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# Cracking the code of biodiversity responses to past climate change

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**How individual species and entire ecosystems will respond to future climate change are among the most pressing questions facing ecologists. Past biodiversity dynamics recorded in the paleoecological archives show a broad array of responses, yet significant knowledge gaps remain. In particular, the relative roles of evolutionary adaptation, phenotypic plasticity, and dispersal in promoting survival during times of climate change have yet to be clarified. Investigating the paleo-archives offers great opportunities to understand biodiversity responses to future climate change. In this review we discuss the mechanisms by which biodiversity responds to environmental change, and identify gaps of knowledge on the role of range shifts and tolerance. We also outline approaches at the intersection of paleoecology, genomics, experiments and predictive models that will elucidate the processes by which species have survived past climatic changes and enhance predictions of future changes in biological diversity.**

### **Looking to the past to understand the future of biodiversity**

Current estimates predict that atmospheric CO<sub>2</sub> levels may rise up to 450-500 ppm by the end of this century, potentially driving an increase in global average temperature on the order of 2 to 5 °C [1]. These projected magnitudes and rates of future climate change, unparalleled in many million years [2], pose major threats to biodiversity [3–6]. The scientific community is struggling to fully comprehend the range of responses of biodiversity to climate change, to anticipate whether species can respond quickly enough, and pinpoint the various roles of life-history properties (e.g., dispersal capacity, genetic diversity, reproductive strategies, phenotypic plasticity, population growth rates) in adapting to a changing environment. To make reliable predictions it is essential to advance our understanding of the underlying principles and mechanisms of biodiversity responses. One fruitful approach is to look to the past by using geo-historical records to learn how individuals, populations, communities and

biomes have responded to previous climatic changes [7–11]. Whether individuals and populations will adapt by evolutionary change or plasticity, whether they will migrate fast enough, and whether those responses will be adequate to forestall collapses of species ranges and prevent widespread species extinctions can be explored using case studies from the past. Indeed, past climate change, whether abrupt or gradual, and whether occurring in deep time or recent history, offers a vast set of unplanned natural experiments to explore biodiversity responses and test ecological and evolutionary theories. Recent years have seen the accumulation of well-documented examples of the influence of climate change on persistence, adaptation and diversification, dispersal, and extinction (e.g. [12–15]). The effects of climate change on rates and routes of range shifts have been intensively studied by biogeographers and paleoecologists, augmented recently by molecular markers and ancient DNA (aDNA; [16,17]). *In situ* tolerance to changing climate conditions has been explored in the fossil record using functional morphology and evolutionary genetics, including recent experimental approaches like ‘resurrection ecology’ [15,18,19]. Finally, paleoecological records of local and global extinctions provide information on the nature and consequences of failure of *in situ* tolerance and range shifts [20,21].

However, key knowledge gaps remain. The relative importance of different mechanisms involved in species tolerance (e.g., evolutionary adaptive change versus phenotypic plasticity), and the nature and rates of climate-driven anagenetic evolution - a transition of one species to another- and cladogenetic evolution - the separation of a species into two or more species or clades- remain poorly understood [22]. The relative efficacy of *in situ* tolerance and range shifting under different rates and magnitudes of climate change is obscure [23,24]. Although much attention has been devoted to paleoecological records of species’ range shifts [25], the speed and underlying controls are not clear except in a few

specific cases [24]. Moreover, significant challenges remain for better integrating knowledge, scales, methods and data from a variety of biological disciplines, from paleoecology to genomics. In this review, we (1) synthesize the main responses of biodiversity to past climate change from deep to recent time (tolerance *in situ*, range shifts, and their simultaneous failure, resulting in extinction), (2) identify key knowledge gaps concerning underlying mechanisms (which span a broad set of biological disciplines), and (3) review and discuss new approaches that integrate multiple methods and disciplines to better understand the strategies by which life adapts to climate change and to better anticipate future responses of biological diversity.

## **Biodiversity responses to climate change**

### **Tolerance, Adaptive Evolution, and Diversification**

Biotic responses to climatic and environmental changes as shown by the fossil records vary from macroevolutionary divergences (at very long ( $10^6$ - $10^7$  yr) time scales), to adaptive evolution ( $10^0$ - $10^5$  yr), to phenotypic adjustments in place ( $10^{-1}$ - $10^3$  yr). Long-term climate change has been considered an important driver of high-order diversification, as clades respond to new climatic regimes [14]. In shorter time spans, many individuals and populations (e.g. long-lived modular organisms like corals and plant genets) can tolerate a high degree of climate change *in situ*. Paleogenetic records suggest also that adaptive evolution can support long-term persistence of species in response to climate change [26]. Adaptations can enable exploitation of new niches: for example, adaptive mutations in woolly mammoth haemoglobin allowed the exploitation of high-latitude cold environments during the Pleistocene [27]. Examples of more recent microevolutionary responses to climatic

change include changes in the body color of owls during warmer winters [28], or adaptive changes in the flowering time of Brassicas in response to drought [29].

Whether adaptive evolutionary change or plasticity are the prevalent strategy to tolerate climatic changes *in situ*, and at what spatial and time scale these two processes play a role, can be difficult to disentangle for extant populations [22,30] and even more challenging for ancient extinct populations, but both are candidate processes in population persistence under climate change. For most reported cases of climate-driven phenotypic changes in the wild, it remains unclear whether they are caused by microevolution or phenotypic plasticity, although recent meta-analyses suggest that most responses to climatic change are mediated by phenotypic plasticity [22,31] (see also [32–35]).

### Range shifts

Range shifting (usually referred as migration in paleo-disciplines) has been a dominant response of species to climate shifts in the past [25]. Past range shifts are typically inferred from spatial and temporal patterns in fossil data [36,37], geographic patterns in genetic markers of extant and extinct populations [17], or both (e.g. [38]). They have shown variable species-specific spatial trajectories, timing and migration rates, ranging from a few tens to a few thousand m/yr, with averages around 2.7 km/decade [39–41]. Overall, there is evidence of both rapid range shifts and community reshuffling [42] as well as many species lagging behind climate [43], which reinforces the high specificity of range shift patterns across taxa.

There are many different mechanisms by which climate change influences range shifts [44]. First, climate change can improve suitability beyond the range limit so that species may establish at formerly unsuitable areas like higher latitudes or altitudes [45,46]. . Second,

climate change could foster colonisation of new areas in several ways: enhanced fecundity of source populations (thus increasing propagule pressure), increased propensity to disperse or emigrate (particularly in animals), or acceleration of dispersal processes [47,48]. Climate change can also enhance establishment of propagules after arrival, both directly [49] and – particularly in rapid climate change– by reducing populations of dominant species, via mortality or disturbance [50]. Finally, climate change could reduce the probability of extinction of leading edge populations, for instance due to extreme climatic events [51]. A variety of processes are involved in species' range shifts, all of which can be directly or indirectly (e.g. mediated by species interactions) influenced by climate change [23,50,52,53]. A challenge for ecologists, biogeographers, and paleoecologists is to identify generalizations, and to understand the role of species-specific, locale-specific, and time-specific contingencies and idiosyncracies in driving patterns and rates of range shifts.

### Extinction

When species cannot tolerate climate change *in situ*, or colonize suitable habitat elsewhere quickly enough, they become extinct. In extreme cases, many high-order clades can be lost in mass extinction events [54,55]. There is strong support for a primary role of climate change, alone or in connection to other factors, in extinction events of different magnitude over the last 500 million years, including the recent extinction of large mammals in the last 50,000 years [21]. Evidence of climate-driven species extinctions in recent centuries is limited [56], with rare exceptions being synergistic functions of both 20<sup>th</sup> century human-induced climate change and other proximate drivers of extinction (including infectious diseases) [57]. However, anthropogenic climate disruption is predicted to soon compete with habitat destruction as the most important driver of contemporary extinctions [58,59].

Climate change may trigger extinctions and local extirpations by surpassing the physiological limits of species, by reducing primary productivity of ecosystems and thereby local population fitness across food webs, and indirectly by disrupting ecological interactions via changes in species distributions or phenology. For instance, coral bleaching, the loss of intracellular endosymbionts due to the increase in prevalence of extreme heating episodes and changes in the carbon cycle, is one of the main supported mechanisms behind coral extinctions during the five mass extinction events [60]. Also, drier and colder climatic conditions during the LGM triggered a reduction in overall primary productivity, provoking losses in genetic diversity and populations of large grazers [13], depleting lineages, for example, of bowhead whales [61], and contributing to local and global extinctions [21]. These pathways to extinctions in different periods of the Earth's history share some commonalities. In particular, climatic changes that exceed in magnitude and speed those experienced during the evolutionary history of species usually trigger extinction events, and climate change has frequently interacted with other extinction drivers [61].



## **Unknowns, challenges and routes ahead**

Our review of the modal responses of biodiversity to past climate change unveils key knowledge gaps concerning the underlying mechanisms. We identify and discuss them here and propose new integrative approaches that show potential to crack the code of how biodiversity responds to climate change.

### Evolutionary adaptation versus plasticity?

Climate-relevant decisions and policies implemented today (e.g., levels of CO<sub>2</sub> emissions) have both short and long-term consequences for future biodiversity, influencing range shifts, divergence, speciation, hybridization, anagenetic evolution and extinction. Paleo-archives reveal that speciation, evolution and phenotypic change have played roles in species responses to past environmental changes. However, the relative roles of those mechanisms in different settings, for different taxa, and across different timespans need clarification and exploration [62].

At deep-time scales, comparative phylogenetics and novel macroevolutionary approaches are offering new insights into speciation and phenotypic change in response to major climatic shifts [63]. For instance, it was found that expected future climate change largely surpass past rates of climatic niche evolution among vertebrate species [64]. Comparative approaches allow fitting various models of phenotypic evolution and diversification to phylogenies in order to estimate evolutionary rates, including speciation and extinction [14]. Recently, models that can explicitly test for the effect of climatic changes on these evolutionary rates have been developed [65–68]. Future studies including genomic level data across thousands of species and climate-dependent evolutionary models will provide deeper insights on the role of climate change on speciation, including bursts, and phenotypic change.

At shorter time spans, from thousands to hundreds of years, comparative analyses of species and populations provide important insights into the evolutionary processes that led to present day genetic and phenotypic diversity. However, when limited to exploring extant genetic patterns, inferences on past processes can be limited. New approaches considering species' traits and explicit scenarios of past range dynamics can bring much deeper insights on the role of phenotypic variation on population persistence, range shifts, and generation of genetic structure [69]. Alternatively, long-term observational studies enable measurement of evolutionary processes by comparing temporal changes in genetic and phenotypic diversity with expectations of neutral and adaptive evolutionary models [70]. Long-term studies, however, may require commitments beyond the career or life spans of individual researchers. 'Resurrection Ecology' (see Glossary, Figure 1 and Anticipating Extinctions section) provides an alternative and complementary path to reconstructing long-term patterns of evolutionary changes and unravelling mechanisms of response to climatic and other environmental changes [15].

### Migrating fast enough?

Although dispersal is a key process underlying range shifts and the spread of native and invasive species, the migration capacity of species under rapid climate change remains uncertain [23]. While some taxa seem unable to shift ranges under changing climates [43], others seem able to migrate at a fast pace [45]. Attempts to explain observed range shifts based on species traits or ecological strategies have obtained modest results [71,72] (but see [73]). Low predictability may be expected given the large number of processes involved in range shifts, as well as the complexity and path-dependence when those processes interact.

The dispersal process itself is highly stochastic and inherently uncertain [74]. Other important processes include size and fecundity of source populations (which determine propagule pressure), gene flow, local adaptation, evolution of dispersal, biotic interactions (competition, facilitation, mutualisms), Allee effects, and so on, all of which are likely to be affected by climate change [23]. Spatial heterogeneity on the landscape plays a role (e.g., dispersal-target size), as does high-frequency climate variability [47,50]. As a result, we may not be able to go much farther than estimating dispersal potentials for different species or populations [74]. A critical challenge is to use paleoecological and ecological data to identify generalizations that can emerge from the location-specific, species-specific, and event-specific particulars of detailed case studies [47,50].

Paleoecology has largely contributed to estimate how fast species migrated under past climatic changes under minimum levels of pre-historic global human intervention. Unfortunately, contemporaneous dispersal rates are likely to be rather different than past rates due to radically different conditions: more fragmented habitats, missing and novel interactions, or nearly unlimited human-mediated dispersal [23]. Hence, estimates of past migrations rates, however informative, may be of limited value when attempting to forecast future range shifts. Instead, a better understanding of the causes of variation in range shift rates may move us forward. Comparative studies of range shifts patterns among tens or even hundreds of species could throw some light into the role of environmental (contingent) factors as well as intrinsic factors that make some species migrate faster, slower or not at all.

#### Anticipating future extinctions

Revealing how the accumulative failure of *in situ* tolerance and dispersal mechanisms leads to population extirpation and ultimately species extinction under climate change is of utmost

importance to provide robust scenarios for future biodiversity and to enhance conservation strategies. Recent insights on the factors correlating with declining genetic diversity, population sizes, and local and global extinctions, have been achieved for megafauna species during the Late Quaternary, highlighting the key role of the integration of disciplines like paleo-genomics and macroecological models [13] to explain range shifts, population collapses and species extinctions under climate change. More recently the application of genomics to historical specimens in biological collections is arising as a novel trend to understand genomic erosion of endangered species [75]. Although past biotic turnover and extinction events have provided better knowledge on extinction dynamics and their relation to climatic changes, paleo-data together with current data has only recently been fully implemented in quantitative assessments of future risk of extinction [76]. Moreover, correlative approaches lacking key biological mechanisms have dominated the forecasting of future responses of biodiversity to climate change. A paradigm shift from correlative models of different complexity to process-based simulations informed by paleo-records will bring deeper insights on the interplay of tolerance and dispersal to explain species range dynamics and extinctions under climate change [77] (Box 2; Figure 2).

#### Integrating experimental approaches, paleorecords and models

A large gap remains between mechanistic experiments at local scales and large-scale macroecological models that forecast the persistence of biological diversity under future global climate change [78]. The integration of experimental paleoecology, resurrection ecology, and large-scale process-based models holds a great potential to shed light on key mechanisms, as the unveiled role of in situ adaptation via evolutionary changes. Their integration can also provide large scales predictions of the magnitude and speed of evolutionary change that species will need to achieve for averting declines and extinction.

Resurrection Ecology (RE) focuses on life forms (zooplankton, insects, algae, fungi, bacteria, plants) producing resting stages as part of their life cycles in response to environmental hardship [18,79,80], and its temporal extent encompasses mainly the last 200 years (but see [81]). When such resting stages can be recovered from ancient sediments and reared in the laboratory, they can reveal molecular targets (genes, metabolites, proteins) that enable evolution and adaptation to changing climate. Resurrecting individuals from such species and populations across documented temporal shifts in the environment uniquely permits simultaneous measurement of both plastic (phenotypic and behavioural) and genetic (evolutionary) responses to climatic change, using common garden or transplant experiments [82,83]. Relative fitness of both historical and modern populations can be measured in response to different climatic regimes, including past, present and future. Such long-term studies, replicated across multiple environments and taxa, can be a powerful resource for building models to forecast species persistence [84] (Figure 1).

A long-standing complement to resurrection ecology might aptly be designated Methuselan ecology (after the biblical character known for his multi-century longevity). Methuselan ecology (ME) focuses on multiple, overlapping generations of living organisms of unusual longevity and studies them to examine demographic, genetic, and ecological responses to environmental change. Tree-rings have long been used to reconstruct growth responses to climate variation over centuries to millennia, and effects of climate variability on demographic patterns over several centuries [85–87]. In a recent set of studies, tree-ring demography has been combined with genetic studies to examine patterns, rates, and controls of colonization of new sites by *Pinus ponderosa* in western North America, revealing interactions among long-distance dispersal, population genetics, climate variability, and Allee effects [88–90]. Although

more difficult to apply to animals, potential exists for simultaneous age- and genetic sampling of animals of unusual longevity that can be independently aged (e.g., certain marine fish, tortoises, corals).

Incipient modelling approaches providing spatially explicit predictions of shifts on species distribution and abundance can now incorporate evolutionary adaptation [91]. These models are however in need of quantitative estimates on the magnitude and speed of adaptation, and both Resurrection Ecology and Methuselan ecology can provide actual values based on historical information. Yet this integration between data and models to forecast future responses at large spatial scales across a variety of taxa faces daunting challenges. Both RE and ME are restricted to a limited set of organisms under a limited set of circumstances (experimental and natural) [80,83]. However, this is not an issue to understanding organismal response to climatic change. Indeed, some species that provide the unique advantage of resurrecting dormant stages are also keystone species in their ecosystem, enabling us to illuminate the links in the causal chain from genes to communities and ecosystems. Ideally, model organisms and systems that feature a comprehensive triad of strong ecological interactions in nature, experimental tractability in diverse contexts and accessibility to modern genomic tools, may be used [92]. The water flea *Daphnia* and the flowering plant *Silene stenophylla*, as well as a number of bacteria are examples of organisms that satisfy these criteria [15]. They can be used as proxies to study the impact of climatic change on different ecosystems.

Resurrection Ecology and Methuselan ecology do not only dig in the past. A forward-in-time approach, involving long-term collection of propagule-banks [79,83] will allow scientists in the future to measure the magnitude and speed of evolutionary changes. Under the Project

Baseline, seeds of several populations across the geographical range of >60 plant species are now stored and will be grown with contemporaneous seeds during the next 50 years, allowing the identification of phenotypic and molecular evolution occurring during the intervening time under different magnitudes of climate change. Similar initiatives in other continents, and a taxonomic expansion of these experiments, would enable a next generation of predictive models incorporating evolutionary adaptation. Joseph Grinnell in 1910 already foresaw that the most significant value of his field work on Californian fauna would be for the students of the future. Today, his and other pioneers' data have served to document the magnitude and rates of species range shifts and local extirpations in the last century [12,93–96].

### **Concluding remarks**

Climate change has triggered large and persistent effects on biological diversity, including speciation, redistribution, local adaptations and extinction events. However, a deeper mechanistic understanding of these dynamics is urgently needed (see also Outstanding Questions). Until recently, most evidence suggested that biotic responses to climate change were dominated by range shifting. It is now clear from both paleoecological and ecological perspectives that *in situ* tolerance, being plasticity or adaptive evolution, are also key responses to climate change. Although adaptation is now an important object of study, we are still lacking sufficient evidence - comparative or experimental - on fundamental questions: How is adaptive evolution shaped by dispersal and range shifting in real ecosystems? Conversely, how is dispersal influenced by adaptive evolution? How do tolerance, adaptive evolution, and dispersal interact in specific circumstances to reduce or amplify risk of extinction? The integration of recorded long-term responses and ecological and evolutionary theories into models will facilitate a deeper understanding of the roles of adaptation and dispersal under climate change. Cracking the code of past biodiversity responses to climate

change will increase the ability to anticipate, adapt and mitigate future declines of biological diversity under climate change.

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## References

- 1 IPCC (2013) *Climate Change 2013 - The Physical Science Basis: Working Group I Contribution to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*, Cambridge University Press.
- 2 Foster, G.L. *et al.* (2017) Future climate forcing potentially without precedent in the last 420 million years. *Nature Communications* 8, 14845
- 3 Moritz, C. and Agudo, R. (2013) The Future of Species Under Climate Change: Resilience or Decline? *Science* 341, 504–508
- 4 Urban, M.C. (2015) Accelerating extinction risk from climate change. *Science* 348, 571–573
- 5 Pecl, G. (2017) Biodiversity redistribution under climate change: Impacts on



ecosystems and human well-being. *Science* 355, 6332

6 Garcia, R.A. *et al.* (2014) Multiple Dimensions of Climate Change and Their Implications for Biodiversity. *Science* 344, 1247579

7 Jackson, S.T. and Overpeck, J.T. (2000) Responses of plant populations and communities to environmental changes of the late Quaternary. *Paleobiology* 26, 194–220

8 Willis, K.J. *et al.* (2010) Biodiversity baselines, thresholds and resilience: testing predictions and assumptions using palaeoecological data. *Trends in Ecology & Evolution* 25, 583–591

9 Dawson, T.P. *et al.* (2011) Beyond Predictions: Biodiversity Conservation in a Changing Climate. *Science* 332, 53–58

10 Fritz, S.A. *et al.* (2013) Diversity in time and space: wanted dead and alive. *Trends in Ecology & Evolution* 28, 509–516

11 Barnosky, A.D. *et al.* (2017) Merging paleobiology with conservation biology to guide the future of terrestrial ecosystems. *Science* 355, eaah4787

12 Moritz, C. *et al.* (2008) Impact of a Century of Climate Change on Small-Mammal Communities in Yosemite National Park, USA. *Science* 322, 261–264

13 Lorenzen, E.D. (2011) Species-specific responses of Late Quaternary megafauna to climate and humans. *Nature* 479, 359–364

14 Condamine, F.L. *et al.* (2013) Macroevolutionary perspectives to environmental change. *Ecology Letters* 16, 72–85

15 Orsini, L. *et al.* (2013) The evolutionary time machine: using dormant propagules to forecast how populations can adapt to changing environments. *Trends in Ecology & Evolution* 28, 274–282

16 Der Sarkissian, C. *et al.* (2014) Ancient genomics. *Philosophical Transactions of the Royal Society B: Biological Sciences* 370, 20130387–20130387

- 17 Hewitt, G.M. (2004) Genetic consequences of climatic oscillations in the Quaternary. *Philosophical Transactions of the Royal Society B: Biological Sciences* 359, 183–195
- 18 Kerfoot, W.C. and Weider, L.J. (2004) Experimental paleoecology (resurrection ecology): Chasing Van Valen’s Red Queen hypothesis. *Limnology and Oceanography* 49, 1300–1316
- 19 Orsini, L. *et al.* (2016) Temporal genetic stability in natural populations of the waterflea *Daphnia magna* in response to strong selection pressure. *Molecular Ecology* 25, 6024–6038
- 20 Jackson, S.T. and Weng, C. (1999) Late Quaternary extinction of a tree species in eastern North America. *Proceedings of the National Academy of Sciences* 96, 13847–13852
- 21 Cooper, A. *et al.* (2015) Abrupt warming events drove Late Pleistocene Holarctic megafaunal turnover. *Science* 349, 602–606
- 22 Merila, J. and Hendry, A.P. (2014) Climate change, adaptation, and phenotypic plasticity: the problem and the evidence. *Evolutionary Applications* 7, 1–14
- 23 Corlett, R.T. and Westcott, D.A. (2013) Will plant movements keep up with climate change? *Trends in Ecology & Evolution* 28, 482–488
- 24 Pearson, R. (2006) Climate change and the migration capacity of species. *Trends Ecol. Evol* 21, 111–113
- 25 Huntley, B. and Webb, T. (1989) Migration: species’ response to climatic variations caused by changes in the earth’s orbit. *J. Biogeogr* 16, 5–19
- 26 Davis, M.B. *et al.* (2005) Evolutionary responses to changing climate. *Ecology* 86, 1704–1714
- 27 Campbell, K.L. *et al.* (2010) Substitutions in woolly mammoth hemoglobin confer biochemical properties adaptive for cold tolerance. *Nature Genetics* 42, 536–540
- 28 Karrel, P. (2011) Climate change drives microevolution in a wild bird. *Nature*

- 29     Franks, S.J. *et al.* (2007) Rapid evolution of flowering time by an annual plant in response to a climate fluctuation. *Proceedings of the National Academy of Sciences* 104, 1278–1282
- 30     Richardson, J.L. *et al.* (2014) Microgeographic adaptation and the spatial scale of evolution. *Trends in Ecology & Evolution* 29, 165–176
- 31     Hendry, A.P. *et al.* (2008) Human influences on rates of phenotypic change in wild animal populations. *Molecular Ecology* 17, 20–29
- 32     Charmantier, A. *et al.* (2008) Adaptive Phenotypic Plasticity in Response to Climate Change in a Wild Bird Population. *Science* 320, 800–803
- 33     Charmantier, A. and Gienapp, P. (2014) Climate change and timing of avian breeding and migration: evolutionary versus plastic changes. *Evol. Appl* 7, 15–28
- 34     Franks, S.J. *et al.* (2013) Evolutionary and plastic responses to climate change in terrestrial plant populations. *Evolutionary Applications* 7, 123–139
- 35     Reusch, T.B.H. (2013) Climate change in the oceans: evolutionary versus phenotypically plastic responses of marine animals and plants. *Evolutionary Applications* 7, 104–122
- 36     Huntley, B. and Birks, H.J.B. (1983) An atlas of past and present pollen maps for Europe, 0-13,000 years ago. Cambridge University Press
- 37     Jackson, S.T. *et al.* (1997) Mapped plant-macrofossil and pollen records of late Quaternary vegetation change in Eastern North America. *Quaternary Science Reviews* 16, 1–70
- 38     Norris, J.R. *et al.* (2015) Late Holocene expansion of ponderosa pine (*Pinus ponderosa*) in the Central Rocky Mountains, USA. *Journal of Biogeography* 43, 778–790
- 39     Blois, J.L. (2013) Narrowing the estimates of species migration rates. *Frontiers*

- 40     Feurdean, A. *et al.* (2013) Tree migration-rates: narrowing the gap between inferred post-glacial rates and projected rates. *PLoS ONE* 8, 71797
- 41     Ordóñez, A. and Williams, J.W. (2013) Climatic and biotic velocities for woody taxa distributions over the last 16 000 years in eastern North America. *Ecology Letters* 16, 773–781
- 42     Tinner, W. and Lotter, A.F. (2001) Central European vegetation response to abrupt climate change at 8.2 ka. *Geology* 29, 551
- 43     Normand, S. *et al.* (2011) Postglacial migration supplements climate in determining plant species ranges in Europe. *Proceedings of the Royal Society B: Biological Sciences* 278, 3644–3653
- 44     Holt, R.D. and Keitt, T.H. (2000) Alternative causes for range limits: a metapopulation perspective. *Ecology Letters* 3, 41–47
- 45     Chen, I.-C. *et al.* (2011) Rapid Range Shifts of Species Associated with High Levels of Climate Warming. *Science* 333, 1024–1026
- 46     Hodgson, J.A. *et al.* (2015) Predicting microscale shifts in the distribution of the butterfly *Plebejus argus* at the northern edge of its range. *Ecography* 38, 998–1005
- 47     Lyford, M.E. *et al.* (2003) Influence of landscape structure and climate variability on a late Holocene plant migration. *Ecological Monographs* 73, 567–583
- 48     Battisti, A. (2006) A rapid altitudinal range expansion in the pine processionary moth produced by the 2003 climatic anomaly. *Glob. Change Biol* 12, 662–671
- 49     Wetthey, D.S. *et al.* (2011) Response of intertidal populations to climate: Effects of extreme events versus long term change. *Journal of Experimental Marine Biology and Ecology* 400, 132–144
- 50     Jackson, S.T. *et al.* (2009) Ecology and the ratchet of events: Climate variability,

niche dimensions, and species distributions. *Proceedings of the National Academy of Sciences* 106, 19685–19692

51 Cavanaugh, K.C. *et al.* (2013) Poleward expansion of mangroves is a threshold response to decreased frequency of extreme cold events. *Proceedings of the National Academy of Sciences* 111, 723–727

52 Anderson, B.J. *et al.* (2009) Dynamics of range margins for metapopulations under climate change. *Proceedings of the Royal Society B: Biological Sciences* 276, 1415–1420

53 Jackson, S.T. and Blois, J.L. (2015) Community ecology in a changing environment: perspectives from the Quaternary. *Proceedings of the National Academy of Sciences* 112, 4915–4921

54 Harnik, P.G. *et al.* (2012) Extinctions in ancient and modern seas. *Trends in Ecology & Evolution* 27, 608–617

55 Barnosky, A.D. *et al.* (2011) Has the Earth's sixth mass extinction already arrived? *Nature* 471, 51–57

56 Young, H.S. *et al.* (2016) Patterns, Causes, and Consequences of Anthropocene Defaunation. *Annual Review of Ecology, Evolution, and Systematics* 47, 333–358

57 Pounds, J.A. (2006) Widespread amphibian extinctions from epidemic disease driven by global warming. *Nature* 439, 161–167

58 Wiens, J.J. (2016) Climate-Related Local Extinctions Are Already Widespread among Plant and Animal Species. *PLOS Biology* 14, e2001104

59 Scheffers, B.R. *et al.* (2016) The broad footprint of climate change from genes to biomes to people. *Science* 354, aaf7671

60 Veron, J.E.N. (2008) Mass extinctions and ocean acidification: biological constraints on geological dilemmas. *Coral Reefs* 27, 459–472

61 Foote, A.D. *et al.* (2011) Out of the Pacific and Back Again: Insights into the

Matrilineal History of Pacific Killer Whale Ecotypes. *PLoS ONE* 6, e24980

62 Chevin, L.-M. *et al.* (2013) Phenotypic plasticity and evolutionary demographic responses to climate change: taking theory out to the field. *Functional Ecology* 27, 967–979

63 Colwell, R.K. and Rangel, T.F. (2010) A stochastic, evolutionary model for range shifts and richness on tropical elevational gradients under Quaternary glacial cycles.

*Philosophical Transactions of the Royal Society B: Biological Sciences* 365, 3695–3707

64 Quintero, I. and Wiens, J.J. (2013) Rates of projected climate change dramatically exceed past rates of climatic niche evolution among vertebrate species. *Ecology Letters* 16, 1095–1103

65 Morlon, H. (2014) Phylogenetic approaches for studying diversification. *Ecology Letters* 17, 508–525

66 Clavel, J. and Morlon, H. (2017) Accelerated body size evolution during cold climatic periods in the Cenozoic. *Proceedings of the National Academy of Sciences* 114, 4183–4188

67 Manceau, M. *et al.* (2017) A unifying comparative phylogenetic framework including traits coevolving across interacting lineages. *Systematic Biology* 66, 551–568

68 Lewitus, E. and Morlon, H. (2018) Detecting environment-dependent diversification from phylogenies: a simulation study and some empirical illustrations. *Systematic Biology* 67, 576–593

69 Papadopoulou, A. and Knowles, L.L. (2016) Toward a paradigm shift in comparative phylogeography driven by trait-based hypotheses. *Proceedings of the National Academy of Sciences* 113, 8018–8024

70 Hoffmann, A.A. and Sgrò, C.M. (2011) Climate change and evolutionary adaptation. *Nature* 470, 479–485

71 Angert, A.L. *et al.* (2011) Do species' traits predict recent shifts at expanding range edges? *Ecology Letters* 14, 677–689

- 72 MacLean, S.A. and Beissinger, S.R. (2017) Species' traits as predictors of range shifts under contemporary climate change: A review and meta-analysis. *Global Change Biology* 23, 4094–4105
- 73 Estrada, A. *et al.* (2016) Usefulness of Species Traits in Predicting Range Shifts. *Trends in Ecology & Evolution* 31, 190–203
- 74 Clark, J.S. (2003) Estimating population spread: what can we forecast and how well? *Ecology* 84, 1979–1988
- 75 Díez-del-Molino, D. *et al.* (2018) Quantifying Temporal Genomic Erosion in Endangered Species. *Trends in Ecology & Evolution* 33, 176–185
- 76 Nogués-Bravo, D. *et al.* (2016) Amplified plant turnover in response to climate change forecast by Late Quaternary records. *Nature Climate Change* 6, 1115–1119
- 77 Fordham, D.A. *et al.* (2014) Better forecasts of range dynamics using genetic data. *Trends in Ecology & Evolution* 29, 436–443
- 78 Nogués-Bravo, D. and Rahbek, C. (2011) Communities Under Climate Change. *Science* 334, 1070–1071
- 79 Franks, S.J. *et al.* (2018) Using the resurrection approach to understand contemporary evolution in changing environments. *Evolutionary Applications* 11, 17–28
- 80 Weider, L.J. *et al.* (2018) Evolutionary aspects of resurrection ecology: Progress, scope, and applications-An overview. *Evolutionary Applications* 11, 3–10
- 81 Frisch, D. *et al.* (2014) A millennial-scale chronicle of evolutionary responses to cultural eutrophication in *Daphnia*. *Ecology Letters* 17, 360–368
- 82 Orsini, L. *et al.* (2012) The role of selection in driving landscape genomic structure of the waterflea *Daphnia magna*. *Molecular Ecology* 22, 583–601
- 83 Etterson, J.R. *et al.* (2016) Project Baseline: An unprecedented resource to study plant evolution across space and time. *American Journal of Botany* 103, 164–173

- 84 Urban, M. (2016) Improving the forecast for biodiversity under climate change. *Science* 353, 8466
- 85 Swetnam, T.W. and Betancourt, J.L. (1998) Mesoscale Disturbance and Ecological Response to Decadal Climatic Variability in the American Southwest. *Journal of Climate* 11, 3128–3147
- 86 Brown, P.M. (2006) Climate effects on fire regimes and tree recruitment in black hills ponderosa pine forests. *Ecology* 87, 2500–2510
- 87 Gray, S.T. *et al.* (2006) Role of multidecadal climate variability in a range extension of pinyon pine. *Ecology* 87, 1124–1130
- 88 Lesser, M.R. and Jackson, S.T. (2012) Contributions of long-distance dispersal to population growth in colonising *Pinus ponderosa* populations. *Ecology Letters* 16, 380–389
- 89 Lesser, M.R. and Jackson, S.T. (2012) Making a stand: five centuries of population growth in colonizing populations of *Pinus ponderosa*. *Ecology* 93, 1071–1081
- 90 Lesser, M.R. *et al.* (2013) Development of genetic diversity, differentiation and structure over 500 years in four ponderosa pine populations. *Molecular Ecology* 22, 2640–2652
- 91 Bush, A. *et al.* (2016) Incorporating evolutionary adaptation in species distribution modelling reduces projected vulnerability to climate change. *Ecology Letters* 19, 1468–1478
- 92 Miner, B.E. *et al.* (2012) Linking genes to communities and ecosystems: *Daphnia* as an ecogenomic model. *Proceedings of the Royal Society B: Biological Sciences* 279, 1873–1882
- 93 Dobrowski, S.Z. *et al.* (2011) Modeling plant ranges over 75 years of climate change in California, USA: temporal transferability and species traits. *Ecological Monographs* 81, 241–257
- 94 Morueta-Holme, N. *et al.* (2015) Strong upslope shifts in Chimborazo’s vegetation



over two centuries since Humboldt. *Proceedings of the National Academy of Sciences* 112, 12741–12745

95 Hédli, R. *et al.* (2017) Resurvey of historical vegetation plots: a tool for understanding long-term dynamics of plant communities. *Applied Vegetation Science* 20, 161–163

96 Stewart, J.A.E. *et al.* (2015) Revisiting the past to foretell the future: summer temperature and habitat area predict pika extirpations in California. *Journal of Biogeography* 42, 880–890

97 McInerney, F.A. and Wing, S.L. (2011) The Paleocene-Eocene Thermal Maximum: A Perturbation of Carbon Cycle, Climate, and Biosphere with Implications for the Future. *Annual Review of Earth and Planetary Sciences* 39, 489–516

98 Prothero, D.R. and Berggren, W.A., eds. (1992) *Eocene-Oligocene Climatic and Biotic Evolution*, Princeton University Press.

99 Erdei, B. *et al.* (2007) Neogene flora and vegetation development of the Pannonian domain in relation to palaeoclimate and palaeogeography. *Palaeogeography, Palaeoclimatology, Palaeoecology* 253, 115–140

100 Svenning, J.-C. (2003) Deterministic Plio-Pleistocene extinctions in the European cool-temperate tree flora. *Ecology Letters* 6, 646–653

101 Davis, M.B. and Shaw, R.G. (2001) Range shifts and adaptive responses to Quaternary climate change. *Science* 292, 673–679

102 Hansen, J. *et al.* (2013) Climate sensitivity, sea level and atmospheric carbon dioxide. *Philosophical Transactions of the Royal Society A: Mathematical, Physical and Engineering Sciences* 371, 20120294–20120294

103 Connolly, S.R. *et al.* (2017) Process, Mechanism, and Modeling in Macroecology. *Trends in Ecology & Evolution* 32, 835–844

104 Fordham, D.A. (2016) Predicting and mitigating future biodiversity loss using long-

term ecological proxies. *Nature Clim. Change* 6, 909–916

## **Glossary**

**Adaptive evolution** –Results from the propagation of advantageous alleles in populations through natural selection, driven by environmental selection pressure acting on genes underlying species traits linked to fitness.

**Dormant propagules** – A still living seed, cyst, spore or egg that has arrested development and is preserved in ice, soil, sediment, permafrost.

**Experimental Paleoecology:** Experimental studies to test sufficiency and necessity of mechanisms (or combinations or sequences of mechanisms) invoked to explain paleoecological phenomena.

**Migration** – Spatial displacement of organisms leading to shifts of species distributions

**Paleogenomics** – The study of ancient genomes to reveal functional genetic patterns through time, supporting inferences concerning evolutionary adaptation, functional traits, population dynamics, domestication, genetic events preceding extirpations or extinctions, and other patterns of interest.

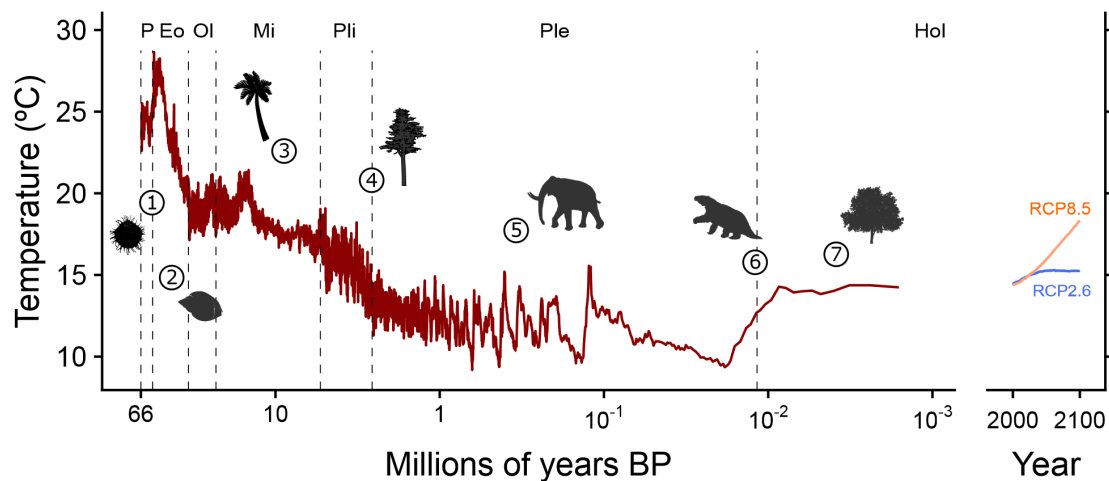
**Phenotypic plasticity** – Ability of individuals of a genotype to alter physiology, morphology, anatomy, phenology, behaviour, or other phenotypic traits in response to environmental change.

**Resurrection ecology** – Study of traits and environmental responses of past populations by hatching or germination of dormant propagules and culturing or cultivation of the organisms.

**Process-based models** - Spatially explicit approaches that simulate the effect of climate and environmental conditions on important vital rates (including population growth, dispersal and plasticity in demographic traits) to explain species distributions and their changes, including range shifts and local extirpations.

**Tolerance** –Ability of a population to persist at a site under environmental change by adaptive evolution, phenotypic plasticity, or both.

### Box 1. Biodiversity responses to past climate change.



**Figure I:** Future climate forcing will surpass those of the previous several million years [2]. Countless individuals in thousands of species across the globe will need to tolerate climate change *in situ*, disperse to more suitable climatic conditions, or undergo extinction. Figure I highlights a number of biodiversity responses directly or indirectly linked to climatic changes along the Cenozoic (last 66 million years).

- 1) During the Paleocene-Eocene Thermal Maximum (~56 million years before present) there were large extinctions in some marine groups (benthic foraminifera), remarkable poleward range shifts in others (dinoflagellates, mammals, reptiles, plants), and high community turnover [97].
- 2) Under a global cooling trend, winters became  $>4^{\circ}\text{C}$  colder across the Eocene-Oligocene boundary, partially driving extinction of many terrestrial mammals in Europe as well as marine invertebrates globally [98].
- 3) Many thermophilous plants shifted their ranges southward and finally went extinct in Europe during the late Miocene global cooling [99].
- 4) More than half (52%) of the cool-temperate European tree genera did not survive the glaciation cycles starting at the end of the Pliocene [100].
- 5) An adaptive mutation of haemoglobin enabled mammoths to tolerate the very low temperatures at high latitudes [27].
- 6) More than 70% of megafauna genera in the Americas and Australia, and 40% in Eurasia, underwent extinction within a relatively brief period of time (5,000-10,000 years) in co-occurrence with climatic changes and human impacts [55].
- 7) Plants in North America migrated northwards between 450 and 2200 km in less than 10,000 years under a warming of 5 degrees [101].

Past temperature data from [102]; future temperature projections (under two greenhouse concentration scenarios: RCP2.5, most benign, and

RCP8.6, most extreme) from [1]. Abbreviations of geological epochs as follows: P = Palaeocene, Eo = Eocene, Ol = Oligocene, Mi = Miocene, Pli = Pliocene, Ple = Pleistocene, Hol = Holocene.

## **Box 2. Correlations are not enough: simulations and process-based models to improve biodiversity forecasts**

Much evidence for the impact of past climate change on biodiversity is based on patterns of co-occurrence between past climatic events and biological responses such as migration, tolerance and extinction. However, the low temporal resolution of available dating techniques for paleorecords often creates difficulties in aligning relevant abiotic dynamics (i.e., climate change, acidification, volcanisms) with biological events. Moving from correlations to causation is challenging because of the co-varying changes in the environment. Recorded biological responses in paleo-records can be used as the testing ground of models deeply rooted in competing ecological and evolutionary theories (Figure 2) [103]. In process-based models [104] these records can serve to inform model parameters, test competing hypotheses and scenarios with the paleorecords, and improve predictions. Nonetheless, predictions may be hindered due to limited data availability that stems from low sampling effort, or because the potential for fossilization is not even across regions and species. Given these constraints, vertebrates and plants from temperate, cold and dry regions of the planet appear by now as the best suited to apply process-based models in the past. The development and further integration of experiments, paleo-records and spatial models on past ecosystems will push the envelope of predictive models of biodiversity and the adequacy of theories and different processes to explain past, and future, biodiversity dynamics under climate change.

**Figure 1. Reconstructing historical patterns of evolutionary change for unravelling mechanisms of genetic and plastic response to anthropogenic environmental changes. a)**

Conceptual framework for the integration of resurrection ecology and predictive models.

Using for example *Daphnia*, dormant propagules can be resurrected (step 1). On resurrected propagules, genetic (G) and phenotypic changes (P) can be quantified over evolutionary time.

Similarly, environmental factors (E) can be inferred from historical records or measured e.g.

via chemical analysis of sediment. The genetic mechanisms (G) underlying phenotypic

changes (P) are identified via a genome wide association analysis (GWAS) (step 2). The

causal link between phenotypic changes (P) and environmental variation (E) is established

via experiments with the support of historical environmental records or reconstruction of

temporal trends in environmental variables (PR). The parameters for predictive models of

phenotypic trajectories are trained on empirical data (G, P and E) from the sedimentary

archive (step 5). Several iterations may be needed to identify the parameters that best fit the

empirical data. Using the optimized parameters, future trajectories of phenotypic (and the

underlying genotypic) trajectories are identified, with a level of uncertainty (step 6). **b)**

Scaling up the approach described in a) to a macroecological scale, across sites -represented

by yellow circle- and biological systems within ecosystems from the tropics to the poles, we

can identify evolutionary and plastic responses of species to global anthropogenic pressures

(within circles from upper left to bottom right: habitat degradation, land-use changes,

invasive species and climate change).

**Figure 2 Cracking the code of biodiversity responses to climate change.** Here we summarize the paleo-data sources, workflow, research challenges and opportunities to incorporate multiple lines of evidence on the magnitude, rate, and processes involved on biodiversity responses to past climate changes for informing biodiversity scenarios. a) Digging in the past (here an example of Late Pleistocene in western Europe) to reconstruct, using a variety of paleo-records (i.e., dated fossil records, ancient molecules) and disciplines (paleoecology, population genomics), the past environmental and biotic conditions and responses to past climate change. b) Main theories and predictions are simulated and tested in process-based models against past recorded trends. c) Opportunities and challenges ahead to ground future biodiversity scenarios in past biological responses and tested biodiversity models.