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Isooctyl-3-(3,5-di-tert-butyl-4-hydroxyphenyl)propionate in silicone matrix

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Rosa Navarro, Ludmila Audouin, Jacques Verdu. Reactions of antioxidants with molecular oxygen. Part II. Isooctyl-3-(3,5-di-tert-butyl-4-hydroxyphenyl)propionate in silicone matrix. *Polymer Degradation and Stability*, 2011, 96 (5), pp.965-973. 10.1016/j.polymdegradstab.2011.01.024 . hal-02456020

HAL Id: hal-02456020

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Reactions of antioxidants with molecular oxygen. Part II. Isooctyl-3-(3,5-di-tert-butyl-4-hydroxyphenyl)propionate in silicone matrix

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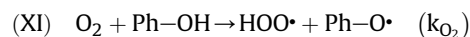
The study of oxidation kinetics of isooctyl-3-(3,5-di-tert-butyl-4-hydroxyphenyl)propionate (Irganox 1135) stabiliser was carried out using silicone oil as an inert substrate. The stabiliser was exposed at 80–120 °C under 0.02–3.0 MPa oxygen pressure. UV/VIS spectrophotometry, gel permeation chromatography and high pressure liquid chromatography were used to follow the stabiliser consumption, whose kinetic parameters were determined by applying a simple kinetic model.

Keywords:

Phenolic antioxidant
Thermal oxidation
Kinetics
UV/VIS
GPC
HPLC

1. Introduction

The reaction between hindered phenols used as polymer antioxidants and molecular oxygen was studied a long time ago [1–3]. In its ground state, oxygen is a biradical potentially able to abstract hydrogen from organic substrates. This reactivity is well known to specialists of high temperature gas phase oxidations, but it is slow in the temperature range of polymer use (typically $T \leq 200$ °C) and practically negligible in hydrocarbon polymers [4]. However, phenols are specially reactive owing to the low dissociation energy (335–355 kJ mol⁻¹) of the O–H bond [5,6]. The whole mechanism can be complex, but the first (and rate controlling) step is always the following:



In air, at atmospheric pressure at $T \leq 150$ °C, this reaction is relatively slow which explains why it has been often ignored in papers on stabilisation mechanisms of hindered phenols [7,8]. However, there are circumstances in which there is no other way to explain the observed behaviour, than the hypothesis of a partial consumption of the stabiliser by its reaction with oxygen. The main characteristic feature of this behaviour is the kinetic curve of

stabiliser consumption which displays its maximum rate at the beginning of exposure where the radical concentration is very low so that radical scavenging cannot significantly contribute to stabiliser consumption [4,9].

The rate of reaction (XI) is proportional to oxygen concentration in the polymer and this latter is proportional to oxygen pressure. In certain applications, for instance geotextiles [4], there is an interest in accelerated aging at low temperature (~ 80 °C) and at high oxygen pressure. Indeed, in such conditions, oxygen–phenol reaction can be a non negligible mode of stabiliser consumption, responsible for a reduction of the stabiliser efficiency. This is a first reason to study oxygen–phenol reactions.

Another reason is that this study can bring some light to polymer stabilisation mechanisms. Molecular oxygen is, no doubt, less reactive than macropoxy radicals formed during polymer oxidation, but the reaction product (phenoxy radical) and thus certain of its further by-products must be the same. Thus oxygen–stabiliser reactions offer the possibility to study secondary stabilising processes without the analytical complications linked to polymer matrices. It has been shown in an earlier study [10] that silicone oil (polydimethylsiloxane) can be a convenient matrix for such experiments, provided that the stabiliser is soluble enough. Its main advantages are its total absence of UV absorption, inertness and compatibility with liquid chromatography.

Here, we have chosen to study a monofunctional phenol: isooctyl-3-(3,5-di-tert-butyl-4-hydroxyphenyl)propionate (Irganox

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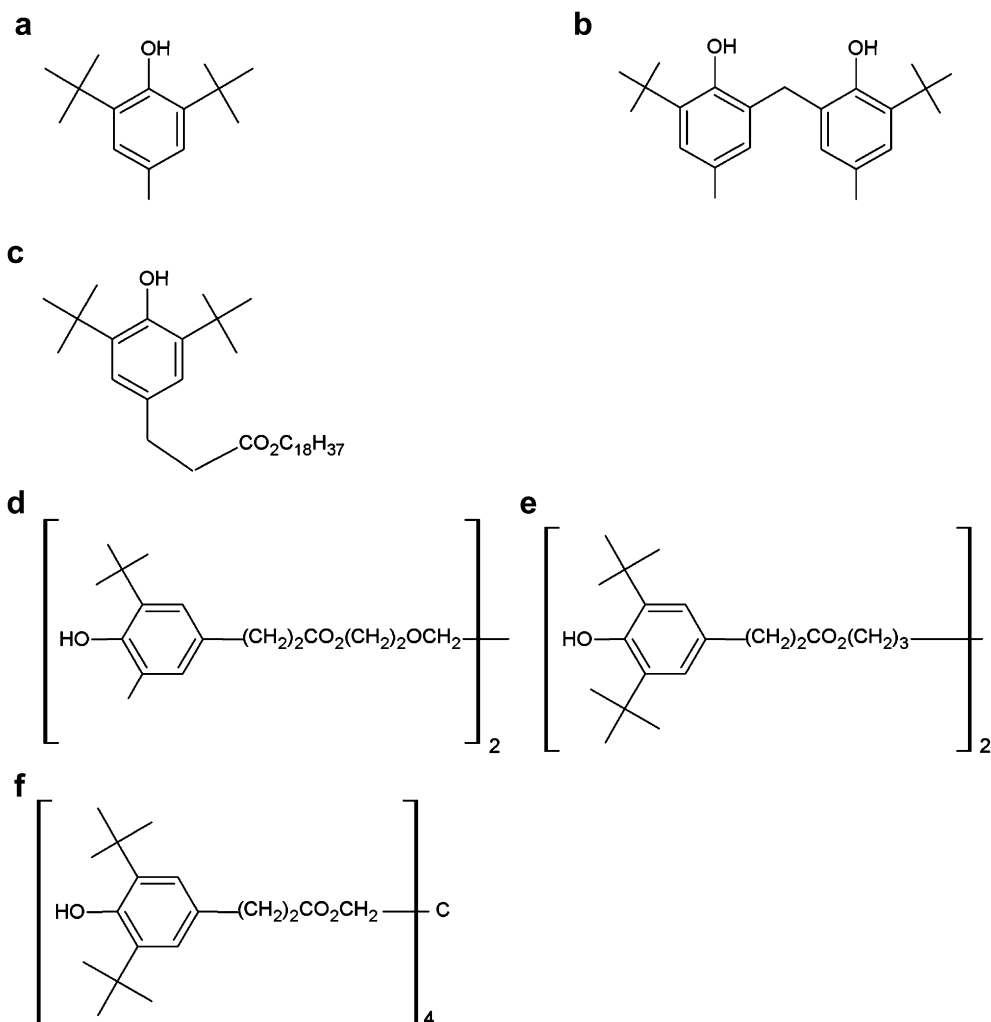


Fig. 1. Structure of calibration compounds: (a) ionol, (b) Irganox 2246, (c) Irganox 1076, (d) Irganox 245, (e) Irganox 259 and (f) Irganox 1010.

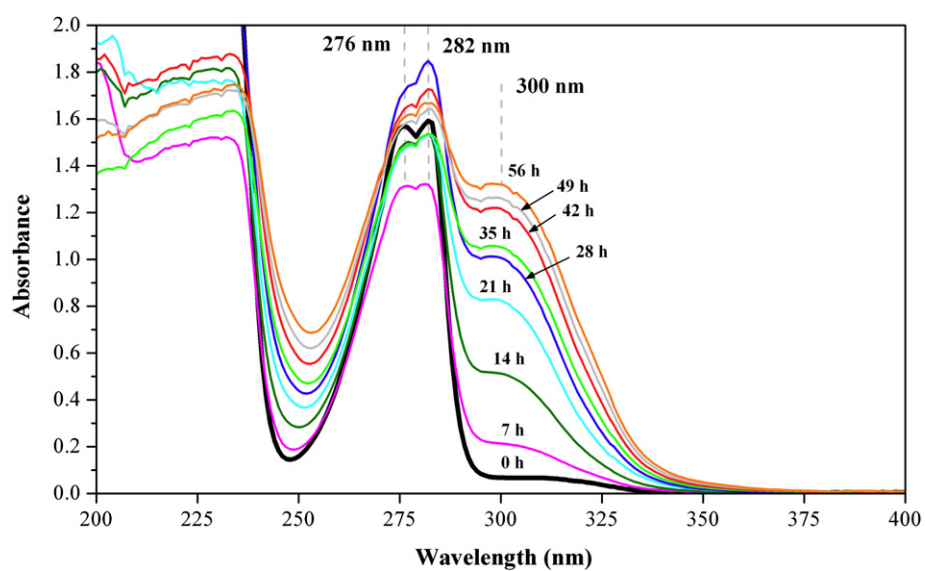


Fig. 2. Series UV/VIS spectra of the AO4 antioxidant after oxidation exposure at 90 °C under 3.0 MPa oxygen pressure.

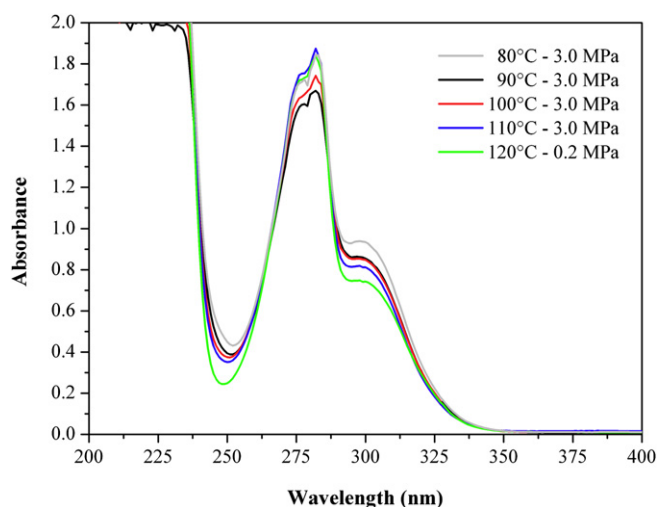


Fig. 3. UV/VIS spectra of AO4 after consuming ~25% of initial stabiliser at different oxidation conditions.

1135), which is well soluble in silicone and belongs to an important antioxidant family characterized by the presence of a propionate ester grafted to the phenyl ring in the *para* position relative to the OH group. Reaction products obtained in the 80–120 °C temperature range, 0.02–3.0 MPa oxygen pressure range were studied by UV/VIS spectrophotometry, gel permeation chromatography and high pressure liquid chromatography.

2. Equipment and experimental procedure

2.1. Materials

The stabiliser used in this study was isooctyl-3-(3,5-di-tert-butyl-4-hydroxyphenyl)propionate (AO4) (commercial name Irganox 1135 from Ciba SC). This antioxidant has a molar mass of 390.6 g mol⁻¹. Silicone oil DC 200 (viscosity 50 mPa s at 25 °C, from Sigma–Aldrich) was employed as an inert substrate.

2.2. Oxidation conditions

The phenolic stabiliser was dissolved in the polydimethylsiloxane to give 1.74 × 10⁻² mol l⁻¹ concentration (0.68% weight). The oxidation of antioxidant solution was studied in an air-ventilated oven at 120 °C, and in autoclaves under 0.2–3.0 MPa oxygen pressure range at 80, 90, 100, 110 and 120 °C. In the air-ventilated oven, the sample was introduced inside a closed container under air to prevent the evaporation of the antioxidant. The way to optimize these exposure conditions was presented in our previous article [10].

2.3. Analytical methods

The direct reaction between the stabiliser and the oxygen was followed by three different analytical methods: UV/VIS spectrophotometry, high pressure liquid chromatography (HPLC) and gel permeation chromatography (GPC).

Absorption spectra between the wavelengths of 200 and 500 nm were recorded in a Perkin–Elmer Lambda 35 UV/VIS spectrophotometer. Two cells of 500 μm thickness were used to perform these measurements.

Both HPLC and GPC were carried out in a WATERS 714 chromatograph equipped with a photodiode array detector (PDA) and a differential refractometer. The PDA detector is an UV/VIS spectrophotometer which operates within a wavelength range of 190–800 nm. The experimental conditions of chromatography varied according to the system used. In HPLC method, a symmetric C18 column (5 μm, 4.6 × 150 mm) was employed. The measurements were carried out under isothermal conditions at 40 °C. The mobile phase consisted of acetonitrile at a flow rate of 1 ml min⁻¹. Several acetonitrile solutions of the phenol under study in known concentrations were prepared and analysed by HPLC in order to obtain a calibration curve. The amplitude of the chromatographic peak was found to be proportional to the phenol concentration. For antioxidant solutions in silicone oil, the following procedure was used: 2 ml of acetonitrile were added to a known quantity (~70 mg) of the silicone oil containing the phenol. The mixture was then vigorously stirred. After a brief rest period, both phases were separated and the acetonitrile phase was analysed by HPLC.

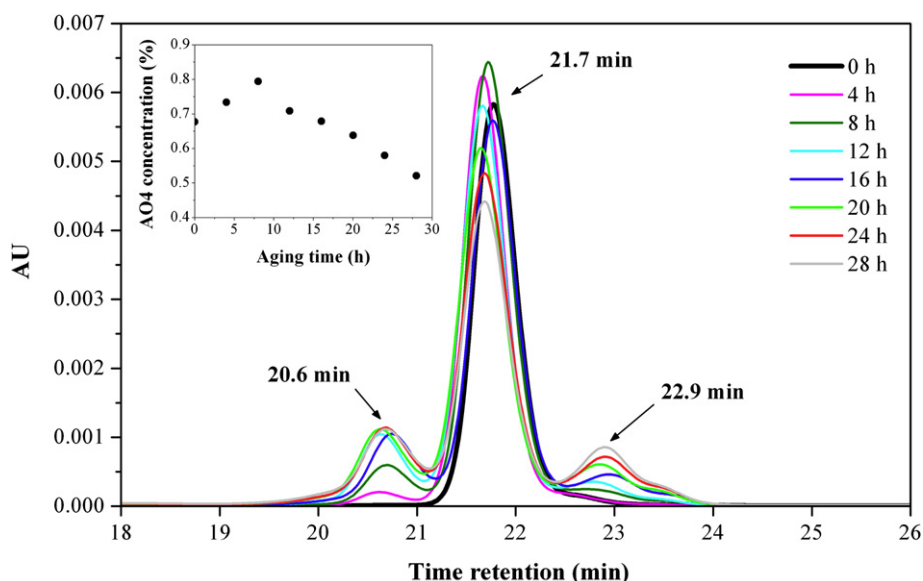


Fig. 4. Series GPC chromatograms of the AO4 antioxidant acquired at 276 nm after oxidation exposure at 110 °C under 3.0 MPa oxygen pressure.

From experiments made in virgin samples, it was shown that almost all the phenol initially present in silicone moved into acetonitrile. Although the phenol subsists, no doubt, in the silicone, it can be considered in a first approach that it is totally extracted, within experimental error, by acetonitrile. Since neither control samples of known concentration nor calibration curve were available for phenol by-products, chromatographic data relative to these species will not be quantitatively interpreted.

In GPC system, the temperature of column was maintained at 40 °C too, but HR1 and HR4E columns (5 μm , 4.6 \times 300 mm), placed in serial order, replaced the previous C18 column. This system enabled us to cover an effective molecular weight range from 50 to 100,000 g mol^{-1} . Tetrahydrofuran (THF) was used as mobile phase at 0.3 ml min^{-1} flow rate. The molar mass calibration curve was obtained with different phenolic and aromatic compounds:

toluene, xylene, ionol, Irganox 2246 (2,2'-methylene-bis(4-methyl-6-tert-butylphenol)), Irganox 1076 (octadecyl-3-(3,5-di-tert-butyl-4-hydroxyphenyl)-propionate), Irganox 245 (ethylenebis(oxyethylene)-bis(3-(5-tert-butyl-4-hydroxy-m-tolyl)-propionate)), Irganox 259 (1,6-hexamethylene-bis(3,5-di-tert-butyl-4-hydroxyhydrocinamate)) and Irganox 1010 (pentaerythritol tetrakis(3-(3,5-di-tert-butyl-4-hydroxyphenyl)propionate)). The compound structures are shown in Fig. 1.

3. Results and discussion

3.1. UV/VIS analysis

The UV/VIS spectrum of the unreacted stabiliser displays a sharp doublet at 276 and 282 nm, as it can be seen in Fig. 2. The 276 nm

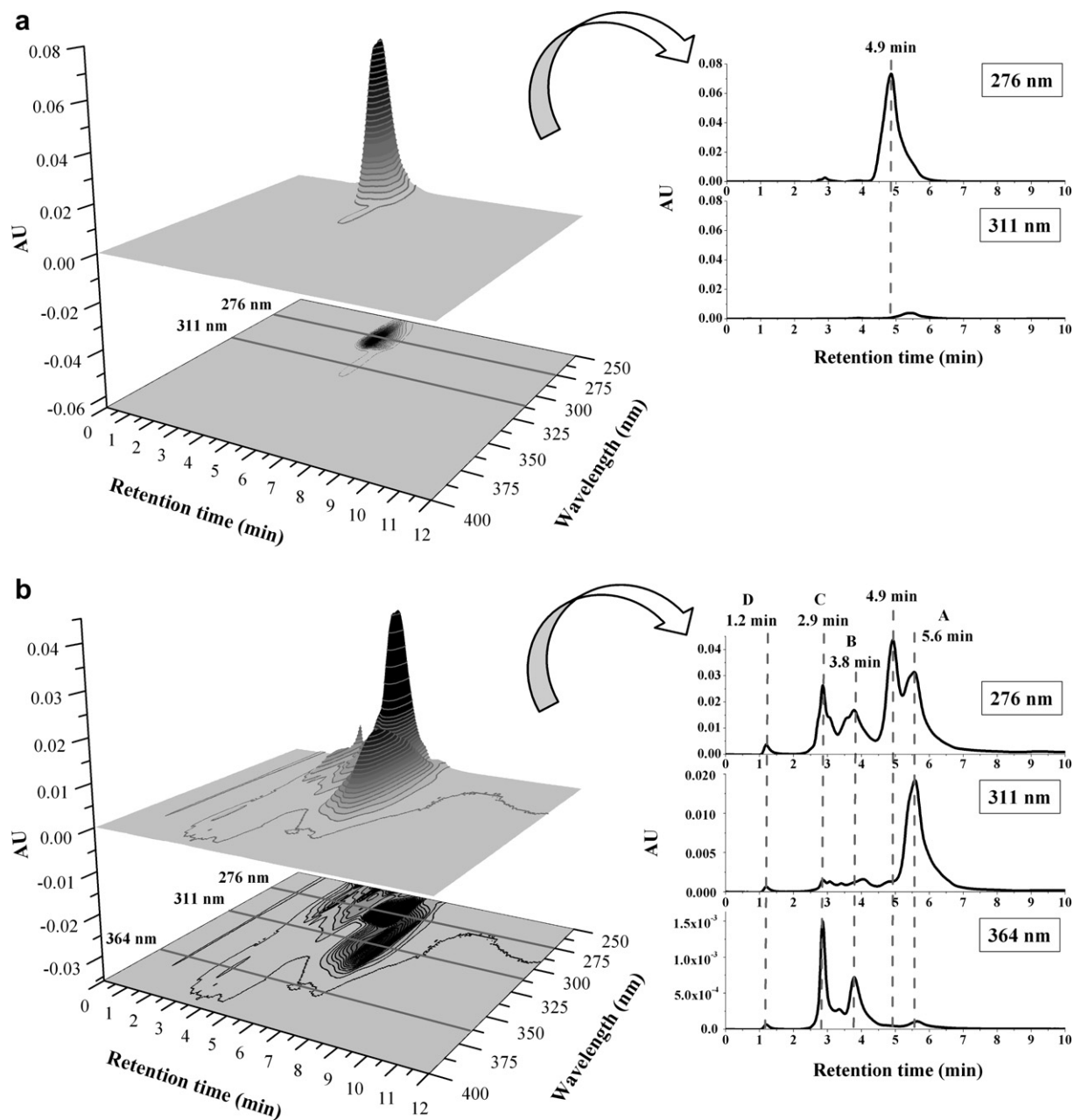


Fig. 5. Tridimensional plot of HPLC chromatograms evolution as a function of the wavelength and the retention time corresponding to the unreacted AO4 (a) and to AO4 exposed at 110 °C under 3.0 MPa oxygen pressure during 28 h (b).

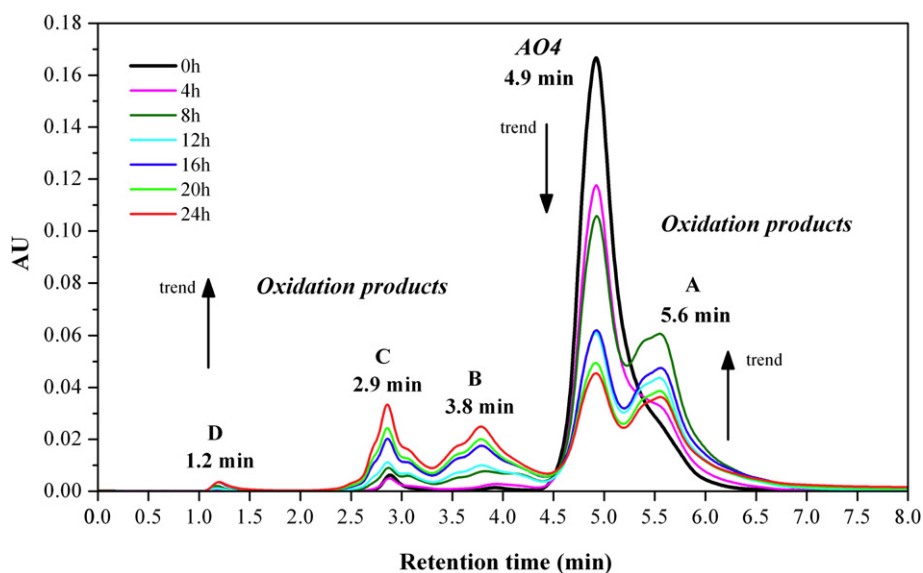


Fig. 6. HPLC chromatograms acquired at 276 nm corresponding to the aging series of the AO4 stabiliser as a function of the reaction time at 110 °C under 3.0 MPa oxygen pressure.

component has been chosen to focus on the kinetic analysis since it is less influenced by the growth of oxidation products absorbing at higher wavelength. Standards with different concentrations of the antioxidant in silicone oil were prepared to determine the molar absorptivity. In the case of AO4, the value of the molar absorptivity was $1742 \text{ l mol}^{-1} \text{ cm}^{-1}$. The unreacted stabiliser absorbs slightly at 300–330 nm range.

Fig. 2 shows the changes of UV/VIS spectra of the AO4 solution with exposure time at 90 °C under 3.0 MPa oxygen pressure. The stabiliser doublet is progressively overlapped by a wide band covering the whole 240–350 nm interval. This band displays a maximum at about 300 nm. Other eventual maxima are masked by overlapping with residual stabiliser bands. The same behaviour was observed in the other experimental conditions under study. This band overlapping hinders the monitoring of the antioxidant consumption and, therefore, a kinetic analysis is impossible using this analytical technique. Despite the complexity of spectra in the oxidation process, according to UV/VIS

analysis, one could say that apparently the same products are formed in the all studied experimental conditions. This is confirmed in Fig. 3, where spectra obtained after nearly ~25% consumption of the initial antioxidant in various conditions are shown.

3.2. GPC analysis

Fig. 4 presents the chromatograms acquired with the UV detector at 276 nm after the exposure of the AO4 antioxidant to oxygen at 110 °C under 3.0 MPa oxygen pressure, as an example of evolution of GPC chromatograms obtained for this antioxidant. The chromatogram of the unreacted stabiliser displays a single peak at 21.7 min, revealing the relative purity of AO4. Oxidation induces the growth of two relatively wide bands respectively centred at 22.9 min and 20.6 min, but extending to 24 min on one side and 19 min on the other side. According to the molar mass calibration, the band maxima would correspond to $\sim 224 \text{ g mol}^{-1}$ (22.9 min)

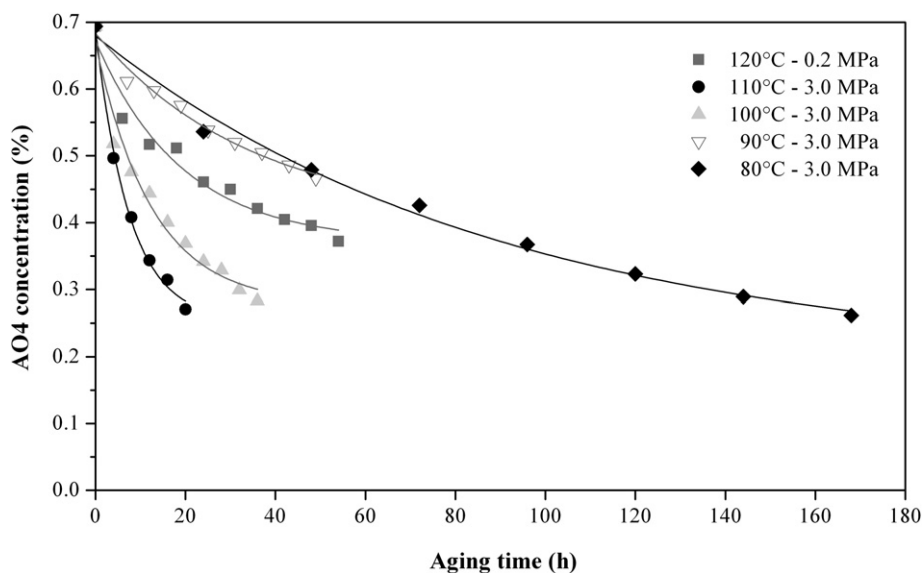


Fig. 7. The consumption curves of the AO4 stabiliser obtained by HPLC analysis at different experimental conditions.

Table 1
Oxygen solubility values calculated in the silicone oil applying Clausius–Clapeyron law.

T – P _{Oxygen}	80 °C	90 °C	100 °C	110 °C	120 °C	120 °C
	3.0 MPa	3.0 MPa	3.0 MPa	3.0 MPa	0.02 MPa	0.2 MPa
S _{O₂} (mol l ⁻¹ Pa ⁻¹) × 10 ⁸	5.00	4.92	4.85	4.78	4.71	4.72

and ~673 g mol⁻¹ (20.6 min), but the molar mass range would extend from ~132 g mol⁻¹ to ~1447 g mol⁻¹.

Both bands of oxidation products increase continuously. Surprisingly, the peak relative to the unreacted stabiliser first increases slightly and then decreases. The simplest explanation of this behaviour is that a by-product having almost the same molar mass as AO4 but with a higher UV absorption “interferes” with AO4 but is more or less rapidly destroyed. This “interference” is not very favourable to the use of GPC for a kinetic study of stabiliser consumption.

3.3. HPLC analysis

The HPLC technique was used to achieve the complete separation between the starting antioxidant and its by-products. Fig. 5 shows the 3D chromatograms of the stabiliser before (a) and after exposure at 110 °C under 3.0 MPa oxygen pressure for 28 h (b). Fig. 5 (a) reveals the presence of two small satellites at retention times respectively close to 3.0 and 4.0 min and a shoulder close to 5.5 min. These three species correspond to the major oxidation products as it is showed in Fig. 5 (b). In other words the stabiliser is initially slightly oxidized, that is not very surprising. Fig. 5 (b) reveals the presence of several by-products in addition to the three aforementioned ones. All these products absorb at 276 nm, but they differ from the initial stabiliser by the following features:

- The by-product A having a retention time close to 5.5 min absorbs strongly at 311 nm but not at 364 nm.
- The by-products B and C having retention times at respectively 3.8 and 2.9 min absorb at 364 nm but not at 311 nm.
- The by-product D having the lowest retention time at 1.2 min, absorbs at 276, 311 and 364 nm.

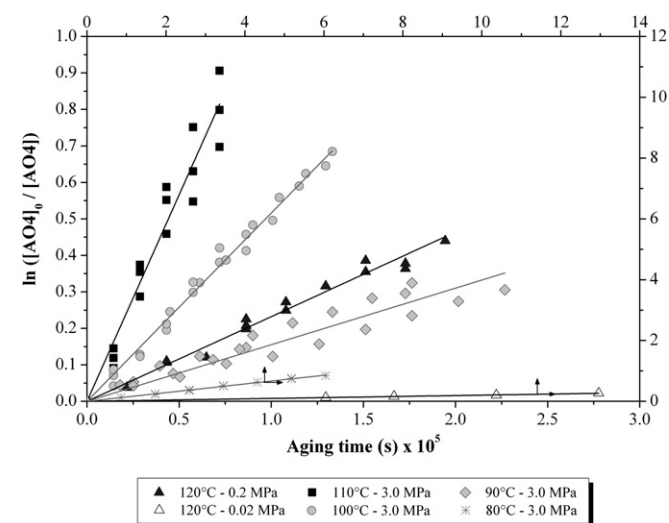
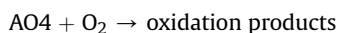


Fig. 8. Obtaining of the pseudo first order constant in different experimental conditions.

UV detection at 276 nm is characteristic for phenolic group absorption and 311 nm for quinone methide structures [11,12]. It is reasonable to suppose that, with this method, the separation is complete and a kinetic study of stabiliser consumption is possible. The study of the growth of by-products is expected to bring interesting complementary information.

Using a single wavelength detection at 276 nm, we have monitored the changes of the HPLC chromatogram in the case of exposure at 110 °C under 3.0 MPa oxygen pressure (Fig. 6). Here, it can be observed that the peak at 4.9 min, corresponding to the starting AO4 molecule, disappears almost completely, that confirms the absence (at least at moderate conversions, typically lower than 50%) of any interference with by-products.

Kinetic curves of the AO4 stabiliser consumption obtained at different experimental conditions are presented in Fig. 7. In all the cases, the kinetic curves seem to correspond to a simple apparent first order process. The same behaviour was found with other phenolic stabilisers in thermal oxidation processes where the oxygen concentration was maintained constant [9,10,13]. As it was seen in a previous study made with 2,2'-methylene-bis(4-methyl-6-tert-butylphenol) antioxidant [10], the true second order rate constant (k_{O_2}) can be estimated graphically from the stabiliser consumption rate using the following equations:



$$r_{AO4} = r_{O_2} = -f_{AO4}k_{O_2}[O_2][AO4] \quad (1)$$

where f_{AO4} is the phenol functionality ($f_{AO4} = 1$ for AO4), $[O_2]$ and $[AO4]$ are the respective oxygen and residual stabiliser concentrations in the silicone oil solution. If $[O_2]$ is considered constant, the system behaves as a pseudo first order process. Therefore,

$$K = k_{O_2}[O_2] \quad (2)$$

$$\frac{d[AO4]}{dt} = -K[AO4] \quad (3)$$

The integration leads to:

$$[AO4] = [AO4]_0 e^{-Kt} \quad (4)$$

where K is the pseudo first order constant and $[O_2] = S_{O_2} \cdot P_{O_2}$, S_{O_2} being the oxygen solubility in the silicone oil (mol l⁻¹ Pa⁻¹), and P_{O_2} the oxygen pressure (Pa). The temperature dependence of the solubility was taken into account through the Clausius–Clapeyron law [14] (Table 1).

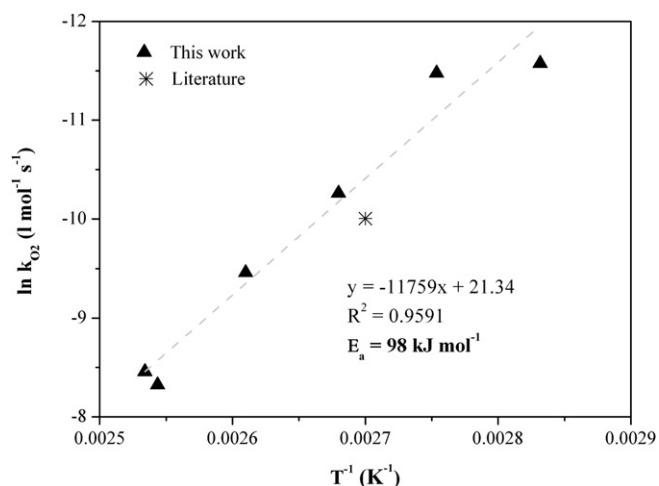
First order plots are shown in Fig. 8. In all the conditions under study they are straight lines passing through the origin, as theoretically expected. The first order constants K can be obtained from the slopes and the true second order rate constants are obtained by dividing K by the oxygen concentration. These data are compiled in Table 2. The true second order rate constant is independent of oxygen pressure, as expected and illustrated by the results obtained at 120 °C for 0.02 and 0.2 MPa oxygen pressures. At 80 °C, (k_{O_2}) = 9.4×10^{-6} l mol⁻¹ s⁻¹ while, at the same temperature but in different experimental conditions, it was found $k_{O_2} = 3.5 \times 10^{-6}$ l mol⁻¹ s⁻¹ for Irganox 1010, a stabiliser belonging to the same family [4]. These values can be considered relatively close owing to the diversity of error sources. These authors [4] obtained this value at 80 °C – 5.0 MPa and using infrared technique to measure the antioxidant concentration.

The values of k_{O_2} were put in an Arrhenius plot together with Irganox 1010 one in Fig. 9. The points are not very far from a straight-line of which the parameters allow us to determine the pre-exponential factor $k_{O_2,0} \approx 1.9 \times 10^9$ l mol⁻¹ s⁻¹ and the activation energy $E_{act} \approx 98$ kJ mol⁻¹.

Table 2

Kinetic parameters corresponding to the direct reaction of the AO4 antioxidant with oxygen.

T – P _{Oxygen}	80 °C	90 °C	100 °C	110 °C	120 °C	120 °C
	3.0 MPa	3.0 MPa	3.0 MPa	3.0 MPa	0.02 MPa	0.2 MPa
$K (s^{-1}) \times 10^6$	1.43 ± 0.01	1.55 ± 0.27	5.16 ± 0.11	11.32 ± 1.61	0.20 ± 0.03	2.32 ± 0.15
$k_{O_2} (l \text{ mol}^{-1} s^{-1}) \times 10^5$	0.94 ± 0.01	1.04 ± 0.18	3.50 ± 0.07	7.79 ± 1.11	21.24 ± 3.78	24.28 ± 1.57

**Fig. 9.** Arrhenius plot of true second order rate constant (k_{O_2}).

3.4. Identification of oxidation products

In order to complete the identification of oxidation products, we have performed GPC analysis with UV detection at 276, 311 and 364 nm for the exposure at 110 °C under 3.0 MPa oxygen pressure (Fig. 10). These results can be briefly resumed as follows. There are three distinct species or groups of species. H in the high molar mass range with a band centred at 20.6 min, i.e. $M_w \sim 673 \text{ g mol}^{-1}$. These species have an UV spectrum covering the three wavelengths investigated: 276, 311 and 364 nm. In other words, they contain presumably the three major kinds of chromophores put in evidence by UV spectra. M at molar masses close to the initial stabiliser one, absorbing at 276 nm as AO4 but absorbing also at 311 nm where AO4 does not absorb or absorbs slightly. L at molar masses lower than the initial stabiliser one, having no absorption at 311 nm but absorbing at 276 and 364 nm. Two components L_1 and L_2 can be in fact distinguished at low molar masses. L_1 absorbs mainly at 276 nm, more slightly at 364 nm, and is eluted at 22.9 min ($\sim 224 \text{ g mol}^{-1}$). L_2 does not absorb at 276 nm but absorbs at 364 nm and is eluted at c.a. 23.5 min, i.e. at a molar mass lower than L_1 ($\sim 168 \text{ g mol}^{-1}$). Products

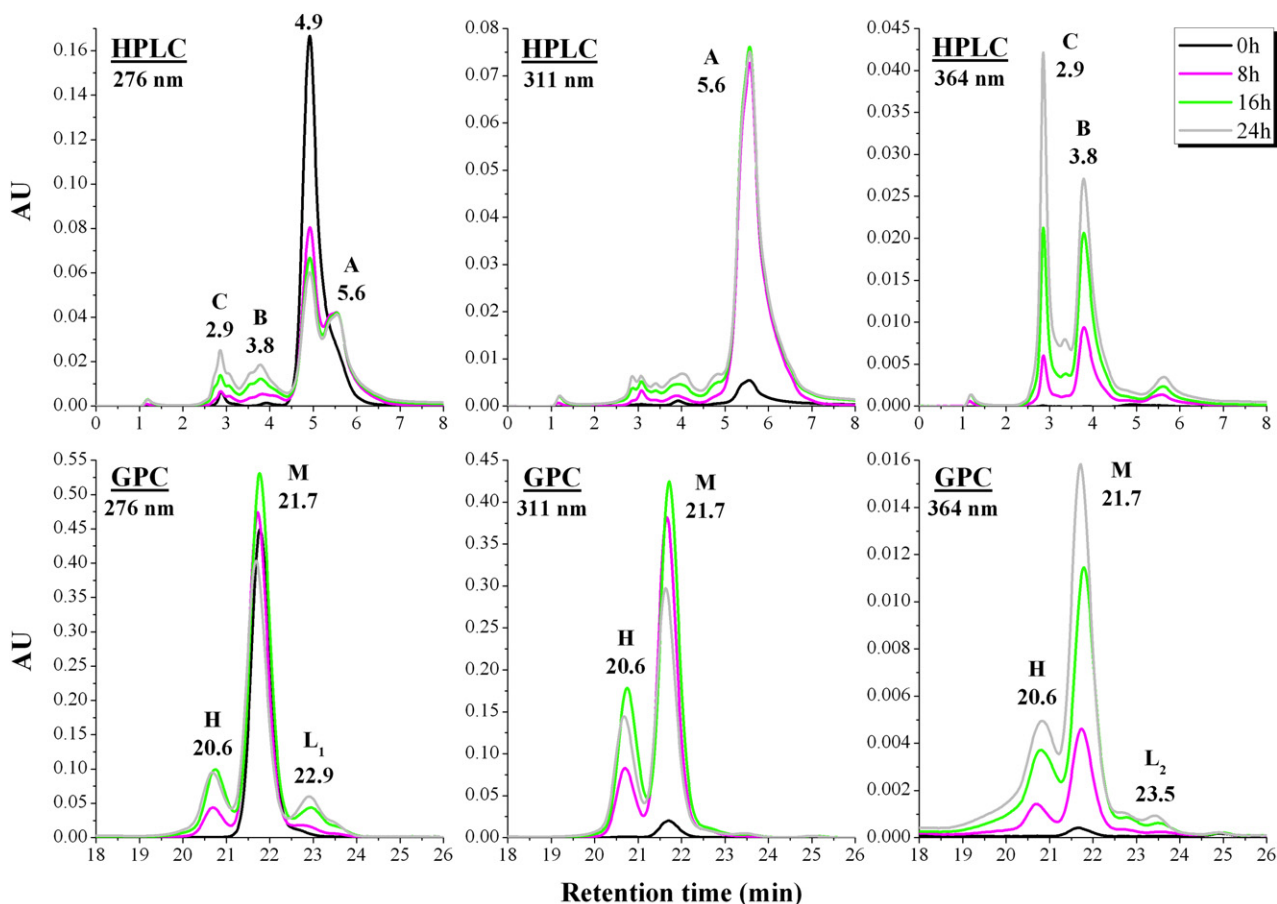
**Fig. 10.** Comparison of HPLC and GPC techniques applied to the aging process of the AO4 stabiliser at 110 °C under 3.0 MPa oxygen pressure.

Table 3
Summary of HPLC and GPC results.

Compound	RT in HPLC (min)	Absorbance			M _w (g mol ⁻¹)	Possible attribution
		276 nm	311 nm	364 nm		
AO4	4.9	✓			390.6	AO4
A	5.6	✓	✓		~398 (M) ~673 (H)	Ci Dimer
B	3.8	✓		✓	~168 (L ₂) ~224 (L ₁) ~398 (M) ~673 (H)	BQ, QM Ci Dimer
C	2.9	✓		✓	~168 (L ₂) ~224 (L ₁) ~398 (M) ~673 (H)	BQ, QM Ci Dimer
D	1.2	✓	✓	✓		

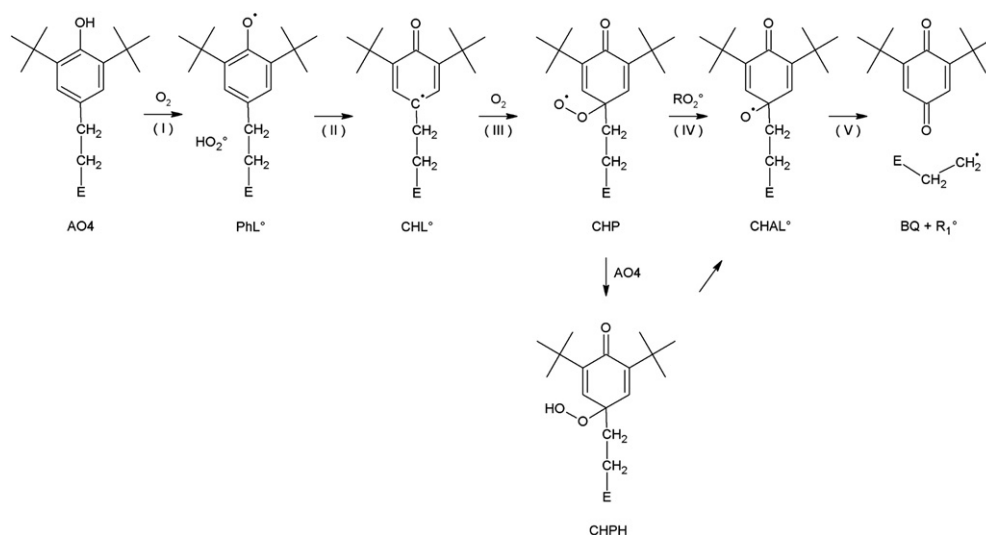
H result obviously from an oligomerisation, the predominant species being dimers. Products M are transformation products of AO4 but only with small mass molar changes. Products L result presumably from the split-off of the aliphatic chain but, since they absorb UV, are constituted of the aromatic part or its transformation products. Table 3 shows a summary of results obtaining by both techniques and the possible attribution of transformation products found.

According to the literature [15,16], the species L of low molar mass would be the benzoquinone (BQ) resulting presumably from the sequence of reactions shown in Scheme 1.

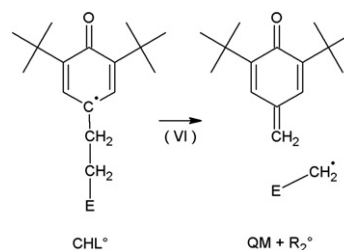
One cannot exclude an eventual rearrangement of the cyclohexadienone radical CHL• by β scission leading to a quinone methide (QM) (Scheme 2).

Whatever the nature of low molecular weight species (L), it must involve radical formation. Process IV–V of peroxide decomposition is a “reinitiation” generating two highly reactive radicals RO• and R₁•. Process VI generates also a highly reactive radical R₂•. Both R₁• and R₂• radicals can react with O₂ generating RO₂• radicals. All these processes tend to decrease the whole stabiliser efficiency.

The species having a molar mass very close to the initial stabiliser one (~398 g mol⁻¹) but with an UV absorption band bathochromically shifted from 276 to 311 nm could be a cinnamate



Scheme 1. Presumed pathway for benzoquinone formation (E = alkyl ester).



Scheme 2. Hypothetic mechanism for quinone methide formation.

(Ci) in which the hindered phenol function is restored [16]. There is no reason to suppose that this new phenol is not reactive towards oxygen (or peroxy radicals). Thus if cinnamate formation was an important secondary product of stabilisation processes, phenols belonging to the family of AO4, i.e. having a dimethylene unit in *para* position of the hydroxyl group, are expected to have a higher stabilising efficiency than the other phenols which do not benefit of this “recycling” of the phenolic group. The difference remains, however, to be experimentally checked.

Concerning now high molar mass species (H), precise analytical studies have been previously published [11,15,17]. They reveal mainly the formation of various types of dimers and tetramers. Our results do not disagree with these interpretations. The most interesting fact appeared here in Fig. 10, is that the oligomers absorb as well at 276 nm as at 311 or 364 nm. It can be deduced that the oligomers contain as well unreacted phenolic groups as cinnamate and/or more conjugated species.

4. Conclusions

The reaction between oxygen and an hindered phenol of the isooctyl-3-(3,5-di-tert-butyl-4-hydroxyphenyl)propionate (AO4) has been studied in an inert solvent (polydimethylsiloxane oil) at temperatures ranging from 80 to 120 °C. UV/VIS spectrophotometry, HPLC and GPC (with UV detection) have been used to characterize the reaction products.

The kinetic study of antioxidant consumption reveals a simple pseudo first order reaction whose rate constant values and activation energy have been determined. Kinetic parameters obtained in this study fit with the values calculated in our previous work [10] and the

ones found by other authors. If the reaction between oxygen and hydrocarbon substrates is negligible at moderate temperatures (<200 °C) and oxygen pressures (<10 MPa), this is not the case for phenolic stabilisers, owing to the very high radical reactivity of the phenol group linked to the low dissociation energy of the O–H bond ($\sim 340 \text{ kJ mol}^{-1}$ against $\sim 390 \text{ kJ mol}^{-1}$ for a saturated hydrocarbon).

We have tried to identify the oxidation products which are expected to be at least partly the same as in polymer stabilisation processes. Three groups of products differing by their average molar mass have been observed. Products of molar mass lower than the starting stabiliser (AO4) result from splitting-off the aliphatic tail of the molecule. The predominant UV absorbing species would be a benzoquinone ($M_w = 220 \text{ g mol}^{-1}$ against $\sim 224 \text{ g mol}^{-1}$ according to GPC) or a quinone methide.

The cinnamate (Ci) parent of the starting stabiliser seems to be formed with a relatively high yield. It results from a sequence of rearrangements only possible in substituted alkyl propionates or phenols having a polymethylene sequence in *para* position relatively to the OH group. This property is interesting because it regenerates a phenol group able, in principle, to participate to a further stabilisation event.

Products of molar mass higher than the antioxidant one were formed at a non negligible yield. They are presumably oligomers with a clear predominance of dimers ($M_w \approx 673 \text{ g mol}^{-1}$ according to GPC).

Reactions leading to low molar mass products are expected to generate new radicals able to initiate oxidation chains. In other words, they are expected to lower the stabiliser efficiency.

In contrast, the reaction responsible for cinnamate formation, which regenerates the phenolic function, is expected to increase the stabiliser efficiency. The balance between all these processes must depend of oxygen pressure and stabiliser concentration. A systematic study of these factors is under progress in our laboratory.

Acknowledgements

Financial support for this investigation has been provided by the Spanish Ministerio de Ciencia e Innovación through the “Ayudas de estancias de movilidad posdoctoral en el extranjero” Program.

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