

Anticancer Activity of Polyoxometalate-Bisphosphonate Complexes: Synthesis, Characterization, In Vitro and In Vivo Results

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ABSTRACT

We synthesized a series of polyoxometalate-bisphosphonate complexes containing $Mo^{VI}O_6$ octahedra, zoledronate or an *N*-alkyl (*n*-C₆ or *n*-C₈) zoledronate analog, and in two cases, Mn as a heterometal. Mo_6L_2 (L = Zol, ZolC₆, ZolC₈) and Mo_4L_2Mn (L = Zol, ZolC₈) were

characterized by using single-crystal X-ray crystallography and/or IR spectroscopy, elemental and EDX analysis and ³¹P NMR. We found promising activity against human non-small cell lung cancer (NCI-H460) cells with IC₅₀ values for growth inhibition of ~5 μ M per bisphosphonate ligand. The effects of bisphosphonate complexation on IC₅₀ decreased with increasing bisphosphonate chain length: C₀ ~6.1x; C₆ ~3.4x and C₈ ~1.1x. We then determined the activity of one of the most potent compounds in the series, Mo₄Zol₂Mn(III), against SK-ES-1 sarcoma cells in a mouse xenograft system finding a ~5x decrease in tumor volume than found with the parent compound zoledronate at the same compound dosing (5 μ g/mouse). Overall, the results are of interest since we show for the first time that heteropolyoxomolybdate-bisphosphonate hybrids kill tumor cells *in vitro* and significantly decrease tumor growth, *in vivo*, opening up new possibilities for targeting both Ras as well as EGFR (epidermal growth factor receptor) driven cancers.

INTRODUCTION

Polyoxometalates (POMs) constitute a class of discrete, anionic, metal-oxygen clusters which can be considered as soluble oxide fragments. They are built from the connection of $\{MO_x\}$ polyhedra, M being a d-block transition metal ion in a high oxidation state, usually W^{VI} , $Mo^{V,VI}$ or $V^{IV,V.1}$ They can incorporate a large range of organic or inorganic species into their structures. This results in a great variety of sizes, nuclearities and shapes² and a vast range of photochemical ³, catalytic ^{4,5}, magnetic ^{6,7} as well as biological properties including antibacterial,⁸ antiviral and antitumoral activities.⁹ POMs can also have activity in tumor cell growth inhibition, e.g. $[NH_3Pr^i]_6[Mo_7O_{24}].3H_2O$ exhibits potent anti-tumor activity against breast, sarcoma, adenocarcinoma and pancreatic cancer cells.¹⁰ The anti-tumor activities *in vitro* of a variety of polyoxotungstates have also been reported and organometallic derivatives with RSn¹¹ or CpTi¹² (Cp = cyclopentadienyl) groups appear to be the most efficient. Recently, the cobalt derivative $\{CoSb_6O_4(H_2O)_3[Co(hmta)SbW_8O_{31}]_3\}^{15-}$ (hmta = hexamethylenetetramine) was also shown to kill various cancer cells.¹³ Polyoxometalates, in particular Dawson-type $[P_2Mo_{18}O_{62}]^{6-}$ anions, were also identified as inhibitors of protein kinase CK2, a multifunctional kinase that is deregulated in many cancers.¹⁴

In order to try and improve activity, one approach is to graft bioactive organic ligands on the inorganic POM core. However, examples of this approach are rare. Organoimidoderivatives of hexamolybdate based on amantadine, an antiviral and anti-Parkinson drug, have been tested *in vitro* against MCF-7 cells and exhibit better activity than do unfunctionalized hexamolybdate and amantadine. ¹⁵ Cholic acid was also grafted onto the *tris*-modified Anderson-type POM [MnMo₆O₁₈{(OCH₂)₃CNH₂}₂]³⁻, resulting in enhanced activity over the non-functionalized POM against breast cancer cells (MCF-7, IC₅₀~56 μ M and MDA-MB-231, IC₅₀~38 μ M).¹⁶ A variant of this approach consists of grafting a cleavable group onto a hexamolybdate POM in order to improve degradability with one such hybrid POM being reported to exhibit good activity against human malignant glioma cells (U251, IC50~25 μ M), the ability to penetrate the blood brain barrier, as well as low toxicity towards rat pheochromocytoma cells (PC12).¹⁷

In our group, we have been investigating polyoxometalate-bisphosphonate complexes. Bisphosphonates (BPs) having the general formula H₂O₃PC(OH)(R)PO₃H₂ have been used clinically for decades to treat bone resorption diseases and some of them exhibit anti-tumor activity. ¹⁸ Once deprotonated, BPs can form complexes with Mo, W or V ions, with nuclearities ranging from 1 to 12.¹⁹ In our first investigation, we reported the anti-cancer activity of hybrid POM/alendronate (Ale) complexes. ²⁰ The dodecanuclear complex $[(Mo^{VI}_{3}O_8)_4(Ale)_4]^{8-}$, noted here Mo₁₂Ale₄, had IC₅₀ values of ~10 µM against human breast cancer cells (MCF-7) *in vitro*. This value represents about 4x the activity of the parent alendronate molecule (i.e. on a per-alendronate basis), indicating a potentiating or synergistic effect. We then extended our study to include zoledronate (Zol; Figure 1), the most potent commercially available bisphosphonate, and synthesized the hexanuclear complex $[(Mo^{VI}_{3}O_8)_2O(Zol)_2]^{6-21}$ We also investigated the incorporation of a heterometal finding that the most active compound was the Mn(III)-containing POM $[(Mo^{VI}_{2}O_6)_2(Zol)_2Mn]^{5-}$, a.k.a. $Mo_4Zol_2Mn(III)$, suggesting a possible role for both the heterometal and the BP in growth inhibition.²²

Here, we describe the synthesis and characterization of a new series of POM/BP complexes and investigate the influence of the oxidation state of the Mn, and the introduction of alkyl chains grafted onto the zoledronate core, on the *in vitro* activity of these compounds against human non-small cell lung cancer (NCI-H460) cells. Lipophilic chains were expected to improve cell uptake of the hybrid POMs since it is known that lipophilic BPs have activities far greater than do non-lipophilic BPs in inhibiting tumor cell growth and invasiveness, both *in vitro* and *in vivo*.²³ Furthermore, a recent study revealed that lipophilic zoledronates inhibit both farnesyl and geranylgeranyldiphosphate synthases, effectively blocking prenylation of KRAS and other small G-proteins critical for tumor growth and cell survival.²⁴ Finally, we report the first results of *in vivo* xenograft tumor experiment with a POM/BP/Mn complex.

RESULTS AND DISCUSSION

In order to test the effect of the introduction of a lipophilic chain onto the BP-ligand on tumor cell growth inhibition, two zoledronate-based bisphosphonates, $ZolC_6$ and $ZolC_8$ (Figure 1), were selected for synthesis of Mo/BP complexes. $ZolC_6$ and $ZolC_8$ are zoledronate analogs that have *N*-alkyl chains (*n*-C₆ or *n*-C₈) on the unsubstituted nitrogen in the parent zoledronate. These ligands were chosen because they were previously found to be the most active (of Zol analogs containing between 0 and 15 carbon substituents) against a range of

tumor cell lines.²⁴ Two families of complexes were investigated: Mo_6L_2 (L = Zol, ZolC₆, ZolC₈), or with Mn(III) as the heterometal and with general formulae $Mo_4L_2Mn(III)$ (L = Zol, ZolC₈). The structures and ligand abbreviations as well as the structures of all of the complexes reported in this study are shown in Figure 1.

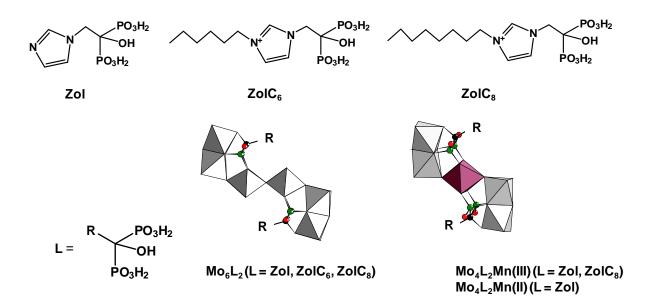


Figure 1. Abbreviations and formulae of the BPs used in this study together with schematic structures of some complexes. Grey polyhedra = MoO_6 , purple polyhedron = MnO_6 , green spheres = P, black spheres = C, red spheres = O.

Synthesis and Characterization.

 Mo_6Zol_2 was synthesized as a reference according to a previously reported procedure.²¹ Mo(VI) complexes with lipophilic bisphosphonates were synthesized by reaction of the BP precursor with Mo(VI) ions in aqueous solution at pH ~5. While it was possible to isolate single crystals suitable for X-ray diffraction by slow evaporation of the reaction mixture for $Mo_6(ZolC_6)_2$, only a poorly crystalline powder could be obtained for the $Mo_6(ZolC_8)_2$ complex. However, the similarity of the IR spectra (Figure S1a) and of the ³¹P NMR spectra (Figure S2) of both compounds, as well as the results of elemental analysis, confirm formation of $Mo_6(ZolC_8)_2$. $Mo_4Zol_2Mn(III)$ was synthesized following a slight modification

of the procedure reported previously.²² Mo₄Zol₂Mn(II) was synthesized by reduction of Mo₄Zol₂Mn(III). Attempts to reduce this complex in solution failed so its reduction was performed by adding ascorbic acid to a suspension of crystals in methanol, as described for a Mn(III)-containing polyoxotungstate.²⁵ A color change of the crystals from gray to yellow was observed. This implies that the Mo(VI) centers have not been reduced, because if they were, a blue coloration would be expected. The $\chi_M T$ vs. *T* curve for this compound is shown in Figure S3 where it can be seen that the $\chi_M T$ product is nearly constant between 300 to 50 K $(\chi_M T = 4.15 \text{ cm}^3 \text{mol}^{-1} \text{K} \text{ at } 300 \text{ K})$, with a decrease at low temperature $(\chi_M T = 2.68 \text{ cm}^3 \text{mol}^{-1} \text{K}$ at 2K) which can be attributed to the zero-field splitting effect. This indicates that Mo₄Zol₂Mn(III) has been fully reduced to Mo₄Zol₂Mn(II) (at room temperature, the calculated $\chi_M T$ values are 2.94 cm³mol⁻¹K for a Mn(III) monomer and 4.29 cm³mol⁻¹K for a Mn(II) monomer assuming g = 1.98). As expected, the IR spectra of the oxidized and reduced POMs are very similar (Figure S1b). The synthesis of $Mo_4(ZolC_8)_2Mn(III)$ was performed by reacting Mn(OAc)₃ and ZolC₈ with a large excess of Na₂MoO₄ in acetate buffer. The reaction with a stoichiometric amount of Mo(VI) ions gave only an insoluble unidentified powder. $Mo_4(ZolC_8)_2Mn(III)$ was characterized by IR spectroscopy (Figure S1c), elemental analysis and EDX (energy dispersive X-ray analysis) measurements. Despite many attempts, we were unable to isolate the analogous compound with the ZolC₆ bisphosphonate.

³¹P NMR spectroscopy was used to determine whether structures characterized in the solid state were maintained in solution. The ³¹P NMR spectrum of $Mo_6(ZolC_6)_2$ dissolved in water at room temperature exhibits two singlets at 17.12 and 16.82 ppm with relative intensities 0.63:1.37 (Figures 2 and S2a). The same behavior is observed for $Mo_6(ZolC_8)_2$ with two singlets at 17.46 and 16.82 ppm with relative intensities 0.89:1.1 (Figure S2b). Only one singlet was expected if the structure observed in the solid state (Figure 3) was retained in solution, corresponding to four equivalent phosphorus nuclei. However the presence of another singlet close to the first one has already been observed for other hexanuclear complexes²¹ and has been attributed to an equilibrium between two conformers which differ by the rotation of the two { Mo_3O_8 } groups around the central oxo bridge (Figure S4). The two resonances are relatively sharp at room temperature and broaden when the temperature increases and finally coalesce at 70°C (Figure 2), indicating that, as expected, dynamic exchange between the two conformers increases on heating.

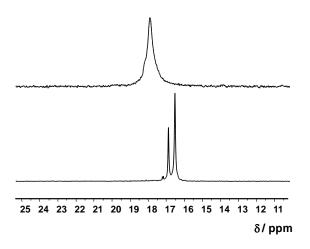


Figure 2. ${}^{31}P{}^{1}H$ NMR spectra of Mo₆(ZolC₆)₂ dissolved in D₂O at two different temperatures.

We also studied the stability of $Mo_6(ZolC_6)_2$ and $Mo_6(ZolC_8)_2$ under more physiological conditions (0.1 M KH₂PO₄ buffer, pH = 7.4, 0.12 M NaCl, 37°C) (Figure S2). The ³¹P NMR spectrum of $Mo_6(ZolC_6)_2$ exhibits two peaks, as observed for the aqueous solution, confirming the stability of the complex. In the spectrum of $Mo_6(ZolC_8)_2$, one of the peaks is split, which may be attributable to the existence of several conformers which could result from different positions of the alkyl chains, as seen in the crystal structure of $Mo_6(ZolC_6)_2$ (*vide infra*). It can also be seen that the ¹H NMR spectra of $Mo_6(ZolC_8)_2$ in water and in the more physiological conditions are identical (Figure S5).

Structures.

The structure of $Mo_6(ZolC_6)_2$ was determined by means of single-crystal X-ray diffraction. There are two crystallographically independent molecules (Figure 3) and the structure of the inorganic core is identical in both molecules. Two { Mo_3O_8 } trimeric units are connected by a central oxygen atom and in each trimer, Mo(VI) ions are connected to a pentadentate BP ligand by P-O-Mo and C-O-Mo bonds. The six Mo(VI) ions are coplanar, as usually observed for hexanuclear Mo(VI) complexes with BP ligands.^{19b} The differences between both complexes lie in the position of the C_6 alkyl chain on the imidazole ring. In one of the molecules (Molecule A, Figure 3a) the chain is bent and there are short C-H⁻⁻O distances (d_{C-H--O} in the 2.33-2.80 Å range).²⁶ In the other molecule (Molecule B, Figure 3b) the alkyl chain is extended away from the cluster. Molecules A and B stack into columns that align in the unit-cell (Figure 3c).

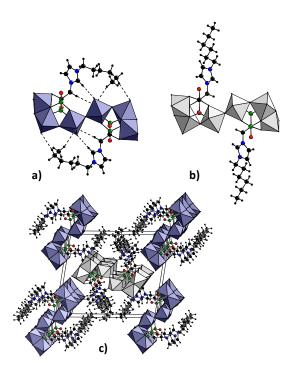


Figure 3. Representation of the two crystallographically independent molecules a) A and b) B in the structure of $Mo_6(ZolC_6)_2$. c) View of the crystal packing. Purple octahedra = MoO_6 octahedra in molecule A, grey octahedra = MoO_6 octahedra in molecule B, green spheres = P, black spheres = C, small black spheres = H, blue spheres = N, red spheres = O, hydrogen bonds are represented as dotted lines.

In the Mo₄L₂Mn complexes, IR, elemental analysis and EDX measurements indicate that the $Mo_4L_2Mn(III)$ (L = Zol, ZolC₈) and $Mo_4Zol_2Mn(II)$ complexes have structures that are similar to Mo_4Zol_2Fe .²² That is, two { Mo_2O_6 } dimeric units with faced-shared Mo(VI) octahedra are bound to an octahedrally coordinated central Mn ion (Figure 1) and two BP ligands are connected to the Mo and Mn ions through P-O-M and C-O-M (M = Mo, Mn) bonds.

In vitro and in vivo results.

We next investigated the effects of the new compounds (together with some compounds reported previously) on tumor cell growth inhibition using the non-small cell human lung cancer cell line NCI-H460. Representative dose-response curves are shown in Figure 4 for zoledronate (Figure 4a), $Mo_4Zol_2Mn(II)$ (Figure 4b), $Mo_6(ZolC_6)_2$ (Figure 4c) and $Mo_4(ZolC_8)_2Mn(III)$ (Figure 4d).

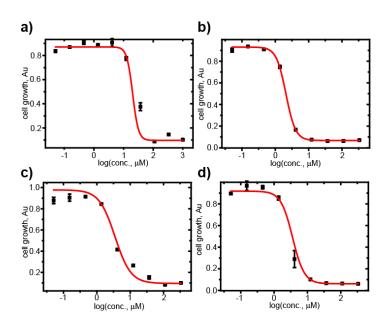


Figure 4. Dose response curves for NCI-H460 cell growth inhibition. a) zoledronate. b) $Mo_4Zol_2Mn(II)$. c) $Mo_6(ZolC_6)_2$. d) $Mo_4(ZolC_8)_2Mn(III)$. Three replicates of duplicates were determined, representative data from one duplicate set are shown.

 IC_{50} values for all compounds are reported in Table 1 on both a molecular weight as well as on a per-bisphosphonate basis. We anticipated that there would be a large increase in activity of the POM/BP complexes containing the more lipophilic bisphosphonates since the lipophilic bisphosphonates acting alone are more active than is zoledronate. With the zoledronate complexes we find that the new $Mo_4Zol_2Mn(II)$ complex has potent activity (on a per bisphosphonate basis), Figure 4b, similar to that of $Mo_4Zol_2Mn(III)$, on average ~6x the activity of zoledronate acting alone and considerably more activity than the Mo_6Zol_2 complex, Table 1. This suggests that the oxidation state of Mn has little influence on the activity of the complex. It can also be seen that zoledronate and the $Mo_4Zol_2Mn(III)$ complex are about 2x more potent in MCF-7 than in NCI-H460 cell lines, as reported previously,²² reflecting not unexpected cell-to-cell variability. With the $ZolC_6$ system we obtained more activity with the $Mo_6(ZolC_6)_2$ system than with $ZolC_6$ alone. However, with the more lipophilic $ZolC_8$ species, the activities of all three species: $ZolC_8$, $Mo_6(ZolC_8)_2$ and $Mo_4(ZolC_8)_2Mn(III)$ (Figure 4d) are quite similar to each other as well as to the Mo_4Zol_2Mn complexes, Table 1.

Formula	IC ₅₀ [µM]	IC ₅₀ [µM] per BP	IC ₅₀ decrease vs BP
Mo ₆ Zol ₂	10±0.9	20±1.8	1.4x
Mo ₄ Zol ₂ Mn(III)	2.6±0.3	5.2 ± 0.6	5.4x
Mo ₄ Zol ₂ Mn(II)	2.0 ± 0.1	4.0±0.2	7.0x
Zol	28±3	28±3	1.0x
$Mo_6(ZolC_6)_2$	3.5±0.1	$7.0{\pm}0.2$	3.4x
ZolC ₆	24 ± 0.8	24±0.8	1.0x
$Mo_6(ZolC_8)_2$	1.9 ± 0.2	3.8±0.4	1.3x
Mo ₄ (ZolC ₈) ₂ Mn(III)	2.9±0.3	5.8 ± 0.6	0.9x
ZolC ₈	5.1±0.2	5.1±0.2	1.0x

Table 1. Growth inhibition of human cancer cell line NCI-H460 by bisphosphonates and POM/BP complexes

We thus next moved to *in vivo* studies which since the pioneering work of Yamase and coworkers^{10a} have been quite rare.²⁷ Since there were only small differences in IC₅₀ values on a per-bisphosphonate basis between the most active species, we chose to investigate the $Mo_4Zol_2Mn(III)$ species reported earlier in our *in vivo* study, for the following reasons: i) it

has good activity in both the H460 (lung cancer) cell line described here, as well as against the MCF-7 (breast cancer) cell line reported previously;²² ii) zoledronate is commercially available while the alkyl zoledronates require several additional synthesis steps; iii) zoledronate is already used clinically while the alkyl zoledronates are not; iv) synthesis of the Mo₄Zol₂Mn(II) complex requires formation of the Mn(III) complex followed by the reduction of the Mn(III) crystals by ascorbate-again an additional synthetic step and v) there is little difference in activity between the Mn(II) and Mn(III) complexes, due perhaps to reduction of the latter in cells.

We used the mouse xenograft (SK-ES-1 sarcoma tumor cell growth) model because in previous work we used this system to investigate the effects of lipophilic bisphosphonate chain-length variation on tumor cell growth where we found that a very lipophilic pyridinium analog of zoledronate (BPH-715, containing an n-C₁₀ alkyloxy side-chain) resulted in a ~5x decrease in tumor volume versus zoledronate.²³ Representative results for PBS control, zoledronate and Mo₄Zol₂Mn(III) treated mice as a function of time are shown in Figures S6ac and tumor cell volumes as a function of time are shown in Figure 5a. As can be seen in Figure 5a, there is a very significant (p = 0.045) decrease in tumor cell volume with Mo₄Zol₂Mn(III) dosed at 5 µg/mouse via intra-tumoral injection once a day for 28 days, corresponding to a ~5x decrease in tumor cell growth versus zoledronate, even though there was only $\sim 1/3$ the amount of zoledronate present in the Mo₄Zol₂Mn(III) treatment (due to the high mass of the POM cluster). There was also only a small difference in mice body weight on zoledronate or Mo₄Zol₂Mn(III) treatment, Figure 5b, showing the absence of any increased toxicity of the complex versus zoledronate alone therapy. These relatively low bisphosphonate concentrations (compared with zoledronate or BPH-715 levels) may make such hybrids in general less toxic than bisphosphonates acting alone, although further work will be needed to investigate this possibility. In the future, it may be possible to carry out

detailed structure-activity relationships investigations of these types of complex, although this will be very challenging since it will be necessary to know cell concentrations of the complexes, their stability, as well as the targets for the Mo-moiety as well as the Mn. Plus, as briefly discussed below, both protein prenylation as well as kinase inhibition can be targeted by bisphosphonates.

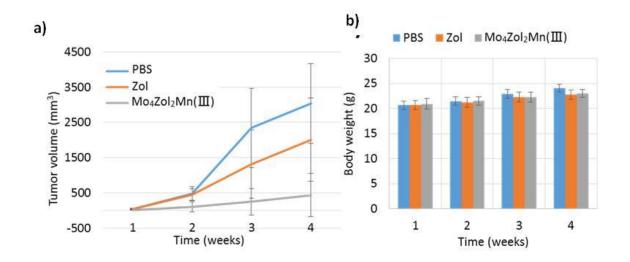


Figure 5. a) Effects of zoledronate and $Mo_4Zol_2Mn(III)$ on SK-ES-1 tumor volume as a function of time (weeks 1, 2, 3 and 4) in 6-week old athymic nude mice treated daily for 28 days with 5 µg compound or phosphate-buffered saline (PBS); 6 mice per group. b) Body weights as a function of time for different treatments.

Conclusions

We synthesized and characterized four new POM/BP complexes: Mo_6L_2 (L=ZolC₆, ZolC₈), $Mo_4(ZolC_8)_2Mn(III)$ and $Mo_4Zol_2Mn(II)$, and tested them, together with previously reported analogs, *in vitro* against a human non-small cell lung cancer cell line NCI-H460. The most active compounds had IC₅₀ values of ~ 4-5 μ M (on a per-bisphosphonate basis), about 6x more active than zoledronate itself (IC₅₀ = 28 μ M). One of the most active compounds tested here *in vitro* was also the most active compound against the MCF-7 breast cancer cell line reported previously, so was investigated *in vivo* against SK-ES-1 sarcoma cells in a mouse xenograft system. There was a five-fold decrease in tumor cell volume *versus* that found with zoledronate after a 28 day treatment period at 5 μ g/mouse dosing (~0.2 mg/kg). At present, the molecular basis for the enhanced activity (over that seen with bisphosphonates alone) remains to be determined since both masking of bisphosphonate charge, increasing cell permeability, as well as potentiation of bisphosphonate activity by the (hetero)polymolybdate moiety may be involved. Nevertheless, the results are of considerable interest because they show for the first time that POM/BP/Mn complexes have potent *in vivo* activity.

What is particularly attractive about the use of bisphosphonates as ligands in POM complexes is that zoledronate, for example, has the potential for targeting both Ras as well as EGF (epidermal growth factor)-driven tumors. Zoledronate inhibits farnesyl diphosphate synthase and this leads to inhibition of Ras prenylation. Moreover, in two recent articles²⁸ zoledronate has also been shown to lead to inhibition of EGFR signaling/phosphorylation. It may thus be possible to develop POM-bisphosphonate compounds that target both EGFR as well as Ras-driven cancers. Zoledronate is also an important POM-ligand since zoledronate itself activates gamma-delta T cells (containing the V γ 2V δ 2 T cell receptor) to kill tumor cells,²⁹ it switches macrophages from an M2 (tumor promoting) to an M1 (tumor killing) phenotype³⁰ and inhibits angiogenesis/invasiveness. There are thus good reasons to believe that POM/bisposphonate/Mn complexes may find utility as drug leads for combination therapies against both Ras as well as EGFR-driven cancers, that also target innate immunity.

EXPERIMENTAL SECTION

Experimental Procedures and Characterization. Zoledronic acid $H_2O_3PC(C_4H_5N_2)(OH)PO_3H_2$ $(H_5Zol)^{31}$ and $(NH_4)_6[(Mo_3O_8)_2O(O_3PC(C_4H_6N_2)OPO_3)_2]$ ·9H₂O $(Mo_6Zol_2)^{21}$ were synthesized according to reported procedures. Li₂H₂(ZolC₆) and Li₂H₂(ZolC₈) were synthesized with a protocol adapted from literature procedures²⁴ (see Supplementary Information).

Synthesis of (NH₄)₆[(Mo₃O₈)₂O(O₃PC(C₄H₅N₂)(C₆H₁₃)(O)PO₃)₂]·8H₂O (Mo₆(ZolC₆)₂): (NH₄)₆Mo₇O₂₄.4H₂O (0.120 g, 0.10 mmol) was dissolved in 5 mL of 1M NH₄OAc/HOAc buffer before addition of Li₂H₂(ZolC₆) (0.106 g, 0.24 mmol). The mixture was sealed in a 23mL Teflon-lined stainless steel reactor before heating to 130°C over a period of 1 h, kept at this temperature for 20 h, then cooled to room temperature over a period of 36 h. The resulting colorless crystals were collected by filtration. Yield: 0.144 g (68% based on Mo). ³¹P NMR (300 MHz, D₂O, 25°C): δ 17.12 (s), 16.76 (s). ¹H NMR (300 MHz, D₂O, 60°C, water saturation): 9.07 (d, 1H, NCHCHN), 7.83 (d, 1H, NCHCHN), 7.63 (s, 1H, NCHN), 5.07 and 4.94 (2m, 2H, NCH₂C), 4.44 (t, 2H, NCH₂CH₂), 2.14 (m, 2H, NCH₂CH₂), 1.59 (m, 6H, CH₂), 1.14 (m, 3H, CH₃). IR (FTR): ν (cm⁻¹) = 1640 (w), 1566 (w), 1433 (vs), 1152 (s), 1127 (s), 1086 (s), 1062 (sh), 1048 (s), 978 (m), 928 (sh), 912 (s), 875 (vs), 730 (vs), 698 (s), 629 (s), 560 (m), 518 (m), 482 (m). Anal. Calc. (found) for C₂₂H₇₆N₁₀Mo₆O₃₉P₄ (1804.4 g mol⁻¹): C 14.64 (14.40), H 4.25 (3.99), N 7.76 (7.69). EDX measurements confirm the Mo/P ratio.

Synthesis of (NH₄)₆[(Mo₃O₈)₂O(O₃PC(C₄H₅N₂)(C₈H₁₇)(O)PO₃)₂]·10H₂O (Mo₆(ZolC₈)₂): Na₂MoO₄.2H₂O (0.096 g, 0.40 mmol) was dissolved in 1.5 mL of water (sol. A); Li₂H₂(ZolC₈) (0.055 g, 0.12 mmol) was dissolved in 1.5 mL of water acidified with 2 drops of 1M hydrochloric acid (sol. B). Sol B was added to sol. A and 1M HCl was added dropwise to pH = 5. NH₄Cl (0.600 g, 11.3 mmol) was added, resulting in the precipitation of a fine white powder. The mixture was allowed to stir at room temperature for 20 min. The powder was filtered and dried with ethanol and diethyl ether. Yield: 0.061 g (48% based on Mo). ³¹P NMR (300 MHz, D₂O, 25°C): δ 17.46 (s), 16.82 (s). ¹H NMR (300 MHz, D₂O, 60°C, water saturation): 9.08 (d, 1H, NCHCHN), 7.86 (d, 1H, NCHCHN), 7.65 (s, 1H, NCHN), 5.05 and 4.96 (m, 2H, NCH₂C), 4.46 (t, 2H, N*CH*₂CH₂), 2.17 (m, 2H, NCH₂*CH*₂CH₂), 1.61 and 1.56 (m, 10H, CH₂), 1.13 (m, 3H, CH₃). IR (FTR): ν (cm⁻¹) = 1560 (w), 1414 (vs), 1153 (sh), 1134 (s), 1083 (m), 1058 (sh), 1048 (s), 980 (m), 917 (s), 876 (vs), 792 (w), 732 (m), 690 (s), 619 (s), 578 (m), 556 (m), 533 (m), 487 (w), 458 (w). Anal. Calc. (found) for $C_{26}H_{88}N_{10}Mo_6O_{41}P_4$ (1896.5 g mol⁻¹): C 16.47 (16.69), H 4.68 (4.51), N 7.39 (7.36). EDX measurements confirm the Mo/P ratio.

Synthesis of $(NH_4)_5[(Mo^{VI}_2O_6)_2(O_3PC(C_4H_6N_2)OPO_3)_2Mn^{III}].10H_2O$ $(Mo_4Zol_2Mn(III))$: To a solution of Na₂MoO₄·2H₂O (0.242 g, 1 mmol) in 10 mL of 1M CH₃COONH₄/CH₃COOH buffer was added Mn(OAc)₃·2H₂O (0.070 g, 0.26 mmol) and zoledronic acid (0.137 g, 0.5 mmol). The solution was stirred for 5 min then NH₃ (33% in water) was added dropwise to pH = 7.5. The solution was left to evaporate and was filtered after 24 h in order to remove a pink precipitate. EDX measurements and IR spectroscopy indicated that the precipitate was a BP complex that did not contain molybdenum. An homogeneous gray crystalline phase of the title compound appeared after three days (see picture in Figure S7). Yield: 0.080 g (21% based on Mo). Anal. Calc. (found) for $C_{10}H_{52}MnMo_4N_9O_{36}P_4$ (1437.2 g mol⁻¹): C 8.36 (8.88), H 3.65 (3.90), N 8.77 (8.78). IR (FTR): v (cm⁻¹) = 1575(m), 1546 (w), 1421(s), 1288 (w), 1136(s), 1112(sh), 1045(s), 1018(sh), 973(m), 915(s), 888(s), 790(s), 699(m), 656(m), 619(w), 560(w), 529(m). EDX measurements confirm the Mo/P and Mo/Mn ratios.

Synthesis of $(NH_4)_5(H_3O)[(Mo^{V_1}_2O_6)_2(O_3PC(C_4H_6N_2)OPO_3)_2Mn^{II}].6H_2O.4CH_3OH$ $(Mo_4Zol_2Mn(II))$: Ascorbic acid (0.150 g, 0.85 mmol) was added to a suspension of $Mo_4Zol_2Mn(III)$ (0.190 g, 0.13 mmol) in 30 mL of MeOH. The solution was stirred for 6 h and was filtered. The resulting pale yellow solid was washed with MeOH and dried. Yield: 0.129 g (68%). IR (FTIR) : v (cm⁻¹) = 1579(m), 1546 (w), 1418(s), 1286 (w), 1137(s), 1112(sh), 1047(s), 1018(sh), 975(m), 911(s), 887(s), 786(s), 699(m), 659(m), 619(w), 561(w), 529(m), 485(w). Anal. Calc. (found) for C₁₄H₆₃N₉MnMo₄O₃₇P₄ (1512.3 g mol⁻¹): C 11.12 (11.02), H 4.20 (3.47), N 8.34 (7.98). EDX measurements confirm the Mo/P and Mo/Mn ratios. Synthesis of $Li_2(NH_4)_3[(Mo^{V_1}_2O_6)_2(O_3PC(C_4H_5N_2)(C_8H_{17})OPO_3)_2Mn^{III}].8H_2O$ (Mo₄(ZolC₈)₂Mn(III)): Na₂MoO₄.2H₂O (0.197 g, 0.82 mmol) was dissolved in 5 mL of 1M NH₄OAc/HOAc and Mn(OAc)₃.2H₂O (0.018 g, 0.048 mmol) was added. A solution of Li₂H₂(ZolC₈) (0.027 g, 0.060 mmol) in 3 mL of H₂O acidified with a few drops of 1M HCl until dissolution was slowly added. The pH was adjusted to 5.8 with 33% NH₃. The solution was stirred for 2 hours and an orange crystalline precipitate which did not contain ZolC₈ was removed by filtration. The pH of the solution was equal to 6.1. The solution was left at room temperature for 5 days and the thin pinkish platelets (see picture in Figure S7) of the title compound were removed by filtration. Yield 0.011 g (23% based on ZolC₈). IR (FTIR) : v (cm⁻¹) = 1639(w), 1661(m), 1420(s), 1123(m), 1046(s), 990(w), 912(m), 871(s), 796(w), 743(w), 691(m), 620(m), 661(m). Anal. Calc. (found) for C₂₆H₆₄Li₂N₇MnMo₄O₃₆P₄ (1627.3 g mol⁻¹): C 19.19 (19.97), H 3.96 (4.18), N 6.03 (5.80). EDX measurements confirm the Mo/P and Mo/Mn ratios.

Infrared spectra were recorded on a Nicolet 30 ATR 6700 FTIR spectrometer.

Crystal Structure Determination. Single crystal X-ray diffraction data for $Mo_6(ZolC_6)_2$ were collected by using a Bruker Nonius X8 APEX 2 instrument equipped with a CCD bidimensional detector using the monochromatised wavelength $\lambda(Mo \ K\alpha) = 0.71073$ Å. Absorption corrections were based on multiple and symmetry-equivalent reflections in the data set using the SADABS program³² based on the method of Blessing.³³ The structure was solved by direct methods and refined by full-matrix least-squares using the SHELX-TL package.³⁴ NH₄⁺ and H₂O could not be distinguished based on the observed electron densities, therefore, all positions were labelled as O and assigned the oxygen atomic diffusion factor. Crystallographic data are given below and the complete data can be found in the cif file CCDC 1547689.

Crystal data for compound Mo₆(ZolC₆)₂: C₂₂H₃₆Mo₆N₄O₄₁P₄, M = 1712.07 g mol⁻¹, Triclinic, space group *P*-1, a = 8.8527(6) Å, b = 16.8404(11) Å, c = 19.6929(13) Å, $\alpha = 81.370(2)^{\circ}$, $\beta = 83.028(2)^{\circ}$, $\gamma = 76.955(2)^{\circ}$, V = 2816.0(3) Å³, T = 296 K, Z = 2, $D_c = 2.019$ g cm⁻³, $\mu = 1.515$ mm⁻¹, GooF = 1.145, Final R indices (I $\ge 2\sigma$ (I)) $R_1 = 0.0406$, $wR_2 = 0.1191$.

Powder X-ray. Powder X-ray diffraction data were obtained on a Bruker D5000 diffractometer using Cu radiation (1.54059 Å). The comparison of the experimental X-ray powder pattern with the powder pattern calculated from the structure solved from single-crystal X-ray diffraction data confirm the homogeneity of $Mo_6(ZolC_6)_2$ (Figure S8).

EDX Measurements. EDX measurements were performed on a JEOL JSM 5800LV instrument.

NMR measurements. ³¹P NMR spectra were recorded in 5 mm tubes with ¹H decoupling by using a Bruker AC-300 spectrometer operating at 121.5 MHz. ³¹P chemical shifts were referenced with respect to an external standard, 85% H₃PO₄, using the convention that low-field, paramagnetic, deshielded values are positive (IUPAC δ -scale). For all compounds 15-20 mg of sample was dissolved in D₂O (500 µL). The concentrations thus varied in the 10-30 mM range.

Magnetic measurements. Magnetic susceptibility measurements were carried out on polycrystalline samples using a Quantum Design MPMS SQUID magnetometer operating in the 300-2 K temperature range and 0-5.5 T. Susceptibility measurements were performed with an applied field of 1000 Oe. Pascal's constants were utilized to estimate diamagnetic corrections, the value in each case being subtracted from the experimental susceptibility data to give the molar magnetic susceptibility (χ_M).

Cell-growth inhibition assays. The human tumor cell line NCI-H460 (non-small cell lung cancer) was obtained from the National Cancer Institute and maintained at 100% humidity and 5% CO₂ at 37 °C. Cell growth inhibition assays were carried out as described previously

 20 A broth microdilution method was used to determine the growth inhibition IC₅₀ values. Briefly, ~5 x 10⁴ cells suspended in 100 μL of DMEM supplemented with 10% fetal bovine serum (FBS), 4.5 g/L glucose and L-glutamine and preserved with 1% penicillin– streptomycin were seeded in 96-well plates (Corning Inc., Corning, NY) and incubated at 37 °C in a 5% CO₂ atmosphere. The cells were incubated with 1000 μM, 333 μM, 111 μM, 37 μM, 12 μM, 4.1 μM, 1.4 μM, 0.46 μM, 0.15 μM, and 0.051 μM of compounds and H₂O as a control for 4 days.²⁰ Then, an MTT ((3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) cell proliferation assay (ATCC, Manassas, VA) was performed to obtain dose– response curves. The IC₅₀ values of the free BP, Mo₆Zol₂ and Mo₄Zol₂Mn(III), which had been measured in MCF-7 cells in our first studies^{21,22} were re-determined here using the same conditions as for the other compounds (Table 1).

In Vivo Tumor Model. Mice experiments were carried out basically as described in Kubo et al.³⁵ Xenografts of human SK-ES-1 cells were initiated by subcutaneous injections of 1.5×10^7 cells into the right flank of four 6-week old athymic nude mice (CLEA, Tokyo, Japan). The mice received daily local injections of bisphosphonates (5 µg, for 28 days), or physiological saline. The smallest and largest diameters of the tumors, were measured weekly. Tumor volumes were calculated using the following formula: volume (mm³) =(smallest diameter)² x (largest diameter)/2. All animal experiments were conducted according to the guidelines of the Institutional Animal Care and Use Committee, and the protocol was approved by the Ethics Committee for Experimental Animals of Hiroshima University. Statistical significance was determined by one-way ANOVA and independent t-test, using SPSSI (version 22, IBM); p < 0.05 was considered to be significant.

ASSOCIATED CONTENT

Supporting Information. Synthesis of ZolC_6 and ZolC_8 , IR and NMR spectra, $\chi_M T = f(T)$ curve for Mo₄Zol₂Mn(II), representation of the *A* and *B* conformers of the Mo₆L₂ compounds, pictures of mice treated with PBS, zoledronate or Mo₄Zol₂Mn(III), SEM images of crystals of Mo₄Zol₂Mn(III) and platelets of Mo₄(ZolC₈)₂Mn(III), powder X-ray diffraction pattern of Mo₆(ZolC₆)₂. This material is available free of charge via the Internet at http://pubs.acs.org.

Accession Codes

CCDC 1547689 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge via www.ccdc.cam.ac.uk/data_request/cif, or by emailing data_request@ccdc.cam.ac.uk, or by contacting The Cambridge Crystallographic Data Centre, 12, Union Road, Cambridge CB2 1EZ, UK; fax: +44 1223 336033.

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