



HAL
open science

Peripheral Routes to Neurodegeneration: Passing Through the Blood–Brain Barrier

Patrizia Giannoni, Sylvie Claeysen, Francesco Noe, Nicola Marchi

► **To cite this version:**

Patrizia Giannoni, Sylvie Claeysen, Francesco Noe, Nicola Marchi. Peripheral Routes to Neurodegeneration: Passing Through the Blood–Brain Barrier. *Frontiers in Aging Neuroscience*, 2020, 12, pp.3. 10.3389/fnagi.2020.00003 . hal-02465771

HAL Id: hal-02465771

<https://hal.science/hal-02465771v1>

Submitted on 19 Feb 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License



Peripheral Routes to Neurodegeneration: Passing Through the Blood–Brain Barrier

Patrizia Giannoni¹, Sylvie Claeyssen², Francesco Noe³ and Nicola Marchi^{2*}

¹ Laboratoire CHROME (EA 7352), Université de Nîmes, Nîmes, France, ² CNRS, INSERM U1191, Institut de Génétique Fonctionnelle, University of Montpellier, Montpellier, France, ³ HiLIFE – Neuroscience Center, University of Helsinki, Helsinki, Finland

A bidirectional crosstalk between peripheral players of immunity and the central nervous system (CNS) exists. Hence, blood–brain barrier (BBB) breakdown is emerging as a participant mechanism of dysregulated peripheral–CNS interplay, promoting diseases. Here, we examine the implication of BBB damage in neurodegeneration, linking it to peripheral brain-directed autoantibodies and gut–brain axis mechanisms. As BBB breakdown is a factor contributing to, or even anticipating, neuronal dysfunction(s), we here identify contemporary pharmacological strategies that could be exploited to repair the BBB in disease conditions. Developing neurovascular, add on, therapeutic strategies may lead to a more efficacious pre-clinical to clinical transition with the goal of curbing the progression of neurodegeneration.

OPEN ACCESS

Edited by:

Daniel Ortuño-Sahagún,
Universidad de Guadalajara, Mexico

Reviewed by:

Annadora Bruce-Keller,
Pennington Biomedical Research
Center, United States
Berislav Zlokovic,
University of Southern California,
United States

*Correspondence:

Nicola Marchi
nicola.marchi@igf.cnrs.fr

Received: 30 July 2019

Accepted: 08 January 2020

Published: 04 February 2020

Citation:

Giannoni P, Claeyssen S, Noe F
and Marchi N (2020) Peripheral
Routes to Neurodegeneration:
Passing Through the Blood–Brain
Barrier. *Front. Aging Neurosci.* 12:3.
doi: 10.3389/fnagi.2020.00003

Keywords: blood–brain barrier, neurodegeneration, peripheral immunity, traumatic brain injury, status epilepticus, autoantibodies, gut–brain axis, inflammation

BRAIN BARRIERS' PATHS, LEAKS, AND NEURODEGENERATION

The term neurodegenerative describes a progressive deterioration of the central nervous system (CNS) that is frequently associated with abnormal accumulation of proteins. Importantly, neurofibrillary tau-protein tangles are not only a major sign of Alzheimer's disease (AD) but are reported in temporal lobe epilepsy and post-traumatic encephalopathies (Tai et al., 2016). Among the emerging disease mechanisms, a peripheral–CNS pathological interplay is proposed to contribute to the neurodegenerative process (Marchi et al., 2014; Engelhardt et al., 2017; Fung et al., 2017; Pavlov and Tracey, 2017; Prinz and Priller, 2017; Le Page et al., 2018). Accordingly, harmful events occurring at the cerebrovascular interface are being examined as key determinants partaking to or even preceding neurodegeneration (Zlokovic, 2011; Nation et al., 2019; Sweeney et al., 2019). At the cerebrovasculature, specialized endothelial cells, mural cells, and astroglia constructs (Abbott et al., 2010; Giannoni et al., 2018; Sweeney et al., 2019) provide physical and biological properties governing the homeostatic–immune interactions between peripheral blood cells, or molecules, and brain neuroglia. The physiological parenchymal milieu composition ensures a healthy neuronal transmission, attainable because of the tightness of the blood–brain barrier (BBB; Zlokovic, 2008; Giannoni et al., 2018; Nation et al., 2019). At the pial arterial and venous level, the cerebrovasculature is permissive to blood cells or molecules, while it becomes impermeable at the arteriole–capillary level where barriers' properties are fully established (Abbott et al., 2010).

BBB vessels also contribute to cerebrospinal and interstitial fluid movements and the elimination of waste products from the interstitial and perivascular spaces (Noé and Marchi, 2019).

It is increasingly recognized that a BBB pathological imprint can provoke a brain pro-inflammatory disequilibrium sufficient to modify neuronal activity in the long term (Marchi et al., 2007, 2014; Nation et al., 2019). Vascular-dependent mechanisms of neurodegeneration can rapidly elicit as a consequence of peripheral infections, head trauma, ischemic stroke, or status epilepticus (**Figure 1**; Nation et al., 2019; Sweeney et al., 2019). These are risk factors for the development of long-term neurodegenerative sequelae and encephalopathies (e.g., post-concussion or head trauma-related chronic traumatic encephalopathy, CTE), cerebral amyloid angiopathy (CAA), AD, and epilepsy. Under conditions of increased BBB permeability, an aberrant bidirectional exchange between the neurovascular unit and the peripheral blood occurs, compounding to neurodegenerative modifications (**Figure 1**; Marchi et al., 2014; Engelhardt et al., 2017; Fung et al., 2017; Pavlov and Tracey, 2017; Prinz and Priller, 2017; Le Page et al., 2018). Completing a vicious cycle, beta-amyloid deposition in the brain can provoke capillaries dysfunction (Thomas et al., 1996; Zhang et al., 1997; Iadecola et al., 1999; Deane et al., 2003, 2012; Nortley et al., 2019). As an example, reactive oxygen species and endothelin-1 production were proposed to elicit vasoconstriction at pericyte locations (Nortley et al., 2019). A question remains regarding whether the endothelin-1 mechanism can directly drive neurodegeneration.

AUTOANTIBODIES AND NEURODEGENERATION: BAD, GOOD, OR NIL?

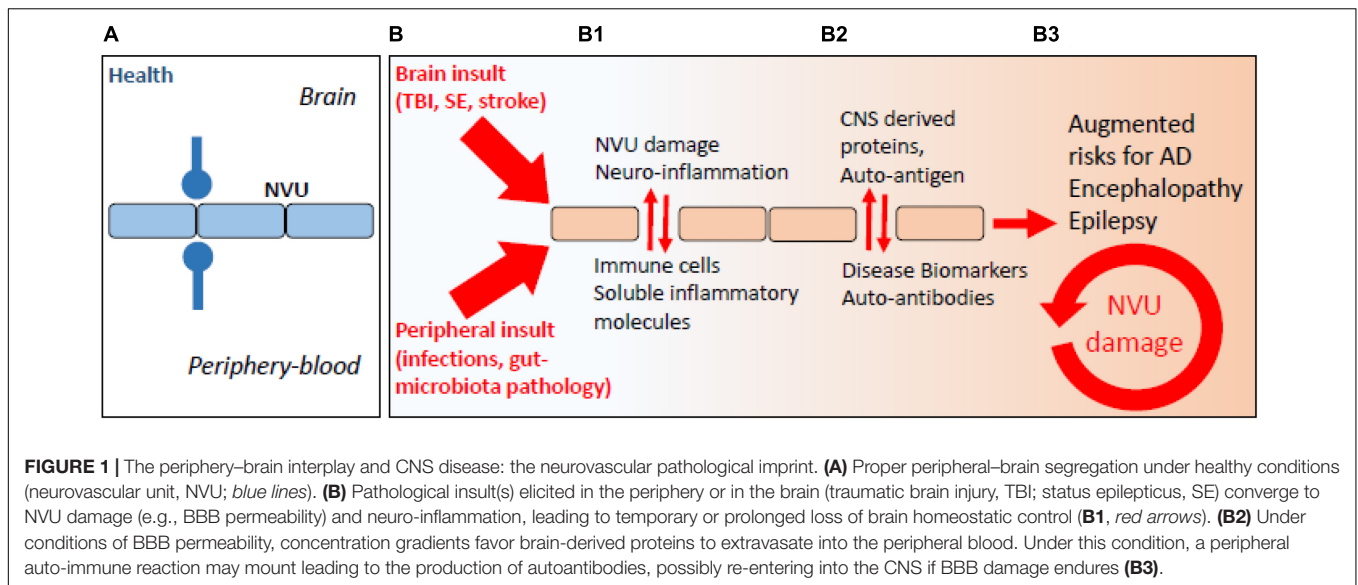
The communication between the peripheral blood and the brain occurs at preferential cerebrovascular sites (Zlokovic, 2011; Noé and Marchi, 2019), e.g., at post-capillary venules or pial vessels, and by a system of lymphatic vessels draining the cerebrospinal and interstitial fluids to cervical lymph nodes (Aspelund et al., 2015; Louveau et al., 2015a,b, 2018). At the intravascular level, moving leukocytes shape a peripheral–brain immune dialog where endothelium activation or permeability, perivascular immune cell homing, and brain entry of immune soluble factors prompt and sustain neuroglia inflammation [**Figure 1**; see Engelhardt et al. (2017) and Ransohoff (2016) for fundamental aspects of endothelial–leukocyte adhesion]. The implication of the cerebrovascular interface to innate and adaptive modalities of immunity is central (Schwartz and Shechter, 2010; Sommer et al., 2017). Adaptive immunity to the brain requires T- and B-cell stimulation at extra-CNS lymphatic organs and by professional antigen-presenting cells (Janeway et al., 2001), thus implying the existence of a peripheral–brain immune dialog, e.g., via the CNS vascular and lymphatic routes (Noé and Marchi, 2019).

A question exists on whether neurodegeneration may result from autoimmune-like processes (**Table 1**). Contingent to a prolonged or recurrent BBB permeability, specific antigens could exit the brain to reach the bloodstream, mounting a peripheral

humoral response. Newly formed brain-directed autoantibodies could be neuropathological upon their entry into the brain across a continuously damaged BBB (Levin et al., 2010). Importantly, autoantibodies and autoreactive T cells were reported in the cerebrospinal fluid (CSF), sera, as well as in the brain of AD patients and experimental models of disease (**Table 1**; Kronimus et al., 2016; Wu and Li, 2016). Anti-A β antibodies (Ig type G) correlated with scores of dementia (Wilson et al., 2009). Intrathecal antibodies against tau filaments were reported in AD patients (Mruthinti et al., 2004) and were proposed as contributors of disease progression (Bartos et al., 2012). Anti-tau autoantibodies are not specific to AD as they are increased in patients suffering from other neurodegenerative diseases, e.g., multiple sclerosis (Fialová et al., 2011).

The significance of peripheral autoantibodies as biomarkers of neurodegenerative conditions also remains to be established. Autoantibodies against the glutamate receptor *N*-methyl-D-aspartate receptor (NMDAR) were detected in plasma of AD patients (Davydova et al., 2007). Levels of antibodies were shown to correlate with clinical severity, as patients affected by moderate and severe dementia presented a twofold autoantibody increase compared with patients suffering from mild dementia (Davydova et al., 2007). The presence of autoantibodies against 5-HT was also reported (Myagkova et al., 2001), with levels increasing during the mild phase of the disease, subsequently reaching a plateau (Myagkova et al., 2001). Similar findings were reported for autoantibodies directed against the receptor for advanced glycation end products (Wilson et al., 2009). In a transgenic model of AD, autoantibodies against the sphingolipid ceramide correlated with amyloid plaque increase (Posse de Chaves and Sipione, 2010; Dinkins et al., 2015). Autoantibodies against ATP synthase (Vacirca et al., 2012), α (1)-adrenergic, and the β (2)-adrenergic receptors were also reported (Karczewski et al., 2012). Autoantibodies against the α (1)-adrenergic and the β (2)-adrenergic receptors may contribute to vascular lesions and increased plaque formation in AD patients (Karczewski et al., 2012).

Importantly, not all autoantibodies are harmful. Brain-reactive natural autoantibodies (NAbs) are protective (Britschgi et al., 2009; Kellner et al., 2009; Dodel et al., 2011; Bach and Dodel, 2012). NAbs are mostly IgM and are spontaneously produced. NAbs are polyreactive with low affinity for self-antigens (Casali and Schettino, 1996). Physiologically, NAbs facilitate phagocytosis of apoptotic cells, inhibit inflammatory pathways, and have a role in maintaining immune tolerance (Elkon and Silverman, 2012). NAbs to A β can inhibit plaque aggregation, block A β toxicity, and catalyze A β clearance (Lindhagen-Persson et al., 2010). Immunotherapies using specific, or aspecific, autoantibodies were tested. Bapineuzumab is the humanized form of a monoclonal anti-A β antibody targeting the N-terminus of A β . In phase II trials, Bapineuzumab administration reduced A β plaques in AD brains (Salloway et al., 2009; Rinne et al., 2010) and was associated with decreased total and phospho-tau levels in the CSF (Asuni et al., 2007). Bapineuzumab was, however, discontinued after a phase III trial and showed no beneficial effects on cognitive or functional outcomes (U.S. National Library of Medicine, 2019a,b). Aducanumab



(BIIB037) is a human monoclonal antibody selectively targeting aggregated A β (oligomers and fibrils) (Sevigny et al., 2016). An Aducanumab phase III trial was terminated as endpoints were not met. The analysis of a larger data set is ongoing. Tau immunotherapies are also being developed, attenuating or preventing functional impairment in experimental models, as reviewed in Sigurdsson (2018).

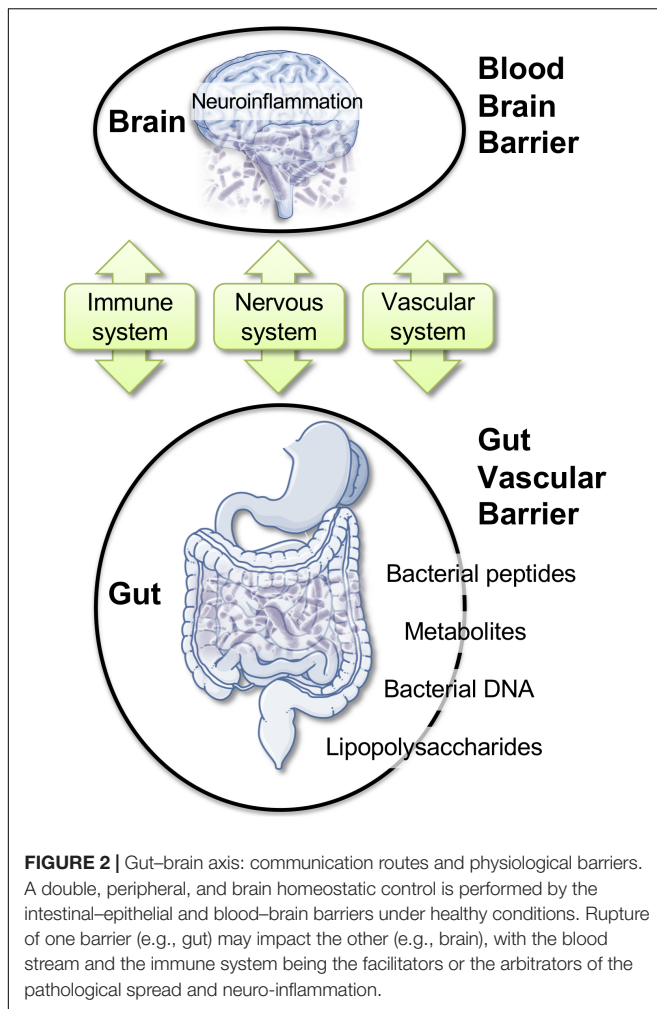
AUTOANTIBODIES AND POST-TRAUMATIC ENCEPHALOPATHY

Resulting from repeated head trauma and BBB damage, chronic traumatic encephalopathy (CTE) presents with accumulation of neurofibrillary tau-protein tangles. In TBI subjects, blood and CSF autoantibodies were suggested as etiological components or as possible biomarkers of neurodegeneration (Raad et al., 2014; Kobeissy, 2015; **Table 1**). Anti-glia fibrillary acidic protein (GFAP) fragments were found in the sera of TBI patients (Zhang et al., 2014). Serum autoantibodies against S100B were reported in American football players when repeated sub-concussive events were associated with BBB damage (Marchi et al., 2013). Autoantibodies against the neuronal $\alpha 7$ -subunit of the acetylcholine receptor (Sorokina et al., 2011) as well as AMPA and NMDA receptors (Goryunova et al., 2007) were detected in TBI subjects, while IgG autoantibodies to neurons and basal lamina were reported in rat serum following experimental head trauma (Rudehill et al., 2006). Autoantibodies to the pituitary gland were reported in TBI subjects 3 years after the trauma (Tanriverdi et al., 2008, 2010). Damage to the pituitary gland is distinctive of the TBI pathology with 20–50% of patients showing some degrees of pituitary dysfunction, which affects growth hormone production (Aimaretti et al., 2005; Tanriverdi et al., 2006). An association between anti-pituitary autoantibodies and pituitary dysfunction was reported in patients suffering from mild TBI, including repetitive concussions (Tanriverdi et al., 2010).

Autoreactive antibodies have been proposed for the treatment of TBI sequelae. The presence of hyper-phosphorylated tau accumulating in neurofibrillary tangles is a characteristic of CTE (Omalu et al., 2010). Even if phospho-tau is detectable only at low levels acutely after TBI (Smith et al., 2003; Blennow et al., 2012; Goldstein et al., 2012; Mannix et al., 2013), a specific form of phospho-tau can be produced in response to TBI (*cis* P-tau) (Kondo et al., 2015). This protein spreads throughout the brain, harming cells and leading to post-traumatic neurodegeneration and dementia. In two animal models of TBI, administration of a monoclonal antibody discriminating between the *cis* and the *trans* forms of the protein and blocking *cis* P-tau prevented the onset of tauopathy and cortical atrophy. These accumulating evidence supports the possible involvement of autoantibodies in post-TBI neurodegenerative conditions, perhaps providing new disease biomarkers and therapeutic entry points.

THE GUT–BRAIN AXIS AND NEURODEGENERATION: IS THERE A BARRIER IMPLICATION?

Here, we discuss a specific framework where alterations of the gut microbiota (GM) could impact BBB permeability, promote neuro-inflammation, and favor neurodegenerative modifications (**Figure 2**; Braniste et al., 2014; Cerovic et al., 2019; Parker et al., 2019; Wang et al., 2019). Bacteria, viruses, parasites, and non-pathogenic fungi constitute the intestinal microbiota. These complex communities of microbes colonizing the gastrointestinal tract are major players in health. Modern life and diets have progressively induced changes in the composition of the GM, perhaps for the worse, as this can contribute to chronic illnesses (Lozupone et al., 2012; Myles, 2014; Kumar and Forster, 2017; Shanahan et al., 2017; Cryan et al., 2019; Pagliai et al., 2019; Reza et al., 2019). Intestinal microbes can influence brain function through a continuous dialog involving the immune,



the vascular, and the nervous systems (Figure 2; Schroeder and Bäckhed, 2016; Cox and Weiner, 2018; Butler et al., 2019; Cryan et al., 2019). Modifications in the composition of the GM was reported in brain disorders, such as autism (Adams et al., 2011; Kang et al., 2019), depression (Kelly et al., 2016; Zheng et al., 2016), Parkinson's disease (Scheperjans et al., 2015; Sampson et al., 2016), and AD (Cattaneo et al., 2017; Vogt et al., 2017; Zhuang et al., 2018). Intriguingly, the extent of the amyloid pathology in AD mice appears to be dependent of the microbial status, which is specific to the animal housing facility. APP/PS1 mice bred in a germ-free facility displays decreased amyloid plaque number compared to mice housed in non-germ-free conditions (Harach et al., 2017). Moreover, the administration of broad-spectrum, combinatorial antibiotics to APP/PS1 mice, either during the peri-natal or the adult stage, reduced brain A β deposition (Minter et al., 2016, 2017).

Existing reports support the hypothesis of a possible infectious origin of AD. A β was proposed as an antimicrobial peptide (Soscia et al., 2010; Moir et al., 2018) responding to pathogens (Kumar et al., 2016; Eimer et al., 2018). Infectious agents, such as *Chlamydia pneumoniae*, *Propionibacterium acne*, *Helicobacter*

pylori, *Porphyromonas gingivalis*, or *spirochetes*, are associated with AD (Kornhuber, 1996; Balin et al., 1998; Kountouras et al., 2006; Miklossy, 2011; Poole et al., 2015). A microbial hypothesis is supported by evidence describing the presence of viruses, such as Herpes simplex virus type I, in the brains of AD patients (Lin et al., 2002; Alonso et al., 2014; Itzhaki et al., 2016).

Within the complex interplay between the gut microbiome and the CNS, a role for brain barriers and neuroinflammation is becoming important (Braniste et al., 2014; Cerovic et al., 2019; Parker et al., 2019; Wang et al., 2019). The impact of the gut microbiome composition on CNS health was reported (Amedei and Boem, 2018; Chu et al., 2019; Sherwin et al., 2019; Virtue et al., 2019). Recent work demonstrated that GM composition controls BBB development and permeability in mice (Braniste et al., 2014). In AD, increased gut permeability due to GM dysbiosis was reported during prolonged stress. In this condition, molecules that are normally secluded in the intestine, e.g., inflammatory mediators, bacteria, or bacterial-derived agents, could leak out and reach the peripheral blood. Bacterial DNA, metabolites, or proteins circulating in the blood stream could, in turn, modify BBB permeability (Braniste et al., 2014; Myles, 2014; Kumar and Forster, 2017; Cerovic et al., 2019; Parker et al., 2019; Wang et al., 2019). Existing reports indicated bacterial DNA in human blood with a possibility for brain access (Lelouvier et al., 2016; Paissé et al., 2016; Schierwagen et al., 2018). Brain entry of *P. gingivalis*, a bacterium associated with periodontal disease, has been described (Dominy et al., 2019). Gingipain inhibitors reduced the bacterial load and the bacteria-induced neuro-inflammation in a mouse model (Dominy et al., 2019). Among Spirochetes, *Borrelia burgdorferi* is a strain associated with Lyme dementia that could enter the brain. In humans, this specific strain can form biofilms similar to senile plaques. A β and bacterial DNA appear as important constituents of these biofilms, suggesting that amyloid plaques may originate in association with or from the spirochetal colonies (Allen, 2016; Miklossy, 2016).

These examples highlight the need of tightly regulated intestinal and brain barriers (Rahman et al., 2018). In AD, a dysbiotic GM may enhance gut permeability and alter BBB integrity, allowing the access of infectious agents or associated molecules into the brain (Martin et al., 2018). Significantly, intestinal and brain barriers are reactive to analogous pro-inflammatory triggers. Circulating inflammatory cytokines IL-17, interferon-gamma (IFN- γ), and the small intestine epithelium protein zonulin can damage the intestinal–epithelia and BBBs (Rahman et al., 2018).

GUT MICROBIOTA AND AUTOANTIBODIES: INITIAL CLUES

Hypotheses linking modifications of the GM and production of autoantibodies are emerging (Petta et al., 2018). Some evidence supports the concept that specific dietary components may affect B-cell maturation and activity, ultimately leading to the formation of autoantibodies (Petta et al., 2018). Obesity was associated with a systemic pro-inflammatory state, characterized by changes in the frequency of B-cell

TABLE 1 | Autoantibodies reported in neurodegenerative disease and post-TBI.

Autoantibodies	Neuro-pathology	Stage	Model investigated	Sample	Observed effects	Isotype	References
Anti-neuronal antibody	TBI	Moderate acute TBI	AM	Serum	–	IgG	Rudehill et al., 2006
Anti-neurofilament	AD	Moderate forms of AD	H	Serum, CSF	–	IgG, IgM	Bartos et al., 2012
Anti-A β	AD	Mild to severe forms of AD, early and late onset	H	Serum, CSF	Suggested to favor A β clearance; correlation with global scores of dementia	IgG {IgG2}, Nab	Myagkova et al., 2001; Bell et al., 2010; Daneman et al., 2010; Schwartz and Shechter, 2010; Armulik et al., 2017; Kisler et al., 2017; Rustenhoven et al., 2017; Sommer et al., 2017; Montagne et al., 2018; Nikolakopoulou et al., 2019
Anti-Tau	AD, TBI	Mid to severe forms of AD	H	Serum, CSF, tissue	Levels correlated with reduced Plaque burden	IgG, Nab	Du et al., 2001; Weksler et al., 2002; Mruthinti et al., 2004; Brettschneider et al., 2005; Rosenmann et al., 2006; Gruden et al., 2007; Gustaw et al., 2008; Britschgi et al., 2009; Wilson et al., 2009; Maffei et al., 2013; Qu et al., 2014
Anti-AMPA receptor	AD, TBI	Moderate to severe AD Mild and repetitive concussion in children	H	Serum	Levels increased in moderate and severe dementia	–	Goryunova et al., 2007
Anti-NMDA receptor	AD, TBI	Moderate to severe AD and dementia, mild and repetitive concussion	H	Serum	Relationship between autoantibody titers and aging	IgG	Goryunova et al., 2007; Busse et al., 2014
Anti-acetyl choline receptor	TBI	TBI to different severity in children	H	Serum	Levels correlate with trauma severity	–	Sorokina et al., 2011
Anti-Dopamine	AD	Mid to severe forms of AD	H	Serum	Match to moderate to severe dementia progression	IgG	Myagkova et al., 2001; Gruden et al., 2007
Anti-5-HT	AD	Mild to severe forms of AD	H	Serum	Levels increased during mild dementia	–	Myagkova et al., 2001
Anti-GFAP	AD, TBI	Pre-senile and senile forms of AD, senile vascular dementia	H	Serum	Relationship between autoantibody titers and aging suggested as a maker of BBB damage	IgG	Tanaka et al., 1989; Gruden et al., 2007

(Continued)

TABLE 1 | Continued

Autoantibodies	Neuro-pathology	Stage	Species model investigated	Sample	Observed effects	Isotype	References
Anti-S100 β	AD, TBI	Mild to severe AD, senile vascular dementia, repeated acute sub-concussion	H	Serum	Match to moderate-severe dementia progression; relationship between autoantibody titers and aging	IgG	Mecocci et al., 1995; Gruden et al., 2007; Marchi et al., 2013
Anti-microglia	AD	Mid to severe forms of AD	H	CSF	–	IgG	McRae et al., 2007
Anti-phospholipid	AD, TBI	Mid cognitive impairment to advanced AD, severe TBI	H	Serum, CSF	Levels correlate with erythrocytes and proteins in CSF	IgG	McIntyre et al., 2007; McIntyre et al., 2015
Anti-ceramide	AD	Chronic pathology in TG mice	AM	Serum	Levels correlate with plaque formation	IgG	Dinkins et al., 2015
Anti-RAGE	AD	Mild cognitive impairment to severe AD	H	Serum	Relationship with anti-A β levels; correlation with global scores of dementia	IgG	Mruthinti et al., 2004; Wilson et al., 2009
Anti-ATP synthase	AD	Mild to severe AD	H	Serum	Induced the inhibition of ATP synthesis	IgG	Vacirca et al., 2012
Anti-pituitary	TBI	Mild to severe TBI, acute and long-term	H	Serum	Association between antibody positivity and hypopituitarism due to head trauma	IgG	Tanriverdi et al., 2008; Tanriverdi et al., 2010; Pani et al., 2019
Anti-adrenergic receptors	AD	Mild to moderate dementia	H	Serum	Suggested contribution to vascular lesions and increased plaque formation	IgG	Karczewski et al., 2012

AM, data derives from animal models only.

TABLE 2 | Available molecules exerting BBB repairing and anti-inflammatory effects.

	Category	Mechanism(s) of action	Reported effects	<i>In vivo /in vitro</i> models	Clinical trials	References
Losartan	Antihypertensive	Angiotensin II antagonist	Improves barrier function	Rats	Antihypertensive drug	Kucuk et al., 2002; Kaya et al., 2003; Hong et al., 2019
Ripamycin	Immunosuppressant	mTOR antagonist	Improves barrier function, promotes claudin-5	Mice	Prevention of transplant rejection	Van Skike et al., 2018
Anakinra	Interleukin-1 receptor antagonist	Human interleukin-1 receptor antagonist (IL-1Ra)	Decreases inflammation	Rats	Anti-inflammatory drug currently used against rheumatoid arthritis cryopyrin-associated periodic syndromes (CAPS) and Still's disease.	Kenney-Jung et al., 2016; Van Skike et al., 2018
IPW	TGFβR1 kinase inhibitor	Inhibition of TGFβR signaling	Reduces brain hyperexcitability, indirect BBB repair	Mice	NA	Senatorov et al., 2019
3K3A-APC	Activated protein C (APC)	BACE-1 inhibition, activation of protease-activated receptor 1	Cytoprotective properties, neovascularization, neurogenesis	Mice	In clinical trial for ischemic stroke (RHAPSODY)	Thiyagarajan et al., 2008; Zhong et al., 2009; Wang et al., 2016; Sinha et al., 2018; Lazic et al., 2019; Lyden et al., 2019
PDGF-BB	PDGFRb agonist	Increased expression of p-Smad2/3	Ameliorates BBB function	<i>In vivo</i>	NA	Arango-Lievano et al., 2018
Imatinib	Kinase inhibitor	Inhibition of PDGFR signaling	anti-inflammatory?	Mice	Precursor cell lymphoblastic leukemia–lymphoma, dermatofibrosarcoma	Su et al., 2015; Klement et al., 2019
Dexamethasone	Glucocorticoid	Decreased JMJD3 gene expression, suppression of MMP-2, MMP-3, and MMP-9 gene activation	Preserves tight junctions integrity	<i>In vitro</i> BBB model	Inflammatory conditions	Hue et al., 2015; Na et al., 2017
Annexin-A1 (ANXA1)	Glucocorticoid anti-inflammatory effector	Binding to FRP2 receptor, inhibition of phospholipase-2	Restores cell polarity, cytoskeleton integrity, and paracellular permeability	<i>In vitro</i> BBB model, <i>Anxa</i> ^{−/−} mice	NA	Cristante et al., 2013; Purvis et al., 2019; Zub et al., 2019
VEGF	Vascular endothelial growth factor	Prevention of Aβ-induced apoptosis	Improves vascular functions	Mice	NA	Religa et al., 2013
Tetramethylpyrazine	Cardiovascular	Blocking JAK/STAT signaling	Reduces BBB damage	Rats	NA	Gong et al., 2019
S-nitrosoglutathione	Nitric oxide donor	Suppression of MMP-9 activity	Prevents BBB damage	Mice	NA	Aggarwal et al., 2015
Cannabidiol	Analgesic, anti-inflammatory, antineoplastic	Activation of PPARγ and 5-HT _{1A} receptors	Prevents BBB damage	<i>In vitro</i> BBB model	NA	Hind et al., 2016

subpopulation [e.g., reduction of the anti-inflammatory IL-10⁺ regulatory B cell (Nishimura et al., 2013)] and by an increase in autoantibody levels (Kosaraju et al., 2017). Diets rich in polyunsaturated fatty acid are associated

with the suppression of pro-inflammatory responses and a reduction of circulating autoantibodies (Pestka et al., 2014; Tomasdottir et al., 2014). Dietary components impact the composition of the gastrointestinal bacterial populations:

consumption of prebiotics increases the intestinal levels of *Bifidobacterium* and *Lactobacillus* (Singh et al., 2017), with a possible link to B-cell differentiation, maturation, and activation (Ouweland et al., 2002). Diet can impact autoantibody production, directly by promoting pro-inflammatory conditions and indirectly by altering the GM. In experimental autoimmune encephalomyelitis (EAE) it was demonstrated that the commensal microbiota composition is a pivotal factor for disease development (Lee et al., 2011) and that modifying the GM impacts the levels of T and B cells or the levels of circulating autoantibodies (Ochoa-Repáraz et al., 2009, 2010).

BBB REPAIRING PHARMACOLOGY: AVAILABLE OPTIONS

The multi-level implication of BBB damage in neurodegenerative disorders has prompted the quest for pharmacological repairing strategies, either directed at the endothelium or by indirect targeting of the cellular players of peripheral and neuro-inflammation. Currently tested drugs are either repurposed or new (Table 2). Examples include losartan, an anti-hypertensive molecule acting as an angiotensin II antagonist. Losartan was shown to reduce BBB permeability in a rat model of hypertension (Kucuk et al., 2002; Kaya et al., 2003) and following pilocarpine-induced status epilepticus (Hong et al., 2019). BBB protection by losartan depends on angiotensin receptor type 1 (AT1) blockade. Another drug is rapamycin, a specific inhibitor of the mammalian target of rapamycin (mTOR) pathway. Rapamycin improved cerebrovascular and cognitive function in a mouse model of AD (Van Skike et al., 2018). Inhibition of mTOR preserved BBB integrity through the upregulation of tight junction proteins and downregulation of matrix metalloproteinase-9. A third option is anakinra, which is the recombinant form of the human IL-1 receptor antagonist (IL-1Ra) that inhibits IL-1 α and IL-1 β binding to the IL-1 receptor type 1. As inflammation comprises BBB dysfunction, the inhibition of IL-1 as proposed is a strategy enabling cerebrovascular protection (Marchi et al., 2009, 2011; Vezzani et al., 2011; Kenney-Jung et al., 2016). Recent strategies include the development of IL-1Ra molecules fused with a cell-penetrating peptide to enhance brain access (Zhang et al., 2017). After transient middle cerebral artery occlusion in rats, IL-1Ra-PEP reduced neuro-inflammation and ischemia (Zhang et al., 2017). The fourth option is IPW-5371, a small molecule blocking the transforming growth factor β receptor (TGF β R) signaling. In a recent study (Senatorov et al., 2019), IPW reduced hyperexcitability in a mouse model, protecting BBB functions. The activated protein C (APC) therapeutic analog 3K3A-APC is a fifth option. This compound has BBB and neuro-protective properties (Thiyagarajan et al., 2008; Zhong et al., 2009; Wang et al., 2016; Sinha et al., 2018; Lazic et al., 2019; Lyden et al., 2019) and it is in clinical trial for stroke treatment (Lyden et al., 2019). Next is platelet-derived growth factor subunits BB (PDGF-BB). Following an acute vascular insult, activation of the PDGF receptor beta (PDGFR β) by

PDGF-BB is beneficial, protecting the endothelium–pericyte structures. The latter was reported in mouse models of status epilepticus (Arango-Lievano et al., 2018) and cerebral ischemia (Marushima et al., 2019). Conversely, in chronic disease settings (e.g., AD, epilepsy, etc.), activation of PDGFR β may participate to inflammation (Rustenhoven et al., 2017; Klement et al., 2019). Under this circumstance, blocking PDGFR β signaling by using the tyrosine kinase inhibitor Imatinib could represent an anti-inflammatory strategy (Rustenhoven et al., 2017; Klement et al., 2019). In general, reducing PDGFR β signaling could lead to contrasting effects, e.g., pericyte deficiency and BBB breakdown (Bell et al., 2010; Daneman et al., 2010; Armulik et al., 2017; Kisler et al., 2017; Montagne et al., 2018; Nikolakopoulou et al., 2019) or anti-inflammatory (Rustenhoven et al., 2017; Klement et al., 2019), depending on disease stage (acute vs. chronic). Another option, Dexamethasone, is a glucocorticoid effective in the formation and maintenance of endothelial tight junctions (Hue et al., 2015; Na et al., 2017). Dexamethasone was proposed to decrease the expression of the Jumonji Domain Containing 3 gene (JMJD3) and metallo-proteinases (MMP-2, MMP-3, and MMP-9). Finally, there is the vascular endothelial growth factor (VEGF). Amyloid accumulation is associated with endothelial apoptosis (Religa et al., 2013) in Alzheimer's patients as well as in mouse models. In AD mice, VEGF administration rescued memory deficits by preventing A β -induced vascular apoptosis (Religa et al., 2013). See Table 2 for complete drug listing, mechanisms and bibliography.

PERSPECTIVES AND CHALLENGES

The importance of cerebrovascular dysfunction in neurodegenerative disorders is twofold: BBB damage is pathophysiological and it allows a diagnostic window, the latter by exploiting specific proteins that shed from the damaged or vascular wall cells to appear into accessible fluids, e.g., blood or CSF. For instance, by dosing soluble PDGFR β in CSF and by using dynamic contrast-enhanced magnetic resonance imaging, a recent study demonstrated BBB breakdown as an early biomarker of human cognitive dysfunction (Montagne et al., 2015; Nation et al., 2019).

Tackling the complex neurodegenerative puzzle requires a continuous sharpening of pharmacological tools. This is important because no efficacious disease-modifying strategy is available to meaningfully delay or prevent disease progression. The problematics here presented may stem from semantic habits as the term *neuro-* indicates, for most, neurons only. Revisiting nomenclature(s) may benefit, if not legitimize, holistic, and neurovascular approaches to CNS disorders since it is evident that considering neuronal circuits insulated from the influence of glio-vascular cells is excessively reductionist.

AUTHOR CONTRIBUTIONS

NM planned, drafted, and corrected most of the manuscript, including figures and tables. FN wrote the parts on auto-immunity and created the table. SC was responsible for the

section “The Gut-Brain Axis and Neurodegeneration: Is There a Barriers’ Implication?”. PG contributed to the section on BBB drugs and to the table, and also contributed to the sections “Gut Microbiota and Autoantibodies Production: Initial Clues” and “References.”

REFERENCES

- Abbott, N. J., Patabendige, A. A. K., Dolman, D. E. M., Yusof, S. R., and Begley, D. J. (2010). Structure and function of the blood-brain barrier. *Neurobiol. Dis.* 37, 13–25.
- Adams, J. B., Johansen, L. J., Powell, L. D., Quig, D., and Rubin, R. A. (2011). Gastrointestinal flora and gastrointestinal status in children with autism—comparisons to typical children and correlation with autism severity. *BMC Gastroenterol.* 11:22. doi: 10.1186/1471-230X-11-22
- Aggarwal, A., Khera, A., Singh, I., and Sandhir, R. (2015). S-nitrosoglutathione prevents blood-brain barrier disruption associated with increased matrix metalloproteinase-9 activity in experimental diabetes. *J. Neurochem.* 132, 595–608. doi: 10.1111/jnc.12939
- Aimaretti, G., Ambrosio, M. R., Di Somma, C., Gasperi, M., Cannavò, S., Scaroni, C., et al. (2005). Residual pituitary function after brain injury-induced hypopituitarism: a prospective 12-month study. *J. Clin. Endocrinol. Metab.* 90, 6085–6092. doi: 10.1210/jc.2005-0504
- Allen, H. B. (2016). Alzheimer’s disease: assessing the role of spirochetes, biofilms, the immune system, and amyloid- β with regard to potential treatment and prevention. *J. Alzheimers Dis.* 27, 1271–1276. doi: 10.3233/jad-160388
- Alonso, R., Pisa, D., Marina, A. I., Morato, E., Rábano, A., and Carrasco, L. (2014). Fungal infection in patients with Alzheimer’s disease. *J. Alzheimers Dis.* 41, 301–311.
- Amedei, A., and Boem, F. (2018). I’ve got a feeling: microbiota impacting the conceptual and experimental perspectives of personalized medicine. *Int. J. Mol. Sci.* 19:E3756. doi: 10.3390/ijms19123756
- Arango-Lievano, M., Boussadia, B., De Terdonck, L. D. T., Gault, C., Fontanaud, P., Lafont, C., et al. (2018). Topographic reorganization of cerebrovascular mural cells under seizure conditions. *Cell Rep.* 24, 1045–1059. doi: 10.1016/j.celrep.2018.03.110
- Armulik, A., Genové, G., Mäe, M., Nisancioglu, M. H., Wallgard, E., Niaudet, C., et al. (2017). Pericytes regulate the blood-brain barrier. *Nature* 468, 557–561.
- Aspelund, A., Antila, S., Proulx, S. T., Karlsen, T. V., Karaman, S., Detmar, M., et al. (2015). A dural lymphatic vascular system that drains brain interstitial fluid and macromolecules. *J. Exp. Med.* 212, 991–999. doi: 10.1084/jem.2014.2290
- Asumi, A. A., Boutajangout, A., Quartermain, D., and Sigurdsson, E. M. (2007). Immunotherapy targeting pathological tau conformers in a tangle mouse model reduces brain pathology with associated functional improvements. *J. Neurosci.* 27, 9115–9129. doi: 10.1523/jneurosci.2361-07.2007
- Bach, J.-P., and Dodel, R. (2012). Naturally occurring autoantibodies against β -Amyloid. *Adv. Exp. Med. Biol.* 750, 91–99. doi: 10.1007/978-1-4614-3461-0_7
- Balin, B. J., Gérard, H. C., Arking, E. J., Appelt, D. M., Branigan, P. J., Abrams, J. T., et al. (1998). Identification and localization of *Chlamydia pneumoniae* in the Alzheimer’s brain. *Med. Microbiol. Immunol.* 187, 23–42.
- Bartos, A., Fialová, L., Svarcová, J., and Ripova, D. (2012). Patients with Alzheimer disease have elevated intrathecal synthesis of antibodies against tau protein and heavy neurofilament. *J. Neuroimmunol.* 252, 100–105. doi: 10.1016/j.jneuroim.2012.08.001
- Bell, R. D., Winkler, E. A., Sagare, A. P., Singh, I., LaRue, B., Deane, R., et al. (2010). Pericytes control key neurovascular functions and neuronal phenotype in the adult brain and during brain aging. *Neuron* 68, 409–427. doi: 10.1016/j.neuron.2010.09.043
- Blennow, K., Hardy, J., and Zetterberg, H. (2012). The neuropathology and neurobiology of traumatic brain injury. *Neuron* 76, 886–899. doi: 10.1016/j.neuron.2012.11.021
- Braniste, V., Al-Asmakh, M., Kowal, C., Anuar, F., Abbaspour, A., Tóth, M., et al. (2014). The gut microbiota influences blood-brain barrier permeability in mice. *Sci. Transl. Med.* 6:263ra158. doi: 10.1126/scitranslmed.3009759
- Brettschneider, S., Morgenthaler, N. G., Teipel, S. J., Fischer-Schulz, C., Bürger, K., Dodel, R., et al. (2005). Decreased serum amyloid beta(1-42) autoantibody levels in Alzheimer’s disease, determined by a newly developed immunoprecipitation assay with radiolabeled amyloid beta(1-42) peptide. *Biol. Psychiatry* 57, 813–816. doi: 10.1016/j.biopsych.2004.12.008
- Britschgi, M., Olin, C. E., Johns, H. T., Takeda-Uchimura, Y., LeMieux, M. C., Rufibach, K., et al. (2009). Neuroprotective natural antibodies to assemblies of amyloidogenic peptides decrease with normal aging and advancing Alzheimer’s disease. *Proc. Natl. Acad. Sci. U.S.A.* 106, 12145–12150. doi: 10.1073/pnas.0904866106
- Busse, S., Busse, M., Brix, B., Probst, C., Genz, A., Bogerts, B., et al. (2014). Seroprevalence of N-methyl-D-aspartate glutamate receptor (NMDA-R) autoantibodies in aging subjects without neuropsychiatric disorders and in dementia patients. *Eur. Arch. Psychiatry Clin. Neurosci.* 264, 545–550. doi: 10.1007/s00406-014-0493-9
- Butler, M. I., Cryan, J. F., and Dinan, T. G. (2019). Man and the microbiome: a new theory of everything? *Annu. Rev. Clin. Psychol.* 15, 371–398. doi: 10.1146/annurev-clinpsy-050718-095432
- Casali, P., and Schettino, E. W. (1996). Structure and function of natural antibodies. *Curr. Top. Microbiol. Immunol.* 210, 167–179. doi: 10.1007/978-3-642-85226-8_17
- Cattaneo, A., Cattane, N., Galluzzi, S., Provasi, S., Lopizzo, N., Festari, C., et al. (2017). Association of brain amyloidosis with pro-inflammatory gut bacterial taxa and peripheral inflammation markers in cognitively impaired elderly. *Neurobiol. Aging* 49, 60–68. doi: 10.1016/j.neurobiolaging.2016.08.019
- Cerovic, M., Forloni, G., and Balducci, C. (2019). Neuroinflammation and the gut microbiota: possible alternative therapeutic targets to counteract Alzheimer’s disease? *Front. Aging Neurosci.* 11:284. doi: 10.3389/fnagi.2019.00284
- Chu, C., Murdock, M. H., Jing, D., Won, T. H., Chung, H., Kressel, A. M., et al. (2019). The microbiota regulate neuronal function and fear extinction learning. *Nature* 574, 543–548. doi: 10.1038/s41586-019-1644-y
- Cox, L. M., and Weiner, H. L. (2018). Microbiota signaling pathways that influence neurologic disease. *Neurother. J. Am. Soc. Exp. Neurother.* 15, 135–145. doi: 10.1007/s13311-017-0598-8
- Cristante, E., McArthur, S., Mauro, C., Maggioli, E., Romero, I. A., Wylezinska-Arridge, M., et al. (2013). Identification of an essential endogenous regulator of blood-brain barrier integrity, and its pathological and therapeutic implications. *Proc. Natl. Acad. Sci. U.S.A.* 110, 832–841. doi: 10.1073/pnas.120936.2110
- Cryan, J. F., O’Riordan, K. J., Cowan, C. S. M., Sandhu, K. V., Bastiaanssen, T. F. S., Boehme, M., et al. (2019). The microbiota-gut-brain axis. *Physiol. Rev.* 99, 1877–2013.
- Daneman, R., Zhou, L., Kebede, A. A., and Barres, B. A. (2010). Pericytes are required for blood-brain barrier integrity during embryogenesis. *Nature* 468, 562–566. doi: 10.1038/nature09513
- Davydova, T. V., Voskresenskaya, N. I., Fomina, V. G., Vetrile, L. A., and Doronina, O. A. (2007). Induction of autoantibodies to glutamate in patients with Alzheimer’s disease. *Bull. Exp. Biol. Med.* 143, 182–183. doi: 10.1007/s10517-007-0044-8
- Deane, R., Du Yan, S., Subramanian, R. K., LaRue, B., Jovanovic, S., Hogg, E., et al. (2003). RAGE mediates amyloid-beta peptide transport across the blood-brain barrier and accumulation in brain. *Nat. Med.* 9, 907–913. doi: 10.1038/nm890
- Deane, R., Singh, I., Sagare, A. P., Bell, R. D., Ross, N. T., LaRue, B., et al. (2012). A multimodal RAGE-specific inhibitor reduces amyloid β -mediated brain disorder in a mouse model of Alzheimer disease. *J. Clin. Invest.* 122, 1377–1392. doi: 10.1172/jci58642

ACKNOWLEDGMENTS

This work was supported by the ANR-Epicyte, ANR-HepatoBrain, Era-Net/ANR Neu-Vasc, Fondation de France, FRC, and Muse Grants to NM.

- Dinkins, M. B., Dasgupta, S., Wang, G., Zhu, G., He, Q., Kong, J. N., et al. (2015). The 5XFAD mouse model of Alzheimer's disease exhibits an age-dependent increase in anti-ceramide IgG and exogenous administration of ceramide further increases anti-ceramide titers and amyloid plaque burden. *J. Alzheimers Dis.* 46, 55–61. doi: 10.3233/jad-150088
- Dodel, R., Balakrishnan, K., Keyvani, K., Deuster, O., Neff, F., Andrei-Selmer, L.-C., et al. (2011). Naturally occurring autoantibodies against beta-amyloid: investigating their role in transgenic animal and in vitro models of Alzheimer's disease. *J. Neurosci.* 31, 5847–5854. doi: 10.1523/jneurosci.4401-10.2011
- Dominy, S. S., Lynch, C., Ermini, F., Benedyk, M., Marczyk, A., Konradi, A., et al. (2019). *Porphyromonas gingivalis* in Alzheimer's disease brains: evidence for disease causation and treatment with small-molecule inhibitors. *Sci. Adv.* 5:eau3333. doi: 10.1126/sciadv.aau3333
- Du, Y., Dodel, R., Hampel, H., Bueger, K., Lin, S., Eastwood, B., et al. (2001). Reduced levels of amyloid beta-peptide antibody in Alzheimer disease. *Neurology* 57, 801–805. doi: 10.1212/wnl.57.5.801
- Eimer, W. A., Vijaya Kumar, D. K., Navalpur Shanmugam, N. K., Rodriguez, A. S., Mitchell, T., Washicosky, K. J., et al. (2018). Alzheimer's disease-associated β -amyloid is rapidly seeded by herpesviridae to protect against brain infection. *Neuron* 11, 56.e3–63.e3.
- Elkon, K. B., and Silverman, G. J. (2012). Naturally occurring autoantibodies to apoptotic cells. *Adv. Exp. Med. Biol.* 750, 14–26. doi: 10.1007/978-1-4614-3461-0_2
- Engelhardt, B., Vajkoczy, P., and Weller, R. O. (2017). The movers and shapers in immune privilege of the CNS. *Nat. Immunol.* 18, 123–131. doi: 10.1038/ni.3666
- Fialová, L., Bartos, A., Svarcová, J., and Malbohan, I. (2011). Increased intrathecal high-avidity anti-tau antibodies in patients with multiple sclerosis. *PLoS One* 6:e27476. doi: 10.1371/journal.pone.0027476
- Fung, T. C., Olson, C. A., and Hsiao, E. Y. (2017). Interactions between the microbiota, immune and nervous systems in health and disease. *Nat. Neurosci.* 20, 145–155. doi: 10.1038/nn.4476
- Giannoni, P., Badaut, J., Dargazani, C., De Maudave, A. F., Klement, W., Costalat, V., et al. (2018). The pericyte-glia interface at the blood-brain barrier. *Clin. Sci. Lond. Engl.* 14, 361–374. doi: 10.1042/CS20171634
- Goldstein, L. E., Fisher, A. M., Tagge, C. A., Zhang, X.-L., Velisek, L., Sullivan, J. A., et al. (2012). Chronic traumatic encephalopathy in blast-exposed military veterans and a blast neurotrauma mouse model. *Sci. Transl. Med.* 4:134ra60.
- Gong, P., Zhang, Z., Zou, Y., Tian, Q., Han, S., Xu, Z., et al. (2019). Tetramethylpyrazine attenuates blood-brain barrier disruption in ischemia/reperfusion injury through the JAK/STAT signaling pathway. *Eur. J. Pharmacol.* 854, 289–297. doi: 10.1016/j.ejphar.2019.04.028
- Goryunova, A. V., Bazarnaya, N. A., Sorokina, E. G., Semenova, N. Y., Globa, O. V., Semenova, Z. B., et al. (2007). Glutamate receptor autoantibody concentrations in children with chronic post-traumatic headache. *Neurosci. Behav. Physiol.* 37, 761–764. doi: 10.1007/s11055-007-0079-3
- Gruden, M. A., Davidova, T. B., Malisaukas, M., Sewell, R. D. E., Voskresenskaya, N. I., Wilhelm, K., et al. (2007). Differential neuroimmune markers to the onset of Alzheimer's disease neurodegeneration and dementia: autoantibodies to A β (25–35) oligomers, S100b and neurotransmitters. *J. Neuroimmunol.* 186, 181–192. doi: 10.1016/j.jneuroim.2007.03.023
- Gustaw, K. A., Garrett, M. R., Lee, H.-G., Castellani, R. J., Zagorski, M. G., Prakasam, A., et al. (2008). Antigen-antibody dissociation in Alzheimer disease: a novel approach to diagnosis. *J. Neurochem.* 106, 1350–1356. doi: 10.1111/j.1471-4159.2008.05477.x
- Harach, T., Marungruang, N., Duthilleul, N., Cheatham, V., Mc Coy, K. D., Frisoni, G., et al. (2017). Reduction of A β amyloid pathology in APPPS1 transgenic mice in the absence of gut microbiota. *Sci. Rep.* 08:41802.
- Hind, W. H., England, T. J., and O'Sullivan, S. E. (2016). Cannabidiol protects an in vitro model of the blood-brain barrier from oxygen-glucose deprivation via PPAR γ and 5-HT $1A$ receptors. *Br. J. Pharmacol.* 173, 815–825. doi: 10.1111/bph.13368
- Hong, S., JianCheng, H., JiaWen, W., ShuQin, Z., GuiLian, Z., HaiQin, W., et al. (2019). Losartan inhibits development of spontaneous recurrent seizures by preventing astrocyte activation and attenuating blood-brain barrier permeability following pilocarpine-induced status epilepticus. *Brain Res. Bull.* 149, 251–259. doi: 10.1016/j.brainresbull.2019.05.002
- Hue, C. D., Cho, F. S., Cao, S., Dale Bass, C. R., Meaney, D. F., and Morrison, B. (2015). Dexamethasone potentiates in vitro blood-brain barrier recovery after primary blast injury by glucocorticoid receptor-mediated upregulation of ZO-1 tight junction protein. *J. Cereb. Blood Flow Metab.* 35, 1191–1198. doi: 10.1038/jcbfm.2015.38
- Iadecola, C., Zhang, F., Niwa, K., Eckman, C., Turner, S. K., Fischer, E., et al. (1999). SOD1 rescues cerebral endothelial dysfunction in mice overexpressing amyloid precursor protein. *Nat. Neurosci.* 2, 157–161. doi: 10.1038/5715
- Itzhaki, R. F., Lathe, R., Balin, B. J., Ball, M. J., Bearer, E. L., Braak, H., et al. (2016). Microbes and Alzheimer's disease. *J. Alzheimers Dis.* 51, 979–984.
- Janeway, C. A., Travers, P., Walport, M., and Shlomchik, M. J. Jr. (2001). *Immunobiology*, 5th Edn. New York, NY: Garland Science.
- Kang, D.-W., Adams, J. B., Coleman, D. M., Pollard, E. L., Maldonado, J., McDonough-Means, S., et al. (2019). Long-term benefit of microbiota transfer therapy on autism symptoms and gut microbiota. *Sci. Rep.* 9:5821. doi: 10.1038/s41598-019-42183-0
- Karczewski, P., Hempel, P., Kunze, R., and Bimmler, M. (2012). Agonistic autoantibodies to the $\alpha(1)$ -adrenergic receptor and the $\beta(2)$ -adrenergic receptor in Alzheimer's and vascular dementia. *Scand. J. Immunol.* 75, 524–530. doi: 10.1111/j.1365-3083.2012.02684.x
- Kaya, M., Kalayci, R., Küçük, M., Arican, N., Elmas, I., Kudat, H., et al. (2003). Effect of losartan on the blood-brain barrier permeability in diabetic hypertensive rats. *Life Sci.* 73, 3235–3244. doi: 10.1016/j.lfs.2003.06.014
- Kellner, A., Matschke, J., Bernreuther, C., Moch, H., Ferrer, I., and Glatzel, M. (2009). Autoantibodies against beta-amyloid are common in Alzheimer's disease and help control plaque burden. *Ann. Neurol.* 65, 24–31. doi: 10.1002/ana.21475
- Kelly, J. R., Borre, Y., O' Brien, C., Patterson, E., El Aidy, S., Deane, J., et al. (2016). Transferring the blues: depression-associated gut microbiota induces neurobehavioural changes in the rat. *J. Psychiatr. Res.* 82, 109–118. doi: 10.1016/j.jpsychires.2016.07.019
- Kenney-Jung, D. L., Vezzani, A., Kahoud, R. J., LaFrance-Corey, R. G., Ho, M.-L., Muskardin, T. W., et al. (2016). Febrile infection-related epilepsy syndrome treated with anakinra. *Ann. Neurol.* 80, 939–945. doi: 10.1002/ana.24806
- Kisler, K., Nelson, A. R., Rege, S. V., Ramanathan, A., Wang, Y., Ahuja, A., et al. (2017). Pericyte degeneration leads to neurovascular uncoupling and limits oxygen supply to brain. *Nat. Neurosci.* 20, 406–416. doi: 10.1038/nn.4489
- Klement, W., Blaquiére, M., Zub, E., deBock, F., Boux, F., Barbier, E., et al. (2019). A pericyte-glia scarring develops at the leaky capillaries in the hippocampus during seizure activity. *Epilepsia* 60, 1399–1411. doi: 10.1111/epi.16019
- Kobeissy, F. H. (2015). *Brain Neurotrauma: Molecular, Neuropsychological, and Rehabilitation Aspects*. Boca Raton, FL: CRC Press.
- Kondo, A., Shahpasand, K., Mannix, R., Qiu, J., Moncaster, J., Chen, C.-H., et al. (2015). Antibody against early driver of neurodegeneration cis P-tau blocks brain injury and tauopathy. *Nature* 523, 431–436. doi: 10.1038/nature14658
- Kornhuber, H. H. (1996). Propionibacterium acnes in the cortex of patients with Alzheimer's disease. *Eur. Arch. Psychiatry Clin. Neurosci.* 246, 108–109. doi: 10.1007/bf02274902
- Kosaraju, R., Guesdon, W., Crouch, M. J., Teague, H. L., Sullivan, E. M., Karlsson, E. A., et al. (2017). B cell activity is impaired in human and mouse obesity and is responsive to an essential fatty acid upon murine influenza infection. *J. Immunol.* 198, 4738–4752. doi: 10.4049/jimmunol.1601031
- Kountouras, J., Tsolaki, M., Gavalas, E., Boziki, M., Zavos, C., Karatzoglou, P., et al. (2006). Relationship between *Helicobacter pylori* infection and Alzheimer disease. *Neurology* 66, 938–940. doi: 10.1212/01.wnl.0000203644.68059.5f
- Kronimus, Y., Albus, A., Balzer-Geldsetzer, M., Straub, S., Semler, E., Otto, M., et al. (2016). Naturally occurring autoantibodies against tau protein are reduced in Parkinson's disease dementia. *PLoS One* 11:e0164953. doi: 10.1371/journal.pone.0164953
- Kucuk, M., Kaya, M., Kalayci, R., Cimen, V., Kudat, H., Arican, N., et al. (2002). Effects of losartan on the blood-brain barrier permeability in long-term nitric oxide blockade-induced hypertensive rats. *Life Sci.* 71, 937–946. doi: 10.1016/s0024-3205(02)01772-1
- Kumar, D. K. V., Choi, S. H., Washicosky, K. J., Eimer, W. A., Tucker, S., Ghofrani, J., et al. (2016). Amyloid- β peptide protects against microbial infection in mouse and worm models of Alzheimer's disease. *Sci. Transl. Med.* 8:340ra72. doi: 10.1126/scitranslmed.aaf1059

- Kumar, N., and Forster, S. C. (2017). Genome watch: microbiota shuns the modern world. *Nat. Rev. Microbiol.* 15, 710–710. doi: 10.1038/nrmicro.2017.136
- Lazic, D., Sagare, A. P., Nikolakopoulou, A. M., Griffin, J. H., Vassar, R., and Zlokovic, B. V. (2019). 3K3A-activated protein C blocks amyloidogenic BACE1 pathway and improves functional outcome in mice. *J. Exp. Med.* 216, 279–293. doi: 10.1084/jem.20181035
- Le Page, A., Dupuis, G., Frost, E. H., Larbi, A., Pawelec, G., Witkowski, J. M., et al. (2018). Role of the peripheral innate immune system in the development of Alzheimer's disease. *Exp. Gerontol.* 01, 59–66.
- Lee, Y. K., Menezes, J. S., Umesaki, Y., and Mazmanian, S. K. (2011). Proinflammatory T-cell responses to gut microbiota promote experimental autoimmune encephalomyelitis. *Proc. Natl. Acad. Sci. U.S.A.* 108(Suppl. 1), 4615–4622. doi: 10.1073/pnas.1000082107
- Lelouvier, B., Servant, F., Païssé, S., Brunet, A.-C., Benyahya, S., Serino, M., et al. (2016). Changes in blood microbiota profiles associated with liver fibrosis in obese patients: a pilot analysis. *Hepatology* 64, 2015–2027. doi: 10.1002/hep.28829
- Levin, E. C., Acharya, N. K., Han, M., Zavareh, S. B., Sedeyn, J. C., Venkataraman, V., et al. (2010). Brain-reactive autoantibodies are nearly ubiquitous in human sera and may be linked to pathology in the context of blood-brain barrier breakdown. *Brain Res.* 1345, 221–232. doi: 10.1016/j.brainres.2010.05.038
- Lin, W.-R., Wozniak, M. A., Cooper, R. J., Wilcock, G. K., and Itzhaki, R. F. (2002). Herpesviruses in brain and Alzheimer's disease. *J. Pathol.* 197, 395–402. doi: 10.1002/path.1127
- Lindhagen-Persson, M., Brännström, K., Vestling, M., Steinitz, M., and Olofsson, A. (2010). Amyloid- β oligomer specificity mediated by the IgM isotype—implications for a specific protective mechanism exerted by endogenous autoantibodies. *PLoS One* 5:e13928. doi: 10.1371/journal.pone.0013928
- Louveau, A., Harris, T. H., and Kipnis, J. (2015a). Revisiting the mechanisms of CNS immune privilege. *Trends Immunol.* 36, 569–577. doi: 10.1016/j.it.2015.08.006
- Louveau, A., Herz, J., Alme, M. N., Salvador, A. F., Dong, M. Q., Viar, K. E., et al. (2018). CNS lymphatic drainage and neuroinflammation are regulated by meningeal lymphatic vasculature. *Nat. Neurosci.* 21, 1380–1391. doi: 10.1038/s41593-018-0227-9
- Louveau, A., Smirnov, I., Keyes, T. J., Eccles, J. D., Rouhani, S. J., Peske, J. D., et al. (2015b). Structural and functional features of central nervous system lymphatic vessels. *Nature* 523, 337–341. doi: 10.1038/nature14432
- Lozupone, C. A., Stombaugh, J. I., Gordon, J. I., Jansson, J. K., and Knight, R. (2012). Diversity, stability and resilience of the human gut microbiota. *Nature* 489, 220–230. doi: 10.1038/nature11550
- Lyden, P., Pryor, K. E., Coffey, C. S., Cudkovic, M., Conwit, R., Jadhav, A., et al. (2019). Final results of the RHAPSODY trial: a multi-center, phase 2 trial using a continual reassessment method to determine the safety and tolerability of 3K3A-APC, a recombinant variant of human activated protein C, in combination with tissue plasminogen activator, mechanical thrombectomy or both in moderate to severe acute ischemic stroke. *Ann. Neurol.* 85, 125–136. doi: 10.1002/ana.25383
- Maftei, M., Thurm, F., Schnack, C., Tumani, H., Otto, M., Elbert, T., et al. (2013). Increased levels of antigen-bound β -amyloid autoantibodies in serum and cerebrospinal fluid of Alzheimer's disease patients. *PLoS One* 8:e68996. doi: 10.1371/journal.pone.0068996
- Mannix, R., Meehan, W. P., Mandeville, J., Grant, P. E., Gray, T., Berglass, J., et al. (2013). Clinical correlates in an experimental model of repetitive mild brain injury. *Ann. Neurol.* 74, 65–75. doi: 10.1002/ana.23858
- Marchi, N., Angelov, L., Masaryk, T., Fazio, V., Granata, T., Hernandez, N., et al. (2007). Seizure-promoting effect of blood-brain barrier disruption. *Epilepsia* 48, 732–742. doi: 10.1111/j.1528-1167.2007.00988.x
- Marchi, N., Bazarian, J. J., Puvanna, V., Janigro, M., Ghosh, C., Zhong, J., et al. (2013). Consequences of repeated blood-brain barrier disruption in football players. *PLoS One* 8:e56805. doi: 10.1371/journal.pone.0056805
- Marchi, N., Fan, Q., Ghosh, C., Fazio, V., Bertolini, F., Betto, G., et al. (2009). Antagonism of peripheral inflammation reduces the severity of status epilepticus. *Neurobiol. Dis.* 33, 171–181. doi: 10.1016/j.nbd.2008.10.002
- Marchi, N., Granata, T., Freri, E., Ciusani, E., Ragona, F., Puvanna, V., et al. (2011). Efficacy of anti-inflammatory therapy in a model of acute seizures and in a population of pediatric drug resistant epileptics. *PLoS One* 6:e18200. doi: 10.1371/journal.pone.0018200
- Marchi, N., Granata, T., and Janigro, D. (2014). Inflammatory pathways of seizure disorders. *Trends Neurosci.* 37, 55–65. doi: 10.1016/j.tins.2013.11.002
- Martin, C. R., Osadchiy, V., Kalani, A., and Mayer, E. A. (2018). The brain-gut-microbiome axis. *Cell Mol. Gastroenterol. Hepatol.* 6, 133–148. doi: 10.1016/j.jcmgh.2018.04.003
- Marushima, A., Nieminen, M., Kremenskaia, I., Gianni-Barrera, R., Woitzik, J., von Degenfeld, G., et al. (2019). Balanced single-vector co-delivery of VEGF/PDGF-BB improves functional collateralization in chronic cerebral ischemia. *J. Cereb. Blood Flow Metab.* 9:271678X18818298. doi: 10.1177/0271678X18818298
- McIntyre, J. A., Chapman, J., Shavit, E., Hamilton, R. L., and Dekosky, S. T. (2007). Redox-reactive autoantibodies in Alzheimer's patients' cerebrospinal fluids: preliminary studies. *Autoimmunity* 40, 390–396. doi: 10.1080/08916930701421020
- McIntyre, J. A., Ramsey, C. J., Gitter, B. D., Saykin, A. J., Wagenknecht, D. R., Hyslop, P. A., et al. (2015). Antiphospholipid autoantibodies as blood biomarkers for detection of early stage Alzheimer's disease. *Autoimmunity* 48, 344–351. doi: 10.3109/08916934.2015.1008464
- McRae, A., Martins, R. N., Fonte, J., Kraftsik, R., Hirt, L., and Miklossy, J. (2007). Cerebrospinal fluid antimicrobial antibodies in Alzheimer disease: a putative marker of an ongoing inflammatory process. *Exp. Gerontol.* 42, 355–363. doi: 10.1016/j.exger.2006.10.015
- Mecucci, P., Parnetti, L., Romano, G., Scarelli, A., Chionne, F., Cecchetti, R., et al. (1995). Serum anti-GFAP and anti-S100 autoantibodies in brain aging, Alzheimer's disease and vascular dementia. *J. Neuroimmunol.* 57, 165–170. doi: 10.1016/0165-5728(94)00180-v
- Miklossy, J. (2011). Alzheimer's disease - a neurospirochetosis. Analysis of the evidence following Koch's and Hill's criteria. *J. Neuroinflamm.* 8:90. doi: 10.1186/1742-2094-8-90
- Miklossy, J. (2016). Bacterial amyloid and DNA are important constituents of senile plaques: further evidence of the spirochetal and biofilm nature of senile plaques. *J. Alzheimers Dis.* 13, 1459–1473. doi: 10.3233/JAD-160451
- Minter, M. R., Hinterleitner, R., Meisel, M., Zhang, C., Leone, V., Zhang, X., et al. (2017). Antibiotic-induced perturbations in microbial diversity during post-natal development alters amyloid pathology in an aged APPSWE/PS1 Δ E9 murine model of Alzheimer's disease. *Sci. Rep.* 7:10411.
- Minter, M. R., Zhang, C., Leone, V., Ringus, D. L., Zhang, X., Oyler-Castrillo, P., et al. (2016). Antibiotic-induced perturbations in gut microbial diversity influences neuro-inflammation and amyloidosis in a murine model of Alzheimer's disease. *Sci. Rep.* 21:30028.
- Moir, R. D., Lathe, R., and Tanzi, R. E. (2018). The antimicrobial protection hypothesis of Alzheimer's disease. *Alzheimers Dement. J. Alzheimers Assoc.* 14, 1602–1614. doi: 10.1016/j.jalz.2018.06.3040
- Montagne, A., Barnes, S. R., Sweeney, M. D., Halliday, M. R., Sagare, A. P., Zhao, Z., et al. (2015). Blood-brain barrier breakdown in the aging human hippocampus. *Neuron* 85, 296–302. doi: 10.1016/j.neuron.2014.12.032
- Montagne, A., Nikolakopoulou, A. M., Zhao, Z., Sagare, A. P., Si, G., Lazic, D., et al. (2018). Pericyte degeneration causes white matter dysfunction in the mouse central nervous system. *Nat. Med.* 24, 326–337. doi: 10.1038/nm.4482
- Mruthinti, S., Buccafusco, J. J., Hill, W. D., Waller, J. L., Jackson, T. W., Zamrini, E. Y., et al. (2004). Autoimmunity in Alzheimer's disease: increased levels of circulating IgGs binding Abeta and RAGE peptides. *Neurobiol. Aging* 25, 1023–1032. doi: 10.1016/j.neurobiolaging.2003.11.001
- Myagkova, M. A., Gavrilova, S. I., Lermontova, N. N., Kalyn, Y. B., Selezneva, N. D., Zharikov, G. A., et al. (2001). Autoantibodies to beta-amyloid and neurotransmitters in patients with Alzheimer's disease and senile dementia of the Alzheimer type. *Bull. Exp. Biol. Med.* 131, 127–129.
- Myles, I. A. (2014). Fast food fever: reviewing the impacts of the Western diet on immunity. *Nutr. J.* 13:61. doi: 10.1186/1475-2891-13-61
- Na, W., Shin, J. Y., Lee, J. Y., Jeong, S., Kim, W.-S., Yune, T. Y., et al. (2017). Dexamethasone suppresses JMJD3 gene activation via a putative negative glucocorticoid response element and maintains integrity of tight junctions in brain microvascular endothelial cells. *J. Cereb. Blood Flow Metab.* 37, 3695–3708. doi: 10.1177/0271678X17701156
- Nation, D. A., Sweeney, M. D., Montagne, A., Sagare, A. P., D'Orazio, L. M., Pachicano, M., et al. (2019). Blood-brain barrier breakdown is an early

- biomarker of human cognitive dysfunction. *Nat. Med.* 25, 270–276. doi: 10.1038/s41591-018-0297-y
- Nikolakopoulou, A. M., Zhao, Z., Montagne, A., and Zlokovic, B. V. (2019). Regional early and progressive loss of brain pericytes but not vascular smooth muscle cells in adult mice with disrupted platelet-derived growth factor receptor- β signaling. *PLoS One* 12:e0176225. doi: 10.1371/journal.pone.0176225
- Nishimura, S., Manabe, I., Takaki, S., Nagasaki, M., Otsu, M., Yamashita, H., et al. (2013). Adipose natural regulatory B cells negatively control adipose tissue inflammation. *Cell Metab.* 18, 759–766. doi: 10.1016/j.cmet.2013.09.017
- Noé, F. M., and Marchi, N. (2019). Central nervous system lymphatic unit, immunity, and epilepsy: is there a link? *Epilepsia Open* 4, 30–39. doi: 10.1002/epi4.12302
- Nortley, R., Korte, N., Izquierdo, P., Hirunpattarasilp, C., Mishra, A., Jaunmuktane, Z., et al. (2019). Amyloid β oligomers constrict human capillaries in Alzheimer's disease via signaling to pericytes. *Science* 365:eav9518. doi: 10.1126/science.aav9518
- Ochoa-Repáraz, J., Mielcarz, D. W., Ditrio, L. E., Burroughs, A. R., Foureau, D. M., Haque-Begum, S., et al. (2009). Role of gut commensal microflora in the development of experimental autoimmune encephalomyelitis. *J. Immunol.* 183, 6041–6050. doi: 10.4049/jimmunol.0900747
- Ochoa-Repáraz, J., Mielcarz, D. W., Haque-Begum, S., and Kasper, L. H. (2010). Induction of a regulatory B cell population in experimental allergic encephalomyelitis by alteration of the gut commensal microflora. *Gut Microbes* 1, 103–108. doi: 10.4161/gmic.1.2.11515
- Omali, B. I., Hamilton, R. L., Kamboh, M. I., DeKosky, S. T., and Bailes, J. (2010). Chronic traumatic encephalopathy (CTE) in a National Football League Player: case report and emerging medicolegal practice questions. *J. Forensic Nurs.* 6, 40–46. doi: 10.1111/j.1939-3938.2009.01064.x
- Ouwehand, A., Isolauri, E., and Salminen, S. (2002). The role of the intestinal microflora for the development of the immune system in early childhood. *Eur. J. Nutr.* 41(Suppl. 1), I32–I37.
- Pagliai, G., Russo, E., Niccolai, E., Dinu, M., Di Pilato, V., Magrini, A., et al. (2019). Influence of a 3-month low-calorie Mediterranean diet compared to the vegetarian diet on human gut microbiota and SCFA: the CARDIVeG study. *Eur. J. Nutr.* doi: 10.1007/s00394-019-02050-0 [Epub ahead of print].
- Païssé, S., Valle, C., Servant, F., Courtney, M., Burcelin, R., Amar, J., et al. (2016). Comprehensive description of blood microbiome from healthy donors assessed by 16S targeted metagenomic sequencing. *Transfusion* 56, 1138–1147. doi: 10.1111/trf.13477
- Pani, F., Di Dalmazi, G., Corsello, A., Oliver, T. G., Livezey, J. R., and Caturegli, P. (2019). MON-450 pituitary antibodies in a cohort of us service members with traumatic brain injury. *J. Endocr. Soc.* 3(Suppl. 1):MON-450.
- Parker, A., Fonseca, S., and Carding, S. R. (2019). Gut microbes and metabolites as modulators of blood-brain barrier integrity and brain health. *Gut Microbes* doi: 10.1080/19490976.2019.1638722 [Epub ahead of print].
- Pavlov, V. A., and Tracey, K. J. (2017). Neural regulation of immunity: molecular mechanisms and clinical translation. *Nat. Neurosci.* 20, 156–166. doi: 10.1038/nn.4477
- Pestka, J. J., Vines, L. L., Bates, M. A., He, K., and Langohr, I. (2014). Comparative effects of n-3, n-6 and n-9 unsaturated fatty acid-rich diet consumption on lupus nephritis, autoantibody production and CD4+ T cell-related gene responses in the autoimmune NZBWF1 mouse. *PLoS One* 9:e100255. doi: 10.1371/journal.pone.0100255
- Petta, I., Fraussen, J., Somers, V., and Kleinewietfeld, M. (2018). Interrelation of diet, gut microbiome, and autoantibody production. *Front. Immunol.* 9:439. doi: 10.3389/fimmu.2018.00439
- Poole, S., Singhrao, S. K., Chukkapalli, S., Rivera, M., Velsko, I., Kesavalu, L., et al. (2015). Active invasion of *Porphyromonas gingivalis* and infection-induced complement activation in ApoE^{-/-} mice brains. *J. Alzheimers Dis.* 43, 67–80. doi: 10.3233/JAD-140315
- Posse de Chaves, E., and Sipione, S. (2010). Sphingolipids and gangliosides of the nervous system in membrane function and dysfunction. *FEBS Lett.* 584, 1748–1759. doi: 10.1016/j.febslet.2009.12.010
- Prinz, M., and Priller, J. (2017). The role of peripheral immune cells in the CNS in steady state and disease. *Nat. Neurosci.* 20, 136–144. doi: 10.1038/nn.4475
- Purvis, G. S. D., Solito, E., and Thiemermann, C. (2019). Annexin-A1: therapeutic potential in microvascular disease. *Front. Immunol.* 10:938. doi: 10.3389/fimmu.2019.00938
- Qu, B.-X., Gong, Y., Moore, C., Fu, M., German, D. C., Chang, L.-Y., et al. (2014). Beta-amyloid auto-antibodies are reduced in Alzheimer's disease. *J. Neuroimmunol.* 274, 168–173. doi: 10.1016/j.jneuroim.2014.06.017
- Raad, M., Nohra, E., Chams, N., Itani, M., Talih, F., Mondello, S., et al. (2014). Autoantibodies in traumatic brain injury and central nervous system trauma. *Neuroscience* 281, 16–23. doi: 10.1016/j.neuroscience.2014.08.045
- Rahman, M. T., Ghosh, C., Hossain, M., Linfield, D., Rezaee, F., Janigro, D., et al. (2018). IFN- γ , IL-17A, or zonulin rapidly increase the permeability of the blood-brain and small intestinal epithelial barriers: relevance for neuro-inflammatory diseases. *Biochem. Biophys. Res. Commun.* 507, 274–279. doi: 10.1016/j.bbrc.2018.11.021
- Ransohoff, R. M. (2016). How neuroinflammation contributes to neurodegeneration. *Science* 353, 777–783. doi: 10.1126/science.aag2590
- Religa, P., Cao, R., Religa, D., Xue, Y., Bogdanovic, N., Westaway, D., et al. (2013). VEGF significantly restores impaired memory behavior in Alzheimer's mice by improvement of vascular survival. *Sci. Rep.* 3, 2053.
- Reza, M. M., Finlay, B. B., and Pettersson, S. (2019). Gut microbes, ageing & organ function: a chameleon in modern biology? *EMBO Mol. Med.* 11:e9872.
- Rinne, J. O., Brooks, D. J., Rossor, M. N., Fox, N. C., Bullock, R., Klunk, W. E., et al. (2010). 11C-PiB PET assessment of change in fibrillar amyloid-beta load in patients with Alzheimer's disease treated with bapineuzumab: a phase 2, double-blind, placebo-controlled, ascending-dose study. *Lancet Neurol.* 9, 363–372. doi: 10.1016/S1474-4422(10)70043-0
- Rosenmann, H., Meiner, Z., Geylis, V., Abramsky, O., and Steinitz, M. (2006). Detection of circulating antibodies against tau protein in its unphosphorylated and in its neurofibrillary tangles-related phosphorylated state in Alzheimer's disease and healthy subjects. *Neurosci Lett.* 410, 90–93. doi: 10.1016/j.neulet.2006.01.072
- Rudehill, S., Muhallab, S., Wennersten, A., von Gertten, C., Al Nimer, F., Sandberg-Nordqvist, A. C., et al. (2006). Autoreactive antibodies against neurons and basal lamina found in serum following experimental brain contusion in rats. *Acta Neurochir.* 148, 199–205. doi: 10.1007/s00701-005-0673-5
- Rustenhoven, J., Jansson, D., Smyth, L. C., and Draganow, M. (2017). Brain Pericytes As Mediators of Neuroinflammation. *Trends Pharmacol. Sci.* 38, 291–304. doi: 10.1016/j.tips.2016.12.001
- Salloway, S., Sperling, R., Gilman, S., Fox, N. C., Blennow, K., Raskind, M., et al. (2009). A phase 2 multiple ascending dose trial of bapineuzumab in mild to moderate Alzheimer disease. *Neurology* 73, 2061–2070. doi: 10.1212/WNL.0b013e3181c67808
- Sampson, T. R., Debelius, J. W., Thron, T., Janssen, S., Shastri, G. G., Ilhan, Z. E., et al. (2016). Gut microbiota regulate motor deficits and neuroinflammation in a model of Parkinson's disease. *Cell* 167, 1469.e12–1480.e12. doi: 10.1016/j.cell.2016.11.018
- Scheperjans, F., Aho, V., Pereira, P. A. B., Koskinen, K., Paulin, L., Pekkonen, E., et al. (2015). Gut microbiota are related to Parkinson's disease and clinical phenotype. *Mov. Disord.* 30, 350–358.
- Schierwagen, R., Alvarez-Silva, C., Madsen, M. S. A., Kolbe, C. C., Meyer, C., Thomas, D., et al. (2018). Circulating microbiome in blood of different circulatory compartments. *Gut* doi: 10.1136/gutjnl-2018-316227 [Epub ahead of print].
- Schroeder, B. O., and Bäckhed, F. (2016). Signals from the gut microbiota to distant organs in physiology and disease. *Nat. Med.* 22, 1079–1089. doi: 10.1038/nm.4185
- Schwartz, M., and Shechter, R. (2010). Systemic inflammatory cells fight off neurodegenerative disease. *Nat. Rev. Neurol.* 6, 405–410. doi: 10.1038/nrneurol.2010.71
- Senatorov, V. V., Friedman, A. R., Milikovskiy, D. Z., Ofer, J., Saar-Ashkenazy, R., Charbash, A., et al. (2019). Blood-brain barrier dysfunction in aging induces hyper-activation of TGF-beta signaling and chronic yet reversible neural dysfunction. *Sci. Transl. Med.* 11:eaaaw8283. doi: 10.1126/scitranslmed.aaw8283
- Sevigny, J., Chiao, P., Bussière, T., Weinreb, P. H., Williams, L., Maier, M., et al. (2016). The antibody aducanumab reduces A β plaques in Alzheimer's disease. *Nature* 01, 50–56.

- Shanahan, F., van Sinderen, D., O'Toole, P. W., and Stanton, C. (2017). Feeding the microbiota: transducer of nutrient signals for the host. *Gut* 66, 1709–1717. doi: 10.1136/gutjnl-2017-313872
- Sherwin, E., Bordenstein, S. R., Quinn, J. L., Dinan, T. G., and Cryan, J. F. (2019). Microbiota and the social brain. *Science* 366:eaar2016.
- Sigurdsson, E. M. (2018). Tau immunotherapies for Alzheimer's disease and related tauopathies: progress and potential pitfalls. *J. Alzheimers Dis.* 64(Suppl.1), S555–S565.
- Singh, R. K., Chang, H.-W., Yan, D., Lee, K. M., Ucmak, D., Wong, K., et al. (2017). Influence of diet on the gut microbiome and implications for human health. *J. Transl. Med.* 15:73. doi: 10.1186/s12967-017-1175-y
- Sinha, R. K., Wang, Y., Zhao, Z., Xu, X., Burnier, L., Gupta, N., et al. (2018). PAR1 biased signaling is required for activated protein C *in vivo* benefits in sepsis and stroke. *Blood* 15, 1163–1171. doi: 10.1182/blood-2017-10-810895
- Smith, C., Graham, D. I., Murray, L. S., and Nicoll, J. A. (2003). Tau immunohistochemistry in acute brain injury. *Neuropathol. Appl. Neurobiol.* 29, 496–502. doi: 10.1046/j.1365-2990.2003.00488.x
- Sommer, A., Winner, B., and Prots, I. (2017). The Trojan horse - neuroinflammatory impact of T cells in neurodegenerative diseases. *Mol. Neurodegener.* 12:78. doi: 10.1186/s13024-017-0222-8
- Sorokina, E. G., Vol'pina, O. M., Semenova, Z. B., Karaseva, O. V., Koroev, D. O., Kamynina, A. V., et al. (2011). [Autoantibodies to $\alpha 7$ -subunit of neuronal acetylcholine receptor in children with traumatic brain injury]. *Zh. Nevrol. Psikiatr. Im. S S Korsakova* 111, 56–60.
- Soscia, S. J., Kirby, J. E., Washicosky, K. J., Tucker, S. M., Ingelsson, M., Hyman, B., et al. (2010). The Alzheimer's disease-associated amyloid beta-protein is an antimicrobial peptide. *PLoS One* 5:e9505. doi: 10.1371/journal.pone.0009505
- Su, E. J., Fredriksson, L., Kanzawa, M., Moore, S., Folestad, E., Stevenson, T. K., et al. (2015). Imatinib treatment reduces brain injury in a murine model of traumatic brain injury. *Front. Cell. Neurosci.* 9:385. doi: 10.3389/fncel.2015.00385
- Sweeney, M. D., Zhao, Z., Montagne, A., Nelson, A. R., and Zlokovic, B. V. (2019). Blood-brain barrier: from physiology to disease and back. *Physiol. Rev.* 01, 21–78. doi: 10.1152/physrev.00050.2017
- Tai, X. Y., Koeppe, M., Duncan, J. S., Fox, N., Thompson, P., Baxendale, S., et al. (2016). Hyperphosphorylated tau in patients with refractory epilepsy correlates with cognitive decline: a study of temporal lobe resections. *Brain J. Neurol.* 139(Pt 9), 2441–2455. doi: 10.1093/brain/aww187
- Tanaka, J., Nakamura, K., Takeda, M., Tada, K., Suzuki, H., Morita, H., et al. (1989). Enzyme-linked immunosorbent assay for human autoantibody to glial fibrillary acidic protein: higher titer of the antibody is detected in serum of patients with Alzheimer's disease. *Acta Neurol. Scand.* 80, 554–560. doi: 10.1111/j.1600-0404.1989.tb03926.x
- Tanriverdi, F., De Bellis, A., Battaglia, M., Bellastella, G., Bizzarro, A., Sinisi, A. A., et al. (2010). Investigation of antihypothalamus and antipituitary antibodies in amateur boxers: is chronic repetitive head trauma-induced pituitary dysfunction associated with autoimmunity? *Eur. J. Endocrinol.* 162, 861–867. doi: 10.1530/EJE-09-1024
- Tanriverdi, F., De Bellis, A., Bizzarro, A., Sinisi, A. A., Bellastella, G., Pane, E., et al. (2008). Antipituitary antibodies after traumatic brain injury: is head trauma-induced pituitary dysfunction associated with autoimmunity? *Eur. J. Endocrinol.* 159, 7–13. doi: 10.1530/EJE-08-0050
- Tanriverdi, F., Senyurek, H., Unluhizarci, K., Selcuklu, A., Casanueva, F. F., and Kelestimur, F. (2006). High risk of hypopituitarism after traumatic brain injury: a prospective investigation of anterior pituitary function in the acute phase and 12 months after trauma. *J. Clin. Endocrinol. Metab.* 91, 2105–2111. doi: 10.1210/jc.2005-2476
- Thiyagarajan, M., Fernández, J. A., Lane, S. M., Griffin, J. H., and Zlokovic, B. V. (2008). Activated protein C promotes neovascularization and neurogenesis in postischemic brain via protease-activated receptor 1. *J. Neurosci.* 28, 12788–12797. doi: 10.1523/JNEUROSCI.3485-08.2008
- Thomas, T., Thomas, G., McLendon, C., Sutton, T., and Mullan, M. (1996). beta-Amyloid-mediated vasoactivity and vascular endothelial damage. *Nature* 380, 168–171. doi: 10.1038/380168a0
- Tomasdottir, V., Thorleifsdottir, S., Vikingsson, A., Hardardottir, I., and Freysdottir, J. (2014). Dietary omega-3 fatty acids enhance the B1 but not the B2 cell immune response in mice with antigen-induced peritonitis. *J. Nutr. Biochem.* 25, 111–117. doi: 10.1016/j.jnutbio.2013.09.010
- U.S. National Library of Medicine, (2019a). *A Long-Term Safety and Tolerability Extension Study Of Bapineuzumab In Alzheimer Disease Patients*. Available from: <https://clinicaltrials.gov/ct2/show/NCT00998764>
- U.S. National Library of Medicine, (2019b). *A Long-Term Safety and Tolerability Study Of Bapineuzumab In Alzheimer Disease Patients 2019*. Available from: <https://clinicaltrials.gov/ct2/show/NCT00996918>
- Vacirca, D., Delunardo, F., Matarrese, P., Colasanti, T., Margutti, P., Siracusano, A., et al. (2012). Autoantibodies to the adenosine triphosphate synthase play a pathogenetic role in Alzheimer's disease. *Neurobiol. Aging* 33, 753–766. doi: 10.1016/j.neurobiolaging.2010.05.013
- Van Skike, C. E., Jahrling, J. B., Olson, A. B., Sayre, N. L., Hussong, S. A., Ungvari, Z., et al. (2018). Inhibition of mTOR protects the blood-brain barrier in models of Alzheimer's disease and vascular cognitive impairment. *Am. J. Physiol. Heart Circ. Physiol.* 314, H693–H703.
- Vezzani, A., French, J., Bartfai, T., and Baram, T. Z. (2011). The role of inflammation in epilepsy. *Nat. Rev. Neurol.* 7, 31–40.
- Virtue, A. T., McCright, S. J., Wright, J. M., Jimenez, M. T., Mowel, W. K., Kotzin, J. J., et al. (2019). The gut microbiota regulates white adipose tissue inflammation and obesity via a family of microRNAs. *Sci. Transl. Med.* 11:eaav1892. doi: 10.1126/scitranslmed.aav1892
- Vogt, N. M., Kerby, R. L., Dill-McFarland, K. A., Harding, S. J., Merluzzi, A. P., Johnson, S. C., et al. (2017). Gut microbiome alterations in Alzheimer's disease. *Sci. Rep.* 7:13537. doi: 10.1242/dmm.041947
- Wang, X., Sun, G., Feng, T., Zhang, J., Huang, X., Wang, T., et al. (2019). Sodium oligomannate therapeutically remodels gut microbiota and suppresses gut bacterial amino acids-shaped neuroinflammation to inhibit Alzheimer's disease progression. *Cell Res.* 29, 787–803. doi: 10.1038/s41422-019-0216-x
- Wang, Y., Zhao, Z., Rege, S. V., Wang, M., Si, G., Zhou, Y., et al. (2016). 3K3A-activated protein C stimulates postischemic neuronal repair by human neural stem cells in mice. *Nat. Med.* 22, 1050–1055. doi: 10.1038/nm.4154
- Wekslar, M. E., Relkin, N., Turkenich, R., LaRusse, S., Zhou, L., and Szabo, P. (2002). Patients with Alzheimer disease have lower levels of serum anti-amyloid peptide antibodies than healthy elderly individuals. *Exp. Gerontol.* 37, 943–948. doi: 10.1016/s0531-5565(02)00029-3
- Wilson, J. S., vainti, S., Buccafusco, J. J., Schade, R. F., Mitchell, M. B., Harrell, D. U., et al. (2009). Anti-RAGE and Abeta immunoglobulin levels are related to dementia level and cognitive performance. *J. Gerontol. A Biol. Sci. Med. Sci.* 64, 264–271. doi: 10.1093/gerona/gln002
- Wu, J., and Li, L. (2016). Autoantibodies in Alzheimer's disease: potential biomarkers, pathogenic roles, and therapeutic implications. *J. Biomed. Res.* 30, 361–372.
- Zhang, F., Eckman, C., Younkin, S., Hsiao, K. K., and Iadecola, C. (1997). Increased susceptibility to ischemic brain damage in transgenic mice overexpressing the amyloid precursor protein. *J. Neurosci.* 17, 7655–7661. doi: 10.1523/jneurosci.17-20-07655.1997
- Zhang, N., Yin, S., Zhang, W., Gong, X., Zhang, N., Fang, K., et al. (2017). Crystal structure and biochemical characterization of an aminopeptidase LapB from *Legionella pneumophila*. *J. Agric. Food Chem.* 65, 7569–7578. doi: 10.1021/acs.jafc.7b02849
- Zhang, Z., Zoltewicz, J. S., Mondello, S., Newsom, K. J., Yang, Z., Yang, B., et al. (2014). Human traumatic brain injury induces autoantibody response against glial fibrillary acidic protein and its breakdown products. *PLoS One* 9:e92698. doi: 10.1371/journal.pone.0092698
- Zheng, P., Zeng, B., Zhou, C., Liu, M., Fang, Z., Xu, X., et al. (2016). Gut microbiome remodeling induces depressive-like behaviors through a pathway mediated by the host's metabolism. *Mol. Psychiatry* 21, 786–796. doi: 10.1038/mp.2016.44
- Zhong, Z., Ilieva, H., Hallagan, L., Bell, R., Singh, I., Paquette, N., et al. (2009). Activated protein C therapy slows ALS-like disease in mice by transcriptionally inhibiting SOD1 in motor neurons and microglia cells. *J. Clin. Invest.* 119, 3437–3449. doi: 10.1172/JCI38476
- Zhuang, Z.-Q., Shen, L.-L., Li, W.-W., Fu, X., Zeng, F., Gui, L., et al. (2018). Gut microbiota is altered in patients with Alzheimer's disease. *J. Alzheimers Dis.* 63, 1337–1346. doi: 10.3233/JAD-180176

- Zlokovic, B. V. (2008). The blood-brain barrier in health and chronic neurodegenerative disorders. *Neuron* 57, 178–201. doi: 10.1016/j.neuron.2008.01.003
- Zlokovic, B. V. (2011). Neurovascular pathways to neurodegeneration in Alzheimer's disease and other disorders. *Nat. Rev. Neurosci.* 12, 723–738. doi: 10.1038/nrn3114
- Zub, E., Canet, G., Garbelli, R., Blaquiére, M., Rossini, L., Pastori, C., et al. (2019). The GR-ANXA1 pathway is a pathological player and a candidate target in epilepsy. *FASEB J.* 33, 13998–14009. doi: 10.1096/fj.201901596R

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Giannoni, Claeysen, Noe and Marchi. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.