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Meaning of xylan acetylation on xylan-cellulose interactions: a quartz crystal microbalance with dissipation (QCM-D) and molecular dynamic study

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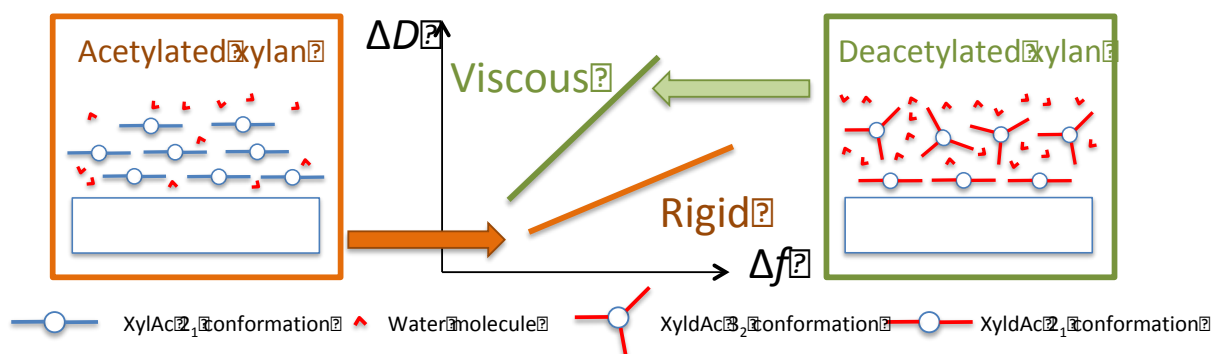
ABSTRACT

In plant cell walls, xylan chains present various substituents including acetate groups. The influence of the acetyl substitution on the organization of xylan-cellulose complexes remains poorly understood. This work combines *in vitro* and *in silico* approaches to decipher the functional role of acetyl groups on the xylan/cellulose interaction. Acetylated xylans were extracted from apple pomace with dimethyl sulfoxide-lithium chloride (DMSO-LiCl) and deacetylated using a mild alkali treatment. The adsorption behavior of acetylated and deacetylated xylan fractions was investigated using quartz crystal microbalance with dissipation (QCM-D) and molecular dynamics. Acetylated xylans form a dense and poorly hydrated and rigid layer on cellulose with xylan chains that have two residues per helical turn conformation, whereas the deacetylated fraction forms a swollen and more viscous layer in which only the xylan chains in direct contact with the cellulose surface have two residues per helical turn conformation. The other chains have three residues per turn conformation.

Keywords:

Xylan; acetyl group; cellulose nanocrystal; quartz crystal microbalance with dissipation (QCM-D); molecular dynamics; hemicellulose.

Graphical abstract



41 INTRODUCTION

42 Plant cell walls are highly complex materials that consist of an intricate network of
43 biopolymers whose main components are polysaccharides. These components include
44 cellulose, pectin and hemicellulose. As a result of their many uses, their structure and function
45 *in planta* has been the focus of many studies (Rojas, 2016; Voragen, Schols, & Visser, 2003).
46 Hemicellulose represents a group of various polysaccharides traditionally extracted by
47 alkaline treatment from plant cell walls. These polysaccharides in dicots are based on a β -(1-
48 4) linked backbone of xylose or glucose and/or mannose, forming xyloglucan, xylan, mannan
49 and glucomannan families (Scheller & Ulvskov, 2010). They exhibit large variations in
50 composition and structure depending on their botanical origin, tissue type and developmental
51 stage. This is particularly true of their backbone substitutions with side chains of saccharide
52 units, hydroxycinnamic acids, methyl ether and acetyl ester groups (Brummell & Schröder,
53 2009; Gatenholm & Tenkanen, 2004; Scheller & Ulvskov, 2010). Specific substitution
54 patterns and their role *in planta* remain unclear. Acetyl esterification is a ubiquitous
55 substitution within hemicellulose families. It usually occurs on O-2 and/or O-3 of xylose in
56 xylan, on O-6 of galactose and/or mannose, and O-2 and/or O-3 of mannose in
57 mannan/glucomannan (Melton, Smith, Ibrahim, & Schröder, 2009; Sims, Craik, & Bacic,
58 1997), and on fucose and/or O-6, O-4 or O-3 of galactose in xyloglucan, although the exact
59 position has yet to be fully established (Quemener et al., 2015; York, Oates, van Halbeek,
60 Darvill, & Albersheim, 1988). The alteration of xylan acetyl esterification *in planta* severely
61 impacts the plant phenotype and its resistance to biotic stresses (Escudero et al., 2017;
62 Grantham et al., 2017; Kumar, Campbell, & Turner, 2016; Yuan, Teng, Zhong, & Ye, 2016).
63 From the applied point of view, decreasing xylan acetyl esterification improves enzymatic

saccharification in the lignocellulosic biomass biorefinery (Kumar et al., 2016; P. M. Pawar et al., 2016).

Accordingly, understanding the role of the hemicellulose structure in terms of its interactions with cellulose and its consequences on cell wall architecture and properties remains a key question in plant development and applications. The present study focuses on the role of acetyl ester decorations on xylan interactions with cellulose by combining *in vitro* and *in silico* experiments to assess the adsorption features of acetylated and deacetylated xylans.

In the case of *in vitro* experiments, a major challenge is the availability of unmodified xylan fractions since acetyl esters are lost when using classical alkaline hemicellulose extractions. However, acetylated xylans can be recovered from wood biomass using DMSO extraction, steam explosion or organosolv pulping (Gatenholm & Tenkanen, 2004). In rich primary cell wall biomass such as fleshy fruit, acetyl-esterified hemicellulose is recovered after the partial removal of pectin in the cell wall via dimethyl sulfoxide doped by lithium chloride (DMSO-LiCl) (Assor, Quemener, Vigouroux, & Lahaye, 2013; Ray, Vigouroux, Quemener, Bonnin, & Lahaye, 2014).

In this study, interactions of acetylated and deacetylated xylan-enriched fractions obtained by this approach from apple pomace were assessed on a cellulose model surface using quartz crystal microbalance and dissipation (QCM-D). QCM-D is a highly effective mass-sensing technique based on the piezoelectric properties of quartz crystals. It has been successfully used to monitor the hemicellulose adsorption on the cellulose nanocrystal model surface. In fact, it provides information about the mass adsorbed on the surface as well as the mechanical behavior of the layer through the measurement of the dissipation factor (Bensselfelt et al., 2016; Eronen, Osterberg, Heikkinen, Tenkanen, & Laine, 2011; Kohnke, Ostlund, & Brelid, 2011; Villares, Bizot, Moreau, Rolland-Sabate, & Cathala, 2017; Villares, Moreau, Capron, & Cathala, 2014; Villares, Moreau, Dammak, Capron, & Cathala, 2015). Recently, Villares et

al. reported the kinetically-driven xyloglucan adsorption on the cellulose surface and described a two-regime mechanism (Villares et al., 2015). A similar approach was applied to investigate the adsorption behavior of acetylated and deacetylated xylan-enriched fractions referred to as XylAc and XyldAc, respectively, which display different behaviors. As a complement to the *in vitro* experiment, molecular dynamics (MD) were used to explore the conformation of xylan chains at the surface of cellulose crystals and to assess their interactions with water.

MATERIAL AND METHODS

Chemicals

Dimethyl sulfoxide (DMSO), lithium chloride (LiCl), 37% hydrochloric acid (HCl), orcinol, standard monosaccharides, sodium tetraborate, 1-methylimidazol, acetic anhydride, sodium borohydride, ammonia 25%, hydrogen peroxide and dichloromethane were provided by Sigma-Aldrich Chimie (Saint-Quentin Fallavier, France). Poly(allylamine hydrochloride) (PAH) was purchased by PolySciences (Mw 120-200 000 g mol⁻¹; sodium hydroxide (50%) (NaOH) by Fluka (Sigma-Aldrich Chimie, Saint-Quentin Fallavier, France); m-hydroxydiphenyl (MHDP, 3 phenylphenol) by Acros Organics (Geel, Belgium); sulfuric acid by Fisher Scientific, Illkirch, France; and ethanol 99% by VWR (Fontenay-sous-Bois, France).

Cellulose Nanocrystal (CNC) Suspension

Cellulose nanocrystals (CNCs) were obtained by sulfuric acid hydrolysis of cotton linters according to Revol et al. (Revol, Bradford, Giasson, Marchessault, & Gray, 1992). Cotton linters were hydrolyzed with 65% wt H₂SO₄ at 65°C for 35 min (12 g cotton linters/170 mL H₂SO₄). Immediately following hydrolysis, the suspension was diluted 10-fold to stop the

reaction and then washed by centrifugation at 10000 rpm for 10 min in order to concentrate the cellulose and to remove the excess aqueous acid. The resulting precipitate was repeatedly rinsed with ultrapure water and recentrifuged until a colloidal suspension was obtained. The colloidal suspension was then dialyzed again with distilled water until neutrality (dialysis membranes MWCO 12-14kDa, Fisher Scientific), after which a mixed-bed research-grade resin (1 g resin/400 g CNC suspension; Sigma-Aldrich TMD-8) was added to the suspension for 48 h to remove residual ions. The final aqueous suspension was 2% concentration by weight (~45% yield). The average crystal dimensions (length and cross-section) measured by tapping-mode atomic force microscopy (AFM) were 100-200 nm x 6-8 nm. Charges determined by conductometric titration were equal to 0.098 e/nm².

Hemicellulose extraction

Native hemicellulose enriched fractions were sequentially extracted from apple pomace obtained from the “Institut Français des Produits Cidricoles” (IFPC, Le Rheu, France), according to the procedure described by Ray et al. (Ray et al., 2014). First, the pectin was partially extracted from fresh apple pomace (15.6 kg) with 37.5 L of a potassium oxalate solution (60 mM, pH 4.7, 16 h) at room temperature (~20 °C) and with 3 x 24 L of water (2 x 3 h and 16 h) at 80 °C. After each extraction, the insoluble residue was recovered by manually pressing through a nylon mesh. The final residue was freeze-dried, weighed and ground (yield: 79% initial DW). The insoluble residue (200 g) was then incubated with 4 L of DMSO-LiCl (8.4 wt%; 5 h at 100 °C) under gentle agitation and nitrogen flow. After centrifugation (11000 g, 15 min, 20 °C), the supernatant was evaporated to dryness using a rotary evaporator at 70 °C and the dried residue was dissolved in 300 mL of water. Native hemicellulose was then precipitated with four volumes of ethanol at 4 °C for 16 h. After centrifugation (11000 g, 15 min, 20 °C), the dried pellet was weighed and the yield of

DMSO-LiCl extraction was calculated as 5% initial DW. Sixteen g of native hemicellulose were fractionated by anion exchange chromatography (DEAE- Sepharose Fast Flow column, 5 cm × 15 cm, GE Healthcare, Uppsala, Sweden) (Ray et al., 2014). Elution was performed at a flow rate of 2 mL min⁻¹ with water (two column volumes, 600 mL) to recover the unbound fraction. Bound polysaccharides were eluted using a gradient (six column volumes) from 0 to 1.0 M NaCl. XylAc was recovered in the second carbohydrate eluting peak (~400 mL elution, ~0.25 M NaCl) determined by colorimetry (see below), dialyzed and freeze-dried (yield: 500 mg, i.e., 0.14% initial DW). An additional removal of pectin was then performed by pectin lyase (0.12 U, 3 h at 40 °C, [EC 4.2.2.10] purified from Peclyve Lyven, France) (Ralet et al., 2012). The XylAc fraction (100 mg) was deacetylated by saponification with 20 mL of 0.5 M NaOH (1 h at 4 °C). After neutralization with HCl, the **deacetylated fraction, referred to as XyldAc**, was desalted by dialysis against deionized water (MWCO 3500 Da, CelluSep H1) and freeze-dried in order to obtain (the yield is greater than 90% DW of the XylAc fraction).

Hemicellulose characterization

The total neutral sugar content was determined by colorimetry with an automated orcinol/sulfuric acid assay (Tollier & Robin, 1979). Glucose was used as a standard.

Uronic acid content was measured by colorimetry using m-hydroxydiphenyl and concentrated sulfuric acid hydrolysis (Blumenkrantz & Asboe-Hansen, 1973; Thibault, 1979). The distinction between galacturonic acid and glucuronic acid was made by comparing colorimetric responses with and without the addition of sodium tetraborate in the acid, according to the procedure used by Filisetti-Cozzi, & Carpita (1991). Galacturonic and glucuronic acids were used as the standard.

Identification and quantification of neutral sugars were performed by gas-liquid chromatography on a TG-225 GC Column (30 x 0.32 mm ID) using a TRACE™ Ultra Gas Chromatograph (Thermo Scientific™; column temperature: 205 °C; split injector temperature: 220 °C; flame ionization detector temperature: 250 °C; carrier gas: H₂) after sulfuric acid degradation and derivatization as alditol acetates (Dheilly et al., 2016).

Acetyl ester content was measured by HPLC as the amount of acetic acid released by saponification of a 5-mg sample in 1 mL 0.5 M NaOH for 1 h at 4 °C. HPLC was carried out on a C18 column (4 mm × 250 mm, Lichrospher 100 RP-18e (5 µm), Interchim, France) thermostated to 25 °C using a refractometric detector (Waters, 2414). An isocratic elution of 4 mM H₂SO₄ was used at a flow rate of 1.0 mL min⁻¹ (Levigne, Thomas, Ralet, Quemener, & Thibault, 2002).

Surface preparation

Piezoelectric AT-cut quartz crystals coated with gold electrodes on each side (QSX301, Q-Sense) were used for QCM-D experiments. Silicon wafers were used for AFM experiments. Quartz crystals and silicon wafers were cleaned with H₂SO₄/H₂O₂ (7:3, v/v), rinsed exhaustively with Milli-Q water, and dried under a nitrogen stream. Prior to use, QCM-D quartz sensors were subjected to a plasma-etching device (Harrick Plasma). CNC surfaces on quartz crystals were prepared by the spin-coating method, as previously described (Villares et al., 2015). A CNC dispersion at 3 g L⁻¹ was dropped onto a pre-coated substrate with poly(allylamine hydrochloride) at 1 g L⁻¹ in water and accelerated from 180 rpm to 3600 rpm after 5 min of adsorption and maintained for 60 s at that speed using an expressly designed spin-coater. The thickness of the CNC layer, evaluated by ellipsometry, was 9 nm ± 5 nm, in agreement with previous reports (Villares et al., 2017; Villares et al., 2015). Xylan solutions

at 1 g L⁻¹ were then dropped onto wafers pre-coated with CNCs in order to perform AFM experiments.

Quartz crystal microbalance with dissipation (QCM-D) experiments

QCM-D experiments were carried out using a Q-Sense E4 instrument (AB, Sweden). Spin-coated CNC surfaces were mounted in the QCM-D cells at 20 °C and equilibrated with water at a flow rate of 50 µL min⁻¹ until the resonance response was stable. The frequency and dissipation signals were then offset to zero just before the measurement.

Frequency ($\Delta f_n/n$) and dissipation (ΔD_n) changes were simultaneously registered at 5 MHz fundamental resonance frequency ($n = 1$) and its several overtones as a function of time ($n = 3, 5, 7$, etc.). The third overtone (15 MHz, $n = 3$) was used to evaluate the QCM-D data.

Xylan solutions at different concentrations (0.3 to 20 µg mL⁻¹) were injected at 50 µL min⁻¹ and allowed to adsorb for 40 min before being exposed again to a water flow in order to remove loosely-bound material. Each xylan solution at the desired concentration was adsorbed on a freshly-prepared CNC modified surface and the experiments were repeated at least three times. All QCM-D experiments were conducted at a constant temperature of 20 °C. The surface concentration (Γ) can be calculated from the mass variation during adsorption obtained from the frequency change ($\Delta f_n/n$) by using the Sauerbrey equation (Sauerbrey, 1959):

$$\Delta m = - C \frac{\Delta f_n}{n}$$

where C is the Sauerbrey constant: constant for the mass sensitivity of the quartz crystal (0.177 mg m⁻² Hz⁻¹ for 5-MHz crystals); and n is the overtone number. Strictly speaking, the Sauerbrey equation is valid for ultrathin rigid films whose frequency and dissipation changes only depend on the mass of the film, especially for adsorbed polymers. A thin film can be considered to be fully elastic and rigid when $\Delta D \leq 1 \cdot 10^{-6}$ and no clear spreading of the

overtones in the $\Delta f_n/n$ and ΔD_n graphs can be detected. Under such conditions, the Sauerbrey equation is valid for estimating the mass changes on the sensor surface.

A complete description of the kinetic modeling approach is described in Appendix 1 of the Supplementary Materials (SM).

Characterization of CNC surfaces coated with xylan fractions

AFM: Topographical images of silicon wafers were recorded by atomic force microscopy (AFM) using an Innova AFM (Bruker). The images were collected in tapping mode under ambient air conditions (temperature and relative humidity) using a monolithic silicon tip (TESPA, Bruker) with a spring constant of 42 N m⁻¹ and a nominal frequency of 320 kHz. Image processing was performed using WSxM 5.0 software.

Molecular dynamics (MD) simulations

The models considered in this study consisted of three molecular species: a cellulose microfibril, xylan chains and water molecules. The cellulose microfibril model contained 18 chains of cellulose (Kubicki et al., 2018) in the I β organization and exposed the (110), (1-10) and (100) surfaces (Mazeau, 2011). The two primitive cell chains of the I β allomorph (Nishiyama, Langan, & Chanzy, 2002) were first duplicated through space, and a P₁ periodic super-crystal was redefined, which consisted of 6x6 cellulose chains. Each individual chain consisted of six anhydroglucose units and was covalently bonded to its own periodic image, mimicking a chain of infinite length. The supercell was then equilibrated using NPT-MD (constant number of particles, pressure and temperature) at 300 °K and 1 atm. The **a** and **b** parameters of the periodic cell were then increased to 50 Å and the excess cellulose chains were removed to retain only 18 chains. The **c** parameter was maintained at its equilibrium value of 31.4 Å. A charged sulfate group (SO₃⁻) was added on an O-6 oxygen atom of the (1-

10) surface to model a cellulose crystal prepared by sulfuric acid, and a Na⁺ cation was also inserted in the simulation box to ensure the neutrality of the system. The model is infinitely long in the longitudinal direction but finite in the perpendicular direction. This model was then hydrated using the simple three-point-charges water model (Berendsen, Postma, Van Gunsteren, & Hermans, 1981) and equilibrated by 50 ns of NVT-MD (constant number of particles, volume and temperature) at 400 °K. This equilibration resulted in a conformational change of the hydroxyl and hydroxymethyl groups exposed to the water environment, the hydroxyl groups created hydrogen bonds with the water molecules, and the surface hydroxymethyl groups rotated from the *tg* orientation to a mixture of the *gg* and *gt* orientations. The water molecules were then removed to generate the initial models of the cellulose-xylan complexes.

Xylan chains consisted exclusively of six skeletal anhydroxylose units. They are either non-substituted or per-acetylated. In the latter case, all of the units were O-acetylated at the O-2 and O-3 oxygen atoms. Two helical conformations, 2_1 and 3_2 , of the xylan skeleton were considered. Their coordinates were extracted from a previous study (Mazeau, Moine, Krausz, & Gloaguen, 2005). For the two helical conformations of the per-acetylated xylan chain, the initial orientation of the acetyl groups corresponded to the lowest energy extracted from a Monte-Carlo sampling of 1000 conformations on an isolated chain with a rigid backbone (SM, Appendix 2).

The xylan chains were inserted in the simulation box around the cellulose microfibril. They were aligned with the microfibril, either parallel or antiparallel at random. Similar to the cellulose chains, the xylan chains were linked to their periodic images by covalent bonds in order to mimic infinitely long chains. The two acetylated models, XylAc, consisted of 20 chains of fully acetylated xylan chains (total mass: 25920 g mol⁻¹), whereas the two non-acetylated models, XyldAc, consisted of 33 chains of non-substituted xylan chains (26136 g

mol⁻¹). Water molecules were then added in the remaining empty spaces of the simulation boxes.

The models were then subjected to energy minimizations followed by MD simulations. The first energy minimization was performed while maintaining the cellulose and xylan chains frozen; only the positions of the water molecules were refined at this step. All the atoms were free to move in the second minimization step. The MD simulations were performed in the NVT ensemble at 400 °K (50 ns), and then at 298 °K (10 ns). The coordinates of the cellulose microfibril were frozen during the MDs. MD simulations were long enough to reach equilibration when the energy, location and conformation of the xylan chains were stable. The last ns of the room-temperature trajectories were used for analysis.

Computational details

Material Studio modeling software (v5.5) was used (Accelrys, Inc., San Diego, CA, USA). Energy calculations were performed using the Universal Force Field (Rappe, Casewit, Colwell, Goddard, & Skiff, 1992). This all-atoms force field was applied in our previous study of cellulose-xylan complexes (Mazeau & Charlier, 2012) and it was also used for the development of a coarse grain force field (Li, Perre, Frank, & Mazeau, 2015). The charge equilibration method was used to calculate partial charges for each atom (Rappe & Goddard, 1991). The long-range interactions were calculated using the particle-mesh Ewald summation method (Darden, York, & Pedersen, 1993) with a cut-off distance of 9 Å.

Energy minimization was performed by the steepest descent method and then by the conjugated gradient method. The convergence criterion was a root-mean-square (rms) force of less than 0.1 kcal.mol⁻¹.Å⁻¹. The integration of Newton's laws of motion in the MD simulations was performed using the standard Verlet algorithm (Verlet, 1967) with a time step

of 0.001 ps. The total external pressure was maintained at 1 atm (Berendsen et al., 1981) and a Hoover algorithm (Evans & Holian, 1985) was used to keep the cell temperature constant.

RESULTS

XylAc and XyldAc fractions extraction and characterization

Investigation of the hemicellulose function in the plant cell wall requires careful extraction in order to provide a structure representative of those naturally occurring *in planta*. However, hemicellulose is classically extracted with alkali, leading to the hydrolysis of the acetyl ester groups. In a previous study, a non-aqueous polar aprotic solvent, namely DMSO doped with LiCl, was used to efficiently extract acetyl-esterified hemicelluloses (Assor et al., 2013; Ray et al., 2014). Applying this extraction procedure to apple pomace after partial removal of pectin followed by anion exchange chromatography allowed the recovery of a xylan-enriched fraction (XylAc). Fig. S1 reviews the overall extraction process. The extract was composed in decreasing order of weight proportion as follows: xylose, uronic acid, arabinose and acetyl esters, as well as low amounts of glucose, galactose and mannose and traces of fucose (Table 1). Mild alkali treatment of the fraction succeeded in removing acetyl esters with limited changes in the sugar composition (Table 1). The extract was thus mainly enriched in arabinoxylan (GAX), whose composition was close to that reported for Gala GAX (Ray et al., 2014) except for the presence of glucuronic acid, though comparatively richer in acetyl esters (7.2% vs. 3.8% in Gala). The presence of low amounts of glucose, mannose, galactose and fucose indicates the likely contamination of the AX by some galactoglucomannan and xyloglucan, whereas the galacturonic acid and rhamnose content suggests the presence of remaining pectin. Assuming that all acetyl esters were related to the xylan and were linked to the xylose residue, there would be one acetyl group for approximately two xylose residues, in

agreement with the average distribution of acetyl ester in plant xylan (Busse-Wicher et al., 2014; Chong et al., 2014; Grantham et al., 2017). Glucuronic acid and arabinose are present in the proportion of one uronic acid for approximately three xyloses, instead of one to six xyloses in the *Arabidopsis* primary wall and one arabinose for approximately 3.4 xylose residues, which could account in part for the pentose unit linked to glucuronic acid in the *Arabidopsis* primary wall xylan (Mortimer et al., 2015).

Table 1. Chemical composition of the acetylated (XylAc) and deacetylated (XyldAc) xylan fractions. Total sugars (TS) and the acetyl group contents are expressed in wt% and monosaccharide content is expressed in mol%.

	<i>Acetyl</i>	<i>TS</i>	<i>Glc</i>	<i>Gal</i>	<i>Man</i>	<i>Xyl</i>	<i>Ara</i>	<i>Fuc</i>	<i>Rha</i>	<i>GalA</i>	<i>GlcA</i>
	(wt%)						(mol%)				
XylAc	7.2	78.3	5.4	5.1	2.3	51.9	16.5	0.2	1.0	16.5	1.0
XyldAc	0	77.6	4.9	5.0	2.7	53.6	16.9	0.3	1.0	14.8	0.9

Adsorption of XylAc and XyldAc fractions on the cellulose model surface

Spin-coated CNCs prepared from cotton linters were used as a model surface. They displayed the expected rod-like morphology with dimensions of approximately 175 nm in length, 15 nm in width and 7 nm in thickness, consistent with previous reports (Cherhal, Cousin, & Capron, 2015). The ability of XylAc and XyldAc-enriched fractions to adsorb on the CNC surface was evaluated and compared by QCM-D. This technique relies on the properties of piezoelectric quartz sensors submitted to an electric field that induces mechanical shear wave of the crystal. Adsorption of any material on the quartz surface induces a shift in the frequency resonance (Δf_n), allowing real-time monitoring of polymer adsorption on the quartz surface. Thanks to its very high sensitivity, the technique has been successfully used to monitor a wide range of

adsorption processes. The second piece of information that can be obtained from the QCM-D analysis is the dissipation factor ΔD . This value provides an indication of the mechanical (viscoelastic) behavior of the layer adsorbed on the quartz since it reflects how the layer will dissipate the energy of the oscillation waves of the quartz sensor. QCM-D has already been used for the investigation of adsorption of xylan and xylan derivatives (Dammak et al., 2013; Kohnke et al., 2011; Sang Hoon Lee, Lee, & Youn, 2014; S. H. Lee, Lee, & Youn, 2015). Prior to QCM-D analysis, the xylan fractions were adsorbed on the CNC surfaces and the morphology was analyzed by atomic force microscopy (Fig. S2). In all the cases, the CNCs were easily visible and the complexes showed no clear differences with the initial CNC surface. This indicates that xylan is adsorbed as an individual polymer chain or, at most, as very small aggregates undetectable by AFM. In the following section, we report the frequency variations for different hemicellulose fractions, followed by an investigation of the dissipation patterns.

Frequency variation during XylAc and XyldAc adsorption and kinetic modeling of the adsorption behaviors

Prior to injection, the CNC surfaces were stabilized with water and the xylan fractions were injected at different concentrations ranging from $0.3 \mu\text{g mL}^{-1}$ up to $20 \mu\text{g mL}^{-1}$ to explore a large panel of ratios between polymers and adsorption sites. For the two fractions, acetylated and deacetylated, the decrease of frequencies observed, regardless of the concentration injected, indicated adsorption of xylan fractions on the cellulose surface. For each xylan concentration, adsorption was repeated at least three times to assess the reproducibility, and a typical frequency variation against time is reported in Fig. 1. Adsorption was irreversible since the rinsing steps applied after adsorption (indicated by arrows on Fig. 1) did not change the frequency values.

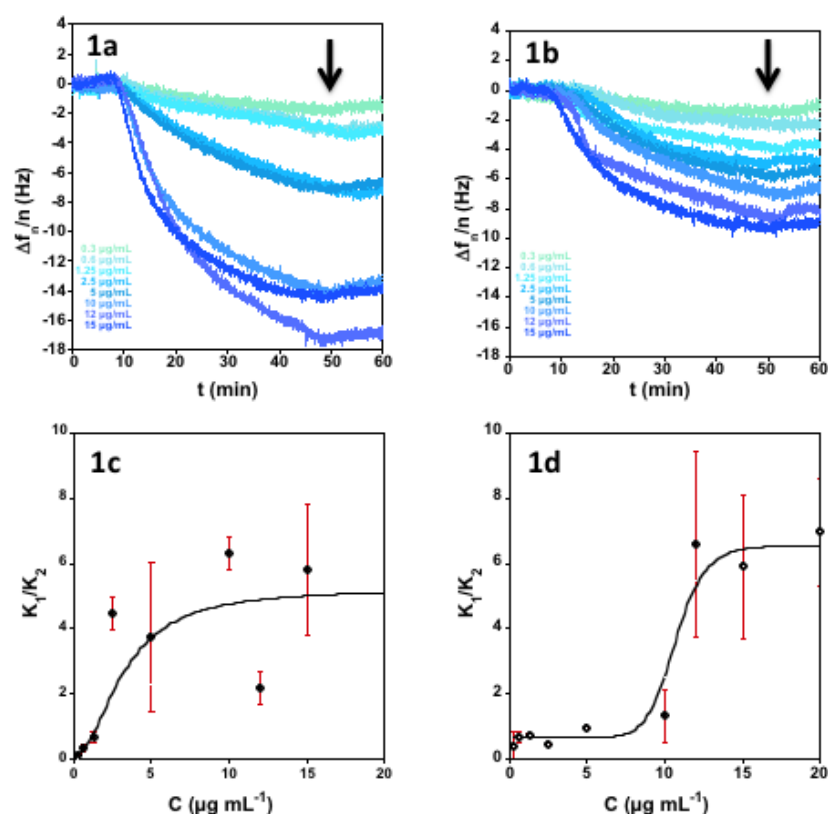


Fig. 1. Frequency (Δf_n) (3rd overtone) for the adsorption of XylAc (1a) and XyldAc (1b) fractions and evolution of the k_1/k_2 ratio as a function of the XylAc (1c) or XyldAc (1d) concentration (solid lines are visual guides). Arrows indicate the beginning of the rinsing step with water.

For the XylAc and XyldAc fractions, different maximal frequency values were obtained for all concentrations studied, indicating that the final amount of xylan adsorbed depended on the sample concentration applied, as previously reported for the xyloglucan adsorption on CNC surfaces (Villares et al., 2017; Villares et al., 2015). Frequency variations were in the same range (tens of Hz) for both xylan fractions, suggesting that the amounts adsorbed are similar, contrary to Kabel *et al* results that reported a higher adsorption of the deacetylated fraction. (Kabel, van den Borne, Vincken, Voragen, & Schols, 2007). This finding can be assigned to

a lower steric hindrance due to the removal of acetyl groups to increase the self-associative capacity of xylans, as reported earlier by Linder (Bosmans et al., 2014; Linder, Bergman, Bodin, & Gatenholm, 2003). Since both the XylAc and XyldAc fractions did not desorb from the surface during the time scale of the experiment, the maximal frequencies cannot be attributed to a dynamic equilibrium between adsorption and desorption of the polymer. Moreover, the continuous flow of XylAc and XyldAc on the surface guaranteed that the maximum coverage possible for each condition was reached. Thus, this pattern suggests that the final amount of polymer adsorbed depended on the competition between adsorption, packing optimization and potential rearrangement, suggesting that the adsorption process is therefore not the result of thermodynamic equilibrium but, instead, is kinetically-driven.

We previously described such behavior by using a kinetic model that comprises two steps: the first one describes the adsorption of hemicellulose onto an uncovered surface associated with the rearrangement of the chains to cover the surface, and the second corresponds to adsorption on a covered surface on which the polymer must diffuse through a pre-adsorbed hemicellulose layer (Villares et al., 2017). This kinetic model was used to determine k_1 and k_2 for the XylAc and XyldAc fractions that are the kinetic constant for the first and second adsorption steps, respectively. The detailed mathematical expressions of the kinetic model and typical fits obtained are given in the Supplementary Material (SM, Appendix 1). The experimental fractional coverage was calculated to obtain values of the kinetic constants k_1 and k_2 , and there was a good fit between experimental and calculated values (Fig. S3). The evolution of the ratio k_1/k_2 against the XylAc and XyldAc fraction concentration is plotted in Fig. 1c and d, respectively. The fractions display different patterns with two domains of concentrations.

At low concentration, the k_1/k_2 ratio is lower or close to one indicating that the kinetic adsorption and lateral rearrangement occur at a similar time scale, leading to large amounts of

extended xylan conformation or trains. At higher concentrations, adsorption occurs through diffusion on the pre-adsorbed layer, and the crowding of the surface limits diffusion and rearrangement mechanisms. Thus, the k_1/k_2 ratio increases since the adsorption process on a bare surface (k_1) dominates, while (k_2) decreases dramatically. The increase of the k_1/k_2 ratio associated with a higher amount of polymer adsorbed can be interpreted either by a denser packing of the molecules when the concentration increases or by limited rearrangement possibilities. A clear difference can be seen between acetylated and deacetylated xylan. In the case of the XylAc fraction, the k_1/k_2 ratio increases very rapidly, suggesting that even at low concentrations, the acetylated sample is able to efficiently cover the surface and the possibility of rearrangement seems to be strongly limited, suggesting efficient interactions between XylAc and the cellulose surface. In the case of XyldAc, a transition between the two regimes (i.e., the concentration at which the k_1/k_2 ratio increases) is clearly seen at around 5-10 $\mu\text{g mL}^{-1}$. This suggests that chain rearrangements can occur at low surface concentrations, whereas surface crowding may limit conformation modification at higher concentrations. Nevertheless, the presence of acetyl groups changes the adsorption behavior of the xylan chains and the acetylated fraction seems to have a greater ability to cover the surface than the deacetylated one.

Comparison of the viscoelasticity behaviors of adsorbed layers

To correlate the evolution in dissipation with changes in frequency, the ΔD was plotted as a function of Δf (hereafter referred to as ΔD vs. Δf plots) to eliminate time as an explicit parameter (Fig. 2). The raw dissipation data for both fractions at different concentrations are illustrated in Figs. 2a and 2b. The ΔD vs. Δf plots revealed different slopes of the curves that are related to the overall viscoelasticity of the layer. A rigid layer will behave similarly to the quartz dissipation and the ΔD will present low values per mass addition and, accordingly, the ΔD vs. Δf slope will be low. In the case of a more viscoelastic layer, the ΔD vs. Δf slope will

display higher values. As shown in Figs. 2c and 2d, the two xylan fractions display strikingly different behaviors. The slope obtained for XylAc is constant ($0.03 \cdot 10^{-6} \text{ Hz}^{-1}$) regardless of the concentration studied, indicating that irrespective of concentration, the adsorbed acetylated xylan layer on cellulose shares a similar organization. Moreover, the very low value of the slope suggests that the adsorbed layer is rigid. This finding is consistent with the kinetic analysis that displays only one regime of adsorption in which the k_l constant is dominant and which suggests that the XylAc layer is strongly adsorbed with limited diffusion and rearrangement processes. It should be emphasized that the slope value of the ΔD vs. Δf of XylAc is significantly lower compared to the values found in the literature. For example, Eeronen et al. (Eronen et al., 2011) reported a value of $0.25 \cdot 10^{-6} \text{ Hz}^{-1}$ for wheat arabinoxylans and of $0.1 \cdot 10^{-6} \text{ Hz}^{-1}$ for spruce galactoglucomanans, which are on the same order of magnitude as the value measured for the XyldAc but almost ten times higher than the value of $0.03 \cdot 10^{-6} \text{ Hz}^{-1}$ determined for the XylAc. This finding suggests that the mechanical properties of the XylAc layer are quite similar to those of the cellulose surface and can probably be attributed to flat and dense layers.

In contrast, the ΔD vs. Δf plot of XyldAc displays different slopes depending on the injection concentration, indicating a more complex behavior (Fig. 2d). The ΔD vs. Δf plot pattern evolved during the adsorption process, suggesting a change in layer organization during adsorption. At low concentrations ($<5 \mu\text{g mL}^{-1}$), the slope value is around $0.25\text{-}0.5 \cdot 10^{-6} \text{ Hz}^{-1}$, whereas at higher concentrations, the value decreases to $0.1\text{-}0.15 \cdot 10^{-6} \text{ Hz}^{-1}$. At low concentrations, the slope value is high, which indicates a high viscosity of the layer, and then starts to decrease to reach a stable value at a concentration of $5\text{-}10 \mu\text{g mL}^{-1}$. This suggests the formation of a loose layer that becomes denser as the concentration increases.

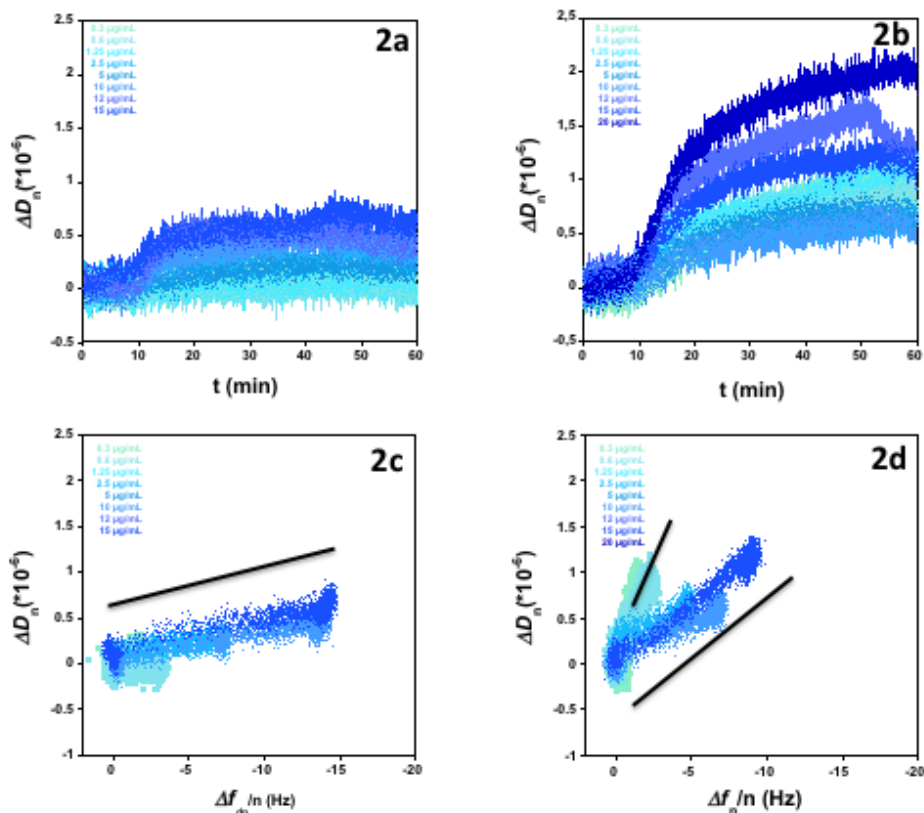


Fig. 2. Dissipation (ΔD) (3^{rd} overtone) for the adsorption of XylAc (2a) and XyldAc (2b) fractions for different concentrations as a function of time and corresponding ΔD vs. Δf plots for XylAc (2c) and XyldAc (2d) (solid lines are visual guides).

Adsorption of XylAc and XyldAc fractions on the cellulose model surface by MD

Four models of the hydrated cellulose xylan complex that differed by the chemical structure of their xylan chains and by their initial conformation were considered. The xylan chains were either acetylated or non-acetylated (corresponding to deacetylated sample). Repeat unit is shown in Fig S4. In addition, to account for the conformational polymorphism of xylan (Chanzy, Dube, & Marchessault, 1979; Larsson, 2004; Teleman, Larsson, & Iversen, 2001), the xylans differed by the initial helical conformation of their backbone: either two units per helical turn when the xylan chains contained three turns of the 2_1 helix in their initial state, or

three units per helical turn when they contained two turns of the left-handed 3_2 helix (Figs. S5 and S6).

The four models were subjected to MD simulations at two temperatures. During the high temperature phase, the xylan chains explored the two helical conformations with rapid inter-conversions between the two forms. This is not surprising since the two conformations have similar energies and the energy barrier between them, estimated from the adiabatic potential energy surface of xylobiose (Mazeau, Moine, Krausz, & Gloaguen, 2005), is lower than 2 kcal mol⁻¹. In contrast, during the room-temperature phase, each xylan chain stabilized in its preferred conformation depending on its immediate environment and remained in that conformation up to the end of the trajectory. As a consequence, the models that differed by the initial conformation of the xylan chains converged into an identical structure of the complex. Fig. 3 provides representative snapshots of the XylAc and XylAc models.

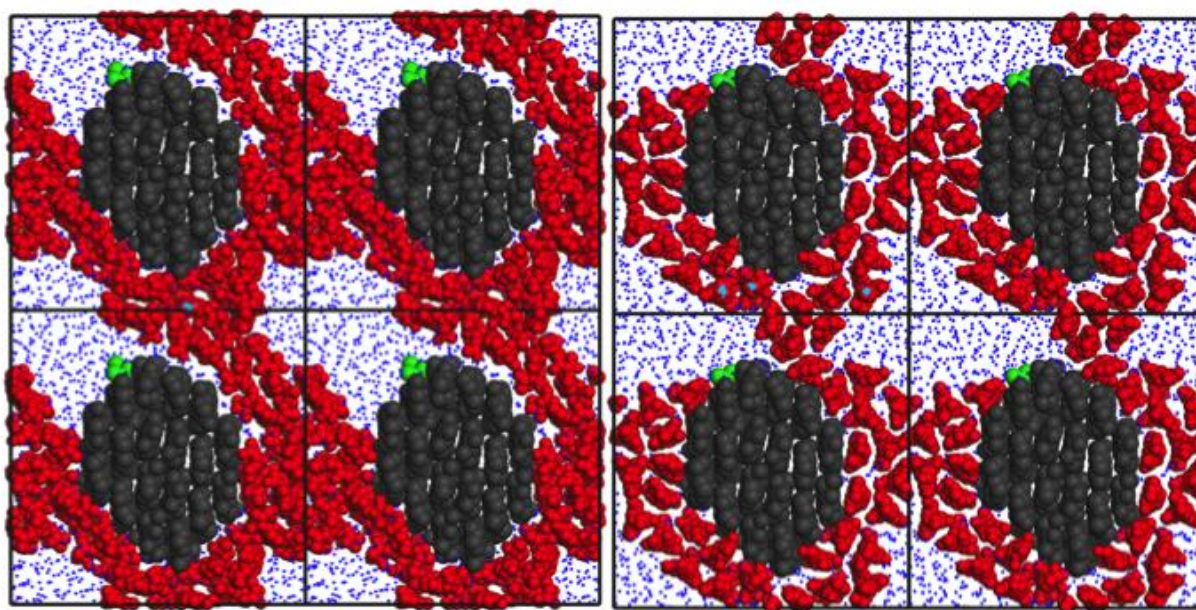


Fig. 3: Cross-sectional views of the models of the hydrated cellulose-xylan complexes. 2x2 basic cells are displayed. Hydrogen atoms are omitted for clarity. Cellulose is in gray. Sulfate group, in green, is located on a chain of the (1-10) surface of cellulose next to its top corner.

471 The xylan chains are in red and the water molecules in blue. Left: XylAc model; right XyldAc
472 model.

473

474 *Conformation of the xylan chains*

475 Analysis of the helical conformations of the xylan chains in the equilibrated models is
476 performed using the turn angle (T), which is defined by the sequence of atoms O-5''-C-
477 5''-C-4-C-5 (Fig. S4). The turn angle describes the orientation of a given unit with respect to
478 its second neighbor for those perfectly stereo-regular helices that were used to build the initial
479 models $T = -50^\circ$ for the 2_1 helix and $T=100^\circ$ for the 3_2 helix.

480 Fig. 4 gives the distributions of the turn angle of the xylan chains in the equilibrated XylAc
481 and XyldAc models. It shows the distributions of all the chains (denoted 'all'), together with
482 that of the chains in close proximity to the cellulose, denoted '1st layer', and that of the chains
483 far from the cellulose, denoted 'other layer'. For the purpose of comparison, the same
484 calculations were performed for arged cellulose, and similar distributions were obtained in all
485 cases (SM, Appendix 3 - Fig. S10 - S11-S12).

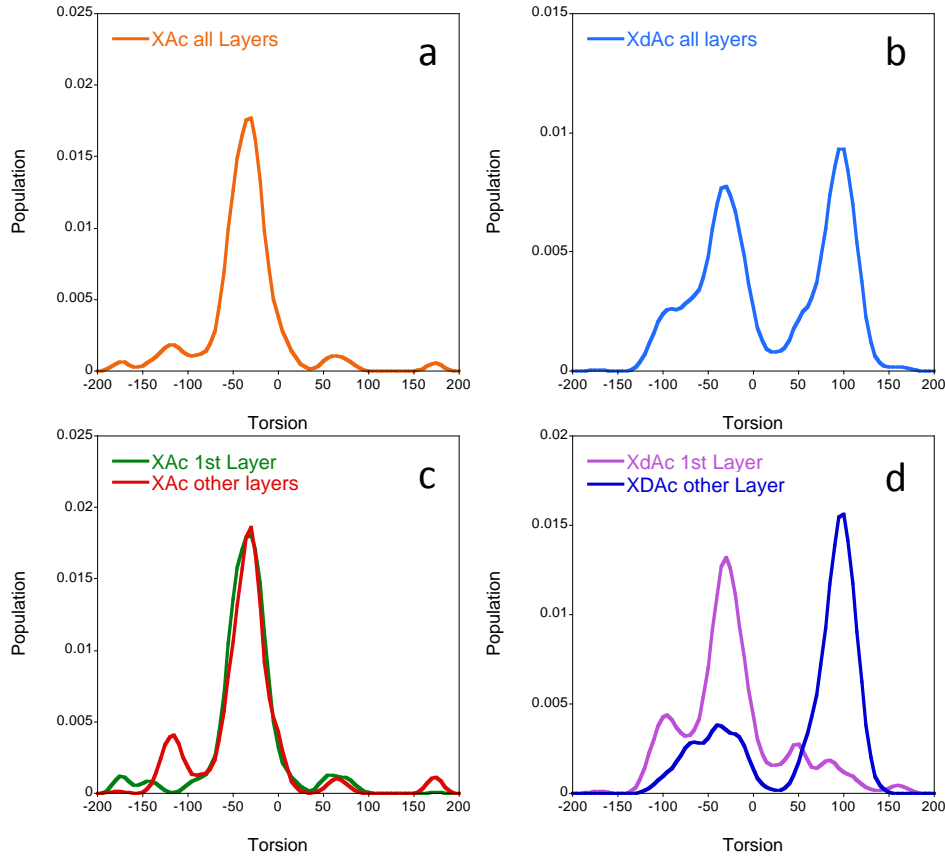


Fig. 4. Distribution of the turn angle in the XylAc (left) and XyldAc (right) simulated systems. Global distributions from all the xylan chains (4a and 4b); and distributions from the xylan chains interacting with cellulose (orange in 4c and light blue in 4d) and from the bulk xylan (red in 4c and dark blue in 4d).

For XylAc, the distribution is almost monomodal and centered at $T = -35^\circ$. Considering the thermal motion and local conformational variability, this peak corresponds to helices with two residues per helical turn. The distribution of T also reveals other forms, non-canonical, less organized and of low occurrence, at $T = -120^\circ$, 60° and 180° . Conformations of the xylan chains in the XyldAc model differ substantially. The bimodal distribution of the turn angle centered at $T = -35^\circ$ and $+95^\circ$ shows that the deacetylated xylan chains explore the two main 2_1 and 3_2 helical forms. It should be noted that most of the chains in direct interaction with

cellulose are mainly in the 2_1 conformation, whereas those far from the cellulose (in the bulk phase of xylan) are in the 3_2 conformation. Here again, a few chains explored alternative disorganized conformations, as suggested by the broad distribution of the turn angle.

Concentration profiles.

The concentration fluctuations of cellulose, xylan and water were determined along the 100 and 010 directions in both the immediate environment of the cellulose and in the bulk xylan (Fig. 5).

At the interface with cellulose, the acetyl groups do not affect the adsorption behavior of xylan. The concentration traces at 40 Å show a maximum for xylan and, concomitantly, a minimum for water, which is particularly visible on the profiles along the 100 direction. This reveals that xylan accumulates at the interface on the uncharged side of the cellulose crystal. The concentration profiles of xylan and water at the interface with cellulose are, however, dissymmetrical. At 10 Å in the plots, i.e., on the charged side of the cellulose, the xylan concentration is minimal, whereas that of water is maximal, which suggests that the charged sulfate group preferentially interacts with the surrounding water molecules rather than with xylans. Additional modeling was performed with a native (uncharged) cellulose crystal in order to estimate the exact effect of the charged sulfate group (see SM, Appendix 3, Figs. S11 and 12). The corresponding concentration traces with uncharged cellulose are in fact more symmetrical and show a high concentration of xylan and a low concentration of water at both interfaces. The charged sulfate group does not interact with the acetylated xylans but can occasionally form a hydrogen bond with a hydroxyl group of the deacetylated xylan. The charged sulfate group, along with its surrounding hydration shell, enhances the cellulose hydrophilicity, increases the porosity of the total system, and lowers the total cellulose surface covered by xylan.

In contrast, within the xylan phase, at around 0 or 50 Å in the concentration profiles, the acetyl groups have a strong influence on xylan organization. For XylAc, xylan concentration is similar to that of water, suggesting that the system is dense and weakly hydrated. This type of organization is obviously facilitated by the dominant 2_1 helical conformation of their backbones whose compact packing, similar to that of cellulose or chitosan, can be easily achieved. However, the multiple conformations explored by the acetyl groups disturb the perfection of the packing at the local level and create small cavities in which few water molecules can penetrate. For XyldAc, in contrast, xylan concentration is much lower than that of water, which suggests that the deacetylated xylan layer is loose and highly hydrated. The two free hydroxyl groups per repeat unit strongly interact with water molecules and the packing of the 3_2 helices promotes uninterrupted columns of water that appear in between the deacetylated xylan chains.

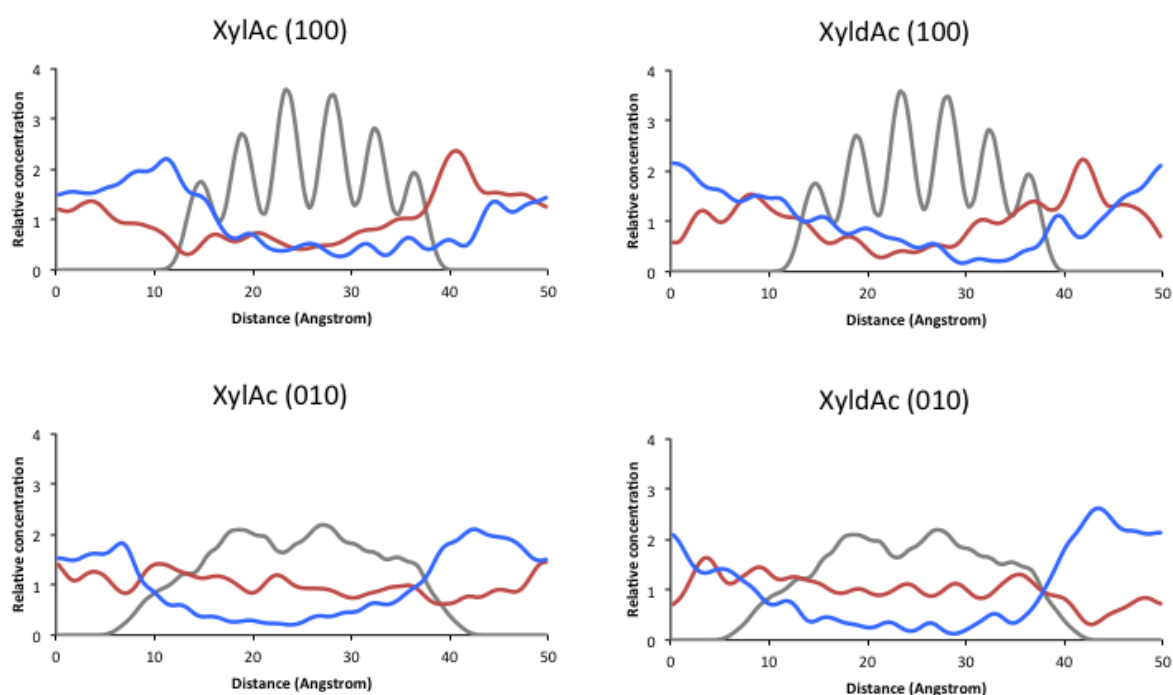


Fig. 5: Concentration profiles of cellulose, xylan and water along directions perpendicular to the exposed crystallographic planes of the cellulose. In gray: cellulose; in red: xylan; in blue: water.

539 Our models are consistent with experimental data from X-ray diffraction and solid-state
540 NMR. The crystal structure of xylan diacetate is anhydrous and made of two-fold helices
541 (Gabbay, Sundararajan, & Marchessault, 1972), whereas that of deacetylated xylan contains
542 between one to three water molecules per xylosyl unit and the chains are in the left-handed
543 three-fold helix (Nieduszynski & Marchessault, 1971; Nieduszynski & Marchessault, 1972).
544 These deacetylated xylan chains also explore the three-fold conformation in a water solution
545 but change their conformation to the two-fold one when interacting with cellulose (Larsson,
546 Hult, Wickholm, Pettersson, & Iversen, 1999). It should be noted that such conformational
547 adaptation is limited to those xylan chains in direct contact with the cellulose surface (Falcoz-
548 Vigne et al., 2017).

549 In a modeling study, the two limiting cases were studied where xylan chains are either per-
550 acetylated or deacetylated, whereas Busse-Wicher et al. modeled the adsorption of a partly-
551 acetylated xylan chain in which the distribution of the acetyl groups is stereoregular, i.e.,
552 where every other xylose residue is acetylated (Busse-Wicher et al., 2014). Comparing the
553 two studies makes it possible to determine the effects of the degree of acetylation in the xylan
554 chain on its interaction with cellulose. The degree of acetylation does not affect the
555 conformation of xylan in direct contact with cellulose, adopting the two residues per helical
556 turn conformation. Acetylation also does not affect the adsorption features of xylan on the
557 hydrophobic surface of cellulose. Instead, it stacks on this surface by its aliphatic C-H groups.
558 The interaction on this particular cellulose surface is dominated by the Van der Waals forces
559 (Martinez-Abad et al, 2017; Mazeau & Charlier, 2012). In contrast, the degree of acetylation
560 affects the adsorption features of xylan on the hydrophilic surfaces of cellulose. The per-
561 acetylated xylan adsorbs flat to maximize the number of Van der Waals contacts. This
562 organization is obviously promoted by the amphiphilicity of the cellulose surfaces (Mazeau,
563 2011). On the contrary, the deacetylated xylan adsorbs in a tilted conformation to establish

hydrogen bonds between its hydroxyl groups and the hydroxyl groups protruding from the cellulose surface. The regularly and alternately substituted xylan considered by Busse-Wicher et al., which forms hydrogen bonds with cellulose, thus follows the adsorption of the non-substituted xylan (Busse-Wicher et al., 2014). This similarity is understandable considering that a xylan polymer in the 2_1 helical conformation in which only one residue out of two is substituted has only one acetylated side. This is the non-substituted side, consisting only of hydroxyl groups, that preferentially interacts with cellulose in the same way as a deacetylated xylan.

DISCUSSION

Xylan presents a broad structural variability and is a major hemicellulose in plant biomass, essentially encountered in secondary cell walls (Brummell & Schröder, 2009; Scheller & Ulvskov, 2010). It is, however, a minor component of primary cell walls and particularly those of fleshy fruit (Brummell & Schröder, 2009) such as apple where it occurs as glucuronoarabinoxylan (GAX) (Ray et al., 2014). The proportions of uronic acids, arabinose and acetyl ester substitutions determined for the apple pomace xylan fractions are similar to those reported for Gala apple (Ray et al., 2014). Considering that apple pomace is not lignified, it is therefore a convenient biological source for native xylan with a preserved substitution pattern that can be used as a model polymer for *in vitro* investigation.

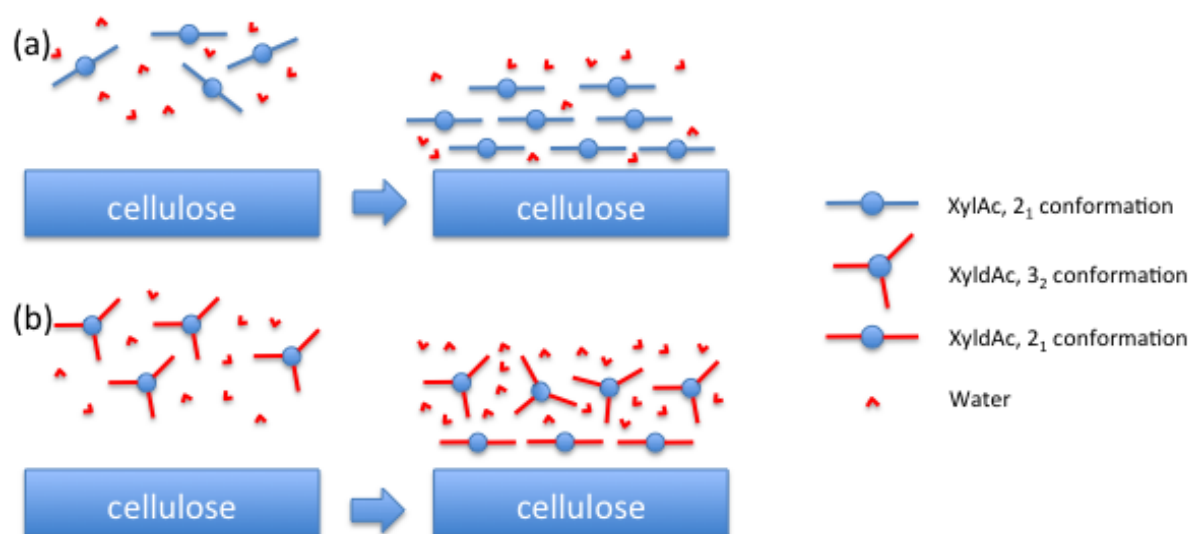
QCM-D experiments complemented by molecular modeling provide an atomistic-scale view of the effect of the acetyl groups attached to xylan on their adsorption on cellulose.

Adsorption of xylan fractions followed by QCM-D demonstrated that the acetylated and deacetylated fractions adsorption processes are both kinetically-driven but display different behaviors. XylAc has an almost homogenous adsorption pattern for all concentrations (k_f

dominated), which suggests limited rearrangement after adsorption, whereas XyldAc displays two clear concentration domains of adsorption (k_1/k_2 increased between 5 and 10 g L⁻¹). These behaviors are also supported by the pattern of the ΔD vs. Δf representation. The ΔD vs. Δf plot for XylAc displays a single slope, whereas that for XyldAc shows two slopes between 2.5 and 10 mg L⁻¹ of xylan injected. The slope change corresponds to the increase of the k_1/k_2 ratio observed in the kinetic modeling analysis. For the higher slope (low concentration), k_1/k_2 is close to one and increase when the slope become lower. The models revealed that adsorption of XylAc is straightforward since the acetylated xylan does not need to modify its solution conformation (two-fold helices). In contrast, adsorption of the very first molecules of XyldAc requires a conformational change to adopt a form complementary to that of cellulose (from three-fold to two-fold), whereas the additional molecules adsorb without conformation change (three-fold).

Additionally, the XylAc ΔD vs. Δf slope displays a very low and homogeneous value (0.03 10⁻⁶ Hz⁻¹) compared to XyldAc fraction values that are higher and variable according to the concentration injected (from 0.5 to 0.1 10⁻⁶ Hz⁻¹). This suggests the formation of a stiff and densely-packed XylAc layer, whereas the XyldAc fraction forms a more viscous layer that can be densified with increasing XyldAc concentration. In agreement with the experimental observations, the models also show that the acetyl groups affect the compactness and hydration of the xylan layer surrounding the cellulose. It is compact and weakly hydrated for acetylated xylan, but loose and highly hydrated for deacetylated xylan. Considering that water plasticizes hemicellulose, complexes made with XylAc would be rigid, as revealed by the low gradient of the ΔD vs. Δf plot, whereas those made with XyldAc would be softer, showing a more viscous character, as suggested by the larger slopes of the ΔD vs. Δf plot. The adsorption process of the acetylated and deacetylated xyans on cellulose can be illustrated with the schematics shown in Fig. 6.

613



614

615 Fig. 6. Schematic representation of the adsorption process of XylAc (a) and XyldAc (b) on
 616 cellulose. Adsorption of XylAc results in the formation of a rigid (almost) anhydrous layer,
 617 whereas XyldAc forms a water-swollen soft layer.

618 Our findings indicated that the presence of acetyl esters influences the supramolecular
 619 organization of the xylan and its interaction with the cellulose surface. Deacetylated and
 620 acetylated xylans display different organization suggesting that the remaining substituent
 621 groups are not enough to keep the xylan interaction with cellulose surface unchanged. Indeed,
 622 in *Arabidopsis* mutants with decreased acetyl ester, compensation is provided by an increase
 623 in glucuronic (Grantham et al., 2017). In our case, the removal of all acetyl groups cannot be
 624 compensated by the other substituents.

625 The functional role of xylan in fleshy fruit remains unknown. In tomato, the trace amount of
 626 xylan was localized in cell walls lining intercellular space (Ordaz-Ortiz, Marcus, & Knox,
 627 2009) and particularly in the skin and inner epidermal cells of the pericarp (Takizawa et al.,
 628 2014). In biomass, it is found in the secondary wall (Carafa, Duckett, Knox, & Ligrone, 2005)
 629 and directly or indirectly contributes to the mechanical properties of the stems (Li et al., 2015;
 630 Dammstrom, Salmen, & Gatenholm, 2009). The biological and technological roles of acetyl

groups are still unclear but it has been hypothesized that they limit the saccharification and fermentation of biomass (Pawar et al., 2017). Besides the steric hindrance induced by acetyl groups to access a xylanase catalytic site, the formation of a poorly hydrated and stiff layer interacting with cellulose, as suggested by our study, can also be an explanation for the limited xylanase accessibility and mechanical properties of plant cell walls.

CONCLUSION

Combined QCM-D and MD approaches were used to illustrate the impact of acetyl decoration on the interaction between xylan and cellulose. Both *in vitro* and *in silico* results showed that acetyl groups attached to the xylan chains significantly influence the conformational behavior of xylan and the supramolecular organization of the cellulose-xylan complex. Acetylated fractions adsorbed on cellulose form a dense and poorly hydrated layer, whereas the deacetylated fraction forms a more hydrated layer. It should be noted that the acetylated fraction has the two residue per turn conformation for all the adsorbed chains, whereas this conformation is limited to the xylan chains in direct contact with the cellulose surface for the deacetylated fraction. Our study therefore supports the assumption that the acetylation pattern promotes different organizations and hydrations of xylan cellulose complexes that can modulate the interaction strength and rheological properties of the cell wall xylan-cellulose supramolecular complexes. Our future studies will focus on the evaluation of the influence of the presence of the acetyl groups on the properties as well as the use of the modulation of acetylation patterns to implement xylan/CNC-based composites.

651 **ASSOCIATED CONTENT**

652 **Supporting Information**

653 DOI:

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