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Time-consistent estimation of LAI by assimilation in GreenLab plant growth model

Thomas Corpetti^{1,2}, Xing Gong^{1,2}, MengZhen Kang¹, BaoGang Hu¹, Laurence
Hubert-Moy²

Abstract

This paper is concerned with the recovery of Leaf Area Index (LAI) time series in intense agriculture areas from moderate resolution remote sensing data (MODIS or SENTINEL). Although their resolution limits an analysis at a parcel level, their high temporal rate enables to monitor land use/land cover through the temporal evolution of key biophysical parameters as LAI. However in practice, frame-by-frame estimation is unsatisfactory since the quality of each single data is subjected to undesirable effects due to atmosphere disturbance, sun geometry, viewing geometry, etc. These effects lead to a lack of temporal consistency of resulting time series. The reconstruction of such time series is delicate using conventional interpolation methods since underlying physical processes are not taken into account. In this paper, we tackle this issue by exploiting the prior information of a plant growth model, namely GreenLab, using stochastic data assimilation techniques. Our experiments on challenging situations, such as few data and fragmented landscapes, demonstrate the approach is robust on various challenging situations and enables to extract additional information about observed fields. Experiments are performed on MODIS data.

Index Terms

Leaf Area Index, GreenLab model, data assimilation, noisy/missing data

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All authors have discussed together about methodological developments. Thomas Corpetti and Gong Xing implement code and make tests. MengZhen Kang and BaoGang Hu gave their skills in GreenLab model and Laurence Hubert-Moy gave her skills about applications on the study site

I. INTRODUCTION

A. Land surface monitoring

Land surface on earth is mainly covered by green plants including forests, grasslands or agricultural fields (Townshend *et al.* (1991)). These vegetation areas are of great importance for global ecological systems and food supply for human beings. Therefore in the past several decades, many satellites have been launched to observe and monitor these areas (Richards (2013); Trotter (1991)). A number of important tools has been developed to extract some crucial information on crops and plants from satellite images. This is indeed a prerequisite in many agricultural applications such as monitoring the plant physiology and/or ecology at different scales (Gao (1996); Huete (1988); Liang (2005)). One of the most popular vegetation index is the **NDVI (Normalized Difference Vegetation Index, Rouse Jr *et al.* (1974))**, based on the spectral reflectance property of green leaves (chlorophyll) at different **spectral bands** and computed using red and near-infrared wavebands. While NDVI is a sensitive indicator of canopy structure and chemical content (plant biomass, leaf area index, chlorophyll content in sparse canopies and foliar nitrogen content), it is still a limited index for total canopy biomass in high-density vegetation areas (Gamon *et al.* (1995)). Some other more sophisticated biophysical indexes have been proposed, such as :

- **fCover (vegetation cover fraction)**: which represents the fraction of ground covered by green vegetation;
- **fAPAR (fraction of photo-synthetically active radiation absorbed by the canopy)**: which represents the fraction of the solar radiation absorbed by live leaves;
- **LAI (Leaf Area Index)** : which represents the total quantity of green leaf area per unit ground surface area.

In practice, fAPAR is sensitive to sun lighting while fCover and LAI are independent from the illumination. In this study we rely on LAI since **the total quantity of leaves is captured (and not only the ground surface); moreover, this parameter is largely used in practice (Baret *et al.* (2007)), for example to estimate productivity for certain crops. Readers can find in Carlson and Ripley (1997); Gamon *et al.* (1995) more details about these biophysical variables.** The estimation of these biophysical variables from remote sensing is a delicate task. Several models, based on light interception by plant canopies and vegetation reflectance in terms of biophysical characteristics, have been designed and inverted to retrieve biophysical information

52 from optical satellite images. Among them, SAIL (Scattering by Arbitrary Inclined Leaves)
53 canopy bidirectional reflectance model Verhoef (1984) and PROSECT (based on leaf optical
54 properties) are widely used Schaepman-Strub *et al.* (2006). Even though the results were globally
55 satisfactory, strong improvements have been obtained when one mixes these two models. The
56 resulting one, PROSAIL, has allowed to describe both the spectral and directional variation of
57 canopy reflectance based on single static images (Jacquemoud *et al.* (2009); Lecerf *et al.* (2008);
58 González-Sanpedro *et al.* (2008)).

59 All these methods have been designed for single images and only few of them exploit the time
60 series observation to derive the plant growth evolution information : in Roerink *et al.* (2000),
61 authors have developed a Fourier analysis method to reconstruct cloud free NDVI composites
62 while in Jonsson and Eklundh (2002), the authors have proposed a technique based on nonlinear
63 least squares. Despite the fact that they provide interesting information, no physical knowledge
64 about plant or vegetation growth is taken into consideration.

65 In this study we suggest to exploit the short revisit period of satellites to improve the quality
66 of estimated biophysical variables by constraining the solution to be consistent with physical
67 dynamic priors. This *data assimilation* procedure, though presented here for LAI estimation, can
68 be applied to any biophysical variable provided that a physical model of its temporal evolution
69 exists.

70 *B. Plant growth modelling*

71 The term *plant modelling* has various meanings depending on scientific communities. One
72 can simulate realistic forms and patterns of plants from an architectural and geometrical point of
73 view (Fisher (1992)), as in earlier developments (see for example Cohen (1967); Honda (1971);
74 Lindenmayer (1968)). This *structural* simulation of plants refers to various tools either related
75 to L-systems (Przemyslaw *et al.* (1988)), fractals (Smith (1984)), particle systems (Reeves and
76 Blau (1985)) or ramified matrix (Viennot *et al.* (1989)) for example.

77 When one is concerned with more physical studies related to plants and crops (agronomy,
78 forestry, biophysics, ...), specific plant growth models have been developed for agronomic pur-
79 poses, harvest prediction or optimum crop management (see for an Brisson *et al.* (2003)) and
80 rely on *physical* properties: they describe the flux of external and internal resources through the
81 element “plant” in order to expect its yield. These *process-based models* (PBM) consider biomass

82 production through the photosynthetic process and the global biomass partitioning among organs
83 (Heuvelink (1999) or Dayan *et al.* (2004)).

84 Between process-based models and architectural models, *functional-structural plant models*
85 (*FSPM*) have emerged since the end of the 1990's (see De Reffye *et al.* (1997); Kurth and
86 Sloboda (1997); Perttunen *et al.* (1996); Vos *et al.* (2007); Yan *et al.* (2004)). They perform
87 efficient and realistic dynamic simulations of plant morphogenesis (see for example Jallas *et al.*
88 (2000); Perttunen *et al.* (1996)) by taking into account both physical and structural properties. The
89 GreenLab plant growth model (Yan *et al.* (2004)) belongs to this family and has been designed to
90 provide dynamic representations of the morphogenesis and architecture of plants. Because of *i*)
91 its efficiency in modelling realistic physical behaviours ; *ii*) its efficiency in designing consistent
92 plant architectures and *iii*) its permanent interaction between structural and functional part, this
93 model is really interesting for plant studies. We therefore rely on it in this paper.

94 *C. Contributions and novelty of the paper*

95 Though generating LAI by assimilating remote sensing data in crop model has already been
96 studied (see for example Launay and Guerif (2005); Zhao *et al.* (2013); Dente *et al.* (2008);
97 Curnel *et al.* (2011)), only a specific culture (sugar, maize, wheat) has been explored in most of
98 these studies. In addition, associated models do not contain a functional part as GreenLab do,
99 and this latter has never been used yet for data assimilation. This is the scope of this paper.

100 The overall article is organised as follows: the next section introduces the principles and
101 required details of GreenLab plant growth model. As this model is designed at a plant level,
102 some manipulations and simplifications are required for its adaptation to medium resolution
103 images. These modifications, detailed in section II-B, enable to finally exploit an easy-to-use
104 version for data assimilation. The assimilation part is presented in section II-C and finally, section
105 III show some experiments on synthetic and real data.

106 II. MATERIAL AND METHODS

107 *A. Introduction to the Greenlab model*

108 GreenLab model aims at simulating plant growth evolution. For each type of plant, this model
109 is calibrated with ad-hoc observations of growth made under controlled environment and where
110 extensive and even destructive measurements of individual plants are made regularly Guo *et al.*
111 (2006); Kang *et al.* (2008). This enables to derive a deterministic description of a plant evolution

112 (in more recent research, some stochasticity has been introduced to bring about plant growth
 113 variations Lopez *et al.* (2008); Wang *et al.* (2011)). The model works sequentially and in each
 114 cycle, two steps are performed:

- 115 1) **Biomass production:** this step generates a biomass pool from some environmental pa-
 116 rameters (sun lighting, photosynthesis efficiency, plant density, ...);
- 117 2) **Biomass repartition:** the generated biomass is shared between some organs (leafs, fruits,
 118 ...).

119 The idea behind this model is that all organs of a plant share a common biomass pool and
 120 are therefore in competition each others. In addition to these two primal operations, a **natural**
 121 **withering** of each plant is modelled. This is schematised in figure 1 and main steps are detailed
 122 below.

123 1) *Biomass production:* The biomass production comes from a large number of factors
 124 as photosynthesis efficiency of plant species, total leaf area, environmental context, etc. In
 125 GreenLab, the production is done over growth cycle through the relation:

$$Q_{t_k} = E \cdot S_p \cdot \frac{1}{r} \left(1 - \exp \left(-k \cdot \frac{S_{t_k}}{S_p} \right) \right) \quad (1)$$

126 where:

- 127 • Q_{t_k} is the Biomass generated at the growth cycle between t_k and t_{k+1} ;
- 128 • E is an empirical environmental factor (sun lighting, temperature, etc);
- 129 • S_p is the land occupation of the plant;
- 130 • k is photosynthesis efficiency;
- 131 • r is an resistance parameters, setting the effect of mutual shading;
- 132 • S_{t_k} is the total leaf area at t_k .

133 From Eq. (1), one observes that biomass generation is largely dependent of the leaf area S_{t_k}
 134 and is also restricted by land occupation S_p , surrounding environment E and species-dependent
 135 coefficient constant r, k .

136 This description is the one that will be used in our data assimilation process. Let us now
 137 describe the biomass repartition step.

138 2) *Biomass repartition:* Once the biomass is produced, it has to be redistributed among the
 139 different organs of the plant following their own demand. To this end, for each kind of organ o
 140 in the set of organs Ω (Ω can contain organs as leaves, branches, sheath tassel, cob, ... as will
 141 be seen later we will focus mainly on branches b , leaves ℓ and fruits f) one has to compute

142 its *biomass increment* $\Delta_{q_o}(t_{k+1})$ at time t_{k+1} following some rules described below (Yan *et al.*
143 (2004)):

$$D_{t_{k+1}} = \sum_{o \in \Omega} \left(N_o(t_{k+1}) \sum_{t=1}^{t_{k+1}} d_o(t) \right) \quad (2)$$

$$d_o(t) = S_o \cdot \phi_o(t) \quad (3)$$

$$\Delta_{q_o}(t_{k+1}) = d_o(t_{k+1}) \cdot \frac{Q_{t_k}}{D_{t_{k+1}}} \quad (4)$$

144 with

- 145 • $D_{t_{k+1}}$ the total biomass demand in the beginning of cycle t_{k+1} depending on all organs
- 146 $o \in \Omega$;
- 147 • $d_o(t)$ the biomass demand of an organ $o \in \Omega$ for time t ;
- 148 • S_o the sink-strength of organ $o \in \Omega$;
- 149 • $\Delta_{q_o}(t_{k+1})$ the increment of biomass related to organ $o \in \Omega$ in the end of cycle cycle k ;
- 150 • $N_o(t_{k+1})$ the active number of organ of type o at time t_{k+1} ;
- 151 • $\phi_o(t)$ a beta law function related to the demand of each organ. This latter is small at the
- 152 beginning and end of organ growth and relatively large during the growth process Guo *et al.*
- 153 (2006). It takes the following form:

$$\phi_o(t) = (t - 0.5)^{a_o-1} \cdot (t_{go} - t + 0.5)^{b_o-1} \quad (5)$$

154 where a_o , b_o and t_{go} are parameters related to an organ. More precisely, t_{go} is called
155 *expansion time* for organ o and indicates the growing period: after this step, it does not join
156 the biomass repartition in Eq. (2) anymore (this models for example the fact that stalks stop
157 to grow after autumn). Parameters a_o and b_o are calibration ones related to each organ.

158 As one can see, the association of the biomass to one specific organ is based on its demand d_o
159 (cf Eq. (3)) and depends on its sink strength S_o and growth age. The strength is plant-dependent:
160 for example concerning fruit trees, the fruit organ tends to have more demand than a tiny root.
161 Therefore S_o in Eq. (3) is bigger for the fruit organ than tiny root one. As for the age component,
162 GreenLab assumes that an organ grows relatively slowly at the beginning and end of his life
163 and accumulate most portion of biomass in the middle part. This is modelled by function $\phi_o(t)$.
164 Finally, the role of Eq. (4) is to distribute the biomass among organs based on individual demands
165 and the total available biomass.

166 3) *Natural withering*: Regarding leaf area index S_{t_k} related to organ *leaf* (noted ℓ , which is
 167 the variable of interest in this paper) and without withering, a natural evolution model for S_{t_k}
 168 would be $S_{t_{k+1}} = S_{t_k} + \Delta_{q_\ell}(t_{k+1})$. To model the fact that each organ naturally dies after a given
 169 time, a withering of all plants is modelled through the following rule:

$$S_{t_{k+1}} = \alpha \cdot (S_{t_k} - \beta(t_k, t_{el})) + \Delta_{q_\ell}(t_{k+1}). \quad (6)$$

170 This equation expresses the fact that in the end of cycle t_k , the next leaf area $S_{t_{k+1}}$ will be
 171 increased by $\Delta_{q_\ell}(t_{k+1})$, the growth of organ *leaf* issued from equation (4). This is altered by
 172 $(\alpha, \beta(t_k, t_{el}))$. The rule of coefficient $\alpha \in [0, 1]$, which stands for the spoilage in natural growth,
 173 is to model a global withering while function $\beta(t_k, t_{el})$ is time-varying and relies on the *life*
 174 *expectance* (roughly speaking the “birth”) t_{el} of leaves ℓ . The value of this function will be
 175 discussed in next section.

176 Equations (1–6) are the core of the GreenLab model for biomass generation, partition and
 177 withering. This model is dependant from many parameters. It is useful to *simulate* plant growth
 178 from known parameters but obviously their *recovery* from image sequences appears impossible,
 179 as pointed out in Yang *et al.* (2008). Fortunately, many simplifications related to medium
 180 resolution remote sensing images are possible and we can design an assimilation tool based
 181 on GreenLab principles. This is the scope of next section.

182 B. Simplified Greenlab for remote sensing data assimilation

183 We suggest to simplify and adapt the initial GreenLab model to remote sensing. Though the
 184 biomass generation process does not change, all details regarding the internal structure of plants
 185 can be simplified. Associated manipulations are detailed bellow.

186 1) *Focus main organs*: We decide to take into consideration only the *three* most important
 187 organs responsible of the majority of biomass production: leaf, fruit and branch (internode),
 188 respectively noted (ℓ, f, b) , yielding a set of organs $\Omega = \{\ell, f, b\}$, the remaining ones being
 189 modelled with noises, as will be shown in equation (10).

190 2) *Plant structure simplification*: In order to describe plant structure evolution, we keep an
 191 empirical *Look-Up-Table*, $U = \{U_i | i = 1, \dots, N\}$ which store statistical number for different
 192 appearing organs. For instance, $U_4 = \{\ell = 3, b = 2, f = 0\}$ means that at 4_{th} growth cycle,
 193 there would be 3 new leaves, 2 new internodes and no new fruits (one also notes $U_4\{\ell\} = 3$,
 194 $U_4\{b\} = 2$ and so on). In practice, available Look-Up-Tables for more than 30 species are

195 already available (see for example Ma *et al.* (2006); Mathieu *et al.* (2007); Wang *et al.* (2009))
 196 and are used in this study.

197 3) *Natural withering modelling*: In equation (6), the withering of leaves at the growth cycle
 198 between t_k and t_{k+1} is modelled with function as $\beta(t_k, t_{eo})$. This function is plant-dependent
 199 and is conditioned by the age of leaves which in practice is available through the structural
 200 sub-model. To prevent from the dependency from this age and on the kind of plants, we rely on
 201 the *active number of organs* $N_o(t_k)$ computed using the Look-Up-Tables U previously defined:

$$N_o(t_k) = \sum_{i=t_k-t_{eo}}^{t_k} U_i\{o\}. \quad (7)$$

202 where t_{eo} is the expectance-time for any organ $o \in \Omega$. Concerning function $\beta(t_k, t_{eo})$ in Eq. (6),
 203 its formulation is based on leaves's appearing time and life expectance t_{eo} as:

$$\beta(t_k, t_{eo}) = \frac{U_{t_k-t_{eo}}\{\ell\}}{N_\ell(t_k)} \cdot S_{t_k} \quad (8)$$

204 with N_ℓ the active number of leaves. Its expression is in practice computed using available
 205 Look-Up-Tables and relation (7).

206 4) *Overall model used for data assimilation*: To model potential errors likely to arise because
 207 of the three aforementioned kind of simplifications, a white gaussian noise of standard deviation
 208 η_{t_k} is introduced for each t_k . Therefore, the overall sequential model to compute Leaf Area
 209 Index $S_{t_{k+1}}$ based on S_{t_k} and greenlab reads:

$$S_{t_{k+1}} = \alpha \cdot (S_{t_k} - \beta(t_k, t_{eo})) + \Delta_{q_\ell}(t_{k+1}) + \eta_{t_k}. \quad (9)$$

210 Using relations (1) to (8), one can reformulate the above plant growth model by:

$$S_{t_{k+1}} = M_k[S_{t_k}] + \eta_{t_k}$$

with:

$$M_k[S_{t_k}] = \underbrace{\alpha \left(1 - \frac{U_{t_k-t_{eo}}\{\ell\}}{N_\ell(t_k)} \right) S_{t_k} - \frac{ES_p d_o(t_{k+1})}{rD_{t_{k+1}}} \exp\left(-k \cdot \frac{S_{t_k}}{S_p}\right)}_{\text{non linear model w.r.t. } S_{t_k}} \quad (10)$$

$$+ \underbrace{\frac{ES_p d_o(t_{k+1})}{rD_{t_{k+1}}}}_{\text{independent from } S_{t_k}}$$

211 where the *unknown parameters* to recover are:

- 212 • S_{t_k} the Leaf Area Index at time t_k ;

- 213 • k the photosynthesis efficiency parameter in equation (1);
- 214 • E/r a coefficient related to the external influence (environmental over resistance parameter)
- 215 in equation (1);
- 216 • α standing for the spoilage in natural growth in equation (6);
- 217 • t_{el} standing for the life expectance of leaves;
- 218 • η standing for the noise due to model errors in equation (10).

219 All other parameters involved in (10) are available through Loop-Up-Tables and from relations
 220 (2) to (5). The second term of $M_k[S_{t_k}]$ in (10) does not depend on the variable of interest S_{t_k}
 221 and the first one is nonlinear because of its exponential part. This model will be used in our
 222 assimilation process.

223 In Figure 2, we give some illustrations of the leaf area index evolution depending on some
 224 parameters. This new version of the model, adapted to medium resolution remote sensing images,
 225 can now be used in data assimilation schemes to combine it with images.

226 C. Data assimilation

227 Combining different sources of information driven by a physical model, commonly named
 228 “data assimilation”, has always been a hot research topic for which a large panel of approaches
 229 exist. Researchers in this area have developed a number of efficient inference tools, such as
 230 stochastic methods Doucet *et al.* (2000), Kalman filter Welch and Bishop (1995) and varia-
 231 tional approaches as the well-known 4DVAR algorithm Bain and Crisan (2009); Le Dimet and
 232 Talagrand (1986); Lions and Mitter (1971); Courtier *et al.* (1994).

233 In all these techniques one can model the problem as follows: the data to estimate (noted S_{t_k}
 234 at time t_k in our application), driven by a dynamical model M , are represented through a system
 235 state discretized into a sequence of hidden states Kitagawa (1996) and are observed directly or
 236 indirectly by O_{t_k} through an observation system H :

$$S_{t_{k+1}} = M_k[S_{t_k}] + \eta_{t_k} \quad (11)$$

$$O_{t_k} = H[S_{t_k}] + \epsilon_{t_k} \quad (12)$$

237 where η_{t_k} (resp. ϵ_{t_k}) are the system noises to estimate that models uncertainties w.r.t the dynamics
 238 M (resp. w.r.t observation operator H). In our application the prediction model M_k is based on
 239 Greenlab and is modelled in Eq. (6). As for the observation operator H , in practice because
 240 of the complexity of the relationship between the image luminance and the LAI, the definition

241 of a complete observation system H that directly links the LAI to data would be tricky. We
 242 therefore prefer to adopt a two stage strategy where at first we estimate the leaf area index
 243 thanks to the PROSAIL model Jacquemoud *et al.* (2009); Lecerf *et al.* (2008) and then use an
 244 identity observation operator in H . This “pseudo-observation” approach is commonly used when
 245 data assimilation systems involve complex relationships between observations and system states
 246 Courtier and Talagrand (1987). In practice, efficient tools based on neural network to inverse
 247 PROSAIL model to estimate biophysical variables from MODIS time series exist and we use
 248 the technique presented in Lecerf *et al.* (2008).

249 Among possible solutions mentioned above to perform model inference, variational and stochas-
 250 tic techniques are the most popular. Here, because of the complexity and non-linearity of our
 251 dynamical model, we prefer to rely on stochastic approaches and more precisely on the **forward-**
 252 **backward smoother** since we consider the whole sequence to recover all data (unlike particle
 253 filter which recovers data with the knowledge of past and current data only) Doucet (2001).
 254 Starting from:

- 255 • An observation sequence $\bar{O}_{t_k} = \{O_{t_1}, O_{t_2}, \dots, O_{t_k}\}$ available sequentially;
- 256 • An initial distribution of the system’s state $p(S_{t_1})$;
- 257 • Transition and observation models: $p(S_{t_k}|S_{t_{k-1}})$ and $p(O_{t_k}|S_{t_k})$ respectively, related to the
 258 stochastic processes M and H presented above;
- 259 • Common assumption of zero-mean time-independent Gaussian noise for η, ϵ in (11-12),
 260 leading to $p(S_{t_k}|S_{t_{k-1}}) \sim \mathcal{N}(M_k(S_{t_{k-1}}), \eta)$ and $p(O_{t_k}|S_{t_k}) \sim \mathcal{N}(H_k(S_{t_k}), \epsilon)$ with \mathcal{N} the
 261 normal distribution,

262 it can be shown that the sequential estimation of (S_{t_k}) can be obtained using a two-fold
 263 prediction/correction system. We refer readers to Doucet (2001) for more details.

264 Following equations (1-6), the set of additional parameters $\{\Theta = E/r, k, \alpha, t_{el}, \eta, \epsilon\}$ has to be
 265 fixed in order to describe properly the LAI evolution of a plant. Parameters $E/r, k$ are related
 266 to environment, leaf mutual shading parameter and photosynthesis efficiency in Eq. (1), α to
 267 natural leaf foliage in Eq. (6), t_{el} the time expectance of leaves in (6), η to the system noise in
 268 Eq. (11) and ϵ to the observation variance in Eq. (12). They are estimated for each time step t_k
 269 by a least square between the mean state \bar{S}_{t_k} of particles $S_{t_k}^i, i = \{1 \dots N\}$ at a given time t_k and
 270 the LAI $S_{t_k}^e$ issued from GreenLab model, depending on $\{r, k, \alpha\}$ (see illustration in Fig. 2):

$$\{r, k, \alpha, t_{el}\} = \arg \min_{r, k, \alpha, t_{el}} \sum_{i=1}^N \|\bar{S}_{t_k} - S_{t_k}^e\|^2. \quad (13)$$

271 As for the system noise η in Eq. (11), its distribution is assumed to be time independent
 272 zero-mean Gaussian. Its variance σ_η can then directly be computed in each time step t_k using
 273 available N particles with the relation:

$$\sigma_\eta = \frac{1}{N} \sum_{i=1}^N \|S_{t_k}^i - S_{t_k}^e\|^2. \quad (14)$$

274 The same holds for the estimation of ϵ : noisy estimation $S_{t_k}^o$ of Leaf Area Index being acquired
 275 from each image O_{t_k} by an inversion of PROSAIL model, the uncertainty σ_ϵ related to the
 276 observation system can be extracted through:

$$\sigma_\epsilon = \frac{1}{N} \sum_{i=1}^N \|S_{t_k}^i - S_{t_k}^o\|^2. \quad (15)$$

277 Concerning the initialization of the system, we estimate noisy LAI sequence $\{S_{t_k}^o | k = 1, \dots, T\}$
 278 from input images. Data with missing observations are approximated with a polynomial regres-
 279 sion from $\{S_{t_k}^o\}$. All these steps result in the following iterative algorithm:

Incremental technique for LAI estimation and parameter inference

- Initializations:
 - From MODIS images $\{O_{t_k}\}$, compute noisy estimations $\{S_{t_k}^o\}$ of LAI following Lecerf *et al.* (2008);
 - Perform a polynomial regression on $\{S_{t_k}^o\}$ in order to initialize $\{S_{t_k}^r | k = 1, \dots, M\}$;
- 1) Estimate hidden parameters $\{E/r, k, \alpha, t_{el}\}$ with Eq. (13)
- 2) Compute associated uncertainties $\{\eta, \epsilon\}$ with Eq. (14-15)
- 3) Perform LAI inference through particle methods described in this section (cf Doucet (2001))
- 4) **Loop** to step (1) until the estimation of LAI converges
- Once converged, the reconstructed Leaf Area Index sequence $\{S_{t_k}^r | k = 1, \dots, M\}$ and associated hidden parameters $\Theta = \{r, k, \alpha, t_{el}, \eta, \epsilon\}$ are available.

281 From this algorithm, we observe that the model and observation uncertainties η and ϵ are
 282 automatically adapted during the process.

III. EXPERIMENTAL RESULTS

283
 284 Our approach has been tested on synthetic, real data sets with ground truth and real data and
 285 in order to analyse the benefits of our method under various situations. Before entering into
 286 details, let us discuss about the experimental setup.

287 A. *Experimental settings*

288 The following experimental conditions have been applied:

- 289 • In each experiment, the number of particles has been set to 200;
- 290 • The efficiency of the reconstruction process presented in this paper has been compared
291 with polynomial regression (5 degrees) and wavelet filter (Daubechies with 5-level). These
292 parameters have also been set by cross-validation. When ground truth is available, only
293 polynomial and linear interpolation has been performed since no enough data were available
294 to test wavelet filters;
- 295 • We have indicated the results with a simulation of GreenLab with estimated parameters
296 $(E/r, k, \alpha, t_{el})$ using the proposed procedure in Section II-C. Then we could iteratively
297 predict the LAI values without the correction steps, as demonstrated in Fig. 2. This could
298 be considered as a model-based regression ;
- 299 • All graphes have been compensated by estimated t_{el} in order to have similar starting points.

300 B. *Data with ground truth measurements*

301 To evaluate the efficiency of our technique, we have used ground LAI measurements of 9 series
302 of wheat and 9 series of rape acquired from January to July 2017 in Brittany, North France (5
303 ground measurements for rape and 4 ground measurements for wheat). These measurements act
304 as ground truth validation. All series have been acquired on different parcels (whose size are
305 larger than one SENTINEL-2 pixel) to sense the variability of our approach. Investigated areas
306 are part of a Long Ecological Research site named “Pleine Fougres”, located on the southern
307 part of the Bay of Mont-Saint-Michel, France, and referenced in the LTER-Europe¹ and ILTER
308 networks². This agricultural landscape is dominated by fields of wheat, maize, rapeseed and
309 grasslands. Field observations and measurements were conducted on the study site during one
310 crop year (2017) in order to calibrate and quantitatively validate the proposed approach. Different
311 parameters were measured and georeferenced once per three weeks using a GPS (5 m accuracy):
312 water saturation, vegetation height and LAI, whereas phenology stages, dates of sowing and
313 harvests, and yields were observed in 18 fields.

¹See: <http://www.lter-europe.net/>

²See: <https://www.ilter.network/>

314 An illustration of the position of on site measurements is visible in figure 4. Corresponding
315 SENTINEL-2 images (9 images during the period) have been downloaded and our process has
316 been applied on pixels associated with parcels. To evaluate the efficiency of the GreenLab model,
317 data from May to September (resp. from February to May) have been removed in the processing
318 of wheat (resp. rape). This choice has been done on the basis of the associated phenological
319 states in order to remove from observations the main informations. Estimated values (one image
320 per day) have been compared with ground truth.

321 Associated errors (mean and standard deviations on 100 realisations) and illustrations of some
322 reconstructions are respectively depicted on table I and figure 5 both for wheat and rape. As
323 one can observe on figure 5, the assimilations with GreenLab (particle filter and smoother) are
324 systematically more in accordance with ground truth than interpolations. For example in all series
325 with wheat, as the maximum of LAI is never observed, interpolation techniques that are not driven
326 by a physical model fail in recovering the growth and decrease of LAI. On the contrary, the use
327 of a physical model enables to better recover the physical evolution of LAI. The same kind of
328 observations stands for rape. Here, as observations during the growing period are missing (from
329 February to May), interpolation techniques fail in recovering the variability of the growth, unlike
330 our assimilation approach. All these observations can be confirmed with quantitative values in
331 table I where assimilations with particle smoother (and then filter) systematically outperform
332 interpolation techniques.

333 These first experiments with ground truth measurements confirm the ability of our technique
334 to recover LAI thanks to the GreenLab model. We now turn to experiments with estimated LAI.

335

336 *C. Data with estimated LAI*

337 Data used are issued from real MODIS reflectance product (a sequence of 17 images) on which
338 LAI estimations, issued from the PROSAIL model, have been reconstructed and validated (see
339 Lecerf *et al.* (2008, 2005)). In addition, associated classifications of large agricultural parcels (we
340 mean by *large* the fact that they recover more than one MODIS pixel) were available, providing
341 us confident LAI time series and associated land cover. In practice three types of agricultural
342 crops were used: colza, maize and wheat. Some LAI profiles are visible in Fig. 3.

343 To model the various artefacts prone to occur in remote sensing images, several processes
344 have been applied on these data:

- 345 • *Noisy observations.* Data are equally subsampled (one image every 10 days) from ground
346 data and a Gaussian white noise has been introduced as shown in Fig. 6.
- 347 • *Sparse observations.* Data for only 6 observations have been kept. The motivation is to
348 evaluate the performance of our algorithm under a short length of remote sensing observation
349 time series (very cloudy regions), as shown in Fig. 7.
- 350 • *Missing observations.* Very few observations were used (7 in practice) but here we simulate
351 the absence of data during a long series. Therefore missing data are consecutive in time as
352 shown in Fig. 8.

353 *1) Result on sequence of noisy observations:* This kind of data represents a great challenge in
354 remote sensing analysis since all observations are in general corrupted by noise and conventional
355 techniques often require to have an estimation of such noise Rabiner and Gold (1975); Burrus
356 *et al.* (1998), unlike our process.

357 In order to make a quantitative evaluation, various levels of Gaussian zero-mean noise, with
358 Signal-to-noise ratio (SNR) ranging from 2 to 10, were tested. The table II represents the
359 root-mean-square error (RMSE) between reconstructed LAI (with particle filter and smoother
360 and other reconstruction techniques introduced in section III-A) and reference data for 100
361 realizations. It is interesting to observe in this table that the introduction of GreenLab plant growth
362 model in the reconstruction process systematically reduces the error residual and outperforms
363 all other methods. Particles smoother almost always gives a better performance than particle
364 filter, which is quite rational as when estimating the LAI at any time point, particle filter only
365 exploits information of up-to-date observations, while particle smoother takes into account all
366 observations in time series. In order to have a visual inspection of the reconstructions, some
367 illustrations are visible in Fig. 6 and confirm that our outputs are of reliable quality. Even if other
368 curves are sometimes consistent with ground truth, particle filter and/or smoother represent the
369 best compromise since strong errors are likely to appear with other techniques (as the polynomial
370 one very sensible to little variations).

371 *2) Sequence with sparse observations:* We have randomly taken only six observations to
372 reconstruct the whole time series. These observations are represented by blue dotted points in
373 Fig. 7. As one can observe, they are more or less issued from a uniform sample along the time
374 period. Quantitative results are depicted in table III. Here again, the performance and the benefit
375 of relying on GreenLab growth model is illustrated. Reconstructed curves are also visible in
376 figure 7 and demonstrate the great ability of our technique to recover proper series.

377 3) *Sequence of missing observations*: We have removed long periods of consecutive data.
378 Some illustrations are visible in Fig. 8 where one can observe that no observations during the
379 growing period are available, which is a very delicate problem. Despite these difficulties, the
380 proposed framework is able to provide very good reconstruction of LAI time series, as shown
381 in Tab. IV and in Fig. 8. Obviously here, only the dynamical model prior is able to estimate
382 consistent values during critical periods of missing data. This is to our opinion a very interesting
383 behaviour of our estimators.

384 In this three experiments, various reconstruction methods have been tested. In all situations, the
385 use of GreenLab as prior plant growth model has enabled to improve the reconstruction of LAI
386 over long time periods. This is to our opinion a very interesting behavior likely to have many
387 potential applications. It should also be outlined that our technique not only reconstructs the
388 LAI but is also able to extract deviations with respect to observations and dynamical model
389 (parameters $\{\eta, \epsilon\}$) and internal parameters of plants $\{E/r, k, \alpha, t_{el}\}$. This point may have
390 interesting applications in the future. *At the moment, three species (wheat, rape, maize) have
391 been studied but the functional part of GreenLab enables this model to adapt to many other kind
392 of cultures. Associated landscapes (Brittany, France) and crops on which our experimentations
393 have been performed are very fragmented, which disturbs the observation with moderate remote
394 sensing images. It is expected that in larger areas (USA, China, ...), the application of our process
395 would be easier. It is also important to point out that some recent domain adaptation techniques
396 Courty *et al.* (2017) issued from machine learning and artificial intelligence (how to fit a model
397 designed for one site to another site) could be tested on these assimilation approaches to adapt
398 this work to other regions or species. This is actually the scope of a current work.*

399 This second series of experiments has quantitatively demonstrated the efficiency of our tech-
400 nique. Let us now present experiments on real MODIS data.

401 *D. Application on large scale study site*

402 We have applied our methods on a large scale study site in Chizé, France. This study site
403 is abundant with various sorts of vegetation, such as wood, grassland, maize, colza, wheat. In
404 practice 16 images from March to October 2006 were available. On these images, the PROSAIL
405 model has been applied to derive LAI time series. An illustration of available data is visible in
406 the top of Fig. 11 where red stands for high values of LAI (indicating dense plant canopy) and

407 blue for low LAI. However at some location, observation data are corrupted, mainly because of
408 cloud coverage, yielding corrupted time series.

409 In the bottom of figure 11, the results from particle smoother under our proposed framework
410 are depicted. In addition we have plotted in Fig. 9 some curves randomly taken from the images.
411 Despite the fact that no ground truth exist here, it appears obviously that our method seems
412 consistent and enables to recover uncorrupted LAI time series.

413 IV. CONCLUSION

414 In this paper, we have proposed a technique that estimates leaf area index time series using
415 data assimilation. The dynamical model is based on the GreenLab plant growth model where
416 a number of modifications have been applied to adapt it in a remote sensing context. LAI
417 observations are issued from the inversion of the PROSAIL model. As shown in our experiments,
418 this technique is efficient in many various situations as very sparse observations or missing/noisy
419 data. Quantitatively, it outperforms other reconstruction strategies that do not rely on a physical
420 prior knowledge, especially when large period of missing data occur. An additional key point of
421 our process is its ability to estimate internal parameters of the plant unobservable from images
422 (environmental parameters, spoilage, ...). This is to our opinion a very good property that open
423 new applications in the future.

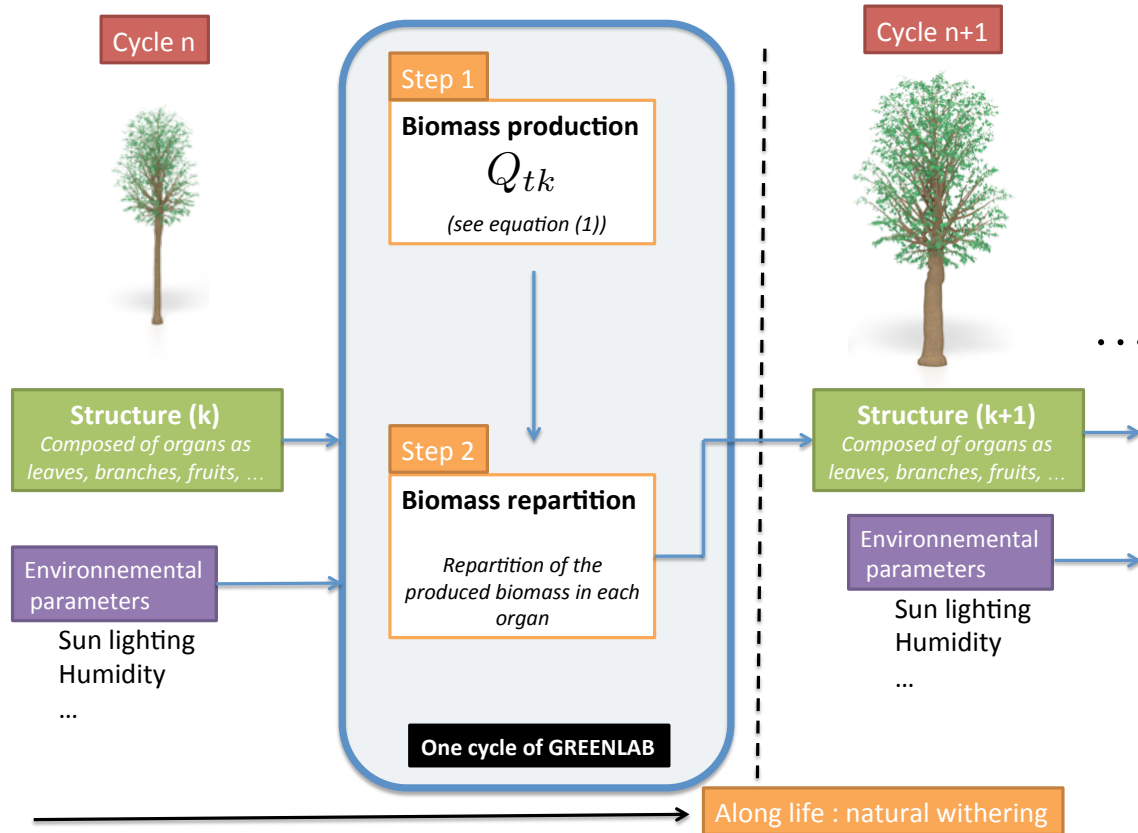


Fig. 1. **Illustration of GreenLab model with two steps:** in each cycle, the **biomass production** step computes the produced biomass based on the current plant structure and environmental parameters. The **biomass repartition** step allocates this biomass in various organs to generate a new plant at cycle $n + 1$. All these operations are balanced by healthy coefficients that model the **natural withering**

424

425

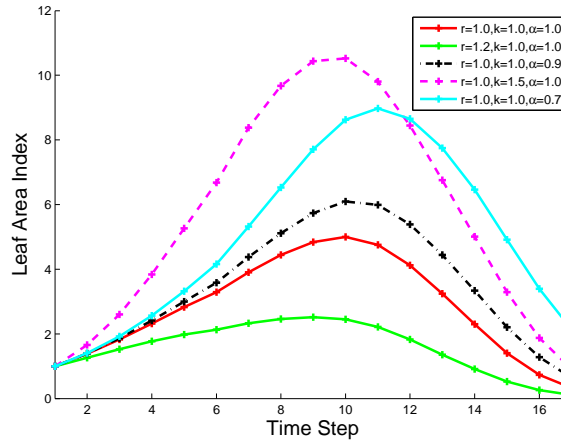


Fig. 2. Leaf area index evolution with different r, k, α

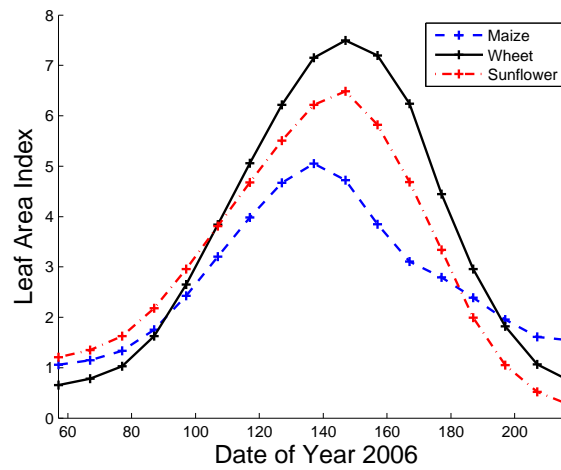


Fig. 3. A demonstration of uncorrupted LAI time series for three sorts of agriculture crops in Brittany, France

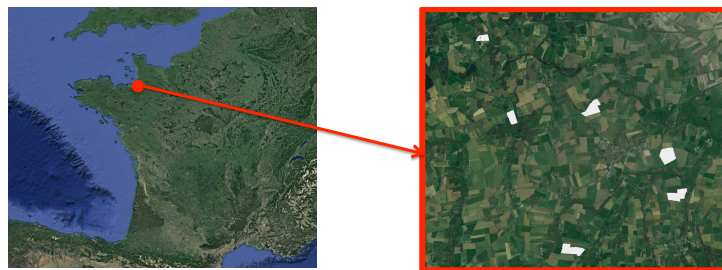


Fig. 4. Localisation of on site measurements for wheat and rape in Brittany, North of France. All parcels are higher than a SENTINEL-2 pixel area

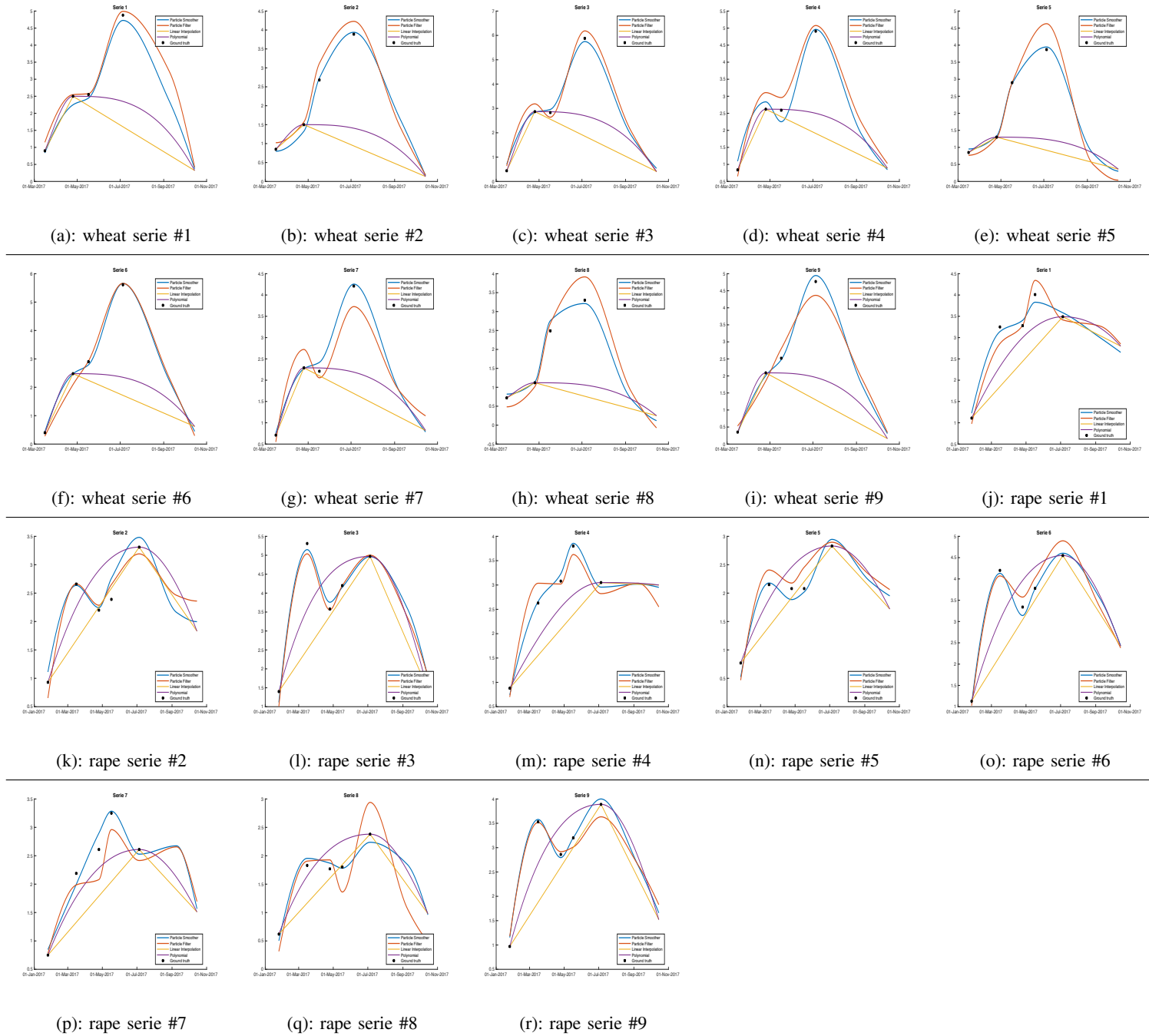


Fig. 5. LAI reconstruction for wheat and rape with ground truth superposition over the year 2017 with missing data from May to September (wheat) and from February to May (rape). Ground truth measurements are block dots, GreenLab assimilation with particle smoother (our technique) is in blue, GreenLab assimilation with particle filter is in red, polynomial interpolation in purple and linear interpolation in yellow.

TABLE I
 ROOT-MEAN-SQUARE ERROR (RMSE) IN RECOVERING LAI FOR ALL SERIES AND IN AVERAGE FOR WHEAT (TOP) AND
 RAPE (BOTTON).

Wheat Series #	Polynomial	Linear	Particle Filter	Particle Smoother
1	1.2656	1.6455	0.2904±0.1056	0.1447±0.0557
2	1.3759	1.6240	0.2722±0.0996	0.1354±0.0541
3	1.5901	2.0144	0.2765±0.1175	0.1419±0.0501
4	1.2049	1.5030	0.2978±0.1117	0.1420±0.0453
5	1.5419	1.7098	0.2948±0.1103	0.1450±0.0540
6	1.6430	1.9742	0.2861±0.1021	0.1458±0.0546
7	1.0115	1.2628	0.2861±0.1047	0.1416±0.0427
8	1.3134	1.4688	0.2821±0.0976	0.1415±0.0565
9	1.4233	1.7686	0.2912±0.0944	0.1434±0.0529
Average	1.3744	1.6634	0.2864±0.1048	0.1424±0.0518
Rape Series #	Polynomial	Linear	Particle Filter	Particle Smoother
1	0.3870	0.4039	0.2934±0.1000	0.1442±0.0529
2	0.2488	0.2025	0.2767±0.1042	0.1359±0.0488
3	0.9274	0.6938	0.2729±0.0961	0.1408±0.0470
4	0.4925	0.5515	0.2909±0.1059	0.1466±0.0524
5	0.0464	0.0354	0.2814±0.0951	0.1384±0.0497
6	0.4558	0.3834	0.2835±0.1046	0.1546±0.0484
7	0.4504	0.5028	0.2669±0.0996	0.1431±0.0501
8	0.0296	0.0175	0.3003±0.1124	0.1443±0.0501
9	0.3396	0.2485	0.2917±0.1107	0.1432±0.0565
Average	0.3753	0.3377	0.2842±0.1032	0.1435±0.0507

TABLE II
 THE ROOT-MEAN-SQUARE ERROR (RMSE) OF DIFFERENT METHODS ON SEQUENCES OF VARIOUS SNR LEVEL.

RMSE	SNR=2	SNR=4	SNR=6	SNR=8	SNR=10
Observations	1.3222±0.2637	0.8712±0.1712	0.7747±0.1327	0.6707±0.1300	0.6313±0.0895
Polynomial	0.7971±0.2126	0.6120±0.1535	0.5760±0.1168	0.5351±0.0893	0.5136±0.0770
Wavelet filter	0.8018±0.1749	0.7040±0.0895	0.6795±0.0793	0.6481±0.0848	0.6365±0.0548
Fitted model	0.7063±0.2477	0.5749±0.1829	0.5259±0.1561	0.5189±0.1699	0.4586±0.0843
Particle filter	0.7755±0.2500	0.6633±0.1822	0.5973±0.1329	0.5452±0.1327	0.5403±0.1175
Particle smoother	0.7072±0.2495	0.5660±0.1634	0.5014±0.1240	0.4837±0.1036	0.4341±0.1057

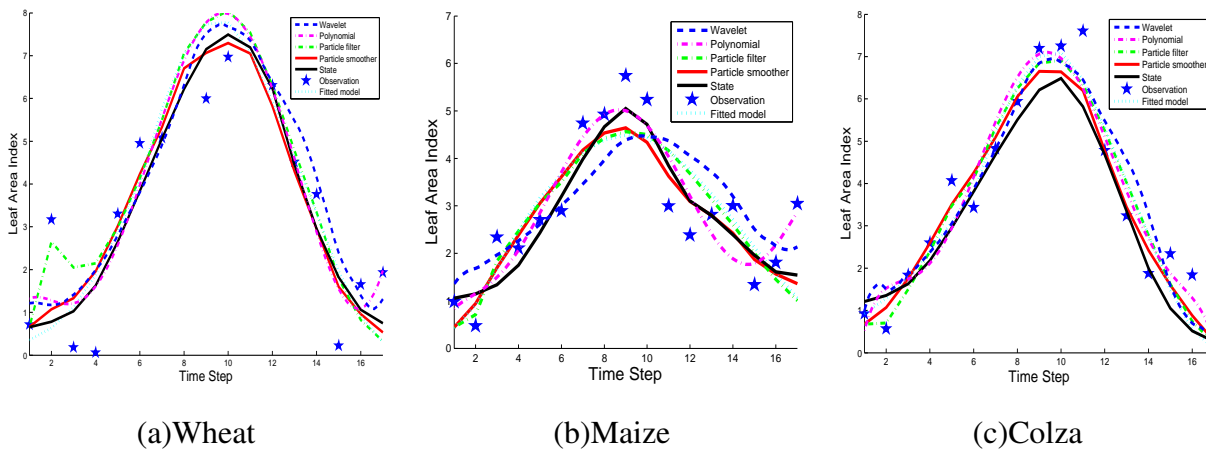


Fig. 6. **LAI reconstruction with various techniques on the sequence of noisy observations (with SNR=6).** Dark line are uncorrupted MODIS observations (ground truth state), blue dots are the input noisy observations, red line is the result of reconstruction with particle smoother, cyan line from particle filter, green line from fitted model, pink line from polynomial regression, blue line from wavelet filter. Even in this situation where the noise is large, the best fitted curves are issued from techniques based on the GreenLab model

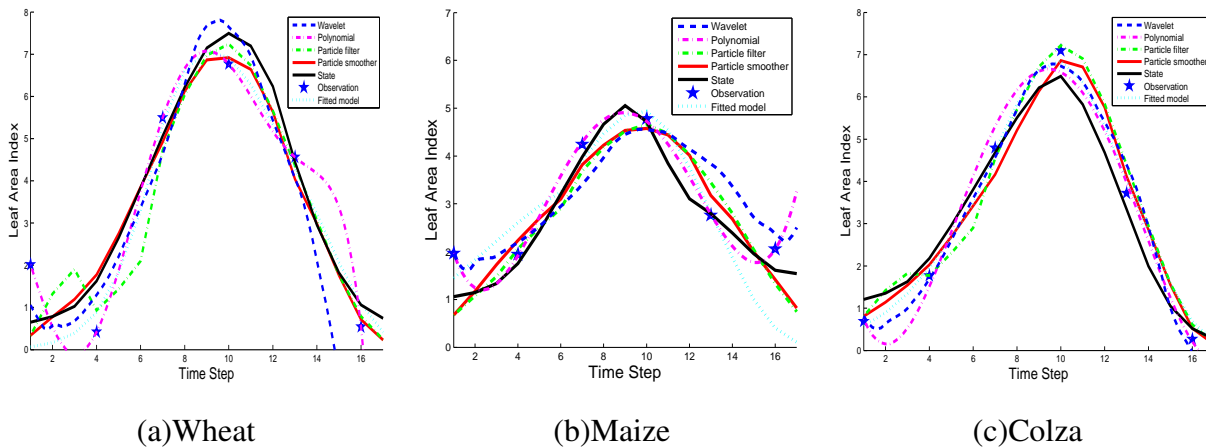


Fig. 7. **LAI reconstruction with various techniques on the sequence of sparse observations (with SNR=6).** Dark line are uncorrupted MODIS observations (ground truth state), blue dots are the input noisy observations, red line is the result of reconstruction with particle smoother, cyan line from particle filter, green line from fitted model, pink line from polynomial regression, blue line from wavelet filter.

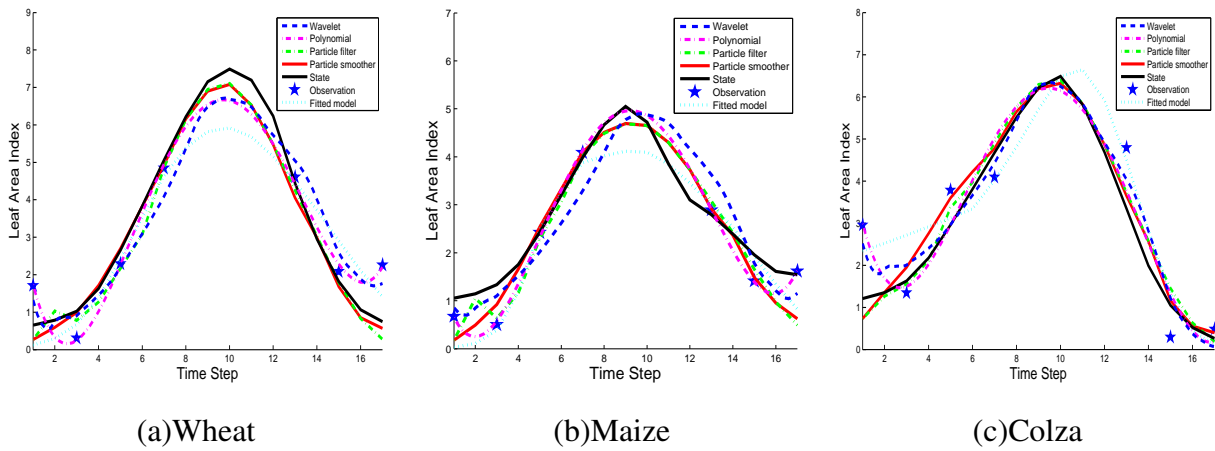


Fig. 8. LAI reconstruction with various techniques on the sequence of missing observations (with SNR=6). Dark line are uncorrupted MODIS observations (ground truth state), blue dots are the input noisy observations, red line is the result of reconstruction with particle smoother, cyan line from particle filter, green line from fitted model, pink line from polynomial regression, blue line from wavelet filter.

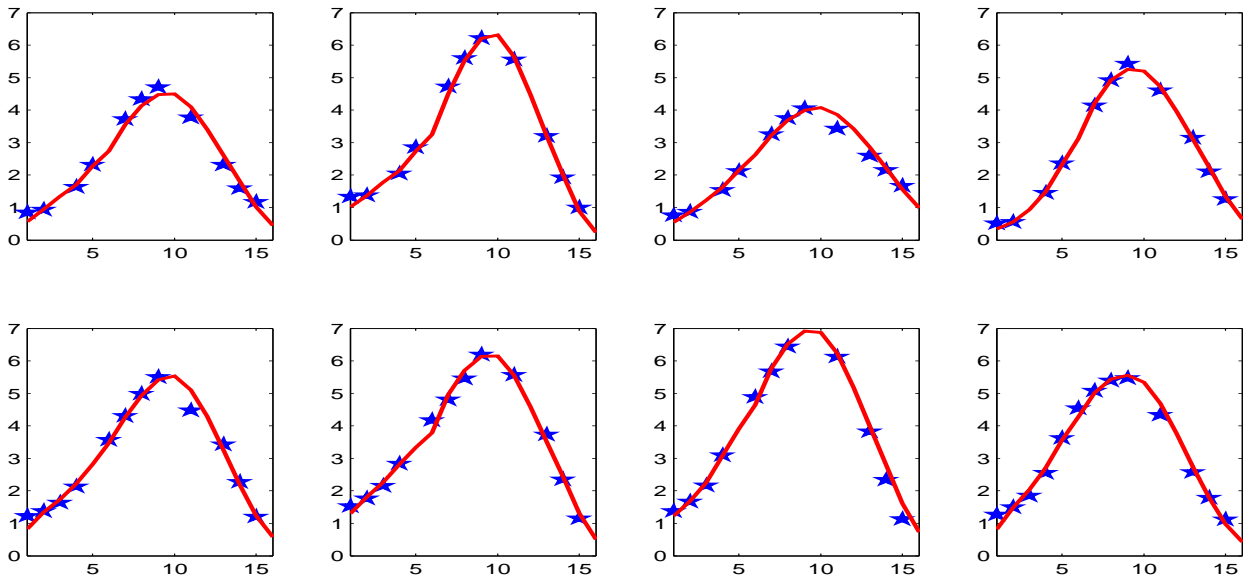


Fig. 9. Several examples of LAI time series reconstruction with particle smoother at a study site in Chizé, France

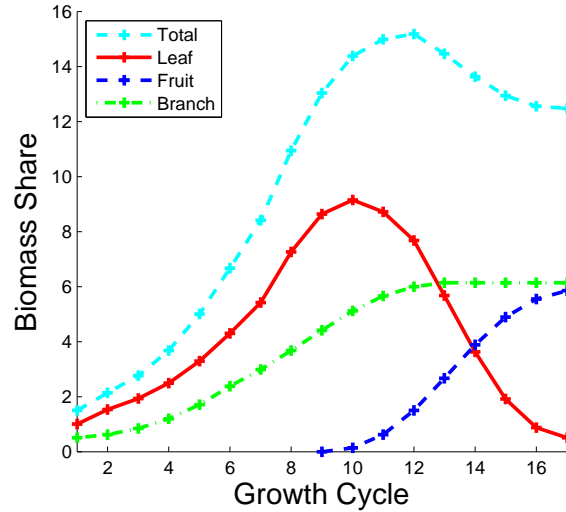


Fig. 10. Estimated biomass production and its partition during continuous growth cycles of unit wheat field.

TABLE III

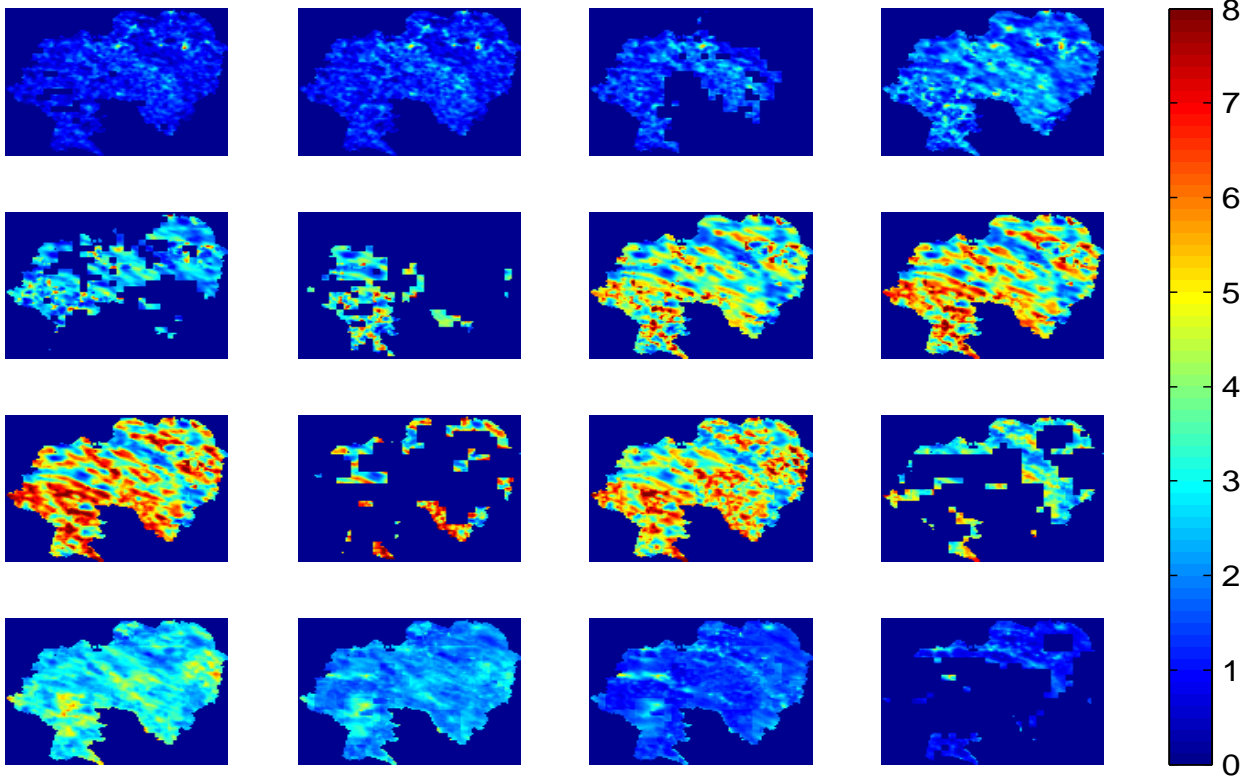
THE ROOT-MEAN-SQUARE ERROR (RMSE) OF DIFFERENT METHODS ON SEQUENCE WITH SPARSE OBSERVATIONS.

RMSE	SNR=2	SNR=4	SNR=6	SNR=8	SNR=10
Observations	1.4425±0.3580	1.2365±0.3881	0.9455±0.3495	0.7818±0.2562	0.6887±0.1431
Polynomial	1.4943±0.5823	1.2852±0.1447	0.1447±0.4577	0.8807±0.3098	0.7867±0.1761
Wavelet filter	1.1932±0.4386	1.1424±0.3814	1.0390± 0.4995	0.7390±0.2517	0.7229±0.1860
Fitted model	1.0346±0.3362	1.0425±0.3267	0.8574±0.3565	0.6767±0.3192	0.7126±0.2611
Particle filter	1.1762±0.3986	1.1233±0.3220	0.8523±0.3780	0.7931±0.3361	0.7549±0.1903
Particle smoother	1.0198±0.3526	0.9622±0.3477	0.9451±0.3330	0.5923±0.2364	0.5919±0.1420

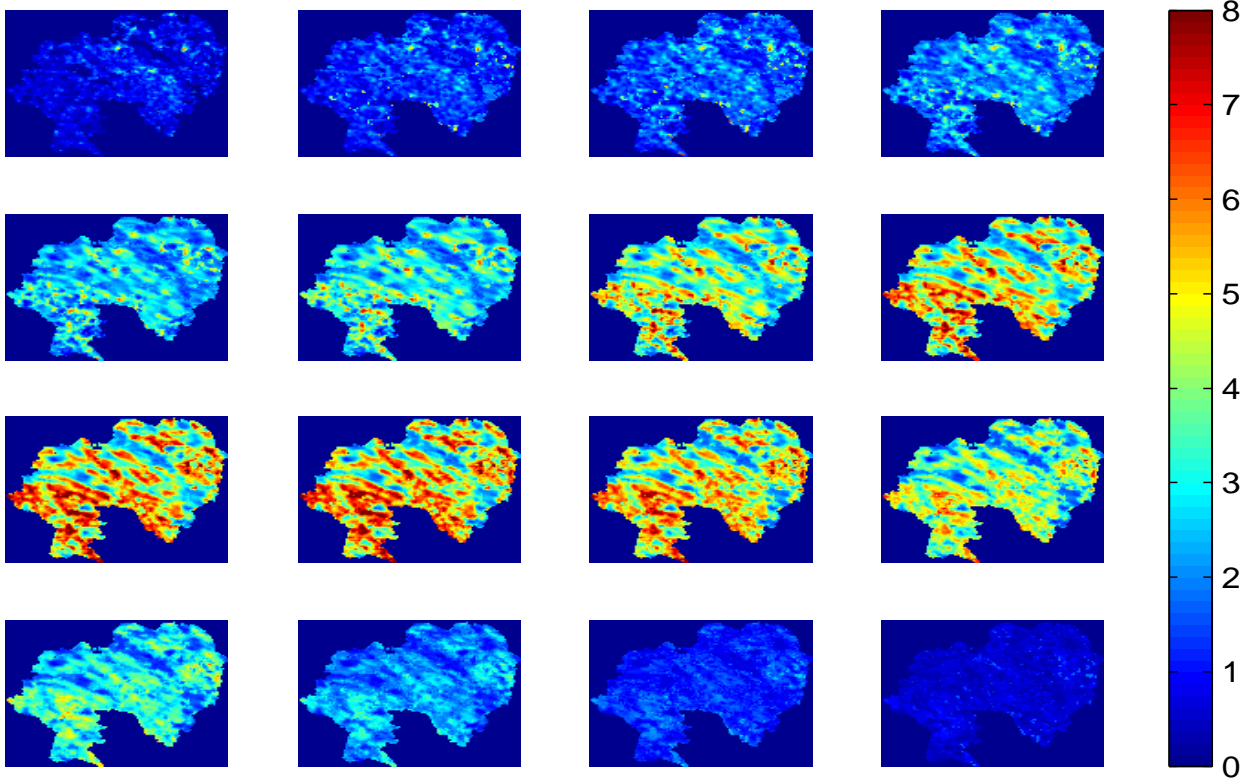
TABLE IV

THE ROOT-MEAN-SQUARE ERROR (RMSE) OF DIFFERENT METHODS ON SEQUENCE OF MISSING OBSERVATIONS.

RMSE	SNR=2	SNR=4	SNR=6	SNR=8	SNR=10
Observations	1.2934±0.3317	1.0424±0.2523	0.9218±0.2468	0.7011±0.1642	0.7118±0.1709
Polynomial	1.2606±0.5608	1.1733±0.4756	1.0649±0.4444	1.1239±0.6996	0.8877±0.5777
Wavelet filter	1.3656±0.6070	1.0531±0.3697	0.8184± 0.2907	0.7741±0.3403	0.7840±0.3407
Fitted model	1.2036±0.6758	1.0689±0.4229	0.8079±0.2906	0.7072±0.3583	0.6894±0.3031
Particle filter	1.2617±0.6158	0.9596±0.3778	0.8526±0.2759	0.7232±0.3553	0.7044±0.3254
Particle smoother	1.2065±0.6876	1.0253±0.4510	0.8055±0.3180	0.7050±0.3578	0.6892±0.3479



(a)



(b)

Fig. 11. Result of particle smoother at a study site in Chizé, France
October 31, 2018

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