

Origin and Evolution of the Neuroendocrine Control of Reproduction in Vertebrates, with Special Focus on Genome and Gene Duplications

Sylvie Dufour, Bruno Quérat, Hervé Tostivint, Catherine Pasqualini, Hubert

Vaudry, Karine Rousseau

▶ To cite this version:

Sylvie Dufour, Bruno Quérat, Hervé Tostivint, Catherine Pasqualini, Hubert Vaudry, et al.. Origin and Evolution of the Neuroendocrine Control of Reproduction in Vertebrates, with Special Focus on Genome and Gene Duplications. Physiological Reviews, 2020, 100 (2), pp.869-943. 10.1152/physrev.00009.2019. hal-02383899

HAL Id: hal-02383899 https://hal.science/hal-02383899v1

Submitted on 15 Nov 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1	
2	Origin and evolution of the neuroendocrine control of reproduction in vertebrates, with
3	special focus on genome and gene duplications
4	
5	Sylvie Dufour ^{1*,} Bruno Quérat ² , Hervé Tostivint ³ , Catherine Pasqualini ⁴ , Hubert Vaudry ⁵ ,
6	Karine Rousseau ¹
7	
8	^{1.} Muséum National d'Histoire Naturelle, BOREA (Biology of Aquatic Organisms and
9	Ecosystems), CNRS, IRD, Sorbonne Université, Université Caen Normandie, Université des
10	Antilles, Paris, France
11	^{2.} Université Paris Diderot, Sorbonne Paris Cite, Biologie Fonctionnelle et Adaptative (BFA),
12	UMR CNRS 8251, Paris, France, INSERM U1133 Physiologie de l'axe gonadotrope, Paris,
13	France
14	^{3.} Muséum National d'Histoire Naturelle, Physiologie moléculaire et adaptation, UMR 7221
15	CNRS, Muséum National d'Histoire Naturelle, Paris, France,
16	^{4.} Université Paris-Saclay, Université Paris-Sud, CNRS, Paris-Saclay Institute of
17	Neuroscience (UMR 9197), Gif-sur-Yvette, France
18	^{5.} Université de Rouen Normandie, INSERM U1239, Rouen, France,
19	
20	Corresponding author: Sylvie Dufour; e-mail: sylvie.dufour@mnhn.fr
21	

23 24	Table of contents
25	
26	1. Introduction
27	
28	2. Looking for the origin and evolution of the neuroendocrine control of reproduction
29	2.1. Objectives
30	2.2 Bases of phylogeny
31	2.2.1. Phylogeny of vertebrates
32	2.2.2. Phylogeny of metazoa
33	2.2.3. Phylogeny of molecular families versus phylogeny of species
34	2.3. Origin and fate of duplicated genes
35	2.3.1. Intra-gene duplication
36	2.3.2. Local gene duplication
37	2.3.3. Whole genome duplication
38	2.3.4. Definition of homologous, orthologous, paralogous, or ohnologous genes
39	2.3.5. Fate of duplicated genes
40	
41	3. Origin and evolution of gonadotropins, the pituitary relay between brain and gonads
42	in vertebrates
43	3.1. Vertebrate gonadotropins
44	3.1.1. Discovery of pituitary gonadotropins
45	3.1.2. Non-pituitary gonadotropins
46	3.1.3. Extra-pituitary expression of LH and FSH
47	3.1.4. Main regulators of gonadal function
48	3.1.5. A role for gonadotropin subunit monomers or homodimers ?
49	3.2. Structural characteristics of gonadotropins
50	3.2.1. Global folding and general characteristics
51	3.2.2. α subunits
52	3.2.3. LHβ subunits
53	3.2.4. CGβ subunit
54	3.2.5. FSH β subunit
55	3.3. Origin and evolution of gonadotropins

- 57 3.3.2. Ancient origin of GPA2/GPB5 before the emergence of vertebrates
- 58 3.3.3. A limited role for vertebrate GPA2/GPB5 in reproduction
- 59 3.3.4. Evolution of the GPH family in vertebrates
- 60

61 4. Origin and evolution of GnRH, brain conductor of reproduction in vertebrates

- 62 **4. 1. Introduction**
- 63 4.2. Origin and evolution of GnRH
- 64 4.2.1. GnRH in mammals
- 65 4.2.2. GnRH in other vertebrates: contribution to the discovery of multiple GnRH
- 66 4.2.3. Evolution of GnRH genes in vertebrates: impact of WGD
- 67 4.3. Origin and evolution of GnRH receptors
- 68 4.3.1. GnRH-R in mammals
- 69 4.3.2. GnRH-R in other vertebrates: contribution to the discovery of multiple GnRH-R
- 70 4.3.3. Evolution of GnRH-R genes in vertebrates: impact of WGD
- 71 4.4. Origin of the GnRH system before the emergence of vertebrates
- 72 4.4.1. GnRH and GnRH-R in non-vertebrate chordates
- 73 4.4.2. GnRH-like peptides and GnRH-R in other non-vertebrates
- 4.4.3. Other GnRH-related peptides (AKH, CZR, ACP) and their receptors
- 75 4.4.4. Evolutionary relationships between GnRH- and AKH-CRZ-ACP-like systems
- 76 4.4.5. GnRH and GnRH-R before bilateria?
- 77 4.5. Functions of GnRH-like peptides: evolutionary perspectives
- 78 4.5.1. Reproductive functions of GnRH in chordates
- 79 4.5.2. Non-reproductive functions of GnRH in chordates
- 80 4.5.3. Roles of GnRH-like peptides in non-chordates
- 81

82 5. Origin and evolution of brain RF-amide peptides in the limelight for reproduction

- 83 5.1. History and evolutionary relationships of the RF-amide peptides
- 84 5.1.1 Discovery of RF-amide peptides in mollusks
- 85 5.1.2. RF- amide peptides superfamily in vertebrates
- **5.2.** GnIH (*farp-3*) and evolutionary-related peptides (NPFF and QRFP) (*farp-1 and -5*)
- 87 5.2.1. Discovery of GnIH in birds: direct inhibitory role on pituitary gonadotropins
- 88 5.2.2. GnIH inhibitory role at the brain and gonadal levels in birds
- 89 5.2.3. GnIH in mammals

- 90 5.2.4. Current studies on the presence and role of GnIH in the reproduction in other 91 vertebrates
-
- 92 5.2.5. Origin and evolution of GnIH and its receptors in vertebrates
- 93 5.2.6. NPFF (PQRF) family (*farp-1*)
- 94 5.2.7. QRFP (26RFa) family (*farp-5*)
- 95 **5.3.** A raising star in mammalian reproduction: Kisspeptin (*farp-4*)
- 96 5.3.1. History of the discovery of Kiss in mammals
- 97 5.3.2. Role of Kiss in the control of reproduction in mammals
- 98 5.3.3. Multiple Kiss and Kiss receptor genes in vertebrates: origin and evolution
- 99 5.3.4. Current studies on the role in reproduction of Kiss in non-mammalian vertebrates
- 100 5.3.5. Kiss before vertebrates
- 101 5.3.6. Kisspeptin, more related to galanin/spexin than to other RF-amide peptides
- 102 **5.4. Prolactin Releasing-Peptide (PrRP) group (***farp-2***)**
- 103 5.5. Conclusion on RF-amide peptides
- 104
- 105 6. Non-peptidic neuromediators and the control of reproduction
- 106 **6.1. Introduction**
- 107 6.1.1. Ancient, multifunctional neuromediators
- 108 6.1.2. Non-peptidic neuromediators produced via enzymatic synthesis: phylogeny of enzymes
- 109 to retrace their evolutionary history
- 110 6.1.3. Origin and evolution of neuromediator receptors: direct phylogeny
- 111 6.2. A key-neuromediator, dopamine
- 112 6.2.1. Major brain neuromediator in human and other mammals
- 113 6.2.2. Evolution of tyrosine hydroxylase, key-enzyme of dopamine synthesis
- 114 6.2.3. Evolution of Dopamine D2 receptors
- 115 6.2.4. Dopamine major inhibitory regulator of gonadotropins secretion in teleosts
- 116 6.2.5. Dopamine inhibitory regulator of gonadotropic axis in other vertebrates
- 117 6.2.6. Dopamine inhibitor of reproduction in other metazoan
- 118 6.3. Many other neuromediators involved in the control of reproduction
- 119 6.3.1. Norepinephrine (NE)
- 120 6.3.2. Serotonin (5-HT)
- 121 6.3.3. Histamine (HA)
- 122 6.3.4. GABA
- 123 6.4. Neuromediators and reproduction: From conservation to disruption

124 6.4.1. Ancient origin of neuromediators and receptors in metazoan

125 6.4.2. Non-peptidic neuromediators as potential targets for neuroendocrine disruption: a

126 phylogenetic approach

127 6.4.3. The "One Health concept"

- 128
- 129

130 Abstract

131 In human, as in the other mammals, the neuroendocrine control of reproduction is ensured by 132 the brain-pituitary gonadotropic axis. Multiple internal and environmental cues are integrated 133 via brain neuronal networks, ultimately leading to the modulation of the activity of 134 gonadotropin-releasing hormone (GnRH) neurons. The decapeptide GnRH is released into the 135 hypothalamic-hypophyseal portal blood system, and stimulates the production of pituitary 136 glycoprotein hormones, the two gonadotropins, luteinizing hormone (LH) and follicle-137 stimulating hormone (FSH). A novel actor, the neuropeptide kisspeptin (Kiss), acting 138 upstream of GnRH, has attracted increasing attention in recent years. Other neuropeptides, 139 such as gonadotropin-inhibiting hormone (GnIH) / RF-amide related peptide (RFRP), and 140 other members of the RF-amide peptide superfamily, as well as various non-peptidic 141 neuromediators such as dopamine and serotonin also provide a large panel of stimulatory or 142 inhibitory regulators. This paper addresses the origin and evolution of the vertebrate 143 gonadotropic axis. Brain-pituitary neuroendocrine axes are typical of vertebrates, the pituitary 144 gland, mediator and amplifier of brain control on peripheral organs, being a vertebrate 145 innovation. The paper reviews, from molecular and functional perspectives, the evolution 146 across vertebrate radiation of some key-actors of the vertebrate neuroendocrine control of 147 reproduction, and traces back their origin along the vertebrate lineage and in other metazoa 148 before the emergence of vertebrates. A focus is given on how gene duplications, resulting 149 from either local events or from whole genome duplication events (WGD), and followed by

paralogous gene loss or conservation, might have shaped the evolutionary scenarios of current
families of key-actors of the gonadotropic axis.

152

153 **1. Introduction**

154 The neuroendocrine control of reproduction has been extensively investigated in mammals 155 (for review: (370)). Two pituitary hormones, the gonadotropins, *i.e.* luteinizing hormone (LH) 156 and follicle-stimulating hormone (FSH), are released in the general blood circulation and 157 regulate gonad activity. The synthesis and release of gonadotropins is mainly controlled by a 158 neuropeptide, gonadotropin-releasing hormone (GnRH), produced by hypothalamic 159 neurosecretory neurons and released into the portal blood system connecting the median 160 eminence to the adenohypophysis. GnRH neuron activity is itself regulated by various brain 161 neuromediators, which allows the integration of multiple environmental and internal cues in 162 the course of developmental or seasonal control of reproduction (Figure 1).

163 Other major physiological functions in mammals are similarly controlled by brain-pituitary-164 peripheral gland axes, such as the thyrotropic axis (brain-pituitary-thyroid) for development 165 and metabolism, the somatotropic axis (brain-pituitary-liver) for body growth, the 166 corticotropic axis (brain-pituitary-adrenal gland) for stress and metabolism. The pituitary 167 gland and its role as a relay between the brain and peripheral glands or peripheral organs, is 168 considered as an innovation of the vertebrate lineage (for review: (641, 738, 740)). The 169 pituitary gland with its production of pituitary hormones under the control of brain 170 neurohormones is a common feature to all vertebrates, even though some anatomical 171 variations in the brain-pituitary relationships are observed. In mammals as in other tetrapods, 172 hypophysiotropic neurons project to the median eminence and neurohormones are released 173 into the pituitary portal system which irrigates the adenohypophysis, while, in teleost fishes, 174 hypophysiotropic neurons directly project up to the adenohypophysis which exhibits a high

175	regionalization of pituitary cell types (231, 287). In cyclostomes (hagfish and lamprey), brain
176	neuropeptides reach the adenohypophysis by diffusion from the third ventricle, which may
177	represent the ancestral pattern of connection between the hypothalamus and the pituitary in
178	vertebrates (for review: (738)).

We review here current information and hypotheses on the origin, conservation and variations of the key actors of the vertebrate brain-pituitary control of reproduction, with a special attention to the impact of gene and genome duplication events in the evolutionary scenario of their molecular families.

183

184 **2.** Looking for the origin and evolution of the neuroendocrine control of reproduction

185 **2.1. Objectives**

186 Comparative biologists have been investigating the control of reproduction as well as of other 187 physiological functions in non-mammalian vertebrates and other metazoa. Comparative 188 studies not only provided specific information on species of interest, but also led to major 189 advances for basic research. A remarkable example concerns the "neurosecretion" process, 190 *i.e.* the ability of certain neurons to synthesize and release hormones, a property exhibited for 191 instance by GnRH neurons. Hypothalamic neurosecretory neurons were first described in 192 1928 in a teleost fish species, *Phoxinus laevis*, by Ernst Scharrer (679), who established the 193 concept of "neurosecretion", a few years after the description by Carl Speidel (1919) (742) of 194 some "glandular" neurons in the spinal cord of chondrichthyan fish, the skates (for reviews: 195 (665, 678)). In the same line, the first RF-amide peptide, designated after the C-terminal 196 arginine (R) and amidated phenylalanine (F) residues, has been isolated in 1977 from the 197 ganglia of a mollusk, the Venus clam, Macrocallista limbosa (623). Since then, an increasing 198 number of RF-amide peptides have been identified in all metazoa, constituting the RF-amide

superfamily and including some brain peptides involved in the neuroendocrine control ofreproduction in vertebrates (see section 5).

201 Comparative and evolutionary reproductive neuroendocrinology is arousing interest for 202 various disciplinary fields, thanks to the variety of biological models investigated and the 203 availability of tools such as genome sequences. There is an increasing attention to teleosts 204 (e.g. zebrafish) as vertebrate models for biomedical research, and an increasing diversity of 205 species investigated worldwide for their relevance to agriculture, aquaculture, conservation, 206 or environmental assessment. Comparative and evolutionary neuroendocrinology aims at 207 tracing back the roots and the evolution of the neuroendocrine systems, from molecular and 208 functional perspectives, and at inferring their roles in the evolution of metazoan species and 209 lifecycles. For agronomic sciences, it contributes to increase basic and applied knowledge for 210 sustainable control of reproductive cycles in captivity, for understanding domestication 211 processes, for diversifying cultured species such as in aquaculture, and promoting sustainable 212 agro/aqua-systems. For conservation biology, it provides new knowledge for understanding 213 the role of neuroendocrine systems in species adaptation and adaptability to environmental 214 factors, a crucial topic in the context of global, climatic and anthropogenic changes. In 215 biomedical sciences, comparative and evolutionary neuroendocrinology contributes to 216 innovative advances by providing alternative vertebrate models suitable for experimentation 217 such as zebrafish, by revealing the evolutionary conservation of key neuroendocrine actors 218 and of their sequences, the mutation of which may lead to severe pathologies, or by 219 highlighting common processes in the impact of environmental stressors and pollutants 220 leading to the disruption of the reproductive capacity of wild species, domesticated animals or 221 humans, in the frame of the "One health" concept.

222

223 **2.2 Bases of phylogeny**

In order to investigate and compare neuroendocrine systems in an evolutionary perspective, it is necessary to relate the data obtained in a given species to the phylogenetical position of this species. Phylogeny of vertebrates and metazoa is by itself an ongoing research topic, based on multiple disciplinary fields such as paleontology, comparative anatomy, embryology, and more recently molecular systematics and phylogenomics. We are using, in the present manuscript, recent and largely consensual advances in vertebrate and metazoa phylogeny.

230

231 <u>2.2.1. Phylogeny of vertebrates</u> (Figure 2)

232 All vertebrate species belong to a monophyletic clade, *i.e.* a group of species that consists of 233 all the descendants of a common ancestor. Among the vertebrate clade, several "subclades" 234 can be distinguished. The most ancient (or most basal) is represented by cyclostomes (such as 235 lampreys and hagfish). Cyclostomes are the sister clade of all other vertebrates characterized 236 by the innovation of the jaws and therefore called gnathostomes. The gnathostome clade is 237 further divided into two sister clades, the chondrichthyans (such as sharks and ray fishes) and 238 the osteichthyans. The osteichthyan clade is further divided into two major sister clades: (i) 239 the actinopterygian clade that encompasses teleost fishes (the largest current group of 240 vertebrates with more than 25,000 species including most fisheries, aquaculture and model 241 fish species) and some other "fish" clades such as holosteans (gar); (ii) the sarcopterygian 242 clade that encompasses the tetrapods including mammals, and also some "fish" species such 243 as the coelacanth, representative species of actinistia, a basal group among sarcopterygians. 244 The current vertebrate phylogeny highlights in particular that the name "fish" is not by itself 245 phylogenetically relevant, as it may apply to species from very distinct vertebrate clades, such 246 as cyclostomes (lampreys), chondrichthyans (sharks), actinopterygians (gar, zebrafish), and 247 sarcopterygians (coelacanth). Thus, human and coelacanth are phylogenetically closer than 248 coelacanth and zebrafish or than human and zebrafish.

249

250 <u>2.2.2. Phylogeny of metazoa</u> (Figure 3)

251 While the broad lines of vertebrate phylogeny remained similar to the traditional classification 252 based on paleontology, comparative anatomy and embryology, the recent advances in 253 molecular systematics and phylogenomics largely challenged the classification of other 254 metazoa (8, 170). We present here a most consensual phylogeny of the major clades of 255 metazoa. Bilateria, the major clade among metazoa, gather animals with a bilateral symmetry 256 (at least at the embryonic stage) and with three tissue layers (endoderm, mesoderm, ectoderm) 257 during embryogenesis so that they are also called triploblastics. Besides bilateria, the 258 respective classification of the basal groups of metazoa, such as cnidaria (corals), porifera 259 (sponges), and others, is still largely under debate. Bilateria encompass two major clades, on 260 the one hand, deuterostomes which include chordates and among chordate the vertebrates, and 261 on the other hand, protostomes which include two major sister clades, ecdysozoa (flatworms, 262 insects, crustaceans,...) and lophotrochozoa (annelids, mollusks, ...) (8, 608).

263

264 2.2.3. Phylogeny of molecular families versus phylogeny of species

265 Phylogenetical analyses of protein families of interest, such as hormones / neuropeptides / 266 receptors / enzymes, potentially involved in the neuroendocrine control of reproduction, are 267 performed in order to compare sequences between various vertebrate or metazoa species. 268 Phylogenetic trees of molecular families highlight protein sequence conservation or 269 divergence and lead to hypotheses on evolutionary relationships between sequences and 270 possible functional variations. The phylogenetic tree of a given protein family does not 271 always follow the phylogenetic tree of the corresponding species, which makes it possible to 272 infer further hypotheses on the duplication, divergence, convergence and loss of 273 corresponding protein-coding genes.

274

275 **2.3. Origin and fate of duplicated genes**

Gene duplications produce so-called paralogous genes, and are considered as major drivers of
evolution, as they provide new gene pools allowing the emergence of novel functions, organs
and regulatory processes.

279

280 <u>2.3.1. Intra-gene duplication</u>

Besides gene duplication, intra-gene duplication of exons also contributes to the evolution of neuroendocrine systems, as exemplified by the multiple copies of thyrotropin-releasing hormone (TRH) peptide coded by the TRH gene (397), or by up to three homologous gonadotropin-inhibiting hormone (GnIH) peptides coded by the GnIH gene (see section 5). Intra-gene peptide duplication may lead to the amplification of peptide production or, after mutations of some of the duplicated sequences, to the production of distinct peptides with potentially differential activities.

288

289 2.3.2. Local gene duplication

290 Gene duplication produces paralogous genes and may originate from local genomic events, 291 targeting either a specific gene or a limited genomic region. If no major genomic 292 restructuration occurs afterwards, local duplication events can be traced back in extant species 293 genomes by tandem position or by a relatively close position of the paralogous genes on the 294 same chromosome. For instance, a local gene duplication in early gnathostomes is 295 hypothesized to be at the origin of the two gonadotropin receptors, LH receptor and FSH 296 receptor, positioned in tandem in tetrapods, including human, as well as in basal 297 sarcopterygian (coelacanth) and basal actinopterygian (gar) (468). The conservation of the 298 tandem position throughout vertebrate evolution may be related to conserved coordination of gene expression, *via* chromatin accessibility and common regulatory factors. In the primate lineage, a specific gene duplication of the LH beta subunit (LH β) gene, that occurred much more recently, is at the origin of the chorionic gonadadotropin beta subunit (CG β), involved in the regulation of an innovative reproductive organ, the placenta; further CG β duplications led to up to six hCG β genes in human arranged in a gene cluster together with the LH β gene on chromosome 19 (for review: (522)) (see section 3).

305

306 2.3.3. Whole genome duplication

307 Gene duplications may also originate from whole genome duplication events (WGD), as 308 proposed by Ohno in vertebrates (548). Gene paralogs stemming from WGD are thus named 309 "ohnologs" by certain authors. It is now consensually admitted that two successive WGD, 310 referred to as "1R" and "2R", for "first round" and "second round" of whole genome 311 duplications, have occurred at the base of the vertebrate lineage (Figure 2), even though the 312 exact timing of the second WGD is still under discussion (for review: (284)). These 1R and 313 2R WGD, which occurred about 450 Million years ago, are still traceable in the current 314 human genome, by fourway paralogous regions (paralogons) covering a large proportion of 315 the human genome (145). It has been advanced that the two rounds of WGD may have 316 contributed to the evolutionary success of the vertebrate lineage.

Genome sequencing of teleost fish models, such as medaka, fugu or zebrafish, revealed an additional WGD, referred to as 3R (for 3rd round of WGD) (for review: (484)) (Figure 2). When occurred this 3R WGD in the actinopterygian lineage event, remained under debate until recently, as more genomes became available including for species of key phylogenetic positions such as a non-teleost actinopterygian, an holostean, the spotted gar (72), or a basal teleost, an elopomorph, the eel (273, 274). These studies concluded that, in the course of actinopterygian evolution, the 3R would have specifically occurred in the teleost lineage, in an early teleost ancestor before the emergence of the basal group of elopomorphs. This 3R
event is thus also referred to as TWGD or TGD (teleost whole genome duplication) (586), and
is hypothesized to have contributed to the diversification of this group, the largest among
vertebrates.

328

329 <u>2.3.4. Definition of homologous, orthologous, paralogous, or ohnologous genes</u>

330 These definitions are important in order to analyze the molecular and functional relationships 331 and diversification processes among molecular families including those involved in 332 neuroendocrine systems. Homologous genes are present in the same or different species, and 333 all derive from a common ancestral gene. Homologous genes include orthologous, paralogous 334 and ohnologous genes. Orthologous genes are genes present in different species and evolved 335 from a common ancestral gene in the course of speciation (emergence of new species over the 336 course of evolution). Paralogous genes are present in the same or in different species and 337 result from the duplication of a common ancestral gene, either via local gene specific 338 duplication or via WGD. Ohnologs are paralogs resulting from WGD.

339

340 <u>2.3.5. Fate of duplicated genes</u>

341 Gene duplication produces two paralogs, which initially exert the same redundant function. In 342 most cases, this redundancy leads to a rapid loss of one of the two paralogs, via accumulation 343 of mutations due to redundancy-related lack of selective pressure. However, both paralogs 344 may be conserved after duplication due to evolutionary selection related to amplification of 345 initial function, or sharing of initial pleiotropic functions (subfunctionalisation), or emergence 346 of a new function (neofunctionalisation). Duplications followed by gene amplification, 347 subfunctionalisation, neofunctionalisation, or loss, are involved in the evolutionary scenarii of 348 the neuroendocrine gonadotropic axis.

349

350 3. Origin and evolution of gonadotropins, the pituitary relay between brain and gonads351 in vertebrates

352 **<u>3.1. Vertebrate gonadotropins</u>**

353 <u>3.1.1. Discovery of pituitary gonadotropins</u>

354 A role of pituitary gland activity on the development of gonads was suggested as far back as 355 1910 (132) by studies showing that hypophysectomy leads to an atrophy of genital organs. 356 The first experiment describing the stimulatory effects of bovine pituitary extracts on rat 357 ovarian growth and luteinization was published in 1922 (180), demonstrating that the anterior 358 part of the pituitary gland contains a gonadotrophic factor. It took a decade before this factor 359 appeared to be composed of two distinct fractions in ewes, one responsible for growth of 360 ovarian follicles and the other for luteinization (190). These two fractions were accordingly 361 named follicle-stimulating hormone (FSH) and luteinizing hormone (LH), respectively. 362 Technical improvement in glycoprotein purification methods, and notably the development of 363 ion-exchange chromatography, made possible a better assignment of the roles of LH and 364 FSH. In hypophysectomised rats, it was found that FSH acts on granulosa cells of the 365 Graafian follicles and LH on the theca interna and ovarian interstitial tissue, the combination 366 of the two stimulating estrogen secretion, ovulation and *corpus luteum* formation (834). The 367 dimeric nature of LH, first suggested by Li and Starman (1964), was confirmed by Papkoff 368 and Samy (1967) (576) who characterized two dissimilar subunits in ovine LH. The 369 biological activity of the reconstituted dimer was finally demonstrated by De la LLosa and 370 Jutisz (1969) (382).

Another factor was also purified from the bovine pituitary gland that stimulates the thyroid
(126). Homology between this thyroid-stimulating hormone (TSH) and the gonadotropins was
suggested after experiments demonstrating that mammalian LH and FSH are able to stimulate

374 the thyroid of teleost fish (203, 204). Soon after, it was established that LH, FSH and TSH 375 have one interchangeable subunit, the glycoprotein hormone (GPH) α subunit, whereas the β 376 subunit determines the hormonal specificity (609). The first amino acid sequence of an LH α 377 subunit was deciphered in 1971 (428) and its identity with the FSH α subunit was 378 demonstrated in 1974 (710). The first sequences of β subunits also appeared in the early 70s 379 ((428) for LH; (417) for TSH; (710) for FSH), confirming the homology between thyrotropin 380 and gonadotropin β subunits. It was thus established by the mid-seventies that the two 381 pituitary gonadotropins and the thyrotropin belong to a family of dimeric glycoproteins that 382 share the α subunit, whereas the three β subunits, which confer hormonal specificity, are 383 derived from a common evolutionary ancestor.

The demonstration of the existence of two pituitary gonadotropins was laboriously extended to all tetrapod groups (for review: (309)); teleosts ((758), for review: (407); chondrostei (628); dipnoi (627); chondrichthyans (629)), showing that all jawed vertebrates are provided with the same pituitary gonadotropic molecules. Up to now, a single gonadotropin has been discovered in the cyclostomes, in hagfish (819) and in lamprey (741) even though, in this later species, the α subunit has still to be identified (739). Glycoprotein hormones (*stricto sensu*), like all other pituitary hormones, are not found before the emergence of vertebrates (83).

391

392 <u>3.1.2. Non-pituitary gonadotropins</u>

The presence of a luteotrophic factor in urine of pregnant women was first observed in 1927 (24), leading to the development of the first pregnancy detection test. This gonadotropic factor was shown to be produced by placental cells in culture (327) and more specifically by syncytiotrophoblast cells (485) and named human chorionic gonadotropin (hCG). The similarity with LH in terms of biological activity was rapidly observed (439) and its ability to bind the same receptor as LH was later demonstrated (92), hence the LH-CG receptor naming.

399 Eventually, the close structural relatedness of hCG to LH was confirmed when sequencing 400 data were made available (765). The human CG β subunit has later been found to be expressed 401 in the pituitary (61) and testis (53, 583). Pregnant mare serum was also shown to contain a 402 gonadotropic factor (123) that was called pregnant mare serum gonadotropin (PMSG) before 403 its origin as a chorionic cell product was recognized and its name changed to equine chorionic 404 gonadotropin (eCG). It was later demonstrated that, unlike their human counterparts, eCG and 405 eLH β subunits are encoded by the same gene (702). LH β subunit expression also occurs in 406 the placenta of a marsupial (482) and possibly in diverse eutherian mammals (119).

407

408 <u>3.1.3. Extra-pituitary expression of LH and FSH</u>

409 LH and FSH are also expressed in the gonads and the brain. The first report of gonadal 410 expression of gonadotropin described the immunological detection of LH in human and rat 411 Leydig cells and that of FSH in Sertoli cells (849). LHB subunit transcripts were further 412 characterized in elongated spermatids in the rat (910), and were found to possess an extended 413 5' UTR end resulting in the same coding sequence as the pituitary transcript apart from the 414 first three amino acids of the signal peptide (909). In female rat, FSH β , LH β and the common 415 α subunits are produced by theca cells, *corpora lutea* and oocytes, whereas granulosa cells 416 only express LH subunits. The ovarian LH β subunit gene uses the same alternative promoter 417 as in the testis (680). Expression of this transcript is reduced by administration of PMSG, 418 which activates both LH and FSH receptors in non-equine species (249, 419). Interestingly, in 419 vitro granulosa cell expression of LHB mRNA is also decreased in response to a GnRH 420 agonist, likely through FSH receptor activation (425). Both LH and FSH are expressed in the 421 Siberian hamster ovary with levels varying according to the estrus cycle but in an opposite 422 fashion as in the pituitary (699). No specific role has been described for these gonadal 423 gonadotropins. Whether gonadal expression of gonadotropins is a common feature in mammals or other tetrapods remains to be clarified but this seems to be the case in teleosts (4, 172, 626, 705, 706, 728, 879, 882). It has to be noted that the seabream ovarian LH β subunit transcript is also extended in its 5' end, suggesting the existence of a conserved alternative and gonadal-specific promoter for the LH β gene. However, the expression of the two ovarian gonadotropins is up-regulated by GnRH in the sea bream (879). Ovarian LH β subunit expression in the sole also varies according to the ovarian maturation stage, like in the pituitary (706).

Another important site of expression of gonadotropins in teleosts is the brain (172, 496, 580, 705). It was notably observed in a cichlid that the two gonadotropins expressed in the preoptic area potentially play a regulatory role on pituitary expression of gonadotropins (574). LH is also expressed in rat hypothalamus in a manner that depends on the presence of a functioning pituitary (177). Brain gonadotropins may be involved in neuronal regulation of steroid synthesis (427).

437

438 <u>3.1.4. Main regulators of gonadal function</u>

439 Genetic mice models where either the FSH β (380) or LH β (442) subunit or the FSH (1) or LH 440 (908) receptor is deleted show that ovarian differentiation and growth are not dependent on 441 pituitary gonadotropin stimulation even if FSH stimulates proliferation of granulosa cells in 442 pre-antral follicles. In teleosts, timing of pituitary gonadotropin expression as compared to 443 gonadal differentiation also pleads for a process that does not involve gonadotropins (518). 444 Mammalian FSH receptors are expressed in granulosa cells from the secondary follicle stage 445 (follicles with at least two layers of granulosa cells) onward and LH receptor expression on 446 theca cells rapidly follows. After puberty onset, FSH is indispensable for the cyclic 447 recruitment of follicles (after oocytes have reached their maximal size) and terminal 448 folliculogenesis. LH is involved in the final steps of pre-ovulatory follicle maturation,

allowing meiosis to resume and inducing ovulation (6, 239, 474). LH is responsible for the
production of aromatizable androgens by theca cells. These androgens then reach granulosa
cells where they are converted into estrogens under the stimulatory action of FSH (371, 708).
At mid-cycle, granulosa cells of recruited follicles acquire LH receptor expression and initiate
progesterone production, the stimulation of which is later relayed by hCG. Progesterone is
essential for ovulation and for preventing further follicle maturation.

As for the ovary, pituitary gonadotropins are not required for testicular differentiation but FSH stimulates proliferation of Sertoli cells in prepubertal testis (399, 448). At puberty onset, FSH promotes Sertoli cell function but its importance for spermatogenesis depends on taxonomic groups (297). LH stimulates proliferation of adult-type Leydig cells responsible for the production of steroid hormones (792) and activates steroidogenesis and therefore, spermatogenesis.

461

462 <u>3.1.5. A role for gonadotropin subunit monomers or homodimers?</u>

463 The gonadotropin receptors are poorly activated by gonadotropin subunit monomers or 464 homodimers. However, the hCG^β subunit is produced in large excess in most epithelial 465 cancers and is believed to play an autocrine angiogenic and anti-apoptotic role, possibly 466 through interacting with components of cysteine-knot growth factor (and notably TGF^β) 467 receptor complexes (124, 303). Free glycoprotein hormone α subunit is also produced in 468 excess by the pituitary and secreted throughout the menstrual cycle (255) and by the placenta 469 (635). In the placenta, the free α subunit is much more potent than the intact hCG in acting 470 synergistically with progesterone to induce decidualization of endometrial stromal cells (512) 471 and stimulates prolactin release from human decidual cells (65). Interestingly, the free α 472 subunit stimulates differentiation and proliferation of prolactin-producing cells in the foetal 473 and postnatal mammalian pituitary gland (31, 42, 95) and prolactin release by frog pituitary 474 cells in culture (544). The action of the free α subunit is not mimicked by LH or hCG 475 heterodimer, demonstrating that the free α subunit, like hCG β , may have a specific role 476 independent on the activation of the gonadotropin receptors.

477

478 **<u>3.2. Structural characteristics of gonadotropins</u>**

479 The glycoprotein hormones are non-covalently linked heterodimers composed of an α subunit 480 that is common to all gonadotropins and TSH and a specific β subunit that confers biological 481 specificity. In all species, the pituitary gonadotropin subunits are transcribed from single 482 genes although some teleost fishes may be provided with two α subunit genes, resulting from 483 genome duplication in recent ancestors (407). The α subunit gene contains four exons, the 484 first of which encoding the 5' untranslated region, the three others coding for a 92-100 amino 485 acid-long protein excluding the signal peptide. All pituitary glycoprotein hormone β subunit 486 genes contain three exons, the first one coding for the 5' untranslated region and, in some 487 species, part of the LH β signal peptide. The second intron-splicing site is located at the same 488 position in all β subunit genes.

489

490 <u>3.2.1. Global folding and general characteristics</u>

491 The most obvious characteristics shared by the α and all β subunits are the presence of one or 492 two glycosylation sites at conserved position and a conserved arrangement of cysteine 493 residues. The crystal structure of hCG was resolved almost simultaneously by two different 494 laboratories (390, 883). The structure of FSH was later deciphered (206) demonstrating that 495 the overall folding pattern of the two glycoprotein hormone dimers is very similar. Taking 496 into account the high degree of conservation of the glycoprotein hormone subunits in 497 vertebrates and in particular the special cysteine arrangements (with the exception of teleost 498 FSH β subunit as related below), it is tempting to assume that this organization is conserved 499 among vertebrate gonadotropins. Both the α and β subunits belong to the large cysteine-knot 500 family of proteins and more specifically to the TGF- β subfamily (13, 28, 271). They present a 501 similar folding producing flat elongated structure organized in a head to tail orientation in the 502 dimer. Each subunit contains four antiparallel β strands tightened with a central cystine-knot 503 motif from which three hairpin loops are deployed so that loops 1 and 3 from one subunit are 504 intertwined with loop 2 of the other subunit. The determining knot motif involves six cysteine 505 residues from each subunit (Figure 4). Four of them bridge β^2 and β^4 strands with two 506 disulphide bonds forming an eight-residue ring through which penetrates a third disulphide 507 bond linking the two other cysteine residues from the $\beta 1$ and $\beta 3$ strands. The α subunits 508 contain a total of 10 cysteine residues, the two additional pairs linking the β 1 to β 2 and β 3 to 509 β 4 strands tying the amino- and carboxy-termini to the core molecule. The β subunits have 510 twelve cysteine residues. In addition to those involved in the knot, two of them form a 511 disulphide bridge linking together the tips of loop 1 and loop 3 and two others, seven amino 512 acids apart from each other, are involved in an intra-chain bonding located at the end of the 513 β 4 strand. The last two cysteine residues form a disulphide bridge between loop 1 and the C-514 terminal extremity that constitutes the buckle of the "fastening belt" that wraps the carboxy-515 end of the β subunit around the α subunit loop 2. This latter structure is of crucial importance 516 for maintaining the stability of the dimer at the low concentrations encountered in the 517 peripheral circulation (13, 888).

518

519 <u>3.2.2. α subunits</u>

520 The α subunit is globally well conserved across vertebrates, a feature that is expected since a 521 mammalian α subunit is able to associate with a teleost LH β subunit to produce a heterodimer 522 that retains most of the biological activity of the native teleost LH (78, 201). Such stability in 523 structure probably results from the constraints imposed on the α subunit that has to efficiently

524 associate with three different β subunits. The α subunits show alternate variable and 525 conserved regions. The variable regions, where insertions/deletions are encountered, are 526 located at the level of loop 1 and loop 3 as well as in the amino-terminal end of the protein. 527 On the contrary, the most conserved regions lie in β_2 , β_3 and β_4 strands and in loop 2. Great 528 apes α subunit sequences are characterized by a 4-amino acid-long deletion in the amino-529 terminal end, at an intron-splicing site. Some teleostean or chondrostean sequences also show 530 deletions at this site. Surprisingly, mutations of cysteine codons occur in some sequences. 531 This is particularly the case in hagfish (*Epatretus atami*) where cysteines C2 and C4 are 532 replaced by an arginine and a tyrosine, respectively. C2 is usually involved in the disulphide 533 bridge with C7 that is part of the cysteine knot motif whereas C4 bridges with C1 to move β 1 534 and $\beta 2$ strands closer to each other (Figure 4). There is only one clone identified and 535 sequenced in hagfish (819), so these apparent mutations might result from artefacts in the 536 cloning or sequencing process. Nevertheless, a different assignment of the cysteine residues 537 may be operating in this subunit with C1 replacing C2 in the bonding with C7 and the absence 538 of bonding between $\beta 1$ and $\beta 2$ strands. Substitutions in cysteine residues are also observed in 539 the α subunit of the Elephant shark (*Callorhinchus milii* - NCBI NM001292594.1) where C9 540 is replaced by an arginine, of the Tiger salamander (Ambystoma tigrinum - CN048018.1) with 541 C6 also replaced by an arginine, and the Arabian camel (Camelus dromedarius -542 XM010987723.1) (but not C. bactarianus - XM 006178591.1 and C. ferus -543 XM 010948966.1) with C2 replaced by an aspartic acid. Whether these substitutions 544 correspond to sequencing errors or to true mutations remains to be clarified. Two 545 glycosylation sites are overall conserved except in crows (Corvus cornix - XM010404527.2, 546 C. brachyrhynchos - XM 08640241.2 and C. macrorhynchos - AB563175.1) where the 547 asparagine at the junction of loop 2 with the β 3 strand is replaced by an aspartic. 548 Carbohydrate side chains are predicted to impact on synthesis and storage, on receptor 549 binding and signal transduction, and eventually on metabolic clearance of gonadotropins 550 (271). Since three species at least out of this taxonomic group share this substitution, a 551 compensatory mechanism should occur for these gonadotropins to assemble and elicit full 552 biological activity.

553

554 <u>3.2.3. LHβ subunits</u>

555 LH β subunits are 112-121 amino acid-long, with the exception of equine LH β where a 10 556 nucleotide-long deletion induces a shift in the reading frame leading to an extension of the 557 carboxy-terminal end in very much the same as for hCG β (702). The lamprey gonadotropin β 558 subunit (Petromyzon marinus - AY730276.1) is also extended at the carboxy-terminal side. If 559 it is an ancestral characteristic, it has not been conserved in the hagfish (Eptatretus atami -560 AB546741.1), another cyclostome species. In comparison to the α subunit, there are no major 561 insertions/deletions within the mature proteins apart for single or two amino acid-long 562 deletion events in some species, occurring in the region of loop 2. The degree of conservation 563 at the amino acid level is lower than in the α subunit. In addition to the twelve cysteine 564 residues, conserved amino acids are found mostly within the β strands but also in the loops 565 that seem to be more constrained than in the α subunit. LH and TSH β subunits usually have 566 one glycosylation site conservatively located in the $\beta 1$ and the $\beta 2$ strand, respectively, 567 whereas FSH β subunits have both of them. One glycosylation site is actually found in a 568 conserved position in almost all LH β subunits. It is to be noted that, in the spotted green 569 pufferfish (Tetraodon nigroviridis - CR706772.2) and in the fugu Takifugu rubripes -570 XM 003963794.2), two related species, the unique glycosylation site is displaced to the 571 carboxy-terminal end, between C10 and C11, two cysteine residues that are linked by a 572 disulphide bridge. The zebrafish (Danio rerio - BC162111.1) LHB subunit has a second site 573 also at this position. Two glycosylation sites are also found in LH β subunits in primates

574 except in human. The two sites are located as in FSHB subunits. Remarkably, the 575 gonadotropin β subunit of cyclostomes, the most basal extant vertebrate species, is provided 576 with the same two glycosylation sites. It is not known whether cyclostomes have only one or 577 two pituitary gonadotropins. All TSH β subunits have two additional amino acids inserted in 578 loop 2 (467) so that the hormone identified in cyclostome that do not have these two 579 additional amino acids most likely belongs to the gonadotropin lineage. It is not known if this 580 GTH is homologous to LH or FSH or is ancestral to LH and FSH, hence the naming GTH for 581 gonadotropic hormone (739, 741, 819). A third potential glycosylation site is found in the 582 lamprey GTH^β subunits in their specific carboxyl extension.

583

584 <u>3.2.4. CGβ subunit</u>

The primate CG β subunit genes arose from a duplication of the LH β subunit gene just before the divergence between New World and Old World Monkeys (see 3.3 Origin and evolution of gonadotropins). These sequences are characterized by a frame shift in the coding sequence due to a single nucleotide deletion, 8 codons before the LH β stop codon. This event generated a carboxyl extension of the encoded protein including several serine residues that are Oglycosylated in trophoblast cells. One of the consequences of this higher glycosylation level is the higher half-life of hCG in the blood when compared to hLH (112).

592 Equine CG β subunits are encoded by the LH β subunit gene expressed in the trophoblast. 593 Surprisingly, a similar carboxyl extension phenomenon occurred in equidae albeit due to a 594 different mechanism. In equidae, a ten nucleotide-long deletion occurred, leading to the same 595 frame shift as in primates (702).

596

597 <u>3.2.5. FSHβ subunit</u>

598 The less conserved subunit is the β subunit of FSH. This is mainly due to their high

599 evolutionary rate in the teleost lineage (627). Up to twelve amino acid residues are deleted in 600 the most derived teleosts, the acanthomorphes, essentially in the loop regions. The most 601 spectacular feature of the teleost FSH β subunits is the rearrangement of the cysteine residues. 602 Although conserved in most basal teleosts including ostariophyses (carp and catfish related 603 species), cysteine C3, which is involved in the latch of the seatbelt loop with C12, is then lost. 604 The seatbelt loop is determinant for the ability of the dimer to keep associated once diluted in 605 the general circulation. Teleosts have developed alternative means to ensure the seatbelt to be 606 fastened. In more recently emerged taxonomic groups (clupeocephala), an additional cysteine 607 (which will be referred to as C0) is found at the amino-terminal end that is extended to the left 608 and includes amino acids aligned to the signal peptide in other taxonomic groups 609 (elopomorphes (341); salmonids (758, 761); percomorphes (225, 226, 373, 553, 866)). It was 610 found by mutating the hCG β subunit so as to reproduce this teleost-type cysteine pattern that 611 the seatbelt could be latched by a disulphide bridge between C0 and C12 (888). In 612 cyprinodontiformes, the mature sequence after cleavage of the signal peptide does not include 613 C0 (709). Instead, another cysteine residue is found either two amino acids downstream of the 614 usual C3 position or just following C5, and it is likely that these subunits use the natural 615 gonadotropin β subunit folding pattern. The cleavage site of the signal peptide has not been 616 characterized so far in ostariophyses. If the mature protein includes C0, their sequence would 617 totalise 13 cysteine residues. Another striking feature in FSH β subunits is the more flexible 618 position and number of glycosylation sites. FSHB subunits usually have two glycosylation 619 sites. The one on the β 1 strand is generally well conserved except in some teleosts. The other 620 site is absent or displaced to variable regions in most teleost species. In an extreme case, the 621 Atlantic halibut (AJ417768), the FSH β subunit is deprived of any glycosylation site. At the 622 opposite, a third site is occasionally found in some sharks, teleosts, turtles, birds and 623 crocodiles. The unique pattern of cysteine arrangements or glycosylation states of teleost FSHs offers remarkable examples to investigate structure-function relationships. It is to be noted that most of these studies have been performed on mammalian hormones expressed in mammalian cells cultured at 37°C and thus, far from the low temperature at which most teleost gonadotropin assemble and act.

628

629 **3.3. Origin and evolution of gonadotropins**

630 <u>3.3.1. Recent discovery of additional members of GPH family in mammals</u>

In search for ligands for orphan G-protein-coupled receptors in the human genome, Hsu and his colleagues identified two new glycoproteins, also present in other vertebrate genomes, that share numerous characteristics with the glycoprotein hormone subunits (293). They were called GPA2 (GPHA2) and GPB5 (GPHB5) as for the second glycoprotein hormone related α subunit and the fifth glycoprotein hormone related β subunit. GPA2 and GPB5 are able to associate into heterodimer and the heterodimer is able to stimulate the TSH receptor but not the gonadotropin receptors (523). The heterodimer was therefore named thyrostimulin.

638

639 3.3.2. Ancient origin of GPA2/GPB5 before the emergence of vertebrates

640 These glycoproteins, which are found in most bilateria (293, 581, 640, 645, 663), are now 641 referred to as GPA and GPB in invertebrates that lack the classical glycoprotein hormone 642 subunits (83). In most investigated invertebrates, the genes encoding GPA and GPB are 643 located close to each other without intervening genes (662), suggesting a functional or 644 regulatory link between them. A leucine-rich repeat-containing receptor related to the 645 glycoprotein hormone receptors (279, 437) was also identified in several invertebrate species 646 (645). The fly receptor (LGR1) binds and is activated by the fly GPA/GPB heterodimer (749). 647 Interestingly, in the parasitic wasp Nasonia (266), the leech Helobdella (832) and the 648 myriapode Strigamia (833), three species that lack GPA and GPB, the homologous receptor is also absent. Altogether, these data strongly suggest that the vertebrate glycoprotein hormone α and β subunits derive from a duplication of GPA and GPB, respectively, early after the emergence of vertebrates.

652

653 <u>3.3.3. A limited role for vertebrate GPA2/GPB5 in reproduction</u>

654 No consensus role for GPA and GPB in invertebrates have been proposed to date even if they 655 may have been involved in specific functions in different species (640). In vertebrates, few 656 data are yet available. Like GPHa, GPA2 harbours two glycosylation sites, the position of the 657 one just after loop 2 being conserved. The other one is located on the $\beta 1$ (by analogy with 658 GPH α) strand. GPA2 lacks the cysteine residues involved in the bonding between β 1 and β 2 659 strands. It contains, in addition to the six residues composing the cystine-knot motif, two 660 cysteine residues that are believed to form a disulphide bridge between loops 1 and 3. These 661 loops appear longer than in GPH α . GPB5 is encoded by two exons with the same intron/exon 662 splicing site as for GPH β subunits. It harbours only one glycosylation site, located on the β 3 663 strand, just following the cysteine knot motif. The main difference between the GPB5 and 664 GPH β subunits is the absence of the carboxy extension that constitutes the seat belt. As 665 mentioned earlier, this seat belt is essential for the stability of the dimer so it is believed that a 666 GPA2/GPB5 dimeric structure may not be maintained once diluted into the general 667 circulation (13). The thyrostimulin naming of the GPA2/GPB5 dimer is based on the ability 668 of the recombinant dimer to stimulate the TSH receptor but not gonadotropin receptors (523, 669 552). It is further supported by the hyperthyroid phenotype observed in mice chronically 670 injected with the recombinant dimer (552) and in mice ubiquitously overexpressing GPB5 671 (443). In the latter case, the thyrotropic effect was attributed to the dimer formed by the 672 overexpressed GPB5 with the GPA2 naturally expressed in the thyroid to activate the TSH 673 receptor. However, the thyrostimulin concept is not supported by the absence of obvious

674 thyroid phenotypes in GPB5 knocked-out mice (443, 552) except for a transient 675 hypothyroxinemia in juveniles (907). Surprisingly, no GPA2-knocked-out mice have been 676 described. The GPA2/GPB5 dimer is 10 to 20 times more active that TSH in stimulating TSH 677 receptor (552). Taking into account the wide variety of tissues expressing GPA2 and/or GPB5 678 (293, 520, 523, 552) and the TSH receptor (874), it is conceivable that the GPA2/GPB5 dimer 679 may act as a local activator of TSH receptor. However, numerous tissues do not seem to co-680 express GPA2 and GPB5 so that, as for the free α and hCG β subunits, other modes of action 681 as monomer or homodimer must be explored for GPA2 and GPB5. In any case, no 682 abnomalities in fertility were mentioned in GPB5 knocked-out mice (443, 552, 906, 907) so 683 that a major role of the GPA2/GPB5 dimer or of GPB5 by itself in the control of reproduction 684 is very unlikely.

685

686 <u>3.3.4. Evolution of the GPH family in vertebrates</u>

687 Taking advantage of the availability of sequenced genomes from species of phylogenetical 688 significance, comparative synteny analysis between amphioxus (a pre-vertebrate chordate) 689 and vertebrate glycoprotein hormone subunit loci allowed the reconstruction of their 690 evolutionary history in vertebrates (467, 663). A first local duplication of a tandemly 691 organized GPA and GPB locus occurred just before the radiation of vertebrates and the 692 emergence of cyclostomes, giving rise to the vertebrate forms of GPA and GPB (GPA2 and 693 GPB5) and to the ancestral glycoprotein hormone (ancGPH) α and β subunits (Figure 5). The 694 ancgpha gene was maintained close to gpa2-gpb5 whereas the $ancgph\beta$ gene was translocated 695 soon after the duplication. After the first round of WGD (1R), one copy each of gpa2 and 696 gpha was lost from one of the duplicated gpa2-gpha-gpb5 loci, leaving two copies of gpb5, 697 *i.e.* gpb5a in the remaining gpa2-gpha locus and gpb5b. After the second WGD (2R), only 698 one copy of the duplicated gpb5b was retained. From the duplicated gpa2-gpb5a-gpha locus,

699 one copy of the duplicated gpha and one copy of the duplicated gpa2-gpb5a were lost, 700 leaving a single $gph\alpha$ encoding gene and the remaining single copies of the tandemly 701 organized gpa2 and gpb5a. These two genes were kept in tandem in teleosts (663) and in the 702 Coelacanth (Ensembl scaffold JH127632; unpublished data). gpb5a was lost in tetrapods that 703 only retained gpb5b. The ancestral form of $gph\beta$ that was somehow parted from its original 704 environment soon after the original, local duplication was duplicated at the 1R giving rise to 705 an ancestral the gonadotropin β subunit gene (*ancgth* β) and to an ancestral TSH β subunit gene 706 (anctsh β) (Figure 5). The 2R duplicated the ancgth β to generate lh β and fsh β genes. One of 707 the duplicated copy of the *anctsh* β gene (encoding TSH β 2) was retained in chondrichthyes 708 and in lungfish and coelacanth but was lost before the radiations of teleosts and tetrapods 709 (467). Of note, after the Teleost-specific WGD (3R), of all duplicated gph genes only the 710 duplicated $tsh\beta 1$ gene was maintained (encoding TSH $\beta 3$ or TSH $\beta 1b$ according to a recent 711 nomenclature), all other duplicated glycoprotein hormone subunit genes being lost (467, 663). 712 As mentioned earlier, two copies of GPH α subunit-encoding genes may have been generated 713 in some teleost species, resulting from recent, specific tetraploidization events.

714 While a tandem duplication was at the origin of the glycoprotein hormone family in 715 vertebrates, the most recently appeared glycoprotein hormone, CG, was also generated by a 716 tandem duplication that occurred during the radiation of primates, just before the divergence 717 between New and Old World monkeys, some 35 to 55 Million years ago (272, 460). 718 Subsequent gene duplications led to a rising number of $CG\beta$ duplicates, that are not all 719 functional (in human, 4 out of 6 CG β genes are functional), up to about 50 copies in gorilla 720 (169). The New World monkeys have only one CG β and one LH β subunit gene. Surprisingly, 721 in the marmoset and probably in all New World monkeys, the LH β gene has lost its pituitary 722 functionality, and it is the CG β gene that is expressed in the pituitary and in the placenta from 723 specific promoters (514). The first duplication event probably affected only the LH β gene but subsequent duplications also concerned the neurotrophin NTF4 gene (also called NTF5 or NTF6) although only one copy retained its functionality. Surprisingly, phylogenetic analysis of LH β and CG β transcripts showed that CG β sequences tend to group with the LH β sequence from the same species instead of with CG β sequences from closely related species. This phenomenon was shown to be due to gene conversion *i.e.* nonreciprocal transfer of sequence information, where the LH β gene sequence and notably its first and second exons act as the parent for *cg\beta* genes conversion (460).

731

732 4. Origin and evolution of GnRH, brain conductor of reproduction in vertebrates

733 <u>4. 1. Introduction</u>

734 In the late 1920s, once the involvement of the pituitary in the processes of reproduction was 735 demonstrated, systematic investigations were initiated to decipher the link between the 736 hypothalamus and pituitary gonadotropin secretion (491, 614). Harris was the first to propose 737 the hypothesis of neurohumoral regulation of gonadotropin secretion (192). The question that 738 remained to be resolved was how information perceived in the CNS would be communicated 739 to the pituitary gland. In the early 1960s, several investigators reported, in crude hypothalamic 740 extracts, the existence of substances that specifically stimulated the secretion of luteinizing 741 hormone (LH) or folliculo-stimulating hormone (FSH), and that were probably polypeptides 742 (367). It was concluded therefore that the hypothalamus regulates the release of LH and FSH 743 by means of two releasing hormones (LH-RH and FSH-RH), transported to the anterior 744 pituitary via the hypothalamic portal circulation. From that moment, several teams devoted 745 their efforts to the isolation and identification of LH- and FSH-releasing hormones. The 746 isolation of the single substance able to stimulate the synthesis and secretion of both 747 gonadotropins was independently achieved in 1971 from porcine and ovine brain by the 748 groups of Schally (465) and Guillemin (16, 77), respectively. It was a decapeptide that was initially termed LH-releasing hormone (LHRH) or LH-releasing factor (LRF), but the
molecule is now generally referred to as gonadotropin-releasing hormone (GnRH). For this
seminal discovery, Schally and Guillemin were awarded the Nobel Prize in Physiology or
Medicine in 1977.

753

754 **4.2. Origin and evolution of GnRH**

755 <u>4.2.1. GnRH in mammals</u>

756 The first GnRH form isolated from pig and sheep is an amidated decapeptide with the 757 sequence pQHWSYGLSPGamide (77, 465). This sequence is identical in almost all 758 mammalian species investigated so far (556). As other neuropeptides, GnRH is synthesized as 759 part of a larger precursor molecule called pre-proGnRH. The full-length cDNA encoding this 760 precursor was initially cloned from a human placental library (687). In human, pre-proGnRH 761 is a protein of 92 amino acids that consists of the decapeptide, extended at the N-terminus by 762 a signal peptide, and at the carboxy-terminus by a Gly-Lys-Arg sequence necessary for 763 enzymatic cleavage and carboxy-terminal amidation of GnRH. The 56 aa-residue sequence 764 located in the C-terminal region of the precursor is called the GnRH-associated peptide 765 (GAP). The architecture of pre-proGnRH is conserved in other mammals.

The mammalian GnRH gene is composed of four exons and three introns (688). Exon 1 contains a part of the 5'-UTR. Exon 2 encodes the rest of the 5'-UTR, the signal peptide, the GnRH decapeptide, the amidation/proteolytic processing motif and the N-terminal part of the GAP. Exon 3 encodes the central portion of the GAP. Exon 4 encodes the C-terminal part of the GAP along with the 3'-UTR.

That GnRH is primarily localized in neurons located in the hypothalamus, particularly in the arcuate-median eminence region and the preoptic area (36, 403). In the median eminence,

GnRH nerve terminals secrete directly into portal capillaries. The peptide is then delivered by the portal circulation to the adenohypophysial gonadotrope cells (614). An important feature of the secretory activity of GnRH neurons is its pulsatile nature (43). The frequency and amplitude of secretory episodes are essential for the release as well as the synthesis of LH and FSH by gonadotrope cells (784).

It was shown that GnRH neurons arise from the nasal placode and migrate to their target regions in the hypothalamus during development (682, 881); for review:(880)) This finding is in agreement with the observations that humans deficient in GnRH also lack their sense of smell and with studies showing a loss of GnRH cells in animals with olfactory placode ablation (871, 880).

In 1994 was reported, for the first time in mammals, the occurrence of a second GnRH gene (347). Details regarding this second gene will be provided in the next section since it encodes a peptide whose sequence was already described in chicken since 1984 (500).

787

788 <u>4.2.2. GnRH in other vertebrates: contribution to the discovery of multiple GnRHs</u>

789 For a long time after it was discovered, GnRH was thought to have an invariant sequence 790 because all mammalian peptides that were characterized were identical (643). This view was invalidated in 1982 when the first GnRH variant, containing the substitution $\text{Arg}^8 \rightarrow \text{Gln}$, was 791 792 isolated from chicken hypothalami (364, 365, 498, 499). Several other molecular isoforms 793 were subsequently identified through peptide purification, cDNA cloning, and more recently 794 data mining. They were primarily found in teleosts, but also in amphibians, elasmobranches 795 and cyclostomes, and even in mammals (for review: (556)). They were generally named 796 according to the species in which they were initially isolated. However, a given form found in 797 a particular species or group can exhibit a much wider distribution, making this nomenclature 798 particularly confusing. For example, salmon GnRH (sGnRH) (703) is present in many other teleost species, including medaka and zebrafish (381, 555). Likewise, the mammalian form of
GnRH (mGnRH) is also found in frogs (127, 269) and fishes (228, 558) but not in the guinea
pig (323).

802 In 1984, the occurrence of a second GnRH was demonstrated in the chicken, called chicken 803 GnRH-II (cGnRH-II) in addition to cGnRH-I (500). Since then, it became evident that almost 804 all vertebrate species are able to produce more than one GnRH form, each encoded by a 805 peculiar gene (for review: (556)): two in most of them, including human (868) and even three, 806 notably in teleosts (870) but also in lampreys (349, 704, 715, 737, 757). Very interestingly, 807 cGnRH-IIs is present in all gnathostomes with exactly the same structure. As a result, in 808 species that express two GnRH forms, the cGnRH-II variant is systematically present in 809 combination with a second one which corresponds to mGnRH in most mammals (870), some 810 amphibians (127) and fishes (228), or cGnRH-I in birds (498, 500) and reptiles (436). In 811 teleost species possessing three forms, cGnRH-II is often associated with the sGnRH variant, 812 plus a third form whose structure is more variable (619).

The peptides belonging to the GnRH family share several characteristics: (i) They all are decapeptides with residues 1, 4, 9 and 10 perfectly conserved as well as modified amino-(pGlu) and carboxy-terminus (NH₂) (248) (Table 1). (ii) They originate from preproGnRH precursors that exhibit the same features as those of mammals (248). (iii) The general organization of their genes is similar (188), suggesting they all share a common ancestral gene.

In the brain of species containing several GnRH forms, each variant exhibits a peculiar distribution. In gnathostomes, three major GnRH neuronal systems were recognized (163). The first one, called the ventral forebrain GnRH system, is composed of neurons mainly located in the preoptic area of the hypothalamus and projecting toward gonadotrope cells in the pituitary. The GnRH molecular form synthesized by these neurons is variable depending

824 on the species: e.g. mGnRH, cGnRH-I, frog GnRH (fGnRH), sea bream GnRH (sbGnRH) or 825 guinea pig GnRH (gpGnRH). Functionally, this peptide corresponds to the hypophysiotropic 826 GnRH form initially characterized by Schally and Guillemin. The second GnRH system, 827 called midbrain GnRH system, is composed of neurons located in the midbrain tegmentum 828 near the third ventricle. These neurons project throughout the brain, especially in the midbrain 829 and hindbrain, but also in the pituitary in some teleost and amphibian species. They produce 830 the cGnRH-II form. The third system, called the terminal nerve GnRH system, has so far been 831 identified only in teleosts. Neurons of this system are mainly located in the terminal nerve 832 ganglion near the olfactory bulb, and project primarily to the telencephalon but also widely 833 into the whole brain, including the retina and the olfactory epithelium. They specifically 834 express the sGnRH variant (556). It is to note that, in some teleost species, cells from the 835 hypothalamic GnRH systems and cells from the terminal nerve GnRH system clearly overlap 836 in the forebrain (232, 233, 502). It was also shown that neurons from both systems share a 837 common embryonic origin: they arise in the olfactory system and migrate into the forebrain 838 (2, 3, 233, 234, 557, 869).

839

840 <u>4.2.3. Evolution of GnRH genes in vertebrates: impact of WGD</u>

841 Because GnRHs are short peptides, their evolutionary relationship was difficult to establish 842 solely on the basis of their primary structure (337). In 1998, a phylogenetic analysis of GnRH 843 prepro-hormones from 23 species of bony fish and mammals revealed the existence of three 844 evolutionarily distinct GnRH groups, named GnRH1, GnRH2 and GnRH3 (869). These three 845 groups globally coincide with the three GnRH forms identified on the basis of their location 846 in the brain: releasing form localized to the hypothalamus (GnRH1), form previously 847 localized solely to midbrain nuclei (GnRH2), and form localized to the telencephalon in 848 teleosts (GnRH3) (188).

Subsequent phylogenetic studies including newly discovered sequences confirmed this categorization, which is now widely accepted and is at the basis of the current nomenclature (248, 337, 556). The occurrence of three GnRH paralogous genes is also supported by synteny analysis (360, 381, 555, 643, 644, 790) since each GnRH gene is positioned in genomic regions containing common neighbors across teleost and tetrapod species.

854 GnRH1 and GnRH2 are clearly two "old" genes since the peptides they encode exhibit a very 855 well conserved structure among gnathostomes. In contrast, GnRH3 has long been considered 856 as a teleost-specific gene, suggesting a more recent emergence, possibly through the teleost-857 specific whole-genome duplication (3R) (248, 337, 556). However, this hypothesis was 858 rejected when it appeared that genes surrounding GnRH3 in teleosts are also present in 859 tetrapods (360, 381, 555, 790). The view that GnRH3 arose by an ancient duplication before 860 the teleost-tetrapod split was recently confirmed by the finding of a GnRH3 ortholog in 861 coelacanth and several species of elasmobranches (219, 644, 903).

862 In lamprey, three members of the GnRH family of neuropeptides have been identified, named 863 lamprey GnRH-I (IGnRH-I), IGnRH-II and IGnRH-III (349, 704, 715, 737, 757). The precise 864 relationship between lamprey peptides and the three subtypes of GnRH in other vertebrates 865 was difficult to resolve. Phylogenetic analysis initially led Sower et al. (2009) (740) to 866 propose that IGnRH-II may represent a vertebrate GnRH2 ancestral form and to classify 867 IGnRH-I and IGnRH-II in a fourth group of vertebrate GnRH peptides. Once again, both 868 assumptions were refuted by synteny analysis showing that IGnRH-II is a GnRH2 ortholog 869 while IGnRHI and IGnRH-III are two copies of GnRH3 that probably arose through tandem 870 duplication (143, 252, 644, 725). It should be noted that the presence of GnRH was suggested 871 in the hagfish hypothalamus by both radioimmunoassay and immunohistochemistry, but the 872 identity of the peptide remains to be determined (541).

34

873 It is now well accepted that the three GnRH paralogs appeared together through the 2R. In 874 strong support of this view is the observation in the genome of many species that GnRH1, 875 GnRH2 and GnRH3 loci, in addition to a fourth locus without GnRH but sharing homologous 876 neighboring genes, are part of a same tetra-paralogon (360, 790, 903). Moreover, a GnRH-877 like gene recently characterized in amphioxus is positioned in a locus surrounded by syntenic 878 homologs of the human GnRH tetra-paralogon (642). Taken together, these data suggest that 879 four GnRH paralogs, including a putative GnRH4, were present in the last common vertebrate 880 ancestor (360, 790) (Figure 6).

After 2R, the history of the GnRH family was driven, across vertebrate evolution, by multiple
and independent gene loss events, leading to the diversity of GnRH currently known.

The GnRH1 gene occurs is all gnathostomes but not in cyclostomes, suggesting gene loss specifically in this latter lineage. The GnRH2 gene is currently the most represented GnRH gene in living vertebrates. No GnRH3 gene has so far been isolated from any tetrapod, while it is present in several groups of fish. It thus appears that this gene was likely deleted during the early evolution of the tetrapod lineage. Finally, the putative GnRH4 gene probably disappeared shortly after 2R without leaving any trace in current species (Figure 6).

889 Apart from these general considerations, additional species-, genus- or family-specific losses 890 of a GnRH form subsequently occurred independently in different vertebrate lineages. For 891 example, some teleosts lost GnRH1 (salmon, trout, goldfish, zebrafish, arowana) or GnRH3 892 (eel, catfish, butterfly fish). In elephant shark, only GnRH1 and GnRH2, but not GnRH3, 893 were identified (536, 644). Also, certain mammalian species lost GnRH2 (rat, mice, cattle, 894 sheep, chimpanzee), and thus now possess only GnRH1 (746). It is to note that no effect of 895 the 3R can be recorded on the current number of GnRH genes in teleosts, suggesting massive 896 gene losses after 3R. In contrast, two GnRH3 are present in sockeye salmon (25), probably

due to the salmon-specific whole-genome duplication (4R) (420), while two tandemly
arranged GnRH1 exist in elephant shark (219) (Figure 6).

899 The detailed evolutionary relationship between the three GnRH paralogs is still not 900 completely understood. Phylogenetic analysis shows that GnRH1 and GnRH3 sequences 901 group into the same clade, suggesting they share a common ancestor (248, 349, 800). 902 Moreover, in teleosts, GnRH1 and GnRH3 neuronal systems exhibit striking similarities 903 regarding both their embryonic origin and their distribution in the brain (337, 556). Taken 904 together, these data support the notion that 1R generated on the one hand the GnRH1-GnRH3 905 ancestor and on the other hand the GnRH2-GnRH4 ancestor. Although appealing, this 906 scenario remains to be confirmed.

907

908 4.3. Origin and evolution of GnRH receptors

909 <u>4.3.1. GnRH-R in mammals</u>

910 GnRH receptors (GnRH-R) were initially characterized in the pituitary and several other 911 tissues such as the brain, placenta and gonads, as specific binding sites with high affinity for 912 mGnRH1 and its potent agonist or antagonist analogs (747). Some immortalized cell lines 913 derived from neoplastic tissues also proved to be an important source of GnRH-R (747). One 914 of them, the mouse pituitary $\alpha T3$ gonadotrope cell line, led to the molecular isolation of the 915 first GnRH receptor sequence, by expression screening for electrophysiological (809) or 916 calcium responses (634) in Xenopus oocytes. This sequence provided the basis for the cloning 917 of GnRH-R in several other species (506, 686).

GnRH-R is mainly expressed in gonadotrope cells of the pituitary, in agreement with its role
in the synthesis and release of gonadotropins, but it is also present in extra-pituitary tissues
including the brain (320), placenta, gonads, prostate, uterus and breast (108, 637). GnRH-R is
also expressed in cancer tissues, either related (*i.e.* prostate, breast, endometrial, and ovarian

922	cancers) or unrelated (i.e. melanoma, glioblastoma, lung, and pancreatic cancers) to the
923	reproductive system. Activation of GnRH-R was shown to inhibit proliferation of transformed
924	cells making this receptor a therapeutic target for direct action of GnRH analogues on certain
925	proliferating cells (422, 450).

926 Analysis of the primary sequence identifies the GnRH receptor as a member of the rhodopsin 927 β -like G protein-coupled receptor (GPCR) family, which contains a characteristic seven 928 transmembrane (TM)-domain structure (686). Unlike most GPCRs, which are intronless, the 929 GnRH-R gene is composed of three exons and two introns (181).

A hallmark feature of the mammalian GnRH-R is the absence of a cytoplasmic carboxyterminal domain, which is unique among GPCRs (686). As a result, this receptor is resistant to rapid desensitization and internalizes very slowly (142, 597). The extracellular loops (EC) and superficial regions of the transmembrane domains (TM) are responsible for receptorligand recognition and binding while the intracellular loops are involved in interacting with G proteins for signal transduction (196, 490).

In gonadotrope cells and gonadotrope-lineage cells, activation of the GnRH receptor stimulates a variety of intracellular signalling pathways (99, 525, 600). However, it is generally accepted that the primary effect of GnRH binding to its receptor is the activation of phospholipase C β (PLC β) through the G_{q/11} G protein and the hydrolysis of phosphatidylinositol 4,5-bisphosphate (PIP2) into diacylgycerol (DAG) and inositol 1,4,5trisphosphate (IP3), which stimulates calcium release and activates protein kinase C (PKC) (247). These events lead to the synthesis and release of gonadotropins (747).

The presence of at least two GnRHs in most vertebrates, including mammals, suggested the existence of several cognate GnRH receptor subtypes (793). In support of this view, the occurrence of a second GnRH-R subtype (called GnRH-RII, as opposed to the first subtype now renamed GnRH-RI) was reported in non-human primates (488, 530). It is noteworthy that the GnRH-RII subtype is either partially or completely silenced in a number of
mammalian species (human, chimpanzee, sheep, cow, rat, and mouse) (487, 506, 507).

A study carried out in marmoset showed that the type II GnRH receptor is more widely distributed than type I receptor (488). As expected, GnRH-RI exhibits greater sensitivity to the hypophysiotropic GnRH form, GnRH1, than to GnRH2. Conversely, the GnRH-RII shows greater sensitivity to GnRH2 than to GnRH1 (304, 449, 488).

953

954 <u>4.3.2. GnRH-R in other vertebrates: contribution to the discovery of multiple GnRH-R</u>

955 The first non-mammalian GnRH-R was isolated from catfish in 1997 (778). Remarkably, this 956 receptor exhibits a 49-amino acid cytoplasmic carboxy terminal domain that is lacking in the 957 mammalian receptors. Moreover, it possesses a much higher affinity for GnRH2 than for 958 GnRH1. A number of subsequent studies described the cloning of GnRH receptors in several 959 other species, including trout (447), Xenopus (794), and chicken (754). In some species, such 960 as in goldfish (305), bullfrog (855) and sea bass (503), the coexistence of two, three and even 961 five distinct GnRH-R subtypes was reported, respectively. Three GnRH-R were also 962 characterized in lamprey (330, 716). GnRH binding sites in GnRH-R type I and II forms are 963 highly conserved but several amino acid residues in the second and fourth transmembrane 964 domains confer selectivity toward GnRH1 and GnRH2 (122, 205).

965 Interestingly, at both the structural and functional levels, non-mammalian GnRH receptors 966 resemble more closely the mammalian type II receptors than the mammalian type I receptors. 967 Indeed, they all have a carboxy-terminal tail and a higher affinity (although sometimes 968 modest) for GnRH2 than for GnRH1 (337). Moreover, in contrast to GnRH-RI, they show an 969 Asp residue in transmembrane domain 2 that is conserved among members of the GPCR 970 family. This Asp residue, that is replaced by an Asn residue in the mammalian GnRH-RI, is 971 required for receptor activation (406, 488). As the mammalian GnRH-RII, non-mammalian GnRH receptors are expressed in numerous tissues, in particular in reproductive organs. In species possessing several GnRH-R subtypes, each receptor exhibits a distinct expression pattern (248). Both mammalian GnRH-RII and non-mammalian GnRH receptors generally couple to the Gq protein and therefore mediate the intracellular production of inositol phosphates (99, 337).

977

978 <u>4.3.3. Evolution of GnRH-R genes in vertebrates: impact of WGD</u>

979 The evolutionary relationship between vertebrate GnRH receptor is still obscur, making it 980 difficult to propose a consensual nomenclature (see (613) for a detailed review of the different 981 nomenclatures proposed over time). As mentioned above, vertebrate GnRH receptors were 982 generally subdivided into two major types (I and II) on the basis of both their structural 983 (presence or not of a C-terminal tail) and functional (selectivity to individual GnRH peptides) 984 features. Type I GnRH-R only contains the mammalian « tail-less » sequences, while type II 985 GnRH-R includes all remaining sequences, from both mammalian and non-mammalian 986 species. Such a categorization is well supported by numerous phylogenetic analyses in which 987 GnRH-RI and GnRH-RII sequences actually group into two separate major clades (302, 613, 988 643, 644, 690, 776, 873).

989 Until recently, GnRH-RI sequences were only found in mammals, suggesting that this 990 receptor evolved after lineage-specific gene duplication in early mammals (337, 643). This 991 concept was eventually invalidated when sequences of GnRH-RI were found in coelacanth, 992 elephant shark and little skate (644, 690, 873), indicating that the gnathostome ancestor 993 already possessed a type I GnRH-R. Thus, functionally, the absence of the cytoplasmic tail in 994 type GnRH-RI can no longer be viewed as an adaptation to mammalian physiology (873), as 995 previously suggested by some authors (142, 794).

996 In contrast to the type I clade, the type II clade is characterized by a wide heterogeneity, since 997 it includes sequences from species of very diverse origins. This heterogeneity is reinforced by 998 the fact that most of these species possess multiple type II receptor forms, e.g. three in 999 lamprey, some teleosts and amphibians, and up to four in coelacanth. Hence, recent studies 1000 subdivided type II clade into two major clades, called IIa and IIb (613, 644, 873). IIa and IIb 1001 can be further split into three and two groups, respectively. The first type IIa GnRH-R clade 1002 contains all three of the lamprey GnRH-R sequences plus one of the coelacanth paralogs. The 1003 second type contains the mammalian type II GnRH-R plus one of the coelacanth paralogs, 1004 while the third type contains GnRH-R sequences from teleosts, coelacanth, amphibians, 1005 reptiles and birds. The first type IIb clade also contains GnRH-R sequences from teleosts, 1006 coelacanth, amphibians, reptiles and birds, while the second type is exclusively composed of 1007 teleost sequences. It is noteworthy that the two IIb subtypes are not discriminated in the 1008 Williams's study.

At first sight, the logic of this breakdown is not obvious. Indeed, phylogenetic analyses, alone, are not perfectly suitable to explain the origin of the different GnRH receptor subtypes, and their evolutionary relationships. Recently, several studies have tried to revisit the history of the GnRH-R family in light of the 2R theory (360, 644, 690).

1013 Following up a previous work (360), Sedifeh et al. (2014) (690) examined the conserved 1014 synteny of chromosomes harboring GnRHR genes. They identified three pairs of GnRH-R 1015 subtypes in three paralogous linkage groups, suggesting that an ancestral pair emerged 1016 through local duplication before 2R. According to their model, 2R thus led to the generation 1017 of 6 subtypes of GnRH-R in the vertebrate ancestor. However, this model is based on a 1018 phylogenetic analysis whose conclusions have been recently strongly challenged (644, 873). 1019 For instance, Roch et al. (2014) proposed that all three major GnRH receptor subtypes (types 1020 I, IIa and IIb) were already present in the vertebrate ancestor before 2R (644). These three

1021 receptors would have been the products of two successive events of tandem duplication. 1022 According to their model, 2R theorically led to the generation of 12 GnRH-R subtypes 1023 (Figure 7). However, due to multiple gene losses, only six of them were preserved. 1024 Thereafter, further losses could occur in the different vertebrate lineages, eventually leading 1025 to the current situation. This model is in good agreement with phylogenetic data, which 1026 identify a total of six GnRH-R subtypes. However, on the basis of practically the same data, 1027 Williams et al. (2014) propose a totally different scenario, in which only three GnRH-R genes 1028 were present in the gnathostome ancestor (873). By constrast, they argue that the current data 1029 do not resolve the number of GnRH receptors in the common ancestor of vertebrates and the 1030 actual impact of 2R.

Taken together, this information shows that further studies, combining both phylogenetic and syntenic data, will be needed to fully understand the evolution of the GnRH-R family in vertebrates.

1034

1035 4.4. Origin of the GnRH system before the emergence of vertebrates

1036 In order to gain more insight into the origin of GnRH, intensive efforts have been made to1037 hunt out GnRH-related peptides in phyla other than vertebrates.

1038

1039 <u>4.4.1. GnRH and GnRH-R in non-vertebrate chordates</u>

The first molecular evidence that GnRH occurs outside vertebrates was provided by the characterization of multiple GnRH-like peptides in tunicates, encoded by one or two different genes, depending on the species (5, 264, 618, 644). In several species examined, such as *Ciona sp.* or *Haloclynthia sp.* (5, 264), pre-proGnRHs are multifunctional precursors, each containing the sequence of two to three tandemly arranged peptides. However, despite the unique organizaton of their precursor, almost all tunicate GnRH forms identified so far are decapeptides including the six invariant amino acids, in positions 1–4, 9 and 10, found in
vertebrates (644) (Table 1). It is noteworthy that the occurrence of both mGnRH and cGnRHI reported by Di Fiore et al. (2000) (193) in *Ciona* could not be confirmed (5).

1049 Search of the Ciona genome also led to the identification of four GnRH receptors (775). All 1050 these receptors possess a C-terminal tail similar to vertebrate type II GnRH-R, as well as 1051 several other structural features shared with vertebrate GnRH-Rs (643). At the functional 1052 level, only one tunicate receptor is able to activate the G_{q/11} pathway and to trigger IP3 1053 accumulation, while the three others provoke an increase of cyclic AMP formation through 1054 stimulation of G_s (775). At the evolutionary level, the four *Ciona* GnRH receptors cluster into 1055 a single clade at the root of the vertebrate GnRH-R one (644). Taken together, these data 1056 demonstrate unequivocally the existence in tunicates of a GnRH system homologous to that in 1057 vertebrates.

1058 The occurrence of a functional GnRH system was also reported in amphioxus. However, the 1059 first Blast searches of the amphioxus genome allowed the identification of only GnRH-like 1060 receptor but not peptide sequences (776). Of the four amphioxus GnRH-R sequences found, 1061 only two, GnRH-R1 and GnRH-R2, cluster as expected, at the root of the chordate GnRH-R 1062 tree (642, 644). In constrat, the two others, GnRH-R3 and GnRH-R4, appear much more 1063 distant and do not group with chordate GnRH-R sequences (see below for more details). 1064 Nevertheless, the four amphioxus GnRH-R genes are positioned in the same locus surrounded 1065 by syntenic homologs of the vertebrate GnRH-R paralogon (642).

In 2009, a peptide with GnRH sequence identical to mammalian GnRH1 was purified from amphioxus (96) but this result proved to be artifactual (642). The occurrence of a genuine amphioxus GnRH-like gene was reported only five years later by using a more sensitive search methodology with hidden Markov models (642). The GnRH identity of this gene was confirmed by synteny analysis. However, the GnRH-like peptide encoded by this gene 1071 exhibits a very atypical structure, pQILCARAFTYTHTWamide, since of its 14 amino acids 1072 (aa) (and not 10), 3 only are conserved at positions 1, 5, and 7 with vertebrate GnRH2 1073 residues (Table 1). It has been proposed that the amphioxus GnRH-like peptide may be 1074 cleaved to form a more active 7 aa-peptide (785). The amphioxus GnRH-like peptide acts as a 1075 ligand for amphioxus GnRH-R3 (642), but does not activate any of the three other amphioxus 1076 GnRH-R subtypes (642), indicating that the natural ligands of the latter receptors remain to be 1077 identified. It is to note that amphioxus GnRH-R1 and GnRHR-2 can be activated by 1078 vertebrate GnRH (642). Interestingly, a second GnRH-like peptide has been identified in the 1079 amphioxus genome (called GnRHv), but this peptide does not activate any of the four known 1080 amphioxus GnRH-R (642).

1081

1082 <u>4.4.2. GnRH-like peptides and GnRH-R in other non-vertebrates</u>

The first non-vertebrate GnRH-like molecule, at least clearly presented as such, was characterized in 2002 from a cephalopod mollusk, the common octopus (313). Octopus GnRH is a dodecapeptide that shares 7 residues with vertebrate GnRH2 from which it mainly differs by the insertion of two amino acids after position 1 (Table 1). The overall organization of the octopus GnRH precursor is the same as that of vertebrate pre-proGnRHs. Its cognate receptor was identified soon after and was demonstrated to have numerous similarities with chordate GnRH receptors (344).

1090 Closely related peptides were subsequently identified in several other species of mollusks (58,

1091 831, 912), annelids (832) and echinoderms (649, 695) (Table 1). All these newly identified 1092 peptides, together with the amphioxus GnRH-like peptide, were proposed to be classified as 1093 invertebrate GnRHs (invGnRHs), as opposed to vertebrate GnRHs (that also include the 1094 tunicate ones) (644). However, recent studies have shown that invGnRHs are probably not 1095 orthologous to GnRHs and should be renamed (see below) (613, 905). 1096

1097 4.4.3. Other GnRH-related peptides (AKH, CZR, ACP) and their receptors

1098 In 1998, the occurrence of a receptor structurally related to vertebrate GnRH receptors was 1099 reported in Drosophila (267). Several years later, the ligand for this Drosophila GnRH-type 1100 receptor was characterized and identified as a member of the adipokinetic hormone (AKH) 1101 family (745). Such a result was totally unexpected since AKH has been well known for a long 1102 time in insects (470, 748) but without apparent relationship with GnRH. However, taking a 1103 closer look, GnRHs and AKHs appear to share the N-terminal pyro-Glu residue and the C-1104 terminal amide plus various identical residues depending on the species, in particular a 1105 conserved Trp at the third last position. It thus appears reasonnable to consider peptides of the 1106 AKH family as possible GnRH homologs.

1107 AKH is an insect peptide first isolated from the locust *corpora cardiaca*, a functional 1108 equivalent of the pituitary gland in insects (835), for its ability to mobilize lipids from the fat 1109 body during flight (748). Many different forms (8-10 residues) of this peptide hormone were 1110 subsequently described in various insect species, with even two or three gene copies in some 1111 of them (216) (Table 1). AKH is also known as the red pigment-concentrating hormone 1112 (RCPH) isolated several years earlier in crustaceans (189) The first RCPH receptor has been 1113 recently characterized from Daphnia (456).

In 1989, an AKH-like peptide was identified in the cockroach and named corazonin (CRZ, ''corazon" being the Spanish word for 'heart') on the basis of its cardioacceleratory properties (830). Although slightly larger in size (11 résidues), CRZ share several structural features with AKH (Table 1). Furthermore, their preprohormones have a similar organization indicating that the two neuropeptide precursor genes must have a common evolutionary origin (829). Like AKH, CRZ is expressed in neurosecretory neurons of the protocerebrum and transported to the storage lobes of the *corpora cardiaca* (622). The CRZ receptor was initally characterized in *Drosophila* by a reverse pharmacology strategy (94, 582). Sequence analyses
showed that CRZ, AKH and GnRH receptors are all evolutionarily related.

1123 In 2006 Belmont et al. (2006) isolated from *Anopheles* a receptor with strong smilarities to

the AKH receptor but which is activated neither by AKH nor by CRZ (45). The natural ligand

1125 for this receptor was subsquently characterized as a new peptide called AKH/corazonin-

related peptide (ACP) due to its intermediate structural features with AKH and CRZ (261).

1127 In all species examined so far, AKH, CRZ and ACP originate from precursors that exhibit the

same overall organization as those of vertebrate GnRHs, with the mature peptide immediatly

1129 after the signal peptide. However, despite their structural similarities with other non-

1130 vertebrate GnRH-like receptors, the AKH receptors are specific for AKH and are not

1131 activated by ACP or CRZ. Similarly, the ACP receptors are specific for ACP and the CRZ

receptors are specific for corazonin, showing that AKH, ACP and CRZ are three independent signaling systems (411).

1134 For a long time, AKH, CRZ and ACP were thought to be confined to the arthopod clade, 1135 where they often (but not systematically) coexist in the same species (411). In 2009 the 1136 existence of an AKH-like peptide and its cognate receptor was reported in the nematode C. 1137 elegans (424), suggesting an older origin of the AKH signaling system. This notion was 1138 backed up soon later when it appeared that a functional AKH system is also present in 1139 mollusks (164, 265, 411), hemichordates (905) and echinoderms (785). Regarding the CRZ 1140 system, the same conclusion can be drawn from a recent study showing the presence of CRZ 1141 and its receptor in hemichordates and echinoderms (644, 785). In contrast, so far, ACP has

1143

1142

1144 <u>4.4.4. Evolutionary relationships between GnRH- and AKH-CRZ-ACP-like systems</u>

been identified only in arthopods (905).

45

1145 The view that AKH, ACP, CRZ and GnRH peptides form a superfamily is generally well 1146 accepted (265, 318, 423, 492, 613, 644, 905). However, the precise relationships between 1147 CRZ, AKH, ACP and GnRH are poorly understood as illustrated by a very confused 1148 nomenclature (265, 613, 644, 801, 905). One of the major issues is whether AKH and CRZ 1149 neuropeptides can be both considered as orthologs to the vertebrate GnRH system. 1150 Unfortunately, this issue is not really addressed by phylogenetic analyses. In contrast, it 1151 appears likely that the invGnRH group previoulsy defined by Roch (2014) (644) is not a 1152 monophyletic group (613, 801). It has thus been proposed to limit the name of GnRH 1153 exclusively to deuterostome GnRHs (613).

1154 Regarding the receptors, the situation is also unclear. Phylogenetic analyses generally agree to 1155 recognize three majors clades (265, 613, 644, 785). The first clade includes all chordate 1156 GnRH-Rs, except amphioxus GnRH-R3 and GnRH-R4, plus GnRH-R-like sequences from 1157 echinoderms, in other words most deuterostome GnRH-Rs. The second clade contains all 1158 arthropod CRZ-R sequences. It also encompasses amphioxus GnRH-R3 and GnRH-R4 plus 1159 some GnRH-R-like sequences from echinoderms and lophotrochozoans (annelids and 1160 mollusks). Surprisingly, among this second clade, are the octopus and Aplysia GnRH 1161 receptors, initially thought to belong to the GnRH-R group, as indicated by their name (344, 1162 348). Finally, the third clade encompasses all arthropod ACP-R plus AKH-R sequences from 1163 arthropods, nematodes and lophotrochozoans. Depending on the authors, different clusterings 1164 of these clades were proposed and formed the basis of two main evolutionary scenarios (654) 1165 (Figure 8). The first scenario states that an ancestral CRZ-R, which was conserved in the 1166 ambulacrarians (echinoderms and hemichordates) and the lophotrochozoans (annelids and 1167 mollusks), duplicated twice independently and thus generated two lineages: the first leading 1168 to CRZ-R and AKH-R, subsequently AKH-R generated artholopod ACP-R in ecdysozoans, 1169 and the second leading to GnRH-R in chordates (Figure 8A). The second scenario states that GnRH-R and CRZ-R might have been arisen via gene duplication in a common ancestor of bilaterians, and that a second gene duplication of GnRH-R might have generated AKH-R and ACP-R during the divergence between lophotrochozoans and ecdysozoans (arthropods and nematods) (Figure 8B). The main difference between the two models is that, in the first one, AKH-R/ACP-R and GnRH-R are not orthologous since they arose independently while, in the second one, they are orthologous. In contrast, both models agree that the CRZ-R gene was probably lost in urochordates, vertebrates and nematodes.

1177 The second scenario was particularly detailed by Zandawala et al. (2018) who thus proposed 1178 to totally reconsider the classical nomenclature of the GnRH- and AKH-CRZ-ACP-like 1179 systems (905). According to these authors, who distinguish two major types of GnRH-like 1180 receptors, *i.e.* the (true) GnRH-R type and the CRZ-R type, the term AKH-R should be left 1181 out and replaced by GnRH-R in all bilaterian species, since AKH-R and GnRH-R are 1182 orthologous. Nevertheless, they suggest keeping AKH-R specifically in arthropods, together 1183 with that of ACP-R, its co-paralog. They also propose to change all the nomenclature of the 1184 invertebrate GnRH-like peptides on the basis of the receptor type that they activate. For 1185 example, the octopus GnRH, as well as its counterparts in other lophotrochozoans, should be 1186 renamed as CRZ because their cognate receptor clearly belongs to the CRZ-R type (344). 1187 Likewise, all AKHs from lophotrochozoans and non-arthropodian ecdysozoans (e.g. 1188 nematodes) should be renamed as (true) GnRHs.

- 1189 As already mentioned, receptors of GnRH-R (GnRH-R1 and GnRH-R2) and CRZ-R (GnRH-
- 1190 R3 and GnRH-R4) types, have been characterized in amphioxus (776). In contrast, only one 1191 GnRH-like peptide was found in this group (642), that problably corresponds, for the reasons
- 1192 given above, to CZR. These data clearly suggest the existence of an authentic amphioxus
- 1193 GnRH that remains to be discovered.

1194 All in all, the Zandawala's scenario appears very attractive, since it is based on the hypothesis 1195 of co-evolution between peptides and receptors, a process generally observed for 1196 neuropeptidergic signaling systems (492). However, further studies are needed to confirm this 1197 model, in particular to characterize the ligand-binding properties of the receptors in a larger 1198 number of species. For example, such studies would help to clarify the phylogenetic status of 1199 the different peptides recently identified in prawn (756), particularly the so-called GnRH, 1200 whose place in the Zandawala's nomenclature remains to be determined. The same remark 1201 applies regarding the so-called AKH2 and GnRH2 found in the marine annelid *Platynereis* 1202 dumerilii (129).

Phylogenetic analyses show that receptors of the GnRH/CRZ superfamily have close evolutionary relationships with receptors of the vasopressin (VP)/ocytocin (OT) family (492, 643). Interestingly, although they are not structurally related, GnRH2, OT and VP genes are tandemly arranged in several vertebral species (252, 360, 643, 790), also suggesting a possible common origin.

1208

1209 <u>4.4.5. GnRH and GnRH-R before bilateria?</u>

The nervous system in cnidarians is well known for its peptide wealth (245). As a matter of fact, a large number of neuropeptides have been isolated from different groups of cnidarians (211). Several reports show that coral tissues contain GnRH-like compounds (18, 812) but they have not been characterized yet. The occurrence of GnRH-like peptides has also been reported in the sea anemone genome (19). However, recent investigations failed to identify any GnRH-R-like sequence in cnidarian genomes (265, 613), suggesting that the GnRH-CRZ signaling system is a bilaterian innovation.

1217 It is noteworthy that an earlier origin of the GnRH system was formerly proposed after the 1218 finding that yeast α -mating factor shows some structural and functional similarities to 1219 vertebrate GnRHs (435). However, evolutionary relationships between both peptides are 1220 unlikely since α -mating factor is known to activate a receptor that does not belong to 1221 rhodopsin β -like GPCRs (329).

1222

1223 4.5. Functions of GnRH-like peptides: evolutionary perspectives

1224 <u>4.5.1. Reproductive functions of GnRH in chordates</u>

1225 In vertebrates, GnRH1 is the main GnRH form targeted to the pituitary. Its stimulatory effect 1226 on gonadotropin release is well established in many species. GnRH1 is thus considered as the 1227 major regulator of the hypothalamo-pituitary-gonadal axis whose stimulation leads to 1228 gametogenesis and steroid production in the gonads (556, 643, 921). Consistent with this 1229 notion, studies conducted in mice and humans show that GnRH1 null mutations result in 1230 hypogonadotropic hypogonadism and infertility (93, 97). In teleosts species lacking GnRH1, 1231 such as zebrafish, GnRH3 is believed to assume the hypophysiotropic role of GnRH1 (3, 1232 744).

1233 The action of GnRH1 is thus closely linked to the presence of a pituitary, which is a 1234 vertebrate innovation (643, 740). However, GnRH1 is also produced in and acting on other 1235 reproductive tissues. This is particularly the case in the gonads where, in numerous species of 1236 fish, amphibians, birds, reptiles and mammals, GnRH1 acts as a paracrine factor involved in 1237 the control of both gametogenesis and steroidogenesis (475, 477). This is also the case in the 1238 placenta (therefore specifically in mammals) where GnRH1 promotes trophoblastic cell 1239 invasion and regulates chorionic gonadotropin biosynthesis (670). In addition to its 1240 physiological actions, GnRH1 also regulates reproductive behaviors, including female 1241 lordosis (656) and male mating behavior (638) in rats.

As mentioned above, GnRH2 is primarily expressed in the vertebrate midbrain and is generally thought to have neuromodulatory actions (337, 556). Thus, intracerebroventricular administration of GnRH2 promotes female reproductive behavior in various mammalian

1245 species (556). Likewise, GnRH2 stimulates female spawning behavior in goldfish (843).

1246 However, in some teleost and amphibian species, GnRH2 fibers originating from the

1247 midbrain were found to project, directly or indirectly, to the pituitary (125, 611, 921). As a

1248 matter of fact, GnRH2, as GnRH1, stimulates gonadotropin synthesis and secretion by

1249 pituitary gonadotrope cells (148, 489, 554).

1250 GnRH3 is implicated in the control of reproductive behavior in teleosts. For instance, in

1251 cichlid fish, GnRH3-immunoneutralization decreases nest-building ability of males (542).

Similarly, in medaka, terminal nerve GnRH3 neurons have an essential role for activatingmating preferences based on familiarity (559).

In tunicates, GnRH induces the release of gametes, probably *via* a direct action on the gonads (779), suggesting that GnRH already exerted reproductive functions in the olfactores ancestor (the common ancestor of urochordates plus vertebrates), before the appearance of the pituitary.

1258

1259 <u>4.5.2. Non-reproductive functions of GnRH in chordates</u>

1260 Both, GnRHs, and their receptors are expressed in numerous non-reproductive tissues in the 1261 central nervous system (cerebral cortex, hippocampus, cerebellum, ...) and in peripheral 1262 organs (bladder, adrenal, heart, ...) (722), suggesting that GnRH may have a variety of non-1263 reproductive activities. In fact, GnRH1 is released in significant concentrations into the 1264 cerebrospinal fluid (CSF) to potentially act outside the hypothalamus (86, 721). GnRH1 can 1265 affect pituitary cells other than gonadotropes (722). In particular, GnRH1 stimulates GH 1266 release in goldfish (455) and chicken (677). A role of GnRH2 and GnRH3 on early brain 1267 regionalization and eye development has also been revealed by knockdown experiments in 1268 zebrafish (886). At the behavioral level, GnRH1 is involved in the control of anxiety and depression was reported both in human and rodents (579), while GnRH2 influences feedingbehavior (461, 777).

1271 Interestingly, a role of GnRH in the control of metamorphosis has been recently observed in 1272 tunicates (342), suggesting that, even in the olfactores ancestor, GnRH is not just a 1273 reproductive hormone.

1274

1275 <u>4.5.3. Roles of GnRH-like peptides in non-chordates</u>

1276 The first two GnRH-like peptides to be characterized in non-chordates were identified from 1277 octopus (313) and C. elegans (424). According to the new Zandawala's nomenclature (see 1278 above, 2018), these peptides correspond to CRZ-type and true GnRH-type forms, 1279 respectively. At the time of their discovery, both peptides were logically investigated for their 1280 reproductive functions. As a matter of fact, the octopus peptide was shown to stimulate 1281 oviduct contraction and gonadal steroidogenesis (312, 344), while the nematode peptide was 1282 described as a regulator of egg-laying behaviour (424). These findings were thus taken as a 1283 strong support to consider both peptides as GnRH homologs (in addition to the fact that they 1284 bind to GnRH-like receptors) (423). However, further studies showed that the reproductive 1285 functions of GnRH-like peptides are often marginal in non-chordate species.

1286 As mentioned above, AKH- and CRZ-type signalling systems were first characterized in 1287 arthropods. Therefore, most of the functional data currently available are from this group, 1288 especially from Drosophila (905). In insects, AKH mainly regulates carbohydrate 1289 homeostasis, and lipid and carbohydrate mobilization, particularly during flight (216). AKH 1290 is also involved in other physiological processes related to locomotion and stress (265), but is 1291 generally not linked to reproduction (905). Only a few studies in the cricket (434) and the 1292 locust (509) indicate that AKH may have a role in reproduction. In crustaceans, RPCH, the 1293 AKH ortholog, modulates gut motility in the lobster (151) and ovarian maturation in the 1294 crayfish, in addition to its role in background color adaption initially reproted in the prawn 1295 (189). The functions of AKH-like peptides in other taxa are still poorly characterized. AKH 1296 has been recently identified in Aplysia (324) and functional assays indicate that it may 1297 modulate metabolism, feeding and gut motility, and also affect some gonadal parameters. 1298 Although this peptide is considered as a true GnRH according to the Zandawala's 1299 nomenclature (2018) (905). While the functions of ACP, the closest paralog of AKH in 1300 arthropods, are still unclear, in a recent study, ACP was shown to play a role in lipid 1301 metabolism and inhibition of oocyte proliferation in prawn (756).

1302 As already indicated, CRZ was originally isolated from cockroach on the basis its 1303 cardiotropic properties (830). However, such an activity has been so far only documented in a 1304 few species. Since then, CRZ has been implicated in a wide range of functions in insects, 1305 including the initiation of ecdysis behaviors in the moth (362), induction of cuticular 1306 pigmentation in the locust (771), reduction of the spinning rate and pupal development in the 1307 silkworm (767), testicular development and spermatogenesis in the prawn (711). In ants, CRZ 1308 controls social behavior and caste identity (236). As for GnRH, the functions of CRZ in other 1309 taxa are still poorly known. The reproductive functions of CRZ in octopus have been 1310 mentioned above, but octopus CRZ (according to the Zandawala's nomenclature) has also 1311 non-reproductive functions, in particular on the heart and muscles (312, 344). In Aplysia, 1312 CRZ affects many processes including motor function, feeding and neuronal activities but has 1313 no significant effect on reproduction (753, 802).

Overall, GnRH-type and CRZ-type peptides are pleiotropic hormones that have both overlapping and specific properties, but do not exhibit clear patterns of functions emerges, thus making difficult to infer their ancestral functions in Urbilateria, the last common ancestor of bilaterians. The only (quasi) certainty is that the critical role of GnRH as a reproductive activator is a characteristic unique to vertebrates (654), likely related to the innovation of thepituitary gland in this group.

1320

1321 5. Origin and evolution of brain RF-amide peptides in the limelight for reproduction

Two types of brain peptides, namely kisspeptin and GnIH/RFRP, have been subjected to most attention in the field of reproductive physiology since the early 2000s. Both belong to the superfamily of RF-amide peptides and have been proposed as new major actors in the neuroendocrine control of reproduction in vertebrates.

1326

1327 <u>5.1. History and evolutionary relationships of the RF-amide peptides</u>

1328 <u>5.1.1 Discovery of RF-amide peptides in mollusks</u>

1329 In 1972, a substance, designated peak C, was isolated in all major classes of the phylum 1330 Mollusca for its cardioexcitatory activity (9). Five years later, peak C was purified from 1331 cerebral ganglion extracts of the sunray venus clam, Macrocallista nimbosa (623). The 1332 neuropeptide identified, FMRFamide, was named after the letter abbreviations of its four 1333 amino acids, phenylalanine, methionine, arginine and phenylalanine, and its amidated C-1334 terminal sequence (623). Since then, FMRFamide along with its structurally related 1335 neuropeptides named FMRFamide-related peptides (FaRPs) or FMRFamide-like peptides 1336 (FLPs) remains the best-investigated family of neuropeptides in mollusks.

1337

1338 <u>5.1.2. RF-amide peptides superfamily in vertebrates</u>

1339 The first vertebrate member of the RF-amide family (peptides ending with Arg-Phe-NH₂) was

1340 isolated and sequenced (Leu-Pro-Leu-Arg-Phe-NH₂: LPLRFamide) from chicken brain in

1341 1983 (152). This peptide was subsequently recognized as a fragment of a longer RF-amide

1342 peptide, gonadotropin-inhibitory hormone (GnIH) (for review: (806). Since 1983, a number

of neuropeptides sharing the typical C-terminal RF-amide (RFa) signature were discovered inall major animal phyla (174, 318, 904).

1345 Dockray (2004) suggested designating the mammalian RF-amide genes, farp-1 to farp-5 in 1346 the chronological order they were described (153): the *farp-1* gene (601, 841) encodes two 1347 peptides, neuropeptide FF (NPFF) and neuropeptide AF (NPAF) (896); the farp-2 gene (280) 1348 encodes prolactin-releasing peptide (PrRP) (280); the farp-3 gene (281, 426) encodes two 1349 peptides, RFRP-1 and RFRP-3 also referred to as neuropeptide VF (NPVF) or GnIH (281); 1350 the *farp-4* gene (227) corresponds to the Kiss-1 gene encoding kisspeptin (or metastin) 1351 peptides (375, 549); the *farp-5* gene (104, 213, 322) encodes 26RFa, also named P518 or 1352 QRFP (104, 213, 322). These five distinct genes thus encode seven peptides in mammals 1353 (153, 630). The *farp* nomenclature based on chronology of discovery has not been followed 1354 by other authors (630). However, a similar classification into five groups has been adopted 1355 (for review: (826)) and further validated by phylogenetical analyses performed on sequences 1356 from various vertebrates including mammals, other tetrapods, teleosts and lamprey (570). 1357 These groups of RF-amide peptides were named by the authors: PQRFa (NPFF) group 1358 corresponding to *farp-1*; PrRP group corresponding to *farp-2*; LPXRFa (GnIH) group 1359 corresponding to *farp-3*; metastin (kisspeptin) group corresponding to *farp-4*; and QRFP 1360 (26RFa) group corresponding to *farp-5* (for review: (805)).

All the RF-amide peptides bind to G protein-coupled receptors (GPCRs). Dockray suggested, as for *farps*, to designate the receptors to the RF-amide peptides in the order they were identified as putative RFamide receptors: accordingly, *rfr-1* for PrRP (*farp-2*) receptor (280); *rfrp-2* for RFRP-1 and RFRP-3/NPVF/GnIH (*farp-3*); *rfr-3* for NPFF and NPAF (*farp-1*) receptors (68, 176, 281); *rfr-4* for metastin/kisspeptin (*farp-4*) receptor (375, 513, 549); *rfr-5* for QRFP/26RF (*farp-5*) receptor (213, 322). Using phylogenetical analyses of more than 800 human GPCR sequences, Fredriksson (2003) showed five main families of GPCRs in human, 1368 which they named glutamate, rhodopsin, adhesion, frizzled/taste2 and secretin (208). The 1369 rhodopsin family was divided into four groups: α , β , γ and δ (208). Kisspeptin receptor (also 1370 named GPR54 or *rfr-4*) belongs to the γ group, more specifically to the SOG receptor cluster, 1371 together with the receptors for non-RFamide peptides, somatostatin (S), opioid (O) and 1372 galanin (G) (208). Receptors of NPFF (*rfr-3*), GnIH/RFRP (*rfr-2*), PrRP (*rfr-1*) and QRFP 1373 (*rfr-5*) cluster within the β group, with receptors for non-RFamide peptides, neuropeptide Y 1374 (NPY), cholecystokinin, GnRH, tachykinin, ghrelin, and neuromedin U (208).

1375 Several studies, based on synteny analysis and phylogenetic relationships of RF-amide 1376 peptides and receptors, provide new insights into the RF-amide peptide superfamily (for 1377 review: (904)). A gene encoding a PQRFa peptide has been identified in a cephalochordate, 1378 the amphioxus, indicating the presence of a *pqrf* gene at least in a chordate ancestor (565). 1379 This ancestral gene likely gave rise to gnih (farp-3) and its sister gene npff (farp-1), via one of 1380 the two rounds of whole genome duplications (1R/2R) that occurred early in the vertebrate 1381 lineage ((565); for reviews: (807, 904)). The authors also reported phylogenetical 1382 relationships between NPFF, GnIH and QRFP (farp-5), suggesting a close evolutionary 1383 relationship between these three groups of RF-amide peptides (904). Kisspeptin (farp-4) has 1384 been proposed to be evolutionary more related to non RF-amide peptides, galanin and spexin, 1385 than to other RF-amide peptides, and proposed to be classified as a member of the 1386 galanin/spexin family ((359); for review: (904)). PrRP receptors likely share a common 1387 ancestry with NPY receptors as reported by Lagerström and collaborators (385), linking the 1388 PrRP group (farp-2) more to a non-RF-amide peptide family than to the other RF-amide 1389 peptide groups. The grouping of the various RF-amide peptide subfamilies/groups in a same 1390 RF-amide superfamily is thus currently a matter of debate (Figure 9).

1391

1392 **5.2.** GnIH (*farp-3*) and evolutionary-related peptides (NPFF and QRFP) (*farp-1 and -5*)

1393 5.2.1. Discovery of GnIH in birds: direct inhibitory role on pituitary gonadotropins

In 2000, Tsutsui and collaborators reported the isolation and characterization, from quail brain, of a novel neuropeptide, having an RFa motif at the C-terminus, which dosedependently inhibits the gonadotropin, luteinizing-hormone (LH), release *in vitro* from quail anterior pituitaries. They named this peptide gonadotropin-inhibitory hormone (GnIH) (808).

- 1398 In the quail, starling and duck, the GnIH precursor mRNA encodes a polypeptide that can
- 1399 potentially be cleaved into three mature peptides with a C-terminal LPXRF-amide sequence
- 1400 (X=L or Q), named GnIH-related peptide-1 (GnIH-RP1), GnIH and GnIH-RP2 (quail: (671);

1401 starling: (814); duck: (207); for review: (378)). GnIH was identified as a mature endogenous

- 1402 peptide by mass spectrometry analyses, in the brain of quail (808), starling *Sturnus vulgaris*
- 1403 (814), zebra finch *Taeniopygia guttata* (788) and chicken (472), while GnIH-RP2 was also
- 1404 indentified in the brain of quail (671).
- 1405 In 2005, Tsutsui and colleagues identified a GnIH receptor (898), using primers designed 1406 from the sequence of GPR147, a novel GPCR, also called NPFF receptor 1 (NPFFR-1) since
- 1407 its discovery together with another NPFF receptor (NPFFR-2 or GPR74) in mammals (68).
- As GnIH has higher affinity for NPFFR-1, the latter is considered to be the primary receptor
- 1409 for GnIH (68, 426).
- The direct inhibitory action of quail GnIH on LH release was confirmed in another bird, the chicken. An inhibitory effect of GnIH on LH release by pituitary slices was reported in the sexually immature female White Leghorn while it was not observed *in vitro* in sexually mature birds and nor on FSH release at both stages (446). In a hybrid of White Leghorn and Silkie Breed, GnIH inhibits LH release (but not mRNA expression), as well as FSH release and mRNA in pituitary explants (117).
- 1416 The direct effect of GnIH at the pituitary level is also supported by the detection of gnihr
- 1417 mRNA in the pituitary of various bird species (quail: (898); starling and quail: (51)) and by

1418 the colocalization of GnIH receptor immunoreactivity with LH β or FSH β mRNA expression 1419 in the chicken pituitary gland (446).

1420

1421 <u>5.2.2. GnIH inhibitory role at the brain and gonadal levels in birds</u>

GnIH may also directly influence GnRH secretion as GnIH-ir fibers are observed in contact with GnRH-ir neurons in many bird species (sparrow: (50); for review: (49); European starling: (814); zebrafinch: (788)). Moreover, GnIH receptor mRNA is expressed in different parts of the brain in quail and starling as shown by PCR (51), and more specifically in both GnRH-I and GnRH-II neurons in European starling *Sturnus vulgaris* as shown by ISH/ICC (814).

Injection of GnIH rapidly reduces plasma LH and attenuates GnRH-induced rise in plasma
LH in sparrow (49, 569). In quail, continuous administration of GnIH for 2 weeks decreases
LHβ mRNA levels (818). In the Pekin duck, icv injection of GnIH reduces plasma LH
concentrations, while administration of GnIH-RP1 has no effect (207). One study reported no
effect of GnIH on LH plasma levels in a bird species: in the Rufous-winged sparrow, GnIH
does not affect basal or GnRH-stimulated plasma LH levels (149).
PCR analyses reveal the presence of GnIH as well as GnIH-R mRNAs in the testis, ovary and

1434 PCK analyses reveal the presence of Ghiff as well as Ghiff-K liktivAs in the testis, ovary and 1435 accessory reproductive organs (oviduct, epididymis and *vas deferens*) in starling and quail 1436 (51). Similarly, transcripts for GnIH and GnIH-R are detected in the testis of house sparrow 1437 *Passer domesticus* ((476); for review: (475)). In this latter species, GnIH decreases 1438 testosterone secretion from cultured testis (476). This indicates possible local, 1439 autocrine/paracrine actions of GnIH in the gonads in birds.

1440

1441 5.2.3. GnIH in mammals

1442 The discovery of GnIH in birds has prompted investigations in mammals. The GnIH 1443 precursor cDNAs identified from mammalian brain encode two GnIH peptides, also known as 1444 RFamide-related peptides (RFRP): RFRP-1 and -3 (for reviews, see (804-806); rat: (281, 1445 822); mouse : (281); Siberian hamster (813); bovine (212, 281, 900); monkey (815); human 1446 (281, 816)). Like avian GnIH and GnIH-RPs, all these peptides possess a common C-terminal 1447 LPXRFamide (X=L or Q) motif, with RFRP-1 being a C-terminal LPLRFamide peptide and 1448 RFRP-3 a LPQRFamide peptide (for review: (807)). In human and bovine (not in rat and 1449 mouse), a third peptide, RFRP-2, is also predicted, but is a LPLRSamide peptide or a 1450 LPLRLamide peptide, respectively (281). This RFRP-2 peptide could not be detected in an 1451 extract of human hypothalamus by using an avian GnIH antibody-conjugated immunoaffinity 1452 column and mass spectrometry, possibly due to its different C-terminal motif or because it is 1453 not processed from the precursor (816). The rat GPR147 does not respond to hRFRP-2, 1454 showing the importance of the C-terminal RFamide for the binding to the receptor (281, 816). 1455 The RFRP-2 sequence is absent from the RFRP precursor in the Siberian hamster (813). Li 1456 and collaborators suggested that the pig GnIH precursor encodes three RFRP peptides, two 1457 with an LPLRFamide C-terminal sequence and one with an LPQRFamide C-terminal motif 1458 (416). The authors reported also similar characteristics in the rabbit and golden hamster GnIH 1459 precursors (416).

In all mammalian species, as in birds, RFRP fiber terminals form close appositions with GnRH neurons (hamsters (377); rats (325, 377); mice (377, 885); sheep (726); monkeys (815); humans (816)). In the mouse, direct application of RFRP-3 on cultured brain slices decreases the firing rate of a subpopulation of GnRH neurons, and this effect is maintained when amino acid transmission is blocked, suggesting a direct suppressive action of RFRP-3 on the excitability of GnRH neurons (165).

1466 In addition to its effects on GnRH neurons, GnIH may also act directly at the pituitary level

1467 via its release in the portal blood in mammals as in birds. GPR147 (also referred to as 1468 NPFFR-1), is expressed in the pituitary in rat (281), Syrian hamster (230) and human (816), 1469 and RFRP-ir fibers are observed in the external layer of the ME in hamsters (230), sheep 1470 (118) and humans (815, 816). However, other studies using a different antiserum failed to 1471 detect RFRP-ir in the ME of rat brain and ip injection of the retrograde tracer Fluorogold did 1472 not label RFRP cell bodies (639). RFRP-3 inhibits GnRH-stimulated gonadotropin synthesis 1473 in pituitary gonadotropes in sheep in vitro (664), supporting a potential action at the pituitary 1474 level. In Artiodactylae, RFRP-3 inhibits GnRH-induced, but not basal, LH and FSH secretion 1475 and expression by primary cultures of pituitary cells (ewe: (118, 664); bovine: (332); pig: 1476 (415)). In rat, Pineda et al (2010) observed an inhibitory effect of RFRP-3 on both basal and 1477 GnRH-stimulated LH secretion (610), while other studies reported no effect (20) or an effect 1478 only on GnRH-stimulated LH release (517). In the Syrian hamster, Ancel et al. (2012) found 1479 no effect on either basal or stimulated LH release (17).

1480 The presence of GnIH and its receptor in the mammalian gonads (Syrian hamster testis (914); 1481 human ovary (550); pig ovary and testis (416, 916)) suggests, like in birds, possible 1482 autocrine/paracrine regulations via a local gonadal GnIH system. Indeed, in the mouse, in vivo 1483 and in vitro studies show that GnIH/RFRP-3 treatment inhibits follicular development and 1484 ovarian steroidogenesis in the female (719), as well as testicular steroidogenesis and 1485 spermatogenesis in the male (22). Similarly, GnIH/RFRP-3 inhibits the synthesis and release 1486 of estradiol by ovarian granulosa cells (415) and of testosterone by Leydig cells (916) in vitro 1487 in pigs. Interestingly, a recent study investigating the role of RFRP-1 on ovarian activities 1488 reported a stimulatory effect of RFRP-1 on ovarian steroidogenesis at proestrus in the mouse, 1489 but an inhibitory action on ovarian folliculogenesis (141).

1490 Using a mouse line with genetic inactivation of NPFFR-1, it has been demonstrated that the

1491 NPFFR-1 pathway is dispensable for preserved puberty and fertility ((401); for review:

(402)). To the best of our knowledge, there is no report on the possible impact of *gnih* (*npvf*)gene inactivation in mice.

1494

1495 <u>5.2.4. Current studies on the presence and role of GnIH in the reproduction in other</u>
1496 vertebrates

1497 <u>a. In other sauropsids</u>

1498 In sauropsids, apart from birds, three GnIH peptides (GnIH, GnIH-RP-1 and GnRH-RP-2) 1499 have been characterized in the red-eared slider turtle Trachemys scripta elegans (821), in the 1500 Chinese softshell turtle and in the anole lizard (for reviews: (566, 807)), and they all possess 1501 the common C-terminal LPXRFamide motif, like the avian and mammalian peptides. 1502 However, few studies have investigated their potential physiological roles. Brain localization 1503 of GnIH by ISH and/or immunohistochemistry has been reported in the Japanese grass lizard 1504 Takidromus tachydromoides (350) and the red-eared slider turtle (821). In the lizard Calotes 1505 versicolor, ovarian localization of GnIH has also been reported and GnIH is able to suppress 1506 ovarian GnRH receptor expression (718).

1507 <u>b- In amphibians: a GH-releasing factor (for review: (315))</u>

1508 In the bullfrog Rana catesbeiana, a RF-amide peptide, isolated from hypothalami, showed 1509 high (75%) sequence identity with quail GnIH (372). It was designated fGRP for "frog GH-1510 releasing peptide" as it exhibits growth hormone-releasing activity (372). In parallel, the same 1511 peptide was isolated from the brain of the European green frog *Rana esculenta*, and named R-1512 RFa for "Rana-RFamide" (104). The cloning of a cDNA for fGRP revealed that the deduced 1513 precursor encodes one fGRP and three putative fGRP-related peptides (fGRP-RP1, -RP2 and 1514 -RP3) (672), These four peptides were all identified as mature peptides and found to have a 1515 common C-terminal LPXRFamide with avian and mammalian GnIHs (824). Two of these 1516 peptides (fGRP and fGRP-RP2) stimulate GH and/or PRL release in vivo and in vitro, while 1517 the other two (fGRP-RP1 and fGRP-RP3) have no effects ((372, 824); for review: (823)). 1518 None of them is able to modify the release of gonadotropins (372, 824). The localization in 1519 the amphibian brain is in agreement with other vertebrate distribution as positive neurons (cell 1520 bodies) are present in the periventricular region of the hypothalamus (372, 612, 673). In 1521 contrast, nerve fibers are widely distributed throughout the brain (612) notably in the 1522 olfactory system ((194, 612). An urodele, the Japanese red-bellied newt Cynops pyrrhogaster 1523 possesses four GnIH peptides (nLPXRFa-1, nLPXRFa-2, nLPXRFa-3 and nLPXRFa-4) but 1524 no functional studies have been performed yet (114). The GnIH system and its functional 1525 significance in amphibians has been recently reviewed (315).

1526 <u>c- In basal sarcopterygian and actinopterygian</u>

1527 In a basal sarcopterygian, the coelacanth Latimeria chalumnae, a GnIH precursor gene has

1528 been identified, encoding three putative LPXRFamide peptides and one LPXRFamide-like

1529 peptide, which align with human RFRP-1/quail GnIH-RP1, human RFRP-2/quail GnIH,

1530 human RFRP-3, and quail GnIH-RP2, respectively (for review: (516)).

1531 In a basal, non-teleost actinopterygian, the spotted gar Lepisosteus oculatus, a GnIH precursor

1532 encodes three putative LPXRFamide peptides, which align with human RFRP-1/quail GnIH-

1533 RP1, human RFRP-2/quail GnIH, and human RFRP-3, respectively (for review: (516)).

1534 <u>d - In teleosts: positive and negative effects on gonadotropins</u>

1535 Up to now, in teleosts, only a single gene for GnIH has been identified as in the spotted gar.

1536 However, owing to the teleost specific WGD (3R), two GnIH genes would have been

1537 expected. This suggests the loss of one of the 3R-GnIH duplicated paralogs shortly after the

1538 3R event.

1539 Most of the teleost GnIH precursors encode three LPXRFamide or LPXRFamide-like

1540 peptides which align with human RFRP-1/quail GnIH-RP1, human RFRP-2/quail GnIH, and

1541 human RFRP-3, respectively (for reviews: (516, 807)). In some species, the precursor

encodes only two peptides which align with human RFRP-1/quail GnIH-RP1, and human
RFRP-2/quail GnIH, respectively. Most GnIHs identified in teleosts possess the common Cterminal LPXRFamide motif observed in other vertebrate GnIHs.

1545 Concerning the physiological role of GnIH in the control of the gonadotropic axis, the 1546 situation appears more complex in teleosts than in other vertebrates. In the goldfish, GnIH 1547 inhibits GnRH-stimulated $lh\beta$ and $fsh\beta$ expression by primary cultures of pituitary cells, with 1548 no effect on basal gonadotropin expression (625), while other studies describe either 1549 stimulatory, inhibitory or no effect on both basal and stimulated LH (release and expression), 1550 depending on the gonadal stage (510, 511). A stimulatory effect of goldfish LPXRa was 1551 observed on LH and FSH release by salmon pituitary cells (14). Similarly, goldfish LPXRa 1552 stimulates *lh* and *fsh* expression by whole pituitaries of grass puffer (700). In tilapia, 1553 treatment of pituitary cells in primary culture with tilapia LPXRFa-2 induces a significant 1554 increase in both FSH and LH release (60). In Cichlasoma dimerus, homologous LPQRFa-1 1555 decreases LH and FSH release, while LPQRFa-2 is able to stimulate FSH release (whole 1556 pituitaries: (899)). In a basal teleost, the eel Anguilla anguilla, ongoing studies indicate that 1557 the three peptides encoded by the eel gnih gene exert an inhibitory effect on $lh\beta$ mRNA 1558 expression by eel pituitary cells.

1559 <u>e- In cyclostomes (agnathans)</u>

1560 Tsutsui and colleagues in collaboration with Sower's laboratory (563) have cloned the 1561 lamprey GnIH precursor cDNA that encodes a precursor encompassing three GnIH peptides 1562 (lamprey LPXRFa-1a, LPXRFa-1b and lamprey LPXRFa-2). The lamprey GnIH peptides 1563 have a C-terminal PQRFamide motif, unlike most GnIHs, but like NPFF, in gnathostomes. *In* 1564 *vivo* administration of one of the lamprey GnIHs (lamprey LPXRFa-2) stimulates the 1565 expression of lamprey GnRH (GnRH3) in the hypothalamus and gonadotropin β mRNA in 1566 the pituitary (563). Thus, the authors suggest that GnIH may have emerged in agnathans as a stimulatory neuropeptide that subsequently diverged to an inhibitory neuropeptide during the
course of evolution from basal vertebrates to later-evolved vertebrates, such as birds and
mammals (563).

1570

1571 <u>5.2.5. Origin and evolution of GnIH and its receptors in vertebrates</u>

1572 Osugi and collaborators showed that agnathans possess both GnIH and NPFF genes, with the 1573 common C-terminal PQRFamide motif (563, 570).

1574 Tsutsui and colleagues hypothesized that vertebrate GnIH and NPFF genes may derive from a 1575 common ancestral gene present in protochordates (for review: (807)). In protochordates, a 1576 precursor cDNA of amphioxus PQRFamide peptides was then identified. This precursor 1577 encodes three mature peptides characterized by immunoaffinity purification and mass 1578 spectrometry (565). The conserved syntenic region around the loci of the amphioxus 1579 PQRFamide peptide gene, as well as of the GnIH gene and NPFF gene in vertebrates (565), 1580 suggests that GnIH and NPFF paralogous genes may originate from WGD in early vertebrates 1581 (for review: (567)) (Figure 9). As amphioxus PQRFamide peptides inhibit forskolin-induced 1582 CRE-luciferase activity in COS-7 cells expressing the amphioxus PQRFa receptor 1 (565), 1583 the authors suggested that the common ancestral peptide of GnIH and NPFF has played an 1584 inhibitory role in reproduction in ancestral chordates (807). The inhibitory role of the 1585 ancestral peptide would have been conserved for GnIHs in mammals, birds and some teleosts, 1586 whereas a change from inhibitory to stimulatory role would have occurred in agnathans and 1587 some teleosts (807). In fact, the GH- and PRL-releasing activity of GnIH in amphibians with 1588 the lack of effect on gonadotropin secretion suggests a larger flexibility in the possible 1589 physiological roles of GnIH peptides in the course of vertebrate evolution.

1590 Concerning GnIH receptors, two GPCRs have been identified in birds and mammals: 1591 (LPXRF-R or NPFFR-1 or GnIHR or OT7T022) considered as the primary GnIH receptor and GPR74 (PQRF-R or NPFFR-2) with a lower affinity for GnIH (for review: (543)).

1593 In teleosts, up to three different GnIH receptor genes (gnihr1, gnihr2 and gnihr3), that share high sequence identity with the mammalian and avian GPR147, have been isolated in 1594 1595 zebrafish (913), goldfish (625) and common carp (599). This may reflect the impact of teleost 1596 specific WGD (3R) or of the additional WGD that occurred in some cyprinids (4R in goldfish 1597 and some carps). However, in most teleosts, only one receptor has been identified so far 1598 (grass puffer LPXRF-R (700); tilapia LPXRF-R (60); orange-spotted grouper GnIHR (858); 1599 clownfish GnIHR (113); tongue sole LPXRFa-R (853)). Further investigations are thus 1600 clearly needed.

Elphick and Mirabeau have identified a GnIH/NPFF-type receptor gene in the genome database of the hemichordate *Saccoglossus kowalevski* (174). Phylogenetic analyses grouped the receptors of vertebrates into NPFFR-1 (GPR147) and NPFFR-2 (GPR74), and the receptors of insects into SIFR (817). Deuterostomian NPFFR genes have already been proposed to share a common evolutionary origin with protostomian SIFR genes (492).

1606

1607 <u>5.2.6. NPFF (PQRF) group (*farp-1*)</u>

1608 The first NPFF (PQRF) peptide to be reported was isolated from the bovine brain and 1609 characterized as a pain modulatory peptide (896). NPFF peptides possess a similar C-terminal 1610 structure (PQRFamide motif) as some GnIH peptides (for review: (577)). As previously 1611 discussed, *npff* and *gnih* genes are paralogous genes that likely emerged from WGD in early 1612 vertebrates. As GnIH precursors, the PQRFa precursor encompasses two to three putative 1613 mature peptides: NPFF and NPAF in mammals, NPFF and NPAF in teleosts, and PQRFa, 1614 PQRFa-RP1 and PQRFa-RP-2 in agnathans (lamprey and hagfish) (for review: (577)). 1615 NPFF peptides have been largely investigated for their role in the control of nociception (for

1616 review: (29)). Other actions have been also reported such as increasing arterial blood pressure

1617 and water intake, while decreasing food intake in mammals (for review: (577)). Compared to 1618 GnIH or kisspeptin, few studies have been carried out to investigate the possible role of NPFF 1619 in reproduction. In teleosts, one study reported that bath application of medaka NPFF inhibits 1620 the pacemaker activity of terminal nerve GnRH neurons in the dwarf gourami (653). In 1621 agnathans, one of the hagfish NPFF peptides directly stimulates pituitary expression of 1622 gonadotropin β mRNA (568), while the lamprey peptides appear to possess a stimulatory 1623 effect only on GnRH2 secretion (140). These data suggest that PQRFamide peptides may play 1624 a role in the reproductive system of lower vertebrates and that they might have adopted new 1625 functions during the course of vertebrate evolution (568).

1626

1627 <u>5.2.7. QRFP (26RFa) group (*farp-5*)</u>

1628 In 2003, a new peptide family was simultaneously identified by three different laboratories 1629 (104, 213, 322). The first research group isolated the peptide from a frog brain extract, and 1630 named it 26RFa after its 26 amino acid sequence and RF terminal motif (104). The other two 1631 research groups identified the peptide while searching for ligands to an orphan GPCR 1632 (GPR103 also referred to as AQ27 or SP9155) (213, 322). Fukusumi and collaborators named 1633 it QRFP, after its pyroglutamylated arginine-phenylalanine-amide signature; the peptide 1634 consists of 43 amino acid residues (213). Jiang and collaborators (2003) named it P52 and its 1635 precursor protein, P518 (322). Comparative studies indicated that amphibian 26RFa and 1636 mammalian QRFP/P52 are orthologs (825). Phylogeny analyses of RF-amide peptides and 1637 receptors suggested a close relationship between QRFP (26RFa) (farp-5) on the one hand and 1638 the GnIH and NPFF groups (*farp-3* and *farp-1*) on the other hand, likely resulting from local 1639 gene duplication, preceding vertebrate WGD (904) (Figure 9). 1640 A ORFP (26RFa) gene has been described in amphioxus (492, 889), suggesting that the gene

To to a second described in ampinoxus (492, 669), suggesting that the gene

1641 existed in chordates before the emergence of vertebrates. As all vertebrates studied so far

1642 possess a single gene, this suggests that duplicated paralogs resulting from the successive 1R,

1643 2R WGD in early vertebrates and from 3R WGD in early teleosts, might have been lost (for1644 review: (404)).

1645 The main role reported for 26RFa/QRFP is the control of feeding behaviour with a potent 1646 or exigenic effect in mammals and birds, while it may also exert other various functions (for 1647 reviews: (102, 103, 105, 404, 825)). Concerning reproductive functions, in vitro and in vivo 1648 studies support the involvement of 26RFa in the control of the gonadotropic axis in mammals 1649 (528). 26RFa dose-dependently stimulates basal and GnRH-induced LH secretion by male 1650 and cyclic female rat pituitaries in vitro (528). Similarly, in vivo (icv or systemic) injection of 1651 26RFa induces a significant increase of serum LH levels in cyclic and ovariectomized females 1652 (528). Patel and collaborators tested the more potent GPR103 ligand, QRFP43, in vivo and in 1653 vitro (594). They reported that icv injection of QRFP43 to male rats increases plasma LH and 1654 FSH levels; in contrast, intraperitoneal administration does not influence circulating 1655 gonadotropin levels (594). In vitro, QRFP43 stimulates GnRH release from hypothalamic 1656 explants from male rats and from the immortalized hypothalamic GnRH-producing neurons 1657 (GT1-7 cells) (594). In teleosts, one study in the goldfish reported that intraperitoneal 1658 injection of 26RFa increases plasma LH levels; the peptide may exert its action at the brain 1659 level, since 26RFa has no direct effect on LH release in vitro by primary cultures of pituitary 1660 cells (431).

1661

1662 **5.3.** A raising star in mammalian reproduction: Kisspeptin (*farp-4*)

1663 <u>5.3.1. Discovery of Kiss in mammals</u>

In 1996, a metastasis suppressor gene for melanoma cells was identified and called Kiss-1... as a tribute to the chocolate speciality of the city of the authors (227). Then, in 1999, the cDNA of a novel GPCR was isolated from rat brain and named GPR54 (396). Since the 1667 highest identity of this orphan GPCR was observed with the galanin receptors and since its 1668 cerebral distribution was similar to those of galanin receptors, the authors suggested that the 1669 endogenous ligand of GPR54 may have a peptidergic nature (396). Two years later, the 1670 natural ligands of GPR54 were isolated from human placenta (375). The Kiss-1 gene encodes 1671 a 54 amino-acid peptide which may be further processed into shorter peptides of 14 and 13 1672 amino-acids (290, 549), all designated as kisspeptins (375). The peptide of 54 amino-acid 1673 residues was also named metastin (549). The 10-amino acid C-terminal fragment of 1674 kisspeptins (or kiss peptides) carries the biological activity (375). The functional importance 1675 of the C-terminal amidation was shown by the lack of binding and functional response for 1676 non-amidated synthetic kisspeptins (375).

1677 A few years later, a pivotal role of kisspeptin in the control of reproduction was revealed. The 1678 discovery by three research groups (214, 648, 693) that inactivation of kisspeptin receptor 1679 (GPR54) leads to hypogonadotropic hypogonadism in human patients (loss-of-function 1680 mutations) and in mice (knock-out, KO) retained attention of reproductive endocrinologists 1681 and initiated intensive research on kisspeptin for many years. Hypogonadotropic 1682 hypogonadism was also observed in Kiss1 KO mice (135). Conversely, mutations leading to 1683 higher resistance of kisspeptin to degradation (714) or prolonged activation of GPR54 (774) 1684 can induce precocious puberty onset in humans (for review: (714)).

1685

1686 5.3.2. Role of Kiss in the control of reproduction in mammals

1687 <u>a- Stimulation of GnRH neurons</u>

Peripheral (subcutaneous, sc) or intracerebroventricular (icv) injection of kisspeptin increases plasma levels of gonadotropins and this response is abolished by the use of a GnRH antagonist (icv, mice: (237); icv, rats: (307); sc, rats: (464); icv, monkeys: (698)), suggesting a stimulatory effect of kisspeptin on gonadotropins via activation of hypothalamic GnRH 1692 neurons. In addition, coexpression of GPR54 mRNA by GnRH neurons is observed in rat 1693 (307) and mouse (483). Icv administration of kisspeptin in sheep also induces a robust release 1694 of GnRH into the cerebrospinal fluid (483). Intraperitoneal (ip) injection of kisspeptin in 1695 wild-type mice stimulates LH and FSH release, but has no effect in gpr54-/- mice (483). All 1696 these data define the kisspeptin system (ligand and receptor) as a major stimulator of the 1697 gonadotropic axis via induction of GnRH in mammals (rodents, sheep and primates). In rats 1698 and mice, two distinct subpopulations of kisspeptin neurons with different roles on 1699 GnRH/gonadotropic axis have been identified: the hypothalamic arcuate nucleus 1700 subpopulation is involved in the pulse mode of GnRH/LH release while the anteroventral 1701 periventricular nucleus subopulation is implicated in the surge mode of GnRH/LH release (for 1702 review: (820). A direct action of kisspeptin on GnRH neurons was demonstrated *in vitro*, as 1703 kisspeptin induces GnRH release from mediobasal hypothalamus (MBH) explants, while this 1704 did not occur in MBH explants from Gpr54 null mice (770). Primary cultures of whole fetal 1705 rat brain respond to exogenous kisspeptin with increased gnrh mRNA expression (750). 1706 Using mouse GnRH-secreting neuronal cell lines (GT1-7 cells: (480); GN11 cells: (631)), two 1707 studies also showed a stimulatory effect of kisspeptin on GnRH secretion and mRNA levels 1708 (GT1-7 and GN11 cells: (540); GT1-7 cells: (780)); in contrast, another study using GT1-7 1709 cells showed that kisspeptin fails to stimulate GnRH expression while increasing GnRH 1710 receptor transcriptional activity ((751); for review: (343)).

1711 <u>b- Direct action on pituitary and gonads</u>

The expression of GPR54 in the pituitary gland and in particular on LH β cells in rat, ovine and human suggests a potential direct action on pituitary cells (for reviews: (218, 636)). In most mammals, kisspeptin-10 is able to stimulate LH secretion *in vitro* (rat: (527); bovine and porcine: (759); ovine: (727); baboon: (440)). However, other studies in rats have been unable to find any detectable action of kisspeptin on LH directly at the pituitary level (464, 783). 1717 Few data are available on *in vitro* FSH regulation by kisspeptin in mammals. Either no effect 1718 on FSH release (baboon: (440)) or moderate stimulation of GnRH-enhanced FSH secretion 1719 (526) has been reported. Concerning the transcriptional regulation of gonadotropins, Witham 1720 and collaborators showed, using both the mouse pituitary gonadotrope cell line L β T2 and 1721 murine primary pituitary cells, that kisspeptin induces LH β and FSH β expression through the 1722 activation of KissR-1 signaling via PKC (876).

- 1723 Expression of kisspeptin, as well as GPR54, in mammalian ovary (rat: (91); swine: (37)) and
- 1725 at the gonadal level. *In vitro* studies on the functional role of the kisspeptin system in gonads

testis (mice: (854); goat: (660); primate: (306)) supports a role of the local kisspeptin system

- have so far given contradictory results in mammals, with no (mouse Leydig cells: (479, 854);
- 1727 Rhesus monkey testes: (769)) or stimulatory effect on steroidogenesis (swine granulosa cells:
- (38); goat Leydig cells: (661)). In addition, kisspeptin was shown to stimulate fertilizationcapacity of mouse spermatozoa (292).
- 1730

1724

1731 <u>5.3.3. Multiple Kiss and Kiss receptor genes in vertebrates: impact of WGD</u>

1732 <u>a- 1R and 2R WGD origin of multiple Kiss genes followed by gene losses during vertebrate</u>

1733 <u>evolution (Figure 10)</u>

1734 While, in rodents, a single gene (*Kiss1*) is present, up to three *Kiss* genes have been identified 1735 in other vertebrate lineages including cyclostomes (2 genes), chondrichthyes (2 or 3 genes), 1736 actinopterygians (1 or 2 genes), basal sarcopterygian (coelacanth: 3 genes), amphibians (up to 1737 3 genes), sauropsids (0, 1 or 2 genes), as well as in non-rodents mammals (1 or 2 genes). 1738 Phylogenic analysis classified the identified Kiss genes into three different clades (i.e., Kiss1, 1739 Kiss2 and Kiss3 clades, leading to a phylogenetically-based nomenclature according to 1740 Pasquier and collaborators (587, 592). The current three Kiss genes may derive from a single 1741 ancestral gene duplicated through the two successive WGD (1R and 2R) in early vertebrates (for review: (587)). These 1R and 2R events should have yielded four different *Kiss* genes in
vertebrates, and syntenic analysis supports the potential existence of a fourth *Kiss* gene
(*Kiss4*) in early vertebrates (587, 592).

1745 The current diversity of Kiss gene numbers (from 0 to 3) in extant vertebrate species is the 1746 result of multiple and independent pseudogenization processes and gene loss events during 1747 vertebrate evolution that occurred after the 1R and 2R events (for review: (587)). The loss of 1748 the putative Kiss4 gene would have occurred early in vertebrate evolution (shortly after the 1749 2R) as it is absent in all extant species of key phylogenetical positions. Additional Kiss gene 1750 loss events would have happened, as only a few extant representative species of 1751 chondrichthyans (elephant shark) and sarcopterygians (coelacanth and Xenopus) possess three 1752 Kiss genes. Kiss would have been lost independently in early actinopterygians and early 1753 amniotes, while Kiss1 would have been lost independently in some teleosts and sauropsids, 1754 and Kiss2 in some sauropsids and some mammals. The Kiss1 and Kiss2 genes are thus the 1755 most frequent Kiss genes in extant vertebrates. Among sauropsids, both Kiss1 and Kiss2 1756 genes are present in some groups such as chelonians (turtle) and crocodilians, while Kiss1 has 1757 apparently been lost independently in some squamates (anoles) and in birds. Furthermore, 1758 Kiss2 has undergone pseudogenisation and loss during bird evolution, leading to the lack of 1759 Kiss in some species such as the chicken, falcon or parrot (591). In mammals, Kiss1 and 1760 Kiss2 are both present in a prototherian mammal (platypus), while Kiss2 has undergone 1761 pseudogenisation and loss in metatherian (opossum) and eutherian mammals (for review: 1762 (587)). Although a *Kiss2*-like gene is present in the genomes of primates including humans, 1763 due to the lack of amidation signal in the precursor, the endogenous Kiss2 peptide may not be 1764 functional or produced (564).

1765 While teleost 3R WGD (TWGD) should have generated additional Kiss paralogs, extant 1766 teleost species do not possess more *Kiss* genes than a non-teleost actinopterygian (spotted gar)

1767 (*Kiss1* and *Kiss2*) (592). The lack of impact of TWGD suggests a massive loss of 3R-Kiss
1768 paralogs shortly after the 3R event. Furthermore, additional loss of *Kiss1* led to the presence
1769 of a single Kiss gene (*Kiss2*) in some teleosts such as the tetraodon and stickleback (587).

1770 b- 1R and 2R WGD origin of multiple Kiss receptor genes followed by gene losses during

1771 <u>vertebrate evolution</u> (Figure 11)

1772 While, in mammals, only a single gene encodes kisspeptin receptor (KissR), several KissR 1773 genes have been identified in other vertebrates including chondrichthyes (2 genes), teleosts (1, 1774 2 or 3 genes), amphibians (3 genes) and sauropsids (0, 1 or 2 genes) (for review: (587)). As 1775 for Kiss genes, the multiple KissR genes may arose from 1R and 2R, so that the potential 1776 existence of four KissR paralogs in vertebrates has been hypothetized (361, 398). Actually, a 1777 fourth KissR has been discovered in extant representative species of basal sarcopterygians 1778 (coelacanth) and actinopterygians (spotted gar). Phylogenetic analysis classified the identified 1779 Kiss genes into four different clades (i.e., KissR1, KissR2, KissR3 and KissR4 clades), 1780 according to the nomenclature by Pasquier and colleagues (587, 589). Phylogenetic and 1781 syntenic analyses indicate that these four paralogs likely resulted from 1R and 2R WGD 1782 ((589, 592). This suggests that a KissR ancestral gene was present before the emergence of 1783 vertebrates. In agreement with this hypothesis, KissR homologous genes have been identified 1784 in non-vertebrate deuterostomes (174). The KissR1 and KissR3 clades are more closely 1785 related to each other than to *KissR2*, and likely represent sister clades stemming from 2R 1786 (587). A phylogenetic reconstruction of a gene family associated with the KissR paralogon, 1787 the PALM family (298), also supports the hypothesis that 2R gave rise to KissR1 and KissR3 1788 sister paralogons on the one hand, and of KissR2 and KissR4 sister paralogons on the other 1789 hand.

1790 As for the *Kiss* genes, after the 1R and 2R events, the history of the KissR family was driven, 1791 throughout vertebrate evolution, by multiple gene loss events leading to the current diversity 1792 of KissR. KissR4 has been lost independently in the actinopterygian and sarcopterygian 1793 lineages. In actinopterygians, this event has likely occurred in a teleost ancestor after the 1794 emergence of holosteans (e.g. the spotted gar still possesses the four KissRs). Among 1795 sarcopterygians, KissR4, which is still present in a basal species (coelacanth) in some 1796 sauropsids (chelonian and squamates) and in prototherian mammals (platypus), has been lost 1797 independently in the amphibian lineage, in some sauropsids (crocodilians) and in the common 1798 ancestor of metatherian and eutherian mammals, including human which possesses a single 1799 KissR (*KissR1*).

1800 As for Kiss, due to massive gene losses, no effect of the teleost-specific TGWD can be 1801 recorded on the current number of *KissR* in teleosts (587, 589, 592). This is further supported 1802 by a recent study in pejerrey *Odonthesthes bonariensis* including phylogenetic analyses (791).

1803 c- Gene conservation or losses and variations in Kiss/KissR couples

1804 Comparison of the evolutionary histories of Kiss and KissR families (Figures 10 and 11) 1805 highlights both common features and divergences (587). Four Kiss and four KissR genes 1806 arose from 1R and 2R early in the vertebrate lineage, followed by repeated independent gene 1807 loss events throughout vertebrate radiation. As a consequence of the independent and 1808 asynchronous losses of Kiss genes, as well as of KissR genes, the extant vertebrate species 1809 present various combinations of putative Kiss and KissR couples. For example, the same two 1810 Kiss genes (Kiss1 and Kiss2) have been conserved in a prototherian mammal (platypus) and 1811 in some teleosts (e.g. zebrafish, seabass) but together with the conservation of different 1812 receptors (KissR1 and KissR4 in platypus, versus KissR2 and KissR3 in zebrafish). Likewise, 1813 unmatching numbers of Kiss and KissR genes are present in some species, such as three Kiss 1814 genes versus four KissR genes in a basal actinopterygian, the spotted gar and a basal 1815 sarcopterygian (coelacanth), or two Kiss genes versus three KissR genes in a basal teleost, the 1816 eel (587, 592). Such variations in the putative couples between Kiss and KissR paralogs reveal a lack of maintenance and co-evolution of specific Kiss/KissR couples during vertebrate evolution. This may have been favored by the promiscuity of Kiss receptors towards various Kiss peptides, related to the high conservation of the 10 amino-acid Cterminal biologically active sequence (kisspeptin-10 or Kp-10) (398, 413, 561). In species where multiple *Kiss* and *KissR* paralogous genes have been conserved, differential physiological roles related to subfonctionalisation or neofunctionalisation may have emerged between paralogs.

1824

1825 <u>5.3.4.</u> Current studies on the role of Kiss in reproduction in non-mammalian vertebrates

1826 <u>a- In amphibians and reptiles</u>

In amphibians and reptiles, few functional studies are available and concern the action of kisspeptin on the gonads. In the anuran amphibian *Rana esculenta, in vitro* Kp-10 treatment induces testicular expression of both *gpr54* and estrogen nuclear receptor α , *ER* α (109). In this species, *ex vivo* treatment of testis with Kp-10 stimulates spermatogenesis (110, 111) and upregulates the expression of testicular *ER* β , *gnrh 1* and 2, as well as the three types of *gnrh*

1832 receptors (110). All the effects of Kp-10 in amphibians are counteracted by Kp-234, a

1833 kisspeptin antagonist (109, 110). In the reptile Uta stansburiana (side-blotched lizard), intra-

1834 coelomical treatment with kisspeptin up-regulates plasma testosterone levels (531).

1835 <u>b- In teleosts: positive and negative effects on reproduction</u>

Even before the demonstration of colocalization of GPR54 on GnRH neurons in mammals (483), Parhar and collaborators reported that kisspeptin receptor (KissR2) localizes to hypothalamic GnRH neurons in tilapia, using laser-captured single GnRH neurons (578). Similarly, in the cichlid fish *Astatotilapia burtoni* (246) and striped bass *Morone saxatilis* (919), GnRH1 neurons express KissR2 mRNA. In the chub mackerel *Scomber japonicus*,

1841 GnRH1 neurons in the preoptic area co-express KissR1, but not KissR2 (545). Kiss1 and

1842 Kiss2 neurons show differential connections with GnRH neurons in the zebrafish brain (697,1843 732).

1844 In agreement with the expression of KissR by GnRH neurons, effects of kisspeptins on GnRH 1845 neurons have been demonstrated in various teleost species. Kiss1 and Kiss2 peptides have 1846 opposite effects on the electrical activity of hypophysiotropic GnRH3 neurons in adult 1847 zebrafish: Kiss1 stimulates, while Kiss2 inhibits neuronal activity (915). In the striped bass 1848 Morone saxatilis, only Kiss2 (but not Kiss1) upregulates the expression of the 1849 hypophysiotropic gnrh1 by in vitro brain slices (920). In contrast, in hypothalamic explants of 1850 the half-smooth tongue sole, Wang and colleagues showed that Kiss2 has no effect on either 1851 gnrh2 or gnrh3 mRNA levels (852). In vivo studies reported either no (zebrafish gnrh 2 and 3: 1852 (366); grouper gnrh3: (707); hybrid bass gnrh 2 and 3: (919); yellowtail kingfish gnrh1: 1853 (535); male chub mackerel gnrh1: (546, 691, 692)), stimulatory (grouper gnrh1: (707)) or 1854 inhibitory (striped bass gnrh1, 2 and 3: (918)) effects of Kiss peptides on brain transcript 1855 levels of gnrh. Depending on the administration routes, in the female chub mackerel, both 1856 stimulatory (sc (692)) and inhibitory (icv (546)) effects of Kiss2 have been reported on brain 1857 gnrh1 mRNA levels. These in vitro and in vivo studies thus indicate variable effects of 1858 kisspeptins on GnRH in teleosts.

1859 Concerning the effect of kisspeptin on the pituitary in teleosts, direct (*in vitro*) stimulatory 1860 action of kisspeptin 2 (Kp2) on pituitary LH secretion was reported in goldfish (895), striped 1861 bass (918, 920) and sea bass (179), but no effect of Kp1 was observed in goldfish (413) and 1862 sea bass (179)). In contrast, in the European eel, heterologous and homologous Kiss peptides 1863 all directly inhibit the expression of lh, with no effect reported on fsh (588, 590). In the 1864 striped bass also, Kp1 is generally ineffective but, at high doses, can inhibit *lh* expression 1865 (918, 920). A few studies also report a stimulatory effect of kisspeptins on FSH (striped bass, 1866 Kp1 and Kp2 (918, 920); sea bass, Kp2 (179)).

1867 In vivo stimulatory (goldfish (413); sea bass (185); seahorse Hippocampus erectus (911)) or 1868 no (goldfish (524)) action of kisspeptins on gonadotropins have been described in teleosts. In 1869 the striped bass, kisspeptin action depends on the stage of development, as intramuscular 1870 injection of Kp1 has no effect on LH at the pubertal stage, while both Kp1 and Kp2 increase 1871 LH levels at gonadal recrudescence (919). In both sexes (adult immature stage) of the chub 1872 mackerel Scomber japonicus, icv administration of Kp2, but not Kp1, increases $fsh\beta$ and $lh\beta$ 1873 mRNA levels (546, 547).

1874 The role of Kiss was further investigated in knocked-out fish models. Two studies reported 1875 normal fertility in KO fishes for kisspeptin-related genes (zebrafish: (768); medaka: (524)). 1876 In KO medaka for kiss1, kiss2, gpr54-1 or gpr54-2, ovaries and testes develop normally as 1877 compared to wild-type fish, with full-grown and late-vitellogenic follicles, and spermatozoa 1878 being observed, respectively (524). There are also no significant differences in the expression 1879 levels of both $lh\beta$ and $fsh\beta$ between the kisspeptin-related gene KO lines and wild-type (524). 1880 Similarly, zebrafish with mutations for kiss1, kiss2, kiss1 or kiss2 show normal 1881 gametogenesis and reproductive capability, as well as normal expression levels of gnrh2, 1882 gnrh3, $lh\beta$ and $fsh\beta$ (768). Double mutation (kiss1-/-; kiss2-/-), which prevents the 1883 compensation between ligands, causes lower expression of gnrh3, $lh\beta$ and $fsh\beta$, supporting 1884 that kisspeptin signalling stimulates the expression of these genes in zebrafish (768). 1885 However, these decreases had no impact on the phenotypes of the fish, as normal reproductive 1886 performance was reported (768). As compared to the induction of hypogonadotropic 1887 hypogonadism in mammals by loss-of-function mutation (human) or KO (mice) of gpr54 or 1888 kiss1 KO, these observations suggest a dispensable role of Kiss in the neuroendocrine control 1889 of reproduction in teleost models (zebrafish and medaka).

1890 Interestingly, a study readdressed this question in mice by using ablation of kisspeptin1891 neurons or of gpr54-expressing neurons (469). Differently from KO experiments, the authors

observed no impact on the timing of puberty, suggesting that kisspeptin neurons play a role in the optimization of reproductive success, by integrating various cues and relaying this information to GnRH neurons, but are dispensable for puberty onset, activation of GnRH pulses and fertility.

1896 <u>c- Loss of kiss function in birds?</u>

1897 As mentioned above (§ 5.2.3 a, b), phylogenomic studies suggest that two Kiss genes (Kiss1 1898 and *Kiss2*) have been inherited from the amniote ancestor. In the course of amniote evolution, 1899 Kiss 1 has likely been lost early in the bird lineage, while it has been conserved in some other 1900 sauropsids and in mammals. The existence of a Kiss2-like gene has been demonstrated in the 1901 genomes of several bird species, mallard duck, zebra finch, and rock pigeon (591). These 1902 avian Kiss2-like sequences diverge from those of other vertebrates and are not found in the 1903 genomes of chicken, falcon and parrot (591), revealing the processes of Kiss2 gene 1904 pseudogenisation and loss in the avian lineage. Together with the loss of Kiss1, these results 1905 support the absence of any functional kisspeptin in birds. Concerning Kiss receptors, two 1906 genes (KissR1 and KissR4) would have been inherited from the amniote ancestor. Synteny 1907 studies demonstrate the loss of KissR1 in the bird lineage, but cannot conclude for the 1908 presence or not of KissR4 with the current databases. Altogether, current phylogenomic 1909 analysis suggests the loss of *Kiss* genes in birds and a possible conservation or loss of a Kiss 1910 receptor (KissR4) (still to be found).

1911 Immunocytochemical studies reported the presence of kisspeptin-like immunoreactive 1912 neurons in the medial preoptic nucleus of the hypothalamus in the mallard drake in close 1913 proximity to GnRH fibers and almost all these neurons co-express aromatase (658). This may 1914 correspond to the derived sequence identified in the duck even though the peptide mutations 1915 and the variations in gene structure suggest a pseudogenisation (591). Kisspeptin 1916 immunoreactivity is also observed in ovarian granulosa cells of hens (887), while genomic 1917 and syntenic analyses support the loss of Kiss in this species (591). In goose, immunoreactive 1918 kisspeptin is also detected in the ovary (granular layer and theca of the follicle) and in the 1919 plasma where concentrations vary during the reproductive cycle (295). This raises the 1920 question as to whether the detection of immunoreactive kisspeptin in birds may result from 1921 cross-reactions with other RF-amide peptides.

1922 A few functional studies using mammalian kiss peptides have been performed in birds. 1923 Central (icv) administration of kisspeptin-52 is able to increase plasma LH levels in the 1924 mallard drake, and pretreatment with a GnRH receptor antagonist prevented this increase in 1925 LH levels (658). Kisspeptin also increases plasma LH levels in the white-crowned sparrow 1926 (cited in (533)). However, in adult male chicken, human Kp-10 has an inhibitory effect, as 1927 intravenous injections of Kp-10 suppress plasma testosterone levels (847). Kisspeptin-10 1928 treatment stimulates progesterone secretion in cultured chicken granulosa cells (887). 1929 Repeated injections of Kp-10 advance the initiation of egg-laying in the juvenile female quail 1930 (533) by increasing lipid synthesis as well as upregulating estrogen receptor protein and 1931 lipogenic gene expression in the liver (884). In the hypothalamus, Kp-10 induces an up-1932 regulation of *cgnrh-I* expression and a down-regulation of *gnih* expression. In the pituitary, 1933 Kp-10 increases $lh\beta$ and gnrh-rII and decreases $fsh\beta$ expression (533). Further studies are 1934 warranted to investigate whether these actions of heterologous kisspeptin in birds are 1935 mediated by a potential Kiss receptor (KissR4; still to be identified in birds) or by other RF-1936 amide peptide receptors.

1937

1938 <u>5.3.5. Kiss before vertebrates</u>

1939 In deuterostomes, in addition to vertebrates, two other sub-phyla constitute the phylum of 1940 chordates: the urochordates and the cephalochordates. Neither kisspeptin nor kisspeptin 1941 receptors have been found in urochordates (such as ascidians). In contrast, four kisspeptin1942 type precursors and 16 kisspeptin-like receptors have been identified in the cephalochordate 1943 amphioxus Branchiostoma floridae (492), suggesting that multiple gene duplication events 1944 specifically targeted Kiss and KissR in this lineage. In B. floridae, one of the reported genes 1945 encodes three putative peptides, suggesting additional intra-gene duplication of peptidic 1946 sequences, while the other genes encode a single peptide (492). More recently, 27 putative 1947 KissR-like genes were identified in B. floridae (856). Further expression and functional 1948 studies were performed in the amphioxus *B. japonicum* on one kiss-like gene (*BjkissL-2*) and 1949 one kissR-like gene (*Bjgpr54L-1*) (856). Using qPCR and immunohistochemistry, the authors 1950 showed that BjkissL-2 mRNA and protein levels are higher in notochord/nerve cord, and 1951 especially in the anterior region of the nerve cord (cerebral vesicle) and Hatschek's pit (856). 1952 Bjgpr54L-1 expression is predominantly observed by qPCR in the notochord/nerve cord, 1953 moderate expression is seen in the testis and ovary, and lower expression in gill, muscle, 1954 hepatic caecum and hind-gut (856). Two amphioxus peptides show functional interaction with 1955 Bigpr54L-1, while the receptor exhibits higher affinity for zebrafish Kiss peptides (856). 1956 Amphioxus synthetic Kiss peptides induce LH release in sexually mature female zebrafish 1957 and, in amphioxus, trigger the expression of gpb5 mRNA, the glycoprotein subunit related to 1958 vertebrate glycoprotein hormones (see section 3), suggesting an ancestral role of Kiss-KissR 1959 in the neuroendocrine control of reproduction (856).

In non-chordate deuterostomes, echinodermata and hemichordata are sister phyla in a clade known as ambulacraria (285). The occurrence of kisspeptin-type receptors has been reported in ambulacrarian species, the sea urchin *S. purpuratus* (phylum echinoderms) and the acorn worm *Saccoglossus kowalevskii* (phylum hemichordates) (318, 492). Recently, kisspeptintype precursors have also been identified in ambulacrarians, the starfish *Asterias rubens*, *S. purpuratus* and *S. kowalevskii* (694, 695). These precursors comprise two putative kisspeptinlike peptides. 1967 Concerning protostomes, some kisspeptin-type receptors, but not yet a peptide precursor, 1968 have been found in lophotrochozoa (mollusca and annelida) (87, 318, 492). Precisely, 1969 kisspeptin-like receptors are present in mollusca, bivalvia (oysters, *Crassostrea gigas* and 1970 *Pinctata fucata* and mussel, *Mytilus galloprovinciallis*) and gastropoda (limpet, *Lottia 1971 gigantean*; slug, *Aplysia californica*), as well as in annelida (worm, *Capitella teleta*) (87). The 1972 occurrence of kisspeptin-like receptors in lophotrochozoan invertebrates suggests the 1973 existence of kisspeptin-like peptides in these animals.

1974 According to Elphick and collaborators, the evolutionary ancestry of kisspeptin-type receptors 1975 can be traced back to the common ancestor of the bilaterians, but with subsequent loss in 1976 urochordate deuterostomes and ecdysozoan protostomes (for reviews: (174, 175)). However, 1977 based on phylogenic and syntenic analyses using Anopheles mosquitoes (ecdysozoan), it 1978 appears that allostatin type A receptors (AST-ARs) and KissR emerged from a common gene 1979 ancestor after the divergence of galanin receptors (GALRs) in the bilaterian genome, 1980 suggesting that invertebrate AST-ARs and ligands shared a common evolutionary origin with 1981 vertebrate kisspeptin/galanin receptors and ligands (186).

1982

1983 <u>5.3.6. Kisspeptin, more related to galanin/spexin than to other RF-amide peptides</u> 1984

1985 Originally, the kisspeptin receptor was discovered for its high degree of sequence similarity 1986 with galanin receptors (396). Kim and collaborators found that spexin (SPX), galanin (GAL) 1987 and kisspeptin (KISS) genes are on the same reconstructed vertebrate ancestral chromosome 1988 (359). Their family genes are distributed among four 2R-generated paralogons (four linkage 1989 groups). Three linkage groups contain SPX1 and KISS2, SPX2 and GAL, and KISS3 and 1990 galanin-like peptide (GALP), respectively. The fourth linkage group has KISS1 alone. The 1991 ancestral forms of KISS, GAL and SPX are proposed to arise by local tandem duplications 1992 before 2R (Figure 9) and to expand their family members through 2R (359). Likewise, 1993 KISSRs and GALRs have probably emerged through local duplications before 2R, producing 1994 three subgroups in vertebrates: KISSRs (4 KISSRs), GALR1 (GALR1a and GALR1b), and 1995 GALR2/3 (GALR2a, GALR2b and GALR3)(for review: (904). SP1 and SPX2 are functional 1996 ligands for GALR2 and GALR3 and are even more potent than GAL in activating GALR3 1997 (359). The evolutionary origin of the KISS (*farp-4*)/SPX/GAL family is far distant from those 1998 of the NPFF (farp-1)/ORFP(farp-3) and PrRP(farp-2)/NPY families, so that some authors 1999 propose that kisspeptin should be classified separately (for review: (904)). This infers that 2000 common peptide sequences such as the C-terminal sequence shared by the RF-amide peptide 2001 super-family may reflect conservation but also convergences of peptide sequences especially 2002 suitable for bioactivity.

2003 The galanin peptide family is constituted of galanin, galanin message-associated peptide 2004 (GMAP; encoded by the same gene as galanin), galanin-like peptide (GALP; encoded by a 2005 distinct gene) and alarin (encoded by a splice variant of GALP gene) (for review: (389)). 2006 Spexin (SPX; now renamed SPX1) was discovered in 2007 in human proteome (493). A 2007 second form, SPX2, has been characterized in all vertebrates except mammals (359). GALR2 2008 and GALR3, but not GALR1, have been shown to be the cognate receptors for SPX (359). 2009 Regarding functional roles of these peptides, they share an involvement in the control of 2010 reproduction, especially as a link between metabolism and reproduction, which have been 2011 previously reviewed in depth by other authors (for reviews: galanin family, (182, 478); 2012 GALP, (393); spexin, (441)).

2013

2014 **5.4. Prolactin releasing peptide (PrRP) group (***farp-2***)**

The search for the ligand of the human orphan pituitary receptor, hGR3, led to the identification of a new peptide from bovine hypothalamic extract that acts as a prolactinreleasing factor in rat anterior pituitary cells and was thus named prolactin releasing peptide, PrRP (280). The prepro-protein generates at least two forms of mature peptide, PrRP31 and 2019 PrRP20 (a C-terminal fragment of PrRP-31) (280)). At the same time, another RF-amide 2020 peptide, with a similar amino acid sequence, was isolated from the brain of a teleost, the 2021 Japanese crucian carp, and named Carassius-RFa (C-RFa) (210). C-RFa homologs have been 2022 characterized in other teleosts as well as in chicken, Xenopus and lamprey, but not in 2023 mammals, and C-RFa had been considered as a teleost PrRP (for review: (762)). However, 2024 while looking for PrRP and C-RFa in chicken, Wang et al (2012) found that PrRP and C-RFa 2025 are encoded by distinct genes in non-mammalian vertebrates, originating through duplication 2026 of an ancestral gene (386). Tachibana and Sakamoto (762) thus proposed that the widely used 2027 name of PrRP/C-RFa in lower vertebrate species be renamed PrRP2, as previously suggested 2028 by Wang and collaborators (859).

2029 Data in non-mammalian vertebrates indicate that PrRP and C-RFa/PrRP2 are able to stimulate 2030 PRL release in vitro (teleosts: C-RFa in tilapia, (685); C-RFa in rainbow trout, (508, 646)) 2031 and in vivo (bird: comparaison C-RFa et mPrRP, (763); amphibian: PrRP, (655); teleost: 2032 (508)). In contrast, after the initial finding of Hinuma and collaborators (280), other studies in 2033 mammals questioned the role of PrRP as a prolactin releasing factor and thus led to the 2034 hypothesis that this role of PrRP may be ancient and even lost in mammals. In fact, there is 2035 now clear evidence that PrRP is involved in the control of feeding behavior and energy 2036 expenditure in mammals as well as in other vertebrates (for reviews: (154, 762)).

Several studies support a role of PrRP in the neuroendocrine control of reproduction in mammals. Icv injection of PrRP to male rats increases plasma LH, FSH and testosterone levels (684), while icv administration of an anti-rat PrRP serum causes a reduction of LH surge (283). As PrRP is not able to induce release of LH and FSH directly from dispersed male pituitary cells *in vitro*, but instead can increase the release of GnRH from hypothalamic explants (684), it was suggested that PrRP could stimulate gonadotropin secretion *via* a hypothalamic mechanism involving the release of GnRH in male rat. The control of GnRH1 neurons by PrRP in rat is further supported by the presence of PrRP fibers in the medial
preoptic area and of PrRP receptors in GnRH1 neurons (187). To the best of our knowledge,
no data are yet available for such a role in other vertebrates.

2047 Phylogenetic, pharmacological and paralogon analyses reveal that prolactin releasing 2048 hormone receptor (PrRP-R) shares a common ancestry with neuropeptide Y (NPY) receptors 2049 (385). In fact, NPY binds to PrRP-R1 and inhibits response of PrRP-R1-expressing cells to 2050 PrRP, behaving as an antagonist. NPY-type peptides have an ancient origin in bilaterians as 2051 they are present both in protostomes (328) and deuterostomes. An ancestral PrRP peptide 2052 probably started coevolving with a redundant NPY receptor, which then became PrRP-R, 2053 approximately 500 Million years ago (385). The distinct PrRP/NPY family genes and their 2054 receptor genes emerged by local gene duplication during early vertebrate evolution (for 2055 review: (904)) (Figure 9). NPY has been largely recognized for its involvement in the 2056 interactions between food intake, metabolism and reproductive function (for reviews: (146, 2057 473, 519)).

2058

2059 5.5. Conclusion on RF-amide peptides

While RF-amide peptides, which all share a conserved C-terminal RF-amide signature, have been described as a family of neuropeptides encoded by five different genes in mammals, phylogeny and synteny analyses suggest that they do not all derive from a common ancestor and perhaps should not be seen as a family.

As reviewed above, RF-amide peptides, such as GnIH and kisspeptin, are critically involved in the control of reproduction and puberty. Most RF-amide peptides (GnIH, kisspeptin, QRFP/26RFa, PrRP) and non-RF-amide peptides which they are related to, such as NPY for PrRP and galanin/spexin for kisspeptin, are also major regulators of food intake and metabolism (for reviews: (153); GnIH, kisspeptin (848); kisspeptin (877); QRFP/26RFa (404); PrRP (762); galanin, spexin (182, 441); NPY (519)), which can be closely linked to
reproduction (452). Modulation of nociception is mainly exerted by NPFF and, to a lesser
extent, by other RF-amide peptides such as QRFP/26Rfa (for reviews: (29, 404)).

2072 Some data point out that RF-amide peptides display low selectivity toward the different RF-2073 amide peptide receptors, within and between receptor families. Cross-activities between RF-

2074 amide peptides and their various receptors have notably been demonstrated in vitro. For

2075 instance, PrRP has high affinity, and higher efficacy than NPFF, at the NPFFR-2 receptor

2076 (178); 26RFa interacts with a receptor distinct from GPR103 (26RFa receptor) that may be

2077 NPFFR-2 (74); Kisspeptin-10 exhibits high binding affinity and receptor activation towards

2078 NPFFR-1 and NPFFR-2 (551). PrRP, kisspeptin and 26RFa activate NPFFR-1 and NPFFR-2

and thus may modulate nociception (171).

2080 Together, these data highlight the complexity of the diversification of neuropeptides and their

2081 receptors via local and whole genome duplications, conservation, losses and convergences,

and the putative variety of interactions between multiple paralogs of ligands and receptors.

2083

2084 6. Non-peptidic neuromediators and the control of reproduction

2085 <u>6.1. Introduction</u>

In addition to GnRH and some other peptides, non-peptidic neurotransmitters such as monoamines (catecholamines, indolamines and histamine), acetylcholine, and amino acids (glutamic acid and gamma-aminobutyric acid, GABA) may also play a more or less prominent role in the neuroendocrine control of reproduction (for reviews: (34, 222, 621, 840, 864, 875, 921)). This chapter will focus on the role of dopamine, norepinephrine, serotonin, histamine, and GABA in the control of reproduction.

2092

2093 <u>6.1.1. Ancient, multifunctional neuromediators</u>

2094 In the nervous system of adult vertebrates, neurotransmitters act as chemical mediators of 2095 intercellular communication by the activation of specific receptors and second messengers in 2096 postsynaptic cells. This specialized role appears to have evolved from more ancestral 2097 functions in primitive organisms where these substances were used as both intra- and 2098 intercellular signals. Indeed, a number of 'classical' neurotransmitters are present in primitive 2099 organisms and early embryos in the absence of a nervous system (79) and it has been 2100 suggested that these molecules "initially functioned as intracellular regulators, then became 2101 local hormones, and finally acquired the role of synaptic transmitters" (80, 811).

2102 It is now clear that in vertebrates, that most known central nervous system neurotransmitter 2103 substances may also participate in the regulation of pituitary secretion and that in some 2104 instances "neurotransmitters" behave like releasing hormones. The best example is dopamine 2105 (DA) which, in mammals, is released from nerve endings of tuberoinfundibular DA (TIDA) 2106 neurons in the median eminence and pituitary stalk to activate DA receptors at the pituitary 2107 level after being transported there by portal blood vessels. Hence, the definitions of 2108 neurotransmitters and neurohormones are partially intermingled (810). Neurotransmitters can 2109 participate in the control of pituitary secretion by serving as a neurotransmitter input, or 2110 releasing factor output, of hypothalamic neuroendocrine transducer cells; they can act also 2111 within these cells, or act as neurotransmitter within a nerve tract that impinges upon the 2112 hypothalamus (23).

2113 <u>a- Monoamines</u>

Monoamines (such as catecholamines, serotonin, histamine) are evolutionary ancient molecules. They are present in metazoa and also in some protozoa (811). They control neural and endocrine systems in both non-vertebrates and vertebrates. In vertebrates, physiological and pharmacological evidences make it clear that monoamines are involved in the regulation of anterior pituitary hormone secretion, including gonadotropins (LH and FSH). -The catecholamines, dopamine (DA), norepinephrine (NE) and epinephrine (E) are major neurotransmitters found in most, if not all, metazoan species, although the nature of the catecholamines may differ between animal phyla. Their role as neuromediators or neurohormones is documented in a limited number of species: While the neuromodulatory functions of DA appears to be conserved in all bilaterians (505), that of NE is clearly demonstrated only in vertebrates. In mammals, both have been implicated in the regulation of GnRH secretion (222, 339).

2126 -The indoleamine, serotonin (5-hydroxytryptamine; 5-HT), first identified as a 2127 neurotransmitter-signaling molecule in mammals, is ubiquitous across all forms of life. Cells-2128 containing 5-HT have been found in representatives from all major metazoan clades, e.g., 2129 cnidarians, plathyhelminths, nematodes, arthropods, annelids, mollusks, echinoderms, tunicates and vertebrates (30, 137, 268) suggesting an early appearance during animal 2130 2131 evolution. As regards reproductive functions, various studies in vertebrates have 2132 demonstrated a role of 5-HT in the secretion of reproductive neurohormones and sexual 2133 behaviors (for review (621)).

-<u>Histamine</u> (HA): Classically established as a "peripherally" important mediator of
inflammation, HA is now accepted to play a role in neurotransmission (254). The brain
histaminergic system is a phylogenetically old group of neurons that project to most regions
of the central nervous system (253).

2138 <u>b- GABA</u>

GABA (gamma amino butyric acid) is known as the main inhibitory neurotransmitter in the brains of mammals (144). Such a function appears ancestrally derived, as GABA exerts similar inhibitory functions in the nervous system of even the most primitive animals (238).

2142 GABA plays a key role in the control of pituitary hormone release, including gonadotropins,

2143 in mammals (414, 839) as in fish (799).

2144

2145 <u>6.1.2. Non-peptidic neuromediators produced via enzymatic synthesis: phylogeny of enzymes</u> 2146 to retrace their evolutionary history

Nonpeptidic neuromediators are decarboxylated derivatives of amino acids: DA and NE are synthesized from tyrosine, 5-HT from tryptophan, HA from histidine, and GABA from glutamic acid, in one or more enzymatically catalysed reactions of which the first serves as the rate-limiting step.

2151 <u>a -Aromatic Amino acid hydroxylases (AAAH)</u>

The first step in catecholamine and indoleamine biosynthesis is the ring hydroxylation of the aromatic amino acid substrates (tyrosine or tryptophan respectively) catalysed by a class of rate-limiting enzymes, the aromatic amino acid hydroxylases (AAAH). These proteins belong to a family of tetrahydrobiopterin (BH4)-dependent monooxygenases, which in vertebrates includes one phenylalanine hydroxylase (PAH), two tyrosine hydroxylases (TH1 and TH2) and two tryptophan hydroxylases (TPH1 and TPH2).

2158 AAAHs are present in phylogenetically distant taxonomic groups such as bacteria, insects and 2159 mammals (85). The TH, TPH, and PAH genes are found in all sequenced genomes of 2160 bilaterians, supporting the presence of each of the AAAH family members long before the 2161 divergence of chordates (595). Phylogenetic studies suggest that the AAAHs originated by 2162 duplication and divergence of a common ancestral gene at the base of the eukaryotic tree. 2163 Some lineage-specific intron loss or gain events have occurred during the expansion of 2164 AAAH genes and generated diversity of gene structures (85). The high level of sequence 2165 identity between different subfamilies suggests evolutionary conserved functions.

In eukaryotes, these enzymes are composed of a homologous central catalytic domain to which is attached discrete N-terminal regulatory domains and a short C-terminal tetramerization domain, whereas the bacterial enzymes lack the N- and C-terminal domains. Therefore it is believed that the C-terminal portions of all three enzymes arised from a common ancestor, and represent the catalytic core (243) while the amino terminal regions are responsible for substrate specificity. The eukaryotic hydroxylases are all homotetramers (see (195, 773).

2173 -TH catalyzes the hydroxylation of tyrosine to L-dihydroxyphenylalanine (L-DOPA), 2174 the first and rate-limiting step in the biosynthesis of catecholamine neurotransmitters DA, NE 2175 and E (409). Two paralogous genes encoding TH are found in teleost fishes, amphibians, 2176 birds and non-eutherian mammals. They emerged as a consequence of a WGD which 2177 occurred before the divergence of jawed vertebrates (probably 2R-WGD), but TH2 was 2178 secondarily lost in eutherian mammals (893). The history of TH gene evolution within 2179 vertebrates may be also shared by the other AAAH (893).

2180 -TPH catalyzes the hydroxylation of tryptophan to 5-hydroxytryptophan, the first and 2181 rate-limiting step in the biosynthesis of serotonin (321). Two distinct molecular isoforms of TPH, TPH1 and TPH2, resulting from a whole genome duplication (WGD) which occurred in 2182 2183 the vertebrate lineage, are present in the brains of zebrafish (44) as well as humans, mice and 2184 rats (851). The TPH isoform discovered first is referred to as TPH1 and is primarily found in 2185 the pineal gland of mammals (243, 421). By contrast, the second TPH isoform, TPH2 first 2186 cloned from the rat pineal gland (139) is strongly expressed in 5-HT-producing neurons of the 2187 raphe nucleus in mammals (593, 851), and also of the hypothalamus in teleosts (632). 2188 Sequence homology analysis of vertebrate TPHs indicates that the sequence identity between 2189 TPH1 and TPH2 within a species is low, whereas the sequence identities between TPH1s or 2190 TPH2s from different species are relatively high. While two TPH genes (tph1 and tph2), are 2191 found in most groups of jawed vertebrates, in teleost fishes (for instance, in zebrafish, 2192 stickleback and medaka) three paralogs (tph1a, tph1b and tph2) that likely result from the 2193 teleost-specific whole genome duplication (TWGD), have been described (for review (421)).

2194 <u>b- Amino acid decarboxylases (ADC)</u>

2195 The aromatic amino acid decarboxylase (AADC), also called DOPA decarboxylase, is 2196 encoded by the *ddc* gene, and transforms L-DOPA into DA. AADC also transforms 5-HTP 2197 into 5-HT. AADC is a member of the large family of decarboxylases which include other 2198 enzymes involved in the synthesis of neurotransmitters, such as histidine decarboxylase 2199 (HDC) for histamine and glutamate decarboxylase (GAD) for GABA (651). HDC is highly 2200 conserved throughout metazoa (254). AADC is present in both protostomes and 2201 deuterostomes, suggesting its presence in ancestral bilaterians. AADC has been duplicated in 2202 insects leading to two paralogous genes (*ddc* and *amd*). In mammals, alternative splicing 2203 generates various *ddc* transcripts, one being specific to the nervous system (894).

2204 Glutamic acid decarboxylase (GAD) catalyzes the decarboxylation of glutamate, the (single 2205 and) rate-limiting step in the biosynthesis of GABA. Throughout the vertebrate taxa, GAD 2206 exists as two major isoforms, GAD65 and GAD67 (for review (799)). They are thought to 2207 derive from 2 paralogous genes that arose in early vertebrates at least in a gnathostome 2208 ancestor as both GAD65 and GAD67 paralogs have been cloned from two chondrichtyan 2209 species; in contrast a single GAD appears to be present in non-vertebrate chordates, 2210 urochordates (ascidians) an cephalochordates (amphioxus) in which the GAD sequence are 2211 highly divergent (70, 391). There is also evidence for a third GAD gene in a teleost, the 2212 armoured grenadier, although this paralog does not classify with either GAD 65 or 67. It is 2213 found throughout the grenadier brain including the hypothalamus but its importance there 2214 remains unexplored (799).

2215

2216 <u>6.1.3.</u> Origin and evolution of neuromediator receptors: direct phylogeny

Neurotransmitter receptors are critical elements in intercellular signaling within the central
nervous system and are divided into two major types, based on their molecular structure and
biophysical properties.

2220 a-<u>G-protein-coupled receptors (GPCRs)</u>

2221 Most of, if not all, the physiological actions of catecholamines, indolamines and histamine, as 2222 well as some actions of GABA (via GABA_B receptors) are mediated by receptors which 2223 belong to the largest membrane protein family known so far, the seven transmembrane (7-2224 TM) segment receptors coupled to heterotrimeric G-protein (G-protein-coupled receptors or 2225 GPCRs). This family of receptors transmits the effect of agonist binding to the various 2226 intracellular effectors by interacting with an intermediary G-protein (532). The diversity of 2227 receptor subtypes inside this neurotransmitter receptor superfamily, observed in many animal 2228 species, is the result of a long and complex evolutionary process consisting in gene 2229 duplications and continuous accumulation of mutations. GPCRs are believed to have evolved 2230 through a series of gene duplication and divergence events that started with a single ancestral 2231 gene (209): the first event was the divergence of the catecholamine receptors from the 2232 muscarinic acetylcholine receptors, which occurred prior to the divergence between 2233 deuterostomes and protostomes, since they are present at least in arthropod and vertebrate 2234 lineages. Subsequently, the ability to activate specific second-messenger pathways diverged 2235 independently in both the muscarinic and the catecholamine receptors (209).

The calculation of phylogenetical distances between bioamine receptors revealed two main periods of diversification: the first one likely occurred in the Precambrian before the separation between the protostomes and deuterostomes and corresponds to the appearance of the main subtypes of bioamine receptors. The second one, might have accompanied the cephalization of the CNS in vertebrates (836).

2241 b- Ligand-gated ion channels

2242 Ligand-gated ion channels, also called ionotropic receptors, are transmembrane ion-channel 2243 proteins, which after binding of their ligands, allow the passing through of permeant ions 2244 (such as sodium, potassium, chloride and calcium), leading to changes in membrane potential. 2245 This superfamily of proteins is found in almost all forms of life (238), with strong structural 2246 similarities despite the variety in ions gated and in ligands (850). Two subtypes of GABA 2247 receptors (GABA_A and GABA_C) and the subtype 3 of serotonin (5-HT3) receptors are 2248 members of this superfamily. They are composed of five homologous subunits. Evolutionary 2249 scenario of the family of GABA_A and glycine-like receptors (GRL), suggests the presence of 2250 an ancestral receptor in bilateria and that a set of local gene duplications, WGD rounds in 2251 early vertebrates (1 R and 2 R) and gene loss led to current set of GLR subunits in extant 2252 vertebrates including human (803).

2253

2254 **<u>6.2. A key-neuromediator: dopamine</u>**

2255 <u>6.2.1. Major brain neuromediator in human and other mammals</u>

2256 Dopamine (DA) is a neurotransmitter broadly distributed in the vertebrate central nervous 2257 system (CNS) (311). After its discovery as a neurotransmitter in the CNS of vertebrates ((88, 2258 89), DA has raised large interest for its various roles in brain functions and its involvement in 2259 severe human pathologies. DA systems regulate sensory perception, motor function, memory, 2260 emotion, food intake, and various hypothalamic functions including the neuroendocrine 2261 control of pituitary hormones. Pathologies related to the multiple functions of DA include 2262 Parkinson's disease (291), schizophrenia, Tourette Syndrome, addiction to drugs, and 2263 pituitary adenoma (for reviews (63, 311, 681)). The involvement of DA in diverse brain 2264 functions is ensured by independent dopaminergic systems (894).

a - <u>Multiple brain dopaminergic systems; multiple brain functions; multiple receptors</u>

2266 The dopamine systems in vertebrates are highly diversified anatomically and also molecularly 2267 as regards DA biosynthesis enzymes and receptors. Gene duplications and losses have 2268 contributed to this diversification (891-893). The mammalian brain contains ten 2269 neuroanatomically distinct dopaminergic systems (named A8 to A17) located in the forebrain 2270 and midbrain (64). The (A8-A10) DA neurons are involved in initiation of movement and 2271 behavioral responses as well as in drug addiction. The A11 DA neurons are involved in 2272 autonomic motor regulation, and the A12-A15 neurons in neuroendocrine regulation. The 2273 A16 DA cell bodies in the olfactory bulb and A17 in the retina are involved in olfactory and 2274 visual perceptions, respectively (63, 64, 724). Comparative anatomical and functional studies 2275 highlight both conservation and divergences of DA forebrain and midbrain systems across 2276 vertebrates (345, 724, 894).

2277 The pleiotropic activity of DA also relies on a diversity of receptors. DA exerts its actions on 2278 target cells by activating specific membrane receptors belonging to two structurally and 2279 functionally different classes of GPCRs, termed D1 and D2, and originally distinguished by 2280 their ability to activate (D1 class) or inhibit (D2 class) adenylate cyclase (351). D1 and D2 2281 receptors also regulate other signal transduction pathways including calcium and potassium 2282 channels and arachidonic acid synthesis [reviewed in (314, 494, 712, 828)]. D1 and D2 2283 receptors both belong to the group A (rhodopsin-like) of the GPCR superfamily but have 2284 different structural features. D1 receptor genes are intronless, and D1 receptor proteins exhibit 2285 a short third cytoplasmic loop and a long C-terminal tail. They are coupled to the Gs family of 2286 G proteins, and D1 receptor agonists promote adenylyl cyclase activity. In contrast, D2 2287 receptor genes contain several introns and encode a long third cytoplasmic loop and a short C-2288 terminal tail. D2 receptors are coupled to the Gi/Go family of G proteins, and D2 receptor 2289 agonists inhibit adenylyl cyclase activity.

2290 Each class of DA receptors comprises two to five subtypes depending on the vertebrate 2291 species: in mammals, the D1 class comprises two receptor subtypes (D1 and D5), while the 2292 D2 class comprises three receptor subtypes (D2, D3 and D4). However, in non-mammalian 2293 species, additional DA receptor genes have been found. Phylogenetic analyses strongly 2294 suggest that the common ancestor of Osteichthyans possessed four subtypes in the D1-class 2295 (D1, D5, D6 and D7), and five subtypes in the D2-class (D2, D3, D4, D8/D2₁ and D9/D4_{rs}; 2296 (891, 892). While multiple gene losses have largely reduced the number of DA receptors in 2297 mammals, many DA receptor genes have been conserved in other vertebrates; moreover, 2298 teleost-specific WGD (3R) further increased the number of DA receptor genes in this lineage 2299 (891).

Notably, D1 and D2 DA receptors are phylogenetically not more related to each other than to other monoamine receptors (82, 131, 892). This suggests that the ability to bind DA, shared by the two receptor classes, was acquired independently reflecting a convergence process during metazoan evolution.

b- <u>Neuroendocrine functions (*e.g.* major inhibitory regulator of pituitary prolactin via</u>
 <u>dopamine D2 receptors</u>)

2306 In mammals, DA represents the major nonpeptidergic agent exhibiting well-defined 2307 neuroendocrine functions and its role as an inhibitor of pituitary lactotroph cell secretion 2308 became recognized in the 1970s: based on the evidence of dopaminergic nerve terminals in 2309 the median eminence (215), McLeod and coworkers (445) proposed that DA may be released 2310 into the pituitary portal system, and thereby act as a hypothalamic hormone (distinct from its 2311 neurotransmitter role in other systems). They demonstrated that dopaminergic agonists were 2312 effective at suppressing prolactin (PRL) secretion in vivo, and in vitro from isolated pituitary 2313 glands (445). DA was subsequently detected in the pituitary portal blood (340), and variations 2314 of DA levels in the portal blood were shown to account for changes in PRL secretion in various physiological conditions (47, 48, 229, 242). DA receptors of the D2 class were identified on anterior pituitary lactotroph cells (453). Finally, the observation that mice lacking the DA D2 receptor are hyperprolactinemic (352, 652), clearly demonstrated the critical role of DA in suppressing endogenous PRL secretion. Interestingly, in all hormonal systems studied so far, whether at the hypothalamic, posterior pituitary, or anterior pituitary level, DA functions as an inhibitor via DA receptors of the D2 class (for reviews (46, 241)).

2321 While DA is now established as the primary regulator of PRL gene expression and release in both mammals and teleost fish (46, 241, 872), it also exerts some negative control on the 2322 2323 release of growth hormone (GH), thyroid-stimulating hormone (TSH) and melanocyte-2324 stimulating hormone (α -MSH) (for review (810)) in the pituitary of both mammals and teleost 2325 fish, despite differences in the neuroanatomical organization of the hypothalamo-pituitary 2326 axis between these vertebrate classes (32). An additional neuroendocrine action of DA, 2327 initially discovered in teleosts and later demonstrated in various vertebrate species, is the 2328 inhibitory control of gonadotropin secretion and reproductive function ((168, 603); see 2329 section 6.2.4).

2330

2331 <u>6.2.2. Evolution of tyrosine hydroxylase, key-enzyme of dopamine synthesis</u>

2332 <u>a- Biosynthetic pathway of dopamine</u>

Dopamine is synthesized from the aromatic amino acid tyrosine through two enzymatic steps. The first and limiting step is catalyzed by an aromatic amino acid hydroxylase (tyrosine hydroxylase [TH]), and the second step by the aromatic amino acid decarboxylase (AADC), often called DOPA-decarboxylase.

TH, as the other AAAHs, has at least one homolog in all bilaterian species studied so far. TH

2338 probably evolved from an ancestral AAAH that did not have a regulatory domain and gave

rise to all three present-day AAAH (394).

2340 b- Tyrosine hydroxylase 1 and 2 in vertebrates

and TH2 DA-synthesizing cells.

2341 The gene coding TH has been identified in species of several vertebrate groups, including 2342 humans (521), rats (244), mice (301), bovines (136), quails (184), chicken (90) and lamprey 2343 (35). Whereas only one gene encodes TH in mammals (with two to four proteic isoforms 2344 generated by alternative splicing), two non-allelic TH genes (th1 and th2) exist in several 2345 teleost species (84). Phylogenetic analyses showed that both th1 and th2 genes are not only 2346 found in teleosts but also in all studied non-eutherian vertebrates, therefore in all vertebrates 2347 except placental mammals (893). It is thus likely that the ancestral vertebrates possessed TH1 2348

2349 The existence of two paralogous TH genes in teleost fishes, amphibians, birds and non-2350 eutherian mammals suggests that the gene duplication giving rise to the two TH genes is 2351 ancient in the vertebrate lineage (35) probably as a consequence of the second round (2R) of 2352 WGD (548, 575, 713). While TH2 is still found in prototherian (platypus) and metatherian 2353 mammals (opossum), it may have been lost in the eutherian mammalian lineage which 2354 possess a single TH, TH1 (893). Genomes of many eutherians are fully sequenced, and the 2355 chromosomal localizations of the TH genes and their syntenic genes support the loss of TH2 2356 in the eutherian mammals. Indeed, apart from the absence of TH2 in the eutherian mammals, 2357 the order of the neighbouring genes (IGF1, PAH, and ASCL1) at the locus is highly 2358 conserved.

2359 c-Tyrosine hydroxylase in metazoa

2360 Tyrosine hydroxylation, and its regulation, is an ancient and critical biochemical pathway. 2361 The hydroxylation of tyrosine to form L-3, 4-dihydroxyphenylalanine is an enzymatic 2362 pathway utilized not only throughout animal phyla but in plants as well. TH, as the other 2363 AAAH, has at least one homolog in all bilaterian species studied so far (505).

2364 The gene coding TH has been identified from various species of non-vertebrates, including schrimps (454), fresh water crab (D pagei; (615)), parasitic helminthes (Schistosoma mansoni 2365 2366 (258); Schistosoma japonicum (294), and insects, [Drosophila (760); moth (300); cricket 2367 (862)]. In several species of insects, as in mammals, different isoforms of TH produced by 2368 alternative splicing have been identified (in the cricket (862), in D. melanogaster and the 2369 armyworm M. separata (62, 534). Since the drosophila TH protein exhibits similar kinetic 2370 parameters and regulatory mechanisms to its mammalian counterpart, it is likely that very 2371 strong evolutionary constraints have been placed on the biochemical mechanism (529).

2372

2373 <u>6.2.3. Evolution of Dopamine D2 receptors</u>

The D2 receptors have been diversified as a consequence of genome duplications in early vertebrates. In mammals, three DA receptors (D2, D3 and D4) are classified as the D2 class. The classification and nomenclature of DA receptors has been initially based on the set of receptors found in eutherian mammals. When more DA receptors have been found in other groups of vertebrates, the necessity of revisiting DA receptor classification was raised by some authors (147, 891, 892).

2380 <u>a- Impact of WGD</u>

The number of DA receptors subtypes has been multiplied by the two major rounds (1R/2R)

of WGD that took place in early vertebrates (286, 296, 438). Additional duplication of DA

- receptor gene occurred in the teleost fish lineage via TWGD (3R) (15, 316, 617, 772).
- 2384 Duplicated paralogous genes were maintained in some vertebrate groups, probably by the
- 2385 acquisition of some differential functional properties, specific of each subtypes, as well as by
- discrete changes in their expression patterns in the brain (for reviews (861,862)).
- 2387 b- <u>Multiple genes in teleosts</u>

2388 Five subtypes of D2 receptor genes $(D2, D3, D4, D8 \text{ (also named } D2_l \text{ for } D2_{like})$ and D9 (also 2389 named $D4_{rs}$ for D4- related sequence) exist in teleosts. The $D2_l$ and $D4_{rs}$ genes would have 2390 originated in the ancestor of gnathostomes (562). Phylogenetic analyses strongly suggest that 2391 the ancestral Osteichthyes (bony jawed vertebrates) possessed five D2-class of DA receptors 2392 (D2, D3, D4, D8 and D9) likely resulting from 1R/2R with a possible additional specific 2393 duplication (891, 892). In eutherian mammals, only three D2-class DA receptors are found 2394 (D2, D3, D4) due to multiple DA receptor gene loss in this lineage (891). In contrast, up to 2395 seven D2 class DA receptors (D2a, D2b, D8, D3, D4a, D4b, D9) have been identified in the 2396 zebrafish (66, 67, 891), the additional receptors (D2a/D2b, D4a/D4b) likely resulting from 2397 TWGD.

2398

2399 <u>6.2.4. Dopamine as a major inhibitory regulator of gonadotropin secretion in teleosts</u>

A major neuroendocrine action of DA, initially discovered in teleosts and later demonstrated in various vertebrate species, is the inhibitory control of gonadotropin secretion and reproductive function (167, 168, 603). This inhibitory role of DA appears to be a powerful counterpart to the well-known stimulatory role of GnRH in the neuroendocrine control of reproduction.

2405 <u>a- Discovery in goldfish</u>

The discovery by Peter and colleagues, using brain lesion approaches, of the presence of a gonadotropin release-inhibiting factor (GRIF (604)) in the hypothalamus of goldfish has been previously reviewed (167). GRIF neurons were subsequently shown to originate from the preoptic area (POA) and project to the pituitary (606). *In vivo* experiments aimed at investigating the effect of various DAergic agonists and antagonists on LH secretion and ovulation strongly suggested that DA could be the GRIF (100). Immunocytochemical and neuroanatomical studies supported this conclusion by revealing DA cell bodies in a specific 2413 region of the POA, the nucleus preopticus anteroventralis (NPOav), with projections in the 2414 proximal pars distalis of the pituitary where gonadotrope cells are located (333, 606). Lesions 2415 of the NPOav induce a surge of LH and subsequent ovulation (336). Finally, in vitro studies 2416 on primary cultures of goldfish pituitary cells revealed that DA inhibits both basal and GnRH-2417 stimulated LH release by acting directly on gonadotropes, through binding to receptors 2418 pharmacologically-related to the mammalian D2 receptors (101). In addition, in goldfish, DA 2419 exerts inhibitory effects on GnRH neurons, blocking the synthesis of the peptide or its release 2420 from the pituitary nerve terminals (602, 901). The discovery of a dual control of reproduction 2421 led to a major application in aquaculture: the development of a spawning induction method 2422 based on combinated treatments with a DA antagonist and a GnRH agonist that has been widely used to induce spawning in captivity of a certain number of species (LINPE method, 2423 2424 named after the scientists R. Peter and HR Lin (for review (603)).

2425 <u>b- Evidence in various teleosts</u>

As a matter of fact, the role of DA as a GRIF, originally demonstrated in the goldfish, a cyprinid, has been later on observed in various other teleost species, including other cyprinids, but also in species belonging to other teleost subgroups such as silurids, salmonids,

2429 percomorphs, mugiliforms, elopomorphs (for review (167)) and pleuronectiforms (251).

2430 In some teleost species, however, this physiological role of DA was not found (for review

2431 (168)). It is worth noting that the intensity of the dopaminergic inhibition varies according to

the species, from a drastic barrier in cyprinids (603, 605) or in prepubertal eel (166, 837) to a

2433 milder control in salmonids (659).

As in the goldfish, DA actions on gonadotrope cells are mediated by receptors of the D2 class

in all species studied (European eel (326, 838), tilapia (408) catfish (26), grey mullet (10) and

2436 zebrafish (202)). In zebrafish, ISH and ICC studies reveal that the TWGD-paralogs D2a and

2437 D2b as well as D8 are expressed in LH-producing cells, suggesting that the three receptors are

involved in mediating DA inhibitory control in this teleost (202). In the eel, the TWGDparalog D2b is predominantly expressed as compared to D2a in LH and FSH cells (326). The expression of D2a and D2b is differentially regulated by sex steroids (585) and the two receptor subtypes are differentially involved in the onset of puberty (319). These data suggested a functional differenciation of the TGWD-D2 paralogs in the eel, with a major role of the D2b receptor in the control of gonadotropic function (326).

2444

2445 <u>6.2.5. Dopamine as an inhibitory regulator of gonadotropic axis in other vertebrates</u>

A dopaminergic inhibition of GnRH-induced pituitary gonadotropin secretion has also been found in a chondrostean fish, the white sturgeon (*Acipenser transmontanus*; (596)). This indicates that the role of DA in the inhibitory control of reproduction may have a more ancient evolutionary origin, in an actinopterygian ancestor, common to teleostean and chondrostean fishes.

The inhibitory influence of DA on gonadotropin secretion and reproduction is not restricted to fish (584, 837): A dopaminergic inhibitory control of reproduction has been shown also in representative species of various other classes of tetrapods as demonstrated in amphibians, birds, and mammals (for review (168)).

2455 In amphibians, some data point to an important role for dopaminergic inhibition in the 2456 regulation of seasonal reproduction. In the frog, Rana temporaria, lesions of the infundibular 2457 hypothalamus in hibernating preovulatory females cause advanced ovulation (734, 735). This 2458 effect is mimicked by a dopaminergic antagonist and blocked by administration of a 2459 dopaminergic agonist (736). More recent studies indicate that the inhibitory effects of DA on 2460 reproduction are conserved in frogs: for instance, in the wood frog, injection of quinpirole in 2461 females delays or inhibits oviposition (844). The release of sperm and eggs is induced in 2462 several anurans and salamanders by the co-injection of a GnRH agonist and the selective DA

receptor D2 antagonist metoclopramide (termed "the amphiplex method" (121, 240, 795, 796,
845). The technique has also been successfully applied in small-scale trials to several other
amphibian species (796).

In birds, there is similar evidence for an inhibitory role of DA on reproduction (for review (168)). For instance, in laying hens, the anovulatory state induced by food withdrawal, is caused by a dopaminergic inhibition of GnRH release at the median eminence level, where a dense concentrations of DA and GnRH-containing processes is observed, providing opportunities for synaptic interactions between them (128).

2471 In mammals, DA is involved in the modulation of pulsatile LH release in some species. This 2472 effect has been extensively studied in seasonal mammals notably in sheep, in which DA 2473 participates in the inhibition of gonadotropic activity before puberty (73) as well as in adults 2474 during seasonal anestrus (for reviews (221, 781)) by acting on GnRH neurons at the median 2475 eminence level (379), through D2 receptors (55). In rat, GnRH neurons are directly innervated 2476 by catecholamine terminals, including DA ones (106), but the participation of the 2477 dopaminergic system in the control of FSH and LH release remains controversial in adults 2478 since raising brain DA activity can either inhibit (41, 130, 222, 432) or stimulate (412, 444) 2479 gonadotropin release. By contrast, in the developing rat, DA clearly exerts an inhibitory 2480 control on both LH and FSH secretion during the second and third weeks of female life, by 2481 influencing GnRH release into the portal blood (384). In some other species, a direct 2482 inhibitory effects of DA on LH release at the pituitary level has been reported: for instance, in 2483 rabbits, DA injected intravenously inhibits GnRH- stimulated LH secretion in normal and also 2484 in stalk-sectioned animals (138). Finally, in humans, several observations suggest that a 2485 direct inhibition by DA on LH release may occur under physiological and pathological 2486 conditions. For instance in men, infusion of DA (which does not cross the blood-brain barrier) 2487 significantly decreases circulating LH levels and attenuates the LH release in response to a 2488 bolus of GnRH (299). This DA-induced inhibition of LH secretion is associated with a 2489 reduction in both pulse frequency and amplitude of LH secretion. Further, the observation that 2490 partial inhibition of LH secretion occurs even in the presence of constant GnRH and DA 2491 infusion suggests that DA may also modulate the gonadotrope response to GnRH. In women 2492 with functional hypothalamic amenorrhea (FHA), a reduction of LH pulse frequency (and of 2493 PRL concentrations) has been documented, which can be reversed by a DA receptor blocker. 2494 In this case, however, a hypothalamic site of DA action was inferred, through inhibition of 2495 GnRH pulse frequency (52).

Recently, in male and female mouse, DA has been shown as the most potent inhibitor of GnRH neuron excitability: using a variety of electrophysiological approaches, the authors demonstrated that approximately around half of all GnRH neurons are inhibited by DA through D1 and D2 receptors, and that the inhibition exerted by DA on GnRH neurons is independent on gonadal steroid status (430).

2501

2502 In summary, DA acts as an inhibitor of reproduction in representatives of all classes of 2503 vertebrates, but this role has been differentially conserved throughout vertebrate evolution. 2504 This may either reflect an ancient regulatory pathway, common to actynopterygian 2505 (chondrosts and teleosts) and sarcopterygian (tetrapods) species, or result from convergent 2506 recruitments of DA neurons for the inhibition of reproduction in various vertebrate taxa (167). 2507 The recruitment of DA neurons for exerting similar functions in different vertebrates suggests 2508 that DA neurotransmission may be especially suitable for the neuroendocrine inhibitory 2509 control of reproduction.

2510

2511 <u>6.2.6. Dopamine as an inhibitor of reproduction in other metazoan</u>

2512 The inhibitory role of DA in the control of reproduction may be more ancient than the origin 2513 of vertebrates. Indeed, a few studies have reported an inhibitory action of DA on reproductive 2514 function in some non-vertebrate species. For example, DA inhibits the growth and maturation 2515 of oocytes in an echinoderm, the sea urchin Strongylocentrotus nudus (358). It is thus likely 2516 that this control was present in the ancestor of deuterostomes. In protostomes, an inhibitory 2517 effect of DA on reproduction has been reported both in some ecdysozoans (such as insects, 2518 crustaceans, nematodes) and lophotrochozoans (such as mollusks). In insects, DA levels are 2519 constantly maintained at a high level in the hemolymph during diapauses (308, 537, 538), and 2520 change with the photoperiod (115). In particular, DA is clearly a key factor for the induction 2521 of egg diapause of the silkworm, Bombyx mori (537). In crustaceans, there is evidence for a 2522 dopaminergic inhibition of testicular maturation in the fiddler crab, Uca pugilutor (666) and 2523 in the red swamp crayfish, Procambarus clarkii (668). Similarly, in the giant freshwater 2524 prawn, Macrobrachium rosenbergii, DA inhibits testicular maturation (711) and inhibits 2525 vitellogenin synthesis and ovarian maturation by inhibiting the release of vitellogenesis-2526 stimulating hormone (VHS) from the thoracic ganglia (107, 787). In a nematode, 2527 Caenorhabditis elegans, genetic and pharmacological analyses reveal that a dopaminergic 2528 pathway, acting through a D2-like DA receptor (755), inhibits egg laying (865). In a mollusk, 2529 zebra mussel, DA has been shown to reduce the intensity of 5-HT-induced spawning (199). 2530 It is thus likely that inhibitory actions of DA on reproductive functions occurred in the 2531 bilaterian ancestor and even possibly predate the emergence of bilateria, since even in a 2532 cnidarian, the coral Acropora tenuis, DA was reported to have an inhibitory effect on

2533 spawning (310).

Therefore, although its physiological functions appear to differ among species, DA plays multiple roles in suppressing the reproductive events in a number of non-vertebrate species (Figure 12). 2537

2538 6.3. Many other neuromediators involved in the control of reproduction

A variety of actors in mammals: Among neurotransmitters, that have been reported to also influence LH release and play a role in the control of reproduction, are found NE, 5-HT, HA as well as GABA.

2542

2543 <u>6.3.1. Norepinephrine (NE)</u>

2544 <u>a- In mammals</u>

2545 Sawyer and colleagues gave the first evidence that noradrenergic neurons may participate in 2546 the control of the gonadotropic axis, by demonstrating that α -adrenergic receptor antagonist 2547 blocks ovulation in rabbits (676). Further studies by various laboratories reported that NE 2548 may modulate plasma LH levels by regulating the activity of GnRH neurons (for review (34). 2549 However, the idea that NE was essential for triggering of the preovulatory release of LH (34) 2550 has been challenged by experiments in rodents (120, 675), and in primates (369, 376), 2551 showing that chronic interruption of the noradrenergic afferents to the hypothalamus does not 2552 impair normal ovulation. This strongly suggests that the noradrenergic system, far from being 2553 essential, exerts a modulatory action on gonadotropin secretion. It is now accepted that NE 2554 has a permissive action within the network of afferences to GnRH neurons, which may be 2555 modulated by sex steroid levels (275).

2556 <u>b- In other vertebrates</u>

2557 The role of NE in the control of reproduction has also been investigated in other vertebrates.

2558 In birds, existing evidence suggests a role for NE in the neuroendocrine control of LH release.

2559 The avian median eminence is known to be well supplied with NE fibers and terminals (81,

2560 701), and the α -adrenoreceptor blocking drug, dibenamine, has been early shown to

suppresses ovulation in the hen (786). Specific blockade of NE synthesis prevents ovulation

2562 in the laying hen (867) and causes a decline in plasma concentrations of LH in young and 2563 adult male domestic fowl (75, 76). In vitro, GnRH secretion from chicken hypothalamus-2564 preoptic area slices (410) or from Japanese quail hypothalamus explants (486) is significantly 2565 elevated in the presence of NE or β adrenergic agonists. However, icv administration of NE in 2566 laying and non-laying turkeys has no effect on LH secretion (262). In non-laying geese, 2567 noradrenergic stimulation is involved in the control of LH secretion, probably through GnRH 2568 neurons (897). In the Chinese geese (Anas cygnoides), the expression of DBH (the enzyme 2569 catalyzing the transformation of DA into NE) undergoes marked periodic variations in 2570 reproductive and endocrine tissues during the reproductive cycle (890). In the Japanese quail, 2571 a noradrenergic pathway is involved in the control of LH release through a modulation of 2572 GnIH gene expression in the hypothalamus (789).

In teleosts, NE appears to play only a slight stimulatory role on gonadotropin secretion, by acting either directly on gonadotrope cells, via α_1 -like adrenergic receptors (98) or on hypothalamic GnRH neurons to stimulate the release of GnRH (607, 902). In fish, as in mammals, the effectiveness of NE is dependent on the stage of gonadal development: *e.g.* in goldfish, NE stimulates LH release mainly during gonadal regression (98, 603).

2578 <u>c- In other metazoa</u>

2579 Few data are available on the role of NE in the reproductive function in other metazoa. In 2580 mollusks, cyclic variations of NE strongly support its participation in the reproductive cycle 2581 of bivalve species. Notably, marked variations in NE content occur in the gonads and pedal 2582 ganglia of mussels, with a 3 to 4-fold increase during gametogenesis and a decline during 2583 spawning (368). Similarly, increased NE levels are measured in gonad of the female Pacific 2584 lion's paw Scallop during gametogenesis stages (developing and maturing stages) while NE 2585 concentrations drop after spawning, suggesting that NE exerts a direct control over 2586 gametogenesis (433).

Collectively, data reveal the recruitment of NE signalling in the control of reproduction both
in protostomes in deuterostomes, as well as in cnidaria, before the emergence of bilateria.
(Figure 12).

2590

2591 <u>6.3.2. Serotonin (5-HT)</u>

2592 <u>a- In mammals</u>

2593 There is strong pharmacological evidence for a role of 5-HT in the regulation of 2594 hypothalamo-pituitary-gonadal axis, through an action either on GnRH neurons or on 2595 gonadotrope cells. Actually, 5-HT neurons located in the raphe region may play inhibitory 2596 roles in gonadotropin secretion: the pharmacological elevation of hypothalamic 5-HT levels 2597 with monoamine oxidase inhibitors suppress spontaneous ovulation in rats (383) and *in vitro*, 2598 5-HT significantly inhibits the release of GnRH from the mediobasal hypothalamus of male 2599 rats (Charli et al, 1978). In sheep (155) and rabbits (624), infusion of 5-HT into the 2600 mediobasal hypothalamus blocks the ovulatory discharge of LH and ovulation. On the other 2601 hand, several lines of evidence support a permissive or facilitatory role for 5-HT in ovulation: 2602 inhibition of 5-HT synthesis with PCPA, suppresses ovulation in rats (383) and several 5-HT 2603 antagonists inhibit spontaneous ovulation (457).

2604 Actually, the hormonal environment of the animal appears to be critical to the expression of 2605 5-HT actions on gonadotropin secretion and ovulation. Indeed, 5-HT system can be 2606 modulated by gonadal steroids (222, 515, 598) and in several mammalian species, 5-HT 2607 receptors are colocalized with estrogen receptors beta (250), indicating that 5-HT and 2608 reproductive endocrine signaling pathways are closely associated. Accordingly, differential 2609 effects are observed in male and female rats: neonatal exposure to androgen is responsible for 2610 the sex difference in the LH response to 5-HTP, a 5-HT precursor, a difference which may be associated with a regulatory action of 5-HT on the cyclic hypothalamic control of 2611

2612 gonadotropin secretion (501). 5-HTP has been shown to stimulate FSH release in normal rats 2613 and to inhibit it in castrated rats, testosterone concentration being a relevant factor in 2614 determining the 'type' of effect produced (331).

2615 The functional interactions between 5-HT and reproductive functions have also been 2616 demonstrated in other vertebrates:

2617 <u>b- In other vertebrates</u>

In birds, 5-HT synthesized in the hypothalamus exerts an inhibitory influence on reproduction: elevation of 5-HT levels directly inhibits GnRH synthesis and LH secretion in female turkey (256) or hen (657) and male domestic fowl (257). Conversely, blockade of the serotonergic system generally stimulates gonadotropin secretion and enhances gonadal development, as shown in young female turkeys (256), or in aging male roosters (27).

2623 In teleosts, by contrast to what is observed in birds, hypothalamic 5-HT neurons exert a 2624 positive stimulatory influence on reproductive neuroendocrine function and gonadotropin 2625 secretion (for review (621)). Specifically, 5-HT stimulates GnRH release, LH release and 2626 socio-sexual behaviors, acting at different levels. At the hypothalamic level, a close 2627 association of 5-HT fibers with GnRH neurons has been observed in the Atlantic croaker 2628 (354). However, central administration of 5-HT has no effect on preoptic GnRH1 mRNA 2629 levels (782), indicating that 5-HT may stimulate GnRH release but not synthesis. As a matter 2630 of fact, 5-HT stimulates GnRH release from the hypothalamus of the goldfish and seabream 2631 (696, 902). In the zebrafish, 5-HT receptors are present in several regions containing GnRH 2632 neurons (539) suggesting possible expression of 5-HT receptors in GnRH neurons, as in 2633 mammals (56).

At the pituitary level, in several teleost species, 5-HT stimulates gonadotropin release *in vivo* and *in vitro* (goldfish (730, 731, 878); Atlantic croacker (353, 355)). In some other teleost species, 5-HT alone has no influence on spontaneous LH release, but potentiates GnRH analog-induced LH secretion when co-administered (Atlantic croacker (878); Prussian carp (729)). Interestingly, the stimulatory action of 5-HT on GnRH release is time- and seasondependent (355), and the mode of action of 5-HT on gonadotropin release also changes as a function of the reproductive stage: in the Atlantic croaker, the effect of 5-HT is more pronounced in mature than in sexually regressed fish (353).

Therefore, as in mammals, the stimulatory action of 5-HT on gonadotropin secretion in teleost appears to be modulated by various factors such as gonadal steroids, environmental conditions and social cues (for review (621)).

2645 <u>c- In other metazoans</u>

2646 Together with DA and NE, 5-HT represents a possible endocrine and neuronal link between 2647 endogenous and exogenous signals allowing non-vertebrates to modulate reproductive 2648 functions in conjunction with environmental constraints (368). In non-vertebrates, the 2649 function and location of 5-HT have been studied mainly in arthropods, mollusks and annelids. 2650 In crustaceans, 5-HT is involved in various physiological processes including sexual behavior 2651 and reproduction (191, 666, 668, 669, 711). Specifically, 5-HT stimulates release of gonad 2652 stimulating hormone (GSH) in the crayfish Procambatus clarkia (668) and in the fresh water 2653 crab (620) and also accelerates testicular maturation in the fiddler crab (666, 667) and the red 2654 swamp crayfish (668) as well as ovarian maturation and spawning in the white shrimp (827). 2655 In several species of mollusks, 5-HT and its receptors play an important role in reproduction, 2656 through stimulation of biosynthesis and release of active egg-laying peptide precursor 2657 proteins, *i.e.* control and initiation of gamete release, and gamete maturation. For instance, 2658 studies on bivalves show that 5-HT levels change during the different stages of the sexual 2659 cycle (resting, developing, maturing and egg-laying) in the gonads and cerebroid ganglia of 2660 mussels, and that the endocrine function of gonads in marine bivalves is regulated through 2661 monoamine neurosecretions arising from the cerebroid ganglia (368). Moreover,

administration of 5-HT induces oocyte maturation and spawning (282, 466, 766). The endogenous 5-HT signal is transmitted to the oocyte through 5-HT receptors (33, 857).

In annelids such as Nereidae, immunopositive staining for 5-HT is observed in neurons of the brain of *Nereis diversicolor* (278) and 5-HT exerts a positive effect on oocyte development,

2666 probably by stimulating a gonadotrophic hormone, present in mature females (392).

Altogether these studies indicate a conserved or convergent role of 5-HT in the control of reproduction in various bilateria, protostomes and deuterostomes (Figure 12). The presence of 5-HT receptors has been demonstrated in cnidarian, thus before the emergence of bilateria (for review (860)), but to our knowledge the effect of 5-HT on reproduction in this phylum

- has not yet been addressed.
- 2672

2673 <u>6.3.3. Histamine (HA)</u>

2674 The possible stimulatory effect of histamine (HA) on LH secretion was first evidenced in the 2675 rabbit by Sawyer (674) who reported that icv injection of HA induces ovulation and increases 2676 plasma LH levels in animals lightly anesthetized with pentobarbital but not awake or ether-2677 anesthetized. Subsequent studies, however, provided inconsistent results, likely depending on 2678 sex steroid environment: icv injection of HA increases plasma LH levels in proestrous or 2679 ovariectomized estrogen and progesterone-primed female rats (157, 160, 418), but has no 2680 effect in males (156, 158). Actually, HA participates in the neuroendocrine regulation of 2681 pituitary hormone secretion by an indirect action at a hypothalamic level: In mammalian 2682 brain, HA cell bodies are located exclusively in the tuberomammillary nucleus (TMN) of the 2683 posterior hypothalamus (253) and send their axons all over the central nervous system. In the 2684 hypothalamus they arborize extensively and may thereby influence the release and function of 2685 various hypothalamic neuropeptides and hormones (254), including GnRH (368, 395):

2686 GnRH-induced secretion of LH in female rats (497). By contrast, it does not affect LH release

2687 from isolated rat pituitary cells (57) or pituitary fragments (497).

2688 Considering the ancient origin of HA, investigation on its potential role on reproductive 2689 function in other vertebrates and other metazoa would deserve to be launched.

2690

2691 <u>6.3.4. GABA</u>

GABA is an inhibitory neurotransmitter extremely abundant in the brain of all vertebrates,that plays a role in the control of reproduction in various vertebrate classes.

2694 a- In mammals, the influence of GABA on LH secretion has long been demonstrated (21, 2695 159, 388, 560, 839). The distribution of hypothalamic GABAergic neurons and their 2696 involvement in the neural control of reproductive processes are now well documented (451). 2697 Although both GABA_A and GABA_B receptors have been identified in rat pituitary 2698 gonadotrope cells (21) and that evidence exists for a possible direct stimulatory effect of GABA on the LH cells (842), the vast majority of studies support a key inhibitory role of 2699 2700 GABA on gonadotropin release, predominantly at the central level, through modulation of the 2701 GnRH neuron activity and secretion. Notably, increasing endogenous GABA concentration 2702 by drugs, GABA or GABA agonist administration, in rats and non-human primates, prevents 2703 ovulation and pulsatile LH release, and blunts the LH surge induced by estrogen or an 2704 estrogen-progesterone combination (162). GABA infusion into the preoptic area or 2705 intraperitoneal injection of muscimol, a GABA_A receptor agonist, blocks the LH surge (7, 2706 277), while the GABA_A receptor antagonist, bicuculline, advances the timing of the LH surge 2707 (363). Moreover, GABA release in the preoptic area decreases prior to and during the time of 2708 the LH surge (317).

2709 GABAergic activity is known to be sensitive to environmental conditions (183) and the 2710 alteration of GABAergic neuronal activity is likely associated with the season-dependent

2711 reproductive cyclicity in seasonal mammals (235). GABAergic activity may also depend on 2712 the stage of reproductive development: in female mice, at puberty, the effect of GABA on 2713 GnRH neurons switches from depolarization to hyperpolarization ((259). Both 2714 autoradiographic and immunohistochemical studies show that a large proportion of GABA 2715 neurons in the diencephalon represents targets for estrogens (197, 277, 388, 471), and it has 2716 been postulated that these neurons may mediate estrogen feedback on GnRH neurons, (388, 2717 752), as suggested by the existence of synaptic contacts between GABAergic terminals and 2718 GnRH neurons (405, 733). Interestingly, it appeared recently that a subpopulation of GnRH 2719 neurons in the adult mouse forebrain is GABAergic and thus that GnRH neurons could 2720 secrete GABA to regulate their own activity (917).

GABA exerts its regulatory effects via two main receptors, the $GABA_A$ and $GABA_B$ subtypes. The origin and evolution of the GABA receptors is highly complex. As some publications exist, focusing on the phylogeny of the multiple $GABA_A$ receptor (458, 650, 803), this topic will not be adressed in the present review.

2725 In vivo experiments undertaken in adult rats, sheep and monkeys all indicate that the 2726 inhibitory effect of GABA on LH secretion is mediated through the GABA_A receptors located 2727 on the soma of GnRH neurons (276, 277, 495, 683). Furthermore, GnRH neurons express 2728 functional GABA_A receptors that may inhibit GnRH neuronal excitability (260, 717, 743). 2729 Much less is known about the possible involvement of GABA_B receptors. Functional GABA_B 2730 receptors are present in various hypothalamic neurons (11, 12, 387, 846) but studies 2731 examining their role suggest that major species differences exist. In the ewe, GABA_B 2732 receptors mediate inhibition of GnRH secretion during the non-breeding season, but not 2733 during the breeding season, this change being possibly part of the mechanism for the seasonal 2734 shift in the negative-feedback effect of estradiol on LH secretion (683). In the adult rat, 2735 GABA_B receptors play an important role in controlling the excitability of GnRH neurons especially in females (429). Interestingly, GABA stimulates GnRH secretion in the adult rat
(59, 161). Yet, the precise physiological role of the excitatory action of GABA on GnRH
neurons remains elusive (861).

2739 <u>b- In other vertebrates</u>

In birds: The domestic turkey is a seasonally breeding bird with a reproductive cycle regulated by photoperiod and terminated by the state of photorefractoriness (720). In this bird, GABAergic neurotransmission in the hypothalamic premammillary nucleus regulates reproductive seasonality and the onset of photorefractoriness (374) through GABA_A receptors (374).

In teleost fish, there is also good evidence for the involvement of GABA in the control of LH release (451, 799, 921). Following the discovery of a heavy GABAergic innervation of the anterior pituitary in goldfish, (334, 335), the effects of GABA have been explored, mainly in goldfish (for review (616)), and to a lesser extent in trout *Oncorhynchus mykiss*. By contrast to its inhibitory effect in mammals, GABA predominantly stimulates LH secretion in adult goldfish and trout, although inhibitory actions have been described in two other species, the Atlantic croacker (356) and the common carp (116).

2752 In goldfish, intraperitoneal injection of GABA increases serum gonadotropin levels (338). 2753 Injection of a GABA transaminase inhibitor, that augments GABA levels within the 2754 hypothalamus and pituitary, also induces an increase in serum gonadotropin levels (723). This 2755 stimulatory effect of GABA on gonadotropins can be accounted for by two major 2756 mechanisms: stimulation of GnRH secretion (338, 723) and inhibition of DA release (723, 2757 797, 798) at the brain level. In vitro, GABA also induces a dose-related stimulation of GnRH 2758 release from pituitary slices, which in teleosts contain GnRH nerve terminals (797). By 2759 contrast, GABA does not directly stimulate LH release from dispersed goldfish pituitary cells 2760 in vitro (338). The in vivo stimulatory effect of GABA varies seasonally and is modulated by

2761 sex steroids (338, 356, 459, 798). Both muscimol (a GABA_A receptor agonist) and baclofen (a 2762 GABA_B receptor agonist) stimulate LH secretion in females, indicating that both receptor 2763 subtypes may mediate the effect of GABA (459, 799).

2764 In rainbow trout, GABA exerts an overall stimulatory effect on basal and GnRH-induced LH, 2765 but also on FSH secretion, both in vivo and in vitro, and these effects are also highly 2766 dependent on the sex and the reproductive stage of the fish (451). As in mammals and 2767 goldfish, a role for GABAergic neurons as target cells for oestrogen feedback on LH release 2768 has been hypothesized in rainbow trout because oestrogen receptors are present in 2769 GABAergic, but not in GnRH neurons. In addition, the stimulatory action of GABA may also 2770 be exerted directly onto the gonadotrope cells of the pituitary, as GABA stimulates basal and 2771 GnRH-induced FSH and LH secretion from dispersed pituitary cells (451).

In the zebrafish brain, the connection between GABAergic and GnRH neurons has been recently evidenced: GnRH3 neurons are directly contacted by GABAergic neurons in the ventral telencephalon and hypothalamus (733). Moreover, as shown also in mouse (917), some GnRH neurons in the anterior part of the telencephalon coexpress GAD67, thus being also GABAergic. Administration of GABA_B receptor agonists and antagonists support the implication of GABA_B receptors in the regulation of GnRH neurons (733).

2778 In agnaths, studies in sea lamprey also indicate that GABA may directly affect GnRH release

2779 in the brain: dual-label in situ hybridization shows that GABA cells are closely apposed to

2780 GnRH cells in the rostral hypothalamus and preoptic region (647). In vitro perfusion or in

vivo injection of GABA and its analogs both significantly increase GnRH in the brain (647),

2782 suggesting that GABAergic control of GnRH is phylogenetically ancient.

2783 <u>c- In other metazoa</u>

2784 GABA may also play some role in the control of reproduction in other metazoa: for instance,

2785 GABA induces spawning in non-chordate deuterostomes, echinoderms (764). GABA also

2786 enhances reproduction in protostomes such as rotifers (220), indicating a role of GABA in the 2787 control of reproduction in both deuterostomes and protostomes. The presence of GABA 2788 receptors demonstrated in cnidaria, attests the early origin of GABA receptors before the 2789 emergence of bilateria (for review (346)), and should inspire future investigation on the 2790 possible role of GABA in reproduction of basal metazoa (Figure 12).

2791

2792 **6.4.** Neuromediators and reproduction: from conservation to disruption

2793 6.4.1. Ancient origin of neuromediators and receptors in metazoan

2794 The neurotransmitter function of DA, NE, 5-HT, HA and GABA in adult organisms is their 2795 more recently developed function. Cell communication through secreted, diffusible signals is 2796 phylogenetically older than neural transmission (223), and the substances known as 2797 neurotransmitters appeared long before the development of the most primitive nervous system 2798 and even multicellular organisms (79): Organisms like protists and animals devoid of nervous 2799 system, as sponges and placozoans, produce a wide array of molecules, which are in some 2800 cases identical to the corresponding compounds found in higher taxa (223). Monoamines 2801 (catecholamines, indoleamines and HA) are found in many protozoans and in almost all 2802 metazoans (811). Non-bilaterian metazoans, with or without neurons, exhibit many 2803 components of the molecular toolkits for synapse function. GABA, catecholamines, and 2804 indoleamines as well as a large number of G protein-coupled receptors for monoamines, and 2805 biogenic amine-synthesizing enzymes can be found in prebilaterian animals such as 2806 cnidarians (69). GABA is used as a signal molecule by non nervous cells of vertebrates (224) 2807 and also in sponges (173) and in plants (71).

2808 The catecholaminergic signalling has an old phylogenetic history and the enzymatic pathway 2809 for the biosynthesis and degradation of monoamine transmitters and the β -adrenergic-2810 mediated mechanisms were conserved throughout evolution from early metazoans to mammals (223). Alpha-adrenergic receptors are found in representatives of two major cladesof bilateria, the protostomes and the deuterostomes (40).

2813 If we take the example of monoamines, according to Turlejski hypothesis (811): their 2814 "earliest role has been as intracellular regulators of cell activity". Produced either inside the 2815 cell or entering it from its environment, they act directly on some of the intracellular signaling 2816 systems, and "cause long-lasting changes in the cell functional state, that allow the organism 2817 to adapt to its environment". "However, very early in evolution, they acquired functions of 2818 hormones", i.e. molecules of external origin acting on specific receptors localized in the cell 2819 membrane and stimulating specific intracellular second messengers to cause long-term 2820 changes in the functional state of cells. "Later on in evolution, biogenic amines acquired 2821 functions of neuromodulators", thus produced and released by specific neurons, and able to 2822 modulate the activity of other neurons, through specific receptors placed within synaptic 2823 contacts. "Finally, preserving some of the evolutionary older functions, monoamines took the 2824 function of neurotransmitters conveying fast changes", through receptors activating ion 2825 channels (811).

2826 The development of a signaling system requires the evolution of a mechanism for producing 2827 signals, receptors and adaptive reactions to the signal. Various data now support the 2828 evolutionary model proposed by Harris and Zahavi (263) to explain the selection of these 2829 precise molecules for intercellular communication: « Classical neurotransmitters » are small 2830 molecules that are catabolitic products of biological molecules invoved in basal cell 2831 metabolism: DA and NE are produced from tyrosine, 5-HT from tryptophan, HA from 2832 histidine, GABA from glutamate,... and the extracellular release of these molecules reflects 2833 the metabolic activity of the secretory cell. Their (i) physico-chemical characteristics, which 2834 are best suited to display reliably the levels of specific metabolic activities, and (ii) toxicity 2835 which induced the selection by neighboring cells of membrane proteins to bind them, may explain why they have been selected for paracrine communication and then for nervouscommunication (263).

2838

2839 <u>6.4.2. Non-peptidic neuromediators as potential targets for neuroendocrine disruption of</u>
 2840 reproduction: a phylogenetic approach

Neuroendocrine disruptors (NEDs) are defined as "pollutants in the environment that are capable of acting as agonists/antagonists- or altering the synthesis and/or metabolism- of neuropeptides, neurotransmitters or neurohormones, which subsequently alter diverse physiological, behavioral or hormonal processes to affect an animal's capacity to reproduce, develop and grow, or deal with stress and other challenges" (863).

2846 While NEDs are now well known to adversely affect reproduction through binding to steroid 2847 receptors (mimicking or antagonizing the activity of steroid hormones), it is important to 2848 mention that they may also affect reproductive functions by acting upon neurotransmitter 2849 systems (such as DA, GABA, NE and 5-HT) that modulate GnRH output (150, 400). To take 2850 only a few examples in mammals: (1) bisphenol A (BPA): When pregnant rats are orally 2851 administered BPA, the turnover of DA and 5-HT in the forebrain are accelerated in both the 2852 mother and the offspring, suggesting an increase in the release of both monoamines from 2853 nerve endings (288). BPA exposure, at even lower doses than environmentally relevant levels, 2854 was shown to have a great impact on monoamine levels in neonatal rat brain over 28 d after 2855 its disappearance (463). In male mice, an increase in anxiety-like behavior induced by 2856 perinatal exposure to BPA seems to be related to decreases in DA metabolites and in the 2857 activity of monoamine oxidase (MAO)-B in the brain. (462). (2) Polychlorinated biphenyls 2858 (PCBs), that are industrial chemical products, also affect DA and 5-HT levels in the brain: 2859 they have been shown to bind to 5-HT, DA, and NE receptors (689). Decreased brain levels of 2860 DA, DOPAC and HVA are observed in the brain of pregnant rats treated with PCBs, but also

in the brain of their offspring (289). PCBs and brominated flame retardants (BFRs) decrease DA turnover, primarily via inhibition of DA transporters DAT and VMAT, and also decrease the uptake of glutamate and GABA, inhibiting GABA signaling, and decreasing 5-HT levels. Several epidemiological studies have shown that PCBs and BFRs may be responsible for similar effects in humans (for review (200)). (3) GABAergic neurons in the preoptic area of the rat brain are known targets of TCDD, a dioxin, and may mediate developmental effects of this contaminant on reproductive function (270).

In birds also, several hypothalamic neural systems proved to be responsive to NEDs (571– 573): for instance, there is evidence in the quail that the catecholamine and indoleamine systems may be altered by perinatal exposure to NEDs (571).

2871 In fish (for review (863)) various pollutants have been shown to impact neurotransmitters 2872 involved in the neuroendocrine control of reproduction. For example, (a) the antidepressant 2873 fluoxetine (FLU), one of the most prevalent environmental pharmaceutical contaminant in 2874 North American and European aquatic ecosystems, that is a selective 5-HT reuptake inhibitor 2875 (SSRI), was reported to affect multiple neuroendocrine pathways and behavioral circuits in 2876 teleost fishes including disruptive effects on reproduction (481). Recently, in zebrafish 2877 embryos, in very much the same as in mammals, FLU was shown to impair the serotonergic, 2878 dopaminergic and adrenergic neurotransmission systems at environmentally relevant 2879 concentrations, by down-regulating transcription patterns of monoamine receptors and 2880 transporters (134). (b) PCBs disrupt the stimulatory 5-HT/GnRH pathway controlling LH 2881 secretion activity by inhibiting hypothalamic TPH and decreasing 5-HT availability in 2882 croakers (357). (c) Mercury, both in its inorganic and organic forms, is capable of impairing 2883 the monoaminergic neurotransmitters involved in the regulation of GnRH and gonadotropins: 2884 after mercury exposure, 5-HT concentrations are decreased in the brains of several fish 2885 species, whereas levels of NE and DA are increased (for review (133)). (d) Pulp and paper 2886 mill effluents contain compounds that interact with key enzymes and receptors that regulate 2887 the GABAergic, glutamatergic and dopaminergic modulation of GnRH and LH, thereby 2888 impairing reproduction in the goldfish (39). (e) Bifenthrin (BF), a pyrethroid insecticide, 2889 alters dopaminergic function in zebrafish embryos and juveniles (54).

2890 While most investigations on NEDs are carried out with vertebrate models, research on non 2891 vertebrate species led to similar results: For instance, (i) in insects, azadirachtin (AZA), an 2892 insecticidal limonoid with antifeeding and sterilizing properties, blocks neurosecretion from 2893 the corpus cardiaca, interacting with the cholinergic, GABAergic and 5-HT systems (504). (ii) 2894 In crustaceans, cadmium and mercury have inhibitory effects on ovarian maturation in the red 2895 swamp crayfish, partly by inhibiting 5-HT-stimulated gonad-stimulating hormone release, and 2896 preventing the ovaries from responding to this hormone (633). Antidepressants, by interfering 2897 with 5-HT, DA or NE neurotransmission, affect freshwater amphipod activity patterns, 2898 marine amphipod photo- and geotactic behavior, and daphnid reproduction and development 2899 (for review (198)). (iii) In mollusks, in the freshwater mussel, exposure to estrogenic 2900 chemicals present in municipal effluents alters the normal metabolism of 5-HT and DA, both 2901 of which are involved in sexual differentiation (217). Antidepressants affect spawning and 2902 larval release in bivalves and reduce fecundity in snails. Exposure to the environmental 2903 contaminant, serotonergic antidepressant fluoxetine, induces spawning in estuarine and 2904 freshwater bivalves (for review (198)).

2905

2906 <u>6.4.3. The "One Health concept"</u>

The neuroendocrine systems, as the links between the brain and peripheral endocrine systems, play key roles in the ability of an organism to respond to its environment. They are thus highly vulnerable to disruption by environmental contaminants such as industrial chemicals, pesticides, pharmaceuticals..., the worldwide presence of which is now demonstrated. NEDs are known to exert some of their adverse effects acting directly upon neurotransmitterpathways that regulate the neuroendocrine systems.

As most neurotransmitter molecules, as well as some of their receptors and transporters are phylogenetically ancient and conserved within the animal phyla, NEDs may interact with them and affect the neurotransmission systems of all metazoan. Chemical disruption of neuromediator signaling may therefore adversely affect health and reproduction in human but also in a diversity of wildlife thus encompassing the whole ecosystem.

2918

2919 In conclusion, molecular families of neuromediators, hormones, their biosynthesis enzymes 2920 and their receptors, involved in the neuroendocrine control of vertebrate reproduction, have an 2921 ancient origin in metazoa, pre-existing the emergence of vertebrates and the innovation of the 2922 brain-pituitary axis (Figure 13). Evolutionary scenarios, based on phylogeny approaches, 2923 enable retracing the origin and evolution of these molecular families, highlighting the impacts 2924 of gene and genome duplications, as well as gene loss events. Combined with physiological 2925 data they allow inferring evolutionary constraints and mechanisms leading to functional 2926 conservation and diversification of the actors of the neuroendocrine control of reproduction.

2927

2928 Call Out Box

Reproduction is the essential biological process of living organisms and is therefore
 submitted to strong evolutionary constraints.

• In human as in other vertebrates, reproduction is controlled by the brain-pituitary gonadotropic (BPG) axis. Evolutionary endocrinology contributes to a better understanding of the neuro-endocrine control of reproduction in living organisms including human, by providing new insights on the origin, conservation and variations of the key actors of the BPG axis. • As revealed by molecular phylogeny analyses, many actors that have been recruited 2937 for the BPG axis in vertebrates (neuromediators, neurohormones, hormones, biosynthesis 2938 enzymes, and receptors) belong to large molecular families of an ancient origin in metazoa.

• The number of actors of the BPG axis has expanded *via* the two rounds of whole genome duplication that occurred at the basis of the vertebrate lineage, which is at the origin for instance of the two gonadotropins LH and FSH and their sister hormone the thyrotropin TSH. Local gene duplications have also contributed to an increase of actor numbers, and such is the origin of human chorionic gonadotropin.

• An additional round of whole genome duplication has occurred at the basis of the 2945 teleost fish lineage, so that some "fish models" currently used for biomedical research, such 2946 as zebrafish or medaka, may present a larger number of actors as compared to human.

• Conversely to gene increase via duplications, gene loss events have also marked the 2948 evolutionary history of some BPG actor families, as is the case for kisspeptin receptors, with a 2949 single gene left in human, as compared to four in basal vertebrate representatives.

• The synthesis and function of the BPG axis actors (neuromediators, neurohormones and hormones) are the targets of disruption by environmental pollutants from industrial, agricultural and pharmaceutical activities. The ancient origin of the BPG actor families makes a variety of living organisms both targets and sentinels of neuro-endocrine disruption. This highlights some of the strong links between ecosystems, biodiversity health and human health, within the framework of the "OneHealth" concept.

2956

Acknowledgements. The authors thank Dr Anne Gaëlle Lafont for her contribution to figuredrawings.

2959

2960 References

2961	1.	Abel MH. The Effect of a Null Mutation in the Follicle-Stimulating Hormone
2962		Receptor Gene on Mouse Reproduction. Endocrinology 141: 1795–1803, 2000.
2963	2.	Abraham E, Palevitch O, Gothilf Y, Zohar Y. The zebrafish as a model system for
2964		forebrain GnRH neuronal development. Gen Comp Endocrinol 164: 151-160, 2009.
2965	3.	Abraham E, Palevitch O, Gothilf Y, Zohar Y. Targeted gonadotropin-releasing
2966		hormone-3 neuron ablation in zebrafish: Effects on neurogenesis, neuronal migration,
2967		and reproduction. Endocrinology 151: 332-340, 2010.
2968	4.	Acharjee A, Chaube R, Joy KP. Molecular cloning and characterization of the
2969		gonadotropin subunits GPa, FSH β , and LH β genes in the stinging catfish
2970		Heteropneustes fossilis: Phylogeny, seasonal expression and pituitary localization. J
2971		Exp Zool Part A Ecol Genet Physiol 323: 567–585, 2015.
2972	5.	Adams BA, Tello JA, Erchegyi J, Warby C, Hong DJ, Akinsanya KO, Mackie
2973		GO, Vale W, Rivier JE, Sherwood NM. Six novel gonadotropin-releasing hormones
2974		are encoded as triplets on each of two genes in the protochordate, Ciona intestinalis.
2975		Endocrinology 144: 1907–1919, 2003.
2976	6.	Adhikari D, Liu K. Molecular mechanisms underlying the activation of mammalian
2977		primordial follicles. Endocr Rev 30: 438–464, 2009.
2978	7.	Adler BA, Crowley WR. Evidence for gamma-aminobutyric acid modulation of
2979		ovarian hormonal effects on luteinizing hormone secretion and hypothalamic
2980		catecholamine activity in the female rat. Endocrinology 118: 91–97, 1986.
2981	8.	Adoutte A, Balavoine G, Lartillot N, Lespinet O, Prud'homme B, de Rosa R. The
2982		new animal phylogeny: reliability and implications. Proc Natl Acad Sci USA 97: 4453-
2983		4456, 2000.
2984	9.	Agarwal RA, Ligon PJB, Greenberg MJ. The distribution of cardioactive agents
2985		among Molluscan species and tissues. Comp Gen Pharmacol 3: 249-260, 1972.

2986	10.	Aizen J, Meiri I, Tzchori I, Levavi-Sivan B, Rosenfeld H. Enhancing spawning in
2987		the grey mullet (Mugil cephalus) by removal of dopaminergic inhibition. Gen Comp
2988		Endocrinol 142: 212–21, 2005.
2989	11.	Akema T, Kimura F. The mode of GABAB receptor-mediated inhibition of the
2990		preovulatory luteinizing hormone surge in female rats. Brain Res 562: 169–172, 1991.
2991	12.	Akema T, Kimura F. Modulation of pulsatile LH secretion by baclofen, a selective
2992		GABAB receptor agonist, in ovariectomized rats. Neuroendocrinology 56: 141-147,
2993		1992.
2994	13.	Alvarez E, Cahoreau C, Combarnous Y. Comparative structure analyses of cystine
2995		knot-containing molecules with eight aminoacyl ring including glycoprotein hormones
2996		(GPH) alpha and beta subunits and GPH-related A2 (GPA2) and B5 (GPB5)
2997		molecules. Reprod Biol Endocrinol 7: 90, 2009.
2998	14.	Amano M, Moriyama S, Ligo M, Kitamura S, Amiya N, Yamamori K, Ukena K,
2999		Tsutsui K. Novel fish hypothalamic neuropeptides stimulate the release of
3000		gonadotrophins and growth hormone from the pituitary of sockeye salmon. J
3001		Endocrinol 188: 417–423, 2006.
3002	15.	Amores A, Force A, Yan YL, Joly L, Amemiya C, Fritz A, Ho RK, Langeland J,
3003		Prince V, Wang YL, Westerfield M, Ekker M, Postlethwait JH. Zebrafish hox
3004		clusters and vertebrate genome evolution. Science 282: 1711–1714, 1998.
3005	16.	Amoss M, Burgus R, Blackwell R, Vale W, Fellows R, Guillemin R. Purification,
3006		amino acid composition and N-terminus of the hypothalamic luteinizing hormone
3007		releasing factor (LRF) of ovine origin. Biochem Biophys Res Commun 44: 205-210,
3008		1971.
3009	17.	Ancel C, Bentsen AH, Sébert ME, Tena-Sempere M, Mikkelsen JD, Simonneaux
3010		V. Stimulatory effect of RFRP-3 on the gonadotrophic axis in the male Syrian hamster:

- 3011 The exception proves the rule. *Endocrinology* 153: 1352–1363, 2012.
- 3012 18. Anctil M. Evidence for gonadotropin-releasing hormone-like peptides in a cnidarian
 3013 nervous system. *Gen Comp Endocrinol* 119: 317–328, 2000.
- 3014 19. Anctil M. Chemical transmission in the sea anemone Nematostella vectensis: A
- 3015 genomic perspective. *Comp Biochem Physiol Part D Genomics Proteomics* 4: 268–
 3016 289, 2009.
- 3017 20. Anderson GM, Relf HL, Rizwan MZ, Evans JJ. Central and peripheral effects of
 3018 RFamide-related peptide-3 on luteinizing hormone and prolactin secretion in rats.
- 3019 *Endocrinology* 150: 1834–1840, 2009.
- 3020 21. Anderson RA, Mitchell R. Effects of gamma-aminobutyric acid receptor agonists on
- 3021 the secretion of growth hormone, luteinizing hormone, adrenocorticotrophic hormone
- and thyroid-stimulating hormone from the rat pituitary gland in vitro. *J Endocrinol*
- 3023 108: 1–8, 1986.
- 3024 22. Anjum S, Krishna A, Tsutsui K. Inhibitory roles of the mammalian GnIH ortholog
- 3025 RFRP3 in testicular activities in adult mice. *J Endocrinol* 223: 79–91, 2014.
- 3026 23. Anton-Tay F, Wurtman R. Brain monoamines and endocrine function. In: Frontiers
- *in Neuroendocrinology*. Martini L and Ganong WF, 1971, p. 45–66.
- 3028 24. Ascheim S, Zondek B. Hypophysenvorderlappenhormon und Ovarialhormon Im Harn
 3029 Von schwangeren. *Klin Wochenschr* 6: 1322–1324, 1927.
- 3030 25. Ashihara M, Suzuki M, Kubokawa K, Yoshiura Y, Kobayashi M, Urano A, Aida
- 3031 K. Two differing precursor genes for the salmon-type gonadotropin-releasing hormone
 3032 exist in salmonids. *J Mol Endocrinol* 15: 1–9, 1995.
- 3033 26. Van Asselt LA, Goos HJ, De Leeuw R, Peter RE, Hol EM, Wassenberg FP, Van
- 3034 **Oordt PG**. Characterization of dopamine D2 receptors in the pituitary of the African
- 3035 catfish, Clarias gariepinus. *Gen Comp Endocrinol* 80: 107–115, 1990.

3036 2	27.	Avital-Cohen N	N, Heiblum	R, I	Rosenstrauch A,	Chaiseha	Y,	Mobarke	γN,	Gumułka
--------	-----	----------------	------------	------	-----------------	----------	----	---------	-----	---------

- 3037 M, Rozenboim I. Role of the serotonergic axis in the reproductive failure associated
 3038 with aging broiler breeder roosters. *Domest Anim Endocrinol* 53: 42–51, 2015.
- 3039 28. Avsian-Kretchmer O, Hsueh AJW. Comparative Genomic Analysis of the Eight-
- 3040 Membered Ring Cystine Knot-Containing Bone Morphogenetic Protein Antagonists.
- 3041 *Mol Endocrinol* 18: 1–12, 2004.
- 3042 29. Ayachi S, Simonin F. Involvement of mammalian RF-amide peptides and their
- 3043 receptors in the modulation of nociception in rodents. *Front Endocrinol (Lausanne)* 5:
- 3044 1–13, 2014.
- 3045 30. Azmitia E. Serotonin and brain: evolution, neuroplasticity, and homeostasis. *Int Rev*3046 *Neurobiol* 77: 31–56, 2007.
- 3047 31. Van Bael A, Denef C. Evidence for a trophic action of the glycoprotein hormone α3048 subunit in rat pituitary. *J Neuroendocr* 8: 99–102, 1996.
- 3049 32. Ball JN. Hypothalamic control of the pars distalis in fishes, amphibians, and reptiles.
 3050 *Gen Comp Endocrinol* 44: 135–170, 1981.
- 3051 33. Bandivdekar AH, Segal SJ, Koide SS. Demonstration of serotonin receptors in
- isolated spisula oocyte membrane. *Invertebr Reprod Dev* 19: 147–150, 1991.
- 3053 34. Barraclough CA, Wise PM. The Role of Catecholamines in the Regulation of
- 3054 Pituitary Luteinizing Hormone and Follicle-Stimulating Hormone Secretion. *Endocr*3055 *Rev* 3: 91–119, 1982.
- 3056 35. Barreiro-Iglesias A, Laramore C, Shifman MI, Anadón R, Selzer ME, Rodicio
- 3057 MC. The sea lamprey tyrosine hydroxylase: cDNA cloning and in situ hybridization
- 3058 study in the brain. *Neuroscience* 168: 659–669, 2010.
- 3059 36. Barry J, Dubois MP, Poulain P. LRF producing cells of the mammalian
- 3060 hypothalamus A fluorescent antibody study. Zeitschrift für Zellforsch und

3061		Mikroskopische Anat 146: 351–366, 1973.
3062	37.	Basini G, Grasselli F, Bussolati S, Ciccimarra R, Maranesi M, Bufalari A, Parillo
3063		F, Zerani M. Presence and function of kisspeptin/KISS1R system in swine ovarian
3064		follicles. Theriogenology 115: 1-8, 2018.
3065	38.	Basini G, Grasselli F, Bussolati S, Ciccimarra R, Maranesi M, Bufalari A, Parillo
3066		F, Zerani M. Presence and function of kisspeptin/KISS1R system in swine ovarian
3067		follicles. <i>Theriogenology</i> 115: 1–8, 2018.
3068	39.	Basu N, Ta CA, Waye A, Mao J, Hewitt M, Arnason JT, Trudeau VL. Pulp and
3069		paper mill effluents contain neuroactive substances that potentially disrupt
3070		neuroendocrine control of fish reproduction. Environ Sci Technol 43: 1635–1641,
3071		2009.
3072	40.	Bauknecht P, Jékely G. Ancient coexistence of norepinephrine, tyramine, and
3073		octopamine signaling in bilaterians. BMC Biol 15: 1–12, 2017.
3074	41.	Beck W, Hancke JL, Wuttke W. Increased sensitivity of dopaminergic inhibition of
3075		luteinizing hormone release in immature and castrated female rats. <i>Endocrinology</i> 102:
3076		837–843, 1978.
3077	42.	Bégeot M, Hemming F, Dubois P, Combarnous Y, Dubois M, Aubert M. Induction
3078		of pituitary lactotrope differentiation by luteinizing hormone β subunit. Science (80-)
3079		226: 566–568, 1984.
3080	43.	Belchetz P, Plant T, Nakai Y, Keogh E, Knobil E. Hypophysial responses to
3081		continuous and intermittent delivery of hypopthalamic gonadotropin-releasing
3082		hormone. Science (80-) 202: 631–633, 1978.
3083	44.	Bellipanni G, Rink E, Bally-Cuif L. Cloning of two tryptophan hydroxylase genes
3084		expressed in the diencephalon of the developing zebrafish brain. Mech Dev 119 Suppl:
3085		S215-220, 2002.

3086 45. Belmont M, Cazzamali G, Williamson M, Hauser F, Grimmelikhuijzen CJP.

- 3087 Identification of four evolutionarily related G protein-coupled receptors from the
- 3088 malaria mosquito Anopheles gambiae. *Biochem Biophys Res Commun* 344: 160–165,
 3089 2006.
- 3090 46. Ben-Jonathan N, Hnasko R. Dopamine as a prolactin (PRL) inhibitor. *Endocr Rev*3091 22: 724–763, 2001.
- 3092 47. Ben-Jonathan N, Neill MA, Arbogast LA, Peters LL, Hoefer MT. Dopamine in
 3093 hypophysial portal blood: relationship to circulating prolactin in pregnant and lactating
 3094 rats. *Endocrinology* 106: 690–696, 1980.
- 309548.Ben-Jonathan N, Oliver C, Weiner HJ, Mical RS, Porter JC. Dopamine in
- 3096 hypophysial portal plasma of the rat during the estrous cycle and throughout
- 3097 pregnancy. *Endocrinology* 100: 452–458, 1977.
- 3098 49. Bentley GE, Jensen JP, Kaur GJ, Wacker DW, Tsutsui K, Wingfield JC. Rapid
- 3099 inhibition of female sexual behavior by gonadotropin-inhibitory hormone (GnIH).
- 3100 *Horm Behav* 49: 550–555, 2006.
- 3101 50. Bentley GE, Perfito N, Ukena K, Tsutsui K, Wingfield JC. Gonadotropin-inhibitory
- 3102 peptide in song sparrows (Melospiza melodia) in different reproductive conditions, and
- 3103 in house sparrows (Passer domesticus) relative to chicken-gonadotropin-releasing
- 3104 hormone. J Neuroendocrinol 15: 794–802, 2003.
- 3105 51. Bentley GE, Ubuka T, McGuire NL, Chowdhury VS, Morita Y, Yano T,
- 3106 Hasunuma I, Binns M, Wingfield JC, Tsutsui K. Gonadotropin-inhibitory hormone
- and its receptor in the avian reproductive system. *Gen Comp Endocrinol* 156: 34–43,
- 3108 2008.
- 3109 52. Berga SL, Loucks AB, Rossmanith WG, Kettel LM, Laughlin GA, Yen SS.
- 3110 Acceleration of luteinizing hormone pulse frequency in functional hypothalamic

3111		amenorrhea by dopaminergic blockade. J Clin Endocrinol Metab 72: 151-156, 1991.
3112	53.	Berger P, Kranewitter W, Madersbacher S, Gerth R, Geley S, Dirnhofer S.
3113		Eutopic production of human chorionic gonadotropin beta (hCG beta) and luteinizing
3114		hormone beta (hLH beta) in the human testis. FEBS Lett 343: 229-33, 1994.
3115	54.	Bertotto L, Richards J, Gan J, Volz D, Schlenk D. Effects of bifenthrin exposure on
3116		the estrogenic and dopaminergic pathways in zebrafish embryos and juveniles. Environ
3117		<i>Toxicol Chem</i> 37: 236–246, 2018.
3118	55.	Bertrand F, Thiery J, Picard S, Malpaux B. Implication of D2-like dopaminergic
3119		receptors in the median eminence during the establishment of long-day inhibition of
3120		LH secretion in the ewe. J Endocrinol 163: 243–254, 1999.
3121	56.	Bhattarai J, Roa J, Herbison A, Han S. Serotonin acts through 5-HT1 and 5-HT2
3122		receptors to exert biphasic actions on GnRH neuron excitability in the mouse.
3123		Endocrinology 155: 513–524, 2014.
3124	57.	Bicknell RJ, Follett BK. A quantitative assay for luteinizing hormone releasing
3125		hormone (LHRH) using dispersed pituitary cells. Gen Comp Endocrinol 26: 141-152,
3126		1975.
3127	58.	Bigot L, Zatylny-Gaudin C, Rodet F, Bernay B, Boudry P, Favrel P.
3128		Characterization of GnRH-related peptides from the Pacific oyster Crassostrea gigas.
3129		Peptides 34: 303–310, 2012.
3130	59.	Bilger M, Heger S, Brann DW, Paredes A, Ojeda SR. A conditional tetracycline-
3131		regulated increase in Gamma amino butyric acid production near luteinizing hormone-
3132		releasing hormone nerve terminals disrupts estrous cyclicity in the rat. Endocrinology
3133		142: 2102–2114, 2001.
3134	60.	Biran J, Golan M, Mizrahi N, Ogawa S, Parhar IS, Levavi-Sivan B. LPXRFa, the
3135		piscine ortholog of GnIH, and LPXRF receptor positively regulate gonadotropin

3136		secretion in tilapia (Oreochromis niloticus). Endocrinology 155: 4391-4401, 2014.
3137	61.	Birken S, Maydelman Y, Gawinowicz MA, Pound A, Liu Y, Hartree A. Isolation
3138		and characterization of human pituitary chorionic gonadotropin. Endocrinology 137:
3139		1402–1411, 1996.
3140	62.	Birman S, Morgan B, Anzivino M, Hirsh J. A novel and major isoform of tyrosine
3141		hydroxylase in Drosophila is generated by alternative RNA processing. J Biol Chem
3142		269: 26559–26567, 1994.
3143	63.	Björklund A, Dunnett S. Dopamine neuron systems in the brain: an update. Trends
3144		Neurosci 30: 194–202, 2007.
3145	64.	Björklund A, Lindvall O. Dopamine-containing systems in the CNS. In: Classical
3146		Transmitters in the CNS. Amsterdam: Björklund, A and Hökfelt, T, 1984, p. 55-122.
3147	65.	Blithe D, Richards R, Skarulis M. Free alpha molecules from pregnancy stimulate
3148		secretion of prolactin from human decidual cells: a novel function for free alpha in
3149		pregnancy. Endocrinology 129: 2257–2259, 1991.
3150	66.	Boehmler W, Carr T, Thisse C, Thisse B, Canfield VA, Levenson R. D4 Dopamine
3151		receptor genes of zebrafish and effects of the antipsychotic clozapine on larval
3152		swimming behaviour. Genes Brain Behav 6: 155–166, 2007.
3153	67.	Boehmler W, Obrecht-Pflumio S, Canfield V, Thisse C, Thisse B, Levenson R.
3154		Evolution and expression of D2 and D3 dopamine receptor genes in zebrafish. Dev
3155		Dyn An Off Publ Am Assoc Anat 230: 481–493, 2004.
3156	68.	Bonini JA, Jones KA, Adham N, Forray C, Artymyshyn R, Durkin MM, Smith
3157		KE, Tamm JA, Boteju LW, Lakhlani PP, Raddatz R, Yao WJ, Ogozalek KL,
3158		Boyle N, Kouranova E V., Quan Y, Vaysse PJ, Wetzel JM, Branchek TA, Gerald
3159		C, Borowsky B. Identification and characterization of two G protein-coupled receptors
3160		for neuropeptide FF. J Biol Chem 275: 39324–39331, 2000.

3161	69.	Bosch TCG, Klimovich A, Domazet-Lošo T, Gründer S, Holstein TW, Jékely G,
3162		Miller DJ, Murillo-Rincon AP, Rentzsch F, Richards GS, Schröder K, Technau U,
3163		Yuste R. Back to the Basics: Cnidarians Start to Fire. Trends Neurosci 40: 92–105,
3164		2017.
3165	70.	Bosma PT, Blázquez M, Collins MA, Bishop JD, Drouin G, Priede IG, Docherty
3166		K, Trudeau VL. Multiplicity of glutamic acid decarboxylases (GAD) in vertebrates:
3167		molecular phylogeny and evidence for a new GAD paralog. Mol Biol Evol 16: 397-
3168		404, 1999.
3169	71.	Bouché N, Lacombe B, Fromm H. GABA signaling: a conserved and ubiquitous
3170		mechanism. Trends Cell Biol 13: 607-610, 2003.
3171	72.	Braasch I, Gehrke A, Smith J, Kawasaki K, Et A. The spotted gar genome
3172		illuminates vertebrate evolution and facilitates human-to-teleost comparisons. Nat
3173		<i>Genet</i> 48: 427–437, 2016.
3174	73.	Brango CW, Whisnant CS, Goodman RL. A role for catecholaminergic neurons in
3175		the suppression of pulsatile luteinizing hormone secretion in the prepubertal ewe lamb.
3176		Neuroendocrinology 52: 448–454, 1990.
3177	74.	Bruzzone F, Lectez B, Alexandre D, Jégou S, Mounien L, Tollemer H, Chatenet
3178		D, Leprince J, Vallarino M, Vaudry H, Chartrel N. Distribution of 26RFa Binding
3179		Sites and GPR103 mRNA in the Central Nervous System of the Rat FEDERICA. J
3180		Comp Neurol 503: 573–591, 2007.
3181	75.	Buonomo FC, Rabii J, Scanes CG. Aminergic involvement in the control of
3182		luteinizing hormone secretion in the domestic fowl. Gen Comp Endocrinol 45: 162-
3183		166, 1981.
3184	76.	Buonomo FC, Scanes CG. Pharmacological studies on the noradrenergic control of
3185		luteinizing hormone secretion in the domestic fowl. Gen Comp Endocrinol 49: 358-

3186 363, 1983.

3187	77.	Burgus R, Butcher M, Ling N, Monahan M, J R, Fellows R, Amoss M, Blackwell
3188		R, Vale W, Guillemin R. Molecular structure of the hypothalamic factor (LRF) of
3189		ovine origin monitoring the secretion of pituitary gonadotropic hormone of
3190		luteinization (LH). C R Acad Sci Hebd Seances Acad Sci D 273: 1611–1613, 1971.
3191	78.	Burzawa-Gérard E, Fontaine Y. Formation d'une molecule hybride douée d'une
3192		activité gonadotrope sur la Grenouille, à partir de la sous-unité α de l'hormone
3193		lutéinisante bovine et d'une sous-unité de l'hormone gonadotrope d'un Poisson
3194		téléostéen. C R Séances Acad Sci III 282: 97–100, 1976.
3195	79.	Buznikov GA, Shmukler YB, Lauder JM. From oocyte to neuron: do
3196		neurotransmitters function in the same way throughout development? Cell Mol
3197		Neurobiol 16: 537–559, 1996.
3198	80.	Buznikov GA, Shmukler YB, Lauder JM. Changes in the physiological roles of
3199		neurotransmitters during individual development. Neurosci Behav Physiol 29: 11-21,
3200		1999.
3201	81.	Calas A. Radioautographic studies of aminergic neurons terminating in the median
3202		eminence. Adv Biochem Psychopharmacol 16: 79–88, 1977.
3203	82.	Callier S, Snapyan M, Le Crom S, Prou D, Vincent J-D, Vernier P. Evolution and
3204		cell biology of dopamine receptors in vertebrates. Biol Cell 95: 489-502, 2003.
3205	83.	Campbell RK, Satoh N, Degnan BM. Piecing together evolution of the vertebrate
3206		endocrine system. Trends Genet 20: 359-366, 2004.
3207	84.	Candy J, Collet C. Two tyrosine hydroxylase genes in teleosts. Biochim Biophys Acta
3208		1727: 35–44, 2005.
3209	85.	Cao J, Shi F, Liu X, Huang G, Zhou M. Phylogenetic analysis and evolution of
3210		aromatic amino acid hydroxylase. FEBS Lett 584: 4775-4782, 2010.

3211 86. Caraty A, Skinner DC . Gonadotropin-releasing hormone in third ventri	3211	86.	Caraty A, Skinner DC	. Gonadotro	pin-releasing	hormone in t	third ventricular
---	------	-----	----------------------	-------------	---------------	--------------	-------------------

- 3212 cerebrospinal fluid: Endogenous distribution and exogenous uptake. *Endocrinology*3213 149: 5227–5234, 2008.
- 3214 87. Cardoso JCR, Félix RC, Bjärnmark N, Power DM. Allatostatin-type A, kisspeptin
- 3215 and galanin GPCRs and putative ligands as candidate regulatory factors of mantle
- 3216 function. *Mar Genomics* 27: 25–35, 2016.
- 3217 88. Carlsson A. The occurrence, distribution and physiological role of catecholamines in
 3218 the nervous system. *Pharmacol Rev* 11: 490–493, 1959.
- 3219 89. Carlsson A, Lindqvist M, Magnusson T, Waldeck B. On the presence of 3-
- 3220 hydroxytyramine in brain. *Science* 127: 471, 1958.
- 3221 90. Carrier A, Devignes MD, Renoir D, Auffray C. Chicken tyrosine hydroxylase gene:
- isolation and functional characterization of the 5' flanking region. *J Neurochem* 61:
- 3223 2215–2224, 1993.
- 3224 91. Castellano JM, Gaytan M, Roa J, Vigo E, Navarro VM, Bellido C, Dieguez C,
- 3225 Aguilar E, Sánchez-Criado JE, Pellicer A, Pinilla L, Gaytan F, Tena-Sempere M.
- 3226 Expression of KiSS-1 in rat ovary: Putative local regulator of ovulation?
- *Endocrinology* 147: 4852–4862, 2006.
- 3228 92. Catt K, Dufau M, Tsuruhara T. Studies on a radioligand-receptor assay system for

3229 luteinizing hormone and chorionic gonadotropin. *J Clin Endocr Metab* : 860–863,

- 3230 1971.
- 3231 93. Cattanach B, Iddon C, Charlton H, Chiappa S, Fink G. Gonadotrophin-releasing
 hormone deficiency in a mutant mouse with hypogonadism. *Nature* 269: 338–340,
 1977.
- 3234 94. Cazzamali G, Saxild N, Grimmelikhuijzen CJP. Molecular cloning and functional
 3235 expression of a Drosophila corazonin receptor. *Biochem Biophys Res Commun* 298:

3236 31–36, 2002.

3237	95.	Chabot V, Magallon T, Taragnat C, Combarnous Y. Two free isoforms of ovine
3238		glycoprotein hormone alpha-subunit strongly differ in their ability to stimulate
3239		prolactin release from foetal pituitaries. J Endocrinol 164: 287–297, 2000.
3240	96.	Chambery A, Parente A, Topo E, Garcia-Fernàndez J, D'Aniello S.
3241		Characterization and putative role of a type 1 gonadotropin-releasing hormone in the
3242		cephalochordate amphioxus. Endocrinology 150: 812-820, 2009.
3243	97.	Chan Y-M, de Guillebon A, Lang-Muritano M, Plummer L, Cerrato F, Tsiaras S,
3244		Gaspert A, Lavoie HB, Wu C-H, Crowley WF, Amory JK, Pitteloud N, Seminara
3245		SB . GNRH1 mutations in patients with idiopathic hypogonadotropic hypogonadism.
3246		Proc Natl Acad Sci 106: 11703–11708, 2009.
3247	98.	Chang JP, Goor F Van, Acharya S. Influences of Norepinephrine, and Adrenergic
3248		Agonists and Antagonists on Gonadotropin Secretion from Dispersed Pituitary Cells of
3249		Goldfish, Carassius auratus. Neuroendocrinology 54: 202–210, 1991.
3250	99.	Chang JP, Pemberton JG. Comparative aspects of GnRH-Stimulated signal
3251		transduction in the vertebrate pituitary – Contributions from teleost model systems.
3252		Mol Cell Endocrinol 463: 142–167, 2018.
3253	100.	Chang JP, Peter RE. Effects of dopamine on gonadotropin release in female goldfish,
3254		Carassius auratus. Neuroendocrinology 36: 351–357, 1983.
3255	101.	Chang JP, Yu KL, Wong AO, Peter RE. Differential actions of dopamine receptor
3256		subtypes on gonadotropin and growth hormone release in vitro in goldfish.
3257		Neuroendocrinology 51: 664–674, 1990.
3258	102.	Chartrel N, Alonzeau J, Alexandre D, Jeandel L, Alvear-Perez R, Leprince J,
3259		Boutin J, Vaudry H, Anouar Y, Llorens-Cortes C. The RFamide neuropeptide
3260		26RFa and its role in the control of neuroendocrine functions. Front Neuroendocrinol

3261 32: 387–397, 2011.

3262	103.	Chartrel N, Bruzzone F, Leprince J, Tollemer H, Anouar Y, Do-Régo JC, Ségalas-
3263		Milazzo I, Guilhaudis L, Cosette P, Jouenne T, Simonnet G, Vallarino M,
3264		Beauvillain JC, Costentin J, Vaudry H. Structure and functions of the novel
3265		hypothalamic RFamide neuropeptides R-RFa and 26RFa in vertebrates. <i>Peptides</i> 27:

- 3266 1110–1120, 2006.
- 3267 104. Chartrel N, Dujardin C, Anouar Y, Leprince J, Decker A, Clerens S, Do-Régo J-
- 3268 C, Vandesande F, Llorens-Cortes C, Costentin J, Beauvillain J-C, Vaudry H.
- 3269 Identification of 26RFa, a hypothalamic neuropeptide of the RFamide peptide family
 3270 with orexigenic activity. *Proc Natl Acad Sci* 100: 15247–15252, 2003.
- 3271 105. Chartrel N, Picot M, El Medhi M, Arabo A, Berrahmoune H, Alexandre D,
- Maucotel J, Anouar Y, Prévost G. The neuropeptide 26RFa (QRFP) and its role in
 the regulation of energy homeostasis: A mini-review. *Front Neurosci* 10, 2016.
- 3274 106. Chen WP, Witkin JW, Silverman AJ. Gonadotropin releasing hormone (GnRH)
- neurons are directly innervated by catecholamine terminals. *Synapse* 3: 288–290, 1989.
- 3276 107. Chen Y, Fan H, Hsieh S, Kuo C. Physiological involvement of DA in ovarian
- 3277 development of the freshwater giant prawn, Macrobrachium rosenbergii. *Aquaculture*3278 228: 383–395, 2003.
- 3279 108. Cheng CK, Leung PCK. Molecular biology of gonadotropin-releasing hormone
- 3280 (GnRH)-I, GnRH-II, and their receptors in humans. *Endocr Rev* 26: 283–306, 2005.
- 3281 109. Chianese R, Ciaramella V, Fasano S, Pierantoni R, Meccariello R. Kisspeptin
- Receptor, GPR54, as a Candidate for the Regulation of Testicular Activity in the Frog
 Rana esculenta1. *Biol Reprod* 88: 1–11, 2013.
- Chianese R, Ciaramella V, Fasano S, Pierantoni R, Meccariello R. Kisspeptin
 drives germ cell progression in the anuran amphibian Pelophylax esculentus: A study

3286		carried out in ex vivo testes. Gen Comp Endocrinol 211: 81-91, 2015.
3287	111.	Chianese R, Ciaramella V, Fasano S, Pierantoni R, Meccariello R. Kisspeptin
3288		regulates steroidogenesis and spermiation in anuran amphibian. Reproduction 154:
3289		403–414, 2017.
3290	112.	Choi J, Smitz J. Luteinizing hormone and human chorionic gonadotropin: Origins of
3291		difference. Mol Cell Endocrinol 383: 203–213, 2014.
3292	113.	Choi YJ, Kim NN, Habibi HR, Choi CY. Effects of gonadotropin inhibitory hormone
3293		or gonadotropin-releasing hormone on reproduction-related genes in the protandrous
3294		cinnamon clownfish, Amphiprion melanopus. Gen Comp Endocrinol 235: 89-99,
3295		2016.
3296	114.	Chowdhury VS, Ubuka T, Osugi T, Shimura T, Tsutsui K. Identification,
3297		localization and expression of LPXRFamide peptides, and melatonin-dependent
3298		induction of their precursor mRNA in the newt brain. J Endocrinol 209: 211-220,
3299		2011.
3300	115.	Chvalova D, Zdechovanova L, Vaneckova H, Hodkova M. Brain norepinephrine
3301		identified by mass spectrometry is associated with reproductive status of females of the
3302		linden bug Pyrrhocoris apterus. Comp Biochem Physiol B-Biochemistry Mol Biol 168:
3303		70–75, 2014.
3304	116.	Chyb J, Sokołowska-Mikołajczyk M, Mikołajczyk T, Socha M, Epler P. The
3305		influence of gabaergic drugs upon LH secretion from the pituitary of the common carp:
3306		an in vitro study. Reprod Biol 1: 51–61, 2001.
3307	117.	Ciccone N, Dunn I, Boswell T, Tsutsui K, Ubuka T, K U, Sharp P. Gonadotropin
3308		inhibitory hormone depresses gonadotropin alpha and follicle-stimulating hormone
3309		beta subunit expression in the pituitary of the domestic chicken. J Neuroendocrinol 16:
3310		999–1006, 2004.

3311	118.	Clarke IJ, Sari IP, Qi Y, Smith JT, Parkington HC, Ubuka T, Iqbal J, Li Q,
3312		Tilbrook A, Morgan K, Pawson AJ, Tsutsui K, Millar RP, Bentley GE. Potent
3313		action of RFamide-related peptide-3 on pituitary gonadotropes indicative of a
3314		hypophysiotropic role in the negative regulation of gonadotropin secretion.
3315		Endocrinology 149: 5811–5821, 2008.
3316	119.	Clerget E, de Sousa N, Bella A, Maghuin-Rogister G, Beckers J. Placental
3317		glycoproteins in the mammals. Ann Endocrinol 69: 18-29, 2008.
3318	120.	Clifton DK, Sawyer CH. Positive and negative feedback effects of ovarian steroids on
3319		luteinizing hormone release in ovariectomized rats following chronic depletion of
3320		hypothalamic norepinephrine. Endocrinology 106: 1099–1102, 1980.
3321	121.	Clulow J, Trudeau VL, Kouba AJ. Amphibian declines in the twenty-first century:
3322		why we need assisted reproductive technologies. Adv Exp Med Biol 753: 275-316,
3323		2014.
3324	122.	Coetsee M, Millar R, Flanagan C, Lu Z. Identification of Tyr(290(6.58)) of the
3325		human gonadotropin-releasing hormone (GnRH) receptor as a contact residue for both
3326		GnRH I and GnRH II: importance for high-affinity binding and receptor activation.
3327		<i>Biochemistry</i> 47: 10305–10313, 2008.
3328	123.	Cole H, Hart G. The potency of blood serum of mares in progressive stages of
3329		pregnancy in effecting the sexual maturity of the immature rat. Am Jour Physiol 93:
3330		57–68, 1930.
3331	124.	Cole LA, Butler S. Hyperglycosylated hCG, hCG β and Hyperglycosylated hCG β :
3332		Interchangeable cancer promoters. Mol Cell Endocrinol 349: 232–238, 2012.
3333	125.	Collin F, Chartrel N, Fasolo A, Michael Conlon J, Vandesande F, Vaudry H.
3334		Distribution of two molecular forms of gonadotropin-releasing hormone (GnRH) in the
3335		central nervous system of the frogRana ridibunda. Brain Res 703: 111-128, 1995.

- 3336 126. Condliffe P, Bates R. Chromatography of thyrotropin on diethyl-aminoethyl cellulose.
 3337 *Arch Biochem Biophys* 68: 229–236, 1957.
- 3338 127. Conlon J, Collin F, Chiang Y, Sower S, Vaudry H. Two molecular forms of
- 3339 gonadotropin-releasing hormone from the brain of the frog, Rana ribibunda:
- 3340 purification, characterization, and distribution. *Endocrinology* 132: 2117–2123, 1993.
- 3341 128. Contijoch AM, Gonzalez C, Singh HN, Malamed S, Troncoso S, Advis JP.
- 3342 Dopaminergic regulation of luteinizing hormone-releasing hormone release at the
- 3343 median eminence level: immunocytochemical and physiological evidence in hens.
- *Neuroendocrinology* 55: 290–300, 1992.
- 3345 129. Conzelmann M, Williams EA, Krug K, Franz-Wachtel M, Macek B, Jékely G.
- The neuropeptide complement of the marine annelid Platynereis dumerilii. *BMC*
- *Genomics* 14: 906, 2013.
- 3348 130. Craven RP, McDonald PG. The effect of intrahypothalamic infusions of dopamine
 3349 noradrenaline on ovulation in the adult rat. *Life Sci I* 10: 1409–1415, 1971.
- 3350 131. Le Crom S, Kapsimali M, Barôme P-O, Vernier P. Dopamine receptors for every
- 3351 species: gene duplications and functional diversification in Craniates. *J Struct Funct*3352 *Genomics* 3: 161–176, 2003.
- 3353 132. Crowe S, Cushing H, Homans J. Experimental hypophysectomy. *Bull Johns Hopkins*3354 *Hosp* 21: 128–169, 1910.
- 3355 133. Crump KL, Trudeau VL. Mercury-induced reproductive impairment in fish. *Environ* 3356 *Toxicol Chem* 28: 895–907, 2009.
- 3357 134. Cunha V, Rodrigues P, Santos MM, Moradas-Ferreira P, Ferreira M. Fluoxetine
- 3358 modulates the transcription of genes involved in serotonin, dopamine and adrenergic
- 3359 signalling in zebrafish embryos. *Chemosphere* 191: 954–961, 2018.
- 3360 135. d'Anglemont de Tassigny X, Fagg LA, Dixon JPC, Day K, Leitch HG, Hendrick

- 3361 AG, Zahn D, Franceschini I, Caraty A, Carlton MBL, Aparicio SAJR, Colledge
- WH. Hypogonadotropic hypogonadism in mice lacking a functional Kiss1 gene. *Proc Natl Acad Sci* 104: 10714–10719, 2007.
- 136. **D'Mello SR, Turzai LM, Gioio AE, Kaplan BB**. Isolation and structural
- characterization of the bovine tyrosine hydroxylase gene. *J Neurosci Res* 23: 31–40,
 1989.
- 3367 137. Dahlström A, Fuxe K. Localization of monoamines in the lower brain stem.
 3368 *Experientia* 20: 398–399, 1964.
- 138. Dailey RA, Tsou RC, Tindall GT, Neill JD. Direct hypophysial inhibition of
- 3370 luteinizing hormone release by dopamine in the rabbit. *Life Sci* 22: 1491–1498, 1978.
- 3371 139. Darmon MC, Guibert B, Leviel V, Ehret M, Maitre M, Mallet J. Sequence of two
- 3372 mRNAs encoding active rat tryptophan hydroxylase. *J Neurochem* 51: 312–316, 1988.
- 3373 140. Daukss D, Gazda K, Kosugi T, Osugi T, Tsutsui K, Sower SA. Effects of lamprey
- 3374 PQRFamide peptides on brain gonadotropin-releasing hormone concentrations and
- 3375 pituitary gonadotropin-β mRNA expression. *Gen Comp Endocrinol* 177: 215–219,
- 3376 2012.
- 3377 141. Dave A, Krishna A, Tsutsui K. Direct effects of RFRP-1, a mammalian GnIH
- 3378 ortholog, on ovarian activities of the cyclic mouse. *Gen Comp Endocrinol* 252: 193–
 3379 199, 2017.
- 3380 142. Davidson J, Wakefield I, Millar R. Absence of rapid desensitization of the mouse
 3381 gonadotropin-releasing hormone receptor. *Biochem J* 300: 299–302, 1994.
- 3382 143. Decatur WA, Hall JA, Smith JJ, Li W, Sower SA. Insight from the lamprey genome:
- 3383 Glimpsing early vertebrate development via neuroendocrine-associated genes and
- 3384 shared synteny of gonadotropin-releasing hormone (GnRH). *Gen Comp Endocrinol*
- 3385 192: 237–245, 2013.

- 3386 144. DeFeudis F V. Amino acids as central neurotransmitters. *Annu Rev Pharmacol* 15:
 3387 105–130, 1975.
- 3388 145. Dehal P, Boore JL. Two rounds of whole genome duplication in the ancestral
 3389 vertebrate. *PLoS Biol* 3, 2005.
- 3390 146. Delgado MJ, Cerdá-Reverter JM, Soengas JL. Hypothalamic integration of
 3391 metabolic, endocrine, and circadian signals in fish: Involvement in the control of food
 3392 intake. *Front Neurosci* 11, 2017.
- 3393 147. Demchyshyn LL, Sugamori KS, Lee FJ, Hamadanizadeh SA, Niznik HB. The
- dopamine D1D receptor. Cloning and characterization of three pharmacologically
- distinct D1-like receptors from Gallus domesticus. *J Biol Chem* 270: 4005–4012, 1995.
- 3396 148. Densmore VS, Urbanski HF. Relative effect of gonadotropin-releasing hormone
- 3397 (GnRH)-I and GnRH-II on gonadotropin release. *J Clin Endocrinol Metab* 88: 2126–
 3398 2134, 2003.
- 3399 149. Deviche P, Small T, Sharp P, Tsutsui K. Control of luteinizing hormone and
- 3400 testosterone secretion in a flexibly breeding male passerine, the Rufous-winged
- 3401 Sparrow, Aimophila carpalis. *Gen Comp Endocrinol* 149: 226–235, 2006.
- 3402 150. Dickerson S, Gore A. Estrogenic environmental endocrine-disrupting chemical effects
- on reproductive neuroendocrine function and dysfunction across the life cycle. *Rev Endocr Metab Disord* 8: 143–159, 2007.
- 3405 151. Dickinson P, Fairfield W, Hetling J, Hauptman J. Neurotransmitter interactions in
- 3406 the stomatogastric system of the spiny lobster: one peptide alters the response of a
- 3407 central pattern generator to a second peptide. *J Neurophysiol* 77: 599–610, 1997.
- 3408152.Dockray G, Reeve JJ, Shively J, Gayton R, Barnard C. A novel active pentapeptide
- from chicken brain identified by antibodies to FMRFamide. *Nature* 305: 328–330,
- 3410 1983.

- 3411 153. Dockray GJ. The expanding family of -RFamide peptides and their effects on feeding
 3412 behaviour. *Exp Physiol* 89: 229–235, 2004.
- 3413 154. Dodd GT, Luckman SM. Physiological roles of GPR10 and PrRP signaling. *Front* 3414 *Endocrinol (Lausanne)* 4: 1–9, 2013.
- 3415 155. Domański E, Przekop F, Skubiszewski B, Wolińska E. The effect and site of action
 3416 of indoleamines on the hypothalamic centers involved in the control of LH release and
 3417 ovulation in sheep. *Neuroendocrinology* 17: 265–273, 1975.
- 3418 156. **Donoso AO**. Induction of prolactin and luteinizing hormone release by histamine in
- male and female rats and the influence of brain transmitter antagonists. *J Endocrinol*76: 193–202, 1978.
- 3421 157. **Donoso AO**, **Banzan AM**. Acute effects of histamine on plasma prolactin and
- 3422 luteininzing hormone levels in male rats. *J Neural Transm* 39: 95–101, 1976.
- 3423 158. Donoso AO, Banzan AM. Failure of histamine to induce the release of luteinizing
- hormone in castrated rats primed with sex steroids. *J Endocrinol* 78: 447–448, 1978.
- 3425 159. Donoso AO, Banzan AM. Effects of increase of brain GABA levels on the
- 3426 hypothalamic-pituitary-luteinizing hormone axis in rats. *Acta Endocrinol (Copenh)*
- 3427 106: 298–304, 1984.
- 3428 160. Donoso AO, Banzan AM, Borzino MI. Prolactin and luteinizing hormone release
- 3429 after intraventricular injection of histamine in rats. *J Endocrinol* 68: 171–172, 1976.
- 3430 161. Donoso AO, López FJ, Negro-Vilar A. Cross-talk between excitatory and inhibitory
- amino acids in the regulation of luteinizing hormone-releasing hormone secretion.
- 3432 *Endocrinology* 131: 1559–1561, 1992.
- 3433 162. Donoso AO, Seltzer AM, Navarro CE, Cabrera RJ, López FJ, Negro-Vilar A.
- 3434 Regulation of luteinizing hormone-releasing hormone and luteinizing hormone
- 3435 secretion by hypothalamic amino acids. *Brazilian J Med Biol Res = Rev Bras Pesqui*

- 3436 *Medicas E Biol* 27: 921–932, 1994.
- 3437 163. Dubois EA, Zandbergen MA, Peute J, Goos HJ. Evolutionary development of three
 3438 gonadotropin-releasing hormone (GnRH) systems in vertebrates. *Brain Res Bull* 57:
 3439 413–418, 2002.
- 3440 164. Dubos MP, Bernay B, Favrel P. Molecular characterization of an adipokinetic
 3441 hormone-related neuropeptide (AKH) from a mollusk. *Gen Comp Endocrinol* 243: 15–
 3442 21, 2017.
- 3443 165. Ducret E, Anderson GM, Herbison AE. RFamide-related peptide-3, a mammalian
- 3444 gonadotropin-inhibitory hormone ortholog, regulates gonadotropin-releasing hormone 3445 neuron firing in the mouse. *Endocrinology* 150: 2799–2804, 2009.
- 3446 166. Dufour S, Lopez E, Le Menn F, Le Belle N, Baloche S, Fontaine YA. Stimulation of
- 3447 gonadotropin release and of ovarian development, by the administration of a
- 3448 gonadoliberin agonist and of dopamine antagonists, in female silver eel pretreated with
- 3449 estradiol. *Gen Comp Endocrinol* 70: 20–30, 1988.
- 3450 167. Dufour S, Sébert M, Weltzien F, Rousseau K, Pasqualini C. Neuroendocrine
- 3451 control by dopamine of teleost reprodcution. *J Fish Biol* 76: 129–160, 2010.
- 3452 168. Dufour S, Weltzien FA, Sebert ME, Le Belle N, Vidal B, Vernier P, Pasqualini C.
- 3453 Dopaminergic inhibition of reproduction in teleost fishes: Ecophysiological and
- evolutionary implications. *Ann N Y Acad Sci* 1040: 9–21, 2005.
- 3455 169. Dumas L, Kim YH, Karimpour-Fard A, Cox M, Hopkins J, Pollack JR, Sikela
- JM. Gene copy number variation spanning 60 million years of human and primate
 evolution. *Genome Res* 17: 1266–1277, 2007.
- 3458 170. Dunn CW, Hejnol A, Matus DQ, Pang K, Browne WE, Smith SA, Seaver E,
- 3459 Rouse GW, Obst M, Edgecombe GD, Sørensen M V., Haddock SHD, Schmidt-
- 3460 Rhaesa A, Okusu A, Kristensen RM, Wheeler WC, Martindale MQ, Giribet G.

- Broad phylogenomic sampling improves resolution of the animal tree of life. *Nature*452: 745–749, 2008.
- 3463 171. Elhabazi K, Humbert JP, Bertin I, Schmitt M, Bihel F, Bourguignon JJ, Bucher 3464 B, Becker JAJ, Sorg T, Meziane H, Petit-Demoulière B, Ilien B, Simonin F. 3465 Endogenous mammalian RF-amide peptides, including PrRP, kisspeptin and 26RFa, 3466 modulate nociception and morphine analgesia via NPFF receptors. 3467 *Neuropharmacology* 75: 164–171, 2013. 3468 172. Elisio M, Soria FN, Fernandino JI, Strüssmann CA, Somoza GM, Miranda LA. 3469 Extrahypophyseal expression of gonadotropin subunits in pejerrey Odontesthes 3470 bonariensis and effects of high water temperatures on their expression. Gen Comp 3471 Endocrinol 175: 329–336, 2012. 3472 173. Elliott GRD, Leys SP. Evidence for glutamate, GABA and NO in coordinating 3473 behaviour in the sponge, Ephydatia muelleri (Demospongiae, Spongillidae). J Exp Biol 3474 213: 2310-2321, 2010. 3475 Elphick MR, Mirabeau O. The evolution and variety of RFamide-type neuropeptides: 174. 3476 Insights from deuterostomian invertebrates. Front Endocrinol (Lausanne) 5: 1-11, 3477 2014. 3478 Elphick MR, Mirabeau O, Larhammar D. Evolution of neuropeptide signalling 175. 3479 systems. J Exp Biol 221: jeb151092, 2018. 3480 Elshourbagy NA, Ames RS, Fitzgerald LR, Foley JJ, Chambers JK, Szekeres PG, 176. 3481 Evans NA, Schmidt DB, Buckley PT, Dytko GM, Murdock PR, Milligan G,
 - 3482 Groarke DA, Tan KB, Shabon U, Nuthulaganti P, Wang DY, Wilson S, Bergsma
 - 3483 DJ, Sarau HM. Receptor for the pain modulatory neuropeptides FF and AF is an
 - 3484 orphan G protein-coupled receptor. *J Biol Chem* 275: 25965–25971, 2000.
 - 3485 177. Emanuele N, Connick E, Howell T, Anderson J, Hojvat S, Baker G, Souchek J,

- 3486 Kirsteins L, Lawrence A. Hypothalamic luteinizing hormone (LH): characteristics 3487 and response to hypophysectomy. *Biol Reprod* 25: 321–326, 1981. 3488 178. Engström M, Brandt A, Wurster S, Savola J, Panula P. Prolactin releasing peptide 3489 has a high affinity and efficacy at neuropeptide FF2 receptors. J Pharmacol Exp Ther 3490 305: 825-832, 2003. 3491 Espigares F, Zanuy S, Gómez A. Kiss2 as a Regulator of Lh and Fsh Secretion via 179. 3492 Paracrine/Autocrine Signaling in the Teleost Fish European Sea Bass (Dicentrarchus 3493 labrax)1. Biol Reprod 93: 1–12, 2015. 3494 180. Evans H, Long J. Characteristic effects upon growth, oestrus and ovulation induced 3495 by the intraperitoneal administration of fresh anterior hypophyseal substance. Proc 3496 Natl Acad Sci 8: 38–39, 1922. 3497 181. Fan NC, Peng C, Krisinger J, Leung PCK. The human gonadotropin-releasing 3498 hormone receptor gene: complete structure including multiple promoters, transcription 3499 initiation sites, and polyadenylation signals. Mol Cell Endocrinol 107, 1995. 3500 182. Fang P, He B, Shi M, Kong G, Dong X, Zhu Y, Bo P, Zhang Z. The regulative 3501 effect of galanin family members on link of energy metabolism and reproduction. 3502 Peptides 71: 240-249, 2015. 3503 Farajnia S, van Westering T, Meijer J, Michel S. Seasonal induction of GABAergic 183. 3504 excitation in the central mammalian clock. Proc Natl Acad Sci USA 111: 9627–9632, 3505 2014. 3506 184. Fauquet M, Grima B, Lamouroux A, Mallet J. Cloning of quail tyrosine 3507 hydroxylase: amino acid homology with other hydroxylases discloses functional 3508 domains. J Neurochem 50: 142-148, 1988.
- 3509 185. Felip A, Zanuy S, Pineda R, Pinilla L, Carrillo M, Tena-Sempere M, Gómez A.
- 3510 Evidence for two distinct KiSS genes in non-placental vertebrates that encode

kisspeptins with different gonadotropin-releasing activities in fish and mammals. *Mol Cell Endocrinol* 312: 61–71, 2009.

3513 186. Felix RC, Trindade M, Pires IRP, Fonseca VG, Martins RS, Silveira H, Power

- 3514 **DM**, **Cardoso JCR**. Unravelling the evolution of the allatostatin-type A, KISS and
- 3515 galanin peptide-receptor gene families in bilaterians: Insights from Anopheles
- 3516 mosquitoes. *PLoS One* 10: 1–30, 2015.
- 3517 187. Feng Y, Zhao H, An XF, Ma SL, Chen BY. Expression of brain prolactin releasing
- 3518 peptide (PrRP) changes in the estrous cycle of female rats. *Neurosci Lett* 419: 38–42,
- 3519 2007.
- 3520 188. Fernald RD, White RB. Gonadotropin-releasing hormone genes: Phylogeny,

3521 structure, and functions. *Front Neuroendocrinol* 20: 224–240, 1999.

- Fernlund P, Josefsson L. Crustacean color-change hormone: amino acid sequence and
 chemical synthesis. *Science (80-)* 177: 173–175, 1972.
- 3524 190. Fevold H, Hisaw F, Leonard S. The gonad-stimulating and the luteinizing hormones

of the anterior lobe of the hypophysis. *Am J Physiol* 97: 291–301, 1931.

- 3526 191. Fingerman F. Roles of neurotransmitters in regulating reproductive hormone release
- and gonadal maturation in decapod crustaceans. *Invert Reprod Dev* 31: 47–54, 1997.
- 3528 192. Fink G. MEMOIR: Harris' neuroendocrine revolution: Of portal vessels and self-
- 3529 priming. *J Endocrinol* 226: T13–T24, 2015.
- 3530 193. Di Fiore M, Rastogi R, Ceciliani F, Messi E, Botte V, Botte L, Pinelli C, D'Aniello
- **B**, **D'Aniello A**. Mammalian and chicken I forms of gonadotropin-releasing hormone
- in the gonads of a protochordate, Ciona intestinalis. Proc Natl Acad Sci USA 97: 2343–
- 3533 2348, 2000.
- 3534 194. Fiorentino M, Pinelli C, D'Aniello B, Iela L, Di Meglio M, Rastogi RK.
- 3535 Development and distribution of FMRFamide-like immunoreactivity in the toad (Bufo

- 3536 bufo) brain. *J Chem Neuroanat* 21: 201–213, 2001.
- 3537 195. Fitzpatrick PF. Tetrahydropterin-Dependent Amino Acid Hydroxylases. *Annu Rev*3538 *Biochem* 68: 355–381, 1999.
- 3539 196. Flanagan CA, Manilall A. Gonadotropin-releasing hormone (GnRH) receptor
- 3540 structure and GnRH binding. *Front Endocrinol (Lausanne)* 8: 1–14, 2017.
- 3541 197. Flügge G, Oertel WH, Wuttke W. Evidence for estrogen-receptive GABAergic
- neurons in the preoptic/anterior hypothalamic area of the rat brain.
- *Neuroendocrinology* **43**: 1–5, 1986.
- Fong PP, Ford AT. The biological effects of antidepressants on the molluscs and
 crustaceans: A review. *Aquat Toxicol* 151: 4–13, 2014.
- 3546 199. Fong PP, Noordhuis R, Ram JL. Dopamine reduces intensity of serotonin-induced
- spawning in the zebra mussel Dreissena polymorpha (Pallas). *J Exp Zool* 266: 79–83,
 1993.
- 3549 200. Fonnum F, Mariussen E. Mechanisms involved in the neurotoxic effects of
- 3550 environmental toxicants such as polychlorinated biphenyls and brominated flame
- 3551 retardants. *J Neurochem* 111: 1327–1347, 2009.
- 3552 201. Fontaine-Bertrand E, Marchelidon J, Salmon C, Fontaine Y. Les sous-unités des
- 3553 lutropines: une activité gonadotrope intrinsèque de α et le role de β dans le
- déterminisme de la spécificité zoologique d'action. *C R Seances Acad Sci III* 292: 507–
- 3555 510, 1981.
- 3556 202. Fontaine R, Affaticati P, Yamamoto K, Jolly C, Bureau C, Baloche S, Gonnet F,
- 3557 Vernier P, Dufour S, Pasqualini C. Dopamine inhibits reproduction in female
- 3558zebrafish (Danio rerio) via three pituitary D2 receptor subtypes. Endocrinology 154:
- 3559 807–18, 2013.
- 3560 203. Fontaine Y. Spécificités des glycoprotéines hypophysaires à activité thyréotrope et

- 3561 hypothèses sur leur phylogénie. *Bull Soc Chim Biol* 49: 647–655, 1967.
- 3562 204. Fontaine Y. La spécificité zoologique des protéines hypophysaires capables de
 3563 stimuler la thyroïde. *Acta Endocrinol* 136: 1–154, 1969.
- 3564 205. Forfar R, Lu ZL. Role of the transmembrane domain 4/extracellular loop 2 junction
- 3565 of the human gonadotropin-releasing hormone receptor in ligand binding and receptor
- 3566 conformational selection. *J Biol Chem* 286: 34617–34626, 2011.
- 3567 206. Fox KM, Dias JA, Van Roey P. Three-Dimensional Structure of Human Follicle3568 Stimulating Hormone. *Mol Endocrinol* 15: 378–389, 2001.
- 3569 207. Fraley GS, Coombs E, Gerometta E, Colton S, Sharp PJ, Li Q, Clarke IJ.
- 3570 Distribution and sequence of gonadotropin-inhibitory hormone and its potential role as
- a molecular link between feeding and reproductive systems in the Pekin duck (Anas

3572 platyrhynchos domestica). *Gen Comp Endocrinol* 184: 103–110, 2013.

- 3573 208. Fredriksson R. The G-Protein-Coupled Receptors in the Human Genome Form Five
- 3574 Main Families. Phylogenetic Analysis, Paralogon Groups, and Fingerprints. *Mol*
- 3575 *Pharmacol* 63: 1256–1272, 2003.
- 3576 209. Fryxell KJ. The evolutionary divergence of neurotransmitter receptors and second3577 messenger pathways. *J Mol Evol* 41: 85–97, 1995.
- 3578 210. Fujimoto M, Takeshita KI, Wang X, Takabatake I, Fujisawa Y, Teranishi H,
- 3579 Ohtani M, Muneoka Y, Ohta S. Isolation and characterization of a novel bioactive
- 3580 peptide, carassius RFamide (C-RFa), from the brain of the Japanese crucian carp.
- *Biochem Biophys Res Commun* 242: 436–440, 1998.
- 3582 211. Fujisawa T, Hayakawa E. Peptide signaling in Hydra. *Int J Dev Biol* 56: 543–550,
 3583 2012.
- 3584 212. Fukusumi S, Habata Y, Yoshida H, Iijima N, Kawamata Y, Hosoya M, Fujii R,
 3585 Hinuma S, Kitada C, Shintani Y, Suenaga M, Onda H, Nishimura O, Tanaka M,

3586		Ibata Y, Fujino M. Characteristics and distribution of endogenous RFamide-related
3587		peptide-1. Biochim Biophys Acta - Mol Cell Res 1540: 221–232, 2001.
3588	213.	Fukusumi S, Yoshida H, Fujii R, Maruyama M, Komatsu H, Habata Y, Shintani
3589		Y, Hinuma S, Fujino M. A New Peptidic Ligand and Its Receptor Regulating Adrenal
3590		Function in Rats. J Biol Chem 278: 46387-46395, 2003.
3591	214.	Funes S, Hedrick JA, Vassileva G, Markowitz L, Abbondanzo S, Golovko A,
3592		Yang S, Monsma FJ, Gustafson EL. The KiSS-1 receptor GPR54 is essential for the
3593		development of the murine reproductive system. Biochem Biophys Res Commun 312:
3594		1357–1363, 2003.
3595	215.	Fuxe K. Cellular localization of monoamines in the median eminence and in the
3596		infundibular stem of some mammals. Acta Physiol Scand 58: 383-384, 1963.
3597	216.	Gäde G. Peptides of the adipokinetic hormone/red pigment-concentrating hormone
3598		family: A new take on biodiversity. Ann N Y Acad Sci 1163: 125–136, 2009.
3599	217.	Gagné F, Blaise C. Effects of municipal effluents on serotonin and dopamine levels in
3600		the freshwater mussel Elliptio complanata. Comp Biochem Physiol Toxicol Pharmacol
3601		<i>CBP</i> 136: 117–125, 2003.
3602	218.	Gahete MD, Vázquez-Borrego MC, Martínez-Fuentes AJ, Tena-Sempere M,
3603		Castaño JP, Luque RM. Role of the Kiss1/Kiss1r system in the regulation of pituitary
3604		cell function. Mol Cell Endocrinol 438: 100–106, 2016.
3605	219.	Gaillard AL, Tay BH, Sirkin DIP, Lafont AG, De Flori C, Vissio PG, Mazan S,
3606		Dufour S, Venkatesh B, Tostivint H. Characterization of gonadotropin-releasing
3607		hormone (GnRH) genes from cartilaginous fish: Evolutionary perspectives. Front
3608		Neurosci 12: 1–14, 2018.
3609	220.	Gallardo WG, Hagiwara A, Snell TW. GABA enhances reproduction of the rotifer
3610		Brachionus plicatilis Muller: Application to mass culture. Aquac Res 31: 713–718,

3611 2000.

- 3612 221. Gallegos-Sánchez J, Malpaux B, Thiéry JC. Control of pulsatile LH secretion during
 3613 seasonal anoestrus in the ewe. *Reprod Nutr Dev* 38: 3–15, 1998.
- 3614 222. Gallo R V. Neuroendocrine regulation of pulsatile luteinizing hormone release in the
 3615 rat. *Neuroendocrinology* 30: 122–131, 1980.
- 3616 223. Gallo V, Accordi F, Chimenti C, Civinini A, Crivellato E. Catecholaminergic
- 3617 System of Invertebrates: Comparative and Evolutionary Aspects in Comparison With
 3618 the Octopaminergic System. *Int Rev Cell Mol Biol* 322: 363–394, 2016.
- 3619 224. Gammelsaeter R, Frøyland M, Aragón C, Danbolt NC, Fortin D, Storm-Mathisen
- 3620 J, Davanger S, Gundersen V. Glycine, GABA and their transporters in pancreatic
- 3621 islets of Langerhans: evidence for a paracrine transmitter interplay. *J Cell Sci* 117:

3622 3749–3758, 2004.

3623 225. García-Hernández MP, Koide Y, Díaz M V., Kawauchi H. Isolation and

3624 characterization of two distinct gonadotropins from the pituitary gland of

- 3625 Mediterranean yellowtail, Seriola dumerilii (Risso, 1810). *Gen Comp Endocrinol* 106:
 3626 389–399, 1997.
- 3627 226. Gen K, Okuzawa K, Senthilkumaran B, Tanaka H, Moriyama S, Kagawa H.

3628 Unique expression of gonadotropin-I and -II subunit genes in male and female red

3629 seabream (Pagrus major) during sexual maturation. *Biol Reprod* 63: 308–319, 2000.

- 3630 227. Gene M, Lee J, Miele ME, Hicks DJ, Karen K, Trent J, Weissman B, Welch DR.
- 3631 KiSS-1, a Novel Human Malignant Melanoma. *J Natl Cancer Inst* 88: 1731–1737,
 3632 1996.
- 3633 228. Gharaei A, Mahboudi F, Esmaili-Sari A, Edalat R, Adeli A, Keyvanshokooh S.
- 3634 Molecular cloning of cDNA of mammalian and chicken II gonadotropin-releasing
- 3635 hormones (mGnRHs and cGnRH-II) in the beluga (Huso huso) and the disruptive

- 3636 effect of methylmercury on gene expression. *Fish Physiol Biochem* 36: 803–817, 2010.
- 3637 229. Gibbs DM, Neill JD. Dopamine levels in hypophysial stalk blood in the rat are
- 3638 sufficient to inhibit prolactin secretion in vivo. *Endocrinology* 102: 1895–1900, 1978.
- 3639 230. Gibson EM, Humber SA, Jain S, Williams WP, Zhao S, Bentley GE, Tsutsui K,
- 3640 Kriegsfeld LJ. Alterations in RFamide-related peptide expression are coordinated with
- the preovulatory luteinizing hormone surge. *Endocrinology* 149: 4958–4969, 2008.
- 3642 231. Golan M, Martin AO, Mollard P, Levavi-Sivan B. Anatomical and functional
 3643 gonadotrope networks in the teleost pituitary. *Sci Rep* 6: 1–8, 2016.
- 3644 232. González-Martínez D, Madigou T, Zmora N, Anglade I, Zanuy S, Zohar Y, Elizur
- 3645 A, Muñoz-Cueto JA, Kah O. Differential Expression of Three Releasing Hormone)
- Messengers in the Brain of the European Sea Bass (Dicentrarchus labrax). J Comp
 Neurol 155: 144–155, 2001.
- 3648 233. González-Martínez D, Zmora N, Mañanos E, Saligaut D, Zanuy S, Zohar Y,
- 3649 Elizur A, Kah O, Muñoz-Cueto JA. Immunohistochemical localization of three
- 3650 different prepro-GnRHs in the brain and pituitary of the European sea bass
- 3651 (Dicentrarchus labrax) using antibodies to the corresponding GnRH-associated
- 3652 peptides. J Comp Neurol 446: 95–113, 2002.
- 3653 234. González-Martínez D, Zmora N, Saligaut D, Zanuy S, Elizur A, Kah O, Muñoz-
- 3654 Cueto JA. New insights in developmental origins of different GnRH (gonadotrophin-
- 3655 releasing hormone) systems in perciform fish: An immunohistochemical study in the
- European sea bass (Dicentrarchus labrax). *J Chem Neuroanat* 28: 1–15, 2004.
- 3657 235. Goodman RL, Jansen HT, Billings HJ, Coolen LM, Lehman MN. Neural systems
 3658 mediating seasonal breeding in the ewe. *J Neuroendocrinol* 22: 674–681, 2010.
- 3659 236. Gospocic J, Shields EJ, Glastad KM, Lin Y, Penick CA, Yan H, Mikheyev AS,
- 3660 Linksvayer TA, Garcia BA, Berger SL, Liebig J, Reinberg D, Bonasio R. The

3661 Neuropeptide Corazonin Controls Social Behavior and Caste Identity in Ants. *Cell*

3662 170: 748-759.e12, 2017.

- 3663 237. Gottsch ML, Cunningham MJ, Smith JT, Popa SM, Acohido B V., Crowley WF,
- 3664 **Seminara S, Clifton DK, Steiner RA**. A role for kisspeptins in the regulation of 3665 gonadotropin secretion in the mouse. *Endocrinology* 145: 4073–4077, 2004.
- 3666 238. Gou Z-H, Wang X, Wang W. Evolution of neurotransmitter gamma-aminobutyric
- acid, glutamate and their receptors. *Dong Wu Xue Yan Jiu = Zool Res* 33: E75-81,
 2012.
- 3669 239. Gougeon A. Human ovarian follicular development: from activation of resting follicles
 3670 to preovulatory maturation. *Ann Endocrinol* 71: 132–143, 2010.
- 3671 240. Graham KM, Langhorne C. J, Vance CK, Willard ST, Kouba AJ. Ultrasound
- 3672 imaging improves hormone therapy strategies for induction of ovulation and in vitro
 3673 fertilization in the endangered dusky gopher frog (Lithobates sevosa). 2018.
- 3674 241. Grattan D. 60 YEARS OF NEUROENDOCRINOLOGY: The hypothalamo-prolactin
 3675 axis. *J Endocrinol* 226: T101-122, 2015.
- 3676 242. De Greef WJ, Neill JD. Dopamine levels in hypophysial stalk plasma of the rat during
- 3677 surges of prolactin secretion induced by cervical stimulation. *Endocrinology* 105:
- 3678 1093–1099, 1979.
- 3679 243. Grenett HE, Ledley FD, Reed LL, Woo SL. Full-length cDNA for rabbit tryptophan
- 3680 hydroxylase: functional domains and evolution of aromatic amino acid hydroxylases.
- 3681 *Proc Natl Acad Sci U S A* 84: 5530–5534, 1987.
- 3682 244. Grima B, Lamouroux A, Blanot F, Biguet NF, Mallet J. Complete coding sequence
 3683 of rat tyrosine hydroxylase mRNA. *Proc Natl Acad Sci U S A* 82: 617–621, 1985.
- 3684 245. Grimmelikhuijzen CJP, Hauser F. Mini-review: The evolution of neuropeptide
 3685 signaling. *Regul Pept* 177: S6–S9, 2012.

3686 246. Grone BP, Maruska KP, Korzan WJ, Fernald RD. Social status regulates kisspeptin
3687 receptor mRNA in the brain of Astatotilapia burtoni. *Gen Comp Endocrinol* 169: 98–
3688 107, 2010.

- 3689 247. Grosse R, Schmid A, Schöneberg T, Herrlich A, Muhn P, Schultz G, Gudermann
- 3690T. Gonadotropin-releasing Hormone Receptor Initiates Multiple Signaling Pathways
- by Exclusively Coupling to Gq/11Proteins. *J Biol Chem* 275: 9193–9200, 2000.
- 3692 248. Guilgur LG, Moncaut NP, Canário AVM, Somoza GM. Evolution of GnRH ligands
 3693 and receptors in gnathostomata. *Comp Biochem Physiol A Mol Integr Physiol* 144:
- 3694 272–283, 2006.
- 3695 249. Guillou F, Combarnous Y. Purification of equine gonadotropins and comparative
 3696 study of their acid-dissociation and receptor-binding specificity. *BBA Gen Subj* 755:
 3697 229–236, 1983.
- 3698 250. Gundlah C, Alves S, Clark J, Pai L-Y, Schaeffer J, Rohrer S. Estrogen receptor-
- beta regulates tryptophan hydroxylase-1 expression in the murine midbrain raphe. *Biol Psychiatry* 57: 938–942, 2005.
- 3701 251. Guzmán J, Cal R, García-López A, Chereguini O, Kight K, Olmedo M,
- 3702 Sarasquete C, Mylonas C, Peleteiro J, Zohar Y, Mañanós E. Effects of in vivo
- 3703 treatment with the dopamine antagonist pimozide and gonadotropin-releasing hormone
- agonist (GnRHa) on the reproductive axis of Senegalese sole (Solea senegalensis).
- 3705 *Comp Biochem Physiol A Mol Integr Physiol* 158: 235–245, 2011.
- 3706 252. Gwee PC, Tay BH, Brenner S, Venkatesh B. Characterization of the
- 3707 neurohypophysial hormone gene loci in elephant shark and the Japanese lamprey:
- 3708 Origin of the vertebrate neurohypophysial hormone genes. *BMC Evol Biol* 9: 1–15,
- 3709 2009.
- 3710 253. Haas H, Panula P. The role of histamine and the tuberomamillary nucleus in the

- 3711 nervous system. *Nat Rev Neurosci* 4: 121–130, 2003.
- 3712 254. Haas H, Sergeeva O, Selbach O. Histamine in the nervous system. *Physiol Rev* 88:
 3713 1183–1241, 2008.
- 3714 255. Hagen C, McNatty K, McNeilly A. Immunoreactive α- and β-subunits of luteinizing
- hormone in human peripheral blood and follicular fluid throughout the menstrual cycle,
- and their effect on the secretion rate of progesterone by human granulosa cells in tissue
- 3717 culture. *J Endocrinol* 69: 33–46, 1976.
- 3718 256. El Halawani ME, Silsby JL, Fehrer SC, Behnke EJ. Reinitiation of ovulatory cycles
- 3719 in incubating female turkeys by an inhibitor of serotonin synthesis, P-
- 3720 chlorophenylalanine. *Biol Reprod* 28: 221–228, 1983.
- 3721 257. Hall TR, Cheung A, Harvey S. Serotoninergic inhibition of LH secretion in the
 3722 domestic fowl. *J Endocrinol* 110: 239–244, 1986.
- 3723 258. Hamdan FF, Ribeiro P. Cloning and characterization of a novel form of tyrosine
- 3724 hydroxylase from the human parasite, Schistosoma mansoni. *J Neurochem* 71: 1369–
 3725 1380, 1998.
- 3726 259. Han S-K, Abraham I, Herbison A. Effect of GABA on GnRH neurons switches from
- 3727 depolarization to hyperpolarization at puberty in the female mouse. *Endocrinology*
- 3728 143: 1459–1466, 2002.
- 3729 260. Han S-K, Todman MG, Herbison AE. Endogenous GABA release inhibits the firing
 3730 of adult gonadotropin-releasing hormone neurons. *Endocrinology* 145: 495–499, 2004.
- 3731 261. Hansen KK, Stafflinger E, Schneider M, Hauser F, Cazzamali G, Williamson M,
- 3732 Kollmann M, Schachtner J, Grimmelikhuijzen CJP. Discovery of a novel insect
- neuropeptide signaling system closely related to the insect adipokinetic hormone and
- 3734 corazonin hormonal systems. *J Biol Chem* 285: 10736–10747, 2010.
- 3735 262. Hargis B. Plasma prolactin and luteinizing hormone levels of female turkeys in

- 3736 response to intraventricular administration of monoaminergic neurotransmitter. *Poult*3737 *Sci* 1661, 1982.
- 3738 263. Harris KD, Zahavi A. The evolution of ACh and GABA as neurotransmitters: a
 3739 hypothesis. *Med Hypotheses* 81: 760–762, 2013.
- 3740264.Hasunuma I, Terakado K. Two Novel Gonadotropin-Releasing Hormones (GnRHs)
- 3741 from the Urochordate Ascidian, *Halocynthia roretzi* : Implications for the Origin of

3742 Vertebrate GnRH Isoforms. *Zoolog Sci* 30: 311–318, 2013.

- 3743 265. Hauser F, Grimmelikhuijzen CJP. Evolution of the AKH/corazonin/ACP/GnRH
- 3744 receptor superfamily and their ligands in the Protostomia. *Gen Comp Endocrinol* 209:
 3745 35–49, 2014.
- 3746 266. Hauser F, Neupert S, Williamson M, Predel R, Tanaka Y, Grimmelikhuijzen C.
- 3747 Genomics and peptidomics of neuropeptides and protein hormones present in the 3748 parasitic wasp Nasonia vitripennis. *J Proteome Res* 9: 5296–5310, 2010.
- 3749 267. Hauser F, Sondergaard L, Grimmelikhuijzen CJP. Molecular cloning, genomic
- 3750 organization and developmental regulation of a novel receptor from Drosophila
- melanogaster structurally related to gonadotropin-releasing hormone receptors from
 vertebrates. *Biochem Biophys Res Commun* 249: 822–828, 1998.
- 3753 268. Hay-Schmidt A. The evolution of the serotonergic nervous system. *Proceedings Biol*3754 Sci 267: 1071–1079, 2000.
- 3755 269. Hayes W, Wray S, Battey J. The frog gonadotropin-releasing hormone-I (GnRH-I)
- 3756 gene has a mammalian-like expression pattern and conserved domains in GnRH-
- associated peptide, but brain onset is delayed until metamorphosis. *Endocrinology* 134:
- 3758 1835–1845, 1994.
- 3759 270. Hays L, Carpenter C, Petersen S. Evidence that GABAergic neurons in the preoptic
 3760 area of the rat brain are targets of 2,3,7,8-tetrachlorodibenzo-p-dioxin during

3761		development. Environ Health Perspect 110 Suppl: 369-376, 2002.
3762	271.	Hearn M, Gomme P. Molecular architecture and biorecognition processes of the
3763		cystine knot protein superfamily: part I. The glycoprotein hormones. J Mol Recognit
3764		13: 223–278, 2000.
3765	272.	Henke A, Gromoll J. New insights into the evolution of chorionic gonadotrophin. Mol
3766		<i>Cell Endocrinol</i> 291: 11–19, 2008.
3767	273.	Henkel C V., Burgerhout E, de Wijze DL, Dirks RP, Minegishi Y, Jansen HJ,
3768		Spaink HP, Dufour S, Weltzien FA, Tsukamoto K, van den Thillart GEEJM.
3769		Primitive duplicate hox clusters in the european eel's genome. PLoS One 7, 2012.
3770	274.	Henkel C V., Dirks RP, de Wijze DL, Minegishi Y, Aoyama J, Jansen HJ, Turner
3771		B, Knudsen B, Bundgaard M, Hvam KL, Boetzer M, Pirovano W, Weltzien FA,
3772		Dufour S, Tsukamoto K, Spaink HP, van den Thillart GEEJM. First draft genome
3773		sequence of the Japanese eel, Anguilla japonica. Gene 511: 195–201, 2012.
3774	275.	Herbison AE. Noradrenergic regulation of cyclic GnRH secretion. Rev Reprod 2: 1-6,
3775		1997.
3776	276.	Herbison AE, Chapman C, Dyer RG. Role of medial preoptic GABA neurones in
3777		regulating luteinising hormone secretion in the ovariectomised rat. Exp Brain Res 87:
3778		345–352, 1991.
3779	277.	Herbison AE, Dyer RG. Effect on luteinizing hormone secretion of GABA receptor
3780		modulation in the medial preoptic area at the time of proestrous luteinizing hormone
3781		surge. Neuroendocrinology 53: 317–320, 1991.
3782	278.	Heuer CM, Loesel R. Immunofluorescence analysis of the internal brain anatomy of
3783		Nereis diversicolor (Polychaeta, Annelida). Cell Tissue Res 331: 713-724, 2008.
3784	279.	Van Hiel MB, Vandersmissen HP, Van Loy T, Vanden Broeck J. An evolutionary
3785		comparison of leucine-rich repeat containing G protein-coupled receptors reveals a

- 3786 novel LGR subtype. *Peptides* 34: 193–200, 2012.
- 3787 280. Hinuma S, Habata Y, Fujii R, Kawamata Y, Hosoya M, Fukusumi S, Kitada C,

3788 Masuo Y, Asano T, Matsumoto H, Sekiguchi M, Kurokawa T, Nishimura O,

- 3789 Onda H, Fujino M. A prolactin-releasing peptide in the brain. *Nature* 393: 272–276,
 3790 1998.
- 3791 281. Hinuma S, Shintani Y, Fukusumi S, Iijima N, Matsumoto Y, Hosoya M, Fujii R,
- Watanabe T, Et A. New neuropeptides containing carboxy-terminal RFamide and
 their receptor in mammals. *Nat Cell Biol* 2: 703–708, 2000.
- 3794 282. Hirai S, Kishimoto T, Kadam AL, Kanatani H, Koide SS. Induction of spawning
- and oocyte maturation by 5-hydroxytryptamine in the surf clam. *J Exp Zool* 245: 318–
 3796 321, 1988.
- 3797 283. Hizume T, Watanobe H, Yoneda M, Suda T, Schiöth HB. Involvement of prolactin3798 releasing peptide in the preovulatory luteinizing hormone and prolactin surges in the
 3799 rat. *Biochem Biophys Res Commun* 279: 35–39, 2000.
- 3800 284. Holland LZ, Ocampo Daza D. A new look at an old question : when did the second
- 3801 whole genome duplication occur in vertebrate evolution ? *Genome Biol* 19: 2–5, 2018.
- 3802 285. Holland P. The Animal Kingdom: A Very Short Introduction. Oxford Uni. Oxford:
- 3803 2011.
- 3804 286. Holland PW, Garcia-Fernàndez J, Williams NA, Sidow A. Gene duplications and
 3805 the origins of vertebrate development. *Dev. Suppl.*.
- 3806 287. Holmes R, Ball J. *The pituitary gland a comparative account*. Cambridge University
 3807 Press, 1974.
- 3808 288. Honma T, Miyagawa M, Suda M, Wang R-S, Kobayashi K, Sekiguchi S. Effects of
- 3809 perinatal exposure to bisphenol A on brain neurotransmitters in female rat offspring.
- *Ind Health* 44: 510–524, 2006.

3811	289.	Honma T, Suda M, Miyagawa M, Wang R-S, Kobayashi K, Sekiguchi S.
3812		Alteration of brain neurotransmitters in female rat offspring induced by prenatal
3813		administration of 16 and 64 mg/kg of 2,2',4,4',5,5'-hexachlorobiphenyl (PCB153). Ind
3814		<i>Health</i> 47: 11–21, 2009.
3815	290.	Hori A, Honda S, Asada M, Ohtaki T, Oda K, Watanabe T, Shintani Y, Yamada
3816		T, Suenaga M, Kitada C, Onda H, Kurokawa T, Nishimura O, Fujino M. Metastin
3817		suppresses the motility and growth of CHO cells transfected with its receptor. Biochem
3818		Biophys Res Commun 286: 958–963, 2001.
3819	291.	Hornykiewicz O. [Dopamine (3-hydroxytyramine) in the central nervous system and
3820		its relation to the Parkinson syndrome in man]. Dtsch Med Wochenschr 87: 1807-
3821		1810, 1962.
3822	292.	Hsu MC, Wang JY, Lee YJ, Jong DS, Tsui KH, Chiu CH. Kisspeptin modulates
3823		fertilization capacity of mouse spermatozoa. Reproduction 147: 835-845, 2014.
3824	293.	Hsu S, Nakabayashi K, Bhalla A. Evolution of glycoprotein hormone subunit genes
3825		in bilateria: identification of two novel human glycoprotein hormone subunit family
3826		genes, GPA2 and GPB5. Mol Endocrinol 16: 1538–1551, 2002.
3827	294.	Hu Y, Shi D, Luo Q, Liu Q, Zhou Y, Liu L, Yu L, Wei W, Shen J. Cloning and
3828		characterization of a novel enzyme: tyrosine hydroxylase from Schistosoma
3829		japonicum. Parasitol Res 109: 1065-1074, 2011.
3830	295.	Hua W, Luo L, Tian Y, Song M, Liu Y, Cui P, Song S, Jiang S, Li F, Fang F.
3831		Analysis of the serum concentrations of kisspeptin and neurokinin B in the geese
3832		during reproductive cycle and their localisation in the ovary. Anim Reprod Sci 151: 78-
3833		84, 2014.
3834	296.	Hughes AL. Phylogenies of developmentally important proteins do not support the
3835		hypothesis of two rounds of genome duplication early in vertebrate history. J Mol Evol

3836 48: 565–576, 1999.

- 3837 297. Huhtaniemi I. A short evolutionary history of FSH-stimulated spermatogenesis.
 3838 *Hormones* 14: 468–478, 2015.
- 3839 298. Hultqvist G, Ocampo Daza D, Larhammar D, Kilimann MW. Evolution of the
- vertebrate paralemmin gene family: Ancient origin of gene duplicates suggests distinct
 functions. *PLoS One* 7, 2012.
- 3842 299. Huseman CA, Kugler JA, Schneider IG. Mechanism of dopaminergic suppression of
 3843 gonadotropin secretion in men. *J Clin Endocrinol Metab* 51: 209–214, 1980.
- 3844 300. Hwang SH, Cho S, Park YC. cDNA cloning and induction of tyrosine hydroxylase
- 3845 gene from the diamondback moth, Plutella xylostella. *Arch Insect Biochem Physiol* 75:
 3846 107–120, 2010.
- 3847 301. Ichikawa S, Nasrin S, Nagatsu T. Expression of mouse tyrosine hydroxylase in
 3848 Escherichia coli. *Biochem Biophys Res Commun* 178: 664–671, 1991.
- 3849 302. Ikemoto T, Enomoto M, Park MK. Identification and characterization of a reptilian
- 3850 GnRH receptor from the leopard gecko. *Mol Cell Endocrinol* 214: 137–147, 2004.
- 303. Iles RK, Delves PJ, Butler SA. Does hCG or hCGβ play a role in cancer cell biology?
 3852 Mol Cell Endocrinol 329: 62–70, 2010.
- 3853 304. Illing N, Jacobs G, Becker I, Flanagan C, Davidson J, Eales A, Zhou W, Sealfon S,
- 3854 Millar R. Comparative sequence analysis and functional characterization of the cloned
- 3855 sheep gonadotropin-releasing hormone receptor reveal differences in primary structure
- and ligand specificity among mammalian receptors. *Biochem Biophys Res Commun*
- 3857 196: 745–751, 1993.
- 3858 305. Illing N, Troskie BE, Nahorniak CS, Hapgood JP, Peter RE, Millar RP. Two
- 3859 gonadotropin-releasing hormone receptor subtypes with distinct ligand selectivity and
- 3860 differential distribution in brain and pituitary in the goldfish (Carassius auratus). *Proc*

- 3861 *Natl Acad Sci U S A* 96: 2526–2531, 1999.
- 3862 306. Irfan S, Ehmcke J, Shahab M, Wistuba J, Schlatt S. Immunocytochemical
 3863 localization of kisspeptin and kisspeptin receptor in the primate testis. *J Med Primatol*
- 386445: 105–111, 2016.
- 3865 307. Irwig M, Fraley G, Smith J, Acohido B, Popa S, Cunningham M, Gottsch M,
- 3866 Clifton D, Steiner R. Kisspeptin activation of gonadotropin releasing hormone
- neurons and regulation of KiSS-1 mRNA in the male rat. *Neuroendocrinology* 80:
 264–272, 2004.
- 3869 308. Isabel G, Gourdoux L, Moreau R. Changes of biogenic amine levels in haemolymph
- 3870 during diapausing and non-diapausing status in Pieris brassicae L. Comp Biochem
- 3871 *Physiol A Mol Integr Physiol* 128: 117–127, 2001.
- 3872 309. Ishii S. Gonadotropins. In: *Vertebrate Endocrinology: Fundamentals and Biochemical* 3873 *Implications*, edited by Pang P, Schreibman M. New York: 1991, p. 33–66.
- 3874 310. Isomura N, Yamauchi C, Takeuchi Y, Takemura A. Does dopamine block the
 3875 spawning of the acroporid coral Acropora tenuis? *Sci Rep* 3: 10–13, 2013.
- 3876 311. Iversen SD, Iversen LL. Dopamine: 50 years in perspective. *Trends Neurosci* 30:
 3877 188–193, 2007.
- 3878 312. Iwakoshi-Ukena E, Ukena K, Takuwa-Kuroda K, Kanda A, Tsutsui K, Minakata
- 3879 **H**. Expression and distribution of octopus gonadotropin-releasing hormone in the
- 3880 central nervous system and peripheral organs of the octopus (Octopus vulgaris) by in
- 3881 situ hybridization and immunohistochemistry. *J Comp Neurol* 477: 310–323, 2004.
- 3882 313. Iwakoshi E, Takuwa-Kuroda K, Fujisawa Y, Hisada M, Minakata H, Ukena K,
- **Tsutsui K**. Isolation and characterization of a GnRH-like peptide from Octopus
- 3884 vulgaris. *Biochem Biophys Res Commun* 291: 1187–1193, 2002.
- 3885 314. Jaber M, Robinson SW, Missale C, Caron MG. Dopamine receptors and brain

3886		function. Neuropharmacology 35: 1503–1519, 1996.
3887	315.	Jadhao AG, Pinelli C, D'Aniello B, Tsutsui K. Gonadotropin-inhibitory hormone
3888		(GnIH) in the amphibian brain and its relationship with the gonadotropin releasing
3889		hormone (GnRH) system: An overview. Gen Comp Endocrinol 240: 69-76, 2017.
3890	316.	Jaillon O, Aury J-M, Brunet F, Petit J-L, Stange-Thomann N, Mauceli E,
3891		Bouneau L, Fischer C, Ozouf-Costaz C, Bernot A, Nicaud S, Jaffe D, Fisher S,
3892		Lutfalla G, Dossat C, Segurens B, Dasilva C, Salanoubat M, Levy M, Boudet N,
3893		Castellano S, Anthouard V, Jubin C, Castelli V, Katinka M, Vacherie B, Biémont
3894		C, Skalli Z, Cattolico L, Poulain J, De Berardinis V, Cruaud C, Duprat S, Brottier
3895		P, Coutanceau J-P, Gouzy J, Parra G, Lardier G, Chapple C, McKernan KJ,
3896		McEwan P, Bosak S, Kellis M, Volff J-N, Guigó R, Zody MC, Mesirov J,
3897		Lindblad-Toh K, Birren B, Nusbaum C, Kahn D, Robinson-Rechavi M, Laudet V,
3898		Schachter V, Quétier F, Saurin W, Scarpelli C, Wincker P, Lander ES,
3899		Weissenbach J, Roest Crollius H. Genome duplication in the teleost fish Tetraodon
3900		nigroviridis reveals the early vertebrate proto-karyotype. Nature 431: 946-957, 2004.
3901	317.	Jarry H, Leonhardt S, Schwarze T, Wuttke W. Preoptic rather than mediobasal
3902		hypothalamic amino acid neurotransmitter release regulates GnRH secretion during the
3903		estrogen-induced LH surge in the ovariectomized rat. Neuroendocrinology 62: 479-
3904		486, 1995.
3905	318.	Jekely G. Global view of the evolution and diversity of metazoan neuropeptide
3906		signaling. Proc Natl Acad Sci 110: 8702-8707, 2013.
3907	319.	Jeng S-R, Yueh W-S, Pen Y-T, Lee Y-H, Chen G-R, Dufour S, Chang C-F.
3908		Neuroendocrine gene expression reveals a decrease in dopamine D2B receptor with no
3909		changes in GnRH system during prepubertal metamorphosis of silvering in wild
3910		Japanese eel. Gen Comp Endocrinol 206: 8–15, 2014.

- 3911 320. Jennes L, Conn PM. Gonadotropin-releasing hormone and its receptors in rat brain.
 3912 *Front. Neuroendocrinol.* 15: 51–77, 1994.
- 3913 321. Jéquier E, Lovenberg W, Sjoerdsma A. Tryptophan hydroxylase inhibition: the
- 3914 mechanism by which p-chlorophenylalanine depletes rat brain serotonin. *Mol*
- *Pharmacol* 3: 274–278, 1967.
- 3916 322. Jiang Y, Luo L, Gustafson EL, Yadav D, Laverty M, Murgolo N, Vassileva G,
- 3917 Zeng M, Laz TM, Behan J, Qiu P, Wang L, Wang S, Bayne M, Greene J, Monsma
- 3918 F, Zhang FL. Identification and characterization of a novel RF-amide peptide ligand
- for orphan G-protein-coupled receptor SP9155. *J Biol Chem* 278: 27652–27657, 2003.
- 3920 323. Jimenez-Liñan M, Rubin BS, King JC. Examination of guinea pig luteinizing
- hormonereleasing hormone gene reveals a unique decapeptide and existence of two
 transcripts in the brain. *Endocrinology* 138: 4123–4130, 1997.
- 3923 324. Johnson JI, Kavanaugh SI, Nguyen C, Tsai PS. Localization and functional
- characterization of a novel adipokinetic hormone in the mollusk, Aplysia californica. *PLoS One* 9, 2014.
- 3926 325. Johnson MA, Tsutsui K, Fraley GS. Rat RFamide-related peptide-3 stimulates GH
- 3927 secretion, inhibits LH secretion, and has variable effects on sex behavior in the adult
 3928 male rat. *Horm Behav* 51: 171–180, 2007.
- 3929 326. Jolly C, Rousseau K, Prézeau L, Vol C, Tomkiewicz J, Dufour S, Pasqualini C.
- 3930 Functional Characterisation of Eel Dopamine D2 Receptors and Involvement in the
- 3931 Direct Inhibition of Pituitary Gonadotrophins. *J Neuroendocrinol* 28, 2016.
- 3932 327. Jones G, Gey G, Gey M. Hormone production by placental cells maintained in
 3933 continuous cultures. *Bull Johns Hopskins Hosp* 72: 26, 1943.
- 3934 328. De Jong-Brink M, Ter Maat A, Tensen CP. NPY in invertebrates: Molecular
- answers to altered functions during evolution. *Peptides* 22: 309–315, 2001.

3936	329.	Josefsson LG. Evidence for kinship between diverse G-protein coupled receptors.
3937		Gene 239: 333–340, 1999.
3938	330.	Joseph NT, Aquilina-Beck A, MacDonald C, Decatur WA, Hall JA, Kavanaugh
3939		SI, Sower SA. Molecular cloning and pharmacological characterization of two novel
3940		GnRH receptors in the lamprey (Petromyzon marinus). Endocrinology 153: 3345-
3941		3356, 2012.
3942	331.	Justo. Effect of serotoninergic system on FSH secretion in male and female rats:
3943		evidence for stimulatory and inhibitory actions. Neuroendocrinology 50: 382-386,
3944		1989.
3945	332.	Kadokawa H, Shibata M, Tanaka Y, Kojima T, Matsumoto K, Oshima K,
3946		Yamamoto N. Bovine C-terminal octapeptide of RFamide-related peptide-3
3947		suppresses luteinizing hormone (LH) secretion from the pituitary as well as pulsatile
3948		LH secretion in bovines. Domest Anim Endocrinol 36: 219–224, 2009.
3949	333.	Kah O, Chambolle P, Thibault J, Geffard M. Existence of dopaminergic neurons in
3950		the preoptic region of the goldfish. Neurosci Lett 48: 293–298, 1984.
3951	334.	Kah O, Dubourg P, Martinoli MG, Geffard M, Calas A. Morphological evidence
3952		for a direct neuroendocrine GABAergic control of the anterior pituitary in teleosts.
3953		Experientia 43: 300–302, 1987.
3954	335.	Kah O, Dubourg P, Martinoli MG, Rabhi M, Gonnet F, Geffard M, Calas A.
3955		Central GABAergic innervation of the pituitary in goldfish: a radioautographic and
3956		immunocytochemical study at the electron microscope level. Gen Comp Endocrinol
3957		67: 324–332, 1987.
3958	336.	Kah O, Dulka JG, Dubourg P, Thibault J, Peter RE. Neuroanatomical substrate for
3959		the inhibition of gonadotrophin secretion in goldfish: existence of a dopaminergic
3960		preoptico-hypophyseal pathway. Neuroendocrinology 45: 451-458, 1987.

- 3961 337. Kah O, Lethimonier C, Somoza G, Guilgur LG, Vaillant C, Larevre JJ. GnRH and
- 3962 GnRH receptors in metazoa: A historical, comparative, and evolutive perspective. *Gen*3963 *Comp Endocrinol* 153: 346–364, 2007.
- 3964 338. Kah O, Trudeau VL, Sloley BD, Chang JP, Dubourg P, Yu KL, Peter RE.
- Influence of GABA on gonadotrophin release in the goldfish. *Neuroendocrinology* 55:
 3966 396–404, 1992.
- 3967 339. Kalra SP, Kalra PS. Neural regulation of luteinizing hormone secretion in the rat.
 3968 *Endocr Rev* 4: 311–351, 1983.
- 3969 340. Kamberi IA, Mical RS, Porter JC. Prolactin-inhibiting activity in hypophysial stalk

blood and elevation by dopamine. *Experientia* 26: 1150–1151, 1970.

- 3971 341. Kamei H, Kawazoe I, Kaneko T, Aida K. Purification of follicle-stimulating
- 3972 hormone from immature Japanese eel, Anguilla japonica, and its biochemical
- 3973 properties and steroidogenic activities. *Gen Comp Endocrinol* 143: 257–266, 2005.
- 3974 342. Kamiya C, Ohta N, Ogura Y, Yoshida K, Horie T, Kusakabe TG, Satake H,
- 3975 Sasakura Y. Nonreproductive role of gonadotropin-releasing hormone in the control
- 3976 of ascidian metamorphosis. *Dev Dyn* 243: 1524–1535, 2014.
- 3977 343. Kanasaki H, Oride A, Mijiddorj T, Sukhbaatar U, Kyo S. How is GnRH regulated
- in GnRH-producing neurons? Studies using GT1-7 cells as a GnRH-producing cell
- 3979 model. *Gen Comp Endocrinol* 247: 138–142, 2017.
- 3980 344. Kanda A, Takahashi T, Satake H, Minakata H. Molecular and functional
- 3981 characterization of a novel gonadotropin-releasing-hormone receptor isolated from the
- 3982 common octopus (Octopus vulgaris). *Biochem J* 395: 125–135, 2006.
- 3983 345. Kaslin J, Panula P. Comparative anatomy of the histaminergic and other aminergic
 3984 systems in zebrafish (Danio rerio). *J Comp Neurol* 440: 342–377, 2001.
- 3985 346. Kass-Simon G, Pierobon P. Cnidarian chemical neurotransmission, an updated

3986		overview. Comp Biochem Physiol A Mol Integr Physiol 146: 9-25, 2007.
3987	347.	Kasten TL, White SA, Norton TT, Bond CT, Adelman JP, Fernald RD.
3988		Characterization of two new preproGnRH mRNAs in the tree shrew: First direct
3989		evidence for mesencephalic GnRH gene expression in a placental mammal. Gen Comp
3990		Endocrinol 104: 7–19, 1996.
3991	348.	Kavanaugh S, Tsai P. Functional Authentication of a Novel Gastropod Gonadotropin-
3992		Releasing Hormone Receptor Reveals Unusual Features and Evolutionary Insight.
3993		<i>PLoS One</i> 11: e0160292, 2016.
3994	349.	Kavanaugh SI, Nozaki M, Sower SA. Origins of gonadotropin-releasing hormone
3995		(GnRH) in vertebrates: Identification of a novel GnRH in a basal vertebrate, the sea
3996		lamprey. Endocrinology 149: 3860-3869, 2008.
3997	350.	Kawano E, Takahata Y, Oishi T, Ukena K, Tsutsui K, Tamotsu S. Neural
3998		Interaction of Gonadotropin-regulating Hormone Immunoreactive Neurons and the
3999		Suprachiasmatic Nucleus with the Paraventricular Organ in the Japanese Grass Lizard
4000		(Takydromus tachydromoides). Zoolog Sci 23: 277-287, 2006.
4001	351.	Kebabian JW, Calne DB. Multiple receptors for dopamine. Nature 277: 93–96, 1979.
4002	352.	Kelly MA, Rubinstein M, Asa SL, Zhang G, Saez C, Bunzow JR, Allen RG,
4003		Hnasko R, Ben-Jonathan N, Grandy DK, Low MJ. Pituitary lactotroph hyperplasia
4004		and chronic hyperprolactinemia in dopamine D2 receptor-deficient mice. Neuron 19:
4005		103–113, 1997.
4006	353.	Khan IA, Thomas P. Stimulatory effects of serotonin on maturational gonadotropin
4007		release in the Atlantic croaker, Micropogonias undulatus. Gen Comp Endocrinol 88:
4008		388–396, 1992.
4009	354.	Khan IA, Thomas P. Immunocytochemical localization of serotonin and
4010		gonadotropin-releasing hormone in the brain and pituitary gland of the Atlantic croaker

4011		Micropogonias undulatus. Gen Comp Endocrinol 91: 167–180, 1993.
4012	355.	Khan IA, Thomas P. Seasonal and daily variations in the plasma gonadotropin II
4013		response to a LHRH analog and serotonin in Atlantic croaker (Micropogonias
4014		undulatus): evidence for mediation by 5-HT2 receptors. J Exp Zool 269: 531-537,
4015		1994.
4016	356.	Khan IA, Thomas P. GABA exerts stimulatory and inhibitory influences on
4017		gonadotropin II secretion in the Atlantic croaker (Micropogonias undulatus).
4018		Neuroendocrinology 69: 261–268, 1999.
4019	357.	Khan IA, Thomas P. Disruption of neuroendocrine control of luteinizing hormone
4020		secretion by aroclor 1254 involves inhibition of hypothalamic tryptophan hydroxylase
4021		activity. Biol Reprod 64: 955–964, 2001.
4022	358.	Khotimchenko Y. Effect of noradrenaline, dopamine and adrenolytics on growth and
4023		maturation of the sea urchin, Strongylocentrotus nudus Agassiz. Int J Invert Rep Dev 4:
4024		369–373, 1982.
4025	359.	Kim D-K, Yun S, Hoon G, Hwang J-I, Park C, Kim J, Kim K, Vaudry H, Seong J.
4026		Coevolution of the spexin/galanin/kisspeptin family: spexin activates galanin receptor
4027		type II and III. <i>Endocrinology</i> 155: 1864–1873, 2014.
4028	360.	Kim DK, Cho EB, Moon MJ, Park S, Hwang JI, Kah O, Sower SA, Vaudry H,
4029		Seong JY. Revisiting the evolution of gonadotropin-releasing hormones and their
4030		receptors in vertebrates: Secrets hidden in genomes. Gen Comp Endocrinol 170: 68-
4031		78, 2011.
4032	361.	Kim DK, Cho EB, Moon MJ, Park S, Hwang JI, Rego JL Do, Vaudry H, Seong
4033		JY. Molecular coevolution of neuropeptides gonadotropin-releasing hormone and
4034		kisspeptin with their cognate G protein-coupled receptors. Front Neurosci 6: 1-8,
4035		2012.

4036 362. Kim Y-J, Spalovska-Valachova I, Cho K-H, Zitnanova I, Park Y, Adams M,

- 4037 Zitnan D. Corazonin receptor signaling in ecdysis initiation. *Proc Natl Acad Sci USA*4038 101: 6704–6709, 2004.
- 4039 363. Kimura F, Jinnai K. Bicuculline infusions advance the timing of luteinizing hormone
 4040 surge in proestrous rats: comparisons with naloxone effects. *Horm Behav* 28: 424–430,
- 4041 1994.
- 4042 364. King JA, Millar RP. Structure of Chicken Hypothalamic Luteinizing Hormone4043 releasing Hormone. II. Isolation and characterization. *J Biol Chem* 257: 10729–10782,
- 4044 1982.
- 4045 365. King JA, Millar RP. Structure of Chicken Hypothalamic Luteinizing Hormone-
- 4046 releasing Hormone. I. Structural detremination on partially purified material. *J Biol*4047 *Chem* 257: 10722–10728, 1982.
- 4048 366. Kitahashi T, Ogawa S, Parhar IS. Cloning and expression of kiss2 in the zebrafish
 4049 and medaka. *Endocrinology* 150: 821–831, 2009.
- 4050 367. Klavdieva M. The History of Neuropeptides II Dicovery of hypothalamic
- 4051 neurohormones, elucidation of the primary structure of substance P and further studies

4052 on neurosecretion. *Front Neuroendocrinol* 17: 126–153, 1996.

- 4053 368. Klouche MS, De Deurwaerdère P, Dellu-Hagedorn F, Lakhdar-Ghazal N,
- **Benomar S**. Monoamine content during the reproductive cycle of Perna perna depends
- 4055 on site of origin on the Atlantic Coast of Morocco. *Sci Rep* 5: 13715, 2015.
- 4056 369. Knobil E. The neuroendocrine control of the menstrual cycle. *Recent Prog Horm Res*4057 36: 53–88, 1980.
- 4058 370. Knobil E, Neill J. The physiology of reproduction. Academic P. 2015.
- 4059 371. Kobayashi M, Nakano R, Ooshima A. Immunocytochemical localization of pituitary
- 4060 gonadotrophins and gonadal steroids confirms the "two-cell, two-gonadotrophin"

- 4061 hypothesis of steroidogenesis in the human ovary. *J Endocrinol* 126: 483–488, 1990.
- 4062 372. Koda A, Ukena K, Teranishi H, Ohta S, Yamamoto K, Kikuyama S, Tsutsui K. A
- 4063 novel amphibian hypothalamic neuropeptide: Isolation, localization, and biological
 4064 activity. *Endocrinology* 143: 411–419, 2002.
- 4065 373. Koide Y, Itoh H, Kawauchi H. Isolation and characterization of two distinct
 4066 gonadotropins, GTHI and GTHII, from bonito (Katsuwonus plelamis) pituitary glands.
- 4067 *Int J Pept Protein Res* 41: 52–65, 1993.
- 4068 374. Kosonsiriluk S, Chaiworakul V, Mauro LJ, El Halawani ME. Enhanced
- 4069 GABAergic inhibition in the premammillary nucleus of photorefractory turkey hens
- 4070 via GABAA receptor upregulation. *Gen Comp Endocrinol* 230–231: 57–66, 2016.
- 4071 375. Kotani M, Detheux M, Vandenbogaerde A, Communi D, Vanderwinden JM, Le
- 4072 Poul E, Brézillon S, Tyldesley R, Suarez-Huerta N, Vandeput F, Blanpain C,
- 4073 Schiffmann SN, Vassart G, Parmentier M. The Metastasis Suppressor Gene KiSS-1
- 4074 Encodes Kisspeptins, the Natural Ligands of the Orphan G Protein-coupled Receptor
 4075 GPR54. *J Biol Chem* 276: 34631–34636, 2001.
- 4076 376. Krey LC, Butler WR, Knobil E. Surgical disconnection of the medial basal
- 4077 hypothalamus and pituitary function in the rhesus monkey. I. Gonadotropin secretion.
- 4078 *Endocrinology* 96: 1073–1087, 1975.
- 4079 377. Kriegsfeld LJ, Mei DF, Bentley GE, Ubuka T, Mason AO, Inoue K, Ukena K,
- 4080 **Tsutsui K**, **Silver R**. Identification and characterization of a gonadotropin-inhibitory
- 4081 system in the brains of mammals. *Proc Natl Acad Sci* 103: 2410–2415, 2006.
- 4082 378. Kriegsfeld LJ, Ubuka T, Bentley GE, Tsutsui K. Seasonal control of gonadotropin4083 inhibitory hormone (GnIH) in birds and mammals. *Front Neuroendocrinol* 37: 65–75,
- 4084 2015.
- 4085 379. Kuljis RO, Advis JP. Immunocytochemical and physiological evidence of a synapse

- 4086 between dopamine- and luteinizing hormone releasing hormone-containing neurons in
 4087 the ewe median eminence. *Endocrinology* 124: 1579–1581, 1989.
- 4088 380. Kumar TR, Wang Y, Lu N, Matzuk MM. Follicle stimulating hormone is required
 4089 for ovarian follicle maturation but not male fertility. *Nat Genet* 15: 201–204, 1997.
- 4090 381. Kuo MW, Lou SW, Postlethwait J, Chung BC. Chromosomal organization,
- 4091 evolutionary relationship, and expression of zebrafish GnRH family members. *J*

4092 *Biomed Sci* 12: 629–639, 2005.

- 4093 382. De la LLosa P, Jutisz M. Reversible dissociation into two subunits and biological
 4094 activity of ovine luteinizing hormone. *Biochem Biophys Acta* 181: 426–436, 1969.
- 4095 383. Labhsetwar AP. Role of monoamines in ovulation: evidence for a serotoninergic
- 4096 pathway for inhibition of spontaneous ovulation. *J Endocrinol* 54: 269–275, 1972.
- 4097 384. Lacau-Mengido IM, Becú-Villalobos D, Thyssen SM, Rey EB, Lux-Lantos VA,
- 4098Libertun C. Antidopaminergic-induced hypothalamic LHRH release and pituitary4099gonadotrophin secretion in 12 day-old female and male rats. J Neuroendocrinol 5:
- 4100 705–709, 1993.
- 4101 385. Lagerström MC, Fredriksson R, Bjarnadóttir TK, Fridmanis D, Holmquist T,
- 4102 Andersson J, Yan YL, Raudsepp T, Zoorob R, Kukkonen JP, Lundin LG, Klovins
- 4103 J, Chowdhary BP, Postlethwait JH, Schiöth HB. Origin of the prolactin-releasing
- 4104 hormone (PRLH) receptors: Evidence of coevolution between PRLH and a redundant
- 4105 neuropeptide Y receptor during vertebrate evolution. *Genomics* 85: 688–703, 2005.
- 4106 386. Lagerström MC, Fredriksson R, Bjarnadottir TK, Schiöth HB. The ancestry of the
 4107 prolactin-releasing hormone precursor. *Ann N Y Acad Sci* 1040: 368–370, 2005.
- 4108 387. Lagrange AH, Wagner EJ, Rønnekleiv OK, Kelly MJ. Estrogen rapidly attenuates a
- 4109 GABAB response in hypothalamic neurons. *Neuroendocrinology* 64: 114–123, 1996.
- 4110 388. Lamberts R, Vijayan E, Graf M, Mansky T, Wuttke W. Involvement of preoptic-

- 4111 anterior hypothalamic GABA neurons in the regulation of pituitary LH and prolactin
 4112 release. *Exp Brain Res* 52: 356–362, 1983.
- 4113 389. Lang R, Gundlach AL, Kofler B. The galanin peptide family: Receptor
- 4114 pharmacology, pleiotropic biological actions, and implications in health and disease.
 4115 *Pharmacol Ther* 115: 177–207, 2007.
- 4116 390. Lapthorn A, Harris D, Littlejohn A, Lustbader J, Canfield R, Machin K, Lorgan
- 4117 F, Isaacs N. Crystal structure of human chorionic gonadotropin. *Nature* 369: 455–461,
 4118 1994.
- 4119 391. Lariviere K, MacEachern L, Greco V, Majchrzak G, Chiu S, Drouin G, Trudeau
- 4120 VL. GAD(65) and GAD(67) isoforms of the glutamic acid decarboxylase gene
- 4121 originated before the divergence of cartilaginous fishes. *Mol Biol Evol* 19: 2325–2329,
 4122 2002.
- 4123 392. Lawrence AJ, Soame JM. The endocrine control of reproduction in Nereidae: a new
- 4124 multi-hormonal model with implications for their functional role in a changing
 4125 environment. *Philos Trans R Soc Lond B Biol Sci* 364: 3363–3376, 2009.
- 4126 393. Lawrence C, Fraley GS. Galanin-like peptide (GALP) is a hypothalamic regulator of
- 4127 energy homeostasis and reproduction. *Front Neuroendocrinol* 32: 1–9, 2011.
- 4128 394. Ledley FD, DiLella AG, Kwok SC, Woo SL. Homology between phenylalanine and
- 4129 tyrosine hydroxylases reveals common structural and functional domains. *Biochemistry*4130 24: 3389–3394, 1985.
- 4131 395. Lee AW, Devidze N, Pfaff DW, Zhou J. Functional genomics of sex hormone-
- 4132 dependent neuroendocrine systems: specific and generalized actions in the CNS. *Prog*
- 4133 Brain Res 158: 243–272, 2006.
- 4134 396. Lee D, Nguyen T, O'Neill G, Cheng R, Liu Y, Howard A, Coulombe N, Tan C,
- 4135 **Tang-Nguyen A-T**, **George S**, **O'Dowd B**. Discovery of a receptor related to the

- 4136 galanin receptors. *FEBS Lett* 446: 103–107, 1999.
- 4137 397. Lee SL, Stewart K, Goodman RH. Structure of the gene encoding rat thyrotropin
 4138 releasing hormone. *J Biol Chem* 263: 16604–16609, 1988.
- 4139 398. Lee Y, Tsunekawa K, Mi JM, Haet NU, Hwang JI, Osugi T, Otaki N, Sunakawa
- 4140 Y, Kim K, Vaudry H, Hyuk BK, Seong JY, Tsutsui K. Molecular evolution of
- 4141 multiple forms of kisspeptins and GPR54 receptors in vertebrates. *Endocrinology* 150:
- 4142 2837–2846, 2009.
- 4143 399. Lejeune H, Habert R, Saez J. Origin, proliferation and differentiation of Leydig cells.
- 4144 *J Mol Endocrinol* 20: 1–25, 1998.
- 4145 400. León-Olea M, Martyniuk CJ, Orlando EF, Ottinger MA, Rosenfeld C,
- 4146 Wolstenholme J, Trudeau VL. Current concepts in neuroendocrine disruption. *Gen*4147 *Comp Endocrinol* 203: 158–173, 2014.
- 4148 401. León S, García-Galiano D, Ruiz-Pino F, Barroso A, Manfredi-Lozano M,
- 4149 Romero-Ruiz A, Roa J, Vázquez MJ, Gaytan F, Blomenrohr M, Van Duin M,
- 4150 Pinilla L, Tena-Sempere M. Physiological roles of gonadotropin-inhibitory hormone
- 4151 signaling in the control of mammalian reproductive axis: Studies in the NPFF1
- 4152 receptor null mouse. *Endocrinology* 155: 2953–2965, 2014.
- 4153 402. Leon S, Tena-Sempere M. Dissecting the roles of gonadotropin-inhibitory hormone in
- 4154 mammals: Studies using pharmacological tools and genetically modified mouse
- 4155 models. *Front Endocrinol (Lausanne)* 6: 1–11, 2016.
- 4156 403. Leonardelli J, Barry J, Dubois M. Demonstration by fluorescent antibody technic of
- 4157 a substance immunologically related to LH-RF in hypothalamus and median eminence
- 4158 in mammals. *C R Acad Sci Hebd Seances Acad Sci D* 276: 2043–2046, 1973.
- 4159 404. Leprince J, Bagnol D, Bureau R, Fukusumi S, Granata R, Hinuma S, Larhammar
- 4160 D, Primeaux S, Sopkova-de Oliveiras Santos J, Tsutsui K, Ukena K, Vaudry H.

- 4161 The Arg–Phe-amide peptide 26RFa/glutamine RF-amide peptide and its receptor:
- 4162 IUPHAR Review 24. Br J Pharmacol 174: 3573–3607, 2017.
- 4163 405. Leranth C, MacLusky NJ, Sakamoto H, Shanabrough M, Naftolin F. Glutamic
 4164 acid decarboxylase-containing axons synapse on LHRH neurons in the rat medial
- 4165 preoptic area. *Neuroendocrinology* 40: 536–539, 1985.
- 4166 406. Lethimonier C, Madigou T, Muñoz-Cueto JA, Lareyre JJ, Kah O. Evolutionary
- 4167 aspects of GnRHs, GnRH neuronal systems and GnRH receptors in teleost fish. *Gen*4168 *Comp Endocrinol* 135: 1–16, 2004.
- 4169 407. Levavi-Sivan B, Bogerd J, Mañanós EL, Gómez A, Lareyre JJ. Perspectives on fish
- 4170 gonadotropins and their receptors. *Gen Comp Endocrinol* 165: 412–437, 2010.
- 4171 408. Levavi-Sivan B, Ofir M, Yaron Z. Possible sites of dopaminergic inhibition of
- 4172 gonadotropin release from the pituitary of a teleost fish, tilapia. *Mol Cell Endocrinol*4173 109: 87–95, 1995.
- 4174 409. Levitt M, Spector S, Sjoerdsma A, Udenfriend S. Elucidation of the rate-limiting
- 4175 step in norepinephrine biosynthesis in the perfused guinea-pig heart. *J Pharmacol Exp*
- 4176 *Ther* 148: 1–8, 1965.
- 4177 410. Li Q, Paciotti GF, Tamarkin L, Ottinger MA. LHRH-I release from quail
- 4178 hypothalamic slices measured by specific EIA. *Gen Comp Endocrinol* 95: 13–24,
 4179 1994.
- 4180 411. Li S, Hauser F, Skadborg SK, Nielsen S V., Kirketerp-Møller N,
- 4181 **Grimmelikhuijzen CJP**. Adipokinetic hormones and their G protein-coupled
- 4182 receptors emerged in Lophotrochozoa. *Sci Rep* 6: 1–13, 2016.
- 4183 412. Li S, Pelletier G. Role of dopamine in the regulation of gonadotropin-releasing
- 4184 hormone in the male rat brain as studied by in situ hybridization. *Endocrinology* 131:
- 4185 395–399, 1992.

4186	413.	Li S, Zhang Y, Liu Y, Huang X, Huang W, Lu D, Zhu P, Shi Y, Cheng CHK, Liu
4187		X, Lin H. Structural and functional multiplicity of the kisspeptin/GPR54 system in
4188		goldfish (Carassius auratus). J Endocrinol 201: 407-418, 2009.
4189	414.	Li X, Shao B, Lin C, O'Byrne KT, Lin Y. Stress-induced inhibition of LH pulses in
4190		female rats: role of GABA in arcuate nucleus. J Mol Endocrinol 55: 9–19, 2015.
4191	415.	Li X, Su J, Fang R, Zheng L, Lei R, Wang X, Lei Z, Jin M, Jiao Y, Hou Y, Guo T,
4192		Ma Z. The effects of RFRP-3, the mammalian ortholog of GnIH, on the female pig
4193		reproductive axis in vitro. Mol Cell Endocrinol 372: 65-72, 2013.
4194	416.	Li X, Su J, Lei Z, Zhao Y, Jin M, Fang R, Zheng L, Jiao Y. Gonadotropin-
4195		inhibitory hormone (GnIH) and its receptor in the female pig: CDNA cloning,
4196		expression in tissues and expression pattern in the reproductive axis during the estrous
4197		cycle. Peptides 36: 176–185, 2012.
4198	417.	Liao T-H, Pierce G. The Primary of Bovine Thyrotropin. J Biol Chem 246: 850–865,
4199		1971.
4200	418.	Libertun C, McCann SM. The possible role of histamine in the control of prolactin
4201		and gonadotropin release. Neuroendocrinology 20: 110-120, 1976.
4202	419.	Licht P, Gallo A, Aggarwal B, Farmer S, Castelino J, Parkoff H. Biological and
4203		binding activities of equine pituitary gonadotrophins and pregnant mare serum
4204		gonadotrophin. J Endocrinol 83: 311-322, 1979.
4205	420.	Lien S, Koop BF, Sandve SR, Miller JR, Kent MP, Nome T, Hvidsten TR, Leong
4206		JS, Minkley DR, Zimin A, Grammes F, Grove H, Gjuvsland A, Walenz B,
4207		Hermansen RA, Von Schalburg K, Rondeau EB, Di Genova A, Samy JKA, Olav
4208		Vik J, Vigeland MD, Caler L, Grimholt U, Jentoft S, Inge Våge D, De Jong P,
4209		Moen T, Baranski M, Palti Y, Smith DR, Yorke JA, Nederbragt AJ, Tooming-
4210		Klunderud A, Jakobsen KS, Jiang X, Fan D, Hu Y, Liberles DA, Vidal R, Iturra

4211		P, Jones SJM, Jonassen I, Maass A, Omholt SW, Davidson WS. The Atlantic
4212		salmon genome provides insights into rediploidization. Nature 533: 200-205, 2016.
4213	421.	Lillesaar C. The serotonergic system in fish. J Chem Neuroanat 41: 294–308, 2011.
4214	422.	Limonta P, Marelli MM, Mai S, Motta M, Martini L, Moretti RM. GnRH
4215		receptors in cancer: From cell biology to novel targeted therapeutic strategies. Endocr
4216		<i>Rev</i> 33: 784–811, 2012.
4217	423.	Lindemans M, Janssen T, Beets I, Temmerman L, Meelkop E, Schoofs L.
4218		Gonadotropin-releasing hormone and adipokinetic hormone signaling systems share a
4219		common evolutionary origin. Front Endocrinol (Lausanne) 2: 1-6, 2011.
4220	424.	Lindemans M, Liu F, Janssen T, Husson S, Mertens I, Gäde G, Schoofs L.
4221		Adipokinetic hormone signaling through the gonadotropin-releasing hormone receptor
4222		modulates egg-laying in Caenorhabditis elegans. Proc Natl Acad Sci USA 106: 1642-
4223		1647, 2009.
4224	425.	Litichever N, Gershon E, Dekel N, Koch Y. Hormonal regulation of GnRH and $LH\beta$
4225		mRNA expression in cultured rat granulosa cells. J Mol Neurosci 39: 78-85, 2009.
4226	426.	Liu Q, Guan XM, Martin WJ, McDonald TP, Clements MK, Jiang Q, Zeng Z,
4227		Jacobson M, Williams DL, Yu H, Bomford D, Figueroa D, Mallee J, Wang R,
4228		Evans J, Gould R, Austin CP. Identification and Characterization of Novel
4229		Mammalian Neuropeptide FF-like Peptides that Attenuate Morphine-induced
4230		Antinociception. J Biol Chem 276: 36961–36969, 2001.
4231	427.	Liu T, Wimalasena J, Bowen RL, Atwood CS. Luteinizing hormone receptor
4232		mediates neuronal pregnenolone production via up-regulation of steroidogenic acute
4233		regulatory protein expression. J Neurochem 100: 1329–1339, 2007.
4234	428.	Liu W, Nahm H, Sweeney C, Baker H, Lamkin W, Ward D. The amino acid
4235		sequence of the S-aminoethylated ovine luteinizing hormone s-subunit (LH- α). Res

- 4236 *Commun Chem Pathol Pharmacol* 2: 168–176, 1971.
- 4237 429. Liu X, Herbison AE. Estrous cycle- and sex-dependent changes in pre- and
- 4238 postsynaptic GABAB control of GnRH neuron excitability. *Endocrinology* 152: 4856–
 4239 4864, 2011.
- 4240 430. Liu X, Herbison AE. Dopamine regulation of gonadotropin-releasing hormone neuron
 4241 excitability in male and female mice. *Endocrinology* 154: 340–350, 2013.
- 4242 431. Liu Y, Zhang Y, Li S, Huang W, Liu X, Lu D, Meng Z, Lin H. Molecular cloning
 4243 and functional characterization of the first non-mammalian 26RFa/QRFP orthologue in

4244 Goldfish, Carassius auratus. *Mol Cell Endocrinol* 303: 82–90, 2009.

- 4245 432. Löfström A, Agnati LF, Fuxe K, Hökfelt T. Evidence for an inhibitory dopaminergic
- 4246 and stimulatory noradrenergic hypothalamic influence on PMS-induced ovulation in
- 4247 the immature rat. II. A pharmacological analysis. *Neuroendocrinology* 24: 289–316,
- 4248 1977.
- 4249 433. López-Sánchez JA, Maeda-Martínez AN, Croll RP, Acosta-Salmón H. Monoamine
- fluctuations during the reproductive cycle of the Pacific lion's paw scallop Nodipecten
 subnodosus. *Comp Biochem Physiol A Mol Integr Physiol* 154: 425–428, 2009.
- 4252 434. Lorenz MW. Adipokinetic hormone inhibits the formation of energy stores and egg
- 4253 production in the cricket Gryllus bimaculatus. *Comp Biochem Physiol B Biochem Mol*4254 *Biol* 136: 197–206, 2003.
- 4255 435. Loumaye E, Thorner J, Catt K. Yeast mating pheromone activates mammalian
 4256 gonadotrophs: evolutionary conservation of a reproductive hormone? *Science (80-)*
- 4257 218: 1323–1325, 1982.
- 4258 436. Lovejoy DA, Fischer WH, Parker DB, McRory JE, Park M, Lance V, Swanson P,
- 4259 **Rivier JE**, **Sherwood NM**. Primary structure of two forms of gonadotropin-releasing
- 4260 hormone from brains of the American alligator (Alligator mississippiensis). *Regul Pept*

4261 33: 105–116, 1991.

- 4262 437. Van Loy T, Vandersmissen HP, Van Hiel MB, Poels J, Verlinden H, Badisco L,
- 4263 Vassart G, Vanden Broeck J. Comparative genomics of leucine-rich repeats
- 4264 containing G protein-coupled receptors and their ligands. *Gen Comp Endocrinol* 155:
- 4265 14–21, 2008.
- 4266 438. Lundin LG. Evolution of the vertebrate genome as reflected in paralogous
- 4267 chromosomal regions in man and the house mouse. *Genomics* 16: 1–19, 1993.
- 4268 439. Lunenfeld B, Eshkol A. Immunology of human chorionic gonadotropin (HCG). *Vitam*4269 *Horm* 25: 137–190, 1967.
- 4270 440. Luque RM, Córdoba-Chacón J, Gahete MD, Navarro VM, Tena-Sempere M,
- 4271 **Kineman RD**, **Castaño JP**. Kisspeptin regulates gonadotroph and somatotroph
- 4272 function in nonhuman primate pituitary via common and distinct signaling
- 4273 mechanisms. *Endocrinology* 152: 957–966, 2011.
- 4274 441. Ma A, Bai J, He M, Wong AOL. Spexin as a neuroendocrine signal with emerging
 4275 functions. *Gen Comp Endocrinol* 265: 90–96, 2018.
- 4276 442. Ma X, Dong Y, Matzuk M, Kumar T. Targeted disruption of luteinizing hormone
- 4277 beta-subunit leads to hypogonadism, defects in gonadal steroidogenesis, and infertility.
- 4278 Proc Natl Acad Sci USA 101: 17294–17299, 2004.
- 4279 443. Macdonald LE, Wortley KE, Gowen LC, Anderson KD, Murray JD, Poueymirou
- 4280 WT, Simmons M V., Barber D, Valenzuela DM, Economides AN, Wiegand SJ,
- 4281 Yancopoulos GD, Sleeman MW, Murphy AJ. Resistance to diet-induced obesity in
- 4282 mice globally overexpressing OGH/GPB5. *Proc Natl Acad Sci* 102: 2496–2501, 2005.
- 4283 444. MacKenzie FJ, Hunter AJ, Daly C, Wilson CA. Evidence that the dopaminergic
- 4284 incerto-hypothalamic tract has a stimulatory effect on ovulation and gonadotrophin
- 4285 release. *Neuroendocrinology* 39: 289–295, 1984.

4286 445. MacLeod RM, Fontham EH, Lehmeyer JE. Prolactin and growth hormone

- 4287 production as influenced by catecholamines and agents that affect brain
- 4288 catecholamines. *Neuroendocrinology* 6: 283–294, 1970.
- 4289 446. Maddineni S, Ocón-grove OM, Krzysik-Walker SM, Hendricks GL, Proudman
- 4290 JA, Ramachandran R. Gonadotrophin-Inhibitory hormone receptor expression in the
- 4291 chicken pituitary gland: Potential influence of sexual maturation and ovarian steroids. *J*

4292 *Neuroendocrinol* 20: 1078–1088, 2008.

- 4293 447. Madigou T, Mañanos-Sanchez E, Hulshof S, Anglade I, Zanuy S, Kah O. Cloning,
- tissue distribution, and central expression of the gonadotropin-releasing hormone
- 4295 receptor in the rainbow trout (Oncorhynchus mykiss). *Biol Reprod* 63: 1857–1866,
- 4296 2000.
- 4297 448. Magre S, Jost A. Sertoli cells and testicular differentiation in the rat fetus. *J Electron*4298 *Microsc Tech* 19: 172–188, 1991.
- 4299 449. Maiti K, Li JH, Wang AF, Acharjee S, Kim WP, Im W, Kwon HB, Seong JY.
- 4300 GnRH-II analogs for selective activation and inhibition of Non-Mammalian and Type-
- 4301 II Mammalian GnRH Receptors. 16: 173–179, 2003.
- 4302 450. Maiti K, Oh DY, Moon J, Acharjee S, Li J, Bai D, Park H, Lee K, Lee Y, Jung N,
- 4303 Kim K, Vaudry H, Kwon H, Seong J. Differential effects of gonadotropin-releasing
- 4304 hormone (GnRH)-I and GnRH-II on prostate cancer cell signaling and death. *J Clin*4305 *Endocrinol Metab* 90: 4287–4298, 2005.
- 4306 451. Mañanos EL, Anglade I, Chyb J, Saligaut C, Breton B, Kah O. Involvement of
- gamma-aminobutyric acid in the control of GTH-1 and GTH-2 secretion in male and
 female rainbow trout. *Neuroendocrinology* 69: 269–280, 1999.
- 4309 452. Manfredi-Lozano M, Roa J, Tena-Sempere M. Connecting metabolism and gonadal
- 4310 function: Novel central neuropeptide pathways involved in the metabolic control of

- 4311 puberty and fertility. *Front Neuroendocrinol* 48: 37–49, 2018.
- 4312 453. Mansour A, Meador-Woodruff JH, Bunzow JR, Civelli O, Akil H, Watson SJ.
- 4313 Localization of dopamine D2 receptor mRNA and D1 and D2 receptor binding in the
- 4314 rat brain and pituitary: an in situ hybridization-receptor autoradiographic analysis. J
- 4315 *Neurosci Off J Soc Neurosci* 10: 2587–2600, 1990.
- 4316 454. Mapanao R, Cheng W. Cloning and characterization of tyrosine hydroxylase (TH)
- 4317 from the pacific white leg shrimp Litopenaeus vannamei, and its expression following
- 4318 pathogen challenge and hypothermal stress. *Fish Shellfish Immunol* 56: 506–516, 2016.
- 4319 455. Marchant T, Chang J, Nahorniak C, Peter R. Evidence that gonadotropin-releasing
- 4320 hormone also functions as a growth hormone-releasing factor in the goldfish.
- 4321 *Endocrinology* 124: 2509–2518, 1989.
- 4322 456. Marco HG, Verlinden H, Vanden Broeck J, Gäde G. Characterisation and
- 4323 pharmacological analysis of a crustacean G protein-coupled receptor: The red pigment-
- 4324 concentrating hormone receptor of Daphnia pulex. *Sci Rep* 7: 1–12, 2017.
- 4325 457. Markó M, Flückiger E. Role of serotonin in the regulation of ovulation. Evidence
- 4326 from pharmacological studies. *Neuroendocrinology* 30: 228–231, 1980.
- 4327 458. Martyniuk CJ, Aris-Brosou S, Drouin G, Cahn J, Trudeau VL. Early evolution of
- 4328 ionotropic GABA receptors and selective regimes acting on the mammalian-specific
 4329 theta and epsilon subunits. *PLoS One* 2, 2007.
- 4330 459. Martyniuk CJ, Chang JP, Trudeau VL. The effects of GABA agonists on glutamic
- 4331 acid decarboxylase, GABA-transaminase, activin, salmon gonadotrophin-releasing
- hormone and tyrosine hydroxylase mRNA in the goldfish (Carassius auratus)
- 4333 neuroendocrine brain. *J Neuroendocrinol* 19: 390–396, 2007.
- 4334 460. **Maston GA**, **Ruvolo M**. Chorionic gonadotropin has a recent origin within primates
- and an evolutionary history of selection. *Mol Biol Evol* 19: 320–335, 2002.

4336	461.	Matsuda K, Nakamura K, Shimakura SI, Miura T, Kageyama H, Uchiyama M,
4337		Shioda S, Ando H. Inhibitory effect of chicken gonadotropin-releasing hormone II on
4338		food intake in the goldfish, Carassius auratus. Horm Behav 54: 83-89, 2008.
4339	462.	Matsuda S, Matsuzawa D, Ishii D, Tomizawa H, Sutoh C, Nakazawa K, Amano
4340		K, Sajiki J, Shimizu E. Effects of perinatal exposure to low dose of bisphenol A on
4341		anxiety like behavior and dopamine metabolites in brain. Prog Neuropsychopharmacol
4342		<i>Biol Psychiatry</i> 39: 273–279, 2012.
4343	463.	Matsuda S, Saika S, Amano K, Shimizu E, Sajiki J. Changes in brain monoamine
4344		levels in neonatal rats exposed to bisphenol A at low doses. Chemosphere 78: 894-906,
4345		2010.
4346	464.	Matsui H, Takatsu Y, Kumano S, Matsumoto H, Ohtaki T. Peripheral
4347		administration of metastin induces marked gonadotropin release and ovulation in the
4348		rat. Biochem Biophys Res Commun 320: 383–388, 2004.
4349	465.	Matsuo H, Baba Y, Nair RMG, Arimura A, Schally A V. Structure of the porcine
4350		LH- and FSH-releasing hormone. I. The proposed amino acid sequence. Biochem
4351		Biophys Res Commun 43: 1334–1339, 1971.
4352	466.	Matsutani T, Nomura T. Induction of spawning by serotonin in the scallop,
4353		Patinopecten yessoensis (Jay). Mar Biol Lett 3: 353-358, 1982.
4354	467.	Maugars G, Dufour S, Cohen-Tannoudji JL, Quérat B. Multiple thyrotropin b-
4355		subunit and thyrotropin receptor-related genes arose during vertebrate evolution. PLoS
4356		<i>One</i> 9, 2014.
4357	468.	Maugars G, Dufour S, Vaudry H. Demonstration of the coexistence of duplicated LH
4358		receptors in teleosts, and their origin in ancestral actinopterygians. PLoS One 10: 1-29,
4359		2015.
4360	469.	Mayer C, Boehm U. Female reproductive maturation in the absence of

- 4361 kisspeptin/GPR54 signaling. *Nat Neurosci* 14: 704–710, 2011.
- 4362 470. Mayer RJ, Candy DJ. Changes in energy reserves during flight of the desert locust,
 4363 schistocerca gregaria. *Comp Biochem Physiol* 31: 409–418, 1969.
- 4364 471. McCarthy MM. Frank A. Beach Award. Functional significance of steroid modulation
- 4365 of GABAergic neurotransmission: analysis at the behavioral, cellular, and molecular
- 4366 levels. *Horm Behav* 29: 131–140, 1995.
- 4367 472. McConn B, Wang G, Yi J, Gilbert ER, Osugi T, Ubuka T, Tsutsui K, Chowdhury
- 4368 VS, Furuse M, Cline MA. Gonadotropin-inhibitory hormone-stimulation of food
- 4369 intake is mediated by hypothalamic effects in chicks. *Neuropeptides* 48: 327–334,
- 4370 2014.
- 4371 473. Mcdonald J k. Role of Neuropeptide Y in Reproductive Function. *Ann N Y Acad Sci*4372 611: 258–272, 1990.
- 4373 474. McGee E, Hsueh AJW. Initial and Cyclic Recruitment of Ovarian Follicles. *Endocr*4374 *Rev 21(2) 200–214* 21: 200–214, 2000.
- 4375 475. McGuire NL, Bentley GE. Neuropeptides in the gonads: From evolution to
- 4376 pharmacology. *Front Pharmacol* SEP: 1–13, 2010.
- 4377 476. McGuire NL, Bentley GE. A functional neuropeptide system in vertebrate gonads:
- 4378 Gonadotropin-inhibitory hormone and its receptor in testes of field-caught house
- 4379 sparrow (Passer domesticus). *Gen Comp Endocrinol* 166: 565–572, 2010.
- 4380 477. Meccariello R, Chianese R, Chioccarelli T, Ciaramella V, Fasano S, Pierantoni R,
- 4381 **Cobellis G.** Intra-testicular signals regulate germ cell progression and production of
- 4382 qualitatively mature spermatozoa in vertebrates. *Front Endocrinol (Lausanne)* 5: 1–11,
- 4383 2014.
- 4384 478. Mechenthaler I. Galanin and the neuroendocrine axes. *Cell Mol Life Sci* 65: 1826–
- 4385 1835, 2008.

4386	479.	Mei H, Doran J, Kyle V, Yeo SH, Colledge WH. Does kisspeptin signaling have a
4387		role in the testes? Front Endocrinol (Lausanne) 4: 1-7, 2013.
4388	480.	Mellon PL, Windle I, Goldsmith PC, Padula CA, Roberts JL, Weiner RI.
4389		Immortalization of hypothalamic GnRH neurons by genetically targeted tumorigenesis.
4390		<i>Neuron</i> 5: 1–10, 2010.
4391	481.	Mennigen JA, Stroud P, Zamora JM, Moon TW, Trudeau VL. Pharmaceuticals as
4392		neuroendocrine disruptors: lessons learned from fish on Prozac. J Toxicol Environ
4393		Health B Crit Rev 14: 387–412, 2011.
4394	482.	Menzies BR, Pask AJ, Renfree MB. Placental expression of pituitary hormones is an
4395		ancestral feature of therian mammals. Evodevo 2: 16, 2011.
4396	483.	Messager S, Chatzidaki EE, Ma D, Hendrick AG, Zahn D, Dixon J, Thresher RR,
4397		Malinge I, Lomet D, Carlton MBL, Colledge WH, Caraty A, Aparicio SAJR.
4398		Kisspeptin directly stimulates gonadotropin-releasing hormone release via G protein-
4399		coupled receptor 54. Proc Natl Acad Sci 102: 1761-1766, 2005.
4400	484.	Meyer A, Schartl M. Gene and genome duplications in vertebrates: The one-to-four (-
4401		to-eight in fish) rule and the evolution of novel gene functions. Curr Opin Cell Biol 11:
4402		699–704, 1999.
4403	485.	Midgley A, Pierce G. Immunohistochemical localization of human chorionic
4404		gonadotropin. J Exp Med 115: 289–294, 1962.
4405	486.	Millam JR, Burke WH, El Halawani ME. Release of gonadotropin-releasing
4406		hormone from the Japanese quail hypothalamus in vitro. Gen Comp Endocrinol 53:
4407		293–301, 1984.
4408	487.	Millar R, Conklin D, Lofton-Day C, Hutchinson E, Troskie B, Illing N, Sealfon S,
4409		Hapgood J. A novel human GnRH receptor homolog gene: abundant and wide tissue

4410 distribution of the antisense transcript. *J Endocrinol* 162: 117–126, 1999.

4411	488.	Millar R, Lowe S, Conklin D, Pawson a, Maudsley S, Troskie B, Ott T, Millar M,
4412		Lincoln G, Sellar R, Faurholm B, Scobie G, Kuestner R, Terasawa E, Katz a. A
4413		novel mammalian receptor for the evolutionarily conserved type II GnRH. Proc Natl
4414		<i>Acad Sci U S A</i> 98: 9636–41, 2001.
4415	489.	Millar RP. Gnrh II and type II GnRH receptors. Trends Endocrinol Metab 14: 35–43,
4416		2003.
4417	490.	Millar RP, Lu Z, Pawson AJ, Flanagan CA, Morgan K, Maudsley SR, Human C,
4418		Sciences R. Gonadotropin-Releasing Hormone Receptors. Endocr Rev 25: 235–275,
4419		2004.
4420	491.	Millar RP, Pawson AJ, Morgan K, Rissman EF, Lu ZL. Diversity of actions of
4421		GnRHs mediated by ligand-induced selective signaling. Front Neuroendocrinol 29:
4422		17–35, 2008.
4423	492.	Mirabeau O, Joly J-S. Molecular evolution of peptidergic signaling systems in
4424		bilaterians. Proc Natl Acad Sci 110: E2028-E2037, 2013.
4425	493.	Mirabeau O, Perlas E, Severini C, Audero E, Gascuel O, Possenti R, Birney E,
4426		Rosenthal N, Gross C. Identification of novel peptide hormones in the human
4427		proteome by hidden Markov model screening. Genome Res 17: 320-327, 2007.
4428	494.	Missale C, Nash SR, Robinson SW, Jaber M, Caron MG. Dopamine receptors:
4429		from structure to function. Physiol Rev 78: 189–225, 1998.
4430	495.	Mitsushima D, Hei DL, Terasawa E. gamma-Aminobutyric acid is an inhibitory
4431		neurotransmitter restricting the release of luteinizing hormone-releasing hormone
4432		before the onset of puberty. Proc Natl Acad Sci USA 91: 395–399, 1994.
4433	496.	Mittelholzer C, Andersson E, Taranger GL, Karlsen Ø, Norberg B. Quantification
4434		of gonadotropin subunits GPa, FSH β , and LH β mRNA expression from Atlantic cod
4435		(Gadus morhua) throughout a reproductive cycle. Comp Biochem Physiol - B Biochem

4436 *Mol Biol* 153: 288–295, 2009.

- 4437 497. Miyake A, Ohtsuka S, Nishizaki T, Tasaka K, Aono T, Tanizawa O, Yamatodani
- 4438 A, Watanabe T, Wada H. Involvement of H1 histamine receptor in basal and
- 4439 estrogen-stimulated luteinizing hormone-releasing hormone secretion in rats in vitro.
- 4440 *Neuroendocrinology* 45: 191–196, 1987.
- 4441 498. Miyamoto K, Hasegawa Y, Igarashi M, Chino N, Sakakibara S, Kangawa K,
- 4442 Matsuo H. Evidence that chicken hypothalamic luteinizing hormone-releasing
- 4443 hormone is [GIn8]-LH-RH. *Life Sci* 32: 1341–1347, 1983.
- 4444 499. Miyamoto K, Hasegawa Y, Minegishi T, Nomura M, Takahashi Y, Igarashi M.
- 4445 Isolation and characterization of chicken hypothalamic luteinizing hormone-releasing
- hormone. *Biochem Biophys Res Commun* 107: 820–827, 1982.
- 4447 500. Miyamoto K, Hasegawa Y, Nomura M, Igarashi M, Kangawa K, Matsuo H.
- 4448 Identification of the second gonadotropin-releasing hormone in chicken hypothalamus:
- 4449 evidence that gonadotropin secretion is probably controlled by two distinct
- 4450 gonadotropin-releasing hormones in avian species. *Proc Natl Acad Sci* 81: 3874–3878,
- 4451
 1984.
- 4452 501. Moguilevsky JA, Faigón MR, Scacchi P, Szwarcfarb B. Role of sexual
- differentiation of the hypothalamus in the differential effect of the serotoninergic
- system on LH in prepubertal male and female rats. *Neuroendocrinology* 45: 274–277,
 1987.
- 4456 502. Mohamed J, Khan I. Molecular cloning and differential expression of three GnRH
 4457 mRNAs in discrete brain areas and lymphocytes in red drum. *J Endocrinol* 188: 407–
 4458 416, 2006.
- 4459 503. Moncaut N, Somoza G, Power D, Canário A. Five gonadotrophin-releasing hormone
 4460 receptors in a teleost fish: isolation, tissue distribution and phylogenetic relationships. J

- 4461 *Mol Endocrinol* 34: 767–779, 2005.
- 4462 504. Mordue A, Blackwell A. Azadirachtin: An update. *J Insect Physiol* 39: 903–924,
 4463 1993.
- 4464 505. Moret F, Guilland JC, Coudouel S, Rochette L, Vernier P. Distribution of Tyrosine
- 4465 Hydroxylase, Dopamine, and Serotonin in the Central Nervous System of Amphioxus
- 4466 (Branchiostoma lanceolatum): Implications for the Evolution of Catecholamine
- 4467 Systems in Vertebrates. *J Comp Neurol* 468: 135–150, 2004.
- 4468 506. Morgan K, Millar RP. Evolution of GnRH ligand precursors and GnRH receptors in
- 4469 protochordate and vertebrate species. *Gen Comp Endocrinol* 139: 191–197, 2004.
- 4470 507. Morgan K, Sellar R, Pawson AJ, Lu ZL, Millar RP. Bovine and ovine
- 4471 gonadotropin-releasing hormone (GnRH)-II ligand precursors and type II GnRH
- receptor genes are functionally inactivated. *Endocrinology* 147: 5041–5051, 2006.
- 4473 508. Moriyama S, Toshihiro ITO, Takahashi A, Amano M, Sower SA, Hirano T,
- 4474 Yamamori K, Kawauchi H. A homolog of mammalian PRL-releasing peptide (fish
- 4475 arginyl-phenylalanyl-amide peptide) is a major hypothalamic peptide of prl release in
- teleost fish. *Endocrinology* 143: 2071–2079, 2002.
- 4477 509. Moshitzky P, Applebaum SW. The role of adipokinetic hormone in the control of
 4478 vitellogenesis in locusts. *Insect Biochem* 20: 319–323, 1990.
- 4479 510. Moussavi M, Wlasichuk M, Chang JP, Habibi HR. Seasonal effect of GnIH on
 4480 gonadotrope functions in the pituitary of goldfish. *Mol Cell Endocrinol* 350: 53–60,
- 4481 2012.
- 4482 511. Moussavi M, Wlasichuk M, Chang JP, Habibi HR. Seasonal Effect of
- 4483 Gonadotrophin Inhibitory Hormone on Gonadotrophin-Releasing Hormone-induced
- 4484 Gonadotroph Functions in the Goldfish Pituitary. *J Neuroendocrinol* 25: 506–516,
- 4485 2013.

4486	512.	Moy E, Kimzey L, Nelson L, Blithe L. Glycoprotein hormone alpha-subunit functions
4487		synergistically with progesterone to stimulate differentiation of cultured human
4488		endometrial stromal cells to decidualized cells: a novel role for free alpha-subunit in
4489		reproduction. Endocrinology 1137: 1332–1339, 1996.
4490	513.	Muir AI, Chamberlain L, Elshourbagy NA, Michalovich D, Moore DJ, Calamari
4491		A, Szekeres PG, Sarau HM, Chambers JK, Murdock P, Steplewski K, Shabon U,
4492		Miller JE, Middleton SE, Darker JG, Larminie CGC, Wilson S, Bergsma DJ,
4493		Emson P, Faull R, Philpott KL, Harrison DC. AXOR12, a Novel Human G Protein-
4494		coupled Receptor, Activated by the Peptide KiSS-1. J Biol Chem 276: 28969–28975,
4495		2001.
4496	514.	Müller T, Simoni M, Pekel E, Luetjens C, Chandolia R, Amato F, Norman R,
4497		Gromoll J. Chorionic gonadotrophin beta subunit mRNA but not luteinising hormone
4498		beta subunit mRNA is expressed in the pituitary of the common marmoset (Callithrix
4499		jacchus). J Mol Endocrinol 32: 115–128, 2004.
4500	515.	Munaro NI. The effect of ovarian steroids on hypothalamic 5-hydroxytryptamine
4501		neuronal activity. Neuroendocrinology 26: 270–276, 1978.
4502	516.	Muñoz-Cueto JA, Paullada-Salmerón JA, Aliaga-Guerrero M, Cowan ME,
4503		Parhar IS, Ubuka T. A journey through the gonadotropin-inhibitory hormone system
4504		of fish. Front Endocrinol (Lausanne) 8: 1–18, 2017.
4505	517.	Murakami M, Matsuzaki T, Iwasa T, Yasui T, Irahara M, Osugi T, Tsutsui K.
4506		Hypophysiotropic role of RFamide-related peptide-3 in the inhibition of LH secretion
4507		in female rats. J Endocrinol 199: 105-112, 2008.
4508	518.	Murata R, Kobayashi Y, Karimata H, Kishimoto K, Kimura M, Shimizu A,
4509		Nakamura M. The role of pituitary gonadotropins in gonadal sex differentiation in the
4510		protogynous Malabar grouper, Epinephelus malabaricus. Gen Comp Endocrinol 178:

4511 587–592, 2012.

- 4512 519. Muroi Y, Ishii T. A novel neuropeptide Y neuronal pathway linking energy state and
 4513 reproductive behavior. *Neuropeptides* 59: 1–8, 2016.
- 4514 520. Nagasaki H, Wang Z, Jackson V, Lin S, Nothacker H, Civelli O. Differential
- 4515 expression of the thyrostimulin subunits, glycoprotein alpha2 and beta5 in the rat
- 4516 pituitary. *J Mol Endocrinol* 37: 39–50, 2006.
- 4517 521. Nagatsu T. The human tyrosine hydroxylase gene. *Cell Mol Neurobiol* 9: 313–321,
 4518 1989.
- 4519 522. Nagirnaja L, Rull K, Uusküla L, Hallast P, Grigorova M, Laan M. Genomics and
- 4520 genetics of gonadotropin beta-subunit genes: Unique FSHB and duplicated LHB/CGB

4521 loci. *Mol Cell Endocrinol* 329: 4–16, 2010.

- 4522 523. Nakabayashi K, Matsumi H, Bhalla A, Bae J, Mosselman S, Hsu S, Hsueh A.
- 4523 Thyrostimulin, a heterodimer of two new human glycoprotein hormone subunits,
- activates the thyroid-stimulating hormone receptor. *EThe J Clin Investig* 109: 1445–
- 4525 1452, 2002.
- 4526 524. Nakajo M, Kanda S, Karigo T, Takahashi A, Akazome Y, Uenoyama Y,
- 4527 Kobayashi M, Oka Y. Evolutionally conserved function of kisspeptin neuronal system
- 4528 is nonreproductive regulation as revealed by nonmammalian study. *Endocrinology*
- 4529 159: 163–183, 2018.
- 4530 525. Naor Z, Huhtaniemi I. Interactions of the GnRH receptor with heterotrimeric G
- 4531 proteins. *Front Neuroendocrinol* 34: 88–94, 2013.
- 4532 526. Navarro VM, Castellano JM, Fernández-Fernández R, Tovar S, Roa J, Mayen A,
- 4533 Barreiro ML, Casanueva FF, Aguilar E, Dieguez C, Pinilla L, Tena-Sempere M.
- 4534 Effects of KiSS-1 peptide, the natural ligand of GPR54, on follicle-stimulating
- hormone secretion in the rat. *Endocrinology* 146: 1689–1697, 2005.

4536	527.	Navarro VM, Castellano JM, Fernández-Fernández R, Tovar S, Roa J, Mayen A,
4537		Nogueiras R, Vazquez MJ, Barreiro ML, Magni P, Aguilar E, Dieguez C, Pinilla
4538		L, Tena-Sempere M. Characterization of the potent luteinizing hormone-releasing
4539		activity of KiSS-1 peptide, the natural ligand of GPR54. Endocrinology 146: 156-163,
4540		2005.
4541	528.	Navarro VM, Fernández-Fernández R, Nogueiras R, Vigo E, Tovar S, Chartrel N,
4542		Le Marec O, Leprince J, Aguilar E, Pinilla L, Dieguez C, Vaudry H, Tena-
4543		Sempere M. Novel role of 26RFa, a hypothalamic RFamide orexigenic peptide, as
4544		putative regulator of the gonadotropic axis. J Physiol 573: 237-249, 2006.
4545	529.	Neckameyer WS, Holt B, Paradowski TJ. Biochemical conservation of recombinant
4546		Drosophila tyrosine hydroxylase with its mammalian cognates. Biochem Genet 43:
4547		425–443, 2005.
4548	530.	Neill JD, Duck LW, Sellers JC, Musgrove LC. A gonadotropin-releasing hormone
4549		(GnRH) receptor specific for GnRH II in primates. Biochem Biophys Res Commun
4550		282: 1012–1018, 2001.
4551	531.	Neuman-Lee L, Grieves T, Hopkins GR, French SS. The role of the kisspeptin
4552		system in regulation of the reproductive endocrine axis and territorial behavior in male
4553		side-blotched lizards (Uta stansburiana). Horm Behav 89: 48-54, 2017.
4554	532.	Neves SR, Ram PT, Iyengar R. G protein pathways. Science 296: 1636–1639, 2002.
4555	533.	Ni Y, Huang Y, Xiao Y, Wu J, Qian F, Grossmann R, Zhao R. Effects of repeated
4556		injection of kisspeptin-10 on the initiation of egg-laying in juvenile quail. Anim Reprod
4557		<i>Sci</i> 134: 203–209, 2012.
4558	534.	Ninomiya Y, Hayakawa Y. Insect cytokine, growth-blocking peptide, is a primary
4559		regulator of melanin-synthesis enzymes in armyworm larval cuticle. FEBS J 274:
4560		1768–1777, 2007.

4561	535.	Nocillado JN, Zohar Y, Biran J, Levavi-Sivan B, Elizur A. Chronic kisspeptin
4562		administration stimulated gonadal development in pre-pubertal male yellowtail
4563		kingfish (Seriola lalandi; Perciformes) during the breeding and non-breeding season.
4564		Gen Comp Endocrinol 191: 168–176, 2013.
4565	536.	Nock TG, Chand D, Lovejoy DA. Identification of members of the gonadotropin-
4566		releasing hormone (GnRH), corticotropin-releasing factor (CRF) families in the
4567		genome of the holocephalan, Callorhinchus milii (elephant shark). Gen Comp
4568		Endocrinol 171: 237–244, 2011.
4569	537.	Noguchi H, Hayakawa Y. Role of dopamine at the onset of pupal diapause in the
4570		cabbage armyworm Mamestra brassicae. FEBS Lett 413: 157-161, 1997.
4571	538.	Noguchi H, Hayakawa Y. Dopamine is a key factor for the induction of egg diapause
4572		of the silkworm, Bombyx mori. Eur J Biochem 268: 774–780, 2001.
4573	539.	Norton WHJ, Folchert A, Bally-Cuif L. Comparative analysis of serotonin receptor
4574		(HTR1A/HTR1B families) and transporter (slc6a4a/b) gene expression in the zebrafish
4575		brain. J Comp Neurol 511: 521–542, 2008.
4576	540.	Novaira HJ, Ng Y, Wolfe A, Radovick S. Kisspeptin increases GnRH mRNA
4577		expression and secretion in GnRH secreting neuronal cell lines. Mol Cell Endocrinol
4578		311: 126–134, 2009.
4579	541.	Nozaki M. Hypothalamic-pituitary-gonadal endocrine system in the hagfish. Front
4580		Endocrinol (Lausanne) 4: 1–8, 2013.
4581	542.	Ogawa S, Akiyama G, Kato S, Soga T, Sakuma Y, Parhar IS.
4582		Immunoneutralization of gonadotropin-releasing hormone type-III suppresses male
4583		reproductive behavior of cichlids. Neurosci Lett 403: 201-205, 2006.
4584	543.	Ogawa S, Parhar IS. Structural and functional divergence of gonadotropin-inhibitory
4585		hormone from jawless fish to mammals. Front Endocrinol (Lausanne) 5: 1–17, 2014.

4586 544. Oguchi A, Tanaka S, Yamamoto K, Kikuyama S. Release of α-subunit of

4587 glycoprotein hormones from the bullfrog pituitary: Possible effect of α -subunit on

4588 prolactin cell function. *Gen Comp Endocrinol* 102: 141–146, 1996.

- 4589 545. Ohga H, Adachi H, Kitano H, Yamaguchi A, Matsuyama M. Kiss1
- 4590 hexadecapeptide directly regulates gonadotropin-releasing hormone 1 in the scombroid
- 4591 fish, chub mackerel. *Biol Reprod* 96: 376–388, 2017.
- 4592 546. Ohga H, Selvaraj S, Adachi H, Imanaga Y, Nyuji M, Yamaguchi A, Matsuyama
- 4593 **M**. Functional analysis of kisspeptin peptides in adult immature chub mackerel
- 4594 (Scomber japonicus) using an intracerebroventricular administration method. *Neurosci*4595 *Lett* 561: 203–207, 2014.
- 4596 547. Ohga H, Selvaraj S, Matsuyama M. The roles of kisspeptin system in the
- reproductive physiology of fish with special reference to chub mackerel studies as main
 axis. *Front Endocrinol (Lausanne)* 9: 1–15, 2018.
- 4599 548. Ohno S, Wolf U, Atkin NB. Evolution from fish to mammals by gene duplication.
 4600 *Hereditas* 59: 169–187, 1968.
- 4601 549. Ohtaki T, Shintani Y, Honda S, Matsumoto H, Hori A, Kanehashi K, Terao Y,
- 4602 Kumano S, Takatsu Y, Masuda Y, Ishibashi Y, Watanabe T, Asada M, Yamada
- 4603 T, Suenaga M, Kitada C, Usuki S, Kurokawa T, Onda H, Nishimura O, Fujino M.
- 4604 Metastasis suppressor gene KiSS-1 encodes peptide ligand of a G-protein-coupled 4605 receptor. *Nature* 411: 613–617, 2001.
- 4606 550. Oishi H, Klausen C, Bentley GE, Osugi T, Tsutsui K, Gilks CB, Yano T, Leung
- 4607 **PCK**. The human gonadotropin-inhibitory hormone ortholog RFamide-related peptide-
- 4608 3 suppresses gonadotropin-induced progesterone production in human granulosa cells.

4609 *Endocrinology* 153: 3435–3445, 2012.

4610 551. Oishi S, Misu R, Tomita K, Setsuda S, Masuda R, Ohno H, Naniwa Y, Ieda N,

4611		Inoue N, Ohkura S, Uenoyama Y, Tsukamura H, Maeda KI, Hirasawa A,
4612		Tsujimoto G, Fujii N. Activation of neuropeptide FF receptors by kisspeptin receptor
4613		ligands. ACS Med Chem Lett 2: 53-57, 2011.
4614	552.	Okada SL, Ellsworth JL, Durnam DM, Haugen HS, Holloway JL, Kelley ML,
4615		Lewis KE, Ren H, Sheppard PO, Storey HM, Waggie KS, Wolf AC, Yao LY,
4616		Webster PJ. A Glycoprotein Hormone Expressed in Corticotrophs Exhibits Unique
4617		Binding Properties on Thyroid-Stimulating Hormone Receptor. Mol Endocrinol 20:
4618		414–425, 2006.
4619	553.	Okada T, Kawazoe I, Kimura S, Sasamoto Y, Aida K, Kawauchi H. Purification
4620		and characterization of gonadotropin I and II from pituitary glands of tuna (Thunnus
4621		obesus). Int J Pept Protein Res 43: 69–80, 1994.
4622	554.	Okada Y. Evidence that Gonadotropin-Releasing Hormone (GnRH) II Stimulates
4623		Luteinizing Hormone and Follicle-Stimulating Hormone Secretion from Monkey
4624		Pituitary Cultures by Activating the GnRH I Receptor. Biol Reprod 69: 1356–1361,
4625		2003.
4626	555.	Okubo K, Mitani H, Naruse K, Kondo M, Shima A, Tanaka M, Asakawa S,
4627		Shimizu N, Yoshiura Y, Aida K. Structural characterization of GnRH loci in the
4628		medaka genome. Gene 293: 181-189, 2002.
4629	556.	Okubo K, Nagahama Y. Structural and functional evolution of gonadotropin-
4630		releasing hormone in vertebrates. Acta Physiol 193: 3-15, 2008.
4631	557.	Okubo K, Sakai F, En LL, Yoshizaki G, Takeuchi Y, Naruse K, Aida K,
4632		Nagahama Y. Forebrain gonadotropin-releasing hormone neuronal development:
4633		Insights from transgenic medaka and the relevance to X-linked Kallmann syndrome.
4634		Endocrinology 147: 1076–1084, 2006.
4635	558.	Okubo K, Suetake H, Aida K. Expression of two gonadotropin-releasing hormone

- 4636 (GnRH) precursor genes in various tissues of the Japanese eel and evolution of GnRH.
 4637 Zoolog Sci 16: 471–478, 1999.
- 4638 559. Okuyama T, Yokoi S, Abe H, Isoe Y, Suehiro Y, Imada H, Tanaka M, Kawasaki
- 4639 T, Yuba S, Taniguchi Y, Kamei Y, Okubo K, Shimada A, Naruse K, Takeda H,
- 4640 Oka, Y Kubo T, H. Takeuchi. A Neural Mechanism Underlying Mating Preferences
 4641 for Familiar Individuals in Medaka Fish. *Science (80-)* 343: 91–94, 2014.
- 4642 560. Ondo JG. Gamma-aminobutyric acid effects on pituitary gonadotropin secretion.
 4643 Science 186: 738–739, 1974.
- 4644 561. Onuma TA, Duan C. Duplicated Kiss1 receptor genes in zebrafish: distinct gene
- 4645 expression patterns, different ligand selectivity, and a novel nuclear isoform with 4646 transactivating activity. *FASEB J* 26: 2941–2950, 2012.
- 4647 562. Opazo JC, Zavala K, Miranda-Rottmann S, Araya R. Evolution of dopamine
 4648 receptors: phylogenetic evidence suggests a later origin of the DRD2l and DRD4rs
- dopamine receptor gene lineages. *PeerJ* 6: e4593, 2018.
- 4650 563. Osugi T, Daukss D, Gazda K, Ubuka T, Kosugi T, Nozaki M, Sower SA, Tsutsui
- 4651 **K**. Evolutionary origin of the structure and function of gonadotropin- inhibitory
- hormone: Insights from lampreys. *Endocrinology* 153: 2362–2374, 2012.
- 4653 564. Osugi T, Ohtaki N, Sunakawa Y, Son YL, Ohkubo M, Iigo M, Amano M, Tsutsui
- 4654 K. Molecular evolution of Kiss2 genes and peptides in vertebrates. *Endocrinology* 154:
 4655 4270–4280, 2013.
- 4656 565. Osugi T, Okamura T, Son YL, Ohkubo M, Ubuka T, Henmi Y, Tsutsui K.
- 4657 Evolutionary origin of GnIH and NPFF in chordates: Insights from novel amphioxus
 4658 RFamide peptides. *PLoS One* 9: 1–11, 2014.
- 4659 566. Osugi T, Ubuka T, Tsutsui K. Review: Evolution of GnIH and related peptides
- 4660 structure and function in the chordates. *Front Neurosci* 8: 1–11, 2014.

4661	567.	Osugi T, Ubuka T, Tsutsui K. An Evolutionary Scenario for Gonadotrophin-
4662		Inhibitory Hormone in Chordates. J Neuroendocrinol 27: 556–566, 2015.
4663	568.	Osugi T, Uchida K, Nozaki M, Tsutsui K. Characterization of novel RFamide
4664		peptides in the central nervous system of the brown hagfish: Isolation, localization, and
4665		functional analysis. Endocrinology 152: 4252–4264, 2011.
4666	569.	Osugi T, Ukena K, Bentley GE, O'Brien S, Moore IT, Wingfield JC, Tsutsui K.
4667		Gonadotropin-inhibitory hormone in Gambel's white-crowned sparrow (Zonotrichia
4668		leucophrys gambelii): cDNA identification, transcript localization and functional
4669		effects in laboratory and field experiments. J Endocrinol 182: 33-42, 2004.
4670	570.	Osugi T, Ukena K, Sower SA, Kawauchi H, Tsutsui K. Evolutionary origin and
4671		divergence of PQRFamide peptides and LPXRFamide peptides in the RFamide peptide
4672		family: Insights from novel lamprey RFamide peptides. FEBS J 273: 1731–1743, 2006.
4673	571.	Ottinger MA, Abdelnabi MA, Henry P, McGary S, Thompson N, Wu JM.
4674		Neuroendocrine and behavioral implications of endocrine disrupting chemicals in
4675		quail. Horm Behav 40: 234–247, 2001.
4676	572.	Ottinger MA, Lavoie E, Thompson N, Barton A, Whitehouse K, Barton M,
4677		Abdelnabi M, Quinn M, Panzica G, Viglietti-Panzica C. Neuroendocrine and
4678		behavioral effects of embryonic exposure to endocrine disrupting chemicals in birds.
4679		Brain Res Rev 57: 376–385, 2008.
4680	573.	Ottinger MA, Lavoie ET, Thompson N, Bohannon M, Dean K, Quinn MJ. Is the
4681		gonadotropin releasing hormone system vulnerable to endocrine disruption in birds?
4682		Gen Comp Endocrinol 163: 104–108, 2009.
4683	574.	Pandolfi M, Pozzi A, Cánepa M, Vissio P, Shimizu A, Maggese M, Lobo G.
4684		Presence of β -follicle-stimulating hormone and β -luteinizing hormone transcripts in the
4685		brain of Cichlasoma dimerus (Perciformes: Cichlidae): effect of brain-derived

- 4686 gonadotropins on pituitary hormone release. *Neuroendocrinology* 89: 27–37, 2009.
- 4687 575. Panopoulou G, Poustka AJ. Timing and mechanism of ancient vertebrate genome
 4688 duplications -- the adventure of a hypothesis. *Trends Genet TIG* 21: 559–567, 2005.
- 4689 576. **Papkoff H**, **Samy T**. Isolation and partial chracterization of the polypeptide chains of
- 4690 ovine interstitial cell-stimulating hormone. *Biochim Biophys Acta* 147: 175–177, 1967.
- 4691 577. Parhar I, Ogawa S, Kitahashi T. RFamide peptides as mediators in environmental

4692 control of GnRH neurons. *Prog Neurobiol* 98: 176–196, 2012.

- 4693 578. Parhar IS, Ogawa S, Sakuma Y. Laser-captured single digoxigenin-labeled neurons
- 4694 of gonadotropin-releasing hormone types reveal a novel G protein-coupled receptor
- 4695 (Gpr54) during maturation in cichlid fish. *Endocrinology* 145: 3613–3618, 2004.
- 4696 579. Parhar IS, Ogawa S, Ubuka T. Reproductive neuroendocrine pathways of social
 4697 behavior. *Front Endocrinol (Lausanne)* 7: 1–8, 2016.
- 4698 580. **Parhar IS**, **Soga T**, **Ogawa S**, **Sakuma Y**. FSH and LH-β subunits in the preoptic
- 4699 nucleus: Ontogenic expression in teleost. *Gen Comp Endocrinol* 132: 369–378, 2003.
- 4700 581. Park J II, Semyonov J, Chia LC, Hsu SYT. Conservation of the heterodimeric
- 4701 glycoprotein hormone subunit family proteins and the LGR signaling system from
 4702 nematodes to humans. *Endocrine* 26: 267–276, 2005.
- 4703 582. Park Y, Kim Y-J, Adams ME. Identification of G protein-coupled receptors for
- 4704 Drosophila PRXamide peptides, CCAP, corazonin, and AKH supports a theory of
 4705 ligand-receptor coevolution. *Proc Natl Acad Sci* 99: 11423–11428, 2002.
- 4706 583. Parrott AM, Sriram G, Liu Y, Mathews MB. Expression of Type II Chorionic
- 4707 Gonadotropin Genes Supports a Role in the Male Reproductive System. *Mol Cell Biol*4708 31: 287–299, 2011.
- 4709 584. Pasqualini C, Vidal B, Le Belle N, Sbaihi M, Weltzien F-A, Vernier P, Zohar Y,
- 4710 **Dufour S**. [An antagonist to GnRH in the control of reproduction in teleost fish:

4711 dopaminergic inhibition. Ancestral origin and differential conservation within

4712 vertebrates?]. *J Soc Biol* 198: 61–67, 2004.

4713 585. Pasqualini C, Weltzien F-A, Vidal B, Baloche S, Rouget C, Gilles N, Servent D,

- 4714 **Vernier P, Dufour S**. Two distinct dopamine D2 receptor genes in the European eel:
- 4715 molecular characterization, tissue-specific transcription, and regulation by sex steroids.
- 4716 *Endocrinology* 150: 1377–1392, 2009.
- 4717 586. Pasquier J, Braasch I, Batzel P, Cabau C, Montfort J, Nguyen T, Jouanno E,
- 4718 Berthelot C, Klopp C, Journot L, Postlethwait JH, Guiguen Y, Bobe J. Evolution
- 4719 of gene expression after whole-genome duplication: New insights from the spotted gar
 4720 genome. *J Exp Zool Part B Mol Dev Evol* 328: 709–721, 2017.
- 4721 587. Pasquier J, Kamech N, Lafont AG, Vaudry H, Rousseau K, Dufour S. Molecular
- 4722 evolution of GPCRs: Kisspeptin/kisspeptin receptors. *J Mol Endocrinol* 52, 2014.
- 4723 588. Pasquier J, Lafont AG, Denis F, Lefranc B, Dubessy C, Moreno-Herrera A,
- 4724 Vaudry H, Leprince J, Dufour S, Rousseau K. Eel Kisspeptins: Identification,
- 4725 functional activity, and inhibition on both pituitary LH and GnRH receptor expression.
- 4726 Front Endocrinol (Lausanne) 8: 1–13, 2018.
- 4727 589. Pasquier J, Lafont AG, Jeng SR, Morini M, Dirks R, van den Thillart G,
- 4728 Tomkiewicz J, Tostivint H, Chang CF, Rousseau K, Dufour S. Multiple Kisspeptin
- 4729 Receptors in Early Osteichthyans Provide New Insights into the Evolution of This
- 4730 Receptor Family. *PLoS One* 7, 2012.
- 4731 590. Pasquier J, Lafont AG, Leprince J, Vaudry H, Rousseau K, Dufour S. First
- 4732 evidence for a direct inhibitory effect of kisspeptins on LH expression in the eel,
- 4733 Anguilla anguilla. *Gen Comp Endocrinol* 173: 216–225, 2011.
- 4734 591. Pasquier J, Lafont AG, Rousseau K, Quérat B, Chemineau P, Dufour S. Looking
- 4735 for the bird Kiss: Evolutionary scenario in sauropsids Genome evolution and

4736		evolutionary systems biology. BMC Evol Biol 14, 2014.
4737	592.	Pasquier J, Lafont AG, Tostivint H, Vaudry H, Rousseau K, Dufour S.
4738		Comparative evolutionary histories of kisspeptins and kisspeptin receptors in
4739		vertebrates reveal both parallel and divergent features. Front Endocrinol (Lausanne) 3:
4740		1–15, 2012.
4741	593.	Patel PD, Pontrello C, Burke S. Robust and tissue-specific expression of TPH2
4742		versus TPH1 in rat raphe and pineal gland. Biol Psychiatry 55: 428-433, 2004.
4743	594.	Patel SR, Murphy KG, Thompson EL, Patterson M, Curtis AE, Ghatei MA,
4744		Bloom SR. Pyroglutamylated RFamide peptide 43 stimulates the hypothalamic-
4745		pituitary- gonadal axis via gonadotropin-releasing hormone in rats. Endocrinology 149:
4746		4747–4754, 2008.
4747	595.	Patton SJ, Luke GN, Holland PW. Complex history of a chromosomal paralogy
4748		region: insights from amphioxus aromatic amino acid hydroxylase genes and insulin-
4749		related genes. Mol Biol Evol 15: 1373–1380, 1998.
4750	596.	Pavlick RJ, Moberg GP. No Title. 105: 218–227, 1997.
4751	597.	Pawson AJ, Faccenda E, Maudsley S, Lu ZL, Naor Z, Millar RP. Mammalian type
4752		I gonadotropin-releasing hormone receptors undergo slow, constitutive, agonist-
4753		independent internalization. Endocrinology 149: 1415-1422, 2008.
4754	598.	Pecins-Thompson M, Brown NA, Kohama SG, Bethea CL. Ovarian steroid
4755		regulation of tryptophan hydroxylase mRNA expression in rhesus macaques. J
4756		Neurosci Off J Soc Neurosci 16: 7021–7029, 1996.
4757	599.	Peng W, Cao M, Chen J, Li Y, Wang Y, Zhu Z, Hu W. GnIH plays a negative role
4758		in regulating GtH expression in the common carp, Cyprinus carpio L. Gen Comp
4759		Endocrinol 235: 18–28, 2016.
4760	600.	Perrett RM, McArdle CA. Molecular mechanisms of gonadotropin-releasing

- 4761 hormone signaling: Integrating cyclic nucleotides into the network. *Front Endocrinol*4762 (*Lausanne*) 4: 1–15, 2013.
- 4763 601. Perry SJ, Huang EYK, Cronk D, Bagust J, Sharma R, Walker RJ, Wilson S,
- 4764 Burke JF. A human gene encoding morphine modulating peptides related to NPFF and
 4765 FMRFamide. *FEBS Lett* 409: 426–430, 1997.
- 4766 602. Peter R, Trudeau V, Sloley B, Peng C, Nahorniak C. Actions of catecholamines,
- 4767 peptides and sex steroids in regulation of gonadotropin-II in the goldfish. In:
- 4768 Reproductive Physiology of Fishes. Sheffield: Scott, A.P., Sumpter, J.P., Kime, D.E. &
- 4769 Rolfe M.S., eds., 1991, p. 30–34.
- 4770 603. Peter RE, Chang JP, Nahorniak CS, Omeljaniuk RJ, Sokolowska M, Shih SH,
- 4771
 Billard R. Interactions of catecholamines and GnRH in regulation of gonadotropin
- 4772 secretion in teleost fish. *Recent Prog Horm Res* 42: 513–48, 1986.
- 4773 604. Peter RE, Crim LW, Goos HJ, Crim JW. Lesioning studies on the gravid female
- 4774 goldfish: neuroendocrine regulation of ovulation. *Gen Comp Endocrinol* 35: 391–401,
 4775 1978.
- 4776 605. Peter RE, Lin HR, Van der Kraak G. Induced ovulation and spawning of cultured
- 4777 fresh water fish in China: advances in application of GnRH analogues and dopamine
 4778 antagonists. *Aquaculture* 74: 1–10, 1988.
- 4779 606. Peter RE, Paulencu CR. Involvement of the preoptic region in gonadotropin release4780 inhibition in goldfish, Carassius auratus. *Neuroendocrinology* 31: 133–141, 1980.
- 4781 607. Peter RE, Yu KL, Marchant TA, Rosenblum PM. Direct neural regulation of the
 4782 teleost adenohypophysis. *J. Exp. Zool.* 4: 84–89, 1990.
- 4783 608. **Philippe H, Lartillot N, Brinkmann H**. Multigene analyses of bilaterian animals
- 4784 corroborate the monophyly of Ecdysozoa, Lophotrochozoa, and protostomia. *Mol Biol*
- 4785 *Evol* 22: 1246–1253, 2005.

4786	609.	Pierce J. The subunits of the pituitary thyrotropin Their relationship to other
4787		glycoprotein hormones. Endocrinology 89: 1331-1334, 1971.
4788	610.	Pineda R, Garcia-Galiano D, Sanchez-Garrido MA, Romero M, Ruiz-Pino F,
4789		Aguilar E, Dijcks FA, Blomenröhr M, Pinilla L, van Noort PI, Tena-Sempere M.
4790		Characterization of the inhibitory roles of RFRP3, the mammalian ortholog of GnIH, in
4791		the control of gonadotropin secretion in the rat: in vivo and in vitro studies. $Am J$
4792		<i>Physiol Metab</i> 299: E39–E46, 2010.
4793	611.	Pinelli C, D'Aniello B, Fiorentino M, Bhat G, Saidapur SK, Rastogi RK.
4794		Distribution of gonadotropin-releasing hormone immunoreactivity in the brain of
4795		Ichthyophis beddomei (Amphibia: Gymnophiona). J Comp Neurol 384: 283–292,
4796		1997.
4797	612.	Pinelli C, Jadhao A, Biswas S, Tsutsui K, D'Aniello B. Neuroanatomical
4798		organization of the brain gonadotropin-inhibitory hormone and gonadotropin-releasing
4799		hormone systems in the frog Pelophylax esculentus. Brain Behav Evol 85: 15-28,
4800		2015.
4801	613.	Plachetzki DC, Tsai PS, Kavanaugh SI, Sower SA. Ancient origins of metazoan
4802		gonadotropin-releasing hormone and their receptors revealed by phylogenomic
4803		analyses. Gen Comp Endocrinol 234: 10–19, 2016.
4804	614.	Plant T. 60 YEARS OF NEUROENDOCRINOLOGY: The hypothalamo-pituitary-
4805		gonadal axis. <i>J Endocrinol</i> 226: T41-54, 2015.
4806	615.	Ponzoni S. Tyrosine hydroxylase protein expression in ventral nerve cord of
4807		Neotropical freshwater crab. Tissue Cell 46: 482-489, 2014.
4808	616.	Popesku JT, Martyniuk CJ, Mennigen J, Xiong H, Zhang D, Xia X, Cossins AR,
4809		Trudeau VL. The goldfish (Carassius auratus) as a model for neuroendocrine
4810		signaling. Mol Cell Endocrinol 293: 43–56, 2008.

- 4811 617. Postlethwait J, Amores A, Force A, Yan YL. The zebrafish genome. *Methods Cell*4812 *Biol* 60: 149–163, 1999.
- 4813 618. Powell JF, Reska-Skinner SM, Prakash MO, Fischer WH, Park M, Rivier JE,
- 4814 Craig AG, Mackie GO, Sherwood NM. Two new forms of gonadotropin-releasing
- 4815 hormone in a protochordate and the evolutionary implications. *Proc Natl Acad Sci US*
- 4816 *A* 93: 10461–10464, 1996.
- 4817 619. Powell JF, Zohar Y, Elizur A, Park M, Fischer WH, Craig AG, Rivier JE,
- 4818 Lovejoy DA, Sherwood NM. Three forms of gonadotropin-releasing hormone
- 4819 characterized from brains of one species. *Proc Natl Acad Sci U S A* 91: 12081–12085,
 4820 1994.
- 4821 620. Prasad GLV, Naik BR, Ko JE, Nagaraju GP. Effects of naloxone, serotonin, and
- 4822 dopamine on reproduction of the freshwater crab Barytelphusa guerini. *J Exp Zool A*4823 *Ecol Genet Physiol* 321: 173–182, 2014.
- 4824 621. **Prasad P, Ogawa S, Parhar IS**. Role of serotonin in fish reproduction. *Front*
- 4825 *Neurosci* 9: 1–9, 2015.
- 4826 622. Predel R, Neupert S, Russell WK, Scheibner O, Nachman RJ. Corazonin in insects.
 4827 Peptides 28: 3–10, 2007.
- 4828 623. **Price D**, **Greenberg M**. Structure of a molluscan cardioexcitatory neuropeptide.
- 4829 *Science (80-)* 197: 1670–671, 1977.
- 4830 624. Przekop F, Skubiszewski B, Domański E. The effects of indoleamines (serotonin and
- 4831 melatonin) on induction of ovulation in rabbits. *Acta Physiol Pol* 26: 395–398, 1975.
- 4832 625. Qi X, Zhou W, Li S, Lu D, Yi S, Xie R, Liu X, Zhang Y, Lin H. Evidences for the
- 4833 regulation of GnRH and GTH expression by GnIH in the goldfish, Carassius auratus.
- 4834 *Mol Cell Endocrinol* 366: 9–20, 2013.
- 4835 626. Qin F, Wang X, Liu S, Zheng Y, Li M, Zhang Y, Wang Z. Gene expression

4836		profiling of key genes in hypothalamus-pituitary-gonad axis of rare minnow
4837		Gobiocypris rarus in response to EE2. Gene 552: 8-17, 2014.
4838	627.	Quérat B, Arai Y, Henry A, Akama Y, Longhurst TJ, Joss JMP. Pituitary
4839		glycoprotein hormone beta subunits in the Australian lungfish and estimation of the
4840		relative evolution rate of these subunits within vertebrates. Biol Reprod 70: 356-63,
4841		2004.
4842	628.	Quérat B, Sellouk A, Salmon C. Phylogenetic analysis of the vertebrate glycoprotein
4843		hormone family including new sequences of sturgeon (Acipenser baeri) beta subunits
4844		of the two gonadotropins and the thyroid-stimulating hormone. Biol Reprod 63: 222-
4845		228, 2000.
4846	629.	Quérat B, Tonnerre-Doncarli C, Géniès F, Salmon C. Duality of gonadotropins in
4847		gnathostomes. Gen Comp Endocrinol 124: 308-314, 2001.
4848	630.	Quillet R, Ayachi S, Bihel F, Elhabazi K, Ilien B, Simonin F. RF-amide
4849		neuropeptides and their receptors in Mammals: Pharmacological properties, drug
4850		development and main physiological functions. <i>Pharmacol Ther</i> 160: 84–132, 2016.
4851	631.	Radovick S, Wrayf S, Lee E, Nicols DK, Nakayamaii Y, Weintraubii BD,
4852		Westphal H, Cutler GB, Wondisford FE. Migratory arrest of gonadotropin-releasing
4853		hormone neurons in transgenic mice. Dev Biol 88: 3402-3406, 1991.
4854	632.	Rahman MS, Thomas P. Molecular cloning, characterization and expression of two
4855		tryptophan hydroxylase (TPH-1 and TPH-2) genes in the hypothalamus of Atlantic
4856		croaker: Down-regulation after chronic exposure to hypoxia. Neuroscience 158: 751-
4857		765, 2009.
4858	633.	Reddy PS, Tuberty SR, Fingerman M. Effects of cadmium and mercury on ovarian
4859		maturation in the red swamp crayfish, Procambarus clarkii. Ecotoxicol Environ Saf 37:
4860		62–65, 1997.

194

4862		the murine gonadotropin-releasing hormone receptor. J Biol Chem 267: 21281-21284,
4863		1992.
4864	635.	Reuter A, Gaspard U, Deville J, Vrindts-Gevaert Y, Franchimont P. Serum
4865		concentrations of human chorionic gonadotropin and its alpha and beta subunits. Clin
4866		Endocrinol 13: 305–317, 1980.
4867	636.	Richard N, Corvaisier S, Camacho E, Kottler ML. KiSS-1 and GPR54 at the
4868		pituitary level: Overview and recent insights. Peptides 30: 123-129, 2009.
4869	637.	Rispoli LA, Nett TM. Pituitary gonadotropin-releasing hormone (GnRH) receptor:
4870		Structure, distribution and regulation of expression. Anim Reprod Sci 88: 57-74, 2005.
4871	638.	Rivier C, Rivier J, Vale W. Effect of a potent GnRH antagonist and testosterone
4872		propionate on mating behavior and fertility in the male rat. Endocrinology 108: 1998-
4873		2001, 1981.
4874	639.	Rizwan MZ, Porteous R, Herbison AE, Anderson GM. Cells expressing RFamide-
4875		related peptide-1/3, the mammalian gonadotropin-inhibitory hormone orthologs, are
4876		not hypophysiotropic neuroendocrine neurons in the rat. Endocrinology 150: 1413-
4877		1420, 2009.
4878	640.	Rocco DA, Paluzzi JP V. Functional role of the heterodimeric glycoprotein hormone,
4879		GPA2/GPB5, and its receptor, LGR1: An invertebrate perspective. Gen Comp
4880		Endocrinol 234: 20–27, 2016.
4881	641.	Roch G, Busby E, Sherwood N. Evolution of Reproductive Neurohormones. In:
4882		Handbook of Neuroendocrinology, edited by Fink G, Pfaff D, Levine J. 2012, p. 73-
4883		94.
4884	642.	Roch G, Tello J, Sherwood N. At the transition from invertebrates to vertebrates, a
4885		novel GnRH-like peptide emerges in amphioxus. Mol Biol Evol 31: 765–778, 2014.

634. Reinhart J, Mertz L, Catt K. Molecular cloning and expression of cDNA encoding

4861

- 4886 643. Roch GJ, Busby ER, Sherwood NM. Evolution of GnRH: Diving deeper. *Gen Comp*4887 *Endocrinol* 171: 1–16, 2011.
- 4888 644. Roch GJ, Busby ER, Sherwood NM. GnRH receptors and peptides: Skating
 4889 backward. *Gen Comp Endocrinol* 209: 118–134, 2014.
- 4890 645. Roch GJ, Sherwood NM. Glycoprotein hormones and their receptors emerged at the
 4891 origin of metazoans. *Genome Biol Evol* 6: 1466–1479, 2014.
- 4892 646. Romero A, Montefusco-Siegmund R, Valenzuela G, Kausel G, Muller M, Brauchi
- 4893 S, Figueroa J. Structurally conserved C-RFa revealed prolactin releasing activity in
- vitro and gene expression changes in pituitary of seasonally acclimatized carp. *Biol Res*4895 45: 183–191, 2012.
- 4896 647. Root AR, Sanford JD, Kavanaugh SI, Sower SA. In vitro and in vivo effects of
- 4897 GABA, muscimol, and bicuculline on lamprey GnRH concentration in the brain of the 4898 sea lamprey (Petromyzon marinus). *Comp Biochem Physiol A Mol Integr Physiol* 138:
- 4899 493–501, 2004.
- 4900 648. de Roux N, Genin E, Carel J-C, Matsuda F, Chaussain J-L, Milgrom E.
- 4901 Hypogonadotropic hypogonadism due to loss of function of the KiSS1-derived peptide
 4902 receptor GPR54. *Proc Natl Acad Sci* 100: 10972–10976, 2003.
- 4903 649. **Rowe ML**, **Elphick MR**. The neuropeptide transcriptome of a model echinoderm, the
- 4904 sea urchin Strongylocentrotus purpuratus. *Gen Comp Endocrinol* 179: 331–344, 2012.
- 4905 650. Russek SJ. Evolution of GABA(A) receptor diversity in the human genome. *Gene*4906 227: 213–222, 1999.
- 4907 651. Sáenz-de-Miera LE, Ayala FJ. Complex evolution of orthologous and paralogous
 4908 decarboxylase genes. *J Evol Biol* 17: 55–66, 2004.
- 4909 652. Saiardi A, Bozzi Y, Baik JH, Borrelli E. Antiproliferative role of dopamine: loss of
- 4910 D2 receptors causes hormonal dysfunction and pituitary hyperplasia. *Neuron* 19: 115–

4911 126, 1997.

- 4912 653. Saito T, Nakane R, Akazome Y, Abe H, Oka Y. Electrophysiological Analysis of the
- 4913 Inhibitory Effects of FMRFamide-Like Peptides on the Pacemaker Activity of
- 4914 Gonadotropin-Releasing Hormone Neurons Electrophysiological Analysis of the
- 4915 Inhibitory Effects of FMRFamide-Like Peptides on the Pacemaker Acti. J
- 4916 *Neurophysiol* 104: 3518–3529, 2010.
- 4917 654. Sakai T, Shiraishi A, Kawada T, Matsubara S, Aoyama M, Satake H. Invertebrate
- 4918 gonadotropin-releasing hormone-related peptides and their receptors: An update. *Front* 4919 *Endocrinol (Lausanne)* 8: 1–11, 2017.
- 4920 655. Sakamoto T, Oda A, Yamamoto K, Kaneko M, Kikuyama S, Nishikawa A,
- 4921 Takahashi A, Kawauchi H, Tsutsui K, Fujimoto M. Molecular cloning and
- 4922 functional characterization of a prolactin-releasing peptide homolog from Xenopus
 4923 laevis. *Peptides* 27: 3347–3351, 2006.
- 4924 656. Sakuma Y, Pfaff D. LH-RH in the mesencephalic central grey acn Can Potentiate
 4925 Lordosis Reflex Of Female Rats. *Nature* 283: 566–567, 1980.
- 4926 657. Sakurai H, Kawashima M, Kamiyoshi M, Tanaka K. Effect of serotonin and beta-
- 4927 endorphin on the release of luteinizing hormone in the hen (Gallus domesticus). *Gen*
- 4928 *Comp Endocrinol* 63: 24–30, 1986.
- 4929 658. Saldanha CJ, Walters BJ, Fraley GS. Neurons that co-localize aromatase- and
- 4930 kisspeptin-like immunoreactivity may regulate the HPG axis of the Mallard drake
- 4931 (Anas platyrhynchos). *Gen Comp Endocrinol* 166: 606–613, 2010.
- 4932 659. Saligaut C, Linard B, Breton B, Anglade I, Bailhache T, Kah O, Jego P. Brain
- 4933 aminergic systems in salmonids and other teleosts in relation to steroid feedback and
- 4934 gonadotropin release. *Aquaculture* 177: 13–20, 1999.
- 4935 660. Samir H, Nagaoka K, Karen A, Ahmed E, Sayed M El, Watanabe G. Investigation

- the mRNA expression of KISS1 and localization of kisspeptin in the testes of Shiba
 goats and its relationship with the puberty and steriodogenic enzymes. *Small Rumin Res* 133: 1–6, 2015.
- 4939 661. Samir H, Nagaoka K, Watanabe G. Effect of kisspeptin antagonist on goat in vitro
 4940 leydig cell steroidogenesis. *Theriogenology* 121: 134–140, 2018.
- 4941 662. Dos Santos S, Bardet C, Bertrand S, Escriva H, Habert D, Querat B. Distinct
- 4942 expression patterns of glycoprotein hormone- α 2 and - β 5 in a basal chordate suggest 4943 independent developmental functions. *Endocrinology* 150: 3815–3822, 2009.
- 4944 663. Dos Santos S, Mazan S, Venkatesh B, Cohen-Tannoudji J, Quérat B. Emergence
- 4945 and evolution of the glycoprotein hormone and neurotrophin gene families in
- 4946 vertebrates. *BMC Evol Biol* 11: 332, 2011.
- 4947 664. Sari IP, Rao A, Smith JT, Tilbrook AJ, Clarke IJ. Effect of RF-amide-related
- 4948 peptide-3 on luteinizing hormone and follicle-stimulating hormone synthesis and
- 4949 secretion in ovine pituitary gonadotropes. *Endocrinology* 150: 5549–5556, 2009.
- 4950 665. Sarnat HB. The Discovery, Proof and Reproof of Neurosecretion. *Can J Neurol Sci*4951 10: 208–212, 1983.
- 4952 666. Sarojini R, Nagabhushanam R, Devi M, Fingerman M. Dopaminergic inhibition of
- 4953 5-hydroxytryptamine-stimulated testicular maturation in the fiddler crab, Uca
- 4954 pugilator. *Comp Biochem Physiol Part C Comp* 111: 287–292, 1995.
- 4955 667. Sarojini R, Nagabhushanam R, Fingerman M. In vivo evaluation of
- 4956 5-hydroxytryptamine stimulation of the testes in the fiddler crab, Uca pugilator: a
- 4957 presumed action on the neuroendocrine system. *Comp Biochem Physiol C* 106: 321–
- 4958 325, 1993.
- 4959 668. Sarojini R, Nagabhushanam R, Fingerman M. In Vivo Effects of Dopamine and
 4960 Dopaminergic Antagonists on Testicular Maturation in the Red Swamp Crayfish,

- 4961 Procambarus clarkii. *Biol Bull* 189: 340–346, 1995.
- 4962 669. Sarojini R, Nagabhushanam R, Fingerman M. Mode of action of the
- 4963 neurotransmitter 5-hydroxytryptamine in stimulating ovarian maturation in the red
- 4964 swamp crayfish, Procambarus clarkii: an in vivo and in vitro study. *J Exp Zool* 271:
- 4965 395–400, 1995.
- 4966 670. Sasaki K, Norwitz E. Gonadotropin-releasing hormone/gonadotropin-releasing
- 4967 hormone receptor signaling in the placenta. *Curr Opin Endocrinol Diabetes Obes* 18:
 4968 401–408, 2011.
- 4969 671. Satake H, Hisada M, Kawada T, Minakata H, Ukena K, Tsutsui K.
- 4970 Characterization of a cDNA encoding a novel avian hypothalamic neuropeptide
- 4971 exerting an inhibitory effect on gonadotropin release. *Biochem J* 354: 379–385, 2001.
- 4972 672. Sawada K, Ukena K, Kikuyama S, Tsutsui K. Identification of a cDNA encoding a
 4973 novel amphibian growth hormone-releasing peptide and localization of its transcript. J
 4974 *Endocrinol* 174: 395–402, 2002.
- 4975 673. Sawada K, Ukena K, Satake H, Iwakoshi E, Minakata H, Tsutsui K. Novel fish
- 4976 hypothalamic neuropeptide: Cloning of a cDNA encoding the precursor polypeptide
- 4977 and identification and localization of the mature peptide. *Eur J Biochem* 269: 6000–
 4978 6008, 2002.
- 4979 674. Sawyer CH. Rhinencephalic involvement in pituitary activation by intraventricular
 4980 histamine in the rabbit under nembutal anesthesia. *Am J Physiol* 180: 37–46, 1955.
- 4981 675. Sawyer CH, Clifton DK. Aminergic innervation of the hypothalamus. *Fed Proc* 39:
 4982 2889–2895, 1980.
- 4983 676. Sawyer CH, Markee JE, Holinshead WH. Inhibition of ovulation in the rabbit by the
 4984 adrenergic-blocking agent dibenamine. *Endocrinology* 41: 395–402, 1947.
- 4985 677. Scanes C, Glavaski-Joksimovic A, Johannsen S, Jeftinija S, Anderson L.

- 4986 Subpopulations of somatotropes with differing intracellular calcium concentration
- 4987 responses to secretagogues. *Neuroendocrinology* 85: 221–231, 2007.
- 4988 678. Scharrer B. Current concepts on the evolution of the neurosecretory neuron. In:
- 4989 *Neurosecretion and neuroendocrine activity.*, edited by Bargmann W, Oksche A,
- 4990 Polenov A, Scharrer B. Springer, 1978, p. 9–14.
- 4991 679. Scharrer E. Die lichtempfindlichkeit blinder elritzen (untersuchungen über das
 4992 zwischenhirn der fische I.). *Z f vergl Physiol.* .
- 4993 680. Schirman-Hildesheim TD, Gershon E, Litichever N, Galiani D, Ben-Aroya N,
- 4994 **Dekel N, Koch Y**. Local production of the gonadotropic hormones in the rat ovary.
- 4995 *Mol Cell Endocrinol* 282: 32–38, 2008.
- 4996 681. Schultz W. Multiple dopamine functions at different time courses. *Annu Rev Neurosci*4997 30: 259–288, 2007.
- 4998 682. Schwanzel-Fukuda M, Pfaff DW. Origin of luteinizing hormone-releasing hormone
 4999 neurons. *Nature* 338: 161–164, 1989.
- 5000 683. Scott CJ, Clarke IJ. Inhibition of luteinizing hormone secretion in ovariectomized
- 5001 ewes during the breeding season by gamma-aminobutyric acid (GABA) is mediated by
- 5002 GABA-A receptors, but not GABA-B receptors. *Endocrinology* 132: 1789–1796, 1993.
- 5003 684. Seal L, Small C, Kim M, Stanley S, Taheri S, Ghatei M, Bloom S. Prolactin
- 5004 Releasin Peptide Stimulates LH and FSH via Hypothalamic Mechanism.
- 5005 *Endocrinology* 141: 1909–1912, 2000.
- 5006 685. Seale AP, Itoh T, Moriyama S, Takahashi A, Kawauchi H, Sakamoto T, Fujimoto
- 5007 M, Riley LG, Hirano T, Grau EG. Isolation and characterization of a homologue of
- 5008 mammalian prolactin releasing peptide from the tilapia brain and its effect on prolactin
- 5009 release from the tilapia pituitary. *Gen Comp Endocrinol* 125: 328–339, 2002.
- 5010 686. Sealfon SC, Millar RP. Functional domains of the gonadotropin-releasing hormone

- 5011 receptor. *Cell Mol Neurobiol* 15: 25–42, 1995.
- 5012 687. Seeburg P, Adelman J. Characterization of cDNA for precursor of human luteinizing
 5013 hormone releasing hormone. *Nature* 311: 666–668, 1984.
- 5014 688. Seeburg P, Mason A, Stewart T, Nikolics K. The mammalian GnRH gene and its
- 5015 pivotal role in reproduction. *Recent Prog Horm Res* 43: 69–98, 1987.
- 5016 689. Seegal R, Schantz S. Biochemical and behavioral sequelae of exposure to dioxins and

5017 PCBs. In: *Dioxins and Health*. New York: Schechter, A, 1994, p. 409–447.

- 5018 690. Sefideh FA, Moon MJ, Yun S, Hong SI, Hwang JI, Seong JY. Local duplication of
- 5019 gonadotropin-releasing hormone (GnRH) receptor before two rounds of whole genome
- 5020 duplication and origin of the mammalian GnRH receptor. *PLoS One* 9, 2014.
- 5021 691. Selvaraj S, Ohga H, Kitano H, Nyuji M, Yamaguchi A, Matsuyama M. Peripheral
- 5022Administration of Kiss1 Pentadecapeptide Induces Gonadal Development in Sexually5023Immature Adult Scombroid Fish. Zoolog Sci 30: 446–454, 2013.
- 5024 692. Selvaraj S, Ohga H, Nyuji M, Kitano H, Nagano N, Yamaguchi A, Matsuyama M.
- 5025 Subcutaneous administration of Kiss1 pentadecapeptide accelerates spermatogenesis in
- 5026 prepubertal male chub mackerel (Scomber japonicus). *Comp Biochem Physiol A Mol*
- 5027 *Integr Physiol* 166: 228–236, 2013.
- 5028 693. Seminara SB, Messager S, Chatzidaki EE, Thresher RR, Acierno JS, Shagoury
- 5029 JK, Bo-Abbas Y, Kuohung W, Schwinof KM, Hendrick AG, Zahn D, Dixon J,
- 5030 Kaiser UB, Slaugenhaupt SA, Gusella JF, O'Rahilly S, Carlton MBL, Crowley
- 5031 WF, Aparicio SAJR, Colledge WH. The GPR54 Gene as a Regulator of Puberty.
- 5032 *Obstet Gynecol Surv* 59: 351–353, 2004.
- 5033 694. Semmens DC, Elphick MR. The evolution of neuropeptide signalling: Insights from
 5034 echinoderms. *Brief Funct Genomics* 16: 288–298, 2017.
- 5035 695. Semmens DC, Mirabeau O, Moghul I, Pancholi MR, Wurm Y, Elphick MR.

- Transcriptomic identification of Starfish Neuropeptide precursors yields new insights
 into neuropeptide evolution. *Open Biol* 6, 2016.
- 5038 696. Senthilkumaran B, Okuzawa K, Gen K, Kagawa H. Effects of serotonin, GABA
- and neuropeptide Y on seabream gonadotropin releasing hormone release in vitro from
- 5040 preoptic-anterior hypothalamus and pituitary of red seabream, Pagrus major. J
- 5041 *Neuroendocrinol* 13: 395–400, 2001.
- 5042 697. Servili A, Le Page Y, Leprince J, Caraty A, Escobar S, Parhar IS, Seong JY,
- 5043 Vaudry H, Kah O. Organization of two independent kisspeptin systems derived from
- 5044 evolutionary-ancient kiss genes in the brain of zebrafish. *Endocrinology* 152: 1527–
- 5045 1540, 2011.
- 5046 698. Shahab M, Mastronardi C, Seminara SB, Crowley WF, Ojeda SR, Plant TM.
- 5047 Increased hypothalamic GPR54 signaling: A potential mechanism for initiation of 5048 puberty in primates. *Proc Natl Acad Sci* 102: 2129–2134, 2005.
- 5049 699. Shahed A, Young KA. Intraovarian expression of GnRH-1 and gonadotropin mRNA
- and protein levels in Siberian hamsters during the estrus cycle and photoperiod induced

5051 regression/recrudescence. *Gen Comp Endocrinol* 170: 356–364, 2011.

- 5052 700. Shahjahan M, Ikegami T, Osugi T, Ukena K, Doi H, Hattori A, Tsutsui K, Ando
- 5053 H. Synchronised expressions of LPXRFamide peptide and its receptor genes: Seasonal,
- 5054 diurnal and circadian changes during spawning period in grass puffer. J
- 5055 *Neuroendocrinol* 23: 39–51, 2011.
- 5056 701. Sharp PJ, Follett BK. The distribution of monoamines in the hypothalamus of the
- 5057 Japanese quail, Coturnix coturnix japonica. Zeitschrift Fur Zellforsch Und
- 5058 *Mikroskopische Anat (Vienna, Austria 1948)* 90: 245–262, 1968.
- 5059 702. Sherman GB, Wolfe MW, Farmerie TA, Clay CM, Threadgill DS, Sharp DC,
- 5060 Nilson JH. A single gene encodes the beta-subunits of equine luteinizing hormone and

- 5061 chorionic gonadotropin. *Mol Endocrinol* 6: 951–959, 1992.
- 5062 703. Sherwood N, Eiden L, Brownstein M, Spiess J, Rivier J, Vale W. Characterization
- 5063 of a teleost gonadotropin-releasing hormone. *Proc Natl Acad Sci* 80: 2794–2798, 1983.
- 5064 704. Sherwood NM, Sower SA, Marshak DR, Fraser BA, Brownstein MJ. Primary
- 5065 structure of gonadotropin-releasing hormone from lamprey brain. *J Biol Chem* 261:
- 5066 4812–4819, 1986.
- 5067 705. Shi B, Liu X, Xu Y, Sun Z, Chen S, Zang K. Molecular and transcriptional
- 5068 characterization of GTHs and mPRa during ovarian maturation in rock bream

5069 Oplegnathus fasciatus. *J Exp Zool* 323A: 430–444, 2015.

- 5070 706. Shi B, Liu X, Xu Y, Wang S. Molecular characterization of three gonadotropin
 5071 subunits and their expression patterns during ovarian maturation in Cynoglossus
 5072 semilaevis. *Int J Mol Sci* 16: 2767–2793, 2015.
- 5073 707. Shi Y, Zhang Y, Li S, Liu Q, Lu D, Liu M, Meng Z, Cheng CHK, Liu X, Lin H.
- 5074 Molecular Identification of the Kiss2/Kiss1ra System and Its Potential Function During
- 5075 17Alpha-Methyltestosterone-Induced Sex Reversal in the Orange-Spotted Grouper,
- 5076 Epinephelus coioides1. *Biol Reprod* 83: 63–74, 2010.
- 5077 708. Shima K, Kitayama S, Nakano R. Gonadotropin binding sites in human ovarian
- 5078 follicles and corpora lutea during the menstrual cycle. *Obs Gynecol* 69: 800–806, 1987.
- 5079 709. Shimizu A, Yamashita M. Purification of mummichog (Fundulus heteroclitus)
- 5080 gonadotropins and their subunits, using an immunochemical assay with antisera raised 5081 against synthetic peptides. *Gen Comp Endocrinol* 125: 79–91, 2002.
- 5082 710. Shome B, Parlow A. Human follicle stimulating hormone (hFSH): first proposal for
- 5083 the amino acid sequence of the alpha-subunit (hFSHa) and first demonstration of its
- 5084 identity with the alpha-subunit of human luteinizing hormone (hLHa). J Clin
- 5085 *Endocrinol Metab* 39: 199–202, 1974.

5086	711.	Siangcham T, Tinikul Y, Poljaroen J, Sroyraya M, Changklungmoa N,
5087		Phoungpetchara I, Kankuan W, Sumpownon C, Wanichanon C, Hanna P,
5088		Sobhon P. The effects of serotonin, dopamine, gonadotropin-releasing hormones, and
5089		corazonin, on the androgenic gland of the giant freshwater prawn, Macrobrachium
5090		rosenbergii. Gen Comp Endocrinol 193: 10-18, 2013.
5091	712.	Sidhu A, Niznik HB. Coupling of dopamine receptor subtypes to multiple and diverse
5092		G proteins. Int J Dev Neurosci Off J Int Soc Dev Neurosci 18: 669–677, 2000.
5093	713.	Sidow A. Gen(om)e duplications in the evolution of early vertebrates. Curr Opin
5094		<i>Genet Dev</i> 6: 715–722, 1996.
5095	714.	Silveira LG, Noel SD, Silveira-Neto AP, Abreu AP, Brito VN, Santos MG, Bianco
5096		SDC, Kuohung W, Xu S, Gryngarten M, Escobar ME, Arnhold IJP, Mendonca
5097		BB , Kaiser UB, Latronico AC. Mutations of the KISS1 gene in disorders of puberty.
5098		J Clin Endocrinol Metab 95: 2276–2280, 2010.
5099	715.	Silver MR, Kawauchi H, Nozaki M, Sower SA. Cloning and analysis of the lamprey
5100		GnRH-III cDNA from eight species of lamprey representing the three families of
5101		Petromyzoniformes. Gen Comp Endocrinol 139: 85–94, 2004.
5102	716.	Silver MR, Nucci N V., Root AR, Reed KL, Sower SA. Cloning and characterization
5103		of a functional type II gonadotropin-releasing hormone receptor with a lengthy
5104		carboxy-terminal tail from an ancestral vertebrate, the sea lamprey. Endocrinology 146:
5105		3351–3361, 2005.
5106	717.	Sim JA, Skynner MJ, Pape JR, Herbison AE. Late postnatal reorganization of
5107		GABA(A) receptor signalling in native GnRH neurons. Eur J Neurosci 12: 3497–3504,
5108		2000.
5109	718.	Singh P, Krishna A, Sridaran R, Tsutsui K. Changes in GnRH I, bradykinin and
5110		their receptors and GnIH in the ovary of Calotes versicolor during reproductive cycle.

- 5111 *Gen Comp Endocrinol* 159: 158–169, 2008.
- 5112 719. Singh P, Krishna A, Tsutsui K. Effects of gonadotropin-inhibitory hormone on
- 5113 folliculogenesis and steroidogenesis of cyclic mice. *Fertil Steril* 95: 1397–1404, 2011.
- 5114 720. Siopes TD. Temporal characteristics and incidence of photorefractoriness in turkey
 5115 hens. *Poult Sci* 80: 95–100, 2001.
- 5116 721. Skinner D, Malpaux B, Delaleu B, Caraty A. Luteinizing hormone (LH)-releasing
- 5117 hormone in third ventricular cerebrospinal fluid of the ewe: correlation with LH pulses
 5118 and the LH surge. *Endocrinology* 136: 3230–3237, 1995.
- 5119 722. Skinner DC, Albertson AJ, Navratil A, Smith A, Mignot M, Talbott H, Scanlan-
- 5120 Blake N. Effects of gonadotrophin-releasing hormone outside the hypothalamic-

5121 pituitary-reproductive axis. *J Neuroendocrinol* 21: 282–292, 2009.

5122 723. Sloley BD, Kah O, Trudeau VL, Dulka JG, Peter RE. Amino acid neurotransmitters
5123 and dopamine in brain and pituitary of the goldfish: involvement in the regulation of

5124 gonadotropin secretion. *J Neurochem* 58: 2254–2262, 1992.

- 5125 724. Smeets WJ, González A. Catecholamine systems in the brain of vertebrates: new
- 5126 perspectives through a comparative approach. *Brain Res Brain Res Rev* 33: 308–379,
 5127 2000.
- 5128 725. Smith JJ, Kuraku S, Holt C, Sauka-Spengler T, Jiang N, Campbell MS, Yandell
- 5129 MD, Manousaki T, Meyer A, Bloom OE, Morgan JR, Buxbaum JD,
- 5130 Sachidanandam R, Sims C, Garruss AS, Cook M, Krumlauf R, Wiedemann LM,
- 5131 Sower SA, Decatur WA, Hall JA, Amemiya CT, Saha NR, Buckley KM, Rast JP,
- 5132 Das S, Hirano M, McCurley N, Guo P, Rohner N, Tabin CJ, Piccinelli P, Elgar G,
- 5133 Ruffier M, Aken BL, Searle SMJ, Muffato M, Pignatelli M, Herrero J, Jones M,
- 5134 Brown CT, Chung-Davidson YW, Nanlohy KG, Libants S V., Yeh CY, McCauley
- 5135 DW, Langeland JA, Pancer Z, Fritzsch B, De Jong PJ, Zhu B, Fulton LL,

5136		Theising B, Flicek P, Bronner ME, Warren WC, Clifton SW, Wilson RK, Li W.
5137		Sequencing of the sea lamprey (Petromyzon marinus) genome provides insights into
5138		vertebrate evolution. Nat Genet 45: 415-421, 2013.
5139	726.	Smith JT, Coolen LM, Kriegsfeld LJ, Sari IP, Jaafarzadehshirazi MR, Maltby M,
5140		Bateman K, Goodman RL, Tilbrook AJ, Ubuka T, Bentley GE, Clarke IJ,
5141		Lehman MN. Variation in kisspeptin and RFamide-related peptide (RFRP) expression
5142		and terminal connections to gonadotropin-releasing hormone neurons in the brain: A
5143		novel medium for seasonal breeding in the sheep. Endocrinology 149: 5770-5782,
5144		2008.
5145	727.	Smith JT, Rao A, Pereira A, Caraty A, Millar RP, Clarke IJ. Kisspeptin is present
5146		in ovine hypophysial portal blood but does not increase during the preovulatory
5147		luteinizing hormone surge: Evidence that gonadotropes are not direct targets of
5148		kisspeptin in vivo. Endocrinology 149: 1951–1959, 2008.
5149	728.	So W-K, Kwok H-F, Ge W. Zebrafish Gonadotropins and Their Receptors: II.
5150		Cloning and Characterization of Zebrafish Follicle-Stimulating Hormone and
5151		Luteinizing Hormone Subunits—Their Spatial-Temporal Expression Patterns and
5152		Receptor Specificity1. Biol Reprod 72: 1382-1396, 2005.
5153	729.	Sokolowska-Mikolajczyk M. Serotonin, GnRHA, and dopamine interaction in the
5154		control of in vivo luteinizing hormone release in Prussian carp(Carassius gibelio
5155		Bloch) at the time of gonad recrudescence. CzechJ Anim Sci: 45-61, 2015.
5156	730.	Somoza GM, Peter RE. Effects of serotonin on gonadotropin and growth hormone
5157		release from in vitro perifused goldfish pituitary fragments. Gen Comp Endocrinol 82:
5158		103–110, 1991.
5159	731.	Somoza GM, Yu KL, Peter RE. Serotonin stimulates gonadotropin release in female
5160		and male goldfish, Carassius auratus L. Gen Comp Endocrinol 72: 374–382, 1988.

732.	Song Y, Duan X, Chen J, Huang W, Zhu Z, Hu W. The distribution of kisspeptin
	(Kiss)1- and Kiss2 - Positive neurones and their connections with gonadotrophin-
	releasing hormone-3 neurones in the zebrafish brain. J Neuroendocrinol 27: 198–211,
	2015.
733.	Song Y, Tao B, Chen J, Jia S, Zhu Z, Trudeau V, Hu W. GABAergic Neurons and
	Their Modulatory Effects on GnRH3 in Zebrafish. Endocrinology 158: 874-886, 2017.
734.	Sotowska-Brochocka J. The stimulatory and inhibitory role of the hypothalamus in
	the regulation of ovulation in grass frog, Rana temporaria L. Gen Comp Endocrinol 70:
	83–90, 1988.
735.	Sotowska-Brochocka J, Licht P. Effect of infundibular lesions on GnRH and LH
	release in the frog, Rana temporaria, during hibernation. Gen Comp Endocrinol 85: 43-
	54, 1992.
736.	Sotowska-Brochocka J, Martyńska L, Licht P. Dopaminergic inhibition of
	gonadotropic release in hibernating frogs, Rana temporaria. Gen Comp Endocrinol 93:
	192–196, 1994.
737.	Sower S, Chiang Y, Lovas S, Conlon J. Primary structure and biological activity of a
	third gonadotropin-releasing hormone from lamprey brain. Endocrinology 132: 1125-
	1131, 1993.
738.	Sower SA. Landmark discoveries in elucidating the origins of the hypothalamic-
	pituitary system from the perspective of a basal vertebrate, sea lamprey. Gen Comp
	<i>Endocrinol</i> 264: 3–15, 2018.
739.	Sower SA, Decatur WA, Hausken KN, Marquis TJ, Barton SL, Gargan J,
	Freamat M, Wilmot M, Hollander L, Hall JA, Nozaki M, Shpilman M, Levavi-
	Sivan B. Emergence of an ancestral glycoprotein hormone in the pituitary of the sea
	lamprey, a basal vertebrate. Endocrinology 156: 3026–3037, 2015.
	 733. 734. 735. 736. 737. 738.

- 5186 740. Sower SA, Freamat M, Kavanaugh SI. The origins of the vertebrate hypothalamic-
- 5187 pituitary-gonadal (HPG) and hypothalamic-pituitary-thyroid (HPT) endocrine systems:
 5188 New insights from lampreys. *Gen Comp Endocrinol* 161: 20–29, 2009.
- 5189 741. Sower SA, Moriyama S, Kasahara M, Takahashi A, Nozaki M, Uchida K,
- **Dahlstrom JM**, **Kawauchi H**. Identification of sea lamprey GTHβ-like cDNA and its
- 5191 evolutionary implications. *Gen Comp Endocrinol* 148: 22–32, 2006.
- 5192 742. Speidel C. Gland cells of internal secretion in the spinal cord of skates. *Pap Dept, Mar*5193 *Biol, Carnegie Inst, Wash* 13: 1–31, 1919.
- 5194 743. Spergel DJ, Krüth U, Hanley DF, Sprengel R, Seeburg PH. GABA- and glutamate-
- 5195 activated channels in green fluorescent protein-tagged gonadotropin-releasing hormone
- 5196 neurons in transgenic mice. *J Neurosci Off J Soc Neurosci* 19: 2037–2050, 1999.
- 5197 744. Spicer OS, Wong TT, Zmora N, Zohar Y. Targeted mutagenesis of the
- 5198 hypophysiotropic Gnrh3 in zebrafish (Danio rerio) reveals no effects on reproductive
- 5199 performance. *PLoS One* 11: 1–22, 2016.
- 5200 745. Staubli F, Jørgensen TJD, Cazzamali G, Williamson M, Lenz C, Søndergaard L,
- 5201 **Roepstorff P, Grimmelikhuijzen CJP**. Molecular identification of the insect
- 5202 adipokinetic hormone receptors. *Proc Natl Acad Sci* 99: 3446–3451, 2002.
- 5203 746. Stewart A, Katz A, Millar R, Morgan K. Retention and silencing of prepro-GnRH-II
- and type II GnRH receptor genes in mammals. *Neuroendocrinology* 90: 416–432,
 2009.
- 5206 747. Stojilkovic S, Reinhart J, Catt K. Gonadotropin-releasing hormone receptors:
- 5207 structure and signal transduction pathways. *Endocr Rev* 15: 462–499, 1994.
- 5208 748. Stone J V., Mordue W, Batley KE, Morris HR. Structure of locust adipokinetic
- 5209 hormone, a neurohormone that regulates lipid utilisation during flight. *Nature* 263:
- 5210 207–211, 1976.

5211	749.	Sudo S, Kuwabara Y, Park J II, Sheau YH, Hsueh AJW. Heterodimeric fly
5212		glycoprotein hormone- α 2 (GPA2) and glycoprotein hormone- β 5 (GPB5) activate fly
5213		leucine-rich repeat-containing G protein-coupled receptor-1 (DLGR1) and stimulation
5214		of human thyrotropin receptors by chimeric fly GPA2 and human GPB5.
5215		Endocrinology 146: 3596–3604, 2005.
5216	750.	Sukhbaatar U, Kanasaki H, Mijiddorj T, Oride A, Hara T, Yamada T, Kyo S.
5217		Expression of GnRH and Kisspeptin in Primary Cultures of Fetal Rat Brain. Reprod
5218		<i>Sci</i> 24: 227–233, 2017.
5219	751.	Sukhbaatar U, Kanasaki H, Mijiddorj T, Oride A, Miyazaki K. Kisspeptin induces
5220		expression of gonadotropin-releasing hormone receptor in GnRH-producing GT1-7
5221		cells overexpressing G protein-coupled receptor 54. Gen Comp Endocrinol 194: 94-
5222		101, 2013.
5223	752.	Sullivan S, Moenter S. GABAergic integration of progesterone and androgen
5224		feedback to gonadotropin-releasing hormone neurons. Biol Reprod 72: 33-41, 2005.
5225	753.	Sun B, Kavanaugh SI, Tsai PS. Gonadotropin-releasing hormone in protostomes:
5226		Insights from functional studies on Aplysia californica. Gen Comp Endocrinol 176:
5227		321–326, 2012.
5228	754.	Sun YM, Flanagan CA, Illing N, Ott TR, Sellar R, Fromme BJ, Hapgood J, Sharp
5229		P, Sealfon SC, Millar RP. A chicken gonadotropin-releasing hormone receptor that
5230		confers agonist activity to mammalian antagonists. Identification of D-LYS6in the
5231		ligand and extracellular loop two of the receptor as determinants. J Biol Chem 276:
5232		7754–7761, 2001.
5233	755.	Suo S, Sasagawa N, Ishiura S. Cloning and characterization of a Caenorhabditis
5234		elegans D2-like dopamine receptor. J Neurochem 86: 869-878, 2003.
5235	756.	Suwansa-ard S, Zhao M, Thongbuakaew T, Chansela P, Ventura T, Cummins S,

5236		Sobhon P. Gonadotropin-releasing hormone and adipokinetic hormone/corazonin-
5237		related peptide in the female prawn. Gen Comp Endocrinol 236: 70-82, 2016.
5238	757.	Suzuki K, Gamble R, Sower S. Multiple transcripts encoding lamprey gonadotropin-
5239		releasing hormone-I precursors. J Mol Endocrinol 24: 365–376, 2000.
5240	758.	Suzuki K, Kawauchi H, Nagahama Y. Isolation and characterization of two distinct
5241		gonadotropins from chum salmon pituitary glands. Gen Comp Endocrinol 71: 292-
5242		301, 1988.
5243	759.	Suzuki S, Kadokawa H, Hashizume T. Direct kisspeptin-10 stimulation on
5244		luteinizing hormone secretion from bovine and porcine anterior pituitary cells. Anim
5245		<i>Reprod Sci</i> 103: 360–365, 2008.
5246	760.	Swanson KD, Ganguly R. Characterization of a Drosophila melanogaster gene similar
5247		to the mammalian genes encoding the tyrosine/tryptophan hydroxylase activator and
5248		protein kinase C inhibitor proteins. Gene 113: 183-190, 1992.
5249	761.	Swanson P, Suzuki K, Kawauchi H, Dickhoff W. Isolation and characterization of
5250		two coho salmon gonadotropins, GTH I and GTH II. Biol Reprod 44: 29-38, 1991.
5251	762.	Tachibana T, Sakamoto T. Functions of two distinct "prolactin-releasing peptides"
5252		evolved from a common ancestral gene. Front Endocrinol (Lausanne) 5: 1-12, 2014.
5253	763.	Tachibana T, Tsukada A, Fujimoto M, Takahashi H, Ohkubo T, Boswell T,
5254		Furuse M. Comparison of mammalian prolactin-releasing peptide and Carassius
5255		RFamide for feeding behavior and prolactin secretion in chicks. Gen Comp Endocrinol
5256		144: 264–269, 2005.
5257	764.	Takahashi N. The spawning of the sea urchin strongylocentrotus intermedius by
5258		gamma-aminobutyric acid. Bull. Japanese Soc. Sci. Fish. 52: 2041, 1986.
5259	765.	Talmadge K, Vamvakopoulos N, Fiddes J. Evolution of the genes for the β subunits
5260		of human chorionic gonadotropin and luteinizing hormone. Nature 307: 37-40, 1984.

5263		gonoduct in the gonad of the Japanese scallop, Patinopecten yessoensis. Gen Comp
5264		Endocrinol 166: 620–627, 2010.
5265	767.	Tanaka Y, Hua YJ, Roller L, Tanaka S. Corazonin reduces the spinning rate in the
5266		silkworm, Bombyx mori. J Insect Physiol 48: 707-714, 2002.
5267	768.	Tang H, Liu Y, Luo D, Ogawa S, Yin Y, Li S, Zhang Y, Hu W, Parhar IS, Lin H,
5268		Liu X, Cheng CHK. The kiss/kissr systems are dispensable for zebrafish reproduction:
5269		Evidence from gene knockout studies. Endocrinology 156: 589-599, 2015.
5270	769.	Tariq AR, Shabab M. Effect of kisspeptin challenge on testosterone and inhibin
5271		secretion from in vitro testicular tissue of adult male rhesus monkey (Macaca mulatta).
5272		Andrologia 49, 2017.
5273	770.	De Tassigny XDA, Fagg LA, Carlton MBL, Colledge WH. Kisspeptin can stimulate
5274		Gonadotropin-Releasing Hormone (GnRH) release by a direct action at GnRH nerve
5275		terminals. <i>Endocrinology</i> 149: 3926–3932, 2008.
5276	771.	Tawfik AI, Tanaka S, De Loof A, Schoofs L, Baggerman G, Waelkens E, Derua R,
5277		Milner Y, Yerushalmi Y, Pener AMP, Bowers WS. Identification of the
5278		gregarization-associated dark- pigmentotropin in locusts through an albino mutant.
5279		Physiology 96: 7083–7087, 1999.
5280	772.	Taylor J, Braasch I, Frickey T, Meyer A, Van de Peer Y. Genome duplication, a
5281		trait shared by 22000 species of ray-finned fish. Genome Res 13: 382-390, 2003.
5282	773.	Teigen K, McKinney J, Haavik J, Martínez A. Selectivity and affinity determinants
5283		for ligand binding to the aromatic amino acid hydroxylases. Curr Med Chem 14: 455-
5284		467, 2007.
5285	774.	Teles MG, Bianco SDC, Brito VN, Trarbach EB, Kuohung W, Xu S, Seminara

766. Tanabe T, Yuan Y, Nakamura S, Itoh N, Takahashi KG, Osada M. The role in

spawning of a putative serotonin receptor isolated from the germ and ciliary cells of the

5261

5262

5286		SB, Mendonca BB, Kaiser UB, Latronico AC. A GPR54 -Activating Mutation in a
5287		Patient with Central Precocious Puberty. N Engl J Med 358: 709-715, 2008.
5288	775.	Tello JA, Rivier JE, Sherwood NM. Tunicate GnRH peptides selectively activate
5289		Ciona intestinalis GnRH receptors and the green monkey type II GnRH receptor.
5290		Endocrinology 146: 4061–4073, 2005.
5291	776.	Tello JA, Sherwood NM. Amphioxus: Beginning of vertebrate and end of invertebrate
5292		type GnRH receptor lineage. Endocrinology 150: 2847–2856, 2009.
5293	777.	Temple JL, Millar RP, Rissman EF. An evolutionarily conserved form of
5294		gonadotropin-releasing hormone coordinates energy and reproductive behavior.
5295		Endocrinology 144: 13–19, 2003.
5296	778.	Tensen C, Okuzawa K, Blomenröhr M, Rebers F, Leurs R, Bogerd J, Schulz R,
5297		Goos H. Distinct efficacies for two endogenous ligands on a single cognate
5298		gonadoliberin receptor. Eur J Biochem 243: 134–140, 1997.
5299	779.	Terakado K. Induction of gamete release by gonadotropin-releasing hormone in a
5300		protochordate, Ciona intestinalis. Gen Comp Endocrinol 124: 277-284, 2001.
5301	780.	Terasaka T, Otsuka F, Tsukamoto N, Nakamura E, Inagaki K, Toma K, Ogura-
5302		Ochi K, Glidewell-Kenney C, Lawson MA, Makino H. Mutual interaction of
5303		kisspeptin, estrogen and bone morphogenetic protein-4 activity in GnRH regulation by
5304		GT1-7 cells. Mol Cell Endocrinol 381: 8–15, 2013.
5305	781.	Thiéry JC, Gayrard V, Le Corre S, Viguié C, Martin GB, Chemineau P, Malpaux
5306		B . Dopaminergic control of LH secretion by the A15 nucleus in anoestrous ewes. J
5307		Reprod Fertil Suppl 49: 285–296, 1995.
5308	782.	Thomas P, Rahman M, Khan I, Kummer J. Widespread endocrine disruption and
5309		reproductive impairment in an estuarine fish population exposed to seasonal hypoxia.
5310		Proceedings Biol Sci 274: 2693–2701, 2007.

- 5311 783. Thompson EL, Patterson M, Murphy KG, Smith KL, Dhillo WS, Todd JF, Ghatei
- 5312 MA, Bloom SR. Central and peripheral administration of kisspeptin-10 stimulates the
 5313 hypothalamic-pituitary-gonadal axis. *J Neuroendocrinol* 16: 850–858, 2004.
- 5314 784. Thompson IR, Kaiser UB. GnRH pulse frequency-dependent differential regulation
 5315 of LH and FSH gene expression. *Mol Cell Endocrinol* 385: 28–35, 2014.
- 5316 785. Tian S, Zandawala M, Beets I, Baytemur E, Slade SE, Scrivens JH, Elphick MR.
- 5317 Urbilaterian origin of paralogous GnRH and corazonin neuropeptide signalling
 5318 pathways. *Sci Rep* 6: 1–7, 2016.
- 5319 786. Van Tienhoven A, Nalbandov A V., Norton HW. Effect of dibenamine on
- 5320 progesterone-induced and spontaneous ovulation in the hen. *Endocrinology* 54: 605–
- 5321611, 1954.
- 5322 787. Tinikul Y, Soonthornsumrith B, Phoungpetchara I, Meeratana P, Poljaroen J,
- 5323 Duangsuwan P, Soonklang N, Mercier AJ, Sobhon P. Effects of serotonin,
- 5324 dopamine, octopamine, and spiperone on ovarian maturation and embryonic
- 5325 development in the giant freshwater prawn, Macrobrachium rosenbergii (De Man,
- 5326 1879). *Crustaceana* 82: 1007–1022, 2009.
- 5327 788. Tobari Y, Iijima N, Tsunekawa K, Osugi T, Okanoya K, Tsutsui K, Ozawa H.
- 5328 Identification of gonadotropin-inhibitory hormone in the zebra finch (Taeniopygia
- 5329 guttata): Peptide isolation, cDNA cloning and brain distribution. *Peptides* 31: 816–826,
 5330 2010.
- 5331 789. Tobari Y, Kansaku N, Tsutsui K. Noradrenergic modulation of gonadotrophin5332 inhibitory hormone gene expression in the brain of Japanese quail. *J Neuroendocrinol*5333 29, 2017.
- Tostivint H. Evolution of the gonadotropin-releasing hormone (GnRH) gene family in
 relation to vertebrate tetraploidizations. *Gen Comp Endocrinol* 170: 575–581, 2011.

5336	791.	Tovar Bohórquez MO, Mechaly AS, Hughes LC, Campanella D, Ortí G, Canosa
5337		LF, Somoza GM. Kisspeptin system in pejerrey fish (Odontesthes bonariensis).
5338		Characterization and gene expression pattern during early developmental stages. Comp
5339		Biochem Physiol -Part A Mol Integr Physiol 204: 146–156, 2017.
5340	792.	Tremblay JJ. Molecular regulation of steroidogenesis in endocrine Leydig cells.
5341		<i>Steroids</i> 103: 3–10, 2015.
5342	793.	Troskie B, Illing N, Rumbak E, Sun Y-M, Hapgood J, Sealfon S, Conklin D,
5343		Millar R. Identification of three putative GnRH receptor subtypes in vertebrates. Gen
5344		Comp Endocrinol 112: 296–302, 1998.
5345	794.	Troskie BE. Complementary Deoxyribonucleic Acid Cloning, Gene Expression, and
5346		Ligand Selectivity of a Novel Gonadotropin-Releasing Hormone Receptor Expressed
5347		in the Pituitary and Midbrain of Xenopus laevis. <i>Endocrinology</i> 141: 1764–1771, 2000.
5348	795.	Trudeau V, Schueler F, Navarro-Martin L, Hamilton C, Bulaeva E, Bennett A,
5349		Fletcher W, Taylor L. Efficient induction of spawning of northern leopard frogs
5350		(Lithobates pipiens) during and outside the natural breeding season. Reprod Biol
5351		<i>Endocrinol RB&E</i> 11: 14, 2013.
5352	796.	Trudeau V, Somoza G, Natale G, Pauli B, Wignall J, Jackman P, Doe K, Schueler
5353		F. Hormonal induction of spawning in 4 species of frogs by coinjection with a
5354		gonadotropin-releasing hormone agonist and a dopamine antagonist. Reprod Biol
5355		<i>Endocrinol RB&E</i> 8: 36, 2010.
5356	797.	Trudeau VL. Neuroendocrine regulation of gonadotrophin II release and gonadal
5357		growth in the goldfish, Carassius auratus. Rev Reprod 2: 55-68, 1997.
5358	798.	Trudeau VL, Sloley BD, Peter RE. GABA stimulation of gonadotropin-II release in
5359		goldfish: involvement of GABAA receptors, dopamine, and sex steroids. Am J Physiol
5360		265: R348-355, 1993.

- 5361 799. Trudeau VL, Spanswick D, Fraser EJ, Larivière K, Crump D, Chiu S, MacMillan
- 5362 **M**, **Schulz RW**. The role of amino acid neurotransmitters in the regulation of pituitary 5363 gonadotropin release in fish. *Biochem Cell Biol* 78: 241–259, 2000.
- 5364 800. Tsai P-S, Zhang L. The Emergence and Loss of Gonadotropin-Releasing Hormone in
- 5365Protostomes: Orthology, Phylogeny, Structure, and Function1. *Biol Reprod* 79: 798–
- 5366
 805, 2008.
- 5367 801. Tsai PS. Gonadotropin-releasing hormone by any other name would smell as sweet.
 5368 *Gen Comp Endocrinol* 264: 58–63, 2018.
- 5369 802. Tsai PS, Sun B, Rochester JR, Wayne NL. Gonadotropin-releasing hormone-like
- 5370 molecule is not an acute reproductive activator in the gastropod, Aplysia californica.
- 5371 *Gen Comp Endocrinol* 166: 280–288, 2010.
- 5372 803. Tsang S-Y, Ng S-K, Xu Z, Xue H. The evolution of GABAA receptor-like genes. *Mol*5373 *Biol Evol* 24: 599–610, 2007.
- 5374 804. Tsutsui K, Bentley GE, Bedecarrats G, Osugi T, Ubuka T, Kriegsfeld LJ.
- 5375 Gonadotropin-inhibitory hormone (GnIH) and its control of central and peripheral
- 5376 reproductive function. *Front Neuroendocrinol* 31: 284–295, 2010.
- 5377 805. Tsutsui K, Bentley GE, Kriegsfeld LJ, Osugi T, Seong JY, Vaudry H. Discovery
- 5378 and evolutionary history of gonadotrophin-inhibitory hormone and kisspeptin: New
- 5379 key neuropeptides controlling reproduction. *J Neuroendocrinol* 22: 716–727, 2010.
- 5380 806. **Tsutsui K**, **Osugi T**. Evolutionary origin and divergence of GnIH and its homologous
- 5381 peptides. *Gen Comp Endocrinol* 161: 30–33, 2009.
- 5382 807. Tsutsui K, Osugi T, Son YL, Ubuka T. Review: Structure, function and evolution of
 5383 GnIH. *Gen Comp Endocrinol* 264: 48–57, 2018.
- 5384 808. Tsutsui K, Saigoh E, Ukena K, Teranishi H, Fujisawa Y, Kikuchi M, Ishii S,
- 5385 **Sharp PJ**. A novel avian hypothalamic peptide inhibiting gonadotropin release.

- 5386 Biochem Biophys Res Commun 275: 661–667, 2000.
- 5387 809. Tsutsumi M, Zhou W, Millar RP, Mellon PL, Roberts JL, Flanagan CA, Dong K,
- Gillo B, Sealfon SC. Cloning and functional expression of a mouse gonadotropinreleasing hormone receptor. *Mol Endocrinol* 6: 1163–9, 1992.
- 5390 810. **Tuomisto J**, **Mannisto P**. Neurotransmitter regulation of Anterior pituitary hormones.
- 5391 *Pharmacol Rev* 37: 250–332, 1985.
- 5392 811. Turlejski K. Evolutionary ancient roles of serotonin: long-lasting regulation of activity
 5393 and development. *Acta Neurobiol Exp (Wars)* 56: 619–636, 1996.
- 5394 812. Twan WH, Hwang JS, Lee YH, Jeng SR, Yueh WS, Tung YH, Wu HF, Dufour S,
- 5395 Chang CF. The presence and ancestral role of gonadotropin-releasing hormone in the
- reproduction of scleractinian coral, Euphyllia ancora. *Endocrinology* 147: 397–406,
 2006.
- 5398 813. Ubuka T, Inoue K, Fukuda Y, Mizuno T, Ukena K, Kriegsfeld LJ, Tsutsui K.
- 5399 Identification, expression, and physiological functions of Siberian hamster

5400 gonadotropin-inhibitory hormone. *Endocrinology* 153: 373–385, 2012.

- 5401 814. Ubuka T, Kim S, Huang YC, Reid J, Jiang J, Osugi T, Chowdhury VS, Tsutsui K,
- 5402 **Bentley GE**. Gonadotropin-inhibitory hormone neurons interact directly with
- 5403 gonadotropin-releasing hormone-I and -II neurons in European starling brain.
- 5404 *Endocrinology* 149: 268–278, 2008.
- 5405 815. Ubuka T, Lai H, Kitani M, Suzuuchi A, Pham V, Cadigan PA, Wang A,
- 5406 Chowdhury VS, Tsutsui K, Bentley GE. Gonadotropin-inhibitory hormone
- 5407 identification, cDNA cloning, and distribution in rhesus macaque brain. *J Comp Neurol*
- 5408 517: 841–855, 2009.
- 5409 816. Ubuka T, Morgan K, Pawson AJ, Osugi T, Chowdhury VS, Minakata H, Tsutsui
- 5410 K, Millar RP, Bentley GE. Identification of human GnIH homologs, RFRP-1 and

- 5411 RFRP-3, and the cognate receptor, GPR147 in the human hypothalamic pituitary axis.
 5412 *PLoS One* 4, 2009.
- 5413 817. Ubuka T, Tsutsui K. Evolution of gonadotropin-inhibitory hormone receptor and its
 5414 ligand. *Gen Comp Endocrinol* 209: 148–161, 2014.
- 5415 818. Ubuka T, Ukena K, Sharp PJ, Bentley GE, Tsutsui K. Gonadotropin-inhibitory
- 5416 hormone inhibits gonadal development and maintenance by decreasing gonadotropin

5417 synthesis and release in male quail. *Endocrinology* 147: 1187–1194, 2006.

- 5418 819. Uchida K, Moriyama S, Chiba H, Shimotani T, Honda K, Miki M, Takahashi A,
- 5419 Sower SA, Nozaki M. Evolutionary origin of a functional gonadotropin in the pituitary
- 5420 of the most primitive vertebrate, hagfish. *Proc Natl Acad Sci* 107: 15832–15837, 2010.
- 5421 820. Uenoyama Y, Tsukamura H, Maeda KI. Kisspeptin/metastin: A key molecule
- controlling two modes of gonadotrophin-releasing hormone/luteinising hormone
 release in female rats. *J Neuroendocrinol* 21: 299–304, 2009.
- 5424 821. Ukena K, Iwakoshi-Ukena E, Osugi T, Tsutsui K. Identification and localization of
- 5425 gonadotropin-inhibitory hormone (GnIH) orthologs in the hypothalamus of the red-
- 5426 eared slider turtle, Trachemys scripta elegans. *Gen Comp Endocrinol* 227: 69–76,
- 5427 2016.
- 5428 822. Ukena K, Iwakoshi E, Minakata H, Tsutsui K. A novel rat hypothalamic RFamide-

related peptide identified by immunoaffinity chromatography and mass spectrometry. *FEBS Lett* 512: 255–258, 2002.

- 5431 823. Ukena K, Koda A, Yamamoto K, Iwakoshi-Ukena E, Minakata H, Kikuyama S,
- 5432**Tsutsui K**. Structures and diverse functions of frog growth hormine-releasing peptide5433(fGRP) and its related peptides (fGRP-RPs): a review. J Exp Zool 305A: 815–821,
- 5434 2006.
- 5435 824. Ukena K, Koda A, Yamamoto K, Kobayashi T, Iwakoshi-Ukena E, Minakata H,

- 5436 Kikuyama S, Tsutsui K. Novel neuropeptides related to frog growth hormone-
- releasing peptide: Isolation, sequence, and functional analysis. *Endocrinology* 144:
 3879–3884, 2003.
- 5439 825. Ukena K, Osugi T, Leprince J, Vaudry H, Tsutsui K. Molecular evolution of
 5440 GPCRs: 26Rfa/GPR103. *J Mol Endocrinol* 52, 2014.
- 5441 826. Ukena K, Tsutsui K. A new member of the hypothalamic rf-amide peptide family,
- 5442 lpxrf-amide peptides: Structure, localization, and function. *Mass Spectrom Rev* 24:
 5443 469–486, 2005.
- 5444 827. Vaca AA, Alfaro J. Ovarian maturation and spawning in the white shrimp, Penaeus
 5445 vannamei, by serotonin injection. *Aquaculture* 182: 373–385, 2000.
- 5446 828. Vallone D, Picetti R, Borrelli E. Structure and function of dopamine receptors.
- 5447 *Neurosci Biobehav Rev* 24: 125–132, 2000.
- 5448 829. Veenstra J. Isolation and structure of the Drosophila corazonin gene. *Biochem Biophys*5449 *Res Commun* 204: 292–296, 1994.
- 5450 830. Veenstra JA. Isolation and structure of corazonin, a cardioactive peptide from the
- 5451 American cockroach. *FEBS Lett* 250: 231–234, 1989.
- 5452 831. Veenstra JA. Neurohormones and neuropeptides encoded by the genome of Lottia
- 5453 gigantea, with reference to other mollusks and insects. *Gen Comp Endocrinol* 167: 86–
 5454 103, 2010.
- 5455 832. Veenstra JA. Neuropeptide evolution: Neurohormones and neuropeptides predicted
 5456 from the genomes of Capitella teleta and Helobdella robusta. *Gen Comp Endocrinol*
- 5457 171: 160–175, 2011.
- 5458 833. Veenstra JA. Neuropeptide evolution: Chelicerate neurohormone and neuropeptide
- 5459 genes may reflect one or more whole genome duplications. *Gen Comp Endocrinol* 229:
- 5460 41–55, 2016.

- 5461 834. Velardo J. Induction of ovulation in immature hypophysectomized rats. Science (80-) 5462 131: 357–359, 1960.
- 5463 835. De Velasco B, Shen J, Go S, Hartenstein V. Embryonic development of the 5464 Drosophila corpus cardiacum, a neuroendocrine gland with similarity to the vertebrate 5465 pituitary, is controlled by sine oculis and glass. Dev Biol 274: 280–294, 2004. 5466 Vernier P, Philippe H, Samama P, Mallet J. Bioamine receptors: evolutionary and 836. 5467 functional variations of a structural leitmotiv. EXS 63: 297-337, 1993. 5468 837. Vidal B, Pasqualini C, Le Belle N, Holland M, Sbaihi M, Vernier P, Zohar Y, 5469 **Dufour S.** Dopamine inhibits luteinizing hormone synthesis and release in the juvenile 5470 European eel: a neuroendocrine lock for the onset of puberty. Biol Reprod 71: 1491-5471 1500, 2004. 5472 838. Vidal B, Pasqualini C, Le Belle N, Holland MC, Sbaihi M, Vernier P, Zohar Y, 5473 **Dufour S.** Dopamine inhibits luteinizing hormone synthesis and release in the juvenile 5474 European eel: a neuroendocrine lock for the onset of puberty. Biol Reprod 71: 1491-5475 500, 2004. 5476 839. Vijavan E, McCann SM. The effects of intraventricular injection of gamma-5477 aminobutyric acid (GABA) on prolactin and gonadotropin release in conscious female 5478 rats. Brain Res 155: 35-43, 1978. 5479 Vijavan E, McCann SM. Effect of blockade of dopaminergic receptors on 840.
 - 5480 acetylcholine (Ach)-induced alterations of plasma gonadotropin and prolactin (Prl) in 5481
 - conscious ovariectomized rats. Brain Res Bull 5: 23-29, 1980.
 - 5482 841. Vilim FS, Aarnisalo AA, Nieminen ML, Lintunen M, Karlstedt K, Kontinen VK,
 - 5483 Kalso E, States B, Panula P, Ziff E. Gene for pain modulatory neuropeptide NPFF:
 - 5484 induction in spinal cord by noxious stimuli. [Online]. Mol Pharmacol 55: 804-11,
 - 1999. http://www.ncbi.nlm.nih.gov/pubmed/10220558. 5485

5486	842.	Virmani MA, Stojilković SS, Catt KJ. Stimulation of luteinizing hormone release by
5487		gamma-aminobutyric acid (GABA) agonists: mediation by GABAA-type receptors and
5488		activation of chloride and voltage-sensitive calcium channels. <i>Endocrinology</i> 126:
5489		2499–2505, 1990.
5490	843.	Volkoff H, Peter RE. Actions of two forms of gonadotropin releasing hormone and a
5491		GnRH antagonist on spawning behavior of the goldfish Carassius auratus. Gen Comp
5492		Endocrinol 116: 347–355, 1999.
5493	844.	Vu M, Trudeau V. Neuroendocrine control of spawning in amphibians and its
5494		practical applications. Gen Comp Endocrinol 234: 28-39, 2016.
5495	845.	Vu M, Weiler B, Trudeau V. Time- and dose-related effects of a gonadotropin-
5496		releasing hormone agonist and dopamine antagonist on reproduction in the Northern
5497		leopard frog (Lithobates pipiens). Gen Comp Endocrinol 254: 86-96, 2017.
5498	846.	Wagner EJ, Ronnekleiv OK, Bosch MA, Kelly MJ. Estrogen biphasically modifies
5499		hypothalamic GABAergic function concomitantly with negative and positive control of
5500		luteinizing hormone release. J Neurosci Off J Soc Neurosci 21: 2085–2093, 2001.
5501	847.	Wahab F, Riaz T, Khan LA, Leprince J, Vaudry H, Tena-Sempere M, Shahab M.
5502		Peripheral administration of the human kisspeptin-10 and 26RFamide inhibits plasma
5503		testosterone levels in the adult male broiler breeder birds (Gallus domesticus). $Pak J$
5504		Zool 44: 7–14, 2012.
5505	848.	Wahab F, Shahab M, Behr R. The involvement of gonadotropin inhibitory hormone
5506		and kisspeptin in the metabolic regulation of reproduction. J Endocrinol 225: R49-
5507		R66, 2015.
5508	849.	Wahlstrom T, Huhtaniemi I, Hovatta O, Seppala M. Localization of luteinizing
5509		hormone, follicle-stimulating hormone, prolactin, and their receptors in human and rat
5510		testis using immunohistochemistry and radioreceptor assay. J Clin Endocrinol Metab

5511 57: 825–830, 1983.

- Walker RJ, Holden-Dye L. Evolutionary aspects of transmitter molecules, their
 receptors and channels. *Parasitology* 102 Suppl: S7-29, 1991.
- 5514 851. Walther D, Bader M. A unique central tryptophan hydroxylase isoform. *Biochem*5515 *Pharmacol* 66: 1673–1680, 2003.
- 5516 852. Wang B, Liu Q, Liu X, Xu Y, Shi B. Molecular characterization of Kiss2 receptor
- and in vitro effects of Kiss2 on reproduction-related gene expression in the
- 5518 hypothalamus of half-smooth tongue sole (Cynoglossus semilaevis). *Gen Comp*
- *Endocrinol* 249: 55–63, 2017.
- 5520 853. Wang B, Yang G, Liu Q, Qin J, Xu Y, Li W, Liu X, Shi B. Characterization of
- 5521 LPXRFa receptor in the half-smooth tongue sole (Cynoglossus semilaevis): Molecular
- 5522 cloning, expression profiles, and differential activation of signaling pathways by
- 5523 LPXRFa peptides. Comp Biochem Physiol -Part A Mol Integr Physiol 223: 23–32,
- 5524 2018.
- Wang JY, Hsu MC, Tseng TH, Wu LS, Yang KT, Chiu CH. Kisspeptin expression
 in mouse Leydig cells correlates with age. *J Chinese Med Assoc* 78: 249–257, 2015.
- 5527 855. Wang L, Bogerd J, Choi H, Seong J, Soh J, Chun S, Blomenröhr M, TRoskie B,
- 5528 Millar R, Yu W, McCann S, Kwon H. Three distinct types of GnRH receptor
- characterized in the bullfrog. *Proc Natl Acad Sci* 98: 361–366, 2001.
- 5530 856. Wang P, Wang M, Ji G, Yang S, Zhang S, Liu Z. Demonstration of a functional
- kisspeptin/kisspeptin receptor system in amphioxus with implications for origin of
 neuroendocrine regulation. *Endocrinology* 158: 1461–1473, 2017.
- 5533 857. Wang Q, He M. Molecular characterization and analysis of a putative 5-HT receptor
- 5534 involved in reproduction process of the pearl oyster Pinctada fucata. *Gen Comp*
- *Endocrinol* 204: 71–79, 2014.

5536	858.	Wang Q, Qi X, Guo Y, Li S, Zhang Y, Liu X, Lin H. Molecular identification of
5537		GnIH/GnIHR signal and its reproductive function in protogynous hermaphroditic
5538		orange-spotted grouper (Epinephelus coioides). Gen Comp Endocrinol 216: 9-23,
5539		2015.
5540	859.	Wang Y, Wang CY, Wu Y, Huang G, Li J, Leung FC. Identification of the receptors
5541		for prolactin-releasing peptide (PrRP) and Carassius RFamide peptide (C-RFa) in
5542		chickens. <i>Endocrinology</i> 153: 1861–1874, 2012.
5543	860.	Watanabe H, Fujisawa T, Holstein TW. Cnidarians and the evolutionary origin of
5544		the nervous system. Dev Growth Differ 51: 167–183, 2009.
5545	861.	Watanabe M, Fukuda A, Nabekura J. The role of GABA in the regulation of GnRH
5546		neurons. Front Neurosci 8: 387, 2014.
5547	862.	Watanabe T, Sadamoto H, Aonuma H. Molecular basis of the dopaminergic system
5548		in the cricket Gryllus bimaculatus. Invertebr Neurosci 13: 107–123, 2013.
5549	863.	Waye A, Trudeau V. Neuroendocrine disruption: more than hormones are upset. J
5550		Toxicol Environ Health B Crit Rev 14: 270–291, 2011.
5551	864.	Weiner R. Role of classic and peptide neuromediators in the neuroendocrine
5552		regulation of LH and prolactin. In: The Physiology of Reproduction. New York:
5553		Knobil, E and Neill, JH, 1988, p. 1235–1281.
5554	865.	Weinshenker D, Garriga G, Thomas JH. Genetic and pharmacological analysis of
5555		neurotransmitters controlling egg laying in C. elegans. J Neurosci Off J Soc Neurosci
5556		15: 6975–6985, 1995.
5557	866.	Weltzien FA, Norberg B, Swanson P. Isolation and characterization of FSH and LH
5558		from pituitary glands of Atlantic halibut (Hippoglossus hippoglossus L.). Gen Comp
5559		Endocrinol 131: 97–105, 2003.
5560	867.	Weppelman RM, Long RA, Van Iderstine A, Taylor JE, Tolman RL, Peterson L,

- 5561 Olson G. Antifertility effects of dithiocarbamates in laying hens. *Biol Reprod* 23: 40–
 5562 46, 1980.
- 868. White RB, Eisen JA, Kasten TL, Fernald RD. Second gene for gonadotropinreleasing hormone in humans. *Proc Natl Acad Sci* 95: 305–309, 1998.
- 5565 869. White RB, Fernald RD. Genomic structure and expression sites of three
- gonadotropin-releasing hormone genes in one species. *Gen Comp Endocrinol* 112: 17–
 25, 1998.
- 5568 870. White SA, Kasten TL, Bond CT, Adelman JP, Fernald RD. Three gonadotropin-
- releasing hormone genes in one organism suggest novel roles for an ancient peptide. *Proc Natl Acad Sci* 92: 8363–8367, 1995.
- 5571 871. Whitlock KE. Origin and development of GnRH neurons. *Trends Endocrinol Metab*5572 16: 145–151, 2005.
- 5573 872. Wigham T, Ball JN. Proceedings: Evidence for dopaminergic inhibition of prolactin
 5574 secretion in the teleost Poecilia latipinna. *J Endocrinol* 63: 46P-47P, 1974.
- 5575 873. Williams BL, Akazome Y, Oka Y, Eisthen HL. Dynamic evolution of the GnRH
- 5576 receptor gene family in vertebrates. *BMC Evol Biol* 14: 1–22, 2014.
- 5577 874. Williams G. Extrathyroidal expression of TSH receptor. *Ann Endocrinol* 72: 68–73,
 5578 2011.
- Wilson CA. Hypothalamic amines and the release of gonadotrophins and other anterior
 pituitary hormones. *Adv Drug Res* 8: 119–204, 1974.
- 5581 876. Witham EA, Meadows JD, Hoffmann HM, Shojaei S, Coss D, Kauffman AS,
- 5582 Mellon PL. Kisspeptin Regulates Gonadotropin Genes via Immediate Early Gene
- 5583 Induction in Pituitary Gonadotropes. *Mol Endocrinol* 27: 1283–1294, 2013.
- 5584 877. Wolfe A, Hussain MA. The emerging role(s) for kisspeptin in metabolism in
- 5585 mammals. *Front Endocrinol (Lausanne)* 9: 1–10, 2018.

5586 878. Wong A, Murphy C, Chang J, Neumann C, Lo A, Peter R. Direct actions of

- serotonin on gonadotropin-II and growth hormone release from goldfish pituitary cells:
- 5588 interactions with gonadotropin-releasing hormone and dopamine and further evaluation
- of serotonin receptor specificity. *Fish Physiol Biochem* 19: 23–34, 1998.
- 5590 879. Wong TT, Zohar Y. Novel expression of gonadotropin subunit genes in oocytes of the
- 5591 gilthead seabream (Sparus aurata). *Endocrinology* 145: 5210–5220, 2004.
- Wray S. From nose to brain: Development of gonadotrophin-releasing hormone -1
 neurones. *J Neuroendocrinol* 22: 743–753, 2010.
- 5594 881. Wray S, Grant P, Gainer H. Evidence that cells expressing luteinizing hormone-
- releasing hormone mRNA in the mouse are derived from progenitor cells in the

olfactory placode. *Proc Natl Acad Sci* 86: 8132–8136, 1989.

- Wu F, Zhang X, Zhang W, Huang B, Liu Z, Hu C, Wang D. Expression of three
 gonadotropin subunits in Southern catfish gonad and their possible roles during early
- gonadal development. *Comp Biochem Physiol A Mol Integr Physiol* 153: 44–48,
 2009.
- 5601 883. Wu H, Lustbader JW, Liu Y, Canfield RE, Hendrickson WA. Structure of human
- chorionic gonadotropin at 2.6 å resolution from MAD analysis of the selenomethionyl
 protein. *Structure* 2: 545–558, 1994.
- 5604 884. Wu J, Fu W, Huang Y, Ni Y. Kisspeptin-10 advanced the initiation of egg-laying in
 5605 quails associated with the increase of triglyceride synthesis in liver. 26: 1–32, 2013.
- 5606 885. Wu M, Dumalska I, Morozova E, Van Den Pol AN, Alreja M. Gonadotropin
- inhibitory hormone inhibits basal forebrain vGluT2-gonadotropin-releasing hormone
 neurons via a direct postsynaptic mechanism. *J Physiol* 587: 1401–1411, 2009.
- 5609 886. Wu S, Page L, Sherwood NM. A role for GnRH in early brain regionalization and eye
 5610 development in zebrafish. *Mol Cell Endocrinol* 257–258: 47–64, 2006.

5611 887. Xiao Y, Ni Y, Huang Y, Wu J, Grossmann R, Zhao R. Effects of kisspeptin-10 on

5612 progesterone secretion in cultured chicken ovarian granulosa cells from preovulatory

5613 (F1-F3) follicles. *Peptides* 32: 2091–2097, 2011.

- 5614 888. Xing Y, Myers R V., Cao D, Lin W, Jiang M, Bernard MP, Moyle WR.
- 5615 Glycoprotein hormone assembly in the endoplasmic reticulum: III. The seatbelt and its
- 5616 latch site determine the assembly pathway. *J Biol Chem* 279: 35449–35457, 2004.
- 5617 889. Xu B, Bergqvist CA, Sundström G, Lundell I, Vaudry H, Leprince J, Larhammar
- 5618 **D**. Characterization of peptide QRFP (26RFa) and its receptor from amphioxus,

5619 Branchiostoma floridae. *Gen Comp Endocrinol* 210: 107–113, 2015.

- 5620 890. Xu Q, Song Y, Liu R, Chen Y, Zhang Y, Li Y, Zhao W, Chang G, Chen G. The
- 5621 dopamine beta-hydroxylase gene in Chinese goose (Anas cygnoides): cloning,
- 5622 characterization, and expression during the reproductive cycle. *Bmc Genet* 17: 48,
- 5623 2016.
- 5624 891. Yamamoto K, Fontaine R, Pasqualini C, Vernier P. Classification of Dopamine
- 5625 Receptor Genes in Vertebrates: Nine Subtypes in Osteichthyes. *Brain Behav Evol* 86:
 5626 164–175, 2015.
- 5627 892. Yamamoto K, Mirabeau O, Bureau C, Blin M, Michon-Coudouel S, Demarque M,
- 5628 Vernier P. Evolution of dopamine receptor genes of the D1 class in vertebrates. *Mol*5629 *Biol Evol* 30: 833–843, 2013.
- 5630 893. Yamamoto K, Ruuskanen J, Wullimann M, Vernier P. Two tyrosine hydroxylase
- 5631 genes in vertebrates New dopaminergic territories revealed in the zebrafish brain. *Mol*5632 *Cell Neurosci* 43: 394–402, 2010.
- 5633 894. Yamamoto K, Vernier P. The evolution of dopamine systems in chordates. *Front*5634 *Neuroanat* 5: 21, 2011.
- 5635 895. Yang B, Jiang Q, Chan T, Ko WKW, Wong AOL. Goldfish kisspeptin: Molecular

5636		cloning, tissue distribution of transcript expression, and stimulatory effects on
5637		prolactin, growth hormone and luteinizing hormone secretion and gene expression via
5638		direct actions at the pituitary level. Gen Comp Endocrinol 165: 60-71, 2010.
5639	896.	Yang H-YT, Fratta W, Majane EA, Costa E. Characterization of Two Brain
5640		Neuropeptides That Modulate the Action of Morphine. Proc Natl Acad Sci USA 82:
5641		7757–7761, 1985.
5642	897.	Yang P, Cheng Z, Watanabe G, Taya K. Effects of third ventricle injection of
5643		norepinephrine analogue on LH secretion of the non-laying SIJI goose. Endocr J 47:
5644		7–12, 2000.
5645	898.	Yin H, Ukena K, Ubuka T, Tsutsui K. A novel G protein-coupled receptor for
5646		gonadotropin-inhibitory hormone in the Japanese quail (Coturnix japonica):
5647		Identification, expression and binding activity. J Endocrinol 184: 257–266, 2005.
5648	899.	Di Yorio MP, Pérez Sirkin DI, Delgadin TH, Shimizu A, Tsutsui K, Somoza GM,
5649		Vissio PG. Gonadotrophin-Inhibitory Hormone in the Cichlid Fish Cichlasoma
5650		dimerus: Structure, Brain Distribution and Differential Effects on the Secretion of
5651		Gonadotrophins and Growth Hormone. J Neuroendocrinol 28, 2016.
5652	900.	Yoshida H, Habata Y, Hosoya M, Kawamata Y, Kitada C, Hinuma S. Molecular
5653		properties of endogenous RFamide-related peptide-3 and its interaction with receptors.
5654		Biochim Biophys Acta - Mol Cell Res 1593: 151–157, 2003.
5655	901.	Yu KL, Peter R. Dopaminergic regulation of brain gonadotropin-releasing hormone in
5656		male goldfish during spawning behavior. Neuroendocrinology 52: 276–283, 1990.
5657	902.	Yu KL, Rosenblum PM, Peter RE. In vitro release of gonadotropin-releasing
5658		hormone from the brain preoptic-anterior hypothalamic region and pituitary of female
5659		goldfish. Gen Comp Endocrinol 81: 256–267, 1991.
5660	903.	Yun S. Furlong M. Sim M. Cho M. Park S. Cho EB. Reves-Alcaraz A. Hwang JI.

5660 903. Yun S, Furlong M, Sim M, Cho M, Park S, Cho EB, Reyes-Alcaraz A, Hwang JI,

5661		Kim J, Seong JY. Prevertebrate local gene duplication facilitated expansion of the
5662		neuropeptide GPCR superfamily. Mol Biol Evol 32: 2803-2817, 2015.
5663	904.	Yun S, Kim DK, Furlong M, Hwang JI, Vaudry H, Seong JY. Does kisspeptin
5664		belong to the proposed RF-amide peptide family? Front Endocrinol (Lausanne) 5: 22-
5665		26, 2014.
5666	905.	Zandawala M, Tian S, Elphick MR. The evolution and nomenclature of GnRH-type
5667		and corazonin-type neuropeptide signaling systems. Gen Comp Endocrinol 264: 64-77,
5668		2018.
5669	906.	van Zeijl CJJ, Surovtseva O V., Kwakkel J, van Beeren HC, Duncan Bassett JH,
5670		Williams GR, Wiersinga WM, Fliers E, Boelen A. Thyrostimulin deficiency does
5671		not alter peripheral responses to acute inflammation-induced nonthyroidal illness. AJP
5672		Endocrinol Metab 307: E527–E537, 2014.
5673	907.	van Zeijl CJJ, Surovtseva O V., Wiersinga WM, Boelen A, Fliers E. Transient
5674		hypothyroxinemia in juvenile glycoprotein hormone subunit B5 knock-out mice. Mol
5675		Cell Endocrinol 321: 231–238, 2010.
5676	908.	Zhang F-P, Poutanen M, Wilbertz J, Huhtaniemi I. Normal Prenatal but Arrested
5677		Postnatal Sexual Development of Luteinizing Hormone Receptor Knockout (LuRKO)
5678		Mice. <i>Mol Endocrinol</i> 15: 172–183, 2001.
5679	909.	Zhang F-P, Rannikko A, Huhtaniemi I. Isolation and characterization of testis-
5680		specific cDNAs for luteinizing hormone beta-subunit in the rat. Biochem Biophys Res
5681		Commun 210: 858–865, 1995.
5682	910.	Zhang F, Markkula M, Toppari J, Huhtaniemi I. Novel expression of luteinizing
5683		hormone subunit genes in the rat testis. Endocrinology 136: 2904–2912, 1995.
5684	911.	Zhang H, Zhang B, Qin G, Li S, Lin Q. The Roles of the Kisspeptin System in the
5685		Reproductive Physiology of the Lined Seahorse (Hippocampus erectus), an

5686		Ovoviviparous Fish With Male Pregnancy. Front Neurosci 12: 1–13, 2018.
5687	912.	Zhang L, Tello JA, Zhang W, Tsai PS. Molecular cloning, expression pattern, and
5688		immunocytochemical localization of a gonadotropin-releasing hormone-like molecule
5689		in the gastropod mollusk, Aplysia californica. Gen Comp Endocrinol 156: 201-209,
5690		2008.
5691	913.	Zhang Y, Li S, Liu Y, Lu D, Chen H, Huang X, Liu X, Meng Z, Lin H, Cheng
5692		CHK. Structural diversity of the gnih/gnih receptor system in teleost: Its involvement
5693		in early development and the negative control of LH release. Peptides 31: 1034–1043,
5694		2010.
5695	914.	Zhao S, Zhu E, Yang C, Bentley GE, Tsutsui K, Kriegsfeld LJ. RFamide-related
5696		peptide and messenger ribonucleic acid expression in mammalian testis: Association
5697		with the Spermatogenic cycle. Endocrinology 151: 617-627, 2010.
5698	915.	Zhao Y, Lin MCA, Mock A, Yang M, Wayne NL. Kisspeptins modulate the biology
5699		of multiple populations of gonadotropin-releasing hormone neurons during
5700		embryogenesis and adulthood in zebrafish (Danio rerio). PLoS One 9: 1-10, 2014.
5701	916.	Zheng L, Su J, Fang R, Jin M, Lei Z, Hou Y, Ma Z, Guo T. Developmental changes
5702		in the role of gonadotropin-inhibitory hormone (GnIH) and its receptors in the
5703		reproductive axis of male Xiaomeishan pigs. Anim Reprod Sci 154: 113–120, 2015.
5704	917.	Zhu J, Xu X, Knight G, He C, Burnstock G, Xiang Z. A subpopulation of
5705		gonadotropin-releasing hormone neurons in the adult mouse forebrain is γ -
5706		Aminobutyric acidergic. J Neurosci Res 93: 1611–1621, 2015.
5707	918.	Zmora N, Stubblefield J, Golan M, Servili A, Levavi-Sivan B, Zohar Y. The
5708		medio-basal hypothalamus as a dynamic and plastic reproduction-related kisspeptin-
5709		gnrh-pituitary center in fish. Endocrinology 155: 1874–1886, 2014.
5710	919.	Zmora N, Stubblefield J, Zulperi Z, Biran J, Levavi-Sivan B, Muñoz-Cueto JA,

5711		Zohar Y. Differential and Gonad Stage-Dependent Roles of Kisspeptin1 and
5712		Kisspeptin2 in Reproduction in the Modern Teleosts, Morone Species1. Biol Reprod
5713		86: 1–12, 2012.
5714	920.	Zmora N, Stubblefield JD, Wong T-T, Levavi-Sivan B, Millar RP, Zohar Y.
5715		Kisspeptin Antagonists Reveal Kisspeptin 1 and Kisspeptin 2 Differential Regulation
5716		of Reproduction in the Teleost, Morone saxatilis1. <i>Biol Reprod</i> 93: 1–12, 2015.
5717	921.	Zohar Y, Muñoz-Cueto J, Elizur A, Kah O. Neuroendocrinology of reproduction in
5718		teleost fish. Gen Comp Endocrinol 165: 438-455, 2010.
5719		

5720

5721 Figure Legends

5722 Figure 1. Neuroendocrine control of reproduction in mammals: the brain-pituitary-5723 gonad axis. Internal and environmental factors are integrated at the brain level and modulate 5724 the activity of gonadotropin-releasing hormone (GnRH) neurons. GnRH is released in the 5725 hypophyseal portal vessel system and controls the production of pituitary gonadotropins, 5726 follicle stimulating hormone (FSH) and luteinizing hormone (LH), which in turn control 5727 gonad activity. A variety of neuromediators, peptidic and non-peptidic, are also involved in 5728 the control of GnRH and gonadotropins, such as Kisspeptin (Kiss), gonadotropin-inhibiting 5729 hormone (GnIH), dopamine (DA), norepinephrine (NE), serotonin (or 5-hydroxytryptamine 5730 5-HT), Histamine (HA), gamma-aminobutyric acid (GABA). The pituitary gland, as a relay 5731 between brain neurons and peripheral glands is considered as a vertebrate innovation.

5732

5733 Figure 2. Vertebrate phylogeny with a special focus on whole genome duplications.

5734 Two rounds (1R and 2R) of whole genome duplication (WGD) occurred at the basis of the

5735 vertebrate lineage. Whether the 2nd round occurred before or after (doted line) the emergence

5736 of the cyclostome lineage is still under debate. A third round (3R) of WGD occurred 5737 specifically in the teleost lineage, and is also referred to as TWGD (teleost-specific whole 5738 genome duplication). Human belong to mammalian, amniote, tetrapod, sarcopterygian 5739 lineages. Coelacanth is a representative species of basal sarcopterygians. Actinopterygians are 5740 the sister clade of sarcopterygians.

5741

5742 Figure 3. Metazoan phylogeny.

5743 Human belong to the vertebrate, chordate, deuterostome, bilateria lineages. Urochordates are 5744 considered as the sister clade of vertebrates. Among bilateria, protostomes are the sister clade 5745 of deuterostomes. The most consensual classifications of protostomes distinguish two sister

5746 clades, lophotrochozoa and ecdysozoa. Cnidaria represent one of the basic groups of metazoa.

5747

5748 Figure 4. Schematic representation of the glycoprotein hormone (GpH) α and β subunits 5749 and comparison with predicted GPA2 and GPB5 structures.

5750 Amino acids are represented by circles. Glycosylation chains are represented by Y shape sign. 5751 Position of intron splicing sites is indicated by V shape sign. The disulfide bonds are 5752 symbolized by bars, red bars between red circles for those responsible for the cystine-knot 5753 structure and in grey for the additional ones. The conserved cysteine residues are numbered. 5754 Location of the β strands (β 1- β 4) and loops (L1-L3) is indicated.

5755

5756 Figure 5. Schematic representation of the gene duplication, genome duplication and 5757 gene loss events leading to the extant repertoire of glycoprotein hormone subunit related 5758 genes in vertebrates.

5759 1R, 2R: first and second rounds of whole genome duplication in early vertebrates; 3R: third 5760 round of whole genome duplication in early teleosts. 1* The GPA2-GPB5(a) tandem was 5761 maintained in teleosts and basal sarcopterygian (coelacanth). GPB5a was lost in tetrapods. 2* 5762 Thyrotropin β 2 subunit (TSH β 2) was maintained in basal sarcopterygians (coelacanth, 5763 lungfish) and chondrichthyes but lost in teleosts and tetrapods. 3* The LH β subunit encoding 5764 gene was duplicated in primates to generate several copies of the chorionic gonadotropin β 5765 (CG β) subunit genes. 4* After 3R, except for the TSH β 1 subunit, only one of the duplicated 5766 glycoprotein hormone related genes was conserved. 5* Some teleosts may have two copies of 5767 the GPH α subunit resulting from recent, specific tetraploidization events.

5768

```
5769 Figure 6. Evolutionary scenario of the GnRH family in vertebrates (adapted from (219)).
```

5770 Anc GnRH: ancestral gonadotropin-releasing hormone gene. 1R, 2R: first and second rounds

5771 of whole genome duplication in early vertebrates; 3R: third round of whole genome 5772 duplication in early teleosts. Black cross: gene loss. Red arrow: local gene duplication.

5773

5774 Figure 7. Evolutionary scenario of the GnRH receptor family in vertebrates (adapted 5775 from (644)).

5776 Anc GnRHR: ancestral gonadotropin-releasing hormone receptor gene. 1R, 2R: first and 5777 second rounds of whole genome duplication in early vertebrates; 3R: third round of whole 5778 genome duplication in early teleosts. Black cross: gene loss. Red arrow: local gene 5779 duplication.

5780

5781 Figure 8. Two evolutionary scenarios of the GnRH-like systems in bilaterians (adapted
5782 from (654)).

5783 Anc: ancestral; GnRH: Gonadotropin-releasing hormone; CRZ: corazonin. AKH: 5784 adipokinetic hormone; ACP: AKH/corazonin-related peptide. Black cross: gene loss. Red 5785 arrow: local gene duplication. 5786

- 5787 Figure 9. Evolutionary relationships between RF-amide and non RF-amide peptide 5788 families.
- 5789 RF-amide peptides are encoded by 5 distinct genes in mammals, which have been named by 5790 Dockray (153) *farp-1* to 5, based on the chronology of their discoveries. Various
- 5791 nomenclatures are used according to authors and phyla such as: Neuropeptide F (NPFF) or
- 5792 PQRF for *farp-1*; Prolactin-releasing peptide (PrRP) for *farp-2*; Gonadotropin-inhibiting
- brmone (GnIH) or Neuropeptide VF (NPVF) or LPXFR for *farp-3*; Kiss for farp-4; QRFP or
- 5794 26RF for *farp-5*.
- 5795 The figure displays some current hypotheses on their evolutionary relationships (for reviews:
- 5796 (359, 567, 807, 904)):
- 5797 NPFF (farp-1) and GnIH (farp-3) would be the closest related RF-amide peptide families,
- 5798 likely having a common PQRF ancestor and resulting from one of the two rounds of whole
- 5799 genome duplication (1R/2R WGD) that occurred in early vertebrates.
- 5800 QRFP (farp-5) would be related to NPFF (farp-1) and GnIH (farp-3) families via a local gene
- 5801 duplication of their common ancestor, before the WGD duplication of NPFF and GnIH.
- 5802 PrRP (farp-2) and Kiss (farp-4) would be more related to non RF-amide peptide families than
- 5803 to the other RF-amide peptide families: PrRP to neuropeptide Y (NPY) and QRFP to galanin
- and spexin, via local gene duplications, respectively.
- 5805

5806 Figure 10. Evolutionary scenario of the Kiss family in vertebrates (adapted from (587)).

- 5807 Anc Kiss: ancestral Kisseptin gene. 1R, 2R: first and second rounds of whole genome
- 5808 duplication in early vertebrates; 3R: third round of whole genome duplication in early
- 5809 teleosts. Black cross: gene loss.
- 5810

5811 Figure 11. Evolutionary scenario of the Kiss receptor family in vertebrates (adapted from5812 (587)).

5813 Anc KissR: ancestral Kisseptin receptor gene. 1R, 2R: first and second rounds of whole 5814 genome duplication in early vertebrates; 3R: third round of whole genome duplication in early 5815 teleosts. Black cross: gene loss.

5816

5817 Figure 12. Ancient origin of neuromediators involved in the neuroendocrine control of5818 reproduction.

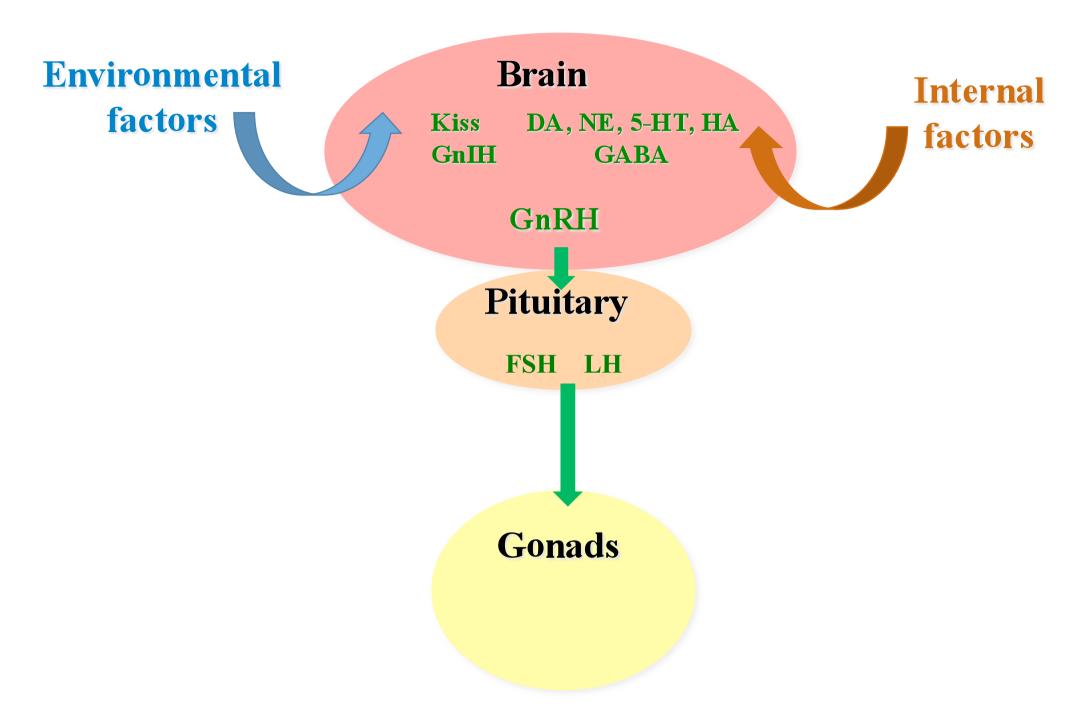
Various neuromediators such as Dopamine (DA), norepinephrine (NE), serotonine (or 5hydrotryptamine, 5-HT), histamine (HA), gamma-aminobutyric acid (GABA) are involved in the brain control of reproduction in mammals. Their presence in other vertebrates, in other deuterostomes, in protosomes and in cnidaria (non-bilateria), suggests an ancient origin in metazoa. They may also have exerted an early role in reproduction in metazoa. Straight letters: neuromediator with a reported role in the control of reproduction. Italic letters with guestion mark: neuromediator present but no information on its putative role in reproduction.

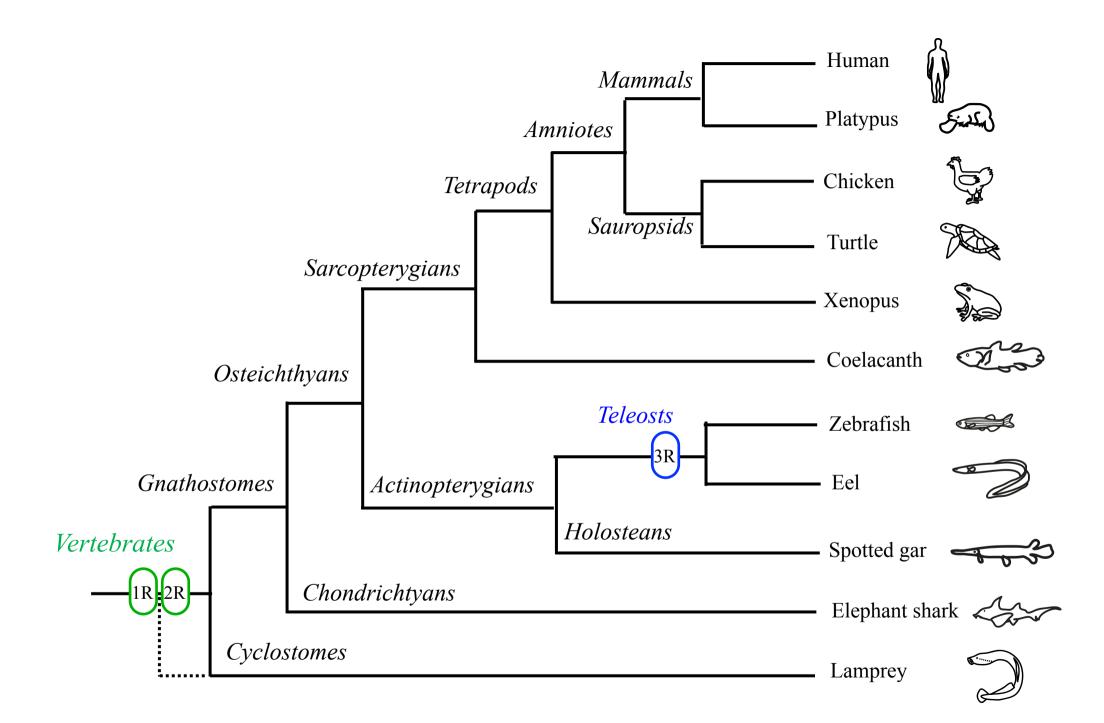
5826

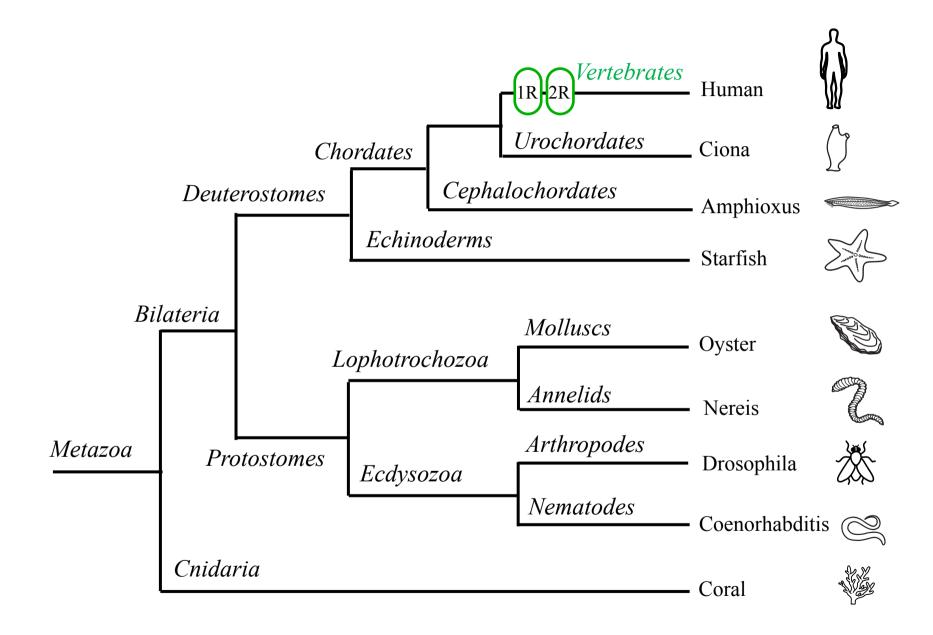
5827 Figure 13. Schematic drawing of the origin of some major actors of the brain-pituitary 5828 control of mammalian reproduction.

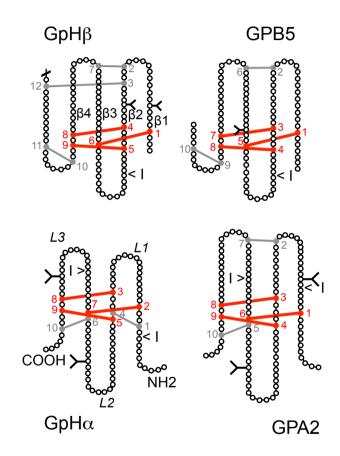
The pituitary gland is a vertebrate innovation ensuring the relay between the brain and peripheral glands and target organs. Molecular families of the various actors of the vertebrate brain-pituitary reproductive neuroendocrine axis have an ancient origin, largely preceding the emergence of the vertebrate lineage. Local and whole genome duplications in vertebrates contributed to the diversification of the Kiss and GnRH (Gonadotropin-releasing hormone) systems and of the glycoprotein (GP) homones, pituitary gonadotropins FSH and LH, and chorionic gonadotropin CG. 1R, 2R: first and second rounds of whole genome duplication in

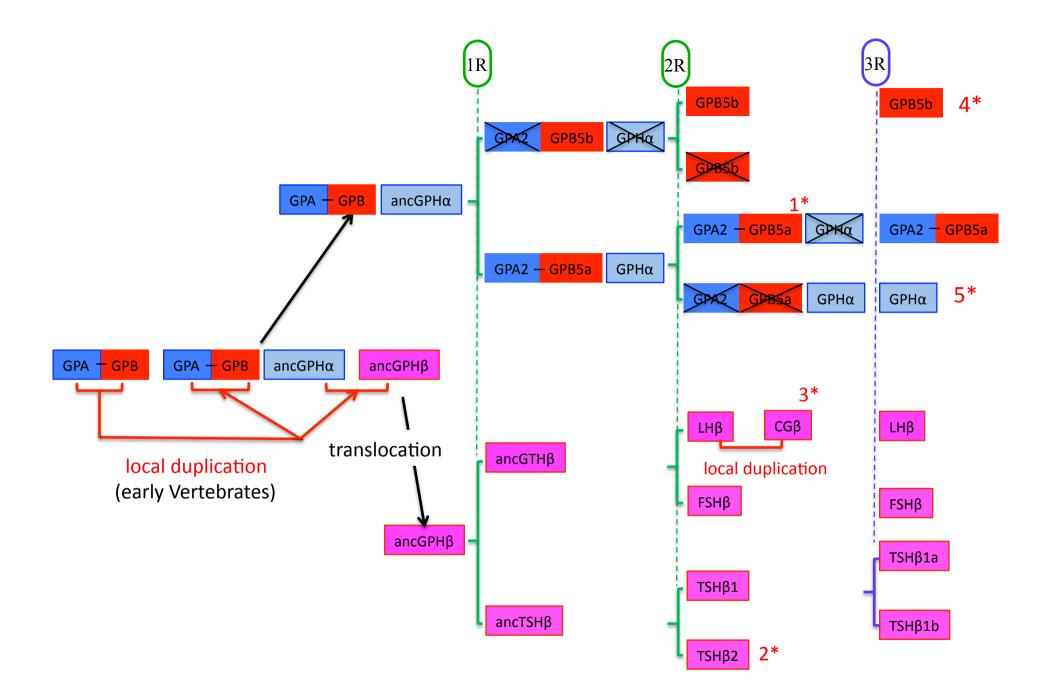
- 5836 early vertebrates; 3R: third round of whole genome duplication in early teleosts. Red bar:
- 5837 local gene duplication.
- 5838
- 5839 Table legends
- 5840 Table 1. Sequences of GnRH-like peptides
- 5841

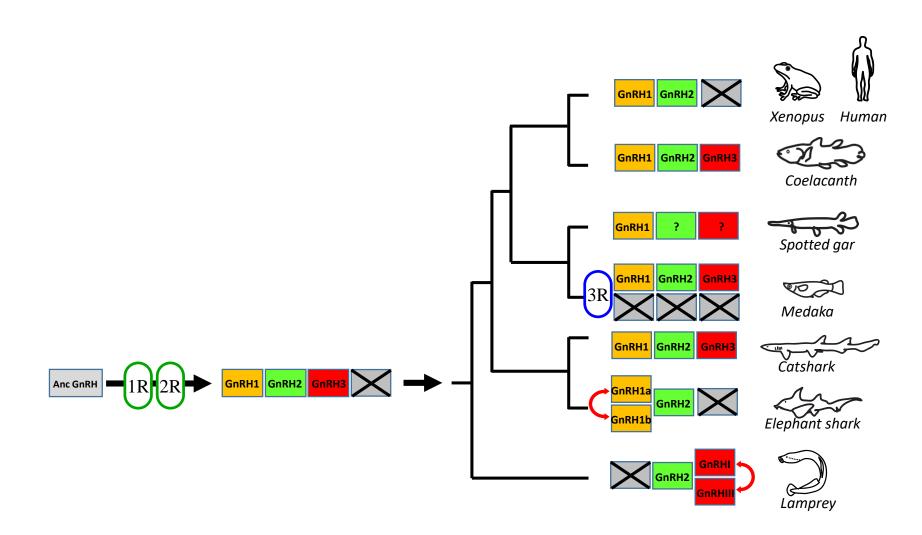




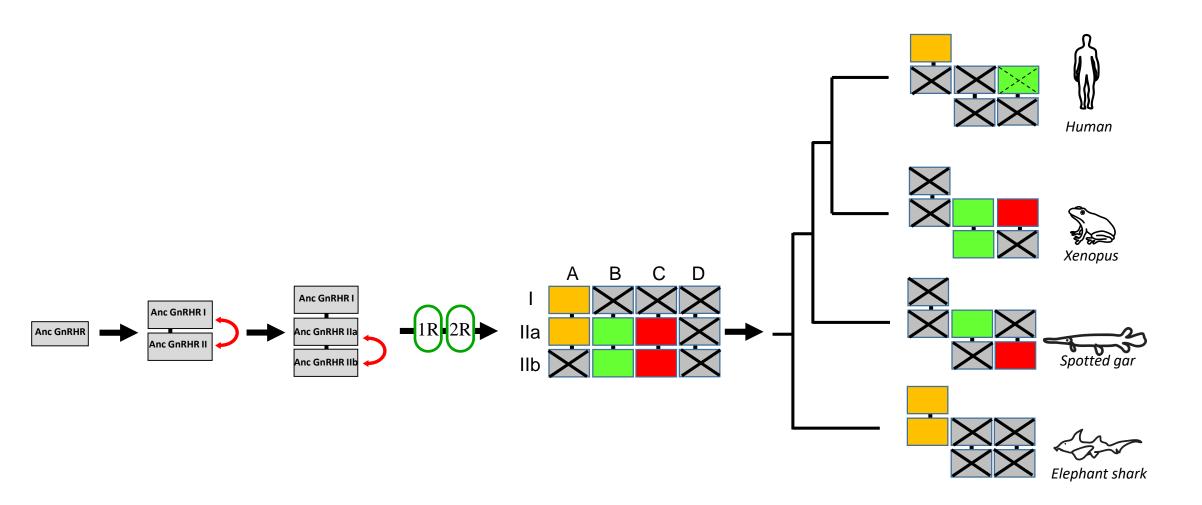


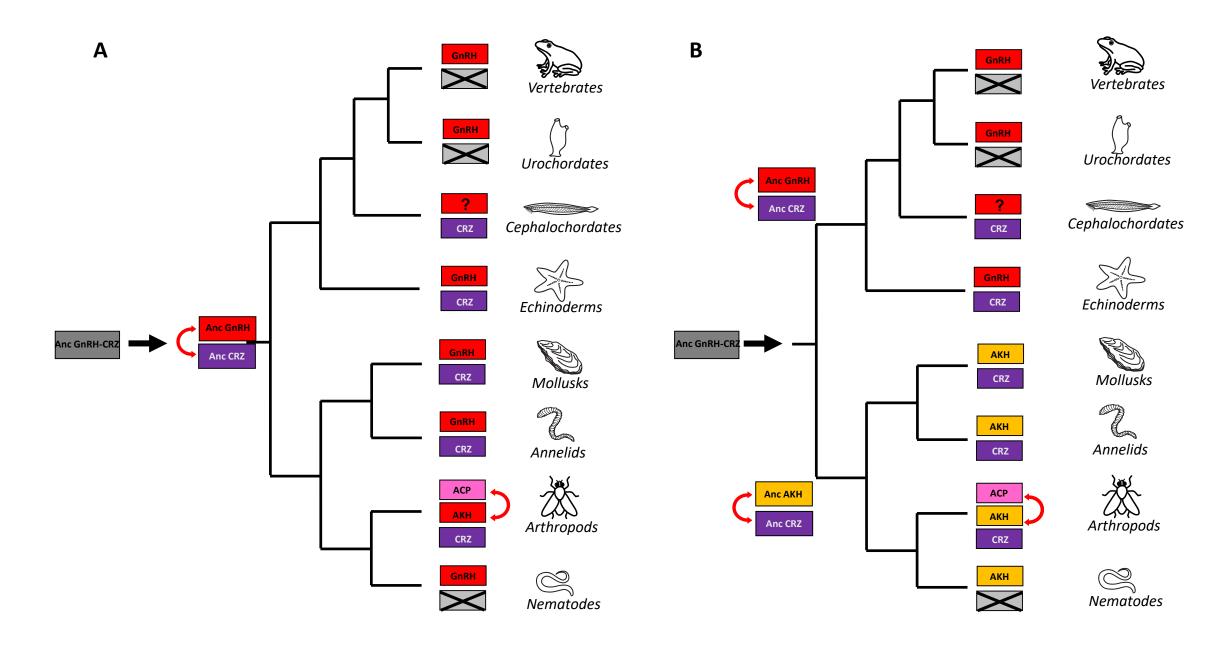






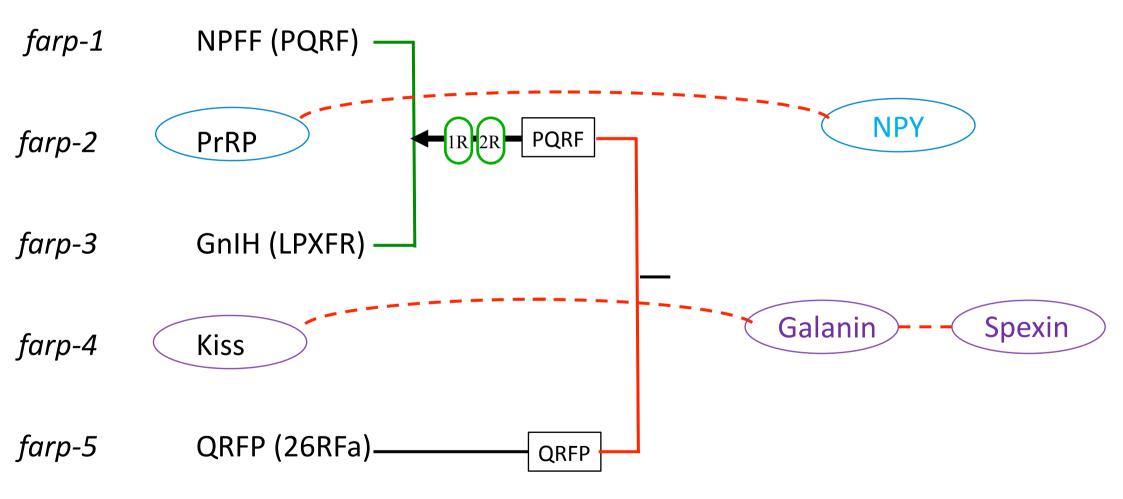
Downloaded from www.physiology.org/journal/physrev at Univ of Massachusetts (128.119.168.112) on October 20, 2019.

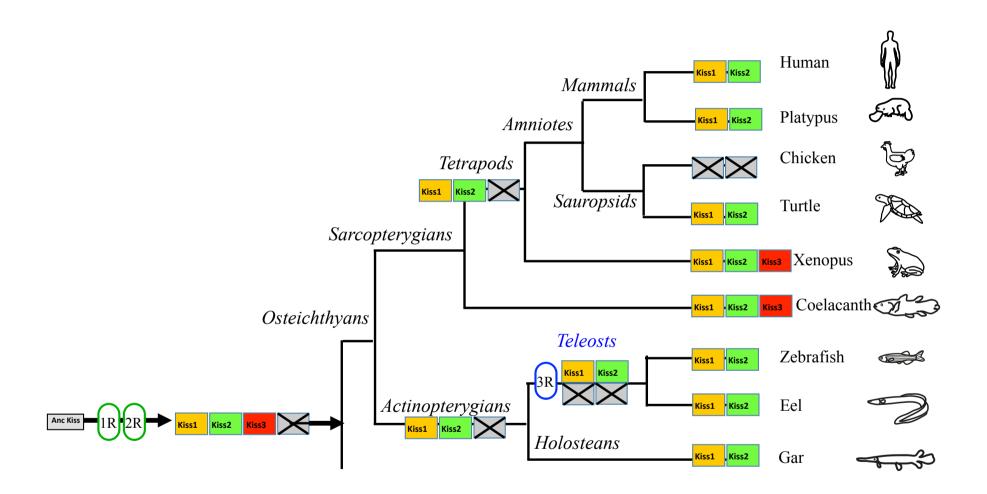


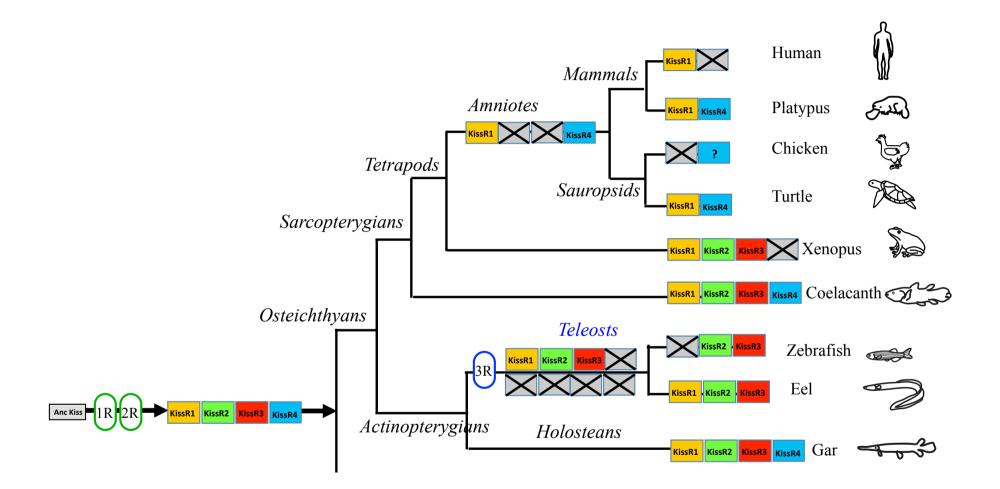


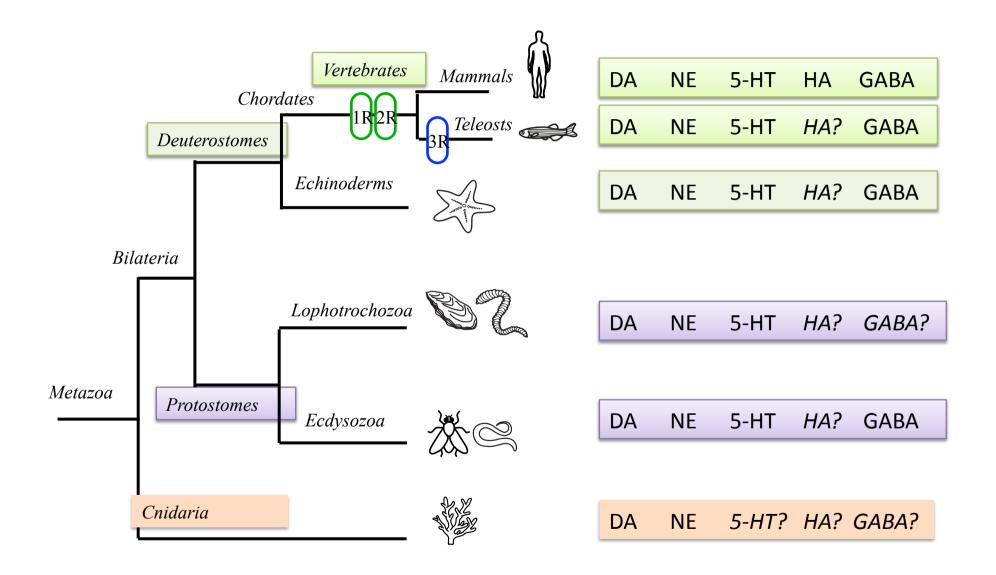
RF-amide peptide families

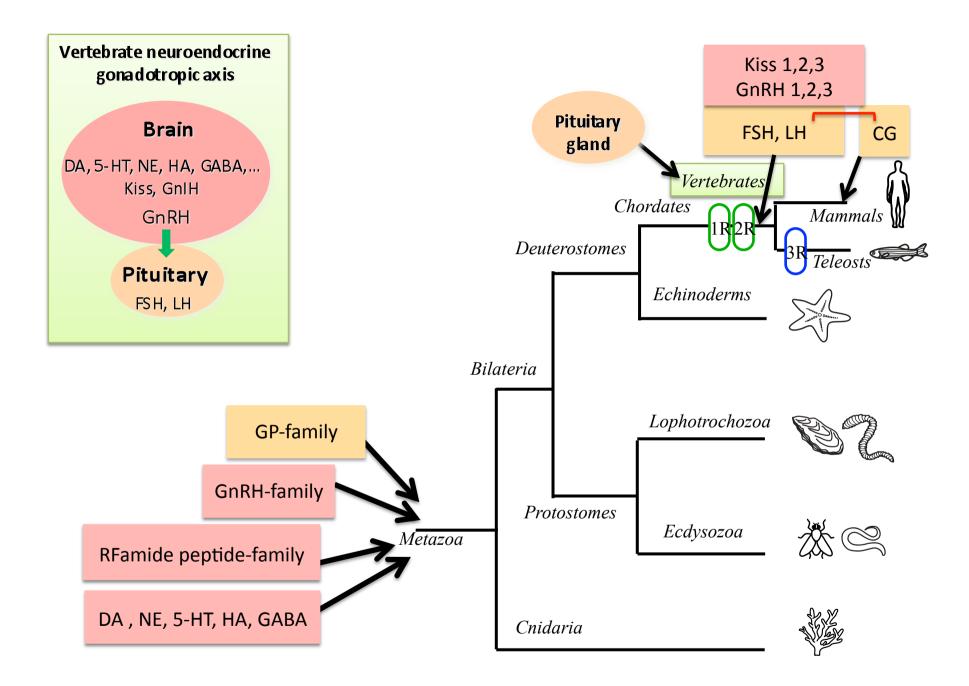
non RF-amide peptide families











Sequences

Species

Original name

Revised name

according to 905

Vertebrate	GnRHs
------------	-------

Vertebrate GnRHs				
	GnRH1	pQHWSYGLRPGa	most mammals	mGnRH
		pQHWSYGVRPGa	Guinea pig	gpGnRH
		pQHWSYGLQPGa	Chicken	cGnRH-I
		pQHWSYGLWPGa	Frog	frGnRH
		pQHWSYGLSPGa	Sea bream	sbGnRH
		pQHWSFGLSPGa	Medaka	mdGnRH
		pQHWSHGLSPGa	Herring	hgGnRH
		pQHWSHGLNPGa	Catfish	cfGnRH
		pQHWSYGMNPGa	Whitefish	whGnRH
	GnRH2	pQHWSHGWYPGa	Chicken	cGnRH-II
		pQHWSHGWFPGa	Lamprey	lGnRH-II
	GnRH3	pQHWSYGWLPGa	Salmon	sGnRH
		pQHWSHGWLPGa	dogfish (catshark)	df-GnRH
		pQHYSLEWKPGa	Lamprey	lGnRH-I
		pQHWSHDWKPGa	Lamprey	lGnRH-III
Tunicate GnF	RHs			
		pQHWSDYFKPGa	Chelysoma	tGnRH-1
		pQHWSLCHAPGa	Chelysoma	tGnRH-2
		pQHWSYEFMPGa	Ciona	tGnRH-3
		pQHWSNQLTPGa	Ciona	tGnRH-4
		pQHWSYEYMPGa	Ciona	tGnRH-5
		pQHWSKGYSPGa	Ciona	tGnRH-6

pQHWSDYFKPGa	Chelysoma	tGnRH-1
pQHWSLCHAPGa	Chelysoma	tGnRH-2
pQHWSYEFMPGa	Ciona	tGnRH-3
pQHWSNQLTPGa	Ciona	tGnRH-4
pQHWSYEYMPGa	Ciona	tGnRH-5
pQHWSKGYSPGa	Ciona	tGnRH-6
pQHWSYALSPGa	Ciona	tGnRH-7
pQHWSLALSPGa	Ciona	tGnRH-8
pQHWSNKLAPGa	Ciona	tGnRH-9
pQHWSYGFSPGa	Halocynthia	tGnRH-10
pQHWSYGFLPGa	Halocynthia	tGnRH-11

Cephalochordate GnRHs

pQILCARAFTYTHTWa	Amphioxus	AmphGnRH	CRZ
pQEHWQYGHWYa	Amphioxus	AmphGnRH-v	?

Hemichordate CRZ			
	pQPHFSLKDRYRWKPa	Acorn worm	skCRZ-like
Echinoderm GnRHs			
	pQVHHRFSGWRPGa	Sea urchin	SpGnRH
	pQIHYKNPGWGPGa	Starfish	ArGnRH
Echinoderm CRZs			
	HNTFSFKGRSRYFPa	Sea urchin	SpCRZ
	HNTFTMGGQNRWKAGa		ArCRZ
	~~~~		
Nematode AKH-GnRH			
		Caenorhabditis	Colur-Capu
	þý miribýwa	Caenornaburtis	CEARC-GIRN
Arthropod AKHs			
		Centipede	
		Fruit fly	
	pQLTFTSSWGa	Silk worm	BmAKH1
	pQLNFTPWWa	Locust	LmAKH
Arthropod ACPs			
	pQVTFSRDWTPAa	Centipede	SmarACP
	pQVTFSRDWNAa	Mosquito	AgamACP
	pQVTFSRDWNPa	Flour bettle	TCACP
	pQITFSRSWVPQa	Prawn	MroACP
Arthropod GnRH			
	pQQHFRTSHFRPDNVa	Prawn	MroGnRH
Arthropod CRZs			
	pQTFQYSKGWEPa	Centipede	SmarCRZ
	pQTFQYSRGWTNa	most insects	CRZ
	pQTFQYSHGWTNa	Locust	LmCRZ

Mollusk AKHs

?

pQVSFSTNWGSa	Oyster	СдАКН	GnRH
pQIHFSPTWGSa	Owl limpet	LgAKH	GnRH
pQHFSPDWGTa	Aplysia	АрАКН	GnRH

#### Mollusk GnRHs

pQNYHFSNGWHPGa	Octopus	OctGnRH	CRZ
pQNYHFSNGWQPa	Oyster	CgGnRH	CRZ
pQNFHFSNGWQPa	Yessi scallop	PwGnRH	CRZ
pQNYHFSNGWYAa	Aplysia	ApGnRH	CRZ
pQNYHFSNGWKSa	Owl limpet	LgGnRH	CRZ

#### Annelid GnRHs

pQAYHFSHGWFPa	Capitella	CpGnRH	CRZ
pQAYHFSNGWMPa	Platynereis	PmGnRH-1	CRZ
pQHAHFSTGWTPGFGIa	Platynereis	PmGnRH-2	?
pQSIHFSRSWQPa	Leech	HrGnRH	CRZ

### Annelid AKHs

pQFSFSLPGKWGNa	Platynereis	PmAKH-1	GnRH
pQLTQSLGWGSAGSSa	Platynereis	PmAKH-2	?

