

Profile of Inhalable Bacteria in PM 2.5 at Mt. Tai, China: Abundance, Community, and Influence of Air Mass Trajectories

Caihong Xu, Min Wei, Jianmin Chen, Chao Zhu, Jiarong Li, Xianmang Xu, Wenxing Wang, Qingzhu Zhang, Aijun Ding, Haidong Kan, et al.

▶ To cite this version:

Caihong Xu, Min Wei, Jianmin Chen, Chao Zhu, Jiarong Li, et al.. Profile of Inhalable Bacteria in PM 2.5 at Mt. Tai, China: Abundance, Community, and Influence of Air Mass Trajectories. Ecotoxicology and Environmental Safety, 2019, 168, pp.110-119. 10.1016/j.ecoenv.2018.10.071. hal-02460329

HAL Id: hal-02460329 https://hal.science/hal-02460329v1

Submitted on 30 Jan 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

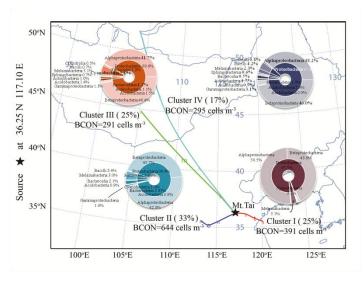
Profile of Inhalable Bacteria in PM_{2.5} at Mt. Tai,

2

China: Abundance, Community, and Influence of Air

3	Mass Trajectories						
4	Caihong Xu ¹ , Min Wei ² , Jianmin Chen ^{1,3,4,5*} , Chao Zhu ¹ , Jiarong Li ¹ , Xianmang Xu ³ ,						
5	Wenxing Wang ³ , Qingzhu Zhang ³ , Aijun Ding ⁴ , Haidong Kan ⁵ , Zhuohui Zhao ⁵ , Abdelwahid						
6	Mellouki ^{1,3,6}						
7	¹ Shanghai Key Laboratory of Atmospheric Particle Pollution and Prevention (LAP ³), Fudan Tyndall						
8	Centre, Department of Environmental Science & Engineering, Fudan University, Shanghai 200433,						
9	China						
10	² College of Geography and Environment, Shandong Normal University, Jinan 250100, China						
11	³ Environment Research Institute, School of Environmental Science and Engineering, Shandong						
12	University, Ji'nan 250100, China						
13	⁴ Institute for Climate and Global Change Research, School of Atmospheric Sciences, Nanjing						
14	University, Nanjing 210023, Jiangsu, China						
15	⁵ School of Public Health, Key Lab of Public Health Safety of the Ministry of Education and NHC Key						
16	Laboratory of Health Technology Assessment, Fudan University, Shanghai 200032, China						
17	⁶ Institut de Combustion, Aérothermique, Réactivité et Environnement, CNRS, 45071 Orléans cedex 02						
18	France						
19	* Corresponding author						
20	E-mail address: <u>jmchen@fudan.edu.cn</u> . (J. M. Chen)						
21	Highlights						
22	1. PM _{2.5} samples exhibited a high microbial biodiversity and complex chemical composition						
23	2. Relationships between bacterial populations and PM composition were investigated.						
24	3. The highest bacterial concentration and more potential pathogens were recorded under the						
25	influence of local air pollutants.						
26	4. The long-range transported air mass exhibited highest richness and diversity with an						
27	increased abundance of dust-borne bacteria.						

28 Graphical abstract



29

30

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

Keywords

31 Bioaerosol, PM, Air mass origin, Diversity

Abstract

Bacteria are ubiquitous in the near-surface atmosphere where they constitute an important component of aerosols with the potential to affect climate change, ecosystems, atmospheric process and human health. Limitation in tracking bacterial diversity accurately has thus far prevented the knowledge of airborne bacteria and their pathogenic properties. We performed a comprehensive assessment of bacterial abundance and diverse community in PM_{2.5} collected at Mt. Tai, via high-throughput sequencing and real-time PCR. The samples exhibited a high microbial biodiversity and complex chemical composition. The dominating populations were gram-negative bacteria including Burkholderia, Delftia, Bradyrhizobium, and PM mass Methylobacterium. The concentration, chemical composition, bacterial concentration and community structure varied under the influence of different air-mass trajectories. The highest mass concentration of PM_{2.5} (61 µg m⁻³) and major chemical components were recorded during periods when marine southeasterly air masses were dominant. The local terrestrial air masses from Shandong peninsula and its adjacent areas harbored highest bacterial concentration loading (602 cells m⁻³) and more potential pathogens at the site. In contrast, samples influenced by the long-distance air flow from Siberia and Outer Mongolia were found to have a highest richness and diversity as an average, which was also marked by the increase of dust-associated bacteria (Brevibacillus and Staphylococcus). The primary research may serve as an important reference for the environmental microbiologist, health workers, and city planners.

52 1. Introduction

The particulate matter with aerodynamic diameters of \leq 2.5 µm (PM_{2.5}) is known as a major health risk for the spread of asthma, stroke burden, and chronic diseases such as nasopharyngitis, bronchitis, and pneumonia¹⁻³. Assessment of air pollution exposure for the global burden of disease revealed that about 29.2% of stroke burden was associated with inhaled particulate matters at a high concentration⁴. The environmental health scholars considered that the toxic effect of PM_{2.5} on the respiratory tract and lungs mainly induced by the suspension and reaction of gases, heavy metals, mineral element, and toxic organic matters (cancerogenic substance) in ambient air^{5, 6}. In addition to these components, there is increasing evidence that pathogenic and allergenic microorganisms (bacteria, eukaryotes, archaea, and viruses) attached to the particles, may deposit in the epithelial cells through respiration, and thus causing various ailments in humans and livestock⁷⁻¹⁰. The pathogenic species (e.g. adenovirus, influenza, SARS, and rhinovirus) and their components (e.g. endotoxins, glucans, and mycotoxins) widely distributed in near-surface atmosphere bring a great threat to the ecosystem balance¹¹⁻¹³.

Bacteria, approximately 10⁴ cells m⁻³ and 0.1 µg m⁻³ in PM, mainly originated from the emission of soil, vegetation, surface of rock, human and animals (excrements and debris from skin and hair), precipitation (surface snow and ice core), and ocean¹⁴. Investigations on the ambient bacteria experienced a recent growth in interest with the aerosol community. To our knowledge, the first study on airborne microorganisms was reported in the mid-19th century¹⁵. The occurrence of high throughput sequencing offers a unique possibility to examine the bacterial, viral, and eukaryotic diversity in the aerosol. While the understanding of airborne microbe is still being developed due to current challenges, e.g., low density of airborne microbes, variability in airborne microbial community structure, and the lack of unified regulations for the research. However, in spite of this, this method has been applied to study the airborne bacteria and the associated pathogens in Japan, USA, Korea, and China in the last few years^{7, 16-18}. Franzetti et al. (2011)¹⁹ described the significant seasonal variation of bacterial community associated with PM₁₀ and PM₂₅ based on the Roche 454 sequencing. Based on the high-throughput sequencing, Bower et al. (2013)²⁰ found the significant seasonal variability of Actinobacteridae and Pseudomonadales across PM_{2.5} and PM_{2.5-10} in Greeley, CO. Cao et al. (2014)²¹ developed a comprehensive framework of the microbial community in PM_{2.5} and PM₁₀ under sever smog events by metagenomics. These findings mainly dressed the scanty composition community of bacteria in the urban air, the identification for the inhaled microbes and their pathogenic members at high altitudes were incomplete. Airborne bacteria with active metabolic have been reported in the mountains and the upper troposphere 16,22. These species dissolving into cloud droplets proposed an important impact on

the regional precipitation distribution, biogeochemical cycling of elements in ecosystem processes, and cloud chemistry by acting as cloud condensation nucleation or ice nucleation^{23,24}. Thus it is necessary to further the understanding for the profile of airborne bacterial community structure at high elevations.

Here, we expound on our initial efforts to advance the knowledge of airborne bacteria over Mt. Tai, the highest site at the North China Plain. It is located on the transport path of the Asian continent outflow and be affected by air mass from the Asian dust area. Mt. Tai is a tilted fault block mountain with height increasing from the north to the south, facing to the East China Sea and back on the yellow river to the west. In this region, the airborne bacterial populations in cloud system and fungal populations in particles have been reported before ²⁴⁻²⁶. The preponderant gram-negative bacteria inhabited in the cold and leaf-surface environments which were involved in the cloud condensation/ice nuclei formation and pathogenicity, demonstrating their importance in ecology, epidemiology and human health²⁴⁻²⁵. The pioneering studies were lack of a comprehensive and compound research on the bacteria in particles. The chemical composition in particles have been proved to be related to the variable airborne microbe. Further, they also highlighted the crucial influence of environmental factors humidity, and temperature on the bacterial community in cloud water and fungal community in particles. Generally the chemical composition of particles may be influenced by different air mass origin with the input of local aerosol or long-range transported aerosol. Whether the different air trajectories do play a role in shaping bacterial community of particles still remains a question. In this study, we aim to: (1) profile the bacterial abundance and community of PM_{2.5} at Mt. Tai, China, (2) to elucidate the environmental variables (temperature, humidity, PM_{2.5}, PM_{2.5} chemistry), and the air mass trajectories during sampling time, (3) to estimate whether the different air mass trajectories effect on the bacterial characteristics.

2. Materials and method

88

89

90 91

92

93

94

95

96

97

98

99

100101

102

103

104

105

106

107

108

109

110

111

112

113

115

116

117118

119

120

121

122

114 2.1 Aerosol collection and DNA Extraction

Fine particulate matter (PM_{2.5}) were sampled on quartz fiber filters (PALL, NY, U.S. 88mm) with a middle-volume sampler (TH-100A, Wuhan Tianhong Instruments Co., Ltd., China) and a flux speed of 100 L min⁻¹. The blank filters were obtained by placing a sterilized quartz microfiber filter inside of the sampler without operation. The sampling campaigns were carried out in 8~13 day periods in summer, autumn, and winter during July 2014 to August 2015. The sampler was located at the summit of Mt. Tai (36.25°N and 117.10°E, 1534 m above sea level), a typical tourist area in the North China Plain (Figure 1). The geographical characteristics of the sampling site have been previously described^{25,26}. Each

collection period lasted 23 h. Before sampling, all the filters were baked at a Muffle furnace at 500°C for 5 h and then placed into sterilized aluminum foil before deposited into a sealed bag. To avoid the contamination, the sampling filter holder and materials used for changing filters were treated with 75% ethanol before sampling every day. After sampling, the samples were stored at -80 °C until the next analysis. The environmental factors including temperature, relative humidity, visibility, water-soluble inorganic ions (NO₃-, SO₄--, NH₄+, Ca²⁺, Na⁺, Mg²⁺, and Cl⁻), meteorological data, the mass concentration of PM_{2.5}, DNA extraction procedure and PCR amplification have been described before²⁶. The final product was ready for resolving for the abundance and community of airborne bacteria by real-time quantitative PCR and Illumina Miseq sequencing.

2.2 Quantification of 16S rRNA gene copy number

The bacterial abundance was determined by the quantification of the copy numbers of the gene 16S rRNA. This value is not directly related to bacterial number due to the presence of multiple ribosomal operons in the bacterial genomes; however, it can be used to look at relative shifts in microbial biomass as the bias in copy number is likely constant across samples. The V4-V5 hypervariable regions were PCR-amplified with universal primers 515F (5'-TGCCAGCMGCCGCGGTAA-3') and 926R (5'-CGTCAATTCMTTTGAGTTT-3'). The real-time quantitive PCR was performed in a total volume of 25 µL reaction mixture including 12.5 Ml TransStart Green qPCR SuperMix (Transgene, AQ101-03), 1µL forward primer, 1µL reverse primer, 5 µL DNA, and 5.5 µL double distilled H₂O. The amplification was run on a BioRad CFX96 Real-Time PCR system under the following conditions: 95 °C for 5 min, followed by 40 cycles of 95 °C for 30 s, 56 °C for 30 s and 72 °C for 30 s, with acquisition of the fluorescence at the end of each 72 °C elongation step. The deionized water was employed as the negative controls and was pipetted into the wells in a 96-well microplate with the DNA extracted from the samples. All the samples and controls were tested in triplicate in each run. Based on the average rRNA gene copy number (3.98), the bacterial cell concentration (cells m⁻³) was calculated using the method described before²⁷⁻²⁹.

2.3 Sequence Analyses

A DNA sequencing library was constructed on the basis of the TruSeq DNA PCR-Free Sample Prep Kit (Illumina) and was sequenced by the MiSeq reagent kit v2 (Illumina) on an Illumina MiSeq instrument (Illumina, San Diego, CA, USA). The raw sequences were analyzed using standard Mothur software. Variable sequence numbers (7044 to 32,293) were obtained after the quality control of raw reads by removing the chimeric sequences, primers, barcodes, and low-quality reads (average quality score<20) using FASTX-ToolKit. We

normalized the sequences number to 7044 for the subsequent analysis. The Operational Taxonomic Units (OTUs) were defined using a distance-based cutoff method by Usearch (version 7.1; http://drive5.com/uparse/). The alpha-diversity estimators Chao1 and Shannon indexes, which indicated the richness and diversity of the bacterial community, were calculated by QIIME (version 1.17; http://qiime.org/scripts/assign_taxonomy.html) after removing the low-abundance OTUs (<0.01%). The taxonomic classification was aligned to the Silva database (http://www.arb-silva.de/) uCLUST using (http://www.drive5.com/uclust/downloads1_2_22q.html) at different similarity levels to determine the diverse bacteria at phylum, class, order, family, genus, and species levels. The sequences obtained in this study have been deposited in the Sequence Read Archive (SRA) under accession numbers SRR51461. The potential pathogenic sequences corresponding to human health were screened referenced to the previous database³⁰⁻³³. The human pathogenic bacteria list, including species names and the diseases caused, was used to find the potential pathogens sequences from total sequences. We selected the operational taxonomic units (OTUs) belonged to these genera mentioned in the reference list. The nearest phylogenetic species were obtained by NCBI BLAST. Finally, the bacterial species belonging to the reference list were determined as the potential pathogens in PM_{2.5}.

2.4 Statistical Analyses

157

158

159

160

161

162

163

164

165

166

167

168169

170

171

172

173

174

175

176

177

178

179

180

181

182

183

184

185

186

The difference in bacterial concentration across different air mass trajectories were assessed by an ANOVA test, followed by post-hoc tests. The ANOVAs test was also conducted to determine whether season had a significant impact on the relative abundances of specific microbial groups. The weighted UniFrac analysis was employed to estimate the significant differences of the microbial community among three seasons. The relationship between the airborne microbial concentrations and environmental factors including PM concentrations and chemical compositions was assessed with a Pearson correlation by SPSS 16.0. The results that Bonferroni adjusted P-value for each pairwise comparison higher than 0.05 were considered to be significant. The correlation coefficient between NMDS plot and environmental elements was calculated via metadata analysis with Mothur software. Statistical values, including Pearson and Spearman correlation coefficients and independent t-tests, were calculated using R software with the R package.

187 3. Results and discussion

188

216

217

218

219

3.1 Bacterial abundance in PM_{2.5}

189 During the sampling period, the PM_{2.5} mass concentration, ranging from 3 to 111 µg m⁻³ with a mean value of 40 µg m⁻³ at Mt. Tai, were much lower than that in summer 2006 190 $(123 \text{ ug m}^{-3})^{32}$ and comparable with that in summer 2007 (59 $\mu \text{g m}^{-3})^{30}$ that conducted at the 191 same site. Table 1 summarizes the corresponding bacterial concentrations in a different size. 192 193 The bacterial concentration of PM_{2.5} varied from 40 to 1728 cells m⁻³ with an average of 490 194 cells m⁻³ at Mt. Tai. It was much lower than those TSP from the high-altitude air, urban Japan, 195 Korea, and Italy, PM_{2.5} and PM₁ from China. The lower bacterial concentration were found to 196 be correlated with PM_{2.5} mass concentration (r = 0.526, p < 0.05), SO₄²⁻ (r = 0.549, p < 0.01), NO_3^- (r=0.458, p < 0.05), NH_4^+ (r=0.570, p < 0.01), K^+ (r=0.508, p < 0.01), Na^+ 197 (r = 0.585, p < 0.01), Mg²⁺ (r = 0.608, p < 0.01), and Ca²⁺ (r = 0.432, p < 0.05) significantly 198 199 based on the Spearman rank correlations. The ambient pollutants attached to the particles may 200 provide nutrients (sulfur, nitrogen, and ammonia) and thus affect the bacterial characteristics. In general, the airborne bacteria, ranging between 10 and 10⁷ cells m⁻³, varied in a degree with 201 many aspects 11,23,29,34, like sampling time, site (altitude, temperature, humidity, etc.), sample 202 203 type (TSP, PM₁₀, PM_{2.5}, PM₁, cloud/fog water, etc.), and analysis methods (culture methods, bio-molecular technology, and quick estimation model)^{14,35,36}. And these indexes effectively 204 represented the situation of bacterial community on particles under different conditions. 205 206 Borodulin et al. (2005)³⁷ observed the varied concentration of culturable microorganisms for 207 about more than two orders of magnitude over an altitude range of 0.5~7 km in southwestern 208 Siberia. They found the overall bacterial concentration in the atmosphere where not only 209 dependent on altitude but also related to the cell size of bacteria and particulate matters. 210 Different bacteria have respective sizes ranging from 0.1~5 µm, and each bacterium has a 211 particular shape feature that plays an important role in the adhesion on PM³. Since the study environments were breeding settings in which bacteria had better conditions to grow and 212 propagate, the bacteria concentration may decrease with decreasing particulate size³⁸. Hence 213 214 more high-throughput sequencing researches for the bacterial community distribution were 215 vital to understanding their diversity fully.

3.2 Inhalable bacterial community in PM_{2.5}

After quality control and normalization, a number of OTUs ranged from 237 to 1579 were detected. The alpha diversity estimators were summarized in Table 2. Similar with Despr´es et al. (2007)⁵⁴, the airborne bacteria at Mt. Tai commonly exhibited lower richness and

diversity compared with those from urban, marine, sand, and dust sites. The diversity estimators can reflect the state of the bacteria and their relationship to the environment, and microbial resources developments, ecosystem functioning, global change and human health are all closely related to microbial diversity. In addition to the changing sampling sites, this discrepancy may be caused by the different analysis platform (Miseq sequencing, pyrosequencing, and clone library). Using the same method (Miseq Sequencing), TSP seems to have higher OTUs, chao1, and Shannon indexes than PM2.5 and PM1, indicating more and various bacterial species in the larger particulate matter. We attribute this increasement to the larger surface area of TSP for bacterial adhesion. As mentioned above, the size of a single bacterial cell is smaller than particles, there is a mass of nutrients and bacterial slime coat on the bacteria that make the bacterial size bigger but still smaller than particles, so there were more bacteria presenting in particles with a larger size. Actually different bacteria had respective variation tendency for particle sizes, although there were some differences on size distribution, bacteria mainly attached to particles with two peaks (1.1~2.1 µm and >7.1 µm)⁵⁵. Except for the bacterial size, the structure of bacterial community can also exert a certain extent influence on the diversity characteristics.

220

221

222

223

224

225

226

227

228

229

230

231

232

233

234

235

236

237

238

239

240

241

242243

244

245

246

247248

249

250

251

252

253

254255

At Mt. Tai, the diverse bacterial community including 40 phyla, 78 classes, 154 orders, 286 families, 724 genera was classified in PM_{2.5}. About 97.82% of OTUs belongs to the phylum Proteobacteria, Cyanobacteria, Firmicutes, and Bacteroidetes. The co-occurrence network were used to reveal the complex bacterial community structure based on the top 100 OTUs (Figure 2). The species of dominant phyla (Proteobacteria, Actinobacteria, Cyanobacteria, Firmicutes, Acidovacteria, Fusobacteria, and Bacterioidetes) were similar in USA^{7,56}, Italy³⁴, and East Asian Regions 15,21,48,49,51,56, but the relative abundance were different. Furthermore, the microbiome is a dynamic aspect of the near-surface layer among extreme weather incidents such as dust, haze, rain, and tropical storms. Proteobacteria members were the dominant species in the atmosphere over mountains and from the troposphere. The relative abundance of this bacterium (91.42%) were richer than that in the dust (69.0%), nondust (53.5%)¹⁸, and haze (16.5%)²⁷ days. It was dominated by a few taxa throughout the whole sampling period, with a marked prevalence of Alphaproteobacteria, Betaproteobacteria, and Gammaproteobacteria. At higher taxonomic levels, a bacterium in the nitrogen fixing, gram-negative pathogenic bacteria genus Burkholderia appeared to be the most abundant. Bowers et al. (2009)⁵⁷ also showed the dominant Burkholderiales in particles collected at the top of Mt. Werner near Steamboat Springs CO, USA at 3220 m above sea level via tagged pyrosequencing. The preponderant gram-negative bacteria in PM_{2.5} were different with Fang et al. (2007)⁵⁸ who reported the 80%~86% culturable bacteria in outdoors environment were gram-positive. They considered the gram-negative bacteria cannot adapt to the higher air

pollutants concentration, stronger solar radiation, and complicated meteorological environment outdoors. We speculate this gap attributed to the different analysis platform. The Illumina Miseq sequencing method gained the comprehensive airborne bacteria no matter culturable or unculturable. Actually, the proportion of culturable bacteria is very small. Another reason is that those gram-negative bacteria in the atmosphere lose cultivability after aerosolization⁵⁹. Hence the majority of airborne bacteria outdoors were gram-negative.

Out of all the gram-negative species, eight opportunistic pathogens were determined based on the human pathogenic bacteria list as described before. *Acinetobacter baumannii*, primarily associated with hospital-acquired infections, commonly found in water and soil. Over the past decades, reports on community-acquired *A. baumannii* infections (wound infection, ventilator-associated pneumonia, and bloodstream infection) have increased and attracted public attention ⁶⁰⁻⁶³. An outbreak of soil-borne bacteria *Pseudomonas putida* was involved in clinic infections in patients with medical devices and immunosuppressed hosts ⁶⁴⁻⁶⁵. Besides, five gram-positive potential pathogens were detected including *Bacillus subtilis*, *Rhodococcus equi*, *Streptococcus anginosus*, *Bacillus pumilus*, *and Rhodococcus fascians*. Some strains of *B. subtilis* cause food poisoning through the ropy spoilage of bread and rots in potatoes ⁶⁶⁻⁶⁷. *R. equi*, mainly thrive on water and soils, were connected with pneumonia in immunocompromised patients and animal hosts ⁶⁸⁻⁶⁹. *S. anginosus* were able to cause diseases including bloodstream infections, pyogenic liver abscess, brain and liver abscesses ⁷⁰⁻⁷¹. These bacterial species suspending in the atmosphere diffuse downwind, yielding a threat to the health of plants and animals in local and downstream area.

Other than the pathogenicity, the diverse bacteria demonstrates a diverse metabolism in the atmospheric process and ecosystem interactions. The bacteria with ice nucleation activity suspending in the atmosphere can take part in the cloud chemistry via act as cloud condensation or ice nuclei. During the same project, we captured some typical cloud events (Event F and G)²⁴, incidentally. The special terrain of Mt. Tai is conducive to the formation of orographic clouds. The tiny droplets could dissolve or adsorb the particles suspending in the atmosphere and then accumulated into the cloud water. Though the same sampling site, the dominant bacterial species varied from *Burkholderia* in PM_{2.5} to the preponderance of *Pseudomonas* in cloud water. The distant discrepancy of dominating bacterial distribution also found between PM_{2.5} of Mt. Tai and PM₁ of Ji'nan (Figure 3A). As to this phenomenon, different bacterial suitable inhabits and growing media (water, soil, atmosphere, etc.) may be important factors. Additionally, Liu et al. (2018)⁷² considered the relative humidity, wind scale, CO, and SO₂ were the main factors, which affected the diversity of total bacteria and the proportion of pathogenic bacteria. The PM's chemical components serve as the

nutritional ingredient, coupled with surrounding environments such as temperature, humidity, wind direction, and atmospheric pressure seems to shift on the bacterial growth and metabolism in atmosphere^{3,10,73}. The chemical and biological data were jointly processed and verified that the varied NH₄⁺, NO₃⁻, Mg²⁺, Ca²⁺, Cl⁻ and Na⁺ explained about 53.7%~63.1% of the variance of species-environment correlations⁷⁴. The chemical composition of PM_{2.5} correlated with the air mass back trajectories. Air mass originating from Mongolia region had strong correlations with dust pollution, along with high concentration of Ca²⁺ in ambient PM_{2.5}. Local emission and air pollutants were more favorable for formation of secondary pollutants (SO₄²⁻, NH₄⁺, and NO₃⁻) and reactions between particulate species. We also detected the obvious correlation between airborne bacterial community and Ca²⁺, Mg²⁺, and temperature at Mt. Tai, while the influence of atmospheric circulation using the backtrajectory analysis was not evaluated fully. Therefore how does the air mass trajectories vary and whether the air mass trajectories evoked the change of concentration and structure of bacterial community is worth discussing.

3.3 Cluster analysis of backward trajectory and PM_{2.5} chemistry

At Mt. Tai, the air mass back trajectories have been categorized into four types, typified by the examples in Figure 4, based on trajectories terminating at the site on the day of measurements. One trajectory per day was, in our experience, adequate to represent the flow patterns which tended to change over timescales of a few days. Inevitably, there were a few days on which assignment of trajectory type was difficult, but the predominant type from the four synoptic hours was adopted in these circumstances. The four types of back trajectory were as follows:

Cluster I (25%): The air mass had passed across the marginal seas in the western Pacific Ocean and had been in contact with the land immediately prior to arrival in the study area. Broadly speaking, the highest mass concentration of $PM_{2.5}$ (61 µg m⁻³) and major water inorganic ions were recorded followed the decreasing order: $SO_4^{2-} > NO_3^{-} > CI^{-} > Na^{+} > Ca^{2+} > K^{+} > NH_4^{+} > Mg^{2+}$. In addition to the secondary inorganic ions, CI^{-} (4.35 µg m⁻³) and Na^{+} (3.73 µg m⁻³) can be assumed mostly to originate from sea salt aerosols. The CI^{-}/Na^{+} molar ratios obtained (0.75) was lower than that of bulk seawater (1.18)⁷⁵, suggesting that the chlorine loss of marine aerosols during transport to some extent. These ions are proved to play essential roles in atmospheric processes. e.g., cloud formation, visibility degradation, solar radiation, acidification of cloud, rain, fog and haze formation and served as the nutrients for the growth and development of airborne bacteria^{24, 45}.

Cluster II (33%): The air mass had come in from a generally western direction bringing it across Shandong peninsula and its adjacent areas, a typical densely polluted areas of northern China. Influenced by the local terrestrial air flow, the average of PM_{2.5} mass was 37 µg m⁻³. The secondary inorganic ions, i.e., $SO_4^{\ 2^-}(9.92~\mu g~m^{-3}),~NO_3^{\ -}(4.23~\mu g~m^{-3}),~and~NH_4^{\ +}(2.29~\mu g~m^{-3})$ m⁻³), which mostly originated from the transformation of their precursors (SO₂, NO₂, and NH₃) were found as the dominant species, followed by Cl⁻ (0.75 µg m⁻³), K⁺ (0.63 µg m⁻³), Ca²⁺ (0.51 µg m^{-3}) , Na⁺ (0.31 µg m^{-3}) , and Mg²⁺ (0.07 µg m^{-3}) . In order to investigate the continental influence, non-sea-salt sulfate (nss-SO₄²⁻) concentrations are estimated as: nss – SO_4^{2-} (µg m⁻³) = [SO_4^{2-}] – 0.245 × [Na⁺]. The mass fraction of nss- SO_4^{2-} ranged from 1.17 μ g m⁻³ to 18.89 μ g m⁻³ (mean 9.85 μ g m⁻³), accounted for 99.02% of total sulfate. SO_4^{2-} mainly originates from anthropogenic and biogenic sources, while nss- SO_4^{2-} originates mainly from stationary sources. We often use the mass ratio of [NO₃⁻]/[nss-SO₄²⁻] as an indicator of the relative importance of stationary versus mobile sources of sulfur and nitrogen in the atmosphere. High [NO₃]/[nss-SO₄²] mass ratios represent the predominance of mobile sources over stationary sources. Due to the extensive use of sulfur-containing coal, the average ratio [NO₃]/[nss-SO₄²] mass ratios in China were ascribed lower value⁷⁶. The measured [NO₃⁻]/[nss-SO₄²] mass ratios at Mt. Tai (0.25) was higher or comparable with those reported in other cities, e.g., 0.17 in Hongkong⁷⁷, 0.20 in Guangzhou⁷⁸ and 0.35 in Oingdao⁷⁹. However, the ratio was lower than those cities where the use of coal was limited, for example, Los Angeles (2.0)⁸⁰ and Seoul (0.73)⁸¹. The result implied that stationary source emissions were likely more important than mobile sources at Mt. Tai under the influence of local terrestrial air flow.

324

325

326

327

328 329

330

331

332

333

334

335336

337

338

339

340 341

342

343344

345

346

347

348

349

350

351

352

353

354

355

356

357

Cluster III (25%) and Cluster IV (17%): The air mass arrived from a northwesterly sector outer Mongolia and having crossed dust area (Inner Mongolia and Shanxi Province) and high anthropogenic emission area (Henan and Shandong Province). This type air mass moves quickly and the corresponding mass concentration of coarse particulate matter was usually high. Typically, these westerly and northwesterly trajectories had not been in contact with any land for several days prior to arriving at the site.

3.4 Influence of air mass trajectories on bacterial populations and community

Obviously, no obvious variation for the bacterial concentration was observed under the influence of air mass trajectories. The terrestrial air masses from the Shandong peninsula and its adjacent areas exhibited a little higher bacterial concentration loading (644 cells m⁻³) and more potential pathogens. While the bacterial concentration of PM influenced by the marine air mass was lower, though the highest chemical composition (Figure 4). In contrast, samples

influenced by the long-distance air flow from Siberia and Outer Mongolia were found to have a highest richness and diversity as an average (Table 2). The effects of long-transported air parcels on the aerosol particles and its composition and sources have been discussed in previous studies⁸²⁻⁸⁵. The Asian dust particles blow from these areas in the westerly wind and play an important role in the leeward ecosystem. The bacterial community at phyla, class, and order levels under the influence of four air mass trajectories was shown in Figure 5. However, the bacterial abundance made no significant difference among the top 20 OTUs (Figure 3B), suggesting the stable community structure of airborne bacteria. Yet, some non-dominant bacterial groups that exhibited pronounced changes in relative abundance across the different air mass trajectories (Figure 6) can be linked to specific source environments of PM_{2.5}. The long-range transported air parcels seem to be able to harbor diverse and dynamic microbial populations⁵³. Kakikawa et al., (2008) verified the injection of dustborne bacteria (Brevibacillus, Staphylococcus, Rhodococcus, Delftia, Pseudomonas, and Agrobacterium tumefaciens) into the atmosphere during the dust event⁸⁶. Maki et al., (2017) also observed the variable microbial community in response to the Asian dust events²². The number of microbial particles at high altitudes (>1200m) increased about 5.5-folds. The corresponding bacterial community was predominantly composed of natural-sand/terrestrial bacteria during dust events instead of anthropogenic/terrestrial bacteria, marine bacteria, and plant-associated bacteria during non-dust events. The prevalent existence of marine bacteria in cloud droplets, the air masses suspended from the Yellow Sea over the marine environment harbored more proteobacterial populations, Pandoraea, and Phyllobacterium associated with plant bodies or surface. The prevalence of *Pandoraea* had a significant positive relationship with the highest content of $SO_4^{2-}(**p<0.01)$. Consistent with these findings, the long-range transported westerly air mass increase the diversity and richness of airborne bacterial community, with the increase of some dustborne species such as genus Brevibacillus, Staphylococcus, and Anoxybacillus (Figure 6A). The Bacilli members obtained from mineral dust particles over downwind areas were positively correlated to temperature and Cl in PM_{2.5} (**p<0.01), excepting the obvious effect of humidity and temperature. They showed a stronger ability to resist ultraviolet light and be able to survive in the atmosphere by the formation of resistant endospores. The natural dust particles from dust areas can be transported to the free troposphere and vertically mixed with the anthropogenic particles during the transportation processes. Together, the airborne bacteria community at Mt. Tai contains a mixture of dustborne bacteria, terrestrial bacteria, marine bacteria, and others. The community of dominant bacteria in PM_{2.5} was relatively stable, and little affected by the sampling environment. In other words, ambient bacterial community structure were stable in atmosphere. Four air mass trajectories from various origin environments may lead to the discrepant composition in PM_{2.5},

358

359

360

361362

363

364

365

366

367

368

369

370

371372

373

374

375

376

377

378

379

380 381

382

383

384

385

386

387

388

389

390

391

392

and then driving the shift on the pronounced changes observed in some non-dominant bacterial groups.

4. Conclusions

394

395

396

397

398

399

400

401

402

403

404

405

406

407

408

409

410

411

412

413

414

415

416

418

In this study, 16S rRNA gene sequencing was conducted to study the bacterial communities at Mt. Tai's PM_{2.5} pollutants from July 2014 to August 2015. Results demonstrated the lower bacterial concentration and diverse bacterial community in PM_{2.5} compared with some aforementioned studies. We also detected thirteen potential pathogenic species, such as A. baumannii, P. putida, B. subtilis, and R. equi which effect on the immunocompromised populations and livestock. In future studies, we intend to analyze the changing bacterial community structure affected by different air mass trajectories, combined with the meteorological data, PM mass concentration, water soluble ion and other physical and chemical properties. The highest mass concentration of PM_{2.5} and major chemical components was recorded during periods when marine southeasterly air masses were dominant. The local terrestrial air masses from Shandong peninsula and its adjacent areas harbored highest bacterial concentration loading and more potential pathogens at the site. In contrast, samples influenced by the long-distance air flow from Siberia and Outer Mongolia were found to have a highest richness and diversity as an average, which was also marked by the increased abundance of dust-associated bacteria (Brevibacillus and Staphylococcus). We conclude that the air flow from different origin environments may lead to the discrepant chemical composition in PM_{2.5}, and then driving the shift on the pronounced changes observed in some non-dominant bacterial groups. We hope our research can provide for some reference value in the global air microbial propagation and air microbial research at higher elevation site.

Conflict of interest

The authors declare that they have no competing financial interests.

Acknowledgements

- 419 This work was supported by the Ministry of Science and Technology of China (No.
- 420 2016YFC0202700), the National Natural Science Foundation of China (No. 91743202,
- 421 21527814). This project has received funding from the European Union's Horizon 2020
- research and innovation programme under grant agreement No 690958.

423 References

- 1. Bell, M. L.; Ebisu, K.; Peng, R. D.; Walker, J.; Samet, J. M.; Zeger, S. L.; Dominici, F.,
- 425 Seasonal and regional short-term effects of fine particles on hospital admissions in 202 US
- 426 Counties, 1999–2005. American Journal of Epidemiology **2008**, 168: 1301-1310.
- 427 2. Laden, F.; Neas, L. M.; Dockery, D. W.; Schwartz, J., Association of fine particulate
- 428 matter from different sources with daily mortality in six U.S. cities. Environmental Health
- 429 *Perspectives* **2000,** *108*: 941-947.
- 430 3. Zhai, Y.; Li, X.; Wang, T.; Wang, B.; Li, C.; Zeng, G., A review on airborne
- 431 microorganisms in particulate matters: composition, characteristics and influence factors.
- 432 *Environment International* **2018,** *113*: 74-90.
- 433 4. Brauer, M.; Freedman, G.; Frostad, J.; van Donkelaar, A.; Martin, R. V.; Dentener, F.; van
- Dingenen, R.; Estep, K.; Amini, H.; Apte, J. S., Ambient air pollution exposure estimation for
- 435 the global burden of disease 2013. *Environmental Science and Technology* **2016,** *50*: 79-88.
- 5. Kleinman, M. T.; Bufalino, C.; Rasmussen, R.; Hyde, D.; Bhalla, D. K.; Mautz, W. J.,
- Toxicity of chemical components of ambient fine particulate matter (PM_{2.5}) inhaled by aged
- 438 rats. *Journal of Applied Toxicology* **2000**, 20: 357-364.
- 439 6. Maaike, S.; Ilse, G.; Maciej, S.; Godri, K. J.; Gerard, H.; Cassee, F. R.; Mudway, I. S.;
- Kelly, F. J.; Harrison, R. M.; Erik, L., In vitro toxicity of particulate matter (PM) collected at
- 441 different sites in the Netherlands is associated with PM composition, size fraction and
- oxidative potential-the RAPTES project. *Particle and Fibre Toxicology* **2011**, 8: 1-15.
- 7. Brodie, E. L.; DeSantis, T. Z.; Parker, J. P.; Zubietta, I. X.; Piceno, Y. M.; Andersen, G. L.,
- 444 Urban aerosols harbor diverse and dynamic bacterial populations. *Proceedings of the National*
- Academy of Sciences of the United States of America **2007**, 104: 299-304.
- 8. Xie, J. E.; Jin, L.; Luo, X. S.; Zhao, Z.; Li, X. D., Seasonal disparities in airborne bacteria
- and sssociated antibiotic resistance Genes in PM2.5 between Urban and Rural Sites.
- 448 Environmental Science and Technology Letters 2018, 5: 74-79.
- 9. Zhou, H.; Wang, X. L.; Li, Z. H.; Kuang, Y.; Mao, D. Q.; Luo, Y., Occurrence and
- 450 distribution of urban dust-associated bacterial antibiotic resistance in Northern China.
- 451 Environmental Science and Technology Letters 2018, 5: 50-55.
- 452 10. Gao, X. L.; Shao, M. F.; Wang, Q.; Wang, L. T.; Fang, W. Y.; Ouyang, F.; Li, J.,
- 453 Airborne microbial communities in the atmospheric environment of urban hospitals in China.
- 454 *Journal of Hazardous Materials* **2018**, *349*: 10-17.
- 455 11. Liu, H.; Zhang, Z. W.; Wen, N. J.; Wang, C., Determination and risk assessment of
- airborne endotoxin concentrations in a university campus. Journal of Aerosol Science 2017,
- **457** *115*: 46-157.

- 458 12. Carty, C. L.; Gehring, U.; Cyrys, J.; Bischof, W.; Heinrich, J., Seasonal variability of
- endotoxin in ambient fine particulate matter. Journal of Environmental Monitoring 2003, 5:
- 460 953-958.
- 461 13. Chen, P. S.; Tsai, F. T.; Lin, C. K.; Yang, C. Y.; Chan, C. C.; Young, C. Y.; Lee, C. H.,
- 462 Ambient influenza and avian influenza virus during dust storm days and background days.
- 463 *Environmental Health Perspectives* **2010**, *118*: 1211-1216.
- 14. Despres, V. R.; Huffman, J. A.; Burrows, S. M.; Hoose, C.; Safatov, A. S.; Buryak, G.;
- 465 Froehlich-Nowoisky, J.; Elbert, W.; Andreae, M. O.; Poeschl, U.; Jaenicke, R., Primary
- 466 biological aerosol particles in the atmosphere: a review. Tellus Series B-Chemical And
- 467 *Physical Meteorology* **2012,** *64*: 145-153.
- 468 15. Ariatti, A.; Comtois, P., Louis Pasteur: the first experimental aerobiologist. Aerobiologia
- **1993,** *9*: 5-14.
- 470 16. Maki, T.; Kakikawa, M.; Kobayashi, F.; Yamada, M.; Matsuki, A.; Hasegawa, H.;
- 471 Iwasaka, Y., Assessment of composition and origin of airborne bacteria in the free
- troposphere over Japan. *Atmospheric Environment* **2013,** 74: 73-82.
- 473 17. Gou, H.; Lu, J. J.; Li, S. M.; Tong, Y. B.; Xie, C. B.; Zheng, X. W., Assessment of
- 474 microbial communities in PM₁ and PM₁₀ of Urumqi during winter. Environmental Pollution
- **2016,** *214*: 202-210.
- 476 18. Cha, S.; Lee, D.; Jang, J. H.; Lim, S.; Yang, D.; Seo, T., Alterations in the airborne
- bacterial community during Asian dust events occurring between February and March 2015 in
- 478 South Korea. *Scientific Reports* **2016**, *6*: 37271.
- 479 19. Franzetti, A.; Gandolfi, I.; Gaspari, E.; Ambrosini, R.; Bestetti, G., Seasonal variability of
- 480 bacteria in fine and coarse urban air particulate matter. Applied Microbiology and
- 481 *Biotechnology* **2011,** *90*: 745-753.
- 482 20. Bowers, R. M.; Clements, N.; Emerson, J. B.; Wiedinmyer, C.; Hannigan, M. P.; Fierer,
- N., Seasonal variability in bacterial and fungal diversity of the near-surface atmosphere.
- 484 Environmental Science and Technology 2013, 47: 2097-12106.
- 485 21. Cao, C.; Jiang, W.; Wang, B.; Fang, J.; Lang, J.; Tian, G.; Jiang, J.; Zhu, T. F., Inhalable
- 486 microorganisms in Beijing's PM_{2.5} and PM₁₀ pollutants during a severe smog event.
- 487 Environmental Science and Technology **2014**, 48: 1499-1507.
- 488 22. Maki, T.; Hara, K.; Iwata, A.; Lee, K. C.; Kawai, K.; Kai, K.; Kobayashi, F.; Pointing, S.
- 489 B.; Archer, S.; Hasegawa, H.; Iwasaka, Y., Variations in airborne bacterial communities at
- 490 high altitudes over the Noto Peninsula (Japan) in response to Asian dust events. *Atmospheric*
- 491 *Chemistry and Physics* **2017** *17*: 11877-11897.

- 492 23. Bowers, R. M.; McCubbin, I. B.; Hallar, A. G.; Fierer, N., Seasonal variability in
- 493 airborne bacterial communities at a high-elevation site. Atmospheric Environment 2012, 50:
- **494** 41-49.
- 495 24. Xu, C. H.; Wei, M.; Chen, J.; Sui, X.; Zhu, C.; Li, J. R.; Zheng, L. L.; Sui, G. D.; Li, W.
- 496 J.; Wang, W. X.; Zhang, Q. Z.; Mellouki, A., Investigation of diverse bacteria in cloud water
- 497 at Mt. Tai, China. Science of the Total Environment 2017, 580: 258-265.
- 498 25. Xu, C. H.; Wei, M.; Chen, J. M.; Zhu, C.; Li, J. R.; Lv, G. L.; Xu, X. M.; Zheng, L. L.;
- 499 Sui, G. D.; Li, W. J.; Chen, B.; Wang, W. X.; Zhang, Q. Z.; Ding, A. J.; Mellouki, A., Fungi
- diversity in PM_{2.5} and PM₁ at the summit of Mt. Tai: abundance, size distribution, and
- seasonal variation. *Atmospheric Chemistry and Physics* **2017**, *17*: 11247-11260.
- 502 26. Lv, G. L.; Sui, X.; Chen, J. M.; Jayaratne, R.; Mellouki, A., Investigation of new particle
- formation at the summit of Mt. Tai, China. Atmospheric Chemistry and Physics 2018, 18:
- **504** 2243-2258.
- 505 27. Xu, C. H.; Wei, M.; Chen, J. M.; Wang, X. F.; Zhu, C.; Li, J. R.; Zheng, L. L.; Sui, G. D.;
- Li, W. J.; Wang, W. X.; Zhang, Q. Z.; Mellouki, A., Bacterial characterization in ambient
- 507 submicron particles during severe haze episodes at Ji'nan, China. Science of the Total
- 508 Environment 2017, 580: 188-196.
- 509 28. An, H. R.; Mainelis, G.; White, L., Development and calibration of real-time PCR for
- quantification of airborne microorganisms in air samples. *Atmospheric Environment* **2006**, *40*:
- 511 7924-7939.
- 512 29. Seunghoon, L.; Hyojeong, L.; Sejin, K.; Hyungmin, L.; Kang, H. J.; Yongpyo, K.,
- 513 Identification of airborne bacterial and fungal community structures in an urban area by T-
- RFLP analysis and quantitative real-time PCR. Science of the Total Environment 2010, 408:
- **515** 1349-1357.
- 516 30. Zhou, Y.; Wang, T.; Gao, X. M.; Xue, L. K.; Wang, X. F.; Wang, Z.; Gao, J.; Zhang, Q.
- 517 Z.; Wang, W. X., Continuous observations of water-soluble ions in PM_{2.5} at Mount Tai
- 518 (1534 m a.s.l.) in central-eastern China. Journal of Atmospheric Chemistry 2009, 64: 107-127.
- 519 31. Guo, F.; Zhang, T., Profiling bulking and foaming bacteria in activated sludge by high
- 520 throughput sequencing. *Water Research* **2012,** *46*: 2772-2782.
- 521 32. Deng, C.; Zhuang, G.; Huang, K.; Li, J.; Zhang, R.; Wang, Q.; Liu, T.; Sun, Y.; Guo, Z.;
- Fu, J. S.; Wang, Z., Chemical characterization of aerosols at the summit of Mountain Tai in
- 523 Central East China. *Atmospheric Chemistry and Physics* **2011**, *11*: 7319-7332.
- 524 33. Ye, L.; Zhang, T., Pathogenic bacteria in sewage treatment plants as revealed by 454
- 525 pyrosequencing. *Environmental Science and Technology* **2011,** *45*: 7173-7179.
- 526 34. Bertolini, V.; Gandolfi, I.; Ambrosini, R.; Bestetti, G.; Innocente, E.; Rampazzo, G.;
- 527 Franzetti, A., Temporal variability and effect of environmental variables on airborne bacterial

- 528 communities in an urban area of Northern Italy. Applied Microbiology and Biotechnology
- **2013,** *97*: 6561-6570.
- 530 35. Fulton John, D., Microorganisms of the upper atmosphere III. Relationship between
- altitude and micropopulation. *Applied Microbiology* **1966**, *14*: 237-240.
- 36. Li, K. J.; Dong, S. F.; Wu, Y.; Yao, M. S., Comparison of the biological content of air
- samples collected at ground level and at higher elevation. *Aerobiologia* **2010**, 26: 233-244.
- 534 37. Borodulin, A. I.; Safatov, A. S.; Shabanov, A. N.; Yarygin, A. A.; Khutorova, O. G.;
- Belan, B. D.; Panchenko, M. V., Physical characteristics of concentration fields of
- tropospheric bioaerosols in the South of Western Siberia. *Journal of Aerosol Science* **2005**, *36*:
- **537** 785-800.
- 538 38. Zheng, W. C.; Zhao, Y.; Xin, H. W.; Li, B. M.; Gates, R. S.; Zhang, Y. H.; Soupir, M. L.,
- 539 Concentrations and size distributions of airborne particulate matter and bacteria in an
- experimental aviary laying-hen chamber. *Transactions of the ASABE* **2013**, *56*: 1493-1501.
- 541 39. Deleon-Rodriguez, N.; Lathem, T. L.; Rodriguez-R, L. M.; Barazesh, J. M.; Anderson, B.
- 542 E.; Beyersdorf, A. J.; Ziemba, L. D.; Bergin, M.; Nenes, A.; Konstantinidis, K. T.,
- Microbiome of the upper troposphere: species composition and prevalence, effects of tropical
- storms, and atmospheric implications. *Proceedings of the National Academy of Science* **2013**,
- **545** *110*: 2575-2580.
- 546 40. Yamaguchi, N.; Ichijo, T.; Baba, T.; Nasu, M., Long-range transportation of bacterial
- 547 cells by Asian dust. *Environmental Mutagen Research* **2014**, *36*: 145-151.
- 548 41. Tanaka, D.; Terada, Y.; Nakashima, T.; Sakatoku, A.; Nakamura, S., Seasonal variations
- in airborne bacterial community structures at a suburban site of central Japan over a 1-year
- time period using PCR-DGGE method. *Aerobiologia* **2015**, *31*: 143-157.
- 551 42. Gandolfi, I.; Bertolini, V.; Bestetti, G.; Ambrosini, R.; Innocente, E.; Rampazzo, G.;
- Papacchini, M.; Franzetti, A., Spatio-temporal variability of airborne bacterial communities
- and their correlation with particulate matter chemical composition across two urban areas.
- 554 Applied Microbiology and Biotechnology **2015**, 99: 4867-4877.
- 555 43. Gao, M.; Yan, X.; Qiu, T. L.; Han, M. L.; Wang, X. M., Variation of correlations
- between factors and culturable airborne bacteria and fungi. Atmospheric Environment 2016,
- **557** *128*: 10-19.
- 44. Gao, J. F.; Fan, X. Y.; Li, H. Y.; Pan, K. L., Airborne bacterial communities of PM_{2.5} in
- Beijing-Tianjin-Hebei Megalopolis, China as revealed by Illumina MiSeq sequencing: A case
- study. Aerosol and Air Quality Research 2017, 17: 788-798.
- 45. Wei, M.; Xu, C. H.; Chen, J. M.; Zhu, C.; Li, J. R.; Lv, G. L., Characteristics of bacterial
- 562 community in cloud water at Mt Tai: similarity and disparity under polluted and non-polluted
- cloud episodes. *Atmospheric Chemistry and Physics* **2017**, 17: 5253–5270.

- 46. Du, P. R.; Du, R.; Lu, Z. D.; Ren, W. S.; Fu, P. Q., Variation of bacterial and fungal
- community structures in PM_{2.5} collected during the 2014 APEC summit periods. Aerosol and
- 566 *Air Quality Research* **2018,** *18*: 444-455.
- 567 47. Jeon, E. M.; Kim, H. J.; Jung, K.; Kim, J. H.; Kim, M. Y.; Kim, Y. P.; Ka, J. O., Impact
- of Asian dust events on airborne bacterial community assessed by molecular analyses.
- 569 *Atmospheric Environment* **2011,** *45*: 4313-4321.
- 48. Maki, T.; Hara, K.; Kobayashi, F.; Kurosaki, Y.; Kakikawa, M.; Matsuki, A.; Chen, B.;
- 571 Shi, G. Y.; Hasegawa, H.; Iwasaka, Y., Vertical distribution of airborne bacterial communities
- in an Asian-dust downwind area, Noto Peninsula. Atmospheric Environment 2015, 119: 282-
- **573** 293.
- 574 49. Katra, I.; Arotsker, L.; Krasnov, H.; Zaritsky, A.; Kushmaro, A.; Ben-Dov, E., Richness
- and diversity in dust stormborne biomes at the southeast mediterranean. Scientific Reports
- **2014,** *4*: 5265.
- 577 50. An, S.; Couteau, C.; Luo, F.; Neveu, J.; DuBow, M. S., Bacterial diversity of surface sand
- samples from the Gobi and Taklamaken deserts. *Microbial Ecology* **2013**, *66*: 850-860.
- 579 51. Seifried, J. S.; Wichels, A.; Gerdts, G., Spatial distribution of marine airborne bacterial
- 580 communities. *MicrobiologyOpen* **2015**, *4*: 475-490.
- 52. Cuthbertson, L.; Amores-Arrocha, H.; Malard, L. A.; Els, N.; Sattler, B.; Pearce, D. A.,
- Characterisation of arctic bacterial communities in the air above Svalbard. *Biology* **2017**, *6*: 1-
- 583 22.
- 53. DeLeon-Rodriguez, N.; Lathem, T. L.; Rodriguez, R. L.; Barazesh, J. M.; Anderson, B.
- E.; Beyersdorf, A. J.; Ziemba, L. D.; Bergin, M.; Nenes, A.; Konstantinidis, K. T.,
- Microbiome of the upper troposphere: species composition and prevalence, effects of tropical
- storms, and atmospheric implications. Proceedings of the National Academy of Sciences of
- 588 the United States of America **2013**, 110: 2575-2580.
- 589 54. Despres, V. R.; Nowoisky, J. F.; Klose, M.; Conrad, R.; Andreae, M. O.; Poschl, U.,
- 590 Characterization of primary biogenic aerosol particles in urban, rural, and high-alpine air by
- 591 DNA sequence and restriction fragment analysis of ribosomal RNA genes. Biogeosciences
- **2007,** *4*: 1127-1141.
- 593 55. Raisi, L.; Aleksandropoulou, V.; Lazaridis, M.; Katsivela, E., Size distribution of viable,
- 594 cultivable, airborne microbes and their relationship to particulate matter concentrations and
- meteorological conditions in a Mediterranean site. *Aerobiologia* **2013**, 29: 233-248.
- 56. Bowers, R. M.; McLetchie, S.; Knight, R.; Fierer, N., Spatial variability in airborne
- 597 bacterial communities across land-use types and their relationship to the bacterial
- 598 communities of potential source environments. *The ISME Journal* **2011,** *5*: 601-612.

- 599 57. Bowers, R. M.; Lauber, C. L.; Wiedinmyer, C.; Hamady, M.; Hallar, A. G.; Fall, R.;
- Knight, R.; Fierer, N., Characterization of airborne microbial communities at a high-elevation
- 601 site and their potential to act as atmospheric ice nuclei. Appled and Environment
- 602 *Microbiology* **2009**, *75*: 5121-5130.
- 58. Fang, Z. G.; Ouyang, Z. Y.; Zheng, H.; Wang, X. K.; Hu, L. F., Culturable airborne
- bacteria in outdoor environments in Beijing, China. *Microbial Ecology* **2007**, *54*: 487-496.
- 605 59. Amato, P.; Parazols, M.; Sancelme, M.; Laj, P.; Mailhot, G.; Delort, A. M.,
- Microorganisms isolated from the water phase of tropospheric clouds at the Puy de Dôme:
- major groups and growth abilities at low temperatures. FEMS Microbiology Ecology 2010, 59:
- 608 242-254.
- 609 60. Peleg, A. Y.; Seifert, H.; Paterson, D. L., Acinetobacter baumannii: emergence of a
- successful pathogen. Clinical Microbiology Reviews **2008**, *21*: 538-582.
- 61. Xie, R.; Zhang, X. D.; Zhao, Q., Analysis of global prevalence of antibiotic resistance in
- 612 Acinetobacter baumannii infections disclosed a faster increase in OECD countries. Emerging
- 613 *Microbes & Infections* **2018**, 7: 31.
- 614 62. Zhang, C.; Ling, B., Treatment options for multidrug-resistant and extensively
- drugresistant Acinetobacter baumannii infections. Journal of Microbes & Infections 2018, 13:
- 616 90-96.
- 617 63. Hawkey, J.; Ascher, D. B.; Judd, L. M.; et al. Evolution of carbapenem resistance in
- Acinetobacter baumannii during a prolonged infection. *Microbial Genomics* **2018**, 4.
- 64. Yang, C. H.; Young, T.; Peng, M., Clinical spectrum of Pseudomonas putida infection.
- *Journal of the Formosan Medical Association* **1996**, 95: 754-761.
- 621 65. Carpenter, R. J.; Hartzell, J. D.; Forsberg, J. A., Pseudomonas putida war wound
- infection in a US Marine: A case report and review of the literature. Journal of Infection 2008,
- *56*: 234-240.
- 624 66. Valerio, F.; De, B. P.; Lonigro, S. L.; Visconti, A.; Lavermicocca, P., Use of
- 625 Lactobacillus plantarum fermentation products in bread-making to prevent *Bacillus subtilis*
- 626 ropy spoilage. *International Journal of Food Microbiology* **2008**, *122*: 328-332.
- 627 67. Apetroaie-Constantin, C.; Mikkola, R.; Andersson, M. A.; Teplova, V.; Suominen, I.;
- Johansson, T.; Salkinoja-Salonen, M., Bacillus subtilis and B. mojavensis strains connected to
- food poisoning produce the heat stable toxin amylosin. Journal of Applied Microbiology 2009
- 630 *106*:1976-1985.
- 631 68. Prescott, J. F., Rhodococcus equi: an animal and human pathogen. Clinical Microbiology
- 632 *Reviews* **1991**, *4*: 20-34.
- 633 69. Yamshchikov, A. V.; Schuetz, A.; Lyon, G. M., Rhodococcus equi infection. Lancet
- 634 Infectious Diseases 2010, 10: 350-359.

- 635 70. Laupland, K. B.; Pasquill, K.; Parfitt, E. C.; Dagasso, G., Steele, L., Streptococcus
- anginosus group bloodstream infections in the western interior of British Columbia, Canada.
- 637 *Infectious Diseases* **2017**, *50*: 423-428.
- 638 71. Murarka, S.; Pranav, F.; Dandavate, V., Pyogenic liver abscess secondary to disseminated
- 639 streptococcus anginosus from sigmoid diverticulitis. Journal of Global Infectious Diseases
- **2011**, *3*: 79-81.
- 72. Liu, H.; Zhang, X.; Zhang, H.; Yao, X.; Zhou, M.; Wang, J., Effect of air pollution on the
- 642 total bacteria and pathogenic bacteria in different sizes of particulate matter. Environmental
- 643 *Pollution* **2018,** 233: 483-493.
- 73. Xie, Z. S.; Li, Y. P.; Lu, R.; Li, W. X.; Fan, C. L.; Liu, P. X.; Wang, J. L.; Wang, W. K.,
- 645 Characteristics of total airborne microbes at various air quality levels. Journal of Aerosol
- 646 Science **2018**, 116: 57-65.
- 74. Innocente, E.; Squizzato, S.; Visin, F.; Facca, C.; Rampazzo, G.; Bertolini, V.; Gandolfi,
- 648 I.; Franzetti, A.; Ambrosini, R.; Bestetti, G., Influence of seasonality, air mass origin and
- particulate matter chemical composition on airborne bacterial community structure in the Po
- 650 Valley, Italy. *Science of the Total Environment* **2017**, *593-594*: 677-687.
- 651 75. Yan, Y.; Fu, P. Q.; Jing, B.; Peng, C.; Boreddy, S. K. R.; Yang, F.; Wei, L. F.; Sun, Y. L.;
- Wang, Z. F.; Ge, M. F., Hygroscopic behavior of water-soluble matter in marine aerosols over
- the East China Sea. *Science of the Total Environment* **2016,** *578*: 307-316.
- 654 76. Huebert, B. J.; Wang, M. X.; Lu, W. X., Atmospheric nitrate, sulfate, ammonium and
- calcium concentrations in China. Tellus Series B-chemical and Physical Meteorology 1988,
- **656** *40*: 260-269.
- 77. Louie, P. K. K.; Watson, J. G.; Chow, J. C.; Chen, A.; Sin, D. W. M.; Lau, A. K. H.,
- 658 Seasonal characteristics and regional transport of PM_{2.5} in Hong Kong. Atmospheric
- 659 Environment 2005, 39: 1695-1710.
- 78. Han, T. T.; Liu, X. G.; Zhang, Y. H.; Gu, J. W.; Tian, H. Z.; Zeng, L. M.; Chang, S. Y.;
- 661 Cheng, Y. F.; Lu, K. D.; Hu, M., Chemical characteristics of PM₁₀ during the summer in the
- mega-city Guangzhou, China. Atmospheric Research 2014, 137: 25-34.
- 663 79. Hu, M.; He, L. Y.; Zhang, Y. H.; Wang, M.; Yong, P. K.; Moon, K. C., Seasonal
- variation of ionic species in fine particles at Qingdao, China. Atmospheric Environment 2002,
- 665 *36*: 5853-5859.
- 80. Kim, B. M.; Teffera, S.; Zeldin, M. D., Characterization of PM_{2.5} and PM₁₀ in the South
- 667 Coast Air basin of Southern California: Part 1-Spatial variations. *Journal of the Air and Waste*
- 668 *Management Association* **2000**, *50*: 2034-2044.

- 81. Kim, H. S.; Huh, J. B.; Hopke, P. K.; Holsen, T. M.; Yi, S. M., Characteristics of the
- 670 major chemical constituents of PM_{2.5} and smog events in Seoul, Korea in 2003 and 2004.
- 671 *Atmospheric Environment* **2007**, *41*: 6762-6770.
- 82. Castañer, R.; Nicolás, J. F.; Crespo, J.; Yubero, E.; Galindo, N.; Caballero, S.; Pastor, C.,
- 673 Influence of air mass origins on optical properties and PM concentrations measured at a high
- 674 mountain station located in the southwestern Mediterranean. *Atmospheric Research* **2017**, *197*:
- 675 244-254.
- 676 83. Masiol, M.; Squizzato, S.; Ceccato, D.; Rampazzo, G.; Pavoni, B., A chemometric
- approach to determine local and regional sources of PM 10 and its geochemical composition
- in a coastal area. *Atmospheric Environment* **2012,** *54*: 127-133.
- 84. Moreda-Piñeiro, J.; Turnes-Carou, I.; Alonso-Rodríguez, E.; Moscoso-Pérez, C.; Blanco-
- Heras, G.; López-Mahía, P.; Muniategui-Lorenzo, S.; Prada-Rodríguez, D., The influence of
- oceanic air masses on concentration of major ions and trace metals in PM_{2.5} fraction at a
- coastal European suburban site. Water Air and Soil Pollution 2015, 226: 1-15.
- 85. Squizzato, S.; Masiol, M., Application of meteorology-based methods to determine local
- and and and action action and action action action action.
- 685 *Atmospheric Environment* **2015**, *119*: 69-81.
- 686 86. Kakikawa, M.; Kobayashi, F.; Maki, T.; Yamada, M.; Higashi, T.; Chen, B.; Shi, G. Y.;
- Hong, C. S.; Tobo, Y.; Iwasaka, Y., Dustborne microorganisms in the atmosphere over an
- Asian dust source region, Dunhuang. Air Quality, Atmosphere and Health 2008, 1: 195-202.

Table 1 Bacterial abundance in PM with different size fraction.

Type	Site	Abundance Range	Average	Method	Reference
		Cells m ⁻³	Cells m ⁻³		
TSP	High-altitude air, Caribbean Sea	$3.0 \times 10^3 \sim 9.0 \times 10^4$	$\frac{\text{Cells m}^{-3}}{5.1 \times 10^3}$	q <i>PCR</i>	39
TSP	High-altitude air, Japan	$1.0 \times 10^3 \sim 2 \times 10^5$		q <i>PCR</i>	40
TSP	Toyama, Japan	$1.1 \times 10^3 \sim 1.3 \times 10^5$	1.8×10^{4}	q <i>PCR</i>	41
TSP	Seoul, Korea	$5.2 \times 10 \sim 4.3 \times 10^3$		q <i>PCR</i>	29
PM_{10}	Milan and Venice, Italy	$2.4 \times 10^2 \sim 4.9 \times 10^{5*}$	$3.0 \times 10^{4*}$	q <i>PCR</i>	42
$PM_{2.5}$	BTH, China	$8.0 \times 10 \sim 5.8 \times 10^{3**}$	$1.1 \times 10^{3**}$	Culture method	43
$PM_{2.5}$	BTH, China	$4.8 \times 10^4 \sim 2.6 \times 10^5$	1.2×10^{5}	q <i>PCR</i>	44
PM_1	Ji'nan, China	$1.8 \times 10^4 \sim 2.6 \times 10^4$	2.2×10^4	q <i>PCR</i>	27
CW***	Mt.Tai, China	$5.8 \times 10^4 \sim 1.6 \times 10^5$	1.2×10^{5}	q <i>PCR</i>	45
$PM_{2.5}$	Mt.Tai, China	$4.0 \times 10 \sim 1.728 \times 10^{3}$	4.9×10^{2}	q <i>PCR</i>	This study

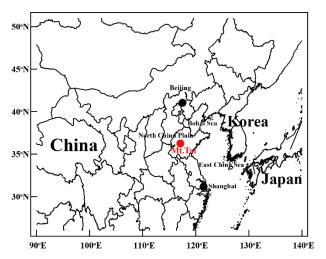
690 * Copies m⁻³; *** CFU m⁻³; **** Cloud Water

Table 2 Statistical summary of alpha diversity estimators from different sample type and study environments.

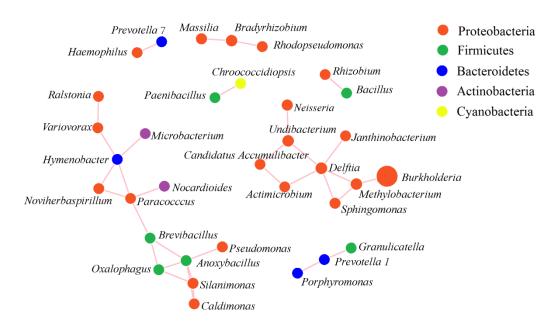
Environment	Sample Type	OTUs	Chao1	Shannon	Method	Reference
Urban/Terrestrial	PM _{2.5}	2457	4915	5.89	Miseq Sequencing	46
Urban/Terrestrial	$PM_{2.5}$	552	571	3.80	Miseq Sequencing	44
Urban/Terrestrial	$PM_{0.56-1}$	222	272	2.44	Miseq Sequencing	27
Dust/Terrestrial	$PM_{2.5}$	62	200	3.51	Clone Library	47
Dust/Terrestrial	TSP		270		Pyrosequencing	48
Dust/Terrestrial	Dust	869	1485		Pyrosequencing	49
Dust/Terrestrial	PM_{10}	733	1265		Pyrosequencing	18
Sand/Terrestrial	Sand	1445	1858	5.99	Pyrosequencing	50
Marine/Coastal	TSP	753			Pyrosequencing	51
Marine/Coastal	TSP	5015		7.66~9.28	Miseq Sequencing	52
High altitude	TSP	410			Pyrosequencing	22
High altitude	TSP	314	241		Miseq Sequencing g	53
High altitude	CW*	664	1302	3.80	Miseq Sequencing	24
High altitude	PM _{2.5}	580	1282	3.01		•
Cluster I	$PM_{2.5}$	491	1151	2.67		
Cluster II	$PM_{2.5}$	567	1322	2.93	Miseq Sequencing	This Study
Cluster III	$PM_{2.5}$	677	1397	3.09		
Cluster IV	PM _{2.5}	625	1234	3.63		

694 * Cloud Water

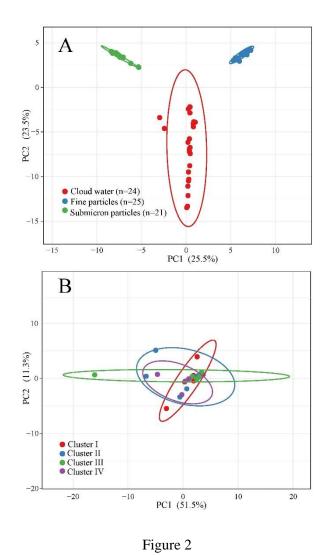
695 **List of Figures** 696 Figure 1 The geographic situation of Mt. Tai in the North China Plain. 697 Figure 2 The co-occurrence network of top 100 OTUs of PM_{2.5} at Mt. Tai, China. The bigger 698 the circle, the greater the number of OTUs. 699 Figure 3 The principal component analysis showed the overall distribution pattern of A 700 bacterial communities in different air samples and B profiles among four air mass cluster. 701 Figure 4 Cluster analysis of air-mass trajectories and corresponding chemical composition 702 and bacterial abundance in PM_{2.5} 703 Figure 5 Bacterial community at phyla, class, order levels under the influence of four air 704 mass trajectories. 705 Figur 6 Variance analysis of genera species based on the Kruskal–Wallis rank sum test (A) 706 and the Spearman's rank correlations with environmental factors (B; *p < 0.05; * * p < 0.01).



708 Figure 1



710 Figure 1



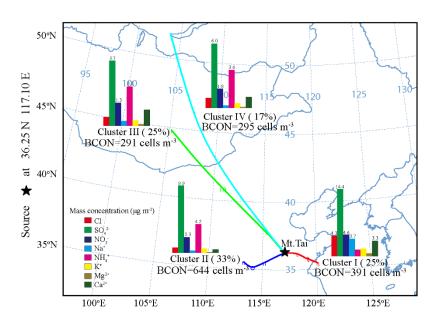


Figure 3

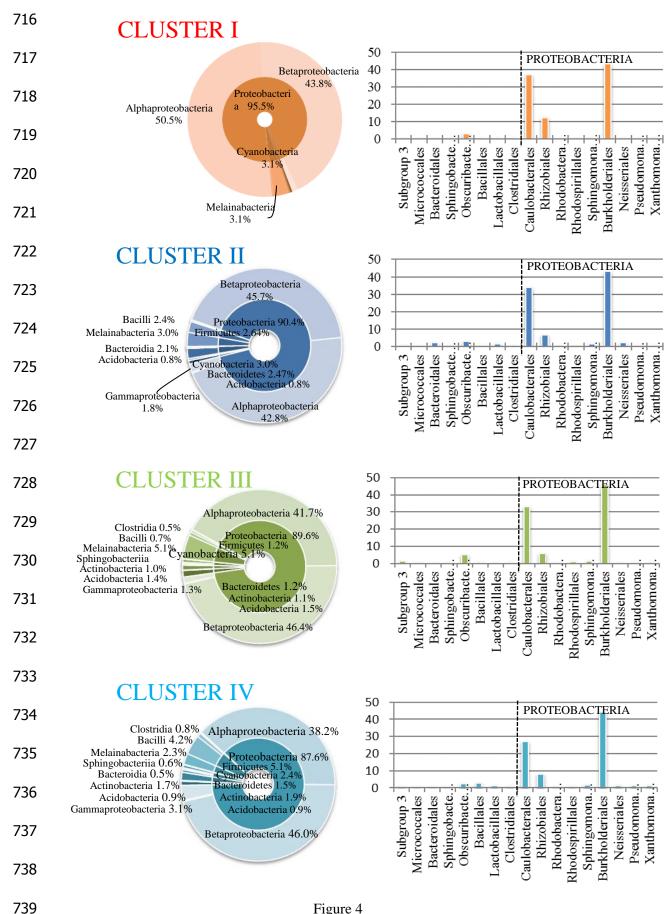
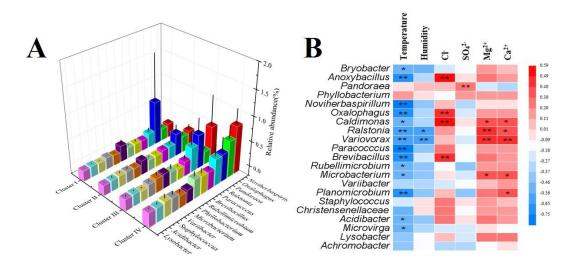


Figure 4



Figur 5