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# The Genetic Legacy of the Indian Ocean Slave Trade: Recent Admixture and Post-admixture Selection in the Makranis of Pakistan

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From the eighth century onward, the Indian Ocean was the scene of extensive trade of sub-Saharan African slaves via sea routes controlled by Muslim Arab and Swahili traders. Several populations in present-day Pakistan and India are thought to be the descendants of such slaves, yet their history of admixture and natural selection remains largely undefined. Here, we studied the genome-wide diversity of the African-descent Makranis, who reside on the Arabian Sea coast of Pakistan, as well that of four neighboring Pakistani populations, to investigate the genetic legacy, population dynamics, and tempo of the Indian Ocean slave trade. We show that the Makranis are the result of an admixture event between local Baluch tribes and Bantu-speaking populations from eastern or southeastern Africa; we dated this event to ~300 years ago during the Omani Empire domination. Levels of parental relatedness, measured through runs of homozygosity, were found to be similar across Pakistani populations, suggesting that the Makranis rapidly adopted the traditional practice of endogamous marriages. Finally, we searched for signatures of post-admixture selection at traits evolving under positive selection, including skin color, lactase persistence, and resistance to malaria. We demonstrate that the African-specific Duffy-null blood group—believed to confer resistance against *Plasmodium vivax* infection—was recently introduced to Pakistan through the slave trade and evolved adaptively in this *P. vivax* malaria-endemic region. Our study reconstructs the genetic and adaptive history of a neglected episode of the African Diaspora and illustrates the impact of recent admixture on the diffusion of adaptive traits across human populations.

The trade of slaves has existed in many cultures since early human history and has left a profound legacy on the social, cultural, and genetic diversity of human populations. Over the last five centuries, more than ten million slaves were transported from Africa to the New World by western European traders.<sup>1</sup> The ancestry of African slaves captured for the transatlantic slave trade has been extensively studied with the use of rich historical records<sup>2,3</sup> and genome-wide surveys of present-day African Americans.<sup>4–6</sup> However, much less is known about the populations who were enslaved for the Indian Ocean slave trade.<sup>7</sup> From the 8<sup>th</sup> to the 19<sup>th</sup> centuries, about four million people were captured from the shores of eastern Africa by Arab Muslim and Swahili traders. It has been suggested that slaves transported before the 16<sup>th</sup> century originated from the Horn of Africa, i.e., Nilotic or Afro-Asiatic speakers from present-day Ethiopia, whereas most Africans enslaved from the 18<sup>th</sup> century onward were Zanj,<sup>8</sup> i.e., Bantu speakers of southeastern Africa. Indeed, the Omani Empire progressively imposed their domination on the Swahili coast and Zanzibar in this time period, leading to an intensified slave trade from these regions. However, direct evidence of the prov-

enance of the African slaves embarked for the Indian Ocean trade remains lacking.

A few present-day populations in South Asia, including the Siddis from western India and the Makranis from Pakistan, are considered to descend from African slaves.<sup>9</sup> Because these populations have not preserved their original African languages and traditions, except perhaps musical culture,<sup>10</sup> studying the genome of South Asian populations of African descent represents a unique opportunity to increase our knowledge about the dynamics and tempo of the Indian Ocean slave trade. Genetic studies of both uniparentally inherited and autosomal markers have estimated that the Siddis have 60%–75% sub-Saharan African genetic ancestry and carry Y chromosome haplogroups characteristic of Bantu-speaking populations.<sup>11,12</sup> By contrast, the African ancestry of the Makranis is limited to 12% ( $\pm 7\%$ ) for the Y chromosome and 40% ( $\pm 9\%$ ) for mtDNA.<sup>13,14</sup> Nevertheless, most of these estimations are based on uniparentally inherited markers, which provide a partial, sex-biased view of past population history, and no studies have appropriately investigated the ancestry sources of African-descent admixed populations in South Asia. Furthermore, their study can inform the

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extent to which traits that were adaptive in parental populations, such as increased resistance to infection, have contributed to the fitness of admixed populations in a different environmental context.<sup>15</sup>

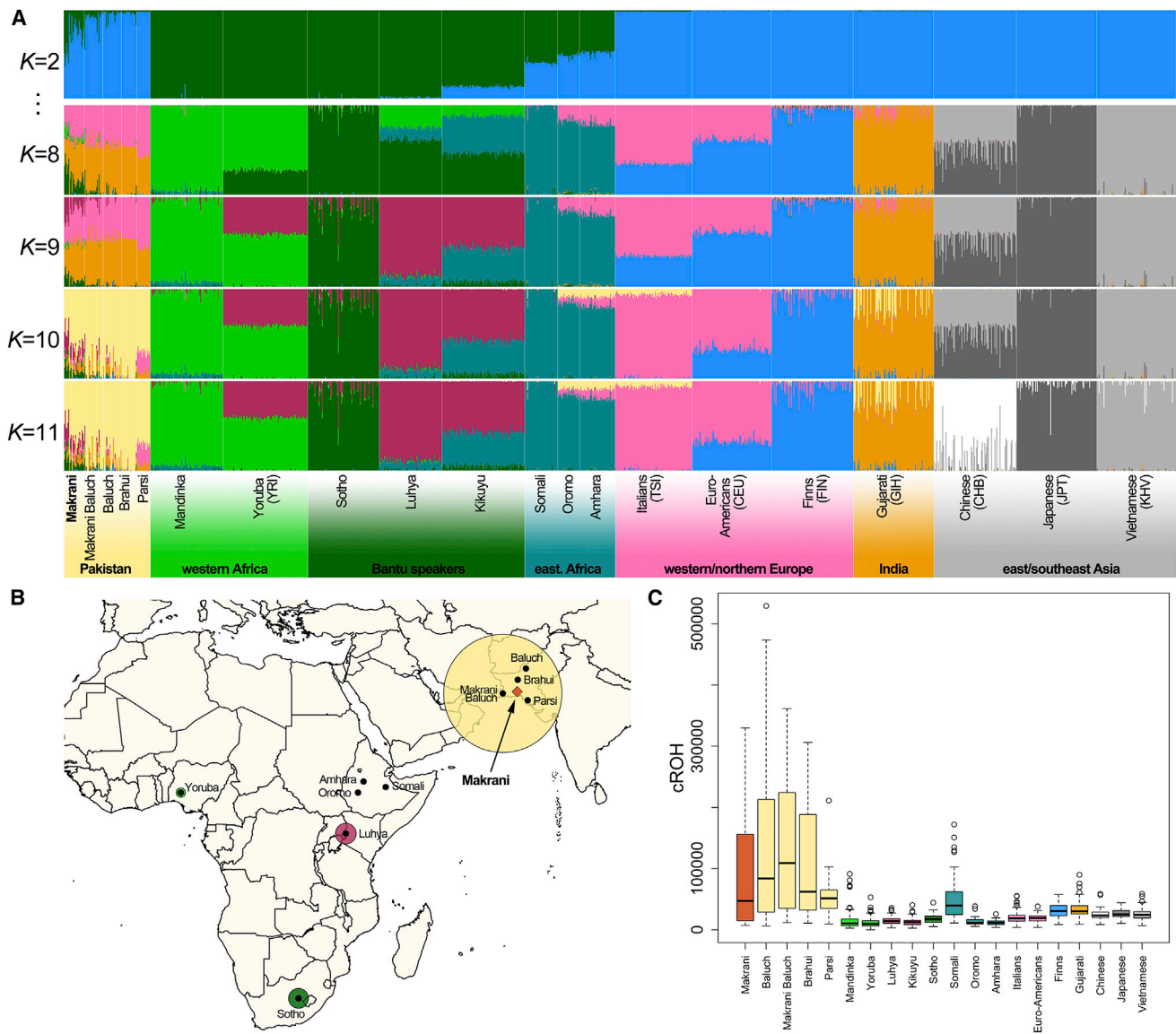
In this study, we aimed to increase our knowledge of the geographic origins, admixture dynamics, and post-admixture selection processes of South Asian African-descent populations that originated from the Indian Ocean slave trade by using a population-genomics approach. To address these questions, we genotyped 118 individuals from five Pakistani populations on the Illumina Omni2.5 array (Figure S1). We excluded 16 individuals who presented with evidence of cryptic relatedness (i.e., kinship coefficient > 0.025 and an identical-by-state statistic < 0.034 estimated from SNP array data with KING<sup>16</sup>), leading to a total of 102 unrelated individuals who were retained for all subsequent analyses. These included 24 African-descent Makranis sampled from different locations near Karachi and on the Makran coast (here referred to as Makranis), 22 Baluch inhabiting the Makran coast (here referred to as Makrani Baluch), 22 Baluch and 18 Brahui from Baluchistan, and 16 Parsi from Karachi, in the Sindh province of Pakistan (Table S1). It is important to note that our sample of African-descent Makranis is distinct from the so-called Makrani of the HGDP-CEPH panel.<sup>17,18</sup> The latter correspond to our Makrani Baluch, who are not considered to descend from African slaves and present with neither anthropological nor cultural features associated with African ancestries.<sup>19</sup> DNA sampling has been described elsewhere.<sup>13,14</sup> Informed consent was obtained from all participants, and the study was approved by the Institut Pasteur (institutional review board no. 2011-54/IRB/6). After standard quality-control filters (Figure S1), a total of 2,263,423 SNPs were retained for subsequent analyses. We merged the newly generated data with relevant, available datasets, including those of the HapMap 3 International Consortium<sup>20</sup> and the African Genome Variation Project,<sup>21</sup> yielding a total of 1,316 individuals genotyped for 1,356,632 SNPs (Table S1).

Unsupervised ADMIXTURE analysis<sup>22</sup> estimated that the Makranis are composed of 25.5% ancestry from sub-Saharan Africa and 74.5% from Pakistan, with a standard deviation (SD) of 16.6% (Figures 1A, S2, and S3; Table S2). The large inter-individual variance in African ancestry among the Makranis is suggestive of recent admixture.<sup>23</sup> We also observed varying levels of African ancestry among the remaining Pakistani populations: 6.7% (SD = 10.7%) in the neighboring Makrani Baluch, followed by a baseline of 1%–2% in the Baluch and Brahui of Baluchistan and the Parsi from Karachi. With increased *K* values in the ADMIXTURE analysis, the African ancestry of the Makranis appeared to be related to that of Bantu-speaking populations from eastern or southeastern Africa, more than that of western or Horn of Africa populations (Figure 1A). We obtained comparable results when merging our newly generated data with two independent datasets that together included 1,111 individuals from 63

worldwide populations but a limited number of SNPs (226,323 SNPs common to all datasets; Figures S4 and S5; Table S1).<sup>6,17</sup>

To formally test whether the Makranis are the result of an admixture event and, if so, to determine when such admixture occurred and which populations were involved, we phased the data with SHAPEIT2<sup>24</sup> and used the haplotype-based GLOBETROTTER approach.<sup>18,25</sup> Admixture linkage-disequilibrium (LD) decay fitted well with expectations under a single admixture event (simulation-based  $p < 0.01$ ; Figure S6) occurring  $300 \pm 25$  years ago ( $12 \pm 1$  generations ago) between a Pakistani population (83% ancestry contribution [bootstrap 95% confidence interval (CI) of the parameter estimate = 82%–84%]) and a sub-Saharan African population (17% ancestry contribution [95% CI = 16%–18%]). The best-matching parental populations were the Baluch (97% relative ancestry contribution [95% CI = 96%–98%] versus 3% [95% CI = 2%–4%] for the Parsi; Figure 1B) and Bantu-speaking populations from eastern or southeastern Africa (42% relative ancestry [95% CI = 10%–42%] for the Luhya of Kenya versus 40% [95% CI = 20%–44%] for the Sotho of South Africa). The contribution from the Horn of Africa, represented here by the Oromo, the Amhara, and the Somali of Ethiopia, was nil across all bootstrap replicates (Table S2). The similar contributions of eastern or southeastern Bantu-speaking groups to present-day Makranis suggest that an intermediate population, such as Mozambicans, could be the most likely unsampled source population. This hypothesis is further supported by haplotype-based principal-component (PC) analysis, which showed that the Makranis are located in an intermediate position between eastern (i.e., the Luhya) and southeastern (i.e., the Sotho) Bantu-speaking populations from PC2 to PC4 (Figure S7). These results indicate that the Makranis probably descend from slaves who were captured on the Swahili coast during the 18<sup>th</sup> century, when the Omani Empire dominated the Indian Ocean slave trade.<sup>8,26</sup>

To learn more about endogamy and marriage practices in the Makranis, we next scanned their genomes for long runs of homozygosity (ROHs), which are indicative of elevated relatedness levels among an individual's ancestors, even when the overall population size is large.<sup>27,28</sup> We detected ROHs with PLINK 1.9<sup>29</sup> by considering 1-Mb homozygous windows that included more than 20 SNPs, allowing for two heterozygous SNPs and five missing genotypes.<sup>30</sup> The mean cumulative ROH in the Makranis was large and not significantly different from that observed in the other Pakistani populations (two-sample *t* test adjusted  $p > 0.05$ ; Figure 1C), who are among the studied populations with the largest levels of parental relatedness worldwide.<sup>28</sup> Notably, we found no significant correlation between cumulative ROHs and Pakistani ancestry in the Makranis (Spearman's  $\rho = 0.25$ ,  $p = 0.24$ ), suggesting that their levels of homozygosity are not simply due to identity by descent in the Pakistani fraction of their genomes. Although future studies with increased sample



**Figure 1. African Origins and Levels of Parental Relatedness of the Admixed Makranis of Pakistan**

(A) Clustering analysis was performed on 1,316 individuals and 819,671 independent SNPs (i.e., 1,356,632 SNPs were LD pruned for SNPs with  $r^2 > 0.5$ ; [Figure S1](#)) with ADMIXTURE.<sup>22</sup> The  $K$  value providing the lowest cross-validation error was  $K = 8$  ([Figure S2](#)). For each  $K$  value, the most supported result across ten iterations is shown. Results for other  $K$  values are shown in [Figure S3](#).

(B) Circle sizes are proportional to the genetic contributions of possible source populations to the Makranis, estimated with GLOBETROTTER<sup>18</sup> on the basis of 16 individuals per source population and 1,356,632 SNPs.

(C) Cumulative runs of homozygosity (cROHs) in the Makranis and other worldwide populations.

sizes are needed to confirm this hypothesis, our results suggest that the descendants of African slaves in Pakistan rapidly adopted the local practices of endogamy.<sup>19</sup>

We next evaluated whether heritable traits that are known to be adaptive in humans and that were brought by African slaves have also conferred a selective advantage in the South Asian environment after the admixture event and led to signatures of post-admixture selection in the Makranis. Specifically, we tested whether variants associated with skin pigmentation (MIM: 227220),<sup>31</sup> lactose tolerance (MIM: 223100),<sup>32</sup> and host resistance to malaria (MIM: 611162)<sup>33</sup> show departures from their expected frequency under an admixture model<sup>15</sup> and/or an excess of

local African ancestry in the Makrani genomes.<sup>6,34,35</sup> We genotyped 18 candidate variants by using TaqMan assays and two variants in the *LCT* (MIM: 603202) regulatory region by using Sanger sequencing ([Figure S1](#)); all were absent from the Omni2.5 SNP array ([Table 1](#)).

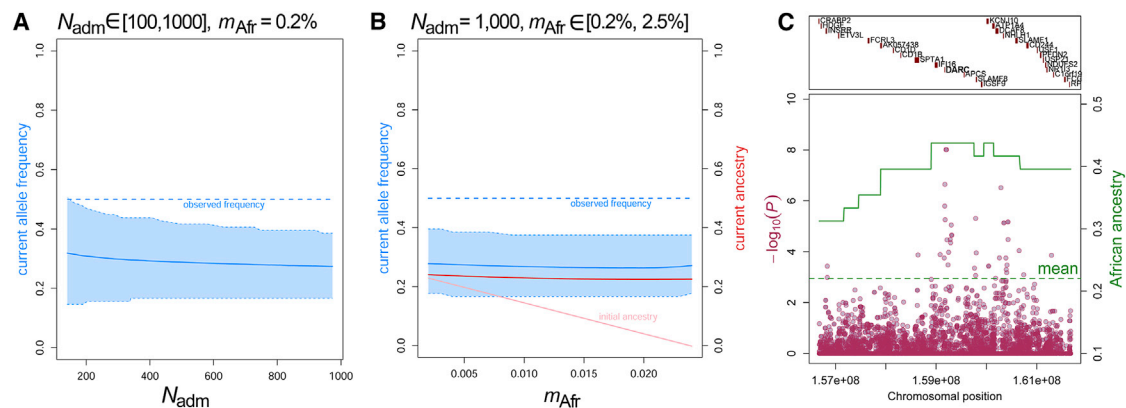
A single variant, rs2814778 in *DARC* (Duffy antigen receptor for chemokines, more recently termed *ACKR1* for atypical chemokine receptor 1 [MIM: 613665]), showed several consistent signals of post-admixture selection ([Figure 2](#)). This variant is responsible for the Duffy-null blood group, which is thought to confer complete resistance to *Plasmodium vivax* malaria.<sup>33</sup> The derived C allele, which reaches near fixation in sub-Saharan Africa and is

**Table 1. Tests for Post-admixture Selection for 20 Candidate Variants in the Makranis**

Variant	Position (GRCh37 <sup>a</sup> )	HUGO Gene Name (MIM No.)	Candidate Allele	Alternative Allele	Luhya	Brahui	Parsi	Baluch	Makrani Baluch	Makrani (Obs.)	Makrani (Exp.)	p Value (Binomial Test)	Local Ancestry	SD from Average	p Value (PCAdapt)
<b>Dark Skin Pigmentation</b>															
rs1426654	chr15: 48,426,484	<i>SLC24A5</i> (609802)	G	A*	0.92	0.09	0.04	0.03	0.14	0.26	0.26	1.0	0.17	-0.71	$3.0 \times 10^{-3}$
rs6058017	chr20: 32,856,998	<i>ASIP</i> (600201)	G	A*	0.76	0.16	0.27	0.24	0.27	0.26	0.37	0.13	0.19	-0.43	0.92
rs2228478	chr16: 89,986,608	<i>MC1R</i> (155555)	G	A*	0.57	0.16	0.12	0.12	0.27	0.30	0.23	0.30	0.20	-0.28	0.12
rs2287499	chr17: 7,592,168	<i>WRAP53</i> (612661)	G	C*	0.86	0.41	0.19	0.41	0.41	0.50	0.52	0.77	0.27	0.71	0.04
rs2467737	chr15: 43,737,188	<i>TP53BP1</i> (605230)	A	G*	0.95	0.59	0.46	0.32	0.30	0.59	0.48	0.18	0.27	0.71	$2.5 \times 10^{-4}$
rs642742	chr12: 89,299,746	<i>KITLG</i> (184745)	T	C*	0.91	0.34	0.35	0.12	0.20	0.33	0.32	1.0	0.23	0.14	$2.6 \times 10^{-3}$
rs16891982	chr5: 33,951,693	<i>SLC45A2</i> (606202)	C	G*	0.99	0.78	0.58	0.91	0.91	0.85	0.93	0.05	0.10	-1.71	0.07
rs26722	chr5: 33,963,870	<i>SLC45A2</i> (606202)	C	T*	0.95	0.81	0.78	0.69	0.75	0.81	0.76	0.50	0.10	-1.71	0.53
rs1667394	chr15: 28,530,182	<i>OCA2</i> (611409), <i>HERC2</i> (605837)	C	T*	0.97	0.61	0.69	0.67	0.70	0.79	0.75	0.62	0.22	0.00	0.04
rs1800404	chr15: 28,235,773	<i>OCA2</i> (611409), <i>HERC2</i> (605837)	C	T*	0.91	0.47	0.44	0.47	0.66	0.56	0.58	0.77	0.27	0.71	0.58
rs1042602	chr11: 88,911,696	<i>TYR</i> (606933)	C	A*	1.00	0.67	0.69	0.83	0.86	0.85	0.87	0.66	0.19	-0.43	0.14
rs9782955	chr1: 236,039,877	<i>LYST</i> (606897)	C	T*	0.95	0.75	0.72	0.75	0.82	0.75	0.80	0.37	0.15	-1.00	0.72
rs3829241	chr11: 68,855,363	<i>TPCN2</i> (612163)	G	A*	0.97	0.85	0.75	0.78	0.89	0.88	0.83	0.56	0.23	0.14	0.51
rs12896399	chr14: 92,773,663	<i>SLC24A4</i> (609840)	G	T*	0.98	0.69	0.66	0.81	0.66	0.81	0.85	0.41	0.23	0.14	0.34
<b>Lactose Tolerance</b>															
rs145946881	chr2: 136,608,746	<i>LCT</i> (603202)	G*	C	0.09	0.00	0.00	0.00	0.00	0.00	0.02	0.629	0.21	-0.14	NA
rs4988235	chr2: 136,608,646	<i>LCT</i> (603202)	A*	G	0.00	0.50	0.00	0.38	0.34	0.24	0.28	0.624	0.21	-0.14	$8.5 \times 10^{-3}$
<b>Malaria Resistance</b>															
rs2814778	chr1: 159,174,683	<i>ACKR1</i> , also known as <i>DARC</i> (613665)	C*	T	1.00	0.03	0.00	0.06	0.25	0.50	0.30	0.0055	0.44	3.13	$2.3 \times 10^{-7}$
rs17047661	chr1: 207,782,889	<i>CRI</i> (120620)	G*	A	0.70	0.03	0.00	0.00	0.07	0.25	0.18	0.190	0.25	0.43	$1.1 \times 10^{-3}$
rs334	chr11: 5,248,232	<i>HBB</i> (141900)	A*	T	0.10	0.00	0.00	0.00	0.00	0.00	0.03	0.633	0.23	0.14	NA
rs2213169	chr11: 5,303,063	<i>HBE1</i> (142100)	A*	G	0.17	0.00	0.00	0.00	0.02	0.02	0.04	0.724	0.23	0.14	NA

Derived alleles are indicated by an asterisk. Observed (obs.) frequencies of candidate alleles are shown together with those expected (exp.) in the Makranis according to their inferred admixture model. Local ancestry at candidate variants, as well as the corresponding number of standard deviations (SDs) from the genome-wide average, was obtained with RFMix 1.5.4.<sup>36</sup> NA stands for not applicable.

<sup>a</sup>UCSC Genome Browser hg19.



**Figure 2. Evidence of Post-admixture Selection of the Duffy-Null Blood Group in Pakistan**

(A and B) Simulation-based evidence for post-admixture selection of the Duffy-null blood group in the Makranis. We performed 100,000 Wright-Fisher simulations of an admixed population resulting from admixture occurring 12 generations ago (A) by using an effective population size ( $N_{adm}$ ) ranging from 100 to 1,000<sup>15</sup> and assuming a fixed, low migration rate ( $m_{Afr}$ ) of 0.2%, corresponding to the proportion of the admixed population that is replaced by individuals from the African parental population at each generation; or (B) by using a fixed  $N_{adm}$  of 1,000 and a  $m_{Afr}$  ranging from 0.2% to 2.5%. The dashed blue line indicates the observed allele frequencies in a sample of 24 individuals. The light-pink line shows the initial, simulated African ancestry in the admixed population, and the red line shows the final African ancestry (in the last generation). We tuned the initial African ancestry so that the final ancestry fit that estimated by ADMIXTURE.

(C) Local signatures of post-admixture selection in the Makranis. The green line indicates the average African local ancestry inferred by RFMix v.1.5.4<sup>36</sup> with two expectation-maximization steps. Maroon points indicate  $-\log_{10}(p)$  values of PCAdapt,<sup>37</sup> which tested for strong genetic differentiation between the Makranis and non-admixed Baluch.

largely absent from Eurasia,<sup>38</sup> was detected at a significantly higher frequency in the Makranis (50%) than expected (30%; adjusted binomial test  $p = 5.5 \times 10^{-3}$ ) given its frequency in the parental populations and admixture proportions estimated by ADMIXTURE (Table S2). This signal was confirmed by 100,000 simulations under the Wright-Fisher neutral model and the assumption of realistic effective population sizes of the Makranis and migration rates from parental populations (simulation-based  $p < 0.05$ ; Figures 2A and 2B). The rs2814778 variant was strongly differentiated between the Makranis and non-admixed Baluch ( $p = 2.3 \times 10^{-7}$ ; Figure 2C).<sup>37</sup> Furthermore, African local ancestry along the genome of the Makranis, inferred by RFMix v.1.5.4,<sup>36</sup> reached 44% in the *DARC* genomic region, 3.13 SDs above the genome-wide average of 22% (Figure 2C; Table S2), indicating that the signal of selection extends to surrounding linked markers. Finally, the C allele was also observed at 25% in the Makrani Baluch (Table 1), whereas their African ancestry was only ~7% (Table S2), suggesting that the variant was also positively selected in Pakistani populations with more limited African ancestry. Together, these results suggest that the Duffy-negative blood group has been under post-admixture selection in South Asia, where 80% of individuals with severe malaria are infected by *P. vivax*.<sup>39</sup>

This study reconstructs the recent genetic and adaptive history of the descendants of African slaves carried by the Indian Ocean trade, a major yet neglected episode of the African Diaspora. Oral traditions suggest that the Makranis descend from Abyssinian (i.e., present-day Ethiopian)

slaves who were transported to Pakistan in the 8<sup>th</sup> century.<sup>40</sup> Our genomic survey indicates instead that most of their African ancestry can be traced back to the so-called Zanj, i.e., Bantu-speaking populations from the Swahili coast. Exhaustive sampling of southeastern Bantu-speaking populations will now be needed to further narrow the geographic origins of the African ancestors of Makrani populations. Despite this potential limitation, the Swahili coastal origin of African slaves is further supported by the estimated date of admixture between the African and Pakistani ancestors of the Makranis in the beginning of the 18<sup>th</sup> century, a period when the majority of slaves were captured in southeast Africa under the rule of the Omani Empire.<sup>8,26</sup> Furthermore, we identified the Baluch as their best-matching source of Pakistani ancestry; Baluch men were historically recruited into the slave trade as soldiers, body guards, and sailors for Omani Muslims mainly during the 18<sup>th</sup> and 19<sup>th</sup> centuries.<sup>19</sup> Our results do not necessarily imply that the Indian Ocean slave trade to Pakistan developed only in this time period, given that the arrival of slaves might not have been accompanied by immediate admixture with local populations. Furthermore, estimating the admixture time on the basis of admixture LD decay informs the period during which the bulk of admixture occurred and can be biased toward more recent times when there is continuous admixture.<sup>41</sup> Nevertheless, our analyses do not support a scenario of admixture mostly occurring in the 8<sup>th</sup> century, as suggested by oral traditions, and suggest instead that the genetic legacy of African slaves in present-day Pakistanis is of more recent origin.

It is also important to highlight that we detected unexpectedly low African ancestry in the Makranis (17%) in relation to that estimated in the African-descent Siddis from India (>60%).<sup>12</sup> Two hypotheses can be put forward to explain this finding: the number of African slaves transported to present-day Pakistan might have been lower than that transported to India, or genetic isolation of African slaves (i.e., segregation) might have been stronger in India than in Pakistan, where co-habitation and intermarriages with slaves were commonly practiced. Consistently with the latter scenario, historical records suggest that female slaves had more chances to intermix with local South Asian populations than male slaves,<sup>26</sup> and genetic analyses suggest that the majority of African slaves who contributed to the gene pool of present-day Pakistanis through admixture were females, whereas those admixing in India were primarily males.<sup>12,14</sup> Indeed, the estimated African ancestry of the Makranis from their mitochondrial genome (40%) is three times that obtained from the Y chromosome (12%), whereas it is only one-third in the Siddis (24% from mtDNA versus 70% from Y chromosome).<sup>12,14</sup>

A seminal study has suggested that natural selection acting in the last few generations can be detected in admixed populations on the basis of signatures of post-admixture selection,<sup>15</sup> but it has been unknown whether adaptive traits such as malaria resistance, skin pigmentation, or lactose tolerance have been selected in admixed populations of South Asia. Our data and analyses indicate that the African-specific Duffy-null blood group has evolved under post-admixture selection since it was introduced by slaves into Pakistan. The Duffy-null phenotype is caused by a variant in the *DARC* promoter region, which abolishes Duffy antigen protein expression on red blood cells<sup>42</sup> and the ability of the malaria parasite *P. vivax* to invade erythrocytes.<sup>43</sup> The extreme frequency differences of the Duffy-null allele between African and non-African populations were interpreted as the result of a strong selective sweep starting ~42 kya in Africans<sup>38,44</sup> as a result of increased resistance to *P. vivax* malaria. However, *P. vivax* infection has been recently documented in African Duffy-null individuals,<sup>45,46</sup> raising the possibility that the *DARC* selection signature results from host resistance to another infectious agent. Given that *P. vivax* malaria is endemic in Pakistan,<sup>39</sup> our results are consistent with the hypothesis that resistance to *P. vivax* is the main evolutionary force driving the frequency of the Duffy-null allele in admixed Pakistanis, in agreement with a genetic study of admixed Malagasy.<sup>15</sup> Admixture mapping of malaria resistance in African-descent admixed populations living in endemic regions, such as Pakistan, could substantiate the major impact of *P. vivax* on the history of human adaptation.

Recent studies have provided empirical evidence that the acquisition by human populations of adaptive traits, including the response to altitude-induced hypoxia, lactase persistence, and HLA-mediated responses, has been facilitated by admixture.<sup>6,34</sup> Our finding of post-admixture

selection targeting the Duffy-null blood group outside Africa provides evidence of adaptive admixture within our species—as opposed to interspecies adaptive introgression. Future theoretical and empirical studies of admixed populations, particularly those resulting from the African Diaspora worldwide, will provide new insights into the recent impact of natural selection on population variation in complex traits and disease susceptibility across a wide range of environmental contexts.

### Accession Numbers

The newly generated SNP genotype data have been deposited in the European Genome-phenome Archive under accession number EGAS00001002558.

### Supplemental Data

Supplemental Data include seven figures and two tables and can be found with this article online at <https://doi.org/10.1016/j.ajhg.2017.09.025>.

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### Web Resources

European Genome-phenome Archive, <https://www.ebi.ac.uk/ega/home>

OMIM, <http://omim.org/>

UCSC Genome Browser, <https://genome.ucsc.edu/>

### References

1. Lovejoy, P.E. (2000). *Transformations in Slavery* (Cambridge University Press).
2. Eltis, D., and Richardson, D. (2010). *Atlas of the Transatlantic Slave Trade* (Yale University Press).
3. Midlo Hall, G. (2007). *Slavery and African Ethnicities in the Americas* (University of North Carolina Press).
4. Bryc, K., Durand, E.Y., Macpherson, J.M., Reich, D., and Mountain, J.L. (2015). The genetic ancestry of African Americans, Latinos, and European Americans across the United States. *Am. J. Hum. Genet.* 96, 37–53.
5. Montinaro, F., Busby, G.B., Pascali, V.L., Myers, S., Hellenthal, G., and Capelli, C. (2015). Unravelling the hidden ancestry of American admixed populations. *Nat. Commun.* 6, 6596.

6. Patin, E., Lopez, M., Grollemund, R., Verdu, P., Harmant, C., Quach, H., Laval, G., Perry, G.H., Barreiro, L.B., Froment, A., et al. (2017). Dispersals and genetic adaptation of Bantu-speaking populations in Africa and North America. *Science* *356*, 543–546.
7. Mirzai, B.A., Montana, I.M., and Lovejoy, P.E. (2009). *Slavery, Islam and Diaspora* (Africa World Press).
8. Vernet, T. (2009). Slave trade and slavery on the Swahili coast (1500-1750). In *Slavery, Islam and Diaspora*, B.A. Mirzai, I.M. Montana, and P.E. Lovejoy, eds. (Africa World Press), pp. 37–76.
9. Shihan de Silva, J., and Pankhurst, R. (2003). *The African Diaspora in the Indian Ocean* (African World Press).
10. Badalkhan, S. (2006). On the Presence of African Musical Culture in Coastal Balochistan. In *Journeys and Dwellings: Indian Ocean Themes in South Asia*, H. Basu, ed. (Orient Longman).
11. Narang, A., Jha, P., Rawat, V., Mukhopadhyay, A., Dash, D., Basu, A., Mukerji, M.; and Indian Genome Variation Consortium (2011). Recent admixture in an Indian population of African ancestry. *Am. J. Hum. Genet.* *89*, 111–120.
12. Shah, A.M., Tamang, R., Moorjani, P., Rani, D.S., Govindaraj, P., Kulkarni, G., Bhattacharya, T., Mustak, M.S., Bhaskar, L.V., Reddy, A.G., et al. (2011). Indian Siddis: African descendants with Indian admixture. *Am. J. Hum. Genet.* *89*, 154–161.
13. Qamar, R., Ayub, Q., Mohyuddin, A., Helgason, A., Mazhar, K., Mansoor, A., Zerjal, T., Tyler-Smith, C., and Mehdi, S.Q. (2002). Y-chromosomal DNA variation in Pakistan. *Am. J. Hum. Genet.* *70*, 1107–1124.
14. Quintana-Murci, L., Chaix, R., Wells, R.S., Behar, D.M., Sayar, H., Scozzari, R., Rengo, C., Al-Zahery, N., Semino, O., Santachiara-Benerecetti, A.S., et al. (2004). Where west meets east: the complex mtDNA landscape of the southwest and Central Asian corridor. *Am. J. Hum. Genet.* *74*, 827–845.
15. Hodgson, J.A., Pickrell, J.K., Pearson, L.N., Quillen, E.E., Prista, A., Rocha, J., Soodyall, H., Shriver, M.D., and Perry, G.H. (2014). Natural selection for the Duffy-null allele in the recently admixed people of Madagascar. *Proc. Biol. Sci.* *281*, 20140930.
16. Manichaikul, A., Mychaleckyj, J.C., Rich, S.S., Daly, K., Sale, M., and Chen, W.M. (2010). Robust relationship inference in genome-wide association studies. *Bioinformatics* *26*, 2867–2873.
17. Li, J.Z., Absher, D.M., Tang, H., Southwick, A.M., Casto, A.M., Ramachandran, S., Cann, H.M., Barsh, G.S., Feldman, M., Cavalli-Sforza, L.L., and Myers, R.M. (2008). Worldwide human relationships inferred from genome-wide patterns of variation. *Science* *319*, 1100–1104.
18. Hellenthal, G., Busby, G.B.J., Band, G., Wilson, J.F., Capelli, C., Falush, D., and Myers, S. (2014). A genetic atlas of human admixture history. *Science* *343*, 747–751.
19. Nicolini, B. (2007). The Baluch role in the Persian gulf during the nineteenth and twentieth centuries. *Comp. Stud. South Asia Afr. Middle East* *27*, 384–396.
20. Altshuler, D.M., Gibbs, R.A., Peltonen, L., Altshuler, D.M., Gibbs, R.A., Peltonen, L., Dermitzakis, E., Schaffner, S.F., Yu, F., Peltonen, L., et al.; International HapMap 3 Consortium (2010). Integrating common and rare genetic variation in diverse human populations. *Nature* *467*, 52–58.
21. Gurdasani, D., Carstensen, T., Tekola-Ayele, F., Pagani, L., Tachmazidou, I., Hatzikotoulas, K., Karthikeyan, S., Iles, L., Pollard, M.O., Choudhury, A., et al. (2015). The African Genome Variation Project shapes medical genetics in Africa. *Nature* *517*, 327–332.
22. Alexander, D.H., Novembre, J., and Lange, K. (2009). Fast model-based estimation of ancestry in unrelated individuals. *Genome Res.* *19*, 1655–1664.
23. Verdu, P., and Rosenberg, N.A. (2011). A general mechanistic model for admixture histories of hybrid populations. *Genetics* *189*, 1413–1426.
24. Delaneau, O., Zagury, J.F., and Marchini, J. (2013). Improved whole-chromosome phasing for disease and population genetic studies. *Nat. Methods* *10*, 5–6.
25. Leslie, S., Winney, B., Hellenthal, G., Davison, D., Boumertit, A., Day, T., Hutnik, K., Royrvik, E.C., Cunliffe, B., Lawson, D.J., et al.; Wellcome Trust Case Control Consortium 2; and International Multiple Sclerosis Genetics Consortium (2015). The fine-scale genetic structure of the British population. *Nature* *519*, 309–314.
26. Nicolini, B. (2009). The Makran-Baluch-African Network in Zanzibar and East Africa during the XIXth Century. In *Uncovering the History of Africans in Asia*, J. Shihan de Silva and J.-P. Angenot, eds. (Brill).
27. McQuillan, R., Leutenegger, A.L., Abdel-Rahman, R., Franklin, C.S., Pericic, M., Barac-Lauc, L., Smolej-Narancic, N., Janicijevic, B., Polasek, O., Tenesa, A., et al. (2008). Runs of homozygosity in European populations. *Am. J. Hum. Genet.* *83*, 359–372.
28. Pemberton, T.J., Absher, D., Feldman, M.W., Myers, R.M., Rosenberg, N.A., and Li, J.Z. (2012). Genomic patterns of homozygosity in worldwide human populations. *Am. J. Hum. Genet.* *91*, 275–292.
29. Chang, C.C., Chow, C.C., Tellier, L.C., Vattikuti, S., Purcell, S.M., and Lee, J.J. (2015). Second-generation PLINK: rising to the challenge of larger and richer datasets. *Giga-science* *4*, 7.
30. Henn, B.M., Gignoux, C.R., Jobin, M., Granka, J.M., Macpherson, J.M., Kidd, J.M., Rodríguez-Botigué, L., Ramachandran, S., Hon, L., Brisbin, A., et al. (2011). Hunter-gatherer genomic diversity suggests a southern African origin for modern humans. *Proc. Natl. Acad. Sci. USA* *108*, 5154–5162.
31. McEvoy, B., Beleza, S., and Shriver, M.D. (2006). The genetic architecture of normal variation in human pigmentation: an evolutionary perspective and model. *Hum. Mol. Genet.* *15*, R176–R181.
32. Tishkoff, S.A., Reed, F.A., Ranciaro, A., Voight, B.F., Babbitt, C.C., Silverman, J.S., Powell, K., Mortensen, H.M., Hirbo, J.B., Osman, M., et al. (2007). Convergent adaptation of human lactase persistence in Africa and Europe. *Nat. Genet.* *39*, 31–40.
33. Kwiatkowski, D.P. (2005). How malaria has affected the human genome and what human genetics can teach us about malaria. *Am. J. Hum. Genet.* *77*, 171–192.
34. Jeong, C., Alkorta-Aranburu, G., Basnyat, B., Neupane, M., Wintonsky, D.B., Pritchard, J.K., Beall, C.M., and Di Rienzo, A. (2014). Admixture facilitates genetic adaptations to high altitude in Tibet. *Nat. Commun.* *5*, 3281.
35. Schlebusch, C.M., Skoglund, P., Sjödin, P., Gattepaille, L.M., Hernandez, D., Jay, F., Li, S., De Jongh, M., Singleton, A., Blum, M.G., et al. (2012). Genomic variation in seven Khoe-San groups reveals adaptation and complex African history. *Science* *338*, 374–379.
36. Maples, B.K., Gravel, S., Kenny, E.E., and Bustamante, C.D. (2013). RFMix: a discriminative modeling approach for rapid



- and robust local-ancestry inference. *Am. J. Hum. Genet.* *93*, 278–288.
37. Luu, K., Bazin, E., and Blum, M.G. (2017). *pcadapt*: an R package to perform genome scans for selection based on principal component analysis. *Mol. Ecol. Resour.* *17*, 67–77.
38. Hamblin, M.T., Thompson, E.E., and Di Rienzo, A. (2002). Complex signatures of natural selection at the Duffy blood group locus. *Am. J. Hum. Genet.* *70*, 369–383.
39. Zubairi, A.B., Nizami, S., Raza, A., Mehraj, V., Rasheed, A.F., Ghanchi, N.K., Khaled, Z.N., and Beg, M.A. (2013). Severe *Plasmodium vivax* malaria in Pakistan. *Emerg. Infect. Dis.* *19*, 1851–1854.
40. Edlefsen, J.B., Shah, K., and Farooq, M. (1960). Makranis, the Negroes of West Pakistan. *Phylon* *21*, 124–130.
41. Loh, P.R., Lipson, M., Patterson, N., Moorjani, P., Pickrell, J.K., Reich, D., and Berger, B. (2013). Inferring admixture histories of human populations using linkage disequilibrium. *Genetics* *193*, 1233–1254.
42. Tournamille, C., Colin, Y., Cartron, J.P., and Le Van Kim, C. (1995). Disruption of a GATA motif in the Duffy gene promoter abolishes erythroid gene expression in Duffy-negative individuals. *Nat. Genet.* *10*, 224–228.
43. Miller, L.H., Mason, S.J., Clyde, D.F., and McGinniss, M.H. (1976). The resistance factor to *Plasmodium vivax* in blacks. The Duffy-blood-group genotype, FyFy. *N. Engl. J. Med.* *295*, 302–304.
44. McManus, K.F., Taravella, A.M., Henn, B.M., Bustamante, C.D., Sikora, M., and Cornejo, O.E. (2017). Population genetic analysis of the DARC locus (Duffy) reveals adaptation from standing variation associated with malaria resistance in humans. *PLoS Genet.* *13*, e1006560.
45. Ménard, D., Barnadas, C., Bouchier, C., Henry-Halldin, C., Gray, L.R., Ratsimbaoa, A., Thonier, V., Carod, J.F., Domarle, O., Colin, Y., et al. (2010). *Plasmodium vivax* clinical malaria is commonly observed in Duffy-negative Malagasy people. *Proc. Natl. Acad. Sci. USA* *107*, 5967–5971.
46. Ryan, J.R., Stoute, J.A., Amon, J., Dunton, R.F., Mtalib, R., Koros, J., Owour, B., Luckhart, S., Wirtz, R.A., Barnwell, J.W., and Rosenberg, R. (2006). Evidence for transmission of *Plasmodium vivax* among a duffy antigen negative population in Western Kenya. *Am. J. Trop. Med. Hyg.* *75*, 575–581.