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Responsiveness to Sugar Solutions in the Moth *Agrotis ipsilon*: Parameters Affecting Proboscis Extension

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Adult moths need energy and nutrients for reproducing and obtain them mainly by consuming flower nectar (a solution of sugars and other compounds). Gustatory perception gives them information on the plants they feed on. Feeding and food perception are integrated in the proboscis extension response, which occurs when their antennae touch a sugar solution. We took advantage of this reflex to explore moth sugar responsiveness depending on different parameters (i.e., sex, age, satiety, site of presentation, and composition of the solution). We observed that starvation but not age induced higher response rates to sucrose. Presentation of sucrose solutions in a randomized order confirmed that repeated sugar stimulations did not affect the response rate; however, animals were sometimes sensitized to water, indicating sucrose presentation might induce non-associative plasticity. Leg stimulation was much less efficient than antennal stimulation to elicit a response. Quinine prevented and terminated sucrose-elicited proboscis extension. Males but not females responded slightly more to sucrose than to fructose. Animals of either sex rarely reacted to glucose, but curiously, mixtures in which half sucrose or fructose were replaced by glucose elicited the same response rate than sucrose or fructose alone. Fructose synergized the response when mixed with sucrose in male but not female moths. This is consistent with the fact that nectars consumed by moths in nature are mixtures of these three sugars, which suggests an adaptation to nectar perception.

Keywords: moth, sugar responsiveness, dose-response curves, nectar, sugar, quinine, gustatory perception, proboscis extension response

INTRODUCTION

Moth reproduction and its regulation by sex pheromone have been largely studied to find ways to control their populations, as their larvae are important crop pests (Cook et al., 2007; Naranjo et al., 2015; Evenden, 2016). While less exploited for managing these pests, adult moths feeding behaviors and gustatory perception are also important for their reproduction; they need energy and nutrients to produce an abundant and healthy offspring (Hill, 1989; Hill and Pierce, 1989; Boggs and Ross, 1993; Wheeler, 1996; Boggs, 1997a,b;

O'Brien et al., 2000, 2004; Fischer et al., 2004; Jervis et al., 2005; Geister et al., 2008; Marchioro and Foerster, 2013; Levin et al., 2017a,b). Their food consists largely in sugars from flower nectars, which contains mainly fructose, glucose, and sucrose as well as a few amino acids and, sometimes, repellent substances (Baker and Baker, 1973; Baker, 1977; Lüttge, 1977; Gottsberger et al., 1984; Adler, 2000; O'Brien et al., 2000, 2004; Pacini et al., 2003; De La Barrera and Nobel, 2004; Adler et al., 2006; Heil, 2011; Nepi et al., 2012; Atwater, 2013; Roy et al., 2017). Thus, gustatory perception is important for moths to assess food quality and discriminate what is edible from what is toxic (Glendinning, 2002; Adler and Irwin, 2005; Tiedeken et al., 2014).

When their legs, antennae, or proboscis contact a sugar solution of sufficient concentration, moths extend their proboscis and use it to imbibe the solution. This proboscis extension response (PER) first reflects the integration of gustatory perception and motivation for sugar and then allows feeding. Sucrose-elicited PER has been described and involved in associative learning in restrained insects including moths (Hartlieb, 1996; Fan et al., 1997; Hartlieb et al., 1999a,b; Fan and Hansson, 2001; Skiri et al., 2005; Jorgensen et al., 2007), butterflies (Kroutov et al., 1999), bees (Menzel, 1999, 2012; Page and Erber, 2002; Sandoz, 2011; Giurfa and Sandoz, 2012; Giurfa, 2015) and flies (Fresquet, 1999; Chabaud et al., 2006); similar feeding-related responses exist in ants (Guerrieri and d'Ettoire, 2010; Perez et al., 2013), crickets (Matsumoto et al., 2015), and bugs (Vinauger et al., 2013; Labrousse et al., 2017). PER has been used by Scheiner and her colleagues to assess responsiveness to sucrose in bees and flies (Scheiner et al., 2004a,b, 2013; Mujagic et al., 2010). These authors demonstrated that sucrose responsiveness is correlated to important parameters of honey bee biology such as division of labor, and foraging decisions are modulated by sugar gustatory perception (see for instance Nachev et al., 2013). Sugar-induced PER has been used in various moth species (Hill and Pierce, 1989; Winkler et al., 2005; Jørgensen et al., 2007; Zhang et al., 2010; Minoli et al., 2012) using different sucrose concentrations and animals in different physiological conditions; however, the impact of the physiological state on sugar responsiveness is hardly known in these insects.

Thus, we aimed for the first time at specifically looking at sugar responsiveness in relation to different physiological parameters (i.e., sex, age, and level of satiety) in the moth species *Agrotis ipsilon*. Testing systematically different concentrations and kinds of sugars composing nectar (i.e., sucrose, fructose, and glucose) further allowed us to find the optimal parameters for releasing the PER. Our work was inspired by extensive work on sucrose responsiveness in bees and flies (Scheiner et al., 2004a, 2013). We tested if sucrose responsiveness can also be induced using other sensory pathways beyond perception by the antennae (i.e., legs) and modulated by other gustatory stimuli (quinine). We also uncovered a non-associative plasticity, sensitization for water.

Abbreviations: PER, Proboscis extension response.

MATERIALS AND METHODS

Animals

This study was performed on adult *A. ipsilon* (Lepidoptera, Noctuidae), which is a native species of France. Animals were reared in our laboratory in INRA, Versailles, France. Males and females were separated at the pupal stage and kept in different climatic chambers (22°C, 60–70% relative humidity) under an inverted photoperiod (16 h of light, starting at 18 h). Newly emerged adults (i.e., animals having just completed the imaginal molt) were collected every day (so that their post-emergence age is known) and grouped by 10 in breeding boxes (20 × 11.5 × 5 cm) with *ad libitum* access to either sucrose solution (fed animals) or tap water (starved animals). The sucrose solution was 12% weight/weight (13.6% weight/volume). We did not use more than 5-day starvation delays as the moths start to be weak or even die after starving for 6 days. Moths provided with sucrose gain weight, whereas moths provided only with water lose weights (unpublished observation), and we often observed that they readily drink the 12% solution in their breeding boxes but hardly react to water. Thus, it is reasonable to assume the feeding statuses of water- and sucrose-fed moths are not the same.

Experiments were conducted under dim red light and started at the middle of the scotophase, when moths are the most active (highest responsiveness of males to the sex pheromone, sex pheromone release by females), under a temperature of 22–24°C and 65% relative humidity. Unless otherwise mentioned, experiments were performed on day 5 post-emergence. Five-day-old *A. ipsilon* are standardly studied because males of this age are sexually mature and respond to the female sex pheromone (Gemeno and Haynes, 2000). In the morning of the experiments, animals were fixed in plastic tubes made from 1 ml pipette cones cut to fit a moth, so that only the head (with antennae and proboscis) protruded. Their position was further assured by inserting a small ball of absorbing paper or cotton behind them in the tube and fixing the top of their body to the tube using adhesive tape. Legs were blocked by the tape or inside the tube, unless otherwise mentioned (section “Proboscis Extension Response Induced by Leg Stimulation” reports the effect of leg stimulation). Once restrained, animals were left to acclimatize in the experimental room until the beginning of the experiment (in the afternoon).

Sugar Responsiveness Assay and Solutions Used

We adapted Scheiner's protocol for honey bees and flies (Scheiner et al., 2004a,b, 2013) to moths. Assessing sugar sensitivity consisted in presenting solutions of increasing sugar concentration; a 10-min interval between each presentation was used, except in section “Sensitization and Presentation Timing,” where a 1-min interval was also used. A solution presentation consisted in touching both antennae of the animal with a wooden toothpick imbued with the sugar solution during 1–4 s; a response was recorded if the animal released a PER, i.e., extended its proboscis beyond its position at the

onset of the stimulation (the initial position varies from one animal to the other, but the occurrence of the proboscis extension can be observed unambiguously). Animals were not fed, and great care was taken to avoid touching the proboscis with the toothpick by coming from behind the head, while the proboscis extends in front of it. We defined PER rate as the proportion of animals releasing a PER upon presentation of a sugar solution on the antennae.

The solution concentrations were 0 (i.e., deionized water), 0.1, 0.3, 1, 3, 10, and 30% (weight/weight in deionized water, e.g., 30% is 3 g of sucrose in 7 ml of deionized water). These sugar concentrations correspond to 0.1, 0.4, 1.1, 4.3, 11.1, and 42.9% weight/volume, respectively. A second presentation of the 0% solution was made afterwise to monitor the occurrence of sensitization, i.e., the fact animals might start to respond to weak stimuli such as 0% solution just because they are excited by previous sugar presentations. Sensitization is discussed in section “Sensitization and Presentation Timing.” This set of concentrations is inspired of previous studies in bees and flies (Scheiner et al., 2004a, 2013) and corresponds to a logarithmic increase in the stimulus intensity (i.e., around -1 , -0.5 , 0 , 0.5 , 1 , 1.5), consistently with Weber’s law in sensory physiology (Waddington and Gottlieb, 1990; Akre and Johnsen, 2014; Shafir and Yehonatan, 2014). In our experiments, the sugar used was usually sucrose, but fructose, glucose, and various mixtures of these sugars were also tested (see section “Response to Various Sugars and Sugar Mixtures”). We focused on these three sugars (obtained from Sigma) as they are the main constituents of flower nectar, the main food of adult moths: typically 10–15% of the fresh mass for each sugar, possibly up to 30% for sucrose (Lüttge, 1977; Gottsberger et al., 1984; Pacini et al., 2003; De La Barrera and Nobel, 2004; Roy et al., 2017). In some experiment (section “The Effect of Quinine on Sucrose Responsiveness”), animals were also presented with 100 mM quinine (Sigma).

To rule out the possibility of having animals responding to water rather than sugar, animals responding to the initial water presentation were not kept in the analysis. However, removing these animals hardly modified the results. Supplementary data report figures including all the animals and the corresponding analysis (**Supplementary Figures S1–S6, Supplementary Tables S13–S24**).

Experiments

Age, Sex, and Feeding Status

First, we investigated the effect of age and feeding status. Sucrose responsiveness was measured for various sucrose concentrations in five groups of males: 3-day-old moths either (1) with *ad libitum* access to sucrose solution (fed) or (2) starved (*ad libitum* access to water only), and 5-day-old moths either (3) fed, (4) starved for 3 days, or (5) starved for 5 days. Five-day-old moths starved for 3 days received sucrose from 0 to 2 days post-emergence and then only water for the following 3 days. Using this group was an attempt to discriminate between age of the animals and starvation duration, as when comparing 3-day-old and 5-day-old unfed moths,

we simultaneously compare age (3-day-old or 5-day-old) and starvation duration (3 days or 5 days), so that the two factors are confused.

To assess the effect of sex, this experiment was replicated with females using the same age/starvation treatments, plus a group of 5-day-old male moths starved for 5 days for comparison purposes. Experiments reported elsewhere (Hostachy et al., submitted) indicated that sucrose responsiveness of males is not affected by the presence of sex pheromone, which allows to test them in parallel with females.

Sensitization and Inter-trial Interval

In a sucrose responsiveness assay, it is important to know whether responses (or lack thereof) to repeated presentations of sucrose of varying concentrations are independent from each other: having already been stimulated by a sucrose solution could affect responsiveness to the next simulations. This could be explained by sensitization (or habituation if we observed a decrease) and/or a time-dependent modification of motivation. To quantify this phenomenon, we performed the sucrose responsiveness assay in two groups of 5-day-old female moths: one group was presented with the sucrose solutions in the same order as previously (ascending concentrations), whereas for the other group, the six sucrose solutions were presented in a randomly determined order for each animal (keeping water as first and last stimulation). If sensitization or motivation variations alter sugar responsiveness when the solutions are presented in ascending order, then sugar responsiveness will be different in the random order group. Within each group, a sub-group was unfed for 5 days, whereas the other sub-group was fed; this was done to evaluate whether the occurrence of sensitization or motivation variations differed between fed and unfed animals.

To further explore the occurrence of sensitization and determine the importance of the inter-presentation interval, we used two inter-trial intervals: either 10 min (as previously) or 1 min. Indeed, a potential sensitization should occur more easily with a shorter inter-trial interval, as this is a form of short-term plasticity. In some animals, we also interleaved water presentations between sucrose presentations, as this was initially done in some experiments with bees to try and prevent sensitization (see for instance, Scheiner et al., 2003). For these groups, the interval between two sucrose presentations was either 2 or 20 min, with a water presentation interleaved at 1 or 10 min, respectively.

Effect of the Stimulation Site

While proboscis extension is classically elicited by antennal stimulation in bees and moths, it can also be triggered by leg stimulation (which is standard for fruit flies). Thus, we performed again the sucrose sensitivity protocol using two groups of animals, stimulating either the antennae (as previously, reference group) or the tibia and tarsi of the front legs. Leg-stimulated animals were restrained slightly differently: one of their front leg was fixed, so that it stayed outside of the tube. During leg stimulations, great care was taken to avoid touching the

antennae. After testing all sucrose concentrations using the legs, the leg-stimulated animals were then assessed a second time using the antennae (i.e., the whole protocol was completed using the legs, then performed again using the antennae).

The proboscis also bears gustatory sensilla, so that stimulating it should elicit a PER. However, in moths, it is quite inconvenient to reach when it is coiled in its rest position; furthermore, it is impossible to prevent the animals to imbibe some of the solution in this case. Thus, evaluating sucrose responsiveness by direct proboscis stimulation is not feasible with this protocol.

Effects of a Bitter Compound: Quinine

As in nature moths consume sugar solutions found in flowers, they can be exposed to alkaloids that might have a deterrent effect. To assess whether alkaloids such as quinine affect PER and sucrose responsiveness, we performed again the sucrose responsiveness protocol on 5-day-old males. Animals releasing a PER had their proboscis immediately touched either with water (control group) or with 100 mM quinine, and we recorded whether the PER would continue after this proboscis stimulation. To complement these observations, we also measured responsiveness to sucrose and sucrose plus 100 mM quinine in 5-day-old males.

Effects of Other Sugars: Fructose, Glucose, and Sugar Mixtures

Sucrose is the reference sugar used in PER experiments in various insects. However, it is well known that nectar can also include fructose and glucose in various proportions. Thus, we measured responsiveness for fructose and glucose besides sucrose, as well as mixtures between these three molecules. For mixtures, the same concentrations were used with sugars in equal amounts (i.e., a 3% sucrose/fructose solution contains 1.5% sucrose, 1.5% fructose, weight/weight; or 2.15% of each sugar, weight/volume). Experiments were done with 5-day-old males unfed for 5 days.

In another experiment, the sugar responsiveness assay was performed in females with glucose, fructose, and sucrose, as well as sucrose/fructose and sucrose/fructose/glucose mixtures. All animals were 5-day-old females unfed for 5 days, and a group of 5-day-old males unfed for 5 days and tested with sucrose was also used for comparison purpose.

Data Analysis

Data were analyzed using R 3.4.0 (R Core Team, 2017) and RStudio 1.1.423, taking an α risk of 0.050. PER rates were computed for each experimental group and are reported in the figures, with the sample sizes reported in parenthesis in the legends. Thus, a dose-response curve was obtained for each experimental group.

For each sugar concentration in a dose-response curve, PER rates were compared between experimental groups using χ^2 test or bilateral Fisher's exact test when χ^2 's assumption (Cochran's rule) was not met. When more than two groups were compared and a significant p was obtained, we performed pairwise

comparisons using the same tests and adjusted p for repeated testing using Holm's method (Holm, 1979); however, it sometimes happened adjusted p could not determine the source of the difference detected in the global test.

In a few cases, response variations within a group were compared using McNemar's test, using Holm's method to adjust p when needed. Logistic regression (binomial generalized linear model) was also used in section "Sensitization and Presentation Timing" to assess whether PER rates to the final water presentation were affected by the sugar concentration presented 10 min before or by the number of PER previously released (preliminary stepwise analysis indicating the interaction term was not significant, so that it was dropped). To avoid tedious lists of p in the results, most of them are only presented in tables in supplementary data (**Supplementary Tables S1–S12**, one table for each figure or figure panel).

RESULTS

The Effect of Age, Starvation Duration, and Sex on Sucrose Responsiveness

We first explored the experimental parameters required for optimally assessing sucrose responsiveness by comparing age and starvation duration. For male moths (**Figure 1A** and **Supplementary Figure S1A**), significant differences were observed for all sucrose solutions of 1% and higher (χ^2 or Fisher's exact test, $p \leq 0.004$): overall, fed animals were less responsive than unfed ones, irrespective of their age, while 5-day-old moths unfed for 5 days were consistently among the highest PER rates (details of the pairwise comparisons are in **Supplementary Table S1**).

For females (**Figure 1B** and **Supplementary Figure S1B**), results were analyzed in two steps: first, 5-day-old males and females unfed for 5 days were compared, and second, all female groups were compared. For the lowest concentrations (0.1 and 0.3%), the PER rates of females were slightly lower than males' (χ^2 or Fisher's exact test, $p \leq 0.022$), suggesting females are slightly less responsive than males for the lowest concentrations. However, the same pattern as for males was observed for the PER rates across treatments, i.e., 5-day-old females unfed for 5 days were consistently among the highest PER rates for 0.3, 1, 3, and 30% concentrations (details of the pairwise comparisons are in **Supplementary Table S2**).

Sensitization and Presentation Timing

When the independence of the stimulations and the occurrence of sensitization was assessed (**Figure 2A** and **Supplementary Figure S2A**), there was a global significant difference between the four groups for 1, 3, and 10% sucrose concentrations (χ^2 or Fisher's exact test, $p \leq 0.001$), replicating the previous finding that fed animals have a lower sucrose responsiveness; by contrast, presenting the solutions in random order or in ascending order did not have any effect (**Supplementary Table S3** reports detailed analysis). As a result, sensitization (or even habituation) or variations of motivations during the assay are unlikely to

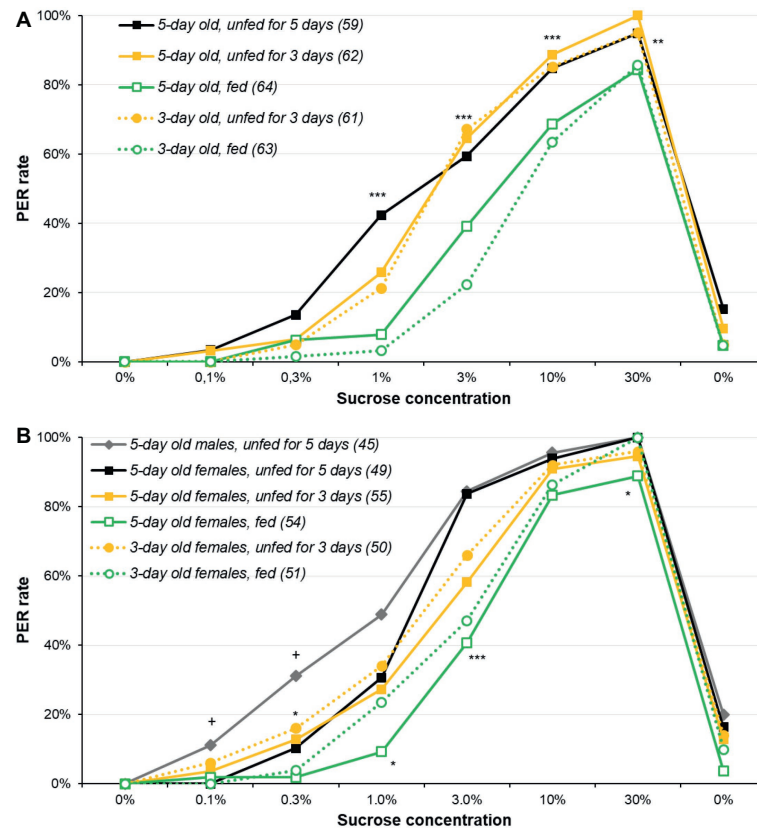


FIGURE 1 | Effect of age and starvation duration on sucrose responsiveness in *Agrotis ipsilon*. The plots present dose-response curves for various sucrose concentrations (x-axis); the y-axis reports the PER rate, i.e., the proportion of insects extending their proboscis when their antennae were briefly touched with a drop of sucrose solution. Each sucrose solution presentation was separated by 10 min. In the legend, numbers in parenthesis correspond to the sample sizes. In part (A), experiments were done with males; in part (B), experiments were done with females and a group of males for comparison purpose. Stars denote significant differences between male or female groups in χ^2 or Fisher's exact tests ($*p < 0.050$; $**p < 0.010$; $***p < 0.001$); in part (B), crosses denote significant differences between 5-day old males and females unfed for 5 days in χ^2 of Fisher's exact test ($*p \leq 0.050$). Details of the analyses are reported in **Supplementary Table S1** for part (A) and **Supplementary Table S2** for part (B).

be an important determinant of the PER rate in the sugar responsiveness assay.

However, even if sensitization does not alter sucrose responsiveness, it could explain the PER rate observed during the final water presentation. To test this, we used the data from the unfed random group and computed a logistic regression to test the effect of two factors on water responsiveness: the total number of responses before the final water test (ranging from 0 to 6, i.e., the number of responses to the 6 sugar concentrations) and the concentration of the sugar solutions used before water (which can be any of the 6 concentrations in this random order group). Both factors were significant (logistic regression $p \leq 0.046$). This indicates that animals responding to most sucrose concentrations tend to respond to the final water, even though they did not respond to the initial water presentation. Moreover, a higher concentration just before the final water presentation (typically, the 3, 10, or 30% solutions) also contributes to this sensitization to water.

Figure 2B and **Supplementary Figure S2B** report sucrose responsiveness with various inter-trial intervals and with or

without interleaved water. Our hypothesis was that sensitization would occur in the 1-min group without interleaved water and be lower in the 10-min group with interleaved water, but this was not the case: the four groups were similarly responsive (χ^2 or Fisher's exact test, $p \geq 0.064$; details of the analysis are in **Supplementary Table S4**). Moreover, responses to the interleaved water presentation were low and did not differ significantly across groups (data not shown; Fisher's exact test, $p \geq 0.386$).

Proboscis Extension Response Induced by Leg Stimulation

Results on the effect of leg stimulation are presented in **Figure 3** and **Supplementary Figure S3** and details of the statistics in **Supplementary Table S5**. Leg stimulation was much less efficient to elicit PER, as it only did so to a meaningful level for the highest concentrations; in all cases, leg-elicited-PER rate was significantly smaller from antennal-elicited-PER rate (χ^2 or Fisher's exact test, $p \leq 0.005$). A second series of tests was made to compare the leg group to the reference group when

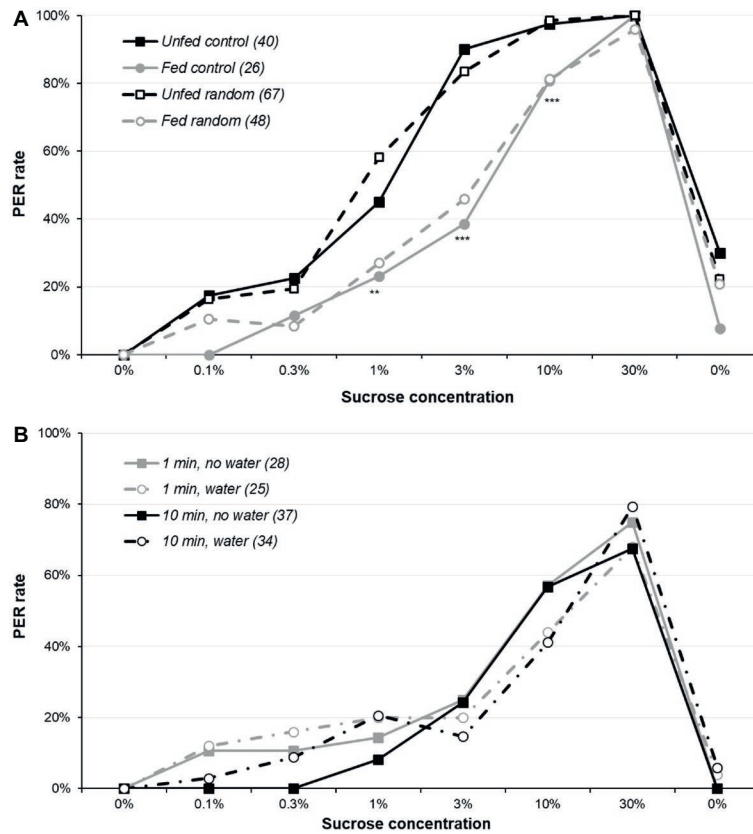


FIGURE 2 | Sensitization and sucrose responsiveness. Part (A) reports sucrose responsiveness in four groups of 5-day-old females: the first two were presented sucrose concentrations in ascending order (as previously), whereas for the other two, the six sucrose solutions were presented in an order randomly determined for each animal. Within each treatment, one of the two groups had been unfed for 5 days, whereas the other was fed. Part (B) plots the sucrose responsiveness in four groups of unfed 5-day-old males: two groups with an inter-trial interval of 10 min (as previously) and two groups with an inter-trial interval of 1 min. For each interval, one of the two groups had a water presentation every other trials, so that for these groups, the interval between two sucrose presentations was either 2 or 20 min, with a water presentation interleaved at 1 or 10 min. Some animals in the two no-water groups were not presented the final water solution. Other details are as in Figure 1. Details of the analyses are reported in Supplementary Table S3 for part (A) and Supplementary Table S4 for part (B).

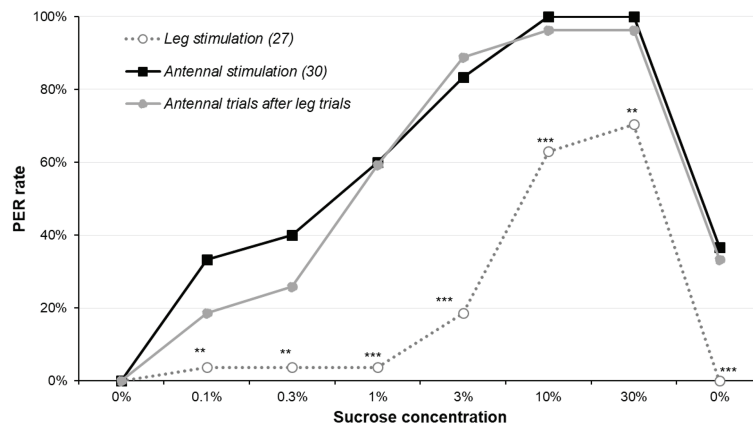


FIGURE 3 | Comparison of antennal stimulation and leg stimulation. The sucrose sensitivity experiment was conducted by stimulating either the antennae (as previously) or the legs of 5-day-old males starved for 5 days. Animal stimulated on the legs for the complete set of sucrose solutions underwent the protocol a second time with the antennae (i.e., the whole set of sucrose solutions was presented twice, one on the legs and the other on the antennae). Stars denote significant difference between the leg stimulated group and the antennal stimulated group run in parallel (χ^2 or Fisher's exact test; ** $p < 0.010$; *** $p < 0.001$). Details of the analyses are reported in Supplementary Table S5. Other details are as in Figure 1.

the leg group was retested on the antennae: this time, there was no difference between the two groups (χ^2 or Fisher's exact test, $p \geq 0.205$). This means that the leg-stimulated group had the same sucrose responsiveness as the control when tested on the antennae. This confirms that the lower PER rate is only due to the different types of stimulation.

The Effect of Quinine on Sucrose Responsiveness

The effects of quinine stimulation are reported in **Figure 4A** and **Supplementary Figure S4A** and the detailed analysis in **Supplementary Table S6**. Before contacting the proboscis with water or quinine, the sucrose responsiveness did not significantly differ between the water- and the quinine-stimulated group (χ^2 or Fisher's exact test, $p \geq 0.123$), confirming the two groups were identical at this step. Touching the proboscis with water did not affect the PER, as no animal retracted its proboscis; by contrast, most animals which proboscis touched quinine interrupted the PER and retracted the proboscis, almost

significantly for sucrose concentrations 1 and 3% (McNemar's test, $p \leq 0.071$), and significantly for concentrations 10 and 30% (McNemar's test, $p \leq 0.005$); no difference was observed for other concentrations, but the PER rates were low for them anyway. We also compared the proportions of animals retracting the proboscis among those initially responding, and the results were the same (data not shown). Interestingly, the fact that the PER rates remain the same as in the control group in ulterior sucrose solution presentations means that even though quinine elicited proboscis retraction, it does not modulate sucrose responsiveness (at least after 10 min). Moreover, when quinine was added to sucrose (**Figure 4B**, **Supplementary Figure S4B**, **Supplementary Table S7**), it strongly inhibited PER (χ^2 or Fisher's exact test; $p \leq 0.024$).

Response to Various Sugars and Sugar Mixtures

In a first experiment, animals were presented with either sucrose, fructose, or a mixture of these sugars (**Figure 5A**

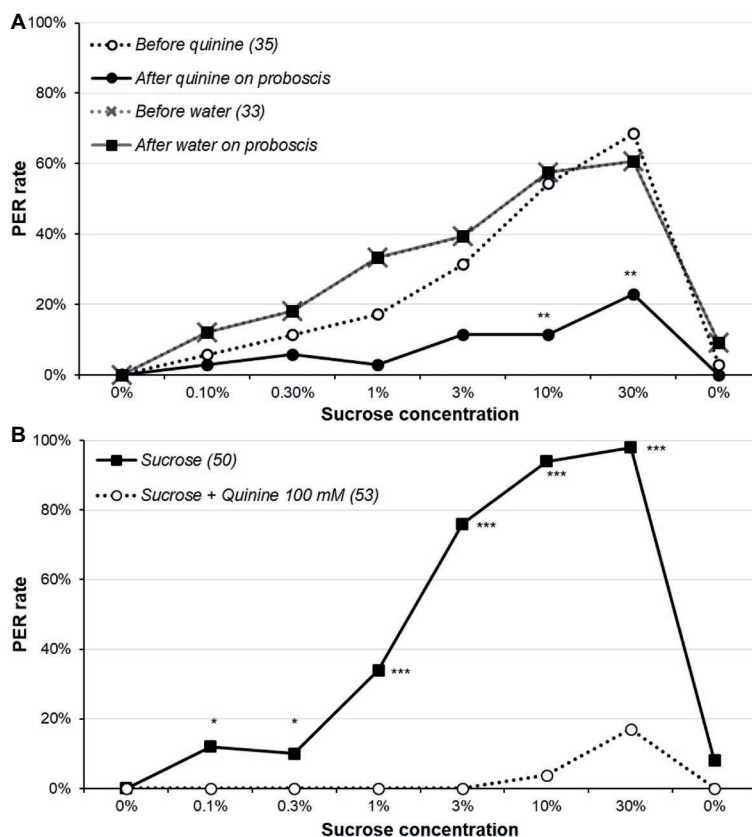
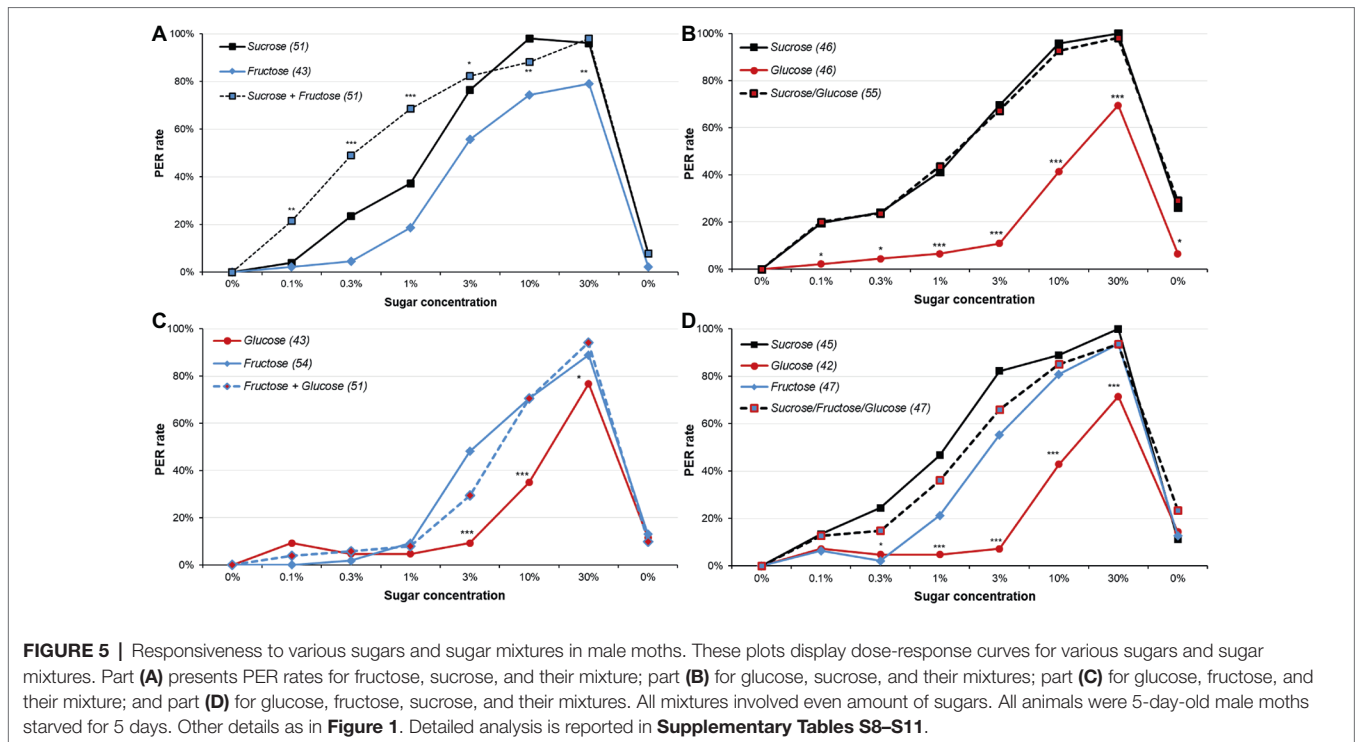


FIGURE 4 | Effect of quinine on sucrose responsiveness. In part (A), animals were presented the series of sucrose solutions (dotted lines), and if they released a PER, they immediately received either 100 mM quinine or water on the extended proboscis; the continuous lines indicate the proportion of animals still releasing a PER after receiving one of these solution. Notice that as touching the proboscis with water had no effect, the two lines (before and after water) are superimposed. Stars denote a significant difference between the PER rate before and after touching the proboscis with quinine, i.e., a significant rate of proboscis retraction (McNemar's test $**p < 0.010$); detailed analysis is in **Supplementary Table S6**. In part (B), animals were presented the series of sucrose solutions either alone as previously or combined with 100 mM quinine. Stars denote a significant difference between the PER rate of the two groups (χ^2 ; $*p < 0.050$, $***p < 0.001$); detailed analysis is in **Supplementary Table S7**. All animals were 5-day-old males starved for 5 days; other details are as in **Figure 1**.



and Supplementary Figure S5A, Supplementary Table S8). Although fructose elicited high PER rates, overall they were significantly lower than for sucrose (χ^2 or Fisher's exact test; adjusted $p = 0.068$ for 3%, $p \leq 0.046$ for 0.3, 1, 10, and 30%). Interestingly, mixtures of fructose and sucrose elicited the same level of PER rate as sucrose alone (χ^2 or Fisher's exact test; adjusted $p \geq 0.217$ for 3, 10, or 30%), or even higher responses for lower concentrations (χ^2 or Fisher's exact test; adjusted $p \leq 0.030$ for 0.1, 0.3, or 1%). Thus, while fructose tended to elicit less responses than sucrose, replacing half the sucrose by fructose in a mixture actually improved the PER rate, indicating a synergistic effect.

In a second experiment, we compared animals' responses to sucrose, glucose, and their mixture (Figure 5B and Supplementary Figure S5B, Supplementary Table S9). Glucose elicited PER much less often than the two other solutions (χ^2 or Fisher's exact test, adjusted $p \leq 0.020$) and induced less sensitization for the final water presentation (χ^2 , adjusted $p \leq 0.022$). Yet, replacing half the sucrose by glucose did not modify the PER rate relatively to the sucrose group (χ^2 or Fisher's exact test, adjusted $p \geq 0.686$); thus, in spite the mixture only includes half sucrose found in the sucrose solution (e.g., the 3% mixture only includes 1.5% sucrose), and so should elicit much lower PER rates, the response is unchanged. This leads to the intriguing conclusion that while glucose did not elicit much response, replacing half of the sucrose by glucose in a solution did not decrease the response, although there was not the synergy observed for sucrose and fructose.

When the responses to fructose, glucose, and their mixture were compared (Figure 5C and Supplementary Figure S5C, Supplementary Table S10), results were similar: glucose elicited

less response than fructose or fructose/glucose mixtures for 3 and 10% (χ^2 , adjusted $p \leq 0.031$); for 30% sugar solution, the global test was significant (χ^2 , $p = 0.038$), but pairwise comparisons were not. Fructose and fructose/glucose mixtures elicited similar PER rates, except for the 3% concentrations, where the mixture was slightly inferior (χ^2 , adjusted $p = 0.049$). To sum up, while animals had lower PER rates for glucose than for fructose, replacing half of the fructose by glucose impaired only slight response. This is comparable to what was seen with sucrose/glucose mixtures.

Finally, we compared PER rates to sucrose, glucose, fructose, and a sucrose/glucose/fructose mixture (Figure 5D and Supplementary Figure S5D, Supplementary Table S11). We replicated the observation that fructose was slightly but significantly less responded to than sucrose (χ^2 or Fisher's exact test, adjusted $p \leq 0.040$ for 0.3, 1 and 3%), and that glucose was much less responded to than fructose or sucrose for 1, 3, 10, and 30% (χ^2 or Fisher's exact test: adjusted $p = 0.068$ for 1% for glucose vs. fructose, adjusted $p \leq 0.046$ for other comparisons). Moreover, glucose also elicited less PER than the mixture for 1, 3, 10, and 30% (χ^2 or Fisher's exact test, adjusted $p \leq 0.045$), whereas sucrose or fructose did not differ from it (χ^2 or Fisher's exact test, adjusted $p \geq 0.151$). Therefore, neither synergy nor inhibition was observed in this case.

When females were tested for sugar mixtures (Figure 6 and Supplementary Figure S6, Supplementary Table S12), contrasting for what was seen in Figure 1B, males and females tested for sucrose did not differ for low concentration of sucrose (χ^2 , $p \geq 0.763$) and males were only slightly higher than females for 10 and 30% sucrose, but without reaching significance (Fisher's exact test, $p = 0.060$ in both cases). While there was no difference between the female groups for 0.1 or 0.3% sugar

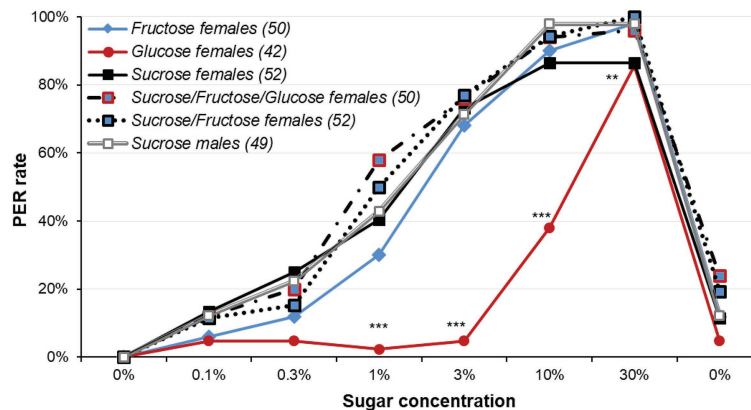


FIGURE 6 | Responsiveness to various sugars and sugar mixtures in female moths. This plot reports dose-response curves for various sugar and sugar mixtures in 5-day old female moths starved for 5 days; a group of 5-day old males starved for 5 days was also used for comparison purpose. Sucrose, fructose, and glucose were used, as well as sucrose/fructose/glucose and sucrose/fructose mixtures. Stars denote significant differences between female groups (χ^2 or Fisher's exact test; ** $p < 0.010$; *** $p < 0.001$). Other details as in **Figure 1**. Detailed analysis is reported in **Supplementary Table S12**.

concentrations (χ^2 or Fisher's exact test, $p \geq 0.080$ in both cases), there were significant differences for 1, 3, and 10% sugar concentrations (χ^2 or Fisher's exact test, $p \leq 0.004$ in all cases). This difference was mainly due to much lower PER rates to glucose solution (χ^2 or Fisher's exact test, adjusted $p \leq 0.042$ in all cases). The other groups did not differ (χ^2 or Fisher's exact test, adjusted $p \geq 0.113$), except Fructose and Sucrose/Fructose/Glucose mixture for 1% (χ^2 , $p = 0.029$).

DISCUSSION

The Sugar Responsiveness Assay: Impact of Various Parameters

This study assessed sucrose responsiveness in relation to different physiological states in the moth *A. ipsilon*. Sucrose responsiveness has been extensively studied in the honey bee (Scheiner et al., 2004a, 2013) and in some moths (Winkler et al., 2005; Zhang et al., 2010; Minoli et al., 2012). We used a broad range of concentrations, consistent with what was used in bees (Scheiner et al., 2004a, 2013) and flies (Scheiner et al., 2004b), and including concentrations found in nectar (10–30% for sucrose: Lüttge, 1977; Gottsberger et al., 1984; Pacini et al., 2003; De La Barrera and Nobel, 2004; Roy et al., 2017).

Our exploration of the parameters associated to sucrose responsiveness has led to various conclusions. Overall, sucrose responsiveness depends upon starvation duration (i.e., motivation for food) and possibly sex but not age; as a result, in most experiments, we used 5-day-old moths starved for 5 days as the standard condition. The inter-trial interval or the interleaving of water are not crucial parameters in the sucrose responsiveness assay, as the response of the animals is not influenced by the previous trials; yet, we could observe some sensitization to water, a new form of behavioral plasticity in *A. ipsilon* (see also Minoli et al., 2012 for a study of sensitization in a related species). Finally, while moths can detect sucrose through their

legs, this pathway is less prone to elicit a PER, either because it does not respond to sucrose very well or because it is less well coupled with the PER motor pathways.

Beyond physiological parameters, the composition of the solution is also essential to observe the PER. In the presence of quinine, moths do not release PER. Thus, quinine is more aversive than sucrose is appetitive, i.e., its ability to elicit a withdrawal of the proboscis is stronger than sucrose's ability to elicit its extension. A simple overshadowing of sucrose is unlikely, as it would not explain PER termination in **Figure 4A**. Concerning the sugars found in nectars, sucrose elicited more responses than fructose, and responses to glucose were much lower than either of them. However, replacing half of the sucrose or half of the fructose by glucose did not lower the PER rate, and mixing sucrose and fructose increased it, suggesting a synergy; interestingly, glucose did not support a synergistic effect in sucrose/fructose/glucose mixture. The results with females are quite different from those observed in males: sucrose and fructose are responded to at the same level, and mixing them (with or without glucose) had no effect. The common point is that glucose alone does not elicit much response while being able to replace the other sugars in mixture.

While simple, the sugar responsiveness assay offers many experimental opportunities. First and foremost, the words "sugar responsiveness" cover various underlying processes, which is why they were preferred to "sugar sensitivity." The dose-response curves used here simultaneously assess three functions: perception (the ability to detect sugar), motivation for sugar (the ability of sugar perception to trigger a PER), and motor response (i.e., if a PER is triggered, the ability to produce it). Detecting sugar and releasing a PER are well-defined phenomena. By contrast, while motivation is associated to them (e.g., **Figures 1** and **2** link hunger to PER rate), its definition is blurrier as it encompasses various situations: not only hunger (i.e., physiological need for food induced by starvation) but also the lack of satiety (i.e., animals' desire to eat). While certainly correlated, these

two situations are not necessarily the same; it would be possible to dissociate them by feeding animals with sweeteners, which induce satiety (i.e., they are palatable) but are physiologically non-nutritive (Schiff et al., 1989; Pszczolkowski and Brown, 2003; Dus et al., 2011; Camacho et al., 2017; Choi et al., 2017; Fisher et al., 2017; Takada et al., 2017; Mustard et al., 2018). The sucrose responsiveness assay would allow to test whether such a dissociation occurs in moth. Similarly, fed animals might have lower response not because they are hungry, but rather because they are habituated to sucrose; to test this hypothesis, animals could be reared (as adult or even as larvae) with fructose rather than sucrose, so that they would not be starved, but still naïve for sucrose. Another possibility offered by the sugar responsiveness assay is to assess lateralization of the response by presenting sugar on a single antennae rather than both; studying lateralization in invertebrate is an emerging field (Jozet-Alves et al., 2012; Baracchi et al., 2018; Niven and Frasnelli, 2018), which complements the classical studies of its role in primate higher cognitive abilities (Prieur et al., 2019). In moths, a study in *Spodoptera littoralis* reported sensitization elicited by pre-exposure to sucrose is a lateralized process (Minoli et al., 2012). All these possible experiments highlight the interest of the sucrose responsiveness assay to investigate precise questions on gustatory responses in moths. Understanding the functioning of sugar-elicited PER could help discriminating between perception, motivation, and PER release mechanisms.

Sensitization is a form of non-associative learning that would make an animal respond to a stimulus (here, the sucrose solution or water) just because it has been excited by a previous presentation of the same or another stimulus (Thompson and Spencer, 1966; Groves and Thompson, 1970; Hammer et al., 1994; Anderson et al., 2007; Anton et al., 2011; Minoli et al., 2012; Blumstein, 2016). Here, we carefully monitored this possibility by using a 10-min interval between each sugar presentation to avoid sensitization, which was hardly considered before in moth sucrose responsiveness. We found sensitization does not affect sucrose responsiveness, although it increases water responsiveness. Interestingly, it was shown in bees and flies that while a forward pairing (presentation of odor, then sugar) promotes associative learning, a backward pairing (sugar than odor) prevents it. This effect seems to be mediated by a desensitization of the PER occurring upon odor presentation at a specific 15-s delay (Dacher and Smith, 2008). Whether this type of desensitization also occurs in moths remains to be determined.

Biological Meaning of Sucrose Responsiveness: Sugar Mixture, Foraging, and Nectar Composition

It is well established in bees that associations between odors and PER/sugar made in restrained conditions can be transferred to free-flying foraging, and vice versa (Gerber et al., 1996; Sandoz et al., 2000; Chaffiol et al., 2005; Gil and De Marco, 2005). Therefore, it is sensible to assume such transfers are possible between the two situations for moths too. Under this hypothesis, sucrose responsiveness is likely to play an important role in modulating foraging behaviors in these insects, as in bees; indeed, it is well established that bees foraging for nectar

are less responsive to sucrose than bees foraging for pollen (Scheiner et al., 2004a). As a result, only the most concentrated nectar sources are exploited by bee nectar foragers. Such a phenomenon could also exist in most foraging moths, which land on flowers (i.e., settling moths but not hovering moths: Oliveira et al., 2004; Makholela and Manning, 2006; Okamoto et al., 2008; Atwater, 2013): it is likely the first appendage assessing sugar concentration in nectar are the legs (or possibly the proboscis) rather than the antennae (see also experiments by Zhang et al., 2010). Thus, only highly concentrated nectar would elicit a PER in moth (**Figure 3**), just like nectar-foraging bees. To explore this hypothesis, the foraging behavior of male and female moths drinking nectar at flowers should be analyzed either in artificial flowers in wind tunnel or in a natural setting. Very little information is available on this topic.

While moths respond quite well to sucrose and fructose, they hardly react to glucose. This is consistent to what was observed in a related species, *Spodoptera littoralis*, during electrophysiological recordings of taste sensilla (Popescu et al., 2013). In spite it is hardly responded to, glucose can replace fructose or sucrose in mixtures without significantly decreasing the PER rate. As nectars are mixtures of sugars rather than single compounded, this result suggests an adaptation to nectar perception. We can make the hypothesis that moths or other animals feeding on only one type of plant (i.e., strictly monophagous) should have a sugar responsiveness specifically tuned to the specific sugar ratio found in this plant's nectar, illustrating plant-pollinator coevolution (e.g., Perret et al., 2001; Nicolson and Fleming, 2003; Lotz and Schondube, 2006). This would have important consequences for the management of pests and pollinators. This hypothesis is also consistent with the fact that learning performance depends on sugar identity in bees (Simcock et al., 2018; Chapuy et al., 2019). By contrast, polyphagous moths such as *A. ipsilon* feed on various types of flowers, possibly with differing nectars (Wynne et al., 1991; Zhu et al., 1993). Nectars can be classified in various broad categories (Percival, 1961), and usually glucose is neither the dominant sugar nor the only one. Thus, it makes sense moths detect it less well. Honey bees are much better at detecting it (Simcock et al., 2018), but it is one of the main constituent of their honey. It would be interesting to compare sucrose responsiveness of other nectar foragers, particularly pollinators.

Honey bees and *Agrotis ipsilon* are sympatric, and both are described as polyphagous (i.e., they are not specialized in a narrow range of plant species but rather forage nectar on various flowers; Wynne et al., 1991; Zhu et al., 1993); thus, they potentially forage on the same plants. However, previous results (Scheiner et al., 2004a, 2013) indicate bees are much more sensitive to low sucrose concentrations than moths and so are likely to explore a wider range of flowers, including those which have nectar less concentrated. A full exploration of the ecological and evolutionary implications of this preliminary observation is beyond the scope of this article, but it certainly deserves to be studied; indeed, nectar foraging insects are often pollinators, and it is relevant to know which plants will or will not be visited by a given insect in a context of pollination

crisis (Senapathi et al., 2015). The sucrose responsiveness assay will be useful to do so.

We found little differences between males and females, and they were not consistently observed. It would be interesting to compare sugar responsiveness in animals before and after reproduction; for males, that would mean before flying toward a female and mating, and for females, before releasing pheromone, mating and laying eggs. It is not unlikely the needs of moths change according to their reproductive status. In particular, females might need amino acids, while eggs are storing reserves (Watt et al., 1974; Heil, 2011; Hendriksma et al., 2014; Simcock et al., 2014; Levin et al., 2017a,b), so that PER rates would be affected by their presence in the tested solutions (but see O'Brien et al., 2000; Marchioro and Foerster, 2013 for contradictory results); Zhang et al. (2010) reported neuronal responses and PER to amino acids. Similarly, males might need sodium and potassium for spermatophore formation (Arms et al., 1974; Adler and Pearson, 1982; Boggs and Jackson, 1991; Smedley and Eisner, 1995, 1996; Beck et al., 1999; Boggs and Dau, 2004; Watanabe and Kamikubo, 2005; Molleman, 2010), so that the presence of salt in the solution might affect the PER rate. The sugar responsiveness assay can easily be modified to test the effect of salt or amino acid on males and females of different reproductive status.

Presenting quinine on the proboscis interrupts the PER, and it inhibits response to sugar. This contrasts to the behavior of bees (which readily drink bitter solutions when they are restrained but not when they are free flying, Ayestaran et al., 2010; De Brito Sanchez et al., 2015; Guiraud et al., 2018) and flies (which accepts bitter compounds after starvation, Ledue et al., 2016). It is established that some plants produce bitter compounds in nectar (Adler, 2000; Adler et al., 2006; Heil, 2011; Roy et al., 2017), possibly to deter predatory insects. It can be expected that *A. ipsilon* would not forage on these plants, but this remains to be verified; this would be consistent with the idea that this moth is a generalist forager, because specialists tend to adapt to deterrent nectar compounds (Berenbaum et al., 1989; Glendinning, 2002; Cornell and Hawkins, 2003; Adler et al., 2006; Reisenman and Riffell, 2015). This also opens the possibility to use the sugar responsiveness assay to evaluate moths' reaction to aversive compounds such as deterrent products released in nectar, or possibly xenobiotics. Beside, some results indicate appetitive and deterrent stimuli are processed in parallel at the antenna level (Kvello et al., 2010); this raises the question of the neurophysiological convergence of these pathways, which must occur at some point as quinine prevents sucrose-induced PER. Interestingly, pre-exposing *S. littoralis* to quinine or sucrose 24 h before sucrose presentation potentiates rather than inhibits the response to 1% sucrose (Minoli et al., 2012). This suggests that previous gustatory experiments are not neutral for sucrose responsiveness.

Neurophysiological Substrates

Gustatory receptors and the downward neurophysiological pathways have been reported on antennae, proboscis, and legs (Jørgensen et al., 2006, 2007; Calas et al., 2009; Kvello et al., 2010; Zhang et al., 2010; Popescu et al., 2013; Agnihotri et al., 2016; Seada

et al., 2018); these neurophysiological results are consistent with our behavioral observations, as they indicated that sugar-induced neuronal responses are lower in the legs than in the proboscis, and glucose alone elicits less responses than sucrose or fructose. Quinine was also well detected but not by the neurons responding to sucrose (Jørgensen et al., 2007). This suggests its effect result from a central integration, rather than an interaction at the level of gustatory receptors; this is different in fruit flies, where bitter compounds can interact at the level of the receptor (Meunier et al., 2003; Jeong et al., 2013; French et al., 2015a,b). Popescu et al. (2013) observed sex-dependent responses, suggesting the differences we observed in **Figure 1B** (but not in **Figure 6**) are supported by differing response levels.

To fully understand how sugars are perceived and responded to by *A. ipsilon* and other moths, it will be necessary to describe the full neuronal pathway of information from antennal gustatory receptors to PER release. At the level of antennae, characterization of the receptors has already been started in moths and involves description of the antennal gustatory sensilla that house gustatory neurons as well as molecular description of the receptors and their interactions with sugars (Balkenius and Kelber, 2006; Popescu et al., 2013; Seada, 2015; Agnihotri et al., 2016; Seada et al., 2018); it would be particularly interesting to describe interactions in mixtures of sugars, quinine, and/or amino acid, specially to find out whether synergy or masking occurs, as for odors (Rospars, 2013); a tempting hypothesis is that glucose would act as a modulator of sugar receptor, explaining it elicits little response while favoring response to other sugars. In turn, gustatory information projects to the central nervous system, particularly the suboesophageal ganglion (Jørgensen et al., 2006; Kvello et al., 2006, 2010; Popescu et al., 2013). How motivation-linked information (hunger and/or other factors) is then integrated with gustatory input to elicit (or not) a PER remains to be determined. While electrophysiological and molecular tools will be needed to reach this goal, the sucrose responsiveness assay provides a solid framework to guide such research.

As previously discussed, stimulating legs with sucrose is much less prone to elicit a PER than stimulating antennae. Assuming motivation to release a PER and the corresponding motor control are central processes, a simple explanation is that legs bear less sugar receptors than antennae and/or that they bear different receptors (i.e., less sensitive to sucrose in the legs). Alternatively, gustatory pathways from the legs could be somehow less connected to the release of PER than those from antennae. Once again, electrophysiology (recording of gustatory sensilla from legs and antennae) will be helpful to distinguish between these two hypotheses, by determining whether gustatory responses of legs and antennae are comparable (Calas et al., 2009; Zhang et al., 2010).

Finally, understanding the central integration of sucrose input that leads to PER will be the ultimate goal. A way to approach this objective will be to understand which neurotransmitter systems are involved. Promising candidates are biogenic amines such as octopamine and dopamine, as they have well-known roles on sucrose sensitivity in other insects (Pankiw and Page, 2003; Schwaerzel et al., 2003; Unoki et al., 2005, 2006;

Scheiner et al., 2006, 2014; Mizunami et al., 2009; Matsumoto et al., 2015). Interestingly, biogenic amines also modulate sex-pheromone-elicited behaviors in male moths (Linn and Roelofs, 1986; Linn et al., 1992; Pophof, 2000; Flecke and Stengl, 2009; Jarriault et al., 2009; Duportets et al., 2010; Abrieux et al., 2014; Hillier and Kavanagh, 2015), while sex pheromones do not affect sucrose responsiveness (Hostachy et al., submitted). If these neurotransmitters modulated sucrose responsiveness, this would imply that they do so through ways different from pheromone-responsiveness modulation.

DATA AVAILABILITY STATEMENT

All datasets generated for this study are included in the article/**Supplementary Material**.

AUTHOR CONTRIBUTIONS

CH, ND, and MD designed the experiments. CH and MD prepared the figures and analyzed the data. MD wrote the first draft of the manuscript. All authors performed the

REFERENCES

- Abrieux, A., Duportets, L., Debernard, S., Gadenne, C., and Anton, S. (2014). The GPCR membrane receptor, DopEcR, mediates the actions of both dopamine and ecdysone to control sex pheromone perception in an insect. *Front. Behav. Neurosci.* 8:312. doi: 10.3389/fnbeh.2014.00312
- Adler, L. S. (2000). The ecological significance of toxic nectar. *Oikos* 91, 409–420. doi: 10.1034/j.1600-0706.2000.910301.x
- Adler, L. S., and Irwin, R. E. (2005). Ecological costs and benefits of defenses in nectar. *Ecology* 86, 2968–2978. doi: 10.1890/05-0118
- Adler, P. H., and Pearson, D. L. (1982). Why do male butterflies visit mud puddles? *Can. J. Zool.* 60, 322–325. doi: 10.1139/z82-043
- Adler, L. S., Wink, M., Distl, M., and Lentz, A. J. (2006). Leaf herbivory and nutrients increase nectar alkaloids. *Ecol. Lett.* 9, 960–967. doi: 10.1111/j.1461-0248.2006.00944.x
- Agnihotri, A. R., Roy, A. A., and Joshi, R. S. (2016). Gustatory receptors in Lepidoptera: chemosensation and beyond. *Insect Mol. Biol.* 25, 519–529. doi: 10.1111/imb.12246
- Akre, K. L., and Johnsen, S. (2014). Psychophysics and the evolution of behavior. *Trends Ecol. Evol.* 29, 291–300. doi: 10.1016/j.tree.2014.03.007
- Anderson, P., Hansson, B. S., Nilsson, U., Han, Q., Sjöholm, M., Skals, N., et al. (2007). Increased behavioral and neuronal sensitivity to sex pheromone after brief odor experience in a moth. *Chem. Senses* 32, 483–491. doi: 10.1093/chemse/bjm017
- Anton, S., Evengard, K., Barrozo, R. B., Anderson, P., and Skals, N. (2011). Brief predator sound exposure elicits behavioral and neuronal long-term sensitization in the olfactory system of an insect. *PNAS* 108, 3401–3405. doi: 10.1073/pnas.1008840108
- Arms, K., Feeny, P., and Lederhouse, R. C. (1974). Sodium: stimulus for puddling behavior by tiger swallowtail butterflies, *Papilio glaucus*. *Science* 185, 372–374. doi: 10.1126/science.185.4148.372
- Atwater, M. (2013). Diversity and nectar hosts of flower-settling moths within a Florida sandhill ecosystem. *J. Nat. Hist.* 47, 2719–2734. doi: 10.1080/00222933.2013.791944
- Ayestaran, A., Giurfa, M., and De Brito Sanchez, M. G. (2010). Toxic but drank: gustatory aversive compounds induce post-ingestional malaise in harnessed honeybees. *PLoS One* 5:e15000. doi: 10.1371/journal.pone.0015000
- Baker, H. G. (1977). Non-sugar chemical constituents of nectar. *Apidologie* 8, 349–356. doi: 10.1051/apido:19770405
- Baker, H. G., and Baker, I. (1973). Amino-acids in nectar and their evolutionary significance. *Nature* 241, 543–545. doi: 10.1038/241543b0
- Balkenius, A., and Kelber, A. (2006). Colour preferences influences odour learning in the hawkmoth, *Macroglossum stellatarum*. *Naturwissenschaften* 93, 255–258. doi: 10.1007/s00114-006-0099-9
- Baracchi, D., Rigosi, E., De Brito Sanchez, G., and Giurfa, M. (2018). Lateralization of sucrose responsiveness and non-associative learning in honeybees. *Front. Psychol.* 9:425. doi: 10.3389/fpsyg.2018.00425
- Beck, J., Mühlenberg, E., and Fiedler, K. (1999). Mud-puddling behavior in tropical butterflies: in search of proteins or minerals? *Oecologia* 119, 140–148. doi: 10.1007/s004420050770
- Berenbaum, M. R., Zangerl, A. R., and Lee, K. (1989). Chemical barriers to adaptation by a specialist herbivore. *Oecologia* 80, 501–506. doi: 10.1007/BF00380073
- Blumstein, D. T. (2016). Habituation and sensitization: new thoughts about old ideas. *Anim. Behav.* 120, 255–262. doi: 10.1016/j.anbehav.2016.05.012
- Boggs, C. L. (1997a). Dynamics of reproductive allocation from juvenile and adult feeding: radiotracer studies. *Ecology* 78, 192–202.
- Boggs, C. L. (1997b). Reproductive allocation from reserves and income in butterfly species with differing adult diets. *Ecology* 78, 181–191.
- Boggs, C. L., and Dau, B. (2004). Resource specialization in puddling Lepidoptera. *Environ. Entomol.* 33, 1020–1024. doi: 10.1603/0046-225X-33.4.1020
- Boggs, C. L., and Jackson, L. A. (1991). Mud puddling by butterflies is not a simple matter. *Ecol. Entomol.* 16, 123–127. doi: 10.1111/j.1365-2311.1991.tb00199.x
- Boggs, C. L., and Ross, C. L. (1993). The effect of adult food limitation on life history traits in *Speyeria mormonia* (Lepidoptera: Nymphalidae). *Ecology* 74, 433–441. doi: 10.2307/1939305
- Calas, D., Marion-Poll, F., and Steinbauer, M. J. (2009). Tarsal taste sensilla of the autumn gum moth, *Mnesampela privata*: morphology and electrophysiological activity. *Entomol. Exp. Appl.* 133, 186–192. doi: 10.1111/j.1570-7458.2009.00921.x
- Camacho, M., Oliva, M., and Serbus, L. R. (2017). Dietary saccharides and sweet tastants have differential effects on colonization of *Drosophila* oocytes by *Wolbachia* endosymbionts. *Biol. Open* 6, 1074–1083. doi: 10.1242/bio.023895
- Chabaud, M., Devaud, J., Pham-Delègue, M., Preat, T., and Kaiser, L. (2006). Olfactory conditioning of proboscis activity in *Drosophila melanogaster*. *J. Comp. Physiol. A* 192, 1335–1348. doi: 10.1007/s00359-006-0160-3

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fphys.2019.01423/full#supplementary-material>

- Chaffiol, A., Laloi, D., and Pham-Delegue, M. H. (2005). Prior classical olfactory conditioning improves odour-cued flight orientation of honey bees in a wind tunnel. *J. Exp. Biol.* 208, 3731–3737. doi: 10.1242/jeb.01796
- Chapuy, C., Ribbens, L., Renou, M., Dacher, M., and Armengaud, C. (2019). Thymol affects congruency between olfactory and gustatory stimuli in bees. *Sci. Rep.* 9:7752. doi: 10.1038/s41598-019-43614-8
- Choi, M.-Y., Tang, S. B., Ahn, S.-J., Amarasekare, K. G., Shearer, P., and Lee, J. C. (2017). Effect of non-nutritive sugars to decrease the survivorship of spotted wing *Drosophila*, *Drosophila suzukii*. *J. Insect Physiol.* 99, 86–94. doi: 10.1016/j.jinsphys.2017.04.001
- Cook, S. M., Khan, Z. R., and Pickett, J. A. (2007). The use of push-pull strategies in integrated pest management. *Annu. Rev. Entomol.* 52, 375–400. doi: 10.1146/annurev.ento.52.110405.091407
- Cornell, H. V., and Hawkins, B. A. (2003). Herbivore responses to plant secondary compounds: a test of phytochemical coevolution theory. *Am. Nat.* 161, 507–522. doi: 10.1086/368346
- Dacher, M., and Smith, B. H. (2008). Olfactory interference during inhibitory backward pairing in honey bees. *PLoS One* 3:e3513. doi: 10.1371/journal.pone.0003513
- De Brito Sanchez, M. G., Serre, M., Avarguès-Weber, A., Dyer, A. G., and Giurfa, M. (2015). Learning context modulates aversive taste strength in honey bees. *J. Exp. Biol.* 218, 949–959. doi: 10.1242/jeb.117333
- De La Barrera, E., and Nobel, P. S. (2004). Nectar: properties, floral aspects, and speculations on origin. *Trends Plant Sci.* 9, 65–69. doi: 10.1016/j.tplants.2003.12.003
- Duportets, L., Barrozo, R. B., Bozzolan, F., Gaertner, C., Anton, S., Gadenne, C., et al. (2010). Cloning of an octopamine/tyramine receptor and plasticity of its expression as a function of adult sexual maturation in the male moth *Agrotis ipsilon*. *Insect Mol. Biol.* 19, 489–499. doi: 10.1111/j.1365-2583.2010.01009.x
- Dus, M., Min, S., Keene, A. C., Lee, G. Y., and Suh, G. S. (2011). Taste-independent detection of the caloric content of sugar in *Drosophila*. *Proc. Natl. Acad. Sci. USA* 108, 11644–11649. doi: 10.1073/pnas.1017096108
- Evenenden, M. L. (2016). “Mating disruption of moth pests in integrated pest management: a mechanistic approach” in *Pheromone communication in moths: Evolution, behavior and application*. eds. J. D. Allison and R. T. Carde (Oakland, CA: University of California Press), 365–393.
- Fan, R. J., Anderson, P., and Hansson, B. (1997). Behavioural analysis of olfactory conditioning in the moth *Spodoptera littoralis* (Boisd.) (Lepidoptera: Noctuidae). *J. Exp. Biol.* 200, 2969–2976.
- Fan, R. J., and Hansson, B. S. (2001). Olfactory discrimination conditioning in the moth *Spodoptera littoralis*. *Physiol. Behav.* 72, 159–165. doi: 10.1016/S0031-9384(00)00394-2
- Fischer, K., O'Brien, D. M., and Boggs, C. L. (2004). Allocation of larval and adult resources to reproduction in a fruit-feeding butterfly. *Funct. Ecol.* 18, 656–663. doi: 10.1111/j.0269-8463.2004.00892.x
- Fisher, M. L., Fowler, F. E., Denning, S. S., and Watson, D. W. (2017). Survival of the house fly (Diptera: Muscidae) on Truvia and other sweeteners. *J. Med. Entomol.* 54, 999–1005. doi: 10.1093/jme/tjw241
- Flecke, C., and Stengl, M. (2009). Octopamine and tyramine modulate pheromone-sensitive olfactory sensilla of the hawkmoth *Manduca sexta* in a time-dependent manner. *J. Comp. Physiol. A* 195, 529–545. doi: 10.1007/s00359-009-0429-4
- French, A., Agha, M. A., Mitra, A., Yanagawa, A., Sellier, M. J., and Marion-Poll, F. (2015a). *Drosophila* bitter taste(s). *Front. Integr. Neurosci.* 9:58. doi: 10.3389/fnint.2015.00058
- French, A. S., Sellier, M.-J., Ali Agha, M., Guigue, A., Chabaud, M.-A., Reeb, P. D., et al. (2015b). Dual mechanism for bitter avoidance in *Drosophila*. *J. Neurosci.* 35, 3990–4004. doi: 10.1523/jneurosci.1312-14.2015
- Fresquet, N. (1999). Effects of aging on the acquisition and extinction of excitatory conditioning in *Drosophila melanogaster*. *Physiol. Behav.* 67, 205–211. doi: 10.1016/S0031-9384(99)00058-X
- Geister, T. L., Lorenz, M. W., Hoffmann, K. H., and Fischer, K. (2008). Adult nutrition and butterfly fitness: effects of diet quality on reproductive output, egg composition, and egg hatching success. *Front. Zool.* 5:10. doi: 10.1186/1742-9994-5-10
- Gemeno, C., and Haynes, K. F. (2000). Periodical and age-related variation in chemical communication system of black cutworm moth, *Agrotis ipsilon*. *J. Chem. Ecol.* 26, 329–342. doi: 10.1023/A:1005468203045
- Gerber, B., Geberzahn, N., Hellstern, F., Klein, J., Kowalsky, O., Wüstenberg, D., et al. (1996). Honeybees transfer olfactory memories established during flower visits to a proboscis extension paradigm in the laboratory. *Anim. Behav.* 52, 1079–1085. doi: 10.1006/anbe.1996.0255
- Gil, M., and De Marco, R. J. (2005). Olfactory learning by means of trophallaxis in *Apis mellifera*. *J. Exp. Biol.* 208, 671–680. doi: 10.1242/jeb.01474
- Giurfa, M. (2015). Learning and cognition in insects. *Wiley Interdiscip. Rev. Cogn. Sci.* 6, 383–395. doi: 10.1002/wcs.1348
- Giurfa, M., and Sandoz, J. C. (2012). Invertebrate learning and memory: fifty years of olfactory conditioning of the proboscis extension response in honeybees. *Learn. Mem.* 19, 54–66. doi: 10.1101/lm.024711.111
- Glendinning, J. I. (2002). How do herbivorous insects cope with noxious secondary plant compounds in their diet? *Entomol. Exp. App.* 104, 15–25. doi: 10.1046/j.1570-7458.2002.00986.x
- Gottsberger, G., Schrauven, J., and Linskens, H. F. (1984). Amino acids and sugars in nectar, and their putative evolutionary significance. *Plant Syst. Evol.* 145, 55–77. doi: 10.1007/BF00984031
- Groves, P. M., and Thompson, R. F. (1970). Habituation: a dual-process theory. *Psychol. Rev.* 77, 419–450. doi: 10.1037/h0029810
- Guerrieri, F. J., and D'Etto, P. (2010). Associative learning in ants: conditioning of the maxilla-labium extension response in *Camponotus aethiops*. *J. Insect Physiol.* 56, 88–92. doi: 10.1016/j.jinsphys.2009.09.007
- Guiraud, M., Hotier, L., Giurfa, M., and De Brito Sanchez, M. G. (2018). Aversive gustatory learning and perception in honey bees. *Sci. Rep.* 8:1343. doi: 10.1038/s41598-018-19715-1
- Hammer, M., Braun, G., and Maelshagen, J. (1994). Food-induced arousal and nonassociative learning in honeybees: dependence of sensitization on the application site and duration of food stimulation. *Behav. Neural Biol.* 62, 210–223. doi: 10.1016/S0163-1047(05)80019-6
- Hartlieb, E. (1996). Olfactory conditioning in the moth *Heliothis virescens*. *Naturwissenschaften* 83, 87–88.
- Hartlieb, E., Anderson, P., and Hansson, B. S. (1999a). Appetitive learning of odours with different behavioural meaning in moths. *Physiol. Behav.* 67, 671–677.
- Hartlieb, E., Hansson, B. S., and Anderson, P. (1999b). Sex or food? Appetitive learning of sex odors in a male moth. *Naturwissenschaften* 86, 396–399.
- Heil, M. (2011). Nectar: generation, regulation and ecological functions. *Trends Plant Sci.* 16, 191–200. doi: 10.1016/j.tplants.2011.01.003
- Hendriksma, H. P., Oxman, K. L., and Shafir, S. (2014). Amino acid and carbohydrate tradeoffs by honey bee nectar foragers and their implications for plant–pollinator interactions. *J. Insect Physiol.* 69, 56–64. doi: 10.1016/j.jinsphys.2014.05.025
- Hill, C. J. (1989). The effect of adult diet on the biology of butterflies. 2. The common crow butterfly, *Euploea core corinna*. *Oecologia* 81, 258–266. doi: 10.1007/BF00379813
- Hill, C. J., and Pierce, N. E. (1989). The effect of adult diet on the biology of butterflies. 1. The common imperial blue, *Jalmenus evagoras*. *Oecologia* 81, 249–257. doi: 10.1007/BF00379812
- Hillier, N. K., and Kavanagh, R. M. B. (2015). Differential octopaminergic modulation of olfactory receptor neuron responses to sex pheromones in *Heliothis virescens*. *PLoS One* 10:e0143179. doi: 10.1371/journal.pone.0143179
- Holm, S. (1979). A simple sequentially rejective multiple test procedure. *Scand. J. Stat.* 6, 65–70.
- Jarriault, D., Barrozo, R. B., Pinto, C. J. D., Greiner, B., Dufour, M. C., Masante-Roca, I., et al. (2009). Age-dependent plasticity of sex pheromone response in the moth, *Agrotis ipsilon*: combined effects of octopamine and juvenile hormone. *Horm. Behav.* 56, 185–191. doi: 10.1016/j.yhbeh.2009.04.005
- Jeong, Y. T., Shim, J., Oh, S. R., Yoon, H. I., Kim, C. H., Moon, S. J., et al. (2013). An odorant-binding protein required for suppression of sweet taste by bitter chemicals. *Neuron* 79, 725–737. doi: 10.1016/j.neuron.2013.06.025
- Jervis, M. A., Boggs, C. L., and Ferns, P. N. (2005). Egg maturation strategy and its associated trade-offs: a synthesis focusing on Lepidoptera. *Ecol. Entomol.* 30, 359–375. doi: 10.1111/j.0307-6946.2005.00712.x
- Jørgensen, K., Almaas, T. J., Marion-Poll, F., and Mustaparta, H. (2007). Electrophysiological characterization of responses from gustatory receptor neurons of sensilla chaetica in the moth *Heliothis virescens*. *Chem. Senses* 32, 863–879. doi: 10.1093/chemse/bjm057
- Jørgensen, K., Kvello, P., Almaas, T. J., and Mustaparta, H. (2006). Two closely located areas in the subesophageal ganglion and the tritocerebrum receive projections of gustatory receptor neurons located on the antennae and the proboscis in the moth *Heliothis virescens*. *J. Comp. Neurol.* 496, 121–134. doi: 10.1002/cne.20908

- Jorgensen, K., Strandén, M., Sandoz, J.-C., Menzel, R., and Mustaparta, H. (2007). Effects of two bitter substances on olfactory conditioning in the moth *Heliothis virescens*. *J. Exp. Biol.* 210, 2563–2573. doi: 10.1242/jeb.004283
- Jozet-Alves, C., Viblan, V. A., Romagny, S., Dacher, M., Healy, S. D., and Dickel, L. (2012). Visual lateralization is task and age dependent in cuttlefish, *Sepia officinalis*. *Anim. Behav.* 83, 1313–1318. doi: 10.1016/j.anbehav.2012.02.023
- Kroutov, V., Mayer, M. S., and Emmel, T. C. (1999). Olfactory conditioning of the butterfly *Agraulis vanillae* (L.) (Lepidoptera, Nymphalidae) to floral but not host-plant odors. *J. Insect Behav.* 12, 833–843. doi: 10.1023/A:1020961211750
- Kvello, P., Almaas, T. J., and Mustaparta, H. (2006). A confined taste area in a lepidopteran brain. *Arthr. Struc. Dev.* 35, 35–45. doi: 10.1016/j.asd.2005.10.003
- Kvello, P., Jørgensen, K., and Mustaparta, H. (2010). Central gustatory neurons integrate taste quality information from four appendages in the moth *Heliothis virescens*. *J. Neurophysiol.* 103, 2965–2981. doi: 10.1152/jn.00985.2009
- Labrousse, C., Lazzari, C. R., and Fresquet, N. (2017). Developmental study of the proboscis extension response to heat in *Rhodnius prolixus* along the life cycle. *J. Insect Physiol.* 98, 55–58. doi: 10.1016/j.jinsphys.2016.11.010
- Ledue, E. E., Mann, K., Koch, E., Chu, B., Dakin, R., and Gordon, M. D. (2016). Starvation-induced depotentiation of bitter taste in *Drosophila*. *Curr. Biol.* 26, 2854–2861. doi: 10.1016/j.cub.2016.08.028
- Levin, E., Mccue, M. D., and Davidowitz, G. (2017a). More than just sugar: allocation of nectar amino acids and fatty acids in a Lepidopteran. *Proc. R. Soc. B* 284:20162126. doi: 10.1098/rspb.2016.2126
- Levin, E., Mccue, M. D., and Davidowitz, G. (2017b). Sex differences in the utilization of essential and non-essential amino acids in Lepidoptera. *J. Exp. Biol.* 220, 2743–2747. doi: 10.1242/jeb.154757
- Linn, C. E., Campbell, M. G., and Roelofs, W. L. (1992). Photoperiod cues and the modulatory action of octopamine and 5-hydroxytryptamine on locomotor and pheromone response in male gypsy moths, *Lymantria dispar*. *Arch. Insect Biochem. Physiol.* 20, 265–284. doi: 10.1002/arch.940200404
- Linn, C. E., and Roelofs, W. L. (1986). Modulatory effects of octopamine and serotonin on male sensitivity and periodicity of response to sex pheromone in the cabbage looper moth, *Trichoplusia ni*. *Arch. Insect Biochem. Physiol.* 3, 161–171. doi: 10.1002/arch.940030206
- Lotz, C. N., and Schondube, J. E. (2006). Sugar preferences in nectar- and fruit-eating birds: behavioral patterns and physiological causes. *Biotropica* 38, 3–15. doi: 10.1111/j.1744-7429.2006.00104.x
- Lüttge, U. (1977). Nectar composition and membrane transport of sugars and amino acids: a review on the present state of nectar research. *Apidologie* 8, 305–319. doi: 10.1051/apido:19770402
- Makholela, T., and Manning, J. C. (2006). First report of moth pollination in *Struthiola ciliata* (Thymelaeaceae) in southern Africa. *South African J. Bot.* 72, 597–603. doi: 10.1016/j.sajb.2006.04.007
- Marchiori, C. A., and Foerster, L. A. (2013). Effects of adult-derived carbohydrates and amino acids on the reproduction of *Plutella xylostella*. *Physiol. Entomol.* 38, 13–19. doi: 10.1111/phen.12000
- Matsumoto, Y., Matsumoto, C.-S., Wakuda, R., Ichihara, S., and Mizunami, M. (2015). Roles of octopamine and dopamine in appetitive and aversive memory acquisition studied in olfactory conditioning of maxillary palpi extension response in crickets. *Front. Behav. Neurosci.* 9:230. doi: 10.3389/fnbeh.2015.00230
- Menzel, R. (1999). Memory dynamics in the honeybee. *J. Comp. Physiol. A* 185, 323–340.
- Menzel, R. (2012). The honeybee as a model for understanding the basis of cognition. *Nat. Rev. Neurosci.* 13, 758–768. doi: 10.1038/nrn3357
- Meunier, N., Marion-Poll, F., Rospars, J.-P., and Tanimura, T. (2003). Peripheral coding of bitter taste in *Drosophila*. *J. Neurobiol.* 56, 139–152. doi: 10.1002/neu.10235
- Minoli, S., Kauer, I., Colson, V., Party, V., Renou, M., Anderson, P., et al. (2012). Brief exposure to sensory cues elicits stimulus-nonspecific general sensitization in an insect. *PLoS One* 7:e34141. doi: 10.1371/journal.pone.0034141
- Mizunami, M., Unoki, S., Mori, Y., Hirashima, D., Hatano, A., and Matsumoto, Y. (2009). Roles of octopaminergic and dopaminergic neurons in appetitive and aversive memory recall in an insect. *BMC Biol.* 7, 1–16. doi: 10.1186/1741-7007-7-46
- Molleman, F. (2010). Puddling: from natural history to understanding how it affects fitness. *Entomol. Exp. Appl.* 134, 107–113. doi: 10.1111/j.1570-7458.2009.00938.x
- Mujagic, S., Sarkander, J., Erber, B., and Erber, J. (2010). Sucrose acceptance and different forms of associative learning of the honey bee (*Apis mellifera* L.) in the field and laboratory. *Front. Behav. Neurosci.* 4:46. doi: 10.3389/fnbeh.2010.00046
- Mustard, J. A., Alvarez, V., Barocio, S., Mathews, J., Stoker, A., and Malik, K. (2018). Nutritional value and taste play different roles in learning and memory in the honey bee (*Apis mellifera*). *J. Insect Physiol.* 107, 250–256. doi: 10.1016/j.jinsphys.2018.04.014
- Nachev, V., Thomson, J. D., and Winter, Y. (2013). The psychophysics of sugar concentration discrimination and contrast evaluation in bumblebees. *Anim. Cogn.* 16, 417–427. doi: 10.1007/s10071-012-0582-y
- Naranjo, S. E., Ellsworth, P. C., and Frisvold, G. B. (2015). Economic value of biological control in integrated pest management of managed plant systems. *Annu. Rev. Entomol.* 60, 621–645. doi: 10.1146/annurev-ento-010814-021005
- Nepi, M., Soligo, C., Nocentini, D., Abate, M., Guarnieri, M., Cai, G., et al. (2012). Amino acids and protein profile in floral nectar: much more than a simple reward. *Flora* 207, 475–481. doi: 10.1016/j.flora.2012.06.002
- Nicolson, S. W., and Fleming, P. A. (2003). Nectar as food for birds: the physiological consequences of drinking dilute sugar solutions. *Plant Syst. Evol.* 238, 139–153. doi: 10.1007/s00606-003-0276-7
- Niven, J. E., and Frasnelli, E. (2018). Insights into the evolution of lateralization from the insects. *Prog. Brain Res.* 238, 3–31. doi: 10.1016/bs.pbr.2018.06.001
- O'Brien, D. M., Boggs, C. L., and Fogel, M. L. (2004). Making eggs from nectar: the role of life history and dietary carbon turnover in butterfly reproductive resource allocation. *Oikos* 105, 279–291. doi: 10.1111/j.0030-1299.2004.13012.x
- O'Brien, D. M., Schrag, D. P., and Del Rio, C. M. (2000). Allocation to reproduction in a hawkmoth: a quantitative analysis using stable carbon isotopes. *Ecology* 81, 2822–2831. doi: 10.1890/0012-9658(2000)081[2822:ATRIAH]2.0.CO;2
- Okamoto, T., Kawakita, A., and Kato, M. (2008). Floral adaptations to nocturnal moth pollination in *Diplomorpha* (Thymelaeaceae). *Plant Species Biol.* 23, 192–201. doi: 10.1111/j.1442-1984.2008.00222.x
- Oliveira, P. E., Gibbs, P. E., and Barbosa, A. A. (2004). Moth pollination of woody species in the Cerrados of Central Brazil: a case of so much owed to so few? *Plant Syst. Evol.* 245, 41–54. doi: 10.1007/s00606-003-0120-0
- Pacini, E., Nepi, M., and Vespri, L. J. (2003). Nectar biodiversity: a short review. *Plant Syst. Evol.* 238, 7–21. doi: 10.1007/s00606-002-0277-y
- Page, R. E. Jr., and Erber, J. (2002). Levels of behavioral organization and the evolution of division of labor. *Naturwissenschaften* 89, 91–106. doi: 10.1007/s00114-002-0299-x
- Pankiw, T., and Page, R. E. (2003). Effect of pheromones, hormones, and handling on sucrose response thresholds of honey bees (*Apis mellifera* L.). *J. Comp. Physiol. A* 189, 675–684. doi: 10.1007/s00359-003-0442-y
- Percival, M. S. (1961). Types of nectar in angiosperms. *New Phytol.* 60, 235–281. doi: 10.1111/j.1469-8137.1961.tb06255.x
- Perez, M., Rolland, U., Giurfa, M., and Dèttorre, P. (2013). Sucrose responsiveness, learning success, and task specialization in ants. *Learn. Mem.* 20, 417–420. doi: 10.1101/lm.031427.113
- Perret, M., Chautems, A., Spichiger, R., Peixoto, M., and Savolainen, V. (2001). Nectar sugar composition in relation to pollination syndromes in Sinningieae (Gesneriaceae). *Ann. Bot.* 87, 267–273. doi: 10.1006/anbo.2000.1331
- Popescu, A., Couton, L., Almaas, T.-J., Rospars, J.-P., Wright, G., Marion-Poll, F., et al. (2013). Function and central projections of gustatory receptor neurons on the antenna of the noctuid moth *Spodoptera littoralis*. *J. Comp. Physiol. A* 199, 403–416. doi: 10.1007/s00359-013-0803-0
- Pophof, B. (2000). Octopamine modulates the sensitivity of silkworm pheromone receptor neurons. *J. Comp. Physiol. A* 186, 307–313. doi: 10.1007/s003590050431
- Prieur, J., Lemasson, A., Barbu, S., and Blois-Heulin, C. (2019). History, development and current advances concerning the evolutionary roots of human right-handedness and language: brain lateralisation and manual laterality in non-human primates. *Ethology* 125, 1–28. doi: 10.1111/eth.12827
- Pszczolkowski, M. A., and Brown, J. J. (2003). Effect of sugars and non-nutritive sugar substitutes on consumption of apple leaves by codling moth neonates. *Phytoparasitica* 31, 283–291. doi: 10.1007/BF02980837
- R Core Team (2017). *R: A language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing. Available at: <https://www.R-project.org/> (Accessed November 13, 2019).
- Reisenman, C. E., and Riffell, J. A. (2015). The neural bases of host plant selection in a neuroecology framework. *Front. Invert. Physiol.* 6:229. doi: 10.3389/fphys.2015.00229

- Rospars, J.-P. (2013). Interactions of odorants with olfactory receptors and other preprocessing mechanisms: how complex and difficult to predict? *Chem. Senses* 38, 283–287. doi: 10.1093/chemse/bjt004
- Roy, R., Schmitt, A. J., Thomas, J. B., and Carter, C. J. (2017). Nectar biology: from molecules to ecosystems. *Plant Sci.* 262, 148–164. doi: 10.1016/j.plantsci.2017.04.012
- Sandoz, J. C. (2011). Behavioral and neurophysiological study of olfactory perception and learning in honeybees. *Front. Syst. Neurosci.* 5:98. doi: 10.3389/fnsys.2011.00098
- Sandoz, J. C., Laloi, D., Odoux, J. F., and Pham-Delegue, M. H. (2000). Olfactory information transfer in the honeybee: compared efficiency of classical conditioning and early exposure. *Anim. Behav.* 59, 1025–1034. doi: 10.1006/anbe.2000.1395
- Scheiner, R., Abramson, C., Brodschneider, R., Craisheim, K., Farina, W., Fuchs, S., et al. (2013). Standard methods for behavioural studies of *Apis mellifera*. *J. Apic. Res.* 52, 1–58. doi: 10.3896/IBRA.1.52.4.04
- Scheiner, R., Barnert, M., and Erber, J. (2003). Variation in water and sucrose responsiveness during the foraging season affects proboscis extension learning in honeybees. *Apidologie* 34, 67–72. doi: 10.1051/apido:2002050
- Scheiner, R., Baumann, A., and Blenau, W. (2006). Aminergic control and modulation of honeybee behaviour. *Curr. Neuropharmacol.* 4, 259–276. doi: 10.2174/157015906778520791
- Scheiner, R., Page, R. E., and Erber, J. (2004a). Sucrose responsiveness and behavioral plasticity in honey bees (*Apis mellifera*). *Apidologie* 35, 133–142. doi: 10.3896/IBRA.1.52.4.04
- Scheiner, R., Sokolowski, M. B., and Erber, J. (2004b). Activity of cGMP-dependent protein kinase (PKG) affects sucrose responsiveness and habituation in *Drosophila melanogaster*. *Learn. Mem.* 11, 303–311. doi: 10.1101/lm.71604
- Scheiner, R., Steinbach, A., Claßen, G., Strudthoff, N., and Scholz, H. (2014). Octopamine indirectly affects proboscis extension response habituation in *Drosophila melanogaster* by controlling sucrose responsiveness. *J. Insect Physiol.* 69, 107–117. doi: 10.1016/j.jinsphys.2014.03.011
- Schiff, N. M., Waldbauer, G. P., and Friedman, S. (1989). Dietary self-selection by *Heliothis zea* larvae: roles of metabolic feedback and chemosensory stimuli. *Ent. Exp. Appl.* 52, 261–270. doi: 10.1111/j.1570-7458.1989.tb01276.x
- Schwaerzel, M., Monastirioti, M., Scholz, H., Friggi-Grelin, F., Birman, S., and Heisenberg, M. (2003). Dopamine and octopamine differentiate between aversive and appetitive olfactory memories in *Drosophila*. *J. Neurosci.* 23, 10495–10502. doi: 10.1523/JNEUROSCI.23-33-10495.2003
- Seada, M. A. (2015). Antennal morphology and sensillum distribution of female cotton leaf worm *Spodoptera littoralis* (Lepidoptera: Noctuidae). *J. Basic Appl. Zool.* 68, 10–18. doi: 10.1016/j.jobaz.2015.01.005
- Seada, M. A., Ignell, R., Al Assiuty, A. N., and Anderson, P. (2018). Functional characterization of the gustatory sensilla of tarsi of the female polyphagous moth *Spodoptera littoralis*. *Front. Physiol.* 9:1606. doi: 10.3389/fphys.2018.01606
- Senapathi, D., Biesmeijer, J. C., Breeze, T. D., Kleijn, D., Potts, S. G., and Carvalheiro, L. G. (2015). Pollinator conservation — the difference between managing for pollination services and preserving pollinator diversity. *Curr. Opin. Insect Sci.* 12, 93–101. doi: 10.1016/j.cois.2015.11.002
- Shafir, S., and Yehonatan, L. (2014). Comparative evaluations of reward dimensions in honey bees: evidence from two-alternative forced choice proboscis-extension conditioning. *Anim. Cogn.* 17, 633–644. doi: 10.1007/s10071-013-0694-z
- Simcock, N. K., Gray, H., Bouchebti, S., and Wright, G. A. (2018). Appetitive olfactory learning and memory in the honeybee depend on sugar reward identity. *J. Insect Physiol.* 106, 71–77. doi: 10.1016/j.jinsphys.2017.08.009
- Simcock, N. K., Gray, H. E., and Wright, G. A. (2014). Single amino acids in sucrose rewards modulate feeding and associative learning in the honeybee. *J. Insect Physiol.* 69, 41–48. doi: 10.1016/j.jinsphys.2014.05.004
- Skiri, H. T., Strandén, M., Sandoz, J. C., Menzel, R., and Mustaparta, H. (2005). Associative learning of plant odorants activating the same or different receptor neurones in the moth *Heliothis virescens*. *J. Exp. Biol.* 208, 787–796. doi: 10.1242/jeb.01431
- Smedley, S. R., and Eisner, T. (1995). Sodium uptake by puddling in a moth. *Science* 270, 1816–1818. doi: 10.1126/science.270.5243.1816
- Smedley, S. R., and Eisner, T. (1996). Sodium: a male moth's gift to its offspring. *Proc. Natl. Acad. Sci. USA* 93, 809–813.
- Takada, T., Sato, R., and Kikuta, S. (2017). A mannitol/sorbitol receptor stimulates dietary intake in *Tribolium castaneum*. *PLoS One* 12:e0186420. doi: 10.1371/journal.pone.0186420
- Thompson, R. F., and Spencer, W. A. (1966). Habituation: a model phenomenon for the study of neuronal substrates of behavior. *Psychol. Rev.* 73, 16–43. doi: 10.1037/h0022681
- Tiedeken, E. J., Stout, J. C., Stevenson, P. C., and Wright, G. A. (2014). Bumblebees are not deterred by ecologically relevant concentrations of nectar toxins. *J. Exp. Biol.* 217, 1620–1625. doi: 10.1242/jeb.097543
- Unoki, S., Matsumoto, Y., and Mizunami, M. (2005). Participation of octopaminergic reward system and dopaminergic punishment system in insect olfactory learning revealed by pharmacological study. *Eur. J. Neurosci.* 22, 1409–1416. doi: 10.1111/j.1460-9568.2005.04318.x
- Unoki, S., Matsumoto, Y., and Mizunami, M. (2006). Roles of octopaminergic and dopaminergic neurons in mediating reward and punishment signals in insect visual learning. *Eur. J. Neurosci.* 24, 2031–2038. doi: 10.1111/j.1460-9568.2006.05099.x
- Vinauger, C., Lallement, H., and Lazzari, C. R. (2013). Learning and memory in *Rhodnius prolixus*: habituation and aversive operant conditioning of the proboscis extension response. *J. Exp. Biol.* 216, 892–900. doi: 10.1242/jeb.079491
- Waddington, K., and Gottlieb, N. (1990). Actual vs perceived profitability: a study of floral choice of honey bees. *J. Insect Behav.* 3, 429–441. doi: 10.1007/BF01052010
- Watanabe, M., and Kamikubo, M. (2005). Effects of saline intake on spermatophore and sperm ejaculation in the male swallowtail butterfly *Papilio xuthus* (Lepidoptera: Papilionidae). *Entomol. Sci.* 8, 161–166. doi: 10.1111/j.1479-8298.2005.00114.x
- Watt, W. B., Hoch, P. C., and Mills, S. G. (1974). Nectar resource use by *Colias* butterflies. *Oecologia* 14, 353–374. doi: 10.1007/BF00384578
- Wheeler, D. (1996). The role of nourishment in oogenesis. *Annu. Rev. Entomol.* 41, 407–431. doi: 10.1146/annurev.en.41.010196.002203
- Winkler, K., Wäckers, F. L., Stingli, A., and Van Lenteren, J. C. (2005). *Plutella xylostella* (diamondback moth) and its parasitoid *Diadegma semiclausum* show different gustatory and longevity responses to a range of nectar and honeydew sugars. *Entomol. Exp. Appl.* 115, 187–192. doi: 10.1111/j.1570-7458.2005.00254.x
- Wynne, J. W., Armon, J. K., Gerhardt, K. O., and Krause, G. F. (1991). Plant species identified as food sources for adult black cutworm (Lepidoptera: Noctuidae) in northwestern Missouri. *J. Kansas Entomol. Soc.* 64, 381–387.
- Zhang, Y. F., Van Loon, J. J., and Wang, C. Z. (2010). Tarsal taste neuron activity and proboscis extension reflex in response to sugars and amino acids in *Helicoverpa armigera* (Hubner). *J. Exp. Biol.* 213, 2889–2895. doi: 10.1242/jeb.042705
- Zhu, Y., Keaster, A. J., and Gerhardt, K. O. (1993). Field observations on attractiveness of selected blooming plants to noctuid moths and electroantennogram responses of black cutworm (Lepidoptera: Noctuidae) moths to flower volatiles. *Environ. Entomol.* 22, 162–166. doi: 10.1093/ee/22.1.162

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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SUPPLEMENTARY DATA

Supplementary Tables S1-12 report the details of statistics for Figure 1-6.

Supplementary Figures S1-6 report the same data as Figures 1-6, but with the animals spontaneously responding to water.

Supplementary Tables S13-24 report the details of statistics for Supplementary Figures S1-6.

		Sucrose concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Global tests	4 degrees of freedom χ^2				37.268	37.01	18.49	15.138	
	χ^2 p-value				< 0.001	< 0.001	< 0.001	0.004	
	Fisher's exact test p-value	1	0.149	0.126					0.163
Pairwise comparisons (adjusted p-values)		0%	0.1%	0.3%	1%	3%	10%	30%	0%
	5days/unfed 5days vs. 5days /unfed 3days				0.175	1	1	0.677	
	5days/unfed 5days vs. 5days /fed				< 0.001	0.124	0.184	0.614	
	5days/unfed 5days vs. 3days/unfed 3days				0.091	1	1	1	
	5days/unfed 5days vs. 3days /fed				< 0.001	< 0.001	0.054	0.677	
	5days /unfed 3days vs. 5days/fed				0.050	0.026	0.051	0.013	
	5days /unfed 3days vs. 3days/unfed 3days				0.880	1	1	0.677	
	5days /unfed 3days vs. 3days/fed				0.002	< 0.001	0.010	0.026	
	5days /fed vs. 3days/unfed 3days				0.166	0.011	0.174	0.614	
	5days /fed vs. 3days/fed				0.880	0.159	1	1	
3days/unfed 3days vs. 3days /fed				0.015	< 0.001	0.051	0.677		

Supplementary Table S1. Detailed analysis of data from Figure 1A.

		Sucrose concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Males vs. females (both 5days/unfed 5days)	1 degree of freedom χ^2			6.358	3.283	0.010			0.213
	χ^2 p-value			0.012	0.070	.919			0.644
	Fisher's exact test p-value	1	0.022				1	1	
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Global tests between females	4 degrees of freedom χ^2				10.398	23.744	4.065		4.970
	χ^2 p-value				0.034	< 0.001	0.397		0.290
	Fisher's exact test p-value	1	0.231	0.046				0.020	
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Pairwise comparisons between females (adjusted p-values)	5days/unfed 5days vs. 5days /unfed 3days			1	1	0.036		1	
	5days/unfed 5days vs. 5days /fed			0.699	0.056	< 0.001		0.272	
	5days/unfed 5days vs. 3days/unfed 3days			1	1	0.258		1	
	5days/unfed 5days vs. 3days /fed			1	1	0.001		1	
	5days /unfed 3days vs. 5days/fed			0.483	0.121	0.275		1	
	5days /unfed 3days vs. 3days/unfed 3days			1	1	0.820		1	
	5days /unfed 3days vs. 3days/fed			0.981	1	0.755		1	
	5days /fed vs. 3days/unfed 3days			0.134	0.020	0.069		1	
	5days /fed vs. 3days/fed			1	0.331	0.820		0.272	
3days/unfed 3days vs. 3days /fed			0.463	1	0.275		1		

Supplementary Table S2. Detailed analysis of data from Figure 1B.

		Sucrose concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Global tests	3 degrees of freedom χ^2		5.672	4.314	15.581	37.939			4.686
	χ^2 p-value		0.129	0.229	0.001	< 0.001			0.196
	Fisher's exact test p-value	1					< 0.001	0.201	
Pairwise comparisons (adjusted p-values)	Unfed control vs. unfed random				0.37	0.71	1		
	Unfed control vs. fed control				0.282	< 0.001	0.094		
	Unfed control vs. fed random				0.282	< 0.001	0.078		
	Unfed random vs. fed control				0.012	< 0.001	0.030		
	Unfed random vs. fed random				0.006	< 0.001	0.010		
	Fed control vs. fed random				0.707	0.710	1		

Supplementary Table S3. Detailed analysis of data from Figure 2A.

		Sucrose concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Global tests	3 degrees of freedom χ^2					1.345	2.666	1.610	
	χ^2 p-value					0.718	0.446	0.657	
	Fisher's exact test p-value	1	0.064	0.068	0.417				.733

Supplementary Table S4. Detailed analysis of data from Figure 2B.

		Sucrose concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Antennal stimulated vs. leg stimulated	1 degree of freedom χ^2		8.011	10.634	20.267	23.945			12.267
	χ^2 p-value		0.005	0.001	< 0.001	< 0.001			< 0.001
	Fisher's exact test p-value	1					0.0002	0.001	
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Antennal stimulation in both groups	1 degree of freedom χ^2		1.609	1.267	0.003				0.069
	χ^2 p-value		0.205	0.260	0.955				0.792
	Fisher's exact test p-value	1				0.709	0.474	0.474	

Supplementary Table S5. Detailed analysis of data from Figure 3.

		Sucrose concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Comparison before proboscis stimulation	1 degree of freedom χ^2				2.375	0.472	0.075	0.472	
	χ^2 p-value				0.123	0.492	0.785	0.492	
	Fisher's exact test p-value	1	0.421	0.507					0.349
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Comparison before and after water stimulation	McNemar's test	0	0	0	0	0	0	0	0
	McNemar's test p-value	1	1	1	1	1	1	1	1
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Comparison before and after quinine stimulation	McNemar's test	0	0.333	0.667	3.571	3.267	9.783	8.000	1.000
	McNemar's test p-value	1	0.564	0.414	0.059	0.071	0.002	0.005	0.317

Supplementary Table S6. Detailed analysis of data from Figure 4A.

		Sucrose concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Global test	1 degree of freedom χ^2				21.582	63.828	83.977	68.646	
	χ^2 p-value				< 0.001	< 0.001	< 0.001	< 0.001	
	Fisher's exact test p-value	1.000	0.011	0.024					0.052

Supplementary Table S7. Detailed analysis of data from Figure 4B.

		Sugar concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Global tests	3 degrees of freedom χ^2			23.811	24.825	8.874	12.002		
	χ^2 p-value			< 0.001	< 0.001	0.012	0.002		
	Fisher's exact test p-value	1.000	0.002					0.003	0.430
Pairwise comparisons (adjusted p-values)		0%	0.1%	0.3%	1%	3%	10%	30%	0%
	Sucrose vs. Fructose		1.000	0.015	0.046	0.068	0.003	0.041	
	Sucrose vs. Sucrose/Fructose		0.030	0.015	0.003	0.463	0.217	1.000	
	Fructose vs. Sucrose/Fructose		0.016	< 0.001	< 0.001	0.015	0.217	0.014	

Supplementary Table S8. Detailed analysis of data from Figure 5A.

		Sugar concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Global tests	3 degrees of freedom χ^2		8.025	8.184	19.17	41.776	50.648		8.673
	χ^2 p-value		0.018	0.017	< 0.001	< 0.001	< 0.001		0.013
	Fisher's exact test p-value	1							< 0.001
Pairwise comparisons (adjusted p-values)		0%	0.1%	0.3%	1%	3%	10%	30%	0%
	Sucrose vs. Glucose		0.017	0.020	< 0.001	< 0.001	< 0.001	< 0.001	0.022
	Sucrose vs. Sucrose/Glucose		0.956	0.974	0.813	0.805	0.686	1.000	0.737
	Glucose vs. Sucrose/Glucose		0.017	0.020	< 0.001	< 0.001	< 0.001	< 0.001	0.012

Supplementary Table S9. Detailed analysis of data from Figure 5B.

		Sugar concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Global tests	3 degrees of freedom χ^2					17.107	16.122	6.52	0.259
	χ^2 p-value					< 0.001	< 0.001	0.038	.879
	Fisher's exact test p-value	1	0.051	.588	.680				
Pairwise comparisons (adjusted p-values)		0%	0.1%	0.3%	1%	3%	10%	30%	0%
	Fructose vs. Glucose					< 0.001	0.001	0.335	
	Glucose vs. Fructose/Glucose					0.031	0.001	0.055	
	Fructose vs. Fructose/Glucose					0.049	0.980	0.490	

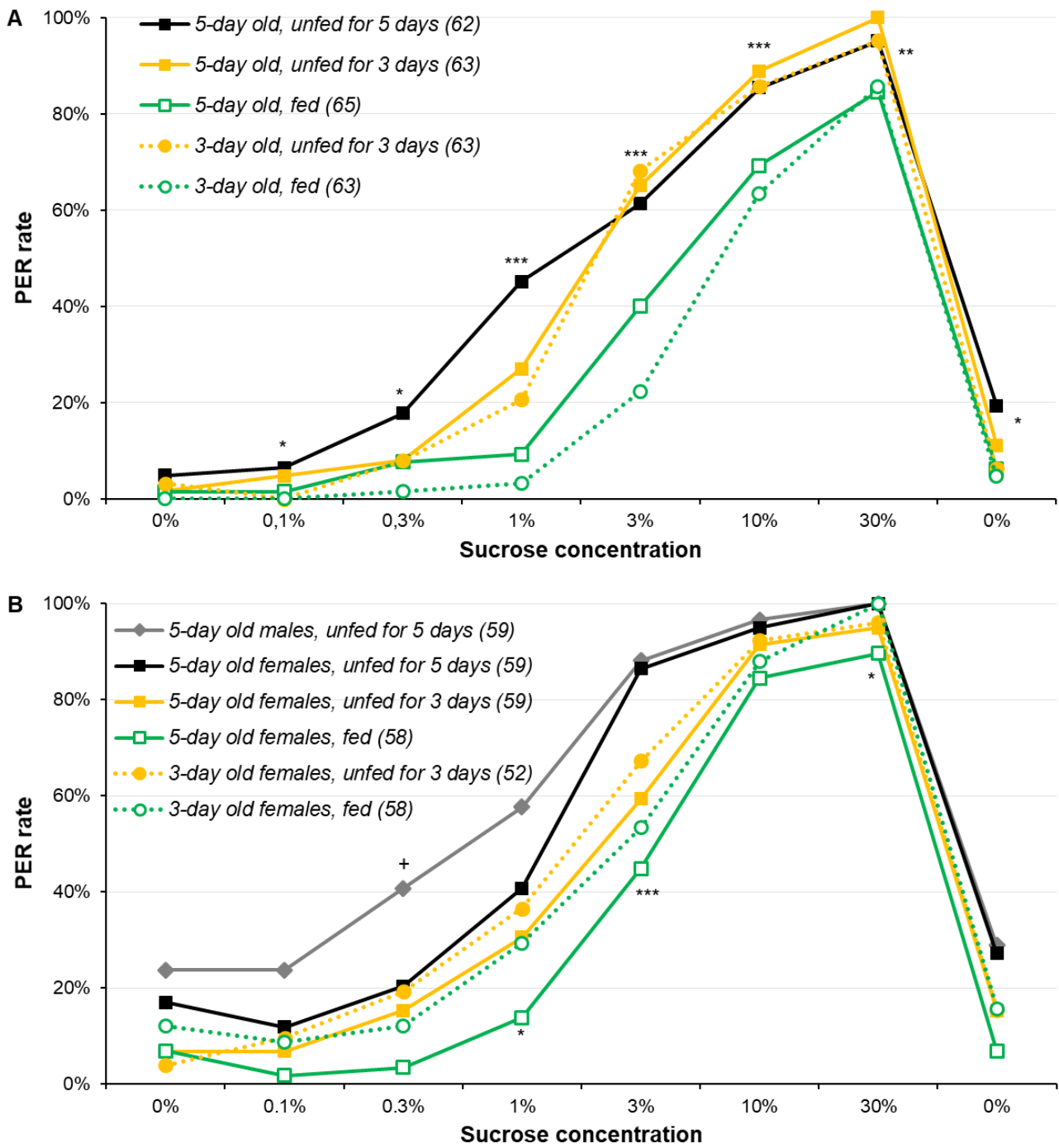
Supplementary Table S10. Detailed analysis of data from Figure 5C.

		Sugar concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Global tests	4 degrees of freedom χ^2			13.762	21.806	54.211	31.305		3.224
	χ^2 p-value			0.003	< 0.001	< 0.001	< 0.001		0.358
	Fisher's exact test p-value	1	0.578						< 0.001
Pairwise comparisons (adjusted p-values)		0%	0.1%	0.3%	1%	3%	10%	30%	0%
	Sucrose vs. Glucose			0.072	< 0.001	< 0.001	< 0.001	< 0.001	
	Sucrose vs. Fructose			0.009	0.040	0.016	0.850	0.726	
	Sucrose vs. Mixture			0.598	0.307	0.151	1.000	0.726	
	Glucose vs. Fructose			0.600	0.068	< 0.001	< 0.001	0.045	
	Glucose vs. Mixture			0.490	0.002	< 0.001	< 0.001	0.045	
	Fructose vs. Mixture			0.235	0.221	0.291	1.000	1.000	

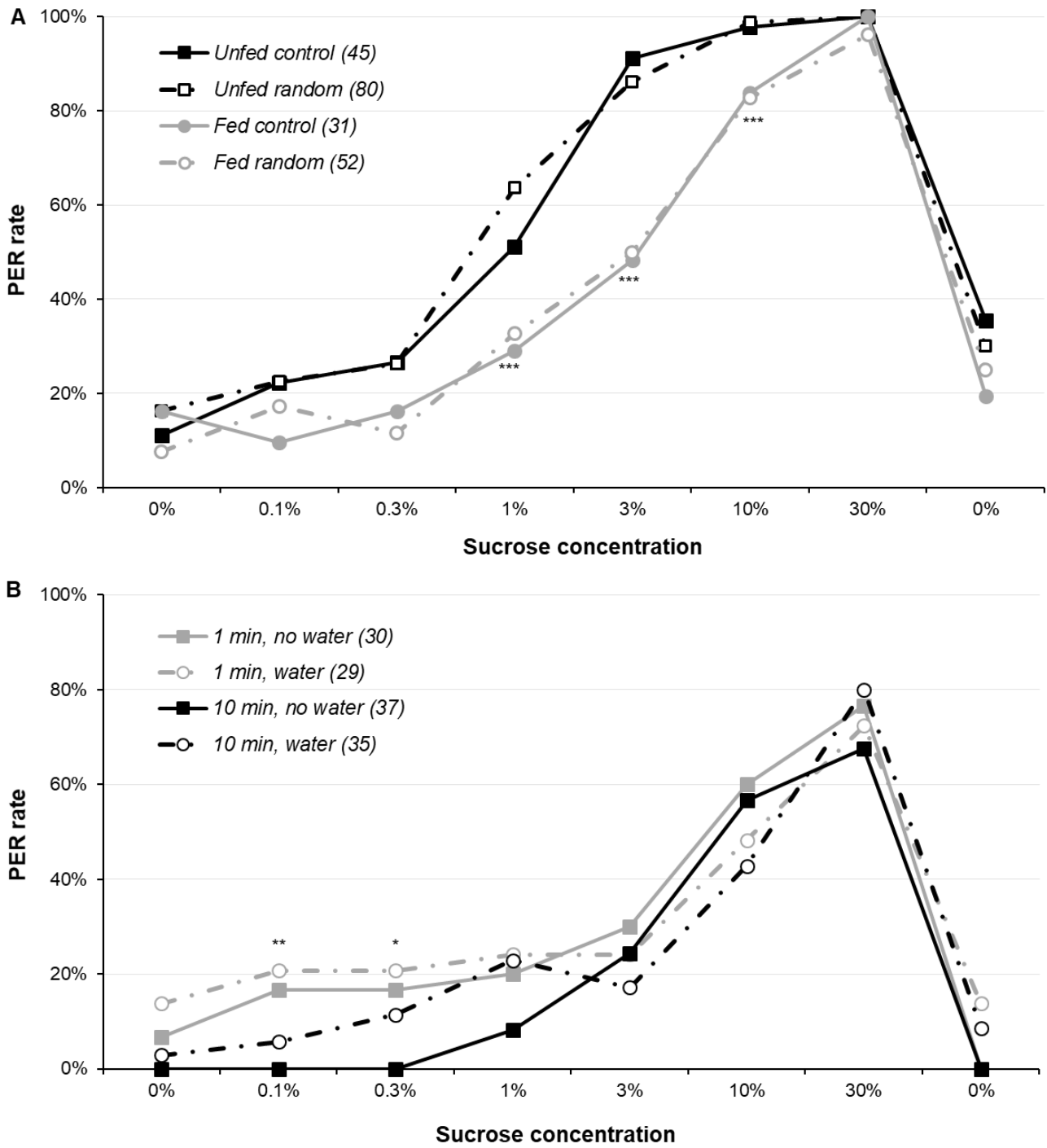
Supplementary Table S11. Detailed analysis of data from Figure 5D.

		Sugar concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Males vs. females (sucrose)	1 degree of freedom χ^2		0.033	0.091	0.063	0.034			0.012
	χ^2 p-value		.855	.763	.801	.853			.913
	Fisher's exact test p-value						0.060	0.060	
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Global tests between females	4 degrees of freedom χ^2			8.344	35.956	70.795	68.228		8.344
	χ^2 p-value			0.080	< 0.001	< 0.001	< 0.001		0.080
	Fisher's exact test p-value		0.524					0.004	
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Pairwise comparisons between females (adjusted p-values)	Sucrose vs. Fructose				0.818	1.000	1.000	0.420	
	Sucrose vs. Sucrose/Fructose				0.818	1.000	1.000	0.113	
	Sucrose vs. Sucrose/Fructose/Glucose				0.301	1.000	1.000	0.814	
	Sucrose vs. Glucose				< 0.001	< 0.001	< 0.001	1.000	
	Fructose vs. Sucrose/Fructose				0.197	1.000	1.000	1.000	
	Fructose vs. Sucrose/Fructose/Glucose				0.029	1.000	1.000	1.000	
	Fructose vs. Glucose				0.003	< 0.001	< 0.001	0.355	
	Sucrose/Fructose vs. Sucrose/Fructose/Glucose				0.818	1.000	1.000	0.951	
	Sucrose/Fructose vs. Glucose				< 0.001	< 0.001	< 0.001	0.064	
	Sucrose/Fructose/Glucose vs. Glucose				< 0.001	< 0.001	< 0.001	0.814	

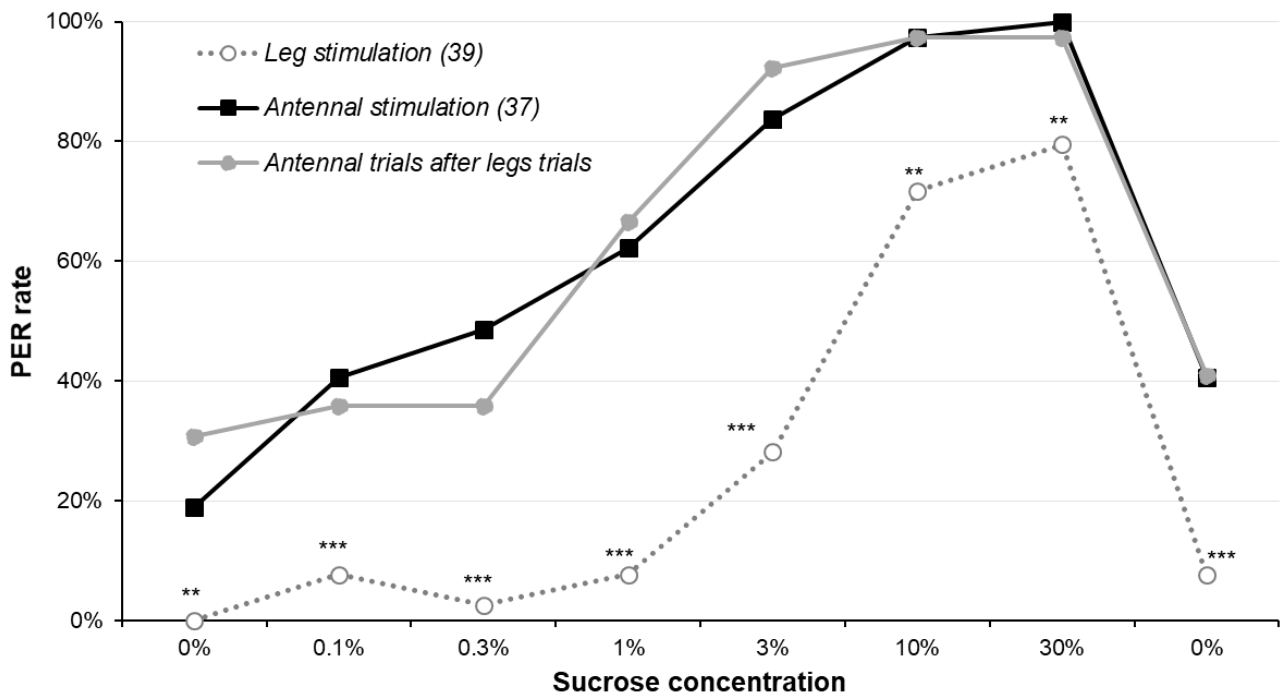
Supplementary Table S12. Detailed analysis of data from Figure 6.



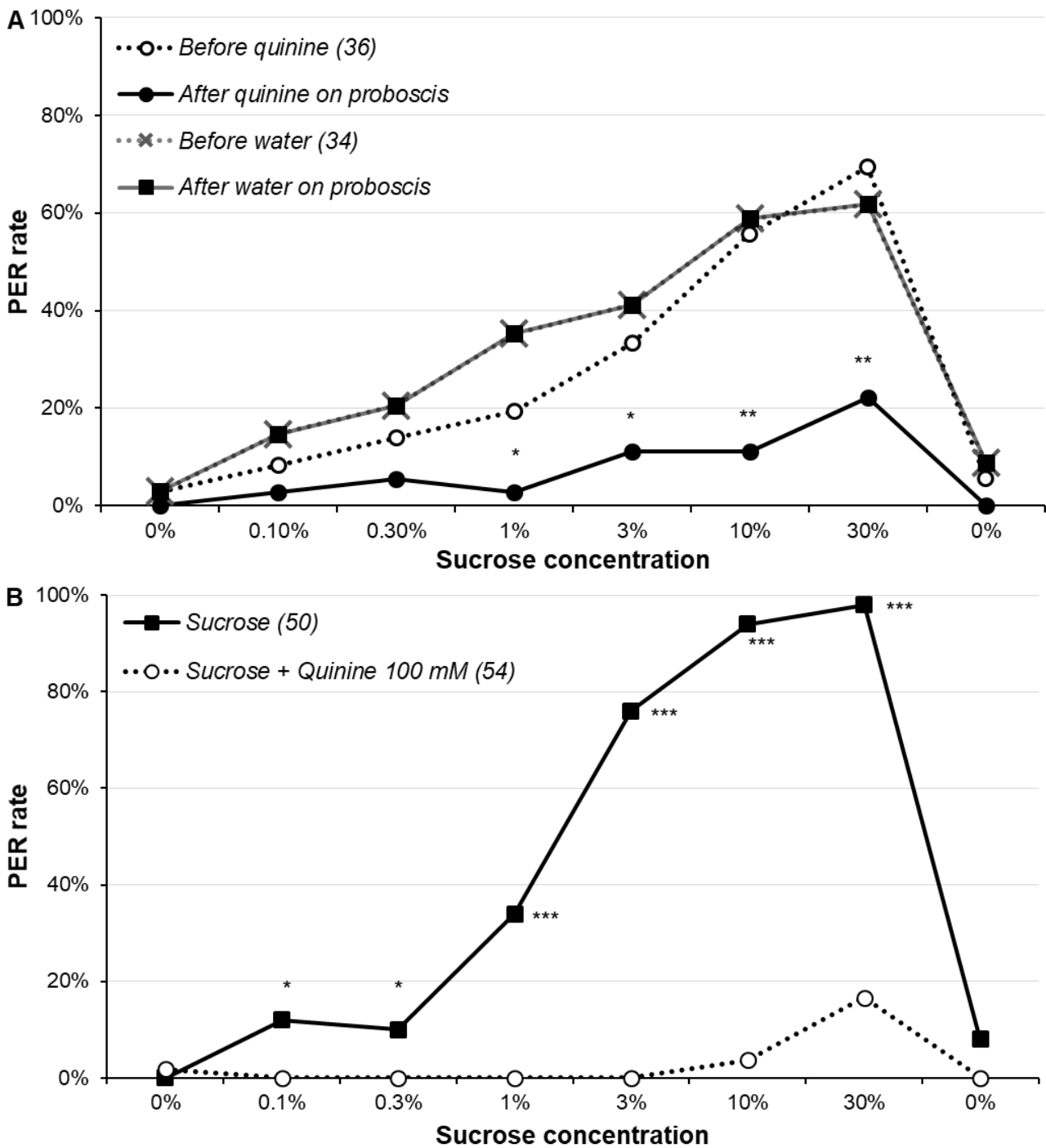
Supplementary Figure S1: data of Figure 1 including animals spontaneously responding to water. Details of the analyses are reported in Supplementary Tables S13 and S14. All other details as in Figure 1.



Supplementary Figure S2: data of Figure 2 including animals spontaneously responding to water. Details of the analyses are reported in Supplementary Tables S15 and S16. All other details as in Figure 2.

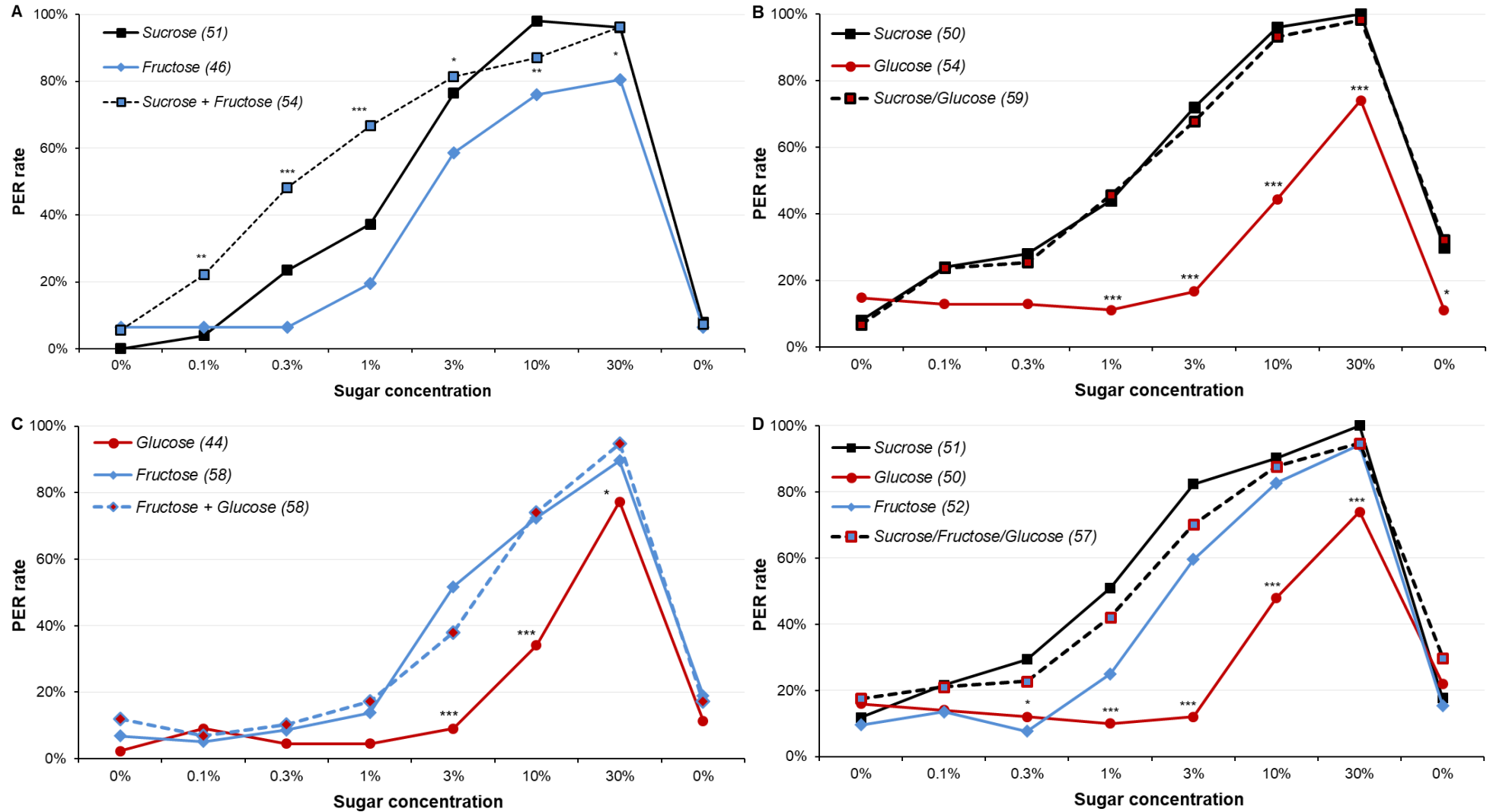


Supplementary Figure S3: data of Figure 3 including animals spontaneously responding to water. Details of the analyses are reported in Supplementary Table S17. All other details as in Figure 3.

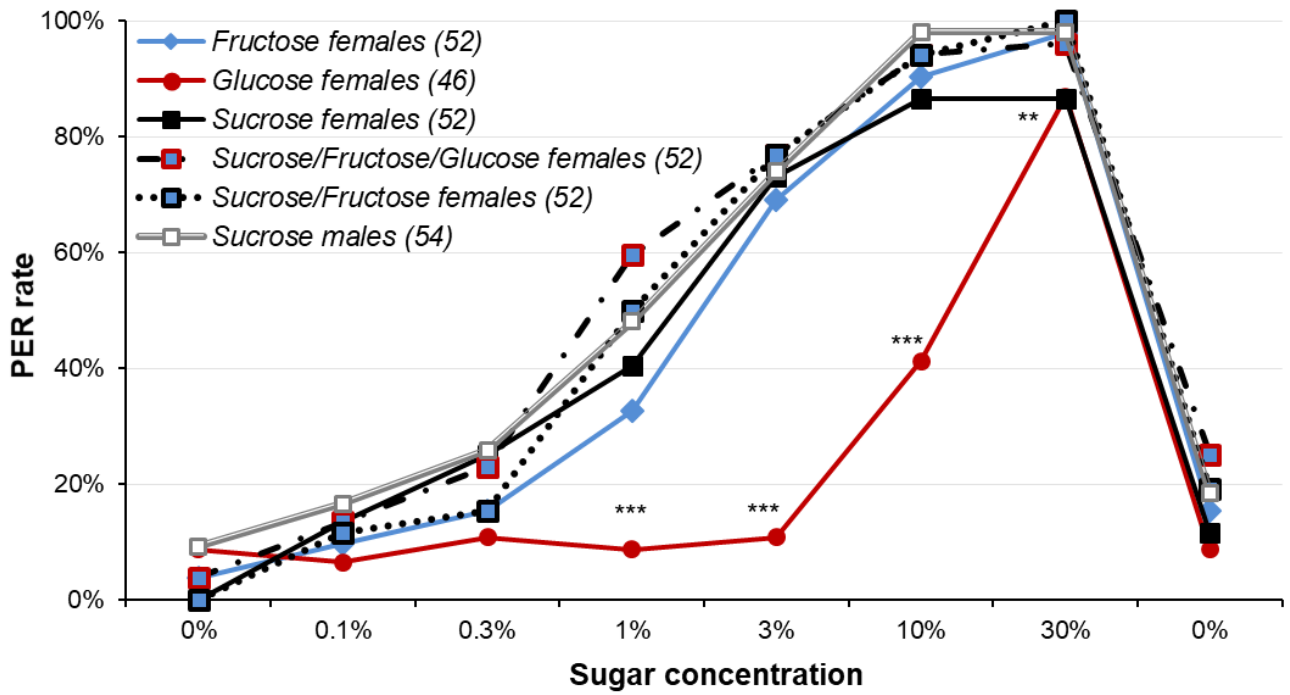


Supplementary Figure S4: data of Figure 4 including animals spontaneously responding to water. In part A, stars denote a significant difference between the PER rate before and after touching the proboscis with quinine, i.e. a significant rate of proboscis retraction (McNemar's test; *: $p < 0.050$, **: $p < 0.010$), and details of the analyses are reported in Supplementary Tables S18. In part B, stars denote a significant difference between the PER rate of the two groups (χ^2 ; *: $p < 0.050$, ***: $p < 0.001$), and details of the analyses are reported in Supplementary Tables S19. All other details as in Figure 4.

Sugar responsiveness in moths



Supplementary Figure S5: data of Figure 5 including animals spontaneously responding to water. Details of the analyses are reported in Supplementary Tables S20 to S23. All other details as in Figure 5.



Supplementary Figure S6: data of Figure 6 including animals spontaneously responding to water. Details of the analyses are reported in Supplementary Tables S24. All other details as in Figure 6.

		Sucrose concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Global tests	4 degrees of freedom χ^2			10.734	40.835	39.155	19.467		10.419
	χ^2 p-value			0.030	< 0.001	< 0.001	< 0.001		0.034
	Fisher's exact test p-value	0.352	0.042						0.002
Pairwise comparisons (adjusted p-values)	5days/unfed 5days vs. 5days /unfed 3days	1	1	0.165	1	1	0.714	1	1
	5days/unfed 5days vs. 5days /fed	1	1	< 0.001	0.082	0.156	0.611	0.291	1
	5days/unfed 5days vs. 3days/unfed 3days	0.576	1	0.030	1	1	1	0.291	0.576
	5days/unfed 5days vs. 3days /fed	0.576	0.021	< 0.001	< 0.001	0.039	0.714	0.139	0.576
	5days /unfed 3days vs. 5days/fed	1	1	0.056	0.027	0.045	0.013	1	1
	5days /unfed 3days vs. 3days/unfed 3days	1	1	0.548	1	1	0.732	1	1
	5days /unfed 3days vs. 3days/fed	1	1	0.002	< 0.001	0.008	0.026	1	1
	5days /fed vs. 3days/unfed 3days	1	1	0.254	0.009	0.156	0.611	1	1
	5days /fed vs. 3days/fed	1	1	0.548	0.120	1	1	1	1
	3days/unfed 3days vs. 3days /fed	1	1	0.030	< 0.001	0.037	0.714	1	1

Supplementary Table S13. Detailed analysis of data from Supplementary Figure S1A.

		Sucrose concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Males vs. females (both 5days/unfed 5days)	1 degree of freedom χ^2	0.837	2.839	5.756	3.391	0.076			0.042
	χ^2 p-value	.360	0.092	0.016	0.066	.782			.838
	Fisher's exact test p-value						1	1	
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Global tests between females	4 degrees of freedom χ^2	7.191		8.778	11.521	24.874	4.349		9.013
	χ^2 p-value	.126		0.067	0.021	< 0.001	.361		0.061
	Fisher's exact test p-value		.256					0.012	
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Pairwise comparisons between females (adjusted p-values)	5days/unfed 5days vs. 5days /unfed 3days				1	0.007		1	
	5days/unfed 5days vs. 5days /fed				0.011	< 0.001		0.129	
	5days/unfed 5days vs. 3days/unfed 3days				1	0.112		1	
	5days/unfed 5days vs. 3days /fed				1	< 0.001		1	
	5days /unfed 3days vs. 5days/fed				0.237	0.583		1	
	5days /unfed 3days vs. 3days/unfed 3days				1	1		1	
	5days /unfed 3days vs. 3days/fed				1	1		1	
	5days /fed vs. 3days/unfed 3days				0.051	0.112		1	
	5days /fed vs. 3days/fed				0.295	1		0.246	
3days/unfed 3days vs. 3days /fed				1	0.583		1		

Supplementary Table S14. Detailed analysis of data from Supplementary Figure S1B

		Sucrose concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Global tests	3 degrees of freedom χ^2	2.456	2.755	5.417	17.47	37.729			2.779
	χ^2 p-value	0.483	0.431	0.144	< 0.001	< 0.001			0.427
	Fisher's exact test p-value						< 0.001	0.129	
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Pairwise comparisons (adjusted p-values)	Unfed control vs. unfed random				0.335	0.844	1		
	Unfed control vs. fed control				0.222	< 0.001	0.115		
	Unfed control vs. fed random				0.222	< 0.001	0.073		
	Unfed random vs. fed control				0.005	< 0.001	0.032		
	Unfed random vs. fed random				0.003	< 0.001	0.006		
	Fed control vs. fed random				0.728	0.887	1		

Supplementary Table S15. Detailed analysis of data from Supplementary Figure S2A.

		Sucrose concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Global tests	3 degrees of freedom χ^2				3.773	1.503	2.437	1.601	
	χ^2 p-value				0.287	0.682	0.487	0.659	
	Fisher's exact test p-value	0.055	0.008	0.016					0.209
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Pairwise comparisons (adjusted p-values)	1 min, no water vs. 1 min, water		0.748	1.000					
	1 min, no water vs. 10 min, no water		0.074	0.074					
	1 min, no water vs. 10 min, water		0.698	1.000					
	1 min, water vs. 10 min, no water		0.031	0.031					
	1 min, water vs. 10 min, water		0.506	1.000					
	10 min, no water vs. 10 min, water		0.698	0.204					

Supplementary Table S16. Detailed analysis of data from Supplementary Figure S2B.

		Sucrose concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Antennal stimulated vs. leg stimulated	1 degree of freedom χ^2		11.335	21.506	25.029	23.720	9.287		11.335
	χ^2 p-value		< 0.001	< 0.001	< 0.001	< 0.001	0.002		< 0.001
	Fisher's exact test p-value	0.005							0.005
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Antennal stimulation in both groups	1 degree of freedom χ^2	1.422	0.173	1.266	0.168				0.002
	χ^2 p-value	0.233	0.677	0.260	0.681				0.966
	Fisher's exact test p-value					.304	1	1	

Supplementary Table S17. Detailed analysis of data from Supplementary Figure S3.

		Sucrose concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Comparison before proboscis stimulation	1 degree of freedom χ^2			0.553	2.221	0.461	0.076	0.458	
	χ^2 p-value			0.457	0.136	0.497	0.782	0.499	
	Fisher's exact test p-value	1	0.4717						0.669
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Comparison before and after water stimulation	McNemar's test	0	0	0	0	0	0	0	0
	McNemar's test p-value	1	1	1	1	1	1	1	1
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Comparison before and after quinine stimulation	McNemar's test	1.000	1.000	1.286	4.500	4.000	10.667	8.758	2.000
	McNemar's test p-value	0.317	0.317	0.257	0.034	0.046	0.0011	0.003	0.157

Supplementary Table S18. Detailed analysis of data from Supplementary Figure S4A.

		Sucrose concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Global test	1 degree of freedom χ^2				21.948	64.669	84.953	69.622	
	χ^2 p-value				< 0.001	< 0.001	< 0.001	< 0.001	
	Fisher's exact test p-value	1	0.010	0.023					0.050

Supplementary Table S19. Detailed analysis of data from Supplementary Figure S4B.

		Sugar concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Global tests	3 degrees of freedom χ^2		10.278	22.271	23.397	7.031	10.607		
	χ^2 p-value		0.006	< 0.001	< 0.001	0.030	0.005		
	Fisher's exact test p-value	0.189						0.012	1.000
Pairwise comparisons (adjusted p-values)		0%	0.1%	0.3%	1%	3%	10%	30%	0%
	Sucrose vs. Fructose		0.666	0.021	0.055	0.122	0.004	0.064	
	Sucrose vs. Sucrose/Fructose		0.025	0.017	0.005	0.528	0.122	1.000	
	Fructose vs. Sucrose/Fructose		0.094	< 0.001	< 0.001	0.037	0.195	0.064	

Supplementary Table S20. Detailed analysis of data from Supplementary Figure S5A.

		Sugar concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Global tests	3 degrees of freedom χ^2	2.325	2.654	4.010	18.537	40.929	52.69		7.935
	χ^2 p-value	.312	.265	.135	< 0.001	< 0.001	< 0.001		0.019
	Fisher's exact test p-value							< 0.001	
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Pairwise comparisons (adjusted p-values)	Sucrose vs. Glucose				< 0.001	< 0.001	< 0.001	< 0.001	0.033
	Sucrose vs. Sucrose/Glucose				0.854	.634	.685	1	0.805
	Glucose vs. Sucrose/Glucose				< 0.001	< 0.001	< 0.001	< 0.001	0.021

Supplementary Table S21. Detailed analysis of data from Supplementary Figure S5B.

		Sugar concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Global tests	3 degrees of freedom χ^2				3.826	20.333	20.935	7.571	1.128
	χ^2 p-value				.148	< 0.001	< 0.001	0.023	.569
	Fisher's exact test p-value	.178	.684	.668					
Pairwise comparisons (adjusted p-values)		0%	0.1%	0.3%	1%	3%	10%	30%	0%
	Fructose vs. Glucose					< 0.001	< 0.001	0.210	
	Glucose vs. Fructose/Glucose					0.002	< 0.001	0.043	
	Fructose vs. Fructose/Glucose					0.135	0.834	0.490	

Supplementary Table S22. Detailed analysis of data from Supplementary Figure S5C.

		Sugar concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Global tests	4 degrees of freedom χ^2	1.817	2.082	10.311	23.251	58.748	34.012		3.958
	χ^2 p-value	0.611	0.556	0.016	< 0.001	< 0.001	< 0.001		0.266
	Fisher's exact test p-value								< 0.001
Pairwise comparisons (adjusted p-values)		0%	0.1%	0.3%	1%	3%	10%	30%	0%
	Sucrose vs. Glucose			0.194	< 0.001	< 0.001	< 0.001	< 0.001	
	Sucrose vs. Fructose			0.031	0.030	0.033	0.800	0.728	
	Sucrose vs. Mixture			1.000	0.360	0.279	0.918	0.728	
	Glucose vs. Fructose			1.000	0.141	< 0.001	< 0.001	0.026	
	Glucose vs. Mixture			0.616	0.001	< 0.001	< 0.001	0.026	
	Fructose vs. Mixture			0.180	0.141	0.279	0.918	1.000	

Supplementary Table S23. Detailed analysis of data from Supplementary Figure S5D.

		Sugar concentration							
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Males vs. females (sucrose)	1 degree of freedom χ^2		0.212	0.012	0.647	0.014			1.007
	χ^2 p-value		.645	.913	.421	.907			.316
	Fisher's exact test p-value	0.057					0.030	0.030	
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Global tests between females	4 degrees of freedom χ^2		1.7	4.676	30.61	65.104	66.16		6.101
	χ^2 p-value		.791	.322	< 0.001	< 0.001	< 0.001		0.192
	Fisher's exact test p-value	0.060							0.005
		0%	0.1%	0.3%	1%	3%	10%	30%	0%
Pairwise comparisons between females (adjusted p-values)	Sucrose vs. Fructose				0.974	1.000	1.000	0.419	
	Sucrose vs. Sucrose/Fructose				0.974	1.000	1.000	0.113	
	Sucrose vs. Sucrose/Fructose/Glucose				0.249	1.000	1.000	0.853	
	Sucrose vs. Glucose				0.003	< 0.001	< 0.001	1.000	
	Fructose vs. Sucrose/Fructose				0.292	1.000	1.000	1.000	
	Fructose vs. Sucrose/Fructose/Glucose				0.035	1.000	1.000	1.000	
	Fructose vs. Glucose				0.027	< 0.001	< 0.001	0.390	
	Sucrose/Fructose vs. Sucrose/Fructose/Glucose				0.974	1.000	1.000	1.000	
	Sucrose/Fructose vs. Glucose				< 0.001	< 0.001	< 0.001	0.089	
	Sucrose/Fructose/Glucose vs. Glucose				0.000	0.000	0.000	0.853	

Supplementary Table S24. Detailed analysis of data from Supplementary Figure S6.