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## Multitasking actors of *Staphylococcus aureus* metabolism and virulence

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## 12 Keywords

13 *Staphylococcus aureus*; metabolism; virulence

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## 15 Abstract (50 words)

16 Recent studies have uncovered the striking commonality of multitasking molecular actors 17 linking metabolism and virulence regulation. Beyond the well-known importance of 18 carbohydrate and amino acid metabolism regulators in coordinating metabolic pathways and 19 pathogenesis, we highlight recent major advances linking lipid and nucleic acid pathways to 20 virulence regulation in *S. aureus*.

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#### 23 Staphylococcus aureus virulence and metabolism

Staphylococcus aureus, which is both a human commensal and pathogen, is mainly found in 24 the anterior nares, but can also colonize the skin and mucosa. The bacterium is responsible for 25 a wide variety of infections, ranging from mild skin infections to life-threatening bacteremia. 26 When facing different host microenvironments, S. aureus must cope with nutrient shifts and 27 modulates its virulence level by well-known interconnected regulatory systems including Agr 28 29 and SaeRS. Agr system controls the switch from expression of cell-surface associated proteins, 30 including fibronectin-binding proteins (FnBPs; FnbA and FnbB), which are involved in biofilm formation and internalization within host cells, to secreted toxins including alpha-31 hemolysin. 32

33 Multiple metabolic factors involved in carbohydrate or amino acid metabolism have been 34 found to moonlight in virulence. We highlight here recent discoveries linking lipid and 35 nucleobase metabolism to *S. aureus* pathogenesis.

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### **37 Purine and pyrimidine biosynthetic pathways**

While investigating the unusual ability of several isolates to form clumps in liquid culture 38 39 (hyper-clumping phenotype), Goncheva et al. identified mutations in the purine biosynthesis repressor gene, *purR* [1]. They discovered that, besides its role in purine pathway regulation, 40 PurR was also involved in virulence modulation. Indeed, purR mutants were hypervirulent in 41 42 a systemic mice infection model. Hypervirulence of the purR mutants correlated to their hyper-clumping phenotype in serum, due to increased expression of genes encoding FnBPs. 43 The clinical relevance of *purR* mutations is supported by the fact that *purR* variants are found 44 in public databases of genomes [1]. These variants could emerge during the course of 45 infection since Goncheva et al. were able to recover purR mutants from the organs of mice 46 47 initially infected with a wild-type USA300 strain [1].

In a paper published a few months later, Sause *et al.* confirmed that, in a *purR* defective mutant, overexpression of FnbA increases bacterial burden in tissues in a murine model of bacteremia [2]. Sause *et al.* demonstrated direct binding of PurR to the promoters of several virulence genes (including *fnbA*, genes coding for leukocidins and alpha-hemolysin) and regulators, highlighting that multiple factors contribute to the hypervirulence of *purR* mutants independently of purine biosynthesis [2].

54 One can hypothesize that a decrease in purine availability during the course of infection 55 requires concomitant PurR derepression of both purine biosynthesis pathway and virulence factor expression. Besides, as shown by Goncheva *et al.* [1], stress induced by the host
immune response selects for *purR*-inactivating mutations, which likely confers metabolic
advantage that incidentally locks these strains in a hypervirulence state.

A role in pathogenesis has also been recently reported for PyrR, the repressor of the pyrimidine biosynthesis operon, by Copin *et al.* while investigating the epidemic success of a USA300 subclone [3]. In this subclone, a *pyrR*-inactivating mutation increased *pyr* operon gene expression, providing a colonization and transmission advantage in a mouse gastrointestinal model [3].

64 Collectively, these findings provide novel links between *S. aureus* pathogenesis and 65 nucleobase metabolism through modulation of virulence and colonization potential (Table 1).

66 The other recent major findings linking pathogenesis and metabolism in S. aureus pertain to

67 the regulation of lipid pathways and more specifically highlight the key roles of fatty acids.

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#### 69 Fatty acid (FA) metabolism

70 FA metabolism has been recently linked to both virulence and colonization potential in S. 71 aureus. Fatty acid kinase A (FakA encoded by fakA aka vfrB) phosphorylates and 72 incorporates exogenous FAs into the S. aureus cell membrane [4]. Whereas a S. aureus strain 73 lacking FakA exhibits an increased virulence in a mouse model of dermonecrosis [5], it has a 74 virulence defect in a mouse model of bacteremia [6]. While the increased skin virulence remained unexplained, Lopez et al. correlated the virulence attenuation observed in the 75 76 bacteremia model with a lack of type VII secretion system (T7SS) activation. The S. aureus 77 T7SS contributes to virulence in murine infection models. The authors suggest that S. aureus 78 modulates the expression of its T7SS in response to environmental levels of host-derived FAs. Indeed, the FakA-mediated incorporation of environmental FAs in the bacterial membrane 79 modifies its fluidity, which could trigger T7SS expression [6]. 80

81 Transcriptional profiling of a *fakA* mutant provided additional explanation for its virulence 82 defect by showing severe attenuation in SaeRS-dependent virulence factor transcription [7]. The SaeRS two-component system is a master regulator of virulence genes in S. aureus. 83 Ericson et al. demonstrated that FAs that fail to be incorporated into the membrane 84 accumulate within *fakA* mutant cells and have a direct inhibitory effect on SaeRS signaling. 85 Thus, by incorporating FAs into S. aureus membranes, FakA modifies both membrane 86 fluidity and the FA intracellular pool, which impact T7SS expression and SaeRS signaling 87 respectively. Interestingly, fakA mutations were identified in clinical isolates from cystic 88

fibrosis patients [8]. Collectively, these studies suggest a key but still underappreciated keyrole for FakA in *S. aureus* pathogenesis.

Studies on bacterial lipid metabolism may further our understanding of the role of FAs in 91 bacterial pathogenesis, since, in addition of being major energy sources and important 92 component of membranes, lipid-related molecules released by bacteria are signal molecules 93 that can be sensed by other bacteria and by host immune cells. Indeed, bacterial lipoproteins 94 and lipopeptides are pathogen-associated molecular patterns (PAMPs), which are major 95 96 agonists for Toll-like receptor (TLR) signaling during the innate immune response. By 97 conducting a screen to identify S. aureus factors of immune evasion, Grayczyk et al. found that a mutation in the lipoic acid synthetase gene (*lipA*) elicited a hyper-inflammatory 98 99 response in macrophages [9]. Lipoic acid (LA) is a fatty acid-related cofactor covalently linked to multiprotein enzymatic complexes such as pyruvate dehydrogenase (PDH), yielding 100 101 lipoyl-PDH [9]. Wild type S. aureus, but not a lipA mutant, releases nanomolar quantities of lipoyl-PDH into the culture supernatant, which is able to blunt TLR2-dependent activation of 102 103 macrophages. To support the immunosuppressive function of protein lipoylation, Grayczyk et 104 al. showed that a synthetic tripeptide containing a lipoyl residue, but not the unmodified tripeptide, was sufficient to suppress TLR1/2-dependent macrophage activation by S. aureus 105 lipoproteins [9]. The authors thus proposed that secreted lipoyl-PDH prevents lipoprotein-106 dependent activation of macrophages through its lipoyl moiety. It is likely that lipoylated 107 proteins antagonize lipoprotein activation of macrophages by competing for TLR binding. 108

More recently, the predominant *S. aureus* secreted lipase, Geh, was demonstrated to also be a potent immunomodulator [10]. Besides liberating long chain FAs from host-derived lipoproteins for nutritional purpose, Geh also hydrolyzes *S. aureus* lipoproteins, which prevents their recognition by host immune cells and fulfills an elegant immune evasion function, which ultimately promotes persistence [10]. Thus, a protein can exhibit several functions by displaying several activities or by performing the same activity on different targets.

LipA and Geh are two striking examples of *S. aureus* enzymes involved in both lipid
metabolism and immune evasion. While LipA seems to prevent lipoprotein activation of TLR,
Geh prevents immune cell activation by directly hydrolyzing lipoproteins. Therefore, FakA,
LipA and Geh represent multitasking enzymes that play a critical role in *S. aureus*pathogenesis in addition to their function in lipid metabolism (Table 1).

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### 122 Concluding Remarks

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The list of multifunctional proteins frequently referred to as "moonlighting proteins" or 124 multitasking enzymes involved in metabolic pathways and bacterial pathogenesis is rapidly 125 expanding. The online MoonProt Database includes around 400 proteins displaying such 126 moonlighting functions and it is likely that many other moonlighting proteins have yet to be 127 found (http://www.moonlightingproteins.org/). Since additional functions can be difficult to 128 predict, multifunctional proteins are often serendipitously discovered during the course of 129 130 proteomic or genomic data analysis. However, while such omics studies may help uncover unexpected new functions of proteins not previously linked to virulence, the well-known 131 classical virulence factors are resourceful and can still surprise us. In particular, it was known 132 133 that when located extracellularly, enzymes of glycolysis could contribute to bacterial adhesion. Strikingly, it is now shown that translocation of cytoplasmic proteins into the extracellular 134 milieu by S. aureus requires a functional agr system through the membrane-damaging activity 135 of phenol-soluble modulin peptides [11]. 136

In addition, *S. aureus* biofilm proteome profiling revealed a high abundance of secreted virulence factors, including hemolysins, leukotoxins, and lipases trapped within the extracellular matrix where they fulfill an additional stabilizing function by generating electrostatic bridges [12]. Altogether, these studies underline how activities of multitasking proteins can be tightly linked with *S. aureus* pathogenicity.

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Table 1. Molecular actors of metabolism with a recently described role in *S. aureus*pathogenesis.

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Gene	Metabolic function	Impact on pathogenesis
purR	Repression of purine biosynthesis	Impacts fibronectin binding proteins (FnBPs),
		leukocidins and $\alpha$ -toxin regulation. <i>purR</i>
		inactivation leads to hyper-clumping,
		hypervirulence and increased biofilm
		formation [1, 2].
pyrR	Repression of pyrimidine	<i>pyrR</i> inactivation provides a colonization and
	biosynthesis	transmission advantage in a mouse
		gastrointestinal model [3].
fakA	Fatty acid phosphorylation and	Impacts type VII secretion system activation
	uptake into S. aureus cell	[6] and SaeRS-dependent virulence factor
	membrane	transcription [7].
		fakA mutants exhibit a virulence defect in a
		mouse model of bacteremia [6] and an

		increased virulence in a mouse model of dermonecrosis [5].
geh	Hydrolysis of environmental long chain lipids and <i>S. aureus</i> lipoproteins released in the extracellular milieu	By hydrolyzing lipoprotein pathogen- associated molecular patterns (PAMPs), Geh prevents their recognition by host immune cells and allows immune evasion [10].
lipA	Synthesis of lipoic acid (LA), a cofactor that covalently modify proteins such as the pyruvate dehydrogenase (PDH) complex yielding lipoyl-PDH	Lipoyl-proteins antagonize lipoprotein activation of macrophage likely by a competition for TLR binding [9].

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