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Body composition and sarcopenia:

the next-generation of personalized oncology and pharmacology?

Marc Hilmi¹, Anne Jouinot¹, Robert Burns², Frédéric Pigneur², Rémi Mounier⁴, Julien Gondin⁴, Cindy Neuzillet^{3*}, François Goldwasser^{1*}

Affiliations :

- 1. Department of Medical Oncology, CAncer Research for PErsonalized Medicine (CARPEM), Paris Centre Teaching Hospitals, Paris Descartes University, USPC, Paris, France
- 2. Department of Radiology, Henri Mondor University Hospital, Créteil, France
- 3. Department of Medical Oncology, Curie Institute, Versailles Saint-Quentin University, Saint-Cloud, France, and GERCOR group, Paris, France
- 4. Institut NeuroMyoGène (INMG) CNRS 5310 INSERM U1217 UCBL, Lyon, France
- * Contributed equally (co-last authors)

Corresponding author:

Dr. Cindy Neuzillet, M.D., Ph.Doc., Department of Medical Oncology, Curie Institute, Versailles Saint-Quentin University (UVSQ), 35 rue Dailly, 92210 Saint-Cloud, France. E-mail: cindy.neuzillet@gmail.com / cindy.neuzillet@curie.fr. Tel: +33 (0)6 82 55 04 92.

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ABSTRACT

Body composition has gained increasing attention in oncology in recent years due to fact that sarcopenia has been revealed to be a strong prognostic indicator for survival across multiple stages and cancer types and a predictive factor for toxicity and surgery complications. Accumulating evidence over the last decade has unraveled the "pharmacology" of sarcopenia. Lean body mass may be more relevant to define drug dosing than the "classical" body surface area or flat-fixed dosing in patients with cancer.

Since sarcopenia has a major impact on patient survival and quality of life, therapeutic interventions aiming at reducing muscle loss have been developed and are being prospectively evaluated in randomized controlled trials. It is now acknowledged that this supportive care dimension of oncological management is essential to ensure the success of any anticancer treatment.

The field of sarcopenia and body composition in cancer is developing quickly, with (i) the newly identified concept of sarcopenic obesity defined as a specific pathophysiological entity, (ii) unsolved issues regarding the best evaluation modalities and cut-off for definition of sarcopenia on imaging, (iii) first results from clinical trials evaluating physical activity, and (iv) emerging body-compositiontailored drug administration schemes.

In this context, we propose a comprehensive review providing a panoramic approach of the clinical, pharmacological and therapeutic implications of sarcopenia and body composition in oncology.

Keywords: cachexia, chemotherapy, sarcopenic obesity, targeted therapy, toxicity.

ABBREVIATIONS

MKI: multikinase inhibitor

- MRI: magnetic resonance imaging
- NLR: neutrophil-to-lymphocyte ratio
- NSCLC: non-small cell lung cancer
- OS: overall survival
- OR: odds ratio
- PK: pharmacokinetics
- PS: performance status
- SCLC: small cell lung cancer
- SMA: skeletal muscle area
- SMI: skeletal muscle index
- TGF-β: transforming growth factor beta
- TMA: total muscle area
- TNF: tumor necrosis factor

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TEXT

1. Introduction

Sarcopenia (*i.e*. loss of muscle mass and function) is a hallmark of cachexia, a multi-organ syndrome characterized by negative protein and energy balance, weight loss (including muscle with or without fat), anorexia, and decreased physical function (Fearon et al., 2011). Cachexia has been defined in the setting of underlying inflammatory disease as: (i) unintended weight loss of > 5% over the past 6 months; or (ii) body mass index (BMI) < 20 kg/m² and any degree of weight loss > 2%; or (iii) muscle atrophy (*i.e.* reduced muscle mass) as determinable by various modalities of body composition analysis and any degree of weight loss > 2% (Fearon et al., 2011). Not all cachectic patients have sarcopenia, and not all sarcopenic patients meet consensus criteria for cachexia, while most cancer patients in this context have both sarcopenia and cachexia (Figure 1). Sarcopenia is observed in more than 50% of patients with metastatic cancers (Morishita et al., 2012). It negatively affects survival and health-related quality of life (HRQoL) of patients due to decreased tolerance to anticancer treatments and increased susceptibility to infections and other complications (Bozzetti, 2017). Therefore, sarcopenia and cachexia represent a major clinical target in oncology.

Cancer-related sarcopenia is multifactorial in origin, involving mainly inflammatory and hypercatabolic syndrome, anxiety/depression, and chemotherapy adverse effects (nausea/vomiting, mucitis, diarrhea, and loss of appetite) (Fearon et al., 2011). Given this multifaceted pathophysiology, a multimodal therapeutic approach to sarcopenia management including nutritional support and exercise on a background of personalized oncology care and family-centered education is advocated for (Morley et al., 2010). Several techniques and markers are used to characterize sarcopenia. Lean body mass (LBM) may be more relevant to adapt drug dosing than the "classical" body surface area or flat-fixed dosing. Sarcopenia and changes in body composition present a rapidly developing field of oncology, with the newly identified concept of sarcopenic defined as a specific pathophysiological entity, unsolved issues regarding evaluation modalities and best cut-off for imaging definition of sarcopenia (probably not a "one-size-fits-all" value), first results

from clinical trials evaluating physical activity, and emerging body-composition-tailored drug administration schemes.

2. Sarcopenia definition and modalities of body composition analysis

2.1 Definition and pathophysiology

Sarcopenia was first described in 1989 as an age-related loss of muscle mass (Rosenberg, 1989). Later, the definition was expanded to a syndrome combining three criteria: (i) loss of skeletal muscle mass, (ii) loss of function, and (iii) loss of physical performance (Rosenberg, 1997; Roubenoff, 2000). Three consensus papers established a definition of sarcopenia based on these three parameters (Cruz-Jentoft et al., 2010; Fielding et al., 2011; Muscaritoli et al., 2010). The European Working Group on Sarcopenia in Older People (EWGSOP) proposed three stages of sarcopenia: presarcopenia (presence of one criterion), sarcopenia (two criteria), and severe sarcopenia (three criteria) (Table 1). The techniques used to diagnose sarcopenia include (i) cross sectional imaging, dual-energy X-ray absorptiometry (DXA), or bioelectrical impedance analysis (BIA) for the assessment of muscle mass, (ii) handgrip test for the assessment of muscle strength, and (iii) a short physical performance battery (*i.e.,* a group of measures that combines gait speed, chair stand, balance tests, and walk speed) (Guralnik et al., 2000) (Figure 2). The muscle function was added into the sarcopenia definition based on the results demonstrating that its decrease is correlated with mortality (Baumgartner et al., 1998). Nevertheless, beside motor functions, skeletal muscle mass plays an important role in the endocrine regulation of metabolism (Janssen & Ross, 2005). Overall the motor function is overly narrow and does not reflect the endocrine function of the muscle. Thus, muscle mass should take the priority over muscle function and be considered as the cornerstone of the definition criteria for sarcopenia (Bulow, Ulijaszek, & Holm, 2018). Sarcopenia is also classified as "primary" when age-related and "secondary" when activity, nutrition, or disease-related such as in cancer or other chronic diseases (Cruz-Jentoft, Landi, Topinková, & Michel, 2010).

Inflammation is a predominant driving mechanism in cancer-related sarcopenia pathophysiology. C reactive protein (CRP) is a marker for inflammation and has been suggested as a

biological marker for sarcopenia (Bano et al., 2017). In a comparable way, neutrophil-to-lymphocyte ratios (NLR) are higher in sarcopenic patients than in those without sarcopenia (Öztürk, Kul, Türkbeyler, Sayıner, & Abiyev, 2018) as well as circulating levels of pro-inflammatory cytokines (*e.g.* tumor necrosis factor [TNF], interleukin-1 [IL-1], and interleukin-6 [IL-6]) (Narsale & Carson, 2014; Schaap, Pluijm, Deeg, & Visser, 2006; Visser et al., 2002). The cytokines of the transforming growth factor beta (TGF-β) family, *i.e.,* TGF-β, myostatin, GDF11, and activins are major atrophic factors widely involved in cancer-induced cachexia (Wakefield & Hill, 2013). Their circulating levels are increased in most cancers, including pancreatic and colorectal cancers (Wildi et al., 2001; Zhao et al., 2016). These molecules trigger intracellular signals leading to loss of contractile proteins linked to a decrease in protein synthesis and a significant increase in the degradation of muscle myofibrillar proteins, inevitably impacting muscle strength production (Cohen, Nathan, & Goldberg, 2015). Preclinical studies have shown that ZIP14, a metal-ion transporter, and Twist1, a transcription factor, are overexpressed in muscle progenitor cells, highlighting the roles of zinc homeostasis and activin/myostatin signaling in cancer-related muscle loss (Parajuli et al., 2018; G. Wang et al., 2018). In addition, muscle proteolysis induces an important efflux of muscle amino acids, which constitutes "bricks and fuel" to boost tumor progression (Mayers et al., 2016). However, the mechanisms underlying cancer-related muscle dysfunction appear to be more complex than just a negative balance between muscle protein synthesis and degradation. It has recently been shown that microenvironmental alterations affecting cells located at the periphery of the muscle fiber also play a key role in cancer-related muscle atrophy. Indeed, muscle fibers are surrounded by muscle stem cells called "satellite cells". During muscle injury, these cells become activated and proliferate to produce myoblasts, which differentiate and fuse to form new myofibers and thus restore muscle tissue integrity. In cancer, inflammation induces muscle damage leading to the activation of satellite cells that are engaged in a myogenic muscle tissue repair program, but cannot differentiate (He et al., 2013). Other mechanisms have recently been shown to be involved in cancer-related sarcopenia, such as epigenetic alterations involving bromodomain protein BRD4 (Segatto et al., 2017) and calcium homeostasis involving ryanodine receptors (A. Agrawal, Suryakumar, & Rathor, 2018;

Waning et al., 2015). Endocrine and metabolic factors also play a pivotal role in the pathogenesis of sarcopenia. Some publications suggests an association between insulin resistance and mitochondrial dysfunction leading to the development of skeletal muscle lipid deposition (Corcoran, Lamon-Fava, & Fielding, 2007) and to reduced muscle oxidative activity (Simoneau & Kelley, 1997) in sarcopenia and cancer-induced cachexia (Abbatecola et al., 2011; van der Ende et al., 2018). Moreover, insulin promotes amino acids transport into cells through nitric oxide synthase (Mann, Yudilevich, & Sobrevia, 2003). Therefore, insulin resistance may lead to poorer protein synthesis due to reduced internalization of amino acids. These hypotheses may explain why patients with type 2 diabetes mellitus are more at risk of developing sarcopenia (Leenders et al., 2013; Morley, Malmstrom, Rodriguez-Mañas, & Sinclair, 2014; Park et al., 2007).

Overall, systemic inflammation, metabolic changes, and secreted "atrophying" cytokines along with the muscle microenvironmental dysfunction alter cell signaling in muscle fibers and lead to imbalance between protein synthesis and degradation (Argilés, Campos, Lopez-Pedrosa, Rueda, & Rodriguez-Mañas, 2016) (Figure 2). These phenomena are very early in some malignancies, particularly in pancreatic cancer, and may precede cancer diagnosis by several years (Agustsson, D'souza, Nowak, & Isaksson, 2011; Mayers et al., 2014). Moreover, sarcopenia itself is affected by chemotherapy, since a variety of common chemotherapy drugs are known to experimentally induce sarcopenia in rodent cancer models (Hojman et al., 2014; Sakai et al., 2014). Data on humans carry some caveats because chemotherapy agents are not given to healthy adults and it seems that muscle wasting is exacerbated by chemotherapy (Awad et al., 2012; Coletti, 2018) through reduced food intakes (Spotten et al., 2017) and induced NF-kB expression (Damrauer et al., 2018). The decrease in food intakes in cancer patients is multifactorial and classically thought to result from changes to appetite, smell and taste and to behavior-regulating regions of brain occurring as a result of inflammatory mediators (Ezeoke & Morley, 2015).

2.2 Modalities of body composition evaluation

In clinical practice, physical performance assessment by short physical performance battery, usual gait speed, and get up and go test (Cruz-Jentoft, Baeyens, et al., 2010; Owusu, Margevicius, Schluchter, Koroukian, & Berger, 2017) and muscle strength evaluation using handgrip test (Kilgour et al., 2013; Veni et al., 2018) are quite standardized. On contrary, there is no consensus on the optimal modality to assess muscle mass in cancer patients.

Anthropometric measurements include weight variation, BMI, waist and hip circumference, waist-to-hip ratio, and skinfold. These are non-invasive, easy to perform, and routinely used to estimate body surface area (BSA; derived from weight and height) for chemotherapy dosing calculation (Griggs, Mangu, Temin, & Lyman, 2012). However, anthropometry is unreliable in some cancer patients, particularly in case of short-term changes in body water composition (*e.g.* in presence of ascites or lymphedema), or in obese patients (Di Sebastiano & Mourtzakis, 2012).

Bioelectrical impedance analysis (BIA) is an additional widely available, non-invasive modality that uses reactance and resistance to determine total body water, fat mass (FM), and fatfree mass. However, BIA is highly dependent on patient hydration state and is biased in case of pathological increase in body water content such as ascites and lymphoedema. Other limitations of BIA is the lack of specific predictive equations for cancer patients leading to frequent inaccuracies (under or overestimations) and the lack of standardization resulting in heterogeneity of sarcopenia prevalence rates across studies (Gonzalez, Barbosa-Silva, & Heymsfield, 2018).

Dual energy X-ray absorptiometry (DXA) uses a three-compartment model comprising FM, fat-free mass, and bone mineral content. This method can detect early lean mass variations, is highly accurate and reproducible (Ellis, 2001; Shiel et al., 2018). However, it cannot discriminate between different types of fat tissues (visceral, subcutaneous, and intramuscular) as only an overall assessment of FM at the molecular and not at compartmental tissue level can be provided by DXA and no differentiation of specific lean tissues like skeletal muscle and the internal organs within the thorax or abdomen can be made (Guglielmi et al., 2016; Prado & Heymsfield, 2014). Consequently, muscle mass using DXA is estimated based on the appendicular skeleton.

Finally, it has been shown that cross sectional muscle surface at the third lumbar vertebrae (L3) best reflects total skeletal muscle mass determined by computed tomography (CT) or magnetic resonance imaging (MRI) (Heymsfield, 2008; Mitsiopoulos et al., 1998; Shen et al., 2004). In practice, total muscle area (TMA, in cm²) is measured at L3 using a semi-automatic segmentation software on a dedicated post-treatment station (with most packages being interchangeable) (Bonekamp et al., 2008). TMA is then normalized to stature (using height² in m^2 , similarly to BMI) (Baumgartner et al., 1998) to obtain the skeletal muscle index (SMI) in cm²/m².

However, uncertainties remain regarding technical aspects of this method. To date, the only technical guideline available for L3 skeletal muscle surface estimation using CT is the one described by Mitsiopoulos *et al.* in 1998 and recommended by Prado *et al.* in 2008 (Mitsiopoulos *et al.*, Prado *et al.*). It recommends measurement of the mean skeletal muscle surface value on two consecutive slices at the L3 level based on Hounsfield unit (HU) thresholds (–29 to +150), followed by manual corrections where necessary. Nevertheless, several technical issues that can influence skeletal muscle surface measurements need clarification such as the effect of intravenous contrast injection (van Vugt et al., 2018), slice thickness (Fuchs et al., 2018), or the influence of tube potential (Morsbach et al., 2018).

Regarding MRI, there are no clear technical recommendations on image acquisition techniques. Most studies used standard T1 and T2-weighted imaging protocols (Heymsfield, 2008; Mitsiopoulos et al., 1998; Yang et al., 2017) and fat-water separated imaging techniques (Dixon, 1984) with manual segmentation to determine L3 skeletal muscle surface, which is time consuming. Very few studies have used or investigated the reproducibility of automated methods applied to MRI (Borga, 2018). This is most likely due to the fact that MRI is not used in a quantitative way because, unlike HU on CT imaging, the intensity levels on MRI are expressed in arbitrary units and are not correlated to tissue composition.

A single-muscle approach to muscle mass quantification using cross-sectional analysis (surface and/or density) of the psoas muscle on abdominal CT and of the pectoralis muscle on thoracic CT in patients who do not undergo abdominal CT (*e.g.* osteosarcomas and head and neck

cancers) has been proposed (Go et al., 2017; Y. S. Kim, Kim, Kang, Ahn, & Kim, 2017). However, these single-muscle, simplified approaches, which hypothesized that one muscle may be representative of total skeletal mass have not been validated by any expert group (Bahat et al., 2016; Baracos, 2017; Cesari et al., 2012). Automated total muscle segmentation on CT imaging has been suggested as an alternative, more accurate method to improve body composition quantification availability and routine application (Baracos, 2017; Popuri, Cobzas, Esfandiari, Baracos, & Jägersand, 2016).

Assessment of muscle quality may provide prognostic information beyond quantity estimation (Sami Antoun et al., 2013; Looijaard et al., 2016; Martin et al., 2013). Skeletal muscle lipid content has been associated with muscle quality *i.e.,* high-lipid content being correlated with poor muscle quality. It can be assessed by measuring muscle density on CT imaging, as the latter decreases with lipid infiltration (Heymsfield, 2008). Goodpaster *et al.* showed that muscle density in elderly patients can account for differences in muscle strength not explained by muscle quantity (Goodpaster, Kelley, Thaete, He, & Ross, 2000). This however was not confirmed by another study of healthy adults (Weeks, Gerrits, Horan, & Beck, 2016). Similarly to TMA estimation, there is no consensus for important technical aspects of CT acquisition such as contrast enhancement, slice thickness, and tube potential influence, which significantly impact HU density values of the muscle. Further research is needed for the standardization of image acquisition and protocols of analysis.

In summary, cancer-related sarcopenia and body composition can be estimated by simple anthropometric values (weight loss and BMI) and L3 SMI using CT or MRI, with CT being the most commonly used modality in clinical practice and oncology research (Kazemi-Bajestani, Mazurak, & Baracos, 2016) (Table 3). Indeed, CT is routinely performed for diagnosis, treatment evaluation, and follow-up in oncological care and TMA assessment. In addition, it can be performed on the same imaging exam without requiring an additional procedure. However, in the foreseeable future, with the development of artificial intelligence, the next gold standard for body composition assessment will most likely rely on 3D body-scanning with automated scoring systems (Cornet et al., 2015; Fang, Berg, Cheng, & Shen, 2018).

2.3 Unsolved questions including cut-off issues

A major unsolved issue in sarcopenia evaluation is the question of the best cut-off value for sarcopenia definition. Sarcopenia is more often considered in a binary fashion for research purposes in the literature, patients being classified as either "sarcopenic" or "non-sarcopenic". Several cut-offs have been proposed (Cornet et al., 2015; Cruz-Jentoft, Baeyens, et al., 2010) for sarcopenia diagnosis, yielding broad variations in sarcopenia prevalence (Bijlsma et al., 2013). The first available cut-off values specific to patients with cancer were published by Prado *et al.* reporting on a population of obese Canadian patients with respiratory and gastrointestinal tract cancers (Prado et al., 2008). They defined cut-off value of 52.4 cm^2/m^2 in men and 38.5 cm^2/m^2 in women associated with mortality. Mourtzakis *et al.* showed that CT-based muscle analysis at L3 was strongly related to appendicular skeletal mass measured with DXA (Mourtzakis et al., 2008). They generated corresponding CT cut-off values of 55.4 cm²/m² in men and 38.9 cm²/m² in women using established DXA cut-off values (7.26 kg/m² and 5.45 kg/m², respectively). In 2011, an international panel of cachexia experts established a new diagnostic criterion for cancer cachexia, using both anthropometric measures and SMI based on cut-off values of 55 cm^2/m^2 in men and 39 cm^2/m^2 in women by CT imaging (Fearon et al., 2011). Following, Martin *et al.* proposed SMI thresholds for sarcopenia in non-obese Caucasians according to sex and BMI (Martin et al., 2013) based on optimal stratification of SMI and survival; selected cut-off values were 43 and 53 cm²/m² in men with a BMI < and > 25 kg/m², respectively, and 41 cm²/m² in women. Several further studies have demonstrated predictive value for toxicity (Sjøblom et al., 2015; Srdic et al., 2016; Stene et al., 2015; B. H. L. Tan et al., 2015), post-surgery complications (Lieffers, Bathe, Fassbender, Winget, & Baracos, 2012; P. Peng et al., 2012; Reisinger et al., 2015), and survival (M. H. Choi, Oh, Lee, Oh, & Won, 2018; Lee et al., 2018) using these thresholds (Table 2).

However, normal amounts of muscle and adipose tissues depend on demographic factors such as age (McCormick & Vasilaki, 2018) and ethnicity (Wells, 2012). It seems that sarcopenia is less prevalent among African-American (Parsons, Baracos, Dhillon, Hong, & Kurzrock, 2012) and Asian

patients (Lau, Lynn, Woo, Kwok, & Melton, 2005) as compared to Caucasians. Therefore, these cutoffs may not be optimal for all patients and may be refined depending on type and stage of the cancer, sex, age, and ethnicity.

Nevertheless, rather than classifying a patient dichotomically as sarcopenic *vs.* nonsarcopenic based on a cut-off value, imaging biomarkers like SMI may be considered as continuous variables as suggested by Voron *et al.* (Voron et al., 2015). In such way, these may be used as the next reference for chemotherapy dosing calculation. This approach is already being tested with promising results (Baracos & Arribas, 2018; Iannessi, Beaumont, Hebert, Dittlot, & Falewee, 2018).

3. Sarcopenia prevalence and prognostic value

Sarcopenia has been reported across all cancer types. Prevalence rates according to primary tumor location and stage are displayed in Table 3. Most studies reported sarcopenia rates above 50% using L3 CT scan, particularly at advanced stages and in pancreatic, lung, bladder, and hematological malignancies. Noticeably, heterogeneity in prevalence rates is observed within single cancer types due to different cut-offs used to define sarcopenia and factors (*e.g.,* clinical stage, ethnicity, age, cancer treatment).

Several studies investigated the prognostic value of sarcopenia in cancer patients. In a metaanalysis of patients with solid tumors a significant association between sarcopenia and shorter overall survival (OS; overall HR = 1.51, *p*<0.001) has been reported, both for advanced and localized stages (Shachar, Williams, Muss, & Nishijima, 2016). However, a recent study suggested that survival may not be different for combined muscle and fat loss compared to fat-only loss in patients with advanced pancreatic cancer (Kays et al., 2018). Besides, a higher risk of post-surgical complications has been identified in several cancers, especially infectious complications following surgery for gastrointestinal (Ida et al., 2015; Krell et al., 2013; Lieffers et al., 2012; P. Peng et al., 2012; Takagi et al., 2017; Zhuang et al., 2016) and lung cancers (Miller et al., 2018; R. Nakamura et al., 2018). Finally, loss of skeletal muscle mass during chemotherapy has also been described as a negative prognostic factor (Daly et al., 2018).

Overall, several studies evaluating body composition using various modalities have consistently reported that sarcopenia is a strong prognostic indicator for localized and advanced cancers.

4. Sarcopenia and toxicity of anticancer treatments

In addition to its prognostic value, sarcopenia is a predictive factor of anticancer drug toxicity and may be more relevant for drug dose calculation than the "classical" body surface area or flat-fixed dosing. Anticancer drugs display various pharmacokinetics (PK) properties but share a narrow therapeutic index. Serious adverse events that may result in toxic death are common in this setting. Furthermore, specific cancer patient populations may exhibit vulnerabilities due to age and comorbidities. Identifying factors that can explain individual variations in treatment efficacy and toxicity is a new challenge of modern oncology. Performance status (PS) has a strong prognostic value (Atkinson et al., 2015) and is predictive of anticancer treatment-related acute toxicity (Sargent et al., 2009). However, PS evaluation is only semi-quantitative and is subjected to inter-observer variability. Thus, additional parameters are needed for the risk assessment of anticancer treatments. Other studies have investigated whether wide variations in body composition could be associated with morbidity in cancer patients. In recent years a large interest has grown in muscle mass due to the development of muscle mass assessments by CT.

We performed systematic search on PubMed using MeSH terms "Neoplasms AND (Sarcopenia OR Body composition OR Malnutrition OR Cachexia) AND Toxicity". Forty-two studies that focused on sarcopenia and toxicity of anticancer treatments were selected (Figure 3).

4.1 Chemotherapy

In total, among 32 studies that investigated the association between sarcopenia and chemotherapy toxicity, 24 showed a significant association between sarcopenia and chemotherapy toxicity (Table 4). Although most of the studies were conducted retrospectively and with a small number of patients (n < 100), this correlation was found consistently across studies regardless of the

time of assessment (*i.e.,* early, within the first months of chemotherapy, or later), the tumor type and stage (*i.e.,* from localized/curative to the metastatic/palliative setting), and the chemotherapy regimen. For example, Prado *et al.* (Prado et al., 2009) evaluated prospectively the tolerance of capecitabine in 55 patients with metastatic breast cancer after the first treatment cycle. Toxicities occurred in 50% of sarcopenic patients *vs.* 20% of non-sarcopenic patients (*p*=0.03). Sarcopenia was also a predictive factor of toxicity in apparently fit patients (*i.e*., PS 0-1) such as those treated in phase I trials (Cousin et al., 2014). Indeed, Cousin *et al.* studied body composition in 93 phase I patients and showed that low SMI was the only factor associated with severe toxicity (Cousin et al., 2014).

Body composition in cancer patients can also be evaluated through renal function assessments in cancer patients. In daily practice, Glomerular Filtration Rate (GFR) is evaluated using the Cockcroft–Gault, Chronic Kidney Disease - Epidemiology Collaboration (CKD-EPI), and Modification of diet in renal disease (MDRD) formulas, which are derived from serum creatinine (Cr) level; this latter is influenced by the total muscle mass (Perrone, Madias, & Levey, 1992). In sarcopenic patients, reduced muscle mass may result in apparently low Cr level and overestimation of renal function by these formulas. Serum cystatin C (CysC) is an alternative marker of GFR, independent from muscle mass (Baxmann et al., 2008; Rule, Bergstralh, Slezak, Bergert, & Larson, 2006). Thus, CysC appears as a better alternative for assessing renal function in patients with low muscle mass. The Cr/CysC ratio can be used as a quantitative surrogate marker of muscle mass (Tetsuka, Morita, Ikeguchi, & Nakano, 2013). In a study including 25 patients with non-small cell lung cancer (NSCLC) (Suzuki et al., 2015), a significant difference was noted in the Cr/CysC ratios of patients exhibiting moderate (grade 1-2) *vs.* severe (grade 3-4) toxicities (mean ratios: 0.84 *vs.* 0.70, respectively; *p*<0.05). Similar results were found in a prospective study of ovarian cancer patients treated with carboplatin (Bretagne et al., 2017). Moreover, renal failure from the early stages exacerbates sarcopenia, explaining the frequent overlap between these two conditions (de Souza et al., 2017).

Contrarily, eight studies showed no association between sarcopenia and chemotherapy toxicity (Blauwhoff-Buskermolen et al., 2016; Jung et al., 2015; Miyata et al., 2017; Palmela et al., 2017; Parsons, Tsimberidou, et al., 2012; Rollins et al., 2016; Srdic et al., 2016; Stene et al., 2015; Versteeg et al., 2017). One explanation could be that not all forms of chemotherapy exhibit sarcopenia-dependent toxicity, even if the studies were adequately powered to detect an association. Finally, one study used liver intra-arterial chemotherapy (Parsons, Tsimberidou, et al., 2012), which displays a specific PK profile.

4.2 Targeted therapy

Eight studies investigated the association between sarcopenia and toxicity of targeted therapies with multikinase inhibitors (MKI), mainly antiangiogenics (Table 5). Overall, patients were metastatic in all studies and had infrequent tumor types in all but one study (four studies of renal carcinoma, two studies of hepatocellular carcinoma, and one study of thyroid carcinoma). The association between sarcopenia and toxicity was reported in seven studies. A meta-analysis pooling the results of four studies of metastatic renal cell carcinoma, showed that dose-limiting toxicities (DLT) of antiangiogenic drugs were more frequent in patients with low SMI (*p*=0.03) (Vrieling et al., 2016). As observed with chemotherapy, increased toxicity in sarcopenic patients was observed from the first cycle of treatment (Huillard et al., 2013) and was associated with high serum concentration of MKI (Massicotte et al., 2013; Mir, Coriat, Blanchet, et al., 2012). In one study of NSCLC patients treated with afatinib (Arrieta et al., 2015), sarcopenia was not found predictive of toxicity using sexspecific cut-offs defined by Prado *et al.* (Prado et al., 2008) and low LBM and malnutrition were associated with severe gastro-intestinal toxicity and DLT.

4.3 Immunotherapy

In recent years, immune checkpoint blockade inhibitors have opened new opportunities in cancer therapy. They have markedly improved the clinical outcomes in several cancers, including metastatic melanoma (Hodi et al., 2010; Robert et al., 2014), renal cancer (Motzer et al., 2018), and

lung cancer (Reck et al., 2016). Severe immune-related adverse events occurred in 10%-20% of patients treated with single-agent immune checkpoint inhibitor (Brahmer et al., 2015; Hodi et al., 2010; Motzer, Escudier, et al., 2015). Predictive factors of toxicity are still poorly known. Unlike for chemotherapy, the use of DLT for immune therapies, and more broadly for monoclonal antibodies (mAb), is inappropriate. Indeed, no maximum tolerated dose was found in phase I trials with bevacizumab (Gordon et al., 2001), rituximab (Ghielmini, 2005), cetuximab, pembrolizumab (Sachs, Mayawala, Gadamsetty, Kang, & de Alwis, 2016), and nivolumab (S. Agrawal, Feng, Roy, Kollia, & Lestini, 2016). Several dosing schemes have been developed based on weight (dose in mg/kg) or flat doses. The Food and Drug Administration (FDA) concluded that response/toxicity of nivolumab is not related to dose/exposure and shifted from body weight dosing (*i.e.* 3 mg/kg) to fixed dosing (240 mg) (www.fda.gov/Drugs/InformationOnDrugs/ApprovedDrugs/ucm520871.htm). However, the rate of severe toxicities seems to increase with ipilimumab dose. In a retrospective study of exposureresponse analysis in advanced melanoma patients, Feng *et al.* (Feng et al., 2013) showed that the occurrence of grade 3-4 immune-related adverse events was related to ipilimumab dose, with 3%, 13%, and 24% of patients developing severe toxicities when treated with 0.3 mg/kg, 3 mg/kg, and 10 mg/kg ipilimumab, respectively. Interestingly, no DLT was observed with tremelimumab despite a similar toxicity profile to ipilimumab (Camacho et al., 2009). A study of 84 patients with metastatic melanoma (Daly et al., 2017) treated with ipilimumab showed that sarcopenic patients developed higher grade adverse events than non-sarcopenic patients (OR=5.34, p=0.033). Similar results were observed in 68 metastatic melanoma patients treated with nivolumab and pembrolizumab (Heidelberger et al., 2017). However, it is hard to discern differences between a direct PK link and a consequence of increased frailty and susceptibility to treatment complications in sarcopenic patients in this setting.

4.4 Summary and discussion

Overall, this literature review shows that sarcopenic patients exhibit high rates of toxicity and highlights the role of body composition in the risk assessment of anticancer treatment in this

setting. However, several questions remain unsolved due to methodological limitations and heterogeneity across reviewed studies. Reporting biases favoring positive over negative studies may have affected the validity of evidence. In addition, most of these studies included limited numbers of patients, with no preliminary estimation of sample size for statistical analysis. Besides, the evaluation of toxicities varied from one study to another: some studies considered all DLT, while other only analyzed some specific toxicities such as hematologic adverse events. The duration of toxicity evaluation ranged from the first cycle only to the entire length of treatment. Moreover, for a single tumor location, the thresholds and techniques used to detect sarcopenia differed between studies. Finally, although several studies using the SMI cut-offs defined for survival by Prado *et al.* showed a positive correlation between sarcopenia and toxicity, it is not certain that these are the best suited for toxicity (Table 2). As it was achieved for prognosis, it would be relevant to conduct a meta-analysis of studies dedicated to toxicity.

5. The pharmacology of sarcopenia

Several hypotheses can be raised to explain the association between cancer-related sarcopenia and toxicity. Evidence has accumulated during the last decade unravelling the "pharmacology" of sarcopenia. We will below develop the main hypotheses through PK and pharmacodynamics.

6.1 Pharmacokinetics

The PK hypothesis assumes that toxicity may be explained by overexposure of anticancer drugs in sarcopenic patients. From a pharmacological perspective, sarcopenia increases drug exposure and several studies showed correlation between LBM, drug exposure, and toxicity (Ali et al., 2016; Arrieta et al., 2015; Prado et al., 2007, 2014, 2011; Sjøblom et al., 2017, 2015; Stanisavljevic & Marisavljevic, 2010; Sucak et al., 2012). Regarding mAb (*e.g.* immune therapies and some of targeted therapies), no data was published so far on sarcopenia and PK. However, mAb are a category of drugs known for larger therapeutic index than cytotoxic chemotherapies or MKI. For

instance, no DLT were observed for the highest doses of anti-programmed-death-1 (PD-1)/antiprogrammed-death-ligand-1 (PD-L1) antibodies in phase I and II trials (Motzer, Rini, et al., 2015; Robert et al., 2014).

6.1.1 Absorption changes

It has been suggested in animal models that cancer-induced cachexia is associated with a gut barrier dysfunction due to microbiota dysbiosis and increased intestinal permeability (Klein, Petschow, Shaw, & Weaver, 2013), possibly leading to overexposure to drugs that are taken orally. The same mechanisms may exist in sarcopenia, but no studies have been conducted to confirm this hypothesis in specific models. This issue would be most pertinent when the drug in question is not typically well-absorbed orally. Absorption of drugs that are intended to be taken with meals may be compromised in patients with low oral intakes.

Obese sarcopenic patients have higher fat-to-lean-mass ratios than obese non-sarcopenic patients. This may explain an overexposure to targeted subcutaneous therapies (*i.e.* trastuzumab) through an increase in (highly vascularized) fat tissue and subcutaneous blood flow. Studies supporting this hypothesis are warranted. Similarly, no studies have been conducted to assess whether intramuscular fat deposits in obese sarcopenic patients alter absorption of intramuscular lipophilic injection drugs (*i.e.,* decapeptyl, fulvestrant). In contrast, drugs with intravenous administration route are not affected by absorption changes.

6.1.2 Distribution changes

Changes in tissue relative proportion are likely to alter PK. Body composition is described as a two compartments model consisting of FM and LBM. LBM is defined as the sum of cellular mass and non-fatty intercellular connective tissue including tendons, ligaments, and bone (Ronenn Roubenoff & Kehayias, 2009). Indeed, muscle mass, being part of LBM, acts as a diffusion compartment for cancer treatments. Thus, decreased LBM in sarcopenia may lead to increased plasma levels of anticancer drugs. Five studies showed that toxicities were increased for higher

doses of chemotherapies per kilogram of LBM (Ali et al., 2016; Prado et al., 2007; Sjøblom et al., 2017, 2015; Stanisavljevic & Marisavljevic, 2010). Other chemotherapy and MKI studies showed increased toxicity in women (Arrieta et al., 2015; Huillard et al., 2013; Prado et al., 2007; Sloan et al., 2002). The most likely explanation is that LBM is lower in women compared to men of the same age (Bredella, 2017), resulting in higher exposure to these drugs. Indeed, BSA, which is used for chemotherapy prescription, does not properly reflect the relative proportion of LBM and FM that are usually higher in men and women, respectively, in body composition (Gusella, Toso, Ferrazzi, Ferrari, & Padrini, 2002). Distribution volumes of mAb are low (*i.e* only extracellular fluids and blood plasma) due to their high hydrophilicity and size (Keizer, Huitema, Schellens, & Beijnen, 2010). As a result, they are less affected by body composition since changes in blood volume are less than proportional with the change in body weight (Boer, 1984). Several studies based on population PK showed that fixed dosing of mAb is more relevant that body weight dosing since it reduces interpatient variability (Hendrikx et al., 2017).

Besides distribution volumes, cancer-related sarcopenia could also be linked to overexposure through its association with hypoalbuminemia (Visser et al., 2005). Plasmatic free fraction increases for highly albumin-bound drugs (*e.g.* carboplatin, etoposide, cisplatin, docetaxel, paclitaxel, irinotecan) in patients with hypoalbuminemia. In a prospective cohort of 100 patients with NSCLC (Srdic et al., 2016), albumin concentration was established as a predictive factor for both chemotherapy toxicity and survival. This finding was confirmed in other studies of NSCLC (Arrieta et al., 2010; X. Wang et al., 2014).

6.1.3 Metabolic and clearance changes

Another explanation for overexposure in sarcopenic patients may be decreased activity of liver cytochromes (CYP) that are involved in metabolism of numerous anticancer drugs. In a PK study conducted to evaluate liver metabolism in rats affected by cancer-induced cachexia (Cvan Trobec et al., 2015), midazolam and propranolol clearances were used as reliable markers of CYP3A4 (Rogers, Rocci, Haughey, & Bertino, 2003) and CYP2D6 (Pirttiaho et al., 1980) activities. In the cachectic

setting, midazolam and propranolol clearances decreased by 80%, leading to higher drug concentrations. Cancer-related inflammation, which is associated with sarcopenia, leads to decreased expression of liver CYP450 (Charles et al., 2006). Beside cytochromes, other enzymes are involved in drug metabolism and may be modulated by nutritional status and cancer-related sarcopenia. An experimental study in rats showed that protein uptake impacted 5-fluorouracil (5-FU) toxicity, with 85% mortality in rats fed with low-protein diet *vs.* 12% with high-protein diet (Davis, Lenkinski, Shinkwin, Kressel, & Daly, 1993). The group of protein-deficient rats showed decreased activity of dihydropyrimidine dehydrogenase, the key enzyme of 5-FU catabolism. Conversely, a high-protein diet was associated with decreased 5-FU toxicity (Flanigen-Roat, Milholland, & Ip, 1985).

In addition, through the reduction of liver enzyme activity, sarcopenia is associated with a lower clearance for drugs which elimination relies on liver metabolism (characterized by low extraction ratio), including anthracyclines (Ballet, Vrignaud, Robert, Rey, & Poupon, 1987). In a study of 24 breast cancer patients, Prado *et al.* (Prado et al., 2011) showed a correlation between LBM and epirubicin clearance. Similar results were found in two other studies including adults and children treated with doxorubicin (Thompson et al., 2009; Wong et al., 2014). Cancer-related inflammation could be a confounding factor by downregulating CYP450 activity and thus detoxification of most of antineoplastic agents (Christmas, 2015). Nevertheless, the epirubicin study of Prado *et al.* included only localized and operated breast cancer cases for which a systemic inflammation level is low, suggesting that sarcopenia may also have a direct, inflammation-independent effect on drug metabolism (Prado et al., 2011). For most MKI, exposure is mainly dependent on liver clearance and LBM was shown to be a good PK predictor. In a study of 40 hepatocellular carcinoma patients treated with sorafenib, median area under the curve (AUC) of sorafenib was twice higher in sarcopenic than in non-sarcopenic patients (Mir, Coriat, Blanchet, et al., 2012). Similar results were found in medullary thyroid cancer patients treated with vandetanib (Massicotte et al., 2013). Moreover, clearance of mAb differs from MKI and does not rely on liver elimination. Indeed, they are primarily eliminated through intracellular degradation after binding to the target, which is fast

and saturable, and to a lesser extent through proteolytic catabolism, a non-specific immunoglobulin G pathway, which is slow and linear (Keizer et al., 2010). As a result, mAb clearance is not related to body weight, but to their affinity, tumor burden, and target expression levels. Overall, except mAb, changes in drug metabolism in sarcopenic patients are related to liver function with an overlap with malnutrition and inflammation.

6.2 Pharmacodynamics

Alternatively, the PD hypothesis postulates that, independently from PK, sarcopenic patients are more sensitive to treatment and may experience toxicities even in the absence of overdose. This is related to the concept of frailty, which has become increasingly recognized as one of the most important issues in health outcomes and is of particular importance in cancer patients (Kumar Pal, Katheria, & Hurria, 2010). Frailty can be defined as a state of diminished physiologic reserve that results in increased vulnerability to stressors (cancer itself, anticancer treatments) and higher risk of adverse events (complications, dependency, death) (Xue, 2011). Frailty, malnutrition, sarcopenia, and cachexia are overlapping entities. Even if frailty is a phenotype that has been primarily described in older adults (≥ 70 years), similar to sarcopenia, it is also observed in younger patients (Smart et al., 2017) and it is associated with increased risk of postoperative complications, chemotherapy toxicities, disease progression, and death (Ethun et al., 2017). Although the Clinical Frailty Scale (Rockwood et al., 2005) is widely used, there is no standardized scale or cut-off to assess frailty, contributing to variable prevalence rate. In a systematic review including 20 studies and nearly 3 000 older cancer patients, the median prevalence of frailty was 42% (range 6%-86%) (Handforth et al., 2015). One explanation is that sarcopenic patients exhibit decreased immunity and slower cell renewal, which results in higher risk of febrile neutropenia and severe mucositis. The association between nutritional status and radiotherapy toxicity (Hill, Kiss, Hodgson, Crowe, & Walsh, 2011) is another argument in favor of the frailty hypothesis since radiation therapy exposure is independent of PK parameters.

6. Sarcopenic obesity: a new entity

Sarcopenic obesity is defined by the association of low muscle mass and obesity (*i.e.,* BMI ≥30 kg/m²). The growing prevalence of obesity worldwide and the recent gain of interest in cancerrelated sarcopenia contributed to identification of a growing number of patients presenting with both the highest ranges of fat mass and the lowest ranges of muscle mass. The mean prevalence of sarcopenic obesity in cancer patients is around 10% (Baracos & Arribas, 2018). However, there is a large variation in prevalence (range 1-27%; Table 3), depending on the methods used for the body composition assessment, the inclusion of sarcopenic overweight patients (*i.e.* BMI \geq 25 kg/m²), and the tumor type and stage. Sarcopenic obesity is observed more frequently in advanced and metastatic tumors than in localized stages (Anandavadivelan, Brismar, Nilsson, Johar, & Martin, 2016; Dalal et al., 2012; Del Fabbro et al., 2012; Heidelberger et al., 2017; Rollins et al., 2016). In comparison to Caucasian patients, the prevalence of sarcopenic obesity seems lower in Asians, probably owing to the low prevalence of obesity in this population (Zhuang et al., 2016).

Sarcopenic obese patients carry the burden of both obesity and sarcopenia, which result in higher health-related risks than any of these conditions alone. Indeed, sarcopenic obesity is associated with shorter survival, worse postoperative outcomes, and increased toxicities of antitumor treatments. Prado *et al.* (Prado et al., 2008) were first to report an association between sarcopenic obesity and survival in cancer patients. They found that sarcopenic obese patients have poorer functional status and shorter survival compared to those with normal SMI (21.6 *vs.* 11.3 months respectively, *p*<0.001). Moreover, sarcopenic obesity was an independent predictor of poorer survival (HR=4.2, *p*<0.0001) in a multivariate analysis adjusted for age, sex, tumor type, tumor stage, and PS. These findings were confirmed by other studies in different cancer types (Table 2), especially pancreatic cancer (Dalal et al., 2012; Rollins et al., 2016; B. H. L. Tan et al., 2015). Obese cancer patients present with usually poor outcomes, with a U-shaped association between BMI and survival, called the "obesity paradox" (Caan et al., 2017; Valentijn et al., 2013), which may be explained by high prevalence of sarcopenic obesity. In a cohort of 1,473 patients with lung or gastrointestinal cancer, Martin *et al.* (Martin et al., 2013) showed that non-sarcopenic obese patients had longer survival (median OS: 35.6 months, *i.e.,* a doubling of overall median OS: 16.7

months) than sarcopenic and weight-losing obese patients (median OS: 8.5 months). Sarcopenic obesity is also associated with worse short and long-term outcomes after cancer surgery. Several studies reported lower survival rates in obese sarcopenic compared to non-obese non-sarcopenic patients after surgery for bladder (Psutka et al., 2015), colon (Malietzis et al., 2016), hepatocellular (Kobayashi et al., 2017), or gastric cancers (Palmela et al., 2017). Sarcopenic obesity has also been associated with surgical complications such as infections and delayed wound healing. Sarcopenic obese patients have 3 to 6-fold higher risk of developing major postoperative complications than non-sarcopenic patients after gastrectomy for gastric cancer (Lou et al., 2017; Nishigori et al., 2016) and after liver (P. D. Peng et al., 2011) or colorectal resection (Berkel et al., 2018; Malietzis et al., 2016) for colorectal cancer. Additionally, sarcopenic obesity is associated with longer hospital stay (Lou et al., 2017; P. D. Peng et al., 2011) and the higher 30-day readmission rates (Lou et al., 2017). However, in another recent study, no relationship was found between sarcopenia or sarcopenic obesity and postoperative complications (Lodewick et al., 2015).

Several hypotheses can be raised to explain the association between sarcopenic obesity and postoperative complications in cancer patients. Obesity is commonly associated with insulinresistance, which could be exacerbated in case of sarcopenia, since skeletal muscle is a major target for insulin-mediated glucose storage (Cleasby, Jamieson, & Atherton, 2016; Srikanthan, Hevener, & Karlamangla, 2010). Moreover, obesity and sarcopenia are both strongly associated with chronic inflammation (Cancello & Clément, 2006; Malietzis et al., 2016; Neves et al., 2016; Reisinger et al., 2015), which could in turn negatively affect the metabolic response and impair the immune response to surgical stress, leading to higher susceptibility to surgical site infections and impaired wound healing (Pierpont et al., 2014). Although the association with chemotherapy toxicity is less documented for sarcopenic obesity than for sarcopenia *per se*, a few recent studies reported an association between sarcopenic obesity and the occurrence of DLT (Table 4). Anandavadivelan *et al.* reported that sarcopenic obese patients have a 5-fold higher early DLT (during cycle 1) rate after neo-adjuvant therapy for esophageal cancer, compared to non-sarcopenic obese patients (Anandavadivelan et al., 2016). Conversely, high BMI or sarcopenia alone were not associated with a

significantly increased risk of DLT. Similar results were found in the neoadjuvant setting in gastric cancer, where all sarcopenic obese patients had to discontinue chemotherapy prematurely due to severe adverse events (Palmela et al., 2017). Heidelberger *et al.* showed that sarcopenic obese women treated with anti-PD1 checkpoints inhibitors for metastatic melanoma had more frequent early DLT (50% *vs.* 7.7% in non-sarcopenic obese, *p*<0.01) (Heidelberger et al., 2017). In contrast, some authors did not find any association between sarcopenic obesity and the occurrence of DLT (Cushen et al., 2016; Grotenhuis et al., 2016).

In conclusion, sarcopenic obesity has an increased prevalence in oncology and is associated with worse functional status, shorter survival, and higher risk of developing postoperative complications or DLT. This new entity reinforces the importance of body composition measurement in daily oncology practice since muscle loss in sarcopenic obese patients is often hidden by the increased BMI and stable body weight.

7. Therapeutic implications

7.1 Muscle mass-guided dose adjustments in daily practice and clinical trials

From the pharmacological point of view, body composition assessment could be helpful for identifying patients at higher risk of complications and severe toxicity and, as a next step, for adjusting dose administration of anticancer treatments. In recent years, many studies highlighted the relationship between body composition and toxicities of anticancer drugs. However, in clinical practice, dose calculation does not take into account body composition. On the one hand, the doses of chemotherapeutic agents are calculated using BSA, which relies on the fact that blood volume – *i.e.,* distribution volume - is correlated to BSA. The BSA equations are based on weight and height and were validated 100 years ago on nine subjects (Du Bois & Du Bois, 1989). Although several formulas have been proposed for BSA calculation since then, none of them seem to fit all patients correctly (Redlarski, Palkowski, & Krawczuk, 2016). On the other hand, oral targeted therapies are prescribed at a flat dose, regardless of body composition. As mentioned above, sarcopenic patients show increased drug exposure. For prescriptions, dose calculations may be adapted to the type of

drug and to the patient body composition. Regarding drugs prescribed for BSA, obese sarcopenic patients are particularly at risk of overdose (La Colla et al., 2007). Indeed, an increase in the distribution volume is expected for lipophilic molecules and thus extending their elimination halflives (Baker, Grochow, & Donehower, 1995) and requiring a dosage adjustment based on the total body weight. Conversely, low lipophilic molecules have limited diffusion in adipose tissue and can be then adjusted to LBM. On the contrary, for MKI that are prescribed in flat doses, underweight sarcopenic patients (BMI <18.5 kg/m²) are more prone to drug overexposure since they display low body mass in addition to low LBM. Patients treated with MKI may also benefit from therapeutic drug monitoring (*i.e.,* assessment of circulating levels). Therapeutic drug monitoring of drugs with strong binding to albumin or subject to liver metabolism is also of interest. Following first cycle administration and assessment of early toxicity, dose adaptation for subsequent cycles may be made according to individual tolerance, as previously proposed (Gurney, 1996).

Beyond the aforementioned "frailty" hypothesis, PK changes may explain the increased incidence of DLT in sarcopenic obese patients treated with chemotherapy. For BSA-based prescriptions, such as cytotoxic chemotherapies, sarcopenic obesity results in high absolute doses (due to large BSA/obesity), while drug distribution volume and metabolism are reduced. Indeed, many cytotoxic chemotherapies are hydrophilic, with distribution and metabolism into the LBM compartment, which is very depleted in case of sarcopenia, resulting in overexposure (La Colla et al., 2007) and a higher incidence of DLT. The distribution volume of lipophilic drugs will also be altered due to decreased plasma protein binding in both obesity and sarcopenia (Feldschuh & Enson, 1977; Hunter et al., 2009). In a study of ovarian cancer patients treated with lipophilic drugs such as liposomal doxorubicin and trabectedin toxicity was strongly associated with a higher FM/LBM (reflecting sarcopenia) in obese and overweight patients (Prado et al., 2014). Drug metabolism may be altered not only by muscle depletion, but also by obesity itself. Indeed, the clearance capacity of liver and kidneys does not grow proportionally with total body weight (Young et al., 2009). For instance, Demirovic *et al.* showed that incorporation of LBM into the Cockcroft-Gault equation provides an accurate estimation of kidney function in obese patients (Demirovic, Pai, & Pai, 2009).

Finally, experimental studies in rats with fatty-liver showed that lipid accumulation results in reduced CYP expression and activity (Su, Sefton, & Murray, 1999), which may partially explain excess toxicity in obese patients.

Studies assessing DLT in obese patients should also be considered in relation to the common practice of "dose-capping", *i.e.*, calculating chemotherapy doses with a maximum BSA of 2.0 m² for obese individuals (Griggs, Mangu, Anderson, et al., 2012; Pai, 2012). Given that dose capping is done without considering body composition, the dose reduction cannot be optimal and may result in under or overdosing (Hunter et al., 2009). Dose capping leads to overestimation of drug dose in sarcopenic obese patients, and to underestimation in non-sarcopenic obese patients. Dignam *et al.* showed that obese patients treated with capped doses of adjuvant chemotherapy for colon cancer, generally tolerate more chemotherapy cycles than normal weighted individuals (Dignam et al., 2006). However, obese patients receiving chemotherapy based on unadjusted BSA develop more severe toxicities (Furlanetto et al., 2016). LBM-based prescriptions would then appear as a better approach for determining chemotherapy doses, although further studies are needed for further clarification of the potential benefit from dose modifications in obese sarcopenic patients. An ongoing LEANOX study (NCT03255434) will compare an impact of LBM-based and BSA-based normalization of oxaliplatin-based chemotherapy on the incidence of severe neurotoxicity in stage III colon cancer patients.

For treatment subjected to renal clearance, we recommend calculation of GFR using CysC since GFR formula based on creatinine may overestimate renal function in sarcopenic patients. In addition, the calculation of carboplatin dose relies directly on GFR estimation and CysC may be considered instead of creatinine for this agent (Schmitt et al., 2009). The relevance of CysC and LBMbased dose adjustment for drugs with renal elimination in clinical practice needs to be confirmed by prospective studies evaluating not only safety parameters, but also response rates, progression-free survival, and OS to ensure that dose adjustments will not translate into decreased drug efficacy. A randomized phase II trial of advanced lung cancer (NCT01624051) comparing cisplatin dosing based on LBM or BSA is ongoing.

Finally, it is now possible to monitor immunotherapies (Puszkiel et al., 2017), even if they display a larger therapeutic index for which the usual definition of DLT is inappropriate (Postel-Vinay, 2015). Thus, our proposal is not to include LBM-based dose adjustment, but only to closely monitor toxicities in sarcopenic patients receiving mAb as these are more vulnerable. If a total body weight is lower than a threshold in which the flat dose was studied, it is necessary to be careful and switch to weight-adjusted dose (*i.e.,* mg/kg).

Given the prognostic and predictive toxicity values of cancer-related sarcopenia, it is preferable in phase I trials to either exclude sarcopenic patients or to perform a subgroup analysis of those patients (*i.e.,* specific cohort or stratification). For malignancies in which sarcopenia is extremely prevalent, it could be argued that including a sarcopenic subanalysis in safety studies is crucial data for determining whether the drug is a generally feasible option for that type of cancer.

7.2 Nutritional interventions, physical activity and drugs targeting muscle loss

Since sarcopenia has a major impact on patient survival and HRQoL, therapeutic interventions aiming at reducing muscle loss (nutritional, physical activity, and pharmacological interventions) have been developed and are prospectively evaluated in randomized controlled clinical trials. It is now acknowledged that this supportive care dimension of oncological management is essential to ensure the success of any anticancer treatment.

7.2.1 Nutritional interventions

A multicenter randomized trial by Cramer et al. evaluating the impact of high-protein oral nutritional supplements during 24 weeks in sarcopenic patients with cancer (Cramer et al., 2016) showed no benefit of nutritional intervention in the severe sarcopenia stage (like in refractory cachexia). In patients with moderate sarcopenia, an improvement was observed only in leg muscle strength with no benefit on muscle mass. Oral solutions enriched with specific amino acids such as glutamine (Ishikawa et al., 2016) and leucine (Deutz et al., 2011) also appeared to improve muscle mass loss in cancer patients. Omega-3 fatty acids such as eicosapentaenoic acid promote muscle

synthesis and decrease muscle degradation by downregulating IL-6 and TNF-alpha production via ubiquitin proteasome pathway and by increasing muscle insulin sensitivity (Pappalardo, Almeida, & Ravasco, 2015). Four randomized trials showed a positive effect of eicosapentaenoic acid on LBM (maintenance or increase) despite the lack of survival benefit (Ryan et al., 2009; Mantovani et al., 2008; Sánchez-Lara et al., 2014; Vasson et al., 2014).

Finally, enteral nutrition appears to provide a significantly greater benefit than parenteral route due to the high infectious risk that is associated with this latter (Koretz, Lipman, Klein, & American Gastroenterological Association, 2001; Zaloga, 2006). In terminally ill patients, it is now clear that implementing artificial nutrition brings no benefit to patients in terms of OS and HRQoL (McCann, Hall, & Groth-Juncker, 1994).

However, nutritional interventions alone are ineffective in improving survival. They may be active and essential as a part of multimodal approaches, including combination with exercise.

7.2.2 Physical activity

In cancer patients, adapted physical activity (APA) reduces disease and/or treatmentinduced symptoms (including pain, fatigue, and anxiety/depression), improves physical fitness, muscle function, and HRQoL (Buffart, Galvão, Brug, Chinapaw, & Newton, 2014; Cramp & Byron-Daniel, 2012), even in advanced-stage disease (Mustian et al., 2017). On contrary, no drug has shown any benefit in the treatment of cancer-related fatigue (Mücke et al., 2015; Mustian et al., 2017).

A randomized controlled trial reported by Cormie et al. showed that patients with prostate cancer benefit from supervised exercise in terms of muscle mass gain and FM loss, preventing treatment toxicity of androgen-deprivation therapy (Cormie et al., 2015). Similarly, positive results were found in breast cancer premenopausal women during endocrine therapy (Hojan, Milecki, Molińska-Glura, Roszak, & Leszczyński, 2013). Two randomized trials also demonstrated a benefit of supervised APA in breast cancer in the adjuvant setting on physical fitness, fatigue and

chemotherapy completion (van Waart et al., 2015), and metabolic syndrome, sarcopenic obesity and circulating biomarkers in overweight/obese patients (Dieli-Conwright et al., 2018), respectively.

Exercise have beneficial effects on tumor outcome (Schmid & Leitzmann, 2014) by modulating various pro-tumoral signaling pathways such as reduced insulin resistance and inflammation (Ashcraft, Peace, Betof, Dewhirst, & Jones, 2016). Besides, combining a high protein diet less than two hours after exercise seems to promote muscle synthesis (Atherton et al., 2010; Pennings et al., 2011). The benefit of physical activity and nutritional interventions for treating sarcopenia in elderly patients by promoting muscle mass, muscle strength, and physical function was confirmed in a meta-analysis reported by Yoshimura et al. (Yoshimura et al., 2017). In cancer patients, the extent of the benefit appears lower because chronic fatigue and a deficient oxidative metabolism are current in advanced stage disease (Argilés, Busquets, López-Soriano, Costelli, & Penna, 2012). In any case, as sarcopenia and fatigue frequently affect patients with metastatic cancer, physical exercise is a promising strategy to improve HRQoL (Focht et al., 2013).

Implementation of an APA program in cancer patients implies a multidisciplinary collaboration between physicians, nurses, dietitians, and the physical activity professionals (Wolin, Schwartz, Matthews, Courneya, & Schmitz, 2012). The APA program should be personalized to the patient characteristics (physical fitness, exercise type preferences, psychological functions, and expectations) and the cancer type and settings (stage, treatments, and tolerance) in order to improve the patient adherence. A combined aerobic exercise and resistance-training program in groups of patients having similar physical capabilities and under the supervision of a physical activity professional seems to be the best setting for exercise intervention efficacy (Cadore & Izquierdo, 2013; Pahor et al., 2014). Besides, cancer patients often present with an elevated basal resting energy expenditure (Jouinot, Vazeille, & Goldwasser, 2018). Given that the APA program is expected to increase energy expenditures, nutritional management is crucial for monitoring and adapting food intakes to ensure that patients meet their nutritional needs. Moreover, the main difficulty is to propose exercise interventions to all patients suffering from sarcopenia, particularly to the most

severely deconditioned (*i.e.,* PS ≥ 2) for whom participation in voluntary exercise sessions in hospital can be challenging.

Therefore, there is a need for developing alternative physical-therapy strategies in these severely deconditioned patients. Neuromuscular electrical stimulation (NMES) consists in generating muscle contractions using portable devices connected to surface electrodes. NMES was proven effective to improve muscle mass and function in sarcopenic patients with chronic obstructive pulmonary disease (Maddocks et al., 2016) and has recently been introduced in cancer patients. NMES is safe, does not require the active cooperation of the patient and can be self-administered at home, thereby providing an acceptable physical therapy for patients with advanced cancer and an altered PS and/or a high-symptom burden for whom attendance to hospital-based exercise training is difficult. However, the effectiveness of NMES in cancer patients remains equivocal (Crevenna, Marosi, Schmidinger, & Fialka-Moser, 2006; Maddocks et al., 2013; O'Connor & Caulfield, 2018; Windholz, Swanson, Vanderbyl, & Jagoe, 2014). Conflicting findings from previous clinical studies may be due to methodological limitations (small sample size, timing of intervention, suboptimal adherence) and heterogeneity of NMES protocols. Indeed, the main determinant of the NMES effectiveness is strength produced in response to stimulation (Gondin, Cozzone, & Bendahan, 2011; Maffiuletti, 2010), which has never been monitored in cancer patients. Overall, there is a need for carefully designed studies in order to draw definitive conclusions on the potential benefits of NMES interventions on cancer-related sarcopenia. It is therefore necessary to better understand the cellular and molecular mechanisms involved in the development of sarcopenia associated with cancer and to propose new complementary and/or alternative therapeutic interventions to APA.

7.2.2 Drugs

7.2.2.1 Hormonal therapy

Testosterone showed a benefit on muscle mass and grip strength in older men (Bakhshi, Elliott, Gentili, Godschalk, & Mulligan, 2000; Ferrando et al., 2002). This improvement is counterbalanced by harmful side effects (*i.e.,* prostate cancer, vascular thrombosis, and sleep apnea)

leading to the development of selective androgen receptor modulators that demonstrate a safer therapeutic profile. Enobosarm is a selective androgen receptor modulator tested *vs.* placebo in 159 pre-cachectic patients with hormone-naive prostate cancer. It showed a significant positive effect on muscle mass (*p*=0.046) with a positive impact on HRQoL, but without difference on tumor progression and muscle strength (Dobs et al., 2013). Similar results were observed in a randomized, prospective, double-blinded study of 170 sarcopenic women without cancer (Papanicolaou et al., 2013). These therapies may behave differently in young versus aged individuals.

Ghrelin is a neurohormone secreted by the stomach that stimulate appetite and muscle anabolism in the hypothalamus (Guillory, Splenser, & Garcia, 2013). Ghrelin analogues such as anamorelin have been developed. More than 450 cachectic advanced NSCLC patients were randomized to receive anamoreline or placebo for 12 weeks in the ROMANA trials (Currow et al., 2017; Temel et al., 2016). The results were in favor of a muscle mass gain in the experimental arm. However, there was no benefit on muscle strength or on OS. In contrast, there was a significant improvement in HRQoL and OS from 9 to 13 months only in patients with increased muscle mass (*p*<0.001). Overall, there is a subgroup of good responders to anamorelin that could benefit from this treatment in terms of survival. Another Japanese study, which enrolled 180 patients, found similar results with an improvement in muscle mass and HRQoL, but with no effect on muscle strength and OS (Takayama et al., 2016).

Vitamin D supplementation reduces the risk of falls in elderly patients (Bischoff-Ferrari et al., 2004) because of its strengthening effects on muscle and bone (Arik & Ulger, 2016). A meta-analysis that included 29 randomized clinical trials found a benefit of vitamin D supplementation on muscle strength, without impact on muscle mass (Beaudart et al., 2014). Even if the benefit seems more important for patients > 65 years and deficient in vitamin D, the precise therapeutic role of vitamin D in cancer-related sarcopenia remains to be defined. Considering its favorable tolerance profile, it would be interesting to combine it with other agents for synergistic effect.

7.2.2.2 Other therapies

As mentioned above, pro-inflammatory cytokines (*e.g.* IL-1, IL-6, TNF-alpha) are involved in muscle wasting in cancer-related sarcopenia. Therefore, counteracting their effects is a relevant strategy to reverse sarcopenia. Infliximab, an anti-TNF agent, has been shown to reverse sarcopenia in inflammatory bowel disease such as Crohn's disease (Subramaniam et al., 2015). However, in a phase II study of 89 patients with pancreatic cancer and cachexia, infliximab failed to significantly improve LBM when compared to placebo (Wiedenmann et al., 2008) . Besides TNF, other cytokines may be more attractive targets. Targeting IL-6 with tocilizumab, an anti-IL-6 receptor antibody, was demonstrated to be effective in a patient with lung cancer cachexia by reversing weight loss, inflammation, and altered PS (Ando et al., 2013). Furthermore, blocking IL-1 pathway with MABp1 has led to a statistical increase of LBM ($p=0.02$) with limited adverse events in a phase I study of 30 metastatic cancer patients (Hong et al., 2014). Randomized clinical trials are warranted to demonstrate the benefit of targeting IL-6 and IL-1 pathways in cancer-related sarcopenia. As an alternative strategy, antagonization of myostatin pathway by targeting activin type II receptor highlighted the potential therapeutic effects in muscle growth in preclinical models (Lach-Trifilieff et al., 2014; Zhou et al., 2010). Clinically, the same positive results (safely increased LBM and improved physical performance) have been found in two phase I studies in cancer patients treated by antimyostatin mAb. (Jameson et al., 2012; Padhi et al., 2014). However, a recent randomized phase II trial in pancreatic cancer patients showed no clinical benefit of antimyostatin mAb, with decreased progression-free survival and OS in patients treated with LY2495655 (Golan et al., 2018). Other cytokines from the TGFβ family may be alternative drug targets.

Other strategies to reduce muscle loss are currently being explored such as mirtazapine, a noradrenergic and specific serotonergic antidepressant (NCT03254173, NCT03283488), and espindolol, a non-selective beta-blocker (Stewart Coats et al., 2016).

Combining physical activity dietary counseling and drugs in a multimodal strategy is expected to be the most effective approach and therefore is recommended (Figure 2). In addition, administration of anticancer drugs must be considered in order to reduce tumor burden and inflammation along with the optimal management of their adverse advents in order to limit

malnutrition and sarcopenia (Yip et al., 2014; Temel et al., 2010). It is therefore necessary to question the risk benefit ratio of each prescription.

8. Conclusions

Overall, body composition has gained increasing attention in oncology practice and research in recent years due to fact that sarcopenia has been revealed as a prognostic and predictive factor across multiple stages and cancer types and responsible for a significant proportion of cancerrelated deaths, increased risk of surgery complications and treatment toxicities, and severe impairment of HRQoL. About 30% of cancer deaths are related to cachexia and sarcopenia (von Haehling & Anker, 2010). Hence, implementing therapeutic strategies to preserve or increase muscle mass in cancer patients, in order to optimize anticancer treatment tolerance and efficacy, is now acknowledged as an essential part of supportive care. Treatment of cancer-related sarcopenia should be multimodal and implemented as early as possible. The assessment of body composition may allow more accurate treatment dose calculation in sarcopenic patients. Prospective studies are needed to evaluate the effect of dose adjustment to LBM for toxicity and for the efficacy outcomes such as response rate and survival. The treatment efficacy may even be improved due to a more favorable toxicity profile and less dose adjustments, leading to a better dose-intensity.

Conflicts of interest

François Goldwasser and Frédéric Pigneur participated to the Fresenius Kabi board of experts. Other authors declare no conflicts of interest.

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REFERENCES

Abbatecola, A. M., Paolisso, G., Fattoretti, P., Evans, W. J., Fiore, V., Dicioccio, L., & Lattanzio, F. (2011). Discovering pathways of sarcopenia in older adults: a role for insulin resistance on mitochondria dysfunction. *The Journal of Nutrition, Health & Aging*, *15*(10), 890–895.

Agrawal, A., Suryakumar, G., & Rathor, R. (2018). Role of defective Ca2+ signaling in skeletal muscle weakness: Pharmacological implications. *Journal of Cell Communication and Signaling*. https://doi.org/10.1007/s12079-018-0477-z

Agrawal, S., Feng, Y., Roy, A., Kollia, G., & Lestini, B. (2016). Nivolumab dose selection: challenges, opportunities, and lessons learned for cancer immunotherapy. *Journal for ImmunoTherapy of Cancer*, *4*(1), 72. https://doi.org/10.1186/s40425-016-0177-2

Agustsson, T., D'souza, M. A., Nowak, G., & Isaksson, B. (2011). Mechanisms for skeletal muscle insulin resistance in patients with pancreatic ductal adenocarcinoma. *Nutrition (Burbank, Los Angeles County, Calif.)*, *27*(7–8), 796–801. https://doi.org/10.1016/j.nut.2010.08.022

Ali, R., Baracos, V. E., Sawyer, M. B., Bianchi, L., Roberts, S., Assenat, E., … Senesse, P. (2016). Lean body mass as an independent determinant of dose-limiting toxicity and neuropathy in patients with colon cancer treated with FOLFOX regimens. *Cancer Medicine*, *5*(4), 607–616. https://doi.org/10.1002/cam4.621

Amini, N., Spolverato, G., Gupta, R., Margonis, G. A., Kim, Y., Wagner, D., … Pawlik, T. M. (2015). Impact Total Psoas Volume on Short- and Long-Term Outcomes in Patients Undergoing Curative Resection for Pancreatic Adenocarcinoma: a New Tool to Assess Sarcopenia. *Journal of Gastrointestinal Surgery: Official Journal of the Society for Surgery of the Alimentary Tract*, *19*(9), 1593–1602. https://doi.org/10.1007/s11605-015-2835-y

Anandavadivelan, P., Brismar, T. B., Nilsson, M., Johar, A. M., & Martin, L. (2016). Sarcopenic obesity: A probable risk factor for dose limiting toxicity during neo-adjuvant chemotherapy in oesophageal cancer patients. *Clinical Nutrition (Edinburgh, Scotland)*, *35*(3), 724–730. https://doi.org/10.1016/j.clnu.2015.05.011

Ando, K., Takahashi, F., Motojima, S., Nakashima, K., Kaneko, N., Hoshi, K., & Takahashi, K. (2013). Possible role for tocilizumab, an anti-interleukin-6 receptor antibody, in treating cancer cachexia. *Journal of Clinical Oncology: Official Journal of the American Society of Clinical Oncology*, *31*(6), e69- 72. https://doi.org/10.1200/JCO.2012.44.2020

Antoun, S., Baracos, V. E., Birdsell, L., Escudier, B., & Sawyer, M. B. (2010). Low body mass index and sarcopenia associated with dose-limiting toxicity of sorafenib in patients with renal cell carcinoma. *Annals of Oncology: Official Journal of the European Society for Medical Oncology*, *21*(8), 1594–1598. https://doi.org/10.1093/annonc/mdp605

Antoun, Sami, Lanoy, E., Iacovelli, R., Albiges-Sauvin, L., Loriot, Y., Merad-Taoufik, M., … Escudier, B. (2013). Skeletal muscle density predicts prognosis in patients with metastatic renal cell carcinoma treated with targeted therapies. *Cancer*, *119*(18), 3377–3384. https://doi.org/10.1002/cncr.28218

Argilés, J. M., Busquets, S., López-Soriano, F. J., Costelli, P., & Penna, F. (2012). Are there any benefits of exercise training in cancer cachexia? *Journal of Cachexia, Sarcopenia and Muscle*, *3*(2), 73–76. https://doi.org/10.1007/s13539-012-0067-5

Argilés, J. M., Campos, N., Lopez-Pedrosa, J. M., Rueda, R., & Rodriguez-Mañas, L. (2016). Skeletal Muscle Regulates Metabolism via Interorgan Crosstalk: Roles in Health and Disease. *Journal of the American Medical Directors Association*, *17*(9), 789–796. https://doi.org/10.1016/j.jamda.2016.04.019

Arik, G., & Ulger, Z. (2016). Vitamin D in sarcopenia: Understanding its role in pathogenesis, prevention and treatment. *European Geriatric Medicine*, *7*(3), 207–213. https://doi.org/10.1016/j.eurger.2015.12.001

Arrieta, O., De la Torre-Vallejo, M., López-Macías, D., Orta, D., Turcott, J., Macedo-Pérez, E.-O., … Baracos, V. E. (2015). Nutritional Status, Body Surface, and Low Lean Body Mass/Body Mass Index Are Related to Dose Reduction and Severe Gastrointestinal Toxicity Induced by Afatinib in Patients With Non-Small Cell Lung Cancer. *The Oncologist*, *20*(8), 967–974. https://doi.org/10.1634/theoncologist.2015-0058

Arrieta, O., Michel Ortega, R. M., Villanueva-Rodríguez, G., Serna-Thomé, M. G., Flores-Estrada, D., Diaz-Romero, C., … Sánchez-Lara, K. (2010). Association of nutritional status and serum albumin levels with development of toxicity in patients with advanced non-small cell lung cancer treated with paclitaxel-cisplatin chemotherapy: a prospective study. *BMC Cancer*, *10*, 50. https://doi.org/10.1186/1471-2407-10-50

Ashcraft, K. A., Peace, R. M., Betof, A. S., Dewhirst, M. W., & Jones, L. W. (2016). Efficacy and Mechanisms of Aerobic Exercise on Cancer Initiation, Progression, and Metastasis: A Critical Systematic Review of In Vivo Preclinical Data. *Cancer Research*, *76*(14), 4032–4050. https://doi.org/10.1158/0008-5472.CAN-16-0887

Atherton, P. J., Etheridge, T., Watt, P. W., Wilkinson, D., Selby, A., Rankin, D., … Rennie, M. J. (2010). Muscle full effect after oral protein: time-dependent concordance and discordance between human muscle protein synthesis and mTORC1 signaling. *The American Journal of Clinical Nutrition*, *92*(5), 1080–1088. https://doi.org/10.3945/ajcn.2010.29819

Atkinson, T. M., Andreotti, C. F., Roberts, K. E., Saracino, R. M., Hernandez, M., & Basch, E. (2015). The level of association between functional performance status measures and patient-reported outcomes in cancer patients: a systematic review. *Supportive Care in Cancer*, *23*(12), 3645–3652. https://doi.org/10.1007/s00520-015-2923-2

Awad, S., Tan, B. H., Cui, H., Bhalla, A., Fearon, K. C. H., Parsons, S. L., … Lobo, D. N. (2012). Marked changes in body composition following neoadjuvant chemotherapy for oesophagogastric cancer. *Clinical Nutrition (Edinburgh, Scotland)*, *31*(1), 74–77. https://doi.org/10.1016/j.clnu.2011.08.008

Bahat, G., Tufan, A., Tufan, F., Kilic, C., Akpinar, T. S., Kose, M., … Cruz-Jentoft, A. J. (2016). Cut-off points to identify sarcopenia according to European Working Group on Sarcopenia in Older People (EWGSOP) definition. *Clinical Nutrition (Edinburgh, Scotland)*, *35*(6), 1557–1563. https://doi.org/10.1016/j.clnu.2016.02.002

Baker, S. D., Grochow, L. B., & Donehower, R. C. (1995). Should anticancer drug doses be adjusted in the obese patient? *Journal of the National Cancer Institute*, *87*(5), 333–334.

Bakhshi, V., Elliott, M., Gentili, A., Godschalk, M., & Mulligan, T. (2000). Testosterone improves rehabilitation outcomes in ill older men. *Journal of the American Geriatrics Society*, *48*(5), 550–553.

Ballet, F., Vrignaud, P., Robert, J., Rey, C., & Poupon, R. (1987). Hepatic extraction, metabolism and biliary excretion of doxorubicin in the isolated perfused rat liver. *Cancer Chemotherapy and Pharmacology*, *19*(3), 240–245.

Bano, G., Trevisan, C., Carraro, S., Solmi, M., Luchini, C., Stubbs, B., … Veronese, N. (2017). Inflammation and sarcopenia: A systematic review and meta-analysis. *Maturitas*, *96*, 10–15. https://doi.org/10.1016/j.maturitas.2016.11.006

Baracos, V. E. (2017). Psoas as a sentinel muscle for sarcopenia: a flawed premise. *Journal of Cachexia, Sarcopenia and Muscle*, *8*(4), 527–528. https://doi.org/10.1002/jcsm.12221

Baracos, V. E., & Arribas, L. (2018). Sarcopenic obesity: Hidden muscle wasting and its impact for survival and complications of cancer therapy. *Annals of Oncology*. https://doi.org/10.1093/annonc/mdx810

Baracos, V. E., Reiman, T., Mourtzakis, M., Gioulbasanis, I., & Antoun, S. (2010). Body composition in patients with non-small cell lung cancer: a contemporary view of cancer cachexia with the use of computed tomography image analysis. *The American Journal of Clinical Nutrition*, *91*(4), 1133S-1137S. https://doi.org/10.3945/ajcn.2010.28608C

Barret, M., Antoun, S., Dalban, C., Malka, D., Mansourbakht, T., Zaanan, A., … Taieb, J. (2014). Sarcopenia is linked to treatment toxicity in patients with metastatic colorectal cancer. *Nutrition and Cancer*, *66*(4), 583–589. https://doi.org/10.1080/01635581.2014.894103

Baumgartner, R. N., Koehler, K. M., Gallagher, D., Romero, L., Heymsfield, S. B., Ross, R. R., … Lindeman, R. D. (1998). Epidemiology of sarcopenia among the elderly in New Mexico. *American Journal of Epidemiology*, *147*(8), 755–763.

Baxmann, A. C., Ahmed, M. S., Marques, N. C., Menon, V. B., Pereira, A. B., Kirsztajn, G. M., & Heilberg, I. P. (2008). Influence of muscle mass and physical activity on serum and urinary creatinine and serum cystatin C. *Clinical Journal of the American Society of Nephrology: CJASN*, *3*(2), 348–354. https://doi.org/10.2215/CJN.02870707

Beaudart, C., Buckinx, F., Rabenda, V., Gillain, S., Cavalier, E., Slomian, J., … Bruyère, O. (2014). The effects of vitamin D on skeletal muscle strength, muscle mass, and muscle power: a systematic review and meta-analysis of randomized controlled trials. *The Journal of Clinical Endocrinology and Metabolism*, *99*(11), 4336–4345. https://doi.org/10.1210/jc.2014-1742

Berkel, A. E. M., Klaase, J. M., de Graaff, F., Brusse-Keizer, M. G. J., Bongers, B. C., & van Meeteren, N. L. U. (2018). Patient's Skeletal Muscle Radiation Attenuation and Sarcopenic Obesity are Associated with Postoperative Morbidity after Neoadjuvant Chemoradiation and Resection for Rectal Cancer. *Digestive Surgery*, 1–8. https://doi.org/10.1159/000490069

Bijlsma, A. Y., Meskers, C. G. M., Ling, C. H. Y., Narici, M., Kurrle, S. E., Cameron, I. D., … Maier, A. B. (2013). Defining sarcopenia: the impact of different diagnostic criteria on the prevalence of sarcopenia in a large middle aged cohort. *Age (Dordrecht, Netherlands)*, *35*(3), 871–881. https://doi.org/10.1007/s11357-012-9384-z

Bischoff-Ferrari, H. A., Dawson-Hughes, B., Willett, W. C., Staehelin, H. B., Bazemore, M. G., Zee, R. Y., & Wong, J. B. (2004). Effect of Vitamin D on falls: a meta-analysis. *JAMA*, *291*(16), 1999–2006. https://doi.org/10.1001/jama.291.16.1999

Blauwhoff-Buskermolen, S., Versteeg, K. S., de van der Schueren, M. A. E., den Braver, N. R., Berkhof, J., Langius, J. A. E., & Verheul, H. M. W. (2016). Loss of Muscle Mass During Chemotherapy Is Predictive for Poor Survival of Patients With Metastatic Colorectal Cancer. *Journal of Clinical Oncology*, *34*(12), 1339–1344. https://doi.org/10.1200/JCO.2015.63.6043

Boer, P. (1984). Estimated lean body mass as an index for normalization of body fluid volumes in humans. *The American Journal of Physiology*, *247*(4 Pt 2), F632-636. https://doi.org/10.1152/ajprenal.1984.247.4.F632

Bonekamp, S., Ghosh, P., Crawford, S., Solga, S. F., Horska, A., Brancati, F. L., … Clark, J. M. (2008). Quantitative comparison and evaluation of software packages for assessment of abdominal adipose tissue distribution by magnetic resonance imaging. *International Journal of Obesity*, *32*(1), 100–111. https://doi.org/10.1038/sj.ijo.0803696

Borga, M. (2018). MRI adipose tissue and muscle composition analysis-a review of automation techniques. *The British Journal of Radiology*, *91*(1089), 20180252. https://doi.org/10.1259/bjr.20180252

Bozzetti, F. (2017). Forcing the vicious circle: sarcopenia increases toxicity, decreases response to chemotherapy and worsens with chemotherapy. *Annals of Oncology: Official Journal of the European Society for Medical Oncology*, *28*(9), 2107–2118. https://doi.org/10.1093/annonc/mdx271

Brahmer, J., Reckamp, K. L., Baas, P., Crinò, L., Eberhardt, W. E. E., Poddubskaya, E., … Spigel, D. R. (2015). Nivolumab versus Docetaxel in Advanced Squamous-Cell Non–Small-Cell Lung Cancer. *New England Journal of Medicine*, *373*(2), 123–135. https://doi.org/10.1056/NEJMoa1504627

Bredella, M. A. (2017). Sex Differences in Body Composition. *Advances in Experimental Medicine and Biology*, *1043*, 9–27. https://doi.org/10.1007/978-3-319-70178-3_2

Bretagne, M., Jouinot, A., Durand, J. P., Huillard, O., Boudou Rouquette, P., Tlemsani, C., … Alexandre, J. (2017). Estimation of glomerular filtration rate in cancer patients with abnormal body composition and relation with carboplatin toxicity. *Cancer Chemotherapy and Pharmacology*. https://doi.org/10.1007/s00280-017-3326-5

Broughman, J. R., Williams, G. R., Deal, A. M., Yu, H., Nyrop, K. A., Alston, S. M., … Muss, H. B. (2015). Prevalence of sarcopenia in older patients with colorectal cancer. *Journal of Geriatric Oncology*, *6*(6), 442–445. https://doi.org/10.1016/j.jgo.2015.08.005

Buffart, L. M., Galvão, D. A., Brug, J., Chinapaw, M. J. M., & Newton, R. U. (2014). Evidence-based physical activity guidelines for cancer survivors: current guidelines, knowledge gaps and future

research directions. *Cancer Treatment Reviews*, *40*(2), 327–340. https://doi.org/10.1016/j.ctrv.2013.06.007

Bulow, J., Ulijaszek, S. J., & Holm, L. (2018). Rejuvenation of the term Sarcopenia. *Journal of Applied Physiology (Bethesda, Md.: 1985)*. https://doi.org/10.1152/japplphysiol.00400.2018

Caan, B. J., Meyerhardt, J. A., Kroenke, C. H., Alexeeff, S., Xiao, J., Weltzien, E., … Prado, C. M. (2017). Explaining the Obesity Paradox: The Association between Body Composition and Colorectal Cancer Survival (C-SCANS Study). *Cancer Epidemiology, Biomarkers & Prevention: A Publication of the American Association for Cancer Research, Cosponsored by the American Society of Preventive Oncology*, *26*(7), 1008–1015. https://doi.org/10.1158/1055-9965.EPI-17-0200

Cadore, E. L., & Izquierdo, M. (2013). New strategies for the concurrent strength-, power-, and endurance-training prescription in elderly individuals. *Journal of the American Medical Directors Association*, *14*(8), 623–624. https://doi.org/10.1016/j.jamda.2013.04.008

Camacho, L. H., Antonia, S., Sosman, J., Kirkwood, J. M., Gajewski, T. F., Redman, B., … Ribas, A. (2009). Phase I/II Trial of Tremelimumab in Patients With Metastatic Melanoma. *Journal of Clinical Oncology*, *27*(7), 1075–1081. https://doi.org/10.1200/JCO.2008.19.2435

Camus, V., Lanic, H., Kraut, J., Modzelewski, R., Clatot, F., Picquenot, J. M., … Jardin, F. (2014). Prognostic impact of fat tissue loss and cachexia assessed by computed tomography scan in elderly patients with diffuse large B-cell lymphoma treated with immunochemotherapy. *European Journal of Haematology*, *93*(1), 9–18. https://doi.org/10.1111/ejh.12285

Cancello, R., & Clément, K. (2006). Is obesity an inflammatory illness? Role of low-grade inflammation and macrophage infiltration in human white adipose tissue. *BJOG: An International Journal of Obstetrics and Gynaecology*, *113*(10), 1141–1147. https://doi.org/10.1111/j.1471- 0528.2006.01004.x

Cesari, M., Fielding, R. A., Pahor, M., Goodpaster, B., Hellerstein, M., van Kan, G. A., … International Working Group on Sarcopenia. (2012). Biomarkers of sarcopenia in clinical trials-recommendations from the International Working Group on Sarcopenia. *Journal of Cachexia, Sarcopenia and Muscle*, *3*(3), 181–190. https://doi.org/10.1007/s13539-012-0078-2

Cespedes Feliciano, E. M., Lee, V. S., Prado, C. M., Meyerhardt, J. A., Alexeeff, S., Kroenke, C. H., … Caan, B. J. (2017). Muscle mass at the time of diagnosis of nonmetastatic colon cancer and early discontinuation of chemotherapy, delays, and dose reductions on adjuvant FOLFOX: The C-SCANS study. *Cancer*, *123*(24), 4868–4877. https://doi.org/10.1002/cncr.30950

Charles, K. A., Rivory, L. P., Brown, S. L., Liddle, C., Clarke, S. J., & Robertson, G. R. (2006). Transcriptional repression of hepatic cytochrome P450 3A4 gene in the presence of cancer. *Clinical Cancer Research: An Official Journal of the American Association for Cancer Research*, *12*(24), 7492– 7497. https://doi.org/10.1158/1078-0432.CCR-06-0023

Chemama, S., Bayar, M. A., Lanoy, E., Ammari, S., Stoclin, A., Goéré, D., … Antoun, S. (2016). Sarcopenia is Associated with Chemotherapy Toxicity in Patients Undergoing Cytoreductive Surgery with Hyperthermic Intraperitoneal Chemotherapy for Peritoneal Carcinomatosis from Colorectal Cancer. *Annals of Surgical Oncology*, *23*(12), 3891–3898. https://doi.org/10.1245/s10434-016-5360- 7

Choi, M. H., Oh, S. N., Lee, I. K., Oh, S. T., & Won, D. D. (2018). Sarcopenia is negatively associated with long-term outcomes in locally advanced rectal cancer. *Journal of Cachexia, Sarcopenia and Muscle*, *9*(1), 53–59. https://doi.org/10.1002/jcsm.12234

Choi, Y., Oh, D.-Y., Kim, T.-Y., Lee, K.-H., Han, S.-W., Im, S.-A., … Bang, Y.-J. (2015). Skeletal Muscle Depletion Predicts the Prognosis of Patients with Advanced Pancreatic Cancer Undergoing Palliative Chemotherapy, Independent of Body Mass Index. *PloS One*, *10*(10), e0139749. https://doi.org/10.1371/journal.pone.0139749

Christmas, P. (2015). Role of Cytochrome P450s in Inflammation. *Advances in Pharmacology (San Diego, Calif.)*, *74*, 163–192. https://doi.org/10.1016/bs.apha.2015.03.005

Cleasby, M. E., Jamieson, P. M., & Atherton, P. J. (2016). Insulin resistance and sarcopenia: mechanistic links between common co-morbidities. *The Journal of Endocrinology*, *229*(2), R67-81. https://doi.org/10.1530/JOE-15-0533

Cohen, S., Nathan, J. A., & Goldberg, A. L. (2015). Muscle wasting in disease: molecular mechanisms and promising therapies. *Nature Reviews. Drug Discovery*, *14*(1), 58–74. https://doi.org/10.1038/nrd4467

Coletti, D. (2018). Chemotherapy-induced muscle wasting: an update. *European Journal of Translational Myology*, *28*(2), 7587. https://doi.org/10.4081/ejtm.2018.7587

Cooper, A. B., Slack, R., Fogelman, D., Holmes, H. M., Petzel, M., Parker, N., … Katz, M. H. G. (2015). Characterization of Anthropometric Changes that Occur During Neoadjuvant Therapy for Potentially Resectable Pancreatic Cancer. *Annals of Surgical Oncology*, *22*(7), 2416–2423. https://doi.org/10.1245/s10434-014-4285-2

Corcoran, M. P., Lamon-Fava, S., & Fielding, R. A. (2007). Skeletal muscle lipid deposition and insulin resistance: effect of dietary fatty acids and exercise. *The American Journal of Clinical Nutrition*, *85*(3), 662–677. https://doi.org/10.1093/ajcn/85.3.662

Cormie, P., Galvão, D. A., Spry, N., Joseph, D., Chee, R., Taaffe, D. R., … Newton, R. U. (2015). Can supervised exercise prevent treatment toxicity in patients with prostate cancer initiating androgendeprivation therapy: a randomised controlled trial. *BJU International*, *115*(2), 256–266. https://doi.org/10.1111/bju.12646

Cornet, M., Lim, C., Salloum, C., Lazzati, A., Compagnon, P., Pascal, G., & Azoulay, D. (2015). Prognostic value of sarcopenia in liver surgery. *Journal of Visceral Surgery*, *152*(5), 297–304. https://doi.org/10.1016/j.jviscsurg.2015.08.001

Cousin, S., Hollebecque, A., Koscielny, S., Mir, O., Varga, A., Baracos, V. E., … Antoun, S. (2014). Low skeletal muscle is associated with toxicity in patients included in phase I trials. *Investigational New Drugs*, *32*(2), 382–387. https://doi.org/10.1007/s10637-013-0053-6

Cramer, J. T., Cruz-Jentoft, A. J., Landi, F., Hickson, M., Zamboni, M., Pereira, S. L., … Mustad, V. A. (2016). Impacts of High-Protein Oral Nutritional Supplements Among Malnourished Men and Women with Sarcopenia: A Multicenter, Randomized, Double-Blinded, Controlled Trial. *Journal of the American Medical Directors Association*, *17*(11), 1044–1055. https://doi.org/10.1016/j.jamda.2016.08.009

Cramp, F., & Byron-Daniel, J. (2012). Exercise for the management of cancer-related fatigue in adults. *The Cochrane Database of Systematic Reviews*, *11*, CD006145. https://doi.org/10.1002/14651858.CD006145.pub3

Crevenna, R., Marosi, C., Schmidinger, M., & Fialka-Moser, V. (2006). Neuromuscular electrical stimulation for a patient with metastatic lung cancer--a case report. *Supportive Care in Cancer: Official Journal of the Multinational Association of Supportive Care in Cancer*, *14*(9), 970–973. https://doi.org/10.1007/s00520-006-0033-x

Cruz-Jentoft, A. J., Baeyens, J. P., Bauer, J. M., Boirie, Y., Cederholm, T., Landi, F., … European Working Group on Sarcopenia in Older People. (2010). Sarcopenia: European consensus on definition and diagnosis: Report of the European Working Group on Sarcopenia in Older People. *Age and Ageing*, *39*(4), 412–423. https://doi.org/10.1093/ageing/afq034

Cruz-Jentoft, A. J., Landi, F., Topinková, E., & Michel, J.-P. (2010). Understanding sarcopenia as a geriatric syndrome. *Current Opinion in Clinical Nutrition and Metabolic Care*, *13*(1), 1–7. https://doi.org/10.1097/MCO.0b013e328333c1c1

Currow, D., Temel, J. S., Abernethy, A., Milanowski, J., Friend, J., & Fearon, K. C. (2017). ROMANA 3: a phase 3 safety extension study of anamorelin in advanced non-small-cell lung cancer (NSCLC) patients with cachexia. *Annals of Oncology: Official Journal of the European Society for Medical Oncology*, *28*(8), 1949–1956. https://doi.org/10.1093/annonc/mdx192

Cushen, S. J., Power, D. G., Murphy, K. P., McDermott, R., Griffin, B. T., Lim, M., … Ryan, A. M. (2016). Impact of body composition parameters on clinical outcomes in patients with metastatic castrateresistant prostate cancer treated with docetaxel. *Clinical Nutrition ESPEN*, *13*, e39–e45. https://doi.org/10.1016/j.clnesp.2016.04.001

Cushen, S. J., Power, D. G., Teo, M. Y., MacEneaney, P., Maher, M. M., McDermott, R., … Ryan, A. M. (2017). Body Composition by Computed Tomography as a Predictor of Toxicity in Patients With Renal Cell Carcinoma Treated With Sunitinib. *American Journal of Clinical Oncology*, *40*(1), 47–52. https://doi.org/10.1097/COC.0000000000000061

Cvan Trobec, K., Kerec Kos, M., Trontelj, J., Grabnar, I., Tschirner, A., Palus, S., … Lainscak, M. (2015). Influence of cancer cachexia on drug liver metabolism and renal elimination in rats: Drug liver metabolism and renal elimination in cancer cachexia. *Journal of Cachexia, Sarcopenia and Muscle*, *6*(1), 45–52. https://doi.org/10.1002/jcsm.12012

Dalal, S., Hui, D., Bidaut, L., Lem, K., Del Fabbro, E., Crane, C., … Bruera, E. (2012). Relationships among body mass index, longitudinal body composition alterations, and survival in patients with locally advanced pancreatic cancer receiving chemoradiation: a pilot study. *Journal of Pain and Symptom Management*, *44*(2), 181–191. https://doi.org/10.1016/j.jpainsymman.2011.09.010

Daly, L. E., Ní Bhuachalla, É. B., Power, D. G., Cushen, S. J., James, K., & Ryan, A. M. (2018). Loss of skeletal muscle during systemic chemotherapy is prognostic of poor survival in patients with foregut cancer. *Journal of Cachexia, Sarcopenia and Muscle*, *9*(2), 315–325. https://doi.org/10.1002/jcsm.12267

Daly, L. E., Power, D. G., O'Reilly, Á., Donnellan, P., Cushen, S. J., O'Sullivan, K., … Ryan, A. M. (2017). The impact of body composition parameters on ipilimumab toxicity and survival in patients with

metastatic melanoma. *British Journal of Cancer*, *116*(3), 310–317. https://doi.org/10.1038/bjc.2016.431

Damrauer, J. S., Stadler, M. E., Acharyya, S., Baldwin, A. S., Couch, M. E., & Guttridge, D. C. (2018). Chemotherapy-induced muscle wasting: association with NF-κB and cancer cachexia. *European Journal of Translational Myology*, *28*(2), 7590. https://doi.org/10.4081/ejtm.2018.7590

Davis, L. E., Lenkinski, R. E., Shinkwin, M. A., Kressel, H. Y., & Daly, J. M. (1993). The effect of dietary protein depletion on hepatic 5-fluorouracil metabolism. *Cancer*, *72*(12), 3715–3722.

de Souza, V. A., Oliveira, D., Barbosa, S. R., Corrêa, J. O. do A., Colugnati, F. A. B., Mansur, H. N., … Bastos, M. G. (2017). Sarcopenia in patients with chronic kidney disease not yet on dialysis: Analysis of the prevalence and associated factors. *PLoS ONE*, *12*(4). https://doi.org/10.1371/journal.pone.0176230

Del Fabbro, E., Parsons, H., Warneke, C. L., Pulivarthi, K., Litton, J. K., Dev, R., … Bruera, E. (2012). The Relationship Between Body Composition and Response to Neoadjuvant Chemotherapy in Women with Operable Breast Cancer. *The Oncologist*, *17*(10), 1240–1245. https://doi.org/10.1634/theoncologist.2012-0169

Demirovic, J. A., Pai, A. B., & Pai, M. P. (2009). Estimation of creatinine clearance in morbidly obese patients. *American Journal of Health-System Pharmacy: AJHP: Official Journal of the American Society of Health-System Pharmacists*, *66*(7), 642–648. https://doi.org/10.2146/ajhp080200

Deutz, N. E. P., Safar, A., Schutzler, S., Memelink, R., Ferrando, A., Spencer, H., … Wolfe, R. R. (2011). Muscle protein synthesis in cancer patients can be stimulated with a specially formulated medical food. *Clinical Nutrition (Edinburgh, Scotland)*, *30*(6), 759–768. https://doi.org/10.1016/j.clnu.2011.05.008

Dhooge, M., Coriat, R., Mir, O., Perkins, G., Brezault, C., Boudou-Rouquette, P., … Chaussade, S. (2013). Feasibility of gemcitabine plus oxaliplatin in advanced hepatocellular carcinoma patients with Child-Pugh B cirrhosis. *Oncology*, *84*(1), 32–38. https://doi.org/10.1159/000342763

Di Sebastiano, K. M., & Mourtzakis, M. (2012). A critical evaluation of body composition modalities used to assess adipose and skeletal muscle tissue in cancer. *Applied Physiology, Nutrition, and Metabolism*, *37*(5), 811–821. https://doi.org/10.1139/h2012-079

Di Sebastiano, K. M., Yang, L., Zbuk, K., Wong, R. K., Chow, T., Koff, D., … Mourtzakis, M. (2013). Accelerated muscle and adipose tissue loss may predict survival in pancreatic cancer patients: the relationship with diabetes and anaemia. *The British Journal of Nutrition*, *109*(2), 302–312. https://doi.org/10.1017/S0007114512001067

Dieli-Conwright, C. M., Courneya, K. S., Demark-Wahnefried, W., Sami, N., Lee, K., Buchanan, T. A., … Mortimer, J. E. (2018). Effects of Aerobic and Resistance Exercise on Metabolic Syndrome, Sarcopenic Obesity, and Circulating Biomarkers in Overweight or Obese Survivors of Breast Cancer: A Randomized Controlled Trial. *Journal of Clinical Oncology*, *36*(9), 875–883. https://doi.org/10.1200/JCO.2017.75.7526

Dignam, J. J., Polite, B. N., Yothers, G., Raich, P., Colangelo, L., O'Connell, M. J., & Wolmark, N. (2006). Body mass index and outcomes in patients who receive adjuvant chemotherapy for colon cancer. *Journal of the National Cancer Institute*, *98*(22), 1647–1654. https://doi.org/10.1093/jnci/djj442

Dixon, W. T. (1984). Simple proton spectroscopic imaging. *Radiology*, *153*(1), 189–194. https://doi.org/10.1148/radiology.153.1.6089263

Dobs, A. S., Boccia, R. V., Croot, C. C., Gabrail, N. Y., Dalton, J. T., Hancock, M. L., … Steiner, M. S. (2013). Effects of enobosarm on muscle wasting and physical function in patients with cancer: a double-blind, randomised controlled phase 2 trial. *The Lancet. Oncology*, *14*(4), 335–345. https://doi.org/10.1016/S1470-2045(13)70055-X

Du Bois, D., & Du Bois, E. F. (1989). A formula to estimate the approximate surface area if height and weight be known. 1916. *Nutrition (Burbank, Los Angeles County, Calif.)*, *5*(5), 303–311; discussion 312-313.

Ellis, K. J. (2001). Selected Body Composition Methods Can Be Used in Field Studies. *The Journal of Nutrition*, *131*(5), 1589S-1595S. https://doi.org/10.1093/jn/131.5.1589S

Ethun, C. G., Bilen, M. A., Jani, A. B., Maithel, S. K., Ogan, K., & Master, V. A. (2017). Frailty and cancer: Implications for oncology surgery, medical oncology, and radiation oncology. *CA: A Cancer Journal for Clinicians*, *67*(5), 362–377. https://doi.org/10.3322/caac.21406

Ezeoke, C. C., & Morley, J. E. (2015). Pathophysiology of anorexia in the cancer cachexia syndrome. *Journal of Cachexia, Sarcopenia and Muscle*, *6*(4), 287–302. https://doi.org/10.1002/jcsm.12059

Falk, A. T., Barrière, J., François, E., & Follana, P. (2015). Bevacizumab: A dose review. *Critical Reviews in Oncology/Hematology*, *94*(3), 311–322. https://doi.org/10.1016/j.critrevonc.2015.01.012

Fang, H., Berg, E., Cheng, X., & Shen, W. (2018). How to best assess abdominal obesity. *Current Opinion in Clinical Nutrition and Metabolic Care*. https://doi.org/10.1097/MCO.0000000000000485

Fearon, K., Strasser, F., Anker, S. D., Bosaeus, I., Bruera, E., Fainsinger, R. L., … Baracos, V. E. (2011). Definition and classification of cancer cachexia: an international consensus. *The Lancet Oncology*, *12*(5), 489–495. https://doi.org/10.1016/S1470-2045(10)70218-7

Feldschuh, J., & Enson, Y. (1977). Prediction of the normal blood volume. Relation of blood volume to body habitus. *Circulation*, *56*(4), 605–612. https://doi.org/10.1161/01.CIR.56.4.605

Feng, Y., Roy, A., Masson, E., Chen, T.-T., Humphrey, R., & Weber, J. S. (2013). Exposure-Response Relationships of the Efficacy and Safety of Ipilimumab in Patients with Advanced Melanoma. *Clinical Cancer Research*, *19*(14), 3977–3986. https://doi.org/10.1158/1078-0432.CCR-12-3243

Ferrando, A. A., Sheffield-Moore, M., Yeckel, C. W., Gilkison, C., Jiang, J., Achacosa, A., … Urban, R. J. (2002). Testosterone administration to older men improves muscle function: molecular and physiological mechanisms. *American Journal of Physiology. Endocrinology and Metabolism*, *282*(3), E601-607. https://doi.org/10.1152/ajpendo.00362.2001

Fielding, R. A., Vellas, B., Evans, W. J., Bhasin, S., Morley, J. E., Newman, A. B., … Zamboni, M. (2011). Sarcopenia: an undiagnosed condition in older adults. Current consensus definition: prevalence, etiology, and consequences. International working group on sarcopenia. *Journal of the American Medical Directors Association*, *12*(4), 249–256. https://doi.org/10.1016/j.jamda.2011.01.003

Flanigen-Roat, D. T., Milholland, R. J., & Ip, M. M. (1985). Effect of a high-protein-low-carbohydrate diet on toxicity and antitumor activity of 5-fluorouracil in mice. *Journal of the National Cancer Institute*, *74*(3), 705–710.

Focht, B. C., Clinton, S. K., Devor, S. T., Garver, M. J., Lucas, A. R., Thomas-Ahner, J. M., & Grainger, E. (2013). Resistance exercise interventions during and following cancer treatment: a systematic review. *The Journal of Supportive Oncology*, *11*(2), 45–60.

Fuchs, G., Chretien, Y. R., Mario, J., Do, S., Eikermann, M., Liu, B., … Fintelmann, F. J. (2018). Quantifying the effect of slice thickness, intravenous contrast and tube current on muscle segmentation: Implications for body composition analysis. *European Radiology*, *28*(6), 2455–2463. https://doi.org/10.1007/s00330-017-5191-3

Fujiwara, N., Nakagawa, H., Kudo, Y., Tateishi, R., Taguri, M., Watadani, T., … Koike, K. (2015). Sarcopenia, intramuscular fat deposition, and visceral adiposity independently predict the outcomes of hepatocellular carcinoma. *Journal of Hepatology*, *63*(1), 131–140. https://doi.org/10.1016/j.jhep.2015.02.031

Fukushima, H., Nakanishi, Y., Kataoka, M., Tobisu, K., & Koga, F. (2016). Prognostic Significance of Sarcopenia in Patients with Metastatic Renal Cell Carcinoma. *The Journal of Urology*, *195*(1), 26–32. https://doi.org/10.1016/j.juro.2015.08.071

Fukushima, H., Yokoyama, M., Nakanishi, Y., Tobisu, K., & Koga, F. (2015). Sarcopenia as a prognostic biomarker of advanced urothelial carcinoma. *PloS One*, *10*(1), e0115895. https://doi.org/10.1371/journal.pone.0115895

Furlanetto, J., Eiermann, W., Marmé, F., Reimer, T., Reinisch, M., Schmatloch, S., … Möbus, V. (2016). Higher rate of severe toxicities in obese patients receiving dose-dense (dd) chemotherapy according to unadjusted body surface area: results of the prospectively randomized GAIN study. *Annals of Oncology: Official Journal of the European Society for Medical Oncology*, *27*(11), 2053–2059. https://doi.org/10.1093/annonc/mdw315

Ghielmini, M. (2005). Multimodality Therapies and Optimal Schedule of Antibodies: Rituximab in Lymphoma as an Example. *ASH Education Program Book*, *2005*(1), 321–328. https://doi.org/10.1182/asheducation-2005.1.321

Go, S.-I., Park, M. J., Song, H.-N., Kang, M. H., Park, H. J., Jeon, K. N., … Lee, G.-W. (2016). Sarcopenia and inflammation are independent predictors of survival in male patients newly diagnosed with small cell lung cancer. *Supportive Care in Cancer*, *24*(5), 2075–2084. https://doi.org/10.1007/s00520- 015-2997-x

Go, S.-I., Park, M. J., Song, H.-N., Kim, H.-G., Kang, M. H., Kang, J. H., … Lee, G.-W. (2017). A comparison of pectoralis versus lumbar skeletal muscle indices for defining sarcopenia in diffuse large B-cell lymphoma - two are better than one. *Oncotarget*, *8*(29), 47007–47019. https://doi.org/10.18632/oncotarget.16552

Golan, T., Geva, R., Richards, D., Madhusudan, S., Lin, B. K., Wang, H. T., … Stemmer, S. M. (2018). LY2495655, an antimyostatin antibody, in pancreatic cancer: a randomized, phase 2 trial: LY2495655 in patients with stage II-IV pancreatic cancer. *Journal of Cachexia, Sarcopenia and Muscle*, *9*(5), 871– 879. https://doi.org/10.1002/jcsm.12331

Gondin, J., Cozzone, P. J., & Bendahan, D. (2011). Is high-frequency neuromuscular electrical stimulation a suitable tool for muscle performance improvement in both healthy humans and athletes? *European Journal of Applied Physiology*, *111*(10), 2473–2487. https://doi.org/10.1007/s00421-011-2101-2

Gonzalez, M. C., Barbosa-Silva, T. G., & Heymsfield, S. B. (2018). Bioelectrical impedance analysis in the assessment of sarcopenia. *Current Opinion in Clinical Nutrition and Metabolic Care*. https://doi.org/10.1097/MCO.0000000000000496

Goodpaster, B. H., Kelley, D. E., Thaete, F. L., He, J., & Ross, R. (2000). Skeletal muscle attenuation determined by computed tomography is associated with skeletal muscle lipid content. *Journal of Applied Physiology (Bethesda, Md.: 1985)*, *89*(1), 104–110. https://doi.org/10.1152/jappl.2000.89.1.104

Gordon, M. S., Margolin, K., Talpaz, M., Sledge, G. W., Holmgren, E., Benjamin, R., … Adelman, D. (2001). Phase I safety and pharmacokinetic study of recombinant human anti-vascular endothelial growth factor in patients with advanced cancer. *Journal of Clinical Oncology: Official Journal of the American Society of Clinical Oncology*, *19*(3), 843–850. https://doi.org/10.1200/JCO.2001.19.3.843

Griggs, J. J., Mangu, P. B., Anderson, H., Balaban, E. P., Dignam, J. J., Hryniuk, W. M., … American Society of Clinical Oncology. (2012). Appropriate chemotherapy dosing for obese adult patients with cancer: American Society of Clinical Oncology clinical practice guideline. *Journal of Clinical Oncology: Official Journal of the American Society of Clinical Oncology*, *30*(13), 1553–1561. https://doi.org/10.1200/JCO.2011.39.9436

Griggs, J. J., Mangu, P. B., Temin, S., & Lyman, G. H. (2012). Appropriate Chemotherapy Dosing for Obese Adult Patients With Cancer: American Society of Clinical Oncology Clinical Practice Guideline. *Journal of Oncology Practice*, *8*(4), e59–e61. https://doi.org/10.1200/JOP.2012.000623

Grotenhuis, B. A., Shapiro, J., van Adrichem, S., de Vries, M., Koek, M., Wijnhoven, B. P. L., & van Lanschot, J. J. B. (2016). Sarcopenia/Muscle Mass is not a Prognostic Factor for Short- and Long-Term Outcome After Esophagectomy for Cancer. *World Journal of Surgery*, *40*(11), 2698–2704. https://doi.org/10.1007/s00268-016-3603-1

Guglielmi, G., Ponti, F., Agostini, M., Amadori, M., Battista, G., & Bazzocchi, A. (2016). The role of DXA in sarcopenia. *Aging Clinical and Experimental Research*, *28*(6), 1047–1060. https://doi.org/10.1007/s40520-016-0589-3

Guillory, B., Splenser, A., & Garcia, J. (2013). The role of ghrelin in anorexia-cachexia syndromes. *Vitamins and Hormones*, *92*, 61–106. https://doi.org/10.1016/B978-0-12-410473-0.00003-9

Guralnik, J. M., Ferrucci, L., Leveille, S. G., Markides, K. S., Ostir, G. V., Studenski, S., … Wallace, R. B. (2000). Lower extremity function and subsequent disability: consistency across studies, predictive models, and value of gait speed alone compared with the short physical performance battery. *The Journals of Gerontology. Series A, Biological Sciences and Medical Sciences*, *55*(4), M221-231.

Gurney, H. (1996). Dose calculation of anticancer drugs: a review of the current practice and introduction of an alternative. *Journal of Clinical Oncology: Official Journal of the American Society of Clinical Oncology*, *14*(9), 2590–2611. https://doi.org/10.1200/JCO.1996.14.9.2590

Gusella, M., Toso, S., Ferrazzi, E., Ferrari, M., & Padrini, R. (2002). Relationships between body composition parameters and fluorouracil pharmacokinetics. *British Journal of Clinical Pharmacology*, *54*(2), 131–139.

Handforth, C., Clegg, A., Young, C., Simpkins, S., Seymour, M. T., Selby, P. J., & Young, J. (2015). The prevalence and outcomes of frailty in older cancer patients: a systematic review. *Annals of Oncology: Official Journal of the European Society for Medical Oncology*, *26*(6), 1091–1101. https://doi.org/10.1093/annonc/mdu540

Harimoto, N., Shirabe, K., Yamashita, Y.-I., Ikegami, T., Yoshizumi, T., Soejima, Y., … Yamanaka, T. (2013). Sarcopenia as a predictor of prognosis in patients following hepatectomy for hepatocellular carcinoma. *The British Journal of Surgery*, *100*(11), 1523–1530. https://doi.org/10.1002/bjs.9258

He, W. A., Berardi, E., Cardillo, V. M., Acharyya, S., Aulino, P., Thomas-Ahner, J., … Guttridge, D. C. (2013). NF-κB-mediated Pax7 dysregulation in the muscle microenvironment promotes cancer cachexia. *The Journal of Clinical Investigation*, *123*(11), 4821–4835. https://doi.org/10.1172/JCI68523

Heidelberger, H., François, G., Nora, K., Anne, J., Olivier, H., Pascaline, B.-R., … Sélim, A. (2017). Sarcopenic overweight is associated with early acute limiting toxicity of anti-PD1 checkpoint inhibitors in melanoma patients. *Investigational New Drugs*. https://doi.org/10.1007/s10637-017- 0464-x

Hendrikx, J. J. M. A., Haanen, J. B. A. G., Voest, E. E., Schellens, J. H. M., Huitema, A. D. R., & Beijnen, J. H. (2017). Fixed Dosing of Monoclonal Antibodies in Oncology. *The Oncologist*, *22*(10), 1212–1221. https://doi.org/10.1634/theoncologist.2017-0167

Heymsfield, S. B. (2008). Development of imaging methods to assess adiposity and metabolism. *International Journal of Obesity (2005)*, *32 Suppl 7*, S76-82. https://doi.org/10.1038/ijo.2008.242

Hill, A., Kiss, N., Hodgson, B., Crowe, T. C., & Walsh, A. D. (2011). Associations between nutritional status, weight loss, radiotherapy treatment toxicity and treatment outcomes in gastrointestinal cancer patients. *Clinical Nutrition (Edinburgh, Scotland)*, *30*(1), 92–98. https://doi.org/10.1016/j.clnu.2010.07.015

Hodi, F. S., O'Day, S. J., McDermott, D. F., Weber, R. W., Sosman, J. A., Haanen, J. B., … Urba, W. J. (2010). Improved Survival with Ipilimumab in Patients with Metastatic Melanoma. *New England Journal of Medicine*, *363*(8), 711–723. https://doi.org/10.1056/NEJMoa1003466

Hojan, K., Milecki, P., Molińska-Glura, M., Roszak, A., & Leszczyński, P. (2013). Effect of physical activity on bone strength and body composition in breast cancer premenopausal women during endocrine therapy. *European Journal of Physical and Rehabilitation Medicine*, *49*(3), 331–339.

Hojman, P., Fjelbye, J., Zerahn, B., Christensen, J. F., Dethlefsen, C., Lonkvist, C. K., … Gehl, J. (2014). Voluntary exercise prevents cisplatin-induced muscle wasting during chemotherapy in mice. *PloS One*, *9*(9), e109030. https://doi.org/10.1371/journal.pone.0109030

Hong, D. S., Hui, D., Bruera, E., Janku, F., Naing, A., Falchook, G. S., … Kurzrock, R. (2014). MABp1, a first-in-class true human antibody targeting interleukin-1α in refractory cancers: an open-label,

phase 1 dose-escalation and expansion study. *The Lancet. Oncology*, *15*(6), 656–666. https://doi.org/10.1016/S1470-2045(14)70155-X

Huang, D.-D., Wang, S.-L., Zhuang, C.-L., Zheng, B.-S., Lu, J.-X., Chen, F.-F., … Yu, Z. (2015). Sarcopenia, as defined by low muscle mass, strength and physical performance, predicts complications after surgery for colorectal cancer. *Colorectal Disease: The Official Journal of the Association of Coloproctology of Great Britain and Ireland*, *17*(11), O256-264. https://doi.org/10.1111/codi.13067

Huillard, O., Mir, O., Peyromaure, M., Tlemsani, C., Giroux, J., Boudou-Rouquette, P., … Goldwasser, F. (2013). Sarcopenia and body mass index predict sunitinib-induced early dose-limiting toxicities in renal cancer patients. *British Journal of Cancer*, *108*(5), 1034–1041. https://doi.org/10.1038/bjc.2013.58

Hunter, R. J., Navo, M. A., Thaker, P. H., Bodurka, D. C., Wolf, J. K., & Smith, J. A. (2009). Dosing chemotherapy in obese patients: actual versus assigned body surface area (BSA). *Cancer Treatment Reviews*, *35*(1), 69–78. https://doi.org/10.1016/j.ctrv.2008.07.005

Iannessi, A., Beaumont, H., Hebert, C., Dittlot, C., & Falewee, M. N. (2018). Computer tomographybased body surface area evaluation for drug dosage: Quantitative radiology versus anthropomorphic evaluation. *PLoS ONE*, *13*(2), 1–12. https://doi.org/10.1371/journal.pone.0192124

Ida, S., Watanabe, M., Yoshida, N., Baba, Y., Umezaki, N., Harada, K., … Baba, H. (2015). Sarcopenia is a Predictor of Postoperative Respiratory Complications in Patients with Esophageal Cancer. *Annals of Surgical Oncology*, *22*(13), 4432–4437. https://doi.org/10.1245/s10434-015-4559-3

Iritani, S., Imai, K., Takai, K., Hanai, T., Ideta, T., Miyazaki, T., … Moriwaki, H. (2015). Skeletal muscle depletion is an independent prognostic factor for hepatocellular carcinoma. *Journal of Gastroenterology*, *50*(3), 323–332. https://doi.org/10.1007/s00535-014-0964-9

Ishihara, H., Kondo, T., Omae, K., Takagi, T., Iizuka, J., Kobayashi, H., & Tanabe, K. (2016). Sarcopenia and the Modified Glasgow Prognostic Score are Significant Predictors of Survival Among Patients with Metastatic Renal Cell Carcinoma Who are Receiving First-Line Sunitinib Treatment. *Targeted Oncology*, *11*(5), 605–617. https://doi.org/10.1007/s11523-016-0430-0

Ishikawa, T., Yasuda, T., Doi, T., Okayama, T., Sakamoto, N., Gen, Y., … Itoh, Y. (2016). The amino acid-rich elemental diet Elental® preserves lean body mass during chemo- or chemoradiotherapy for esophageal cancer. *Oncology Reports*, *36*(2), 1093–1100. https://doi.org/10.3892/or.2016.4877

Jameson, G. S., Von Hoff, D. D., Weiss, G. J., Richards, D. A., Smith, D. A., Becerra, C., … Lin, B. K. (2012). Safety of the antimyostatin monoclonal antibody LY2495655 in healthy subjects and patients with advanced cancer. *Journal of Clinical Oncology*, *30*(15_suppl), 2516–2516. https://doi.org/10.1200/jco.2012.30.15_suppl.2516

Janssen, I., & Ross, R. (2005). Linking age-related changes in skeletal muscle mass and composition with metabolism and disease. *The Journal of Nutrition, Health & Aging*, *9*(6), 408–419.

Joglekar, S., Asghar, A., Mott, S. L., Johnson, B. E., Button, A. M., Clark, E., & Mezhir, J. J. (2015). Sarcopenia is an independent predictor of complications following pancreatectomy for adenocarcinoma. *Journal of Surgical Oncology*, *111*(6), 771–775. https://doi.org/10.1002/jso.23862 Jouinot, A., Vazeille, C., & Goldwasser, F. (2018). Resting energy metabolism and anticancer treatments. *Current Opinion in Clinical Nutrition and Metabolic Care*, *21*(3), 145–151. https://doi.org/10.1097/MCO.0000000000000457

Jung, H.-W., Kim, J. W., Kim, J.-Y., Kim, S.-W., Yang, H. K., Lee, J. W., … Kim, J. H. (2015). Effect of muscle mass on toxicity and survival in patients with colon cancer undergoing adjuvant chemotherapy. *Supportive Care in Cancer: Official Journal of the Multinational Association of Supportive Care in Cancer*, *23*(3), 687–694. https://doi.org/10.1007/s00520-014-2418-6

Kamachi, S., Mizuta, T., Otsuka, T., Nakashita, S., Ide, Y., Miyoshi, A., … Anzai, K. (2016). Sarcopenia is a risk factor for the recurrence of hepatocellular carcinoma after curative treatment. *Hepatology Research: The Official Journal of the Japan Society of Hepatology*, *46*(2), 201–208. https://doi.org/10.1111/hepr.12562

Kays, J. K., Shahda, S., Stanley, M., Bell, T. M., O'Neill, B. H., Kohli, M. D., … Zimmers, T. A. (2018). Three cachexia phenotypes and the impact of fat-only loss on survival in FOLFIRINOX therapy for pancreatic cancer. *Journal of Cachexia, Sarcopenia and Muscle*. https://doi.org/10.1002/jcsm.12307

Kazemi-Bajestani, S. M. R., Mazurak, V. C., & Baracos, V. (2016). Computed tomography-defined muscle and fat wasting are associated with cancer clinical outcomes. *Seminars in Cell & Developmental Biology*, *54*, 2–10. https://doi.org/10.1016/j.semcdb.2015.09.001

Keizer, R. J., Huitema, A. D. R., Schellens, J. H. M., & Beijnen, J. H. (2010). Clinical pharmacokinetics of therapeutic monoclonal antibodies. *Clinical Pharmacokinetics*, *49*(8), 493–507. https://doi.org/10.2165/11531280-000000000-00000

Kilgour, R. D., Vigano, A., Trutschnigg, B., Lucar, E., Borod, M., & Morais, J. A. (2013). Handgrip strength predicts survival and is associated with markers of clinical and functional outcomes in advanced cancer patients. *Supportive Care in Cancer: Official Journal of the Multinational Association of Supportive Care in Cancer*, *21*(12), 3261–3270. https://doi.org/10.1007/s00520-013-1894-4

Kim, E. Y., Kim, Y. S., Park, I., Ahn, H. K., Cho, E. K., & Jeong, Y. M. (2015). Prognostic Significance of CT-Determined Sarcopenia in Patients with Small-Cell Lung Cancer. *Journal of Thoracic Oncology: Official Publication of the International Association for the Study of Lung Cancer*, *10*(12), 1795–1799. https://doi.org/10.1097/JTO.0000000000000690

Kim, Y. S., Kim, E. Y., Kang, S. M., Ahn, H. K., & Kim, H. S. (2017). Single cross-sectional area of pectoralis muscle by computed tomography - correlation with bioelectrical impedance based skeletal muscle mass in healthy subjects. *Clinical Physiology and Functional Imaging*, *37*(5), 507–511. https://doi.org/10.1111/cpf.12333

Klein, G. L., Petschow, B. W., Shaw, A. L., & Weaver, E. (2013). Gut barrier dysfunction and microbial translocation in cancer cachexia: a new therapeutic target. *Current Opinion in Supportive and Palliative Care*, *7*(4), 361–367. https://doi.org/10.1097/SPC.0000000000000017

Kobayashi, A., Kaido, T., Hamaguchi, Y., Okumura, S., Shirai, H., Yao, S., … Uemoto, S. (2017). Impact of Sarcopenic Obesity on Outcomes in Patients Undergoing Hepatectomy for Hepatocellular Carcinoma. *Annals of Surgery*. https://doi.org/10.1097/SLA.0000000000002555

Koretz, R. L., Lipman, T. O., Klein, S., & American Gastroenterological Association. (2001). AGA technical review on parenteral nutrition. *Gastroenterology*, *121*(4), 970–1001.

Krell, R. W., Kaul, D. R., Martin, A. R., Englesbe, M. J., Sonnenday, C. J., Cai, S., & Malani, P. N. (2013). Association between sarcopenia and the risk of serious infection among adults undergoing liver transplantation. *Liver Transplantation: Official Publication of the American Association for the Study of Liver Diseases and the International Liver Transplantation Society*, *19*(12), 1396–1402. https://doi.org/10.1002/lt.23752

Kumar Pal, S., Katheria, V., & Hurria, A. (2010). Evaluating the Older Patient with Cancer: Understanding Frailty and the Geriatric Assessment. *CA: A Cancer Journal for Clinicians*, *60*(2), 120– 132. https://doi.org/10.3322/caac.20059

La Colla, L., La Colla, G., Albertin, A., Poli, D., Baruffaldi Preis, F. W., & Mangano, A. (2007). The use of propofol and remifentanil for the anaesthetic management of a super-obese patient. *Anaesthesia*, *62*(8), 842–845. https://doi.org/10.1111/j.1365-2044.2007.05102.x

Lach-Trifilieff, E., Minetti, G. C., Sheppard, K., Ibebunjo, C., Feige, J. N., Hartmann, S., … Glass, D. J. (2014). An antibody blocking activin type II receptors induces strong skeletal muscle hypertrophy and protects from atrophy. *Molecular and Cellular Biology*, *34*(4), 606–618. https://doi.org/10.1128/MCB.01307-13

Lanic, H., Kraut-Tauzia, J., Modzelewski, R., Clatot, F., Mareschal, S., Picquenot, J. M., … Jardin, F. (2014). Sarcopenia is an independent prognostic factor in elderly patients with diffuse large B-cell lymphoma treated with immunochemotherapy. *Leukemia & Lymphoma*, *55*(4), 817–823. https://doi.org/10.3109/10428194.2013.816421

Lau, E. M. C., Lynn, H. S. H., Woo, J. W., Kwok, T. C. Y., & Melton, L. J. (2005). Prevalence of and risk factors for sarcopenia in elderly Chinese men and women. *The Journals of Gerontology. Series A, Biological Sciences and Medical Sciences*, *60*(2), 213–216.

Lee, J., Chang, C.-L., Lin, J.-B., Wu, M.-H., Sun, F.-J., Jan, Y.-T., … Chen, Y.-J. (2018). Skeletal Muscle Loss is an Imaging Biomarker of Outcome After Definitive Chemoradiotherapy for Locally Advanced Cervical Cancer. *Clinical Cancer Research: An Official Journal of the American Association for Cancer Research*. https://doi.org/10.1158/1078-0432.CCR-18-0788

Leenders, M., Verdijk, L. B., van der Hoeven, L., Adam, J. J., van Kranenburg, J., Nilwik, R., & van Loon, L. J. C. (2013). Patients with type 2 diabetes show a greater decline in muscle mass, muscle strength, and functional capacity with aging. *Journal of the American Medical Directors Association*, *14*(8), 585–592. https://doi.org/10.1016/j.jamda.2013.02.006

Levolger, S., van Vledder, M. G., Muslem, R., Koek, M., Niessen, W. J., de Man, R. A., … Ijzermans, J. N. M. (2015). Sarcopenia impairs survival in patients with potentially curable hepatocellular carcinoma. *Journal of Surgical Oncology*, *112*(2), 208–213. https://doi.org/10.1002/jso.23976

Lieffers, J. R., Bathe, O. F., Fassbender, K., Winget, M., & Baracos, V. E. (2012). Sarcopenia is associated with postoperative infection and delayed recovery from colorectal cancer resection surgery. *British Journal of Cancer*, *107*(6), 931–936. https://doi.org/10.1038/bjc.2012.350

Lodewick, T. M., van Nijnatten, T. J. A., van Dam, R. M., van Mierlo, K., Dello, S. A. W. G., Neumann, U. P., … Dejong, C. H. C. (2015). Are sarcopenia, obesity and sarcopenic obesity predictive of outcome in patients with colorectal liver metastases? *HPB: The Official Journal of the International Hepato Pancreato Biliary Association*, *17*(5), 438–446. https://doi.org/10.1111/hpb.12373

Looijaard, W. G. P. M., Dekker, I. M., Stapel, S. N., Girbes, A. R. J., Twisk, J. W. R., Oudemans-van Straaten, H. M., & Weijs, P. J. M. (2016). Skeletal muscle quality as assessed by CT-derived skeletal muscle density is associated with 6-month mortality in mechanically ventilated critically ill patients. *Critical Care (London, England)*, *20*(1), 386. https://doi.org/10.1186/s13054-016-1563-3

Lou, N., Chi, C.-H., Chen, X.-D., Zhou, C.-J., Wang, S.-L., Zhuang, C.-L., & Shen, X. (2017). Sarcopenia in overweight and obese patients is a predictive factor for postoperative complication in gastric cancer: A prospective study. *European Journal of Surgical Oncology: The Journal of the European Society of Surgical Oncology and the British Association of Surgical Oncology*, *43*(1), 188–195. https://doi.org/10.1016/j.ejso.2016.09.006

Maddocks, M., Halliday, V., Chauhan, A., Taylor, V., Nelson, A., Sampson, C., … Wilcock, A. (2013). Neuromuscular electrical stimulation of the quadriceps in patients with non-small cell lung cancer receiving palliative chemotherapy: a randomized phase II study. *PloS One*, *8*(12), e86059. https://doi.org/10.1371/journal.pone.0086059

Maddocks, M., Nolan, C. M., Man, W. D.-C., Polkey, M. I., Hart, N., Gao, W., … Higginson, I. J. (2016). Neuromuscular electrical stimulation to improve exercise capacity in patients with severe COPD: a randomised double-blind, placebo-controlled trial. *The Lancet. Respiratory Medicine*, *4*(1), 27–36. https://doi.org/10.1016/S2213-2600(15)00503-2

Maffiuletti, N. A. (2010). Physiological and methodological considerations for the use of neuromuscular electrical stimulation. *European Journal of Applied Physiology*, *110*(2), 223–234. https://doi.org/10.1007/s00421-010-1502-y

Malietzis, G., Currie, A. C., Athanasiou, T., Johns, N., Anyamene, N., Glynne-Jones, R., … Jenkins, J. T. (2016). Influence of body composition profile on outcomes following colorectal cancer surgery. *The British Journal of Surgery*, *103*(5), 572–580. https://doi.org/10.1002/bjs.10075

Mann, G. E., Yudilevich, D. L., & Sobrevia, L. (2003). Regulation of amino acid and glucose transporters in endothelial and smooth muscle cells. *Physiological Reviews*, *83*(1), 183–252. https://doi.org/10.1152/physrev.00022.2002

Mantovani, G., Macciò, A., Madeddu, C., Gramignano, G., Serpe, R., Massa, E., … Floris, C. (2008). Randomized phase III clinical trial of five different arms of treatment for patients with cancer cachexia: interim results. *Nutrition (Burbank, Los Angeles County, Calif.)*, *24*(4), 305–313. https://doi.org/10.1016/j.nut.2007.12.010

Martin, L., Birdsell, L., Macdonald, N., Reiman, T., Clandinin, M. T., McCargar, L. J., … Baracos, V. E. (2013). Cancer cachexia in the age of obesity: skeletal muscle depletion is a powerful prognostic factor, independent of body mass index. *Journal of Clinical Oncology: Official Journal of the American Society of Clinical Oncology*, *31*(12), 1539–1547. https://doi.org/10.1200/JCO.2012.45.2722

Massicotte, M.-H., Borget, I., Broutin, S., Baracos, V. E., Leboulleux, S., Baudin, E., … Antoun, S. (2013). Body Composition Variation and Impact of Low Skeletal Muscle Mass in Patients With

Advanced Medullary Thyroid Carcinoma Treated With Vandetanib: Results From a Placebo-Controlled Study. *The Journal of Clinical Endocrinology & Metabolism*, *98*(6), 2401–2408. https://doi.org/10.1210/jc.2013-1115

Mayers, J. R., Torrence, M. E., Danai, L. V., Papagiannakopoulos, T., Davidson, S. M., Bauer, M. R., … Vander Heiden, M. G. (2016). Tissue of origin dictates branched-chain amino acid metabolism in mutant Kras-driven cancers. *Science (New York, N.Y.)*, *353*(6304), 1161–1165. https://doi.org/10.1126/science.aaf5171

Mayers, J. R., Wu, C., Clish, C. B., Kraft, P., Torrence, M. E., Fiske, B. P., … Wolpin, B. M. (2014). Elevation of circulating branched-chain amino acids is an early event in human pancreatic adenocarcinoma development. *Nature Medicine*, *20*(10), 1193–1198. https://doi.org/10.1038/nm.3686

McCann, R. M., Hall, W. J., & Groth-Juncker, A. (1994). Comfort care for terminally ill patients. The appropriate use of nutrition and hydration. *JAMA*, *272*(16), 1263–1266.

McCormick, R., & Vasilaki, A. (2018). Age-related changes in skeletal muscle: changes to life-style as a therapy. *Biogerontology*. https://doi.org/10.1007/s10522-018-9775-3

Meza-Junco, J., Montano-Loza, A. J., Baracos, V. E., Prado, C. M. M., Bain, V. G., Beaumont, C., … Sawyer, M. B. (2013). Sarcopenia as a prognostic index of nutritional status in concurrent cirrhosis and hepatocellular carcinoma. *Journal of Clinical Gastroenterology*, *47*(10), 861–870. https://doi.org/10.1097/MCG.0b013e318293a825

Miller, J. A., Harris, K., Roche, C., Dhillon, S., Battoo, A., Demmy, T., … Yendamuri, S. (2018). Sarcopenia is a predictor of outcomes after lobectomy. *Journal of Thoracic Disease*, *10*(1), 432–440. https://doi.org/10.21037/jtd.2017.12.39

Mir, O., Coriat, R., Blanchet, B., Durand, J.-P., Boudou-Rouquette, P., Michels, J., … Goldwasser, F. (2012). Sarcopenia Predicts Early Dose-Limiting Toxicities and Pharmacokinetics of Sorafenib in Patients with Hepatocellular Carcinoma. *PLoS ONE*, *7*(5), e37563. https://doi.org/10.1371/journal.pone.0037563

Mir, O., Coriat, R., Boudou-Rouquette, P., Ropert, S., Durand, J.-P., Cessot, A., … Goldwasser, F. (2012). Gemcitabine and oxaliplatin as second-line treatment in patients with hepatocellular carcinoma pre-treated with sorafenib. *Medical Oncology (Northwood, London, England)*, *29*(4), 2793–2799. https://doi.org/10.1007/s12032-012-0208-x

Mitsiopoulos, N., Baumgartner, R. N., Heymsfield, S. B., Lyons, W., Gallagher, D., & Ross, R. (1998). Cadaver validation of skeletal muscle measurement by magnetic resonance imaging and computerized tomography. *Journal of Applied Physiology (Bethesda, Md.: 1985)*, *85*(1), 115–122. https://doi.org/10.1152/jappl.1998.85.1.115

Miyamoto, Y., Baba, Y., Sakamoto, Y., Ohuchi, M., Tokunaga, R., Kurashige, J., … Baba, H. (2015). Sarcopenia is a Negative Prognostic Factor After Curative Resection of Colorectal Cancer. *Annals of Surgical Oncology*, *22*(8), 2663–2668. https://doi.org/10.1245/s10434-014-4281-6

Miyata, H., Sugimura, K., Motoori, M., Fujiwara, Y., Omori, T., Yanagimoto, Y., … Yano, M. (2017). Clinical Assessment of Sarcopenia and Changes in Body Composition During Neoadjuvant

Chemotherapy for Esophageal Cancer. *Anticancer Research*, *37*(6), 3053–3059. https://doi.org/10.21873/anticanres.11660

Morishita, S., Kaida, K., Tanaka, T., Itani, Y., Ikegame, K., Okada, M., … Domen, K. (2012). Prevalence of sarcopenia and relevance of body composition, physiological function, fatigue, and health-related quality of life in patients before allogeneic hematopoietic stem cell transplantation. *Supportive Care in Cancer: Official Journal of the Multinational Association of Supportive Care in Cancer*, *20*(12), 3161–3168. https://doi.org/10.1007/s00520-012-1460-5

Morley, J. E., Argiles, J. M., Evans, W. J., Bhasin, S., Cella, D., Deutz, N. E. P., … Anker, S. D. (2010). Nutritional Recommendations for the Management of Sarcopenia. *Journal of the American Medical Directors Association*, *11*(6), 391–396. https://doi.org/10.1016/j.jamda.2010.04.014

Morley, J. E., Malmstrom, T. K., Rodriguez-Mañas, L., & Sinclair, A. J. (2014). Frailty, sarcopenia and diabetes. *Journal of the American Medical Directors Association*, *15*(12), 853–859. https://doi.org/10.1016/j.jamda.2014.10.001

Morsbach, F., Zhang, Y.-H., Nowik, P., Martin, L., Lindqvist, C., Svensson, A., & Brismar, T. B. (2018). Influence of tube potential on CT body composition analysis. *Nutrition (Burbank, Los Angeles County, Calif.)*, *53*, 9–13. https://doi.org/10.1016/j.nut.2017.12.016

Motzer, R. J., Escudier, B., McDermott, D. F., George, S., Hammers, H. J., Srinivas, S., … CheckMate 025 Investigators. (2015). Nivolumab versus Everolimus in Advanced Renal-Cell Carcinoma. *The New England Journal of Medicine*, *373*(19), 1803–1813. https://doi.org/10.1056/NEJMoa1510665

Motzer, R. J., Rini, B. I., McDermott, D. F., Redman, B. G., Kuzel, T. M., Harrison, M. R., … Hammers, H. J. (2015). Nivolumab for Metastatic Renal Cell Carcinoma: Results of a Randomized Phase II Trial. *Journal of Clinical Oncology: Official Journal of the American Society of Clinical Oncology*, *33*(13), 1430–1437. https://doi.org/10.1200/JCO.2014.59.0703

Motzer, R. J., Tannir, N. M., McDermott, D. F., Arén Frontera, O., Melichar, B., Choueiri, T. K., … Escudier, B. (2018). Nivolumab plus Ipilimumab versus Sunitinib in Advanced Renal-Cell Carcinoma. *New England Journal of Medicine*, *378*(14), 1277–1290. https://doi.org/10.1056/NEJMoa1712126

Mourtzakis, M., Prado, C. M. M., Lieffers, J. R., Reiman, T., McCargar, L. J., & Baracos, V. E. (2008). A practical and precise approach to quantification of body composition in cancer patients using computed tomography images acquired during routine care. *Applied Physiology, Nutrition, and Metabolism*, *33*(5), 997–1006. https://doi.org/10.1139/H08-075

Mücke, M., Mochamat, Cuhls, H., Peuckmann-Post, V., Minton, O., Stone, P., & Radbruch, L. (2015). Pharmacological treatments for fatigue associated with palliative care. *The Cochrane Database of Systematic Reviews*, (5), CD006788. https://doi.org/10.1002/14651858.CD006788.pub3

Murimwa, G. Z., Venkat, P. S., Jin, W., Leuthold, S., Latifi, K., Almhanna, K., … Frakes, J. M. (2017). Impact of sarcopenia on outcomes of locally advanced esophageal cancer patients treated with neoadjuvant chemoradiation followed by surgery. *Journal of Gastrointestinal Oncology*, *8*(5), 808– 815. https://doi.org/10.21037/jgo.2017.06.11

Muscaritoli, M., Anker, S. D., Argilés, J., Aversa, Z., Bauer, J. M., Biolo, G., … Sieber, C. C. (2010). Consensus definition of sarcopenia, cachexia and pre-cachexia: joint document elaborated by Special Interest Groups (SIG) "cachexia-anorexia in chronic wasting diseases" and "nutrition in geriatrics." *Clinical Nutrition (Edinburgh, Scotland)*, *29*(2), 154–159. https://doi.org/10.1016/j.clnu.2009.12.004

Mustian, K. M., Alfano, C. M., Heckler, C., Kleckner, A. S., Kleckner, I. R., Leach, C. R., … Miller, S. M. (2017). Comparison of Pharmaceutical, Psychological, and Exercise Treatments for Cancer-Related Fatigue: A Meta-analysis. *JAMA Oncology*, *3*(7), 961–968. https://doi.org/10.1001/jamaoncol.2016.6914

Nakamura, N., Hara, T., Shibata, Y., Matsumoto, T., Nakamura, H., Ninomiya, S., … Tsurumi, H. (2015). Sarcopenia is an independent prognostic factor in male patients with diffuse large B-cell lymphoma. *Annals of Hematology*, *94*(12), 2043–2053. https://doi.org/10.1007/s00277-015-2499-4

Nakamura, R., Inage, Y., Tobita, R., Yoneyama, S., Numata, T., Ota, K., … Terashima, T. (2018). Sarcopenia in Resected NSCLC: Effect on Postoperative Outcomes. *Journal of Thoracic Oncology: Official Publication of the International Association for the Study of Lung Cancer*, *13*(7), 895–903. https://doi.org/10.1016/j.jtho.2018.04.035

Narsale, A. A., & Carson, J. A. (2014). Role of interleukin-6 in cachexia: therapeutic implications. *Current Opinion in Supportive and Palliative Care*, *8*(4), 321–327. https://doi.org/10.1097/SPC.0000000000000091

Nault, J.-C., Pigneur, F., Nelson, A. C., Costentin, C., Tselikas, L., Katsahian, S., … Decaens, T. (2015). Visceral fat area predicts survival in patients with advanced hepatocellular carcinoma treated with tyrosine kinase inhibitors. *Digestive and Liver Disease*, *47*(10), 869–876. https://doi.org/10.1016/j.dld.2015.07.001

Neves, R. X., Rosa-Neto, J. C., Yamashita, A. S., Matos-Neto, E. M., Riccardi, D. M. R., Lira, F. S., … Seelaender, M. (2016). White adipose tissue cells and the progression of cachexia: inflammatory pathways. *Journal of Cachexia, Sarcopenia and Muscle*, *7*(2), 193–203. https://doi.org/10.1002/jcsm.12041

Nishigori, T., Tsunoda, S., Okabe, H., Tanaka, E., Hisamori, S., Hosogi, H., … Sakai, Y. (2016). Impact of Sarcopenic Obesity on Surgical Site Infection after Laparoscopic Total Gastrectomy. *Annals of Surgical Oncology*, *23*(Suppl 4), 524–531. https://doi.org/10.1245/s10434-016-5385-y

O'Connor, D., & Caulfield, B. (2018). The application of neuromuscular electrical stimulation (NMES) in cancer rehabilitation: current prescription, pitfalls, and future directions. *Supportive Care in Cancer: Official Journal of the Multinational Association of Supportive Care in Cancer*. https://doi.org/10.1007/s00520-018-4269-z

Owusu, C., Margevicius, S., Schluchter, M., Koroukian, S. M., & Berger, N. A. (2017). Short Physical Performance Battery, usual gait speed, grip strength and Vulnerable Elders Survey each predict functional decline among older women with breast cancer. *Journal of Geriatric Oncology*, *8*(5), 356– 362. https://doi.org/10.1016/j.jgo.2017.07.004

Öztürk, Z. A., Kul, S., Türkbeyler, İ. H., Sayıner, Z. A., & Abiyev, A. (2018). Is increased neutrophil lymphocyte ratio remarking the inflammation in sarcopenia? *Experimental Gerontology*, *110*, 223– 229. https://doi.org/10.1016/j.exger.2018.06.013

Padhi, D., Higano, C. S., Shore, N. D., Sieber, P., Rasmussen, E., & Smith, M. R. (2014). Pharmacological inhibition of myostatin and changes in lean body mass and lower extremity muscle size in patients receiving androgen deprivation therapy for prostate cancer. *The Journal of Clinical Endocrinology and Metabolism*, *99*(10), E1967-1975. https://doi.org/10.1210/jc.2014-1271

Pahor, M., Guralnik, J. M., Ambrosius, W. T., Blair, S., Bonds, D. E., Church, T. S., … LIFE study investigators. (2014). Effect of structured physical activity on prevention of major mobility disability in older adults: the LIFE study randomized clinical trial. *JAMA*, *311*(23), 2387–2396. https://doi.org/10.1001/jama.2014.5616

Pai, M. P. (2012). Drug dosing based on weight and body surface area: mathematical assumptions and limitations in obese adults. *Pharmacotherapy*, *32*(9), 856–868. https://doi.org/10.1002/j.1875- 9114.2012.01108.x

Palmela, C., Velho, S., Agostinho, L., Branco, F., Santos, M., Santos, M. P. C., ... Baracos, V. E. (2017). Body Composition as a Prognostic Factor of Neoadjuvant Chemotherapy Toxicity and Outcome in Patients with Locally Advanced Gastric Cancer. *Journal of Gastric Cancer*, *17*(1), 74–87. https://doi.org/10.5230/jgc.2017.17.e8

Papanicolaou, D. A., Ather, S. N., Zhu, H., Zhou, Y., Lutkiewicz, J., Scott, B. B., & Chandler, J. (2013). A phase IIA randomized, placebo-controlled clinical trial to study the efficacy and safety of the selective androgen receptor modulator (SARM), MK-0773 in female participants with sarcopenia. *The Journal of Nutrition, Health & Aging*, *17*(6), 533–543. https://doi.org/10.1007/s12603-013-0335 x

Pappalardo, G., Almeida, A., & Ravasco, P. (2015). Eicosapentaenoic acid in cancer improves body composition and modulates metabolism. *Nutrition (Burbank, Los Angeles County, Calif.)*, *31*(4), 549– 555. https://doi.org/10.1016/j.nut.2014.12.002

Parajuli, P., Kumar, S., Loumaye, A., Singh, P., Eragamreddy, S., Nguyen, T. L., … Atfi, A. (2018). Twist1 Activation in Muscle Progenitor Cells Causes Muscle Loss Akin to Cancer Cachexia. *Developmental Cell*, *45*(6), 712-725.e6. https://doi.org/10.1016/j.devcel.2018.05.026

Park, S. W., Goodpaster, B. H., Strotmeyer, E. S., Kuller, L. H., Broudeau, R., Kammerer, C., … Health, Aging, and Body Composition Study. (2007). Accelerated loss of skeletal muscle strength in older adults with type 2 diabetes: the health, aging, and body composition study. *Diabetes Care*, *30*(6), 1507–1512. https://doi.org/10.2337/dc06-2537

Parsons, H. A., Baracos, V. E., Dhillon, N., Hong, D. S., & Kurzrock, R. (2012). Body composition, symptoms, and survival in advanced cancer patients referred to a phase I service. *PloS One*, *7*(1), e29330. https://doi.org/10.1371/journal.pone.0029330

Parsons, H. A., Tsimberidou, A. M., Pontikos, M., Fu, S., Hong, D., Wen, S., … Kurzrock, R. (2012). Evaluation of the clinical relevance of body composition parameters in patients with cancer metastatic to the liver treated with hepatic arterial infusion chemotherapy. *Nutrition and Cancer*, *64*(2), 206–217.

Peng, P. D., van Vledder, M. G., Tsai, S., de Jong, M. C., Makary, M., Ng, J., … Pawlik, T. M. (2011). Sarcopenia negatively impacts short-term outcomes in patients undergoing hepatic resection for colorectal liver metastasis. *HPB: The Official Journal of the International Hepato Pancreato Biliary Association*, *13*(7), 439–446. https://doi.org/10.1111/j.1477-2574.2011.00301.x

Peng, P., Hyder, O., Firoozmand, A., Kneuertz, P., Schulick, R. D., Huang, D., … Pawlik, T. M. (2012). Impact of sarcopenia on outcomes following resection of pancreatic adenocarcinoma. *Journal of Gastrointestinal Surgery: Official Journal of the Society for Surgery of the Alimentary Tract*, *16*(8), 1478–1486. https://doi.org/10.1007/s11605-012-1923-5

Pennings, B., Koopman, R., Beelen, M., Senden, J. M. G., Saris, W. H. M., & van Loon, L. J. C. (2011). Exercising before protein intake allows for greater use of dietary protein-derived amino acids for de novo muscle protein synthesis in both young and elderly men. *The American Journal of Clinical Nutrition*, *93*(2), 322–331. https://doi.org/10.3945/ajcn.2010.29649

Perrone, R. D., Madias, N. E., & Levey, A. S. (1992). Serum creatinine as an index of renal function: new insights into old concepts. *Clinical Chemistry*, *38*(10), 1933–1953.

Pierpont, Y. N., Dinh, T. P., Salas, R. E., Johnson, E. L., Wright, T. G., Robson, M. C., & Payne, W. G. (2014). Obesity and Surgical Wound Healing: A Current Review. *ISRN Obesity*, *2014*. https://doi.org/10.1155/2014/638936

Pirttiaho, H. I., Sotaniemi, E. A., Pelkonen, R. O., Pitkänen, U., Anttila, M., & Sundqvist, H. (1980). Roles of hepatic blood flow and enzyme activity in the kinetics of propranolol and sotalol. *British Journal of Clinical Pharmacology*, *9*(4), 399–405.

Popuri, K., Cobzas, D., Esfandiari, N., Baracos, V., & Jägersand, M. (2016). Body Composition Assessment in Axial CT Images Using FEM-Based Automatic Segmentation of Skeletal Muscle. *IEEE Transactions on Medical Imaging*, *35*(2), 512–520. https://doi.org/10.1109/TMI.2015.2479252

Postel-Vinay, S. (2015). Redefining dose-limiting toxicity. *Clinical Advances in Hematology & Oncology: H&O*, *13*(2), 87–89.

Prado, C. M. M., Baracos, V. E., McCargar, L. J., Mourtzakis, M., Mulder, K. E., Reiman, T., … Sawyer, M. B. (2007). Body composition as an independent determinant of 5-fluorouracil-based chemotherapy toxicity. *Clinical Cancer Research: An Official Journal of the American Association for Cancer Research*, *13*(11), 3264–3268. https://doi.org/10.1158/1078-0432.CCR-06-3067

Prado, C. M. M., Baracos, V. E., McCargar, L. J., Reiman, T., Mourtzakis, M., Tonkin, K., … Sawyer, M. B. (2009). Sarcopenia as a determinant of chemotherapy toxicity and time to tumor progression in metastatic breast cancer patients receiving capecitabine treatment. *Clinical Cancer Research: An Official Journal of the American Association for Cancer Research*, *15*(8), 2920–2926. https://doi.org/10.1158/1078-0432.CCR-08-2242

Prado, C. M. M., Baracos, V. E., Xiao, J., Birdsell, L., Stuyckens, K., Park, Y. C., … Sawyer, M. B. (2014). The association between body composition and toxicities from the combination of Doxil and trabectedin in patients with advanced relapsed ovarian cancer. *Applied Physiology, Nutrition, and Metabolism = Physiologie Appliquee, Nutrition Et Metabolisme*, *39*(6), 693–698. https://doi.org/10.1139/apnm-2013-0403

Prado, C. M. M., & Heymsfield, S. B. (2014). Lean tissue imaging: a new era for nutritional assessment and intervention. *JPEN. Journal of Parenteral and Enteral Nutrition*, *38*(8), 940–953. https://doi.org/10.1177/0148607114550189

Prado, C. M. M., Lieffers, J. R., McCargar, L. J., Reiman, T., Sawyer, M. B., Martin, L., & Baracos, V. E. (2008). Prevalence and clinical implications of sarcopenic obesity in patients with solid tumours of the respiratory and gastrointestinal tracts: a population-based study. *The Lancet. Oncology*, *9*(7), 629–635. https://doi.org/10.1016/S1470-2045(08)70153-0

Prado, C. M. M., Lima, I. S. F., Baracos, V. E., Bies, R. R., McCargar, L. J., Reiman, T., … Sawyer, M. B. (2011). An exploratory study of body composition as a determinant of epirubicin pharmacokinetics and toxicity. *Cancer Chemotherapy and Pharmacology*, *67*(1), 93–101. https://doi.org/10.1007/s00280-010-1288-y

Psutka, S. P., Boorjian, S. A., Moynagh, M. R., Schmit, G. D., Costello, B. A., Thompson, R. H., … Tollefson, M. K. (2016). Decreased Skeletal Muscle Mass is Associated with an Increased Risk of Mortality after Radical Nephrectomy for Localized Renal Cell Cancer. *The Journal of Urology*, *195*(2), 270–276. https://doi.org/10.1016/j.juro.2015.08.072

Psutka, S. P., Boorjian, S. A., Moynagh, M. R., Schmit, G. D., Frank, I., Carrasco, A., … Tollefson, M. K. (2015). Mortality after radical cystectomy: impact of obesity versus adiposity after adjusting for skeletal muscle wasting. *The Journal of Urology*, *193*(5), 1507–1513. https://doi.org/10.1016/j.juro.2014.11.088

Puszkiel, A., Noé, G., Boudou-Rouquette, P., Cossec, C. L.-, Arrondeau, J., Giraud, J.-S., … Blanchet, B. (2017). Development and validation of an ELISA method for the quantification of nivolumab in plasma from non-small-cell lung cancer patients. *Journal of Pharmaceutical and Biomedical Analysis*, *139*, 30–36. https://doi.org/10.1016/j.jpba.2017.02.041

Reck, M., Rodríguez-Abreu, D., Robinson, A. G., Hui, R., Csőszi, T., Fülöp, A., … Brahmer, J. R. (2016). Pembrolizumab versus Chemotherapy for PD-L1–Positive Non–Small-Cell Lung Cancer. *New England Journal of Medicine*, *375*(19), 1823–1833. https://doi.org/10.1056/NEJMoa1606774

Redlarski, G., Palkowski, A., & Krawczuk, M. (2016). Body surface area formulae: an alarming ambiguity. *Scientific Reports*, *6*, 27966. https://doi.org/10.1038/srep27966

Reisinger, K. W., van Vugt, J. L. A., Tegels, J. J. W., Snijders, C., Hulsewé, K. W. E., Hoofwijk, A. G. M., … Poeze, M. (2015). Functional compromise reflected by sarcopenia, frailty, and nutritional depletion predicts adverse postoperative outcome after colorectal cancer surgery. *Annals of Surgery*, *261*(2), 345–352. https://doi.org/10.1097/SLA.0000000000000628

Research, C. for D. E. and. (n.d.). Approved Drugs - Modification of the Dosage Regimen for Nivolumab [WebContent]. Retrieved September 9, 2018, from https://www.fda.gov/Drugs/InformationOnDrugs/ApprovedDrugs/ucm520871.htm

Robert, C., Ribas, A., Wolchok, J. D., Hodi, F. S., Hamid, O., Kefford, R., … Daud, A. (2014). Antiprogrammed-death-receptor-1 treatment with pembrolizumab in ipilimumab-refractory advanced melanoma: a randomised dose-comparison cohort of a phase 1 trial. *The Lancet*, *384*(9948), 1109– 1117. https://doi.org/10.1016/S0140-6736(14)60958-2

Rockwood, K., Song, X., MacKnight, C., Bergman, H., Hogan, D. B., McDowell, I., & Mitnitski, A. (2005). A global clinical measure of fitness and frailty in elderly people. *CMAJ: Canadian Medical Association Journal = Journal de l'Association Medicale Canadienne*, *173*(5), 489–495. https://doi.org/10.1503/cmaj.050051

Rogers, J. F., Rocci, M. L., Haughey, D. B., & Bertino, J. S. (2003). An evaluation of the suitability of intravenous midazolam as an in vivo marker for hepatic cytochrome P4503A activity. *Clinical Pharmacology and Therapeutics*, *73*(3), 153–158. https://doi.org/10.1067/mcp.2003.23

Rollins, K. E., Tewari, N., Ackner, A., Awwad, A., Madhusudan, S., Macdonald, I. A., … Lobo, D. N. (2016). The impact of sarcopenia and myosteatosis on outcomes of unresectable pancreatic cancer or distal cholangiocarcinoma. *Clinical Nutrition (Edinburgh, Scotland)*, *35*(5), 1103–1109. https://doi.org/10.1016/j.clnu.2015.08.005

Rosenberg, I. H. (1989). Summary comments. *The American Journal of Clinical Nutrition*, *50*(5), 1231– 1233. https://doi.org/10.1093/ajcn/50.5.1231

Rosenberg, I. H. (1997). Sarcopenia: origins and clinical relevance. *The Journal of Nutrition*, *127*(5 Suppl), 990S-991S. https://doi.org/10.1093/jn/127.5.990S

Roubenoff, R. (2000). Sarcopenia: a major modifiable cause of frailty in the elderly. *The Journal of Nutrition, Health & Aging*, *4*(3), 140–142.

Roubenoff, Ronenn, & Kehayias, J. J. (2009). The Meaning and Measurement of Lean Body Mass. *Nutrition Reviews*, *49*(6), 163–175. https://doi.org/10.1111/j.1753-4887.1991.tb03013.x

Rule, A. D., Bergstralh, E. J., Slezak, J. M., Bergert, J., & Larson, T. S. (2006). Glomerular filtration rate estimated by cystatin C among different clinical presentations. *Kidney International*, *69*(2), 399–405. https://doi.org/10.1038/sj.ki.5000073

Ryan, A. M., Reynolds, J. V., Healy, L., Byrne, M., Moore, J., Brannelly, N., … Flood, P. (2009). Enteral nutrition enriched with eicosapentaenoic acid (EPA) preserves lean body mass following esophageal cancer surgery: results of a double-blinded randomized controlled trial. *Annals of Surgery*, *249*(3), 355–363. https://doi.org/10.1097/SLA.0b013e31819a4789

Sachs, J. R., Mayawala, K., Gadamsetty, S., Kang, S. P., & de Alwis, D. P. (2016). Optimal Dosing for Targeted Therapies in Oncology: Drug Development Cases Leading by Example. *Clinical Cancer Research: An Official Journal of the American Association for Cancer Research*, *22*(6), 1318–1324. https://doi.org/10.1158/1078-0432.CCR-15-1295

Sakai, H., Sagara, A., Arakawa, K., Sugiyama, R., Hirosaki, A., Takase, K., … Narita, M. (2014). Mechanisms of cisplatin-induced muscle atrophy. *Toxicology and Applied Pharmacology*, *278*(2), 190–199. https://doi.org/10.1016/j.taap.2014.05.001

Sánchez-Lara, K., Turcott, J. G., Juárez-Hernández, E., Nuñez-Valencia, C., Villanueva, G., Guevara, P., … Arrieta, O. (2014). Effects of an oral nutritional supplement containing eicosapentaenoic acid on nutritional and clinical outcomes in patients with advanced non-small cell lung cancer: randomised trial. *Clinical Nutrition (Edinburgh, Scotland)*, *33*(6), 1017–1023. https://doi.org/10.1016/j.clnu.2014.03.006

Sargent, D. J., Köhne, C. H., Sanoff, H. K., Bot, B. M., Seymour, M. T., de Gramont, A., … Goldberg, R. M. (2009). Pooled safety and efficacy analysis examining the effect of performance status on outcomes in nine first-line treatment trials using individual data from patients with metastatic colorectal cancer. *Journal of Clinical Oncology: Official Journal of the American Society of Clinical Oncology*, *27*(12), 1948–1955. https://doi.org/10.1200/JCO.2008.20.2879

Schaap, L. A., Pluijm, S. M. F., Deeg, D. J. H., & Visser, M. (2006). Inflammatory markers and loss of muscle mass (sarcopenia) and strength. *The American Journal of Medicine*, *119*(6), 526.e9-17. https://doi.org/10.1016/j.amjmed.2005.10.049

Schmid, D., & Leitzmann, M. F. (2014). Association between physical activity and mortality among breast cancer and colorectal cancer survivors: a systematic review and meta-analysis. *Annals of Oncology: Official Journal of the European Society for Medical Oncology*, *25*(7), 1293–1311. https://doi.org/10.1093/annonc/mdu012

Schmitt, A., Gladieff, L., Lansiaux, A., Bobin-Dubigeon, C., Etienne-Grimaldi, M.-C., Boisdron-Celle, M., … Chatelut, E. (2009). A universal formula based on cystatin C to perform individual dosing of carboplatin in normal weight, underweight, and obese patients. *Clinical Cancer Research: An Official Journal of the American Association for Cancer Research*, *15*(10), 3633–3639. https://doi.org/10.1158/1078-0432.CCR-09-0017

Segatto, M., Fittipaldi, R., Pin, F., Sartori, R., Dae Ko, K., Zare, H., … Caretti, G. (2017). Epigenetic targeting of bromodomain protein BRD4 counteracts cancer cachexia and prolongs survival. *Nature Communications*, *8*. https://doi.org/10.1038/s41467-017-01645-7

Shachar, S. S., Deal, A. M., Weinberg, M., Nyrop, K. A., Williams, G. R., Nishijima, T. F., … Muss, H. B. (2017). Skeletal Muscle Measures as Predictors of Toxicity, Hospitalization, and Survival in Patients with Metastatic Breast Cancer Receiving Taxane-Based Chemotherapy. *Clinical Cancer Research: An Official Journal of the American Association for Cancer Research*, *23*(3), 658–665. https://doi.org/10.1158/1078-0432.CCR-16-0940

Shachar, S. S., Deal, A. M., Weinberg, M., Williams, G. R., Nyrop, K. A., Popuri, K., … Muss, H. B. (2017). Body Composition as a Predictor of Toxicity in Patients Receiving Anthracycline and Taxane-Based Chemotherapy for Early-Stage Breast Cancer. *Clinical Cancer Research: An Official Journal of the American Association for Cancer Research*, *23*(14), 3537–3543. https://doi.org/10.1158/1078- 0432.CCR-16-2266

Shachar, S. S., Williams, G. R., Muss, H. B., & Nishijima, T. F. (2016). Prognostic value of sarcopenia in adults with solid tumours: A meta-analysis and systematic review. *European Journal of Cancer*, *57*, 58–67. https://doi.org/10.1016/j.ejca.2015.12.030

Sharma, P., Zargar-Shoshtari, K., Caracciolo, J. T., Fishman, M., Poch, M. A., Pow-Sang, J., … Spiess, P. E. (2015). Sarcopenia as a predictor of overall survival after cytoreductive nephrectomy for metastatic renal cell carcinoma. *Urologic Oncology*, *33*(8), 339.e17-23. https://doi.org/10.1016/j.urolonc.2015.01.011

Sharma, P., Zargar-Shoshtari, K., Caracciolo, J. T., Richard, G. J., Poch, M. A., Pow-Sang, J., … Spiess, P. E. (2015). Sarcopenia as a predictor of complications in penile cancer patients undergoing inguinal

lymph node dissection. *World Journal of Urology*, *33*(10), 1585–1592. https://doi.org/10.1007/s00345-014-1471-6

Shen, W., Punyanitya, M., Wang, Z., Gallagher, D., St-Onge, M.-P., Albu, J., … Heshka, S. (2004). Total body skeletal muscle and adipose tissue volumes: estimation from a single abdominal cross-sectional image. *Journal of Applied Physiology (Bethesda, Md.: 1985)*, *97*(6), 2333–2338. https://doi.org/10.1152/japplphysiol.00744.2004

Shiel, F., Persson, C., Furness, J., Simas, V., Pope, R., Climstein, M., … Schram, B. (2018). Dual energy X-ray absorptiometry positioning protocols in assessing body composition: A systematic review of the literature. *Journal of Science and Medicine in Sport*. https://doi.org/10.1016/j.jsams.2018.03.005

Simoneau, J. A., & Kelley, D. E. (1997). Altered glycolytic and oxidative capacities of skeletal muscle contribute to insulin resistance in NIDDM. *Journal of Applied Physiology (Bethesda, Md.: 1985)*, *83*(1), 166–171. https://doi.org/10.1152/jappl.1997.83.1.166

Sjøblom, B., Benth, J. Š., Grønberg, B. H., Baracos, V. E., Sawyer, M. B., Fløtten, Ø., … Jordhøy, M. (2017). Drug Dose Per Kilogram Lean Body Mass Predicts Hematologic Toxicity From Carboplatin-Doublet Chemotherapy in Advanced Non-Small-Cell Lung Cancer. *Clinical Lung Cancer*, *18*(2), e129– e136. https://doi.org/10.1016/j.cllc.2016.09.008

Sjøblom, B., Grønberg, B. H., Benth, J. Š., Baracos, V. E., Fløtten, Ø., Hjermstad, M. J., … Jordhøy, M. (2015). Low muscle mass is associated with chemotherapy-induced haematological toxicity in advanced non-small cell lung cancer. *Lung Cancer*, *90*(1), 85–91. https://doi.org/10.1016/j.lungcan.2015.07.001

Sloan, J. A., Goldberg, R. M., Sargent, D. J., Vargas-Chanes, D., Nair, S., Cha, S. S., … Loprinzi, C. L. (2002). Women experience greater toxicity with fluorouracil-based chemotherapy for colorectal cancer. *Journal of Clinical Oncology: Official Journal of the American Society of Clinical Oncology*, *20*(6), 1491–1498. https://doi.org/10.1200/JCO.2002.20.6.1491

Smart, R., Carter, B., McGovern, J., Luckman, S., Connelly, A., Hewitt, J., … Moug, S. (2017). Frailty Exists in Younger Adults Admitted as Surgical Emergency Leading to Adverse Outcomes. *The Journal of Frailty & Aging*, *6*(4), 219–223. https://doi.org/10.14283/jfa.2017.28

Spotten, L. E., Corish, C. A., Lorton, C. M., Ui Dhuibhir, P. M., O'Donoghue, N. C., O'Connor, B., & Walsh, T. D. (2017). Subjective and objective taste and smell changes in cancer. *Annals of Oncology: Official Journal of the European Society for Medical Oncology*, *28*(5), 969–984. https://doi.org/10.1093/annonc/mdx018

Srdic, D., Plestina, S., Sverko-Peternac, A., Nikolac, N., Simundic, A.-M., & Samarzija, M. (2016). Cancer cachexia, sarcopenia and biochemical markers in patients with advanced non-small cell lung cancer-chemotherapy toxicity and prognostic value. *Supportive Care in Cancer: Official Journal of the Multinational Association of Supportive Care in Cancer*. https://doi.org/10.1007/s00520-016-3287-y

Srikanthan, P., Hevener, A. L., & Karlamangla, A. S. (2010). Sarcopenia exacerbates obesityassociated insulin resistance and dysglycemia: findings from the National Health and Nutrition Examination Survey III. *PloS One*, *5*(5), e10805. https://doi.org/10.1371/journal.pone.0010805 Stanisavljevic, N. S., & Marisavljevic, D. Z. (2010). Weight and body composition changes during R-CHOP chemotherapy in patients with non-Hodgkin's lymphoma and their impact on dose intensity and toxicity. *Journal of B.U.ON.: Official Journal of the Balkan Union of Oncology*, *15*(2), 290–296.

Stene, G. B., Helbostad, J. L., Amundsen, T., Sørhaug, S., Hjelde, H., Kaasa, S., & Grønberg, B. H. (2015). Changes in skeletal muscle mass during palliative chemotherapy in patients with advanced lung cancer. *Acta Oncologica*, *54*(3), 340–348. https://doi.org/10.3109/0284186X.2014.953259

Stewart Coats, A. J., Ho, G. F., Prabhash, K., von Haehling, S., Tilson, J., Brown, R., … for and on behalf of the ACT-ONE study group. (2016). Espindolol for the treatment and prevention of cachexia in patients with stage III/IV non-small cell lung cancer or colorectal cancer: a randomized, double-blind, placebo-controlled, international multicentre phase II study (the ACT-ONE trial). *Journal of Cachexia, Sarcopenia and Muscle*, *7*(3), 355–365. https://doi.org/10.1002/jcsm.12126

Su, G. M., Sefton, R. M., & Murray, M. (1999). Down-regulation of rat hepatic microsomal cytochromes P-450 in microvesicular steatosis induced by orotic acid. *The Journal of Pharmacology and Experimental Therapeutics*, *291*(3), 953–959.

Subramaniam, K., Fallon, K., Ruut, T., Lane, D., McKay, R., Shadbolt, B., ... Taupin, D. (2015). Infliximab reverses inflammatory muscle wasting (sarcopenia) in Crohn's disease. *Alimentary Pharmacology & Therapeutics*, *41*(5), 419–428. https://doi.org/10.1111/apt.13058

Sucak, G. T., Suyanı, E., Baysal, N. A., Altındal, Ş., Çakar, M. K., Akı, Ş. Z., … Şanlıer, N. (2012). The role of body mass index and other body composition parameters in early post-transplant complications in patients undergoing allogeneic stem cell transplantation with busulfan-cyclophosphamide conditioning. *International Journal of Hematology*, *95*(1), 95–101. https://doi.org/10.1007/s12185- 011-0980-y

Suzuki, K., Furuse, H., Tsuda, T., Masaki, Y., Okazawa, S., Kambara, K., … Tobe, K. (2015). Utility of creatinine/cystatin C ratio as a predictive marker for adverse effects of chemotherapy in lung cancer: A retrospective study. *The Journal of International Medical Research*, *43*(4), 573–582. https://doi.org/10.1177/0300060515579116

Takagi, K., Yoshida, R., Yagi, T., Umeda, Y., Nobuoka, D., Kuise, T., & Fujiwara, T. (2017). Radiographic sarcopenia predicts postoperative infectious complications in patients undergoing pancreaticoduodenectomy. *BMC Surgery*, *17*(1), 64. https://doi.org/10.1186/s12893-017-0261-7

Takayama, K., Atagi, S., Imamura, F., Tanaka, H., Minato, K., Harada, T., … Watanabe, K. (2016). Quality of life and survival survey of cancer cachexia in advanced non-small cell lung cancer patients-Japan nutrition and QOL survey in patients with advanced non-small cell lung cancer study. *Supportive Care in Cancer: Official Journal of the Multinational Association of Supportive Care in Cancer*, *24*(8), 3473–3480. https://doi.org/10.1007/s00520-016-3156-8

Tamandl, D., Paireder, M., Asari, R., Baltzer, P. A., Schoppmann, S. F., & Ba-Ssalamah, A. (2016). Markers of sarcopenia quantified by computed tomography predict adverse long-term outcome in patients with resected oesophageal or gastro-oesophageal junction cancer. *European Radiology*, *26*(5), 1359–1367. https://doi.org/10.1007/s00330-015-3963-1

Tan, B. H. L., Brammer, K., Randhawa, N., Welch, N. T., Parsons, S. L., James, E. J., & Catton, J. A. (2015). Sarcopenia is associated with toxicity in patients undergoing neo-adjuvant chemotherapy for oesophago-gastric cancer. *European Journal of Surgical Oncology: The Journal of the European Society of Surgical Oncology and the British Association of Surgical Oncology*, *41*(3), 333–338. https://doi.org/10.1016/j.ejso.2014.11.040

Tan, Benjamin H. L., Birdsell, L. A., Martin, L., Baracos, V. E., & Fearon, K. C. H. (2009). Sarcopenia in an overweight or obese patient is an adverse prognostic factor in pancreatic cancer. *Clinical Cancer Research: An Official Journal of the American Association for Cancer Research*, *15*(22), 6973–6979. https://doi.org/10.1158/1078-0432.CCR-09-1525

Tegels, J. J. W., van Vugt, J. L. A., Reisinger, K. W., Hulsewé, K. W. E., Hoofwijk, A. G. M., Derikx, J. P. M., & Stoot, J. H. M. B. (2015). Sarcopenia is highly prevalent in patients undergoing surgery for gastric cancer but not associated with worse outcomes. *Journal of Surgical Oncology*, *112*(4), 403– 407. https://doi.org/10.1002/jso.24015

Temel, J. S., Abernethy, A. P., Currow, D. C., Friend, J., Duus, E. M., Yan, Y., & Fearon, K. C. (2016). Anamorelin in patients with non-small-cell lung cancer and cachexia (ROMANA 1 and ROMANA 2): results from two randomised, double-blind, phase 3 trials. *The Lancet. Oncology*, *17*(4), 519–531. https://doi.org/10.1016/S1470-2045(15)00558-6

Temel, J. S., Greer, J. A., Muzikansky, A., Gallagher, E. R., Admane, S., Jackson, V. A., … Lynch, T. J. (2010). Early Palliative Care for Patients with Metastatic Non–Small-Cell Lung Cancer. *New England Journal of Medicine*, *363*(8), 733–742. https://doi.org/10.1056/NEJMoa1000678

Tetsuka, S., Morita, M., Ikeguchi, K., & Nakano, I. (2013). Creatinine/cystatin C ratio as a surrogate marker of residual muscle mass in amyotrophic lateral sclerosis. *Neurology and Clinical Neuroscience*, *1*(1), 32–37. https://doi.org/10.1002/ncn3.11

Thompson, P. A., Rosner, G. L., Matthay, K. K., Moore, T. B., Bomgaars, L. R., Ellis, K. J., … Berg, S. L. (2009). Impact of body composition on pharmacokinetics of doxorubicin in children: a Glaser Pediatric Research Network study. *Cancer Chemotherapy and Pharmacology*, *64*(2), 243–251. https://doi.org/10.1007/s00280-008-0854-z

Thoresen, L., Frykholm, G., Lydersen, S., Ulveland, H., Baracos, V., Birdsell, L., & Falkmer, U. (2012). The association of nutritional assessment criteria with health-related quality of life in patients with advanced colorectal carcinoma. *European Journal of Cancer Care*, *21*(4), 505–516. https://doi.org/10.1111/j.1365-2354.2012.01327.x

Thoresen, Lene, Frykholm, G., Lydersen, S., Ulveland, H., Baracos, V., Prado, C. M. M., … Falkmer, U. (2013). Nutritional status, cachexia and survival in patients with advanced colorectal carcinoma. Different assessment criteria for nutritional status provide unequal results. *Clinical Nutrition (Edinburgh, Scotland)*, *32*(1), 65–72. https://doi.org/10.1016/j.clnu.2012.05.009

Valentijn, T. M., Galal, W., Tjeertes, E. K. M., Hoeks, S. E., Verhagen, H. J., & Stolker, R. J. (2013). The obesity paradox in the surgical population. *The Surgeon: Journal of the Royal Colleges of Surgeons of Edinburgh and Ireland*, *11*(3), 169–176. https://doi.org/10.1016/j.surge.2013.02.003

van der Ende, M., Grefte, S., Plas, R., Meijerink, J., Witkamp, R., Keijer, J., & van Norren, K. (2018). Mitochondrial dynamics in cancer-induced cachexia. *Biochimica Et Biophysica Acta*. https://doi.org/10.1016/j.bbcan.2018.07.008

van Vledder, M. G., Levolger, S., Ayez, N., Verhoef, C., Tran, T. C. K., & Ijzermans, J. N. M. (2012). Body composition and outcome in patients undergoing resection of colorectal liver metastases. *The British Journal of Surgery*, *99*(4), 550–557. https://doi.org/10.1002/bjs.7823

van Vugt, J. L. A., Braam, H. J., van Oudheusden, T. R., Vestering, A., Bollen, T. L., Wiezer, M. J., … Boerma, D. (2015). Skeletal Muscle Depletion is Associated with Severe Postoperative Complications in Patients Undergoing Cytoreductive Surgery with Hyperthermic Intraperitoneal Chemotherapy for Peritoneal Carcinomatosis of Colorectal Cancer. *Annals of Surgical Oncology*, *22*(11), 3625–3631. https://doi.org/10.1245/s10434-015-4429-z

van Vugt, J. L. A., Coebergh van den Braak, R. R. J., Schippers, H. J. W., Veen, K. M., Levolger, S., de Bruin, R. W. F., … Willemsen, F. E. J. A. (2018). Contrast-enhancement influences skeletal muscle density, but not skeletal muscle mass, measurements on computed tomography. *Clinical Nutrition (Edinburgh, Scotland)*, *37*(5), 1707–1714. https://doi.org/10.1016/j.clnu.2017.07.007

van Waart, H., Stuiver, M. M., van Harten, W. H., Geleijn, E., Kieffer, J. M., Buffart, L. M., … Aaronson, N. K. (2015). Effect of Low-Intensity Physical Activity and Moderate- to High-Intensity Physical Exercise During Adjuvant Chemotherapy on Physical Fitness, Fatigue, and Chemotherapy Completion Rates: Results of the PACES Randomized Clinical Trial. *Journal of Clinical Oncology: Official Journal of the American Society of Clinical Oncology*, *33*(17), 1918–1927. https://doi.org/10.1200/JCO.2014.59.1081

Vasson, M.-P., Talvas, J., Perche, O., Dillies, A.-F., Bachmann, P., Pezet, D., … Bouteloup, C. (2014). Immunonutrition improves functional capacities in head and neck and esophageal cancer patients undergoing radiochemotherapy: a randomized clinical trial. *Clinical Nutrition (Edinburgh, Scotland)*, *33*(2), 204–210. https://doi.org/10.1016/j.clnu.2013.06.008

Veni, T., Boyas, S., Beaune, B., Bourgeois, H., Rahmani, A., Landry, S., … Morel, B. (2018). Handgrip fatiguing exercise can provide objective assessment of cancer-related fatigue: a pilot study. *Supportive Care in Cancer: Official Journal of the Multinational Association of Supportive Care in Cancer*. https://doi.org/10.1007/s00520-018-4320-0

Versteeg, K. S., Blauwhoff-Buskermolen, S., Buffart, L. M., de van der Schueren, M. A. E., Langius, J. A. E., Verheul, H. M. W., … Konings, I. R. (2017). Higher Muscle Strength Is Associated with Prolonged Survival in Older Patients with Advanced Cancer. *The Oncologist*. https://doi.org/10.1634/theoncologist.2017-0193

Visser, M., Kritchevsky, S. B., Newman, A. B., Goodpaster, B. H., Tylavsky, F. A., Nevitt, M. C., & Harris, T. B. (2005). Lower serum albumin concentration and change in muscle mass: the Health, Aging and Body Composition Study. *The American Journal of Clinical Nutrition*, *82*(3), 531–537. https://doi.org/10.1093/ajcn/82.3.531

Visser, M., Pahor, M., Taaffe, D. R., Goodpaster, B. H., Simonsick, E. M., Newman, A. B., … Harris, T. B. (2002). Relationship of interleukin-6 and tumor necrosis factor-alpha with muscle mass and muscle strength in elderly men and women: the Health ABC Study. *The Journals of Gerontology. Series A, Biological Sciences and Medical Sciences*, *57*(5), M326-332.

von Haehling, S., & Anker, S. D. (2010). Cachexia as a major underestimated and unmet medical need: facts and numbers. *Journal of Cachexia, Sarcopenia and Muscle*, *1*(1), 1–5. https://doi.org/10.1007/s13539-010-0002-6

Voron, T., Tselikas, L., Pietrasz, D., Pigneur, F., Laurent, A., Compagnon, P., … Azoulay, D. (2015). Sarcopenia Impacts on Short- and Long-term Results of Hepatectomy for Hepatocellular Carcinoma. *Annals of Surgery*, *261*(6), 1173–1183. https://doi.org/10.1097/SLA.0000000000000743

Vrieling, A., Kampman, E., Knijnenburg, N. C., Mulders, P. F., Sedelaar, J. P. M., Baracos, V. E., & Kiemeney, L. A. (2016). Body Composition in Relation to Clinical Outcomes in Renal Cell Cancer: A Systematic Review and Meta-analysis. *European Urology Focus*. https://doi.org/10.1016/j.euf.2016.11.009

Wakefield, L. M., & Hill, C. S. (2013). Beyond TGFβ: roles of other TGFβ superfamily members in cancer. *Nature Reviews. Cancer*, *13*(5), 328–341. https://doi.org/10.1038/nrc3500

Wang, G., Biswas, A. K., Ma, W., Kandpal, M., Coker, C., Grandgenett, P. M., … Acharyya, S. (2018). Metastatic cancers promote cachexia through ZIP14 upregulation in skeletal muscle. *Nature Medicine*, *24*(6), 770–781. https://doi.org/10.1038/s41591-018-0054-2

Wang, S.-L., Zhuang, C.-L., Huang, D.-D., Pang, W.-Y., Lou, N., Chen, F.-F., … Yu, Z. (2016). Sarcopenia Adversely Impacts Postoperative Clinical Outcomes Following Gastrectomy in Patients with Gastric Cancer: A Prospective Study. *Annals of Surgical Oncology*, *23*(2), 556–564. https://doi.org/10.1245/s10434-015-4887-3

Wang, X., Han, H., Duan, Q., Khan, U., Hu, Y., & Yao, X. (2014). Changes of serum albumin level and systemic inflammatory response in inoperable non-small cell lung cancer patients after chemotherapy. *Journal of Cancer Research and Therapeutics*, *10*(4), 1019–1023. https://doi.org/10.4103/0973-1482.137953

Waning, D. L., Mohammad, K. S., Reiken, S., Xie, W., Andersson, D. C., John, S., … Guise, T. A. (2015). Excess TGF-β mediates muscle weakness associated with bone metastases in mice. *Nature Medicine*, *21*(11), 1262–1271. https://doi.org/10.1038/nm.3961

Weeks, B. K., Gerrits, T. A. J., Horan, S. A., & Beck, B. R. (2016). Muscle Size Not Density Predicts Variance in Muscle Strength and Neuromuscular Performance in Healthy Adult Men and Women. *Journal of Strength and Conditioning Research*, *30*(6), 1577–1584. https://doi.org/10.1519/JSC.0000000000001241

Wells, J. C. K. (2012). Ethnic variability in adiposity, thrifty phenotypes and cardiometabolic risk: addressing the full range of ethnicity, including those of mixed ethnicity: Ethnicity, body composition and cardiometabolic risk. *Obesity Reviews*, *13*, 14–29. https://doi.org/10.1111/j.1467- 789X.2012.01034.x

Wendrich, A. W., Swartz, J. E., Bril, S. I., Wegner, I., de Graeff, A., Smid, E. J., … Pothen, A. J. (2017). Low skeletal muscle mass is a predictive factor for chemotherapy dose-limiting toxicity in patients with locally advanced head and neck cancer. *Oral Oncology*, *71*, 26–33. https://doi.org/10.1016/j.oraloncology.2017.05.012

Wiedenmann, B., Malfertheiner, P., Friess, H., Ritch, P., Arseneau, J., Mantovani, G., … Von Hoff, D. (2008). A multicenter, phase II study of infliximab plus gemcitabine in pancreatic cancer cachexia. *The Journal of Supportive Oncology*, *6*(1), 18–25.

Wildi, S., Kleeff, J., Maruyama, H., Maurer, C. A., Büchler, M. W., & Korc, M. (2001). Overexpression of activin A in stage IV colorectal cancer. *Gut*, *49*(3), 409–417.

Windholz, T., Swanson, T., Vanderbyl, B. L., & Jagoe, R. T. (2014). The feasibility and acceptability of neuromuscular electrical stimulation to improve exercise performance in patients with advanced cancer: a pilot study. *BMC Palliative Care*, *13*, 23. https://doi.org/10.1186/1472-684X-13-23

Wolin, K. Y., Schwartz, A. L., Matthews, C. E., Courneya, K. S., & Schmitz, K. H. (2012). Implementing the exercise guidelines for cancer survivors. *The Journal of Supportive Oncology*, *10*(5), 171–177. https://doi.org/10.1016/j.suponc.2012.02.001

Wong, A. L., Seng, K. Y., Ong, E. M., Wang, L. Z., Oscar, H., Cordero, M. T., … Lee, S. C. (2014). Body fat composition impacts the hematologic toxicities and pharmacokinetics of doxorubicin in Asian breast cancer patients. *Breast Cancer Research and Treatment*, *144*(1), 143–152. https://doi.org/10.1007/s10549-014-2843-8

Xue, Q.-L. (2011). The Frailty Syndrome: Definition and Natural History. *Clinics in Geriatric Medicine*, *27*(1), 1–15. https://doi.org/10.1016/j.cger.2010.08.009

Yang, Y. X., Chong, M. S., Lim, W. S., Tay, L., Yew, S., Yeo, A., & Tan, C. H. (2017). Validity of estimating muscle and fat volume from a single MRI section in older adults with sarcopenia and sarcopenic obesity. *Clinical Radiology*, *72*(5), 427.e9-427.e14. https://doi.org/10.1016/j.crad.2016.12.011

Yip, C., Goh, V., Davies, A., Gossage, J., Mitchell-Hay, R., Hynes, O., … Mason, R. (2014). Assessment of sarcopenia and changes in body composition after neoadjuvant chemotherapy and associations with clinical outcomes in oesophageal cancer. *European Radiology*, *24*(5), 998–1005. https://doi.org/10.1007/s00330-014-3110-4

Yoshimura, Y., Wakabayashi, H., Yamada, M., Kim, H., Harada, A., & Arai, H. (2017). Interventions for Treating Sarcopenia: A Systematic Review and Meta-Analysis of Randomized Controlled Studies. *Journal of the American Medical Directors Association*, *18*(6), 553.e1-553.e16. https://doi.org/10.1016/j.jamda.2017.03.019

Young, J. F., Luecke, R. H., Pearce, B. A., Lee, T., Ahn, H., Baek, S., … Taylor, S. J. (2009). Human Organ/Tissue Growth Algorithms that Include Obese Individuals and Black/White Population Organ Weight Similarities from Autopsy Data. *Journal of Toxicology and Environmental Health, Part A*, *72*(8), 527–540. https://doi.org/10.1080/15287390802647203

Zaloga, G. P. (2006). Parenteral nutrition in adult inpatients with functioning gastrointestinal tracts: assessment of outcomes. *Lancet (London, England)*, *367*(9516), 1101–1111. https://doi.org/10.1016/S0140-6736(06)68307-4

Zhao, J., Liang, Y., Yin, Q., Liu, S., Wang, Q., Tang, Y., & Cao, C. (2016). Clinical and prognostic significance of serum transforming growth factor-beta1 levels in patients with pancreatic ductal

adenocarcinoma. *Brazilian Journal of Medical and Biological Research*, *49*(8). https://doi.org/10.1590/1414-431X20165485

Zhou, X., Wang, J. L., Lu, J., Song, Y., Kwak, K. S., Jiao, Q., … Han, H. Q. (2010). Reversal of cancer cachexia and muscle wasting by ActRIIB antagonism leads to prolonged survival. *Cell*, *142*(4), 531– 543. https://doi.org/10.1016/j.cell.2010.07.011

Zhuang, C.-L., Huang, D.-D., Pang, W.-Y., Zhou, C.-J., Wang, S.-L., Lou, N., … Shen, X. (2016). Sarcopenia is an Independent Predictor of Severe Postoperative Complications and Long-Term Survival After Radical Gastrectomy for Gastric Cancer: Analysis from a Large-Scale Cohort. *Medicine*, *95*(13), e3164. https://doi.org/10.1097/MD.0000000000003164

Tables

Table 1. Consensus definitions of sarcopenia

Abbreviations: BIA: bioelectrical impedance analysis, DXA: dual-energy X-ray absorptiometry

Table 2. Correlation of sarcopenia with clinical outcomes according to different cut-offs measured by computed tomography at the third lumbar vertebrae.

Table 3. Prevalence of sarcopenia

Abbreviations: BIA: bioelectrical impedance analysis, Cr: creatinine, CysC: cystatin C, L3 CT scan: computed tomography at the third lumbar vertebra level, LBM: lean body mass.

* Male; [(79.5 – 0.24 × mass (kg) – 0.15 × age (years)] × mass (kg) + 73.2 which is divided by square of height (m²).

Female; [(69.8 – 0.26 × mass (kg) – 0.12 × age (years)] × mass (kg) + 73.2 which is divided by square of height (m²).

Table 4. Sarcopenia and chemotherapy toxicity

Abbreviations: 5FU: 5-fluorouracil, ACF: 5FU, cisplatin, and adriamycin, BIA: bioelectrical impedance analysis, Cr: creatinine, CMF: cyclophosphamide, methotrexate, and 5FU, CysC: cystatin C, DCF: 5FU, cisplatin, and docetaxel, FOLFOX: 5FU, folinic acid, and oxaliplatin, L3 CT scan: computed tomography at the third lumbar vertebra level, LBM: lean body mass, N: number of patients included in the study, NLR: neutrophil lymphocyte ratio, Pts: patients, SCLC: small cell lung cancer, SMI: skeletal muscle index, OR: odds ratio.

* Male; [(79.5 – 0.24 × mass (kg) – 0.15 × age (years)] × mass (kg) + 73.2 which is divided by square of height (m²). Female; [(69.8 – 0.26 × mass (kg) – 0.12 × age (years)] × mass (kg) + 73.2 which is divided by square of height (m²).

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Table 5. Sarcopenia and targeted therapy toxicity

