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Marina Volostnykh, Sergey Borisov, Mikhail Konovalov, Anna Sinelshchikova, Yulia Gorbunova, et al.. Platinum(II) and palladium(II) complexes with electron-deficient meso -diethoxyphosphorylporphyrins: synthesis, structure and tuning of photophysical properties by varying peripheral substituents. Dalton Transactions, 2019, 48 (24), pp.8882-8898. 10.1039/c9dt01577a . hal-02351865

HAL Id: hal-02351865

<https://hal.science/hal-02351865>

Submitted on 6 Dec 2019

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Platinum(II) and palladium(II) complexes with electron-deficient *meso*-diethoxyphosphorylporphyrins: synthesis, structure and tuning of photophysical properties by varying peripheral substituents

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^e *Electronic Supplementary Information (ESI) available: 1H, 13C, and 31P NMR spectra, MALDI-TOF and ESI-HR mass spectra of all prepared complexes, X-ray crystal data, emission spectra and photophysical properties of the Pt and Pd porphyrinates, photodegradation kinetic data. See DOI: 10.1039/x0xx00000x*

A series of electron-deficient platinum(II) and palladium(II) *meso*-(diethoxyphosphoryl)porphyrins, namely [10-(diethoxyphosphoryl)-5,15-bis(*p*-tolyl)porphyrinato]palladium(II) (**PdDTolPP**), {10-(diethoxyphosphoryl)-5,15-bis[*p*-(methoxycarbonyl)phenyl]porphyrinato}palladium(II) [**PdD(CMP)PP**], [10-(diethoxyphosphoryl)-5,15-dimesitylporphyrinato]palladium(II) (**PdDMesPP**), [5-(diethoxyphosphoryl)-10,15,20-trimesitylporphyrinato]palladium(II) (**PdTMesPP**) and the corresponding platinum(II) compounds, were synthesized and structurally characterized in solution by means of ¹H, ¹³C, ³¹P NMR spectroscopies and in the solid state by single crystal X-ray diffraction [**PdDTolPP**, **PdD(CMP)PP** and **PtD(CMP)PP**]. Their optical and photophysical properties (UV–vis absorption, luminescence and excitation spectra, phosphorescence quantum yields and lifetimes) were also determined. The complexes under investigation emit at room temperature in the near-infrared region (670–770 nm). Phosphorescence quantum yields of the palladium(II) *meso*-phosphorylated porphyrins lie in the range of 3.4 to 5.8%, with lifetimes of 633 to 858 μs in deoxygenated toluene solutions at room temperature. The corresponding platinum(II) complexes exhibit phosphorescence quantum yields in the range of 9.2 to 11%, with luminescence decay times of 56 to 69 μs. Moreover, effective homogeneous oxygen quenching and the good sensitivity in toluene (~155 Pa⁻¹ s⁻¹) were observed for the platinum(II) complexes with phosphorylporphyrins in solution. Investigations of the photostability of porphyrinylphosphonates and related complexes lacking a phosphoryl group in DMF under irradiation in air using a 400 W vis–NIR lamp demonstrated

that photobleaching is strongly dependent on the substituents at the periphery of the macrocycle. Platinum and palladium trimesitylphosphorylporphyrins **PdTMesPP** and **PtTMesPP** exhibit a high photostability in DMF solution and seem to be the most potentially interesting derivatives of the series for oxygen sensing in biological samples and the covalent immobilization on solid supports to prepare sensing devices including optic fibers.

Platinum(II) and palladium(II) porphyrins are very attractive luminophores due to their strong room temperature phosphorescence and large Stokes shifts.¹ They have found useful applications in OLEDs,² photovoltaics,³ as singlet oxygen sensitizers for photodynamic therapy⁴ and photocatalysis,⁵ as well as labels in living cells.⁶⁻⁸ Efficient quenching of their phosphorescence by molecular oxygen enables the optical oxygen sensing.⁸⁻¹⁰ The quantitative measurement of dioxygen levels by these compounds relies on monitoring the changes of either the emission intensity or the lifetime of the long-lived triplet excited state induced by the interactions of the excited chromophore with O₂ molecules.^{11,12} When phosphorescent compounds are used for oxygen sensing, their photostability is of particular importance because the mechanisms of oxygen signalling and chromophore photobleaching involve several common processes. Reactive oxygen species (ROS: superoxide anion radical and singlet oxygen) generated during the detection process can react with the porphyrin molecules leading to their degradation. Alternatively, the triplet state itself can be involved in various photo-induced reactions.¹³ These chemical processes could impair the accurate determination of the dioxygen concentration in the studied sample.

Pt(II) and Pd(II) complexes with octaethylporphyrin (**OEP**) are commonly used phosphorescent indicators among the tetrapyrrolic compounds, since they possess high brightness.¹⁴⁻¹⁶ However the stability of OEP complexes toward oxidative degradation is quite low. It was shown that the presence of electron-withdrawing groups (Br, CN, and NO₂) has a pronounced effect on the electro-chemical properties of porphyrin rings,¹⁷⁻¹⁹ in particular, it makes them easily reducible and difficult to oxidize relative to the unsubstituted porphyrin. Therefore, for most applications Pt(II) and Pd(II) complexes with halogen-substituted TPP ligands such as, **H₂F₂₀TPP** [5,10,15,20-tetra(pentafluorophenyl)porphyrin] or **H₂F₂₈TPP** [3,4,7,8,12,13,17,18-octafluoro-5,10,15,20-tetra(pentafluorophenyl)porphyrin], which are robust towards oxidation, are successfully used.^{11,20-22} π -Extended porphyrins [*e.g.* tetrabenzoporphyrins (**TBP**)²³ or tetranaphthoporphyrins (**TNP**)²⁴] have also been intensively investigated in the field of optical sensing materials. These compounds are excitable in the red and NIR part of the

electromagnetic spectrum and possess a moderate to high brightness. For increasing the photostability of TBP and TNP to oxidation by $^1\text{O}_2$ and other ROS, the general strategy of incorporating electron-withdrawing groups into the luminophore structure can also be applied.^{25,26}

To the best of our knowledge, no photophysical data for electron-deficient luminescent porphyrins substituted with phosphoryl groups have been reported yet, because convenient synthetic methods for their preparation were developed only recently.^{27–30} Nevertheless, in a few studies it was demonstrated that the introduction of one or more electron-withdrawing $\text{P}(\text{O})(\text{OR})_2$ (where R is an alkyl group) groups directly attached to the porphyrin core at either β -pyrrolic or *meso*- positions leads to positive shifts of the redox potentials.^{31–34} It makes platinum(II) and palladium(II) complexes with phosphorylporphyrins promising candidates for the development of photostable oxygen sensing materials. Moreover, *meso*-(diethoxyphosphoryl)porphyrinates are of particular interest for the preparation of functional materials, because their chemical modifications allow to obtain water-soluble derivatives²⁹ and compounds with anchoring groups for subsequent grafting onto SiO_2 surfaces and metal oxide supports.^{35–40} In this work, we describe the synthesis, structural characterization, and photophysical properties of the novel phosphorescent *meso*-substituted Pt(II) and Pd(II) phosphorylporphyrins. We examine the impact of introducing an electron-withdrawing $\text{P}(\text{O})(\text{OEt})_2$ group and aryl substituents at *meso*-positions on the photophysical characteristics of the platino- and palladoporphyrins, including their photostability, as well as their sensitivity of molecular oxygen detection in solution.

Results and discussion

Synthesis and characterization of the Pt(II) and Pd(II) complexes

The phosphorescent Pt(II) and Pd(II) complexes with *meso*-phosphorylated porphyrins [$\text{MR}^1\text{PP}-\text{MR}^4\text{PP}$, with $\text{R}^1 = \text{DTol}$, $\text{R}^2 = \text{D}(\text{CMP})$, $\text{R}^3 = \text{DMes}$, and $\text{R}^4 = \text{TMes}$; M = Pt(II) and Pd(II)] (Chart 1) were prepared in three steps: formation of the zinc(II) phosphorylated complexes, removal of Zn^{2+} in acidic media, and subsequent complexation with the appropriate metal cations. The zinc(II) phosphorylporphyrins ($\text{ZnR}^1\text{PP}-\text{ZnR}^4\text{PP}$) were obtained using a described procedure,²⁹ with subsequent treatment with HCl for metal removing. For

palladium(II) insertion, the synthetic procedure was slightly modified^{41–44} by refluxing the solution of free-base porphyrin with Pd(OAc)₂ in acetonitrile instead of the CHCl₃/MeOH mixture,⁴⁵ in which reduction of Pd(OAc)₂ by MeOH occurs.⁴⁶ The replacement of MeOH by acetonitrile enabled to obtain all targeted complexes **PdR¹PP–PdR⁴PP** in nearly quantitative yields (91–100%), although the reduction of palladium acetate is still observed to some extent and an excess of the palladium salt should be employed to achieve a complete complex formation.

Platinum(II) complexes with the phosphorylated porphyrins **PtR¹PP–PtR²PP** were prepared according to a published procedure,^{47–49} by reacting the corresponding free-base porphyrins **H₂R¹PP–H₂R⁴PP** with PtCl₂ in boiling benzonitrile under argon. Complexes **PtDTolPP** and **PtD(CMP)PP** were obtained in 85 and 70% yield, respectively. Surprisingly, **H₂DMesPP** and **H₂TMesPP** were less reactive than **H₂DTolPP** and **H₂D(CMP)PP** porphyrins. In spite of prolonged heating and larger amounts of PtCl₂, **PtDMesP** could only be isolated in 18% yield. When complexation of **H₂TMesPP** was performed under the same conditions, the starting compound was also completely consumed, but the non-phosphorylated complex **PtTMesP** was only isolated in low yield (15%), because of C–P bond breaking. This side reaction has already been observed for porphyrinylphosphonate diesters under acidic conditions.^{34,49} When sodium acetate was added to the reaction mixture to prevent a pH increase during the course of the metal insertion, complexation proceeded smoothly even at 150 °C, affording the desired product in 89% yield.

The reference compounds, namely the Pt(II) and Pd(II) complexes with the non-phosphorylated porphyrins (**MR¹P–MR⁴P**, Chart 1), have not been described in the literature so far, excepted for the **PdDTolP** complex that was presented without full characterization by Ouyang *et al.*⁵⁰ as a side product of the oxidative coupling of metal chlorins. Herein, these complexes were obtained for the first time from the free-base A₂-type porphyrins by the aforementioned procedures for complexation (see the Experimental Section).

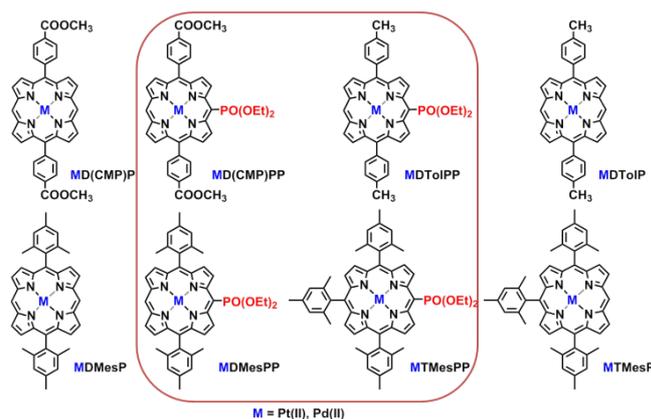


Chart 1. Chemical structures of the investigated phosphorylated Pt(II) and Pd(II) porphyrins **MR¹PP–MR⁴PP** and of the corresponding reference compounds **MR¹P–MR⁴P**.

All isolated platinum(II) and palladium(II) porphyrinates were characterized in solution by ¹H and ³¹P NMR and UV–vis spectroscopies, as well as mass spectrometry (MALDI-TOF and HR-ESI; see the Experimental Section and ESI). ¹H, ¹³C, and ³¹P NMR spectra of **MR¹P–MR⁴P** and **MR¹PP–MR⁴PP** in CDCl₃ or CDCl₃/CD₃OD mixture (2:1 v/v) were in accordance with the proposed structures of the compounds (Figures S3–S6, S11–S16, S19–S22, S27–S32 in the ESI). In the IR spectra of the **MR¹PP–MR⁴PP** porphyrins (Figures S63–S66, S71–S74 in the ESI), intense signals at ~ 950, ~1050, and ~1250 cm⁻¹ were observed in addition to the basic characteristic bands of the porphyrin macrocycles and aryl substituents. Since these features are absent in the spectra of the respective reference compounds **MR¹P–MR⁴P** (Figures S59–S62, S67–S70 in the ESI), they are assigned to the P=O and P–O vibrations of the phosphonate group (see the Experimental Section).

Single crystals for three complexes, namely **PdDTolIP**, **PdD(CMP)PP**, and **PtD(CMP)PP**, were grown by slow diffusion of *n*-hexane into a solution of the corresponding compound in dichloromethane or chloroform. The structures were determined by single crystal X-ray diffraction analysis performed at 100 K. The summary of the crystallographic data is presented in Table S1 (ESI). All complexes crystallized as solvates in the triclinic *P*-1 space group. The asymmetric unit includes one porphyrin molecule and one solvent molecule: CH₂Cl₂ for **PdDTolIP**, or CHCl₃ for **PdD(CMP)PP** and **PtD(CMP)PP** (Fig. 1). Platinum(II) and palladium(II) complexes with the same porphyrin [**PdD(CMP)PP** and **PtD(CMP)PP**] are isostructural with very close unit cell parameters (Fig. 1 and Table S1, ESI). The molecular structures of the

three characterized complexes are typical for palladium(II) and platinum(II) porphyrins. The metal centers have a square-planar coordination geometry formed by four pyrrole nitrogen atoms. The palladium and platinum atoms are nearly coplanar with the N₄ plane of the porphyrin core with a slight displacement of 0.0124(9), 0.011(1), and 0.011(1) Å for **PdDTolPP**, **PdD(CMP)PP**, and **PtD(CMP)PP**, respectively. The mean metal–nitrogen distances are 2.016, 2.018, and 2.019 Å for **PdDTolPP**, **PdD(CMP)PP**, and **PtD(CMP)PP**, respectively, which is comparable to the metal–nitrogen distances reported for other palladium(II) and platinum(II) porphyrins.^{21,45,51,52} The molecular structure of the palladium complex with *p*-tolyl substituted porphyrin **PdDTolPP** reveals a slightly ruffled conformation of the macrocycle, in contrast to the virtually planar porphyrin core found for **PdD(CMP)PP** and **PtD(CMP)PP**. Perspective views along the porphyrin plane are shown in Fig. 1. The largest deviations from the porphyrin N₄ mean plane are observed for the *meso*-carbon C(14) [0.268(3) Å] and β -carbon C(2) [0.248(4) Å] atoms in the **PdDTolPP** crystal structure. In case of **PdD(CMP)PP** and **PtD(CMP)PP**, the macroheterocycle is more flat and the largest deviations are observed for the α -carbon atoms closest to the phosphoryl substituent [0.151(4) Å and 0.141(4) Å, respectively].

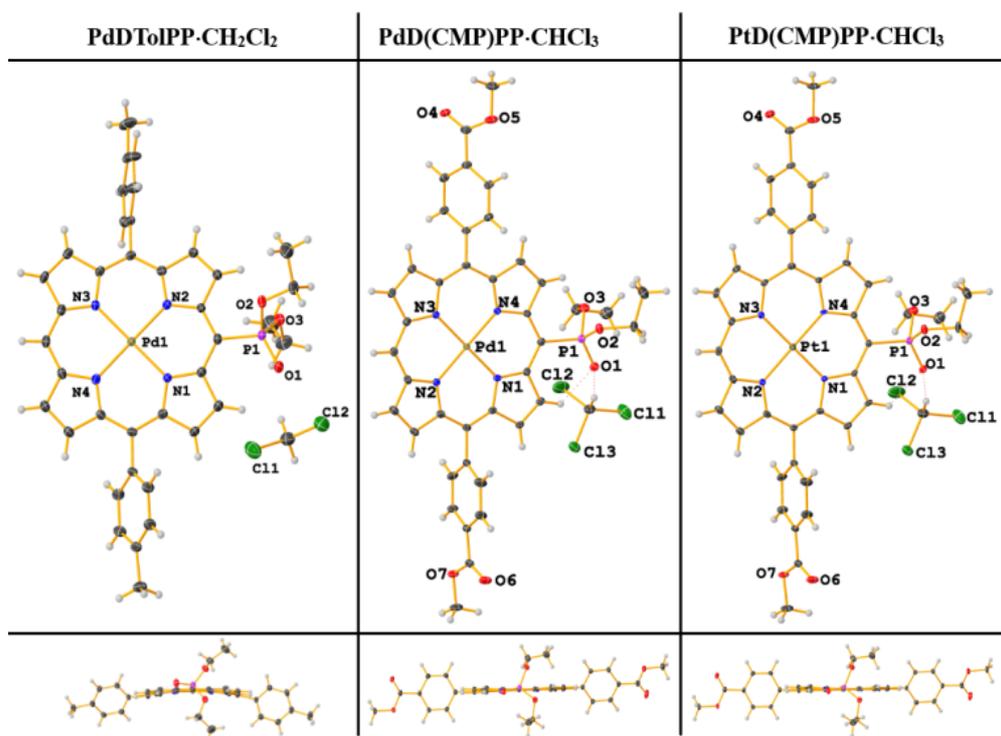


Fig. 1 The asymmetric unit and respective view along the porphyrin plane of **PdDTolPP·CH₂Cl₂** (left), **PdD(CMP)PP·CHCl₃** (middle) and **PtD(CMP)PP·CHCl₃** (right) and respective view along the porphyrin plane. Thermal ellipsoids are at the 50% probability level.

In all structures, the phosphoryl group is twisted with respect to the N₄ plane in such a way that the P=O bond lies almost in the mean porphyrin plane, forming an intramolecular C_β-H...O hydrogen bond [**PdDTolPP**: C(3)...O(1) = 2.857 Å, angle C(3)-H(3)...O(1) = 124.9°; **PdD(CMP)PP**: C(3)...O(1) = 2.851 Å, angle C(3)-H(3)...O(1) = 127.2°; **PtD(CMP)PP**: C(3)...O(1) = 2.857 Å, angle C(3)-H(3)...O(1) = 127.2°]. The ethoxy groups point out above and below the porphyrin plane (Fig. 1). The torsion angles O(1)-P(1)-C(1)-C(2) are 1.63, 14.1, and 13.8° for **PdDTolPP**, **PdD(CMP)PP**, and **PtD(CMP)PP**, respectively. The P-O bond distances of 1.467(2)-1.472(2) Å (for P = O) and 1.568(2)-1.586(2) Å (for P-O) are typical for *meso*-substituted phosphorylporphyrins.^{31,45,53} The *meso*-phenyl rings are twisted with respect to the porphyrin core by the following angles: 60.0 and 78.9° for **PdDTolPP**, 85.6 and 63.7° for **PdD(CMP)PP**, 84.3 and 63.7° for **PtD(CMP)PP**.

In all three structures, solvent molecules are involved in intermolecular C-H...O hydrogen bonds with phosphoryl oxygen atoms. In addition, we identified Cl...O and Cl...C contacts that are shorter than the sum of Bondi's van der Waals radii⁵⁴ (Table S8, ESI).

The crystal packing for **PdDTolPP**·CH₂Cl₂, **PdD(CMP)PP**·CHCl₃, and **PtD(CMP)PP**·CHCl₃ is discussed in details in the ESI. It is formed mainly by □□□ interactions between porphyrin molecules (Figure S75, Table S9-10, ESI). However, such interactions should not play any role for photophysical measurements in solutions owing to the low porphyrin concentrations (*c* < 10⁻⁵ M) used for these experiments.

Photophysical properties

The UV-vis absorption spectra of palladium(II) and platinum(II) complexes with the phosphorylated porphyrins **MR¹PP**-**MR⁴PP** (Fig. 2, Table 1 and Table S11 in the ESI) were measured in dilute toluene solutions at room temperature and compared to those of the respective complexes formed with the non-phosphorylated A₂-type porphyrins **MR¹P**-**MR⁴P**.

Changing aryl substituents from the electron-donating *p*-tolyl to the electron-accepting *p*-(carbomethoxy)phenyl group has only a little effect on the electronic spectra of the porphyrins. The spectrum of both Pd(II) complexes [**PdDTolPP** and **PdD(CMP)PP**] exhibit a Soret band

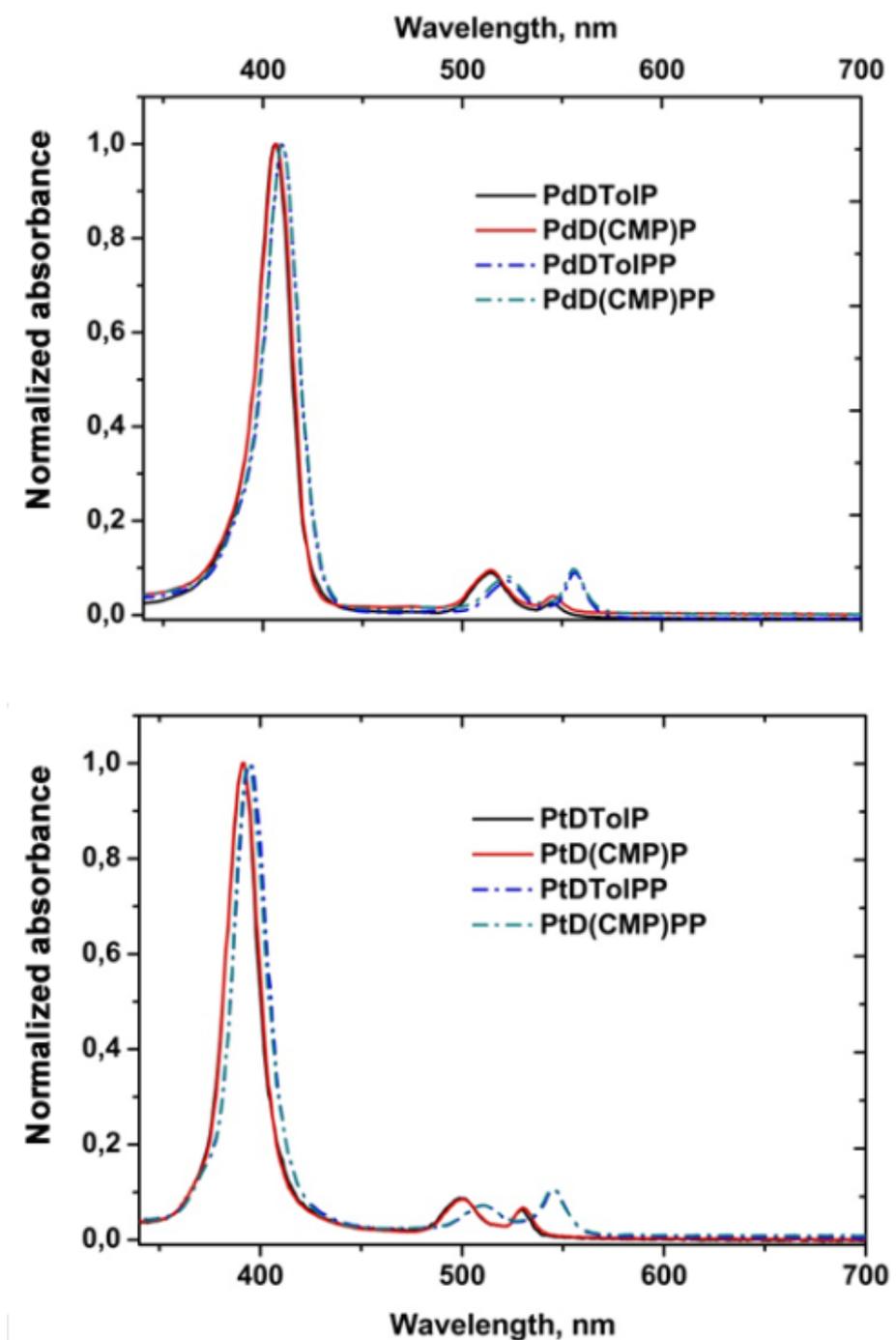


Fig. 2 UV-vis absorption spectra of the Pd(II) and Pt(II) phosphorylated **MR¹PP–MR⁴PP** porphyrins and corresponding non-phosphorylated **MR¹P–MRR⁴P** counterparts in toluene.

at 409 nm and two Q(1,0) and Q(0,0) bands at 522 and 556 nm, respectively. The absorption maxima of the corresponding platinum(II) complexes **PtDTolIP** and **PtD(CMP)PP** are hypsochromically shifted by ~15 nm compared to those of the palladium(II)

phosphorylporphyrins. The maximum of the Soret band of these complexes is located at 395 nm, while that of the Q(1,0) and Q(0,0) bands is centred at 511 and 546 nm, respectively.

Substitution in the *meso*-position of the porphyrin macrocycle with an electron-withdrawing phosphoryl group results in bathochromic shifts of all absorption maxima. Thus, the Soret band for the **MR¹PP–MR⁴PP** [M = Pt(II), Pd(II)] porphyrins was shifted by 3–5 nm and the Q bands by ~8–15 nm, in comparison with the corresponding **MR¹P–MR⁴P** complexes that have no phosphoryl group. The intensity of the Q(0,0) band for both Pt(II) and Pd(II) phosphorylated porphyrins is higher than that for the **MR¹P–MR⁴P** counterparts. This behavior can be explained by the Gouterman four-orbital model,⁵⁵ namely that electron-withdrawing phosphoryl group at the *meso*-position may increase the a_{1u} – a_{2u} orbital energy spacing and thus the intensity of the Q(0,0) transition.

The Pt(II) and Pd(II) complexes with phosphorylated porphyrins **MR¹PP–MR⁴PP** show moderate phosphorescence in deoxygenated solution at room temperature; their photophysical data are listed in Table 1 and Table S11 (ESI). Fig. 3 shows representative examples of normalized excitation and emission spectra of the complexes under investigation [**MR¹PP** and **MR²PP**, with M = Pt(II) and Pd(II)] together with those corresponding to the reference compounds [**MR¹P** and **MR²P**, with M = Pt(II) and Pd(II)] in toluene at 298 K. There is a close resemblance between the excitation and ground-state absorption spectra of all studied complexes. No fluorescence is detectable for the platinum(II) complexes and very little fluorescence ($\tau = 3.75$ ns, $\phi_F < 0.1\%$) for the respective palladium(II) complexes. In general, the emission profile of phosphorylated Pt(II) and Pd(II) porphyrinates is nearly identical to that of non-phosphorylated derivatives, but as can be observed, the insertion of the electron-withdrawing phosphoryl group results in a bathochromic shift of the emission band by *ca.* 25–30 nm compared to the parent A₂-type porphyrins. A narrow Q(0,0) emission band [full width at half-maximum intensity (FWHM) = 35 nm] at 692 nm with a weak Q(1,0) band at 765 nm is observed for phosphorylated Pd(II) porphyrinates (Fig. 3a).

Absolute quantum yields (ϕ_{em}) and luminescence lifetimes (τ) of the complexes were measured in deoxygenated toluene solutions. For the palladium(II) *meso*-phosphorylporphyrins, the observed ϕ_{em} and τ values are 4.3% and 640 μ s for **PdDTolPP**, 5.8% and 633 μ s for **PdD(CMP)PP**, 3.4% and 770 μ s for **PdDMesPP**, and 4.1% and 858 μ s for **PdTMesPP**. The corresponding platinum(II) complexes exhibit a

hypsochromically shifted ($\sim 20\text{--}25\text{ nm}$) [compared to Pd(II) porphyrinates] narrow Q(0,0) emission band (FWHM = 30 nm) at 670 nm with a weak Q(1,0) band at 739 nm (Fig. 3b). They also possess 2.5–3-fold higher luminescence quantum yields (9.2–11%) and 10-fold lower luminescence decay times [65, 69, 64, and 56 μs for **PtDTolPP**, **PtD(CMP)PP**, **PtDMesPP** and **PtTMesPP**, respectively].

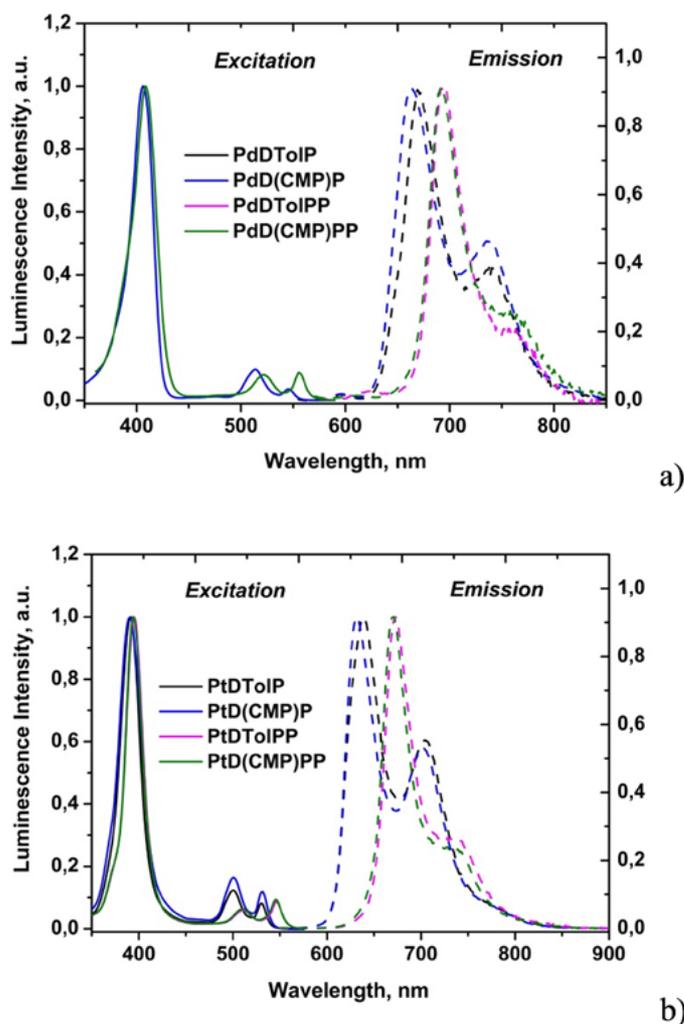


Fig. 3 Excitation [$\lambda_{\text{em}} = \lambda_{\text{max}}$ (nm)] and emission [$\lambda_{\text{ex}} = \lambda_{\text{abs}}$ Q(1,0) nm] spectra of the Pd(II) (a) and Pt(II) (b) complexes **MR¹PP–MR²PP** in deoxygenated toluene at 298 K.

Furthermore, the emission spectra and decay times were recorded at 77 K, because the data obtained at low temperature are more accurate than those at 298 K due to the absence of competing processes (Table 1; Fig. S76–77 and Table S11 in the ESI). At low-temperature, the ratio of the emission intensities for the red to NIR emission peaks is much smaller for the dyes without a phosphoryl group (especially in the case of platinum(II) complexes) than for the phosphorylated porphyrins. The obtained luminescence maxima at 77 K were used for estimation of the triplet excited-state energy (E_T), which are summarized in Table 1 and Table S11 (ESI). Introduction of the electron-withdrawing group leads to a decrease in energy of the triplet excited state, bringing the E_T values of the dye closer to the energy of the $^1\Sigma_g^+$ level of dioxygen ($E = 13121 \text{ cm}^{-1}$).⁵⁶ According to the data obtained at 77 K, decay times of phosphorylated Pt(II) and Pd(II) complexes are slightly reduced with respect of those found for the corresponding **MR¹P–MR⁴P** diarylporphyrins. Among the phosphorylated complexes, the following trend is observed for the phosphorescence decay time τ at 77 K: **MD(CMP)PP** > **MDTolPP** > **MDMesPP** > **MTMesPP**. A similar tendency is observed in the case of the luminescence quantum yields determined at room temperature. Thus, the combination of electron-withdrawing phosphoryl and *p*-(carbomethoxy)phenyl substituents leads to the most favorable photophysical parameters. The phosphorylation of the porphyrin macrocycle results in reduction of both ϕ_{em} and τ , which can be explained by the increased probability of the non-radiative deactivation of the triplet excited state. However, the obtained values are consistent with those known for the most photostable palladium(II) and platinum(II) porphyrinates (**PtF₂₀TPP** and **PdF₂₀TPP**),^{20,57} presented in the literature, so metalloporphyrins presented in this work are suitable for elaborating oxygen sensing devices.

Table 1. Photophysical properties of the phosphorylated **PtR¹PP–PtR⁴PP** porphyrins and of the non-phosphorylated **PtR¹P–PtR⁴P** counterparts in toluene

Dye	λ_{abs} ($\epsilon \times 10^{-3}$) (nm, M ⁻¹ cm ⁻¹)	λ_{em}^a (nm)	ϕ_{em}^b	τ^a (μ s)	λ_{em} (77 K) ^c (nm)	τ (77 K) ^c (μ s)	E_T^d (cm ⁻¹)
PtTMesPP	402 (136), 515 (12), 549 (9)	674	11	56	666, 741	94	15020
PtDMesPP	394 (211), 510 (13), 545 (21)	670	9.2	64	662, 734	117	15100
PtDTolPP	396 (257), 511 (17), 546 (26)	672, 740(sh.)	11	65	663, 737	119	15080
PtD(CMP)PP	395 (325), 510 (28), 546 (41)	671, 740(sh.)	11	69	660, 735	122	15150
PtTMesP	397 (184), 506 (18), 536 (9)	650	9.3	83	643, 712	122	15550
PtDMesP	391 (241), 501 (19), 532 (16)	637	13	82	633, 701	122 ^e	15800
PtDTolP	393 (143), 500 (24), 529 (18)	639, 705(sh.)	15	87	635, 703	133	15748
PtD(CMP)P	392 (206), 502 (18), 531 (14)	632, 702(sh.)	16	82	634, 700	137	15773
PtF₂₀TPP^f	390 (323), 504 (23), 538 (29)	647, 710 (sh)	8.8	60	-	-	-

^a Measured in diluted toluene solutions at 298 K deoxygenated by nitrogen bubbling. ^b The absolute emission quantum yields were determined with an integrating sphere. ^c Measured in frozen glasses (toluene/tetrahydrofuran 4:6 v/v). ^d Calculated from the emission spectra at 77 K. ^e Non mono-exponential signal with a much longer component (1.56 ms) is present. ^f Data in CH₂Cl₂ are taken from ref.²⁰

Quenching by molecular oxygen in solution

Phosphorescence of all the investigated complexes is effectively quenched in solution by molecular oxygen. Studies were conducted with **PdDTolPP**, **PdD(CMP)PP**, **PtDTolPP**, **PtD(CMP)PP** and their non-phosphorylated counterparts. The results are presented in Fig. 4 and Fig. S78 (ESI). However, the Pd(II) complexes show significantly lower luminescence brightness and very high sensitivity even at low dioxygen partial pressure in solution (pO_2), making precise measurements challenging. Thus, the oxygen quenching processes in solution were investigated in detail only for the phosphorylated **PtDTolPP** and **PtD(CMP)PP** porphyrins, and for the corresponding non-phosphorylated **PtDTolP** and **PtD(CMP)P** counterparts.

The dioxygen quenching of phosphorescence intensity (Fig. 4a) and decay time (Fig. 4b) can be interpreted in terms of the Stern–Volmer relation, $I_0/I = \tau_0/\tau = 1 + k_q \square_0[O_2] = 1 + K_{sv}pO_2$, where I_0 and I denote the phosphorescence intensity in the absence and the presence of dioxygen, respectively; τ_0 and τ correspond to the related decay lifetimes, k_q is the bimolecular quenching constant; K_{sv} is the Stern-Volmer constant.

As expected, all the studied complexes show good linearity of τ_0/τ as a function of pO_2 between 0 and 1.95 kPa, which indicates an effective homogeneous oxygen quenching in solution. The calculated K_{sv} values for both platinum(II) phosphorylated porphyrins **PtDTolPP** and **PtD(CMP)PP** are similar [10.42 kPa^{-1} for **PtDTolPP** and 10.53 kPa^{-1} for **PtD(CMP)PP**], but lower than the corresponding K_{sv} values for the Pt(II) complexes lacking the phosphoryl group [15.08 kPa^{-1} for **PtDTolP** and 12.36 kPa^{-1} for **PtD(CMP)P**]. This is mostly due to the longer decay times of the latter. On the other hand, the bimolecular quenching constants $k_q = K_{sv}/\tau_0$ are also slightly smaller in the case of phosphorylated complexes [$k_q = 173, 160, 156, 153 \text{ Pa}^{-1} \text{ s}^{-1}$ for **PtDTolP**, **PtD(CMP)P**, **PtDTolPP**, and **PtD(CMP)PP**, respectively]. This may be due to the slow diffusion rates in solution of the bulky phosphorylated porphyrins, or to the proximity of the E_T value of the dyes with the $^1\Sigma_g^+$ energy level of dioxygen. Nevertheless, the sensitivity of the platinum(II) phosphorylated porphyrin complexes is comparable to that of the commonly used **PtOEP** ($210 \text{ Pa}^{-1} \text{ s}^{-1}$ in toluene).²⁵ Thus, it can be concluded that these dyes and their derivatives are suitable indicators for developing optical oxygen sensors.

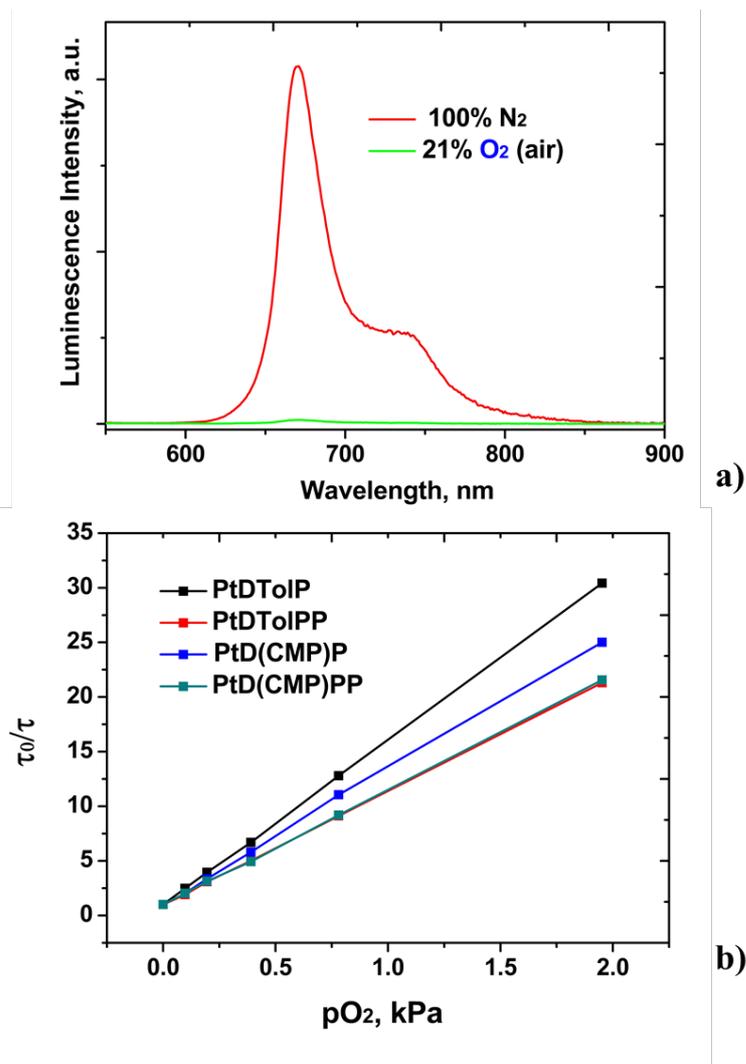


Fig. 4. Luminescence spectra of **PtDTolIP** in toluene at 298 K in the absence and in the presence of dioxygen ($\lambda_{\text{ex}} = 511 \text{ nm}$) (a); Stern-Volmer plots for **PtR¹,R²P** and **PtR¹,R²PP** in toluene ($\sim 7 \times 10^{-6} \text{ M}$, 295 K) (b).

Photostability

Increasing interest in the development of oxygen sensors^{10,58} and imaging agents,^{7,8} which should be stable under different environmental conditions and useful for measurements in biosamples and *in vivo*, explains the particular attention paid on improving their photostability.^{59–61} While the photoinduced degradation pathways of many organic fluorophores, such as coumarins, rhodamines, and cyanines, have been explored to some extent,¹³ the reactions involved in the photobleaching of porphyrins and phthalocyanines are less well understood. Various porphyrins have been considered as potential dioxygen sensors,^{41,62–67} and for some of the series the influence of their peripheral substituents on the efficiency of O₂ detection has been investigated.^{20,22,49,68} However, discussions of their photostability are commonly limited to a general

statement that the introduction of electron-withdrawing substituents at the tetrapyrrolic macrocycle or at peripheral aryl groups increases the photostability, as typically observed for other organic fluorophores.^{8,20,22} To the best of our knowledge, mechanisms of the photoinduced degradation of porphyrins in solution have still not been reported and data accumulated for photobleaching of phthalocyanines⁶⁹ are also insufficient for suggesting any general reaction pathway.

In this work, we compared the photostability of different porphyrinylphosphonate diesters (**MR¹PP–MR⁴PP**) and analogous compounds (**MR¹P–MR⁴P**) that lack the phosphorous substituent, to get a deeper insight on the influence of peripheral substituents at the tetrapyrrolic macrocycle on the photostability of porphyrins.

The photodegradation of all platinum(II) and palladium(II) complexes was studied at first in DMF under the same conditions. The porphyrin solutions ($c = 5 \mu\text{M}$) were irradiated at 25 °C in air using a 400 W lamp, which emits in the visible and IR range (350–800 nm; Figure S79, ESI). The photoinduced degradation kinetics of each tested compound was monitored by absorption spectrophotometry at the maximum of the Soret band until nearly or complete disappearance. The electronic absorption spectrum of the resulting solution was recorded at the end of the irradiation period. In all cases, the final solution did not absorb visible light anymore. No further efforts to identify the products were undertaken.

Although we have followed a commonly used methodology for studying the photobleaching of organic compounds,⁷⁰ it should be stressed that these experiments allow only a semi-quantitative comparison within a defined series of analogous chromophores. Indeed, the degradation rate depends, among other factors, on the concentration of the chromophore, the photon absorption cross section, the intersystem crossing efficiency, and the triplet state lifetime. For comparison purposes, the concentration and the total amount of absorbed photons should be identical for all investigated substances, which is in practice an unachievable situation as the light absorptivity ($\int \epsilon(\lambda) d\lambda$) of even closely structurally related molecules might significantly vary with the nature of peripheral substituents born by the aromatic moiety.

The time course of the normalized absorbance decay recorded over more than 80 h of continuous irradiation for seven platinum porphyrinylphosphonate diesters is depicted in Fig. 5. The inset of Fig. 5 clearly shows an induction period during which the absorbance remains nearly constant over about 25–35 min, followed by an abrupt multiphasic decrease associated to photobleaching. The systematic observation of an

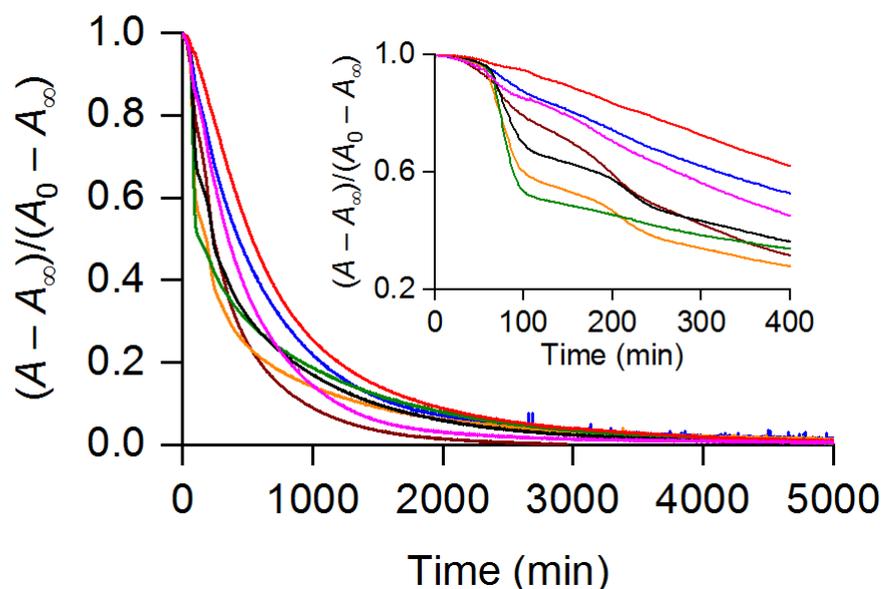


Fig. 5. Photodegradation of platinum porphyrins ($c = 5 \mu\text{M}$) in DMF at $25.0(5)^\circ\text{C}$ under vis–NIR irradiation with an Osram Powerstat HQI BT 400 W lamp. Colour code: **PtDTolP** (brown), **PtDTolPP** (orange), **PtD(CMP)PP** (green), **PtDMesP** (magenta), **PtDMesPP** (black), **PtTMesP** (blue), **PtTMesPP** (red).

initiation phase strongly supports the photoinduced build-up of radicals, which are presumably responsible for different degradation processes occurring consecutively or simultaneously as suggested by the non-monotonous decay curves of several studied compounds.

As shown in Fig. 5, the rate of the porphyrin photobleaching depends strongly on the substituents attached at the periphery of the tetrapyrrolic core. For example, the extend of degradation during the first 100 min of irradiation varies from 5 to 45%, depending on the porphyrin structure. Moreover, the number and nature of peripheral substituents have a most significant influence on the peculiar shape of the kinetic traces. Accordingly, the compounds studied herein can be divided into two sets. The first group is formed by the mesityl-substituted porphyrins **PtDMesP**, **PtTMesP**, and **PtTMesPP**, which are decomposed according to one predominant pathway, like most of the previously studied organic phosphores.^{13,70}

The absorbance decline can be approximated by a single exponential decay function. Moreover, no noticeable band shift could be detected over time when the degradation of **PtTMesPP** was monitored over the entire 350–600 nm range (Fig. S80, ESI). The photobleaching rates of these compounds can be compared by considering the pseudo-first-order rate constants summarized in Table 2 derived by nonlinear least squares

adjustment of the experimental data after cutting the induction period (Fig. S83–S85, ESI).

Table 2. Pseudo-first-order rate constants (k_{obs}) for the photodegradation of platinum porphyrins in DMF at 25.0(5) °C

Compound	k_{obs} (s ⁻¹)	$k_{\text{obs}} / k_{\text{obs}}(\text{PtDMesP})$
PtDMesP	3.66×10^{-5}	1
PtTMesP	2.83×10^{-5}	0.77
PtTMesPP	2.66×10^{-5}	0.73

The introduction of a third mesityl substituent at the *meso* position of the macrocycle increases the photostability of platinum 5,15-dimesitylporphyrinate (**PtDMesP**), as **PtTMesP** decomposes 1.3-fold more slowly than **PtDMesP**. Comparison of the photobleaching rate constants of **PtTMesP** and **PtTMesPP** reveals that the introduction of the diethoxyphosphoryl group at *meso* position of the macrocycle improves the photostability of the porphyrin only slightly, in spite of the strong electron-withdrawing character of this group.

All the other platinum porphyrinates [**PtDMesPP**, **PtDTolP**, **PtDTolPP**, and **PtD(CMP)PP**] belong to the second group of studied complexes, for which the intricate decomposition kinetics proceeds according to several pathways, as highlighted by a first slowdown of the degradation rate some minutes only after the initial boost, which is followed by a second boost before the final rate deceleration. In addition, the degradation of **PtDTolPP** was also monitored over the entire 350–600 nm range (Fig. S81, ESI). Although no noticeable band shift could be detected during the first 40 h of irradiation ($\lambda_{\text{abs}} = 395, 511, \text{ and } 546 \text{ nm}$), the Soret band undergoes a slight bathochromic shift of ca. 7 nm during the final stage of the degradation (40–116 h). This particular succession of fast and slow regimes hampers a quantitative analysis. All compounds belonging to this second category are less stable than those of the first series, even if the electron-withdrawing diethoxyphosphoryl substituent is present at the periphery of the macrocycle. Regarding the kinetics of decomposition of **PtDMesPP**, **PtDTolPP**, and **PtD(CMP)PP**, we can note that the replacement of both mesityl substituents by *p*-methylphenyl or *p*-methoxycarbonylphenyl groups accelerates photobleaching. Also, the introduction of the phosphorous substituent on **PtDMesP** and **PtDTolP** has negative influence on the porphyrin degradation, at least in the early stage of the process. Considering the electronic effects of all these substituents, it can be concluded that the electron-withdrawing character does not always increase the photostability in this series

of porphyrins, as claimed earlier for porphyrins bearing pentafluorophenyl or other strongly electron-withdrawing substituents.^{1,8}

Interestingly, the introduction of a diethoxyphosphoryl group can change the photodegradation pathways in DMF, as clearly revealed by the comparison of the bleaching curves of **PtDMesP** and **PtDMesPP**. While the degradation of **PtDMesP** can be considered in first approximation as a pseudo-first order process, the decomposition of the phosphorylated derivative involves several pathways. This might be reasonably explained by the reactivity of the *meso* site at which a bulky diethoxyphosphoryl group is attached. We have already reported on the unusual C–P bond cleavage undergone by porphyrinylphosphonate diesters under acidic conditions.^{30,45} It seems that irradiation also accelerates the cleavage of C–P bond.

Another conclusion is that the compounds bearing mesityl groups are the most stable ones, while an increasing number of hydrogen atoms in either *meso* and/or in *ortho* positions of *meso* aryl substituents accelerates the degradation of platinum porphyrinates. These less stable macrocycles are decomposed according to several pathways, presumably because these *meso* and *ortho* hydrogen atoms take part in photo-induced redox processes upon irradiation.

Next, the photostability of the **PdD(CMP)PP**, **PdDMesPP**, **PdTmesP**, **PdTmesPP** palladium complexes and of the reference compound **PdF₂₀TPP** was investigated under the same conditions. The results are summarized in Fig. 6 and compared to those of the corresponding platinum complexes in Fig. S82 (ESI). For a given substitution pattern, the palladium porphyrin is significantly less stable than the platinum counterpart. Within the palladium series, the trimesitylphosphorylporphyrin **PdTmesPP** displays a higher photostability compared to that of dimesityl- (**PdDMesPP**) and bis[*p*-methoxycarbonyl]phenyl] **PdD(CMP)PP** phosphorylporphyrins. Surprisingly, **PdF₂₀TPP**, known for its high photostability,^{1,8} was significantly more reactive than the mesityl-substituted derivatives under the conditions employed in this work.

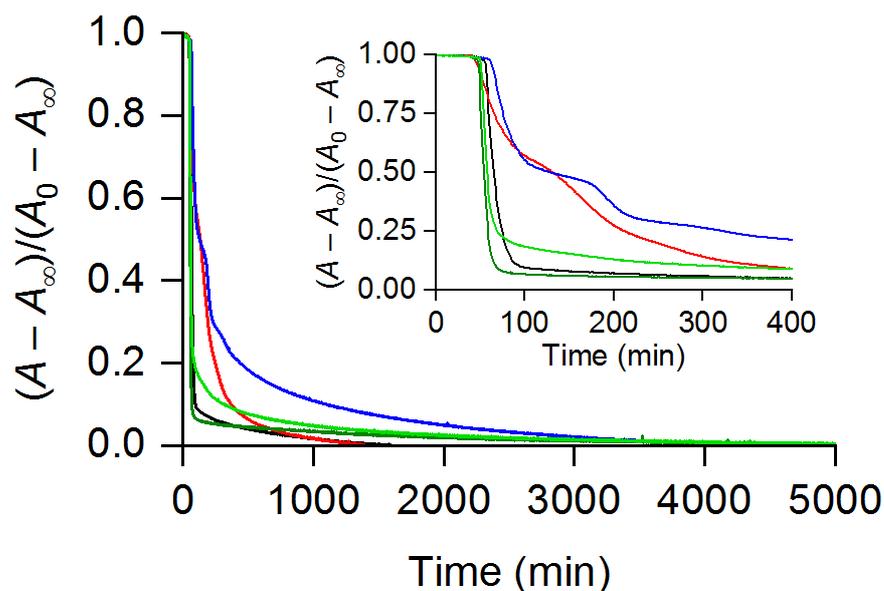


Fig. 6. Photodegradation of platinum porphyrins ($c = 5 \mu\text{M}$) in DMF at $25.0(5)^\circ\text{C}$ under vis-NIR irradiation with an Osram Powerstar HQI BT 400 W lamp. Colour code: **PdD(CMP)PP** (dark green), **PdDMesPP** (black), **PdTMesP** (blue), **PdTMesPP** (red), **PdF₂₀TPP** (green).

Next, the photodegradation kinetics of the most stable palladium complexes **PdTMesPP** and **PdDMesPP**, together with the platinum porphyrin **PtTMesP** were also investigated in toluene under identical irradiation conditions. In these experiments, the porphyrin concentrations slightly varied ($4.3\text{--}5.2 \mu\text{M}$) because the initial absorbance at the maximum of the Soret band was fixed to $A = 1.0$ (Fig. 7).

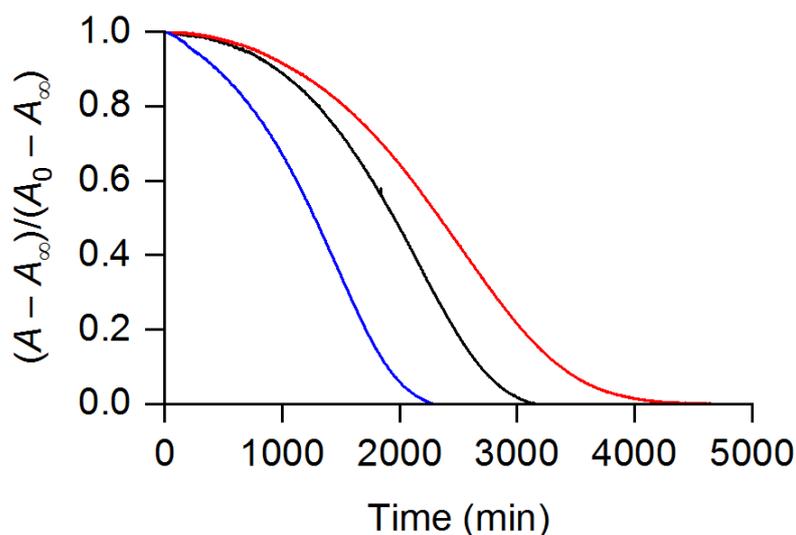


Fig. 7. Photodegradation of **PdDMesPP** ($c = 4.6 \mu\text{M}$, black), **PdTMesPP** ($c = 4.3 \mu\text{M}$, red), and **PtTMesP** ($c = 5.2 \mu\text{M}$, blue) in toluene at $25.0(5)^\circ\text{C}$ under vis-NIR irradiation with an Osram Powerstar HQI BT 400 W lamp.

Absorbance decay curves of nearly sigmoidal shapes were observed, presumably because of the occurrence of side reactions involving photooxidation products of toluene and palladium phosphorylporphyrins. Nevertheless, it has been noted that the trimesityl-substituted derivative **PdTMesPP** resists better to photobleaching in toluene compared to **PdDMesPP**, in agreement with the results obtained in DMF. Remarkably, the platinum complex **PtTMesP** exhibit higher reactivity in toluene than the two other investigated palladium porphyrins, whereas the reverse trend was observed in DMF.

Finally, the photostability of four palladium(II) [**PdDTolIP**, **PdDTolPP**, **PdD(CMP)PP**, and **PdTPP** (TPP = 5,10,15,20-tetraphenylporphyrin)] (Fig. S86, ESI) and two platinum(II) complexes [**PtDTolIP** and **PtD(CMP)PP**] (Fig. S87, ESI) was investigated in air-saturated toluene solutions stored under day light during 2 months. It was observed that the platinum(II) derivatives **PtDTolIP** and **PtD(CMP)PP** are less stable compared to their palladium(II) counterparts [**PdDTolIP** and **PdD(CMP)PP**], in accordance with the aforementioned investigations performed in the same solvent. Concomitantly to the absorbance decrease, new absorption bands in the case of platinum(II) phosphorylated porphyrinates were observed at higher wavelengths after two months of exposure to sunlight. Moreover, the phosphorylated derivatives **PdDTolPP** and **PdD(CMP)PP** degraded more slowly (25–35%) than **PdDTolIP** and **PdTPP** (75–90%). So, under these conditions the introduction of the electron-withdrawing phosphoryl group in the palladium(II) porphyrins seems to increase their photostability, as it was observed for **PtTMesP** and **PtTMesPP** in DMF.

It turns out that the photobleaching rate of phosphorylporphyrins is highly solvent dependent. Likewise, the central metal atom plays a crucial role, palladium complexes being more reactive than platinum ones in DMF, why the trend is reversed in toluene. Not surprisingly, the number and nature of *meso* substituents have also a significant impact on the degradation pathways and rates. Peripheral aromatic groups with *ortho* hydrogen atoms are less protective than those possessing methyl groups (*e.g.* mesityl) at the same position. Among the series of compounds considered in this work, the trimesitylporphyrinylphosphonate diesters **PdTMesPP** and **PtTMesPP** exhibit the highest photostability in air-saturated DMF solution and toluene. Thus, they are presumably the most appealing phosphorylated derivatives for dioxygen sensing in biological samples or covalent immobilization on solid supports to prepare sensing devices, including optical fibers.

Conclusions

In conclusion, Pd(II) and Pt(II) complexes with electron-deficient *meso*-(diethoxyphosphoryl)porphyrins have been conveniently prepared and characterized. The complexes under investigation emit at room temperature in the near-infrared region (670–770 nm). Absorption and emission spectrophotometric studies indicate that the incorporation of diethoxyphosphoryl group at the *meso* positions of the porphyrin ring results in bathochromic shifts of the absorption (~5 nm for Soret band and ~15 nm for Q bands) and the emission (~25–30 nm) bands of complexes compared to those of the parent A₂-type porphyrins. Phosphorescence quantum yields of the palladium(II) *meso*-phosphorylated porphyrins lie in the range of 3.4 to 5.8%, with lifetimes of 633 to 858 μs in deoxygenated toluene solutions at room temperature. The corresponding platinum(II) complexes exhibit quantum yields in the range of 9.2 to 11%, with luminescence decay time constants of 56 to 69 μs. The obtained values are consistent with those known for the most photostable palladium(II) and platinum(II) porphyrinates (**PtF₂₀TPP** and **PdF₂₀TPP**) reported so far. Luminescent oxygen sensing properties of the Pt(II) complexes were studied in solution. Our results demonstrate effective homogeneous oxygen quenching and good sensitivities, with bimolecular quenching constants (k_q) of ~155 Pa⁻¹ s⁻¹ in toluene for the platinum(II) phosphorylated porphyrinates. It was also observed that the photostability of porphyrinylphosphonates strongly depends on both the solvent and the substituents occurring at the periphery of the macrocycle. Among the studied compounds, platinum(II) and palladium(II) trimesitylphosphorylporphyrins **PdTMesPP** and **PtTMesPP** exhibit a high photostability in air-saturated DMF solution. Thus, phosphorylated metalloporphyrins presented in this work are suitable for preparing dioxygen-sensing devices.

Experimental section

General methods and characterization of compounds

Unless otherwise noted, all chemicals and starting materials were obtained commercially from Acros® or Aldrich®, and used without further purification. Solvents were dried using standard procedures.⁷¹ Chloroform (analytical grade) was purchased from Merck and was distilled over CaH₂. DMA was dried over molecular sieves. Toluene (HPLC-grade) was dried over alumina cartridges using the solvent purification system PureSolv PS-MD-5 model from Innovative Technology. Analytical thin-layer chromatography

(TLC) was carried out using Merck silica gel 60 plates (precoated sheets, 0.2 mm thick, with the fluorescence indicator F254). Column chromatography purifications were carried out on silica gel (MN Kieselgel 60, 63–200 μm , Macherey-Nagel or Silica 60, 63–200 μm , Aldrich). Gel permeation chromatography was carried out using Bio-BeadsTM S-X3 Support (Bio-Rad Laboratories). ¹H, ¹³C, and ³¹P NMR spectra were acquired on Bruker Avance III (600 MHz), Avance III (500 MHz), or Avance II (300 MHz) spectrometers. Chemical shifts are expressed in parts per million (ppm), referenced on the δ scale by using residual non-deuterated solvent signals as internal standard for ¹H and ¹³C NMR spectroscopies and external phosphoric acid (H₃PO₄) for ³¹P NMR spectroscopy. The coupling constants are expressed in units of frequency (Hz). MALDI-TOF mass-spectra were obtained on a Bruker Ultraflex II LRF 2000 mass-spectrometer in positive ion mode with a dithranol matrix. Accurate mass measurements (HRMS) were recorded on a Thermo LTQ Orbitrap XL apparatus equipped with electrospray ionization (ESI) source. Solutions in CHCl₃/methanol (1:1) were used for the analysis. The reported *m/z* values correspond to the most intense peak of the isotopic pattern that were simulated with the Xcalibur software (Thermo). FT-IR spectra were collected at 4 cm⁻¹ resolution on either a FT-IR Nexus (Nicolet) or a Vertex 70v (Bruker) spectrometer equipped with an universal micro-ATR (Pike) or an A225 (Bruker) ATR sampling accessory, respectively. The UV–vis spectra were recorded with a Helios Alpha (Thermo Electron), CARY 50 (Varian), CARY 5000 (Varian) and a Shimadzu 2450 spectrophotometers by using rectangular quartz cells (1–10 mm).

The measurements were performed at the Shared Facility Centers of the Institute of Physical Chemistry and Electrochemistry, RAS, and at the "Plateforme d'Analyses Chimiques et de Synthèse Moléculaire de l'Université de Bourgogne - Pôle Chimie Moléculaire", the technological platform for chemical analysis and molecular synthesis (<http://www.wpcm.fr>).

X-ray crystallography

Single crystals of **PdDTolPP·CH₂Cl₂** were obtained by slow diffusion of *n*-hexane into a solution of the Pd(II) complex in dichloromethane. Single crystals of **PdD(CMP)PP·CHCl₃** and **PtD(CMP)PP·CHCl₃** were grown by slow diffusion of *n*-hexane into solutions of the corresponding Pd(II) and Pt(II) complexes in chloroform. Crystals mounted onto a glass needle using silica oil, were cooled to the data collection

temperature of 100 K. X-ray diffraction experiments for single crystals were performed on a Bruker Kappa Apex II automatic four-circle diffractometer equipped with an area detector (Mo K α sealed-tube X-ray source, $\lambda = 0.71073 \text{ \AA}$, graphite monochromator). The unit cell parameters were refined over whole dataset using SAINT-Plus.⁷² The experimental reflection intensities were corrected for absorption using the SADABS program.⁷³ The structures were solved with the ShelXT⁷⁴ program and refined by the full-matrix least-squares method (SHELXL-2014⁷⁵) on F^2 over the whole dataset in the anisotropic approximation for all nonhydrogen atoms, both routines being implemented in the Olex2 environment.⁷⁶ The H atoms were placed in the geometrically calculated positions with the isotropic temperature factors set at 1.2 times (CH groups) or 1.5 times (CH₃ group) the equivalent isotropic temperature factor of their bonded C atoms. Table S1 (ESI) lists the crystallographic characteristics and details of the diffraction experiments.

X-ray diffraction experiments were performed at the Center for Shared Use of Physical Methods of Investigation at the Frumkin Institute of Physical Chemistry and Electrochemistry, RAS.

Atomic coordinates have been deposited in the Cambridge Crystallographic Data Centre [CCDC deposition codes are 1904438 for **PdDTolPP**, 1904436 for **PdD(CMP)PP**, 1904437 for **PtD(CMP)PP**] and can be obtained on request at www.ccdc.cam.ac.uk/data_request/cif.

Synthesis

Dipyrromethane was prepared by the published procedure.^{77,78} Non-phosphorylated free-base A₂-type porphyrins, namely 5,15-bis(*p*-tolyl)porphyrin (**H₂DTolP**),^{79,80} 5,15-bis[*p*-(methoxycarbonyl)phenyl]porphyrin [**H₂D(CMP)P**],⁸¹ 5,15-dimesitylporphyrin (**H₂DMesP**),^{82,83} and 5,10,15-trimesitylporphyrin (**H₂TMesP**),⁸⁴ were obtained by literature methods.

The starting free-base phosphorylated porphyrins 10-(diethoxyphosphoryl)-5,15-bis(*p*-tolyl)porphyrin (**H₂DTolPP**) and 10-(diethoxyphosphoryl)-5,15-bis[*p*-(methoxycarbonyl)phenyl]porphyrin [**H₂D(CMP)PP**], 10-(diethoxyphosphoryl)-5,15-dimesitylporphyrin (**H₂DMesPP**), and 5-(diethoxyphosphoryl)-10,15,20-trimesitylporphyrin (**H₂TMesPP**) were prepared according to the published procedures.²⁹

General procedure for preparation of Pd(II) complexes. A solution of the free-base porphyrin **H₂R¹P–H₂R⁴P** or **H₂R¹PP–H₂R⁴PP** in a CHCl₃/CH₃CN mixture was stirred and heated. During refluxing, Pd(OAc)₂ (4-16 equiv.) was added into the flask. The reaction mixture was further stirred until complete conversion of starting porphyrin. The degree of consumption of the porphyrin was monitored by MALDI-TOF mass-spectrometry and UV–vis spectroscopy. After cooling to room temperature, the solvent was removed under reduced pressure. The residue was purified by column chromatography on silica gel and gel permeation chromatography to afford **PdR¹P–PdR⁴P** or **PdR¹PP–PdR⁴PP**.

[5,15-Bis(*p*-tolyl)porphyrinato]palladium(II) (PdDTolP) was prepared from 20 mg (0.041 mmol) of **H₂DTolP** and 37 mg (0.163 mmol) of Pd(OAc)₂ dissolved in 11 mL of a CHCl₃/CH₃CN (80:20 v/v) mixture that was refluxed for 15 min. The resulting solid was purified by column chromatography on silica gel using CH₂Cl₂ as eluent to give **PdDTolP** as a pink purple crystalline powder (25.4 mg, 100%). ¹H NMR (600 MHz, CDCl₃, 25 °C): δ_H 2.73 (s, 6H, Ph-CH₃), 7.59 (d, ³J_{H,H} = 7.6 Hz, 4H, *m*-Ph), 8.10 (d, ³J_{H,H} = 7.8 Hz, 4H, *o*-Ph), 9.04 (d, ³J_{H,H} = 4.7 Hz, 4H, Hβ), 9.28 (d, ³J_{H,H} = 4.7 Hz, 4H, Hβ), 10.26 (s, 2H, H_{meso}). MS (MALDI-TOF): *m/z* = 594.08 ([M]⁺, calcd. for C₃₄H₂₄N₄Pd *m/z* = 594.10). HRMS (ESI): *m/z* = 594.10468 ([M]⁺, calcd. for C₃₄H₂₄N₄Pd *m/z* 594.10303). UV–vis [toluene; λ_{max}, nm (log ε)]: 406 (5.50), 514 (4.23), 544 (3.96). IR (neat, cm⁻¹): ν_{max} 2952 (w), 2914 (m), 2849 (m), 1729 (m), 1537 (w), 1509 (w), 1457 (w), 1418 (w), 1391 (w), 1376 (w), 1335 (w), 1302 (w), 1234 (w), 1216 (w), 1194 (w), 1179 (m), 1102 (w), 1068 (m), 1046 (m), 1006 (s), 944 (w), 901 (w), 853 (s), 797 (m), 779 (s), 721 (s), 693 (s), 586 (w), 554 (w).

{5,15-Bis[*p*-(methoxycarbonyl)phenyl]porphyrinato}palladium(II) [PdD(CMP)P] was prepared from 20 mg (0.035 mmol) of **H₂D(CMP)P** and 31 mg (0.138 mmol) of Pd(OAc)₂ dissolved in 9 mL of a CHCl₃/CH₃CN (80:20 v/v) mixture. The reaction mixture was refluxed for 15 min. The resulting solid was purified by column chromatography on silica gel using CH₂Cl₂ as eluent to give **PdD(CMP)P** as a pink purple crystalline powder (19.2 mg, 83%). ¹H NMR (600 MHz, CDCl₃, 25 °C): δ_H 4.14 (s, 6H, Ph-COOCH₃), 8.31 (d, ³J_{H,H} = 8.3 Hz, 4H, *o*-Ph), 8.48 (d, ³J_{H,H} = 8.3 Hz, 4H, *m*-Ph), 8.96 (d, ³J_{H,H} = 4.7 Hz, 4H, Hβ), 9.32 (d, ³J_{H,H} = 4.8 Hz, 4H, Hβ), 10.32 (s, 2H, H_{meso}). MS (MALDI-TOF): *m/z* = 682.11 ([M]⁺, calcd. for C₃₆H₂₄N₄O₄Pd *m/z* = 682.08). HRMS (ESI): *m/z* = 682.08240 ([M]⁺, calcd. for C₃₆H₂₄N₄O₄Pd *m/z* = 682.08269). UV–vis [toluene; λ_{max}, nm (log ε)]: 406 (4.97), 514 (3.94), 544 (3.54). IR

(neat, cm^{-1}): ν_{max} 2917 (w), 2849 (w), 1715 (s, C=O), 1603 (w), 1540 (w), 1507 (w), 1457 (w), 1427 (w), 1397 (w), 1337 (w), 1305 (w), 1270 (br, m), 1194 (w), 1176 (w), 1152 (w), 1107 (m), 1095 (m), 1071 (m), 1009 (s), 969 (w), 919 (w), 874 (w), 847 (s), 821 (m), 780 (s), 754 (s), 723 (s), 693 (s), 638 (w), 592 (w), 569 (w), 554 (m).

[10-(Diethoxyphosphoryl)-5,15-bis(*p*-tolyl)porphyrinato]palladium(II)

(PdDTolPP) was prepared from 7 mg (0.011 mmol) of **H₂DTolPP** and 10 mg (0.045 mmol) of Pd(OAc)₂ dissolved in 3 mL of a CHCl₃/CH₃CN (80:20 v/v) mixture. The reaction mixture was refluxed for 15 min. The resulting solid was purified by column chromatography on silica gel using a CHCl₃/*n*-hexane (1:1 v/v) as eluent to give **PdDTolPP** as a pink purple crystalline powder (8.5 mg, 100%). ¹H NMR (600 MHz, CDCl₃/CD₃OD 2:1 v/v, 25 °C): δ_{H} 1.16 (t, ³*J*_{H,H} = 7.0 Hz, 6H, OCH₂CH₃), 2.53 (s, 6H, Ph-CH₃), 3.98–4.05 (m, 2H, OCH₂CH₃), 4.24–4.31 (m, 2H, OCH₂CH₃), 7.39 (d, ³*J*_{H,H} = 7.6 Hz, 4H, *m*-Ph), 7.83 (d, ³*J*_{H,H} = 7.7 Hz, 4H, *o*-Ph), 8.70 (d, ³*J*_{H,H} = 4.7 Hz, 2H, H β), 8.78 (d, ³*J*_{H,H} = 5.1 Hz, 2H, H β), 9.02 (d, ³*J*_{H,H} = 4.6 Hz, 2H, H β), 10.02 (s, 1H, H_{meso}), 10.08 (d, ³*J*_{H,H} = 5.1 Hz, 2H, H β). ³¹P{¹H} NMR (600 MHz, CDCl₃/MeOD 2:1 v/v, 25 °C): δ_{P} 23.17. MS (MALDI-TOF): *m/z* = 730.17 ([M]⁺, calcd. for C₃₈H₃₃N₄O₃PPd *m/z* = 730.13). HRMS (ESI): *m/z* = 730.13491 ([M]⁺, calcd. for C₃₈H₃₃N₄O₃PPd *m/z* = 730.13196). UV–vis [toluene; λ_{max} , nm (log ϵ): 410 (5.32), 523 (4.28), 557 (4.37). IR (neat, cm^{-1}): ν_{max} 2976 (w), 2918 (w), 1608 (w), 1539 (w), 1468 (w), 1433 (w), 1391 (w), 1369 (w), 1329 (w), 1295 (w), 1249 (m), 1214 (w), 1179 (w), 1160 (w), 1089 (w), 1067 (m), 1043 (w), 1010 (s), 969 (s), 959 (s), 889 (m), 873 (m), 849 (m), 790 (s), 756 (m), 734 (m), 712 (m), 696 (m), 593 (s), 569 (s).

{10-(Diethoxyphosphoryl)-5,15-bis(*p*-

(methoxycarbonyl)phenyl]porphyrinato}palladium(II) [PdD(CMP)PP] was prepared from 70 mg (0.098 mmol) of **H₂D(CMP)PP** and 88 mg (0.392 mmol) of Pd(OAc)₂ dissolved in 25 mL of a CHCl₃/CH₃CN (80:20 v/v) mixture. The reaction mixture was refluxed for 15 min. The resulting solid was purified by column chromatography on silica gel using a CHCl₃/MeOH (100:1 v/v) mixture as eluent to give **PdD(CMP)PP** as a pink purple crystalline powder (70.4 mg, 91%). ¹H NMR (600 MHz, CDCl₃/CD₃OD 2:1 v/v, 25 °C): δ_{H} 1.17 (t, ³*J*_{H,H} = 7.0 Hz, 6H, OCH₂CH₃), 3.93 (t, 6H, PhCOOCH₃), 4.01–4.05 (m, 2H, OCH₂CH₃), 4.27–4.31 (m, 2H, OCH₂CH₃), 8.03 (d, ³*J*_{H,H} = 8.0 Hz, 4H, *o*-Ph), 8.25 (d, ³*J*_{H,H} = 8.0 Hz, 4H, *m*-Ph), 8.58 (d, ³*J*_{H,H} = 4.7 Hz, 2H, H β), 8.68 (d, ³*J*_{H,H} = 5.2 Hz, 2H, H β), 9.01 (d, ³*J*_{H,H} = 4.7 Hz, 2H, H β), 10.01 (s, 1H, H_{meso}), 10.12 (d, ³*J*_{H,H} = 5.2 Hz, 2H, H β). ³¹P{¹H} NMR (600 MHz, CDCl₃/CD₃OD 2:1

v/v, 25 °C): δ_P 22.55. MS (MALDI-TOF): $m/z = 818.28$ ($[M]^+$, calcd. for $C_{40}H_{33}N_4O_7PPd$ $m/z = 818.11$). HRMS (ESI): $m/z = 818.11239$ ($[M]^+$, calcd. for $C_{40}H_{33}N_4O_7PPd$ $m/z = 818.11162$), 841.09803 ($[M+Na]^+$, calcd. for $C_{40}H_{33}N_4O_7PPdNa$ $m/z = 841.10139$). UV-vis [toluene; λ_{max} , nm (log ϵ): 410 (5.31), 523 (4.25), 557 (4.31). IR (neat, cm^{-1}): ν_{max} 2984 (w), 2917 (w), 2850 (w), 1719 (s, C=O), 1607 (m), 1541 (w), 1433 (m), 1401 (w), 1391 (w), 1308 (w), 1274 (s), 1253 (s), 1217 (w), 1179 (w), 1162 (w), 1109 (m), 1097 (m), 1072 (w), 1044 (w), 1011 (s), 965 (s), 893 (m), 864 (m), 822 (w), 785 (m), 761 (s), 729 (s), 710 (s), 694 (m), 585 (m), 565 (m), 551 (m).

(5,15-Dimesitylporphyrinato)palladium(II) (PdDMesP) was prepared from 15 mg (0.027 mmol) of **H₂DMesP** and 98 mg (0.437 mmol) of Pd(OAc)₂ in 8 mL of a CHCl₃/CH₃CN (50:50 v/v) mixture. The reaction mixture was refluxed for 96 h. The resulting solid was purified by silica gel column chromatography and gel permeation chromatography using CH₂Cl₂ and CHCl₃ as eluents, respectively, to give **PdDMesP** as an orange crystalline powder (6 mg, 34%). ¹H NMR (600 MHz, CDCl₃, 25 °C): δ_H 1.81 (s, 12H, *o*-CH₃), 2.65 (s, 6H, *p*-CH₃), 7.31 (br s, 4H, *m*-Ph), 8.84 (d, ³*J*_{H,H} = 4.6 Hz, 4H, H β), 9.24 (d, ³*J*_{H,H} = 4.6 Hz, 4H, H β), 10.20 (s, 2H, H_{meso}). ¹³C {¹H} NMR (150 MHz, CDCl₃, 25 °C): δ_C 21.50 (2C, *p*-CH₃), 21.55 (4C, *o*-CH₃), 106.49 (2C, *meso*-C), 118.91 (2C, *meso*-C), 127.84 (4C, *m*-C_{Mes}), 130.27 (4C, β -C), 131.42 (4C, β -C), 137.78 (2C, C_{Mes}), 137.85 (2C, C_{Mes}), 139.30 (4C, *o*-C_{Mes}), 141.04 (4C, α -C), 141.09 (4C, α -C). HRMS (ESI): $m/z = 650.16779$ ($[M]^+$, calcd. for $C_{38}H_{32}N_4Pd$ $m/z = 650.16563$). UV-vis [toluene; λ_{max} , nm (log ϵ): 405 (5.33), 514 (4.31), 546 (3.95). IR (neat, cm^{-1}): ν_{max} 2953 (m), 2920 (s), 2852 (m), 1717 (br, w), 1609 (w), 1571 (w), 1536 (w), 1456 (br, m), 1396 (w), 1375 (m), 1338 (w), 1320 (w), 1301 (w), 1260 (w), 1242 (w), 1183 (w), 1165 (w), 1144 (w), 1060 (m), 1036 (w), 1011 (s), 970 (w), 912 (w), 853 (m), 835 (m), 787 (m), 729 (m), 721 (m), 698 (m), 568 (w).

(5,15,20-Trimesitylporphyrinato)palladium(II) (PdTMesP) was prepared from 19 mg (0.029 mmol) of **H₂TMesP** and 104 mg (0.463 mmol) of Pd(OAc)₂ in 8 mL of a CHCl₃/CH₃CN (50:50 v/v) mixture. The reaction mixture was refluxed for 96 h. The resulting solid was purified by silica gel column chromatography and gel permeation chromatography using CH₂Cl₂ and CHCl₃ as eluent, respectively, to give **PdTMesP** as an orange crystalline powder (7 mg, 32%). ¹H NMR (600 MHz, CDCl₃, 25 °C): δ_H 1.84 (s, 12H, *o*-CH₃), 1.85 (s, 6H, *o*-CH₃), 2.63 (s, 3H, *p*-CH₃), 2.65 (s, 6H, *p*-CH₃), 7.27 (br s, 2H, *m*-Ph), 7.30 (br s, 4H, *m*-Ph), 8.66 (d, ³*J*_{H,H} = 4.8 Hz 2H, H β), 8.67 (d, ³*J*_{H,H} = 4.8 Hz, 2H, H β), 8.79 (d, ³*J*_{H,H} = 4.6 Hz, 2H, H β), 9.22 (d, ³*J*_{H,H} = 4.6 Hz, 2H, H β), 10.12 (s,

¹H, H_{meso}). ¹³C{¹H} NMR (150 MHz, CDCl₃, 25 °C): δ_c 21.49 (1C, *p*-CH₃), 21.50 (2C, *o*-CH₃), 21.61 (4C, *o*-CH₃), 21.72 (2C, *p*-CH₃), 106.04 (1C, *meso*-C), 119.06 (2C, *meso*-C), 119.71 (1C, *meso*-C), 127.75 (2C, *m*-C_{Mes}), 127.80 (4C, *m*-C_{Mes}), 130.06 (2C, β-C), 130.22 (4C, β-C), 131.25 (2C, β-C), 137.70 (1C, C_{Mes}), 137.75 (2C, C_{Mes}), 137.93 (2C, C_{Mes}), 138.06 (1C, C_{Mes}), 139.32 (6C, *o*-C_{Mes}), 140.87 (2C, α-C), 141.00 (2C, α-C), 141.14 (2C, α-C), 141.29 (2C, α-C). HRMS (ESI): *m/z* = 768.24534 ([M]⁺, calcd. for C₄₇H₄₂N₄Pd *m/z* = 768.24388). UV–vis [toluene; λ_{max}, nm (log ε)]: 412 (5.40), 519 (4.37), 550 (3.68). IR (neat, cm⁻¹): ν_{max} 2914 (w), 2851 (w), 1726 (w), 1609 (w), 1570 (w), 1538 (w), 1447 (br, m), 1388 (w), 1373 (w), 1341 (w), 1332 (w), 1305 (w), 1242 (w), 1214 (w), 1184 (w), 1166 (w), 1152 (w), 1065 (m), 1034 (w), 1010 (s), 952 (w), 902 (w), 864 (w), 843 (m), 834 (m), 796 (m), 776 (m), 755 (m), 718 (m), 692 (m), 557 (w).

[10-(diethoxyphosphoryl)-5,15-dimesitylporphyrinato]palladium(II)

(PdDMesPP) was prepared from 25 mg (0.037 mmol) of **H₂DMesPP** and 66 mg (0.295 mmol) of Pd(OAc)₂ in 9 mL of a CHCl₃/CH₃CN (50:50 v/v) mixture. The reaction mixture was refluxed for 2 h. The resulting solid was purified by silica gel column chromatography using a CH₂Cl₂/MeOH (95:5 v/v) mixture as eluent to give **PdDMesPP** as a pink crystalline powder (27 mg, 97%). ¹H NMR (600 MHz, CDCl₃, 25 °C): δ_H 1.38 (t, ³J_{H,H} = 7.0 Hz, 6H, OCH₂CH₃), 1.83 (s, 12H, *o*-CH₃), 2.66 (s, 6H, *p*-CH₃), 4.21–4.29 (m, 2H, OCH₂CH₃), 4.51–4.59 (m, 2H, OCH₂CH₃), 7.32 (br s, 4H, *m*-Ph), 8.75 (d, ³J_{H,H} = 4.6 Hz, 2H, Hβ), 8.83 (d, ³J_{H,H} = 5.1 Hz, 2H, Hβ), 9.18 (d, ³J_{H,H} = 4.6 Hz, 2H, Hβ), 10.16 (s, 1H, H_{meso}), 10.38 (d, ³J_{H,H} = 5.1 Hz, 2H, Hβ). ³¹P{¹H} NMR (600 MHz, CDCl₃, 25 °C): δ_P 22.59. ¹³C{¹H} NMR (150 MHz, CDCl₃, 25 °C): δ_c 16.48 (d, ²J_{C,P} = 7.2 Hz, 2C, OCH₂CH₃), 21.49 (2C, *p*-CH₃), 21.55 (4C, *o*-CH₃), 62.77 (d, ²J_{C,P} = 4.4 Hz, 2C, OCH₂CH₃), 102.03 (d, ²J_{C,P} = 185.6 Hz, 1C, *meso*-C), 109.34 (1C, *meso*-C), 120.27 (2C, *meso*-C), 127.90 (4C, *m*-C_{Mes}), 129.99 (2C, β-C), 131.48 (2C, β-C), 132.21 (2C, β-C), 133.11 (2C, β-C), 137.61 (2C, C_{Mes}), 138.01 (2C, C_{Mes}), 139.13 (4C, *o*-C_{Mes}), 140.49 (2C, α-C), 140.74 (2C, α-C), 142.18 (2C, α-C), 144.21 (d, ²J_{C,P} = 18.3 Hz, 2C, α-CP). HRMS (ESI): *m/z* = 787.19982 ([M+H]⁺, calcd. for C₄₂H₄₂N₄O₃PPd *m/z* = 787.20239), 809.18205 ([M+Na]⁺, calcd. for C₄₂H₄₁N₄O₃PPdNa *m/z* = 809.18433). UV–vis [toluene; λ_{max}, nm (log ε)]: 408 (5.34), 522 (4.23), 555 (4.33). IR (neat, cm⁻¹): ν_{max} 2919 (w), 2851 (w), 1730 (w), 1608 (w), 1561 (w), 1539 (w), 1476 (w), 1436 (br, m), 1388 (w), 1374 (w), 1327 (w), 1295 (w), 1250 (m), 1217 (w), 1165 (w), 1086 (w), 1063 (m), 1043 (m), 1011 (s), 954 (br, m), 883 (m), 853 (m), 834 (w), 800 (m), 782 (m), 736 (m), 725 (m), 713 (m), 695 (m), 600 (m), 578 (w), 560 (m), 538 (m).

[10-(diethoxyphosphoryl)-5,15,20-trimesitylporphyrinato]palladium(II)

(PdTMesPP) was prepared from 24 mg (0.030 mmol) of **H₂TMesPP** and 54 mg (0.241 mmol) of Pd(OAc)₂ in 8 mL of a CHCl₃/CH₃CN (50:50 v/v) mixture. The reaction mixture was refluxed for 2 h. The resulting solid was purified by silica gel column chromatography using a CH₂Cl₂/MeOH (95:5 v/v) mixture as eluent to give **PdTMesPP** as a pink crystalline powder (26 mg, 96%). ¹H NMR (600 MHz, CDCl₃, 25 °C): δ_H 1.37 (t, ³J_{H,H} = 7.0 Hz, 6H, OCH₂CH₃), 1.85 (s, 12H, *o*-CH₃), 1.86 (s, 6H, *o*-CH₃), 2.61 (s, 3H, *p*-CH₃), 2.64 (s, 6H, *p*-CH₃), 4.19–4.27 (m, 2H, OCH₂CH₃), 4.48–4.57 (m, 2H, OCH₂CH₃), 7.26 (br s, 2H, *m*-Ph), 7.29 (br s, 4H, *m*-Ph), 8.56 (d, ³J_{H,H} = 4.8 Hz, 2H, Hβ), 8.61 (d, ³J_{H,H} = 4.8 Hz, 2H, Hβ), 8.76 (d, ³J_{H,H} = 5.2 Hz, 2H, Hβ), 10.31 (d, ³J_{H,H} = 5.2 Hz, 2H, Hβ). ³¹P{¹H} NMR (600 MHz, CDCl₃, 25 °C): δ_P 22.72. ¹³C{¹H} NMR (150 MHz, CDCl₃, 25 °C): δ_C 16.46 (d, ²J_{C,P} = 6.6 Hz, 2C, OCH₂CH₃), 21.46 (1C, *p*-CH₃), 21.48 (2C, *o*-CH₃), 21.58 (4C, *o*-CH₃), 21.63 (2C, *p*-CH₃), 62.71 (d, ²J_{C,P} = 4.4 Hz, 2C, OCH₂CH₃), 101.35 (d, ²J_{C,P} = 187.1 Hz, 1C, *meso*-C), 120.50 (2C, *meso*-C), 122.92 (1C, *meso*-C), 127.81 (2C, *m*-C_{Mes}), 127.85 (4C, *m*-C_{Mes}), 129.79 (2C, β-C), 131.00 (2C, β-C), 131.47 (2C, β-C), 132.88 (2C, β-C), 137.40 (1C, C_{Mes}), 137.72 (2C, C_{Mes}), 137.92 (2C, C_{Mes}), 137.94 (1C, C_{Mes}), 139.04 (2C, *o*-C_{Mes}), 139.11 (4C, *o*-C_{Mes}), 140.44 (2C, α-C), 140.60 (2C, α-C), 142.18 (2C, α-C), 144.48 (d, ²J_{C,P} = 18.2 Hz, 2C, α-CP). HRMS (ESI): *m/z* = 904.27887 ([M]⁺, calcd. for C₅₁H₅₁N₄O₃PPd *m/z* = 904.27281), 905.28173 ([M+H]⁺, calcd. for C₅₁H₅₂N₄O₃PPd *m/z* = 905.28064), 927.26365 ([M+Na]⁺, calcd. for C₅₁H₅₁N₄O₃PPdNa *m/z* = 927.26258). UV–vis [toluene; λ_{max}, nm (log ε)]: 415 (5.38), 527 (4.29), 560 (4.28). IR (neat, cm⁻¹): ν_{max} 2918 (w), 2852 (w), 1729 (w), 1610 (w), 1561 (w), 1540 (w), 1435 (br, m), 1376 (w), 1348 (w), 1333 (w), 1302 (w), 1252 (m), 1207 (w), 1161 (w), 1088 (w), 1069 (m), 1044 (m), 1011 (s), 953 (br, m), 884 (m), 865 (w), 851 (m), 830 (w), 799 (m), 755 (w), 727 (m), 711 (m), 598 (m), 575 (w), 561 (m), 538 (m).

General Procedure for Platinum(II) Metalation. A solution of the free-base porphyrin **H₂R¹P–H₂R⁴P** or **H₂R¹PP–H₂R⁴PP** and PtCl₂ (2–10 equiv.) were refluxed in benzonitrile. During the heating, the color of the reaction mixture became brown-orange. The degree of conversion was monitored by MALDI-TOF mass-spectrometry and UV–vis spectroscopy. After complete conversion of the starting porphyrin, the reaction mixture was cooled to room temperature and *n*-hexane was added. The resulting precipitate was filtrated, washed two times with *n*-hexane and dissolved in chloroform.

After evaporation of the solvent, the residue was purified by column chromatography on silica gel to afford **PtR¹P–PtR⁴P** or **PtR¹PP–PtR⁴PP**.

[5,15-Bis(*p*-tolyl)porphyrinato]platinum(II) (PtDTolP) was prepared from 20 mg (0.041 mmol) of **H₂DTolP** and 33 mg (0.123 mmol) of PtCl₂ dissolved in 7 mL of benzonitrile. The reaction mixture was stirred under reflux for 4 h. The resulting solid was purified by column chromatography on silica gel using CH₂Cl₂ as eluent to give **PtDTolP** as an orange crystalline powder (20.6 mg, 72%). ¹H NMR (600 MHz, CDCl₃, 25 °C): δ_H 2.73 (s, 6H, Ph-CH₃), 7.58 (d, ³J_{H,H} = 7.6 Hz, 4H, *m*-Ph), 8.08 (d, ³J_{H,H} = 7.8 Hz, 4H, *o*-Ph), 8.97 (d, ³J_{H,H} = 4.8 Hz, 4H, Hβ), 9.21 (d, ³J_{H,H} = 4.9 Hz, 4H, Hβ), 10.15 (s, 2H, H_{meso}). MS (MALDI-TOF): *m/z* = 684.18 ([M+H]⁺, calcd. for C₃₄H₂₄N₄Pt *m/z* = 684.17). HRMS (ESI): *m/z* = 683.16445 ([M]⁺, calcd. for C₃₄H₂₄N₄Pt *m/z* = 683.16460). UV–vis [toluene; λ_{max}, nm (log ε)]: 393 (5.16), 502 (4.38), 531 (4.25). IR (neat, cm⁻¹): ν_{max} 2954 (w), 2916 (w), 2850 (w), 1729 (w), 1541 (w), 1509 (w), 1455 (w), 1395 (w), 1341 (w), 1326 (w), 1308 (m), 1179 (w), 1103 (w), 1069 (m), 1042 (w), 1012 (s), 943 (w), 897 (w), 854 (s), 799 (m), 778 (s), 721 (s), 693 (s), 625 (w), 615 (w), 590 (w), 568 (w), 554 (w).

{5,15-bis[*p*-(methoxycarbonyl)phenyl]porphyrinato}platinum(II) [PtD(CMP)P] was prepared from 40 mg (0.07 mmol) of **H₂D(CMP)P** and 56 mg (0.208 mmol) of PtCl₂ dissolved in 12 mL of benzonitrile. The reaction mixture was stirred under reflux for 4 h. The resulting solid was purified by column chromatography on silica gel using CH₂Cl₂ as eluent to give **PtD(CMP)P** as an orange crystalline powder (25 mg, 46%). ¹H NMR (600 MHz, CDCl₃, 25 °C): δ_H 4.14 (s, 6H, Ph-CH₃), 8.30 (d, ³J_{H,H} = 7.9 Hz, 4H, *o*-Ph), 8.47 (d, ³J_{H,H} = 8.0 Hz, 4H, *m*-Ph), 8.90 (d, ³J_{H,H} = 4.8 Hz, 4H, Hβ), 9.26 (d, ³J_{H,H} = 4.8 Hz, 4H, Hβ), 10.21 (s, 2H, H_{meso}). MS (MALDI-TOF): *m/z* = 771.15 ([M]⁺, calcd. for C₃₆H₂₄N₄O₄Pt *m/z* = 771.14). HRMS (ESI): *m/z* = 771.14461 ([M]⁺, calcd. for C₃₆H₂₄N₄O₄Pt *m/z* = 771.14428). UV–vis [CHCl₃; λ_{max}, nm (log ε)]: 393 (5.31), 502 (4.25), 531 (4.14). IR (neat, cm⁻¹): ν_{max} 2953 (m), 2915 (s), 2849 (m), 1716 (s, C=O), 1604 (w), 1541 (w), 1492 (w), 1457 (w), 1428 (w), 1398 (w), 1376 (w), 1363 (w), 1310 (w), 1222 (w), 1195 (w), 1179 (m), 1096 (m), 1075 (m), 1047 (w), 1013 (m), 968 (w), 928 (w), 855 (w), 848 (w), 821 (w), 782 (m), 755 (m), 723 (s), 694 (m), 646 (w), 615 (w), 594 (w), 569 (w), 554 (w).

[10-(Diethoxyphosphoryl)-5,15-bis(*p*-tolyl)porphyrinato]platinum(II) (PtDTolPP) was prepared from 7 mg (0.011 mmol) of **H₂DTolPP** and 7.5 mg (0.022 mmol) of PtCl₂ dissolved in 2 mL of benzonitrile. The reaction mixture was stirred under

reflux for 4 h. The resulting solid was purified by column chromatography on silica gel using CHCl_3/n -hexane (1:1 v/v) as eluent to give **PtDTolPP** as an orange red crystalline powder (7.8 mg, 85%). ^1H NMR (600 MHz, $\text{CDCl}_3/\text{CD}_3\text{OD}$ 2:1 v/v, 25 °C): δ_{H} 1.16 (t, $^3J_{\text{H,H}} = 7.0$ Hz, 6H, OCH_2CH_3), 2.53 (s, 6H, Ph- CH_3), 3.98–4.05 (m, 2H, OCH_2CH_3), 4.24–4.30 (m, 2H, OCH_2CH_3), 7.39 (d, $^3J_{\text{H,H}} = 7.6$ Hz, 4H, *m*-Ph), 7.83 (d, $^3J_{\text{H,H}} = 7.7$ Hz, 4H, *o*-Ph), 8.68 (d, $^3J_{\text{H,H}} = 4.8$ Hz, 2H, H β), 8.74 (d, $^3J_{\text{H,H}} = 5.2$ Hz, 2H, H β), 8.99 (d, $^3J_{\text{H,H}} = 4.8$ Hz, 2H, H β), 9.96 (s, 1H, H_{meso}), 10.06 (d, $^3J_{\text{H,H}} = 5.3$ Hz, 2H, H β). $^{31}\text{P}\{^1\text{H}\}$ NMR (600 MHz, $\text{CDCl}_3/\text{CD}_3\text{OD}$ 2:1 v/v, 25 °C): δ_{P} 22.52. MS (MALDI-TOF): $m/z = 820.23$ ($[\text{M}+\text{H}]^+$, calcd. for $\text{C}_{38}\text{H}_{34}\text{N}_4\text{O}_3\text{PPt}$ $m/z = 820.19$). HRMS (ESI): $m/z = 819.19732$ ($[\text{M}]^+$, calcd. for $\text{C}_{38}\text{H}_{33}\text{N}_4\text{O}_3\text{PPt}$ $m/z = 819.19357$). UV–vis [toluene; λ_{max} , nm (log ϵ): 396 (5.41), 510 (4.23), 546 (4.42). IR (neat, cm^{-1}): ν_{max} 2952 (m), 2920 (m), 1609 (w), 1541 (w), 1439 (w), 1391 (w), 1375 (w), 1363 (w), 1333 (w), 1311 (w), 1246 (m, P=O), 1216 (w), 1179 (w), 1162 (w), 1088 (w), 1069 (m), 1042 (w), 1015 (s), 966 (s), 963 (m), 892 (m), 873 (m), 850 (w), 800 (s), 792 (m), 781 (m), 710 (m), 694 (m), 664 (w), 640 (w), 603 (s), 571 (s), 552 (w).

{10-(Diethoxyphosphoryl)-5,15-bis[*p*-(methoxycarbonyl)phenyl]porphyrinato}platinum(II) [PtD(CMP)PP] was prepared from 15 mg (0.021 mmol) of **H₂D(CMP)PP** and 11.2 mg (0.042) of PtCl_2 dissolved in 4 mL of benzonitrile. The reaction mixture was stirred under reflux for 4 h. The resulting solid was purified by column chromatography on silica gel using CHCl_3/n -hexane (1:1 v/v) as eluent to give **PtD(CMP)PP** as a red orange crystalline powder (12.3 mg, 70%). ^1H NMR (600 MHz, $\text{CDCl}_3/\text{CD}_3\text{OD}$ (2:1 v/v), 25 °C): δ_{H} 1.15 (t, $^3J_{\text{H,H}} = 7.0$ Hz, 6H, OCH_2CH_3), 3.92 (s, 6H, PhCOO CH_3), 3.99–4.06 (m, 2H, OCH_2CH_3), 4.24–4.30 (m, 2H, OCH_2CH_3), 8.01 (d, $^3J_{\text{H,H}} = 7.9$ Hz, 4H, *o*-Ph), 8.23 (d, $^3J_{\text{H,H}} = 7.9$ Hz, 4H, *m*-Ph), 8.53 (d, $^3J_{\text{H,H}} = 4.8$ Hz, 2H, H β), 8.62 (d, $^3J_{\text{H,H}} = 5.3$ Hz, 2H, H β), 8.95 (d, $^3J_{\text{H,H}} = 4.8$ Hz, 2H, H β), 9.91 (s, 1H, H_{meso}), 10.08 (d, $^3J_{\text{H,H}} = 5.3$ Hz, 2H, H β). $^{31}\text{P}\{^1\text{H}\}$ NMR (600 MHz, $\text{CDCl}_3/\text{CD}_3\text{OD}$ 2:1 v/v, 25 °C): δ_{P} 21.91. MS (MALDI-TOF): $m/z = 908.04$ ($[\text{M}+\text{H}]^+$, calcd. for $\text{C}_{40}\text{H}_{34}\text{N}_4\text{O}_7\text{PPt}$ $m/z = 908.17$). HRMS (ESI): $m/z = 907.17703$ ($[\text{M}]^+$, calcd. for $\text{C}_{40}\text{H}_{33}\text{N}_4\text{O}_7\text{PPt}$ $m/z = 907.17324$). UV–vis [toluene; λ_{max} , nm (log ϵ): 396 (5.51), 510 (4.44), 546 (4.61). IR (neat, cm^{-1}): ν_{max} 2980 (w), 2921 (w), 2849 (w), 1717 (s, C=O), 1607 (m), 1566 (w), 1545 (w), 1433 (m), 1392 (w), 1307 (w), 1274 (s, P=O), 1252 (s, P=O), 1217 (w), 1178 (w), 1162 (w), 1110 (m), 1089 (m), 1074 (w), 1043 (w), 1013 (s), 960 (s), 893 (m), 865 (m), 822 (w), 786 (m), 761 (s), 728 (s), 709 (s), 694 (m), 586 (s), 561 (s), 551 (m).

(5,15-Dimesitylporphyrinato)platinum(II) (PtDMesP) was prepared from 21 mg (0.038 mmol) of **H₂DMesP** and 102 mg (0.384 mmol) of PtCl₂ in 6 mL of benzonitrile. The reaction mixture was stirred under reflux for 18 h. The resulting solid was purified by silica gel column chromatography and gel permeation chromatography using CHCl₃ as eluent to give **PtDMesP** as an orange crystalline powder (22 mg, 78%). ¹H NMR (600 MHz, CDCl₃, 25 °C): δ_H 1.86 (s, 12H, *o*-CH₃), 2.66 (s, 6H, *p*-CH₃), 7.32 (br s, 4H, *m*-Ph), 8.80 (d, ³J_{H,H} = 4.8 Hz, 4H, Hβ), 9.18 (d, ³J_{H,H} = 4.8 Hz, 4H, Hβ), 10.11 (s, 2H, H_{meso}). ¹³C{¹H} NMR (150 MHz, CDCl₃, 25 °C): δ_c 21.49 (4C, *o*-CH₃), 21.50 (2C, *p*-CH₃), 107.24 (2C, *meso*-C), 119.48 (2C, *meso*-C), 127.90 (4C, *m*-C_{Mes}), 129.96 (4C, β-C), 131.35 (4C, β-C), 137.37 (2C, C_{Mes}), 137.91 (2C, C_{Mes}), 139.23 (4C, *o*-C_{Mes}), 140.34 (4C, α-C), 140.42 (4C, α-C). HRMS (ESI): *m/z* = 739.22774 ([M]⁺, calcd. for C₃₈H₃₂N₄Pt *m/z* = 739.22723), 762.21649 ([M+Na]⁺, calcd. for C₃₈H₃₂N₄PtNa *m/z* = 762.21669). UV-vis [toluene; λ_{max}, nm (log ε)]: 391 (5.38), 501 (4.27), 532 (4.19). IR (neat, cm⁻¹): ν_{max} 2954 (m), 2921 (m), 2852 (m), 1828 (w), 1786 (w), 1736 (w), 1711 (w), 1606 (w), 1589 (w), 1570 (w), 1521 (m), 1447 (br, m), 1398 (w), 1371 (m), 1341 (w), 1324 (w), 1306 (m), 1260 (w), 1242 (w), 1217 (w), 1164 (w), 1144 (w), 1070 (m), 1059 (m), 1041 (w), 1013 (s), 915 (w), 853 (s), 836 (m), 789 (m), 778 (s), 758 (m), 720 (s), 698 (s), 672 (w), 646 (w), 567 (w).

(5,15,20-Trimesitylporphyrinato)platinum(II) (PtTMesP) was prepared from 10 mg (0.015 mmol) of **H₂TMesP** and 40 mg (0.150 mmol) of PtCl₂ in 3 mL of benzonitrile. The reaction mixture was stirred under reflux for 50 h. The resulting solid was purified by silica gel column chromatography and gel permeation chromatography using CHCl₃ as eluent to give **PtTMesP** as an orange-brown crystalline powder (11 mg, 85%). ¹H NMR (600 MHz, CDCl₃, 25 °C): δ_H 1.87 (s, 18H, *o*-CH₃), 2.62 (s, 3H, *p*-CH₃), 2.64 (s, 6H, *p*-CH₃), 7.27 (br s, 2H, *m*-Ph), 7.30 (br s, 4H, *m*-Ph), 8.61 (d, ³J_{H,H} = 4.8 Hz, 2H, Hβ), 8.62 (d, ³J_{H,H} = 4.8 Hz, 2H, Hβ), 8.75 (d, ³J_{H,H} = 4.8 Hz, 2H, Hβ), 9.14 (d, ³J_{H,H} = 4.8 Hz, 2H, Hβ), 10.03 (s, 1H, H_{meso}). ¹³C{¹H} NMR (150 MHz, CDCl₃, 25 °C): δ_c 21.48 (1C, *p*-CH₃), 21.49 (2C, *o*-CH₃), 21.53 (4C, *o*-CH₃), 21.65 (2C, *p*-CH₃), 106.75 (1C, *meso*-C), 119.68 (2C, *meso*-C), 120.37 (1C, *meso*-C), 127.81 (2C, *m*-C_{Mes}), 127.86 (4C, *m*-C_{Mes}), 129.78 (2C, β-C), 129.93 (2C, β-C), 129.99 (2C, β-C), 131.14 (2C, β-C), 137.51 (2C, C_{Mes}), 137.62 (1C, C_{Mes}), 137.77 (1C, C_{Mes}), 137.82 (2C, C_{Mes}), 139.26 (6C, *o*-C_{Mes}), 140.23 (2C, α-C), 140.28 (2C, α-C), 140.50 (2C, α-C), 140.55 (2C, α-C). HRMS (ESI): *m/z* = 857.30273 ([M]⁺, calcd. for C₄₇H₄₂N₄Pt *m/z* = 857.30555). UV-vis [toluene; λ_{max}, nm (log ε)]: 397 (5.26), 506 (4.25), 536 (3.97). IR (neat, cm⁻¹): ν_{max} 2916 (w), 2852

(w), 1727 (w), 1698 (w), 1610 (w), 1571 (w), 1542 (w), 1452 (br, m), 1390 (w), 1375 (w), 1348 (w), 1337 (w), 1311 (m), 1269 (w), 1243 (w), 1224 (w), 1215 (w), 1187 (w), 1168 (w), 1067 (m), 1015 (s), 953 (w), 908 (w), 865 (w), 845 (m), 836 (s), 795 (s), 776 (m), 718 (s), 693 (m), 672 (m), 648 (w), 595 (w).

[10-(diethoxyphosphoryl)-5,15-dimesitylporphyrinato]platinum(II)

(PtDMesPP) was prepared from 25 mg (0.037 mmol) of **H₂DMesPP** and 97 mg (0.366 mmol) of PtCl₂ in 6 mL of benzonitrile. The reaction mixture was stirred under reflux for 8 h. The resulting solid was purified by silica gel column chromatography and gel permeation chromatography using a CH₂Cl₂/MeOH (95:5 v/v) mixture and CHCl₃ as eluent, respectively, to give **PtDMesPP** as a pink crystalline powder (6 mg, 18%). ¹H NMR (600 MHz, CDCl₃, 25 °C): δ_H 1.36 (t, ³J_{H,H} = 7.0 Hz, 6H, OCH₂CH₃), 1.85 (s, 12H, *o*-CH₃), 2.65 (s, 6H, *p*-CH₃), 4.20–4.28 (m, 2H, OCH₂CH₃), 4.49–4.57 (m, 2H, OCH₂CH₃), 7.31 (br s, 4H, *m*-Ph), 8.71 (d, ³J_{H,H} = 4.7 Hz, 2H, H_β), 8.77 (d, ³J_{H,H} = 5.3 Hz, 2H, H_β), 9.12 (d, ³J_{H,H} = 4.7 Hz, 2H, H_β), 10.06 (s, 1H, H_{meso}), 10.34 (d, ³J_{H,H} = 5.1 Hz, 2H, H_β). ³¹P{¹H} NMR (600 MHz, CDCl₃, 25 °C): δ_P 21.98. ¹³C{¹H} NMR (150 MHz, CDCl₃, 25 °C): δ_C 16.46 (d, ²J_{C,P} = 7.2 Hz, 2C, OCH₂CH₃), 21.49 (6C, *p+o*-CH₃), 62.83 (d, ²J_{C,P} = 4.5 Hz, 2C, OCH₂CH₃), 103.19 (d, ²J_{C,P} = 185.7 Hz, 1C, *meso*-C), 109.93 (1C, *meso*-C), 120.71 (2C, *meso*-C), 127.95 (4C, *m*-C_{Mes}), 129.57 (2C, β-C), 131.25 (2C, β-C), 132.23 (2C, β-C), 132.85 (2C, β-C), 137.22 (2C, C_{Mes}), 138.07 (2C, C_{Mes}), 139.07 (4C, *o*-C_{Mes}), 139.68 (2C, α-C), 140.20 (2C, α-C), 141.71 (2C, α-C), 143.21 (d, ²J_{C,P} = 17.8 Hz, 2C, α-CP). HRMS (ESI): *m/z* = 898.24236 ([M+Na]⁺, calcd. for C₄₂H₄₁N₄O₃PPtNa *m/z* = 898.24597). UV–vis [toluene; λ_{max}, nm (log ε)]: 394 (5.32), 510 (4.12), 545 (4.32). IR (neat, cm⁻¹): ν_{max} 2976 (w), 2919 (w), 2855 (w), 1726 (w), 1610 (w), 1563 (w), 1543 (w), 1454 (m), 1440 (m), 1390 (w), 1377 (w), 1361 (w), 1333 (w), 1301 (w), 1254 (m), 1218 (w), 1165 (w), 1087 (w), 1067 (m), 1044 (m), 1017 (s), 961 (br, m), 884 (m), 856 (m), 837 (w), 800 (m), 736 (w), 725 (w), 712 (w), 696 (w), 603 (m), 578 (w), 562 (m), 542 (w).

[10-(diethoxyphosphoryl)-5,15,20-trimesitylporphyrinato]platinum(II)

(PtTMesPP) was prepared from 10 mg (0.013 mmol) of **H₂TMesPP**, 33 mg (0.124 mmol) of PtCl₂ and 10 mg (0.122 mmol) NaOAc in 3 mL of benzonitrile according to the general procedure. The reaction mixture was stirred at 150 °C for 1 h. The resulting solid was purified by silica gel column chromatography and gel permeation chromatography using CH₂Cl₂/MeOH (95:5 v/v) mixture and CHCl₃ as eluent, respectively, to give **PtTMesPP** as pink crystalline powder (11 mg, 89%). ¹H NMR (600

MHz, CDCl₃, 25 °C): δ_{H} 1.35 (t, $^3J_{\text{H,H}} = 6.9$ Hz, 6H, OCH₂CH₃), 1.87 (s, 12H, *o*-CH₃), 1.88 (s, 6H, *o*-CH₃), 2.62 (s, 3H, *p*-CH₃), 2.64 (s, 6H, *p*-CH₃), 4.17–4.25 (m, 2H, OCH₂CH₃), 4.47–4.55 (m, 2H, OCH₂CH₃), 7.28 (br s, 6H, *m*-Ph), 8.53 (d, $^3J_{\text{H,H}} = 4.8$ Hz, 2H, H β), 8.56 (d, $^3J_{\text{H,H}} = 4.8$ Hz, 2H, H β), 8.71 (d, $^3J_{\text{H,H}} = 5.2$ Hz, 2H, H β), 10.28 (d, $^3J_{\text{H,H}} = 5.2$ Hz, 2H, H β). $^{31}\text{P}\{^1\text{H}\}$ NMR (600 MHz, CDCl₃, 25 °C): δ_{P} 22.09. $^{13}\text{C}\{^1\text{H}\}$ NMR (150 MHz, CDCl₃, 25 °C): δ_{C} 16.44 (d, $^2J_{\text{C,P}} = 6.6$ Hz, 2C, OCH₂CH₃), 21.46 (1C, *p*-CH₃), 21.47 (2C, *o*-CH₃), 21.51 (4C, *o*-CH₃), 21.56 (2C, *p*-CH₃), 62.75 (d, $^2J_{\text{C,P}} = 4.9$ Hz, 2C, OCH₂CH₃), 102.54 (d, $^2J_{\text{C,P}} = 185.6$ Hz, 1C, *meso*-C), 121.01 (2C, *meso*-C), 123.41 (1C, *meso*-C), 127.86 (2C, *m*-C_{Mes}), 127.90 (4C, *m*-C_{Mes}), 129.38 (2C, β -C), 130.86 (2C, β -C), 131.24 (2C, β -C), 132.59 (2C, β -C), 137.00 (1C, C_{Mes}), 137.33 (2C, C_{Mes}), 137.99 (3C, C_{Mes}), 139.03 (2C, *o*-C_{Mes}), 139.07 (4C, *o*-C_{Mes}), 139.67 (2C, α -C), 140.01 (2C, α -C), 141.74 (2C, α -C), 143.44 (d, $^2J_{\text{C,P}} = 18.5$ Hz, 2C, α -CP). HRMS (ESI): $m/z = 993.33999$ ([M]⁺, calcd. for C₅₁H₅₁N₄O₃PPt $m/z = 993.33448$), 1016.32463 ([M+Na]⁺, calcd. for C₅₁H₅₁N₄O₃PPtNa $m/z = 1016.32387$). UV–vis [toluene; λ_{max} , nm (log ϵ): 402 (5.13), 515 (4.07), 549 (3.97). IR (neat, cm⁻¹): ν_{max} 2956 (s), 2924 (s), 2857 (m), 1726 (s), 1610 (w), 1579 (w), 1542 (w), 1457 (br, m), 1377 (w), 1354 (w), 1338 (w), 1273 (br, m), 1209 (w), 1162 (w), 1122 (m), 1072 (m), 1041 (m), 1019 (s), 962 (br, m), 885 (w), 865 (w), 853 (w), 833 (w), 801 (m), 741 (m), 728 (w), 712 (m), 651 (w), 601 (m), 563 (m), 541 (w).

Photophysical measurements

Photophysical solution studies were performed for dye concentrations ranging between 3 and 7 μM . Luminescence spectra were recorded on a FluoroLog® 3 spectrofluorometer (Horiba Scientific) equipped with a NIR-sensitive R2658 photomultiplier from Hamamatsu (300–1050 nm). All dye solutions were deoxygenated in a screw-cap cuvette (Hellma) by bubbling high purity nitrogen (99.9999%) through the solution for at least 15 min. Absolute quantum yields at room temperature were measured with an integrating sphere from Horiba. The luminescence decay times were acquired in the time domain on the FluoroLog® 3 spectrofluorometer equipped with a DeltaHub module (Horiba Scientific) controlling a SpectraLED-392 ($\lambda = 392$ nm) and using DAS-6 Analysis software for data analysis. For characterization of the oxygen sensing properties, the composition of the gas was adjusted with a custom-build gas-mixing device based on mass-flow controllers from Voegtlin (www.red-y.com) by

mixing compressed air and dinitrogen. The luminescence spectra and phosphorescence decay times at 77 K were recorded for the dye solutions in a mixture of toluene and tetrahydrofuran (4:6 v/v), which produces good glasses at this temperature. The measurements were made in the Institute of Analytical Chemistry and Food Chemistry, Graz University of Technology.

Photostability studies

Photostability measurements were conducted in a 100 mL double-jacketed glass vessel equipped with a magnetic stirring bar and fitted to a Lauda RE106 water circulator ensuring a constant temperature of 25.0(5) °C. The cell was sealed with a 5-necked lid from Metrohm (reference 6.1414.010) fitted with a thermometer and an immersion probe having a 1 cm optical path length made of Suprasil 300 (Hellma, reference 6610202). The probe was connected to a Cary 50 (Varian) spectrophotometer through fiber guides and a fiber optic coupler. Prior to each experiment, the reference spectrum was recorded with the pure solvent and the absorbance set to zero at the wavelength used to monitor the time course of the reaction. Visible absorption spectra were recorded over the 350–800 nm range by taking a point at each nanometer at a scan rate of 400 nm/min. Kinetic traces were recorded at the maximum of the Soret band with an average time of 0.1 s by taking one point every minute until the end of the reaction (typically, 5000–7000 min). A spectrum was recorded systematically before and after irradiation. The light source was an Osram Powerstat HQI BT 400 W lamp (metal halogenide lamp with UV filter) emitting in the visible range (the spectrum of the lamp provided by the manufacturer is displayed in Figure S79, ESI) and was placed in a fixed position 20 cm away from the glass reactor, both devices being aligned with respect to their centers. The setup remained untouched during all experiments to ensure reproducibility, while the lamp was allowed to warm up and to stabilize for at least half an hour before starting the experiments. In most cases however, it remained switched on in between the irradiations. During filling/emptying operations and before exposing the porphyrin solution to the light once it was introduced in the cell, the latter was protected by an UV filtering Plexiglas screen covered with an aluminum foil.

All solutions of porphyrins were freshly prepared by carefully weighing a few milligrams of solid with a Mettler-Toledo XP205 Delta Range balance (resolution: 0.01 mg) into a 5 mL volumetric flask. Compounds were dissolved in analytical grade DMF

(Sigma-Aldrich) dried over molecular sieves or toluene (Sigma-Aldrich) dried over Al₂O₃ and the resulting solutions were stored in the dark. In few instances, short (less than 1 min) sonication was required. The reactor was filled with 50 mL of solvent using a glass pipet and allowed to reach the equilibrium temperature. After performing the zeroing operations of the spectrophotometer, an aliquot of the porphyrin mother solution was injected in the reactor with a 1000 μ L Eppendorf micropipette; the introduced volume was calculated so that the final concentration after dilution was 5 μ M unless otherwise noted. Immediately after, a spectrum was measured while protecting the cell with the screen. At the end of the data collection, the cell was exposed to the light source while starting the recording of the absorbance decay curve. Data manipulation and nonlinear least-squares fitting to a monoexponential decay function [$y = a \exp(-k_{obs}t) + bt + c$] were performed with Origin 8.0 (Figure S83-S85, ESI).

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was supported by the Russian Foundation for Basic Research (grant number 18-33-00734-mol-a), the Russian Academy of Sciences (RAS), and the Centre National de la Recherche Scientifique (CNRS). It was carried out in the framework of the International Associated French–Russian Laboratory of Macrocyclic Systems and Related Materials (LAMREM) of CNRS and RFBR (RFBR grant number 17-53-16028). Some measurements were performed using equipment of CKP FMI IPCE RAS. Prof. Anthony Romieu is warmly acknowledged for helpful discussions concerning this work. We are also grateful to Marie-José Penouilh and Quentin Bonnin for their technical assistance.

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