

One for each ocean: revision of the Bursa granularis (Röding, 1798) species complex (Gastropoda: Tonnoidea: Bursidae)

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1	One for each ocean: revision of the Bursa granularis (Röding, 1798) species complex
2	(Gastropoda: Tonnoidea: Bursidae)
3	
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13	
14	Running title: BURSA GRANULARIS COMPLEX
15	
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17	ABSTRACT
18	Bursa granularis (Röding, 1798) is a tonnoidean gastropod viewed as broadly
19	distributed throughout the Indo-Pacific and tropical western Atlantic. Because of its variable
20	shell it received no less than thirteen names, now all synonymized under the name B.
21	granularis. We sequenced a fragment of the cox1 gene for 82 specimens covering a large part
22	of its distribution and most type localities. Two delimitation methods were applied, one based
23	on genetic distance (ABGD) and one based on phylogenetic trees (GMYC). All analyses
24	suggest that specimens identified as B. granularis comprise four distinct species: one limited
25	to the tropical western Atlantic; another to southwestern Western Australia; and two in the
26	Indo-Pacific, from the Red Sea to the open Pacific, partly sympatric - but not syntopic - in
27	Japan, the Philippines, Vanuatu and New Caledonia. Based on shells characters, we applied
28	available names to each of the four species, respectively B. cubaniana (d'Orbigny, 1841), B.
29	elisabettae Nappo, Pellegrini & Bonomolo, 2014, B. granularis s.s., and B. affinis Broderip,
30	1833, and we provide new standardized conchological descriptions for all of them. Our results
31	demonstrate that a long planktotrophic larval stage, common among Tonnoidea, does not
32	necessarily ensure a circum-tropical species distribution.
33	
34	INTRODUCTION

Tonnoideans are a rather small superfamily of Caenogastropoda (~357 species, WoRMS 35 2017) related to the neogastropods, either as its sister group (Zou *et al.* 2011) or belonging in 36 Neogastropoda (Colgan et al., 2007; Cunha et al., 2009; Hayashi, 2005; Williams et al., 37 2014, Osca *et al.*, 2015). Notwithstanding the non-planktotrophic development of a few 38 Australian species, one of the characteristic features of tonnoidean gastropods is their long to 39 extremely long planktonic larval stages, named teleplanic larvae [based on the Greek *tele*, 40 distant, and *planos*, wandering; Scheltema (1971)]. The record has been observed for a larva 41 42 of Fusitriton oregonensis Redfield, 1846 (Ranellidae), which lived in an aquarium for 4.5 years without metamorphosing (Strathmann & Strathmann, 2007). An indirect estimation of 43 the duration of larval life was also proposed by Scheltema (1972) who, based on the extent of 44 the distribution area and the speed of ocean currents, inferred the time it would take for the 45 larvae to cross ocean basins — generally several months. Conversely, and in quite circular 46 47 reasoning, the duration of this larval time led to the hypothesis that some species may have trans-oceanic dispersal capabilities (e.g., Scheltema, 1966, 1968, 1971, 1972, 1986a, 1986b, 48 49 1988; Laursen, 1981; Pechenik et al., 1984), resulting in a cosmopolitan distribution. Scheltema (1971, table 1) also kept larvae alive in an aquarium that had been collected in the 50 plankton, and found that some taxa lived as planktonic veliger larvae for several months [e.g., 51 Monoplex nicobaricus (Röding, 1798) lived for 390 days in captivity]. 52

As in most other marine gastropods, tonnoidean species were described based on 53 features of the teleoconch, using a limited number of specimens and characters. The available 54 material of many species of Bursidae described before 1960 rarely exceeded three specimens. 55 Because of this limited evaluation of the intraspecific variability, numerous new species were 56 described for every newly recognized morphological form. When additional material became 57 available, malacologists realized that they may have greatly underestimated the intraspecific 58 59 shell variability, since supposedly geographically restricted species were actually morphologically highly similar to other nominal species from other localities, moreover 60 sharing identical protoconchs. This was the first step towards an important synonymization 61 62 trend in tonnoidean systematics. Also, taking into account the expected great dispersal abilities, modern authors (e.g., Beu, 1998, Nappo et al., 2014) followed the path led by 63 Scheltema to the point where some very well-defined morphs were ranked as no more than 64 subspecies. They thus recognized a smaller number of species, but with trans-oceanic 65 66 distributions.

Among the Tonnoidea, the family Bursidae includes 54 Recent species (WoRMS
2017), among which several are potentially species complexes, i.e., species for which

alternative hypotheses of delimitation have been proposed in the literature. In particular, the 69 Bursa granularis complex, already identified as such by Castelin et al. (2012), is typically 70 recognized as a single species by modern authors (Beu, 1998, 2005, 2010), although 71 sometimes with subspecies (Nappo et al., 2014). According to both the literature and the 72 GBIF database (2016) (Fig. 1), B. granularis has a subtropical and tropical distribution 73 throughout the Indo-West Pacific, the eastern Pacific, and the tropical western Atlantic. [In 74 the eastern Atlantic, it has only been recorded from the Cape Verde Islands (Garcia-Talavera, 75 1983, cited by Beu 1998), but this record is unconfirmed by Rolán (2005).] This distribution 76 77 would make it one of the most cosmopolitan species among the Tonnoidea. However, this apparently cosmopolitan species has received no fewer than ten names (Beu, 1998). WoRMS 78 79 (2017) lists 13 synonyms of Tritonium granulare Roding, 1798: Tritonium jabick Roding, 1798, Biplex rubicola Perry, 1811, Ranella granifera Lamarck, 1816, Ranella affinis 80 81 Broderip, 1833, Ranella cubaniana d'Orbigny, 1841, Ranella livida Reeve, 1844, Bursa 82 cumingiana Dunker, 1862, Bursa alfredensis Turton, 1932, Bursa kowiensis Turton, 1932, B. 83 cubaniana intermedia Nowell-Usticke, 1959, Bursa corrugata lineata Nowell-Usticke, 1959 and Bursa granularis elisabettae Nappo, Pellegrini & Bonomolo, 2014. The numerous 84 alternative species hypotheses led to various usages of these names in the literature; whereas 85 19th century authors (e.g. Reeve, 1844b) recognized up to four different species, the last 86 revision proposed to group them all under the name Bursa granularis (Beu, 1998). Two of 87 these, however — B. granularis cubaniana (d'Orbigny, 1841) and Bursa granularis 88 elisabettae Nappo, Pellegrini & Bonomolo, 2014 — are accepted by Nappo et al. (2014) at 89 the rank of subspecies besides the nominate subspecies Bursa granularis granularis (Röding, 90 1798). 91

92 Using several molecular markers, Castelin et al. (2012) identified two morphologically 93 distinct clades within the Bursa granularis complex. Interestingly, these two clades were found in sympatry in Vanuatu, at similar depths, and the authors suggested that additional 94 sampling would be needed to clarify species delimitations within the group. We increased the 95 96 size of the dataset, including both additional species from the same geographical region sampled by Castelin et al. (2012) and samples from other localities, with a particular focus on 97 98 type localities, covering a large part of the global distribution of the Bursa granularis 99 complex. Our goal is to clarify the species delimitation within the *Bursa granularis* complex 100 and to test the hypothesis that this species has a world-wide geographical distribution. The specimens were first separated tentatively into morphospecies based on shell characters. 101 102 In a second step, all the specimens were sequenced (cox1 gene) to test whether the recognized

- 103 morphogroups correspond to distinct molecular clusters. Finally, we attributed the different
- 104 genetic and morphological groups identified within the complex to names available in the
- 105 literature and discuss the implication in term of their geographical distribution. .
- 106

107 MATERIAL AND METHODS

108 Sampling

109 The material for this study was collected from various localities during a series of shallow-

- 110 water and deep-sea expeditions to Saudi Arabia (UF 2013), Viet Nam (NT 2014), Vanuatu
- 111 (UF 2005, Santo Marine biodiversity survey 2006), Mozambique (MAINBAZA 2009,
- 112 INHACA 2011), Madagascar (UF 2008), the Philippines (PANGLAO 2004, UF 2015),
- 113 Mariana Island (UF 2008), Micronesia (UF 2008), Okinawa Japan (UF 2010), Guam (UF
- 114 2010), Marquesas Islands (Pakaihi I Te Moana 2012), Papua New Guinea (PAPUA NIUGINI
- 115 2012), Marshall Islands (UF 2008), New Caledonia (TERRASSES 2008, UF 2013), Taiwan
- 116 (UF 2005), Florida, USA (UF 2010), Guadeloupe (KARUBENTHOS 2012), and Western
- 117 Australia (UF 2009, WESTERN AUSTRALIA 2011). MNHN specimens collected before
- 118 2012 were anaesthetized with an isotonic solution of MgCl₂ and fixed in 96% ethanol.
- 119 Specimens collected after 2012 were processed with a microwave oven (Galindo *et al.* 2014);
- the living molluscs in small volumes of sea water were exposed to microwaves for ca. 30
- seconds. Bodies were immediately removed from shells and dropped into 96% ethanol. UF
- specimens were directly put alive into 75 or 95% ethanol. The analysed material includes 82
- specimens, 42 registered in the MNHN and 40 in the UF collections (Supplementary Material
- 124 Table 1). The specimens and the corresponding sequences are also registered in the Barcode
- 125 of Life Datasystem (project BURSA) and GenBank. Maps with the sampling locality of all
- the specimens were generated using Qgis 2.16.3 (Fig. 1).
- 127

128 DNA sequencing

- 129 DNA was extracted using the Epmotion 5075 robot (Eppendorf), following the
- 130 manufacturers' recommendations. A fragment of the cytochrome oxidase subunit I (cox1)
- 131 gene was amplified using the universal primers LCO1490/HCO2198 (Folmer *et al.*, 1994).
- 132 PCR reactions were performed in volumes of 20 μ l, containing 3 ng DNA, 1X reaction buffer,
- 133 2.5 mM MgCl2, 0.26 mM dNTP, 0.3 mM of each primer, 5% DMSO and 1.5 units of
- 134 Qbiogene Q-Bio Taq. The amplification consisted of an initial denaturation step at 94°C for 5
- min, followed by 40 cycles of denaturation at 94°C for 30 s, annealing at 47°C for 30 s,

- followed by extension at 72° C for 1 min. The final extension was at 72° C for 5 min. PCR
- 137 products were purified and sequenced (both strands) by the Eurofins sequencing facility.
- 138

139 Species delimitation

All specimens were first separated into morphogroups, using characters of the teleoconch and 140 in particular the outer lip. In a second step, the cox1 sequences were analysed. Alignment was 141 done by eye. Pairwise genetic distances were calculated using MEGA v. 6 (Tamura et al., 142 143 2013). Phylogenetic trees were reconstructed using maximum likelihood (ML) and Bayesian 144 inference approaches (BI), with MEGA v. 7 (with Bootstraps 100 replications and default parameters for tree inference options) for ML analysis, and with MrBayes (Huelsenbeck, 145 Ronquist & Hall, 2001) (six Markov chains and 10,000,000 generations, five chains, three 146 swaps at each generation, a sampling frequency of one tree each 1,000 generations, chain 147 148 temperature set at 0.02) and BEAST v1.8.3 (Drummond et al., 2012) (with uncorrelated relaxed clock, and 10,000,000 generations) for the BI analysis. In all analyses the three codon 149 150 positions of the cox1 gene were treated as independent partitions and the substitution model was set to GTR+G and GTR+I+G for the ML and BI analyses, respectively. Convergence for 151 152 both BI analysis was evaluated using Tracer v. 1.4.1 (Rambaut et al., 2014) to check that all effective sample size values exceeded 200. Consensus trees were calculated after omitting the 153 first 25% trees as burn-in. Nodal support was assessed as posterior probability (PP) in the BI 154 analyses and as bootstraps (BS) for the ML analysis. Closely related bursid species were used 155 as outgroups: Bursa latitudo Garrard, 1961, Bursina ignobilis (Beu, 1987) and Tutufa bufo 156 (Röding, 1798), following Castelin et al. (2012). 157

Species delimitations were performed using the online versions of the Automatic 158 Barcode Gap Discovery (ABGD; Puillandre *et al.*, 2012a) using a simple distance (p-distance) 159 160 model and default parameters. ABGD automatically detects the barcode gap in the pairwise distribution of genetic distances between low and high genetic distances, hypothesized to 161 correspond to intra- and inter-specific genetic distances, respectively. The General Mixed 162 163 Yule Coalescent (GMYC) method (Pons et al., 2006; Monaghan et al., 2009) was also used, with default parameters (both single and multiple methods). GMYC defines the transition 164 165 (unique in the single version, but variable between lineages in the multiple version) between 166 speciation and coalescent events in the ultrametric tree obtained with BEAST, inferred from 167 the rate of splits in the tree.

168

169 *Shell description*

Once the species were delimitated based on molecular data, shells were described using the 170 methods developed by Merle (2001, 2005) for primary homologies definition. Primary cords 171 are spiral ornamentations present from the first teleoconch whorl. Secondary cords (S) are 172 spiral ornamentations appearing later during shell ontogeny. The shoulder cord is designated 173 as P1. Cords of the convex part of the whorl are added from adapical to abapical (P2 to P6, 174 Fig. 2A). The primary cord of the infrasutural ramp is designated IP and the secondary cord 175 IS. Secondary cords on the convex part of the whorl are designated S1to S6. Cords of the Fig. 2 177 siphonal canal were not considered. Denticles inside the outer lip are named D1 to D6, starting under the shoulder cord and added from adapical to abapical; denticles above the 178 shoulder are designed as ID1 and ID2 added from abapical to adapical (ID2 is not shown in 179 Fig. 2). Denticles are bifid on most specimens but can merge or further bifurcate. In order to 180 describe the outer lip we introduce here the outer lip denticles formula (OLDF). It is a four-181 182 digit sequence representing the number of visible denticles (D) between a notch and the

preponderant primary cord (P1,P3 and P5) or between two preponderant primary cords ,
starting from the anal notch towards the siphonal one. For example, in the specimen of *B*. *granularis* figured in Fig. 2, the OLDF should be read as AN,2, P1, 4, P3, 4, P5, 4, SC or

- 186 2,4,4,4 for short.
- 187 188

189 ABBREVIATIONS

190 Depositaries

- 191 MAC. ML Museo di Storia Naturale Aquilegia, Cagliari
- 192 MHNG Muséum d'Histoire naturelle, Geneva, Switzerland
- 193 MNHN Muséum National d'Histoire Naturelle, Paris, France
- 194 NHMUK Natural History Museum, London, United Kingdom
- 195 RASM Russian Academy of Science, Moscow
- 196 UF Florida Natural History Museum, University of Florida, Gainesville, United States
- 197
- 198 Shell characters
- AN anal notch
- 200 SC siphonal canal
- 201 H height
- 202 P Primary cord
- 203 IP: infrasutural primary cord

204	P1 Shoulder cord
205	P2-6 primary cords of the convex part of the whorl
206	S secondary cord
207	IS infrasutural secondary cord
208	S1-6 secondary cords of the convex part of the whorl
209	D denticle of the outer lip
210	ID1-2 denticle of the infrasutural ramp
211	D1-6 denticles of the convex part of the whorl
212	OLDF outer lip denticles formula
213	
214	
215	
216	
217	RESULTS
218	Among the 82 specimens, excluding outgroups, three morphogroups were initially
219	recognized. The ABGD (both initial and recursive partitions) and GMYC (single) methods
220	consistently found 4 clusters. The only exception is the multiple method of GMYC: five

clusters are proposed, one specimen (IM-2013-19496) from the Caribbean cluster (see below) 221 being considered different. Given that this specimen is very close in molecular sequences to 222 other Caribbean specimens, we conclude this to be an artefact of the multiple method of 223 GMYC, which is known to oversplit (Fujisawa and Barraclough, 2013; Kekkonen and Hebert, 224 2014). Among the four clusters recognized with the cox1 gene, two correspond to two 225 morphogroups. One morphogroup is thus divided into two genetic clusters, corresponding to 226 227 the two clades revealed by Castelin et al. (2012). A posteriori re-examination of the shells revealed stable teleoconch characters distinguishing the two genetic clusters. The 228 morphological characters used to recognize first the three morphogroups, and, after the 229 molecular analyses, the final four morphogroups, are described in the SYSTEMATICS 230 231 section. The four genetic clusters also correspond to highly supported clades in both BI and ML analyses (PP > 0.99 and BS > 80) (Fig. 3). Pairwise genetic distances between clades 232 233 were never less than 6.4%, while within cluster genetic distances never exceeded 1.8%, even 234 between geographically distant localities such as Saudi Arabia and Western Australia. 235 Remarkably, all clades are structured geographically: Clade 1 ranges from Mozambique to the Red Sea, Vietnam, Japan, the Philippines, Western Australia, Vanuatu and New Caledonia, 236 237 suggesting an occurrence throughout the whole Indian Ocean and western Pacific; Clade 2 is

Fig. 3	found in sympatry with Clade 1 in Okinawa, the Philippines, Vanuatu and New Caledonia,
	but is also present in Taiwan and Papua New Guinea and extends further east to Guam and
240	French Polynesia; although Clades 1 and 2 can be found in sympatry they never occur in
241	syntopy. Clade 3 is circumscribed to the Caribbean; and Clade 4 is limited to southernmost
242	Western Australia.
243	Based on the original descriptions and type localities, we have attributed the name
244	Bursa granularis to Clade 1; Bursa affinis to Clade 2; Bursa cubaniana to Clade 3; and Bursa
245	elisabettae to Clade 4.
246	
247	SYSTEMATIC DESCRIPTIONS
248	
249	Superfamily TONNOIDEA Suter, 1913 (1825)
250	Family BURSIDAE Thiele, 1925
251	Bursa Röding, 1798
252	Type species: Bursa monitata Roding, 1798, by subsequent designation (Jousseaume, 1881:
253	174) (junior synonym of Murex bufonius Gmelin, 1791, by First Reviser's action of
254	Winckworth, 1945: 137).
255	
256	Bursa granularis (Röding, 1798)
Fig. 4 7	(Fig. 4 A-D, J)
258	Tritonium granulare Roding, 1798: 127. (Red Sea, in accordance with ICZN Art. 76.3;
259	original types lost. Neotype MHNG 1098/85/1, designated by Beu, 1998: 150).
260	Bursa granularis — H. & A. Adams, 1852: 106; Barnard, 1963: 17; Hinton, 1972: 12, pl. 6,
261	fig. 22; Hinton, 1978: 32, fig. 8; Kilburn & Rippey, 1982: 73, pl. 16, fig. 14, Drivas & Jay,
262	1988: 62, pl. 16, fig. 4; Wilson, 1993: 226, pl. 43, figs 11a-b, 12; Kubo in Kubo & Kurozumi,
263	1995: 74, 78, fig. 7; Beu, 1998: 150, figs 48a-e, 58d; Beu, 1999: 44, fig. 85; Beu, 2005: 19,
264	figs 27-28; Lee & Chao, 2003: 40, pl. 4, fig. 93; Castelin et al., 2012: 4843, fig. 4; Dolorosa et
265	<i>al.</i> , 2013: 8, fig. 3D.
266	Colubrellina granularis — Habe, 1961: 47, pl. 24, fig. 5; Okutani, 1986: 116-117, top left fig.
267	Colubrellina (Dulcerana) granularis — Habe, 1964: 76, pl. 24, fig. 5; Wilson & Gillett,
268	1971: 80, pl. 54, figs 7-7b.
269	Bursa (Colubrellina) granularis granularis. — Beu, 1985: 64; Cossignani, 1994: 75-77;
270	Nappo et al., 2014: pl. 3, figs 1-3, pl. 4, figs 1-4, pl. 5, fig. 1, pl. 7, fig. 1.
271	Bursa (Bufonariella) granularis — Bosch et al., 1995: 102, fig. 737.

- 272 Bursa (Colubrellina) granularis Okutani, 2000: 269, pl. 133, fig. 1; Zhang & Ma, 2004:
- 273 182, text-figs 113a-c, pl. 5, figs 1-3.
- 274 Tritonium jabick Röding, 1798: 127 (Red Sea, in accordance with ICZN Art. 76.3; original
- types lost. Neotype MHNG 1098/85/1, designated by Beu, 1998: 150).
- 276 *Dulcerana jabick* Iredale, 1931: 213.
- 277 Colubrellina jabick Oyama & Takemura, 1960: Colubrellina pl., figs 3-4.
- 278 Biplex rubicola Perry, 1811: pl. 5, fig. 4 (Red Sea, in accordance with ICZN Art. 76.3;
- original types lost. Neotype MHNG 1098/85/1, designated by Beu, 1998: 150).
- 280 Ranella granifera Lamarck, 1816: pl. 414, fig. 4, "Liste des objets": 4; 1822: 153 (Red Sea,
- designated by Beu 1998:150; Lectotype MHNG 1098/85/1, designated by Beu, 1998:150);
- 282 Kiener, 1841: 16, pl. 11, fig. 1; Deshayes, 1843: 548; Reeve, 1844a: pl. 6, fig. 30; Küster,
- 283 1871 [in Küster & Köbelt, 1870-1878]: 143, pl. 39, fig.1.
- 284 Bursa cumingiana Dunker, 1862: 238; New Caledonia; Lectotype NHMUK 1968530
- 285 designated by Beu 1998:150 Dunker, 1863: 59, pl. 19, figs 7-8.
- 286 Ranella (Lampas) granifera Tryon, 1880: 41, pl. 22, figs 35-40.
- *Ranella semigranosa* Reeve, 1844b: pl. 6, fig. 25; Krauss, 1848: 113 (not of Lamarck, 1822).
- 289
- 290 *Material examined*: See Supplementary material table 1.
- 291 Diagnosis: Shell biconic, varices strictly aligned, numerous nodules on all primary, secondary
- and tertiary cords, OLDF: 2.4.4.4, or 2.4.4.3 in large specimens
- 293 Description: Protoconch of 3.75 whorls (following the counting method of Bouchet &
- Kantor, 2004), rather tall with weakly impressed suture, with finely reticulate protoconch I of
- 295 0.5 whorls, reticulate sculpture on first whorl of protoconch II, fading out before end of last
- 296 protoconch whorl. Teleoconch: tall and relatively narrow with 4-5 whorls (up to 6 in very
- large specimens) Height (H) around 40mm, up to 60mm. Spire angle 50°. proportion of
- aperture 53. % of the shell width. Proportion of the last whorl 54% of the height. Varices
- strictly aligned for most of spire height, becoming slightly displaced only on last whorl or two
- 300 of large specimens. Abapertural face of each varix slightly excavated, buttressed by spiral
- 301 cords. Spiral ornamentation of 7 primary cords, one on infrasutural ramp (IP) and 6 on convex
- part of whorl (P1-6). After whorl 2.5 (varix 5) IP becoming very reduced, noticeable only
- 303 where it intersects varices. Secondary cords in interspaces between primary cords. Primary
- 304 cord above IP remaining reduced throughout shell ontogeny whereas others growing in
- 305 similar fashion to primary cords. On last whorl, spiral ornamentation composed of 9 nodules

per primary cord between two varices (and secondary cords of infrasutural ramp) and double
this number for secondary ones. Apertural lip well flared, bearing numerous prominent,
narrow, transverse denticles. Outer lip bearing numerous denticles; ID1 bifid, ID2 absent, D14 bifid, D5 bifid but merged in large specimens, D6 bifid (Fig. 5).

309 310

306

Fig.5

Fig. 6

7

308

Distribution and habitat: Bursa granularis has a tropical distribution throughout the IndoWest Pacific province (Fig.1), from the Mozambique Channel throughout East Africa and the
Red Sea to the northern Indian Ocean, in the western Pacific from Kii Peninsula and
Yamaguchi Prefecture, Honshu, Japan (Beu, 1999), south to Rottnest Island in Western
Australia (Wilson, 1993). Its eastward distribution in the Pacific extends from Japan to
Australia through the Philippines, Vanuatu and New Caledonia. Bursa granularis is
commonly found in very shallow water but some specimens were found alive around 100m
deep.

Remarks: The nomenclatural history of *Bursa granularis* is a particularly complex one. The

319 320

types of Tritonium granulare, T. jabick and Biplex rubicola are lost. Beu (1998) designated 321 322 the lectotype of *Ranella granifera* as neotype of the three other names. As a consequence these formerly subjective synonyms became objective synonyms and following ICZN Art. 323 76.3 they also all share the same type locality, the Red Sea. There is only one species 324 belonging to the complex in this locality, to which the name B. granularis can thus 325 unambiguously be attributed. Furthermore the type specimen of *Ranella granifera* displays 326 characters congruent with the description above (Fig. 4,A). 327 Bursa cumingiana (type locality New Caledonia) is another available name in the B. 328 granularis complex. There are two (and only two) Molecular Operational Taxonomic Unit 329

330 (MOTU) living sympatrically in this region and the lectotype of *Ranella cumingiana*

331 (NHMUK 1968530) is a well-preserved adult specimen, which distinctively displays four

denticles between P5 and the anterior canal, identifying it as *Bursa granularis*. The color

pattern of the teleoconch of *B. granularis* is highly variable, from off-white, pale brown or

pale orange to deep reddish brown. Some specimens bear the same striped color pattern as *B*.

cubaniana. A few young specimens display the angular profile of *B. affinis*, especially in the

336 Mozambique Channel, but it never occurs in fully grown specimens.

- 337
- 338

339 *Bursa affinis* (Broderip, 1833)

340 (Fig. 4 E-I,K)

- 342 *Ranella affinis* Broderip, 1833: 179 ("Annaa," Tuamotu Islands, Lectotype NHMUK
- 343 1950.11.28.4, designated by Beu, 1998:150); G. B. Sowerby II, 1835: pl. 89, fig. 12; Reeve,
- 1844b: pl. 4, fig. 19; Küster, 1871 [in Küster & Köbelt, 1870-1878]: 142, pl. 38a, fig. 5.
- 345 Bursa granularis affinis Hertlein & Allison, 1960: 15.
- 346 Ranella livida Reeve, 1844a: pl. 6, fig. 28; ("Annaa," Tuamotu Islands, Lectotype NHMUK
- 347 1967657_1, designated by Beu, 1998:150); 1844b: 138; Krauss, 1848: 113.
- 348 *Dulcerana granularis* Iredale, 1931: 213; Rippingale & McMichael, 1961: 69, pl. 7, fig. 19
 349 (not of Röding, 1798).
- Bursa granularis —Hertlein & Allison, 1960: 15; Salvat & Rives, 1975: 307, fig. 179; Kay,
- 3511979: 227, fig. 80A; Severns 2011: 150, pl. 59, fig. 6; Beu *et al.*, 2012: 67, fig. E2 (not of
- 352 Röding, 1798).
- Bursa (Colubrellina) granularis Kaiser, 2007: 39, pl. 26, figs 7a-b (not of Röding, 1798).
 354
- 355 *Material examined:* See Supplementary material table 1.
- 356 *Diagnosis:* Shell biconic, varices strictly aligned, numerous nodules on all primary, secondary
- and tertiary cords, shoulder angular prominent, OLDF: 2,3,4,3 or 3,4,4,3 in particularly large
 specimens.
- 359 *Description:* Protoconch as in *B. granularis.* Teleoconch tall and relatively narrow with 4–5
- whorls (up to 6 on large specimens). H 40 to 50 mm, up to 80 mm. Spire angle 55°.
- Proportion of aperture, 50% of the shell width. Proportion of last whorl 52% of the height.
- 362 Varices strictly aligned for most of spire height, becoming a little displaced only on last whorl
- 363 or two of large specimens. Abapertural face of each varix deeply excavated, buttressed by
- spiral cords. Spiral ornamentation as in *B. granularis*. After whorl 2 (varix 4) IP grows in
- importance to equalother cords, only to reduce again after varix 6, leaving room for
- development of secondary cord above it. Every primary cord surrounded by secondary cords,
- 367 growing in importance similarly to primary cords. Spiral ornamentation composed, on last
- 368 whorl, of 9 nodules per primary cord between two varices with the exception of P1, which
- 369 bears much bigger nodules, pointing slightly upwards, and decreasing in number with
- ontogeny, from 9 to 5. Secondary cords bear same number of small nodules as *B. granularis*.
- Nodules on varices where they meet P1 angulating the shoulder. Apertural lips well flared,
- bearing numerous prominent, narrow, internal transverse denticles. ID2 present in large

specimens, ID1 bifid, D1 bifid, D2 single but bifid in large specimens, D3-4 bifid, D5 single,
D6 bifid (Fig. 5).

375

376 *Distribution and habitat: Bursa affinis* has a tropical distribution throughout the Pacific

377 Ocean (Fig. 1), from Okinawa, Japan to Sydney Harbour, New South Wales, Australia, and

throughout Polynesia to Hawaii (Beu, 1998). In the eastern Pacific, particularly large

specimens are common at Clipperton Island (Hertlein & Allison, 1960; Kaiser, 2007) and

specimens were recorded by Emerson (1991) from the Revillagigedo Islands, Cocos Island,

and the mainland coast at Bahia Chamela, Jalisco, western Mexico (Beu, 2010). Its westward

distribution in the Pacific extends from Japan to Australia through the Philippines, Papua New

383 Guinea, Vanuatu and New Caledonia. *Bursa affinis* is commonly found in very shallow water

but specimens are occasionally found alive in around 40 m deep.

385

386 *Remarks: Bursa affinis* and *Bursa livida* have the same type locality; "Annaa," [now Anaa]

Tuamotu Islands. The type specimens (Fig. 4 E,G) of both have the same OLDF, and we

consider the latter to be a junior synonym of the former. Considering that there is only one

MOTU occurring in Outer Oceania in general (Fig. 1), and French Polynesia in particular, we attribute to it the name *affinis*.

201

Fig.6

Bursa cubaniana (d'Orbigny, 1841)

- 393 (Fig. 6 A-D,L)
- 394

Ranella cubaniana d'Orbigny, 1841: 165, pl. 23, fig. 24 (Ste Lucie, West Indies, Holotype

396 NHMUK 1854.10.4.412); Mörch, 1877: 24.

397 *Gyrineum affine var. cubanianum* — Dall, 1889: 224.

398 *Gyrineum afine (sic) cubanianum* — Morretes, 1949: 92.

399 Bursa (Colubrellina) cubaniana — Abbott, 1958: 57, text-fig. 2, pl. 1, fig. k. Warmke &

400 Abbott, 1961: 103, pl. 18, fig. i; Rios, 1970: 70; Coelho & Matthews, 1971: 52, figs 11-12;

401 Humfrey, 1975: 126, pl. 14, figs 4, 4a; H. & E. Vokes, 1983: 23, pl. 11, fig. 14.

402 Bursa cubaniana — Nowell-Usticke, 1959: 61. Work, 1969: 663; Kaufmann & Gotting,

403 1970: 372, fig. 87; Matthews & Kempf, 1970: 28; de Jong & Coomans, 1988: 214, pl. 16, fig.
404 382.

405 Bursa cubaniana var. intermedia Nowell-Usticke 1959: 61-62, pl. 3, fig. 13 (North of

406 Fredericksted, St. Croix, Virgin Islands).— Boyko & Cordeiro, 2001: 24.

- 407 Bursa corrugata var. lineata Nowell-Usticke 1959: 61-62, pl. 3, fig. 12 (North of
- 408 Fredericksted, St. Croix, Virgin Islands).— Boyko & Cordeiro, 2001:24.
- 409 Bursa granularis cubaniana Abbott, 1974: 167, pl. 7, fig. 1781 (as B. granularis in pl.
- 410 caption). Rios, 1975: 81, pl. 23, fig. 331; Finlay, 1978: 149; Bandel, 1984: 102, pl. 10, figs 3,
- 411 8; Diaz & Gotting, 1988: 156; Diaz, 1990: 19; Diaz & Puyana, 1994: 174, fig. 650; Redfern,
- 412 2001: 58, pl. 29, figs 247A-B. Nappo *et al.*, 2014: pl. 5, figs. 2-3, pl. 6, figs. 1-3, pl. 7, fig. 2
- 413 Bursa (Colubrellina) granularis cubaniana Rios, 1975: 81, pl. 23, fig. 331; Beu, 1985: 64;
- 414 Cossignani, 1994: 78.
- 415 Bursa (Colubrellina) granularia (sic) cubaniana Rios, 1994: 92, pl. 31, fig. 365.
- 416 Bursa granularis Leal, 1991: 111, pl. 16, figs C-D; Lipe & Abbott, 1991: 14-15, illus ; Beu
- 417 2010: 48, pl. 3, figs 1, 3; Redfern, 2013: 83, figs 241 (not Röding, 1798).
- 418 Bursa (Colubrellina) granularia (sic) Rios, 1985: 79, pl. 28, fig, 347 (not of Röding,
- 419 1798).
- 420 *Gyrineum affine* Dall, 1903: 132 (not of Broderip, 1833).
- 421 Ranella (Bursa) affinis Trechmann, 1933: 39 (not of Broderip, 1833).
- 422
- 423 *Material examined:* See Supplementary Material table 1.
- 424 *Diagnosis:* Shell biconic, varices strictly aligned, varices abaperturally deeply excavated,
- relatively numerous nodules on all primary cords, particularly large one on P1, beige stripes
- following primary cords, OLDF: 3,5,4,3 or 3,5,5,4 in particularly large specimens.
- 427 *Description:* Protoconch similar to other species in the complex (see above). Teleoconch tall
- 428 and relatively narrow with 4-5 whorls (up to six for larger specimens). H around 50 mm, up to
- 429 60 mm. Spire angle 55°; proportion of aperture, 50% of the shell width; proportion of last
- 430 whorl 47% of the height. Varices strictly aligned for most of spire height, becoming a little
- displaced only on last whorl or two in large specimens. Abapertural face of each varix deeply
- 432 excavated, buttressed by spiral cords. Spiral ornamentation much reduced compared to *Bursa*
- 433 granularis and B. affinis. P1, 3 and 5 much more pronounced than others. Primary cords (IP
- 434 included) bearing 4 rather large nodules, with small nodules overlying large ones on large
- 435 specimens. Secondary cords not visible on shell after whorl 1, except slightly visible where
- 436 intersecting varices. Nodules on varices where they meet P1, angulating shoulder, not as sharp
- 437 as on *B. affinis*. Apertural lips well flared, bearing numerous prominent, narrow transverse
- 438 internal denticles;s ID2 single, ID1 bifid, trifurcated in large specimens, D1 bifid, trifurcated
- 439 in large specimens, D2 bifid, D3 bifid, trifurcated in large specimens, D4 bifid, D5 single,

110	hifid in large specimens. D6 hifid (Fig. 5) Shalls distinctively bearing beigg group strings
440	following the spinel cords
441	Di cil ci di la li constanzia di ante anno in the mastern Atlantic (Ei a la December)
442	Distribution and nabitat: Bursa cubaniana occurs in the western Atlantic (Fig. 1; Rosenberg
443	2009), where it is recorded from southeastern Florida (Abbott, 1974; this study), Texas and
444	Louisiana (Garcia, 1999) westwards to Mexico, Costa Rica, Panama and Colombia
445	(Massemin et al. 2009), the Bahamas (Redfern 2013), Guadeloupe (this study), Jamaica, Cuba
446	and St Lucia (d'Orbigny, 1841) south to French Guiana (Massemin et al. 2009), Bahia and
447	Pernambuco, Brazil (Matthews, 1968; Rios, 1985, 1994, Mello & Perrier 1986). In the eastern
448	Atlantic, it has been recorded only from the Cape Verde Islands, but this record in
449	unconfirmed (Rolán, 2005).
450	Remarks: The type locality of Bursa cubaniana is Cuba and there is only one MOTU in the
451	Caribbean region (Fig. 1). So the name Bursa cubaniana is unambiguously attributed to this
452	MOTU.
453	
454	Bursa elisabettae Nappo, Pellegrini & Bonomolo, 2014
455	(Fig. 6 E-G,M)
456	
457	Bursa granularis elisabettae Nappo, Pellegrini & Bonomolo, 2014: 5, pl. 1, figs 1- 3; pl. 2,
458	figs 1-3; pl. 3, figs 1-3 (Flinders Bay, Cape Leeuwin, southern Western Australia, 12 m,
459	MAC.ML 1536)
460	Bursa cf. nodosa — Bozzetti, 1991:1, fig.1 (not of Borson, 1823)
461	Bursa granularis "affinis" form — Wilson, 1993: pl. 43, fig. 12 (not of Röding, 1798).
462	
463	Material examined: See Supplementary Material table 1.
464	Diagnosis: Shell biconic, varices strictly aligned, varices abaperturally rather excavated,
465	nodules weak on the first primary cords, brown stripes following primary cords, outer lip
466	denticles small, OLDF:4,3,3,3 or 4,3,2,3 in large specimens.
467	Description: Protoconch similar to that of other species in the complex (see above).
468	Teleoconch tall, relatively narrow with 4-5 whorls, H around 50 mm. Spire angle 45°; % of
469	aperture, 52% of the shell width; % of last whorl 52% of the height. Varices strictly aligned,
470	although low and hardly visible in some specimens. Spiral ornamentation very reduced to
471	absent from some specimens. Primary cords P1, 3 and 5 are the only ones visible; each bears
472	4 nodules between two varices, especially visible on P1. No secondary cords present. Nodules
473	on varices where they meet P1 angulate shoulder, although not as sharply as in B affinis

Apertural lips well flared, bearing numerous prominent, narrow transverse internal denticles;
ID2-1 bifid, D1 single, D2 bifid, D3 single, D4 bifid, merged in large specimens, D5 single,
D6 bifid (Fig. 5).

477

478 *Distribution and habitat:* The known distribution of *Bursa elisabettae* is restricted to
479 southwestern Western Australia, in shallow water.

480

Remarks: Bursa elisabettae is the last species described in the complex. There is only MOTU
occurring around Cape Leeuwin, southwestern Australia (type locality) to which we attribute
the name *B. elisabettae*. The possible occurrence of *B. elisabettae* in South Africa is discussed
below.

485

486 DISCUSSION

Bursa granularis is considered to be easy to identify, although it is most variable, hence the 487 488 thirteen names it received through history and the innumerable subsequent references in the literature. Previous works failed to provide a comprehensive characterization of such 489 490 variability, leading to the aggregation of the whole morphological spectrum under a single name, Bursa granularis (Beu, 1998, 2005, 2010). An integrated taxonomic approach based on 491 extensive sampling (geographical, bathymetrical, and type locality of many nominal taxa), 492 coupled with a detailed analysis of the shell variability and the sequencing of the cox1 gene, 493 analysed with two methods (ABGD and GMYC), revealed four groups within the Bursa 494 granularis complex, here considered to be distinct species. However, our species hypotheses 495 are supported by one gene and morphological data only; an independent genetic marker would 496 497 help to determine whether the partition obtained with the Cox-1 gene corresponds to the species limits. 498

Among the four delimited species, one pair, Bursa granularis and B. affinis, is barely 499 distinguishable morphologically, and without molecular evidence they would hardly be 500 501 suspected to be separate species. Furthermore, the intraspecific morphological variability sometimes exceeds the interspecific disparity, particularly in shell coloration; the pinkish 502 503 coloration of the first whorls, formerly used to discriminate B. affinis (see Reeve, 1844) can 504 be found in specimens of both species, and some specimens present the striped pattern usually 505 attributed to B. cubaniana (for which this pattern is stable). We were able to discriminate species on morphology only through the detailed observation of the spiral ornamentation of 506 507 the shells. The importance of such ornamentation for systematics was previously recognized

in, e. g., Muricidae (Merle & Houart, 2003; Merle, 2005; Merle *et al.*, 2001), Mathildidae

509 (Bieler, 1995) and Calliostomatidae (Marshall, 1995). Although Muricidae and Bursidae are

510 not closely related, they have a similar morphological organization of the sculptural elements

and the methodology applies as well to Bursidae as to Muricidae. Other morphological

512 characters, e.g. apertural color, spire height, and sculptural prominence (considering one cord

513 at a time), are of very little value.

514 Other anatomical characters and radulae were not considered in the present study but 515 may perhaps provide useful information. Abbott (1958) pointed out differences between *B*. 516 *cubaniana* and *B. granularis* in denticle size and number on the central teeth (Abbott, 1958: 517 figs 1d, 2c), but radular studies are often based on too small a sample size and their results 518 may not be significant. For good pictures of the radula of *B. granularis*, see Barbakova *et al.* 519 (2016) and for insight on the general anatomy of *B. cubaniana* see Simone (2011).

520 The integrated taxonomic approach followed here was thus efficient to propose robust species hypotheses. It is an additional example of the value of molecular characters when 521 522 species cannot readily be distinguished morphologically, a common pattern in gastropods (e. g. Puillandre et al., 2012b, Duda et al., 2008). However, if proposing putative species using 523 524 DNA sequences is now a common practice, linking the species hypotheses to available names attached to non-sequenced name-bearing types remains daunting. Indeed all the species in the 525 Bursa granularis complex were described based on shell characters only, including B. 526 elisabettae, described as recently as 2014. More generally, most species of molluscs were 527 described before the molecular era, and the application of historical names remain based on 528 non-sequenced, sometimes problematical, dry material (Bouchet & Strong, 2010). 529

530 Because we did not sample the entire distribution area of the complex, additional species possibly may be recognized in the future. Bursa alfredensis and B. kowiensis (Fig 6 I-531 K, N)were both described from Port Alfred, South Africa and are the only two names that we 532 have refrained from associating with one of the MOTU of this study. The types of both are 533 specimens displaying a loss of the fine sculpture early in ontogenesis. The adult South African 534 535 specimen in Fig. 6 I (for which we have no molecular data) certainly corresponds to the nominal species described as B. kowiensis by Turton (1932). This loss of ornamentation 536 537 resembles the one displayed by *B. cubaniana*, but even more by *B. elisabettae*. If *B.* elisabettae, B. alfredensis and B. kowiensis were found to be a single species, then B. 538 539 kowiensis or B. alfredensis, both names established in 1932, would be the valid name. Such a scenario would be congruent with the hypothesis of a Pleistocene colonization of the 540 541 Caribbean (which happens to be the stratigraphic distribution of *B. cubaniana* in the region;

Beu, 2010) from the Indian Ocean, around South Africa. This hypothesis was discussed by
Vermeij & Rosenberg (1993) for 17 Indo-West-Pacific species.

South African collectors (R. Aiken, pers. com.) have also drawn our attention to a
small form of *Bursa granularis* that is dredged in "deep water" (Fig.6 H). We have not seen
this variation from anywhere else, and the deep water (100 meters?) habitat is very unusual
from *B. granularis* s.s., which has been exceptionnally dredged alive in 87-90 m (specimen
MNHN IM-2009-5148, sequenced). The status of this small, South African, deep-water form
will have to be evaluated by sequencing.

550 Recent work (Nappo et al., 2014) suggested that Bursa granularis, B. cubaniana, and B. elisabettae were subspecies of B. granularis. We rank them here as full species. Firstly, as 551 552 we showed above, the two pairs B. granularis-affinis and B. cubaniana-elisabettae seem to have very distinct biogeographical histories. Secondly, Castelin et al. (2012, fig. 7) showed 553 554 that the pair B.granularis-affinis diverged from each other over 11 MYA (the distance between B. cubaniana and B. elisabettae would suggest a comparable time of divergence, 555 556 although it was not tested). The divergence between the two pairs is necessarily older. The estimated divergence time (11 MYA; Castelin et al., 2012) between Bursa granularis and 557 558 B. affinis corresponds to the Seravallian/Tortonian stages, a period with a particularly low sea level (around 100 m lower than today; Scotese, 2014). Australia, Papua New Guinea, 559 Indonesia, Japan and the Philippines were all connected to Eurasia by a series of land bridges 560 or very shallow seas, reducing to nearly zero all marine connectivity between the Pacific and 561 Indian Oceans. Given the sympatric distribution of *B. granularis* and *B. affinis* in an area 562 extending from Japan to eastern Australia through the Philippines, Papua New Guinea and 563 564 New Caledonia, this speciation event would represent an example of allopatric speciation with secondary contact subsequent to recolonisation events. However, because our results are 565 566 based only on mitochondrial data, we cannot rule out an alternative scenario in which both groups are actually still genetically connected. In this scenario, the pattern observed with the 567 cox1 gene would only be the result of an ancient allopatric divergence, the secondary contact 568 569 being too recent to homogenize both mitochondrial gene pools. Although the number of sequenced specimens and the high genetic divergence between *B. affinis* and *B. granularis* do 570 571 not support this hypothesis, nuclear data would help support one or the other of these scenarios. In a forthcoming molecular phylogeny of the Tonnoidea (Strong et al., in prep.), 572 573 nuclear gene sequences are available for a limited number of specimens in the Bursa granularis complex but they do provide confirmation of the present cox-1 based species 574 575 delimitations.

One of the main consequences of the partition into four species proposed here is that 576 larval duration alone does not explain geographical distributions. Other factors, such as 577 availability of suitable habitats, may limit distribution areas, in spite of a species' large 578 579 dispersal abilities. A striking example is *Fusitriton oregonensis*; it is the only species for which the larval duration has been directly measured (although under laboratory conditions), 580 with an estimated larval stage of 5 years (Strathmann & Strathmann, 2007), but its 581 distribution area is limited to the North Pacific from central Japan to California, extending 582 583 south to off Panama in deep water (Smith, 1970).

584 While the distribution area of Bursa granularis remains large, we have shown that it was greatly overestimated in the classic taxonomic extension of the species. It is not unlikely 585 586 that the same pattern – one very variable species with an extremely large distribution being in fact several species with smaller distributions – could be found in other tonnoideans. For 587 588 Bursidae only, Bursa corrugata (Perry, 1811) (8 subjective synonyms and recorded from East Pacific, Eastern Atlantic Ocean and the Caribbean), Bursa ranelloides (Reeve, 1844) (7 589 590 subjective synonyms, recorded from the Indian and Atlantic Oceans, and Japan and Hawaii in the Pacific) and Bursa rhodostoma (G. B. Sowerby II, 1835) (6 subjective synonyms, 591 592 recorded from the whole Indo-Pacific and the Caribbean) are good candidates for an integrative taxonomy-based revision. However, achieving a representative sampling for 593 molecular taxonomy becomes increasingly difficult, considering the multiplicity of permits 594 necessitated by sampling distribution areas that span numerous countries. The future will tell 595 whether Article 8a of the Nagoya Protocol to the Convention of Biological Diversity 596 (encouraging the Parties to the protocol to "create conditions to promote and encourage 597 research [...] through simplified measures on access for noncommercial research purposes") 598 will reverse the ebb of the tide (Bouchet et al., 2016). For the time being, complying with 599 regulations on "Access and Benefit Sharing" and "Prior Informed Consent" is a formidable 600 601 obstacle on the road of academic researchers investigating species limits and their names. 602

603

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- 996 phylogenetic framework. *Molecular Phylogenetics and Evolution*, **61**: 425–435.
- 997
- 998 CAPTIONS
- 999 **Figure 1.** Distribution of the *Bursa granularis* complex recorded by the Global Biodiversity
- 1000 Information Facility (GBIF 2016). Records from Senegal were removed after being re-identified as
- 1001 *Bursa scrobilator* (Linnaeus, 1758). Type localities of corresponding species are marked by arrows.
- 1002 Figure 2. Shell characters of *Bursa granularis*: A. global apertural view. B. outer lip.
- 1003 (Specimen figured: UF-423792, UF cruise 2008 (Madagascar), Nosy Komba, NW side). IP:
- 1004 infrasutural ramp primary cord, IS: infrasutural ramp secondary cord, P1-6: primary cords, S1-6:
- secondary cords, AN: anal notch, SC: syphonal canal. ID1: infrasutural denticle; , D1-6 Denticles of
- the convex part of the whorl.

- 1007 Figure 3. Molecular tree for the cox1 gene. Bursa granularis (Röding, 1798) is represented by the
- shell of UF-423792 (marked by a black square on the tree); *Bursa affinis* (Broderip, 1833) is
- represented by the shell of IM-2007-43056 (marked by a white square on the tree); Bursa cubaniana
- 1010 (d'Orbigny, 1841) is represented by the shell of IM-2009-23406 (marked by a black hexagon) and
- 1011 Bursa elisabettae Nappo, Pellegrini & Bonomolo, 2014 is represented by the shell of IM-2009-23322
- 1012 (marked by a black star). Support for nodes is given as PP/BS, where these exceed 0.95 and 95,
- 1013 respectively. PNG: Papua New Guinea.
- 1014 Figure 4. Shells of the Bursa granularis complex. A-D. Bursa granularis (Röding, 1798): A. MHNG
- 1015 1098/85/1, Lectotype of *Ranella granifera* Röding, 1798, Neotype of *Tritonium granulare* Röding,
- 1016 1798, of *T. jabick* Röding, 1798, and of *Biplex rubicola* Perry, 1811, from the Red Sea H: 64 mm. B.
- 1017 IM-2009-5148, MAINBAZA 2009 (Mozambique Channel), stn DW3168, H: 26 mm. C. UF-423792,
- 1018 UF 2008 (Madagascar), Nosy Komba, NW side.H: 40 mm. D. IM-2007-43071, Santo Marine
- 1019 Biodiversity Survey 2006 (Vanuatu), stn VM62, H: 25 mm. E-I. Bursa affinis (Broderip, 1833): E.
- 1020 MNHUK 1950.11.28.4, Lectotype of *Ranella affinis* Broderip, 1833, supposedly from "Annaa,"
- 1021 Tuamotu Islands, H: 62.5 mm F. IM-2007-43056, Santo Marine Biodiversity Survey 2006 (Vanuatu),
- 1022 stn VM02, H: 37 mm. G. NHMUK 1967657 Lectotype of *Ranella livida* Reeve, 1844, supposedly
- 1023 from "Annaa," Tuamotu Islands, H: 56.8 mm H. UF-422918, UF 2008 (Federated States of
- 1024 Micronesia), Kosrae Letu, in front of Nautilus Hotel, H: 25 mm. I. IM-2007-43039, PANGLAO 2004
- 1025 (Philippines), stn R24, H: 41mm. J. IM-2009-5148 Bursa granularis protoconch close-up. K. IM-
- 1026 2007-43056 *Bursa affinis* protoconch close-up. P1, P3, P5 preponderant primary cords; numbers
- 1027 indicates the number of denticles
- 1028

Figure 5. Synthetic diagrams showing the sequences of appearance of the primary and secondary
cords in the four species of the *Bursa granularis* complex. In all ontogenies, the primary cords (black
lines) are numbered following an adapical to abapical order.

- 1032
- 1033 Figure 6. Shells of the Bursa granularis complex. A-D. Bursa cubaniana (d'Orbigny, 1841): A.
- 1034 NHMUK 1854.10.4.412, Holotype of *Ranella cubaniana* d'Orbigny, 1841, St. Lucie, West Indies, H:
- 1035 51.7 mm **B.** IM-2009-23406, Port Louis, Pointe Antisu, Guadeloupe, H: 41mm. **C.** IM-2013-20184,
- 1036 KARUBENTHOS 2012 (Guadeloupe), stn GM09, H: 51mm. **D.** UF 437626, UF 2010 (Florida, USA),
- 1037 Florida, Monroe County, Florida Keys, Tennessee Reef, H: 3.5 cm. E-G. Bursa elisabettae Nappo,
- 1038 Pellegrini & Bonomolo, 2014 E. MAC.ML 1536 Holotype of Bursa granularis elisabettae Nappo,
- 1039 Pellegrini & Bonomolo, 2014, Flinders Bay, Cape Leeuwin, South West Australia, H: 35 mm. F. IM-
- 1040 2009-23319, WESTERN AUSTRALIA 2011, stn WA17, H: 31mm. G. IM-2009-23313, WESTERN
- 1041 AUSTRALIA 2011, stn WB28, H: 28 mm. H "deep sea" form of south African Bursa granularis
- 1042 MNHN ex coll. Aiken, off Richard bay, South Africa, dredge H: 24.8mm-I Bursa aff.kowiensis
- 1043 MNHN ex coll. Aiken, Coffee Bay, South Africa H: 40 mm. dived J. Holotype of Bursa kowiensis

- Bursa cubaniana protoconch close up. M. IM-2009-23313, Bursa elisabettae protoconch close-up. N.
- MNHN ex coll. Aiken, Bursa aff. kowiensis protoconch close-up. P1, P3, P5 preponderant primary
- cords; numbers indicates the number of denticles









