

# First report on the prevalence and subtype distribution of Blastocystis sp. in dairy cattle in Lebanon and assessment of zoonotic transmission

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1	First report on the prevalence and subtype distribution of <i>Blastocystis</i> sp. in dairy cattle
2	in Lebanon and assessment of zoonotic transmission

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### 24 Abstract

Blastocystis sp. is frequently identified in a wide range of animal hosts, including 25 bovids. Because of its burden and zoonotic potential, this parasite has been sought in 26 27 domestic cattle from various countries, since this livestock may also represent a possible reservoir of human infection. However, epidemiological data regarding the prevalence and 28 ST distribution of *Blastocystis* sp. in this animal group is lacking in Lebanon. Therefore, 29 faecal samples were collected from a total of 254 dairy cattle raised on 55 farms located in 30 31 the North Lebanon region and screened for the presence of the parasite by quantitative real-32 time PCR. The overall prevalence of Blastocystis sp. was shown to reach 63.4% in cattle livestock. Sequence analysis of positive samples indicated the presence of seven STs, with 33 34 predominance of ST10 (44.0%) and ST14 (36.8%) and lower proportions of ST2 (8.0%), ST1 35 (7.2%), ST5 (2.4%), ST3 and ST7 (0.8% each). This survey was the first conducted worldwide reporting ST2 and ST7 in domestic cattle and confirmed that ST10 and ST14 36 represent cattle-adapted STs in view of their high prevalence. Faecal samples from in-contact 37 38 dairy farmers and patients hospitalised in the same Lebanese governorate who reported no contact with cattle livestock were also analysed for the presence of *Blastocystis* sp. The same 39 three STs were identified in both human cohorts, with predominance of ST3, followed either 40 by ST1 or ST2 depending of the group. No other STs, including ST10 or ST14, have been 41 reported. Moreover, even though ST1, ST2 and ST3 were found to be common to dairy cattle 42 43 and farmers cohorts, only one ST3 isolate showed 100% sequence identity between both hosts. Consequently, the presence and low prevalence of ST1, ST2, ST3, ST5 and ST7 44 identified herein in domestic cattle, most of which exhibit low host specificity, could be 45 46 derived from occasional direct exposure to faecal material from human and non-human hosts or by ingestion of contaminated drinking water or food in the enclosure of the farms. 47

- 48 Together with the absence of ST10 and ST14 in the human population, these data suggest that
- 49 cattle play a negligible role as zoonotic reservoirs of *Blastocystis* sp.

- 51 Keywords: *Blastocystis* sp., Dairy cattle, Intestinal parasite, Molecular epidemiology,
- 52 Transmission, Zoonosis

### 54 **1.** Introduction

*Blastocystis* sp. is currently the most common single-celled intestinal parasite found in 55 humans, since its prevalence can greatly exceed 50% in developing countries (El Safadi et al., 56 57 2014; Forsell et al., 2016) and reach an average of around 20% in industrialised regions of Europe (Bart et al., 2013; El Safadi et al., 2016). This protozoan has also been frequently 58 found in many groups of animals, including non-human primates and various groups of 59 60 mammals, birds, reptiles, amphibians and insects (Tan, 2004; Stensvold and Clark, 2016; Yoshikawa et al., 2016; Cian et al., 2017). The principal mode of transmission of *Blastocystis* 61 62 sp. is the faecal-oral route, essentially through consumption of water or food contaminated by environmentally resistant cystic forms of the parasite (Tan, 2008). Due to the very common 63 64 asymptomatic carriage of the parasite in the human population worldwide, its pathogenic 65 potential and clinical significance is remained uncertain (Andersen and Stensvold, 2016). However numerous recent in vitro studies have identified the mechanisms and molecules 66 involved in the virulence of the parasite (Clark et al., 2013; Wawrzyniak et al., 2013; 67 68 Ajjampur and Tan, 2016; Stensvold and Clark, 2016).. Blastocystis sp. infection would thus be associated with various gastrointestinal disorders (Tan, 2008; Tan et al., 2010) and/or 69 70 urticaria (Lepczynska et al., 2015) in numerous human clinical cases. 71 An extensive genetic diversity has been demonstrated within the genus *Blastocystis* 72 based on the comparison of small subunit (SSU) rRNA gene sequences (Noël et al., 2005; 73 Stensvold et al., 2007). Indeed, 17 subtypes (STs) have been identified so far in mammalian and avian hosts (Alfellani et al., 2013a), ten of which infect humans with varying prevalence 74 (Alfellani et al., 2013b; Clark et al., 2013; Ramirez et al., 2016; Stensvold and Clark, 2016). 75 76 Briefly, more than 90% of human isolates belong to ST1 to ST4, in large part likely related to human-to-human transmission (Alfellani et al., 2013b; Clark et al., 2013). Most other STs 77

found in the human population are considered to be of animal origin, with low host

79 specificity and are potentially zoonotic. For instance, ST5 frequently infects pigs (Wang et 80 al., 2014), whereas ST6 and ST7 are predominant in birds (Cian et al., 2017). Based on largescale epidemiological surveys, these last STs were considered to be pig- and avian-adapted 81 82 STs respectively, and corresponding isolates were shown to be possibly transmitted from these animals to their in-contact workers at intensive commercial piggeries and poultry 83 slaughterhouses (Wang et al., 2014; Greige et al., 2018). In addition, ST8, which is common 84 85 in non-human primates, was identified with unexpected high prevalence in their keepers (Stensvold et al., 2009). In view of these data, it became necessary to determine the infection 86 87 rate and ST distribution of *Blastocystis* sp. in other animal groups, such as bovids and particularly livestock cattle. In the main recent molecular epidemiological surveys including a 88 large number of specimens (Zhu et al., 2017; Lee et al., 2018; Masuda et al., 2018; Wang et 89 90 al., 2018; Maloney et al., 2019), the prevalence of the parasite in domestic dairy/beef cattle varied between 2.9% and 54.1%, depending on the country. The potential significant 91 92 occurrence of *Blastocystis* sp. in livestock cattle could thus generate a risk of zoonotic 93 transmission, especially in dairy farmers. Moreover, the same molecular studies and a few others identified ST10 and ST14 as the predominant STs in cattle, likely reflecting bovids as 94 95 natural hosts for both STs (Cian et al., 2017; Zhu et al., 2017; Masuda et al., 2018). Strikingly, ST10 and ST14 have never yet been reported in human infections, suggesting 96 97 minimal risk of zoonotic transmission (Cian et al., 2017; Wang et al., 2018). However, 98 neither of these STs has so far been sought in cattle and their in-contact farmers to confirm this hypothesis. 99

100 The objectives of this study was thus to obtain the first molecular data regarding the 101 prevalence and ST distribution of *Blastocystis* sp. in dairy cattle in Lebanon. A particular 102 purpose was to confirm that bovids represent natural hosts of ST10 and ST14. In addition, the 103 potential zoonotic transmission of the parasite was assessed through a comparative analysis

of the ST distribution of isolates identified in dairy cattle and in-contact farmers, as well as inindividuals reporting no contact with these animals.

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### 107 2. Materials and methods

108 2.1. Ethics statement

The protocol of this study was approved by the Lebanese Minister of Public Health 109 with reference number 2014/4/39716 and by the research ethics committee of Hamidi 110 Medical Centre in Tripoli, Lebanon, with reference number Hamidi-7-2017. Oral and written 111 112 informed consent was obtained from all subjects included in this study after a clear explanation of the research objectives prior to enrolment. The subjects' data were collected 113 114 anonymously (with encryption of the identity of individuals). This study was conducted in 115 accordance with the Code of Ethics of the World Medical Association (Declaration of Helsinki). All dairy cattle samples were collected under the supervision of the farmers. No 116 approval from the Institutional Animal Care and Use Committee or ethics committee was 117 necessary, as no invasive sampling approach was performed. 118

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120 2.2. Study sites and sample collection

The study was conducted in the two governorates comprising the North Lebanon 121 122 region, namely Akkar and North governorates, the latter including the regional capital Tripoli 123 (Figure 1). All healthy adult dairy cattle included in this study were older than 12 months and were of the Holstein-Friesian breed (Bos taurus). In Akkar governorate, 157 faecal specimens 124 were randomly collected from 22 villages and 36 farms from July to November 2016. In 125 126 order to obtain epidemiological data from dairy cattle in different geographical areas in Lebanon, 97 samples were also randomly obtained from 15 villages and 19 farms from July 127 to November 2017 in North governorate, for an overall total of 254 cattle samples screened in 128

129 this survey (one specimen per animal). Briefly, faecal samples were collected in the early 130 morning with the assistance of the breeders and veterinarians. Specimens were either collected directly from the rectum of the animal with a gloved hand or directly from the 131 132 ground immediately after defecation to prevent potential contamination between animals. Each fresh faecal sample was recovered in a sterile plastic container, respecting the 133 conditions of asepsis and hygiene. The containers were stored in ice and then immediately 134 transported to the Laboratoire Microbiologie Santé et Environnement (LMSE) at the AZM 135 Centre in Tripoli. To evaluate the zoonotic potential of *Blastocystis* sp. isolates identified in 136 137 dairy cattle, human stool samples were obtained during the same period from 40 farmers working at the same 19 dairy cattle farms located in North governorate and from 40 138 individuals followed up for different pathologies or who presented for routine medical check-139 140 ups at Hamidi Medical Centre in Tripoli who reported no contact with cattle (control 141 population). A standardised questionnaire was designed to collect data of interest, such as age, sex, residency and gastrointestinal symptoms (abdominal pain, bloating, constipation, 142 diarrhoea and/or vomiting) for each volunteer, as well as number of years as in-contact 143 worker with cattle for the cohort of dairy farmers. 144

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146 2.3. DNA extraction

Total genomic DNA was extracted directly from approximately 250 mg of animal or
human faecal samples using the QIAamp DNA Stool Mini Kit (Qiagen GmbH, Hilden,
Germany) according to the manufacturer's recommended procedures. The DNA was eluted in
200 μl of elution buffer and stored at -20°C at the LMSE in Tripoli. The DNA samples were
then transported to the Pasteur Institute in Lille (France) for molecular screening and
subtyping of *Blastocystis* sp.

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154 2.4. Detection and molecular subtyping of *Blastocystis* sp. isolates

SSU rRNA gene detection of the parasite was performed by quantitative real-time 155 PCR (qPCR) using 2 µl of extracted DNA and the Blastocystis-specific primer pair 156 157 BL18SPPF1 / BL18SR2PP, as described previously by Poirier et al. (2011). DNA extraction controls (isolation of DNAs without stool and from a *Blastocystis* sp.-negative stool) 158 subsequently used in qPCR assays and positive (DNA obtained from Blastocystis sp. ST7 159 160 strain B axenic culture maintained in the laboratory) and negative (DNA matrix replaced by water) qPCR controls were performed. The qPCR product from each positive sample was 161 162 purified and sequenced in both strands by Genoscreen (Lille, France). For 3 human samples obtained from breeders and 3 others collected from patients followed at Hamidi Medical 163 Centre in Tripoli, sequence chromatogram analysis revealed the presence of double traces, 164 165 suggesting mixed infection by different STs. Mixed signals were also generated for numerous qPCR products obtained from dairy cattle samples. All 6 human samples and a selection of 9 166 animal samples corresponding to mixed infections were thus re-analysed by non-qPCR using 167 168 the same primer pair as for qPCR. Non-qPCR amplification, as well as purification and cloning of the non-qPCR product, were performed as described previously (Cian et al., 2017). 169 Briefly, the purified non-qPCR product cloned in the T-vector, pCR 2.1-TOPO (Invitrogen, 170 Carlsbad, USA) was amplified in Escherichia coli One Shot TOP10 competent cells and 171 minipreparations of plasmid DNA were done using the NucleoSpin Plasmid kit (Macherey-172 173 Nagel, Düren, Germany). Five positive clones containing inserts of the expected size were selected arbitrarily and sequenced on both strands. The SSU rRNA gene sequences obtained 174 in this study were deposited in GenBank under accession numbers MH883051 to MH883225. 175 176 The sequences obtained were compared with all Blastocystis sp. homologous sequences available from the National Centre for Biotechnology Information (NCBI) using the 177 nucleotide Basic Local Alignment Search Tool (BLAST) program. The STs were identified 178

by determining the exact match or closest similarity to all known mammalian and avian *Blastocystis* sp. STs according to the most recent classification of the parasite (Alfellani et al.,
2013a).

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183 2.5. Statistical analyses

Statistical analyses were performed using GraphPad Prism software, version 6
(GraphPad Software, La Jolla, CA, USA). The chi-squared test was used to compare the *Blastocystis* sp. prevalence between the Akkar and North governorate dairy cattle groups.
Fisher's exact test was used to compare the ST distribution of the parasite between the two
human cohorts, as well as those of the two animal groups. All tests were two-sided and the
general significance level was set at a *P* value of below 0.05.

190

## 191 **3. Results and discussion**

A total of 254 single dairy cattle samples collected from 37 villages and 55 farms located 192 in two governorates of North Lebanon were screened in this study (Table 1). Of these 193 samples, 157 were obtained at Akkar governorate from cattle raised on 36 different farms. Of 194 these 157 samples, 78.3% (n = 123) were identified as positive for *Blastocystis* sp. by qPCR. 195 Of the 97 dairy cattle samples collected from 19 different farms in North governorate, 38 196 (39.2%) were also positive for the parasite by qPCR. The statistically significant difference of 197 198 Blastocystis sp. prevalence in cattle between the two geographical regions (chi-squared test, Z = 4.498, P < 0.05) may possibly be due to the lower quality hygiene and sanitary conditions 199 at farms in the rural governorate of Akkar, thus facilitating the transmission of the parasite 200 201 between livestock. By combining the molecular data obtained from the two governorates, the prevalence of *Blastocystis* sp. reached 63.4% (161/254) among dairy cattle samples. This 202 high infection rate in Lebanon was roughly similar to that observed using molecular methods 203

204 in other countries, including Japan (54.1%) (Masuda et al., 2018), Thailand (50.0%) (Udonsom et al., 2018), Colombia (80.0%) (Ramirez et al., 2014) and Libya (41.7%) 205 (Alfellani et al., 2013a) but was significantly higher than that reported in Korea (6.7%) (Lee 206 207 et al., 2018), China (9.5% and 10.3%) (Zhu et al., 2017; Wang et al., 2018, respectively), Iran (9.6%) (Badparva et al., 2015), the United Kingdom (22.6%) (Alfellani et al., 2013a) and the 208 United States (19.1% and 2.9%) (Fayer et al., 2012; Maloney et al., 2019, respectively). 209 210 Additional surveys also revealed a prevalence of 71.0%, 21.4% and 1.8% in Japan (Abe et al., 2002), Brazil (Franco Moura et al., 2018) and Spain (Quilez et al., 1995) respectively, 211 212 using direct light microscopy of faecal smears, in the knowledge that this identification method has been shown to be less sensitive than PCR (Poirier et al., 2011). Although the 213 214 prevalence of the parasite in domestic cattle may vary between geographical areas, it is 215 nevertheless common worldwide, thus generating a potential risk of transmission to incontact humans. 216

To start assessing the zoonotic potential of the parasite, single stool samples were 217 collected from a total of 80 volunteers divided into two cohorts (Table 1). The first cohort 218 included 40 individuals (26 males and 14 females), all working on dairy cattle farms in North 219 220 governorate for a period of between 6 months and 30 years, with a median of 8 years, 221 indicating a potentially long period of contact with animals. The age of the individuals in this 222 cohort was between 22 and 65 years (mean age of  $36 \pm 10$  years) and 21 of these 40 breeders 223 (52.5%) were shown to be infected with *Blastocystis* sp. by qPCR. Moreover, only 5 breeders presented gastrointestinal symptoms and 3 of them were infected by the parasite. The second 224 cohort (or control group) consisted of 40 patients (12 males and 28 females) aged 21 to 70 225 226 years (mean age of  $24 \pm 12$  years), hospitalised at Hamidi Medical Centre in Tripoli for various pathologies or medical check-ups, who reported no contact with cattle. 62.5% of the 227 228 patients (25/40) presented one or more gastrointestinal symptoms. Using qPCR, the

229 prevalence of *Blastocystis* sp. in this second human cohort was not significantly different to 230 that of the dairy farmers, i.e. 57.5% (23/40). In addition, 52.0% of symptomatic patients (13/25) were infected by the parasite. Interestingly, the infection rate observed in each of 231 232 these two groups (52.5% and 57.5% respectively) was very similar to that recently reported in North Lebanon region in a large cohort of schoolchildren (63.0%) (Osman et al., 2016) and 233 two groups of individuals working in a poultry slaughterhouse (56.0%) or hospitalised at 234 Hamidi Medical Centre in Tripoli (54.0%) (Greige et al., 2018). On the other hand, the 235 similar prevalence observed between all these Lebanese groups of individuals suggested that 236 237 people working closely with animals, including for instance domestic cattle or poultry (Greige et al., 2018) but also pigs (Wang et al., 2014), did not systematically have a higher 238 risk of acquiring Blastocystis sp. infection as previously proposed (Rajah Salim et al., 1999), 239 240 without affecting the possibility that some STs are of zoonotic origin.

In a second step, the genetic diversity of Blastocystis sp. isolates was explored in dairy 241 cattle and human cohorts. In this study, all partial SSU rRNA gene sequences obtained from 242 243 animal or human samples showed 99 to 100% identity with homologous sequences available in databases allowing the direct subtyping of the corresponding isolates (Table 1). Regarding 244 245 the 123 positive dairy cattle samples identified in Akkar governorate, 84 corresponded to single infections by either ST1 (n = 4, 3.3%), ST5 (n = 3, 2.4%), ST7 (n = 1, 0.8%), ST10 (n246 = 40, 32.5%) and ST14 (n = 36, 29.3%). The 39 remaining positive dairy cattle samples 247 248 (31.7%) from this governorate presented mixed infections with at least 2 unidentified STs according to the resulting sequence chromatograms. Of the 38 positive animal specimens 249 identified in North governorate, 21 corresponded to single infections with the detection of 250 251 ST2 (n = 1, 2.6%), ST3 (n = 1, 2.6%), ST10 (n = 12, 31.6%) and ST14 (n = 7, 18.4%) and the 17 other samples (44.8%) to mixed infections. Given the large number of mixed infections 252 identified in animal samples collected in both governorates, only a selection of them was thus 253

254 re-analysed by non-qPCR to have a partial overview of the STs found in these specimens. Therefore, one sample from each farm presenting mixed infection in North governorate was 255 selected. Cloning of the non-qPCR products was performed from these 9 representative 256 257 animal specimens for subtyping, and 5 positive clones were selected arbitrarily and sequenced for each cloning. Seven of the 9 selected samples showed mixed infections with 258 two different STs (ST2/ST1, n = 4; ST2/ST10, n = 1; ST2/ST14, n = 2) whereas the two 259 remaining samples harboured 3 different STs (ST1/ST2/ST10 and ST2/ST10/ST14 260 respectively). Taking into account these 9 mixed infections consisting of either two or three 261 262 different STs, the ST distribution in dairy cattle from this governorate was as follows: ST1 (n = 5), ST2 (n = 10), ST3 (n = 1), ST10 (n = 15) and ST14 (n = 10). The major difference 263 observed between these two animal cohorts in terms of distribution of STs was the presence 264 265 of ST2 only in the group of North dairy cattle, while it was absent from the Akkar group of animals (Fisher's exact test, 24.4% vs. 0%, P < 0.05). By merging all molecular data 266 obtained from dairy cattle samples in North Lebanon region, a total of 125 isolates were 267 subtyped and belonged to 7 different STs with, in order of prevalence, ST10 (55/125, 44.0%), 268 ST14 (46/125, 36.8%), ST2 (10/125, 8.0%), ST1 (9/125, 7.2%), ST5 (3/125, 2.4%), ST3 and 269 ST7 (1/125 each, 0.8%). Herein, ST10 and ST14 were thus largely predominant by 270 accounting for more than 80% of the subtyped isolates. As shown in Table 2 reporting all the 271 272 isolates subtyped from domestic cattle in various geographical areas, ST10 and ST14 also 273 globally represented the most widely distributed STs (64.5% of total isolates). This was clearly the case, for instance, in Denmark (Stensvold et al., 2009), Japan (Masuda et al., 274 2018), Korea (Lee et al., 2018), China (Zhu et al., 2017) and Libya (Alfellani et al., 2013a). 275 276 Strikingly, ST10 and ST14 were the only two STs identified in cattle in two studies conducted in the USA (Santin et al., 2011; Fayer et al., 2012). Therefore, all of these data 277 confirmed that domestic cattle would be natural hosts for *Blastocystis* sp. ST10 and ST14. In 278

279 addition to these two predominant STs, five minor STs (ST1, ST2, ST3, ST5 and ST7) were identified in the livestock investigated in this study. Briefly, it was the first report of the 280 presence of ST2 and ST7 in domestic cattle, while the three other STs had previously been 281 282 detected with varying prevalence in the same animal group in different countries (Table 2). Regarding the dairy farmers cohort, 18 individuals presented single infections by the 283 parasite, and 3 others had mixed infections with two STs. With the addition of these mixed 284 285 infections, a total of 24 isolates was subtyped. As shown in Table 1, ST3 was predominant (12/24, 50.0%), followed by ST2 (7/24, 29.2%) and ST1 (5/24, 20.8%). In the case of the 286 287 second human cohort comprising patients enrolled at Hamidi Medical Centre in Tripoli, 20 and 3 of them presented single and mixed infections with two STs respectively. Therefore, a 288 total of 26 isolates was subtyped and three different STs were identified as follows: ST3 289 290 (13/26, 50.0%), ST1 (8/26, 30.8%) and ST2 (5/26, 19.2%). Interestingly, no other ST was identified in these two human groups. Moreover, although the number of samples analysed in 291 each cohort remained limited, the distribution of the different STs was not significantly 292 293 different between the two human groups (Fisher's exact test, P = 0.67). Strikingly, this distribution with predominance of ST3 followed by either ST1 or ST2 was also similar to that 294 295 observed in previous studies conducted recently among patients at the same hospital in Tripoli, staff members working in poultry slaughterhouses in North Lebanon (Greige et al., 296 297 2018) and among schoolchildren in Tripoli (Osman et al., 2016).

By analysing all the molecular data obtained herein, the zoonotic potential of *Blastocystis* sp. was evaluated through the comparison of the ST distribution reported in animal and human cohorts in North Lebanon. Despite the high prevalence of ST10 and ST14 in cattle livestock, neither ST was identified especially in dairy farmers despite these workers having intimate, long-term and repeated contact with these animals. To our knowledge, these two STs have also never been documented in human infections worldwide (Clark et al., 2013;

304 Stensvold and Clark, 2016), suggesting that humans would not be susceptible to their infection. Another hypothesis that remains to be further confirmed could be that these two 305 STs produce non-resistant cystic environmental forms, thus limiting possible transmission of 306 307 the parasite to another host through indirect contact. ST5 was also infrequently identified in dairy cattle, as in previous studies conducted in various countries (Table 2) and herein only in 308 animals raised at Akkar governorate. Recently, ST5 was proposed as a pig-adapted ST (Wang 309 310 et al., 2014) and consequently cannot likely be assigned to domestic cattle at this time. Its zoonotic potential was confirmed through the frequent infection of piggery staff by this ST 311 312 (Wang et al., 2014) and pig-rearing villagers (Yan et al., 2007). However, despite the large number of samples screened, ST5 has not yet been identified in this study as in other 313 314 Lebanese human cohorts (Osman et al., 2016; Greige et al., 2018). The only known case of 315 ST7 infection in cattle was identified in this study at a farm located at Akkar governorate. This ST was proposed as an avian-adapted ST with potential zoonotic risk (Alfellani et al., 316 2013a; Cian et al., 2017) and its sporadic prevalence in cattle livestock likely reflected 317 opportunistic infection through contact with bird faeces. Again, ST7 has never been yet 318 identified in the Lebanese human population (Osman et al., 2016; Greige et al., 2018), as 319 confirmed herein. Interestingly, three STs (ST1, ST2 and ST3) were found in common in 320 dairy cattle and their in-contact farmers in North governorate, suggesting potential 321 322 transmission between the hosts. However, only six animal isolates belonging to these STs and 323 as part of single or mixed infections were identified on 4 different farms at which dairy farmers were infected by the same STs. By comparing the corresponding dairy cattle and in-324 contact farmer isolate sequences, 100% identity was shown for only one ST3 isolate infecting 325 326 both hosts. Since ST3 is known to be predominantly harboured by the human population (Clark et al., 2013; Stensvold and Clark, 2016) including Lebanon and not frequently found 327 in domestic cattle as shown in Table 2, this case of Blastocystis sp. ST3 infestation in 328

329 livestock can likely be explained by reverse zoonosis from human handlers to animals, as suggested in previous studies focused on various animal hosts (Ramirez et al., 2014) and pigs 330 (Wang et al., 2014). Regarding the two others, ST1 and ST2, considered to be STs linked to 331 human infection (Clark et al., 2013; Stensvold and Clark, 2016), their origin in Lebanese 332 domestic cattle remains uncertain according to our sequence data. Globally, the low 333 prevalence of ST1, ST2, ST3, ST5 and ST7 observed in the Lebanese cattle population 334 335 suggests that these animals are unlikely to be natural hosts of these STs and are potentially opportunistically infected through direct exposure to faecal material from human and non-336 337 human hosts or by ingestion of contaminated drinking water or food in their environment, as proposed in both stray and household dogs (Wang et al., 2013; Osman et al., 2015). The 338 difference in the prevalence of ST2 observed between dairy cattle raised in the two Lebanese 339 340 governorates, while this ST is known to infect humans and numerous animal groups (Clark et al., 2013; Stensvold and Clark, 2016; Cian et al., 2017), reinforced this hypothesis by 341 highlighting different infection sources in two different Lebanese geographical areas. 342 However, since domestic cattle carry potential zoonotic STs, these animals are thus capable 343 of shedding the corresponding isolates and may therefore represent secondary zoonotic 344 reservoirs of infection. 345

346

### 347 4. Conclusion

This study presents the first molecular epidemiological data on *Blastocystis* sp. infection in cattle livestock in Lebanon. Through both the high number of samples screened and isolates subtyped, it provided new insights into the prevalence and ST distribution of the parasite in this animal host. This survey conducted in North Lebanon demonstrated that dairy cattle were frequently infected by *Blastocystis* sp., with an overall prevalence of the parasite exceeding 60%. This clearly raised the question of the impact of this infection in the bovid

354	sector, which is of major importance in this country. The variety of STs colonising this
355	animal group was also highlighted, with a total of seven STs identified, including ST1, ST2,
356	ST3, ST5, ST7, ST10 and ST14. ST10 and ST14 were largely predominant in this animal
357	population, confirming that both STs could be considered cattle-adapted STs. Others STs
358	were all identified with low prevalence and their presence was suggested to be related to
359	opportunistic contamination from various animal or even human and environmental sources.
360	Finally, by comparing the ST distribution of the parasite in cattle and in-contact dairy
361	farmers, as well as in individuals without contact with bovids, the risk of zoonotic
362	transmission of Blastocystis sp. was highlighted to be likely minimal. However, further large-
363	scale studies should be conducted to fully understand the exact role of domestic cattle in the
364	epidemiology of Blastocystis sp. and as potential sources of transmission of the parasite to
365	humans, at least as secondary reservoirs.
366	
367	Conflicts of interest
368	The authors declare that there are no competing interests.
369	
370	Authors' contributions
371	D.E.S., M.O., G.C., M.H. and E.V. conceived the study and designed the experiments.
372	S.G., D.E.S., S.K., N.G. and M.O. collected the samples and/or performed the experiments.
373	M.B. performed the statistical analyses. S.G., D.E.S., S.K., S.B.V., M.Che, M.Cha, G.C.,
374	M.H. and E.V. participated in the interpretation of data. S.G., D.E.S., S.K. and E.V. wrote the
375	manuscript. All authors have read and approved the final version of the manuscript.
376	
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## 523 **Table 1**

<sup>524</sup> Prevalence of *Blastocystis* sp. infection and ST distribution in animal and human cohorts in

Infection	Akkar dairy cattle	North dairy cattle	North breeders	North hospital patients			
	( <i>n</i> = 157)	( <i>n</i> = 97)	( <i>n</i> = 40)	( <i>n</i> = 40)			
% positive	78.3% (123)	39.2% (38 <sup>a</sup> )	52.5% (21 <sup>b</sup> )	57.5% (23 <sup>c</sup> )			
ST1	4	5	5	8			
ST2	0	10	7	5			
ST3	0	1	12	13			
ST5	3	0	0	0			
ST7	1	0	0	0			
ST10	40	15	0	0			
ST14	36	10	0	0			
$\mathbf{MI}^{\mathrm{d}}$	39	8	0	0			

525 North Lebanon

527 characterisation of a total of 41 isolates

<sup>528</sup> <sup>b</sup>Mixed infections by 2 STs identified in 3 samples resulting in the molecular characterisation

- 529 of a total of 24 isolates
- <sup>c</sup>Mixed infections by 2 STs identified in 3 samples resulting in the molecular characterisation
- 531 of a total of 26 isolates
- <sup>532</sup> <sup>d</sup>MI, Mixed infections with unidentified STs

<sup>&</sup>lt;sup>a</sup>Mixed infections by 2 or 3 STs identified in 9 selected samples resulting in the molecular

# **Table 2**

536														
537	Country	No. of isolates identified		Blastocystis sp. STs									Reference	
538		by sequencing												
539			ST1	ST2	ST3	ST4	ST5	ST6	ST7	ST10	ST12	ST14	Others	
540														
541	Japan	45	0	0	0	0	0	0	0	1	0	44	0	Masuda et al., 2018
542	Japan <sup>a</sup>	8	1	0	1	0	6	0	0	0	0	0	0	Yoshikawa et al., 2004
543	Japan <sup>a</sup>	3	1	0	2	0	0	0	0	0	0	0	0	Abe et al., 2003
544	Korea	30	6	0	0	0	5	0	0	9	0	10	0	Lee et al., 2018
545	China	14	0	0	2	0	0	0	0	10	0	2	0	Wang et al., 2018
546	China	54	0	0	0	2	1	0	0	41	0	10	0	Zhu et al., 2017
547	Thailand	6	0	0	0	0	0	0	0	2	4	0	0	Udonsom et al., 2018
548	Iran <sup>a</sup>	17	0	0	4	0	11	2	0	0	0	0	0	Badparva et al., 2015
549	Libya	10	0	0	0	0	2	0	0	6	0	2	0	Alfellani et al., 2013a

535 ST distribution of *Blastocystis* sp. in domestic cattle around the world

550	Lebanon	125	9	10	1	0	3	0	1	55	0	46	0	Present study
551	UK	5	1	0	0	0	1	0	0	3	0	0	0	Alfellani et al., 2013a
552	Denmark	25	0	0	0	0	3	0	0	22	0	0	0	Stensvold et al., 2009
553	Colombia	20	12	0	8	0	0	0	0	0	0	0	0	Ramirez et al., 2014
554	USA	7	0	0	0	0	0	0	0	7	0	0	0	Santin et al., 2011
555	USA	11	0	0	0	0	0	0	0	8	0	3	0	Fayer et al., 2012
556	USA	76	0	0	4	18	27	0	0	5	0	8	14 <sup>b</sup>	Maloney et al., 2019
557	Total	456	30	10	22	20	59	2	1	169	4	125	14	
558														

<sup>559</sup> <sup>a</sup>PCR using ST-specific primers allowing the detection of only ST1 to ST7

<sup>560</sup> <sup>b</sup>Including STs 17, 21, 23, 24, 25 and 26 proposed by Maloney et al., 2019

# **Figure captions**

- **Fig. 1.** Study area in Lebanon and description of the animal and human cohorts screened in
- this epidemiological survey.

