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Characterization of intrinsic membrane properties of vestibulo-spinal neurons through xenopus development

Introduction

In vertebrates, central vestibular neurons (2°OVN) are subdivided in distinct populations according to their projection and their function (vestibulo-ocular/spinal - commissural...). In amphibian, vestibulospinal neurons (VSN), involved in postural control, are organized into lateral vestibulospinal tract (LVST) and tangential (TAN) nuclei projecting to ipsiand contralateral spinal cord, respectively (Straka et al., 2001). Here, we focused on the LVST neurons, largest group (and common to other vertebrates lineages) localized mostly in rhombomeres 4, at the VIIIth nerve entrance.



The metamorphosis in *Xenopus laevis* toad, a purely aquatic anuran, offers a unique opportunity to unravel integrative properties of central vestibular neurons in relation to a specific vestibular sensory reference frame (aquatic vs terrestrial, otolith vs canal) and to a given locomotor system (tail- vs limb based swimming).

This study aims to investigate:



In adult terrestrial frog, two groups of 2°OVN were described according to their discharge dynamic and their intrinsic membrane properties (Beraneck et al., 2007). In response to injection of positive current steps phasic neurons exhibit a high-frequency burst of 1-5 spikes, with monophasic AHP, whereas tonic neurons fire continuously spikes with a biphasic AHP. Such membrane properties tune phasic and tonic 2°OVN in band-pass and low-pass neuronal filters, respectively.

Consequently, phasic neurons could encode motion-related high dynamic sensory-motor signals whereas tonic neurons could encode position-related slow dynamic signals in vestibulo-motor behaviors. Nonetheless this characterization was based on 2°OVN that were not related to a specific vestibular functional pathway



> Intrinsic membrane properties exhibited by 2°OVN from identified LVST pathway at both larval and adult-like stages.

> Underlying developmental plasticity mechanisms related to the posturo-locomotor system re-modeling.

Materials and methods



Experiments were performed on post-metamorphosis adult-like (juvenile) stage 65-66 and larval stage 53-56 of south African clawed toad xenopus laevis. Stages were identified according to external body criteria (Nieuwkoop and Faber, 1956). Briefly, consecutive to anesthesia in a 0.05% MS-222 water solution and after forebrain removal, the CNS was dissected in cold Ringer's saline (93.5 mM NaCl, 3 mM KCl, 30 mM NaHCO3, 0.5 mM NaH2PO4, 2.6 mM CaCl2, 1 mM MgCl2, and 11 mM glucose, pH 7.4). Rhodamine dextran (RDA) crystals were applied in a tiny incision performed at the ventral surface of the rostral hemi-cord. CNS in vitro preparation was incubated in circulating ringer's saline at 16°C for at least 3h to allow retrograde labeling of vestibulospinal neurons.

norizontal slice transversal slice





Patch-clamp recordings of RDA+ LVST neurons were performed in whole-cell configuration on 350µm transversal or horizontal brainstem slices continuously perfused with oxygenated Ringer's solution at 18-20°C. Patch electrodes were filled with intracellular solution (in mM= 115 K-gluconate, 2 MgCl2, 2 EGTA, 10 HEPES, 2 MgATP, 0, 2 NaGTP and 0, 01% biocytin). Different protocols in current clamp (-200 to +300pA current step serie and current ramp) and voltage clamp configuration (Ih, IV curve) were applied to measure discharge dynamics and basic membrane properties of RDA+ LVST neurons .



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Thoraco-lumbar spinal network

Patch-clamp recorded RDA+ threshold LVST neurons were intracellularly filled with biocytin. After recordings slices were fixed in PFA 4% and revealed with Alexa fluor 488 streptavidin. Slices were imaged with a confocal microscope at 10x and 40x to allow a good medio lateral and rostro-caudal localization of recorded neurons within the entire LVST population.



