

# Does pollution influence small mammal diet in the field? A metabarcoding approach in a generalist consumer

Shinji Ozaki, Clémentine Fritsch, Benoît Valot, Frédéric Mora, Renaud

Scheifler, Francis Raoul, Thierry Cornier

# ▶ To cite this version:

Shinji Ozaki, Clémentine Fritsch, Benoît Valot, Frédéric Mora, Renaud Scheifler, et al.. Does pollution influence small mammal diet in the field? A metabarcoding approach in a generalist consumer. Molecular Ecology, 2018, 27 (18), pp.3700-3713. 10.1111/mec.14823 . hal-02368039

# HAL Id: hal-02368039 https://hal.science/hal-02368039v1

Submitted on 7 Feb 2022

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1	Does pollution influence small mammal diet in the field? A metabarcoding approach
2	in a generalist consumer.
3	
4	Shinji Ozaki <sup>*1</sup> , Clémentine Fritsch <sup>1</sup> , Benoit Valot <sup>1</sup> , Frédéric Mora <sup>2</sup> , Thierry Cornier <sup>3</sup> ,
5	Renaud Scheifler <sup>1#</sup> and Francis Raoul <sup>1#</sup>
6	
7	<sup>1</sup> Laboratoire Chrono-environnement, UMR CNRS 6249 UsC INRA, Université
8	Bourgogne Franche-Comté, 16 route de Gray, 25030 Besançon cedex, France
9	<sup>2</sup> Conservatoire Botanique National de Franche-Comté, Observatoire Régional des
10	Invertébrés, 7 rue Voirin, 25000 Besançon, France
11	<sup>3</sup> Centre régional de phytosociologie agréé Conservatoire Botanique National de Bailleul,
12	Hameau de Haendries, F-59270 Bailleul, France
13	
14	* Corresponding author: Shinji Ozaki
15	Phone number: +33 (0)3 81 66 65 98
16	E-mail address: shinji.ozaki@univ-fcomte.fr
17	# both authors contributed equally to supervising this work
18	

19 Abstract:

20 Mammals are mainly exposed to trace metals via consuming contaminated food. Several 21 studies have demonstrated relationships between metal concentrations in food and in animal 22 tissues. However, potential effects of trace metals on feeding behaviour of wildlife have been 23 poorly documented under field conditions, despite experimental evidence showing that food 24 selection is impacted by resource contamination. Here we test the hypothesis that the diet of 25 a generalist rodent, the wood mouse (Apodemus sylvaticus), is altered by soil trace metal 26 contamination in the field. Wood mice were sampled in spring and in autumn along a 27 gradient of soil contamination in the surroundings of a former smelter located in northern France. Available resources in the field were inventoried and the diet of the animals was 28 29 analysed using DNA "metabarcoding". We demonstrated that (i) relationship between the 30 resource richness in the diet and their richness in the field was altered by soil metal 31 contamination. Wood mice specialised their diet along the gradient of soil metal 32 contamination for both plant and invertebrate resources in spring. We also showed that (ii) 33 preference for Salicaceae, a plant family accumulating metals, decreased when soil 34 contamination increased. These results suggest that environmental trace metal pollution 35 could act as a force modulating trophic interactions in terrestrial food webs, thereby affecting 36 wildlife exposure to contaminants by trophic route.

37

#### 38 Keywords:

39 Biodiversity; Diet diversity; Food selection; Trace metals; Apodemus sylvaticus

#### 41 Introduction

42 Feeding behaviour is determined by life history traits and life stages of animals (Schoener, 43 1971; Pyke et al., 1977), abundance and distribution of food resources in nature (White, 44 2008; Crampton et al., 2011), and food quality (i.e. nutrient and toxin content) (Butet, 1986a; 45 Kerley & Erasmus 1991; Schmidt, 2000; Lewis et al., 2001). All these factors are driven by 46 ecological determinants like season, climate, community interactions or landscape features 47 (Polis et al., 1997; Visser & Both 2005; White, 2008). The optimal foraging theory assumes 48 that animals consume resources providing high energy intake per unit time (Stephens & 49 Krebs, 1986) and that the diet of animals tends to be specialized when food density is high (MacArthur & Pianka, 1966). However, animals require not only energy but also nutrients 50 51 like vitamins or essential metals. The nutrient requirements can thus hardly be satisfied from 52 a single plant species and thus animals seek a variety of diet for nutrient balance (i.e. nutrient 53 balance hypothesis) (Westoby, 1978). Likewise, Freeland and Janzen (1974) proposed that 54 generalist mammalian herbivores are unable to detoxify large amounts of similar plant 55 secondary metabolites (i.e. those from a single plant species) and thus a variety of plant 56 sources allow them to avoid similar plant toxin load (i.e. detoxification limitation 57 hypothesis). When the use of a resource is disproportional to its availability in the field, the 58 feeding process by which an animal chooses the resource is called "selection" (Johnson, 59 1980; Manly et al., 2002). The nature of selection (i.e. preference or avoidance) reflects the 60 likelihood of a given resource for being chosen when this resource is offered with others 61 (Johnson, 1980). The food selection hence manifests a relationship between individual 62 requirements and resource constraints.

63 Trace metals (TM) are environmental pollutants of concern for both human health and 64 wildlife. They occur often as a legacy of past industrial activities. Even if the emissions of 65 many of them have been reduced in the past decades thanks to environmental regulation and

66 technical improvements at least in so-called developed countries, the pollution still persists 67 over vast surfaces due to non-degradability of metals and to their current emissions in many 68 countries (Kabata-Pendias, 2011; EMEP, 2013). TM pollution may have effects on plants and invertebrates at several levels of organisation, from sub-individual physiological effects 69 70 reducing survival or reproduction to the modification of community composition and 71 structure (Walker et al., 2012). For instance, a recent meta-analysis about the responses of 72 terrestrial biota to TM pollution -dealing with 206 pollution point sources worldwide-73 showed a decrease of diversity for most of the taxa (producers, secondary consumers and 74 decomposers) (Kozlov & Zvereva, 2011). In addition, Dazy et al. (2009) demonstrated that 75 plant species richness correlated negatively to TM concentrations in soils in eastern France. 76 Metal pollution can also reduce species richness in earthworm communities (Spurgeon & Hopkin, 1999; Nahmani et al., 2003) and impact the structure and composition of soil macro-77 78 fauna communities (Nahmani & Lavelle, 2002). Trophic transfer, i.e. the transfer of matter 79 via food consumption, is generally considered to be the main route for TM from the 80 environment to animals of higher trophic levels (Shore & Rattner, 2001; Smith et al., 2007). 81 For small mammals, several studies have demonstrated relationships between soil pollution 82 by TM, contamination of food, and subsequently, high concentrations of TM in wild animals' 83 tissues (Hunter & Johnson, 1982; Hunter et al., 1987a; Torres & Johnson, 2001; Rogival et 84 al., 2007; van den Brink et al., 2010). Avoidance of TM contaminated food can be a 85 mechanism that mitigates organisms' exposure to contaminants (Gillet & Ponge, 2003). This 86 has been reported for the wood mouse (Apodemus sylvaticus) under laboratory settings but 87 not confirmed in natural conditions (Beernaert et al., 2008). To the best of our knowledge, 88 no study has reported variations of richness in the diet and selection as a response to both 89 the variation of resource availability and the contamination of the environment in natura in 90 terrestrial ecosystems.

91 One of the main difficulties in assessing wildlife diet is the identification of items actually 92 consumed. Identifying items with the highest taxonomic resolution is almost impossible with 93 classical macro- or microscopy-based methods, which then makes it difficult to match diet 94 with field records of resources. Recently, DNA-based techniques have arisen as a powerful 95 approach for the taxonomic identification. Although DNA-based identification is not novel, 96 current technological developments such as DNA metabarcoding approach enable to 97 simultaneously identify different taxa in environmental samples, like soils or faeces, 98 containing multiple specimens and potentially highly degraded DNA (reviewed in Valentini 99 et al., 2009a; Taberlet et al., 2012; Pompanon et al., 2012). The efficiency of this approach 100 for analysing the diet has been demonstrated in various species including herbivorous 101 mammals, birds and invertebrates (Soininen et al., 2009; Valentini et al., 2009b), 102 insectivorous bats (Bohmann et al., 2011), or omnivorous animals like the brown bear (De 103 Barba et al., 2013).

104 In the present study, we focused on the wood mouse (Apodemus sylvaticus), which is a 105 generalist small mammal (Watts, 1968; Hansson, 1985; Butet, 1990) and therefore a model 106 of choice when studying diet plasticity as a response to environmental variations. Indeed, 107 provided its opportunistic feeding behaviour, the diet richness (i.e. total number of resources' 108 species in the diet) of the wood mouse is expected to be positively correlated with resource 109 richness in the field. We hypothesised that: (i) diet richness of the wood mouse would be 110 negatively correlated to TMs contamination of soils given the reported negative effects of 111 pollution on plant and invertebrate richness in community, and (ii) food selection by the 112 wood mouse would be affected by TM contamination in soils possibly due to avoidance 113 behaviour.

114

#### 115 Materials and methods

116 Study sites

117 This study was carried out in the surroundings of the former lead (Pb) and zinc (Zn) smelter 118 named "Metaleurop Nord" located in northern France (Noyelles Godault, Hauts-de-France, 119 France). The surrounding soils were highly contaminated by TM, mainly cadmium (Cd), Pb, 120 and Zn, over a large area of about 120 km<sup>2</sup> because of metal-contaminated dust released for 121 decades from this smelter and another one named "Umicore", located about 4 km east of the 122 first (Sterckeman et al., 2002; Douay et al., 2008; Douay et al., 2009; Waterlot et al., 2016). A study area of 40 km<sup>2</sup> was defined and divided into 160 sites of 25 ha (500m x 500m) 123 124 (Fritsch et al., 2010), from which seven are used in the present study (see below). Total TM concentrations in the soil of woody patches ranged from 0.1 to 236.5 mg kg<sup>-1</sup> of dry soil for 125 Cd, from 16 to 7331 mg kg<sup>-1</sup> of dry soil for Pb, and from 44 to 7264 mg kg<sup>-1</sup> of dry soil for 126 Zn in an area of 40 km<sup>2</sup> in the surroundings of the former smelter (Fritsch et al., 2010). These 127 128 values largely exceed background TM concentrations in soils of the region Hauts-de-France 129 (Sterckeman et al., 2007). Those TM concentrations in woody patches were used in our 130 modelling (cf. below). The seven sites are located along a soil pollution gradient and 131 composed of three types of dominant habitats (woodland, urban features, arable lands) to 132 provide a wide range of resource richness in the field (for details see Supporting Information 133 SI1).

134

#### 135 **Rodent trapping**

At each site, we trapped wood mice in spring (April) and in autumn (September and October) 2012 according to Fritsch et al. (2011) and in accordance with current French legislation about ethics and use of animals in research. Briefly, small break-back traps were used with peanut as rodent's bait. Ten lines, composed of 10 traps 3 m spaced each, were used per site and per season. All trap lines were set in woody habitats of the sites and their location was 141 geo-referenced. The trap lines were checked in the morning for three consecutive days and 142 re-set and/or re-baited, if necessary. After being weighed in the field, animals were 143 immediately frozen at -20°C for further analyses.

Two hundred and forty-six rodents (117 in spring and 129 in autumn; 116 females and 127 males) were used in the study. Age was estimated using body weight according to four classes (approximately, class I < 50 days < class II < 130 days < class III < 230 days < class IV), referring to Vandorpe & Verhagen (1980) and Tête et al. (2014). Age structure differed between seasons: no mouse of age class I was found in spring, and the proportions of animals in the other 3 classes were significantly different ( $\chi^2 = 31.04$ , p-value < 0.001) such that animals in spring were older than in autumn.

151

#### 152 Stomach content samples

153 Consumed items were identified by biomolecular analysis of stomach contents (SC). Wood 154 mice were thawed at room temperature. Their SC was extracted from the body with a spatula, 155 which was thoroughly cleaned with disposable tissue, then washed with ultra-pure water 156  $(18.2 \text{ M}\Omega/\text{cm}^2)$  and wiped off with other tissue between each extraction. Remaining bait was 157 removed from SC. Each SC was then homogenized and split into two aliquots: one (about 158 10 mg) was stored in 95% ethanol for metabarcoding analysis (see below), the other was 159 reserved for another study. For 13 mice among the 246 rodents, faeces taken from intestines 160 were used as an alternative material for food identification because their stomach was empty. 161 The 13 mice were equitably distributed according to the explanatory factors tested in this 162 study.

163

#### 164 Metabarcoding analysis

165 The DNA extraction, amplification and the PCR purification were performed at SPYGEN

166 facilities (www.spygen.com). DNeasy Blood and Tissue Kit (Qiagen GmbH) was used for extracting total DNA, following the manufacturer's instruction. DNA amplification was 167 168 performed with three sets of primers (Table 1): the P6 loop of the chloroplast trnL (UAA) 169 intron g/h (Taberlet et al., 2007) was used for identifying general plant species; primer 170 targeting a short fragment of mitochondrial 16S gene (16S mtDNA) was used for arthropods 171 and molluscs DNA (16SMAV-F/16SMAV-R) (De Barba et al., 2013); and primer targeting 172 short region of 16S mtDNA for earthworms (ewD/ewE) (Bienert et al., 2012). PCR 173 amplifications were carried out on Applied Biosystems Veriti 96 Wells (Life Technologies). 174 The amplification was realized in a final volume of 25 µL using 3 µL of DNA extract. Two 175 PCR replicates were performed per each sample. For the amplification of arthropod and 176 mollusc DNA, 2 µM of a blocking primer for mammal's DNA (MamMAVB1; De Barba et 177 al., 2013) was added in the PCR mix. The amplification mixture contained 1 U of AmpliTaq 178 Gold DNA Polymerase (Applied Biosystems), 10 mM of Tris-HCl, 50 mM of KCl, 2.5 mM 179 of MgCl<sub>2</sub>, 0.2 mM of each dNTP, 0.2 µM of group-specific primers, 0.2 µg/µL of bovine 180 serum albumin (BSA, Roche Diagnostic) and ultra-pure water (18.2 M $\Omega$  /cm<sup>2</sup>) to bring each 181 sample to the final volume. The mixture was denatured at 95°C for 10 min, followed by 45 182 cycles of 30 s at 95°C, 30 s at 50°C for trnL-g/h, at 55°C for 16SMAV-F/16SMAV-R and at 183 58°C for ewD/ewE and 1 min at 72°C, followed by a final elongation at 72°C for 7 min. 184 Extraction and PCR negative controls were analysed in parallel in order to monitor potential 185 contamination. After amplification, the samples were titrated using capillary electrophoresis 186 (QIAxcel; Qiagen GmbH) and purified using a MinElute PCR purification kit (Qiagen 187 GmbH). Before sequencing, purified DNA was titrated again using capillary electrophoresis. 188 The purified PCR products were pooled in equal volumes, to achieve an expected sequencing 189 depth of 10,000 reads per sample. Libraries were prepared using TruSeq Nano DNA genomic 190 kit (Illumina) and a pair-end sequencing (2x100 bp) was carried out with an Illumina HiSeq sequencer (Illumina) using TruSeq SBS Kit v3 (Illumina) following the manufacturer's
instructions. Library preparation and sequencing were performed at Fasteris facilities
(www.fasteris.com).

194 Reads were handled by the software Mothur pipeline (Schloss et al., 2009). Briefly, forward 195 and reverse reads were assembled in contig sequences. Then, sequences were filtered based 196 on length (20-90bp for trnL-g/h; 36-38bp for 16SMAV; 69-81bp for ewD/ewE), homo-197 polymer (less than 10 nucleotides) and no ambiguous nucleotides. After de-replication 198 (occurrence count of each different sequence), only unique sequences with a minimum count 199 of 10 (sum of all samples) were used for further analysis.

200

# 201 Field inventory of plant and invertebrate resources

Vegetation survey was realized once per site from 4<sup>th</sup> June to 5<sup>th</sup> September 2012. Three 202 203 different strata were defined based on height: tree stratum (woody species > 8 m high), shrub 204 stratum (woody species < 8 m high), and herbaceous stratum. Taxa were identified at species 205 level in the field. Nomenclature of species followed Lambinon et al. (2004) and Dudman & 206 Richards (1997). Cover-abundance of vascular plant species was visually estimated as the 207 vertically projected area following Braun-Blanquet et al. (1952). Vegetation habitats were 208 determined by plants' composition of one or more strata and delineated as polygons with the 209 aid of aerial pictures. The polygons were geo-referenced and digitalized using the QGIS 210 software (http://www.qgis.org/). We listed 236 different plant taxa in the field (see SI1 for 211 the taxonomic richness in the field per site and SI2 for the list of the taxa and their abundance 212 per site).

Invertebrate fauna was sampled at the same time as the rodent trapping (i.e. in the two seasons). Ground-dwelling invertebrates were captured near rodent traps using three pitfall traps per line. Flying invertebrates were captured with yellow pan traps containing a mix of

216 soap and water. Four yellow pan traps were set in each site. Yellow colour attracts especially 217 flower visiting insects. The traps were checked every morning for three consecutive days. 218 All traps were geo-referenced. Captured invertebrates were conserved in ethanol or in freezer 219 at -20°C and identified to the highest possible taxonomic resolution by morphological 220 characteristics (the main references are Coulon, 2003; Forel & Leplat, 2001; Jeannel, 1941; 221 Trautner & Geigenmueller, 1987). Captured ground-dwelling and flying invertebrates 222 consisted of 88 and 95 taxa, respectively (see SI1 for the taxonomic richness in the field per 223 site and SI2 for the list of the taxa and their abundance per site). Because of the different 224 taxonomic resolution, we used mainly family level (and higher levels) for invertebrate 225 richness in the field (cf below): 24 and 95 families (or higher) in ground-dwelling and flying 226 invertebrates, respectively.

227 We then estimated available resources for wood mice near their trapping location. Although 228 notion of "availability" is distinct from "abundance" in selection studies (Manly et al., 2002), 229 we considered them equivalent for wood mice given its generalist diet behaviour. Plant 230 resource availability was estimated by the cover-abundance of each plant species found 231 within a buffer of 10 m around trap lines (referred to as "buffer" thereafter). This buffer is equivalent to an area of about 1000 m<sup>2</sup>, considered as the average size of wood mouse vital 232 233 domain (Quéré & Le Louarn, 2011). We considered the data set collected between June and 234 September to be representative of food availability in spring and autumn. Since the traps 235 used to capture invertebrates (pitfall trap and yellow pan trap) were inappropriate for 236 catching earthworms and molluscs, both earthworm and mollusc richness in the field were 237 unknown. Resource availability of ground-dwelling invertebrates in buffers both in spring 238 and in autumn was estimated by the abundance of each taxa found per trap line (sum of the 239 three traps per line), whereas availability of flying invertebrates was based on abundance in 240 the yellow pan trap nearest to the trap line.

241

#### 242 Diet diversity analyses

## 243 <u>Sequence data for diet diversity analyses</u>

244 Diet diversity analyses were performed by employing Molecular Operational Taxonomic 245 Units (MOTUs): groups of DNA sequences clustered according to similarity, which are thus 246 independent of any reference database (Sun et al., 2012). Sequences were clustered for each 247 primer with average neighbour algorithm using Needlman-Wunsch distance. Distances 248 cutting clusters were arbitrarily chosen as 0.032 for sequences obtained from primer trnL-249 g/h, as 0.042 for 16SMAV and as 0.034 for ewD/ewE (see SI3 for sequence data of plant, 250 arthropod and earthworm MOTUs). Deagle et al. (2013) showed that the number of 251 sequences obtained from degraded DNA is likely not proportional to the biomass of each 252 taxa really eaten. The quantitative sequence information data were therefore converted into 253 presence/absence of MOTUs using an occurrence threshold of 100 to remove background 254 sequencing data.

255

#### 256 *Diet richness and modelling*

257 Richness in the diet of each mouse was calculated as the total number of MOTUs per mouse 258 by three types of food: plant, arthropod and mollusc (referred to as "arthropod" thereafter), 259 or earthworm. We assumed that plants potentially consumed by the invertebrates would not 260 artificially increase the number of plant MOTUs consumed by the rodent (i.e. there may be 261 no secondary predation, for detail see Harwood et al., 2001) because the number of plant 262 MOTUs was not significantly different between SCs containing or not containing 263 invertebrate food (arthropod and/or earthworm) (data not shown). The diet richness per 264 mouse was compared among seasons, wood mouse sexes and age-classes, and landscape 265 features using the nonparametric Wilcoxon-Mann-Whitney (for comparison of two categories) or Kruskal-Wallis tests (for more than two categories), due to skewed distribution
of MOTUs data. Frequency of occurrence was also compared by chi-square goodness-of-fit
test.

269 Relationship between diet richness, richness in the field and TM concentrations in soil was 270 assessed by statistical modelling. In order to estimate the richness in the diet of all mice in a 271 buffer, the total number of MOTUs of all mice captured over a given buffer (referred to as 272 "diet richness" thereafter) was calculated, per type of food or as total diet richness by 273 summing plant, arthropod and earthworm MOTUs. The relationship was analysed using 274 multimodel inference on the basis of the information-theoretic approaches (Burnham & 275 Anderson, 2002). A set of candidate models was established by combinations of several 276 explanatory variables. Second order Akaike Information Criterion (AICc: corrected AIC for 277 small sample size in relation to the number of model parameters) was used for ranking the 278 proposed models. Probability of the "best" model over the set was estimated by Akaike 279 weight (w). In situations where the best model was uncertain (i.e. w of the first ranked model 280 < 0.9), we applied a model averaging approach for reducing model selection uncertainty. A 281 confidence set of candidate models, whose  $\Delta AICc < 6$  (difference of AICc from the first 282 ranked model < 6, which is equivalent to a likelihood ratio from the first ranked model > 283 0.05), was re-selected, and each parameter of those re-proposed models was averaged by re-284 calculated Akaike weight (Burnham & Anderson, 2002). When a given parameter was not 285 included in a proposed model, the value of the parameter was considered as 0 in this model 286 (Burnham & Anderson, 2002; Lukacs et al., 2010).

All candidate models were established as generalized linear models (GLM) with logarithm link function for Poisson distribution. Models were built separately per season, given potential seasonal diet variation of the animal (Butet, 1986b; Abt & Bock, 1998), as well as per TM, considering their different effects on organisms and transfer pathways (Eisler, 2000). We used the diet richness in buffer as response variable and number of analysed mice per trap line as one of the explanatory variables. Soil TM concentrations of woody patch characterised by Fritsch et al. (2010) were applied to buffer set in or near a given woody patch. Mice of age class I were discarded from modelling because diet of juveniles could differ from adults (Watts, 1968; Montgomery & Montgomery, 1990). The modelling was carried out on 38 buffers in spring and 37 buffers in autumn. Number of mice used in the analysis was 181 and their number per line ranged from 1 to 6.

298 For plant richness in the diet, combinations of three variables and one interaction were 299 considered as candidate models: number of mice, TM concentrations in soils of woody 300 habitat (referred by the data of Fritsch et al., 2010; see "Study site" above) with a natural 301 logarithm transformation, and plant richness in the field, as well as an interaction between 302 the TM concentrations and the plant richness in the field (i.e. 10 models including the 303 intercept only model). For arthropod richness in the diet, four variables and two interactions 304 were used: mouse number, TM concentrations, both ground-dwelling and flying invertebrate 305 richness in the field, as well as two interactions between the TM concentration and the 306 richness in the field (i.e. 26 models). For earthworm richness in the diet, we used only two 307 variables: mouse number and TM concentrations (i.e. four models). For total diet richness, 308 we used three variables and one interaction: mouse number, TM concentrations, and total 309 richness in the field (sum of plant and both ground-dwelling and flying invertebrate richness 310 at family level), as well as interaction between the TM concentrations and the total richness 311 in the field (i.e. 10 models). Overdispersion (observed variance higher than theoretical one) 312 was checked according to Cameron & Trivedi (1990). Explanatory value of our final models were estimated by deviance  $R^2$  ( $R^2_D$ : proportion of deviance explained by the given model) 313 314 (Zuur, 2009).

#### 316 Food selection analyses

## 317 Sequence data for selection analysis and association with field survey data

318 Reference sequences of the species recorded in our study area were taken from the GenBank 319 sequence database (www.ncbi.nlm.nih.gov/genbank). Each sequence extracted from SCs 320 (i.e., query sequence) was compared to the reference sequences for checking their similarity, 321 where we accepted difference of only one nucleotide. When query sequences could match with several reference sequences, they were gathered into one group composed of the 322 323 corresponding species, called "sequence group (Grp)" (see SI4 for details of sequence 324 groups). Non assigned query sequences were excluded. Query sequences with occurrence 325 lower than 100 were not considered. Food Grp data were finally converted into 326 presence/absence. We represented Grps using both ID number and family name(s) 327 corresponding to their component species. We applied such method only to plant food 328 because almost all invertebrate resources in the field were identified to family level only.

329

# 330 *Plant food selection and modelling*

331 Frequency of occurrence of each plant food item (i.e. percentage of number of occurrences 332 of a given item compared to the total number of occurrence of all items, referring to Klare 333 et al., 2011) was calculated at family level using Grp. Food selection was assessed, for each 334 plant food item, per season, comparing the frequency of occurrence and relative proportion 335 of resources in the field (i.e. cover-abundance of a given plant food of all strata compared to 336 the total cover-abundance of all plants of all strata). Plant resource taxa in the field were 337 gathered into corresponding Grp and then into the same plant family. Resource availability 338 was estimated based on the abundance of the plant families in each buffer, summed up per 339 family for all mice in each season. Significance of food selection was evaluated per plant 340 family using Bailey's confidence intervals at p-value < 0.05 (Bailey, 1980; Cherry, 1996):

when relative availability of a given resource was located below or above the confidence
interval of its frequency of occurrence, this resource is considered as preferred or avoided,
respectively.

344 Preference change according to richness in the field and/or TM concentrations in soil was 345 then assessed by multimodel inference approaches. First, food selection was estimated in 346 each buffer as described above. Buffers where selection for a given resource was observed 347 were considered 1 (i.e. selection for the resource is present), whereas the other buffers were 348 considered 0 (i.e. selection for the resource is absent). We used GLM with logit link function 349 for assessing change of proportion of buffers where preference was observed. There were 350 only one plant family (Salicaceae) for which selection was significantly observed in more 351 than five buffers, allowing statistical analyses. Salicaceae was preferred in spring in six 352 among 38 buffers. Candidate models were established by combinations of three variables 353 and one interaction: mice number in buffer, plant richness in the field, and TM concentrations, 354 as well as an interaction between the richness in the field and the TM concentrations. The 355 modelling was performed for each TM. Model selection methods were the same as described 356 above.

All statistical analyses were computed using the statistical software R (ver. 3.4.2; R Development Core Team). GLM was handled by "glm" function in "lme4" package and model averaging by "model.avg" function in "MuMIn" package. Overdispersion checking was carried out by "dispersiontest" function in "AER" package.

361

362 **Results** 

## 363 Resources richness in the field, trace metals and diet richness

Plant items occurred in the diet of 96.3% of wood mice, arthropod items in 65.9% and
earthworm items in 29.3%. The occurrence of earthworm items was significantly lower in

forest landscape and significantly higher in urban landscape ( $\chi^2 = 7.619$ , p-value = 0.022; for details see SI5). The variations of the total richness in the diet and of the relative proportion of the three food item categories (plants, arthropods and earthworms) along the soil contamination gradient are shown in SI6 and SI7, respectively.

Number of plant MOTUs per mouse ranged from 0 to 7 and median was 2 MOTUs. It was significantly higher in autumn than in spring (W = 5394.5, p-value < 0.001). Number of arthropod MOTUs ranged from 0 to 9 and median was 1 MOTUs. In autumn, the number was significantly lower in age class I than in class II (Kruskal-Wallis  $\chi^2 = 7.972$ , p-value = 0.047) (N.B. no mouse of class I was found in spring). Number of earthworm MOTUs ranged from 0 to 3 and median was 0 MOTUs. The number was significantly higher in urban area than in forest area in spring only (Kruskal-Wallis  $\chi^2 = 8.068$ , p-value = 0.018) (see SI8).

No single "best" model was evidenced and thus model averaging was applied for all sets of 377 378 models. Complete results of modelling are shown in SI9. The final model for plant richness 379 in the diet in buffer included mouse number, soil TM concentrations, richness in the field 380 and the interaction between the last two for the three TM in the two seasons. Figure 1a and 381 1b show plant richness in the diet according to plant richness in the field and Cd 382 concentration in soils in spring, respectively. The pattern was similar for the other metals. Influence of richness in the field on the diet richness was tiny in spring (Figure 1a), whereas 383 384 diet richness decreased according to TM in soils regardless of the richness in the field (e.g. from 6 to 4 plant MOTUs along the range of TM concentration; Figure 1b). R<sup>2</sup><sub>D</sub> was 0.301, 385 386 0.313 and 0.307 for Cd, Pb and Zn, respectively. Such tendency changed in autumn (Figure 387 1c and 1d for Cd, similar for other TM). Diet richness was negatively correlated to plant 388 richness when soil contamination was low, while this correlation disappeared, or even 389 reversed, when TM concentration increased (Figure 1c). A similar tendency was predicted 390 for the relationship between diet richness and TM contamination in soils (Figure 1d): diet richness was negatively correlated to TM concentration when plant richness was low and positively correlated when the plant richness was high.  $R^2_D$  was 0.624, 0.492 and 0.601 for Cd, Pb and Zn, respectively.

394 The model for invertebrate richness in the diet included all proposed explanatory variables 395 in the two seasons, except the interaction between TMs in soils and richness of ground-396 dwelling invertebrates for the models for Cd and Zn in autumn. In spring, arthropod richness 397 in the diet increased according to richness of both ground-dwelling and flying invertebrates 398 when soil contamination was low. This correlation disappeared, then reversed, when TM 399 concentration in soils increased. For example, our model for Cd predicted that arthropod 400 MOTUs increased from 2 to 5 along the gradient of ground-dwelling invertebrate richness 401 in the field when Cd concentration in soils was low, whereas arthropod MOTUs decreased 402 from 5 to 3 when Cd concentration was high (Figure 2a; the pattern was similar for flying 403 invertebrates). The relationship between arthropod richness in the diet and TM concentration 404 in soils followed the same tendency (Figure 2b). In autumn, arthropod richness in the diet 405 was explained almost only by mouse number. Neither TM concentration in soils nor richness 406 of both types of invertebrates had sensible influence on diet richness (Figure 2c and 2d).  $R^{2}_{D}$ 407 for Cd, Pb and Zn was respectively 0.502, 0.472 and 0.495 in spring, and 0.495, 0.504 and 408 0.497 in autumn.

The averaging model for earthworm richness in the diet included mouse number and TM concentrations in soils in both seasons. Earthworm richness in the diet slightly decreased along TM gradient (e.g. difference of about 0.5 MOTUs; Figure 3a and 3b), and mouse number on trap line had a larger influence in autumn than in spring.  $R^2_D$  for Cd, Pb and Zn was respectively 0.101, 0.056 and 0.075 in spring, and 0.169, 0.158, 0.192 in autumn.

414

#### 415 **Plant food selection**

We found 48 Grps in the food of the 246 mice. Sapindaceae, Salicaceae, Cornaceae, Adoxaceae, Asteraceae, and Rosaceae were plant taxa frequently eaten in our area (see SI10 for details). Selection for some plant families was observed at the scale of all study sites together and at the two seasons: preference for Fagaceae and avoidance of Urticaceae. Furthermore, selected plant resources changed between the two seasons, for example preference for Salicaceae, Sapindaceae and Rosaceae in spring and preference for Poaceae, Adoxaceae, Asteraceae and Cornaceae in autumn (see SI11 for details).

423 Model averaging was used for the final model for Salicaceae preference in spring (see SI12 424 for complete results). The model included all variables: mouse number, plant richness in the 425 field, TM concentration in soils and the interaction. Preference for Salicaceae was negatively 426 correlated to plant richness in the field when soil contamination was low (e.g. from 0.6 to 427 0.4; Figure 4a). On the other hand, preference for Salicaceae disappeared along the gradient 428 of TM concentration in soils, whatever plant richness in the field was (e.g. preference 429 probability for Salicaceae decreased from 0.6 to 0; Figure 4b). R<sup>2</sup><sub>D</sub> of the final model was 430 0.280, 0.165 and 0.271 for Cd, Pb and Zn, respectively.

431

## 432 Discussion

433 We first hypothesised that soil TM contamination would reduce diet richness of wood mouse 434 due to adverse effect of TM on resources' richness in the field. Our study revealed negative 435 correlations between diet richness and TM in soils. This was true for both earthworms and 436 plants in spring and in autumn (although only at low and medium levels of plant richness in 437 autumn), and for arthropods in spring when richness in the field was high. On the contrary, 438 positive correlations were observed, for plants in autumn when plant richness in the field 439 was high, and for arthropods in spring when arthropod richness in the field was low. A 440 negative relationship between diet richness and pollution by TM is expected if resource

441 richness in the field is negatively affected by TM concentrations. Literature about the effects 442 of pollution on various taxonomic groups suggests a global negative effect of pollution on 443 the richness of organisms, and an absence or a positive correlation for a few taxa groups or 444 situations. Although decreasing richness of earthworms with increasing TM concentrations 445 gradients in soils has been widely reported (Spurgeon & Hopkin, 1996, Spurgeon & Hopkin, 446 1999; Pérès et al., 2011), several studies have reported that relationships between species 447 richness of soil macro-fauna and TM contamination is not straightforward (Nahmani & 448 Lavelle, 2002; Gillet & Ponge, 2003; Migliorini, et al., 2004). Moreover, several studies 449 showed that community composition was affected by soil TM concentrations. Tolerance to, 450 and/or accumulation capacity of metals differ according to taxa, organs, and metals (Das et 451 al., 1997; Broadley et al., 2007; Pourrut et al., 2011). Lower resistance to toxic effects can 452 make metal sensitive plants disappear in contaminated areas, which may favour metal 453 tolerant ones, sometimes without modification of taxonomic richness. Such modification of 454 community composition has been reported by several authors in plants (Banásová et al., 455 2006; Strandberg et al., 2006; Dazy et al., 2009) and in invertebrates (Nahmani & Lavelle, 456 2002; Migliorini et al., 2004; Babin-Fenske & Anand, 2011). In our study area, we found no 457 correlation or even positive trends between TM concentration in soils and both plant and 458 invertebrate richness in the field (unpublished data). However, there were confounding 459 factors in our site like the anthropogenic increase of habitat diversity (see Douay et al., 2009) 460 and possibly of plant richness along the gradient of contamination on the Metaleurop Nord 461 site. Community composition also differed according to soil contamination levels or other 462 soil properties affecting metal bioavailability (unpublished data). Difference in resource 463 composition could alter resource availability and thus modify feeding behaviour and diet 464 composition. Few studies documented the effects of pollution on the diet of terrestrial 465 vertebrates, most of them dealing with diet composition rather than richness or diversity.

466 Clare et al. (2014), describing the diet of insectivorous bats Myotis lucifugus in different 467 locations, showed a trend of lower diet species richness in area of lower habitat quality 468 (acidification or organic pollution). Eeva et al. (2005) showed a significant difference in diet 469 composition of nestlings of two insectivorous birds, Parus major and Ficedula hypoleuca, 470 between unpolluted and TM polluted sites, reflecting differences in resources composition, 471 phenology and feeding ecology among the two bird species: in polluted areas where 472 caterpillars were less abundant, both species ate more beetles and different larvae of flying 473 insects but less moth. The study of Heroldová (2002) demonstrated that the diet of field vole 474 Microtus agrestis in forest clearings caused by air pollution was dominated by grasses, 475 reflecting higher abundance of grasses and a poorer diversity of food supply in clearings. 476 Thus, it is suggested that diet richness (as demonstrated in the present study) and/or 477 composition can be affected by pollution induced changes in trophic resources.

478 We also hypothesized that TM contamination would affect food selection by wood mouse. 479 Our model predicts that preference probability for Salicaceae in spring in the highest 480 contaminated sites decreases to become almost null. Avoidance of TM contaminated soil or 481 food has been shown in invertebrates but has rarely been investigated in vertebrates, and is 482 particularly hard to prove under field conditions in comparison with controlled experiments. 483 This behaviour has been observed under laboratory conditions for some invertebrates like 484 isopods or small moths (Odendaal & Reinecke, 1999; Zidar et al., 2004; Scheirs et al., 2006), 485 and for the wood mouse (Beernaert et al., 2008), but mechanisms of such aversion are not 486 yet elucidated. In Beernaert et al. (2008), wood mice chose acorns (Quercus robur, 487 Fagaceae) sampled from unpolluted sites rather than ones from metal polluted sites (mainly 488 by Cd, copper, Pb, and Zn), but such avoidance behaviour was no longer detected under field 489 conditions, perhaps due to ecological co-factors such as food shortage, and/or the presence 490 of competitors and predators. Avoidance of contaminated food, leading to shifts in diet, can 491 be one of the mechanisms allowing animals to mitigate their exposure in highly polluted 492 environments (Gillet & Ponge, 2003). Actually, Salicaceae family is considered as TM 493 accumulator and resistant to highly TM contaminated soils (Pulford & Watson, 2003). 494 Migeon et al. (2009) showed that Salicaceae family in the surroundings of Metaleurop Nord, 495 composed of two genera Salix and Populus, accumulated larger amount of Cd and Zn in their 496 shoot parts than the other families like Fagaceae or Sapindaceae. These two last are also 497 preferred plant families in our study and known as common food of wood mice (i.e. Watts, 498 1968). However, mechanisms of changes in preference remains unclear. The nutritional 499 quality of metal contaminated plant organs has been shown to be lowered, with for instance 500 a decrease of sugar or protein concentrations (Scheirs et al., 2006; Beernaert et al., 2008), 501 features which play an important role for food choice in rodents (Butet, 1990; Kerley & 502 Erasmus, 1991). It is also possible that animals might identify metal contaminated plants due 503 to metal-induced metabolic compounds (Poschenrieder et al., 2006), largely known as 504 compounds limiting feeding (Foley & Moore, 2005). Dearing et al (2000) reported that plant 505 secondary compounds might play more important role than nutrient level in the diet of the 506 generalist woodrat, Neotoma albigula. It is therefore supposed that mice could directly or 507 indirectly detect noxious substances (plant toxins or TM) and avoid ingesting such items. 508 Also, Behmer et al. (2005) demonstrated from their experiments using nutritionally balanced 509 synthetic food that high Zn concentration per se altered the feeding behaviour of the desert 510 locust Schistocerca gregaria. Whatever the mechanism, the hyperaccumulation of metals in 511 plant tissues contribute to defensive effects of plants against natural enemies, including 512 invertebrate herbivory (Poschenrieder et al., 2006; Boyd, 2007). Such defensive effect has 513 not been reported against vertebrates so far, but the disappearance of the preference for 514 Salicaceae family along TM contamination in soils could be associated to its high metal 515 accumulation capacity. Furthermore, plant richness in the diet in autumn increased along the

516 soil TM concentration gradient when richness in the field was high. Apart from changes of 517 resource composition, this pattern could be explained, at least partly, by the nutrient balance 518 hypothesis or the detoxification limitation hypothesis: if nutritional quality of all plant 519 resources was altered (i.e. if nutritional quality was not clearly contrasted among resources) 520 diet richness of animals could increase for completing nutrient and/or for avoiding loads of 521 similar secondary compounds.

522 Contrary to plants, selection for invertebrate food could not be analysed in part because of 523 the primer used for arthropods and molluscs: its short size is more reliable for the analysis 524 of degraded DNA in the case of diet (Taberlet et al., 2012) but the precision in taxonomic 525 assignment was low. We also did not sample earthworms in the field. Moreover, it is difficult 526 to determine selection for invertebrate resources because of their mobility. Foraging strategy 527 of predators can be modified according to mobility of preys (Sih & Christensen, 2001), 528 suggesting that relative quantity in the field is not the only factor to consider when analysing 529 mobile prey selection. Even though we supposed that decreased earthworm diet richness 530 might be linked to their richness in the field, invertebrate food selection in wood mice 531 remains as a further challenge.

532 An important feature of our results is the strong seasonal difference of patterns, for both food 533 selection and correlations with TM contamination. In temperate ecosystems, seasons have a 534 key influence on the dynamics of ecosystems and on the various aspects of biology and 535 ecology of living organisms. Seasonal difference of patterns might result from the adaptation 536 of wood mice feeding behaviour to phenology of resources. Indeed, wood mice consume 537 mainly seeds and other reproductive organs (Watts, 1968; Butet, 1986a), seeking high energy 538 or protein-rich tissues (Butet, 1990). Although parts of plant resources actually consumed 539 could not be distinguished by using DNA metabarcoding, we supposed that parts of plants 540 preferably consumed have changed between spring and autumn. Moreover, the nutritional

541 composition and quality of plants and invertebrates are likely to vary according to seasons. 542 Also, resources functionally important for wood mice might have a strong phenology, such 543 as mushrooms in autumn. However, the molecular primers we used did not target mushrooms, 544 which prevents further conclusion. Finally, TM accumulation in organisms is also subject to 545 seasonality at contaminated site (i.e. Hunter et al, 1987b; Migeon et al., 2009). Seasonal 546 variation of TM accumulation pattern can be related to growth season or physiological changes of each taxa (Kabata-Pendias, 2011; Hussein et al., 2006). Disentangling the 547 548 combined effects of these factors on the patterns of interest in this study is beyond our present 549 scope, but it is clear that seasonality is a key feature of the interplay between pollution, 550 resource dynamics and consumer feeding behaviour.

551 Our study sheds light on the effects of soil TM contamination on patterns of feeding 552 behaviour of a generalist consumer, by applying DNA metabarcoding for diet identification. 553 We underline potential altering effects of TM on the diet richness, which possibly partly 554 results from modifying food selection, at least for plant resources. Such modification of diet 555 related to habitat contamination might lead to a different pattern of contaminant exposure in 556 the animals. We hence suggest that ecosystem pollution should now be considered as a driver 557 of changes in food web structure and dynamics in time and space.

558

# 559 Acknowledgements

560 This study was financially supported by the project BIOTROPH, co-funded by the *Agence* 

561 De l'Environnement et de la Maîtrise de l'Energie (ADEME; contract No.1172C0030) and

the Conseil Régional du Nord-Pas de Calais (CRNPC; orders No.12000921 and 14001044;

563 joint call with the Fondation pour la Recherche sur la Biodiversité). The first author was

- also financially supported by a grant from the *Conseil Régional de Franche-Comté* (contract
- 565 No. 2015C-06107). The authors gratefully thank Cécile Grand from ADEME for fruitful

- 566 scientific discussions. We also thank Eva Bellemain and Alice Valentini from SPYGEN
- 567 Company for their help in DNA analyses and interpretation. We finally thank François Gillet,
- 568 Nadia Crini, Dominique Rieffel and Anne-Sophie Prudent for their precious assistance.

569 **References:** 

- Abt, K. F., & Bock, W. F. (1998). Seasonal variations of diet composition in farmland field
  mice *Apodemus* spp. and bank voles *Clethrionomys glareolus*. *Acta Theriologica*, *43*,
  379–389.
- Babin-Fenske, J., & Anand, M. (2011). Patterns of insect communities along a stress gradient
  following decommissioning of a Cu–Ni smelter. *Environmental Pollution*, 159,
  3036–3043.
- 577 Bailey, B. J. R. (1980). Large sample simultaneous confidence intervals for the multinomial
  578 probabilities based on transformation of the cell frequencies. *Technometrics*, 22,
  579 583–589.
- Banásová, V., Horak, O., Čiamporová, M., Nadubinská, M., & Lichtscheidl, I. (2006). The
  vegetation of metalliferous and non-metalliferous grasslands in two former mine
  regions in Central Slovakia. *Biologia*, *61*, 433–439.
- 583 Beernaert, J., Scheirs, J., Van Den Brande, G., Leirs, H., Blust, R., De Meulenaer, B., ...
- Verhagen, R. (2008). Do wood mice (*Apodemus sylvaticus* L.) use food selection as
  a means to reduce heavy metal intake? *Environmental Pollution*, *151*, 599–607.
- 586 Behmer, S. T., Lloyd, C. M., Raubenheimer, D., Stewart-Clark, J., Knight, J., Leighton, R.
- 587 S., ... Smith, J. A. C. (2005). Metal hyperaccumulation in plants: mechanisms of
  588 defence against insect herbivores. *Functional Ecology*, *19*, 55–66.
- Bienert, F., De Danieli, S., Miquel, C., Coissac, E., Poillot, C., Brun, J.-J., & Taberlet, P.
  (2012). Tracking earthworm communities from soil DNA. *Molecular Ecology*, *21*, 2017–2030.
- Bohmann, K., Monadjem, A., Lehmkuhl Noer, C., Rasmussen, M., Zeale, M. R. K., Clare,
  E., ... Gilbert, M. T. P. (2011). Molecular Diet Analysis of Two African Free-Tailed
  Bats (Molossidae) Using High Throughput Sequencing. *PLoS ONE*, *6*, e21441.

595	Boyd, R.	S. (2	2007).	The	defense	hypothesis	of	elemental	hyperaccumulation:	status,
596	cha	alleng	ges and	new	direction	s. Plant and	Sol	il, 293, 153	—176.	

- 597 Braun-Blanquet, J., Roussine, N., & Nègre, R. (1952). Les groupements végétaux de la
  598 France méditerranéenne. CNRS: Direction de la Carte des Groupements Végétaux
  599 de l'Afrique du Nord.
- Broadley, M. R., White, P. J., Hammond, J. P., Zelko, I., & Lux, A. (2007). Zinc in plants. *New Phytologist*, *173*, 677–702.
- Burnham, K. P., & Anderson, D. R. (2002). Model selection and multimodel inference: a
   *practical information-theoretic approach* (2<sup>nd</sup> ed). New York: Springer.
- Butet, A. (1986a). Régime alimentaire d'une population de mulots sylvestres (*Apodemus sylvaticus* L., 1758), dans une lande xéro-mésophile en cours de recolonisation
  végétale. *Bulletin D'Ecologie*, 17, 21–37.
- Butet, A. (1986b). Stratégie d'utilisation et déterminisme des choix alimentaires d'un
  rongeur polyphage (*Apodemus sylvaticus*, L.): l'approche énergétique. *Acta Oecologica*, 7, 243–262.
- Butet, A. (1990). Teneur azotée des ressources et choix trophiques du Mulot sylvestre
  (*Apodemus sylvaticus*) dans un système oligotrophe. *Canadian Journal of Zoology*,
  68, 26–31.
- Cameron, A. C., & Trivedi, P. K. (1990). Regression-based Tests for Overdispersion in the
  Poisson Model. *Journal of Econometrics*, *46*, 347–364.
- 615 Cherry, S. (1996). A comparison of confidence interval methods for habitat use-availability
  616 studies. *The Journal of Wildlife Management*, 60, 653–658.
- 617 Clare, E. L., Symondson, W. O. C., Broders, H., Fabianek, F., Fraser, E. E., MacKenzie, A.,
  618 ... Reimer, J. P. (2014). The diet of *Myotis lucifugus* across Canada: assessing
  619 foraging quality and diet variability. *Molecular Ecology*, 23, 3618–3632.

- 620 Coulon, J. (2003). Les Bembidiina de la faune de France. Clés d'identification commentées
  621 (Coleoptera Carabidae Trechinae) (Première partie). *Bulletin mensuel de la Société*622 *linnéenne de Lyon*, 72, 256–272.
- 623 Crampton, L. H., Longland, W. S., Murphy, D. D., & Sedinger, J. S. (2011). Food abundance
  624 determines distribution and density of a frugivorous bird across seasons. *Oikos*, *120*,
  625 65–76.
- Das, P., Samantaray, S., & Rout, G. R. (1997). Studies on cadmium toxicity in plants: a
  review. *Environmental Pollution*, *98*, 29–36.
- Dazy, M., Béraud, E., Cotelle, S., Grévilliot, F., Férard, J.-F., & Masfaraud, J.-F. (2009).
  Changes in plant communities along soil pollution gradients: Responses of leaf
  antioxidant enzyme activities and phytochelatin contents. *Chemosphere*, 77, 376–
  383.
- De Barba, M., Miquel, C., Boyer, F., Mercier, C., Rioux, D., Coissac, E., & Taberlet, P.
  (2013). DNA metabarcoding multiplexing and validation of data accuracy for diet
  assessment: application to omnivorous diet. *Molecular Ecology Resources*, *14*, 306–
  323.
- Deagle, B. E., Thomas, A. C., Shaffer, A. K., Trites, A. W., & Jarman, S. N. (2013).
  Quantifying sequence proportions in a DNA-based diet study using Ion Torrent
  amplicon sequencing: which counts count? *Molecular Ecology Resources*, *13*, 620–
  633
- Dearing, M. D., Mangione, A. M., & Karasov, W. H. (2000). Diet breadth of mammalian
  herbivores: nutrient versus detoxification constraints. *Oecologia*, *123*, 397–405.
- Douay, F., Pruvot, C., Waterlot, C., Fritsch, C., Fourrier, H., Loriette, A., ... Scheifler, R.
  (2009). Contamination of woody habitat soils around a former lead smelter in the
  North of France. *Science of The Total Environment*, 407, 5564–5577.

- 645 Douay, F., Pruvot, C., Roussel, H., Ciesielski, H., Fourrier, H., Proix, N., & Waterlot, C.
- 646 (2008). Contamination of Urban Soils in an Area of Northern France Polluted by
  647 Dust Emissions of Two Smelters. *Water, Air, and Soil Pollution, 188,* 247–260.
- Dudman, A. A., & Richards, A. J. (1997). Dandelions of Great Britain and Ireland,
   Handbook 9. London: Botanical Society of the British Isles.
- Eeva, T., Ryömä, M., & Riihimäki, J. (2005). Pollution-related changes in diets of two
  insectivorous passerines. *Oecologia*, 145, 629–639.
- Eisler, R. (2000). Handbook of chemical risk assessment: health hazards to humans, plants,
  and animals. Volume 1, metals. Boca Raton, Florida: Lewis.
- EMEP (2013). Long term Changes of Heavy Metal Transboundary Pollution of the *Environment (1990-2010)* (No. 2/2013).
- 656 Foley, W. J., & Moore, B. D. (2005). Plant secondary metabolites and vertebrate herbivores
- 657 from physiological regulation to ecosystem function. *Current Opinion in Plant*658 *Biology*, 8, 430–435.
- 659 Forel, J., & Leplat, J. (2001). Faune des carabiques de France (Vol. 1). Magellanes.
- Freeland, W. J., & Janzen, D. H. (1974). Strategies in herbivory by mammals: the role of
  plant secondary compounds. *The American Naturalist*, *108*, 269–289.
- Fritsch, C., Cœurdassier, M., Giraudoux, P., Raoul, F., Douay, F., Rieffel, D., ... Scheifler,
   R. (2011). Spatially Explicit Analysis of Metal Transfer to Biota: Influence of Soil
- 664 Contamination and Landscape. *PLoS ONE*, 6, e20682.
- 665 Fritsch, C., Giraudoux, P., Cœurdassier, M., Douay, F., Raoul, F., Pruvot, C., ... Scheifler, R.
- 666 (2010). Spatial distribution of metals in smelter-impacted soils of woody habitats:
- Influence of landscape and soil properties, and risk for wildlife. *Chemosphere*, *81*,
  141–155.
- 669 Gillet, S., & Ponge, J.-F. (2003). Changes in species assemblages and diets of Collembola

- 670 along a gradient of metal pollution. Applied Soil Ecology, 22, 127-138. 671 Hansson, L. (1985). The food of bank voles, wood mice and yellow-necked mice. 672 Symposium of the Zoological Society of London, 55, 141–168. 673 Harwood, J. D., Phillips, S. W., Sunderland, K. D., & Symondson, W. O. C. (2001). 674 Secondary predation: quantification of food chain errors in an aphid-spider-carabid 675 system using monoclonal antibodies. *Molecular Ecology*, 10, 2049–2057. Heroldová, M. (2002). Food selection of *Microtus agrestis* in air-pollution affected clearings 676 677 in the Beskydy Mts, Czech Republic. Folia Zoologica, 51, 83-91. 678 Hunter, B. A., & Johnson, M. S. (1982). Food chain relationships of copper and cadmium in 679 contaminated grassland ecosystems. Oikos, 38, 108-117. 680 Hunter, B. A., Johnson, M. S., & Thompson, D. J. (1987a). Ecotoxicology of copper and 681 cadmium in a contaminated grassland ecosystem III. Small mammals. Journal of 682 Applied Ecology, 24, 601–614. 683 Hunter, B. A., Johnson, M. S., & Thompson, D. J. (1987b). Ecotoxicology of copper and 684 cadmium in a contaminated grassland ecosystem II. Invertebrates. Journal of Applied 685 Ecology, 24, 587-599. 686 Hussein, M. A., Obuid-Allah, A. H., Mohammad, A. H., Scott-Fordsmand, J. J., & Abd El-687 Wakeil, K. F. (2006). Seasonal variation in heavy metal accumulation in subtropical 688 population of the terrestrial isopod, Porcellio laevis. Ecotoxicology and 689 Environmental Safety, 63, 168–174. 690 Jeannel, R. (1941). Coléoptères carabiques. Paris: Librairie de la faculté des sciences. 691 Johnson, D. H. (1980). The Comparison of Usage and Availability Measurements for 692 Evaluating Resource Preference. Ecology, 61, 65–71. Kabata-Pendias, A. (2011). Trace elements in soils and plants (4th ed). Boca Raton: CRC 693
- 694 Press.

- Kerley, G. I. H., & Erasmus, T. (1991). What do mice select for in seeds? *Oecologia*, 86,
  261–267.
- Klare, U., Kamler, J. F., & Macdonald, D. W. (2011). A comparison and critique of different
  scat-analysis methods for determining carnivore diet: Comparison of scat-analysis
  methods. *Mammal Review*, *41*, 294–312.
- Kozlov, M. V., & Zvereva, E. L. (2011). A second life for old data: Global patterns in
  pollution ecology revealed from published observational studies. *Environmental Pollution*, 159, 1067–1075.
- Lambinon, J., Delvosalle, L., & Duvigneaud, J. (2004). Nouvelle flore de la Belgique, du *Grand-Duché de Luxembourg, du Nord de la France et des Régions voisines* (5th ed).
  Meise: Jardin botanique national de Belgique.
- Lewis, C. E., Clark, T. W., & Derting, T. L. (2001). Food selection by the white-footed mouse
   (*Peromyscus leucopus*) on the basis of energy and protein contents. *Canadian Journal of Zoology*, *79*, 562–568.
- Lukacs, P. M., Burnham, K. P., & Anderson, D. R. (2010). Model selection bias and
  Freedman's paradox. *Annals of the Institute of Statistical Mathematics*, 62, 117–125.
- MacArthur, R. H., & Pianka, E. R. (1966). On optimal use of a patchy environment. *The American Naturalist*, *100*, 603–609.
- Manly, B. F. L., McDonald, L., Thomas, D., McDonald, T. L., & Erickson, W. P. (2002).
   *Resource selection by animals: statistical design and analysis for field studies* (2<sup>nd</sup>)
- 715 ed). Dordrecht: Kluwer Academic.
- 716 Migeon, A., Richaud, P., Guinet, F., Chalot, M., & Blaudez, D. (2009). Metal Accumulation
- by Woody Species on Contaminated Sites in the North of France. *Water, Air, and Soil Pollution, 204, 89–101.*
- 719 Migliorini, M., Pigino, G., Bianchi, N., Bernini, F., & Leonzio, C. (2004). The effects of

- heavy metal contamination on the soil arthropod community of a shooting range. *Environmental Pollution*, *129*,
- Montgomery, S. S. J., & Montgomery, W. I. (1990). Intrapopulation variation in the diet of
  the wood mouse *Apodemus sylvaticus*. *Journal of Zoology*, 222, 641–651.
- Nahmani, J., & Lavelle, P. (2002). Effects of heavy metal pollution on soil macrofauna in a
  grassland of Northern France. *European Journal of Soil Biology*, *38*, 297–300.
- Nahmani, J., Lavelle, P., Lapied, E., & van Oort, F. (2003). Effects of heavy metal soil
  pollution on earthworm communities in the north of France: The 7<sup>th</sup> international
  symposium on earthworm ecology. Cardiff. Wales. 2002. *Pedobiologia*, 47, 663–
  669.
- Odendaal, J. P., & Reinecke, A. J. (1999). Short-Term Toxicological Effects of Cadmium on
  the Woodlouse, *Porcellio laevis* (Crustacea, lsopoda). *Ecotoxicology and Environmental Safety*, 43, 30–34.
- Pérès, G., Vandenbulcke, F., Guernion, M., Hedde, M., Beguiristain, T., Douay, F., ...
  Cluzeau, D. (2011). Earthworm indicators as tools for soil monitoring,
  characterization and risk assessment. An example from the national Bioindicator
  programme (France). *Pedobiologia*, *54S*, S77–S87.
- Polis, G. A., Anderson, W. B., & Holt, R. D. (1997). Toward an integration of landscape and
  food web ecology: the dynamics of spatially subsidized food webs. *Annual Review of Ecology and Systematics*, 28, 289–316.
- 740 Pompanon, F., Deagle, B. E., Symondson, W. O. C., Brown, D. S., Jarman, S. N., & Taberlet,
- P. (2012). Who is eating what: diet assessment using next generation sequencing. *Molecular Ecology*, *21*, 1931–1950.
- Poschenrieder, C., Tolrà, R., & Barceló, J. (2006). Can metals defend plants against biotic
  stress? *Trends in Plant Science*, *11*, 288–295.

- 745 Pourrut, B., Shahid, M., Dumat, C., Winterton, P., & Pinelli, E. (2011). Lead Uptake, Toxicity,
- and Detoxification in Plants. *Reviews of Environmental Contamination and Toxicology*, 213, 113–136.
- Pulford, I. D., & Watson, C. (2003). Phytoremediation of heavy metal-contaminated land by
  trees—a review. *Environment International*, *29*, 529–540.
- Pyke, G. H., Pulliam, H. R., & Charnov, E. L. (1977). Optimal foraging: a selective review
  of theory and tests. *Ouarterly Review of Biology*, *52*, 137–154.
- Quéré, J.-P., & Le Louarn, H. (2011). *Les rongeurs de France : Faunistique et biologie* (2<sup>nd</sup>
  ed). Versailles: Quae.
- Rogival, D., Scheirs, J., & Blust, R. (2007). Transfer and accumulation of metals in a soil–
  diet–wood mouse food chain along a metal pollution gradient. *Environmental Pollution*, 145, 516–528.
- Scheirs, J., Vandevyvere, I., Wollaert, K., Blust, R., & De Bruyn, L. (2006). Plant-mediated
  effects of heavy metal pollution on host choice of a grass miner. *Environmental Pollution*, 143, 138–145.
- Schloss, P. D., Westcott, S. L., Ryabin, T., Hall, J. R., Hartmann, M., Hollister, E. B., ...
  Weber, C. F. (2009). Introducing mothur: Open-Source, Platform-Independent,
  Community-Supported Software for Describing and Comparing Microbial
  Communities. *Applied and Environmental Microbiology*, *75*, 7537–7541.
- Schmidt, K. A. (2000). Interactions between food chemistry and predation risk in fox
  squirrels. *Ecology*, *81*, 2077–2085.
- Schoener, T. W. (1971). Theory of feeding strategies. *Annual Review of Ecology and Systematics*, 2, 369–404.
- Shore, R. F., & Rattner, B. A. (2001). *Ecotoxicology of Wild Mammals*. New York: John
  Wiley and Sons.

- Sih, A., & Christensen, B. (2001). Optimal diet theory: when does it work, and when and
  why does it fail? *Animal Behaviour*, *61*, 379–390.
- 572 Smith, P. N., Cobb, G. P., Godard-Codding, C., Hoff, D., McMurry, S. T., Rainwater, T. R.,
- 773 & Reynolds, K. D. (2007). Contaminant exposure in terrestrial vertebrates.
  774 *Environmental Pollution*, 150, 41–64.
- Soininen, E. M., Valentini, A., Coissac, E., Miquel, C., Gielly, L., Brochmann, C., ...
  Taberlet, P. (2009). Analysing diet of small herbivores: the efficiency of DNA
  barcoding coupled with high-throughput pyrosequencing for deciphering the
  composition of complex plant mixtures. *Frontiers in Zoology*, *6*, 16.
- Spurgeon, D. J., & Hopkin, S. P. (1996). The effects of metal contamination on earthworm
  populations. *Applied Soil Ecology*, *4*, 147–160.
- Spurgeon, D. J., & Hopkin, S. P. (1999). Seasonal variation in the abundance, biomass and
  biodiversity of earthworms in soils contaminated with metal emissions from a
  primary smelting works. *Journal of Applied Ecology*, *36*, 173–183.
- Stephens, D. W., & Krebs, J. R. (1986). *Foraging Theory* (Princeton University Press).
  Princeton, New Jersey.
- Sterckeman, T., Douay, F., Fourrier, H., & Proix, N. (2007). Référentiel pédo-géochimique
  du Nord-Pas de Calais. *Conseil Régional Du Nord-Pas de Calais*.
- Sterckeman, T., Douay, F., Proix, N., Fourrier, H., & Perdrix, E. (2002). Assessment of the
  contamination of cultivated soils by eighteen trace elements around smelters in the
  North of France. *Water, Air, and Soil Pollution, 135*, 173–194.
- Strandberg, B., Axelsen, J. A., Pedersen, M. B., Jensen, J., & Attrill, M. J. (2006). Effect of
  a copper gradient on plant community structure. *Environmental Toxicology and*
- 793 *Chemistry*, *25*, 743–753.
- 794 Sun, Y., Cai, Y., Huse, S. M., Knight, R., Farmerie, W. G., Wang, X., & Mai, V. (2012). A

- large-scale benchmark study of existing algorithms for taxonomy-independent
  microbial community analysis. *Briefings in Bioinformatics*, *13*, 107–121.
- 797 Taberlet, P., Coissac, E., Pompanon, F., Gielly, L., Miquel, C., Valentini, A., ... Willerslev,
- E. (2007). Power and limitations of the chloroplast *trn*L (UAA) intron for plant DNA
  barcoding. *Nucleic Acids Research*, *35*(3), e14.
- Taberlet, P., Coissac, E., Hajibabaei, M., & Rieseberg, L. H. (2012). Environmental DNA. *Molecular Ecology*, 21, 1789–1793.
- Tête, N., Afonso, E., Crini, N., Drouhot, S., Prudent, A.-S., & Scheifler, R. (2014). Hair as a
  noninvasive tool for risk assessment: Do the concentrations of cadmium and lead in
  the hair of wood mice (*Apodemus sylvaticus*) reflect internal concentrations? *Ecotoxicology and Environmental Safety*, 108, 233–241.
- Torres, K. C., & Johnson, M. L. (2001). Bioaccumulation of metals in plants, arthropods,
  and mice at a seasonal wetland. *Environmental Toxicology and Chemistry*, *20*, 2617–
  2626.
- 809 Trautner, J., & Geigenmueller, K. (1987). *Tiger Beetles, Ground Beetles (Illustrated Key to*810 *the Cicindelidae and Carabidae of Europe)*. Aichtal: Joseph Margraf.
- 811 Valentini, A., Pompanon, F., & Taberlet, P. (2009a). DNA barcoding for ecologists. *Trends*812 *in Ecology & Evolution*, 24, 110–117.
- Valentini, A., Miquel, C., Nawaz, M. A., Bellemain, E., Coissac, E., Pompanon, F., ...
  Taberlet, P. (2009b). New perspectives in diet analysis based on DNA barcoding and
- 815 parallel pyrosequencing: the trnL approach. *Molecular Ecology Resources*, *9*, 51–60.
- 816 van den Brink, N., Lammertsma, D., Dimmers, W., Boerwinkel, M.-C., & van der Hout, A.
- 817 (2010). Effects of soil properties on food web accumulation of heavy metals to the
  818 wood mouse (*Apodemus sylvaticus*). *Environmental Pollution*, *158*, 245–251.
- 819 Vandorpe, E., & Verhagen, R. (1980). An age reference model for the Wood mouse

- *Apodemus sylvaticus* (Linnaeus, 1758), by use of the lens technique. *Annales de La Société Royale Zoologique de Belgique*, 109, 133–140.
- Visser, M. E., & Both, C. (2005). Shifts in phenology due to global climate change: the need
  for a yardstick. *Proceedings of the Royal Society B: Biological Sciences*, 272, 2561–
  2569.
- Walker, C. H., Hopkin, S. P., Sibly, R. M., & Peakall, D. B. (2012). *Principles of Ecotoxicology* (4<sup>th</sup> ed). Boca Raton: CRC Press.
- 827 Waterlot, C., Pruvot, C., Bidar, G., Fritsch, C., De Vaufleury, A., Scheifler, R., & Douay, F.
- 828 (2016). Prediction of Extractable Cd, Pb and Zn in Contaminated Woody Habitat
  829 Soils Using a Change Point Detection Method. *Pedosphere*, *26*, 282–298.
- Watts, C. H. S. (1968). The Foods Eaten by Wood Mice (*Apodemus sylvaticus*) and Bank
  Voles (*Clethrionomys glareolus*) in Wytham Woods, Berkshire. *Journal of Animal Ecology*, 37, 25–41.
- Westoby, M. (1978). What are the biological bases of varied diets? *The American Naturalist*, *112*, 627–631.
- White, T. C. R. (2008). The role of food, weather and climate in limiting the abundance of
  animals. *Biological Reviews*, *83*, 227–248.
- Zidar, P., Drobne, D., Štrus, J., Van Gestel, C. A. M., & Donker, M. (2004). Food selection
  as a means of Cu intake reduction in the terrestrial isopod *Porcellio scaber* (Crustacea,
  Isopoda). *Applied Soil Ecology*, 25, 257–265.
- Zuur, A. F. (2009). *Mixed effects models and extensions in ecology with R.* New York:
  Springer.
- 842
- 843 Data Accessibility:

<sup>DNA sequence data, soil TM concentration in buffers, resources availability in buffers, and
wood mouse data are available from the metadata portal "dat@osu";
doi:10.25666/DATAOSU-2018-07-17.</sup> 

847

# 848 <u>Supporting Information:</u>

- 849 SI1: Information for each study site about TM contamination and dominant landscape 850 feature, as well as richness of plants and invertebrates observed in the field.
- 851 SI2: Inventory of plant and invertebrate taxa observed in each site.
- 852 SI3: Sequence data of plant, arthropod and earthworm MOTUs for each mouse.
- 853 SI4: Sequence data of sequence groups for each mouse.
- 854 SI5: Frequency of occurrence of plant, arthropod and earthworm items in wood mouse food 855 across several environmental and biological factors.
- 856 SI6: Variation of the total richness in the diet along the soil contamination gradient.
- 857 SI7: Relative proportion of the three food item categories (plants, arthropods and 858 earthworms) along the soil contamination gradient.
- 859 SI8: Plant, arthropod and earthworm richness in the diet of wood mouse across several 860 environmental and biological factors.
- 861 SI9: Complete data on models of richness in the diet.
- 862 SI10: Complete data on number and frequency of occurrence of plant food item (sequence 863 group).
- 864 SI11: Information on food selection for plant families corresponding to sequence groups in 865 the wood mouse in spring and in autumn.
- 866 SI12: Complete results on plant food selection modelling at buffer scale.
- 867

# 868 Author contributions:

- F.R. and R.S. conceived the study; S.O., R.S., C.F. and F.R. designed the study; R.S., C.F.,
- 870 F.M., T.C. and F.R. performed field samplings and laboratory treatments; B.V. performed
- bioinformatics treatments; S.O. performed the statistical analyses; S.O., F.R., R.S. and C.F.
- interpreted the results. S.O. wrote the manuscript under the supervision of F.R, R.S. and C.F.

*Table 1: Characteristics of the three primers used to amplify wood mouse stomach content DNA samples.* 

Taxonomic group	DNA type	DNA region	Primer name	Primer sequence 5'-3'	Reference
Plants	Chloroplast	trnL (UAA)	g (forward)	GGGCAATCCTGAGCCAA	Taberlet et al., 2007
			h (reverse)	CCATTGAGTCTCTGCACCTATC	
Arthropods & molluscs	Mitochondrial	16S mtDNA	16SMAV-F	CCAACATCGAGGTCRYAA	De Barba et al., 2013
			16SMAV-R	ARTTACYNTAGGGATAACAG	
Earthworms	Mitochondrial	16S mtDNA	ewD (forward)	ATTCGGTTGGGGCGACC	Bienert et al., 2012
			ewE (reverse)	CTGTTATCCCTAAGGTAGCTT	



The patterns are similar for Pb and Zn concentrations in soils.







o

. .

 $[Cd]_{soils} (mg.kg^{-1})$ 

 $\bigcirc$ 



Ô

Ground-dwelling invertebrate richness in the field

0

o



- 901 Figure 3: Earthworm richness in the diet of the wood mouse along the gradient of Cd
   902 concentration in soils in spring (a) and in autumn (b). Each point represents a buffer and
   903 point size is proportional to the number of mice captured in a given buffer. Values predicted
   904 from our model are indicated by a line. Mouse number is fixed to the median value for
   905 drawing the line (three mice in spring and two mice in autumn). The patterns are similar for
- *Pb and Zn concentrations in soils.*



