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# Biosensors to diagnose Chagas disease: A review

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**Abstract:** Chagas disease (CD), which mostly affects underprivileged people, has turned into one of Latin America's main public health problems. Prevention of the disease requires early diagnosis, initiation of therapy, and regular blood monitoring of the infected individual. However, the majority of the infections go undiagnosed because of general mild symptoms and lack of access to medical care. Therefore, more affordable and accessible detection technologies capable of providing early diagnosis and parasite load measurements in settings where CD is prevalent are needed to enable enhanced intervention strategies. This review discusses currently available detection technologies and emerging biosensing technologies for a future application to CD. Even if biosensing technologies still require further research efforts to develop portable systems, we arrive to the conclusion that biosensors could improve diagnosis and the patients' treatment follow-up, in terms of rapidity, small sample volume, high integration, ease of use, real-time and low cost detection compared to current conventional technologies.

Keywords: Chagas disease, biosensors, detection technologies, diagnosis, neglected diseases.

#### 1. Introduction

Chagas disease (CD), discovered in 1909 by the Brazilian physician Carlos Chagas (1), nowadays has turned into one of Latin America's main public health problems (2). Based in disability-adjusted life-years as a measure of disease burden, CD figure as the most important parasitic vector-borne illness in the Region of Americas, seven times higher than Malaria and up to three times more than Dengue (3), and yet it is still absent in the agenda of the public health policies and practices of many endemic countries (4, 5). More generally, ranks fourth in mortality and eighth in morbidity

among world neglected tropical diseases (6), and is estimated that between eight and eleven million people are infected, while 100 million are at risk of acquiring the disease; the main cause being to live in proximity with disease vectors (7). CD is caused by the parasitic presence of the Trypanosoma cruzi in the organism, which is mainly transmitted by contamination with infected feces of blood-sucking triatomine vectors during a human blood meal. Nevertheless, it can also be transmitted through blood transfusions, organ transplants, infected mothers to their unborn children and ingestion of contaminated food (i. e. Oral transmission; WHO, 2002). Although disease progression can be associated with the mechanism of infection, with oral transmission causing the most severe outbreaks (9), people living at risk regions are susceptible to polyparasitism (i.e. Coinfections and superinfections with different strains of T. cruzi), with unknown effects in the variability of the disease progression and response to treatments (10).

CD mostly affects underprivileged people and the majority of the *T. cruzi* infections go undiagnosed because of general mild symptoms and lack of access to medical care (11). Due to this fact, CD is considered as a Neglected Tropical Disease whose improvement in diagnosis and treatment today requires research and development efforts with non-profit interests. The highest prevalence of Chagas disease has been reported in Bolivia (6.75–15.4 %), followed by Paraguay (0.69–9.3 %), Panama (0.01-9.02 %), Brazil (0.8-1.30 %), Mexico (0.5-6.8 %) and Argentina (4.13-8.2 %) (12). Citing a case, this disease causes almost 6% of the annual deaths in Mexico and the seroprevalence can roughly be estimated at least in 3% due to a lack of active epidemiological surveillance (most cases are detected during blood screening procedures in blood banks). Yet, less than 0.5% of the infected individuals have access to treatment in this country as a result of anachronisms in the normativity, among other failures in the public health system (13).

Even if CD mainly affects tropical countries, with nowadays ease of traveling and migration, other countries are also being affected by this infection (14). Several cases have been reported in USA, Canada, Europe and in Western Pacific regions like Japan and Australia (14-16). Notably, CD continues to be an inconspicuous public health problem, with limited medical awareness, either because it is commonly targeted in people with relatively low medical access or because it can currently occur in unexpected regions. Thus, the treatment of CD urgently needs to generalize and standardize diagnostic procedures.

Biosensors are relatively new analytical devices that can help to detect the presence of specific compounds and pathogens in liquid environments and complex mixtures like: water and blood

serum. Although these devices have been formerly used in the alimentary industry (mostly to detect toxins and infectious pathogens), they are been increasingly used to diagnose human diseases (17). Therefore, these devices can be employed for the diagnosis of CD. The development of biosensors requires a biological active component to be immobilized onto the surface of a transducer. The selective recognition layer, towards *T. cruzi* specific antigens present in patients' blood serum, can selectively detect the target analyte generating a signal response in the sensor (see Figure 1). Depending on their transducing principle biosensors can be electrochemical, acoustic or optic.

In this work, we firstly introduce a brief description of the disease. Secondly, we present a review of biosensor technologies whose applicability to diagnose CD has been investigated. Finally, we mention the benefits and drawbacks of applying biosensors as solutions to this major public health issue and the infrastructure required to conduct biosensor experiments for this application.

#### 2. Brief description of CD and current needs

CD passes through two successive stages: an acute phase and a chronic phase. The acute phase occurs at the following 6-8 weeks after infection. The acute phase is followed by the chronic phase of CD, which lasts for the rest of the life of the infected individual, and has different forms. In the

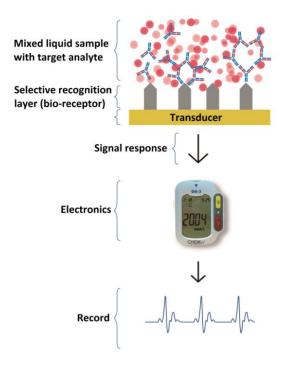


Figure 1. General scheme of a biosensor detection strategy. A biosensor is composed of a biochemical interface where specific bio-species are absorbed; a transducer which translates the recognition event to another physical response that can be measured and an electronic system which acquires and records the signal.

indeterminate form, an equilibrium between the parasite and the immunological response of the infected individual is reached and most infected patients appear healthy, with no evidence of organ damage that could be found by current standard methods of clinical diagnosis (8). About 50-70% of infected individuals will remain in this condition for the rest of their lives. However, several years after the chronic phase has started, 10-40% of infected individuals will pass to the cardiac form of the disease and will develop injuries of various organs, mainly the heart, the digestive system, and occasionally, the peripheral nervous system (8, 18). These important symptomatic changes occur 10-20 years after the acute phase of the disease and include a broad range of types of damage. The clinical manifestations vary from mild symptoms to heart failure and, frequently, sudden cardiac death (18). The acute phase of CD is recognized only in an estimated 1-2% of all individuals acquiring the infection (8) due to a lack of access to sufficient medical care. Thus, more than any other parasitic disease, CD is closely related to social and economic development. Paradoxically, acute phase is the most appropriate period for drug treatment, showing relatively high levels of sero-conversion, while organ damage is prevented (19). Conversely, it has been shown that drug treatment with Trypanocidal therapy in patients with established Chagas cardiomyopathy, can cause seroconversion, but does not stop cardiac clinical deterioration (20).

Given that no vaccine is currently available to prevent CD, vector control, diagnosis tests, opportune drug treatment, and clinical follow-up are the most effective methods to fight against the disease (21). Nevertheless, all these measures suffer of several hindrances imposed by the synergistic negative effects of diverse vulnerability components of CD risk, such as ecological factors (i.e. land-use changes) -that are broadening the contact zones between humans and parasites (Lopez-Cancino et al., 2015)-, the chronic failure of health care policies hindering the reduction of CD incidence (5), the limited awareness of physicians (22), and several socio-cultural practices that perpetuate CD exposure in endemic regions (23). Health policies to control/reduce vectorial exposure to CD in Latin America are challenged for the relatively high diversity of vectors that shows a broad environmental tolerance. Likewise, a wide territory of America is suitable for CD vectorial transmission (24-27).

In order to face the epidemiological challenges due to the increasing complexity of interactions among the transmission routes of *T. cruzi* in endemic and non-endemic countries, access to early diagnostic and treatment seems as the most cost-efficient ways to reduce the CD burden (11). Several paths of scientific advances and discoveries envisage an optimistic future to reduce CD

 burden coming from a better understanding of CD transmission and management toward its interruption (28). The fact that the World Health Organization (WHO) has made a commitment with Bill and Melinda Gates Foundation and other stakeholders to control the most neglected diseases by 2020, including CD among them, opens an opportunity window to orientate research priorities. This commitment, launched in 2012, called the London Declaration was (http://www.unitingtocombatntds.org/). However, claims have been raised to urge implicated organizations to set the needed measures to reach these goals, among which improved diagnosis (i.e. precision and accessibility) seems as one of the most important first steps (5).

Blood banks in endemic countries require fast and secure screening method for small and medium health facilities, since screening for CD is mandatory in some endemic countries (8). Several countries do not have an active program for CD detection and depend only for the blood bank reports. The most important reason of this fact is the low feasibility to detect Chagas in patients. Since, CD do not produce particular symptoms, there are not incentives from society. Nevertheless, the bug bite is quite notorious and people can suspect that they are infected with *T. cruzi* when a *chinchoma*<sup>1</sup> appears. In such cases, people can search medical assistance and be subjects of a blood test.

Climate change and global warming increase the risk of rising CD burden in some regions. Climate change impacts on vector-borne diseases (29) and is undoubtedly detonating variables that make the CD transmission become potentially dangerous, as the WHO points out (30). Nevertheless, the incidence of CD can be greatly reduced by residual insecticide-based vector control programs that decrease the populations of the transmitting vectors and by improving housing (31).

#### 3. Current detection technologies and their limitations

Currently, laboratory methods are employed to diagnose CD. Depending on the patients' phase of infection some are more convenient than others. During the acute phase of CD, a large number of parasites are present in the peripheral blood and can be diagnosed by direct microscopical observation of fresh blood (parasitological test). However, for the chronic phase of CD the diagnosis is not possible, due to the scarce parasitemia. Therefore, the immunodiagnosis is widely used since nearly all *T. cruzi*-infected individuals in the chronic phase develop antibodies against the complex antigenic mixture of the parasite (8).

<sup>&</sup>lt;sup>1</sup> Name of the inflammatory injury after a bug bite.

 Several immunodiagnosis tests are available, but mainly three conventional tests are widely used: indirect haemagglutination (IHA), indirect immunofluorescence (IIF) and Enzyme Linked Immuno Assay (ELISA). These tests present several limitations such as: (i) cross-reactivity with other parasites; (ii) not 100% of sensitivity; (iii) the need to be performed in a laboratory; and (iv) a long time is required to obtain the results. IHA test results can be obtained in about two hours, whereas IIF results can be obtained after numerous steps in two hours and ELISA takes several hours to carry out, including prior sensitization of microplates with *T. cruzi* antigens for about 12 hours (6). All these tests have to be performed in centralized laboratories; some of them require sophisticated equipment and skilled technicians. Since none of these tests have a sensitivity of 100%, the WHO recommends conducting at least two conventional tests for a definitive diagnosis of *T. cruzi* infection (8).

More recently, non-conventional tests, like rapid lateral flow (RLF) tests, are commercially available in the market to detect T. cruzi infection using whole blood, serum or plasma (Sánchez-Camargo et al. 2014). These tests are based in different tests principles: immunochromatography, particle agglutination, immunofiltration or immunodot. They provide fast results (between 5 to 60 min reading times) without the need of electrical equipment and they require low volume samples (5 to 150  $\mu$ l). However, the sensitivities and specificities of such tests are lower than that of conventional tests and they only provide qualitative or semi-quantitative results, which prevents obtaining important test information like genetic lineage of the T. cruzi (32) and the immunoreaction kinetics.

From a prospective point of view, it is important to discuss about another kind of technology, which could be deployed for Chagas Disease diagnosis in the near future. Over the last decade, Shear-Horizontal (SH) Surface Acoustic Wave (SAW) immunosensors (Love-SAW with a guiding layer) have been developed for the diagnosis of various diseases (33). Such immunosensors exhibit a high sensitivity and a very low limit of detection (in the order of pg/µl of blood serum). This technology give very relevant results for the detection of antibodies, specific to certain diseases (34). The only drawback of this technology is that a residual frequency or phase shift always remains, which is induced by non-specific mass effects, i.e. a shift which does not correspond to specific antigen-antibody interactions on the sensor surface (34). In order to drastically reduce or avoid cross-reactions with other type of interactions that may lead to false positives, Rayleigh-SAW generation, on the same piezoelectric substrate, seems to be very promising (35). In addition,

these waves can be used to generate fast fluid actuation to improve mixing and desorption promoting faster molecular interactions. Indeed, Rayleigh-SAW liquid effects can induce intense recirculation, actuation, heating or atomization, depending on the mechanical power conveyed by these acoustic waves (36, 37). This recirculation can allow to re-suspend all non-specific species that could settle and lead to non-specific responses.

#### 4. Biosensing research efforts for Chagas diagnosis

Biosensors that have been investigated for the diagnosis of CD can be classified into electrochemical -where amperometric (38-41) and impedimetric (42) sensors can be found- and optical -where mainly Surface Plasmon Resonance (SPR) transducers (6) are found. Biosensors could provide the benefits presented in Table 1 in comparison with other currently employed techniques for the diagnosis of CD.

Pumpin-Ferreira et al., in 2005, reported a biosensor for the diagnosis of CD (38). It consisted on an amperometric immunosensor. This biosensor required an electrochemical interaction and, therefore, a potensiostat-galvanostat was required to conduct the measurements. Potensiostats are powerful equipment, but they are large and heavy for a final portable biosensing system. Hence, other electronics for biosensors characterization should be developed which provide higher miniaturization and integration capabilities for portable systems.

Recently, Luz et al. (2015) presented the first biosensor for the diagnosis of CD based on SPR transducers (6). They obtained the parameter related to the presence of antibodies anti-T. Cruzi found in human serum in approximately 20 min. SPR transducing principle requires an optical source for the laser generation and the integration of this source to the equipment, currently, leads to high volume and heavy apparatus, only suitable for laboratory tests. Moreover, even if optical biosensors can be very sensitive, the cost of SPR equipment is higher than USD \$50,000 and for this reason, not many researchers can afford such systems (43).

Table 1. Some methods for the diagnosis of CD.

Methods	Drawbacks	Benefits	References
Selective Media	Microorganism needs to overgrowth fast	+ Cheap	(44, 45)
	Long time to yield results	+ Easy to perform	
	<ul> <li>Needs a laboratory</li> </ul>		
	Needs an aseptic work area		
	<ul> <li>Needs trained personnel</li> </ul>		
	- Tedious procedure		
ELISA	Requires highly qualified personnel	+ High selectivity and sensitivity	(44-49)
	Consumes a lot of time	+ Improves the time required to yield results	
	- Needs a laboratory	+ It works well for samples without interfering	
	- Expensive	molecules	
Quantitative PCR	– Expensive	+ High selectivity and sensitivity	(45)
	Needs trained personnel	+ Improving the time required to yield results	(47–49)
	– Needs a laboratory		(47-42)
	– Difficult to perform		
Rapid test	- Specificity 96.8%.	+ 15-25 min	(7, 50)
	- Just qualitative results.	+ High sensitivity of 99.5 %	
	- The method needs a tube, a measured	+ Low cost (less than \$2 to the end user)	
	volume of sample and reagent		
	Can present false positive		

Biosensors	Not commercially available	+ No need of an aseptic working area	(6)
	<ul> <li>Large dimensions (currently)</li> </ul>	+ Fast (real-time)	(38-42)
	- Needs a laboratory	+ Easy to perform. Not need of trained personnel	(17)
	Further research and development is	+ In situ simple preparation	
	required for portable systems	+ High analytical specificity	
	High research cost	+ Reduction of reagents consumption	
		+ Reduced analysis time	
		+ High reliability	
		+ High sensitivity	
		+ Integration of multiple processes in a single device	
		+ Possible automation	
		+ Low cost of fabrication	

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#### 5. Biosensors and their contribution to reduce CD burden

An ideal serological test should be easy to perform in a single step, fast, cheap, require no special equipment or refrigeration reagents and should have a sensitivity and specificity of 100%. Such a test does not exist currently for the diagnosis of CD. Hence, new technologies, which combine, robustness, simplicity, portability and rapidity with an effective sensitivity and selectivity could contribute to more efficiently diagnose CD. There are evidences that show that biosensors could meet most of these attributes for this application (6, 38). Biosensors could improve the diagnosis and the patients' treatment follow-up, in terms of rapidity, real-time and low cost detection compared to current detection technologies like Polymerase Chain Reaction (PCR) and ELISA. In addition, the use of biosensors offers significant advantages like: small fluid volume manipulation, a high integration capability that facilitates the development of portable devices and ease of use. This should allow their use by non-specialized personnel in non-centralized laboratories (17). Nevertheless, further research efforts are needed to achieve a biosensing portable device for CD diagnosis.

The expected features of biosensors are high selectivity and sensitivity, real-time label-free monitoring, easy to use, reliability, high miniaturization capabilities and low cost. Biosensors based on optical and acoustic wave sensing technologies could meet these requirements in a near future and seem to be very promising tools for this application. Such devices will lead to more sensitive tests at lower reagent concentrations, allowing biosensing system users to: i) reduce the cost of reagents; ii) obtain valuable quantitative information; and iii) extend the measurement range of the assays.

#### 6. Infrastructure requirements

To develop a portable biosensor system for the rapid diagnosis of CD, first of all, it is necessary to integrate a transducer with a suitable sensitive bio-chemical layer. Some authors have already achieved this milestone, as stated in Section 4. Additionally, the system requires the integration of:
i) an electronic read-out system, for the interrogation and signal acquisition; ii) a microfluidic system, to handle bio-fluids; and iii) a thermal control unit, to keep the temperature stable during the sample analysis. This last point could be avoided if it is proven that the temperature sensitivity of the sensor in use is negligible for the experiments we are conducting or if a differential measurement setup for a temperature compensation is employed (51). Furthermore, it is

 important to mention, that a fully-automated feature is desirable for the complete system, in order to run the sample analysis as comfortably as possible.

Nowadays, some companies offer commercial solutions for integrated biosensing systems that could be employed to diagnose CD. However, most of this systems are still of considerable size, weight and price, which prevents their wide use for field applications in low income communities. Table 2 shows some integrated biosensing platforms currently available in the market and some of their features. As can be appreciated in the table, all these systems require to be operated in a laboratory due to their dimensions. If researchers choose a non-commercial solution, they require to design and develop a system according to their needs. Nevertheless, this might allow them to pursuit a more compact, cost-effective and portable system.

Table 2. Different biosensing systems currently available in the market.

Product	Company Name	Transducer Technology	Dimensions (cm)	Weight (kg)
Q-Sense Omega Auto®	Q-sense	Acoustic	70×67×57	83
Biacore X100®	General Electric	SPR	59.6×56.3×59.3	47
AWS A20-F20®	AWsensors	Acoustic	77×75×45	60
OpenPlex®	Horiba	SPRi	49×30.4×48	15.6

#### 7. Conclusions

Since there is not vaccine for CD, currently, the vector control and diagnosis tests are the most effective methods for preventing the disease and apply effective drug treatments. Even if a prophylactic or therapeutic vaccine could be achievable in next years, this would need to be part of integrated efforts that include better diagnostic means, since a vaccine is unlikely to be enough to stop the parasite transmission. Therefore, highly predictive diagnostic tests are required, not only to estimate the real size of CD problem, but also to assess the effectiveness of every action conducted towards a disease burden reduction.

Currently, there are three conventional tests to diagnose Chagas in its chronic phase: IHA, IIF and ELISA. All of these tests have sensitivities under 100%. Therefore, the WHO recommends performing at least two of these tests for a conclusive diagnosis, leading to a bottleneck of parasite detection, caused by limited local availability of laboratories in which such tests can be performed.

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In addition, the diagnosis of the disease is generally delayed due to logistic restrictions of potential patients to access diagnostic centers.

Non-conventional qualitative tests, like RLF tests, are currently commercially available. Some of such tests can lead results within minutes, but cannot be considered as conclusive tests by themselves. Biosensors could be employed to support RLF test results in the future, diminishing the overall time to achieve definitive quantitative results. Moreover, biosensors could exceptionally contribute to a fast and secure screening method for blood banks in small and medium health facilities. Hence, biosensors could improve CD diagnosis and the patients' treatment follow-up, in terms of rapidity, small sample volume, high integration, ease of use, real-time and low cost detection compared to current conventional tests. Pursuing these goals is of considerable importance and interest to diminish CD burden and to reduce the risk of disease spreading intensified due to the climate change. Nevertheless, further research efforts are still needed to develop portable biosensing systems in order to effectively employ this technology for CD diagnosis.

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- M. I. Rocha-Gaso, F. Sarry and D. Beyssen thank the Agence National de la Recherche (ANR), for its financial support by granting the international AWESOM project.
- 9. References
- Chagas C. 1909. Nova tripanozomiaze humana: estudos sobre a morfolojia e o ciclo evolutivo do 1. Schizotrypanum cruzi n. gen., n. sp., ajente etiolojico de nova entidade morbida do homem. Mem Inst Oswaldo Cruz 1:159-218.
- Hotez PJ, Bottazzi ME, Franco-Paredes C, Ault SK, Periago MR. 2008. The neglected tropical diseases of 2. Latin America and the Caribbean: a review of disease burden and distribution and a roadmap for control and elimination. PLoS Negl Trop Dis 2:e300.
- World Health Organization. 2014. Global burden of disease estimates for 2000-2012. Geneva. 3.
- 276 4. Gascon J, Vilasanjuan R, Lucas A. 2014. The need for global collaboration to tackle hidden public health crisis of Chagas disease. Expert Rev Anti Infect Ther 12:393-5.
  - Tarleton RL, Gürtler RE, Urbina J a, Ramsey J, Viotti R. 2014. Chagas disease and the london 5. declaration on neglected tropical diseases. PLoS Negl Trop Dis 8:e3219.

1	280	6.	Luz JGG, Souto DEP, Machado-Assis GF, de Lana M, Kubota LT, Luz RCS, Damos FS, Martins HR. 2015.
1 2 3	281		Development and Evaluation of a SPR-based Immunosensor for Detection of Anti-Trypanosoma cruzi
3 4 5	282		Antibodies in Human Serum. Sensors Actuators B Chem.
6 7	283	7.	Sánchez-Camargo CL, Albajar-Viñas P, Wilkins PP, Nieto J, Leiby D a., Paris L, Scollo K, Flórez C,
8	284		Guzmán-Bracho C, Luquetti AO, Calvo N, Tadokoro K, Saez-Alquezar A, Palma PP, Martin M, Flevaud
10 11	285		L. 2014. Comparative evaluation of 11 commercialized rapid diagnostic tests for detecting Trypanosoma
12 13	286		cruzi antibodies in serum banks in areas of endemicity and nonendemicity. J Clin Microbiol <b>52</b> :2506–
	287		2512.
	288	8.	WHO. 2002. CONTROL OF CHAGAS DISEASE Second report of the WHO Expert Committee World Health
	289		OrganizationWHO Technical Report Series.
<ul><li>20</li><li>21</li><li>22</li></ul>	290	9.	Coura JR. 2015. The main sceneries of chagas disease transmission. The vectors, blood and oral
23 24	291		transmissions - A comprehensive review. Mem Inst Oswaldo Cruz <b>110</b> :277–282.
25 26	292	10.	Perez CJ, Lymbery AJ, Thompson RCA. 2014. Chagas disease: The challenge of polyparasitism? Trends
27 28	293		Parasitol <b>30</b> :176–182.
29 30	294	11.	Ramsey JM, Elizondo-Cano M, Sanchez-González G, Peña-Nieves A, Figueroa-Lara A. 2014.
	295		Opportunity cost for early treatment of Chagas disease in Mexico. PLoS Negl Trop Dis 8:e2776.
33 34 35	296	12.	Liu Q, Zhou X-N. 2015. Preventing the transmission of American trypanosomiasis and its spread into
	297		non-endemic countries. Infect Dis poverty <b>4</b> :60.
38 39	298	13.	Manne JM, Snively CS, Ramsey JM, Salgado MO, Bärnighausen T, Reich MR. 2013. Barriers to
40 41	299		treatment access for Chagas disease in Mexico. PLoS Negl Trop Dis <b>7</b> :e2488.
	300	14.	Schmunis GA, Yadon ZE. 2010. Chagas disease: a Latin American health problem becoming a world
	301		health problem. Acta Trop <b>115</b> :14–21.
46 47	302	15.	Conners EE, Vinetz JM, Weeks JR, Brouwer KC. 2016. A global systematic review of Chagas disease
48 49 50	303		prevalence among migrants. Acta Trop <b>156</b> :68–78.
51	304	16.	Requena-Méndez A, Aldasoro E, de Lazzari E, Sicuri E, Brown M, Moore DAJ, Gascon J, Muñoz J. 2015.

Prevalence of Chagas Disease in Latin-American Migrants Living in Europe: A Systematic Review and

Sin MLY, Mach KE, Wong PK, Liao JC. 2014. Advances and challenges in biosensor-based diagnosis of

Meta-analysis. PLoS Negl Trop Dis **9**:1–15.

infectious diseases. Expert Rev Mol Diagn 14:225–44.

<sub>56</sub> 306

58 307

60 308 

17.

	309	18.	Prata A. 2001. Clinical and epidemiological aspects of Chagas disease. Lancet Infect Dis 1:92–100.
1 2	310	19.	Pinto AY das N, Valente V da C, Coura JR, Valente SA da S, Junqueira ACV, Santos LC, Ferreira AG, de
3 4	311		Macedo RC. 2013. Clinical Follow-Up of Responses to Treatment with Benznidazol in Amazon: A Cohort
5 6	312		Study of Acute Chagas Disease. PLoS One <b>8</b> :1–9.
7 8 9	313	20.	Morillo CA, Marin-Neto JA, Avezum A, Sosa-Estani S, Rassi AJ, Rosas F, Villena E, Quiroz R, Bonilla R,
10 11	314		Britto C, Guhl F, Velazquez E, Bonilla L, Meeks B, Rao-Melacini P, Pogue J, Mattos A, Lazdins J, Rassi A,
12 13	315		Connolly SJ, Yusuf S, BENEFIT Investigators. 2015. Randomized Trial of Benznidazole for Chronic
14 15	316		Chagas' Cardiomyopathy. N Engl J Med <b>373</b> :1295 – 1306.
	317	21.	Aparecido A. PF, Carolina VU, Michelle DSC, Naira CP, Marcos VF, Glauco P dos S, Cecílio SF, Assis VB,
	318		Hideki Y. 2013. Amperometric Biosensor for Diagnosis of DiseaseState of the Art in Biosensors -
20 21 22	319		Environmental and Medical Applications.
23 24	320	22.	Stimpert KK, Montgomery SP. 2010. Physician Awareness of Chagas Disease, USA. Emerg Infect Dis
25 26	321		<b>16</b> :871–872.
27 28	322	23.	Ventura-Garcia L, Roura M, Pell C, Posada E, Gasc??n J, Aldasoro E, Mu??oz J, Pool R. 2013. Socio-
29 30	323		Cultural Aspects of Chagas Disease: A Systematic Review of Qualitative Research. PLoS Negl Trop Dis 7.
	324	24.	Fergnani N, Ruggiero A, Ceccarelli S, Menu F, Rabinovich J. 2013. Large-scale patterns in morphological
33 34	325		diversity and species assemblages in Neotropical Triatominae (Heteroptera: Reduviidae). Mem Inst
35 36 37	326		Oswaldo Cruz <b>108</b> :997–1008.

25. Ibarra-Cerdeña CN, Zaldivar-Riveron A, Peterson AT, Sanchez-Cordero V, Ramsey JM. 2014. Phylogeny and niche conservatism in North and Central American triatomine bugs (Hemiptera: Reduviidae: Triatominae), vectors of Chagas' disease. PLoS Negl Trop Dis. 8(10): e3266

37 38

39 40 327

328 41 42

43 329 44

45 330 46 47 331

333

334

48 49 332

50 51

52 53

54 55

57

59 60 337

61 62

63 64 65

26. Ramsey JM, Peterson a T, Carmona-Castro O, Moo-Llanes D a, Nakazawa Y, Butrick M, Tun-Ku E, la Cruz-Félix K De, Ibarra-Cerdeña CN. 2015. Atlas of Mexican Triatominae (Reduviidae: Hemiptera) and vector transmission of Chagas disease. Mem Inst Oswaldo Cruz 110:339-52.

27. Gurgel-Gonçalves R, Galvão C, Costa J, Peterson a T. 2012. Geographic distribution of chagas disease vectors in Brazil based on ecological niche modeling. J Trop Med 2012:705326.

<sub>56</sub> 335 28. Lescure F-X, Le Loup G, Freilij H, Develoux M, Paris L, Brutus L, Pialoux G. 2010. Chagas disease: 58 336 Changes in knowledge and management. Lancet Infect Dis 10:556–570.

> 29. Sutherst RW. 2004. Global Change and Human Vulnerability to Vector-Borne Diseases Global Change

338	3	and Human Vulnerability to Vector-Borne Diseases. Clin Microbiol Rev <b>17</b> :136–173.
1 2 <b>33</b> 9 3	30.	Githeko AK, Lindsay SW, Confalonieri UE, Patz JA. 2000. Climate change and vector-borne diseases: a
4 <b>340</b> 5	)	regional analysis. Bull World Health Organ <b>78</b> :1136–1147.
6 <b>341</b>	l 31.	Coura JR. 2013. Chagas disease: Control, elimination and eradication. Is it possible? Mem Inst Oswaldo
8 9 342	2	Cruz <b>108</b> :962–967.
10 11 343	32.	Bhattacharyya T, Falconar AK, Luquetti AO, Costales J a, Grijalva MJ, Lewis MD, Messenger L a, Tran
12 13 344	1	TT, Ramirez J-D, Guhl F, Carrasco HJ, Diosque P, Garcia L, Litvinov S V, Miles M a. 2014. Development
14 15 345	5	of peptide-based lineage-specific serology for chronic Chagas disease: geographical and clinical
16 17 <b>34</b> 6	5	distribution of epitope recognition. PLoS Negl Trop Dis <b>8</b> :e2892.
18 19 <b>347</b> 20	7 33.	Rocha-Gaso M-I, March-Iborra C, Montoya-Baides A, Arnau-Vives A. 2009. Surface generated acoustic
21 21 22	3	wave biosensors for the detection of pathogens: a review. Sensors (Basel) <b>9</b> :5740–69.
23 24 349	9 34.	Lee H, Lee S, Namkoong K, Ko C. 2008. SAW immunosensors for HBsAb detection. Sensors, 2008.
25 26 <b>35</b> 0	35.	Rocha-Gaso MI, Renaudin A, Sarry F, Beyssen D. 2015. Lab-on-a-chip based integrated hybrid
27 28 <b>351</b>	L	technologies for biofluids manipulation and characterization. Procedia Eng <b>120</b> :687–690.
29 30 <b>35</b> 2	2 36.	Beyssen D, Brizoual L Le, Elmazria O. 2006. Microfluidic device based on surface acoustic wave. Sensors
31 32 <b>35</b> 3	3	Actuators B.
33 34 35	<b>1</b> 37.	Roux-Marchand T, Beyssen D, Sarry F, Elmazria O. 2015. Rayleigh Surface Acoustic Wave As an
35 36 37 355	5	Efficient Heating System for Biological Reactions: Investigation of Microdroplet Temperature
38 39 <b>35</b> 6	5	Uniformity. IEEE Trans Ultrason Ferroelectr Freq Control <b>62</b> :729–735.
40 41 357	7 38.	Ferreira a. a P, Colli W, Da Costa PI, Yamanaka H. 2005. Immunosensor for the diagnosis of Chagas'
42 43 <b>35</b> 8	3	disease. Biosens Bioelectron <b>21</b> :175–181.
44 45 <b>35</b> 9	9 39.	Belluzo MS, Ribone MÉ, Camussone C, Marcipar IS, Lagier CM. 2011. Favorably orienting recombinant
46 47 360	)	proteins to develop amperometric biosensors to diagnose Chagas' disease. Anal Biochem <b>408</b> :86–94.
48 49 50	L 40.	Ferreira AAP, Colli W, Alves MJM, Oliveira DR, Costa PI, Güell AG, Sanz F, Benedetti AV, Yamanaka H.
51 52 362	<u>)</u>	2006. Investigation of the interaction between Tc85-11 protein and antibody anti-T. cruzi by AFM and
53 54 363	3	amperometric measurements. Electrochim Acta <b>51</b> :5046–5052.
55 56 <b>36</b> 4	41.	Vinicius Foguel M, Pilondos Santos G, Aparecido Pupim Ferreira A, Yamanaka H, Vicente Benedetti A.
57 58 <b>36</b> 5	5	2011. Amperometric immunosensor for Chagas' disease using gold CD-R Transducer. Electroanalysis
59 60 <b>36</b> 6	5	<b>23</b> :2555–2561.
61 62		15
63 64		15
65		

	367	42.	Erdmann CA, Kovalczuk E, Inaba J, Viana AG, Pessoa CA, Wohnrath K, Garcia JR. 2013. Development of
1 2	368		a Nano-particle enhanced impedimetric biosensor for Chagas' Disease DiagnosisXLII Annual Meeting of
3 4	369		SBBq.
5 6 7	370	43.	Coltro WKT, Neves RDS, Motheo ADJ, Da Silva JAF, Carrilho E. 2014. Microfluidic devices with
8 9	371		integrated dual-capacitively coupled contactless conductivity detection to monitor binding events in
10 11	372		real time. Sensors Actuators, B Chem <b>192</b> :239–246.
12 13	373	44.	Sachse K, Frey J. 2002. PCR Detection of Microbial Pathogens: Methods and Protocols, p <i>In</i> Totowa, N
	374		(ed.), Methods in Molecular Biology, 1st ed. Humana Press Inc, Springer.
	375	45.	Diguta C, Rousseaux S, Weidmann S, Bretin N, Vincent B, Guilloux-Benatier MAH. 2010. Development
18 19 20	376		of a qPCR assay for specific quantification of Botrytis cinerea on grapes. FEMS Microbiol Lett <b>313</b> :81–
	377		87.
23 24	378	46.	Thacker J, Casale E, Tucker C. 1996. Immunoassays (ELISA) for rapid, quantitative analysis in the food-
25 26	379		processing industry. J Agric Food Chem <b>44</b> :2680–2685.
27 28	380	47.	Iqbal S, Mayo M, Bruno J, Bronk B, Batt C, Chambers P. 2000. A review of molecular recognition
	381		technologies for detection of biological threat agents. Biosens Bioelectron <b>15</b> :549–578.
<ul><li>31</li><li>32</li><li>33</li></ul>	382	48.	Andreotti P, Ludwig G, Peruski A, Tuite J, Morse S, Peruski L. 2003. Immunoassay of infectious agents.
34 35	383		Biotechniques <b>35</b> :850–859.
36 37	384	49.	Peruski A, Peruski L. 2003. Immunological methods for detection and identification of infectious
38 39	385		disease and biological warfare agents. Clin Diagn Lab Immunol <b>10</b> :506–513.
40 41	386	50.	Barfield CA, Barney RS, Crudder CH, Wilmoth JL, Stevens DS, Mora-Garcia S, Yanovsky J. 2010. A
	387		Highly Sensitive Rapid Diagnostic Test for Chagas Disease That Utilizes a Recombinant Trypanosoma
44 45 46	388		cruzi Antigen. IEEE Trans Bio-Medical Eng <b>58</b> :814–817.
	389	51.	Balcerzak A, Rejmund F, Gutkiewicz P, Zienkiewicz B, Zhavnerko G. 2014. Ultrasonic chemical sensor
49	390		with organic monomolecular layer. Arch Acoust <b>31</b> :47–52.
51 52 53 54 55 56 57	391		
58 59 60			
61 62			
63 64			16
65			