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Flavagline analog FL3 induces cell cycle arrest in urothelial carcinoma cell of the bladder by inhibiting the Akt/PHB interaction to activate the GADD45α pathway

Gangjun Yuan^{1,2†}, Xin Chen^{1,2†}, Zhuowei Liu^{2†}, Wensu Wei^{1,2}, Qinghai Shu³, Hussein Abou-Hamdan⁴, Lijuan Jiang¹, Xiangdong Li¹, Rixin Chen², Laurent Désaubry^{4,5*}, Fangjian Zhou^{1,2*} and Dan Xie^{1*}

Abstract

Background: Prohibitin 1 (PHB) is a potential target for the treatment of urothelial carcinoma of the bladder (UCB). FL3 is a newly synthesized agent that inhibits cancer cell proliferation by targeting the PHB protein; however, the effect of FL3 in UCB cells remains unexplored.

Methods: FL3 was identified to be a potent inhibitor of UCB cell viability using CCK-8 (cell counting kit-8) assay. Then a series of *in vitro* and *in vivo* experiments were conducted to further demonstrate the inhibitory effect of FL3 on UCB cell proliferation and to determine the underlying mechanisms.

Results: FL3 inhibited UCB cell proliferation and growth both *in vitro* and *in vivo*. By targeting the PHB protein, FL3 inhibited the interaction of Akt and PHB as well as Akt-mediated PHB phosphorylation, which consequently decreases the localization of PHB in the mitochondria. In addition, FL3 treatment resulted in cell cycle arrest in the G2/M phase, and this inhibitory effect of FL3 could be mimicked by knockdown of PHB.

Through the microarray analysis of mRNA expression after FL3 treatment and knockdown of PHB, we found that the mRNA expression of the growth arrest and DNA damage-inducible alpha (*GADD45a*) gene were significantly upregulated. When knocked down the expression of *GADD45a*, the inhibitory effect of FL3 on cell cycle was rescued, suggesting that FL3-induced cell cycle inhibition is *GADD45a* dependent.

Conclusion: Our data provide that FL3 inhibits the interaction of Akt and PHB, which in turn activates the *GADD45a*-dependent cell cycle inhibition in the G2/M phase.

Keywords: FL3, PHB, Urothelial carcinoma of the bladder, *GADD45a*, Cell cycle

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Background

Urothelial carcinoma of the bladder (UCB) is the most common malignancy involving in the genitourinary system, it presents a high risk of recurrence and mortality [1]. Especially, muscle-invasive bladder cancer (MIBC) patients suffer from a poor 5-year survival rate of less than 35% [2]. Adjuvant chemotherapy is helpful to delay the progression of this disease, however, presents limited effects and serious side-effects [3]. Thus, there is an urgent need to develop novel drugs that are both effective and safe.

Prohibitin 1 (PHB) is an evolutionarily conserved adaptor protein involved in diverse biological processes such as transcriptional regulation, cell proliferation, mitochondrial function, and resistance to apoptosis [4, 5]. It is ubiquitously expressed in diverse cellular compartments including the nucleus, cytoplasm and mitochondria [6]. PHB is critical of PHB localization within mitochondria by that PHB involves in and maintains the integrity of mitochondria [7]. Recently, numerous studies have revealed that PHB is largely involved in the pathogenesis of several types of human cancers including prostate, breast, lung and gastric cancers [3, 8–10]. Our previous study also demonstrated that PHB is over-expressed in UCB tissues and correlates with a poor prognosis in UCB patients [3, 11]. Moreover, PHB localization in mitochondria is critical for promoting UCB cell proliferation and this process is in the requirement of Akt-mediated phosphorylation [11]. These data, taken together, suggest that PHB is a potential target for the treatment of UCB.

Flavaglines are natural products that newly isolated from medicinal plants in Asia and were found to have anti-tumor effects in human cancer cell lines without being toxic to healthy cells [12–14]. A recent study reported that Flavaglines target the PHB protein and thus, substantially inhibit cell proliferation in leukemic Jurkat cells by repressing activation of the CRaf-MEK-ERK signaling pathway [12]. Similar effects were observed in several human cancer cell lines such as melanoma, colon cancer, breast cancer, leukemia, and lung cancer [15]. To date, however, the potential inhibitory effect of Flavaglines and its underlying mechanisms in UCB cells remain unexplored.

FL3, a derivative analogue of Flavaglines, is identified to be a potent inhibitor of cancer cell proliferation [16]. To date, however, its biological functions and underlying mechanisms in UCB pathogenesis has not been elucidated. Thus, in the present study, by using a series in vitro and in vivo assays, we investigated the potential inhibitory activity of FL3 in UCB cells and studied if FL3 could also target PHB protein to block the Akt/PHB interaction and consequently influence certain critical signal pathways in UCB proliferation.

Methods

Materials

Synthetic PHB ligands and FL3-conjugated compound 6 (an amino linker) were provided from The Laboratory for Therapeutic Innovation, University of Strasbourg (Strasbourg France). All of the drugs were stored at -20°C as solids, and were dissolved in dimethylsulfoxide (DMSO) before use.

Cell culture

UCB cell lines T24, BIU and 5637, and normal bladder uroepithelial cell line SV-HUC-1 were cultured at 37°C with 5% CO_2 in RPMI 1640 media supplemented with 10% fetal bovine serum, penicillin (100 U/ml), and streptomycin (100 $\mu\text{g}/\text{ml}$).

Cell viability assay

Cell viability was determined using the Cell Counting Kit-8 (CCK8) Kit (Dojindo, Kumamoto, Japan). After incubation with concentrations of FL3 and paclitaxel for indicated hours, absorbance was measured at 450 nm according to the manufacture's introduction. Cell viability was expressed as the percentage of absorbance of cells treated with FL3 or paclitaxel compared with cells treated with DMSO.

Colony formation

Cells were plated as a density of 500 cells/well in 6-well plates and incubated overnight. Then different concentrations of FL3 were added to each well followed by incubation for 4–5 days. After a growth period, colonies were fixed in methanol for 30 min and stained with 0.1% crystal violet for 1 h.

Western blot analysis

Cells were harvested and lysed, followed by isolation of the supernatant and measurement of the protein concentration using the bicinchoninic method. Then the protein extracts were separated on 10% sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) gels, and electrotransferred to a PVDF membrane at 250 mA for 2 h at room temperature. Then, the membrane was blocked in 5% bovine serum albumin, and incubated overnight at 4°C with primary antibodies (Akt, phospho-Akt Ser 473, prohibitin-1, phospho (Ser/Thr)-Akt substrate, and cyclin-dependent kinase antibodies were from Cell Signaling Technology, Danvers, MA, USA; REA antibody was from Millipore, Billerica, MA, USA; histone H3, Cox IV, GADD45 α , and GAPDH antibodies were from Proteintech Group Inc., Rosemont, IL, USA). The membrane was washed and incubated with secondary antibody for 1 h at room temperature. The signal was measured using the ECL detection system (Tanon, Shanghai, China).

Immunoprecipitation

Cells (1×10^8) were harvested after incubation with 0.5 μ M FL3 or DMSO for 24 h. Then the cells were lysed with immunoprecipitation (IP) lysate buffer (Beyotime, Shanghai, China) and the protein extracts were isolated. A volume of 20 μ l Protein G-agarose beads/tube were washed three times with buffer, followed by incubation with anti-PHB antibody, anti-phospho-Akt antibody or IgG antibody at 4 °C with rotation for 3 h. Equal amounts of protein (21 μ g) were incubated with beads overnight at 4 °C. After incubation, The beads were re-suspended in 1 \times LDS Sample Buffer and boiled for 5 min. Then the proteins were resolved on 10% SDS-PAGE gels for Western blot analysis.

Generation of the FL3-Affigel

FL3-Affigel beads were generated according to the protocol by Thuaud [16]. Briefly, a volume of 1 ml packed Affigel 10 (Bio-Rad Laboratories, Hercules, CA, USA) was washed on a glass filter with acetonitrile (ACN) and added to a solution of FL3-conjugated compound 6 (3 mg) in ACN (0.8 ml) and triethylamine (20 μ l). This suspension was gently rotated for 12 h at room temperature. Ethanolamine (50 μ l) was added and after 4 h, the resin was thoroughly washed with ACN, ethanol and water. Coupled AffiGel-10 agarose beads were stored in PBS containing 0.02% NaN₃ at 4 °C until further use. PBS was used as the negative control (NC).

Identification of FL3 binding to PHB protein

T24 cells (1×10^8) cells were harvested to obtain total protein extracts. 30 μ l FL3- or NC-Affigel beads/tube were incubated with total cell extracts at 4 °C with gentle rotation overnight. After incubation, the bound mixtures were re-suspended in 1 \times LDS buffer and subjected to SDS-PAGE for Western blot analysis with primary PHB antibody.

Detection of PHB protein in the subcellular compartments

Approximately 1×10^8 cells were harvested after incubation with DMSO (control) or FL3 0.5 μ M for 24 h. Using the Nuclear and Cytoplasmic Protein Extraction Kit and Cell Mitochondria Isolation Kit (Beyotime), proteins of the nucleus, cytoplasm, and mitochondria were separately extracted according to the manufacturer's introductions. Then the extracts were subjected for Western blot analysis.

Knockdown experiment

The Lenti-Pac HIV Expression Packaging Kit (GeneCopoeia, Rockville, MD, USA) was used to construct stable low-expressing PHB T24 cell lines according to the manufacturer's protocol. Stable PHB-silenced cells were

selected with 0.1% puromycin (Invitrogen, Darmstadt, Germany).

Apoptosis assay

The percentage of apoptotic cells was determined by using an Annexin V-PE apoptosis detection kit (KeyGen Biotech, Jiangsu, China) according to the manufacturer's introduction. In brief, a total of 10^5 T24 and BIU cells were harvested after incubation with FL3 for 24 h, then stained these cells with Annexin V and PE subsequently. Finally, the cells were subjected to apoptosis analysis with the ACEA NovoExpress System (ACEA Biosciences Inc., CA, USA).

Cell cycle analysis

The Cell Cycle Assay Kit (Beyotime) was used to analyze the cell cycle distribution. According to the manufacturer's introduction, FL3-treated or control T24 cells were harvested and dealt with PI, RNase A, Triton X-100 subsequently, followed by cell cycle analysis with the MoFlo XDP Flow Cytometry System (Beckman Coulter, CA, USA).

Subcutaneous and orthotopic xenografts in Balb/c nude mice

Four-week-old Balb/c nude mice were purchased from Charles River Laboratories (Beijing, China). All of the animal experiments were approved by the Animal Ethics Committee of Sun Yat-sen University (Guangzhou, China). A total of 5×10^5 cells were subcutaneously inoculated into the right flank of mice. When the tumor sizes reached to about 4–6 mm³, the mice were randomly divided into four groups with seven mice each, and intraperitoneally injected with FL3 (2 mg/kg, 5 mg/kg), palicetaxel (10 mg/kg, positive control), or DMSO (NC) every 2 days for a total of eight times. Body weights and tumor volume were measured and recorded at the time of administration. At the end of the study, the tumors and main organs including livers, kidneys, lungs, and hearts were removed for further immunohistochemistry experiments. The following formula was used to measure tumor volumes: tumor volume = $1/2 L \times W^2$, where L represents length and W denotes width.

Immunohistochemistry

The removed organs and tumors were fixed in formalin and then embedded in paraffin. Sections (4 μ m thick) were cut and stained with hematoxylin and eosin (H & E). For further immunohistochemical analysis, sections were de-paraffinized in xylene, hydrated in graded alcohol, and blocked in 3% hydrogen peroxide to inhibit endogenous peroxidase activity. Antigen retrieval was completed by incubating the slides for

5 min in Ethylene Diamine Tetraacetic Acid (EDTA) buffer (pH 8.0). After incubation with 10% goat serum, the slides were incubated with anti-PHB antibody (1:400; Santa Cruz) overnight at 4 °C, followed by incubation with secondary goat anti-rabbit antibody at 37 °C for 30 min. Then, the slides were stained with DAB staining solution for less than 5 min, and re-stained with hematoxylin for 1 min followed by polarization for less than 10 s.

Statistical analysis

All statistical analyses were performed with IBM SPSS Statistics 19.0 (SPSS Inc., Chicago, IL, USA). All data both in vitro and in vivo are presented as mean ± S.D. and assessed by two-tailed Student’s *t*-test with three

independent experiments. *P* values of < 0.05 was considered statistically significant.

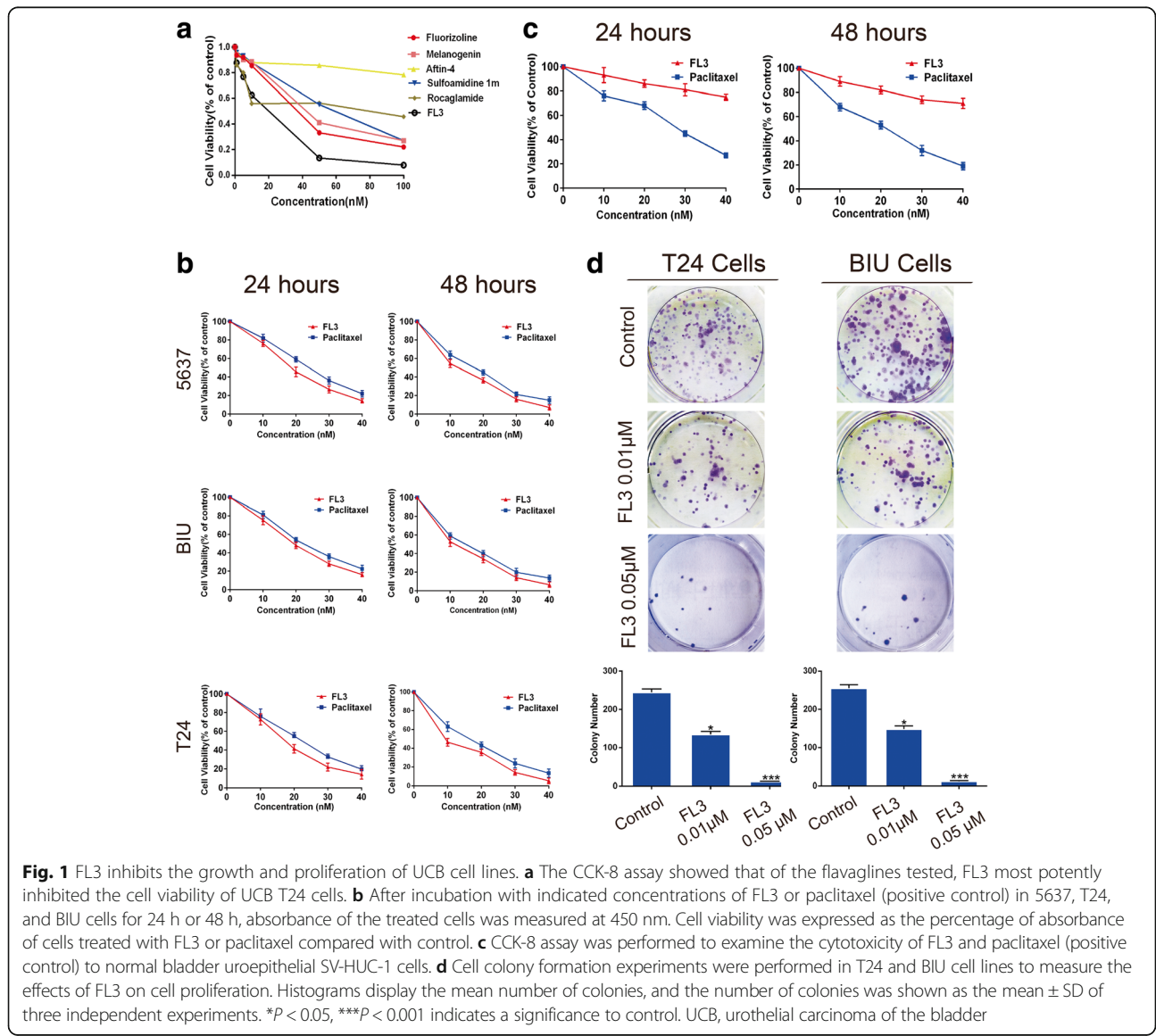
Results

FL3 is a potent inhibitor of UCB cell growth

To determine if Flavaglines had anti-tumor effects in UCB cells, we measured the cell viability of UCB T24 cells after treatment with various PHB ligands for 24 h. As shown in Fig. 1a, all of PHB ligands used decreased cell viability of T24 cells, in which FL3 exhibited the most potent effect to inhibit cell growth.

FL3 suppresses UCB cell proliferation in vitro

Next, we explored the inhibitory effect of FL3 on UCB cell proliferation in vitro. To this end, the cell viability



assay was performed in three UCB cell lines after FL3 treatment. Paclitaxel, a well-known anticancer agent for some human cancers including UCB, was used as a positive control. As shown in Fig. 1b, FL3 dominantly inhibited the cell viability of three UCB cell lines in a time- and dose-dependent manner. Compared to paclitaxel, FL3 presented less cytotoxicity to normal bladder uroepithelial SV-HUC-1 cells (Fig. 1c). The colony formation assay showed that FL3 treatment significantly inhibited the colony formation ability of T24 and BIU cells compared to the control (Fig. 1d). Taken together, these

results showed that FL3 suppressed UCB cell proliferation in vitro.

FL3 binds to the PHB protein

Because PHB is the direct target of Flavaglines in Jurkat T cells [12], FL3-Affigel-10 beads were used to determine whether FL3 targets PHB protein in UCB cell lines using immunoprecipitation assay. As shown in Fig. 2a, PHB protein was pulled down by the FL3-Affigel-10 beads, but there was no PHB binding to the

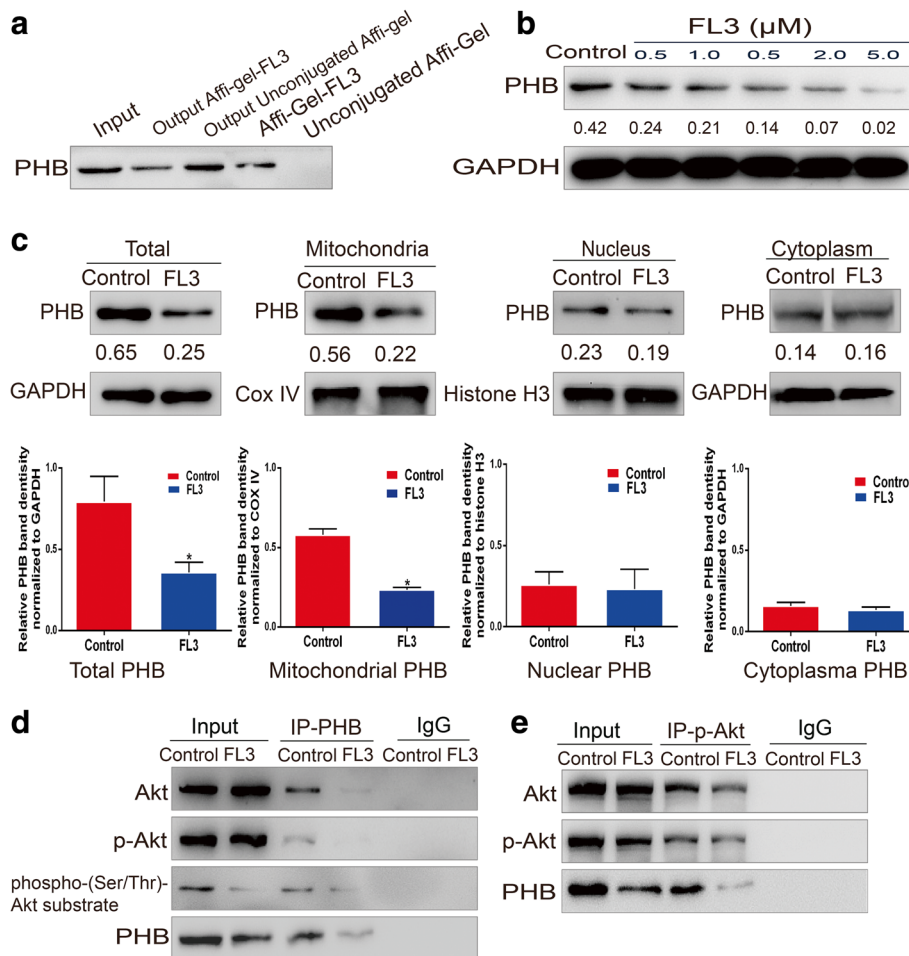


Fig. 2 FL3 binds to PHB protein and inhibits the Akt/PHB interaction. **a** FL3-conjugated or unconjugated beads were incubated with total cell lysates from T24 cells. The eluted proteins were resolved on 10% SDS-PAGE gels for Western blotting with primary PHB antibody. **b** After incubation with indicated concentrations of FL3 for 24 h, total proteins of T24 cells were extracted and subjected to Western blot with primary PHB antibody. The Western blots have been quantified by densitometry and the quantitative values have been incorporated below the Western blot bands. **c** Subcellular fractions including cytoplasm, nucleus and mitochondria were isolated from the FL3-treated or control T24 cells. Then these fractions were lysed to obtain their total proteins, followed by Western blot analysis. Histograms show PHB protein intensity normalized to GAPDH, COX IV and Histone H3, respectively. The values represent the mean ± SD of three independent experiments. *P denotes < 0.05. The Western blots have been quantified by densitometry and the quantitative values have been incorporated below the Western blot bands. **d** After treatment with 0.5 μM FL3, total cell lysates of T24 cells were immunoprecipitated with primary PHB antibody. Then the eluted proteins were subjected to Western blot analysis with primary antibodies as indicated (left panel). **e** After treatment with 0.5 μM FL3, total cell lysates of T24 cells were immunoprecipitated with primary phospho-Akt (p-Akt) antibody. Then the eluted proteins were subjected to Western blot analysis with primary antibodies as indicated (left panel)

unconjugated-Affigel-beads, indicating that FL3 binds to PHB protein.

FL3 decreases PHB localization in the mitochondria

A recent study showed that FL3 treatment could influence the localization of PHB in cardiomyocytes by promoting PHB translocation from nucleus to mitochondria [13], however, it remains to be determined in UCB cells. We first measured the total PHB protein levels of FL3-treated T24 cells, and the result showed that the protein levels of PHB was slowly decreased by FL3 treatment (Fig. 2b). Although ubiquitously existed in multiple cellular compartments and each location stands for different functions, PHB mainly distributes in the mitochondria and participates in the stability of mitochondrial structure and functions [4, 7, 17–19]. We detected the change of PHB protein levels in main cellular fractions after FL3 treatment, and the results revealed that mitochondrial PHB levels was dramatically decreased, whereas nuclear PHB expression and cytoplasmic PHB expression were not significantly changed (Fig. 2c), suggesting that FL3 dose not promote PHB translocation from mitochondria to nucleus and cytoplasm.

FL3 inhibits the Akt/PHB interaction and decreases PHB phosphorylation

PHB is a substrate of Akt, it is activated and transferred to the mitochondria after phosphorylation by Akt at Thr-258 site [11, 20]. FL3 might impede PHB localization within the mitochondria by blocking the interaction of PHB with Akt, which subsequently inhibits the phosphorylation process of PHB. To test this, total proteins of FL3-treated T24 cells were subjected to immunoprecipitation assay with primary PHB or phospho-Akt (Ser 473) antibody followed by western blot analysis. When proteins were immunoprecipitated with PHB antibody, FL3 treatment resulted in significant decrease of pulled-down PHB, Akt and phospho-Akt (p-Akt) compared to the control (Fig. 2d). If the proteins were immunoprecipitated with p-Akt antibody, pulled-down Akt and phospho-Akt levels were not significantly different, whereas the pulled-down PHB was significantly decreased compared to control (Fig. 2e).

Due to the shortage of available antibody recognizing phosphorylated PHB protein, we indirectly detected phosphorylated levels of PHB with a phospho-(Ser/Thr) Akt substrate (PAS) antibody, which can preferentially recognize peptides and proteins containing phospho-Ser/Thr preceded by Lys/Arg at positions – 5 and – 3 in a manner largely independent of other surrounding amino acids. As shown in Fig. 2d, a protein between 25 and 35 kDa was pulled down by PAS, and its expression was clearly reduced by FL3 compared to the control.

This result indirectly showed that FL3 treatment decreased PHB phosphorylated levels.

Taken together, these data showed that FL3 inhibited the interaction of Akt and PHB, which in turn decreased PHB phosphorylation and localization in the mitochondria.

FL3 does not induce cell apoptosis in UCB cells

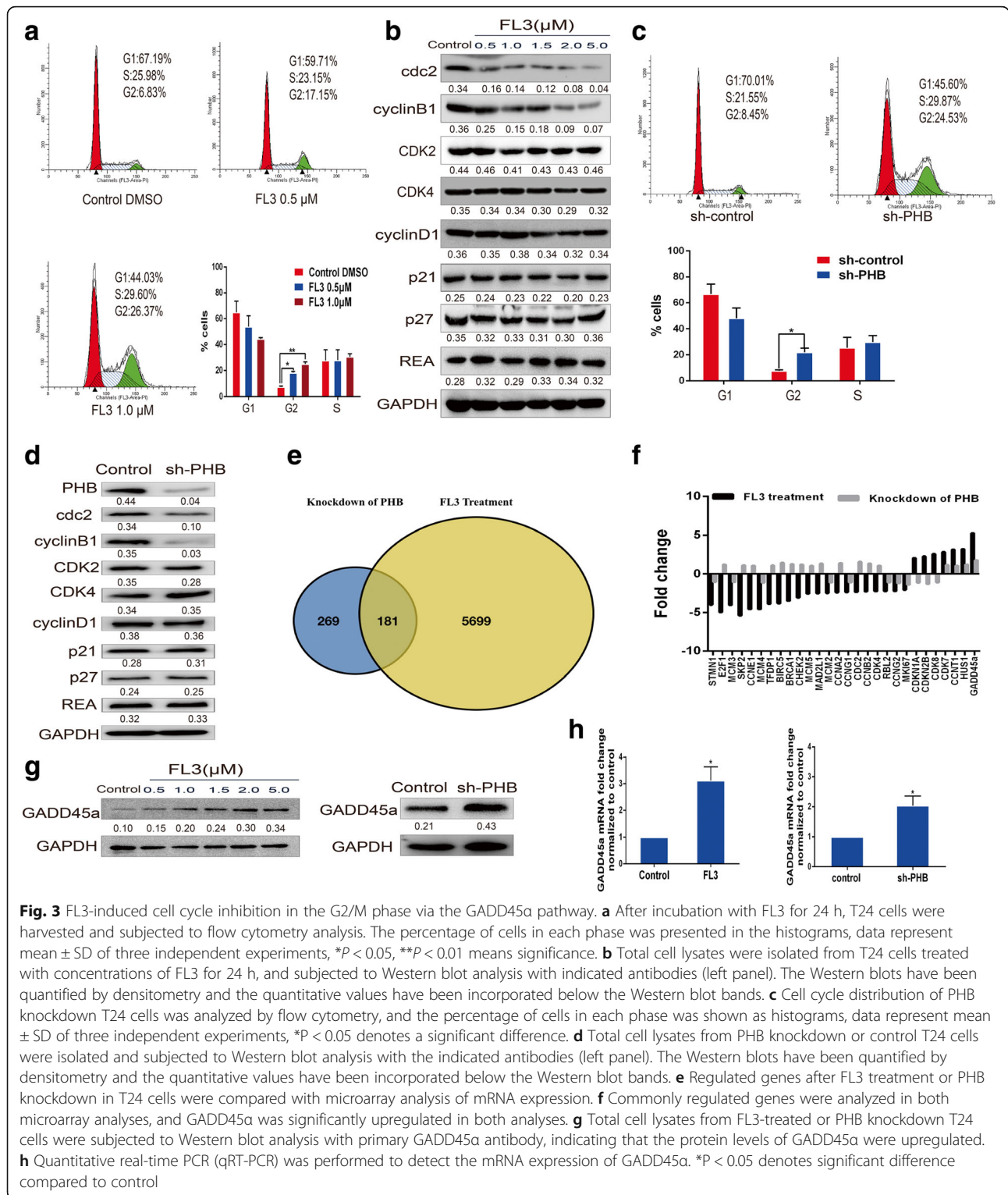
A recent study has revealed the apoptotic activity of FL3 in certain cancer cells, such as HL60 and Hela cells, by triggering the translocation of Apoptosis Inducing Factor (AIF) and caspase-12 to the nucleus [16]. We also examined the apoptotic activity of FL3 in UCB T24 and BIU cells by flow cytometry assay. However, our results did not show a similar result that FL3 treatment did not significantly increase the apoptotic number of both T24 and BIU cells, suggesting that FL3 does not induce cell apoptosis in UCB cells (Additional file 1: Figure S1).

FL3 induces cell cycle arrest in the G2/M phase

It has been shown that FL3 affects cell cycle distribution in Jurkat T cells [12], so we determined whether it has similar effects in UCB cells. As Fig. 3a shown, FL3 resulted in cell cycle arrest in the G2/M phase by increasing the percentage of cells in the G2 phase from 6.83% in control cells to 17.15% and 26.37% in FL3 -treated T24 cells, and the percentage of cells in S phase nonsignificantly ranged from 25.98% to 23.15% and 29.60%. In addition, protein expressions of cdc2 and cyclin B1, which are implicated with G2-M transition [21], were dramatically decreased (Fig. 3b). In contrast, protein expressions of G1-S transition related cyclins such as CDK2, CDK4, and cyclin D1 were unchanged. Also, several key regulators of cell cycle including p21, p27 and REA (repressor of estrogen receptor activity) [22–24] were also analyzed, the results showed that their protein expressions were not significantly changed after FL3 treatment (Fig. 3b).

PHB knockdown mimics the inhibitory effect of FL3 on the cell cycle progression in UCB T24 cells

To clarify the action of FL3 on PHB signaling, we knocked down PHB expression in T24 cells to determine whether PHB knockdown would mimic the inhibitory effect of FL3 on cell cycle progression. Flow cytometry assay was performed and the results showed that PHB knockdown increased the percentage of cells in the G2 phase from 8.45% to 24.53%, accompanying with slight and insignificant increase of cells in the S phase from 21.55% to 29.87% (Fig. 3c). Meanwhile, the protein levels of cdc2 and cyclin B1 were decreased, whereas that of CDK2, CDK4, cyclin D1, p21, p27 and REA were not significantly changed (Fig. 3d). These data suggests that knockdown of PHB mimics the inhibitory effect of FL3 on cell cycle progression.



GADD45a is upregulated after FL3 treatment and PHB knockdown

To identify the common pathways regulated by FL3 treatment or PHB knockdown, microarray analyses of mRNA expression were performed in FL3-

treated or PHB knockdown T24 cells. Totally regulated genes were compared and presented as shown in Fig. 5e, a total of 181 genes were commonly regulated in both analyses. Since FL3 and PHB knockdown inhibits cell cycle progression in the G2/M

phase, we tried to explore the underlying mechanism of this effect. A total of 84 cell cycle-related genes [25] were compared with each other in both analyses. Significantly regulated genes (more than 1.7-fold) were selected out in Additional file 2: Table S1, 28 genes were significantly regulated by FL3. Comparing the 28 genes in microarray analysis of PHB knockdown cells, 11 genes were commonly regulated by FL3 and PHB knockdown (Fig. 3f), among of which GADD45α (the growth arrest and DNA damage-inducible alpha), a checkpoint gene during the G2-M transition [26], was verified as the only significantly regulated gene (5.26 vs. 1.76-fold). Upregulation of GADD45α was further validated by western blotting and quantitative real time-PCR (qRT-PCR) (Fig. 3g-h). The results indicated that FL3-induced cell cycle arrest might be GADD45α-dependent.

FL3-mediated cell cycle inhibition is GADD45α-dependent

To determine whether FL3-induced cell cycle arrest is GADD45α-dependent, a rescue experiment was performed by repressing GADD45α expression with siRNA in T24 cells. Flow cytometry assay demonstrated that FL3 treatment did not significantly change the percentage of cells in the G2 phase (from 7.70% to 11.36%) in GADD45α-silenced cells, whereas the percentage of cells in this phase increased from 10.58% to 24.42% in the control cells (Fig. 4a, b). Thus, repression of GADD45α rescued the inhibitory effect of FL3 on cell cycle progression. In addition, repression of GADD45α did not change the protein levels of PHB, cdc2 and cyclinB1 (Fig. 4c). Moreover, repression of GADD45α attenuated the inhibitory effect of FL3 on the colony formation ability of T24 cells (Fig. 4d), indicating that GADD45α is critical for UCB cell proliferation. Taken together, FL3-induced cell cycle inhibition is GADD45α-dependent.

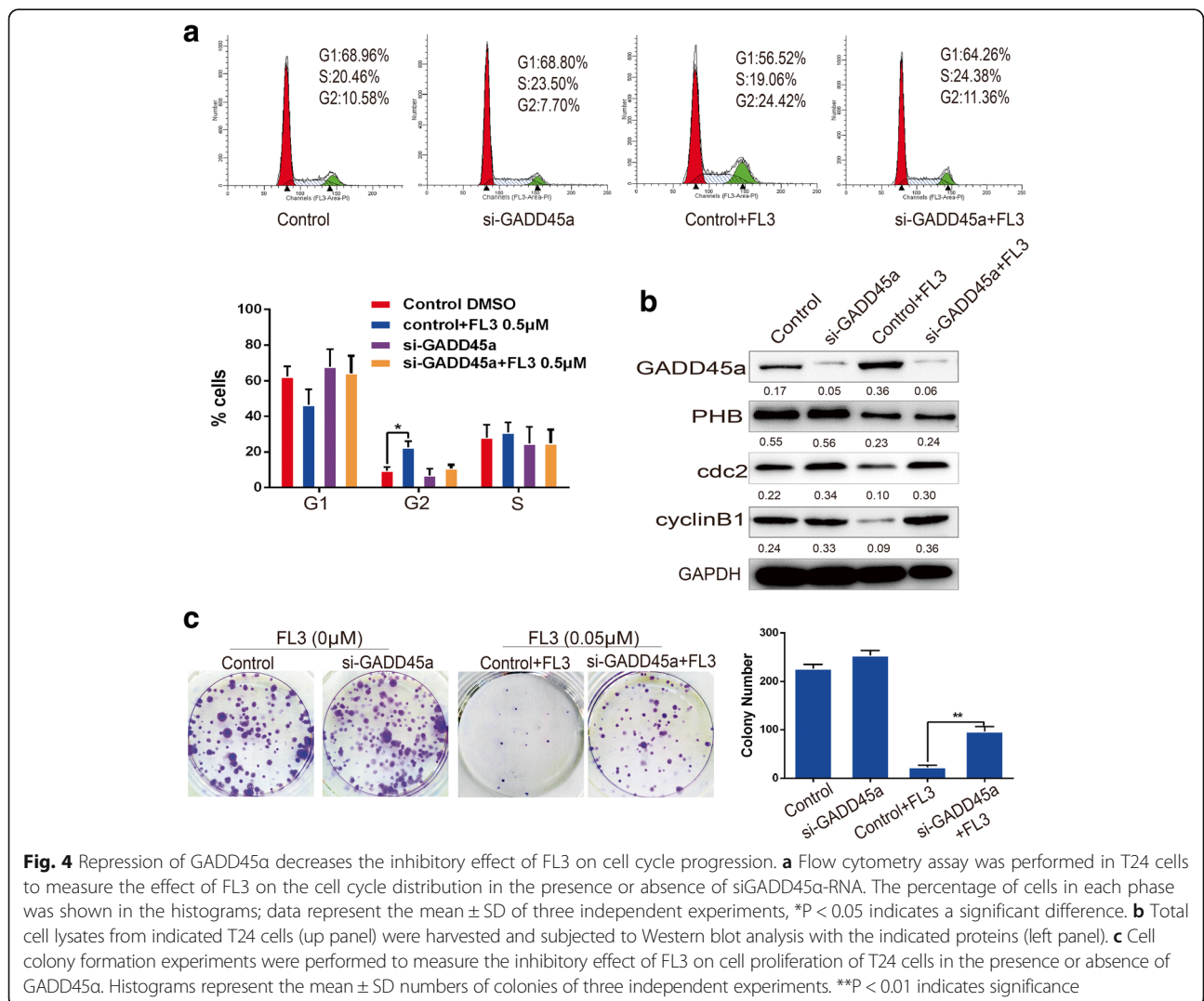


Fig. 4 Repression of GADD45α decreases the inhibitory effect of FL3 on cell cycle progression. **a** Flow cytometry assay was performed in T24 cells to measure the effect of FL3 on the cell cycle distribution in the presence or absence of siGADD45α-RNA. The percentage of cells in each phase was shown in the histograms; data represent the mean ± SD of three independent experiments, *P < 0.05 indicates a significant difference. **b** Total cell lysates from indicated T24 cells (up panel) were harvested and subjected to Western blot analysis with the indicated proteins (left panel). **c** Cell colony formation experiments were performed to measure the inhibitory effect of FL3 on cell proliferation of T24 cells in the presence or absence of GADD45α. Histograms represent the mean ± SD numbers of colonies of three independent experiments. **P < 0.01 indicates significance

FL3 suppresses xenograft tumor growth in vivo

To further verify the inhibitory role of FL3 on cell proliferation in vivo, xenograft tumor models were constructed (Fig. 5a). Both FL3 and paclitaxel treatments significantly inhibited the volumes and weights of tumors compared to the control (Fig. 5b, c). Furthermore, immunohistochemistry assay presented that FL3 treatment resulted in slightly decreased expression of PHB protein compared to the control (Fig. 5d). In contrast, the expression of GADD45 α was slightly elevated. Meaningfully, there was no significant body weight loss in FL3-treated mice compared to the control (Fig. 5e). H&E staining of the livers, lungs, hearts, and kidneys showed no major organ-related toxicities from FL3 treatment compared to the control group (Fig. 5f).

Discussion

Current chemotherapeutic drugs have limited effectiveness and intolerable toxicities in most cases of UCB. Therefore, there is an urgent need for developing novel therapeutic modalities of UCB. It has been widely reported that PHB is critical for many cellular biologic responses including senescence, development and tumorigenesis [4, 7, 16, 27–30]. Our previous study revealed that in UCBs, PHB is frequently overexpressed in >80% clinical patients, and UCB with high expression of PHB predicts a poor disease-free survival of the patient, suggesting a potential therapeutic target of PHB in UCB [11].

Recent studies have shown that flavaglines can directly binds to PHB to inhibit the activation of PHB-mediated signaling pathways, consequently leading to inhibition of protein synthesis, cell cycle progression, and cell growth

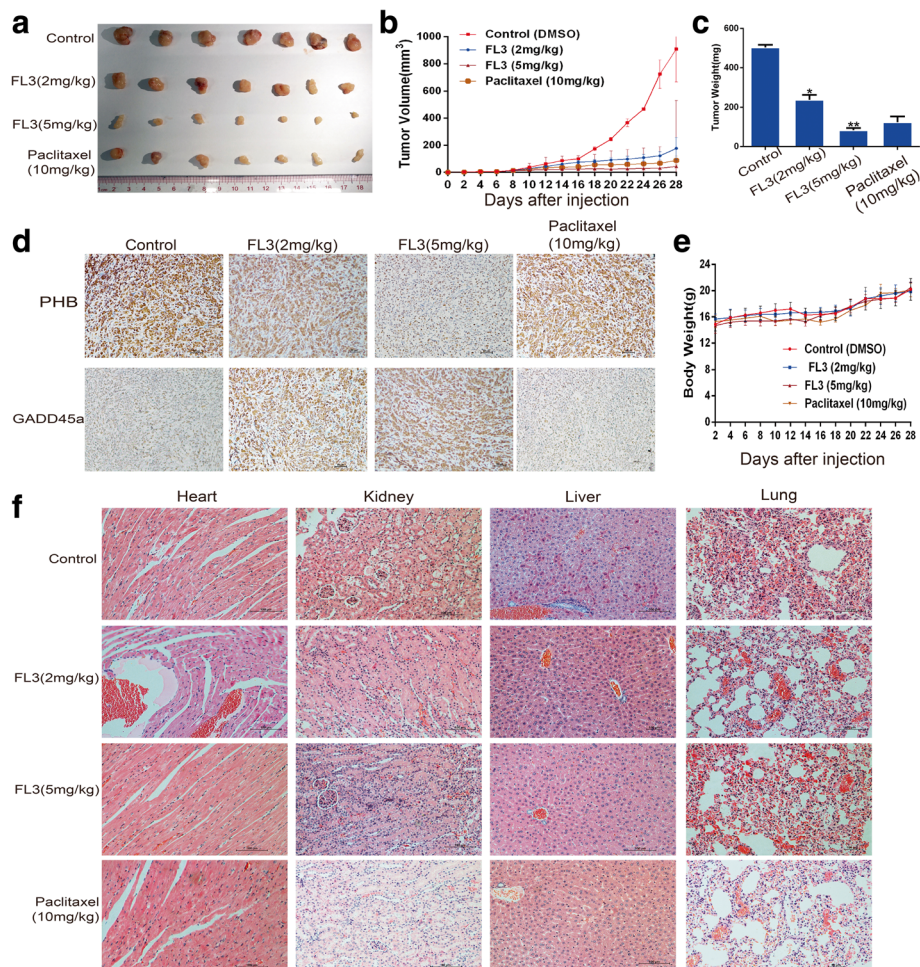


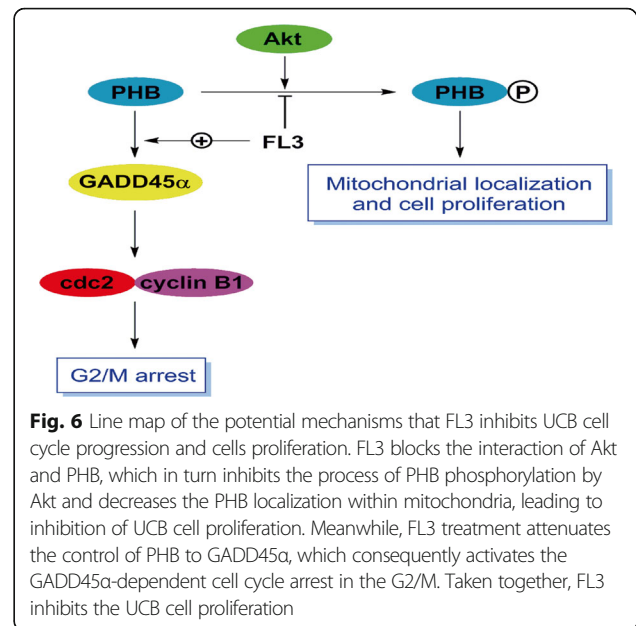
Fig. 5 FL3 inhibits growth of UCB tumor xenografts in vivo. **a** The xenograft tumors were isolated from mice at the end of study. **b** Tumor volumes were recorded from the date of injection to the end of the study (mean, $n = 7$). **c** Histograms present the mean tumor weight in each group, means \pm SD ($n = 7$). ****** $P < 0.01$, ******* $P < 0.001$ indicates a significant difference between FL3-treated mice and control mice. **d** Tumors were embedded in paraffin and 5 μ m thick sections were used for immunohistochemistry analysis with PHB or GADD45 α antibody. **e** Body weights of mice were recorded along with the records of tumor volumes as dashed lines (mean, $n = 7$). **f** Main organs including heart, kidney, liver, and lung were removed from mice and embedded in paraffin for further hematoxylin eosin staining

in cancer cells [12, 31, 32]. Meanwhile, flavaglines present little cytotoxicity to healthy cells at nanomolar concentrations [13, 16, 33, 34]. Therefore, the discovery of flavaglines brings new hope for the treatment of UCB. In our present study, we have demonstrated that FL3, a derivative of flavaglines, directly targets PHB and inhibits UCB cell proliferation both in vitro and in vivo.

The PI3K/Akt pathway has been implicated in many cellular biological responses and the development of carcinogenesis [35–38]. We found that both Akt and PHB were upregulated in UCB tissues and correlated with the poor prognosis of UCB patients [11]. Recent studies have demonstrated that PHB is a substrate of Akt, and Akt-mediated phosphorylation is required for PHB localization within the mitochondria as well as UCB cell proliferation [11, 39]. Mitochondrial PHB is closely associated with mitochondrial functions including mitochondrial stabilization, resistance of apoptosis, the respiratory chain, and components of mitochondrial DNA [40, 41]. Correlating with cell proliferation, reduction of PHB expression in the mitochondria might lead to mitochondria-related apoptosis and cell cycle inhibition. In this study, our results showed that FL3 treatment decreased the interaction of Akt and PHB as well as PHB localization within mitochondria. However, we did not observe the similar effect of inducing apoptosis in UCB cells as FL3 dose in HL60 and Hela cells. We thought that the functions of FL3 to induce cellular apoptosis might be tumor type-specific.

Since the activity of FL3 is based on its binding with PHB protein, knockdown of PHB could mimic the inhibitory effect of FL3 on UCB cell cycle progression. We found that the expression of GADD45 α , a regulator of cell cycle, was significantly upregulated after either FL3 treatment or PHB knockdown. GADD45 α is a member of the growth arrest and DNA damage 45 (GADD45) gene family, which encodes three highly homologous proteins GADD45 α , GADD45 β , and GADD45 γ [37]. GADD45 α localizes to the nucleus and involves in the inhibition of cell cycle G2-M transition by inhibiting the activation of cdc2/cyclin B1 kinases, leading to the initiation of the G2/M checkpoint mechanism and subsequently arrests cell cycle progression in the G2/M phase [21, 26, 42, 43]. Consistent with these studies, the expression of GADD45 α expression was upregulated while the expression of cdc2 and cyclin B1 were decreased after FL3 treatment in UCB cells. If the expression of GADD45 α is repressed, the inhibitory effect of FL3 on cell cycle would be rescued. Thus, our results has strongly suggested that FL3-induced G2/M cell cycle inhibition is GADD45 α -dependent.

GADD45 α involved cell cycle regulation is controlled by Akt/FOXO3A pathway by that Akt represses the activity of GADD45 α and promotes cell cycle progression [44]. In our results, the expression of GADD45 α is also



controlled by PHB. Taking together, GADD45 α expression is negatively regulated by Akt/PHB. By directly targeting with PHB protein, FL3 blocks the interaction of Akt and PHB and in turn attenuates the control of Akt/PHB to GADD45 α , with a consequence of upregulation of GADD45 α and cell cycle inhibition in the G2/M phase.

Conclusions

This study provides data to elucidate the mechanism of FL3 inhibiting UCB cell proliferation (Fig. 6), suggesting that FL3 might be a therapeutic agent for the treatment of UCB in the future.

All data in this study have been recorded at Sun Yat-sen University Cancer Center (Number RDDB2017000176).

Additional files

Additional file 1: Figure S1. FL3 does not induce apoptosis in UCB T24 and BIU cells. After incubation with 0.5 μ M and 1.0 μ M FL3 for 24 h, T24 and BIU cells were harvested and stained with Annexin V-FITC/PE to count the numbers of apoptotic cells. Q3–1, dead cells; Q3–2, late apoptotic cells; Q3–3, live cells; Q3–4, early apoptotic cells. This experiment was independently repeated for three times. Histograms display the number of cells as the mean \pm SD of three independent experiments. (TIFF 67307 kb)

Additional file 2: Table S1. List of cell cycle-related genes regulated by FL3 treatment or PHB knockdown in T24 cells using microarray analysis of mRNA expression. (DOC 94 kb)

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Authors' contributions

ZWL, FJZ and DX involved in the study concept and design; LD and H A-H provided the PHB ligands and involved in the study concept; XC designed the study and drafted the manuscript; WSW performed flow cytometry analysis; QHS performed Generation of the FL3-Affigel; XDL, LJJ, and RXC performed microarray analysis. All authors read and approved the final manuscript.

Ethics approval

All animal experiments were approved by the Animal Ethics Committee of Sun Yat-sen University cancer center.

Competing interests

The authors declare that they have no competing interests.

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References

- Chen W, Zheng R, Baade PD, et al. Cancer statistics in China, 2015. *CA Cancer J Clin*. 2016;66(2):115–32.
- DeSantis CE, et al. Cancer treatment and survivorship statistics, 2014. *CA Cancer J Clin*. 2014;64:252–71.
- Mitsui Y, Yasumoto H, Arichi N, Honda S, Shiina H, Igawa M. Current chemotherapeutic strategies against bladder cancer. *Int Urol Nephrol*. 2012; 44(2):431–41.
- Mishra S, Murphy LC, Nyomba BL, Murphy LJ. Prohibitin: a potential target for new therapeutics. *Trends Mol Med*. 2005;11:192–7.
- McClung JK, et al. Isolation of a cDNA that hybrid selects antiproliferative mRNA from rat liver. *Biochem Biophys Res Commun*. 1989;164:1316–22.
- Koushyar S, Jiang WG, Dart DA. Unveiling the potential of prohibitin in cancer. *Cancer Lett*. 2015;369:316–22.
- Peng YT, Chen P, Ouyang RY, Song L. Multifaceted role of prohibitin in cell survival and apoptosis. *Apoptosis*. 2015;20:1135–49.
- Gamble SC, et al. Androgens target prohibitin to regulate proliferation of prostate cancer cells. *Oncogene*. 2004;23:2996–3004.
- Canevari RA, et al. Identification of novel biomarkers associated with poor patient outcomes in invasive breast carcinoma. *Tumour Biol*. 2016;37:13855–70.
- Kang X, et al. Prohibitin: a potential biomarker for tissue-based detection of gastric cancer. *J Gastroenterol*. 2008;43:618–25.
- Jiang L, Dong P, Zhang Z, Li C, Li Y, Liao Y, Li X, Wu Z, Guo S, Mai S, et al. Akt phosphorylates Prohibitin 1 to mediate its mitochondrial localization and promote proliferation of bladder cancer cells. *Cell Death Dis*. 2015;6: e1660.
- Polier G, et al. The natural anticancer compounds rocaglamides inhibit the Raf-MEK-ERK pathway by targeting prohibitin 1 and 2. *Chem Biol*. 2012;19: 1093–104.
- Qureshi R, Yildirim O, Gasser A, Basmadjian C, Zhao Q, Wilmet JP, Désaubry L, Nebigil CG. FL3, a synthetic Flavagline and ligand of Prohibitins, protects cardiomyocytes via STAT3 from doxorubicin toxicity. *PLoS One*. 2015;10(11): e0141826.
- Basmadjian C, Thuaud F, Ribeiro N, Désaubry L. Flavaglines: potent anticancer drugs that target prohibitins and the helicase eIF4A. *Future Med Chem*. 2013;5(18):2185–97.
- Kim S, Salim AA, Swanson SM, Kinghorn AD. Potential of cyclopenta[b]benzofurans from *Aglaia* species in cancer chemotherapy. *Anti Cancer Agents Med Chem*. 2006;6:319–45.
- Thuaud F, Bernard Y, Türkeri G, Dirr R, Aubert G, Cresteil T, Baguet A, Tomasetto C, Svitkin Y, Sonenberg N, et al. Synthetic analogue of rocaglaol displays a potent and selective cytotoxicity in cancer cells: involvement of apoptosis inducing factor and caspase-12. *J Med Chem*. 2009;52(16):5176–87.
- McClung JK, Jupe ER, Liu XT, Dell'Orco RT. Prohibitin: potential role in senescence, development, and tumor suppression. *Exp Gerontol*. 1995;30: 99–124.
- Thuaud F, Ribeiro N, Nebigil CG, Désaubry L. Prohibitin ligands in cell death and survival: mode of action and therapeutic potential. *Chem Biol*. 2013;20: 316–31.
- Zhou TB, Qin YH. Signaling pathways of prohibitin and its role in diseases. *J Recept Signal Transduct Res*. 2013;33:28–36.
- Ande SR, Mishra S. Prohibitin interacts with phosphatidylinositol 3,4,5-triphosphate (PIP3) and modulates insulin signaling. *Biochem Biophys Res Commun*. 2009;390:1023–8.
- Malumbres M, Barbacid M. Cell cycle, CDKs and cancer: a changing paradigm. *Nat Rev Cancer*. 2009;9:153–66.
- Chen RY, Fan YM, Zhang Q, Liu S, Li Q, Ke GL, Li C, You Z. Estradiol inhibits Th17 cell differentiation through inhibition of ROR γ T transcription by recruiting the ER α /REA complex to estrogen response elements of the ROR γ T promoter. *J Immunol*. 2015;194(8):4019–28.
- Dai L, Liu Y, Liu J, Wen X, Xu Z, Wang Z, Sun H, Tang S, Maguire AR, Quan J, et al. A novel cyclinE/cyclinA-CDK inhibitor targets p27(Kip1) degradation, cell cycle progression and cell survival: implications in cancer therapy. *Cancer Lett*. 2013;333(1):103–12.
- Park HY, Kim GY, Moon SK, Kim WJ, Yoo YH, Choi YH. Fucoidan inhibits the proliferation of human urinary bladder cancer T24 cells by blocking cell cycle progression and inducing apoptosis. *Molecules*. 2014;19(5):5981–98.
- Cell Cycle of PCR Arrays in QIAGEN Website. SABiosciences Corporation, Maryland, US. 2017. <https://www.sabiosciences.com/cn/shop/pcr/primer-sets/rt2-profiler-pcr-arrays/?catno=PAHS-020Z#geneglobe>. Accessed 21 Aug 2017.
- Wang XW, et al. GADD45 induction of a G2/M cell cycle checkpoint. *Proc Natl Acad Sci U S A*. 1999;96:3706–11.
- Ande SR, Nguyen KH, Nyomba BL, Mishra S. Prohibitin in adipose and immune functions. *Trends Endocrinol Metab*. 2016;27:531–41.
- Chowdhury I, Thomas K, Thompson WE. Prohibitin (PHB) roles in granulosa cell physiology. *Cell Tissue Res*. 2016;363:19–29.
- Li L, Guo JD, Wang HD, Shi YM, Yuan YL, Hou SX. Prohibitin 1 gene delivery promotes functional recovery in rats with spinal cord injury. *Neuroscience*. 2015;286:27–36.
- Chowdhury I, Thompson WE, Thomas K. Prohibitins role in cellular survival through Ras-Raf-MEK-ERK pathway. *J Cell Physiol*. 2014;229:998–1004.
- Luan Z, He Y, Alattar M, Chen Z, He F. Targeting the prohibitin scaffold-CRAF kinase interaction in RAS-ERK-driven pancreatic ductal adenocarcinoma. *Mol Cancer*. 2014;13:38.
- Yurugi H, Marini F, Weber C, David K, Zhao Q, Binder H, Désaubry L, Rajalingam K. Targeting prohibitins with chemical ligands inhibits KRAS-mediated lung tumours. *Oncogene*. 2017;
- Ribeiro N, et al. Flavaglines as potent anticancer and cytoprotective agents. *J Med Chem*. 2012;55:10064–73.
- Callahan KP, et al. Flavaglines target primitive leukemia cells and enhance anti-leukemia drug activity. *Leukemia*. 2014;28:1960–8.
- Bahrami A, et al. The potential value of the PI3K/Akt/mTOR signaling pathway for assessing prognosis in cervical cancer and as a target for therapy. *J Cell Biochem*. 2017;
- Nan Y, et al. Combinatorial therapy with adenoviral-mediated PTEN and a PI3K inhibitor suppresses malignant glioma cell growth in vitro and in vivo by regulating the PI3K/AKT signaling pathway. *J Cancer Res Clin Oncol*. 2017;
- Lien EC, Dibble CC, Toker A. PI3K signaling in cancer: beyond AKT. *Curr Opin Cell Biol*. 2017;45:62–71.
- Mayer IA, Arteaga CL. The PI3K/AKT pathway as a target for cancer treatment. *Annu Rev Med*. 2016;67:11–28.

39. Han EK, Mcgonigal T, Butler C, Giranda VL, Luo Y. Characterization of Akt overexpression in MiaPaCa-2 cells: prohibitin is an Akt substrate both in vitro and in cells. *Anticancer Res.* 2008;28:957–63.
40. Berger KH, Yaffe MP. Prohibitin family members interact genetically with mitochondrial inheritance components in *Saccharomyces Cerevisiae*. *Mol Cell Biol.* 1998;18:4043–52.
41. Bogenhagen DF, Wang Y, Shen EL, Kobayashi R. Protein components of mitochondrial DNA nucleoids in higher eukaryotes. *Mol Cell Proteomics.* 2003;2:1205–16.
42. Zhan Q, et al. Association with Cdc2 and inhibition of Cdc2/cyclin B1 kinase activity by the p53-regulated protein Gadd45. *Oncogene.* 1999;18:2892–900.
43. Malumbres M, Barbacid M. Mammalian cyclin-dependent kinases. *Trends Biochem Sci.* 2005;30:630–41.
44. Salvador JM, Brown-Clay JD, Fornace AJ. Gadd45 in stress signaling, cell cycle control, and apoptosis. *Adv Exp Med Biol.* 2013;793:1–19.

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