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Thrombolytic Therapy Based On Fucoidan-Functionalized Polymer Nanoparticles Targeting P-selectin

Maya Juenet^{1,2}, Rachida Aid-Launais^{1,2,3†}, Bo Li^{1,2†}, Alice Berger^{1,2}, Joël Aerts³, Véronique Ollivier^{1,2}, Antonino Nicoletti^{1,2}, Didier Letourneur^{1,2†} and Cédric Chauvierre^{1,2†*}

¹INSERM, U1148, Laboratory for Vascular Translational Science, X. Bichat Hospital, 46 rue Henri Huchard, 75018, Paris, France

²Paris Diderot University, Paris 13 University, Sorbonne Paris Cité, Paris, France

³FRIM, INSERM UMS 034 Paris Diderot University, X. Bichat Hospital, 75018, Paris France

†Equal contribution

*Corresponding author: cedric.chauvierre@inserm.fr Phone: +(33)140257538

Abstract:

Injection of recombinant tissue plasminogen activator (rt-PA) is the standard drug treatment for thrombolysis. However, rt-PA shows risk of hemorrhages and limited efficiency even at high doses. Polysaccharide-poly(isobutylcyanoacrylate) nanoparticles functionalized with fucoidan and loaded with rt-PA were designed to accumulate on the thrombus. Fucoidan has a nanomolar affinity for the P-selectin expressed by activated platelets in the thrombus. Solid spherical fluorescent nanoparticles with a hydrodynamic diameter of 136 ± 4 nm were synthesized by redox radical emulsion polymerization. The clinical rt-PA formulation was successfully loaded by adsorption on aminated nanoparticles and able to be released *in vitro*. We validated the *in vitro* fibrinolytic activity and binding under flow to both recombinant P-selectin and activated platelet aggregates. The thrombolysis efficiency was demonstrated in a mouse model of venous thrombosis by monitoring the platelet density with intravital microscopy. This study supports the hypothesis that fucoidan-nanoparticles improve the rt-PA efficiency. This work establishes the proof-of-concept of fucoidan-based carriers for targeted thrombolysis.

Keywords:

Nanomedicine; Thrombolysis; Fucoidan; P-selectin; Drug delivery

1 **Introduction**

2 Since the 1990's, intravenous injection of recombinant tissue plasminogen activator (rt-PA) has
3 remained the recommended treatment to induce vessel recanalization in acute thrombotic events
4 such as ischemic stroke [1, 2]. rt-PA is a serine protease that catalyzes the conversion of
5 plasminogen to plasmin. Plasmin degrades the fibrin network of the thrombus thereby promoting
6 thrombolysis. rt-PA preferentially acts by forming a ternary complex in which both rt-PA and
7 plasminogen are bound to fibrin. However, because of its short half-life, high doses of rt-PA
8 need to be injected which generate a cascade of events in the circulation leading to serious side
9 effects. Intracranial hemorrhages are indeed observed in 6% of the treated patients causing 50%
10 of subsequent mortality [3]. Due to the bleeding complications and reported neurotoxic effects,
11 the benefit-to-risk ratio of rt-PA administration rapidly decreases with time and less than 10% of
12 patients end up receiving treatment [4].

13 The most significant improvements in interventional management of stroke include the
14 association of rt-PA with endovascular methods [5]. These strategies have enabled to increase
15 the therapeutic window from 3.5 to 6 hours after the first symptoms onset. However, the use of
16 endovascular devices is limited to “easy-to-reach” thrombi and requires highly trained neuro-
17 interventionist as well as specific clinical facilities [6]. Thus, there is still a dire need for safe and
18 non-invasive treatments. In addition to the development of other fibrinolytic agents [2, 7],
19 nanomedical approaches for local or targeted delivery of thrombolytic drugs arouse a growing
20 interest [8-10]. Lipid and/or PEGylated nanostructures encapsulating a thrombolytic drug were
21 first reported and showed benefit from increasing the drug half-life [11-14]. More recently, this
22 strategy was combined with active targeting approaches, mostly based on magnetic guidance
23 [15-19]. Once accumulated on the thrombus, nanocarriers improve thrombolytic efficiency while
24 reducing hemorrhagic complications, in preclinical models of thrombosis. The use of an external
25 magnet makes these nanostructures hardly scalable yet in a general clinical setting. Another
26 approach to promote the specific accumulation of the drug relies on functionalizing nanocarriers
27 with antibodies and/or peptides directed against the fibrin network [20-22], against the collagen
28 IV of the vascular basement membrane [23] or against the glycoprotein IIb/IIIa (GPIIb/IIIa)
29 expressed by activated platelets [24-26]. Unlike GPIIb/IIIa, P-selectin has been poorly explored
30 with respect to targeted thrombolysis, although this transmembrane glycoprotein is not expressed
31 by resting platelets in the circulation and is extensively expressed by activated platelets localized

1 in the thrombus [27]. In addition, P-selectin is overexpressed by activated endothelial cells,
2 notably in ischemic tissues [28].

3 In this work, P-selectin was chosen as the molecular target. It has been previously demonstrated
4 *in vivo* by us and other groups, that fucoidan-functionalized nanoparticles or microparticles
5 targeting P-selectin can be used for the diagnosis of endothelial activation and intraluminal
6 thrombosis, and for tumor-selective drug delivery [29-31]. Fucoidan refers to a type of highly
7 sulfated polysaccharide containing L-fucose groups [32]. This natural compound exhibits a high
8 affinity for P-selectin, mimicking its main ligand the P-selectin Glycoprotein Ligand 1 (PSGL-1)
9 [33]. We report herein the first nanoparticle preparation combining a thrombolytic drug and
10 fucoidan.

11 Polymer nanoparticles, and especially polysaccharide-based nanoparticles, are promising for the
12 combination of fucoidan with rt-PA [34]. The nanoparticles (NPs) designed in this study were
13 based on polysaccharide-poly(isobutylcyanoacrylate) NPs produced by redox radical emulsion
14 polymerization (RREP). In contrast to the anionic emulsion polymerization, RREP produces
15 stealth NPs with polysaccharides located at the surface in a brush-like structure [35, 36]. The
16 drug can be loaded by adsorption on the polysaccharide shell to promote its direct availability at
17 the thrombus site and the full retention of its activity. To mimic the free primary amines present
18 in the natural fibrin binding sites, or the L-arginine used to stabilize rt-PA in Actilyse[®], dextran
19 modified with amino groups was incorporated into the nanoparticle formulation [37]. After
20 demonstrating here that fucoidan can effectively improve *in vitro* the interaction of fucoidan-
21 functionalized NPs (Fuco-NPs) with P-selectin under flow, we also evidenced the efficiency of
22 rt-PA-Fuco-NPs *in vivo*.

23

24 **Materials and Methods**

25 **Study design**

26 The aim of this study was to test the therapeutic efficacy of targeted NPs containing a
27 thrombolytic drug, rt-PA in its Actilyse[®] form. Animal experiments were conducted in C57BL/6
28 male mice (weight 20-23 g) in respect of the applicable regulation for animal experimentation
29 using protocols approved by the animal care and use committee of the Claude Bernard Institute
30 (APAFIS #8724, Paris, France). The treatment groups included buffer, vehicle control, free rt-
31 PA, non-targeted NPs loaded with rt-PA, and targeted NPs loaded with rt-PA. Unless otherwise

1 indicated, 8 to 10 animals per group were investigated and were randomly assigned to the study
2 groups. The experimentalists were blinded to the identity of the study groups while assaying the
3 multiple endpoints in this study.

4 5 **Products**

6 Isobutylcyanoacrylate (IBCA) monomers were provided by Orapi (Saint Vulbas, France).
7 Several polysaccharides were used: dextran 40 was purchased from PharmaCosmos (Holbaek,
8 Denmark) and carboxymethyl-dextran 40 (carboxyl content of 5% w/w) from TdB Consultancy
9 (Uppsala, Sweden). Medium molecular weight fucoidan (Mn = 18 kDa / Mw = 104 kDa) was a
10 gift from Algues & Mer (Ouessant, France). NPs were mixed with commercially available rt-PA
11 provided by Boehringer Ingelheim (Ingelheim am Rhein, Germany) in its Actilyse[®] form. It was
12 reconstituted according to the manufacturer's recommendations and then aliquoted and stored at
13 -80°C for up to 12 months.

14 15 **Chemical modification of dextran with amino groups**

16 Amino groups were grafted to dextran chains according to a method described by Prigent-
17 Richard *et al.* [38]. Dextran was dissolved at 0.2 g/L in 6.2 M NaOH and cooled at 4°C for 40
18 minutes and 766 mg of 3-Bromopropylamine hydrobromide (Sigma-Aldrich, Saint-Quentin
19 Fallavier, France) was added, which corresponded to an equivalent of 0.5 molecules per glucose
20 unit. The solution was left to react 4 hours at room temperature under stirring and then
21 neutralized. Modified dextran chains were precipitated in ethanol, solubilized in distilled water
22 and extensively dialyzed against water using a Biotech CE tubing dialysis tubing (MWCO 1,000
23 Da, Spectrum Europe B.V., Breda, The Netherlands). The polysaccharide was finally freeze-
24 dried before use. The amine content was determined by isotope-ratio mass spectrometry of
25 nitrogen (IRMS Nu Horizon, Nu instruments). The N content of the aminated dextran was
26 determined to be 0.4 w/w.

27 28 **Polysaccharide-poly(isobutylcyanoacrylate) nanoparticle synthesis**

29 137.5 mg of polysaccharides were dissolved in 8 mL of a nitric acid solution (0.2 M) and kept 10
30 minutes at 40°C under nitrogen bubbling and magnetic stirring. To synthesize fucoidan-
31 functionalized NPs (Fuco-NPs), polysaccharides were introduced in the following proportions:

1 10% w/w fucoidan, 40% w/w amino-dextran and 50% w/w dextran. For Control-NPs, the
2 anionic fucoidan was replaced by anionic carboxymethylated dextran. Ammonium nitrate cerium
3 (IV) ions at 0.08 M in nitric acid 0.2 M were added to the polysaccharide solution, which
4 initiated the formation of radicals. 500 μ L of IBCA monomers were consequently added,
5 according to the method described by Chauvierre *et al.* [35]. Nile Red, a hydrophobic fluorescent
6 dye (Sigma Aldrich, Saint Quentin Fallavier, France), dissolved in acetone at 0.5 mg/mL was
7 incorporated in the medium just after the IBCA to label the hydrophobic core based on the work
8 of Lira *et al.* [39]. To synthesize similar non-fluorescent NPs, acetone alone was added. The
9 medium was left to react for 50 minutes. 1.25 mL of a trisodium citrate dihydrate solution at 1.02
10 M was added dropwise to inhibit the reaction and the medium was neutralized by NaOH 1 M.
11 The final suspension was centrifuged at 2,500 g to remove any large aggregates coming from the
12 block polymerization of IBCA and further dialyzed against water (MWCO 100 kDa). In the rest
13 of the study, the term “unloaded NPs” refers to either fluorescent or non-fluorescent Fuco-NPs
14 and Control-NPs that were not loaded with rt-PA.

15

16 **Physical nanoparticle characterization**

17 Size and Zeta potential were measured by dynamic light scattering (DLS) and electrophoretic
18 light scattering (ELS) respectively (Zetasizer NanoZS, Malvern Instruments SARL, Orsay,
19 France). Samples were dissolved in distilled water and KCl 1 mM for size and zeta potential
20 determination respectively. Particles morphology was visualized by Scanning Electron
21 Microscopy (SEM) (Philips XL 30 ESEM-FEG, Amsterdam, The Netherlands). For all
22 experiments described in this study, NPs were normalized by mass concentration determined
23 after freeze-drying.

24 Finally, the fucoidan content was determined by a semi-quantitative solid-phase colorimetric
25 assay described by Lee *et al.* [40]. 4 μ L of NPs in suspension at a concentration of 20 mg/mL
26 were dropped off a 1 x 1 cm² piece of Whatman Chromatography paper grade 1. This was
27 repeated 6 times on the same point. The paper was left to dry between each drop. The paper was
28 first soaked into a methanol/acetone (6:4) solution for 2 minutes and then into a
29 methanol/acetone/water (6:4:15) solution with 50 mM HCl and 0.1% w/w methylene blue for 10
30 minutes. Finally, the paper was extensively washed with acetic acid/methanol/acetone/water
31 (5:6:4:75) until no coloration was seen in the washing solution. The dye adsorbed on the paper

1 was extracted with Sodium Dodecyl Sulfate at 2% w/v in methanol for 15 minutes at 50°C and
2 its concentration was determined by absorbance reading at 663 nm on an Infinite[®] 200 PRO
3 microplate reader (TECAN Group Ltd., Männedorf, Switzerland). Standard curves were obtained
4 from fucoidan in solution with known concentrations.

6 **Nanoparticle radiolabeling for biodistribution study**

7 Fuco-NPs were radiolabeled with ^{99m}Tc to assess their biodistribution. 200 MBq of sodium
8 pertechnetate [^{99m}Tc]Na was mixed with 2.8 µg of stannous chloride and 500 µg of Fuco-NPs in
9 NaCl 0.9%. Stannous chloride is required to reduce pertechnetate and enable an interaction of the
10 reduced radioactive species with the polysaccharide shell of the nanoparticles [41]. The
11 suspension was incubated for 7 minutes at room temperature. Instant thin-layer chromatography
12 (ITLC) was performed to control that the amount of free ^{99m}Tc was ≤ 5%. 2 µL of the suspension
13 were dropped off a silica gel layer (ITLC-SG Varian) and migration was allowed to occur with
14 methyl-ethyl-ketone (minimum 7 cm). Thus, no supplementary purification step was performed
15 to eliminate free pertechnetate from radiolabeled Fuco-NPs before injection. In parallel, 1 mL of
16 the radiolabeled nanoparticles was centrifuged at 6,400 g for 5 minutes to remove any
17 technetium aggregates. The activities of the supernatant (containing the radiolabeled NPs) and of
18 the pellet (containing large aggregates) were measured with an ionization chamber (Medi 40,
19 Medisystem, Guyancourt, France).

21 ***In vivo* SPECT/CT imaging**

22 The animal study was performed in respect of the applicable regulation for animal
23 experimentation and with approval of the animal care and use committee of the Claude Bernard
24 Institute (Autor. APAFIS #8724, Paris, France).

25 C57BL/6 male mice (weight 20-23 g) (EJ, Le Genest, St-Berthevin, France) were used for *in*
26 *vivo* imaging studies. Mice were kept fully sedated with 1.5-2% isoflurane during the injection
27 step and the SPECT/CT imaging. 200 µL of the radiolabeled suspension (30 MBq)
28 corresponding to 100 µg of nanoparticles was injected through the retro-orbital route. SPECT/CT
29 images were acquired 80 minutes after injection, using a nanoSPECT/CT apparatus (Mediso
30 medical imaging systems, Hungary) with 4 detectors. Ultra-high resolution (0.5 mm) multi-
31 pinhole whole body mouse collimators were used. In addition, 20 angular projections with 50

1 s/projection and a peak energy window of 140 keV \pm 20% were used. Data were reconstructed
2 using the TeraTomo software (Mediso medical imaging systems, Hungary) and a 3D Monte
3 Carlo-based algorithm (64 \times 64 matrix, voxel size of 0.47 \times 0.47 \times 0.47 mm³, 3 subsets and 48
4 iterations). Finally, slides were visualized in 3 planes: sagittal, coronal and axial. A 3D movie
5 was also reconstructed.

6

7 **Tissue biodistribution**

8 Biodistribution of radiolabeled nanoparticles was performed in 3 mice C57Bl/6 (20–23 g,
9 Janvier, CERJ, Laval, France). Mice were anesthetized with intraperitoneal injection of ketamine
10 (Vétoquinol SA, Lure, France) and xylazine (Bayer SAS, La Garenne-Colombes, France) at 100
11 mg/kg and 10 mg/kg respectively. Mice were injected through the retro-orbital route, with an
12 activity of 10 MBq, corresponding to 100 μ g of radiolabeled nanoparticles. Mice were sacrificed
13 on average 80 minutes after injection with an overdose of anesthesia. Liver, spleen, kidneys,
14 heart, lungs, brain, testis, a sample of skin (plus hairs), skeletal muscle, bone (femur), blood and
15 urine, were sampled. All samples, as well as the injected solution, were put in pre-weighed
16 plastic vials. The samples were weighed, and their radioactivity was measured in an automated
17 gamma counter (PerkinElmer 1480 Wizard™3'', Villebon-sur-Yvette, France) collecting 171
18 and 245 keV gamma rays (window width: 135-300 keV). The results were calculated as percent
19 injected dose per gram of organ (% ID/g). It is worth highlighting that after 80 minutes, urine
20 had been partially excreted. Therefore the % ID/g value associated to it is an underestimate of
21 what had been eliminated by this way.

22

23 **Flow binding assay of nanoparticles on recombinant P-selectin**

24 To evaluate the interaction of unloaded NPs with their molecular target, an *in vitro* binding assay
25 under flow was developed [42]. Micro-channels of Vena8 Fluoro⁺ chambers (Cellix Ltd, Dublin,
26 Ireland) were coated overnight with recombinant human P-selectin (R&D systems France, Lille,
27 France) at 50 μ g/mL and rinsed with NaCl 0.9% just before use.

28 NPs diluted at 1 mg/mL in NaCl 0.9% were perfused through channels for 5 minutes either at
29 venous or arterial flow with an ExiGo™ pump (Cellix Ltd, Dublin, Ireland). Venous and arterial
30 conditions corresponded to a shear stress of 6.75 dyne/cm² and of 67.5 dyne/cm², respectively.

31 The binding of NPs was visualized in real time under fluorescence microscopy (Axio Observer,

1 Carl Zeiss Microimaging GmbH, Iena, Germany). The quantitative analysis was limited to NP
2 clusters that formed onto the coated channel because of the resolving power of the fluorescence
3 microscope. After rinsing with NaCl 0.9%, 10 fields per channel (area of 1230 μm x 105 μm)
4 were imaged and analyzed with the image analysis software ImageJ (NIH, Bethesda, U.S.) to
5 quantify the number of fluorescent NPs clusters. Unspecific binding was controlled on uncoated
6 channels and on channels coated with Bovine Serum Albumin at 50 $\mu\text{g}/\text{mL}$.

7 8 **rt-PA loading onto nanoparticles**

9 rt-PA at 0.4 mg/mL was put in contact with NPs at 0.3 mg/mL in a total volume of 500 μL for 2
10 hours at 4°C under gentle agitation in PBS buffer with 0.01% v/v Tween 20. The suspensions
11 were then purified by ultrafiltration at 15,000 g with Vivaspin 500 device (Sartorius France SAS,
12 Dourdan, France). A Molecular Weight Cut Off of 300 kDa was chosen to eliminate free rt-PA
13 (68 kDa). Three cycles of washes were performed with the Tween-PBS buffer.

14 Samples were reconstituted to 500 μL with buffer. The protein content of the purified
15 suspensions was determined by Pierce BCA protein assay (Life Technologies SAS, Courtaboeuf,
16 France). A supplementary step to eliminate any artifact coming from the NPs was applied
17 according to the manufacturer's protocol for eliminating interfering substances. 200 μL of cold
18 acetone (-20°C) was added to 50 μL of the purified samples and incubated for 30 minutes at -
19 20°C to cause protein precipitation. Samples were then centrifuged at 1,000 g for 10 minutes.
20 The supernatant was poured off and the remaining acetone was allowed to evaporate by leaving
21 the samples 30 minutes at room temperature. 50 μL of ultrapure water was added to the protein
22 pellet. 1 mL of the Pierce BCA protein assay working solution was then added and left 30
23 minutes at 37°C. The samples were cooled to room temperature before their absorbance at 562
24 nm was read on an Infinite[®] 200 PRO microplate reader (TECAN Group Ltd., Männedorf,
25 Switzerland). Standard curves were obtained by diluting Actilyse[®] in the Tween-PBS buffer and
26 all steps including protein precipitation were followed in parallel.

27 28 **rt-PA loading efficiency assessed by flow cytometry**

29 The loading efficiency of rt-PA onto Fuco-NPs was assessed by flow cytometry. Briefly, FITC-
30 rt-PA (Abcam) at 0.04 mg/mL was placed in contact with Nile Red Fuco-NPs at 0.03 mg/mL for
31 2 hours at 4°C under gentle agitation in PBS buffer with 0.01% v/v Tween 20. The suspensions

1 were added to a PBS solution and were analyzed with a BD FACSAria™ III flow cytometer
2 (Becton Dickinson). The Nile Red, excited by a 561 nm laser was detected on a PMT set at
3 610/20 nm and the FITC rt-PA, excited by a 480 nm, was detected on a PMT set at 530/30 nm.
4 The analysis was performed with the Diva software (Becton Dickinson).

6 ***In vitro* rt-PA release**

7 The release of rt-PA from the Fuco-NPs was assessed by flow cytometry. Briefly, FITC-rt-PA
8 (Abcam) at 0.04 mg/mL was placed in contact with Nile Red Fuco-NPs at 0.03 mg/mL for 2
9 hours at 4°C under gentle agitation in PBS buffer with 0.01% v/v Tween 20. The suspensions
10 were added to tubes pre-filled with 300 µL of PBS buffer at 37°C and placed under gentle
11 agitation at 37°C during 0, 15, 30, 45, 60 and 90 minutes. At each time point, they were analyzed
12 at 37°C with a BD FACSAria™ III flow cytometer (Becton Dickinson). The Nile Red, excited
13 by a 561 nm laser, was detected on a 610/20 nm PMT while the FITC rt-PA, excited at 480 nm,
14 was detected on a 530/30 nm PMT. The analysis was performed with the Diva software (Becton
15 Dickinson).

17 ***In vitro* amidolytic and fibrinolytic activities of rt-PA-loaded nanoparticles**

18 Amidolytic and fibrinolytic activities of rt-PA were measured just after loading and compared to
19 free rt-PA normalized at the same concentration based on the Pierce BCA protein assay.

20 Amidolytic activity of rt-PA loaded NPs was assessed with the fluorogenic substrate PefaFluor®
21 tPA (Cryopep, Montpellier, France). Samples were reconstituted after purification. 90 µL of
22 purified samples diluted by 1/400 v/v were put in contact with 10 µL of PefaFluor® at 5 mM.
23 After addition of PefaFluor®, a kinetic profile was obtained by measuring the fluorescence level
24 every 30 seconds during 80 minutes at 37°C with Infinite® 200 PRO microplate reader. Increase
25 of fluorescence corresponded to the formation of a fluorescent product coming from the substrate
26 hydrolysis by rt-PA. Enzymatic velocity was determined from the initial slope of the resulting
27 kinetic profile and compared to that of free rt-PA at the same concentration.

28 Fibrinolytic activity was assessed with a fibrin lysis assay adapted from the work of Liang *et al.*
29 [43]. Low melting agarose (Carl Roth GmbH + Co. KG, Karlsruhe, Germany) solution at 3%
30 w/v in TRIS buffer was heated at 65°C. The solution was cooled at 37°C and 2.5 U of thrombin
31 (Sigma-Aldrich) were added. 5 mL of a fibrinogen (Sigma-Aldrich) solution in TRIS buffer were

1 heated at 37°C and added. The reaction mixture was poured into 9 cm culture dish and cooled at
2 4°C for 30 min. On the solidified agar plate, 3 mm wells were created as sample reservoir. 5 µl
3 of reconstituted purified samples diluted by 1/15 v/v were added into the wells and incubated
4 overnight at 37°C in a humid atmosphere. After incubation, the lysis area was quantified with
5 ImageJ and compared to that of free rt-PA at the same concentration.

6

7 **Flow binding assay of nanoparticles on activated platelet aggregates**

8 To further investigate the binding efficiency of unloaded and loaded NPs to activated platelets,
9 channels of Vena8 Fluoro⁺ chambers were coated with 50 µg/mL of fibrillar type I collagen
10 Horm[®] (Takeda, Linz, Austria) overnight and rinsed with NaCl 0.9% before use. Human whole
11 blood (EFS, Hôpital Bichat, Paris, France) collected on 75 µM PPACK (Cryoep, Montpellier,
12 France) was perfused at arterial shear rate for 3.5 to 5 minutes to induce platelet activation and
13 aggregation.

14 In a first set of experiments, the expression of P-selectin by platelet aggregates was controlled by
15 perfusing whole blood containing 20 µg/mL of a FITC-labeled anti-P-selectin antibody or a
16 FITC-labeled matched isotype IgG (Ancell, Bayport, U.S.). After rinsing, 10 fields per channel
17 were imaged and analyzed with the image analysis software ImageJ to compare the fluorescence
18 uptake of aggregates.

19 In a second set of experiments, PPACK-anticoagulated whole human blood was mixed with 5
20 µM of DIOC₆ (Life Technologies SAS, Saint-Aubin, France), a green fluorescent dye that labels
21 mitochondria of live cells, and perfused at arterial shear stress (67.5 dyne/cm²) to allow platelet
22 aggregation. After rinsing, suspensions of Nile Red-NPs at 1 mg/mL were then perfused over
23 preformed platelet aggregates at venous shear stress (6.75 dyne/cm²) for 5 minutes. Their
24 accumulation onto activated aggregates was monitored by fluorescence microscopy in real time.
25 After rinsing, 10 fields per channel (area of 1230 µm x 105 µm) were imaged and analyzed with
26 ImageJ to quantify the level of fluorescent NPs bound to aggregates. Intensity settings were kept
27 the same for both types of NPs. Aggregates were detected using ImageJ by thresholding the
28 green fluorescence level generated by the platelets. The Mean Fluorescence Intensity (MFI) of
29 the NPs on these selected areas was determined using ImageJ. One MFI value was obtained per
30 channel averaged over more than 25 aggregates.

1 In a third set of experiments, rt-PA at 0.4 mg/mL was put in contact with fluorescent NPs at 0.3
2 mg/mL for 2 hours at 4°C under gentle agitation in PBS buffer. The suspensions were then
3 purified with three cycles of ultrafiltration at 15,000 g with Vivaspin 500 device (Sartorius
4 France SAS, Dourdan, France). Finally, loaded NPs were resuspended in PBS just before being
5 perfused at venous shear rate onto activated platelets aggregates marked with DIOC₆. Channels
6 were rinsed after 12 minutes of NPs perfusion and 10 fields per channel were imaged and
7 analyzed with ImageJ. One MFI value was obtained per channel by averaging the NPs
8 fluorescence intensity on several aggregates.

9

10 ***In vivo* model of thrombosis on mouse mesenteric vein**

11 FeCl₃-induced thrombosis model on mouse mesenteric vein was developed based on the work of
12 Bonnard *et al.* [44]. C57BL/6 male mice (EJ, Le Genest, St-Berthevin, France) aged from 5 to 8
13 weeks were chosen for the study. Mice were anesthetized with intraperitoneal injection of
14 ketamine (Vétoquinol SA, Lure, France) and xylazine (Bayer SAS, La Garenne-Colombes,
15 France) at 100 mg/kg and 10 mg/kg respectively. A midline abdominal incision was performed
16 to expose the mesentery. The mesentery was gently laid out over a transparent Petri dish lid and
17 vessels were visualized by intravital microscopy (Leica MacroFluo™, Leica Microsystems SAS,
18 Nanterre Cedex, France) using Orca Flash 4.0 scientific CMOS camera (Hamamatsu Photonics
19 France SARL, Massy, France). Retro-orbital injection of 30 µL of Rhodamine 6G (Sigma-
20 Aldrich) prepared at 0.3% w/v was performed. Rhodamine 6G fluorescently labeled the platelets
21 and the leukocytes.

22 A 1 mm large Whatman chromatography paper soaked into a 10% w/v iron chloride (Sigma-
23 Aldrich) solution prepared in saline was then left 1 minute on one of the mesentery veins. After
24 the paper was removed, the vein was washed twice with a saline solution. The veins were
25 measured to have a mean diameter of 171 ± 31 µm (mean \pm SD, n = 41 mice). The thrombus
26 formation was monitored in real-time by fluorescence microscopy by following the accumulation
27 of fluorescently labeled platelets.

28

29 **Evaluation of *in vivo* thrombolysis by intravital microscopy**

30 46 mice were randomly set into 5 groups for the study receiving buffer, Fuco-NPs, reconstituted
31 Actilyse® at 2.5 mg/kg (rt-PA), Control-NPs associated with Actilyse® normalized to a dose of

1 2.5 mg/kg (rt-PA-Control-NPs), and Fuco-NPs associated with Actilyse[®] normalized to a dose of
2 2.5 mg/kg (rt-PA-Fuco-NPs).

3 Samples were prepared on the day of animal experiments. Non-fluorescent Fuco-NPs and
4 Control-NPs at 0.3 mg/mL in Tween-PBS were mixed with Actilyse[®] at 0.4 mg/mL during 2
5 hours at 4°C under gentle agitation as previously detailed. Suspensions were washed 3 times by
6 ultrafiltration using Vivaspin 500 devices (MWCO 300 kDa, Sartorius France SAS, Dourdan,
7 France). After the third ultrafiltration cycle, concentrated samples were used for injection.
8 Amidolytic activity of the concentrated samples was consequently measured with the fluorogenic
9 substrate PefaFluor[®] tPA and used to normalize the injected dose. Samples were stored on ice
10 and used within 2 hours.

11 Retro-orbital injection of the different treatments and controls was performed once fluorescently
12 labeled platelets started to aggregate in the injured site and formed a visible thrombus, on
13 average 7 ± 3 minutes after thrombosis induction (mean \pm SD, n = 41 mice). This injection
14 method was chosen because it is fast, simple, does not require moving the mouse and ensures a
15 distribution in the circulation similar to that obtained with standard intravenous injection [45].
16 The injected volumes were of 125 ± 36 μ L (mean \pm SD), which has been reported to be adapted
17 to the mouse [46]. The thrombus evolution was monitored in real-time by fluorescence imaging
18 (excitation 545 nm – emission 610 nm). Images were acquired just before injection and regularly
19 during the first 15 minutes after injection, then every 5 minutes up to 30 minutes. The thrombus
20 was detected over time using ImageJ by thresholding the fluorescence level. The thrombus
21 density was defined as the total intensity of the detected area. The thrombus relative density was
22 normalized to the peak platelet accumulation for each animal. This parameter was used to assess
23 the benefit of the different treatments compared to the group receiving buffer. It allowed to
24 alleviate the variability between thrombi and was a better estimate of the global thrombolysis
25 trend.

26 On few additional mice, images were taken continuously over 40 minutes (one image every 10
27 seconds) to provide typical examples of thrombus formation followed by thrombolysis. In these
28 mice, DIOC₆ (Life Technologies SAS, Saint-Aubin, France) was injected at 25 μ M instead of
29 Rhodamine 6G. These movies were taken for the following groups: rt-PA, rt-PA-Control-NPs,
30 and rt-PA-Fuco-NPs. The nanoparticles used were fluorescently labeled in red with Nile Red and

1 dual fluorescence imaging was performed (excitation 480 nm – emission 527 nm / excitation 545
2 nm – emission 610 nm).

3

4 **Statistics**

5 Results are presented as mean \pm standard error of the mean ($n \geq 3$). Statistical significance
6 between experimental groups was assessed using the software GraphPad Prism (GraphPad
7 Software, Inc., La Jolla, U.S.) with 95% confidence level: ns non-significant, * $p < 0.05$;
8 ** $p < 0.01$; *** $p < 0.001$. For all tests, data were assumed to be normally distributed. Unpaired
9 non-parametric test (Mann Whitney) was used for samples that contain ≤ 6 elements. One-way
10 ANOVA with following *Post hoc* Turkey's test was applied for the *in vivo* experiments. Grubbs'
11 test was performed on all the data to identify the significant outliers ($p > 0.01$).

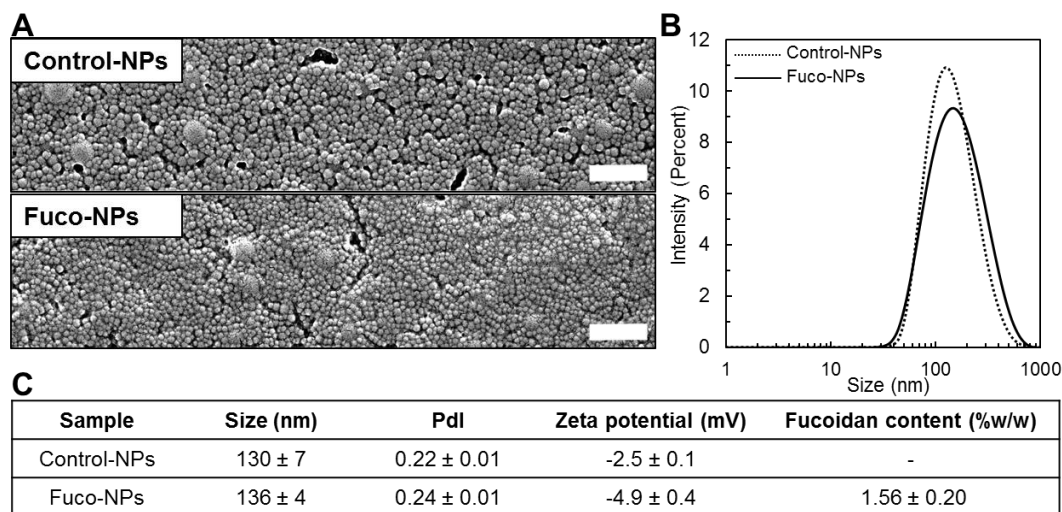
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14

1 Results

2 Nanoparticle synthesis and characterization



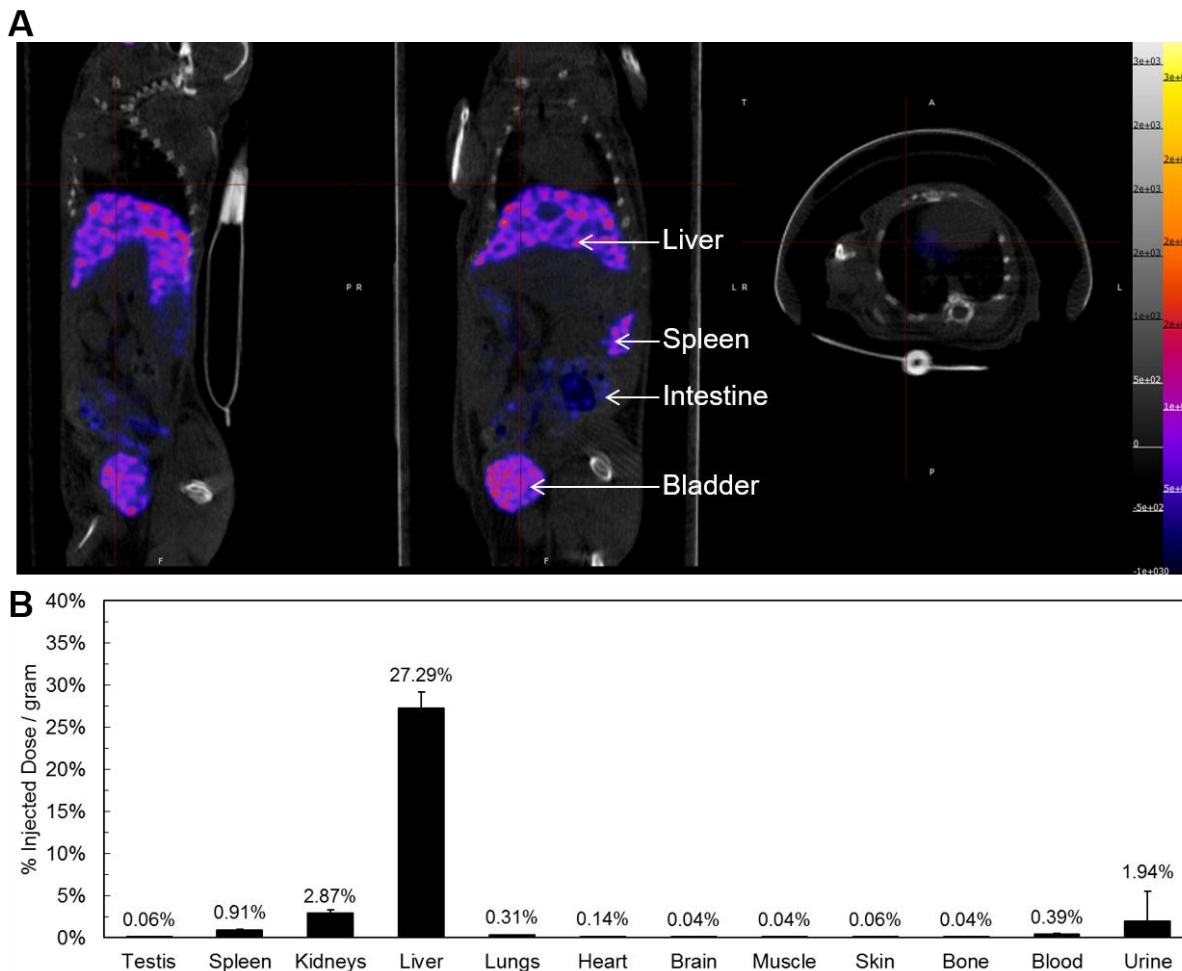
3 **Fig 1. Physico-chemical characterization of Control-NPs and Fuco-NPs.** **A)** Scanning Electron
4 Microscopy (SEM) (scale bar = 500 nm). **B)** Diffusion Light Scattering (DLS) measurement in
5 intensity. **C)** Size values and Polydispersity Index (PdI) determined by DLS and zeta potential values
6 measured by Electrophoretic Light Scattering (n = 5 independent batches per type of nanoparticle;
7 mean ± standard error of the mean). Fucoidan content of Fuco-NPs in mass percent of the total
8 nanoparticle weight determined by a colorimetric assay (n = 3).
9

10
11 NPs made of dextran, aminated dextran and isobutylcyanoacrylate (IBCA) were synthesized by
12 redox radical emulsion polymerization in the presence of either fucoidan (Fuco-NPs) or
13 carboxymethyl-dextran (Control-NPs). Solid spherical NPs were obtained, as shown in SEM
14 images (Figure 1A). The size distribution determined by DLS was similar for both types of NPs,
15 with an average size in intensity of 130 ± 7 nm for Control-NPs and 136 ± 4 nm for Fuco-NPs
16 (Figures 1B, 1C). Samples exhibited similar polydispersity indexes of 0.22 ± 0.01 for Control-
17 NPs and of 0.24 ± 0.01 for Fuco-NPs. ELS measurements provided slightly negative zeta
18 potential values of -2.5 ± 0.1 mV for Control-NPs and of -4.9 ± 0.4 mV for Fuco-NPs (Figure
19 1C) due to the presence of anionic polysaccharides (Fucoidan or carboxymethyl-dextran). The
20 DLS graphs in intensity, number and volume and the ELS graphs corresponding to Figure 1 are
21 reported in supplementary Figure S1. Physical properties of Control-NPs were therefore similar

1 to those of Fuco-NPs. In addition, Fuco-NPs were composed of 1.56 ± 0.20 % of fucoidan in
2 mass percent of the total nanoparticle weight (Figure 1C).

3

4 **Fucoidan-functionalized nanoparticle biodistribution**



5

6 **Fig 2. SPECT/CT imaging and tissue biodistribution in mice determined 80 minutes after**

7 **injection of Fuco-NPs labeled with ^{99m}Tc .** A) Representative whole body SPECT/CT imaging: from

8 left to right: sagittal, coronal and axial planes. B) Tissue biodistribution determined by tissue activity

9 measurement normalized to the tissue weight and expressed as a percentage of the injected dose (n=3

10 mice).

11 Fuco-NPs were radiolabeled with ^{99m}Tc and injected in mice for SPECT/CT imaging and tissue

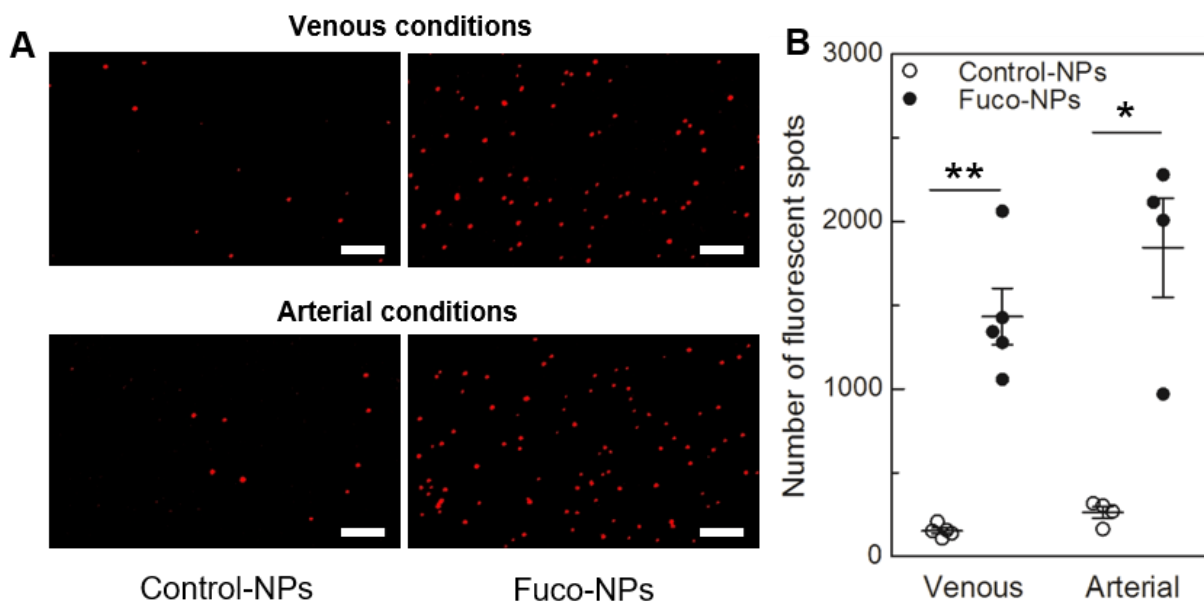
12 biodistribution. SPECT/CT profiles at 80 minutes illustrated in Figure 2 and in supplementary

13 movie S1 showed a classical nanoparticle biodistribution with main accumulation in the bladder

1 and liver. Some activity was also detected in kidneys and in urine and no signal was detected on
2 SPECT/CT images in the stomach wall and in the thyroid corroborating a stable nanoparticle
3 labeling. No significant accumulation was noticed in the lungs, the heart and the brain.

4

5 **Binding of unloaded nanoparticles to P-selectin under flow**



6

7 **Fig 3. Evaluation of NPs binding on recombinant P-selectin under flow.** A) Fluorescently labeled
8 nanoparticles accumulated as clusters on channels coated at 50 $\mu\text{g/mL}$ with P-selectin. Nile Red-labeled
9 Fuco-NPs and Control-NPs were infused in micro-channels at either venous or arterial shear stress for 5
10 minutes (6.75 and 67.50 dyne/cm^2 , respectively) (scale bar = 10 μm). B) Corresponding quantitative
11 analysis of the number of fluorescent dots on a surface of 1230 μm x 105 μm (* $p < 0.05$; ** $p < 0.01$; $n = 5$
12 channels in venous conditions; $n = 4$ channels in arterial conditions).

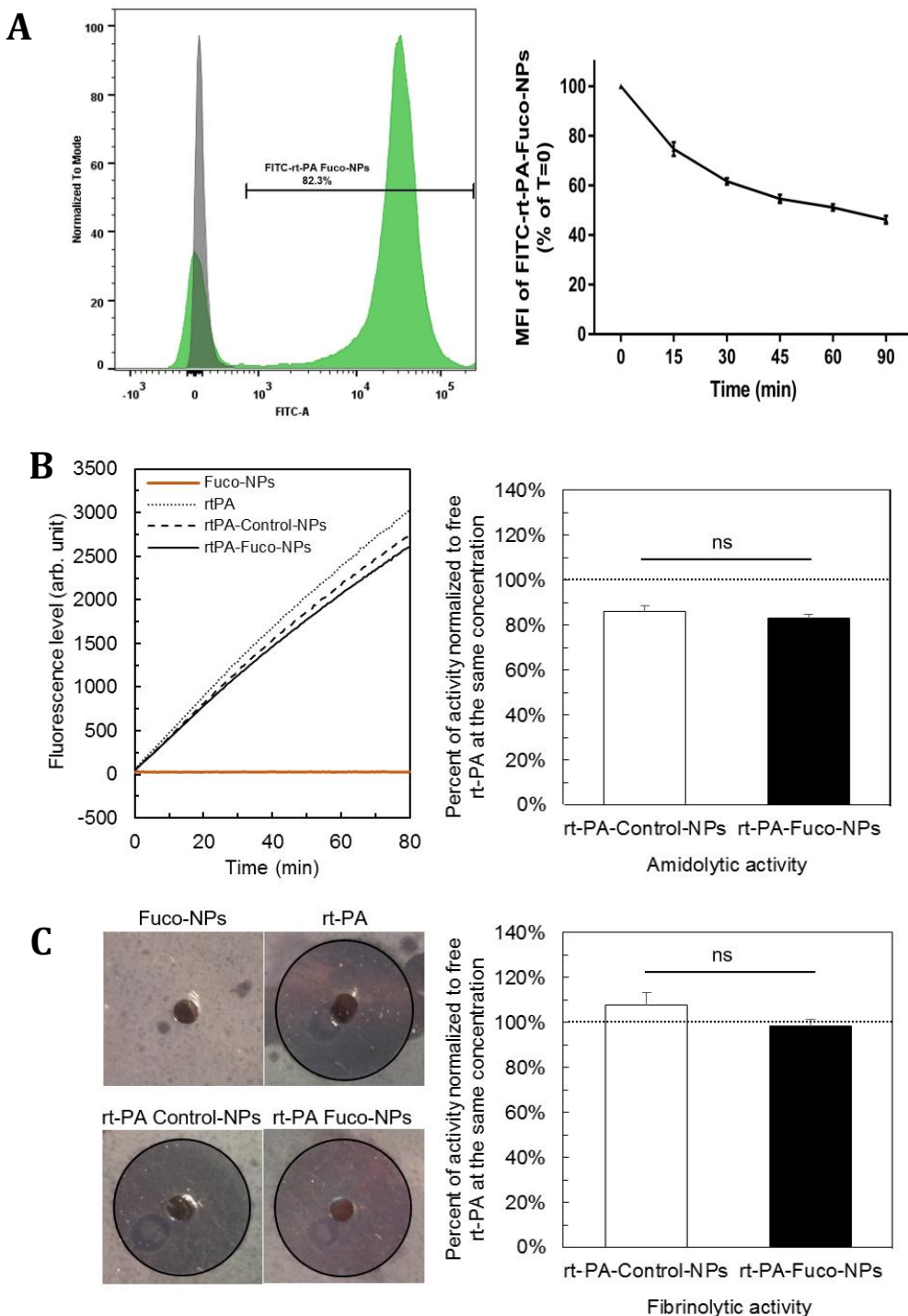
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14 Unloaded Fuco-NPs were first evaluated to evidence the capacity of fucoidan to interact with its
15 molecular target (Figure 3A). Under venous and arterial conditions, fluorescently labeled Fuco-
16 NPs accumulated significantly more to P-selectin than Control-NPs, with 1434 ± 169 clusters
17 versus 154 ± 17 clusters under venous shear stress, and 1844 ± 297 clusters versus 264 ± 34
18 clusters under arterial shear stress (Figure 3B).

19

20

1 **rt-PA loading and *in vitro* release and activity of rt-PA-loaded nanoparticles**



2
 3 **Figure 4. A) Loading efficiency and drug release assessed by flow cytometry.** Left: Nile Red Fuco-
 4 NPs were loaded either with FITC-rt-PA (green line) or left blank (grey line). More than 80% of the
 5 Fuco-NPs were loaded with FITC-rt-PA. Right: Flow cytometry evaluation of FITC-rt-PA release from
 6 Nile Red Fuco-NPs at 37°C. The MFI (Median Fluorescence Intensity) of FITC-rt-PA-Fuco-NPs at

1 different time points in PBS buffer. The fluorescence value was normalized over the average value of
2 MFI of FITC-rtPA-Fuco-NPs at time 0 min. (n=3). **Amidolytic and fibrinolytic activities. B)**
3 Amidolytic activities of Fuco-NPs, free rt-PA, rt-PA-Control-NPs and rt-PA-Fuco-NPs measured by the
4 PefaFluor[®] fluorogenic assay. Left: The tangent gradient is correlated to the enzymatic velocity. Right:
5 Corresponding quantitative analysis normalized to free rt-PA at the same concentration (n ≥ 3
6 experiments; ns non-significant). **C)** Fibrinolytic activities of the same samples determined by a fibrin-
7 plate agarose assay. Left: The fibrinolytic potential is obtained by measuring the lysed zone area after
8 overnight incubation at 37°C. Right: Corresponding quantitative analysis normalized to free rt-PA at the
9 same concentration (n ≥ 3 experiments; ns non-significant).

10
11 To further evidence the presence of rt-PA on Fuco-NPs, we used FITC-labeled rt-PA and loading
12 efficiency was assessed by flow cytometry analysis. When FITC-rt-PA-loaded Nile Red Fuco-NPs
13 (Figure 4A Left panel, green line) were compared to blank Nile Red Fuco-NPs (grey line),
14 82.3% of the fluorescently-conjugated Fuco-NPs were positive. In addition, the restricted
15 coefficient of variation (65.9%) indicated that the NPs had a uniform coverage of rt-PA.

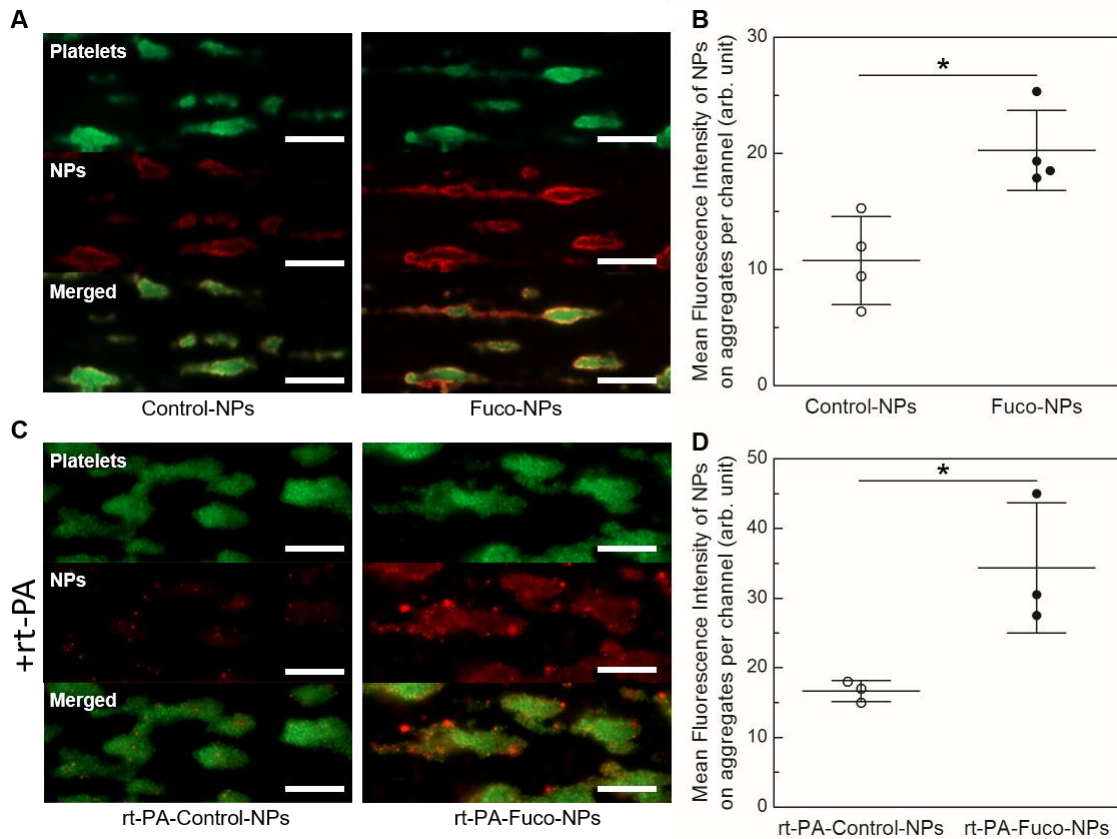
16 To further evidence the *in vitro* release of rt-PA from Fuco-NPs, we used FITC-labeled rt-PA
17 and the drug release was assessed by flow cytometry analysis. As shown in Figure 4A in the
18 right panel, the MFI (Median fluorescence intensity) of the FITC-rtPA loaded onto Fuco-NPs
19 decreased with time. These results showed that rt-PA in PBS buffer at 37°C under gentle
20 agitation is gradually released *in vitro* from Fuco-NPs up to 54% at 90 minutes.

21 Quantification of rt-PA adsorbed on the NPs showed that the same amount of drug was retained
22 in rt-PA-Fuco-NPs and in rt-PA-Control-NPs corresponding to 0.35 ± 0.02 mg of protein per mg
23 of NPs. Results of the loaded protein enzymatic activity and its activity in contact with fibrin are
24 presented in Figure 4. The amidolytic activity of the rt-PA in contact with Control-NPs and
25 Fuco-NPs decreased by $14 \pm 3\%$ and $17 \pm 2\%$ respectively (Figure 4B). The capacity of loaded
26 rt-PA to induce fibrinolysis in the fibrin plate assay was maintained, with values of $108 \pm 6\%$ for
27 Control-NPs and of $99 \pm 3\%$ for Fuco-NPs (Figure 4C). Thus, rt-PA loaded onto the NPs kept its
28 full activity in contact with fibrin. No significant difference was observed between both types of
29 NPs enabling to compare them in following experiments. In addition, unloaded Fuco-NPs
30 induced no effect (Figures 4B, 4C).

31

32

1 **Nanoparticles binding efficiency to activated platelets aggregates under flow**

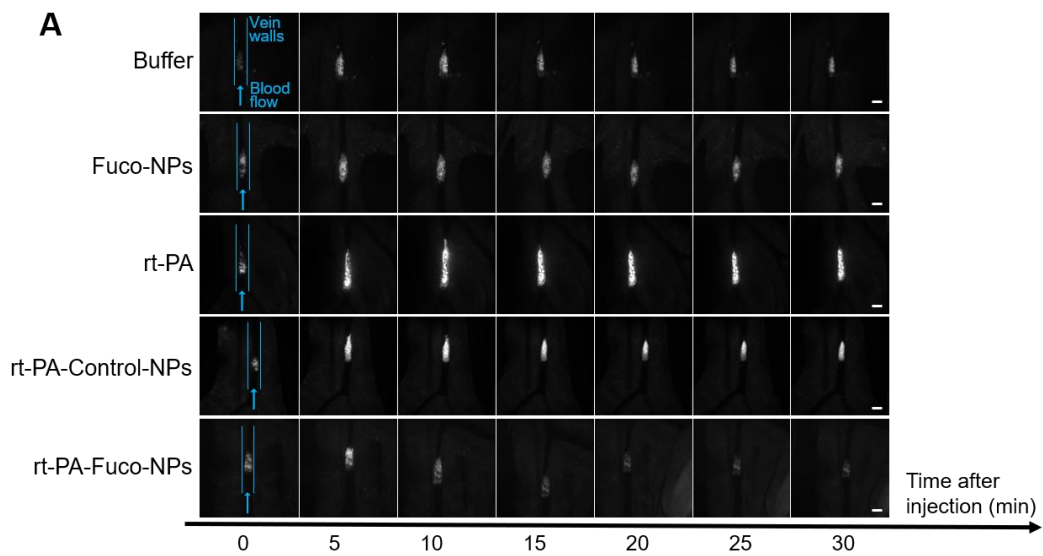


2
3 **Fig 5. Evaluation of NPs binding on activated platelet aggregates under flow.** A) Fluorescence image
4 after infusion for 5 minutes of unloaded Nile Red-nanoparticles (red) onto activated platelets aggregates
5 in venous conditions. Human whole blood mixed with 5 μ M of DIOC₆ (green) was previously infused
6 during 3.5 minutes in collagen coated micro-channels to induce platelet aggregates (scale bar = 50 μ m).
7 **B)** Corresponding quantitative analysis of the Mean Fluorescence Intensity averaged on platelet
8 aggregates. One MFI value was obtained per channel averaged over more than 25 aggregates. Values of 4
9 independent experiments are reported (* p <0.05). **C)** Fluorescence image after infusion for 12 minutes of
10 Nile Red-nanoparticles (red) loaded with rt-PA under similar conditions (scale bar = 50 μ m). **D)**
11 Corresponding quantitative analysis of rt-PA-loaded nanoparticles on platelet aggregates (* p <0.05).

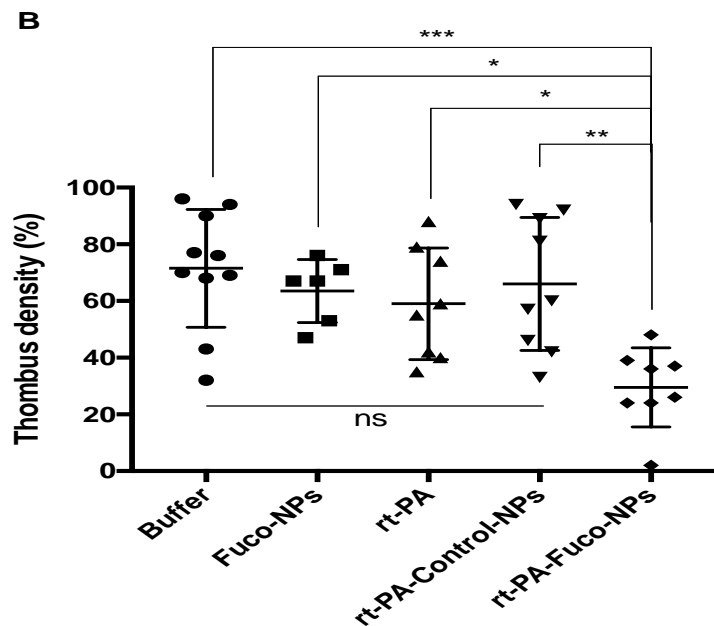
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13 The Fuco-NPs binding efficiency to P-selectin expressed by activated human platelets is
14 illustrated in Figure 5. Perfusion of human whole blood on collagen induced formation of
15 platelet aggregates that expressed P-selectin (supplementary data Figure S2). After perfusion of
16 either Nile Red-Fuco-NPs or Nile Red-Control-NPs on preformed aggregates at venous shear
17 rate, an accumulation of red fluorescence over time was observed at the surface of the platelets.

1 After perfusion, the fluorescence intensity induced by Fuco-NPs was two-fold higher than that of
 2 Control-NPs showing that Fuco-NPs accumulated significantly more onto activated platelets
 3 ($p < 0.05$) (Figures 5A, 5B). As shown in Figures 5C and 5D, NPs loaded with rt-PA showed the
 4 same trend. After perfusion, a strong fluorescence was observed on the platelet aggregates with
 5 rt-PA-Fuco-NPs (Figure 5C). Their accumulation was significantly higher than that of rt-PA-
 6 Control-NPs ($p < 0.05$) (Figure 5D).

7
 8 ***In vivo* thrombolytic efficiency**
 9



10



11

1 **Fig 6. Evaluation of thrombolysis efficiency in a mouse model of venous thrombosis. A)**
2 **Fluorescence imaging of Rhodamine 6G-labeled platelets at site of thrombosis over time.**

3 Representative examples of thrombus evolution as determined by platelet accumulation at different
4 times after injection of buffer, Fuco-NPS, rt-PA, rt-PA-Control-NPs and rt-PA-Fuco-NPs, respectively
5 (scale bar = 200 μ m). Prior to injection, circulating platelets were fluorescently labeled with
6 Rhodamine 6G and thrombosis was generated on a mesenteric vein by applying a FeCl₃-soaked paper
7 for 1 minute. **B) Thrombus density at 30 minutes after injection.** The thrombus density was defined
8 as the fluorescence intensity of the thrombus area normalized to the peak platelet accumulation after
9 injection. Five groups were analyzed (n = 41 mice). The dose of rt-PA was normalized by the protein
10 amount. The significance is defined with respect to the control group that received buffer (ns: non-
11 statistically significant; *p<0.05; **p<0.01; ***p<0.001).

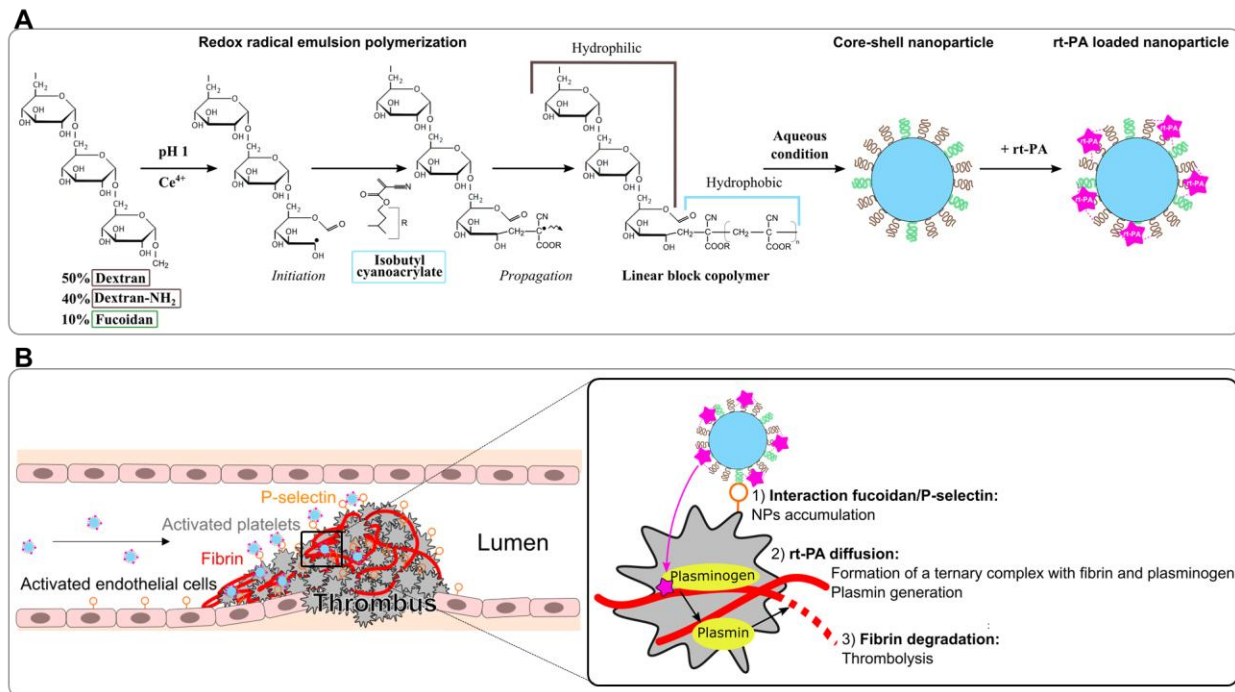
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13 All treatments were administered during thrombus formation. All thrombi experienced a growing
14 phase. In 68% of mice, the maximum intensity was reached in the first 10 minutes after injection.
15 Figure 6A provides representative examples of thrombus evolution with the different treatments.
16 The thrombus density normalized to the peak intensity for each animal was similar for mice that
17 received buffer as a control (n = 10) and for mice receiving unloaded Fuco-NPs (n = 6), as
18 shown by the quantitative analysis depicted in Figure 6B. In both groups, the thrombus became
19 rapidly stable after reaching a plateau and the relative thrombus density at 30 minutes was
20 around 67.5% (71.5 \pm 6.6% for Buffer and 63.5 \pm 4.5% for Fuco-NPs). In mice receiving rt-PA-
21 based treatments (rt-PA, rt-PA-Control-NPs and rt-PA-Fuco-NPs), a competition between
22 thrombosis and thrombolysis was observed in the first 15 minutes. However, the overall trend
23 was a fast platelet recruitment followed by a decrease and a stabilization at 30 minutes.
24 Examples of platelet accumulation over time after injection of buffer, Fuco-NPs rt-PA, rt-PA-
25 Control-NPs and rt-PA-Fuco-NPs are depicted in Figure 6A. Kinetics can be seen in
26 supplementary data for rt-PA-Fuco-NPs, rt-PA-Control-NPs and rt-PA alone (supplementary
27 movies S2, S3 and S4). rt-PA injection alone at a dose of 2.5 mg/kg (n = 8) decreased the
28 thrombus density down to 59.0 \pm 7.0%, which was not statistically different from the control
29 group. At this dose, free rt-PA was thus not sufficient to generate a pronounced thrombolytic
30 effect. For rt-PA-Control-NPs injected into 9 mice, the thrombus density was on average 66.0 \pm
31 7.8% at 30 minutes, which is not statistically different from the rt-PA alone. In contrast, injection
32 of rt-PA-Fuco-NPs (n = 8) at a similar dose generated a significant thrombus reduction. The

1 density decreased down to $29.5 \pm 4.9\%$ after 30 minutes. Furthermore, the treatment based on rt-
 2 PA-Fuco-NPs was the only one to induce more than 70% of reduction (density <30%) in 4 mice
 3 out of 8.

4

5 Discussion

6 In this study, a new strategy for targeted thrombolysis based on polymer NPs functionalized with
 7 fucoidan was evaluated (Figure 7). The preclinical study demonstrated the potential of such NPs
 8 in acute thrombosis. Fucoidan is an abundant natural polysaccharide that presents no
 9 immunogenic risk. It is therefore of great interest in comparison with P-selectin antibodies and
 10 recombinant PSGL-1 used to functionalize nano- and microsystems [47, 48]. Fucoidan-based
 11 NPs are highly promising nanocarriers to be used in pathologies characterized by overexpression
 12 of P-selectin, such as thrombosis. In addition, injection of rt-PA alone to induce thrombolysis has
 13 a limited efficiency and is associated to serious side effects. Fucoidan-functionalized NPs were
 14 developed as carriers for rt-PA to improve the drug efficiency with the intention to promote its
 15 specific accumulation and to decrease the injected dose and side effects.



16

17 **Figure 7. A) Nanoparticle formulation.** Polysaccharide chains are oxidized by Cerium (IV) ions. The
 18 free radical initiates the radical emulsion polymerization of isobutylcyanoacrylate monomers, which
 19 creates amphiphilic linear blocks copolymers. In aqueous conditions, they assemble into core-shell

1 nanoparticles. rt-PA is mixed with the suspension and interacts with the aminated nanoparticles by
2 electrostatic interactions. **B) Nanoparticle interaction with the thrombus.** 1) Nanoparticles specifically
3 accumulated on the thrombus via the fucoidan/P-selectin interaction. 2) rt-PA may diffuse within the
4 fibrin network where it forms a ternary complex with plasminogen linked to fibrin. This initiates the
5 formation a plasmin. 3) Plasmin generates fibrinolysis.

6
7 Physico-chemical characterizations and biodistribution studies in mice were first performed on
8 unloaded NPs. Spherical NPs with a hydrodynamic diameter of around 130 nm were obtained
9 (Figure 1). Unspecific NPs (Control-NPs) were synthesized with a negatively charged
10 polysaccharide instead of fucoidan (Figure 1). Carboxymethyl-dextran was chosen as a control to
11 obtain NPs with a slightly negative surface charge but without the sulfate groups of fucoidan that
12 are involved in its interaction with P-selectin [27, 41]. The physical properties of unloaded
13 Control-NPs and Fuco-NPs were similar. The NPs size was in the range of other types of NPs
14 developed for thrombolysis [13-16]. Although the size could be an important parameter, the
15 optimal NP size for interaction with a fibrin network is unknown. The radiolabeled Fuco-NPs
16 biodistribution profile in mice shown by both SPECT images and tissue biodistribution
17 highlighted a partial elimination by the kidneys with a high adsorbed dose in the bladder,
18 followed by a capture of Fuco-NPs by the reticuloendothelial system in the liver. These results
19 are in agreement with a study performed with nanoparticles synthesized by RREP but with a
20 different polysaccharide coating [49]. In our case, no signal was found in the heart, brain or in
21 the lungs. The *in vitro* binding assays performed on recombinant P-selectin confirmed that
22 fucoidan could efficiently recognize and bind P-selectin, even when incorporated in the
23 polysaccharide shell (Figure 3). Interestingly, this experiment showed that the affinity of Fuco-
24 NPs for P-selectin was observed both at low shear stress (6.75 dyne/cm^2) and at high shear stress
25 (67.5 dyne/cm^2), whereas most *in vitro* affinity assays described in the literature are performed in
26 static conditions [21, 24] or at very low ($<6.75 \text{ dyne/cm}^2$) or uncontrolled shear stress [50, 51].
27 The analysis of NPs by flow cytometry revealed that $>80\%$ of NPs were loaded with rt-PA and
28 that the amount of rt-PA per NP was quite homogeneous (Figure 4A). Still, we also found that a
29 minor proportion of NPs was not loaded which indicated that the loading step could be
30 improved. This is a critical issue since non-loaded NPs can potentially compete with loaded ones
31 for P-selectin binding and may have lowered the thrombolytic activity of rt-PA-NP preparations.
32 The *in vitro* results described in Figure 5 revealed that Fuco-NPs interacted significantly more

1 with activated platelet aggregates than Control-NPs. In addition, adsorption of rt-PA did not
2 seem to impair the Fuco-NPs binding efficiency. Thus, these results suggested that the use of rt-
3 PA-Fuco-NPs could induce more rt-PA to accumulate into the thrombus.

4 Fuco-dan-functionalized NPs associated with rt-PA were also tested *in vitro* to evaluate the drug
5 release. At 37°C in PBS buffer, the rt-PA could be released continuously from the Fuco-NPs.
6 Then, they were tested *in vivo* in a mouse model of venous thrombosis induced by iron chloride.
7 This model has the advantage to create a dense and reproducible thrombus within few minutes,
8 in comparison with other standard venous models that require hours or days to develop, and
9 display lack of reproducibility [52]. Buffer alone was injected in 10 mice to evaluate thrombosis
10 standard evolution. In all mice, the maximum platelet accumulation was reached within the first
11 15 minutes after injection. At 30 minutes the thrombus was therefore considered stable. Figure 6
12 summarized the preclinical study results. Unloaded Fuco-NPs did not improve significantly the
13 outcome compared with buffer. This result confirmed that unloaded Fuco-NPs have no intrinsic
14 effect on the thrombosis process even if they potentially accumulate on the thrombus. The
15 thrombi in mice receiving free rt-PA or rt-PA-Control-NPs were not significantly different from
16 the ones in the control group. At the used dose, free rt-PA was not sufficient to display a
17 pronounced thrombolytic effect. Moreover, these results indicate that associating rt-PA with a
18 non-targeting nanocarrier had no significant effect on the drug efficiency. In our work, rt-PA is
19 loaded by adsorption post-synthesis, whereas entrapment strategies during the NP synthesis
20 process have mainly been described with non-targeting nanocarriers [12-14]. While covalent
21 binding strategies may avoid premature drug release, it could decrease the protein fibrinolytic
22 activity once on site [53]. In our design, the presence of NPs only slightly affected the drug
23 amidolytic activity and full fibrinolytic activity of the protein was recovered in contact with
24 fibrin (Figures 4B & 4C). Positively charged aminated dextran was incorporated into the
25 polysaccharide shell to promote electrostatic interaction, mimicking the natural fibrin binding
26 sites. Interestingly, L-arginine is used to stabilize rt-PA pharmaceutical formulations [14, 54].
27 Alternative functional groups such as lysine could be explored in future studies to load the drug
28 since lysine can bind the kringle domains of rt-PA [55, 56].

29 We found that targeted nanoparticles loaded with rt-PA (rt-PA-Fuco-NPs) significantly improved
30 the outcome at 30 minutes as the thrombus density decreased down to 29.5%. In addition, 4 mice
31 out of 8 showed a decrease >70%. Despite the risk of desorption from the nanoparticles [57],

1 these *in vivo* results indicated that the loaded protein remained active. Together, these results
2 strongly support the fact that rt-PA and Fuco-NPs had a potent synergistic effect. In addition, the
3 beneficial effect of the targeting strategy was clearly evident after 30 minutes post-induction
4 whereas other targeted nanocarriers described in the literature required more time to improve the
5 drug efficiency [25, 26].

6 The results of this preclinical study suggested that the fucoidan present on the surface of the NPs
7 enhances the thrombolytic efficiency. The *in vitro* binding assays supported the hypothesis that
8 this is achieved by promoting the accumulation of the NPs on the platelet surface. Injection of
9 100 μg of fluorescent NPs did not allow us to observe any uptake with the limited sensibility of
10 the imaging technique. Once injected at a 20 times higher dose, Fuco-NPs were clearly identified
11 on the edge of the thrombus from the first minute and accumulated for more than 30 minutes
12 (Supplementary Figure S3). This observation is in accordance with other targeting studies with
13 nanoparticles and is promising for future developments of Fuco-NPs [21, 58].

14 We have used an appropriate but limited number of animals and the mouse model that we have
15 used is far from the complexity and variety of human thrombi. Nevertheless, the present study is
16 a first proof-of-concept for the valuable association of rt-PA with fucoidan-functionalized
17 nanocarriers. The nanocarriers that were used were chosen according to the presence of
18 polysaccharides at the surface and to their stealth properties [59]. The injected concentration was
19 calculated to be about 50 μg NPs per mL of blood. At this concentration, these functionalized
20 nanoparticles exhibit an appropriate cytocompatibility [60, 61]. Their biodegradability needs to
21 be considered for future clinical developments. Even if intravital microscopy is a powerful and
22 precise technique in mice to assess thrombosis evolution in terms of platelet recruitment [62],
23 complementary experiments to monitor the blood flow, to evaluate treatment side effects, like
24 bleeding time, and to assess the targeting capacity of rt-PA-Fuco-NPs *in vivo* in small and large
25 animals, are needed [24, 51].

27 **Conclusions**

28 An efficient thrombolytic agent based on fucoidan-functionalized polymer nanoparticles
29 targeting P-selectin was developed. A flow assay was designed to validate the role of fucoidan at
30 the surface of the nanoparticle. As compared to other strategies that involve grafting antibodies
31 or the use external magnetic force, fucoidan is of great interest for promoting the specific

1 accumulation of nanocarriers at site of thrombosis. An *in vivo* proof-of-concept showed that rt-
2 PA loaded fucoidan-nanoparticles improved the thrombolysis efficiency of the drug in
3 thrombosis acute phase. These results highlighted for the first time the relevance of targeting P-
4 selectin for the treatment of thrombosis with rt-PA. Optimization steps of the interactions
5 between the drug and nanoparticles, as well as in-depth studies of the *in vivo* behavior of loaded
6 nanoparticles should be performed before moving towards clinical implementation.

7

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32

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1 MJ (PhD student) performed the synthesis and characterizations of nanoparticles, the rt-PA
2 loading, the *in vivo* thrombolysis evaluation, participated to all experiments and wrote the draft
3 of the manuscript. RA (Engineer) performed the *in vivo* model of thrombosis, the *in vivo*
4 SPECT/CT imaging and the tissue biodistribution. BL (PostDoc) performed the chemical
5 modification of dextran and the *in vitro* drug release experiments. AB (Trainee in Master)
6 realized the *in vitro* amidolytic and fibrinolytic tests. JA (Lecturer) carried out the radiolabeling
7 of the nanoparticles. VO (Engineer) performed the flow binding assays. AN (Professor) realized
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9 results and corrected the manuscript. CC (Researcher) supervised the thesis of MJ, discussed the
10 results and corrected the manuscript.

11 The authors have declared that no competing interest exists.

12 **Supplementary Materials:**

13
14 Figure S1. Physico-chemical characterization of Control-NPs and Fuco-NPs. A) Diffusion Light
15 Scattering (DLS) measurement in intensity, number and volume. B) Electrophoretic Light
16 Scattering (ELS) measurement.

17 Figure S2. Expression of P-selectin by activated platelet aggregates evidenced by perfusion of
18 FITC-labeled anti-P-selectin. FITC-labeled anti-P-selectin or FITC-labeled IgG with matched
19 isotype were mixed with human whole blood. Blood was perfused in collagen coated micro-
20 channels at arterial shear stress (67.5 dyne/cm^2). Activation and aggregation of platelets was
21 visualized by phase contrast microscopy and fluorescence microscopy (scale bar = $50 \mu\text{m}$).

22 Figure S3. Accumulation of Fuco-NPs in the thrombus monitored by intravital microscope. 2 mg
23 of Nile Red-labeled Fuco-NPs were injected in mice where thrombosis was previously induced
24 and platelets were labeled in green with DIOC₆. Platelet and NP accumulation at site of injury
25 were monitored in real-time up to 40 minutes. Images shown here were taken every 100 seconds.

26

27 Movie S1: SPECT/CT imaging at 80 minutes.

28 Movie S2: Thrombus density over 30 minutes after treatment with rt-PA-Fuco-NPs in a mouse.

29 Movie S3: Thrombus density over 30 minutes after treatment with rt-PA-Control-NPs in a
30 mouse.

- 1 Movie S4: Thrombus density over 30 minutes after treatment with rt-PA in a mouse.
- 2
- 3 Graphical abstract: Core-shell polymer nanoparticles are functionalized with fucoidan to target
- 4 P-selectin and promote the specific accumulation of loaded rt-PA at the thrombus.
- 5