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# 1 Neonicotinoid-induced mortality risk for bees foraging on oilseed rape nectar persists despite

# 2 EU moratorium

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## 16 ABSTRACT

The implication of neonicotinoids in bee declines led in 2013 to an EU moratorium on three neonicotinoids in bee-attractive crops. 17 However, neonicotinoids are frequently detected in wild flowers or untreated crops suggesting that neonicotinoids applied to cereals 18 19 can spread into the environment and harm bees. Therefore, we quantified neonicotinoid residues in nectar from winter-sown oilseed 20 rape in western France collected within the five years under the EU moratorium. We detected all three restricted neonicotinoids. Imidacloprid was detected in all years with no clear declining trend but a strong inter- and intra-annual variation and maximum 21 22 concentrations exceeding reported concentrations in treated crops. No relation to non-organic winter-sown cereals was identified even though these were the only crops treated with imidacloprid, but residue levels depended on soil type and increased with rainfall. 23 24 Simulating acute and chronic mortality suggests a considerable risk for nectar foraging bees. We conclude that persistent imidacloprid 25 soil residues diffuse on a large-scale in the environment and substantially contaminate a major mass-flowering crop. Despite the limitations of case-studies and risk simulations, our findings provide additional support to the recent extension of the moratorium to a 26 permanent ban in all outdoor crops. 27

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29 Keywords: Neonicotinoid, imidacloprid, foraging bees, oilseed rape nectar, risk assessment, environmental fate

#### 31 **1. Introduction**

Neonicotinoids comprise the dominant class of insecticides in the world with a market share of over 25% in 2015 (Casida, 2018). At their launch in the 1990s, they were considered more environmentally friendly than the prevailing insecticides, due to lower application rates and higher specificity to insects (i.e. lower toxicity to vertebras, including humans; Tomizawa and Casida, 2011). Neonicotinoids ensure efficient and lasting protection from insect pests, because the persistent and highly toxic active compounds translocate throughout the plant (Jeschke et al., 2011; Simon-Delso et al., 2015; Van der Sluijs et al., 2013). Relative specificity to insects results mostly from a higher affinity of the active compounds to the nicotinic acetylcholine receptors of insects than those of vertebrates (Tomizawa and Casida, 2005).

Paradoxically, the characteristics that initially contributed to the economic rise and environment-friendly perception of neonicotinoids are also the cause of environmental concerns. High persistence and water solubility do not only ensure systemic protection but also cause neonicotinoids to accumulate in the environment and contaminate ground and surface waters including rivers, lakes and puddles (Limay-Rios et al., 2016; Samson-Robert et al., 2014; Sánchez-Bayo et al., 2016; Schaafsma et al., 2015; Van der Sluijs et al., 2013). Neonicotinoids caused, however, most concern due to the chronic exposure of bees to sublethal doses in pollen and nectar, which led in December 2013 to a European Union-wide moratorium on the use of three neonicotinoids – imidiacloprid, thiametoxam and clothianidin – in bee-attractive crops.

46 The moratorium did, however, not ban these neonicotinoids in crops such as winter-sown cereals or sugar beets from which residues can spread to nearby non-target plants. The contamination of wild flowers at field borders is widespread (Botías et al., 2015; David et 47 48 al., 2016; Long and Krupke, 2016; Tsvetkov et al., 2017) with concentrations sometimes exceeding those of the treated crop (David et 49 al., 2016). Wild flowers can in fact be the main source of neonicotinoid in bee-collected pollen (David et al., 2016; Tsvetkov et al., 50 2017). Hazardous neonicotinoid concentrations were also found in flowers and honeybee-collected nectar of untreated oilseed rape 51 (Henry et al., 2015; Thompson et al., 2013; Woodcock et al., 2017). In fact, in the year after the moratorium came into effect, 52 maximum neonicotinoid prevalence in UK honey samples coincided with oilseed rape flowering and residue concentrations increased 53 with the area of oilseed rape surrounding the regarded honeybee hives (Woodcock et al., 2018) indicating that the crop took up soil 54 residues that persisted for more than a year.

Contamination of non-target plants can originate from treated crops in the surroundings or from the cultivation of a treated crop on the same field in previous years (Henry et al., 2015; Wood and Goulson, 2017). Neonicotinoid seed-dressed crops take up only a small portion of the active ingredient (~5%); the remainder stays on the field unless it is transported by wind or water (Wood and Goulson, 2017). Neonicotinoids are persistent in the environment with half-lives in aerobic soil conditions (Van der Sluijs et al., 2013) ranging from a few months to years (Bonmatin et al., 2015). Consequently, neonicotinoids are often found in soil, sometimes even several years after applications ceased (Hladik et al., 2018; Jones et al., 2014). Neonicotinoids in agricultural soils do not only pose a direct threat to ground-nesting bees (Chan et al., 2019), but can also be taken up by plants and then threaten foraging bees.

62 To assess neonicotinoid spread in the environment more extensively, we examined nectar collected from a total of 291 winter-sown oilseed rape fields over the five years under the EU moratorium (2014-2018) using neonicotinoid residue analyses with particularly 63 low limits of detection and quantification (Martel et al., 2013). The fields, located in a 435 km<sup>2</sup>-large agricultural study area in western 64 France (Fig. 1), could not be sampled in multiple years due to crop successions, but were sampled 1-6 times during one oilseed rape 65 flowering period. We related the prevalence (presence/absence) and concentration of imidacloprid, the most prevalent neonicotinoid in 66 67 our study, to temporal and environmental (soil type, weather, land use) variables to identify the main drivers of accidental imidacloprid contamination in a mass-flowering crop. Finally, we simulated imidacloprid-induced acute and chronic mortality for 68 nectar foraging honeybees, bumblebees and solitary bees using a risk assessment scheme adapted from the one used by the European 69 70 Food Safety Authority (EFSA) in order to identify whether the EU restrictions on neonicotinoid use in bee-attractive crops have 71 eliminated the risk of imidacloprid-induced mortality for bees feeding on oilseed rape nectar.

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## 73 **2.** Materials and methods

#### 74 **2.1.** *The study site*

The study was conducted in the 435 km<sup>2</sup>-large Long-Term Socio-Ecological Research (LTSER) Zone Atelier "Plaine & Val de Sèvre" site, central western France (46°23'N, 0°41'W), which is characterized by a high proportion of arable land and an oceanic climate with mean monthly temperatures ranging from 5-20 °C and 820 mm of precipitation well distributed over the year. In the LTSER site the broad soil type (Fig. 1) and the precise land use have been documented and mapped on vector-based GIS shapefiles since 1994

(Bretagnolle et al., 2018). Within the study period, between 2014 and 2018, the land area was covered on average to 33.0% with wheat
6.1% barley, 1.4% other cereals, 9.7% maize, 9.3% sunflower, 7.2% oilseed rape, 8.0% grassland, 7.1% legumes (mostly alfalfa:
3.0%) and 4.6% other crops. Organic farmland covered 6.5% of the area.

### 82 **2.2.** Oilseed rape nectar sampling

83 Between 2014 and 2018, a total of 291 winter-sown oilseed rape fields were selected for nectar sampling. The field selection criteria differed between years and included flower timing (early and late flowering oilseed rape fields) and crop choice in previous years. 84 Specifically, we strived for uniform distributions of the number of times winter-sown cereals were cultivated in the previous 10 years 85 86 (i.e. from 0-8) or the number of years between the last wheat and the sampled oilseed rape cultivation. To estimate repeatability and 87 temporal variation in neonicotinoid residue concentrations, fields were sampled 1-6 times per year (Fig. S1) on different days between 23 March and 10 June (Table S1) and between 9 a.m. and 7 p.m. (Fig. S2), yielding a grand total of 536 nectar samples. The number 88 89 of samples per field depended on the number of workers available for nectar collection, funding and weather conditions (sampling could not be done during rainy weather or drought). Using 5.0 µL capillaries (Drummond Scientific, Broomall, PA, USA), 90 91 approximately 25 µL of nectar per sample was collected directly from randomly selected open flowers that were at least 10 m from 92 field margins to avoid edge effects.

#### 93 **2.3.** *Neonicotinoid residue analysis*

Around 2  $\mu$ L of nectar were used to determine the sugar content by hand-held refractometers (BS Eclipse BS 45-81 / 45-82, Bellingham + Stanley Ltd., UK). The remaining nectar was analysed using liquid chromatography with electrospray tandem mass spectrometry by the EU reference laboratory for neonicotinoid multi-residual analyses (ANSES, Sophia-Antipolis, France; Martel et al., 2013) to identify (limit of detection (LOD) = 0.1 ng mL<sup>-1</sup>) and quantify (limit of quantification (LOQ) = 0.3 ng mL<sup>-1</sup>) the five neonicotinoids that were approved in the EU during the study period: acetamiprid, clothianidin, imidacloprid, thiacloprid, thiamethoxam. In this study, we focus on the three neonicotinoids that were banned in bee-attractive crops (clothianidin, imidacloprid & thiamethoxam) to determine whether the EU moratorium has eliminated their risk for bees foraging on oilseed rape nectar.

Daily minimum and maximum air temperature and precipitation data were obtained from a weather station held by Météo France south of Niort and within the LTSER site, with data made available at the US National Oceanic and Atmospheric Administration. The total precipitation and growing degree days (oilseed rape: base temperature = 5 °C; wheat: base temperature = 0 °C (Ruiz Castillo and Gaitán Ospina, 2016)) over the growing seasons of the sampled oilseed rape and of wheat cultivated in the preceding year were calculated and related to imidacloprid prevalence. The oilseed rape growing season was estimated to range from 1 August of the previous year to the mean sampling day of the sampling year. The wheat growing season ranged from 1 October two years before nectar sampling to 1 July of the year before nectar sampling.

110 **2.5.** *Statistical analyses* 

111 Among the neonicotinoids restricted from use in bee-attractive crops, only imidacloprid was detected frequently enough to allow for 112 meaningful statistical analyses. Both the prevalence (absence/presence) and concentration of imidacloprid were analysed. For the latter, only the concentrations of positive samples were included (i.e. samples <LOD were excluded) to assess effects on concentration 113 independently of effects on prevalence and to avoid zero-inflated datasets. However, to exclude artefacts that may arise when residues 114 below the LOD are undetected and prevalence and concentration in positive samples show opposing trends, we refit the selected 115 116 model on concentration with the whole dataset (i.e. including negative samples) and report these results in Table S2 and Fig. S3. Samples  $\leq$  LOQ but > LOD were set to 0.2 ng mL<sup>-1</sup> as determined by the equation (LOD+LOQ)/2, which is a more conservative 117 assumption than the one used in the EFSA first-tier risk assessment in which LOQ is assigned to samples < LOQ but > LOD (EFSA, 118 2013). In the analyses on the whole dataset, samples  $\langle LOD \rangle$  were set to 0.05 ng mL<sup>-1</sup> (i.e. LOD/2). Analyses of repeatability of 119 imidacloprid prevalence and concentration of different samples from the same field were restricted to the years 2016, 2017 and 2018 120 121 as typically only one sample per field was taken in 2014 and 2015. Repeatability was estimated by the intraclass coefficient (ICC) using the rpt function of the rptR package in R (Stoffel et al., 2017). ICCs were estimated on the logit link scale for prevalence and on 122 123 the normal scale for logarithmically transformed  $(log_{10})$  concentration in positive samples.

The relationship between imidacloprid residues and temporal parameters (sampling year, sampling Julian day number, sampling time of day) and environmental parameters (soil type, precipitation, mean air temperature, non-organic winter-sown cereal cultivation in

126 the surrounding of the focal fields or on the same field in previous years) were analysed using generalized linear mixed-effects models 127 with field identity as a random factor, a logit-link function and a binomial error distribution. For the concentration data, a linear model 128 without random factors (LM, with Gaussian error distribution) was used, as the number of positive samples per field was too low to 129 allow for the inclusion of a random effect for field identity. Akaike Information Criterion (AIC) was used to select the most 130 parsimonious models. First, full models for prevalence and concentration were built that contained sampling year, sampling Julian day 131 number (incl. a quadratic effect), sampling time of day (incl. a quadratic effect) as fixed effects and the percentage of area covered by non-organic winter cereals in 20 m buffers surrounding the sampled fields in the sampling year. The full models contained additional 132 fixed effects describing (i) precipitation, (ii) mean temperature, (iii) soil type, and (iv) non-organic winter cereal cultivation in 133 134 previous years on the sampled fields, which were pre-selected in a comparison of a range of univariate models (i.e. with only one 135 fixed effect) with a null model (i.e. without fixed effects). The compared variables were (i) the amount of precipitation in periods of 1-136 10 days that ended 0-4 days before sampling day (hereafter 'lag'), (ii) mean air temperature in periods of 1-5 days with a lag of 0-3 137 days, (iii) soil type distinguishing between red and calcareous soil and soil type distinguishing between red, shallow calcareous and 138 deep calcareous soil and (iv) the number of years passed since the last non-organic cereal cultivation and the number of non-organic cereal cultivations in the previous 1-5 years. For each of the four categories, one variable was included in the full models as long as 139 140 the variables yielded a lower AIC than the regarded null model. In addition, two-way interactions between soil type and weather 141 variables were included in the full models. The two full models were then compared to all possible reduced models with the same 142 random effect (i.e. field identity for prevalence and none for concentration in positive samples) except that models with quadratic 143 terms or two-way interactions were only considered if they also contained the non-quadratic term or the main effects of these 144 variables. Explained variability of selected models was determined by coefficients of determination or their equivalent for models fit on (restricted) maximum likelihood. These pseudo  $R^2$  values were determined using Nagelkerke's method. 145

All analyses were done in R version 3.5.0. GLMMs for model selection were fit using glmmTMB of the glmmTMB package. However, for illustrations the selected model was re-fit using the glmer function of the lme4 package because glmmTMB does currently not allow to set random effects to zero. Pseudo coefficients of determination for GLMMs were obtained using the r.squaredGLMM function of the lmtest package. Both marginal and conditional pseudo coefficients of determination are reported to show the theoretical variance explained by fixed factors only  $(R^2_m)$  and by the entire model  $(R^2_c)$ , respectively.

This risk assessment was restricted to nectar foragers feeding on imidacloprid residues as thiamethoxam and clothianidin were only infrequently detected. Hereby, samples <LOD were set to 0.05 ng mL<sup>-1</sup> (i.e. LOD/2). To determine the sensitivity of the risk estimates to this assumption, we report in Supplementary Data 1 also risk estimates when samples <LOD were set to zero or to the value of the LOD.

156 As EFSA is responsible for assessing the risk of plant protection products in the EU, we adapted their first-tier assessment for bees using our measured field-level imidacloprid residue and sugar content values rather than theoretical values. Similarly to EFSA, we 157 158 computed exposure toxicity ratios (ETRs) for acute and chronic toxicity for 1000 hypothetical individual honeybees, bumblebees or 159 solitary bees per field (EFSA, 2014, 2013). ETRs constitute the quotient of an expected environmental dose (i.e. the imidacloprid 160 residue intake) and a median lethal dose. However, to obtain easily interpretable risk estimates, we refrained from comparing ETRs to 161 EFSA's trigger values that indicate potentially unsafe levels in a worst-case scenario and are based on the lowest observed background mortality and arbitrary assessment factors. Instead, we related ETRs to a probability of death, which we used to simulate acute and 162 163 chronic mortality for individual honeybees, bumblebees and solitary bees foraging for 10 days on oilseed rape nectar to determine whether imidacloprid exposure through oilseed rape can shorten their lifespans. To obtain estimates for each field, we assumed that 164 165 bees would forage throughout the regarded timeframe on the same oilseed rape field. In fact, honeybees forage up to 10 days in their 166 lives (Schippers et al., 2006), while bumblebees and solitary bees can forage over even longer periods (Evans et al., 2017; Michener, 2007). Honeybees, bumblebees and some solitary bees forage intensively on oilseed rape during its bloom (Baron et al., 2017; 167 168 Holzschuh et al., 2016; Magrach et al., 2018; Perrot et al., 2018; Rollin et al., 2013; Stanley et al., 2013) and exhibit flower constancy 169 (Amaya-Márquez, 2009; Gegear and Laverty, 2005; Grüter et al., 2011).

In a first step, neonicotinoid concentrations were converted to mass fractions in ppb (i.e.  $\mu g k g^{-1}$ ), by dividing them by the density of the nectar sample,  $\rho_{nectar}\rho_{nectar}$  (in kg L<sup>-1</sup>), which was estimated by the sugar content  $\omega_{sugar}\omega_{sugar}$  (in kg kg<sup>-1</sup>), and assumed densities of water (1 kg L<sup>-1</sup>) and sugar (1.6 kg L<sup>-1</sup>) as follows:

173 
$$\rho_{\text{nectar}} = (1 + 0.6 \,\omega_{\text{sugar}}) \,\rho_{\text{nectar}} = (1 + 0.6 \,\omega_{\text{sugar}})$$
 (equation 1).

For each of the three regarded bee types (*Apis mellifera*, bumblebees and solitary bees), a theoretical normal distribution of daily sugar consumption amounts was derived from reported ranges of daily sugar consumption (in mg; Fig. S4; EFSA, 2013; Rortais et al., 2005). The normal distributions were centred on the mean of the reported minimum and maximum sugar consumption amounts and standard deviations were estimated by a quantile function (qnorm) with alpha being set to 0.01, so that 99% of the estimated daily sugar consumption amounts were within the reported ranges (EFSA, 2014):

180 s. d. = 
$$\frac{\text{mean-min}}{\text{qnorm}(1-\text{alpha})}$$
 s. d. =  $\frac{\text{mean-min}}{\text{qnorm}(1-\text{alpha})}$  (equation 2).

For 1000 bees per field and bee type, we randomly selected daily sugar consumption amounts from these probabilistic distributions and imidacloprid concentrations with corresponding sugar content values from the available measures (if a field was more than once sampled) to calculate daily residue intake ( $\mu$ g bee<sup>-1</sup>) as follows (EFSA, 2013):

184 
$$daily residue intake = \frac{neonicotinoid mass fraction}{sugar content} \times \frac{daily sugar consumption}{\frac{10^6 mg}{kg}} daily residue intake = \frac{neonicotinoid mass fraction}{sugar content} \times \frac{10^6 mg}{\frac{10^6 mg}{kg}}$$
  
185  $\frac{daily sugar consumption}{\frac{10^6 mg}{kg}}$  (equation 3).

186 Acute and chronic ETRs for each bee type and field were calculated for the periods in which the corresponding lethal doses were 187 determined (i.e. one day for the acute  $LD_{50}$  on bumblebees, two days for the acute  $LD_{50}$  on honeybees and solitary bees and 10 days for chronic LDD<sub>50</sub> on any bee type; Table 1). For this, residue intake over the regarded period was divided by the corresponding 188 189 median lethal dose (expressed as an absolute amount for the regarded period rather than a daily amount). Bee mortality was subsequently simulated based on an assumed relationship between probability of death and ETR. For ETR >0.1, probability of death 190 191 was assumed to follow a logistic regression with ETR=0.1, corresponding to 10% mortality (Sanchez-Bayo and Goka, 2014) and 192 ETR=0.5 corresponding to 50% mortality, while for ETR<0.1 no mortality was assumed (Fig. S5). Mortality was simulated in ten 1-193 day periods for acute mortality of bumblebees, in five 2-day periods for acute mortality of honeybees and solitary bees and in one 10-194 day period for chronic mortality. Acute and chronic mortality were combined by considering any bee dead that was simulated to die within any of the ten 1-day periods / five 2-day periods due to acute toxicity or within the 10-day period due to chronic mortality. 195 Finally, we calculated the proportion of bees that would die a premature death due to imidacloprid-induced toxicity per field and 196 197 determined then the proportion of fields that had mortality rates higher than 50%.

## 198 **3. Results**

### 199 **3.1.** Prevalence and concentration of neonicotinoids in oilseed rape nectar

200 All three neonicotinoids that were banned in oilseed rape and other bee-attractive crops throughout the study period were detected at 201 least once within the five years (Fig. 2a). Imidacloprid was detected in all years and in 43% samples overall and was therefore clearly 202 more prevalent than other neonicotinoids including thiacloprid, which was allowed for spray applications on oilseed rape during the study period and detected in 6.7% of samples (Fig. S6). Imidacloprid prevalence varied strongly between years. In 2014 and 2016, 203 204 imidacloprid was detected in over 60% of the samples (Fig. 2a). Due to repeated sampling on the same field, the proportion of imidacloprid-positive fields was even higher, exceeding 90% in 2016. In contrast, in 2015, imidacloprid was detected in only 5.4% of 205 206 the samples/fields. Imidacloprid concentrations spanned also a wide range. Although 96.5% of all samples (i.e. 91.8% of positive samples) contained less than 1 ng mL<sup>-1</sup> imidacloprid, extremely high concentrations of up to 70 ng mL<sup>-1</sup> were detected in a few 207 208 samples (Fig 2b). Imidacloprid prevalence was generally fairly repeatable while concentrations in positive samples of the same field varied more (Table 2). In fact, all of the 19 samples with imidacloprid concentrations above 1 ng mL<sup>-1</sup> stemmed from different fields 209 210 although these fields were sampled on average three times and tested positive in 79% of the cases.

## 211 **3.2.** Relation between neonicotinoid residues and weather, time, soil type or cereal cultivation

212 We expected that temperature and precipitation influence the degradation and uptake of neonicotinoids. However, there was no clear link between inter-annual differences in imidacloprid prevalence and weather conditions (growing degree days and precipitation) in 213 214 the oilseed rape growing season or the preceding wheat growing season (Fig. S7). To determine which factors influenced imidacloprid prevalence and concentration (in positive samples only), we conducted model selection analyses using variables on sampling time 215 216 (year, Julian day number and time of day), non-organic cereal cultivation (in the surrounding of the focal fields or on the focal fields 217 but in previous years), weather (precipitation and air temperature), soil type and two-way interactions between weather variables and soil type. The selected models explained a substantial amount of variation for both prevalence ( $R^2_m$ =0.42 (only fixed effects),  $R^2_c$ =0.61 218 219 (including field identity as random factor)) and concentration in positive samples ( $R^2=0.27$ ). Imidacloprid prevalence tended to 220 decrease with mean air temperature and both imidacloprid prevalence and concentration (in positive samples) were related to year, 221 tended to decrease with Julian day and increase with precipitation in the days before sampling, although the regarded period of rainfall

222 was considerably longer for prevalence than for concentration (Fig. 3, Table S2). Both imidacloprid prevalence and concentration 223 were also higher on red soil (i.e. brunisol on ferralitic clay) than on calcareous soil with no differences between shallow and deep 224 calcareous soil. In addition, the increase in imidacloprid concentration with precipitation was particularly pronounced in oilseed rape 225 grown on red soil, as indicated by the two-way interaction between soil type and precipitation (Fig. 3; Table S2). However, this was 226 only true for concentration in positive samples. Including samples below the limit of detection also suggests that imidacloprid residues 227 were higher on red than on calcareous soil, but only on calcareous soil imidacloprid residues increased with rainfall in the regarded 228 period before sampling (Table S2, Fig. S3). Despite being the only crops treated with imidacloprid, non-organic winter cereals cultivated in the surroundings of the regarded oilseed rape fields or on the same fields but in previous years had no measurable impact 229 230 on imidacloprid prevalence or concentration in those samples tested positive for the substance.

231 **3.3.** *Risk to foragers* 

232 We simulated the risk of imidacloprid-induced mortality for individual honeybees, bumblebees and solitary bees foraging on oilseed rape nectar over a period of 10 days using a scheme that has been adapted from EFSA's first-tier risk assessment (EFSA, 2014, 2013) 233 234 to obtain easily interpretable risk estimates per field based on our measured data. For honeybees, the risk peaked in 2014 and 2016 (Fig. 4), with an estimated 50% of nectar foragers likely to die due to imidacloprid in 12% of fields in 2014 and 2016 (i.e. 9 and 7 235 236 fields, respectively), and 5% of fields in the whole study period. For nectar-foraging bumblebees, we determined in 2014 and 2016 at 237 least ten fields (13% and 20% of fields, respectively) and in 2018 one field with an estimated mortality of above 50% (Fig. 4, Supplementary Data 1). For solitary bees, the risk assessment indicated that in one field in 2016 half of the bees would die due to 238 239 acute toxicity and in above 10% of fields in 2014 and 2016 (6 and 9 fields, respectively) due to chronic toxicity. Our estimates suggest 240 that unlike honeybee mortality, wild bee mortality was mostly driven by chronic toxicity (Fig. 4). However, estimates of chronic 241 mortality in bumblebees and solitary bees are based on dietary lethal doses that have been extrapolated from honeybees by EFSA 242 using a safety factor of 10 (Table 1; EFSA, 2018). In addition, acute mortality of solitary bees was estimated using a lethal dose for 243 contact rather than oral exposure (Uhl et al., 2018).

### 244 **4.** Discussion

We found that neonicotinoid contamination of oilseed rape nectar is widespread despite EU-wide restrictions on the application of clothianidin, imidacloprid and thiamethoxam in bee-attractive crops. In particular imidacloprid was found in a varying and sometimes considerable fraction of fields in all years. The prevalence of neonicotinoid contamination varied strongly between years, but with no clear decline over time since the EU moratorium came into effect in December 2013. We detected imidacloprid not only frequently but sometimes also in very high concentrations. In two samples collected in 2016, over 45 ppb (52 ng mL<sup>-1</sup> and 70 ng mL<sup>-1</sup>) imidacloprid was found, which is more than five times the expected maximum concentration in nectar of imidacloprid-treated oilseed rape (Wood and Goulson, 2017).

During the study period, imidacloprid use was restricted in the study site to the seed-treatment of non-organic winter-sown cereals and even before the moratorium, imidacloprid was never used in oilseed rape and banned for seed-treatments in sunflower (1999) and maize (2004) in France (Maxim and van der Sluijs, 2007). However, we did not find any relation between imidacloprid residues in oilseed rape and the cultivation of non-organic winter-sown cereals in previous years on the sampled fields or in the sampling year on surrounding fields.

257 This suggests that imidacloprid spreads on a large scale in the environment, contaminating not only wild flowers at field borders of 258 treated crops but also other crops that were planted outside the immediate vicinity or several years after the application of the 259 insecticide. Neonicotinoids can travel from treated crops to insect-pollinated plants at the moment of sowing through contaminated 260 dust (Girolami et al., 2013; Greatti et al., 2006; Krupke et al., 2012; Pistorius et al., 2010; Tapparo et al., 2012) or later on, through wind eroded soil (Limay-Rios et al., 2016; Schaafsma et al., 2015). Contaminated dust can contain extremely high concentrations of 261 262 neonicotinoids (Bonmatin et al., 2015; Krupke et al., 2017; Wood and Goulson, 2017). Therefore, it is conceivable that dust drift 263 caused the extremely high imidacloprid concentrations detected in two oilseed rape nectar samples of our study, although winter 264 cereals in neighbouring fields were sown at least half a year before sampling. However, only a small proportion of neonicotinoid is 265 released as dust (<2% in maize seed-coating; Tapparo et al., 2012) and dust drift from cereals is small compared to maize, which was 266 not allowed to be treated with imidacloprid (Wood and Goulson, 2017). In addition, the adoption of improved seed drills in recent 267 years has effectively limited dust drift (Wood and Goulson, 2017).

268 A more likely mechanism of large-scale imidacloprid spread is transport by water in leachate, run-off or contaminated irrigation water 269 (Bradford et al., 2018; Huseth and Groves, 2014; Kurwadkar et al., 2014), which neonicotinoids are prone to as they have to be water-270 soluble to be systemic (Bonmatin et al., 2015; Giorio et al., 2017). This is supported by our finding that imidacloprid prevalence and 271 concentration increased with rainfall in the days before sampling and was higher on red soil (i.e. unsaturated brunisol on ferralitic 272 clay) than on calcareous soil. Imidacloprid may be better retained on red soil because of the finer texture, higher content of the clay 273 mineral kaolinite and higher water-holding capacity compared to the calcareous soil. Clay minerals and organic matter increase imidacloprid adsorption (Liu et al., 2002) and also soil texture affects the leaching potential of neonicotinoids, which is highest in 274 275 sandy soils and lowest in loams (Wood and Goulson, 2017). Neonicotinoids are much more persistent under aerobic than anoxic 276 conditions (Giorio et al., 2017). In fact, imidacloprid can persist several years after application as the half-life in soil ranges from 100 277 to 1230 days (Giorio et al., 2017). Large-scale neonicotinoid spread and uptake from contaminated soil is supported by a Switzerland-278 wide survey that showed that neonicotinoids are prevalent in long-standing organic farmland and ecological focus areas with 279 concentrations in soil and plant samples being correlated (Humann-Guilleminot et al., 2019). At a much smaller spatial scale, lateral 280 spread of imidacloprid residues around a horticultural crop, which affected ground-nesting insects, was associated with precipitation in 281 a Japanese study (Sánchez-Bayo et al., 2007). However, although moisture content and soil temperature can influence degradation and 282 leaching in soils (Bonmatin et al., 2015) and imidacloprid residues correlated in our study with rainfall in the days before sampling, 283 inter-annual differences in imidacloprid prevalence could not be explained by air temperature or precipitation in the beginning of the year or in previous years. Although we cannot exclude differences in the cultivation of imidacloprid seed-treated non-organic winter-284 sown cereals between red and calcareous soils, this was likely a minor factor, as imidacloprid was exclusively used as seed-treatment 285 in winter-sown cereals. Since the seeds were commercially sold with an imidacloprid coating, it is unlikely that large differences in the 286 287 application rate of imidacloprid occurred.

We found that the detected imidacloprid concentrations pose a substantial risk to bees foraging on oilseed rape nectar. At 16 of 291 fields, imidacloprid toxicity was estimated to kill 50% of honeybees foraging on oilseed rape nectar for 10 days. For solitary bees and bumblebees such a high mortality rate was determined at 15 and 23 fields, respectively. We acknowledge that our mortality estimates are rough expectations of risk levels rather than empirically supported predictions. We assumed that bees would forage for 10 days exclusively on single oilseed rape fields, for which we took nectar samples at 1-6 dates. This means we did not capture the whole

293 variability of imidacloprid residues, as neonicotinoid levels can vary strongly between fields and within a field on different or even the 294 same date (see results and e.g. Osterman et al., 2019). In addition, not accounting for a potential preference (Kessler et al., 2015) for or 295 avoidance against (Kang and Jung, 2017) imidacloprid may have affected the results (but see also Gels et al., 2002; Larson et al., 296 2013). Besides, many bees likely do not forage exclusively on oilseed rape. However, it is well-established that mass-flowering 297 oilseed rape is attractive to honeybees (Requier et al., 2015; Rollin et al., 2013; Stanley et al., 2013), generalist bumblebees (Baron et 298 al., 2017; Holzschuh et al., 2016; Magrach et al., 2018; Stanley et al., 2013) and solitary bees (Holzschuh et al., 2016; Magrach et al., 299 2018; Perrot et al., 2018) and blooms at a time in the season when honeybee and bumblebee colonies are growing rapidly with consequently high food demands (Requier et al., 2015; Westphal et al., 2009). In addition, individuals of many bee species exhibit 300 301 high flower constancy (Amaya-Márquez, 2009; Gegear and Laverty, 2005; Grüter et al., 2011). Moreover, foraging on alternative 302 floral resources does not necessarily reduce the risk of neonicotinoid exposure. Neonicotinoids are frequently found in wild flowers 303 (Botías et al., 2015; David et al., 2016; Long and Krupke, 2016; Tsvetkov et al., 2017) and in our study oilseed rape fields were 304 contaminated with neonicotinoids that they have not been treated with.

305 The assumed foraging timespan of 10 days is a worst-case scenario for honeybees (Rortais et al., 2005; Visscher and Dukas, 1997). 306 However, wild bees can forage over longer periods (Evans et al., 2017; Michener, 2007), which may imply even higher probabilities of premature death. To translate estimated exposure to mortality rates, we assumed a logistic dose-response curve with 10% mortality 307 308 at ETR=0.1 and 50% mortality at ETR=1, which is typical for pesticides in many species (Sanchez-Bayo and Goka, 2014) and this seems to fit well imidacloprid toxicity for honeybees (Cresswell, 2011). Thereby, we relied largely on the same lethal doses that the 309 310 European Food Safety Authority (EFSA) uses for their risk assessments (EFSA, 2018). However, there is some variability in dose responses and median lethal doses (Cresswell, 2011; Decourtye and Devillers, 2010). In addition, toxicity can vary considerably 311 between taxa and even between different subspecies of Apis mellifera (Suchail et al., 2001). This may further increase the mortality 312 313 risk.

Our risk assessment suggests an even higher risk for individual wild bees than for honeybees. However, we used a contact rather than an oral acute lethal dose for solitary bees and chronic lethal dietary doses for bumblebees and solitary bees that were extrapolated from values derived from honeybees using a safety factor of 10 by EFSA (EFSA, 2018). This likely caused an over-estimation of mortality rates, although neonicotinoids tend to be more toxic to bees via oral than contact exposure (EFSA, 2018; Sanchez-Bayo and

Goka, 2014). Therefore, the mortality risk associated with nectar consumption is not necessarily higher for individual wild bees, but ground-nesting wild bees may also be exposed to neonicotinoid residues in agricultural soils (Chan et al., 2019). In addition, implications of losses of individual bees may be more severe for wild bee populations than for honeybees, as elevated forager losses translate for solitary bees directly into population declines. In contrast, social bees can compensate for the loss of individual foragers. This is particularly true for honeybees due to their large colony size as field studies showing more severe effects of neonicotinoids on bumblebee than on honeybee colonies suggest (Henry et al., 2015; Osterman et al., 2019; Rundlöf et al., 2015; Wintermantel et al., 2018; Woodcock et al., 2017).

325 Nonetheless, our risk estimates suggest an impact on honeybee colony functioning. Typically, honeybees forage 6.5-10 days in their lives (Khoury et al., 2011; Rortais et al., 2005; Visscher and Dukas, 1997) and a reduction of the average foraging lifespan by 2.8 days 326 has been estimated to precipitate colony failure (Khoury et al., 2011). We estimated that in eight fields (four fields each in 2014 and 327 328 2016) 50% mortality of nectar foragers due to acute toxicity was reached after 4 days. Even in fields with more moderate loss rates, long-term effects may occur. Elevated forager mortality may trigger for instance bees to forage at a younger age with negative 329 330 consequences for brood care, foraging efficiency and worker longevity (Khoury et al., 2011; Perry et al., 2015), causing colonies to enter a positive feedback loop accelerating colony weakening (Perry et al., 2015). For instance, reduced foraging efficiency can result 331 332 in increased forager recruitment and consequently reduced brood care (Gill et al., 2012). All these aspects can also be affected by 333 exposure to sublethal levels of neonicotinoids (Godfray et al., 2015; Pisa et al., 2017). In fact, a 6-year survey of Greek apiaries indicated that sublethal neonicotinoid concentrations in nearby nectar sources led to a substantial decline in colony size and honey 334 335 production (Chambers et al., 2019) and field experiments showed that neonicotinoids in nearby oilseed rape plants can lead to substantial reproductive impairments for bumblebees and solitary bees (Rundlöf et al., 2015; Wintermantel et al., 2018; Woodcock et 336 al., 2017). 337

We found that both imidacloprid prevalence and concentration in positive samples increased with rainfall in the previous days, likely due to increased uptake from the soil. Bees, in particular honeybees, avoid foraging during rainy days (Javorek et al., 2002). Therefore, they may forage more intensively when rainfall ceases, which implies that intensive foraging and high prevalence and concentration would coincide, which further increases the risk. We regard here only the risk of imidacloprid-induced mortality and

neglect potential interactions with other factors affecting bees such diseases or lack of flowers (Chambers et al., 2019; Goulson et al.,
2015).

344 Despite the limitations of our risk simulation, we conclude that the EU moratorium restricting imidacloprid, thiamethoxam and clothianidin use in bee-attractive crops has not eliminated the risk for bees foraging on oilseed rape nectar. This questions the concept 345 of banning harmful pesticides only for the use in insect-pollinated crops. In fact, both systemic and non-systemic pesticides, including 346 neonicotinoids, have been found in bee-collected pollen from untreated plants and were shown to pose a risk to bees (McArt et al., 347 2017; Nicholls et al., 2018; Tosi et al., 2018). For bumblebees, neonicotinoid exposure through pollen and nectar declined post-ban 348 349 (2015) in rural areas, but not in peri-urban areas (Nicholls et al., 2018). These and our results, provide support to the total ban of all neonicotinoids in France and the EU-wide ban of the outdoor use of imidacloprid, thiamethoxam and clothianidin in all crops that 350 351 have come into effect in September and December 2018, respectively. Neonicotinoids are, however, still used extensively outside the 352 European Union as well as in permanent greenhouses within the European Union (except in France), from which they may leach or be 353 discharged into nearby water systems. Therefore, there remains a need to examine the movement of neonicotinoids in the 354 environment. The extent by which neonicotinoids spread through leaching, runoff and dust drift as well as the factors governing neonicotinoid uptake by plants should be studied in more detail. The large variability in neonicotinoid prevalence and concentration 355 that we determined suggests that pesticides, especially systemic pesticides, which are water-soluble and mobile, should be extensively 356 357 assessed under differing field conditions particularly in respect to their transportation pathways and fate before conclusions on their safety can be drawn. 358

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- 577 **7.** Tables
- 578 **Table 1.** Acute median lethal doses (LD<sub>50</sub>) and chronic dietary median lethal doses (LDD<sub>50</sub>) used in the risk assessment for nectar-
- 579 foraging bees foraging on imidacloprid residues in oilseed rape nectar.

Endpoint	Honeybee	Bumblebee	Solitary b580	
			581	
Acute LD <sub>50</sub> (in µg bee <sup>-1</sup> )			582	
	0.0037 (48 h) <sup>a,b</sup>	0.04 (24 h) <sup>b</sup>	0.03 (48 h) <sup>5,83</sup>	
			584	
Chronic 10 d-LDD <sub>50</sub> (in $\mu$ g bee <sup>-1</sup> d <sup>-1</sup> )	0.00282 <sup>a</sup>	0.000282 <sup>e</sup>	$0.000282 \frac{e}{585}$	<sup>a</sup> (EFSA, 2018)

<sup>b</sup> (Marletto et al., 2003)

<sup>c</sup> (Uhl et al., 2018)

588 <sup>d</sup> Contact rather than oral exposure

<sup>e</sup> The endpoint has been extrapolated from the endpoint for honeybees using a safety factor of ten.

590

- 592 **Table 2.** Repeatability in imidacloprid prevalence and log<sub>10</sub> concentrations in positive samples. Repeatability refers to the closeness of
- 593 the agreement among imidacloprid residues from samples of the same field and was measured as intraclass coefficient (ICC). Only
- 594 fields with at least two (positive) measurements were considered.

Year <sup>a</sup>	Prevalence		Log <sub>10</sub> concentration		
	<i>ICC</i> <sup>b</sup> (95% CI <sup>c</sup> )	<b>P</b> <sup>b,d</sup>	<i>ICC</i> (95% CI °)	<b>P</b> <sup>d</sup>	
2016	0.13 (0-0.26)	0.014	0 (0-0.19)	1	
2017	0.73 (0.34-0.98)	<0.001	0 (0-0.60)	0.44	
2018	0.09 (0-0.44)	0.24	0.24 (0-0.88)	0.31	
all	0.32 (0.13-0.41)	<0.001	0 (0-17)	1	

<sup>a</sup> In 2014 and 2015, fields were (typically) sampled only once and could therefore not be separately assessed.

<sup>b</sup> Estimation was done on the logit scale.

<sup>c</sup> 95% CI were obtained by bootstrapping with 1500 simulations.

<sup>d</sup> *P*-values were computed by permutation tests with 1500 simulations.

599





Figure 1. Study site. Location and main soil types of the Long-Term Socio-Ecological Research (LTSER) site '*Plaine & Val de Sèvre*' and the oilseed rape fields (black) that nectar was collected from within the study period (2014-2018).



607

**Figure 2.** Prevalences and concentrations of clothianidin, imidacloprid and thiamethoxam in oilseed rape nectar by year. (a) Prevalences are shown per sample (grey) and per field (white). Error bars denote 95% confidence intervals computed by binomial tests. (b) Tukey boxplots show neonicotinoid concentrations in positive samples on a  $log_{10}$ -scale with horizontal lines denoting median values and triangles mean values. The bottom and the top of the boxes show the first and the third quartiles, respectively. Whiskers illustrate minimum and maximum values within 1.5 interquartile ranges. The dots denote outliers. The number of positive samples is shown above the boxplots. Samples < LOQ but > LOD were set to 0.2 ng mL<sup>-1</sup> as determined by the equation (LOD+LOQ)/2.



616 Figure 3. Imidacloprid prevalence and concentration in positive samples in oilseed rape nectar in relation to selected predictors. Solid lines and bars denote estimates and dotted lines and error bars 95% intervals derived from (a) a generalized linear mixed-effects model 617 (GLMM) with a binomial error distribution and a log-link for prevalence or (b) a linear model (LM) with a Gaussian error distribution 618 for concentration in samples in which imidacloprid was detected. The GLMM for prevalence contained field identity as random factor 619 and sampling Julian day number, precipitation in a 10-day period ending 4 days before sampling, mean air temperature in a 4-day 620 621 period ending 3 days prior to nectar sampling and soil type as fixed factors. The LM for concentration contained sampling Julian day 622 number both as linear and quadratic terms, and an interaction (including main effects) between precipitation in a 1-day period ending 3 days before sampling and soil type as predictors. 623



Figure 4. Mortality risk for nectar foragers from imidacloprid exposure through oilseed rape. Percentage of fields at which over 50% of honeybees, bumblebees and solitary bees are estimated to die due to acute (red circle) and chronic (blue square) imidacloprid toxicity over 10 days. Combined mortality (due to acute or chronic toxicity) is indicated by black crosses and black lines (chronic mortality is only estimated after 10 days).

