

Biosensors to diagnose Chagas disease: A review

María-Isabel Rocha-Gaso, Luis-Jesús Villarreal-Gómez, Denis Beyssen, Frédéric Sarry, M.-A Reyna-Carranza, Carlos-Napoleón Ibarra-Cerdeña

▶ To cite this version:

María-Isabel Rocha-Gaso, Luis-Jesús Villarreal-Gómez, Denis Beyssen, Frédéric Sarry, M.-A Reyna-Carranza, et al.. Biosensors to diagnose Chagas disease: A review. Sensors, 2017, 17 (11), pp.2629. 10.3390/s17112629 . hal-02392999

HAL Id: hal-02392999 https://hal.science/hal-02392999

Submitted on 4 Dec 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés. Elsevier Editorial System(tm) for Diagnostic Microbiology and Infectious Disease Manuscript Draft

Manuscript Number: DMID-16-758

Title: Biosensors to diagnose Chagas disease: A review

Article Type: Review article

Corresponding Author: Dr. Carlos N. Ibarra-Cerdeña, Ph.D.

Corresponding Author's Institution: Centro de Investigación y Estudios Avanzados del IPN (Cinvestav) Unidad Mérida

First Author: Maria I Rocha-Gaso, Ph. D.

Order of Authors: Maria I Rocha-Gaso, Ph. D.; Luis J Villarreal-Gómez, Ph. D.; Denis Beyseen, Ph. D.; Frederic Sarry, Ph. D.; Marco A Reyna-Carranza, Ph. D.; Carlos N. Ibarra-Cerdeña, Ph.D.

1	1	Biosensors to diagnose Chagas disease: A review
2 3 4	2	MI. Rocha-Gaso ¹ , LJ. Villarreal-Gomez ² , D. Beyssen ³ , F. Sarry ³ , MA. Reyna-Carranza ² , C. N.
5 6	3	Ibarra-Cerdeña ^{4,*}
7 8	4	¹ Universidad de Quintana Roo, Unidad Cancún, Cancun, Mexico;
9 10	5	² Universidad Autónoma de Baja California, Mexico;
11 12	6	³ Institut Jean Lamour, Vandoeuvre les Nancy, France;
13 14 15	7	⁴ Departamento de Ecología Humana, Cinvestav Unidad Mérida, Merida, Mexico.
16 17	8	*Corresponding author's e-mail: cibarra@cinvestav.mx
19 20	9	Abstract: Chagas disease (CD), which mostly affects underprivileged people, has turned into one
21 22	10	of Latin America's main public health problems. Prevention of the disease requires early diagnosis,
23 24	11	initiation of therapy, and regular blood monitoring of the infected individual. However, the majority
25	12	of the infections go undiagnosed because of general mild symptoms and lack of access to medical
∠o 27 28	13	care. Therefore, more affordable and accessible detection technologies capable of providing early
29	14	diagnosis and parasite load measurements in settings where CD is prevalent are needed to enable
31	15	enhanced intervention strategies. This review discusses currently available detection technologies
32 33 24	16	and emerging biosensing technologies for a future application to CD. Even if biosensing
35	17	technologies still require further research efforts to develop portable systems, we arrive to the
36 37 29	18	conclusion that biosensors could improve diagnosis and the patients' treatment follow-up, in terms
30 39	19	of rapidity, small sample volume, high integration, ease of use, real-time and low cost detection
40 41 42	20	compared to current conventional technologies.
43 44	21	Keywords: Chagas disease, biosensors, detection technologies, diagnosis, neglected diseases.

1. Introduction

Chagas disease (CD), discovered in 1909 by the Brazilian physician Carlos Chagas (1), nowadays has turned into one of Latin America's main public health problems (2). Based in disability-adjusted life-years as a measure of disease burden, CD figure as the most important parasitic vector-borne illness in the Region of Americas, seven times higher than Malaria and up to three times more than Dengue (3), and yet it is still absent in the agenda of the public health policies and practices of many endemic countries (4, 5). More generally, ranks fourth in mortality and eighth in morbidity

among world neglected tropical diseases (6), and is estimated that between eight and eleven million people are infected, while 100 million are at risk of acquiring the disease; the main cause being to live in proximity with disease vectors (7). CD is caused by the parasitic presence of the Trypanosoma cruzi in the organism, which is mainly transmitted by contamination with infected feces of blood-sucking triatomine vectors during a human blood meal. Nevertheless, it can also be transmitted through blood transfusions, organ transplants, infected mothers to their unborn children and ingestion of contaminated food (i. e. Oral transmission; WHO, 2002). Although disease progression can be associated with the mechanism of infection, with oral transmission causing the most severe outbreaks (9), people living at risk regions are susceptible to polyparasitism (i.e. Coinfections and superinfections with different strains of T. cruzi), with unknown effects in the variability of the disease progression and response to treatments (10).

CD mostly affects underprivileged people and the majority of the T. cruzi infections go undiagnosed because of general mild symptoms and lack of access to medical care (11). Due to this fact, CD is considered as a Neglected Tropical Disease whose improvement in diagnosis and treatment today requires research and development efforts with non-profit interests. The highest prevalence of Chagas disease has been reported in Bolivia (6.75-15.4 %), followed by Paraguay (0.69-9.3 %), Panama (0.01-9.02 %), Brazil (0.8-1.30 %), Mexico (0.5-6.8 %) and Argentina (4.13-8.2 %) (12). Citing a case, this disease causes almost 6% of the annual deaths in Mexico and the seroprevalence can roughly be estimated at least in 3% due to a lack of active epidemiological surveillance (most cases are detected during blood screening procedures in blood banks). Yet, less than 0.5% of the infected individuals have access to treatment in this country as a result of anachronisms in the normativity, among other failures in the public health system (13).

Even if CD mainly affects tropical countries, with nowadays ease of traveling and migration, other countries are also being affected by this infection (14). Several cases have been reported in USA, Canada, Europe and in Western Pacific regions like Japan and Australia (14–16). Notably, CD continues to be an inconspicuous public health problem, with limited medical awareness, either because it is commonly targeted in people with relatively low medical access or because it can currently occur in unexpected regions. Thus, the treatment of CD urgently needs to generalize and standardize diagnostic procedures.

58 Biosensors are relatively new analytical devices that can help to detect the presence of specific 59 compounds and pathogens in liquid environments and complex mixtures like: water and blood

serum. Although these devices have been formerly used in the alimentary industry (mostly to detect toxins and infectious pathogens), they are been increasingly used to diagnose human diseases (17). Therefore, these devices can be employed for the diagnosis of CD. The development of biosensors requires a biological active component to be immobilized onto the surface of a transducer. The selective recognition layer, towards T. cruzi specific antigens present in patients' blood serum, can selectively detect the target analyte generating a signal response in the sensor (see Figure 1). Depending on their transducing principle biosensors can be electrochemical, acoustic or optic.

In this work, we firstly introduce a brief description of the disease. Secondly, we present a review of biosensor technologies whose applicability to diagnose CD has been investigated. Finally, we mention the benefits and drawbacks of applying biosensors as solutions to this major public health issue and the infrastructure required to conduct biosensor experiments for this application.

72 2. Brief description of CD and current needs

CD passes through two successive stages: an acute phase and a chronic phase. The acute phase occurs at the following 6-8 weeks after infection. The acute phase is followed by the chronic phase of CD, which lasts for the rest of the life of the infected individual, and has different forms. In the



Figure 1. General scheme of a biosensor detection strategy. A biosensor is composed of a biochemical interface where specific bio-species are absorbed; a transducer which translates the recognition event to another physical response that can be measured and an electronic system which acquires and records the signal.

indeterminate form, an equilibrium between the parasite and the immunological response of the infected individual is reached and most infected patients appear healthy, with no evidence of organ damage that could be found by current standard methods of clinical diagnosis (8). About 50-70% of infected individuals will remain in this condition for the rest of their lives. However, several years after the chronic phase has started, 10-40% of infected individuals will pass to the cardiac form of the disease and will develop injuries of various organs, mainly the heart, the digestive system, and occasionally, the peripheral nervous system (8, 18). These important symptomatic changes occur 10-20 years after the acute phase of the disease and include a broad range of types of damage. The clinical manifestations vary from mild symptoms to heart failure and, frequently, sudden cardiac death (18). The acute phase of CD is recognized only in an estimated 1-2% of all individuals acquiring the infection (8) due to a lack of access to sufficient medical care. Thus, more than any other parasitic disease, CD is closely related to social and economic development. Paradoxically, acute phase is the most appropriate period for drug treatment, showing relatively high levels of sero-conversion, while organ damage is prevented (19). Conversely, it has been shown that drug treatment with Trypanocidal therapy in patients with established Chagas cardiomyopathy, can cause seroconversion, but does not stop cardiac clinical deterioration (20).

Given that no vaccine is currently available to prevent CD, vector control, diagnosis tests, opportune drug treatment, and clinical follow-up are the most effective methods to fight against the disease (21). Nevertheless, all these measures suffer of several hindrances imposed by the synergistic negative effects of diverse vulnerability components of CD risk, such as ecological factors (i.e. land-use changes) -that are broadening the contact zones between humans and parasites (Lopez-Cancino et al., 2015)-, the chronic failure of health care policies hindering the reduction of CD incidence (5), the limited awareness of physicians (22), and several socio-cultural practices that perpetuate CD exposure in endemic regions (23). Health policies to control/reduce vectorial exposure to CD in Latin America are challenged for the relatively high diversity of vectors that shows a broad environmental tolerance. Likewise, a wide territory of America is suitable for CD vectorial transmission (24-27).

103 In order to face the epidemiological challenges due to the increasing complexity of interactions 104 among the transmission routes of *T. cruzi* in endemic and non-endemic countries, access to early 105 diagnostic and treatment seems as the most cost-efficient ways to reduce the CD burden (11). 106 Several paths of scientific advances and discoveries envisage an optimistic future to reduce CD

burden coming from a better understanding of CD transmission and management toward its interruption (28). The fact that the World Health Organization (WHO) has made a commitment with Bill and Melinda Gates Foundation and other stakeholders to control the most neglected diseases by 2020, including CD among them, opens an opportunity window to orientate research priorities. This commitment, launched in 2012, called the London Declaration was (http://www.unitingtocombatntds.org/). However, claims have been raised to urge implicated organizations to set the needed measures to reach these goals, among which improved diagnosis ¹³ 114 (i.e. precision and accessibility) seems as one of the most important first steps (5).

16 115 Blood banks in endemic countries require fast and secure screening method for small and medium health facilities, since screening for CD is mandatory in some endemic countries (8). Several 18 116 20 117 countries do not have an active program for CD detection and depend only for the blood bank $_{22}$ 118 reports. The most important reason of this fact is the low feasibility to detect Chagas in patients. Since, CD do not produce particular symptoms, there are not incentives from society. Nevertheless, the bug bite is quite notorious and people can suspect that they are infected with T. cruzi when a chinchoma¹ appears. In such cases, people can search medical assistance and be subjects of a blood test.

Climate change and global warming increase the risk of rising CD burden in some regions. Climate change impacts on vector-borne diseases (29) and is undoubtedly detonating variables that make the CD transmission become potentially dangerous, as the WHO points out (30). Nevertheless, the ³⁸ 126 incidence of CD can be greatly reduced by residual insecticide-based vector control programs that 40 127 decrease the populations of the transmitting vectors and by improving housing (31).

43 128 3. Current detection technologies and their limitations

Currently, laboratory methods are employed to diagnose CD. Depending on the patients' phase of infection some are more convenient than others. During the acute phase of CD, a large number of parasites are present in the peripheral blood and can be diagnosed by direct microscopical observation of fresh blood (parasitological test). However, for the chronic phase of CD the diagnosis is not possible, due to the scarce parasitemia. Therefore, the immunodiagnosis is widely used since nearly all T. cruzi-infected individuals in the chronic phase develop antibodies against the complex antigenic mixture of the parasite (8).

б

¹ Name of the inflammatory injury after a bug bite.

Several immunodiagnosis tests are available, but mainly three conventional tests are widely used: indirect haemagglutination (IHA), indirect immunofluorescence (IIF) and Enzyme Linked Immuno Assay (ELISA). These tests present several limitations such as: (i) cross-reactivity with other parasites; (ii) not 100% of sensitivity; (iii) the need to be performed in a laboratory; and (iv) a long time is required to obtain the results. IHA test results can be obtained in about two hours, whereas IIF results can be obtained after numerous steps in two hours and ELISA takes several hours to carry out, including prior sensitization of microplates with T. cruzi antigens for about 12 hours (6). All these tests have to be performed in centralized laboratories; some of them require sophisticated equipment and skilled technicians. Since none of these tests have a sensitivity of 100%, the WHO recommends conducting at least two conventional tests for a definitive diagnosis of T. cruzi infection (8).

More recently, non-conventional tests, like rapid lateral flow (RLF) tests, are commercially available in the market to detect T. cruzi infection using whole blood, serum or plasma (Sánchez-Camargo et al. 2014). These tests are based in different tests principles: immunochromatography, particle agglutination, immunofiltration or immunodot. They provide fast results (between 5 to 60 min reading times) without the need of electrical equipment and they require low volume samples (5 to 150 µl). However, the sensitivities and specificities of such tests are lower than that of conventional tests and they only provide qualitative or semi-quantitative results, which prevents obtaining important test information like genetic lineage of the T. cruzi (32) and the immunoreaction kinetics.

From a prospective point of view, it is important to discuss about another kind of technology, which 42 157 could be deployed for Chagas Disease diagnosis in the near future. Over the last decade, Shear-44 158 Horizontal (SH) Surface Acoustic Wave (SAW) immunosensors (Love-SAW with a guiding layer) 46 159 have been developed for the diagnosis of various diseases (33). Such immunosensors exhibit a $_{48}$ 160 high sensitivity and a very low limit of detection (in the order of $pg/\mu l$ of blood serum). This ₅₀ 161 technology give very relevant results for the detection of antibodies, specific to certain diseases (34). The only drawback of this technology is that a residual frequency or phase shift always remains, which is induced by non-specific mass effects, i.e. a shift which does not correspond to specific antigen-antibody interactions on the sensor surface (34). In order to drastically reduce or avoid cross-reactions with other type of interactions that may lead to false positives, Rayleigh-SAW generation, on the same piezoelectric substrate, seems to be very promising (35). In addition,

these waves can be used to generate fast fluid actuation to improve mixing and desorption promoting faster molecular interactions. Indeed, Rayleigh-SAW liquid effects can induce intense recirculation, actuation, heating or atomization, depending on the mechanical power conveyed by these acoustic waves (36, 37). This recirculation can allow to re-suspend all non-specific species that could settle and lead to non-specific responses.

4. Biosensing research efforts for Chagas diagnosis

Biosensors that have been investigated for the diagnosis of CD can be classified into electrochemical -where amperometric (38–41) and impedimetric (42) sensors can be found- and optical -where mainly Surface Plasmon Resonance (SPR) transducers (6) are found. Biosensors could provide the benefits presented in Table 1 in comparison with other currently employed techniques for the diagnosis of CD.

Pumpin-Ferreira et al., in 2005, reported a biosensor for the diagnosis of CD (38). It consisted on an amperometric immunosensor. This biosensor required an electrochemical interaction and, therefore, a potensiostat-galvanostat was required to conduct the measurements. Potensiostats are powerful equipment, but they are large and heavy for a final portable biosensing system. Hence, other electronics for biosensors characterization should be developed which provide higher miniaturization and integration capabilities for portable systems.

Recently, Luz et al. (2015) presented the first biosensor for the diagnosis of CD based on SPR transducers (6). They obtained the parameter related to the presence of antibodies anti-*T. Cruzi* found in human serum in approximately 20 min. SPR transducing principle requires an optical source for the laser generation and the integration of this source to the equipment, currently, leads to high volume and heavy apparatus, only suitable for laboratory tests. Moreover, even if optical biosensors can be very sensitive, the cost of SPR equipment is higher than USD \$50,000 and for this reason, not many researchers can afford such systems (43).

8 192

Table 1. Some methods for the diagnosis of CD.

9 193	Table 1. Some methods for the diagnosis of CD.				
11	Methods	Drawbacks	Benefits	References	
12 13 14 15 16 17 18 19 20	Selective Media	 Microorganism needs to overgrowth fast Long time to yield results Needs a laboratory Needs an aseptic work area Needs trained personnel 	 + Cheap + Easy to perform 	(44, 45)	
20 21 22 23 24 25 26 27	ELISA	 Tedious procedure Requires highly qualified personnel Consumes a lot of time Needs a laboratory Expensive 	 + High selectivity and sensitivity + Improves the time required to yield results + It works well for samples without interfering molecules 	(44-49)	
28 29 30 31 32 33	Quantitative PCR	 Expensive Needs trained personnel Needs a laboratory Difficult to perform 	 + High selectivity and sensitivity + Improving the time required to yield results 	(45) (47–49)	
34 35 36 37 38 39 40	Rapid test	 Specificity 96.8%. Just qualitative results. The method needs a tube, a measured volume of sample and reagent Can present false positive 	 + 15-25 min + High sensitivity of 99.5 % + Low cost (less than \$2 to the end user) 	(7, 50)	

1					
3					
4 F					
5 6	Biosensors	 Not commercially available 	+ No need of an aseptic working area	(6)	
7		 Large dimensions (currently) 	+ Fast (real-time)	(38–42)	
8 9		 Needs a laboratory 	+ Easy to perform. Not need of trained personnel	(17)'	
10		– Further research and development is	+ In situ simple preparation		
11 12		required for portable systems	+ High analytical specificity		
13		 High research cost 	+ Reduction of reagents consumption		
14 15			+ Reduced analysis time		
16			+ High reliability		
17 18			+ High sensitivity		
19			+ Integration of multiple processes in a single device		
20 21			+ Possible automation		
22			+ Low cost of fabrication		
23 24 194					
25					
$\frac{26}{27}$ 195					
²⁸ 196					
29					
30 197 31					
32 33					
34					
35 36					
37					
38					
40					
41					
42					
44					
45 46					
47					9
48 49					

6 7

8 9

10

12

14

16

18

19 20

21 22

23 24

25 26

27 28 29

30

32

34

36

38

40

41 42 43

45 46

47

49

51

53

55

57

59

61 62

63 64 65

5. Biosensors and their contribution to reduce CD burden

199 An ideal serological test should be easy to perform in a single step, fast, cheap, require no special 200 equipment or refrigeration reagents and should have a sensitivity and specificity of 100%. Such a 201 test does not exist currently for the diagnosis of CD. Hence, new technologies, which combine, 202 robustness, simplicity, portability and rapidity with an effective sensitivity and selectivity could contribute to more efficiently diagnose CD. There are evidences that show that biosensors could 11 203 13 204 meet most of these attributes for this application (6, 38). Biosensors could improve the diagnosis 15 **205** and the patients' treatment follow-up, in terms of rapidity, real-time and low cost detection ₁₇ 206 compared to current detection technologies like Polymerase Chain Reaction (PCR) and ELISA. In 207 addition, the use of biosensors offers significant advantages like: small fluid volume manipulation, 208 a high integration capability that facilitates the development of portable devices and ease of use. 209 This should allow their use by non-specialized personnel in non-centralized laboratories (17). 210 Nevertheless, further research efforts are needed to achieve a biosensing portable device for CD 211 diagnosis.

The expected features of biosensors are high selectivity and sensitivity, real-time label-free 212 31 213 monitoring, easy to use, reliability, high miniaturization capabilities and low cost. Biosensors based 33 214 on optical and acoustic wave sensing technologies could meet these requirements in a near future 35 **215** and seem to be very promising tools for this application. Such devices will lead to more sensitive tests at lower reagent concentrations, allowing biosensing system users to: i) reduce the cost of 37 216 39 217 reagents; ii) obtain valuable quantitative information; and iii) extend the measurement range of 218 the assays.

44 219 6. Infrastructure requirements

220 To develop a portable biosensor system for the rapid diagnosis of CD, first of all, it is necessary to ⁴⁸ 221 integrate a transducer with a suitable sensitive bio-chemical layer. Some authors have already 50 222 achieved this milestone, as stated in Section 4. Additionally, the system requires the integration of: 52 223 i) an electronic read-out system, for the interrogation and signal acquisition; ii) a microfluidic 54 224 system, to handle bio-fluids; and iii) a thermal control unit, to keep the temperature stable during 56 225 the sample analysis. This last point could be avoided if it is proven that the temperature sensitivity 58 226 of the sensor in use is negligible for the experiments we are conducting or if a differential 60 227 measurement setup for a temperature compensation is employed (51). Furthermore, it is

important to mention, that a fully-automated feature is desirable for the complete system, in order to run the sample analysis as comfortably as possible.

Nowadays, some companies offer commercial solutions for integrated biosensing systems that could be employed to diagnose CD. However, most of this systems are still of considerable size, weight and price, which prevents their wide use for field applications in low income communities. Table 2 shows some integrated biosensing platforms currently available in the market and some of their features. As can be appreciated in the table, all these systems require to be operated in a laboratory due to their dimensions. If researchers choose a non-commercial solution, they require to design and develop a system according to their needs. Nevertheless, this might allow them to pursuit a more compact, cost-effective and portable system.

Table 2. Different biosensing systems currently available in the market.

Product	Company Name	Transducer Technology	Dimensions (cm)	Weight (kg)
Q-Sense Omega Auto®	Q-sense	Acoustic	70×67×57	83
Biacore X100®	General Electric	SPR	59.6×56.3×59.3	47
AWS A20-F20®	AWsensors	Acoustic	77×75×45	60
OpenPlex®	Horiba	SPRi	49×30.4×48	15.6

7. Conclusions

Since there is not vaccine for CD, currently, the vector control and diagnosis tests are the most effective methods for preventing the disease and apply effective drug treatments. Even if a prophylactic or therapeutic vaccine could be achievable in next years, this would need to be part of integrated efforts that include better diagnostic means, since a vaccine is unlikely to be enough to stop the parasite transmission. Therefore, highly predictive diagnostic tests are required, not only to estimate the real size of CD problem, but also to assess the effectiveness of every action conducted towards a disease burden reduction.

Currently, there are three conventional tests to diagnose Chagas in its chronic phase: IHA, IIF and ELISA. All of these tests have sensitivities under 100%. Therefore, the WHO recommends performing at least two of these tests for a conclusive diagnosis, leading to a bottleneck of parasite detection, caused by limited local availability of laboratories in which such tests can be performed.

251 In addition, the diagnosis of the disease is generally delayed due to logistic restrictions of potential 252 patients to access diagnostic centers.

253 Non-conventional qualitative tests, like RLF tests, are currently commercially available. Some of 254 such tests can lead results within minutes, but cannot be considered as conclusive tests by themselves. Biosensors could be employed to support RLF test results in the future, diminishing the 255 256 overall time to achieve definitive quantitative results. Moreover, biosensors could exceptionally ¹² 257 contribute to a fast and secure screening method for blood banks in small and medium health 14 258 facilities. Hence, biosensors could improve CD diagnosis and the patients' treatment follow-up, in 16 259 terms of rapidity, small sample volume, high integration, ease of use, real-time and low cost detection compared to current conventional tests. Pursuing these goals is of considerable 18 260 20 261 importance and interest to diminish CD burden and to reduce the risk of disease spreading ₂₂ 262 intensified due to the climate change. Nevertheless, further research efforts are still needed to 263 develop portable biosensing systems in order to effectively employ this technology for CD 26 **264** diagnosis.

29²265 8. Acknowledgments

31 266 M. I. Rocha-Gaso, F. Sarry and D. Beyssen thank the Agence National de la Recherche (ANR), for 33 267 its financial support by granting the international AWESOM project.

36 268 9. References

1

2 3 4

5 6

7 8

9 10

11

13

15

17

19

21

23

24 25

27 28

30

32

34 35

37 38

39 40

41 42

44

57

62

63 64 65

269 Chagas C. 1909. Nova tripanozomiaze humana: estudos sobre a morfolojia e o ciclo evolutivo do 1. 270 Schizotrypanum cruzi n. gen., n. sp., ajente etiolojico de nova entidade morbida do homem. Mem Inst 43 271 Oswaldo Cruz 1:159–218.

45 272 Hotez PJ, Bottazzi ME, Franco-Paredes C, Ault SK, Periago MR. 2008. The neglected tropical diseases of 2. 46 47 273 Latin America and the Caribbean: a review of disease burden and distribution and a roadmap for 48 49 274 control and elimination. PLoS Negl Trop Dis 2:e300. 50

- 51 275 World Health Organization. 2014. Global burden of disease estimates for 2000-2012. Geneva. 3. 52
- 53 276 4. Gascon J, Vilasanjuan R, Lucas A. 2014. The need for global collaboration to tackle hidden public health 54 55 56 **277** crisis of Chagas disease. Expert Rev Anti Infect Ther 12:393–5.
- 58 **278** Tarleton RL, Gürtler RE, Urbina J a, Ramsey J, Viotti R. 2014. Chagas disease and the london 5. 59 60 279 declaration on neglected tropical diseases. PLoS Negl Trop Dis 8:e3219. 61

Luz JGG, Souto DEP, Machado-Assis GF, de Lana M, Kubota LT, Luz RCS, Damos FS, Martins HR. 2015.
 Development and Evaluation of a SPR-based Immunosensor for Detection of Anti-Trypanosoma cruzi
 Antibodies in Human Serum. Sensors Actuators B Chem.

5

16

20

33

46

57

62

63 64 65

6 283 7. Sánchez-Camargo CL, Albajar-Viñas P, Wilkins PP, Nieto J, Leiby D a., Paris L, Scollo K, Flórez C, 7 8 284 Guzmán-Bracho C, Luquetti AO, Calvo N, Tadokoro K, Saez-Alquezar A, Palma PP, Martin M, Flevaud 9 10 285 L. 2014. Comparative evaluation of 11 commercialized rapid diagnostic tests for detecting Trypanosoma 11 12 286 cruzi antibodies in serum banks in areas of endemicity and nonendemicity. J Clin Microbiol 52:2506-13 14 15 **287** 2512.

- 288
 8. WHO. 2002. CONTROL OF CHAGAS DISEASE Second report of the WHO Expert Committee World Health
 19
 289
 OrganizationWHO Technical Report Series.
- 21
22
22290
229.Coura JR. 2015. The main sceneries of chagas disease transmission. The vectors, blood and oral
transmissions A comprehensive review. Mem Inst Oswaldo Cruz 110:277–282.
- 25
 292 10. Perez CJ, Lymbery AJ, Thompson RCA. 2014. Chagas disease: The challenge of polyparasitism? Trends
 27
 293 Parasitol 30:176–182.
- 29
 30 294 11. Ramsey JM, Elizondo-Cano M, Sanchez-González G, Peña-Nieves A, Figueroa-Lara A. 2014.
 31
 32 295 Opportunity cost for early treatment of Chagas disease in Mexico. PLoS Negl Trop Dis 8:e2776.
- ³⁴ 296 12. Liu Q, Zhou X-N. 2015. Preventing the transmission of American trypanosomiasis and its spread into
 ³⁶ 297 non-endemic countries. Infect Dis poverty 4:60.
- ³⁸ 39
 ³⁹ 298
 ⁴⁰ 41
 ⁴⁰ 299
 ⁴⁰ Treatment access for Chagas disease in Mexico. PLoS Negl Trop Dis 7:e2488.
- 42
 43 300 14. Schmunis GA, Yadon ZE. 2010. Chagas disease: a Latin American health problem becoming a world
 44
 45 301 health problem. Acta Trop 115:14–21.
- 4730215.Conners EE, Vinetz JM, Weeks JR, Brouwer KC. 2016. A global systematic review of Chagas disease4849303prevalence among migrants. Acta Trop 156:68–78.
- 51
5230416.Requena-Méndez A, Aldasoro E, de Lazzari E, Sicuri E, Brown M, Moore DAJ, Gascon J, Muñoz J. 2015.53
54305Prevalence of Chagas Disease in Latin-American Migrants Living in Europe: A Systematic Review and55
56306Meta-analysis. PLoS Negl Trop Dis 9:1–15.
- 58 307 17. Sin MLY, Mach KE, Wong PK, Liao JC. 2014. Advances and challenges in biosensor-based diagnosis of
 infectious diseases. Expert Rev Mol Diagn 14:225–44.

- 309 18. **Prata A**. 2001. Clinical and epidemiological aspects of Chagas disease. Lancet Infect Dis **1**:92–100.
- Pinto AY das N, Valente V da C, Coura JR, Valente SA da S, Junqueira ACV, Santos LC, Ferreira AG, de
 Macedo RC. 2013. Clinical Follow-Up of Responses to Treatment with Benznidazol in Amazon: A Cohort
 Study of Acute Chagas Disease. PLoS One 8:1–9.
- 8 313 Morillo CA, Marin-Neto JA, Avezum A, Sosa-Estani S, Rassi AJ, Rosas F, Villena E, Quiroz R, Bonilla R, 20. 9 10 314 Britto C, Guhl F, Velazquez E, Bonilla L, Meeks B, Rao-Melacini P, Pogue J, Mattos A, Lazdins J, Rassi A, 11 12 Connolly SJ, Yusuf S, BENEFIT Investigators. 2015. Randomized Trial of Benznidazole for Chronic 315 13 14 15 316 Chagas' Cardiomyopathy. N Engl J Med **373**:1295 – 1306.
- 17
 317
 21.
 Aparecido A. PF, Carolina VU, Michelle DSC, Naira CP, Marcos VF, Glauco P dos S, Cecílio SF, Assis VB,

 18
 19
 318
 Hideki Y. 2013. Amperometric Biosensor for Diagnosis of DiseaseState of the Art in Biosensors

 20
 21
 319
 Environmental and Medical Applications.
- 23
2432022.Stimpert KK, Montgomery SP. 2010. Physician Awareness of Chagas Disease, USA. Emerg Infect Dis25
2632116:871-872.
- 27
 28
 322
 23. Ventura-Garcia L, Roura M, Pell C, Posada E, Gasc??n J, Aldasoro E, Mu??oz J, Pool R. 2013. Socio30
 323 Cultural Aspects of Chagas Disease: A Systematic Review of Qualitative Research. PLoS Negl Trop Dis 7.
- 31
 32 324 24. Fergnani N, Ruggiero A, Ceccarelli S, Menu F, Rabinovich J. 2013. Large-scale patterns in morphological
 33
 34 325 diversity and species assemblages in Neotropical Triatominae (Heteroptera: Reduviidae). Mem Inst
 36 326 Oswaldo Cruz 108:997–1008.
- 38 39
 327
 25. Ibarra-Cerdeña CN, Zaldivar-Riveron A, Peterson AT, Sanchez-Cordero V, Ramsey JM. 2014. Phylogeny
 40 41
 328
 43
 329
 Triatominae), vectors of Chagas' disease. PLoS Negl Trop Dis. 8(10): e3266
- 45 330 26. Ramsey JM, Peterson a T, Carmona-Castro O, Moo-Llanes D a, Nakazawa Y, Butrick M, Tun-Ku E, la
 47 331 Cruz-Félix K De, Ibarra-Cerdeña CN. 2015. Atlas of Mexican Triatominae (Reduviidae: Hemiptera) and
 48 49 332 vector transmission of Chagas disease. Mem Inst Oswaldo Cruz 110:339–52.
- 51
5233327.Gurgel-Gonçalves R, Galvão C, Costa J, Peterson a T. 2012. Geographic distribution of chagas disease53
54334vectors in Brazil based on ecological niche modeling. J Trop Med 2012:705326.
- Lescure F-X, Le Loup G, Freilij H, Develoux M, Paris L, Brutus L, Pialoux G. 2010. Chagas disease:
 Changes in knowledge and management. Lancet Infect Dis 10:556–570.
- 60 337 29. Sutherst RW. 2004. Global Change and Human Vulnerability to Vector-Borne Diseases Global Change 61
- 63 64 65

59

44

7

- 338 and Human Vulnerability to Vector-Borne Diseases. Clin Microbiol Rev 17:136–173. 1 2 339 30. Githeko AK, Lindsay SW, Confalonieri UE, Patz JA. 2000. Climate change and vector-borne diseases: a 3 4 340 regional analysis. Bull World Health Organ 78:1136–1147. 5 6 341 31. Coura JR. 2013. Chagas disease: Control, elimination and eradication. Is it possible? Mem Inst Oswaldo 7 8 342 Cruz 108:962-967. 9 10 343 Bhattacharyya T, Falconar AK, Luquetti AO, Costales J a, Grijalva MJ, Lewis MD, Messenger L a, Tran 32. 11 12 TT, Ramirez J-D, Guhl F, Carrasco HJ, Diosque P, Garcia L, Litvinov S V, Miles M a. 2014. Development 344 13 14 15 **345** of peptide-based lineage-specific serology for chronic Chagas disease: geographical and clinical 16 distribution of epitope recognition. PLoS Negl Trop Dis 8:e2892. 17 346 18 19 347 33. Rocha-Gaso M-I, March-Iborra C, Montoya-Baides A, Arnau-Vives A. 2009. Surface generated acoustic 20 21 348 wave biosensors for the detection of pathogens: a review. Sensors (Basel) 9:5740-69. 22 23 349 34. Lee H, Lee S, Namkoong K, Ko C. 2008. SAW immunosensors for HBsAb detection. Sensors, 2008. 24 25 350 35. Rocha-Gaso MI, Renaudin A, Sarry F, Beyssen D. 2015. Lab-on-a-chip based integrated hybrid 26 27 ₂₈ 351 technologies for biofluids manipulation and characterization. Procedia Eng **120**:687–690. 29 30 352 36. Beyssen D, Brizoual L Le, Elmazria O. 2006. Microfluidic device based on surface acoustic wave. Sensors 31 32 353 Actuators B. 33 ³⁴ 354 37. Roux-Marchand T, Beyssen D, Sarry F, Elmazria O. 2015. Rayleigh Surface Acoustic Wave As an 35 36 355 Efficient Heating System for Biological Reactions: Investigation of Microdroplet Temperature 37 38 356 Uniformity. IEEE Trans Ultrason Ferroelectr Freq Control 62:729–735. 39 40 357 38. Ferreira a. a P, Colli W, Da Costa PI, Yamanaka H. 2005. Immunosensor for the diagnosis of Chagas' 41 42 43 358 disease. Biosens Bioelectron 21:175-181. 44 Belluzo MS, Ribone MÉ, Camussone C, Marcipar IS, Lagier CM. 2011. Favorably orienting recombinant 45 359 39. 46 47 360 proteins to develop amperometric biosensors to diagnose Chagas' disease. Anal Biochem 408:86–94. 48 49 361 Ferreira AAP, Colli W, Alves MJM, Oliveira DR, Costa PI, Güell AG, Sanz F, Benedetti AV, Yamanaka H. 40. 50 51 362 2006. Investigation of the interaction between Tc85-11 protein and antibody anti-T. cruzi by AFM and 52 53 amperometric measurements. Electrochim Acta 51:5046-5052. 363 54 55 ₅₆ 364 41. Vinicius Foguel M, Pilondos Santos G, Aparecido Pupim Ferreira A, Yamanaka H, Vicente Benedetti A. 57 58 **365** 2011. Amperometric immunosensor for Chagas' disease using gold CD-R Transducer. Electroanalysis 59 60 366 **23**:2555-2561. 61 62 15 63
- 64 65

- 367 42. Erdmann CA, Kovalczuk E, Inaba J, Viana AG, Pessoa CA, Wohnrath K, Garcia JR. 2013. Development of
 a Nano-particle enhanced impedimetric biosensor for Chagas' Disease DiagnosisXLII Annual Meeting of
 369 SBBq.
- 637043.Coltro WKT, Neves RDS, Motheo ADJ, Da Silva JAF, Carrilho E. 2014. Microfluidic devices with8371integrated dual-capacitively coupled contactless conductivity detection to monitor binding events in10372real time. Sensors Actuators, B Chem 192:239–246.
- ¹²₁₃
 ³⁷³ 44. Sachse K, Frey J. 2002. PCR Detection of Microbial Pathogens: Methods and Protocols, p. . *In* Totowa, N
 ¹⁴₁₅
 ³⁷⁴ (ed.), Methods in Molecular Biology, 1st ed. Humana Press Inc, Springer.
- 23
2437846.Thacker J, Casale E, Tucker C. 1996. Immunoassays (ELISA) for rapid, quantitative analysis in the food-25
26379processing industry. J Agric Food Chem 44:2680–2685.
- 27
 28 380 47. Iqbal S, Mayo M, Bruno J, Bronk B, Batt C, Chambers P. 2000. A review of molecular recognition
 30 381 technologies for detection of biological threat agents. Biosens Bioelectron 15:549–578.
- 32 382 48. Andreotti P, Ludwig G, Peruski A, Tuite J, Morse S, Peruski L. 2003. Immunoassay of infectious agents.
 33
 34 383 Biotechniques 35:850–859.
- ³⁶ 384 49.
 ³⁷ ³⁸ 49.
 ³⁸ 385 disease and biological warfare agents. Clin Diagn Lab Immunol **10**:506–513.
- ⁴⁰
 ⁴¹ 386 50. Barfield CA, Barney RS, Crudder CH, Wilmoth JL, Stevens DS, Mora-Garcia S, Yanovsky J. 2010. A
 ⁴²
 ⁴³ 387 Highly Sensitive Rapid Diagnostic Test for Chagas Disease That Utilizes a Recombinant Trypanosoma
 ⁴⁴
 ⁴⁵ 388 cruzi Antigen. IEEE Trans Bio-Medical Eng 58:814–817.
- 47 389 51. Balcerzak A, Rejmund F, Gutkiewicz P, Zienkiewicz B, Zhavnerko G. 2014. Ultrasonic chemical sensor
 48
 49 390 with organic monomolecular layer. Arch Acoust 31:47–52.
- ⁵¹ **391**