



HAL
open science

The relationship between membrane fatty acid content and mitochondrial efficiency differs within- and between-omega-3 dietary treatments

Karine Salin, Margaux Mathieu-Resuge, Nicolas Graziano, Emmanuel Dubillot, Fabienne Le Grand, Philippe Soudant, Marie Vagner

► To cite this version:

Karine Salin, Margaux Mathieu-Resuge, Nicolas Graziano, Emmanuel Dubillot, Fabienne Le Grand, et al.. The relationship between membrane fatty acid content and mitochondrial efficiency differs within- and between- omega-3 dietary treatments. *Marine Environmental Research*, 2020, 163, pp.105205. 10.1016/j.marenvres.2020.105205 . hal-02409224

HAL Id: hal-02409224

<https://hal.science/hal-02409224v1>

Submitted on 22 Aug 2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 **TITLE:** The relationship between membrane fatty acid content and mitochondrial efficiency differs
2 within- and between- omega-3 dietary treatments

3

4 **AUTHORS:**

5 Karine Salin^{1*}, Margaux Mathieu-Resuge², Nicolas Graziano^{1,3}, Emmanuel Dubillot³, Fabienne Le
6 Grand¹, Philippe Soudant¹, Marie Vagner^{1,3}

7

8 **AFFILIATIONS:**

9 ¹ Univ Brest, Ifremer, CNRS, IRD, LEMAR, F-29280, Plouzané, France

10 ² WasserCluster Lunz — Inter-University Centre for Aquatic Ecosystem Research, Dr. Carl Kupelwieser
11 Promenade 5 A-3293 Lunz am See, Austria

12 ³ UMR 7266 LIENSs, 2 rue Olympe de Gouges 17000 La Rochelle, France

13

14 ***CORRESPONDING AUTHOR:** Karine Salin; Email: Karine.salin@ifremer.fr

15

16 **RUNNING TITLE:** n-3 HUFA levels alter mitochondrial efficiency

17

18 **KEYWORDS:** ATP/O ratio, docosahexaenoic acid (DHA), eicosapentaenoic acid (EPA), food quality,
19 *Chelon auratus*, global change.

20 **ABSTRACT**

21 An important, but underappreciated, consequence of climate change is the reduction in
22 crucial nutrient production at the base of the marine food chain: the long-chain omega-3 highly
23 unsaturated fatty acids (n-3 HUFA). This can have dramatic consequences on consumers, such as fish
24 as they have limited capacity to synthesise n-3 HUFA *de novo*. The n-3 HUFA, such as
25 docosahexaenoic acid (DHA, 22:6n-3) and eicosapentaenoic acid (EPA, 20:5n-3), are critical for the
26 structure and function of all biological membranes. There is increasing evidence that fish will be
27 badly affected by reductions in n-3 HUFA dietary availability, however the underlying mechanisms
28 remain obscure. Hypotheses for how mitochondrial function should change with dietary n-3 HUFA
29 availability have generally ignored ATP production, despite its importance to a cell's total energetics
30 capacity, and in turn, whole-animal performance. Here we (i) quantified individual variation in
31 mitochondrial efficiency (ATP/O ratio) of muscle and (ii) examined its relationship with content in
32 EPA and DHA in muscle membrane of a primary consumer fish, the golden grey mullet *Chelon*
33 *auratus*, receiving either a high or low n-3 HUFA diet. Mitochondria of fish fed on the low n-3 HUFA
34 diet had higher ATP/O ratio than those of fish maintained on the high n-3 HUFA diet. Yet,
35 mitochondrial efficiency varied up about 2-fold among individuals on the same dietary treatment,
36 resulting in some fish consuming half the oxygen and energy substrate to produce the similar amount
37 of ATP than conspecific on similar diet. This variation in mitochondrial efficiency among individuals
38 from the same diet treatment was related to individual differences in fatty acid composition of the
39 membranes: a high ATP/O ratio was associated with a high content in EPA and DHA in biological
40 membranes. Our results highlight the existence of interindividual differences in mitochondrial
41 efficiency and its potential importance in explaining intraspecific variation in response to food chain
42 changes.

43 1. Introduction

44 Fish provide critical sustenance for millions of people worldwide and have far reaching impacts on
45 the productivity of ecosystems (McIntyre et al., 2016). Yet, ongoing and future climate change
46 threatens the persistence of fish populations globally (Pörtner and Knust, 2007). An important, but
47 underappreciated, consequence of climate change is the reduction in production at the base of the
48 food chain of essential nutrient: the long-chain omega-3 highly unsaturated fatty acids (n-3 HUFA)
49 (da Motta Pacheco et al., 2014; Galloway and Winder, 2015; Hixson and Arts, 2016). **Water warming**
50 **(Hixson and Arts, 2016), acidification (Bermudez et al., 2015), or UV irradiation (Kang, 2011), all affect**
51 **n-3 HUFA primary producers physiology and community assemblages, leading to a dominance of n-3**
52 **HUFA-impooverished taxa (Galloway and Winder, 2015).** The n-3 HUFA such as eicosapentaenoic acid
53 (EPA, 20:5n-3) and docosahexaenoic acid (DHA, 22:6n-3) are essential to the structure and function
54 of all biological membranes and are thus considered to be important drivers of organism
55 performance (Hulbert et al., 2005; Ishizaki et al., 2001; Mazorra et al., 2003; Vagner et al., 2015).
56 Endogenous biosynthesis of n-3 HUFA from precursors is limited in most vertebrates, including
57 marine fish (Alimuddin et al., 2005; Arts and Kohler, 2009; Oboh et al., 2017). Changes in the n-3
58 HUFA availability in the fish diet causes strongly correlated changes in the fatty acid composition of
59 their biological membranes (Guderley et al., 2008; Nogueira et al., 2001; Ramsey et al., 2005). Fish on
60 n-3 HUFA-deficient diets can perform badly: reductions in EPA and DHA dietary content **reduced**
61 **growth (Norambuena et al., 2015; Vagner et al., 2015) and thermal tolerance (Vagner et al., 2014),**
62 **and altered whole-organism metabolic traits (Vagner et al., 2015; Vagner et al., 2014).** Determining
63 what causes animal performance to vary with n-3 HUFA diet constitutes a fundamental step in
64 predicting the impacts of projected oceanic changes on fish resilience (Kang, 2011).

65 Consideration of mitochondrial capacity to make ATP may improve our understanding of the
66 links between dietary n-3 HUFA availability and whole-animal performance. Mitochondrial ATP is
67 produced via oxidative phosphorylation, a process through which energy substrates are oxidized to
68 generate a protonmotive force that drives the phosphorylation of ADP to ATP. Hypotheses for how
69 mitochondrial function should change with dietary n-3 HUFA availability have focused on variation in
70 respiratory capacities and have generally ignored variation in ATP production (Kraffe et al., 2007;
71 Ramsey et al., 2005; but see Herbst, 2014; Vagner et al., 2015; Vagner et al., 2014). Although ATP
72 production depends on the rate of substrate oxidation, the number of ATP molecules produced for
73 each atom of oxygen consumed by the mitochondria during substrate oxidation (ATP/O ratio) can
74 vary (Brand, 2005; Salin et al., 2015). A fraction of the protonmotive force that is generated from
75 substrate oxidation is dissipated through proton leak across mitochondrial inner membranes and this
76 leakage can decrease the protonmotive force available to produce ATP (Brand, 2005; Kadenbach,
77 2003). Thus, the **greater** the mitochondria leak, the less efficiently an animal converts its metabolic

78 substrates into ATP, and the **lower** the ATP/O ratio. The proportion of energy dissipated in the
79 proton leak and the efficiency to make ATP vary among conspecific (Bottje and Carstens, 2009;
80 Robert and Bronikowski, 2010; Salin et al., 2016b) and can be influenced by environmental factors
81 including diet (Fontaine et al., 1996; Salin et al., 2018). A number of studies have found positive links
82 between intraspecific heterogeneity in efficiency to produce mitochondrial ATP and the whole-
83 organism performance, such as locomotory performance (Coen et al., 2012; Distefano et al., 2018;
84 but see Jahn and Seebacher, 2019), developmental rate (Salin et al., 2012), growth efficiency (Bottje
85 and Carstens, 2009; Salin et al., 2019) and reproductive output (Robert and Bronikowski, 2010),
86 suggesting that it might be a trait of ecological relevance.

87 Recent research has recognized the importance of accounting for individual heterogeneity in
88 predicting responses to global changes (Hamel et al., 2018). Because some individuals perform much
89 better than others **within** the same environment, individual heterogeneity is likely to directly
90 influence the potential for species to evolve adaptations for a reduced n-3 HUFA availability. A
91 number of studies have found positive links between n-3 HUFA content in membranes and
92 mitochondrial proton leak when comparing among species (Brand et al., 1994; Brookes et al., 1998),
93 but contradictory results were found when comparing treatment groups with studies reporting
94 positive (Martin et al., 2013), negative (Fontaine et al., 1996; Guderley et al., 2008) or no
95 relationships (Guderley et al., 2008) between content in n-3 HUFA in mitochondrial membranes and
96 leak respiration. N-3 HUFA are thought to have important effects on the mitochondrial capacity to
97 make ATP; but until now, there has, to our knowledge, been no assessment of whether membrane n-
98 3 HUFA content could explain variation in mitochondrial metabolism among individuals.

99 The present experiment integrates measurements of mitochondrial efficiency and
100 mitochondrial proton leak to determine whether reductions in EPA and DHA availability in food led to
101 changes in energy metabolism of a primary consumer fish. We first examined the effect of dietary n-
102 3 HUFA content on mitochondrial function, in particular on the efficiency to produce ATP (ATP/O
103 ratio) and the respiratory capacities to offset the proton leak (LEAK respiration). Secondly, we tested
104 whether differences between individuals in mitochondrial efficiency and mitochondrial LEAK
105 respiration vary with membrane n-3 HUFA content. To address this, we experimentally manipulated
106 the quantity of n-3 HUFA in food for wild-caught juvenile golden grey mullet (*Chelon auratus*). Fish
107 were fed either a high n-3 HUFA or low n-3 HUFA **diet**, and their membrane fatty acid composition
108 and mitochondrial functioning were determined in skeletal muscle. We choose juvenile golden grey
109 mullet as our study organism because they are likely to be the first levels of the food chain to face a
110 decline in availability of dietary n-3 HUFA, as this fish fed mainly on primary producers (Lebreton et
111 al., 2011; Mourente and Tocher, 1993). We analysed mitochondrial properties in the skeletal muscle
112 because the mitochondrial function of this tissue is known to influence whole-animal performance

113 (Coen et al., 2012; Salin et al., 2016a), and that fatty acid dietary content influences muscle
114 membrane fatty acid composition (McKenzie et al., 1998; Vagner et al., 2015).

115

116 **2. Materiel and methods**

117 *2.1. Fish origin and care*

118 Wild juvenile golden grey mullets (n = 23) were netted from the marshes of L'Houmeau France
119 (46°12'14.4''N 1°11'43.7''W) in November 2017 and transported to the laboratory Littoral
120 Environment Society (LIENSs), France, where all the experiments were conducted. Fish were kept in a
121 common thermoregulated 300 L-tank supplied with aerated recirculated sand-filtered natural
122 seawater and equipped with an external biological filter (Eheim, Deizisau, Germany). Fish were
123 maintained under a 12 L : 12 D photoperiod, and fed daily with commercial pellets (Le Gouessant®,
124 Lamballe, France). Temperature ($13.8 \pm 0.2^\circ\text{C}$) and salinity (28.7 ± 0.1) were monitored daily
125 (TetraCon® 325, Laboratoires Humeau, La Chapelle-sur-Erdre, France) and were kept similar to that
126 of the sampling site. Oxygen ($87.9 \pm 2.6\%$ air saturation) was monitored once a week. Fish were
127 acclimated for seven weeks to these conditions. The collection and handling of the animals were
128 carried out under the jurisdiction of the Departmental Service of Fisheries and the Animal Care
129 Committee of France (# 12886), respectively.

130 In January 2018, fish were anesthetized in 0.1 g L^{-1} MS-222 (Ethyl 3-aminobenzoate
131 methanesulfonate) in seawater, measured for body mass ($34.3 \pm 6.5 \text{ g}$), individually pit-tagged
132 (Biolog-id, Bernay, France), and randomly assigned to one of the four replicate 300-L holding tanks
133 (n=5-6 fish per tank). All fish were fasted for 24h beforehand to ensure their guts were empty. Fish
134 were acclimated for 4 weeks in their new tanks, during which time they were fed commercial pellets
135 twice daily to a total of 2% of their biomass (Le Gouessant®, Lamballe, France).

136 In February 2018, fish were gradually acclimated to 20°C by means of a sequence of 3 step
137 increases of 2°C over a two-week period. This temperature was chosen because it is the temperature
138 at which whole-organism performance declined in mullet fed on a low n-3 HUFA diet, but is within
139 their natural thermal range as 20°C reflects the mean summer temperature in mullet natural
140 environment (Vagner et al., 2015; Vagner et al., 2014). Fish were left another 2 weeks period of
141 acclimation to initiate physiological adjustment to the change in temperature (Bouchard and
142 Guderley, 2003). Temperature ($19.9 \pm 1^\circ\text{C}$) and salinity (29.6 ± 1.1) were daily measured in the four
143 experimental tanks.

144

145 *2.2. Diet treatment and tissue sampling*

146 Differences in dietary n-3 HUFA content were achieved by replacing fish oil (rich in n-3 HUFA) of the
147 High n-3 HUFA diet with rapeseed oil (poor in n-3 HUFA) in the Low n-3 HUFA diet (See ingredients in

148 Table S1 (a)). Experimental diets were isocaloric and isolipidic. High and Low n-3 HUFA diets
149 contained 17.5 % and 1.2 % EPA+DHA, respectively, per total fatty acid mass of the diets, which
150 represents about 15-fold difference in EPA and DHA content between diets (See Table S1 (a) for fatty
151 acid composition of the diets). The EPA and DHA content in High n-3 HUFA diet cover the needs of
152 several fish species (Robin and Skalli, 2007), **although** the needs of the golden grey mullet are not
153 known. The Low n-3 HUFA diet was estimated to significantly reduce n-3 HUFA content in biological
154 membranes and has been shown to impair aerobic performance of golden grey mullet at 20°C
155 (Vagner et al., 2015).

156 Following the period of acclimation to water temperature of 20°C, fish were switched to the
157 experimental diets for about two months (mean \pm SEM = 61 ± 2 days, range: 45 – 73 days). This
158 duration was chosen because it is sufficient to detect differences in the membrane fatty acid
159 composition between diet treatments (Robin and Skalli, 2007). Because only two fish per day could
160 be analysed for their mitochondrial function at the end of the experiment, the duration of the diet
161 treatment differed between processing batches. Fish were randomly allocated to the treatment: fish
162 had their food progressively switched to either the High n-3 HUFA diet (n = 11) or the Low n-3 HUFA
163 diet (n = 12). The experimental diet to commercial diet ratio was increased every second day from
164 25%:75% to 50%:50%, 75%:25% and finally to 100%:0%. Body mass did not differ between fish
165 groups subsequently assigned to the two food treatments (High n-3 HUFA diet: 31.3 ± 1.8 g; Low n-3
166 HUFA diet: 34.9 ± 1.5 g, *t*-test: $t_{21} = -1.510$, *p* = 0.146). Body mass was re-measured (as above) every
167 2-3 weeks and rations were recalculated to adjust for growth.

168 At the end of the food treatment period, fish were fasted for 24 h before being anesthetized
169 and culled. Fish were weighed, measured and two samples of skeletal muscle were immediately
170 dissected, taken dorsally from the lateral line (to have both red and white muscle) and just behind
171 the head. One aliquot was collected from one side of the fish and kept in ice-cold isolation buffer
172 (sucrose 250 mM, EGTA 1mM, Tris-HCl 20mM, pH 7.4 at 4°C) for immediate mitochondrial assay,
173 while the other aliquot (≈ 1 g) was collected from the other side and immediately flash-frozen for
174 subsequent fatty acid analysis.

175

176 2.3. Determination of fatty acid composition

177 2.3.1. Lipid extraction

178 Lipids from muscle of fish were extracted following the method used in Mathieu-Resuge et al. (2019).
179 Muscle aliquots were ground into a fine homogeneous powder in liquid nitrogen. Lipids from 200-
180 250 mg of muscle powder were then extracted in 6 ml of mixture chloroform/methanol (2:1; v/v). To
181 ensure complete lipid extraction, samples were sonicated at 4 °C during 2 x 5 min. Lipids from
182 experimental diets were extracted following the same method. Pellets were ground in a mortar and

183 powder was transferred in chloroform/methanol (2:1; v/v). All lipid extracts were then stored at -20
184 °C under nitrogen atmosphere until further analysis.

185

186 *2.3.2. Purification of polar lipids*

187 Fatty acids of biological membranes are connected to a polar head group, altogether named polar
188 lipid (PL), unlike fatty acids of energy stores that constitute triglyceride, a neutral lipid. To analyse the
189 FA composition of biological membrane, PL were separated from neutral FA and FA from only PL
190 were considered. PL were separated from neutral one by solid-phase extraction following the
191 method described in (Martin et al., 2013). Briefly, an aliquot of muscle total lipid extract (1/6) was
192 evaporated to dryness under nitrogen, recovered with 3 washings of 0.5 ml of chloroform/methanol
193 (98:2; v/v) and deposited at the top of a silica gel column (40 mm × 4 mm, silica gel 60A 63–200 µm)
194 previously heated at 450°C and deactivated with 6 weight % H₂O. After elution of neutral lipids with
195 10 mL of chloroform/methanol (98:2; v/v), PL were eluted with 20 mL of methanol, transferred into a
196 vial containing 2.3 µg of internal standard (tricosanoic acid C23:0) and stored at -20°C under nitrogen
197 atmosphere.

198

199 *2.3.3. Polar lipid transesterification*

200 Polar lipid transesterification was realized as described in Mathieu-Resuge et al. (2019). PL fractions
201 were evaporated to dryness under nitrogen. Fatty acid methyl esters (FAME) were obtained by
202 addition of 800 µL of H₂SO₄ / methanol (3.4 %; v/v) and heating at 100°C for 10 min. FAME were then
203 extracted in 800 µL of hexane and washed three times with 1.5 ml of distilled water saturated in
204 hexane.

205

206 *2.3.4. Gas chromatography analysis of FAME*

207 FA composition was analysed by GC coupled with Flame-Ionization Detector (GC-FID) as described by
208 Mathieu-Resuge et al. (2019) with a Varian CP8400 gas chromatograph. After splitless-mode
209 injection, FAME were separated in parallel on two different columns (DBWAX 30 m × 0.25 mm ID x
210 0.2 µm and DB5 30 m × 0.25 mm ID x 0.2 µm, Agilent). FAME were identified by comparisons of their
211 retention time with those of commercial standards (Supelco 37 Component FAME Mix, PUFA No.1
212 and No.3, and Bacterial Acid Methyl Ester Mix, Sigma) and lab-made standard mixtures. The internal
213 standard (C23:0) allowed to calculate the FA content (µg mg⁻¹ muscle wet mass). Fatty acid content
214 was then determined as the percentage of fatty acid mass per total mass of fatty acids in membrane
215 lipids from the polar lipids fraction. The sum of EPA and DHA content in membrane lipids was
216 calculated as:

$$\Sigma \text{ EPA + DHA content} = \frac{\text{mass of EPA} + \text{mass of DHA}}{\text{total mass of fatty acids}} \times 100$$

217

218 2.4. Mitochondrial function

219 2.4.1. Isolation of mitochondria

220 Isolation of muscle mitochondria and measurement of mitochondrial efficiency were adjusted from
221 published protocols (Ghanizadeh Kazerouni et al., 2016; Salin et al., 2016c). Briefly, mitochondria
222 were isolated from 1 g (mean \pm SEM: 1.07 \pm 0.03 g) of muscle. Tissue was finely chopped on ice and
223 homogenized in 10 mL of isolation buffer by four gentle passes in a Potter-Elvehjem homogenizer
224 and centrifuged at 500 g for 5 min at 4°C. The supernatant was transferred in a clean tube and
225 centrifuged at 1000 g for 5 min at 4°C and then centrifuged at 9000 g for 10 min at 4°C. The pellet
226 containing mitochondria was gently resuspended in 500 μ L assay buffer at 4°C (20mM Taurine,
227 10mM KH₂PO₄, 20 mM HEPES, 110 mM D-sucrose, 60 mM K-lactobionate, 1g L⁻¹ BSA fatty acid free,
228 pH 7.2 at 20°C).

229

230 2.4.2 Mitochondrial ATP/O ratio and LEAK respiration measurements

231 Mitochondrial efficiency has been quantified through the measurement of the ATP/O ratio, which is
232 the amount of ATP generated per unit of oxygen consumed. Mitochondrial LEAK respiration was
233 determined as the rate of oxygen consumption when ATP synthesis was inhibited. We used a
234 protocol that simultaneously measures both mitochondrial ATP production and the oxygen
235 consumption related to that ATP production, as in Salin et al. (2016c). Assays were conducted at
236 20°C.

237 Oxygen and magnesium green fluorescence signals were detected simultaneously using two
238 respirometry chambers equipped with fluorescent sensors and recorded using DatLab software
239 (Oroboros Instruments, Innsbruck Austria). To estimate ATP production we used the magnesium-
240 sensitive fluorescent probe, Magnesium Green, to determine changes in [Mg²⁺] (Szmackinski and
241 Lakowicz, 1996). ATP production was then calculated from the rate of change in [Mg²⁺] and is based
242 on the unequal affinities of ATP and ADP for Mg²⁺ (Chinopoulos et al., 2009).

243 The oxygen electrodes were calibrated at two points: air-saturated assay buffer (daily) and
244 zero oxygen after sodium dithionite addition (fortnightly). Stepwise additions of MgCl₂ at each run
245 were performed for calibration of the fluorescent signal into Mg²⁺. The two binding affinity (K_d)
246 values of ATP and ADP for Mg²⁺ were determined in presence of isolated mitochondria and calculated
247 as in (Chinopoulos et al., 2014) ; the values were K_{d-ATP} = 0.197 mM and K_{d-ADP} = 1.609 mM.

248 Isolated mitochondria from each fish were added to one of the two measurement chambers
249 of the oxygraph immediately following isolation; fish from a processing pair (i.e. a Low and a High n-3
250 HUFA) were measured in parallel. The remaining part of the isolated mitochondria was preserved on

251 ice for use in a replicate trial. Assays contained 200 μl of isolated mitochondria with 1.8 mL of assay
252 buffer in the presence of complex I substrates (5 mM pyruvate and 0.5 mM malate) and complex II
253 substrate (10 mM succinate). Magnesium green (2.2 μM), EGTA (0.1 mM), EDTA (5 μM) and MgCl_2 (1
254 mM) were added to determine changes in $[\text{Mg}^{2+}]$ and so to calculate the rate of ATP production as in
255 (Chinopoulos et al., 2014).

256 The rate of oxygen consumption to support ATP production was assessed by adding a
257 saturating concentration of ADP (2 mM, Mg^{2+} free) to the chamber. The raw rate of ATP production
258 was also measured in this condition. The rate of oxygen consumption to offset the leakage of proton
259 – LEAK respiration - was then measured after inhibition of mitochondrial ATP synthesis (with 4 μM
260 carboxyatractyloside). The rate of ATP hydrolysis was also measured in this condition. The rate of
261 ATP hydrolysis was then added to the raw rate of ATP production to obtain the corrected rate of ATP
262 production. Addition of complex I inhibitor (0.5 μM rotenone) and complex III inhibitor (2.5 μM
263 antimycin A) allowed determination of residual oxygen consumption, which was then subtracted
264 from all other oxygen consumption values. The second replicated trial was used to control for
265 repeatability of the assay. It was identical to the first one, but started an hour and half later, using
266 the remaining isolated mitochondrial and the same measurement chamber. Every second day, the
267 measurement chamber associated with a treatment group was reversed to control for any inter-
268 respirometry chamber difference in readings. No effect of the choice of measurement chamber on
269 mitochondrial function was detected.

270

271 2.4.3. Analysis of Mitochondrial ATP/O ratio and LEAK respiration

272 We expressed respiration rate as pmoles of $\text{O}_2 \text{ sec}^{-1} \text{ ml}^{-1}$ of assay buffer and ATP production
273 as pmoles of $\text{ATP sec}^{-1} \text{ ml}^{-1}$ of assay buffer for each replicate. Finally, the ATP/O ratio was calculated
274 as the ratio of corrected ATP production to two-fold respiration that supported ATP production; the
275 rate of respiration is doubled since each molecule of oxygen is comprised of two oxygen atoms.
276 Replicated ATP/O ratios were highly correlated (Pearson's $r = 0.749$, $P < 0.001$). However, we found a
277 consistent shift in the values of the measurements between the first and the second trials of ATP/O
278 ratio (drift between trials: Paired $t = -6.194$, $P < 0.001$). The data from the second trial of muscle
279 assay were excluded because the mitochondrial integrity may have been impaired with time post-
280 isolation, as previously found in liver mitochondria (Salin et al., 2016c). Only data of the first LEAK
281 respiration and ATP/O ratio trial were kept for the main analyses. Protein concentration was
282 measured at 595 nm using the Bradford method with bovine serum albumin as a standard. We
283 expressed LEAK respiration as pmoles of $\text{O}_2 \text{ sec}^{-1} \text{ mg}^{-1}$ of mitochondrial protein. See descriptive
284 statistics of the mitochondrial function in Table S2.

285

286 2.5. Statistical analysis

287 We first examined the effect of the treatments (High and Low dietary n-3 HUFA content) on
288 the membrane fatty acid composition (Σ EPA + DHA content). Normality (Shapiro-Wilk test) and
289 homoscedasticity (Bartlett-test) were tested. When normality and homoscedasticity were not
290 satisfied, non-parametric analyses were carried out. We used Wilcoxon tests to determine whether
291 muscle membrane content of EPA and DHA, and their sum (Σ EPA + DHA content) differed between
292 dietary treatment groups. To examine potential differences in final body mass between treatments
293 that could explain differences in mitochondrial function, a *t*-test was employed. The effect of the
294 dietary treatment on mitochondrial function (ATP/O ratio and LEAK respiration) were also analysed
295 using *t*-tests.

296 We then used linear model **analyses** to test whether ATP/O ratio and LEAK respiration were
297 correlated with membrane lipid content in Σ EPA + DHA. The models included mitochondrial function
298 (ATP/O or LEAK respiration) as the dependant variable, dietary treatments as a categorical effect, and
299 Σ EPA + DHA content as continuous predictor. The normality of the linear model residuals was
300 validated for all models using Shapiro–Wilk tests. We also tested the interactions between dietary
301 treatment and membrane lipid content in Σ EPA + DHA. All interactions were non-significant, so they
302 were removed from the models.

303 The duration of the dietary treatment **was included as potential covariate**, the processing
304 batch and fish tanks were included as potential **random factor** in **initial analysis**. **Each of these**
305 **variable** were not significant, so were removed from all the **final models**. All statistical analyses were
306 performed with the free software R (R Core Team, 2017), with R Version 3.6.1, with significance level
307 set to $p < 0.05$. Data are presented as means \pm standard error of the mean (SEM).

308

309 3. Results

310 3.1. Effect of the dietary treatment on membrane fatty acid composition

311 Muscle of mullets fed on Low n-3 HUFA diet showed significant differences in their membrane
312 fatty acid composition compared to those of fish fed on High n-3 HUFA diet (Table S3). EPA and DHA
313 content in muscle membranes were significantly lower in Low n-3 HUFA fish compared to High n-3
314 HUFA fish (EPA: $W = 119$, $p < 0.001$; DHA: $W = 132$, $p < 0.001$; Table 1). Not surprisingly, fish on
315 average had a lower content of Σ EPA+ DHA in their muscle when fed the Low n-3 HUFA diet ($W =$
316 130 , $p < 0.001$, Table 1). Final body mass did not differ between High and Low n-3 HUFA fish ($t = -$
317 1.668 ; $p = 0.112$), excluding body mass as confounding sources of differences between dietary
318 treatments.

319

320 3.2. Effect of the dietary treatment on mitochondrial function

321 ATP/O ratio was significantly higher in the mitochondria of Low compared to High n-3 HUFA
322 mullet ($t = -3.107$, $p = 0.005$; Fig. 1A). However, there was no effect of dietary n-3 HUFA content on
323 LEAK respiration in muscle mitochondria ($t = -0.423$, $p = 0.677$; Fig. 1B).

324

325 *3.3. Relationships across individuals between mitochondrial function and membrane fatty acid* 326 *composition*

327 The ATP/O ratio ranged from 1.46 to 3.33 for mitochondria of mullet eating the High n-3
328 HUFA diet and from 2.05 to 3.67 for mitochondria of fish on Low n-3 HUFA diet (Fig. 2A). **Variation in**
329 **mitochondrial function between individuals was mainly explained by differences in n-3 HUFA content**
330 **in muscle (Figure S1)**. Regardless of the food treatment, the muscle ATP/O ratio of a fish was strongly
331 and positively related to its Σ EPA + DHA content: individuals that had the higher mitochondrial
332 efficiency under either diet had the highest content in EPA and DHA in their membranes ($t = 2.513$, p
333 $= 0.021$; Fig. 2A and Table 2). There was no relationship between a fish's mitochondrial LEAK
334 respiration and its Σ EPA + DHA content among individuals with the same dietary treatment ($t = -$
335 0.104 , $p = 0.919$, Fig. 2B and Table 2). We found no correlation between ATP/O ratio and LEAK
336 respiration from the same mitochondria (High n-3 HUFA diet: Pearson's $r = 0.229$, $p = 0.499$; Low n-3
337 HUFA diet: Pearson's $r = -0.157$, $p = 0.627$).

338

339 **4. Discussion**

340 We asked whether a decline in dietary n-3 HUFA content leads to changes in mitochondrial
341 metabolic phenotype for a model of primary consumer fish, the golden grey mullet. We manipulated
342 dietary content in n-3 HUFA and assessed membrane FA composition and mitochondrial function in
343 mullet. We found that diet strongly influenced membrane fatty acid composition: mullets on a Low
344 n-3 HUFA diet had lower levels of n-3 HUFA in muscle biological membranes, which suggested that
345 the mitochondrial membranes contained less n-3 HUFA. Previous studies have indeed demonstrated
346 that modification of the fatty acid composition of the diet causes strongly correlated changes in the
347 membrane fatty acid composition of mitochondria (Herbst et al., 2014; Jeromson and Hunter, 2014;
348 Ramsey et al., 2005), including those of fish species (Guderley et al., 2008; Morash et al., 2009). Our
349 findings reveal that ATP/O ratio increased significantly in mitochondria of fish fed a low n-3 HUFA
350 diet, so that a decline in dietary content in n-3 HUFA elicits greater mitochondrial efficiency to
351 produce ATP. **Surprisingly, the differences in membrane n-3 HUFA content between individuals had**
352 **an opposite effect on mitochondrial efficiency**: under the same dietary treatment, **individuals** that
353 displayed higher EPA and DHA **membrane content** had mitochondria with the highest ATP/O ratio.

354 Aerobic performance of marine fish will not only be challenged in a near future by increased
355 oxygen needs and low oxygen availability at high temperatures but probably also by a shift in ocean

356 productivity (Behrenfeld et al., 2006), where EPA and DHA availability might decline considerably
357 (Hixson and Arts, 2016). Mitochondrial aerobic capacity has traditionally been determined in terms
358 of oxygen consumption, enzymatic activities or density of mitochondria (Brookes et al., 1998;
359 Guderley et al., 2008; Martin et al., 2013; Morash et al., 2009; Ramsey et al., 2005; Yu et al., 2014).
360 However, oxygen consumption provides only an indirect measure of ATP production, and the
361 relationship between oxygen consumption and ATP production can vary both among and within
362 individuals. Our results clearly show that when evaluating mitochondrial adjustment in response to a
363 reduction in n-3 HUFA availability in diet, variation in the amount of ATP generated per molecule of
364 oxygen consumed by mitochondria can have major importance in explaining associated changes in
365 whole-animal performance. Past work on mullets found that fish fed on a low n-3 HUFA diet
366 consumed less oxygen compared to the fish fed a high n-3 diet to reach a similar swimming speed
367 (Vagner et al., 2015; Vagner et al., 2014). Our results suggest that higher mitochondrial efficiency
368 may result in lower oxygen requirement for fish to produce ATP and sustain muscle contraction for
369 locomotory performance, as previously demonstrated in human (Coen et al., 2012). Mitochondria
370 with high efficiency can be beneficial for energy-demanding cellular processes (Salin et al., 2015), yet
371 there could also be a cost. Mitochondria are a major producer of reactive oxygen species (ROS) and
372 mitochondrial efficiency can be positively related to ROS production (Brand, 2000; Salin et al., 2015),
373 an area which would require further study.

374 Here, we found no significant differences in LEAK respiration of mullet muscle across dietary
375 treatments. In contrast, LEAK respiration in trout muscle were higher for mitochondria from fish fed
376 a low n-3 HUFA diet than for mitochondria from fish fed a high n-3 HUFA diet (Guderley et al., 2008).
377 Another experiment in trout has shown the opposite effect: animals fed a lower n-3 HUFA diet
378 displayed lower LEAK respiration in muscle mitochondria (Martin et al., 2013). We stress that it is
379 entirely possible for the mitochondrial LEAK response to dietary content in n-3 HUFA to vary among
380 individuals, species and other environmental factors, but also that the consequence on the proton
381 leakage might depend on the magnitude and duration of the dietary treatment. Equivocal support
382 may indicate that the responses of LEAK to changes in membrane fatty acid composition does not fall
383 along a linear response curve, as typically assumed. Instead, the effect of dietary content in n-3 HUFA
384 on membrane composition, and in turn proton permeability might be biphasic (Abbott et al., 2010)
385 and might promote non-linear response of LEAK. While testing a large range of n-3 HUFA dietary
386 content was beyond the scope of the present study, it may only be possible to explain contrasted
387 pattern of LEAK response if many dietary levels of n-3 HUFA are considered.

388 In our study, mullets from the same food treatment displayed important individual variation
389 in mitochondrial efficiency that covary with membrane fatty acid composition: individual that had
390 higher EPA and DHA content in their membranes had mitochondria that synthesized ATP more

391 efficiently than those of individuals with lower EPA and DHA content. Previous studies suggested that
392 variation in FA composition of the mitochondrial inner membrane can influence the mitochondrial
393 membrane conductance of protons, and in turn the proportion of oxygen used to offset the proton
394 leakage (Hulbert et al., 2007), leading to variation in the efficiency of ATP production (Brand, 2005).
395 However, this does not appear to be the case in the present study, as LEAK did not covary with
396 ATP/O ratio and membrane fatty acid composition. An explanation for this discrepancy might lie in
397 the fact that mitochondrial efficiency also depends on protein abundance and activity of the
398 mitochondrial inner membrane-bound complexes involved in oxidative phosphorylation (i.e. electron
399 transport complexes and ATP synthase). This alternative explanation is based on the fact that fatty
400 acid composition of membrane lipids may affect lipid–protein interactions and therefore the function
401 of embedded proteins (Brenner, 1984). As well as varying with membrane FA composition and
402 protein function, mitochondrial ATP/O ratio can shift in response to energy substrate use (Brand,
403 2005). Previous work in Atlantic salmon has demonstrated that individual variation in DHA and EPA in
404 muscle content was associated with differences in the expression of genes involved in lipid
405 catabolism and carbohydrate metabolism (Horn et al., 2019). However, further research is needed to
406 determine whether differences in energy substrates utilization determine individual variation in
407 mitochondrial efficiency in mullet.

408 Some mullets on the low n-3 HUFA dietary treatment had actually higher EPA and DHA
409 contents than others on the high n-3 HUFA dietary treatment that were eating almost 15 times as
410 much EPA and DHA. Individual variation in membrane fatty acid composition is likely to be a
411 complex, integrative characteristics influenced by several metabolic pathways. If individual
412 differences in fatty acid membrane composition covary with the rates of lipid assimilation,
413 biosynthesis and degradation, it is important to recognize that fatty acid composition may vary with
414 pathways that are generally neglected. For example, ability to biosynthesis EPA and DHA in marine
415 fish is very limited, as it is generally insufficient to compensate dietary deficiency (Tocher, 2003), but
416 the link between individual variation in n-3 HUFA content and their rate of biosynthesis might only
417 appear across individuals eating the same amount of n-3 HUFA. Another explanation might be an
418 individual heterogeneity in the rate of degradation of EPA and DHA, while also considered minor in
419 fish (McKenzie, 2001). Individual variation in the rate of assimilation may also be significant for n-3
420 HUFA content in membrane. Regardless of their food intake, individual fish that have higher levels of
421 EPA and DHA in membrane lipids may have preferentially retained these fatty acids from their diet.
422 Our observations illustrate that an understanding of individual variability in membrane fatty acid
423 composition can be gained only through consideration of differential metabolic pathways of fatty
424 acids.

425 The individual covariation between mitochondrial efficiency and membrane fatty acid
426 composition we observed might be because higher EPA and DHA contents in the membrane promote
427 higher efficiency of the mitochondria, as explained above. However, given that our study is
428 correlative, it is perhaps high mitochondrial efficiency that may promote membranes with relatively
429 high levels of EPA and DHA, because more ATP is available to fuel metabolic pathways that retain
430 dietary n-3 HUFA. Assimilation of fatty acids in the intestine can be energetically costly (Mansbach
431 and Gorelick, 2007), and for example, fish that have higher efficiency to make ATP may actually have
432 higher assimilation rate of EPA and DHA compared with fish at lower mitochondrial efficiency. The
433 variation in mitochondrial efficiency of muscle tissue studied here might be representative of the
434 mitochondrial efficiency in other tissues, including intestine, although previous studies looking at
435 correlation of mitochondrial efficiency across tissues in the same individual have shown equivocal
436 results (Salin et al., 2019). Further study looking at mitochondrial metabolism in different tissues will
437 be necessary to determine if golden grey mullets are able to improve their mitochondrial efficiency
438 across multiple tissues.

439 Flexibility in mitochondrial efficiency may be particularly important since the capacity of the
440 mitochondria to produce ATP can set limits on the capacity of an organism to respond to
441 environmental changes (Blier et al., 2013; Sokolova et al., 2012). Our data imply that the greater
442 mitochondrial efficiency induced by the Low n-3 HUFA diet might benefit for the ability of the
443 mitochondrial to make ATP for energy-demanding cellular processes when reduction in EPA and DHA
444 availability in food web. Information on the consequences of individual heterogeneity in
445 mitochondrial efficiency on fish performance would allow a better understanding of the effect of
446 decline in EPA and DHA availability in marine food web. Further research should focus on identifying
447 the individual variation in metabolic pathway of EPA and DHA. This type of variation may be very
448 important in an evolutionary context as well, not only generating phenotypic variation among
449 individuals, but also allowing animals to reduce the energy costs of making ATP by increasing the
450 mitochondrial efficiency.

451
452 ACKNOWLEDGEMENTS: We thank Salomé Ducos for help with fish handling and José-Luis Zambonino
453 Infante for designing the diets. We thank the three referees for helpful comments that improved the
454 manuscript.

455
456 [CRediT author statement](#): KS, MMR, FLG, PS and MV conceived the ideas and designed methodology.
457 KS, MMR, NG, ED, FLG and MV collected the data. KS, MMR, NG, FLG, PS and MV analysed the data.
458 KS led the writing of the manuscript except for section 2.3 written by FLG; MMR, FLG, PS and MV
459 revised the manuscript and added comments. All authors gave final approval for publication.

460

461 DECLARATION OF INTEREST: The authors declare that they have no known competing financial
462 interests or personal relationships that could have appeared to influence the work reported in this
463 paper.

464

465 DATA ACCESSIBILITY: The dataset supporting this article will be made available by archiving it in the
466 Dryad Digital Repository.

467

468 FUNDING: This work is a contribution to the project ECONAT Axe 1 - Ressources Marines Littorales:
469 qualité et éco-valorisation, funded by the Contrat de Plan Etat-Région and the CNRS and the
470 European Regional Development Fund.

471

472 **References**

473 Abbott, S.K., Else, P.L., Hulbert, A.J., 2010. Membrane fatty acid composition of rat skeletal muscle is
474 most responsive to the balance of dietary n-3 and n-6 PUFA. *British Journal of Nutrition* 103, 522-
475 529.

476 Alimuddin, Yoshizaki, G., Kiron, V., Satoh, S., Takeuchi, T., 2005. Enhancement of EPA and DHA
477 biosynthesis by over-expression of masu salmon Delta 6-desaturase-like gene in zebrafish. *Transgenic*
478 *Res.* 14, 159-165.

479 Arts, M.T., Kohler, C.C., 2009. Health and condition in fish: the influence of lipids on membrane
480 competency and immune response, in: Kainz, M., Brett, M.T., Arts, M.T. (Eds.), *Lipids in Aquatic*
481 *Ecosystems*. Springer New York, New York, NY, pp. 237-256.

482 Behrenfeld, M.J., O'Malley, R.T., Siegel, D.A., McClain, C.R., Sarmiento, J.L., Feldman, G.C., Milligan,
483 A.J., Falkowski, P.G., Letelier, R.M., Boss, E.S., 2006. Climate-driven trends in contemporary ocean
484 productivity. *Nature* 444, 752-755.

485 Bermudez, R., Feng, Y., Roleda, M.Y., Tatters, A.O., Hutchins, D.A., Larsen, T., Boyd, P.W., Hurd, C.L.,
486 Riebesell, U., Winder, M., 2015. Long-term conditioning to elevated pCO₂ and warming influences the
487 fatty and amino acid composition of the diatom *Cylindrotheca fusiformis*. *Plos One* 10.

488 Blier, P.U., Lemieux, H., Pichaud, N., 2013. Holding our breath in our modern world: will mitochondria
489 keep the pace with climate changes? *Canadian Journal of Zoology* 92, 591-601.

490 Bottje, W.G., Carstens, G.E., 2009. Association of mitochondrial function and feed efficiency in
491 poultry and livestock species. *Journal of Animal Science* 87, E48-E63.

492 Bouchard, P., Guderley, H., 2003. Time course of the response of mitochondria from oxidative muscle
493 during thermal acclimation of rainbow trout, *Oncorhynchus mykiss*. *J. Exp. Biol.* 206, 3455-3465.

494 Brand, M.D., 2000. Uncoupling to survive? The role of mitochondrial inefficiency in ageing.
495 *Experimental Gerontology* 35, 811-820.

496 Brand, M.D., 2005. The efficiency and plasticity of mitochondrial energy transduction. *Biochemical*
497 *Society Transactions* 33, 897-904.

498 Brand, M.D., Couture, P., Hulbert, A.J., 1994. Liposomes from mammalian liver mitochondria are
499 more polyunsaturated and leakier to protons than those from reptiles. *Comparative Biochemistry*
500 *and Physiology Part B: Comparative Biochemistry* 108, 181-188.

501 Brand, M.D., Nicholls, D.G., 2011. Assessing mitochondrial dysfunction in cells. *Biochemical Journal*
502 435, 297-312.

503 Brenner, R.R., 1984. Effect of unsaturated-acids on membrane-structure and enzyme-kinetics.
504 *Progress in Lipid Research* 23, 69-96.

505 Brookes, P.S., Buckingham, J.A., Tenreiro, A.M., Hulbert, A.J., Brand, M.D., 1998. The proton
506 permeability of the inner membrane of liver mitochondria from ectothermic and endothermic
507 vertebrates and from obese rats: correlations with standard metabolic rate and phospholipid fatty
508 acid composition. *Comparative Biochemistry and Physiology B-Biochemistry & Molecular Biology*
509 *119*, 325-334.

510 Chinopoulos, C., Kiss, G., Kawamata, H., Starkov, A.A., 2014. Chapter seventeen - Measurement of
511 ADP-ATP exchange in relation to mitochondrial transmembrane potential and oxygen consumption,
512 in: Lorenzo, G., Guido, K. (Eds.), *Methods in Enzymology*, Elsevier academic press inc, San Diego, pp.
513 333-348.

514 Chinopoulos, C., Vajda, S., Csanády, L., Mándi, M., Mathe, K., Adam-Vizi, V., 2009. A novel kinetic
515 assay of mitochondrial ATP-ADP exchange rate mediated by the ANT. *Biophysical Journal* *96*, 2490-
516 2504.

517 Coen, P.M., Jubrias, S.A., Distefano, G., Amati, F., Mackey, D.C., Glynn, N.W., Manini, T.M.,
518 Wohlgenuth, S.E., Leeuwenburgh, C., Cummings, S.R., Newman, A.B., Ferrucci, L., Toledo, F.G.S.,
519 Shankland, E., Conley, K.E., Goodpaster, B.H., 2012. Skeletal muscle mitochondrial energetics are
520 associated with maximal aerobic capacity and walking speed in older adults. *The Journals of*
521 *Gerontology Series A: Biological Sciences and Medical Sciences* *68*, 447-555.

522 da Motta Pacheco, L.F.C., Uribe, E., Pino, J., Troncoso, J., Quiróz, A., 2014. The effect of UV light and
523 CO₂ in the production of polyunsaturated aldehydes in *Skeletonema costatum* (Bacillariophyceae).
524 *American Journal of Plant Sciences* *5*, 3632.

525 Distefano, G., Standley, R.A., Zhang, X.L., Carnero, E.A., Yi, F., Cornell, H.H., Coen, P.M., 2018.
526 Physical activity unveils the relationship between mitochondrial energetics, muscle quality, and
527 physical function in older adults. *J. Cachexia Sarcopenia Muscle* *9*, 279-294.

528 Fontaine, E.M., Moussa, M., Devin, A., Garcia, J., Ghisolfi, J., Rigoulet, M., Leverve, X.M., 1996. Effect
529 of polyunsaturated fatty acids deficiency on oxidative phosphorylation in rat liver mitochondria.
530 *Biochim. Biophys. Acta-Bioenerg.* *1276*, 181-187.

531 Galloway, A.W.E., Winder, M., 2015. Partitioning the Relative Importance of Phylogeny and
532 Environmental Conditions on Phytoplankton Fatty Acids. *PLOS ONE* *10*, e0130053.

533 Ghanizadeh Kazerouni, E., Franklin, C.E., Seebacher, F., 2016. UV-B exposure reduces locomotor
534 performance by impairing muscle function but not mitochondrial ATP production. *J. Exp. Biol.* *219*,
535 96-102.

536 Guderley, H., Kraffe, E., Bureau, W., Bureau, D.P., 2008. Dietary fatty acid composition changes
537 mitochondrial phospholipids and oxidative capacities in rainbow trout red muscle. *Journal of*
538 *Comparative Physiology B-Biochemical Systemic and Environmental Physiology* *178*, 385-399.

539 Hamel, S., Gaillard, J.-M., Yoccoz, N.G., 2018. Introduction to: Individual heterogeneity – the causes
540 and consequences of a fundamental biological process. *Oikos* *127*, 643-647.

541 Herbst, E.A.F., Paglialunga, S., Gerling, C., Whitfield, J., Mukai, K., Chabowski, A., Heigenhauser, G.J.F.,
542 Spriet, L.L., Holloway, G.P., 2014. Omega-3 supplementation alters mitochondrial membrane
543 composition and respiration kinetics in human skeletal muscle. *The Journal of Physiology* *592*, 1341-
544 1352.

545 Hixson, S.M., Arts, M.T., 2016. Climate warming is predicted to reduce omega-3, long-chain,
546 polyunsaturated fatty acid production in phytoplankton. *Global Change Biology* *22*, 2744-2755.

547 Horn, S.S., Sonesson, A.K., Krasnov, A., Moghadam, H., Hillestad, B., Meuwissen, T.H.E., Ruyter, B.,
548 2019. Individual differences in EPA and DHA content of Atlantic salmon are associated with gene
549 expression of key metabolic processes. *Scientific Reports* *9*.

550 Hulbert, A.J., Pamplona, R., Buffenstein, R., Buttemer, W.A., 2007. Life and death: metabolic rate,
551 membrane composition, and life span of animals. *Physiol Rev* *87*, 1175-1213.

552 Hulbert, A.J., Turner, N., Storlien, L.H., Else, P.L., 2005. Dietary fats and membrane function:
553 implications for metabolism and disease. *Biological Reviews* *80*, 155-169.

554 Ishizaki, Y., Masuda, R., Uematsu, K., Shimizu, K., Arimoto, M., Takeuchi, T., 2001. The effect of
555 dietary docosahexaenoic acid on schooling behaviour and brain development in larval yellowtail.
556 *Journal of Fish Biology* *58*, 1691-1703.

557 Jahn, M., Seebacher, F., 2019. Cost of transport is a repeatable trait but is not determined by
558 mitochondrial efficiency in zebrafish (*Danio rerio*). The Journal of Experimental Biology 222,
559 jeb201400.

560 Jeromson, S., Hunter, D.J., 2014. Influencing mitochondrial membrane composition and
561 bioenergetics through omega-3 supplementation. The Journal of Physiology 592, 1913-1914.

562 Kadenbach, B., 2003. Intrinsic and extrinsic uncoupling of oxidative phosphorylation. Biochim.
563 Biophys. Acta-Bioenerg. 1604, 77-94.

564 Kang, J.X., 2011. Omega-3: A link between global climate change and human health. Biotechnology
565 Advances 29, 388-390.

566 Kraffe, E., Marty, Y., Guderley, H., 2007. Changes in mitochondrial oxidative capacities during thermal
567 acclimation of rainbow trout *Oncorhynchus mykiss*: roles of membrane proteins, phospholipids and
568 their fatty acid compositions. J. Exp. Biol. 210, 149-165.

569 Lebreton, B., Richard, P., Parlier, E.P., Guillou, G., Blanchard, G.F., 2011. Trophic ecology of mullets
570 during their spring migration in a European saltmarsh: A stable isotope study. Estuarine Coastal and
571 Shelf Science 91, 502-510.

572 Mansbach, C.M., II, Gorelick, F., 2007. Development and Physiological Regulation of Intestinal Lipid
573 Absorption. II. Dietary lipid absorption, complex lipid synthesis, and the intracellular packaging and
574 secretion of chylomicrons. American Journal of Physiology-Gastrointestinal and Liver Physiology 293,
575 G645-G650.

576 Martin, N., Bureau, D.P., Marty, Y., Kraffe, E., Guderley, H., 2013. Dietary lipid quality and
577 mitochondrial membrane composition in trout: responses of membrane enzymes and oxidative
578 capacities. Journal of Comparative Physiology B-Biochemical Systemic and Environmental Physiology
579 183, 393-408.

580 Mathieu-Resuge, M., Kraffe, E., Le Grand, F., Boens, A., Bideau, A., Lluch-Cota, S.E., Racotta, I.S.,
581 Schaal, G., 2019. Trophic ecology of suspension-feeding bivalves inhabiting a north-eastern Pacific
582 coastal lagoon: Comparison of different biomarkers. Marine Environmental Research 145, 155-163.

583 Mazon, C., Bruce, M., Bell, J.G., Davie, A., Alorend, E., Jordan, N., Rees, J., Papanikos, N., Porter, M.,
584 Bromage, N., 2003. Dietary lipid enhancement of broodstock reproductive performance and egg and
585 larval quality in Atlantic halibut (*Hippoglossus hippoglossus*). Aquaculture 227, 21-33.

586 McIntyre, P.B., Reidy Liermann, C.A., Revenga, C., 2016. Linking freshwater fishery management to
587 global food security and biodiversity conservation. Proceedings of the National Academy of Sciences
588 113, 12880-12885.

589 McKenzie, D.J., 2001. Effects of dietary fatty acids on the respiratory and cardiovascular physiology of
590 fish. Comparative Biochemistry and Physiology a-Molecular & Integrative Physiology 128, 607-621.

591 McKenzie, D.J., Higgs, D.A., Dosanjh, B.S., Deacon, G., Randall, D.J., 1998. Dietary fatty acid
592 composition influences swimming performance in Atlantic salmon (*Salmo salar*) in seawater. Fish
593 Physiol. Biochem. 19, 111-122.

594 Morash, A.J., Bureau, D.P., McClelland, G.B., 2009. Effects of dietary fatty acid composition on the
595 regulation of carnitine palmitoyltransferase (CPT) I in rainbow trout (*Oncorhynchus mykiss*).
596 Comparative Biochemistry and Physiology B-Biochemistry & Molecular Biology 152, 85-93.

597 Mourente, G., Tocher, D.R., 1993. Incorporation and metabolism of C-14-labeled polyunsaturated
598 fatty-acids in wild-caught juveniles of golden gray mullet, *liza-aurata*, *in vivo*. Fish Physiol. Biochem.
599 12, 119-130.

600 Nogueira, V., Piquet, M.A., Devin, A., Fiore, C., Fontaine, E., Brandolin, G., Rigoulet, M., Leverve, X.M.,
601 2001. Mitochondrial adaptation to *in vivo* polyunsaturated fatty acid deficiency: Increase in
602 phosphorylation efficiency. Journal of Bioenergetics and Biomembranes 33, 53-61.

603 Norambuena, F., Morais, S., Emery, J.A., Turchini, G.M., 2015. Arachidonic acid and eicosapentaenoic
604 acid metabolism in juvenile *Atlantic salmon* as affected by water temperature. Plos One 10, 25.

605 Oboh, A., Kabeya, N., Carmona-Antoñanzas, G., Castro, L.F.C., Dick, J.R., Tocher, D.R., Monroig, O.,
606 2017. Two alternative pathways for docosahexaenoic acid (DHA, 22:6n-3) biosynthesis are
607 widespread among teleost fish. Scientific Reports 7, 3889.

608 Pörtner, H.O., Knust, R., 2007. Climate change affects marine fishes through the oxygen limitation of
609 thermal tolerance. Science 315, 95-97.

610 Ramsey, J.J., Harper, M.E., Humble, S.J., Koomson, E.K., Ram, J.J., Bevilacqua, L., Hagopian, K., 2005.
611 Influence of mitochondrial membrane fatty acid composition on proton leak and H₂O₂ production in
612 liver. *Comparative Biochemistry and Physiology B-Biochemistry & Molecular Biology* 140, 99-108.
613 Robert, K.A., Bronikowski, A.M., 2010. Evolution of senescence in nature: Physiological evolution in
614 populations of garter snake with divergent life histories. *American Naturalist* 175, E47-159.
615 Robin, J.H., Skalli, A., 2007. Incorporation of dietary fatty acid in European sea bass (*Dicentrarchus*
616 *labrax*) - A methodological approach evidencing losses of highly unsaturated fatty acids. *Aquaculture*
617 263, 227-237.
618 Salin, K., Auer, S.K., Anderson, G.J., Selman, C., Metcalfe, N.B., 2016a. Inadequate food intake at high
619 temperatures is related to depressed mitochondrial respiratory capacity. *J. Exp. Biol.* 219, 1356-1362.
620 Salin, K., Auer, S.K., Rey, B., Selman, C., Metcalfe, N.B., 2015. Variation in the link between oxygen
621 consumption and ATP production, and its relevance for animal performance. *Proc. R. Soc. B-Biol. Sci.*
622 282, 20151028.
623 Salin, K., Auer, S.K., Rudolf, A.M., Anderson, G.J., Selman, C., Metcalfe, N.B., 2016b. Variation in
624 metabolic rate among individuals is related to tissue-specific differences in mitochondrial leak
625 respiration. *Physiological and Biochemical Zoology* 89, 511-523.
626 Salin, K., Luquet, E., Rey, B., Roussel, D., Voituron, Y., 2012. Alteration of mitochondrial efficiency
627 affects oxidative balance, development and growth in frog (*Rana temporaria*) tadpoles. *J. Exp. Biol.*
628 215, 863-869.
629 Salin, K., Villasevil, E.M., Anderson, G.J., Auer, S.K., Selman, C., Hartley, R.C., Mullen, W., Chinopoulos,
630 C., Metcalfe, N.B., 2018. Decreased mitochondrial metabolic requirements in fasting animals carry an
631 oxidative cost. *Functional Ecology* 32, 2149-2157.
632 Salin, K., Villasevil, E.M., Auer, S.K., Anderson, G.J., Selman, C., Metcalfe, N.B., Chinopoulos, C., 2016c.
633 Simultaneous measurement of mitochondrial respiration and ATP production in tissue homogenates
634 and calculation of effective P/O ratios. *Physiological Reports* 4, e13007.
635 Salin, K., Villasevil Eugenia, M., Anderson Graeme, J., Lamarre Simon, G., Melanson Chloé, A.,
636 McCarthy, I., Selman, C., Metcalfe Neil, B., 2019. Differences in mitochondrial efficiency explain
637 individual variation in growth performance. *Proceedings of the Royal Society B: Biological Sciences*
638 286, 20191466.
639 Sokolova, I.M., Frederich, M., Bagwe, R., Lannig, G., Sukhotin, A.A., 2012. Energy homeostasis as an
640 integrative tool for assessing limits of environmental stress tolerance in aquatic invertebrates.
641 *Marine Environmental Research* 79, 1-15.
642 Szmajcinski, H., Lakowicz, J.R., 1996. Fluorescence lifetime characterization of magnesium probes:
643 Improvement of Mg²⁺ dynamic range and sensitivity using phase-modulation fluorometry. *Journal of*
644 *Fluorescence* 6, 83-95.
645 Team, R.C., 2017. R: A language and Environment for Statistical, 3.6.1. ed. R Foundation for Statistical
646 Computing, Vienna, Austria.
647 Tocher, D.R., 2003. Metabolism and functions of lipids and fatty acids in teleost fish. *Reviews in*
648 *Fisheries Science* 11, 107-184.
649 Vagner, M., Lacoue-Labarthe, T., Zambonino-Infante, J.-L., Mazurais, D., Dubillot, E., Le Delliou, H.,
650 Quazuguel, P., Lefrançois, C., 2015. Depletion of essential fatty acids in the food source affects
651 aerobic capacities of the golden grey mullet *Liza aurata* in a warming seawater context. *Plos One* 10,
652 19.
653 Vagner, M., Zambonino-Infante, J.-L., Mazurais, D., Imbert-Auvray, N., Ouillon, N., Dubillot, E., Le
654 Delliou, H., Akbar, D., Lefrançois, C., 2014. Reduced n-3 highly unsaturated fatty acids dietary content
655 expected with global change reduces the metabolic capacity of the golden grey mullet. *Marine*
656 *Biology* 161, 2547-2562.
657 Yu, L., Fink, B., Herlein, J., Oltman, C., Lamping, K., Sivitz, W., 2014. Dietary fat, fatty acid saturation
658 and mitochondrial bioenergetics. *Journal of Bioenergetics and Biomembranes* 46, 33-44.
659

660 **Table 1:** Mean \pm SEM eicosapentaenoic acid (EPA), docosahexaenoic acid (DHA) content and sum of
 661 EPA and DHA content (Σ EPA + DHA) in muscle membrane fatty acids (percentage of the EPA and DHA
 662 mass per total fatty acids mass in membrane, %), of juvenile golden grey mullet given High (n=11) or
 663 Low (n=12) omega-3 highly unsaturated fatty acids (n-3 HUFA) diet. Different letters within a row
 664 indicate significant differences between diet treatments (Wilcoxon tests, at the significant level $p <$
 665 0.05). More details on FA composition of muscle membranes are provided Table S3.

666

<i>Muscle membrane FA composition (%)</i>	High n-3 HUFA fish	Low n-3 HUFA fish
<i>EPA</i>	9.8 \pm 0.3 ^a	5.7 \pm 1.0 ^b
<i>DHA</i>	20.8 \pm 0.7 ^a	10.1 \pm 2.0 ^b
Σ EPA + DHA	30.6 \pm 0.8 ^a	15.8 \pm 3.0 ^b

667

668

669 **Table 2:** Results from linear model analyses of muscle mitochondrial efficiency (ATP/O ratio) and
 670 respiratory capacities to offset the proton leak (LEAK respiration) in juvenile golden grey mullet as a
 671 function of the sum of eicosapentaenoic acid (EPA), docosahexaenoic acid (DHA) content - Σ EPA +
 672 DHA in muscle (percentage of the EPA and DHA mass per total fatty acid mass in membrane). Bold
 673 denotes significant results.

674

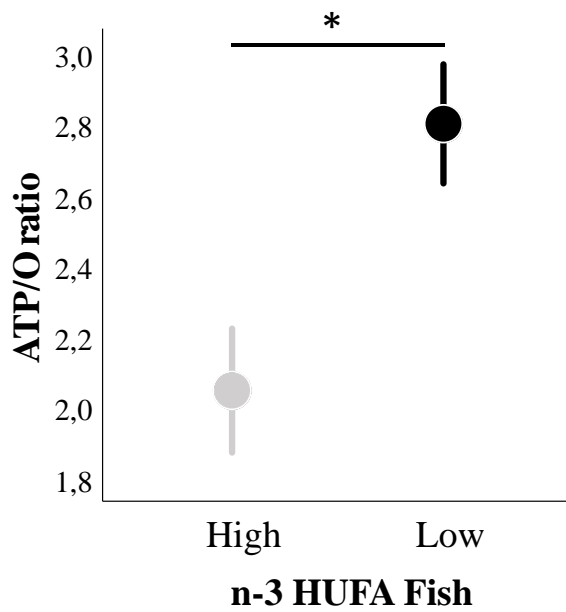
<i>Dependant variable</i>	<i>Source of variation</i>	<i>Parameter estimate \pm SEM</i>	<i>t-value</i>	<i>p-value</i>
ATP/O ratio	Intercept	2.21 \pm 0.23	7.978	< 0.001
	Σ EPA + DHA	0.04 \pm 0.01	2.513	0.021
	diet	1.30 \pm 0.31	4.232	< 0.001
LEAK respiration	Intercept	56.59 \pm 6.45	8.769	<0.001
	Σ EPA + DHA	0.04 \pm 0.34	-0.104	0.919
	diet	2.61 \pm 7.17	0.364	0.720

675

676 FIGURES

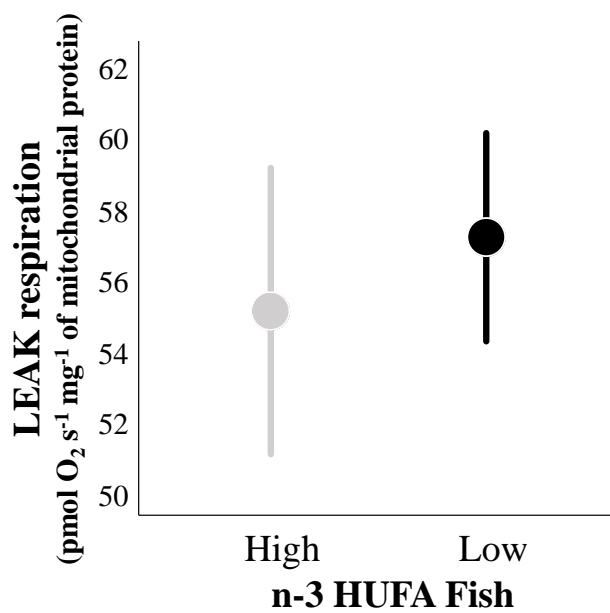
677 **Figure 1.** Effect of omega-3 highly unsaturated fatty acids (n-3 HUFA) dietary content on (a)
678 mitochondrial efficiency estimated as ATP/O ratio ($t_{21} = -3.107$, $p = 0.005$) and (b) mitochondrial LEAK
679 respiration ($t_{21} = -0.423$, $p = 0.677$) of muscle of golden grey mullet that were kept either on High
680 (n=11) or Low (n=12) n-3 HUFA diet. Data are plotted as mean \pm SEM. * denotes significant effect.

681 (a)



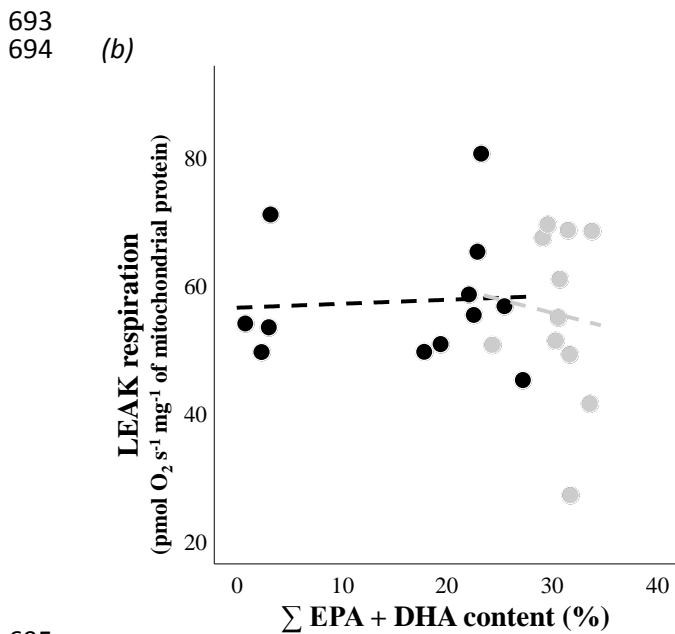
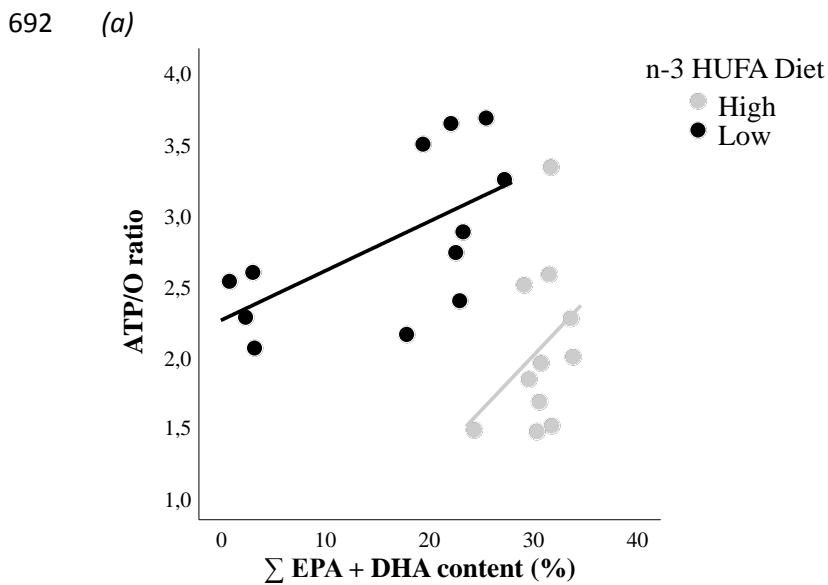
682

683 (b)



684

685 **Figure 2.** Relationships between the mitochondrial function and membrane lipid content in
 686 eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA) content (expressed as Σ EPA + DHA
 687 content, percentage of the sum of EPA and DHA mass per total fatty acid mass in membranes) of
 688 muscle of juvenile golden grey mullet fed on a Low (n=12) versus High (n=11) omega-3 highly
 689 unsaturated fatty acid (n-3 HUFA) diet. Mitochondrial efficiency (ATP/O ratio) in relation to (a) Σ EPA
 690 + DHA content, (b) LEAK respiration in relation to Σ EPA + DHA content. Continuous lines show
 691 significant effects, dashed lines show non-significant effects (see Table 2 for details).



695
696

697 **ELECTRONIC SUPPLEMENTARY MATERIAL**

698 **Table S1:** (a) Ingredient (in g 100 g⁻¹) and (b) fatty acid (FA) composition expressed as a percentage
 699 of FA mass per total FA mass (% mean ± standard error of the mean; n = 3) in the High and Low
 700 omega-3 highly unsaturated fatty acid (n-3 HUFA) diets (made at INRA, Donzag, France). Only FA that
 701 occur above 1 % in at least a treatment group are represented. Bold denotes significant results of
 702 one-way ANOVA. Assumptions of normal distribution and homoscedasticity of residuals were met.

703 (a)

Ingredients ^a	High n-3 HUFA diet	Low n-3 HUFA diet
Fish meal LT 94	17	17
Casein	30	30
Rapeseed oil	2	10
Fish oil	8	0
Precooked starch	30	30
Vitamin mixture ^b	8	8
Mineral mixture ^c	4	4
Betaine	1	1

704

705 ^a Sources: fish meal LT 94: Norse (Fyllingsdalen, Norway); casein: Sigma-Aldrich (Germany); rapeseed oil:
 706 Système U (Créteil, France); fish oil: pure cod oil Cooper (Melun, France); precooked starch: Prégéflo Roquette
 707 Frères (Lestrem, France); vitamin mixture (INRA Jouy-en-Josas, France)

708 ^b Vitamin mixture (g kg⁻¹ vitamin mix): retinyl acetate 1; cholecalciferol 2.5; D α -tocopheryl acetate 5;
 709 menadione 1; thiamine-HCl 0.1; riboflavin 0.4; D-calcium pantothenate 2; pyridoxine-HCl 0.3; cyanocobalamin
 710 1; niacin 1; choline 200; ascorbic acid (ascorbyl polyphosphate) 5; folic acid 0.1; D-biotin 1; mesoinositol 30.

711 ^c Mineral mixture (g kg⁻¹ mineral mix): KCl 90; KI 0,04; CaHPO₄·2H₂O 500; NaCl 40; CuSO₄·5H₂O 3; ZnSO₄·7H₂O 4;
 712 CoSO₄ 0.02; FeSO₄·7H₂O 20; MnSO₄·H₂O 3; CaCO₃ 215; MgOH 124; Na₂SeO₃ 0 03; NaF 1.

713

714 (b)

Diet FA composition (%)	High n-3 HUFA	Low n-3 HUFA	F	Df	p-value
14 :0	4.30 ± 0.20	0.41 ± 0.02	362.2	1	< 0.001
16 :0	18.67 ± 0.80	6.10 ± 0.26	223.6	1	< 0.001
18 :0	4.59 ± 0.21	1.91 ± 0.09	142.7	1	< 0.001
16 :1n-7	4.83 ± 0.11	0.44 ± 0.01	1630	1	< 0.001
18 :1n-9	25.04 ± 0.23	56.74 ± 0.30	7004	1	< 0.001
18 :1n-7	2.56 ± 0.36	0.05 ± 0.05	46.2	1	< 0.01
20 :1n-9	1.82 ± 0.08	4.94 ± 2.09	2.2	1	0.21
18 :2n-6	5.06 ± 0.36	15.13 ± 1.82	29.6	1	< 0.01
18 :3n-3	2.42 ± 0.25	6.49 ± 1.03	14.7	1	0.02
20 :4n-6	1.38 ± 0.07	0.27 ± 0.04	205.1	1	< 0.001
20 :5n-3	6.63 ± 0.60	0.39 ± 0.07	106.3	1	< 0.001
22 :5n-3	1.64 ± 0.09	0.88 ± 0.31	5.5	1	0.07
22 :6n-3	10.95 ± 0.63	0.78 ± 0.14	246.3	1	< 0.001

715

716 **Table S2:** Mean \pm SEM mitochondrial function of muscle of golden grey mullet fed with High (n=11)
 717 and Low omega-3 highly unsaturated fatty acid (n-3 HUFA) diet. Rate of ATP production, rate of
 718 oxygen consumption to support ATP production (OXPHOS respiration), and to offset the proton
 719 leakage (LEAK respiration). Fluxes are expressed in pmol ATP produced and oxygen consumed $s^{-1} mg^{-1}$
 720 of mitochondrial protein. The respiratory control ratio (RCR), an index of mitochondrial coupling
 721 (Brand and Nicholls, 2011) was calculated as the ratio of OXPHOS respiration to the LEAK respiration.

Muscle mitochondrial function **High n-3 HUFA fish** **Low n-3 HUFA fish**

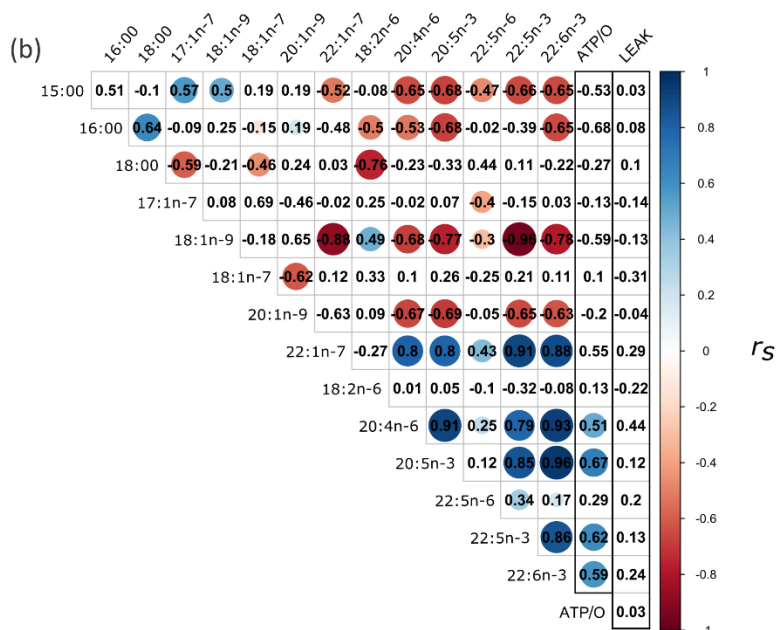
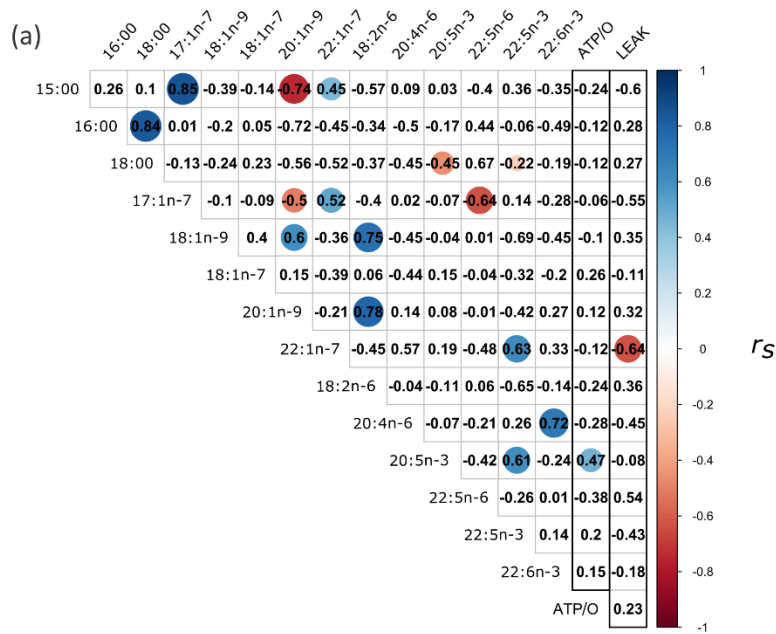
<i>ATP production</i>	3267.6 \pm 445.8	3869.7 \pm 237.4
<i>OXPHOS respiration</i>	836.8 \pm 112.0	733.6 \pm 60.8
<i>LEAK respiration</i>	55.1 \pm 4.0	57.1 \pm 2.9
<i>RCR</i>	14.6 \pm 1.5	13.0 \pm 1.0

722
 723 **Table S3:** Results from Kruskal-Wallis tests comparing fatty acid contents (percent of FA mass per
 724 total FA mass) in membrane lipids of the muscle of golden grey mullet (*Chelon auratus*) that were
 725 either fed a High or Low omega-3 highly unsaturated fatty acid (n-3 HUFA) diet. Only FA that occur
 726 above 1 % in at least a treatment group are represented. Means are expressed \pm standard error of
 727 the mean. Bold denotes significant results. N = 11-12 per treatment group.

<i>Muscle membrane FA composition (%)</i>	<i>High n-3 HUFA</i>	<i>Low n-3 HUFA</i>	<i>Chi²</i>	<i>p-value</i>
15 :0	0.97 \pm 0.08	1.68 \pm 0.29	4.125	0.04
16 :0	18.49 \pm 0.75	21.15 \pm 2.20	0.015	0.902
18 :0	9.78 \pm 0.39	9.91 \pm 1.44	0.379	0.54
17 :1n-7	1.29 \pm 0.09	1.66 \pm 0.17	1.83	0.18
18 :1n-9	11.42 \pm 0.46	19.32 \pm 1.98	15.51	< 0.001
18 :1n-7	2.95 \pm 0.07	2.81 \pm 0.08	1.67	0.20
20 :1n-9	1.03 \pm 0.06	1.41 \pm 0.09	8.72	0.003
22 :1n-7	1.42 \pm 0.10	1.40 \pm 0.19	0.034	0.85
18 :2n-6	1.70 \pm 0.24	4.95 \pm 0.72	11.05	< 0.001
20 :4n-6	4.16 \pm 0.08	2.31 \pm 0.42	16.5	< 0.001
20 :5n-3	9.82 \pm 0.32	5.70 \pm 1.03	10.64	< 0.001
22 :5n-6	1.35 \pm 0.08	0.89 \pm 0.14	8.37	< 0.01
22 :5n-3	5.58 \pm 0.15	3.97 \pm 0.62	2.76	0.09
22 :6n-3	20.78 \pm 0.66	10.08 \pm 1.96	16.5	< 0.001

729
 730

731 **Figure S1:** Spearman's correlation analyses between the membrane lipid fatty acids and the
 732 mitochondrial function of muscle of golden grey mullet fed with (a) High (n=11) and (b) Low omega-3
 733 highly unsaturated fatty acid (n-3 HUFA) diet. FA composition were calculated as percent of FA mass
 734 per total FA mass. Only FA that occur above 1 % in at least a treatment group are represented.
 735 Mitochondrial function were expressed as the mitochondrial efficiency (ATP/O) and the LEAK
 736 respiration. N = 11-12 per treatment group. The coloured circles represent significant correlations
 737 between two variables ($p < 0.05$). The numbers inside cells are the associated Spearman's correlation
 738 (r_s).



739