] cluster upon reconstitution in a homodimer but a SAXS-derived structural model highlighted the existence of a trimer of dimers (44, 45). These *in vitro* observations are in agreement with the *in vivo* investigations which showed that only the maturation of [4Fe-4S] proteins, including respiratory complexes, aconitase and lipoate synthase, is affected in a yeast *nfu1* mutant or in human patients (8, 45–47). Hence, the best, not to say the sole documented example where both *in vitro* and *in vivo* results support the requirement of a NFU protein in the maturation of [2Fe-2S] proteins is for the plant NFU2-DHAD couple (23, 28). Concerning Arabidopsis NFU1, we have observed that it was purified as an apo-form after heterologous expression in *E. coli* and that a [4Fe-4S] cluster holo-dimeric loaded form of NFU1 was uniquely formed after an *in vitro* anaerobic IscS-mediated reconstitution. NFU1 homodimerization was also visible in plant cells using BiFC but not in the yeast nuclear context during Y2H assays. Overall, this suggests that NFU1 should not participate to the maturation of [2Fe-2S] clusters, but only to the maturation of [4Fe-4S] and possibly of [3Fe-4S] clusters.

The question of the NFU1 partners among SUF components was unsolved so far. We have obtained evidence for possible interactions with SUFD, SUFA1, GRXS16 and HCF101. As already discussed, the detection in co-IP experiments of HCF101 and GRXS16 as putative NFU1 partners whereas they were not found to physically interact by Y2H and/or BiFC raises some doubt about the existence of a direct contact. For this reason, these interactions are not represented in the updated model proposed in Fig. 7. On the other hand, the interaction with SUFD and SUFA1 seen in Y2H and in Y2H and BiFC experiments respectively, suggests that NFU1 directly receives its cluster from the SUFBCD scaffold complex, before exchanging its cluster with SUFA1, the sole A-type transfer protein in chloroplasts. Accordingly, *E. coli* NfuA is able to receive its Fe-S cluster from both ISCU and $SUFBC_2D$ scaffold and to interact with all A-type transfer proteins, having the ability to transfer its cluster *in vitro* to SufA, IscA and ErpA, and forming a complex with ErpA stabilizing its Fe-S cluster (43, 48, 49). However, the model is that ErpA and NfuA to a lesser extent are the final Fe-S cluster donors to client apo-proteins (49). A [4Fe-4S] cluster transfer from the NFU domain of *A. vinelandii* NifU to ^{NIF}IscA is also documented (50). Noteworthy, *E. coli* SufA does also receive an Fe-S cluster from $SUFBC_2D$ (51) and is competent for instance for the maturation of both [2Fe-2S] and [4Fe-4S] containing proteins as shown using ferredoxin and biotin synthase (52, 53). This raises the possibility that, in chloroplasts, SUFA1 may obtain its cluster independently of NFU1 and on the contrary deliver it to NFU1. This sequence order would fit with the mitochondrial model in which NFUs likely act downstream of the A-type transfer proteins, ISCA1/ISA1 and ISCA2/ISA2. *In vitro* experiments provided evidence that these A-type transfer proteins exist as homodimers, as ISCA1/2 heterodimers but also as ISCA-IBA57 heterodimers (6, 54, 55). In fact, it seems that an even higher level of flexibility and adaptability exists for A-type transfer proteins as most of the characterized homo- or heterodimers are able to incorporate either [2Fe-2S] or [4Fe-4S] clusters. Hence, whereas the interaction between NFUs and A-type transfer proteins is a robust observation made in both prokaryote and eukaryote organisms (8, 43, 49, 56), which of NFU1 or SUFA1 act upstream remains uncertain in the chloroplastic SUF system. So far, SUFA1 was characterized as a [2Fe-2S] cluster containing protein able to receive *in vitro* a Fe-S cluster from GRXS14 and to transfer it to a ferredoxin (17, 31, 57). Hence, a Fe-S cluster exchange from NFU1 to SUFA1 would necessitate an oxidative conversion from the [4Fe-4S]-loaded NFU1 into [2Fe-2S] loaded SUFA1 forms, unless SUFA1 also binds [4Fe-4S] clusters. Alternatively, a reductive Fe-S cluster conversion from a [2Fe-2S]-loaded SUFA1 to [4Fe-4S]-loaded NFU1 might be possible. Further experiments are needed to address in which direction these exchanges occur.

The question of whether NFU1 has direct client Fe-S proteins was also the purpose of this study. By combining several complementary approaches, we have identified 9 NFU1 partners, if we consider the interaction with IPMI as too uncertain. They incorporate either a [3Fe-4S] cluster in the case of GLU2 or one or several [4Fe-4S] clusters in the case of SUFE3, APR2, cLIP, DWARF27.1, HCAR, ISPG, NIR and THIC. Although we could not test or obtain evidence for all these interactions by all methods, most of the interactions have been observed using at least two different approaches. Whereas the chloroplastic lipoate synthase cLIP has not been much investigated so far, the requirement of NFU1 as a maturation factor is consistent with NFU-type proteins being required for the maturation or repair of Fe-S clusters in lipoate synthase present in bacteria, or in yeast and human mitochondria (8, 58). The interactions of NFU1 with THIC and ISPG were evidenced both *in vivo* by Y2H and

BiFC and *in vitro* from the observation of an intact [4Fe-4S] cluster transfer from a holodimeric NFU1. The fact that THIC was also retrieved from the co-IP experiments makes little doubt about the validity of this interaction. Concerning ISPG, the result is consistent with the observation that *E. coli* NfuA is also a required maturation factor for bacterial IspG/H enzymes as recently demonstrated by a genetic approach (49). Taken together, these results indicate that the plastidial NFU1 of Arabidopsis is involved in maturation pathways that are conserved throughout evolution. For other proteins specific to plants, such as DWARF27.1 and HCAR, obtaining data about their maturation will be crucial as no extrapolation is possible from other model organisms. Only HCAR was analyzed before by western blots in *sufb*, *sufc*, *sufd*, *nfu2* and *hcf101* Arabidopsis mutants (11). The protein level was diminished in *sufb*, *sufc*, *sufd* RNAi lines, although not totally absent, but did not vary much in *nfu2* and *hcf101*. This leads us to discuss the physiological consequences of the described interactions and possible redundancies with other SUF maturation factors. The *sufa1* and *nfu1* Arabidopsis mutants have no or weak growth phenotypes under standard conditions, which is also the case of *apr2* or *hcar* mutants (59, 60). Nevertheless, mutants for other identified proteins have either strong(er) growth phenotypes, for instance the albino phenotype of an *ispg* mutant (61), or even embryo- or seedling-lethal phenotypes as observed for *sufe3* and *thic* mutants, respectively (12, 33). These phenotypic differences indicate indeed the existence of back-up system(s) for NFU1, meaning a certain level of redundancy with other maturation factors. This would be totally in line with the *E. coli* model, in which several factors serve for the maturation of a single target and their nature may depend on the growth conditions (62). For instance, in *E. coli*, depending on stress conditions, either NfuA or ErpA is involved in the maturation of ISPG (49). Hence, determining whether the functions of NFU1 are restricted to stress conditions or more prominent in this context is likely now required. In plant chloroplasts, the most likely candidates for ensuring functions similar to NFU1 are obviously NFU2, NFU3 but also HCF101 because they assemble the same type of cluster and they are critical for plant growth (22, 29, 40). From a phylogenetic point of view, NFU1 and NFU2/3 form two separate phylogenetic clades with the gene duplication generating *NFU2* and *NFU3* having occurred in an ancestor of angiosperms. For instance, a single *NFU2/3* representative is present in chlorophyceae, bryophytes and lycophytes whereas there are two in monocots and dicots (3, 23). Assuming that NFU1/2/3 originate from a single ancestral gene, these proteins may have diverged to some extent but also conserved common biochemical and structural properties. In support of this conclusion, all three plastidial NFUs were able to restore the growth defect of a yeast mutant for the mitochondrial Nfu1, despite the difference in protein organization between NFUs present in both organelles (21). However, the functions of NFU1 and NFU2/3 diverged and are clearly not fully overlapping. Unlike NFU1, NFU2 and NFU3 are involved in PSI maturation together with HCF101 but are also biochemically competent for the maturation of the [2Fe-2S] cluster in DHAD (23). Additional functions of NFU2/3 may actually have been masked by the strong effect on PSI and by their redundancy (a double *nfu2 nfu3* mutant is lethal, Touraine et al., 2019). Hence, to address the question of redundancy among NFUs, it would be mandatory to perform similar experiments with NFU2 and NFU3 in order to identify their set of client proteins.

Experimental procedures

Heterologous expression in E. coli and purification of recombinant proteins

The sequences coding for the presumed mature forms (*i.e.*, devoid of N-terminal targeting sequences) of Arabidopsis NFU1, THIC and ISPG were cloned respectively into the *Nde*I and *Bam*HI restriction sites of pET12a, the *Nco*I-*Xho*I and *Nde*I-*Xho*I restriction sites of pET28a (Table S2), in order to produce an untagged NFU1, an N-terminal His-tagged THIC and a C-terminal His-tagged ISPG.

NFU1 was expressed in the *E. coli* BL21 (DE3) strain containing the pSBET plasmid (63). Protein expression in 3.2 L was induced by adding of 100 µM of isopropyl β-D-thiogalactopyranoside (IPTG) during exponential growth phase. After 4 h at 37°C, cells were collected by centrifugation for 20 min at 6,318 g and the cell pellets were resuspended in about 25 mL of TN buffer (30 mM Tris-HCl pH 8.0, 200 mM NaCl). Bacteria cells were lysed by sonication (3 x 1 min) and cell debris removed at 4°C by centrifugation for 30 min at 27,216 g. The soluble fraction was sequentially precipitated by ammonium sulfate to 40% and then to 80% of the saturation. NFU1 was recovered mainly in the 0-40 % ammonium sulfate fraction. This fraction was subjected to gel filtration chromatography (ACA44) equilibrated with TN buffer. NFU1-containing fractions were pooled, concentrated and dialyzed against 30 mM Tris-HCl pH 8.0 buffer by ultrafiltration (YM10 membrane) under nitrogen pressure using an Amicon cell. The sample was loaded on an ion exchange chromatography (DEAE sepharose column) equilibrated in 30 mM Tris-HCl pH 8.0 buffer, before applying a linear 0-0.4 M NaCl gradient. The purest fractions containing NFU1 as judged by SDS-PAGE analysis were pooled and dialyzed against 30 mM Tris-HCl pH 8.0 buffer by ultrafiltration. Finally, the fractions were concentrated and stored at -20°C until further use.

The His-tagged ISPG and THIC were expressed in the *E. coli* Rosetta2 (DE3) strain. Protein expression was achieved in a 3.2 L culture. After an initial growth phase at 37°C up to the exponential phase (OD600 = 0.6-0.8), flasks were placed two hours at 4°C in the presence of 0.5% ethanol before induction by 100 µM IPTG. The cultures were further grown for about 18 h at 20°C. Cells were collected by centrifugation for 20 min at 6,318 g and resuspended in about 25 mL of TN buffer plus imidazole 20 mM (TNI20). Bacteria cells were lysed by sonication (3 x 1 min) and soluble and insoluble fractions cell were separated by centrifugation for 30 min at 27,216 g and 4°C. The soluble fraction was then loaded onto a Ni-NTA affinity column (Qiagen) equilibrated in TNI20 buffer. After extensive washing, recombinant proteins were eluted in TN buffer containing 250 mM imidazole. Proteins were then concentrated and dialyzed against TN buffer by ultrafiltration under nitrogen pressure (Amicon, YM10 membrane) and stored at -20°C. The purity of each recombinant protein was checked on SDS-PAGE. The concentrations of apo-proteins were determined spectrophotometrically using the theoretical molecular extinction coefficient at 280 nm of 4,595 $M^{-1}cm^{-1}$ for NFU1, 41,425 $M^{-1}cm^{-1}$ for ISPG and 92,290 $M^{-1}cm^{-1}$ for THIC.

In vitro IscS-mediated reconstitution of Fe-S cluster

All experiments were done at room temperature under anaerobic atmosphere using a Jacomex glovebox (O_2 < 2 ppm). Reduced apo-NFU1, ISPG and THIC were obtained by treating purified proteins with a 20-fold excess TCEP and a 50-fold excess EDTA for 1 h or overnight in the case of NFU1 which allows to get rid of residual polysulfides visible from the presence of a shoulder at 320 nm (64), before desalting on a G-25 column pre-equilibrated with 30 mM Tris-HCl pH 8.0 buffer. The Fe-S cluster reconstitution was performed in 500 µL of 37.5 mM Tris-HCl pH 8.0, 37.5 mM NaCl buffer, using 50 µM protein, a 20-fold excess of L-cysteine and ammonium iron (II) sulfate hexahydrate and catalytic

amount of *E. coli* cysteine desulfurase IscS purified as described previously (36). The reaction was initiated by the addition of IscS and followed by monitoring the UV-visible absorption spectrum. After a 1 h reaction, the Fe-S cluster-loaded proteins were desalted on a G-25 column equilibrated with TN buffer.

Determination of the oligomerization state of NFU1

The oligomerization state of apo- and holo-NFU1 was determined by size-exclusion chromatography. Samples containing about 200 µg of protein were loaded onto Sephadex S200 10/300 column equilibrated in TN buffer and connected to an Akta purifier system (GE Healthcare). Proteins were detected by recording absorption at 280 nm and 420 nm. The column was calibrated using a molecular weight standard from Sigma. Elution volume, protein name and molecular weight are as follows: 8.47 mL, thyroglobulin, 669 kDa; 10.51 mL, apo-ferritin, 443 kDa; 11.92 mL, β-amylase, 200 kDa; 13.89 mL, bovine serum albumin, 66 kDa; 15.74 mL, carbonic anhydrase, 29 kDa; 17.01 mL, cytochrome c, 12.4 kDa, and 18.28 mL, aprotinin, 6.5 kDa.

In vitro Fe-S cluster transfer experiments

Under strictly anaerobic conditions in a Jacomex glovebox (O_2 < 2 ppm), two molar equivalents of reconstituted holo-NFU1 (50 or 100 µM) respective to reduced apo-ISPG or apo-THIC (25 or 50 µM) were mixed for 1 h in the absence or presence of a 10-fold excess EDTA to ensure that the Fe-S cluster is transferred intact and not upon degradation and reassembly. Untagged NFU1 and tagged acceptor proteins (ISPG or THIC) were then separated on Ni-NTA affinity column (IMAC-Qiagen) equilibrated in TNI20 buffer. The column was then washed with 6 column volumes of TNI20 buffer, and elution was performed using TN buffer containing 250 mM imidazole. Fractions corresponding to washing and elution steps were concentrated to a volume of 500 µL using Vivaspin 500 centrifugal filters, before recording UV-visible absorption spectra and quantifying the contents in iron and acid-labile sulfide. Aliquots of each fraction were also analyzed by SDS-PAGE.

Spectroscopic methods

UV-visible absorption spectra were recorded using Shimadzu UV-3101 PC scanning or Agilent Cary 60 spectrophotometers. CD spectra were recorded using a JASCO J-715 spectropolarimeter. Septa-sealed quartz cuvette cells with either a 1-mm or 1-cm pathlength were used for both absorption and CD spectroscopies. Resonance Raman spectra were acquired using a Ramanor U1000 scanning spectrometer (Instruments SA, Edison, NJ) fitted with a cooled photomultiplier tube and photon-counting electronics (Instruments SA, Edison, NJ), using excitation lines from a Sabre argon laser (Coherent, Santa Clara, CA). A droplet of concentrated sample (approximately 2 mM in Fe-S clusters) was frozen at 17 K on a gold-plated sample holder mounted to a cold finger of a Displex Model CSA-202E closed cycle helium refrigerator (Air Products, Allentown, PA).

Iron and acid-labile sulfide quantification

Protein concentrations were determined using the colorimetric bicinchoninic acid assay (BCA) kit as recommended (Interchim). For iron quantification, different volumes of protein (25, 50 and 100 µL) were diluted in 130 µL of water. Proteins were precipitated by adding 90 µL of 70% (v/v) perchloric acid for 10 min at room temperature after strong shaking. After centrifugation (10 min, 11,600 g), 180 µL of supernatant were mixed with 144 µL of 3.2 mM bathophenanthroline disulfate, 72 µL of 192 mM sodium ascorbate and 152 µL of 6.2 M ammonium acetate. The mixture was homogenized with 30 sec of vortex before 30 min of incubation at room temperature. Iron amounts were determined by

subtracting the non-specific absorbance at 680 nm from the specific absorbance of the ferrous iron-chelator complex measured at 535 nm relatively to a standard curve obtained with ammonium iron (II) sulfate (0 to 20 nmol).

For acid-labile sulfide quantification, 25, 50 and 100 µL of proteins (eventually brought to 100 µL with water) were mixed with 300 µL of 1% (w/v) zinc acetate and 15 µL of 3 M NaOH. The mixture was incubated for 10 min at room temperature before adding 50 µL of 20 mM N,N-dimethyl-p-phenylenediamine (prepared in 7.2 M HCl) and 50 µL of 30 mM $FeCl_3$ (prepared in 1.2 M HCl). After 30 sec shaking and incubation at 4°C for 3 h, the mixture was centrifuged for 5 min at 11,600 *g* and the presence of methylene blue in the supernatant was recorded at 670 nm. Lithium sulfide (0 to 20 nmol) was used for the calibration curve.

Binary yeast two-hybrid assays

The experiments have been performed with the Gal4-based yeast two-hybrid reporter strain CY306 (Vignols et al., 2005). The sequences coding for proteins devoid of their chloroplastic targeting sequences were cloned in both pGADT7 and pGBKT7 vectors (Clontech Laboratories) using *Nco*I-, *Nde*I-, *Bam*HI-, or *Xho*I-containing primers (Table S2). Binary interactions were tested using both AD (Activator domain for Gal4) and BD (DNA binding domain for Gal4) fusion combinations. Cells co-transformed with pGAD- and pGBK-based constructs were selected on minimal YNB medium (0,7% yeast extract w/o amino acids, 2% glucose, 2% agar) containing required amino acids and bases (histidine, adenine, lysine, uracil and methionine). Interactions were assessed through cell growth on selective YNB media, in the absence of histidine and in the absence or presence of 2 to 10 mM 3-AT to get rid of some trans-activating constructs and estimate the strength of each interaction. Negative controls were performed using co-transformations of AD- or BD- fusions with empty vectors. Each dot is a 7 µL drop of cultures adjusted at an OD_{600}=0.05. Representative images shown here were taken 5 days post-dotting.

Bimolecular fluorescence complementation

All proteins selected for BiFC analyses were cloned as full-length open reading frames upstream of the C-terminal and N-terminal regions of the YFP protein into the pUC-SPYCE and pUC-SPYNE vectors, abbreviated -C and -N in figures, respectively), using a restriction site-based strategy and primers listed in Table S3 (66). Leaf protoplasts were prepared from 21- to 28-day old *Arabidopsis* plantlets grown in growth chambers in short day conditions (8h light / 23°C / 217 µmol.m^{-2}.s^{-1}, 16h dark / 20°C, 65% humidity) and transfected according to (67) using 10 µg of each pUC-SPYCE and pUC-SPYNE constructs expressing selected proteins. YFP fluorescence in *Arabidopsis* protoplast cells was recorded 20-24h post-transfection by using a Leica TCS SP8 confocal laser-scanning microscope. YFP was excited with an argon laser at 514 nm and detected at 520-550 nm, whereas chlorophyll auto-fluorescence was monitored at 680-720 nm after excitation at 561 nm. Images were obtained using a LAS X software and treated with Adobe Photoshop CS3 at high resolution. Results are representative of at least two independent transfection experiments including the analysis of around 20 cells per transformation event.

DNA constructs

To generate *ProNFU1::gNFU1-GFP* expressing plants, the *NFU1* locus (2000 bp prior to the start codon until the end of the coding sequence without the STOP) was amplified with AttB1ProNFU1 (5’ GGGGACAAGTTTGTACAAAAAAGC AGGCTCGCAGTACCCTAAACCATTG 3’) and AttB2NFU1 (5’ GGGGACCACTTTGTACAAGAAAGCT GGGCTTGTAAAGGTTAC 3’) primers,

cloned in pDONR207 vector and recombined in pGWB4 (68). To generate plants expressing a stroma-targeted GFP (*Pro35S::CTP$_{NFU3}$-GFP*), the *NFU3* chloroplastic peptide signal was amplified with AttB1NFU3 (5' GGGGACAAGTTTGTACAAAAAAGC AGGCTATGGGTTCTGTTTCGGGTC 3') and AttB2-PS-NFU3 (5' GGGGACCACTTTGTACAAGAAAGCT GGGCAGCTCACGTGACCAAATAC 3') primers, cloned in pDONR207 and recombined in pGWB505 (derived from pGWB series (68)). All the PCR products were obtained using high-fidelity Phusion DNA polymerase and each construct in pDONR207 was sequenced to ensure its integrity.

Co-immunoprecipitation experiments

ProNFU1::gNFU1-GFP and *Pro35S::CTP$_{NFU3}$-GFP* expressing seedlings were germinated and grown under long day condition (16 h/8 h light/dark) on half strength Murashige and Skoog medium (MS/2) with 0.05% MES, 1% sucrose, 0.7% agar. One g of aerial tissues from two weeks-old seedlings was cross-linked in 1% formaldehyde in PBS buffer two times (7 min under vacuum). The reaction was blocked by adding 300 mM glycine (30 min under vacuum). Fixed tissues were rinsed three times with water, dried and frozen in liquid nitrogen prior grinding. Powder was resuspended in 2 mL of RIPA buffer (50 mM Tris-HCl, pH 7.5, 1 mM EDTA, 1% Nonidet P-40, 1% sodium-deoxycholate) and centrifuged two times 10 min at 14000 g to remove cell debris. 50 µL of antibodies raised against GFP coupled to magnetic beads (Miltenyi Biotec®) were added to the supernatant and mixed (wheel rotation) for 30 min at 4°C. Tubes were then placed on magnetic rack and after 4 washes with RIPA buffer, proteins were eluted with 100 µL of 1X Laemmli solution (65 mM Tris-HCl pH 7.5, 5% glycerol, 2% SDS, 125 mM DTT). These manipulations were done on 4 biological replicates per genotype.

Mass spectrometry analysis

To analyze co-IP samples by mass spectrometry, eluted proteins were loaded on a 10% precast acrylamide gel (biorad) for a short run (15 min, 100 kV). The whole band was manually excised from the gel and cut in small pieces. After sequential washes with 25 mM ammonium bicarbonate, 50% acetonitrile in 25 mM ammonium bicarbonate and 100% acetonitrile, thiol groups of cysteines were reduced with 10 mM DTT for 45 min and alkylated for 30 min with 55 mM iodoacetamide. The bands were then sequentially washed with 50% acetonitrile in 25 mM ammonium bicarbonate and with 100% acetonitrile. The proteins were then digested with 0.25 µg of trypsin (Sequencing Grade Modified, Promega) in 25 mM ammonium bicarbonate overnight at 37°C. Peptides were eluted first with 2% formic acid and twice with 80% acetonitrile in 2% formic acid. Supernatants were pooled and evaporated in a vacuum centrifuge. Peptides were resuspended in 8 µL of 2% formic acid and 6 µL were injected for LC-MS/MS analyses. They were performed using an Ultimate 3000 RSLC nano system (Thermo Fisher Scientific Inc, Waltham, MA, USA) interfaced online with a nano easy ion source and a Q Exactive Plus Orbitrap mass spectrometer (Thermo Fisher Scientific Inc, Waltham, MA, USA). The samples were analyzed in Data Dependent Acquisition (DDA). Protein digests were first loaded onto a pre-column (Thermo Scientific PepMap 100 C18, 5 µm particle size, 100 Å pore size, 300 µm i.d. x 5 mm length) at a flow rate of 10 µL/min for 3 min.

The peptides were separated on a reverse-phase column (Thermo Scientific PepMap C18, 2 µm particle size, 100 Å pore size, 75 µm i.d. × 50 cm length) at a flow rate of 300 nL/min. Loading buffer (solvent A) was 0.1% formic acid in water and elution buffer (solvent B) was 0.1% formic acid in 80% acetonitrile. The linear gradient employed was 2-25% of solvent B for 103 min, then 25-40% of solvent B from 103 to 123 min, finally 40-90% of solvent B from 123 to 125

min. The total run time was 150 min including a high organic wash step and re-equilibration step. The Q Exactive Plus mass analyzer was operated in positive ESI mode at 1.8 kV. In DDA, the top 10 precursors were acquired between 375 and 1500 m/z with a 2 Th (Thomson) selection window, dynamic exclusion of 40 s, normalized collision energy (NCE) of 27 and resolutions of 70,000 for MS and 17,500 for MS2. Raw mass spectrometric data were analyzed in the Maxquant environment (69) v.1.5.0.0 and Andromeda was employed for database search (70). The MS/MS data were matched against the TAIR10 + GFP database (35417 entries). The “Trypsin/P” criterion was chosen as digestion enzyme. Up to two missed cleavages were allowed for protease digestion. For protein identification and quantification, cysteine carbamidomethylation was set up as a fixed modification and oxidation of methionine as a variable modification. Mass tolerance for precursor ions was 20 and 4.5 ppm for the first and the main searches respectively, and it was 20 ppm for the fragment ions. At least two peptides are necessary for protein identification and quantification. A peptide-spectrum match (PSM) false discovery rate (FDR) and a protein FDR below 0.01 were required. Using the above criteria, the rates of false peptide sequence assignment and false protein identification were lower than 1%. For the other characteristics, Maxquant default parameters were used. Intensities without normalization were considered and a protein was considered as a putative interactant if it was found in at least 3 replicates in *ProNFU1::NFU1g-GFP* lines and not in 3 or 4 replicates of *ProNFU3::GFP* lines. The values of Pearson correlation coefficients calculated between each biological replicate, were between 0.75 and 0.85. The mass spectrometry proteomics data have been deposited to the ProteomeXchange Consortium via the PRIDE (71) partner repository with the dataset identifier PXD015295.

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Conflict of interest
The authors declare no conflict of interest.

Author contributions
MR, JPT, FV, NB, TA, LC, HCW, TD performed the experiments, FV, NB, VS, MKJ, CD, JC, NR have conceived and supervised the experiments. All authors have read and approved the manuscript, the initial draft having been written by MR, JC and NR.

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Figure legends

Figure 1. Spectroscopic and oligomeric state characterization of reconstituted Arabidopsis NFU1.
UV-visible absorption and CD spectra (A), resonance Raman spectrum (B) and analytical gel filtration studies (C) of reconstituted Arabidopsis NFU1. The ε and Δε values for the UV-visible absorption and CD spectra (A) are based on NFU1 monomer concentration. The resonance Raman spectrum (B) was recorded using a droplet of NFU1 (~2 mM in clusters) frozen at 17 K, using 457.9 nm laser excitation. The spectrum is the sum of 100 scans with each scan involving counting protons for 1 s every 0.5 cm^{-1}, with 7 cm^{-1} spectral resolution. Bands due to the frozen buffer solution have been subtracted. For analytical gel filtration (C), apo-NFU1 (grey line) and reconstituted holo-NFU1 (black line) proteins were loaded onto a Sephadex S200 10/300 column. Absorbance of the eluted fractions was recorded at 280 nm (solid line) and 420 nm (dashed line), a wavelength characteristic of Fe-S cluster absorption. The apparent molecular masses of apo- and holo-NFU1 were determined from the elution volumes relatively to those of standard proteins, as described in the experimental procedures.

Figure 2. Binary Y2H assays between Arabidopsis NFU1 and SUF components.
The co-transformed yeast cells were plated at an OD_{600} of 0.05 on a control plate containing histidine (+His) and on test plates without histidine (-His) and containing 2 or 5 mM 3-amino triazol (3AT). Yeast cells were grown for 5 days at 30°C. Empty pGAD/pGBK-NFU1 and pGAD-NFU1/empty pGBK co-transformed yeast cells do not grow without histidine (not shown). The interaction between NFU1 and SUFD is not visible when the other combination of chimeric constructs is used.

Figure 3. Binary Y2H assays between Arabidopsis NFU1 and putative client proteins known to incorporate [4Fe-4S] clusters.
The co-transformed yeast cells were plated at an OD_{600} of 0.05 on a control plate containing histidine (+His) and on test plates without histidine (-His), eventually containing 2, 5 or 10 mM 3-amino triazol (3AT). Yeast cells were grown for 5 days at 30°C. Empty pGAD /pGBK-NFU1 co-transformed yeast cells do not grow without histidine (not shown).

Figure 4. BiFC assays between Arabidopsis NFU1 and its putative interactors in Arabidopsis leaf protoplasts.
Arabidopsis protoplasts were transfected with combinations of two vectors expressing NFU1 with itself (A) or with its potential interactors upstream of the N- or C-terminal halves of YFP (B). Results shown are representative of at least two independent transfection assays for at least 20 cells per transfection. Scale bar: 10 µm. In A, the replacement of one of these constructs by an empty vector as negative controls gives no signal. In B, NFU1-C was assayed individually with THIC-N, ISPG-N, SUFA1-N, SUFE3-N and cLIP-N. Negative controls using an empty-C vector are shown separately in Fig. S4. The mid values of argon laser intensities used are indicated. Protoplast transfections with vectors expressing the same combinations of proteins fused to the other YFP halves (NFU1-N and selected protein-C fusions) provided the same results with similar laser intensities (not shown).

Figure 5. *In vitro* Fe-S cluster transfer from holo-NFU1 to apo-ISPG monitored by UV-visible absorption spectroscopy.
The transfer reaction was initiated by mixing two molar equivalents of holo-NFU1 (50 or 100 µM) relative to a reduced apo-ISPG (25 or 50 µM). After 1h incubation, the untagged NFU1

was separated from the His-tagged ISPG on IMAC. Insert at the top shows an SDS-PAGE of the unbound (U) and eluted (E) fractions. M: molecular weight marker.
In A, UV-visible absorption spectra of IscS-reconstituted holo-NFU1 prior (dashed line) and after incubation with apo-ISPG and IMAC separation (solid line). In B, UV-visible absorption spectra of apo-ISPG prior (dashed line) and after incubation with holo-NFU1 and IMAC separation (solid line). The ε values are based on NFU1 and ISPG monomer concentrations.

Figure 6. *In vitro* Fe-S cluster transfer from holo-NFU1 to apo-THIC monitored by UV-visible absorption spectroscopy.
The transfer reaction was initiated by mixing two molar equivalents of holo-NFU1 (50 or 100 µM) relative to a reduced apo-THIC (25 or 50 µM). After 1h incubation, the untagged NFU1 was separated from the His-tagged THIC on IMAC. Insert at the top shows an SDS-PAGE of the unbound (U) and eluted (E) fractions. M: molecular weight marker.
In A, UV-visible absorption spectra of IscS-reconstituted holo-NFU1 prior (dashed line) and after incubation with apo-ISPG and IMAC separation (solid line). In B, UV-visible absorption spectra of apo-THIC prior (dashed line) and after incubation with holo-NFU1 and IMAC separation (solid line). The ε values are based on NFU1 and THIC monomer concentrations.

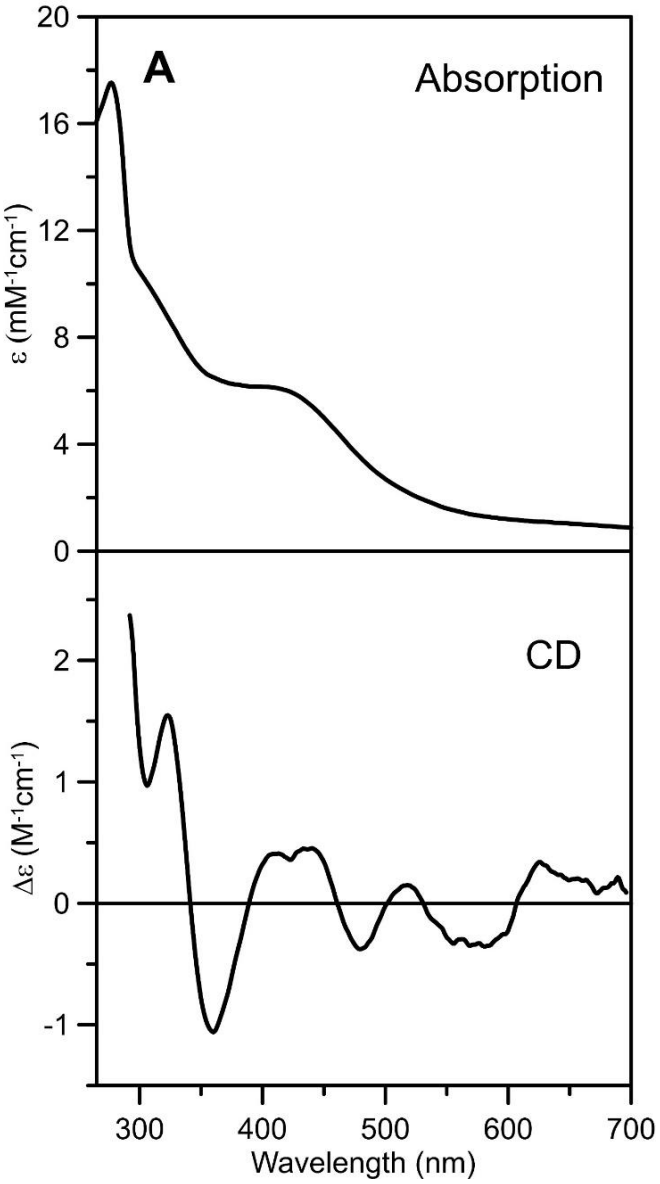
Figure 7. Positioning of NFU1 and its partner proteins in the current model of the SUF machinery.
The previous model (23) for the plastidial SUF machinery was implemented with recent results including those reported in this paper for NFU1. The approaches used for defining these interactions are depicted with a color code, as visible on the scheme. A question mark remains for the interaction between NFU1 and IPMI large subunit, because we could not firmly establish it. For the sake of clarity, other candidate SUF components *i.e.*, BOLA1/4, GRXS14/16 and IBA57.2 have not been represented here because their position is unclear.

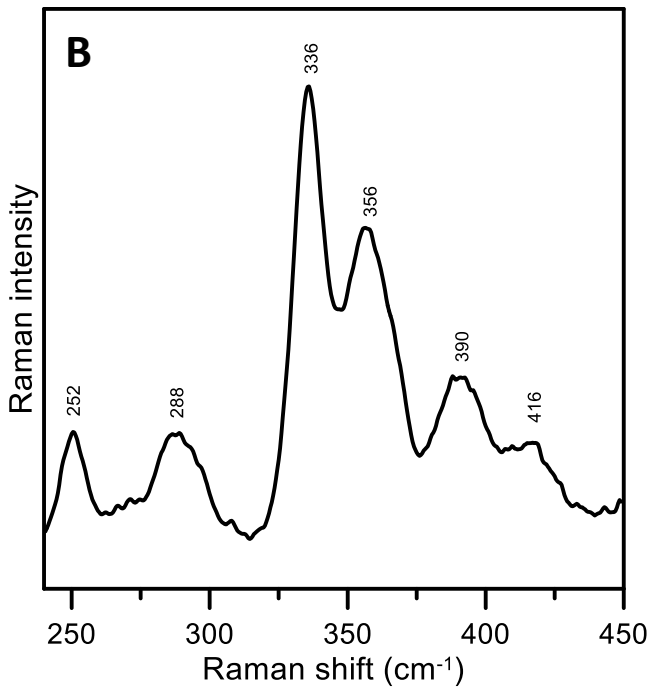
Gene IDs	moy_intensity *Pro35S::CTP NFU3-GFP*	moy_intensity *ProNFU1::gNFU1-GFP*	Number of peptides	Symbol	Protein name	Function	Localization	Fe-S cluster type	Sequence coverage [%]
AT5G50210	0,00	1,87E+07	8	SUFE3	quinolinate synthase SUFE3	NAD synthesis	plastid	4Fe-4S	17.3
AT1G62180	0,00	1,09E+07	7	APR2	5'adenylyl-phosphosulfate reductase 2	sulfate assimilation	plastid	4Fe-4S	30
AT2G29630*	7,80E+04	1,05E+07	5	THIC	thiamin C biosynthesis	thiamine biosynthesis	plastid	4Fe-4S	11.3
AT2G41220*	1,17E+05	1,10E+06	8	GLU2	glutamate synthase 2	nitrogen assimilation	plastid	3Fe-4S	6.4
AT2G38270*	8,86E+04	2,64E+06	2	GRXS16	glutaredoxin S16	Fe-S cluster assembly machinery	plastid	2Fe-2S, 4Fe-4S	16.7
AT3G24430*	1,61E+05	1,83E+07	9	HCF101	high-chlorophyll fluorescence 101	Fe-S cluster assembly machinery	plastid	4Fe-4S	27.3
AT2G43100	0,00	6,92E+06	6	IPMI2	isopropylmalate isomerase 2	glucosinolate and leucine biosynthesis	plastid	none	29.3
AT3G58990	0,00	5,76E+06	4	IPMI1	isopropylmalate isomerase 1	glucosinolate and leucine biosynthesis	plastid	none	23.3
AT2G43090*	1,54E+05	1,18E+07	8	IPMI3	isopropylmalate isomerase 3	glucosinolate and leucine biosynthesis	plastid	none	42.6
AT5G10920	0,00	1,64E+06	2	none	argininosuccinate lyase	amino acid biosynthesis (arginine)	plastid	none	6.6
AT1G58080	0,00	8,08E+06	3	ATP-PRT1	ATP phosphoribosyl transferase 1	amino acid biosynthesis (histidine)	plastid	none	12.9
AT5G05590	0,00	1,40E+06	3	PAI2	phosphoribosylanthranilate isomerase 2	amino acid biosynthesis (tryptophan)	plastid	none	18.2
AT4G14210	0,00	7,17E+06	3	PDS3	phytoene desaturase 3	carotenoid biosynthesis	plastid	none	6.4
AT5G59370	0,00	2,63E+07	6	ACT4	actin4	cellular architecture	cytosol	none	17
AT4G16390	0,00	1,14E+07	9	SVR7	suppressor of variegation 7	chloroplast biogenesis	plastid	none	15.7
AT1G69200	0,00	1,54E+06	2	FLN2	fructokinase-like 2	chloroplast organization	plastid	none	4.9
AT1G79050	0,00	5,65E+06	6	RECA1	homolog of bacterial RecA	DNA metabolism	plastid	none	19.6
AT2G43710	0,00	1,19E+06	5	SSI2	suppressor of SA insensitive 2	fatty acid desaturation	plastid	none	18.7
AT4G15560	0,00	1,44E+06	2	DXS	1-deoxy-D-xylulose 5-phosphate synthase	isoprenoid biosynthesis	plastid	none	5.9
AT4G21210	0,00	8,84E+06	5	RP1	PPDK regulatory protein 1	kinase activity	plastid	none	16.1
AT1G70070	0,00	1,65E+06	3	ISE2	increased size exclusion limit 2	plasmodesmata formation	plastid, cytosol, nucleus	none	4
AT2G21280	0,00	4,09E+06	2	GC1	giant chloroplast 1	plastid division	plastid	none	7.8

AT5G24020	0,00	1,54E+06	2	ARC11	accumulation and replication of chloroplasts 11	plastid division	plastid	none	7.1
AT1G80480	0,00	1,49E+07	7	PTAC17	plastid transcriptionally active 17	plastid gene expression	plastid	none	26.1
AT4G13670	0,00	3,57E+07	10	PTAC5	plastid transcriptionally active 5	plastid gene expression	plastid	none	39.5
AT5G13630	0,00	7,04E+06	5	GUN5	genomes uncoupled 5	plastid-to-nucleus signal transduction	plastid	none	6.1
AT1G02560	0,00	1,96E+07	4	CLPP5	nuclear encoded CLP protease 5	protease	plastid	none	15.4
AT4G20960	0,00	3,19E+06	4	PYRD	pyrimidine deaminase	riboflavin biosynthesis	plastid	none	16
ATCG00810	0,00	3,44E+06	2	RPL22	ribosomal protein L22	ribosomal protein	plastid	none	17.5
AT1G78630	0,00	9,60E+06	2	emb1473	embryo defective 1473	ribosomal protein	plastid, cytosol, mitochondrion	none	13.3
AT3G12930	0,00	4,72E+06	2	DG238	delayed greening 238	ribosome availability	plastid	none	10.9
AT3G62910	0,00	2,19E+06	3	CPRF1	chloroplast ribosome release factor 1	ribosome availability	plastid	none	10.4
ATCG00180	0,00	1,48E+06	2	RPOC1	RNA polymerase beta' subunit-1	RNA polymerase	plastid	none	3.8
ATCG00190	0,00	2,78E+06	3	RPOB	RNA polymerase subunit β	RNA polymerase	plastid	none	3.8
ATCG00740	0,00	2,87E+06	3	RPOA	RNA polymerase subunit α	RNA polymerase	plastid	none	10.9
AT5G13030	0,00	5,78E+06	7	SELO	selenoprotein O	ROS regulation	plastid	none	12
AT1G32900	0,00	5,81E+06	8	GBSS1	granule bound starch synthase 1	starch biosynthesis	plastid	none	19.7
AT5G24300	0,00	7,06E+06	5	SS1	starch synthase 1	starch biosynthesis	plastid	none	10.1
AT5G43780	0,00	2,67E+06	7	ATPS4	ATP sulfurylase 4	sulfate assimilation	plastid	none	19
AT3G21200	0,00	2,47E+06	2	PGR7	proton gradient regulation 7	tetrapyrrole biosynthesis	plastid	none	11.4
AT1G22940	0,00	5,07E+05	2	TH1	thiamine requiring 1	thiamine biosynthesis	plastid	none	5
AT3G06730	0,00	2,07E+06	2	TRXZ	thioredoxin Z	thioredoxin	plastid	none	15.8
AT4G29670	0,00	4,71E+06	2	ACHT2	atypical cys his rich thioredoxin 2/TRX-lilium2	thioredoxin	plastid	none	16.6
AT5G65840	0,00	6,78E+06	4	none	none	thioredoxin	plastid	none	14.9
AT2G42220	0,00	8,84E+06	5	STR9	sulfurtransferase 9	thiosulfate metabolism	plastid	none	26.1
AT3G08920	0,00	3,14E+06	3	STR10	sulfurtransferase 10	thiosulfate metabolism	mitochondrion	none	17.8
AT5G19370	0,00	1,46E+07	9	STR12	sulfurtransferase 12	thiosulfate metabolism	plastid	none	37.8
AT4G27700	0,00	8,26E+06	4	STR14	sulfurtransferase 14	thiosulfate metabolism	plastid	none	24.1
AT1G03160	0,00	1,58E+06	3	FZL	FZO-like	thylakoid organization	plastid	none	4.8

AT4G33760	0,00	3,88E+06	4	OKI	okina kuki	tRNA synthesis	plastid, cytosol, mitochondrion	none	8.9
AT1G23180	0,00	1,97E+06	3	none	ARM repeat superfamily protein	unknown	plastid, nucleus	none	6.6
AT3G62530	0,00	7,67E+06	4	none	ARM repeat superfamily protein	unknown	plastid, mitochondrion	none	23.5
AT2G13440	0,00	1,59E+06	2	none	glucose-inhibited division family A protein	unknown	plastid, mitochondrion	none	4.8
AT5G07190	0,00	5,14E+06	3	ATS3	seed gene 3	unknown	extracellular	none	27.6

Table 1. Potential NFU1 partners obtained with co-immunoprecipitation experiments and their assigned or presumed function.
Proteins were considered as interactors if they were identified in coIP experiments in at least 3 replicates using ProNFU1::gNFU1-GFP lines and not identified in the 4 replicates using Pro35S::CTPNFU3-GFPlines. The five hits labelled with an asterisk are Fe-S proteins or Fe-S associated proteins found in only one replicate using Pro35S::CTPNFU3-GFPlines. Additional non Fe-S proteins selected using this criterion are listed in Table S1.





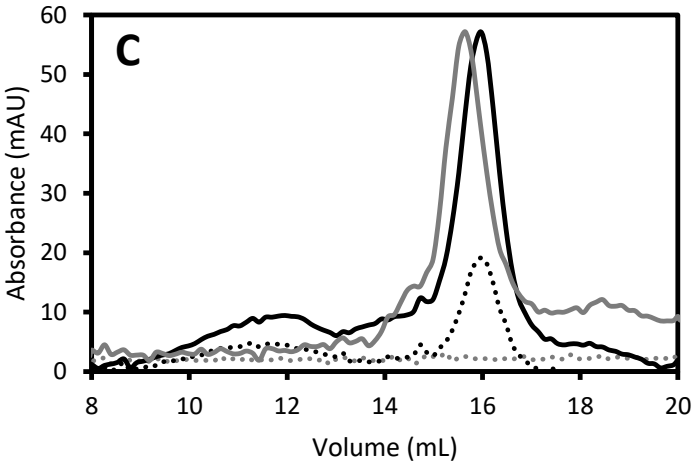


Figure 1

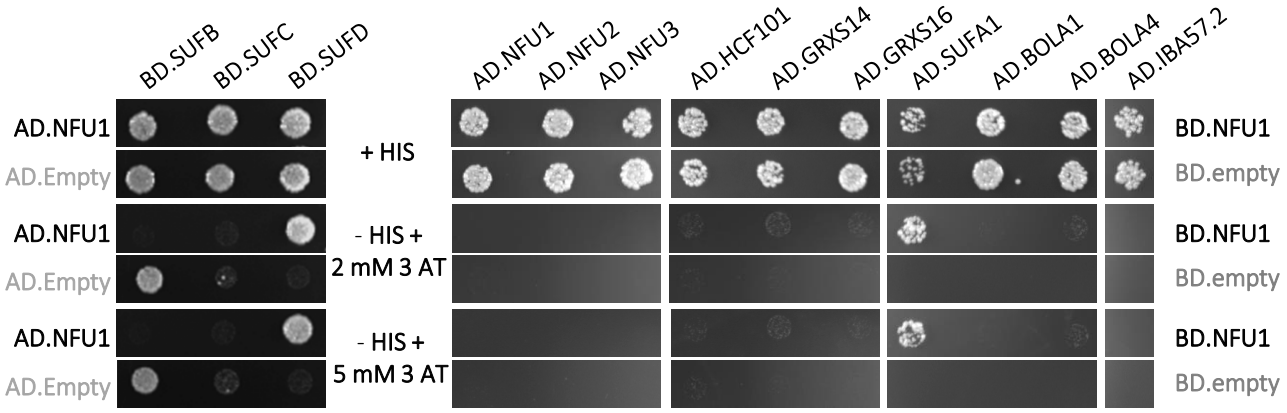


Figure 2

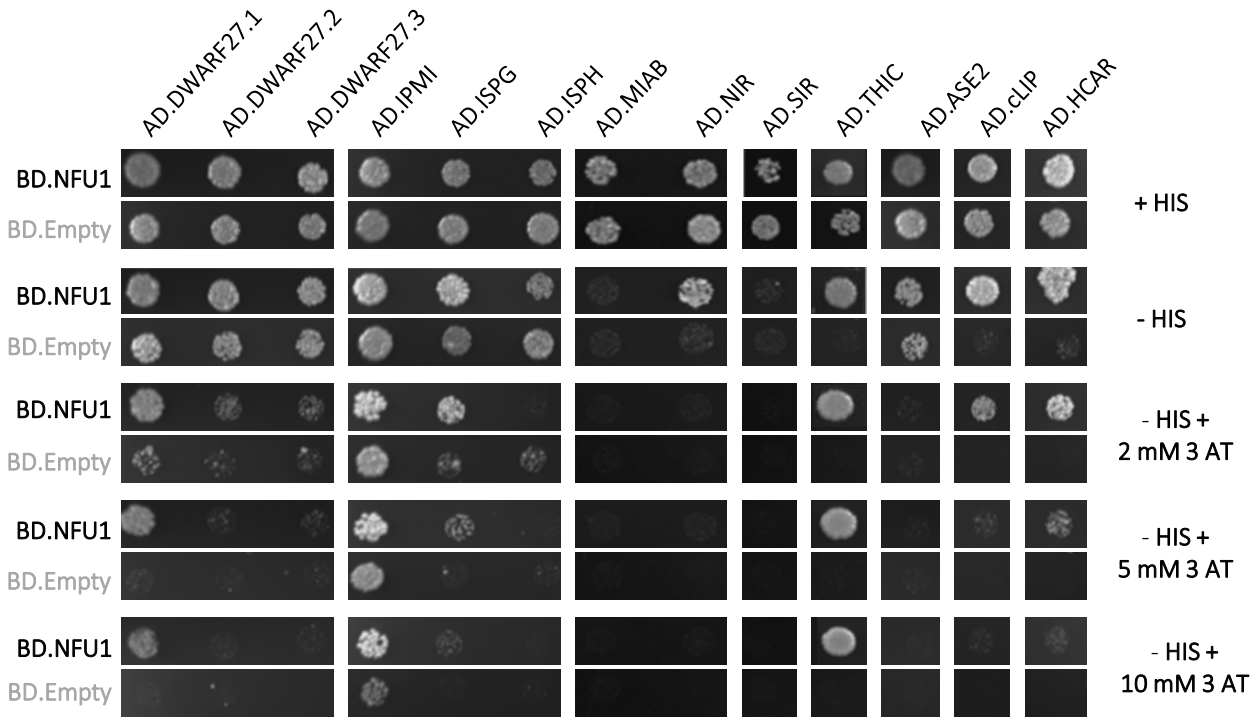


Figure 3

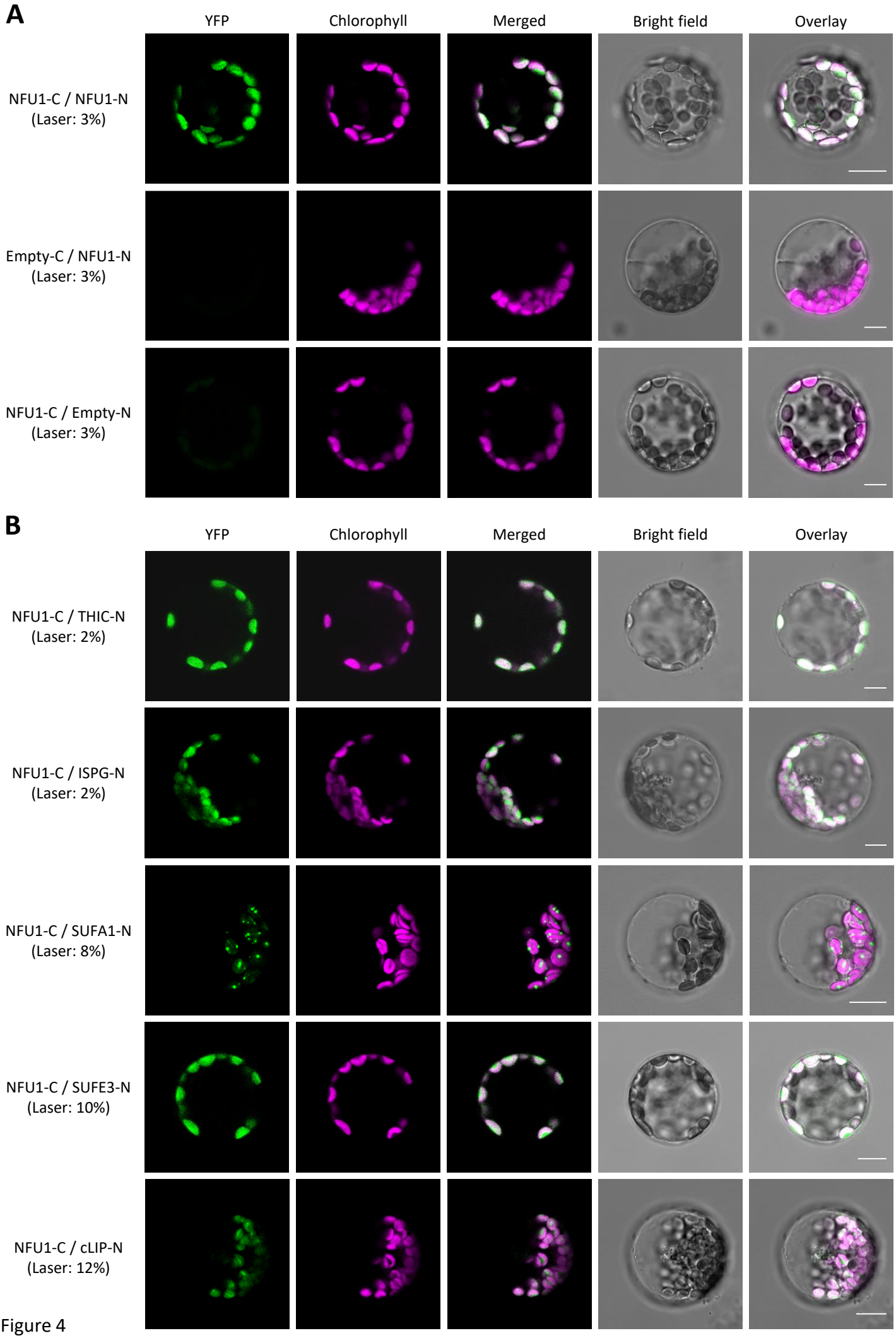


Figure 4

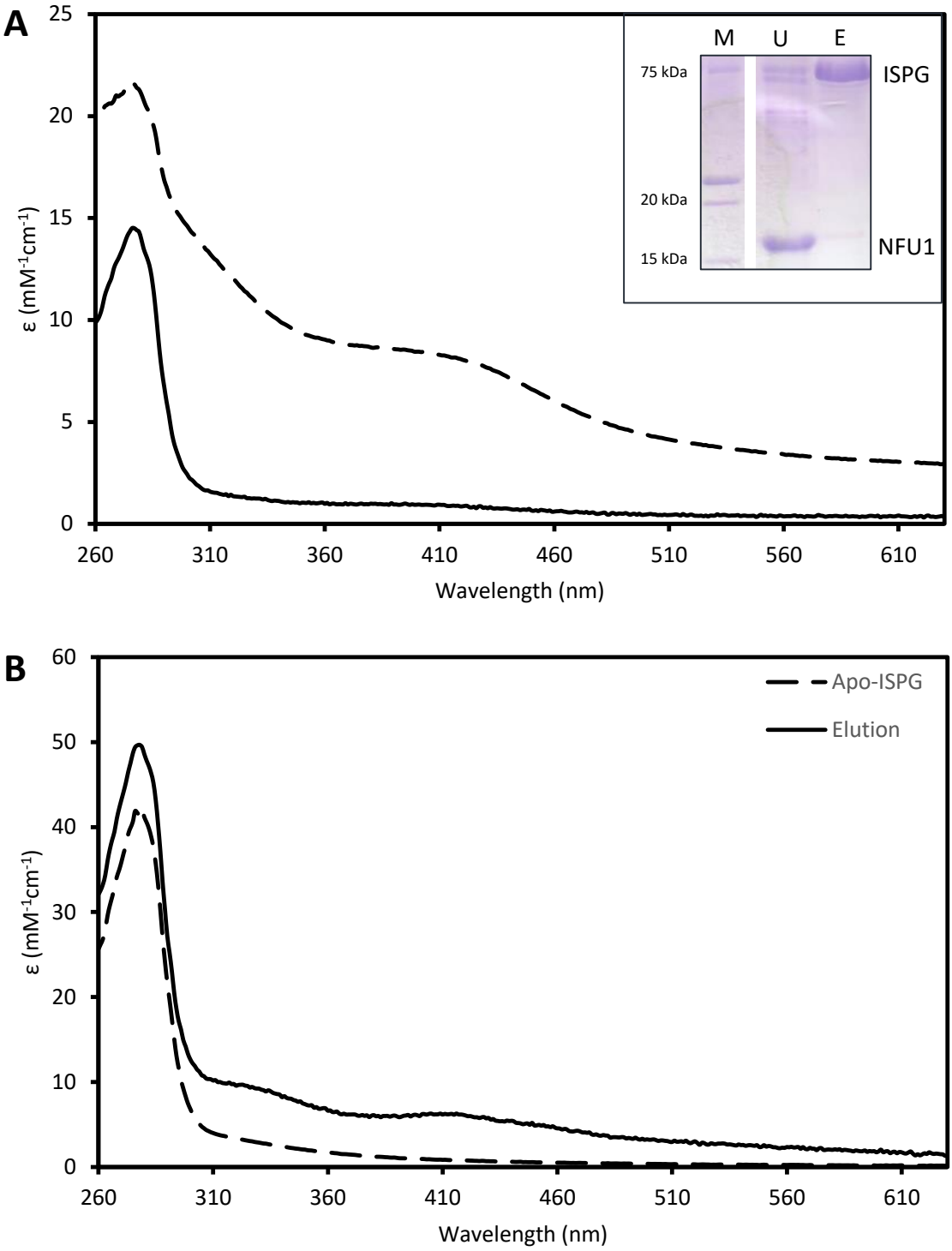


Figure 5

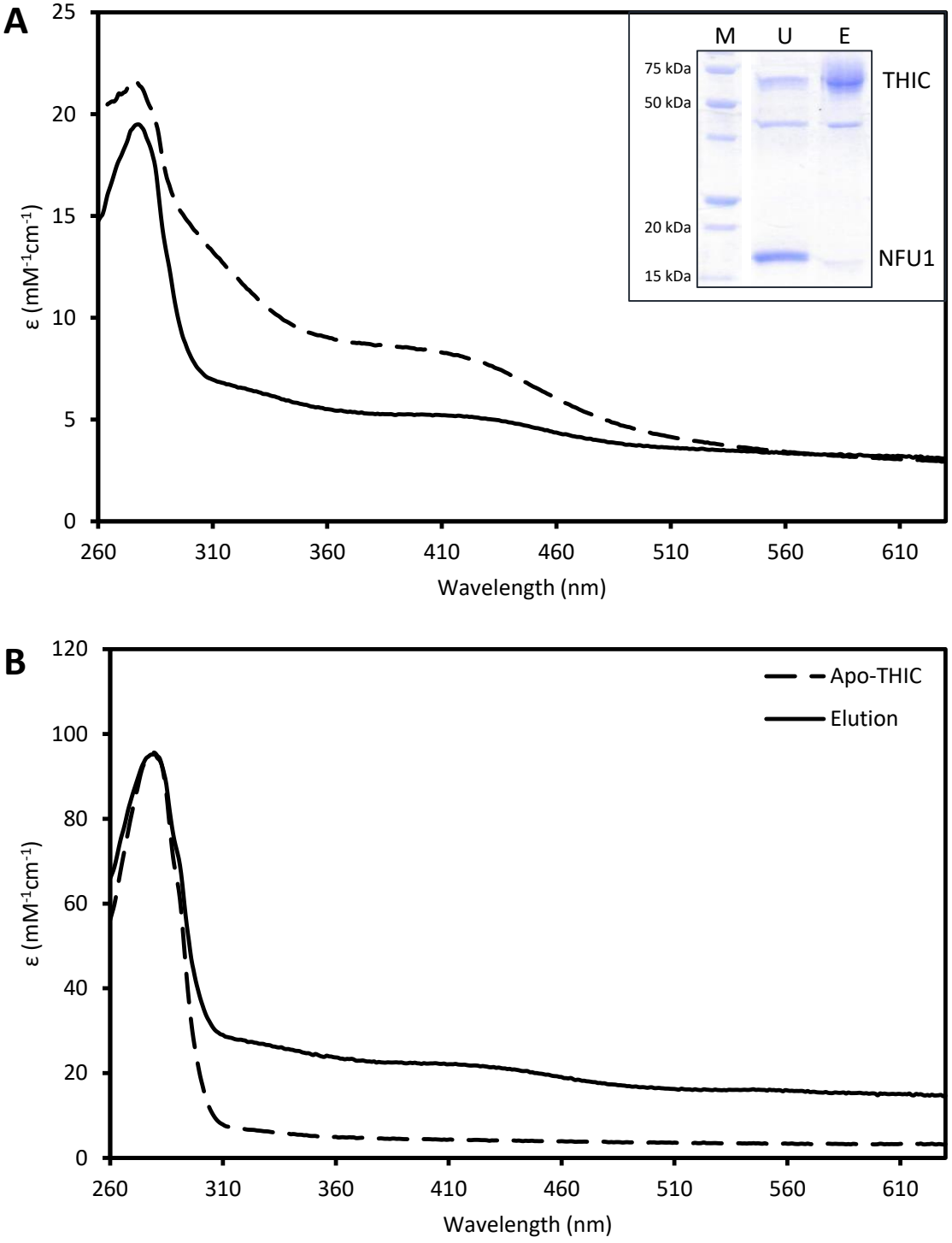


Figure 6

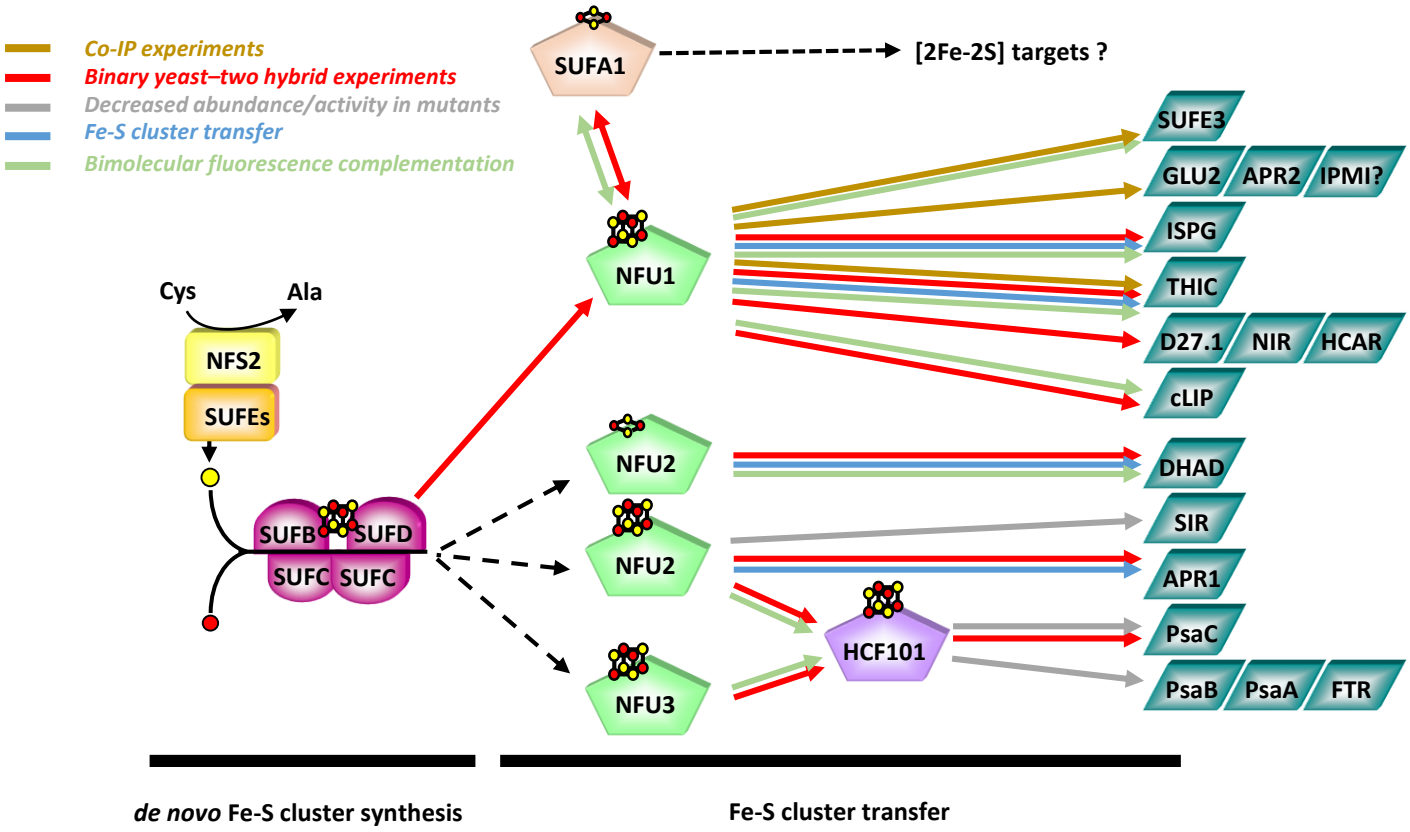


Figure 7

The plastidial Arabidopsis thaliana NFU1 protein binds and delivers [4Fe-4S] clusters to specific client proteins

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