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Developing Foram-AMBI for biomonitoring in the Mediterranean: Species assignments to ecological categories

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1. Abstract

Most environmental bio-monitoring methods using the species composition of marine faunas define the Ecological Quality Status of soft bottom ecosystems based on the relative proportions of species assigned to a limited number of ecological categories. In this study we analyse the distribution patterns of benthic foraminifera in the Mediterranean as a function of organic carbon gradients on the basis of 15 publications and assign the individual species to five ecological categories. Our categories (of sensitive, indifferent and 3rd, 2nd and 1st order opportunists) are very similar to the ecological categories commonly used for macrofauna, but show some minor differences. In the 15 analyzed publications, we considered the numerical data of 493 taxa, of which 199 could be assigned. In all 79 taxa were classified as sensitive, 60 as indifferent, 46 as 3rd order, 12 as 2nd order and 2 as 1st order opportunists. The remaining 294 taxa are all accessory, and will only marginally contribute to biotic indices based on relative species proportions. In this paper we wanted also to explain the methodology we used for these species assignments, paying particular attention to all complications and problems encountered. We think that the species list proposed here will constitute a highly useful tool for foraminiferal bio-monitoring of soft bottoms in the Mediterranean Sea, which can be used in different ecological indices (Foram-AMBI and similar methods). With additional information coming available in the next few years, it will be possible to expand the list, and, if necessary, to apply some minor corrections. As a next step, we intend to test this species list using several biotic indices, in a number of independent data sets, as soon as these will become available.

2. Introduction

The increasing concern for marine ecosystem health has led to a strong demand for suitable bio-indicator methods, capable of quantitatively assessing the quality of marine habitats and the biotic response to various types of anthropogenic impact. In Europe, this

demand is even stronger because of two decisions of the European Community, which enforce member states to define the Ecological Quality Status (EQS) of their marine water bodies. The European Water Framework Directive (WFD, 2000/60/EC) obliges all countries to achieve a good status of all water bodies, including marine waters up to one nautical mile offshore, by 2015. Similarly, the Marine Strategy Framework Directive (MSFD, 2008/56/EC) aims to obtain Good Environmental Status (GES) for Europe's marine waters by 2020.

Because of these far-reaching decisions, a large number of monitoring tools have been developed. It is important not only to evaluate the physico-chemical characteristics of the concerned water bodies, but also the eventual impact of eutrophication, pollutants and physical disturbance on the living biota. In order to do so, several bio-indicators have been developed. For soft-bottom marine habitats, macrofauna is traditionally used as a bio-indicator of EQS, and a wide range of different biotic indices have been developed (overview in Borja et al., 2016). Among these, many are based on the relative proportions of indicator species (e.g., Word, 1979; Bellan, 1980; Bellan-Santini, 1980; Roberts et al., 1998; Borja et al., 2000; Gomez-Gesteira & Dauvin, 2000; Eaton, 2001; Smith et al., 2001; Simboura & Zenetos, 2002).

Most of the biotic indices using macrofauna are based on changes in faunal composition and/or biodiversity in response to organic enrichment, due to the different ecological strategies of the concerned species. The underlying thought is that although anthropogenic pollution may have many aspects (such as different chemical pollutants), an increased organic load introduced into the marine environment can be considered as a common trait. In more extreme cases, increased oxygen demand may lead to the development of hypoxia at the sediment-water interface. It is implicitly assumed that the faunal response to organic enrichment, eventually accompanied by hypoxia, is representative for most types of pollution.

Therefore, in most macrofaunal indices, macrofaunal taxa are classified as a function of their response to enrichment, either in a rather arbitrary way, or on the basis of more or less elaborated statistical procedures (e.g., Hily, 1984; Glémarec et al., 1986; Borja et al., 2000; Rosenberg et al., 2004; Muxika et al., 2007). In most of these biotic indices, the relative proportions of a number of previously defined ecological groups are used to quantitatively define the EQS. At present, the AZTI Marine Biotic Index (AMBI, Borja et al., 2000) is probably the most commonly used. It is largely based on the early works of Glémarec and Hily (1981), Hily (1984) and Grall and Glémarec (1997), and uses five different ecological groups.

The use of meiofauna, occurring in substantially higher densities, is much less developed (Kennedy & Jacoby, 1999). Among these, benthic foraminifera (BF) appear particularly suitable for bio-monitoring. BF faunas typically contain high numbers of individuals in small areas (typically hundreds to thousands of individuals per 100 cm²), present high species diversity, with various microhabitats and ecological niches being occupied and individual species showing a wide range of reactions to anthropogenic impact. Because of their short life spans (3 months to 2 years; Murray, 1991), they react very rapidly to anthropogenic disturbance, and thus give an integrated picture for a relevant period of time. Most importantly, the shells of most species are preserved in the sediment, thus offering the possibility of reconstructing the historical development of pollution, and obtaining a more precise idea of base-line conditions and faunas.

The international FOraminiferal BioMonitoring (FOBIMO) Group, a consortium of scientists developing the use of foraminifera as bio-indicators, was founded in 2011, with the objective of developing a standardised foraminiferal biomonitoring tool, and making it available to a wider community. As a first step, a standardised protocol was proposed for

sampling and sample treatment (Schönfeld et al., 2012). The next step was to develop a standardised biotic index based on foraminifera.

Since the AMBI-index is widely used for macrofauna, easy to apply, and apparently yields coherent results (e.g., Salas et al., 2004; Muniz et al., 2005; Muxika et al., 2005; Hutton et al., 2015), the FOBIMO-Group decided to investigate the possibility of adapting this index to foraminifera. During the early stages of this process, it appeared that individual species might not show the same response to organic enrichment in different climatologic and oceanographic settings. For this reason, four working groups were created, studying NE Atlantic and Arctic ecosystems, transitional environments, tropical environments and Mediterranean ecosystems, respectively. The working group concerned with NE Atlantic and Arctic ecosystems presented the Foram-AMBI index (Alve et al., 2016), and tested it by comparing the Foram-AMBI scores with organic carbon content (TOC) and Shannon diversity in four independent data sets.

This paper presents the first results of the working group on Mediterranean ecosystems. Because of the particular characteristics of their habitats (high temperature and salinity, overall oligotrophy contrasting with coastal eutrophication), Mediterranean faunas may have different ecological requirements than Atlantic faunas, maybe related to some cryptic endemism. Consequently, there was a consensus that at the first stage, ecological species assignments should be limited to the Mediterranean area, and should be exclusively based on observations made within the Mediterranean. At a later phase, it will be interesting to compare species assignments between the Mediterranean and other basins, in order to investigate whether ecological strategies of individual species are indeed different between basins, and if so, whether these differences are important.

Evidently, the assignment of individual species to various ecological groups is crucial for all biotic indices, which use the relative proportions of these groups to quantitatively define the EQS. In most previous studies, species assignments to ecological groups have been made rather arbitrarily, more or less based on expert knowledge, for macrofauna as well as for foraminifera (e.g., Borja et al., 2000; Dimiza et al., 2016). The aim of the FOBIMO-Group was to base the species assignments on the objective study of a maximal number of suitable data sets, whereby assignments of individual taxa are made as a function of their distribution along well described organic enrichment gradients.

In the paper of Alve et al. (2016), which introduces the Foram-AMBI index, the process of species assignment was done as described in the previous paragraph, but is not described in great detail. Here, we present a list with 223 taxa occurring in the Mediterranean, which we have assigned to five ecological groups, on the basis of a careful study of 15 data sets. We wanted especially to show in detail: 1) how individual species were assigned to the ecological groups, and 2) the complications we encountered, which made this process sometimes particularly difficult. We think that the species list presented here is the best result that can be obtained today, on the basis of an objective study of published data. However, additional data sets will become available in future, and will allow assigning more species, and eventually, apply some moderate corrections. The validation of Foram-AMBI using the species list presented here, and the comparison of the Atlantic and Mediterranean species lists are objectives for further studies.

3. Criteria for the foraminiferal data sets used in this study

The aim of this study was to describe the behaviour of Mediterranean BF taxa in response to various levels of natural and/or anthropogenic organic enrichment. Among the

many published articles dealing with the recent ecology of BF faunas in the Mediterranean, we retained only 15 studies for the assignment of species to ecological categories. Our selection was based on three main criteria: 1) the presence of a gradient in organic carbon content, 2) the nature of the studied samples (living, dead or total faunas), and additionally 3) the availability of grain size data.

Since sediment organic carbon content is probably the most practical descriptor of organic enrichment, we decided to use only data sets in which this parameter had been measured. In fact, sediment organic carbon is also used as environmental reference parameter for biomonitoring methods using macrofauna (e.g., Borja et al., 2003).

Living BF faunas mirror present environmental conditions (e.g., Schönfeld et al., 2012), whereas total faunas (live + dead individuals) tend to give an averaged picture for a (much) longer period (Murray, 1982). For this reason, we decided that our ecological species assessments had to be based as much as possible on benthic foraminiferal biocoenoses. However, in the Mediterranean, living (rose Bengal stained) faunas have only been collected over the last 25 years, and studies of living assemblages are still rare, and often do not include organic carbon data.

Sediment grain size is another important parameter controlling BF distribution in marine environments (e.g., Basso and Spezzaferri, 2000; Celia Magno et al., 2012). It is not necessarily sediment grain size itself that influences BF faunas, but a complex of other factors related to it, such as organic content, pore water oxygen concentration, vegetation, current velocity (e.g., Jorissen, 1987). Unfortunately, grain size has only rarely been quantified in BF ecological studies. Nevertheless, we privileged data sets including this parameter.

Only eight data sets found in the literature on recent Mediterranean foraminifera respected the two main criteria retained for this study: living faunas and organic carbon data. In order to increase the number of data sets, seven supplementary studies were added, in spite of the fact that one of the two conditions was not respected. In fact, the studies of Jorissen (1988), Samir and El-Din (2001), Hyams-Kaphzan et al. (2008), Romano et al. (2009; 2013) and Ferraro et al. (2012) are all based on total faunas. Nevertheless, in view of the size of the respective data sets, and the presence of reliable organic carbon measurements for all stations, we expected that these studies would add essential information about the ecological preferences of many Mediterranean BF species. The same holds for the study of Ernst et al. (2005), which is based on a laboratory experiment.

All fifteen retained studies (Fig. 1, Table 1, Appendix A) concerned open marine shelf environments, some of them in supposedly unpolluted environments, with natural C_{org} gradients, and others in clearly polluted settings. For all studies, only samples with a minimum of 40 specimens have been considered for further analysis. A more detailed description of the 15 retained data sets is added as supplementary material (Appendix A).

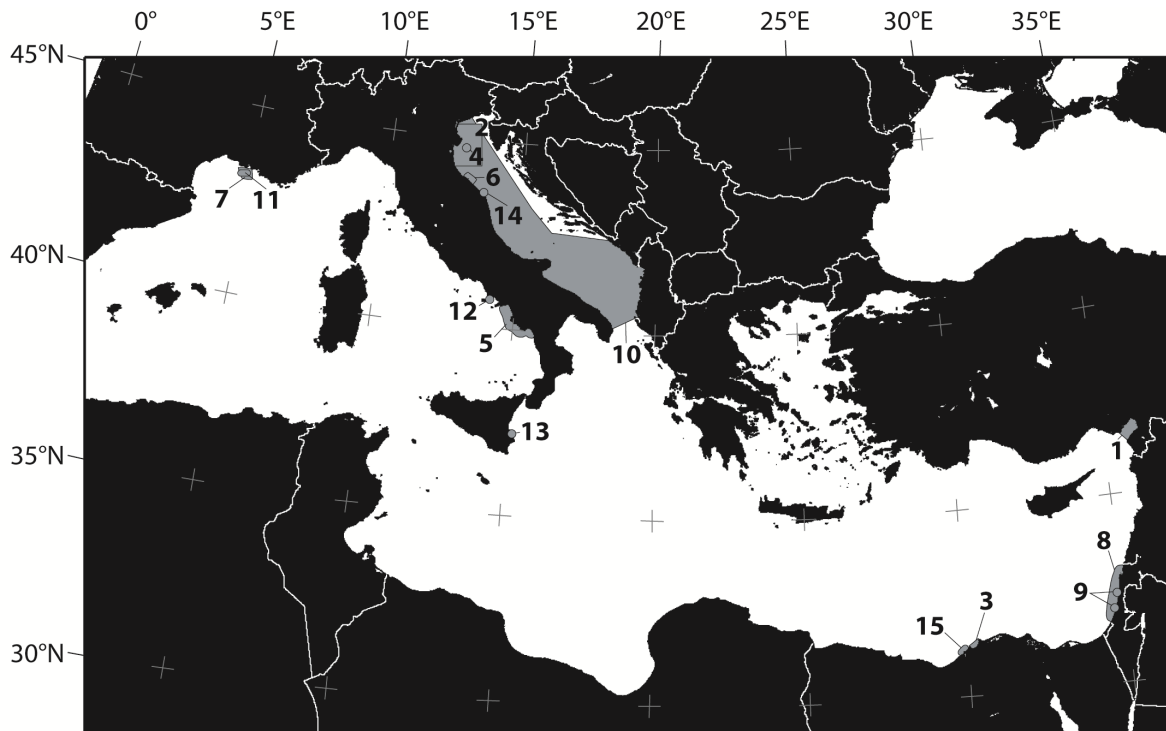


Fig. 1. Map showing the 15 previous studies on Mediterranean BF ecology used in this paper. 1. Basso and Spezzaferri, 2000; 2. Donnici and Serandrei-Barbero, 2002; 3. Elshanawany et al., 2011; 4. Ernst et al., 2005; 5. Ferraro et al., 2012; 6. Frontalini et al., 2011; 7. Goineau et al., 2011; 8. Hyams-Kaphzan et al., 2008; 9. Hyams-Kaphzan et al., 2009; 10. Jorissen, 1988; 11. Mojtahid et al., 2009; 12. Romano et al., 2009; 13. Romano et al., 2013; 14. Sabbatini et al., 2012; 15. Samir and El-Din, 2001.

Careful inspection of Table 1 reveals some major methodological differences between the 15 studies. Probably the most important bias in the data sets is due to different methods to measure organic carbon content; this topic is further discussed in paragraph 5.4. Unfortunately, the large majority of the studies are based on samples taken with Van Veen grabs, which presents the risk of losing part of the more or less liquid superficial sediment (Schönfeld et al., 2012). For the studies based on living foraminifera, different rose Bengal concentrations and

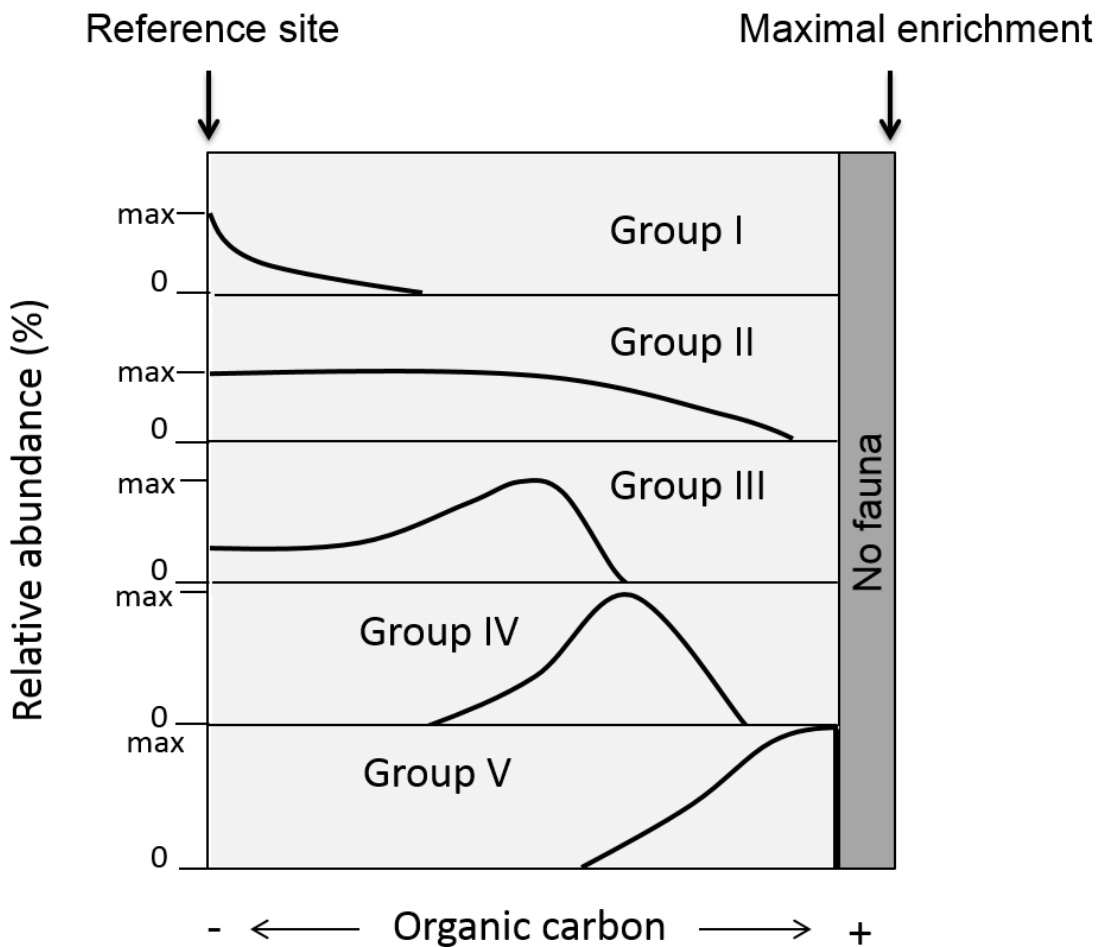
staining periods have been used.

Concerning size fraction, about half of the studies are based on the >63 μm fraction, the others on the >125 μm or >150 μm fraction. We think that in spite of these differences, ecological responses to organic carbon gradients can be perceived in all studies. However, it is evident that opportunistic reactions or sensitivity to increased organic input of small-sized species can only be observed in studies of the >63 μm fraction.

4. Ecological groups

In this study, we have classified the BF taxa in five ecological groups, with different responses to organic enrichment (Fig. 2). A similar subdivision has been used in many previous studies dealing with macrofauna. The five ecological groups traditionally used for macrofauna (sensitive, indifferent, tolerant, 2nd and 1st order opportunists) are largely based on the faunal successions described in the classical study of Pearson and Rosenberg (1978) around the sewage dump site in the Firth of Clyde, and have been summarised by Grall and Glémarec (1997). Borja et al. (2000) implemented them in their widely used AMBI-index.

Initially, the FOBIMO-Group attempted to use the same five ecological categories for BF. However, the ecological patterns revealed in the 19 studies considered by Alve et al. (2016), and the 15 studies presented here, suggested that the definitions of some of the groups were not entirely satisfying for BF. For this reason, slightly modified, more precise descriptions of the five groups were presented by Alve et al. (2016). In order to avoid confusion with the ecological groups described for macrofauna (e.g., Grall and Glémarec, 1997; Borja et al., 2000), we decided to change the name of the third category from “Tolerant” to “3rd order opportunists” (Fig; 2). We think that this new name better describes the distributional pattern of this group. Examples for each of the five groups are presented in Figure 3.



Conceptual graph showing the changes of the cumulative relative abundance of all species belonging to each of the 5 ecological groups along an organic enrichment gradient. Pristine natural conditions are situated on the left, increasingly enriched conditions are found towards the right, until finally azoic conditions are reached on the right of the diagram. See text for further explanation. Modified after Alve et al., 2016.

Group I contains all “**Sensitive species**”. This concerns taxa which are (very) sensitive to organic enrichment, and mainly occur in natural, oligotrophic, unpolluted ecosystems. This group of species is prominent at the reference site(s), where natural conditions are found, characterised by low to moderate organic matter contents. It disappears, or shows a clear decrease (ideally in absolute as well as relative abundance) in case of increasing organic enrichment. The concerned species are absent at strongly enriched sites. This group contains many different taxa, with individual species mostly being present with low relative densities (below 2%). Unfortunately, it is very difficult to observe clear trends for species occurring with such low relative densities, and consequently, many rare species which probably belong to this group, could not be assigned. .

Group II consists of “**Indifferent species**”. These species are indifferent to the first stages of organic enrichment, but disappear in case of (strongly) increased organic supplies. They tend to be present with fairly low relative densities, and do not show a clear trend in absolute and/or relative abundance towards moderately enriched sites.

Group III is composed of “**Third-order opportunists**”. This concerns species which are present at the reference sites, in natural conditions, which are tolerant to the first stages of organic enrichment, and are relatively favoured by such conditions, as is shown by their abundance increase (absolute as well as relative) towards more enriched areas. However, their density maximum is usually fairly distant from the areas of maximum enrichment, where they tend to be absent. The species of this group have previously been labelled as “tolerant” (e.g., Grall and Glémarec, 1997; Alve et al., 2016). However, since the species of Group II are also tolerant to the early stages of ecosystem enrichment, and the species of Group III show a clear opportunistic response to enrichment, we have preferred to name them “third-order opportunists”.

Group IV consists of “**Second-order opportunists**”. These taxa are absent, or occur in low frequency (<2%) at the reference station(s), with natural conditions, and low to moderate organic matter content, and strongly increase towards sites of maximum organic enrichment, with maximum abundance between Groups III and V.

Group V is composed of “**First-order opportunists**”. These species are also absent or rare (<2%) at the reference site(s), their density strongly increases towards the organic enrichment source, but their maximum abundance is situated closer to the site(s) of maximum enrichment than species of the Group IV. These are the last species present before azoic conditions are encountered (Fig. 2). Dense populations of these highly opportunistic taxa can only be observed in strongly eutrophicated areas.

During the analysis of individual data sets, we sometimes felt the need to use intermediate categories. This was for instance the case when we hesitated between two groups, or when we observed a clear opportunistic response, but it was difficult to determine where exactly the concerned station was situated along the overall enrichment gradient. In the first case we used mixed categories (I-II, II-III, etc.), whereas in the second case we indicated only the opportunistic response by assigning a III-IV-V label. For the final assignments (Table 2, Appendix C, right side), for each species all individual assignments (Appendix C, left side; for many species, assignments were available for more than one study) were carefully assessed, and each species was assigned to a single category, so that the intermediate categories disappeared.

Finally, in individual studies, a large number of taxa could not be assigned (NA in Appendix C). This was for instance the case when percentages were very low, when the species was only present in few samples, or showed strongly varying percentages without a clear trend.

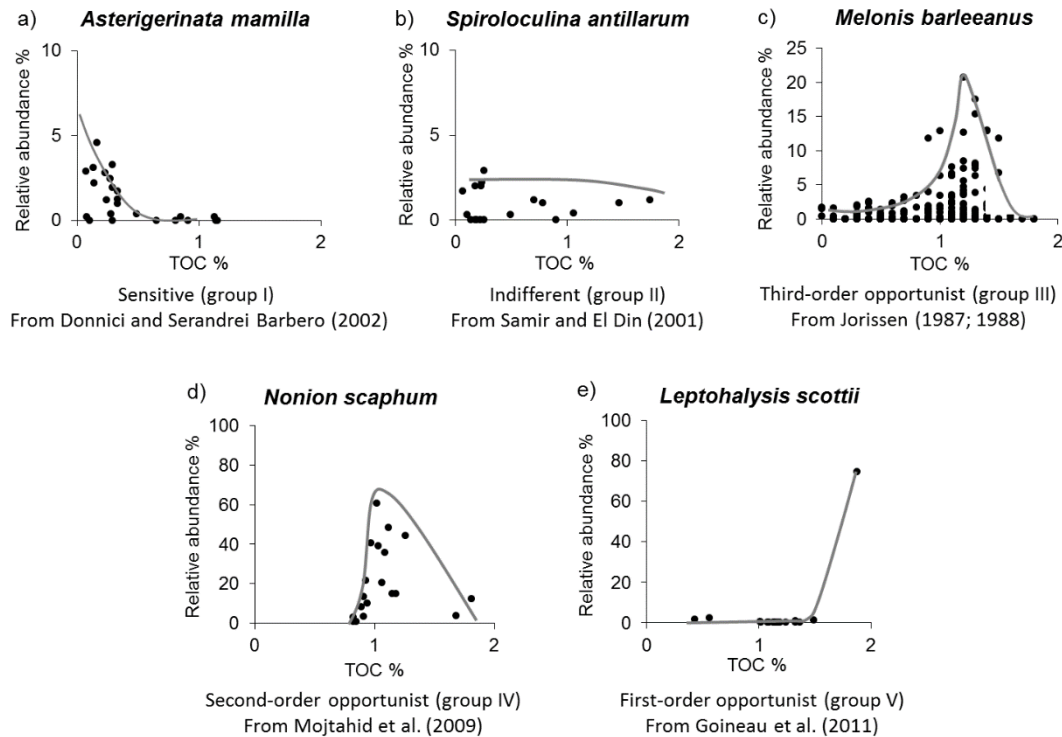


Fig. 3. Examples of species assigned to each of the 5 ecological groups. All data are taken from individual studies, which are indicated below each of the five panels. The schematic enveloping curves indicate the maximum relative frequencies found as a function of %TOC, when all other conditions are optimal.

5. Difficulties encountered in the analyses of the 15 data sets

While analysing the 15 selected data sets, complications of very different nature have been encountered. Some of these issues concern the comparison of sites within a single data set, whereas others concern the comparison of species distribution between different data sets. The following seven subchapters will briefly discuss the main difficulties we encountered.

5.1. Comparing sites with different substrate types

Most foraminiferal taxa have a preference for a particular type of substrate, some species prefer sandy sediments, whereas others prefer a silty to clayey sea floor. In natural coastal settings, sediment grain size tends to be strongly correlated with organic matter because, especially in fine-grained sediments, organic compounds may be adsorbed on the surface of clay minerals (smectite, illite) and within the clay mineral interlayers (e.g., Kennedy et al., 2002). The consequence is that in natural conditions, clayey sediments usually have a higher TOC content than sandy sediments. In natural conditions, BF faunas found on sandy sediments with low TOC contents usually have a high contribution of epiphytic/epifaunal taxa, which are generally considered as pollution-sensitive (e.g., Barras et al., 2014). Conversely, the faunas of clayey substrates often contain high proportions of stress-resistant taxa (e.g., Jorissen, 1987; Celia Magno et al., 2012).

In many nearshore settings, inner littoral sandy sediments present a rather sharp boundary with clayey sediments of a coast-parallel mud belt, which has developed in the Holocene on a global scale (e.g., Van der Zwaan and Jorissen, 1991). Onshore to offshore transects which cross this major biofacial limit will show a sudden increase in TOC, accompanied by an important shift in faunal composition, towards faunas with a higher

percentage of stress-tolerant taxa. Of course, this faunal shift mainly reflects a change in substrate type, and not an increased anthropogenic enrichment. This observation clearly shows that the faunal successions along TOC gradients can only be correctly understood if sediment grain size is taken into account. Preferably, the organic gradient under consideration should not be accompanied by a change in sediment grain size, although this is only rarely the case.

A clear example of this problem is given by *Asterigerina adriatica* in the data set of Donnici and Serandrei Barbero (2002). In fact, in this data set, the transition from sand to clay bottoms coincides with a shift in TOC from values between 0.1 and 0.5% to values between 0.6 and 1.2% (Fig. 4). If we would consider the whole range of TOC values, this species would probably be classified as “indifferent” (Group II), since no clear trend in relative abundance is visible in response to increasing %TOC. However, if we consider only the sandy substrates and no longer take into account changes induced by the abrupt change in sediment grain size at about 0.55% TOC, a clear positive correlation with TOC becomes evident, suggesting that this species is initially favoured by increased TOC values. Consequently, in this study, the species was classified as a 3rd order opportunist (Group III).

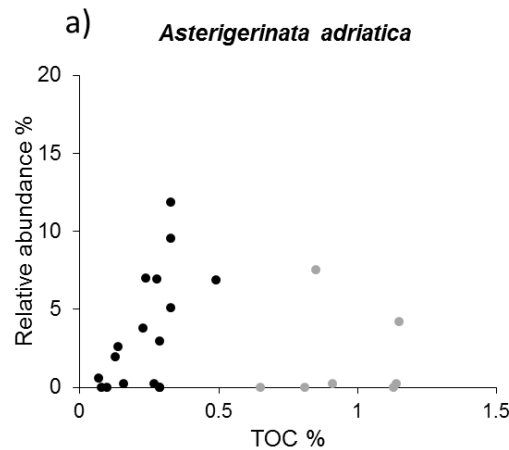


Fig. 4. Relative abundance of *Asterigerinata adriatica* vs. TOC%, data from Donnici and Serandrei Barbero (2002). The diagram includes all the studied samples. Black symbols indicate samples from sandy sediments, grey symbols samples from muddy sediments.

5.2. Taxonomical issues

The taxonomy of BF is complex. First, very different species and genus concepts are used, some based on typological approach, with only a limited amount of morphological variability within the taxon, whereas others admit a much wider morphological range in a single species or genus. Next, various taxonomical schools still exist, which do not give the same taxonomical importance to some of the morphological criteria. For instance, very different taxonomical schemes are used for non-costate buliminids. Finally, some species names are only used regionally, also in cases where the endemic nature of the concerned species has never been shown. The consequence is that in many cases, different species names are in use for what apparently is a single species. In the last decades, molecular studies have partly solved some of these problems (e.g., Hayward et al., 2004; Tsuchiya et al., 2008; Darling et al., 2016), but much taxonomical confusion remains.

In the case of this inventory of Mediterranean foraminifera, we tried as far as possible to avoid taking decisions in case of taxonomical disagreements. In cases where we thought that in the analysed papers, different names were used for the same species, we systematically listed the data of all papers under all species names used, leading to several entries for what is apparently a single species (for instance, *Nonion fabum* and *Nonion scaphum* in Table 2 and

Appendix C). Only in a few cases of evidently wrong determinations, which could only be recognised as such when SEM photos were available, we allowed ourselves to correct the name. However, since many of the analysed papers did not present plates, this was only done in a very limited number of cases. A similar strategy was followed at the genus level; in case of more than one genus name used for the same species, all data were listed under both genus names (for instance, *Bolivina alata* and *Brizalina alata* in Table 2 and Appendix C). Appendix B lists all taxa considered as synonymous (at the genus as well as the species level).

A final difficulty was the fact that in several of the investigated studies, many taxa were listed in open nomenclature, without species name. In this case, it was impossible to compare species patterns between studies. Details for such taxa can be found in Appendix C. In Table 2, we decided not to list such taxa, with three exceptions. *Triloculinella* sp. 1 (data from Hyams-Kaphsam et al., 2009) was added because it was attributed to ecological group IV, which has only few representatives, and because it was the only representative of this genus. *Fursenkoina* sp. 1 and sp. 2 (of the same authors), which were attributed to groups III and I, respectively, were added to show that not all representatives of this genus have a distribution suggesting an opportunistic and/or stress-tolerant character.

5.3. Data on living faunas versus total/dead faunas

Most of the data sets we used are based on living (Rose Bengal stained) faunas. However, five studies based on total faunas (thanatocoenoses) were used as well, because of the very large data sets they contain. Living and total faunas do not give exactly the same information. The living fauna represents a snapshot in a particular environmental context that may vary considerably over short timescales. Conversely, total assemblages give an averaged picture for a much longer period of time (years to centuries), and their composition has been transformed by taphonomical losses (e.g., Murray and Alve, 1999; Denne and Sen Gupta, 1989; Jorissen and Wittling, 1999). This concerns especially agglutinated taxa, many of which are rapidly decomposed after the death of the organisms (Bizon and Bizon, 1985; Schröder, 1988). However, taphonomical processes may also affect various taxa with calcareous tests, with important interspecific differences (Murray, 1991). In spite of these taphonomic losses, which become more severe towards deeper sediment layers, total faunas still may contain useful ecological information, as is shown by the abundant use of fossil assemblage composition in paleoceanographical studies. As explained before, because of the scarceness of suitable data sets on living foraminifera, we decided to add five studies based on total assemblages in the topmost cm of the sediment. However, in these cases, all samples were critically scrutinised, and those with an indication of important taphonomical changes (bad preservation, reworked specimens, uncommonly high densities, etc.) were removed from the data sets before further examination.

5.4. Different methods for OM content measurements

Marine sediments often contain a mixture of organic carbon from terrestrial and marine sources. Organic matter in aquatic systems is a complex mixture of molecules such as carbohydrates, amino acids, hydrocarbons, fatty acids and phenols, natural macromolecules and colloids, originating from living phytoplankton and other plant material, soil organic matter, faunal remains, sewage and industrial discharge. There are numerous methods to quantify OM content, two of which are most often used: elemental analysis, and the loss on ignition method (e.g., Buchanan, 1984). While the first method (i.e., TOC measurement) is more accurate and

has been widely documented (e.g., Luczak et al., 1997), the loss of weight on ignition method is still largely used in benthic ecology because it is quick and cheap, although it has a larger analytical error, especially in clayey sediments (e.g., Mook and Hoskin, 1982).

Table 1 shows that, generally, TOC determined by elemental analysis yields maximal values of 1 to 5%, whereas in the loss on ignition method, maximum values are above 10%. It appears therefore that there is a strong methodological bias, and consequently, the TOC data of the 15 data sets cannot be compared at face value.

5.5. Abundant vegetation debris leading to high TOC values

Organic carbon compounds are delivered to the marine environment from three main sources: fluvial supply of particulate organic matter, nearshore production of benthic plants and algae, and phytoplankton production. The various organic compounds are more or less labile, and have widely varying nutritional values for benthic organisms. Consequently, several methods have been proposed to describe the bioavailability of organic matter (e.g., Dauwe and Middelburg, 1998; Grémare et al., 2003). It is generally assumed that most BF taxa depend mainly on labile, easily metabolisable organic matter (e.g., De Rijk et al., 2000; Mojtahid et al., 2009). It is evident, that in our data set, the TOC values will present a mixture of various types of organic matter, possibly with very different nutritional values. We suspect that the very high TOC values found in some of the data sets (more than one order of magnitude higher than usual (e.g., Basso and Spezzaferri, 2000) are not only due to a methodological bias (elemental analysis versus loss on ignition, see previous paragraph), but may be caused by the presence of abundant remains of macroalgae and *Posidonia*. This hypothesis is corroborated by the abundant presence of epiphytic species in many of the samples. Such organic remains have probably a low nutritional value for foraminifera, and therefore, in these cases the high TOC values are probably not indicative of a food-enriched environment. Consequently, species found with high percentages in these samples are not favoured by enrichment, but are rather associated with macroalgae, for instance because of their epiphytic lifestyle. It is clear that preferences or absences of species at sites, with high TOC values due to phytal macrodetritus, should not be interpreted as a response to ecosystem eutrophication. All data sets have been scrutinised very carefully for this potential bias.

5.6. Comparing naturally eutrophicated and polluted areas

Another potential problem is the fact that the analysed data sets represent a mixture of studies of natural sites, without evident anthropogenic pollution, and sites from polluted areas, some of them even strongly polluted. In both types of setting, the BF response to varying TOC concentrations is used to characterise the various species, and to attribute them to one of the ecological categories. However, there is a fundamental difference between natural and anthropogenically enriched sites. In most natural ecosystems, organic enrichment is not accompanied by chemical pollution. Therefore, samples from naturally enriched sites basically show a faunal response to organic enrichment (and eventually hypoxia/anoxia) alone. Conversely, at polluted sites, organic supplies are usually accompanied by a more or less wide range of toxic chemical compounds, and the faunas potentially show a response to a multiple stressor context. In many cases of anthropogenic pollution there is a positive correlation between TOC and the concentrations of chemical pollutants (e.g., Romano et al. 2009; 2013 and by Elshanawany et al. 2011), and TOC values can be used as an integrative descriptor of pollution. However, in some studies, the correlation between TOC and other pollutants is much less evident (e.g., Samir and El Din, 2001), and TOC may not be a good descriptor for

ecosystem stress. Such different situations may explain the observed differences in faunal behaviour between sites with comparable TOC values.

5.7. Positioning the data sets on the overall organic enrichment gradient

The final problem we encountered was the position of each of the investigated data sets along the ecological continuum used to define the five ecological groups (Fig. 2), which extends from pristine natural environments to sites which are so heavily enriched, that BF have disappeared. Although natural environments may be enriched in organic matter, especially when they are under strong fluvial influence, eutrophication will not attain the same levels as in sites with strong anthropogenic pollution, such as sewage outlets, drill mud disposal sites or harbours. Since advanced stages of organic enrichment only rarely develop in natural coastal sites, it is highly improbable to find maximal percentages of the opportunistic species of ecological categories IV and V (2nd and 1st order opportunists), or even more extreme azoic conditions. Most cases of opportunistic species responses in natural environments concern type III species (3rd order opportunists). In order to take into account this complicated factor, all 15 data sets were very carefully evaluated, and we attempted to define for each study its range along the overall organic enrichment, and made species assignments accordingly.

6. Constructing the Master Table

Initially, each of the 15 data sets was studied independently by at least two researchers, who, whenever possible, assigned species to the five ecological categories in function of the relationship between their relative frequencies and TOC, eventually using the intermediate categories, as explained in paragraph 4. Next, the results of these first analyses were presented in a plenary session (FOBIMO Meeting, 28-30 October 2014, Angers), where each species assignment has been motivated and discussed, in order to verify that the same criteria had been used in all studies. This resulted in a spreadsheet with all individual species assignments made in the 15 studies (Appendix C, left side).

The next step was to assign each individual species to an ecological category on the basis of a comparison and careful appreciation of the data of all studies in which the species was identified. This resulted in the final “Master Table” (Table 2, Appendix C, right side). In Table 2, only the final assignments are listed, whereas all detailed information is given in Appendix C, for assigned as well as non assigned taxa.

When comparing the results for all 15 data sets, a wide range of different situations were encountered, for instance:

- 1) All individual species assignments (Appendix C, left side) agreed, and the assigned ecological category was retained for the final Master Table. Examples are *Peneroplis planatus*, *Bolivina subaenariensis* and *Quinqueloculina agglutinans*.
- 2) Some species (e.g., *Miliolinella semicostata* or *Textularia conica*) could only be assigned unambiguously in a single study, which was considered sufficient for a final assignment.
- 3) In some cases, the species could not be assigned in any of the 15 studies (because of an absence of a clear pattern, or very low densities), and the final assignment was NA (Not Assigned). This was for instance the case for *Elphidium pulvereum*, *Amphistegina lobifera* and *Pyrgo elongata*. Such species are absent in the list of assigned species (Table 2) but are listed in Appendix C.
- 4) The species could be assigned in several studies, but not in other ones, because the pattern was not clear enough. In such cases the ultimate assignment (Appendix C, right side) was based on the studies in which the species could be assigned. This was for

instance the case for *Haynesina depressula*, *Ammonia beccarii* and *Adelosina mediterraneensis*.

- 5) Other species, such as *Ammonia parkinsoniana*, *Cibicides lobatulus* or *Miliolinella subrotunda*, could be classified in a large number of studies, but often with slightly different results. In such cases, we generally privileged data sets which showed the clearest trends, either decreasing percentages (sensitivity) or increasing percentages toward higher TOC (opportunistic response to enrichment). The most typical cases were:
 - a. The individual assignments of a species varied from sensitive (group I) to indifferent (group II). Since we decided to privilege the clearer trends, we assigned such taxa to group I. Examples are *Planorbulina mediterraneensis*, *Miliolinella subrotunda* and *Cibicides lobatulus*.
 - b. The individual assignments varied from indifferent (group II) to opportunistic (group III). As in the previous case, the more explicit trends were privileged, and such species were generally placed in category III. This was for instance the case for *Bulimina marginata*, *Porosonion granosum* and *Bolivina seminuda*.
- 6) In some rare cases we were faced with contradictive assignments, with a species showing a sensitive behaviour in some studies, and an opportunistic behaviour in other ones. In such cases we generally decided to place the species in category II (indifferent species) or III, in function of the position of the studied data set on the overall enrichment scale (see paragraph 5.7).
- 7) For several genera, some species could, but others could not be assigned, mostly due to low relative densities and/or non-diagnostic patterns. In the case of *Adelosina*, *Miliolinella* and *Rosalina*, nearly all species which could be assigned confidently, and were mostly placed in group I (sensitive species). Since the less common species of these genera together showed also a distribution typical of group I, we supposed that they had similar ecological requirements. Therefore, for these two genera, we added an entry in the Master Table for the genus as a whole (e.g., *Adelosina* spp.). In this way, all rare species belonging to these genera were implicitly assigned as well.
- 8) When in individual studies, an opportunistic behaviour was observed, but we could not decide whether the concerned sites were moderately, strongly or extremely enriched in organic matter, we initially considered these species only as "opportunistic", and assigned them to a lump category "III-IV-V". As usual, the final assignment of such species was made by comparing the assignments of all studies in which the species was found. Once again, in general the clearest responses were privileged, leading to assignments in the highest possible category.

According to the methodologies used and the range of environmental conditions, the observations on species distribution of some studies were considered as slightly more reliable compared to other ones. Reasons for this could be the fact that living (Rose Bengal stained) faunas had been studied, more samples had been considered, or the TOC gradient was larger. In cases of contradictory evidence for a given species, results from studies considered as more reliable were privileged. Finally, in spite of these general rules, in some rare cases it was extremely difficult to reach a decision on the basis of the available data, mostly because of the presence of clearly contradictory information. In such cases, the final assignment was sometimes partly based on expert knowledge, after extensive discussion in the general assembly.

7. The Master Table of ecological assignments

In the 15 investigated studies, 493 taxa (synonyms only counted once) had absolute and relative frequencies high enough to be considered. Of these, 199 could be classified in one of the five ecological categories (Table 2), because they showed at least in one study a clear response to organic enrichment, either in anthropogenic or naturally eutrophicated settings. The remaining 294 taxa occurred mostly with low relative frequencies, so that their contribution to most bio-indication methods based on relative taxon frequencies is probably rather limited.

Of the 199 taxa which could be classified:

- 79 have been placed in ecological group I (sensitive)
- 60 in ecological group II (indifferent),
- 46 in group III (3rd order opportunists)
- 12 in group IV (2nd order opportunists), and
- 2 in group V (1st order opportunists).

Species which were determined under different names in the fifteen investigated studies have been listed two (or more) times in the Master Table, but have only been counted once to obtain the numbers given above. This concerns for instance the 1st order opportunist *Leptohalysis scotti*, which is also listed as “*Reophax scotti*”. Appendix B lists all recognised synonyms occurring in the Master Table. 294 of the 493 considered taxa could not yet be classified, because of inconclusive data. However, more suitable studies will doubtlessly become available in the future, which should ultimately make it possible to classify part of these taxa as well.

When on the basis of the final assignments, the appearances of representatives of the 5 groups are considered in each of the studies (Table 3), it appears that in most cases representatives of all ecological categories are present, including opportunistic Groups III, IV and V (e.g., Basso and Spezzaferri, 2000; Goineau et al., 2011). This suggests that a rather complete ecological gradient has been sampled, from natural, non-enriched to heavily eutrophicated sites. In some studies, only very few representatives of Groups III, IV and V were observed. This concerned the studies of Samir and El Din (2001), Hyams-Kaphzan et al. (2008), Romano et al. (2009) and Elshanawany et al. (2011). These studies apparently did not include heavily organic matter enriched sites. Conversely, the studies of Mojtahid et al. (2009), Frontalini et al. (2011), Sabbatini et al. (2012), and, to a lesser degree, Goineau et al. (2011) contained only small numbers of Group I (and Group II) taxa, suggesting that all investigated stations were enriched in TOC, either naturally or anthropogenically, and that these studies lack the pristine, non-enriched side of the spectrum.

8. Discussion and perspectives

In this paper we present assignments of 199 common Mediterranean BF taxa to 5 ecological categories. The final list of species assignments is the fruit of a very thorough and objective inspection of all existing Mediterranean BF data sets until 2014. We think that this list is an essential tool for all bio-indication methods, which use the relative proportions of stress-tolerant and/or stress-sensitive taxa to obtain a quantified measure of the EQS, either by Foram-AMBI (Alve et al., 2016) or by a comparable method (e.g., Barras et al., 2014; Dimiza et al., 2016).

For macrofauna, very similar lists have been constructed. However, with the exception of the list used to define the Benthic Quality Index (BQI, Rosenberg et al., 2004) which is defined in a very objective way, the way most other species lists have been constructed is not

very transparent. In view of the decisive importance of the species assignments to ecological categories, it appeared essential to us to present our list for Mediterranean species, together with a clear overview of how it was constructed, and of the many problems and complications which were encountered while constructing it. We think that it is important that all researchers who use such lists, realise that they are obtained by painstaking literature analyses, and unavoidably contain an element of subjectivity. The present list represents at best our current knowledge of Mediterranean BF ecology. Additional future studies will allow to complete it, and to apply corrections for some taxa, if needed.

As explained before, like most existing lists for macrofauna, our species assignment list is mainly based on the faunal response to organic matter enrichment, which is used as a descriptor for anthropogenic pollution. An important question is whether species which are resistant to, or even favoured by organic enrichment, are also tolerant to other pollutants. Since in marine ecosystems, stress parameters are mostly co-occurring, the answer to this question can probably only come from experimental studies (e.g., Denoyelle et al., 2012; Nardelli et al., 2013).

Finally, after constructing this list, the next step is to test it on independent data sets, containing also TOC measurements, either by using Foram-AMBI (Alve et al., 2016) or similar methods (e.g., Barras et al., 2014; Dimiza et al., 2016). Another objective for further studies is the comparison of ecological assignments of Mediterranean and Atlantic species (Alve et al., 2016). An important question is whether the eventual differences between the two lists are due to different responses to organic enrichment in different climate regimes.

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Table 1: Overview of the 15 studies used in this paper, with an indication of the sampling area and period, water depth range, sampling tool, staining method, studied size fraction, the nature of the studied samples (living, dead or total faunas), the number of samples taken into account for the assignment and the method used to determine OC contents.

Article reference/ (no. reported in Fig. 1)	Location/ Sampling period	Water depth range (m)	Sampling device/Sampled interval	Staining treatment/Mesh sieve	Assemblage	No. of considered samples	TOC method	TOC range variations	Comments
Basso and Spezzaferri, 2000. (1)	Turkey- Iskenderun Bay/ June 1993	7-85	10l Van Veen Grab/0-2cm	4% formalin solution with sea water buffered with 20g of Na ₂ B ₄ O ₇ /l and 2 g Rose Bengal /63µm	Living (Rose Bengal stained)	18	Loss on ignition	2.7-15.8%	
Donnici et al., 2002. (2)	Italy-Northern Adriatic continental shelf/July-August 1996	5-46	Van Veen grab & box corer (total association) & box corer (living association)/0-7 cm	1hour in a Rose Bengal and ethanol solution 5gr 1l/wet sieving 63µm; dry sieving >125µm	Living (Rose Bengal stained) and total	25	Elemental analyser (Mod 2400)	0.07-1.15 %	
Elshanawany et al., 2011. (3)	Egypt-Abu-Qui Bay/May-November, 2005	0.83-8.8	Grab/0-3 cm	Solution Rose Bengal and ethanol/63µm	Living (Rose Bengal stained)	9	Carbon analyser (C-200, version 2.6)	0.1-5.9%	
Ernst et al., 2005. (4)	Italy-Northern Adriatic Sea/February, 2000	32	Van Veen Grab/0-3 cm	NA/500µm	Living	NA	Elemental analyser Fison NA 1500 NCS		Experimental research on living BF in oxic/anoxic conditions
Ferraro et al., 2012. (5)	Italy-Southern Tyrrhenian continental shelf/October-November, 2003	12-195	Van Veen Grab/0-5 cm	NA/>63 and 125 µm	Total assemblage	72	Elemental analyser ThermoElectron Flash EA 1112	0.33-5.73%	Frozen samples
Frontalini et al., 2011. (6)	Italy-Adriatic Sea/coast along Gabice-Pesaro-Fano/May-July, 2002	5-13	15l Van Veen Grab/upper centimetres	solution of Rose Bengal and ethanol (2g/l)/ >63µm	Living (Rose Bengal stained)	6	Loss on ignition	3-13%	Dried at 60°C for 6h

Table 1, continued

Article reference/ (no. reported in Fig. 1)	Location/ Sampling period	Water depth range (m)	Sampling device/Sampled interval	Staining treatment/Mesh sieve	Assemblage	No. of considered samples	TOC method	TOC range variations	Comments
Basso and Spezzaferri, 2000. (1)	Turkey- Iskenderun Bay/ June 1993	7-85	10l Van Veen Grab/0-2cm	4% formalin solution with sea water buffered with 20g of Na ₂ B ₄ O ₇ /l and 2 g Rose Bengal /63µm	Living (Rose Bengal stained)	18	Loss on ignition	2.7-15.8%	
Donnici et al., 2002. (2)	Italy-Northern Adriatic continental shelf/July-August 1996	5-46	Van Veen grab & box corer (total association) & box corer (living association)/0-7 cm	1hour in a Rose Bengal and ethanol solution 5gr 11l/wet sieving 63µm; dry sieving >125µm	Living (Rose Bengal stained) and total	25	Elemental analyser (Mod 2400)	0.07-1.15 %	
Elshanawany et al., 2011. (3)	Egypt-Abu-Qui Bay/May-November, 2005	0.83-8.8	Grab/0-3 cm	Solution Rose Bengal and ethanol/63µm	Living (Rose Bengal stained)	9	Carbon analyser (C-200, version 2.6)	0.1-5.9%	
Ernst et al., 2005. (4)	Italy-Northern Adriatic Sea/February, 2000	32	Van Veen Grab/0-3 cm	NA/500µm	Living	NA	Elemental analyser Fison NA 1500 NCS		Experimental research on living BF in oxic/anoxic conditions
Ferraro et al., 2012. (5)	Italy-Southern Tyrrhenian continental shelf/October-November, 2003	12-195	Van Veen Grab/0-5 cm	NA/>63 and 125 µm	Total assemblage	72	Elemental analyser ThermoElectron Flash EA 1112	0.33-5.73%	Frozen samples
Frontalini et al., 2011. (6)	Italy-Adriatic Sea/coast along Gabice-Pesaro-Fano/May-July, 2002	5-13	15l Van Veen Grab/upper centimetres	solution of Rose Bengal and ethanol (2g/l)/ >63µm	Living (Rose Bengal stained)	6	Loss on ignition	3-13%	Dried at 60°C for 6h

Table 1, continued

Article reference/ (no. reported in Fig. 1)	Location/ Sampling period	Water depth range (m)	Sampling device/Sampled interval	Staining treatment/Mesh sieve	Assemblage	No. of considered samples	TOC method	TOC range variations	Comments
Sabbatini et al., 2012. (14)	Italy-Adriatic Sea (Ancona)/May, July, October, 2008	15	Van Veen Grab/0-1 cm	10% formalin solution with sea water buffered with Na ₂ B ₄ O ₇ /I and 2ml of Rose Bengal/>63 and 150 µm	Living (Rose Bengal stained)	4	Fluorimetry (Chl-a) and spectrophotometry (BCP)	1.6-17.5 µg/g Chl-a; 0.6-1.96 mg/g BCP	BCP= sum of lipids, proteins and carbohydrates (bioavailable C). Samples stored at -20C° until investigated. Only used to classify <i>L. scottii</i>
Samir and El Din, 2001. (15)	Egypt- El Mex and Miami Bays/March, 1994	1-19	Petterson grab/0-2 cm	20% formalin solution and stain with buffered Rose Bengal (1g/l distilled water)/>63µm	Total assemblages	18	NA	0.22-3.15%	TOM published in El-Sabrouti et al. (1997)

Table 2: List of the 256 taxa (including 33 synonyms) which have been assigned to ecological categories. The asterisks in the left column indicate species listed under several names, which are considered synonymous here. These synonyms are listed in Appendix B.

Synonyms	Species	Final assignment
	<i>Adelosina cf. carinatastriata</i>	1
	<i>Acostata mariae</i>	3
	<i>Adelosina cliarensis</i>	1
	<i>Adelosina longirostra</i>	1
	<i>Adelosina mediterraneensis</i>	1
	<i>Adelosina spp.</i>	1
	<i>Adercotryma glomeratum</i>	2
	<i>Affinetrina planciana</i>	2
	<i>Agglutinella compressa</i>	2
	<i>Ammonia beccari f. inflata</i>	2
	<i>Ammonia beccarii</i>	2
	<i>Ammonia compacta</i>	3
*	<i>Ammonia falsobeccarii</i>	3
*	<i>Ammonia inflata</i>	2
*	<i>Ammonia parkinsoniana</i>	1
*	<i>Ammonia perlucida</i>	3
	<i>Ammonia tepida</i>	4
	<i>Ammoscalaria foliaris</i>	2
	<i>Amphicoryna scalaris</i>	2
	<i>Amphistegina radiata</i>	1
*	<i>Articulina mucrunata</i>	1
*	<i>Asterigerinata adriatica</i>	3
	<i>Asterigerinerata mamilla</i>	1
	<i>Astrononion stelligerum</i>	1
	<i>Aubignyna perlucida</i>	3

	<i>Bigenerina nodosaria</i>	2
	<i>Biloculinella labiata</i>	2
	<i>Bolivina aenariensis</i>	2
	<i>Bolivina alata</i>	2
	<i>Bolivina catanensis</i>	1
	<i>Bolivina difformis</i>	2
	<i>Bolivina dilatata</i>	2
	<i>Bolivina dilatata spathulata</i>	1
	<i>Bolivina pseudoplicata</i>	2
	<i>Bolivina seminuda</i>	3
	<i>Bolivina spathulata</i>	3
	<i>Bolivina striatula</i>	3
	<i>Bolivina subaenariensis</i>	2
*	<i>Bolivina variabilis</i>	2
	<i>Brizalina alata</i>	2
	<i>Brizalina difformis</i>	2
	<i>Brizalina laevigata</i>	3
	<i>Brizalina striatula</i>	3
	<i>Buccella frigida granulata</i>	1
	<i>Buccella granulata</i>	1
	<i>Buccella pustulosa</i>	2
	<i>Bulimina aculeata</i>	3
	<i>Bulimina costata</i>	2
*	<i>Bulimina denudata</i>	4
*	<i>Bulimina elongata</i>	3
	<i>Bulimina gibba</i>	3
	<i>Bulimina marginata</i>	3
	<i>Caronia silvestrii</i>	3
*	<i>Cassidulina carinata</i>	4
*	<i>Cassidulina laevigata</i>	4
*	<i>Cassidulina oblonga</i>	2
	<i>Cibicidella variabilis</i>	1

*	<i>Cibicides lobatulus</i>	1
	<i>Cibicides refulgens</i>	1
	<i>Clavulina cylindrica</i>	3
	<i>Cornuspira involvens</i>	3
	<i>Coscinospira hemprichii</i>	1
	<i>Cribroelphidium oceanensis</i>	3
	<i>Cribroelphidium poeyanum</i>	3
	<i>Cycloforina contorta</i>	1
	<i>Cycloforina polygona</i>	2
	<i>Cycloforina quinquecarinata</i>	1
	<i>Discorbinella bertheloti</i>	1
	<i>Discorbis bertheloti</i>	1
	<i>Discorbis mirus</i>	1
	<i>Eggerella scabra</i>	3
	<i>Eggerelloides advenus</i>	3
	<i>Eggerelloides scaber</i>	3
*	<i>Elphidium aculeatum</i>	1
*	<i>Elphidium advenum</i>	2
*	<i>Elphidium complanatum</i>	1
*	<i>Elphidium crispum</i>	1
	<i>Elphidium decipiens</i>	2
	<i>Elphidium depressulum</i>	2
	<i>Elphidium granosum</i>	3
	<i>Elphidium lidoensis</i>	2
	<i>Elphidium macellum</i>	1
	<i>Elphidium poeyanum</i>	3
	<i>Elphidium punctatum</i>	2
	<i>Elphidium striatopunctatum</i>	1
	<i>Elphidium translucens</i>	2
	<i>Epistominella vitrea</i>	4
	<i>Eponides concameratus</i>	1
	<i>Eratidus foliaceus</i>	2

	<i>Fissurina orbignyana caribaea</i>	2
*	<i>Fursenkoina</i> sp. 1 (Hyams-Kaphzan et al. 2009)	3
	<i>Fursenkoina</i> sp. 2 (Hyams-Kaphzan et al. 2009)	1
*	<i>Gavelinopsis praegeri</i>	1
*	<i>Gavelinopsis translucens</i>	1
	<i>Glabratella erecta</i>	1
	<i>Glabratella hexacamerata</i>	1
	<i>Globobulimina affinis</i>	2
	<i>Globocassidulina subglobosa</i>	2
	<i>Globotextularia anceps</i>	1
*	<i>Globulina gibba</i>	1
*	<i>Guttulina lactea</i>	3
	<i>Guttulina problema</i>	2
	<i>Gyroidina umbonata</i>	2
	<i>Haynesina depressula</i>	2
	<i>Haynesina germanica</i>	3
*	<i>Heterostegina depressa</i>	1
*	<i>Hopkinsina pacifica</i>	3
	<i>Hyalinea balthica</i>	2
	<i>Lachlanella planciana</i>	2
	<i>Lachlanella variolata</i>	1
	<i>Laevipeneroplis karreri</i>	2
	<i>Lagenamma atlantica</i>	2
	<i>Lagenamma difflugiformis</i>	2
*	<i>Lagenamma fusiformis</i>	3
	<i>Leptohalysis scottii</i>	5
	<i>Lobatula lobatula</i>	1
	<i>Massilina paronai</i>	2
	<i>Massilina secans</i>	2
	<i>Melonis barleeanus</i>	3
	<i>Miliolinella labiosa</i>	1
	<i>Miliolinella perplexa</i>	3

	<i>Miliolinella semicostata</i>	1
	<i>Miliolinella</i> spp.	1
*	<i>Miliolinella subcircularis</i>	1
	<i>Miliolinella subrotunda</i>	1
*	<i>Morulaepecta bulbosa</i>	3
*	<i>Neoconorbina posidonicola</i>	1
	<i>Neoconorbina terquemi</i>	1
	<i>Nonion depressulum</i>	2
	<i>Nonion fabum</i>	4
	<i>Nonion scaphum</i>	4
*	<i>Nonionella atlantica</i>	3
	<i>Nonionella opima</i>	4
*	<i>Nonionella turgida</i>	5
	<i>Nonionoides grateloupi</i>	2
	<i>Nouria polymorphinoides</i>	3
	<i>Nubecularia lucifuga</i>	1
	<i>Nubeculina divaricata</i>	1
	<i>Pararotalia calcariformata</i>	1
	<i>Pararotalia spinigera</i>	1
	<i>Peneroplis karreri</i>	2
	<i>Peneroplis pertusus</i>	1
	<i>Peneroplis planatus</i>	1
	<i>Planorbulina mediterraneensis</i>	1
	<i>Planorbulina variabilis</i>	1
	<i>Porosononion granosum</i>	3
	<i>Porosononion subgranosum</i>	3
	<i>Pseudoeponides falsobeccarii</i>	3
	<i>Pseudotriloculina brongniartiana</i>	2
*	<i>Pseudotriloculina laevigata</i>	2
*	<i>Pyrgo oblonga</i>	2
	<i>Quinqueloculina agglutinans</i>	1
	<i>Quinqueloculina agglutinata</i>	1

	<i>Quinqueloculina annectens</i>	1
	<i>Quinqueloculina aspera</i>	1
	<i>Quinqueloculina auberiana</i>	1
*	<i>Quinqueloculina badenensis</i>	3
	<i>Quinqueloculina bosciana</i>	2
*	<i>Quinqueloculina bradyana</i>	2
*	<i>Quinqueloculina candeiana</i>	2
	<i>Quinqueloculina contorta</i>	1
*	<i>Quinqueloculina costata</i>	1
	<i>Quinqueloculina disparilis</i>	1
	<i>Quinqueloculina inaequalis</i>	2
*	<i>Quinqueloculina laevigata</i>	1
	<i>Quinqueloculina lata</i>	3
*	<i>Quinqueloculina milletti</i>	1
	<i>Quinqueloculina padana</i>	3
	<i>Quinqueloculina parvula</i>	2
	<i>Quinqueloculina pygmaea</i>	3
	<i>Quinqueloculina seminula</i>	3
*	<i>Quinqueloculina seminula f. longa</i>	4
	<i>Quinqueloculina seminulum</i>	3
	<i>Quinqueloculina seminulum f. longa</i>	4
	<i>Quinqueloculina stelligera</i>	3
*	<i>Quinqueloculina subpolygona</i>	1
*	<i>Quinqueloculina tenuicollis</i>	4
	<i>Quinqueloculina tropicalis</i>	4
*	<i>Quinqueloculina viennensis</i>	2
*	<i>Quinqueloculina vulgaris</i>	1
*	<i>Rectuvigerina phlegeri</i>	3
	<i>Recurvoides trochamminiformis</i>	2
	<i>Reophax fusiformis</i>	3
	<i>Reophax nana</i>	3
	<i>Reophax scorpiurus</i>	2

*	<i>Reophax scotti</i>	5
	<i>Reussella spinulosa</i>	1
	<i>Rosalina bradyi</i>	1
*	<i>Rosalina candeiana</i>	1
	<i>Rosalina floridana</i>	1
	<i>Rosalina globularis</i>	1
*	<i>Rosalina macropora</i>	1
	<i>Rosalina obtusa</i>	2
*	<i>Rosalina</i> spp.	1
*	<i>Sigmoilina costata</i>	1
	<i>Sigmoilina edwarsi</i>	2
*	<i>Sigmoilinita costata</i>	1
	<i>Sigmoilopsis schlumbergeri</i>	3
*	<i>Siphonaperta agglutinans</i>	1
*	<i>Siphonaperta aspera</i>	1
	<i>Sorites orbiculus</i>	1
*	<i>Spirillina vivipara</i>	3
*	<i>Spiroloculina angulata</i>	1
*	<i>Spiroloculina antillarum</i>	2
	<i>Spiroloculina dilatata</i>	2
	<i>Spiroloculina excavata</i>	1
	<i>Spiroloculina nummiformis</i>	1
	<i>Spiroloculina rotundata</i>	1
	<i>Spiroplectammina earlandi</i>	2
	<i>Stainforthia complanata</i>	3
	<i>Stainforthia fusiformis</i>	3
	<i>Textularia agglutinans</i>	3
	<i>Textularia bocki</i>	3
*	<i>Textularia calva</i>	3
	<i>Textularia conica</i>	1
	<i>Textularia earlandi</i>	2
	<i>Textularia truncata</i>	1

	<i>Trifarina angulosa</i>	2
	<i>Triloculina affinis</i>	2
*	<i>Triloculina laevigata</i>	1
*	<i>Triloculina marioni</i>	2
	<i>Triloculina plicata</i>	3
*	<i>Triloculina schreiberiana</i>	3
*	<i>Triloculina tricarinata</i>	1
	<i>Triloculina trigonula</i>	1
	<i>Triloculinella</i> sp. 1 (Hyams-Kaphzan et al. 2009)	4
	<i>Trochammina globigeriniformis</i>	2
	<i>Uvigerina mediterranea</i>	2
*	<i>Uvigerina phlegeri</i>	3
	<i>Valvulineria bradyana</i>	4
*	<i>Vertebralina striata</i>	1

Table 3: Number of taxa finally assigned to each of the 5 ecological categories in each of the 15 studies.

	Basso and Spezzaferri 2000	Donnici and Serandrei 2002	Elshawayany et al. 2011	Ernst et al. 2005	Ferraro et al. 2012	Frontalini et al. 2011	Goineau et al. 2011	Hyams-Kaphzan et al. 2008	Hyams-Kaphzan et al. 2009	Jorissen 87-88	Mojtahid et al. 2009	Romano et al. 2009	Romano et al. 2013	Sabbatini et al. 2012	Samir and El-Din 2001	Total assigned
Group I	25	26	39	0	14	3	10	11	16	10	3	16	9	3	33	79
Group II	12	14	7	0	6	1	23	3	14	13	18	8	4	5	17	60
Group III	15	17	6	6	10	11	18	5	22	12	11	9	12	11	2	46
Group IV	2	4	1	1	4	1	8	1	3	5	5	0	3	2	2	12
Group V	1	0	0	1	0	1	2	0	1	1	2	0	0	1	0	2
Total assigned	55	61	53	8	34	17	61	20	56	41	39	33	28	22	54	199
Not assigned	23	49	5	0	0	2	37	5	37	11	17	1	2	5	16	294
Total no. of species	78	110	58	8	34	19	98	25	93	52	56	34	30	27	70	493

Appendix A. Short description of all retained data sets

The location of the 15 investigated sites is shown in Figure 1 and more detailed information is listed in Table 1. For all studies, only samples with a minimum of 40 specimens have been used.

The first nine studies present data based on living (Rose Bengal stained) assemblages:

The benthic foraminiferal and TOC data, published by Basso and Spezzaferri (2000), concern Iskenderun Bay (Eastern Turkey). Their comprehensive data set of 139 samples (Fig. 1, n° 1) includes TOC gradients related to several pollution sources (e.g., oil pipe terminal, fertiliser industry and cities), measured along a transect from the coast to the centre of the Bay, where pollution is diluted by mixing with open sea-water. Data on grain size are available and its influence on the faunal distribution was critically evaluated. The problem of the data set is that, because of intense pollution, abundances of benthic foraminifera are generally very low and only 18 samples had a minimum of 40 living specimens, and were used here.

Donnici and Serandrei Barbero (2002) investigated faunas from 25 stations in the northern Adriatic Sea, around the Po Delta (Fig. 1, n° 2), which is densely populated and affected by anthropogenic disturbance. Their study considered both living and total faunas, with a total of 500 living specimens counted per sample. Grain size analysis data were also available.

The study of Elshanawany et al. (2011) of Abu-Qir Bay, located on the Egyptian coast near Alexandria (Fig. 1, n° 3), concerns 9 stations, which were sampled in two different seasons (May and November 2005). The region suffers from pollution from different sources, and is considered by the authors as a heavily impacted area. TOC values are presented for all stations, and do not show clear seasonal differences. Two samples were excluded from the data set because we suspected that the very high organic matter content was caused by macroalgae remains, as corroborated by the high relative abundance of epiphytic species. In this study, TOC and metal concentrations are positively correlated.

Frontalini et al. (2011) investigated 6 stations in a shallow basin along the Central Adriatic Sea coast, between Gabicce and Fano (Fig. 1, n° 6). The Tavollo, Foglia and Metauro Rivers flow into the Adriatic Sea here, affecting the marine ecosystem. Macrobenthic communities are poor in the area, supposedly due to pollution and/or anthropogenic pressure. The data set includes grain size measurements, which have been reported in Colantoni et al. (2003). The organic carbon contents in the original datasets were expressed as organic matter (mg/g). As this made it difficult to directly compare the values with other data sets, which all express organic carbon contents as TOC (%), we converted the organic matter values into TOC % according to Brady (1984) and Boyd (1995).

The investigation of Goineau et al. (2011) was conducted in the Rhône prodelta (northwestern Mediterranean Sea); it investigated 20 stations more or less influenced by river outflow (Fig. 1, n° 7). Information about a wide range of environmental parameters is presented, such as pore water chemistry (oxygen, nitrate, nitrite and ammonia), sedimentary particulate organic matter (organic carbon content) and its bioavailability (amino acids, lipid contents, carbon and nitrogen stable isotopes) as well as grain size.

The study of Hyams-Kaphzan et al. (2009) presents data for two stations, one located 200 m north of the Shafdan outfall (activated sewage sludge) near Palmahim and the other located 5.5 km to the Northeast, outside the influence of sludge disposal (Fig. 1, n° 9). The two stations

were sampled bi-monthly for a period of 17 months (January 2003 - May 2004). Samples collected from the polluted station in March, July, September and November 2003 were removed from the data set because they had very low abundances of benthic foraminifera, which is probably due to the development of hypoxia in summer. The dataset includes TOC and chlorophyll *a* data, which showed a strong positive correlation, as well as sediment oxygen penetration that correlates negatively with temperature and grain size. Some species showed different patterns when compared to TOC and chlorophyll. In such cases we privileged to base the ecological assignments on the latter factor, because pigments should better describe the food sources available for foraminifera than TOC.

The investigation of Mojtahid et al. (2009) was also conducted in the Rhône prodelta (Fig. 1, n° 11), the same region as Goineau et al. (2011). The sediments from this region are naturally enriched in organic matter due to river outflow. The benthic foraminiferal faunas at most of the 33 stations are characterised by a fairly low diversity, and contain mainly stress-tolerant taxa. The data set includes data on $\delta^{13}\text{C}$ and C/N-ratios, which were used to evaluate the quality of the organic matter, as well as oxygen concentration at the sea floor and oxygen penetration into the sediments.

The data set from Sabbatini et al. (2012) concerns a single station in Portonovo Bay (Ancona, Adriatic Sea, Fig. 1, n° 14), affected by Po River discharge, and sampled in four seasons (May, July, October 2008 and February 2009). The data set includes biopolymeric C, chlorophyll *a* and phaeopigments analyses. The organic carbon contents in the first three sampling periods were very similar, which precluded the assignment of any species to ecological categories. Conversely, a strongly increased organic matter content was measured in February 2009, when the BF fauna was characterised by a very high absolute and relative abundance of *Leptohalysis scotti*, interpreted as an opportunistic response to strongly increase organic matter input. No other species assignments could be made on the basis of this study, but ultimately, after integration of all 15 studies, 24 of the species observed here could be assigned.

Six other studies were based on total assemblages. Because of the extent of the data sets, and the availability of organic carbon measurements, we decided to use these data as well. They are as follows:

The benthic foraminiferal dataset published in Ferraro et al. (2012), which includes grain size, has been complemented with TOC measures reported earlier (Sprovieri et al., 2006). The study concerns 153 stations from the southern Tyrrhenian Sea, between the Gulfs of Salerno and Policastro (Fig. 1, n° 5). The main focus of the study was to assess the relation between benthic foraminiferal faunas, water depth, substrate, and organic matter. We reduced the original dataset from 153 to 72 stations, because some stations did not have organic matter values, and others had anomalously high TOC values, possibly due to *Posidonia* and/or macroalgae remains.

The dataset of Hyams-Kaphzan et al. (2008) presents data on total benthic foraminifera assemblages from the Israel Mediterranean coast (Fig. 1, n° 8). 57 samples were sampled along 7 transects perpendicular to the coast, between Akhziv in the North and Ashqelon in the South, from 3 to 40 m depth. The study revealed that environmental conditions were in general good, as suggested by presence of large symbiont-bearing benthic foraminifera. The data set includes grain size data as well as TOC, generally with low values (Almogi-Labin et al., 2012), possibly related to the damming of the Nile River that has reduced nutrient input and has induced enhanced oligotrophy.

The extensive data set reported in Jorissen (1988) presents data for 285 stations from the Adriatic Sea (Fig. 1, n° 10), of which 249 were retained, from 7 to 302 m depth. Grain size is available. A problem is the presence of reworked benthic foraminifera (of Pleistocene age) in many samples of the sediment-starved eastern part of the study area. To avoid the bias of reworking, samples with very high number of foraminifera per gram of sediments were removed from the dataset.

Romano et al. (2009) collected 37 samples in front of a dismissed industrial site at Bagnoli, along the highly urbanised coast of the Pozzuoli Gulf (Naples, Italy, Fig. 1, n° 12). Anomalously high concentrations of several heavy metals were found in the sediment, as a consequence of anthropogenic activity. The data set includes grain size analyses, heavy metal concentration, polycyclic aromatic hydrocarbons and TOC. TOC values show a very wide range and are correlated with trace element concentrations.

13) Romano et al. (2013) investigated 37 samples from the Augusta Harbour in Sicily (Fig. 1, n° 13). This site is highly contaminated by harbour and chemical/petrochemical activities. Data set includes grain size, heavy metal, Polycyclic Aromatic Hydrocarbons (PAHs) and Polychlorinated Biphenyls (PCBs) concentrations. Very high contamination degree was recognised in the southern part of the harbour, mainly due to Hg, often associated to very high PCBs levels. Organic matter values show a very wide range and are correlated to anthropogenic contaminant concentrations.

Samir and El-Din (2001) have studied total BF faunas in 18 stations from two bays close to Alexandria, Egypt (Fig. 1, n° 15). El-Mex Bay is affected by pollution with metals, whereas Miami Bay is rather affected by domestic sewage. The environmental data set includes Corg, TOC, pH, bottom water dissolved oxygen, grain size as well as heavy metal concentrations (Cd, Cr, Cu, Pb, Zn).

Finally, a single data set based on an experimental study was included:

15) Ernst et al. (2005 (Fig. 1, n° 4) experimentally assessed the effects of variations in oxygen and labile organic matter flux on BF microhabitats using mesocosms. Different concentrations of a mixture of green algae and diatoms were added to open and sealed (to generate anoxic conditions) microcosms. Oxygen concentrations in the overlying water and in the pore waters, salinity, TOC and Total Organic Nitrogen were measured monthly. The response to organic matter input and oxygen concentration was quantified, with special focus on the migration potential of the selected species. On the basis of the presented data, we could attribute ecological assignments to four of the nine studied species.

Appendix B. List of all taxa (genera as well as species) considered synonymous in these studies. In Table 2, these taxa have systematically been listed under all names that are listed here.

<i>Acostata (cf.) mariae</i>	<i>Reophax nana</i>	
<i>Ammonia beccari f. inflata</i>	<i>Ammonia inflata</i>	
<i>Ammonia perlucida</i>	<i>Aubignyna perlucida</i>	
<i>Bolivina alata</i>	<i>Brizalina alata</i>	
<i>Bolivina difformis</i>	<i>Brizalina difformis</i>	
<i>Bolivina striatula</i>	<i>Brizalina striatula</i>	
<i>Buccella frigida granulata</i>	<i>Buccella granulata</i>	
<i>Caronia silvestrii</i>	<i>Moruloplecta bulbosa</i>	
<i>Cibicidella variabilis</i>	<i>Planorbulina variabilis</i>	
<i>Cibicides lobatulus</i>	<i>Lobatula lobatula</i>	
<i>Criboelphidium poeyanum</i>	<i>Elphidium poeyanum</i>	
<i>Crirostomoides jeffreysii</i>	<i>Labrospira jeffreysii</i>	
<i>Discorbis bertheloti</i>	<i>Discorbinella bertheloti</i>	
<i>Eggerelloides scaber</i>	<i>Eggerella scabra</i>	
<i>Elphidium pauciloculum</i>	<i>Haynesina paucilocula</i>	<i>Nonion pauciloculum</i>
<i>Elphidium depressulum</i>	<i>Haynesina depressula</i>	<i>Nonion depressulum</i>
<i>Lachlanella planciana</i>	<i>Affinetrina planciana</i>	
<i>Laevipeneroplis karreri</i>	<i>Peneroplis karreri</i>	
<i>Lagenammina fusiformis</i>	<i>Reophax fusiformis</i>	
<i>Leptohalysis scottii</i>	<i>Reophax scottii</i>	
<i>Nonion fabum</i>	<i>Nonion scaphum</i>	
<i>Pararotalia spinigera</i>	<i>Pararotalia calcariformata</i>	
<i>Porosonion granosum</i>	<i>Elphidium granosum</i>	
<i>Pseudoeponides falsobeccarii</i>	<i>Ammonia falsobeccarii</i>	
<i>Quinqueloculina agglutinans</i>	<i>Quinqueloculina agglutinata</i>	<i>Siphonaperta agglutinans</i>
<i>Quinqueloculina aspera</i>	<i>Siphonaperta aspera</i>	
<i>Quinqueloculina badenensis</i>	<i>Quinqueloculina padana</i>	
<i>Quinqueloculina seminulum</i>	<i>Quinqueloculina seminula</i>	
<i>Quinqueloculina seminulum f. longa</i>	<i>Quinqueloculina seminula f. longa</i>	
<i>Rectuvigerina phlegeri</i>	<i>Uvigerina phlegeri</i>	
<i>Sigmoilina costata</i>	<i>Sigmoilinita costata</i>	
<i>Spiroplectammina earlandi</i>	<i>Textularia earlandi</i>	

Appendix C, continued

Synonyms	Species	Basso and Spezzaferri 2000	Donnici and Serandrei 2002	Elsharawy et al. 2011	Ernst et al. 2005	Ferraro et al. 2012	Frontalini et al. 2011	Goineau et al. 2011	Hyams-Kaphzan et al. 2008	Hyams-Kaphzan et al. 2009	Jorissen 87-88	Mojtahid et al. 2009	Romano et al. 2009	Romano et al. 2013	Sabbatini et al. 2012	Samir and El-Din 2001	Final assignment	Number of studies
	<i>Bolivina aenariensis</i>													2			2	1
*	<i>Bolivina alata</i>							2				2					2	2
	<i>Bolivina catanensis</i>		NA										2	1			1	2
*	<i>Bolivina difformis</i>												NA			2	2	1
	<i>Bolivina dilatata</i>			2				NA				NA			NA		2	1
	<i>Bolivina dilatata spathulata</i>									1							1	1
	<i>Bolivina pseudoplicata</i>		NA											2			2	1
	<i>Bolivina seminuda</i>						3			2				23			3	3
	<i>Bolivina spathulata</i>		3	23			NA	NA			2	NA			NA		3	3
*	<i>Bolivina striatula</i>	NA	NA	23			3	NA		NA					NA		3	2
	<i>Bolivina subaenariensis</i>							2				2					2	2
	<i>Bolivina variabilis</i>			NA									2	2			2	2
	<i>Bolivina</i> sp. 1									NA							NA	0
	<i>Bolivina</i> sp. 2									NA							NA	0
	<i>Bolivina</i> sp. 3									NA							NA	0
	<i>Bolivina</i> sp. 4									NA							NA	0
	<i>Bolivina</i> sp. 5									NA							NA	0
	<i>Bolivina</i> sp. 6									NA							NA	0
	<i>Bolivina</i> sp. 7			NA													NA	0
*	<i>Brizalina alata</i>							2				2					2	2
*	<i>Brizalina difformis</i>												NA			2	2	1
	<i>Brizalina laevigata</i>		3														3	1
*	<i>Brizalina striatula</i>	NA	NA	23			3										3	2
*	<i>Buccella frigida granulata</i>		1	1		2			1		1						1	5
*	<i>Buccella granulata</i>		1	1		2			1	NA	1		NA				1	5
	<i>Buccella pustulosa</i>		2														2	1
	<i>Buccella</i> sp. 1									NA							NA	0
	<i>Bulimina aculeata</i>	NA				2				NA	3	3	2				3	5
	<i>Bulimina costata</i>							NA			NA	2					2	1
	<i>Bulimina denudata</i>										4						4	1
	<i>Bulimina elongata</i>	3	NA	NA		12	2	NA		345			2	345	NA		3	6
	<i>Bulimina gibba</i>		3					NA		345							3	2
	<i>Bulimina marginata</i>	345	3		NA	2	NA	3		345	NA	23		345	NA		3	7
	<i>Bulimina</i> sp. 1							NA									NA	0
	<i>Buliminella elegantissima</i>		NA				NA			NA					NA		NA	0
	<i>Buliminella</i> sp. 1									NA							NA	0
*	<i>Caronia silvestrii</i>				345					2							3	2
	<i>Cassidulina carinata</i>		NA			2		4		NA	NA	3		NA			4	3
	<i>Cassidulina crassa</i>			NA				NA		NA	NA						NA	0
	<i>Cassidulina laevigata</i>		NA			2		4			NA	3		NA			4	3
	<i>Cassidulina oblonga</i>							NA			2	2					2	2
	<i>Chilostomella ovoidea</i>											NA					NA	0
	<i>Chilostomella oolina</i>							NA									NA	0
*	<i>Cibicidella variabilis</i>		1	1												2	1	3
*	<i>Cibicides lobatulus</i>		1	12		12		NA			1		1	2		2	1	7
	<i>Cibicides refulgens</i>		1	1												12	1	3
	<i>Cibicidina walli</i>	NA															NA	0
	<i>Cibicides</i> sp. 1			1													1	1
	<i>Clavulina cylindrica</i>							345				23					3	2
	<i>Cornuspira foliacea</i>							NA									NA	0
	<i>Cornuspira involvens</i>									NA				3			3	1
	<i>Coccospira henrichii</i>	1															1	1
	<i>Criboelphidium oceanensis</i>						3										3	1
*	<i>Criboelphidium poeyanum</i>					23				NA	23						3	2
	<i>Criboelphidium</i> sp. 1								23								2	1
*	<i>Crirostomoides jeffreysi</i>									NA		NA					NA	0
	<i>Crirostomoides</i> sp. 1							2									2	1
	<i>Crirostomoides subglobosus</i>							NA				NA					NA	0

Appendix C, continued

Synonyms	Species	Basso and Spezzafèri 2000	Donnici and Serandrei 2002	Eikhanawany et al. 2011	Ernst et al. 2005	Ferraro et al. 2012	Frontalini et al. 2011	Goineau et al. 2011	Hyams-Kaphzan et al. 2008	Hyams-Kaphzan et al. 2009	Jorissen 87-88	Mojtahid et al. 2009	Romano et al. 2009	Romano et al. 2013	Sabbatini et al. 2012	Samir and El-Din 2001	Final assignment	Number of studies
	<i>Laevidentalina</i> sp. 1									NA							NA	0
*	<i>Laevipeneroplis karrieri</i>															2	2	1
	<i>Lagena</i> cf. <i>sulcata</i>		NA														NA	0
	<i>Lagena laevis</i>		NA														NA	0
	<i>Lagena oceanica</i>									NA							NA	0
	<i>Lagena striata</i>	NA	NA														NA	0
	<i>Lagena striata strumosa</i>		NA														NA	0
	<i>Lagena strumosa</i>									NA							NA	0
	<i>Lagena substriata</i>									NA							NA	0
	<i>Lagena</i> sp. 1											NA					NA	0
	<i>Lagena</i> sp. 2									NA							NA	0
	<i>Lagena</i> sp. 3									NA							NA	0
	<i>Lagena</i> sp. 4									NA							NA	0
	<i>Lagena</i> sp. 5							NA									NA	0
	<i>Lagenammina atlantica</i>					12				NA							2	1
	<i>Lagenammina difflugiformis</i>		NA				23				2						2	2
*	<i>Lagenammina fusiformis</i>	345	NA				2		NA	NA							3	1
	<i>Lagenammina</i> sp. 1						2										2	1
	<i>Lagenammina</i> sp. 2													NA			NA	0
	<i>Lenticulina atlantica</i>							NA				NA					NA	0
	<i>Lenticulina cultrata</i>	NA															NA	0
	<i>Lenticulina limbosa</i>		NA														NA	0
	<i>Lenticulina peregrina</i>							NA				NA					NA	0
	<i>Lenticulina</i> sp. 1									NA							NA	0
	<i>Lenticulina</i> sp. 2										2						2	1
	<i>Lenticulina</i> sp. 3			NA													NA	0
*	<i>Leptohalysis scottii</i>							345		1	345				345		5	4
*	<i>Lobatula lobatula</i>		1	12		12					1		1	2		2	1	7
	<i>Loxostomina africana</i>									NA							NA	0
	<i>Loxostomina limbata</i>									NA							NA	0
	<i>Marginulina bacheii</i>		NA														NA	0
	<i>Massilina disciformis</i>		NA														NA	0
	<i>Massilina gualtieriana</i>															NA	NA	0
	<i>Massilina paronai</i>															2	2	1
	<i>Massilina secans</i>															2	2	1
	<i>Melonis barleeanus</i>					2		2			3	2					3	4
	<i>Melonis pompilioides</i>	NA															NA	0
	<i>Milammina fusca</i>									NA							NA	0
	<i>Miliolinella labiosa</i>			1													1	1
	<i>Miliolinella perplexa</i>															3	3	1
	<i>Miliolinella semicostata</i>			1													1	1
	<i>Miliolinella subcircularis</i>		1														1	1
	<i>Miliolinella subrotunda</i>	NA	1	1			NA		2			2	2				1	5
	<i>Miliolinella</i> sp. 1								2								2	1
	<i>Miliolinella</i> sp. 2													NA			NA	0
	<i>Miliolinella</i> sp. 3			1													1	1
	<i>Miliolinella</i> sp. 4						NA										NA	0
	<i>Miliolinella</i> sp. 5									NA							NA	0
*	<i>Morulaepecta bulbosa</i>				345					2							3	2
	<i>Neoconorbina posidonicola</i>													12			1	1
	<i>Neoconorbina terquemi</i>		NA	1		1			NA		1						1	3
	<i>Neolenticulina variabilis</i>							NA									NA	0
	<i>Nodophtalmidium</i> sp.									NA							NA	0
	<i>Nodosaria perversa</i>		NA														NA	0
	<i>Nonion commune</i>			NA													NA	0
*	<i>Nonion depressulum</i>	NA	NA			NA					2		2	2		NA	2	3
*	<i>Nonion fabum</i>							4				4					4	2
*	<i>Nonion pauciloculum</i>		NA														NA	0
*	<i>Nonion scaphum</i>							4				4					4	2

Appendix C, continued

Synonyms	Species	Basso and Spezzafèri 2000	Donnici and Serandrei 2002	Elshantawy et al. 2011	Ernst et al. 2005	Ferraro et al. 2012	Frontalini et al. 2011	Goineau et al. 2011	Hyams-Kaphzan et al. 2008	Hyams-Kaphzan et al. 2009	Jorissen 87-88	Mojtabid et al. 2009	Romano et al. 2009	Romano et al. 2013	Sabbatini et al. 2012	Samir and El-Din 2001	Final assignment	Number of studies
	<i>Spiroloculina</i> sp. 1														NA		NA	0
	<i>Spiroloculina</i> sp. 2									NA							NA	0
	<i>Spiroloculina</i> sp. 3							NA									NA	0
	<i>Spiroloculina</i> sp. 4			2													2	1
*	<i>Spiroplectammina earlandi</i>									2					NA		2	1
	<i>Spiroplectammina</i> sp. 1									NA							NA	0
	<i>Stainforthia complanata</i>	3						NA									3	1
	<i>Stainforthia concava</i>											NA					NA	0
	<i>Stainforthia fusiformis</i>		NA				3										3	1
	<i>Stainforthia</i> sp. 1														NA		NA	0
	<i>Subedentostomina lavelansis</i>									NA							NA	0
	<i>Technitella legumen</i>							NA									NA	0
	<i>Technitella melo</i>							NA									NA	0
	<i>Textularia agglutinans</i>		2				3	2	NA	2	2				NA		3	5
	<i>Textularia bocki</i>	3															3	1
	<i>Textularia calva</i>					23											3	1
	<i>Textularia conica</i>		1							NA					NA		1	1
*	<i>Textularia earlandi</i>									2					NA		2	1
	<i>Textularia porecta</i>							NA									NA	0
	<i>Textularia sagittula</i>						NA	NA		NA							NA	0
	<i>Textularia</i> sp. 1							NA									NA	0
	<i>Textularia truncata</i>												1			NA	1	1
	<i>Tretomphalus bulloides</i>									NA							NA	0
	<i>Trichohyalus lacunae</i>		NA														NA	0
	<i>Trifarina angulosa</i>		NA								2					2	2	2
	<i>Triloculina affinis</i>												2				2	1
	<i>Triloculina austriaca</i>														NA	NA	NA	0
	<i>Triloculina laevigata</i>			1													1	1
	<i>Triloculina marioni</i>	2								NA							2	1
	<i>Triloculina plicata</i>	NA											12	23			3	2
	<i>Triloculina schreiberiana</i>								23	NA							3	1
	<i>Triloculina</i> sp. 1														NA		NA	0
	<i>Triloculina</i> sp. 2									NA							NA	0
	<i>Triloculina</i> sp. 3									NA							NA	0
	<i>Triloculina</i> sp. 4									NA							NA	0
	<i>Triloculina</i> sp. 5			1													1	1
	<i>Triloculina tricarinata</i>	NA		1				NA		NA						2	1	2
	<i>Triloculina trigonula</i>	23	1	1				NA			2					2	1	5
	<i>Triloculinella</i> sp. 1									345							4	1
	<i>Trochammina globigeriniformis</i>							23							NA		2	1
	<i>Trochammina inflata</i>		NA														NA	0
	<i>Trochammina</i> sp. 1									2							2	1
	<i>Trochamminula fissuraperta</i>							NA									NA	0
	<i>Trochamminula</i> sp. 1							NA									NA	0
	<i>Uvigerina bifurcata</i>							NA									NA	0
	<i>Uvigerina elongatastriata</i>							NA									NA	0
	<i>Uvigerina mediterranea</i>	NA						NA		NA	2						2	1
	<i>Uvigerina peregrina</i>	NA	NA					NA		NA							NA	0
*	<i>Uvigerina phlegeri</i>	345				23		3			3	12					3	5
	<i>Uvigerina</i> sp. 1									NA							NA	0
	<i>Uvigerina</i> sp. 2							NA									NA	0
	<i>Valvulineria bradyana</i>	NA				23		3			34	34					4	4
	<i>Valvulineria complanata</i>							NA									NA	0
	<i>Varidentella</i> sp. 1									NA							NA	0
	<i>Vertebralina striata</i>	NA	NA	1					NA							2	1	2
	<i>Webbiana</i> sp. 1									NA							NA	0
	<i>Wellmanellina striata</i>															NA	NA	0