

Influence of soil type on TiO2 nanoparticle fate in an agro-ecosystem

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1	Influence of soil type on TiO ₂ nanoparticle fate in an agro-
2	ecosystem
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17	Abstract:
18	Nanoparticles (NPs) and in particular TiO ₂ -NPs are increasingly included in commercial goods leading
19	to their accumulation in sewage sludge which is spread on agricultural soils as fertilizers in many
20	countries. Crop plants are thus a very likely point of entry for NPs in the food chain up to humans. So
21	far, soil influence on NP fate has been under-investigated. In this article, we studied the partitioning of
22	TiO ₂ -NPs between soil and soil leachate, their uptake and biotransformation in wheat seedlings and
23	their impact on plant development after exposure on 4 different types of soil with different
24	characteristics: soil texture (from sandy to clayey), soil pH, cationic exchange capacity, organic matter
25	content. Results suggest that a NP contamination occurring on agricultural soils will mainly lead to NP
26	accumulation in soil (increase of Ti concentration up to 302 % in sand) but to low to negligible transfer
27	to soil leachate and plant shoot. In our experimental conditions, no sign of acute phytotoxicity has
28	been detected (growth, biomass, chlorophyll content). Clay content above 6% together with organic
29	matter content above 1.5% lead to translocation factor from soil to plant leaves below 2.5% (<i>i.e.</i> below

- 30 13 mg Ti.kg⁻¹ dry leaves). Taken together, our results suggest low risk of crop contamination in an agro-
- 31 ecosystem.
- 32
- 33 <u>Keywords</u>: agriculture, bioavailability, ecotoxicology, food safety, plant, transfer

34 **1. Introduction**:

For more than a decade, **nanotechnologies** have attracted the interest of industrials and scientists throughout the world. The manipulation of matter at the nanometer scale opens up new possibilities in many different fields, such as industry, medicine and sustainable development. Nanotechnologies imply the use of a huge amount of nanoparticles (NPs), *i.e.* particles with at least one dimension below 100 nm. The new properties exhibited by NPs are also a source of questions about a potentially different behavior and toxicity.

41 The number of **commercial goods** available on the market and (officially) containing NPs has increased from 54 in 2005 to 2850 in 2016 in all fields of everyday life (cosmetics, food containers, 42 sporting goods, personal care, medicine, construction, agriculture)^{1,2}. TiO₂-NPs are one of the most 43 used NPs, appearing in about 25% of the products according to nano-databases^{1,2}. One can estimate a 44 45 yearly production of TiO₂-NPs comprised between 1 000 and 10 000 tons ³. Under their nanometric 46 form, they are mainly used for their photocatalytic properties for instance in construction materials (self-cleaning materials) or for their UV-adsorbing properties in cosmetics (sunscreens)³. Besides, they 47 can be found in small proportions in food additives ^{4,5}. 48

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50 This expansion of the use of TiO_2 -NPs is inevitably leading to an increased dissemination in the 51 environment. The release may be accidental, during the production or transport of NPs, or chronic, 52 during the life cycle of products containing NPs. It has been shown that, TiO₂-NPs contained in certain 53 textiles and paints are detached from their matrix during laundry or weathering and finally end up in wastewater treatment plants⁷⁻¹⁰ and in particular in sewage sludge, where they can reach a 54 concentration of 2 g.kg^{-1 11}. Moreover, the spreading of sewage sludge as soil fertilizer is a common 55 practice in agriculture with quantities reaching 3 tons (dry weight)/ha/year and is regulated for 56 57 instance in the US, in Australia and in Europe for sludge heavy metal content but nothing about NPs ^{12–15}. This leads to the dissemination of TiO₂-NPs in agricultural soils. In addition, the expected 58 environmental concentrations could be much higher locally with the intentional introduction of NPs in 59

the environment. For instance, thanks to their unique UV properties, TiO₂-NPs are used for purification
 purposes in both soil and water^{16–19}. Another critical case is the use of nanopesticides^{20,21}. Indeed, TiO₂ NPs are used in crop protection products for some years in order to optimize the product efficiency
 and then to eventually reduce the amount of inputs into the environment (sustainable development).
 The introduction of these products on the market is also expected to increase in the coming years^{22–}
 ²⁵.

66 Until very recently, most of the NP risk assessment studies have been carried out on a single 67 organism in simplified conditions (hydroponics), exposed to high concentrations of NPs during short 68 time, simulating acute contaminations ^{26,27}. Over the last years, studies related to the fate of NPs in the 69 environment evidenced a trend towards more realistic exposure scenario: soil exposure, longer 70 exposure period and lower NP concentrations ^{26,27}.

Studying the fate of TiO₂-NPs in soil is challenging. Indeed, soil is a very complex matrix. Moreover, the contribution of TiO₂ engineered nanomaterials is challenging to measure in the soil because of the high geogenic Ti background ($\approx 0.6\%$ of the terrestrial crust). Accordingly, to date, only modelling studies can help approximate the amount of engineered TiO₂-NPs that is ending up in the environment. The last article estimation forecasts the presence of 61 mg of TiO₂.kg⁻¹ of soil amended with sewage sludge ¹¹, making **agricultural soil** the main sink for nanomaterials in the environment and crop plants a privileged point of entry for NPs in the food chain up to humans.

Currently, knowledge about the interactions of NPs with soil is very limited. Once NPs reach the soil, they can be either physically retained or chemically adsorbed onto the surface of soil particles ²⁷. These interactions could mitigate NP phytotoxicity and bioavailability ²⁷, meaning that a same contamination could lead to very distinct consequences according to soil type. The literature published on plants exposed in soils suggested a very limited transfer of NPs to aerial parts. For a better risk assessment, it is thus of primary importance to understand how soil characteristics control NP fate in a crop system. For this purpose we exposed wheat, a widely grown crop over the world, to TiO₂-NPs (0, 100, 500 mg.kg⁻¹) in 4 different types of soil already containing geogenic Ti at different
concentrations. Ti quantification in soil, soil leachates and wheat leaves was assessed. Ti distribution
in roots and leaves and speciation in leaves were also determined using large scale research facilities.
Finally NP phytotoxicity was evaluated through plant development parameters: height, fresh and dry
weights and chlorophyll content.

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91 2. Material and Methods

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93 **2.1. Experimental set-up**

TiO₂-NPs with a nominal diameter of 25 nm and a crystalline phase mainly anatase (89%) were used
(see Figure S1 for more details).

A sand and 3 agricultural soils provided by Lufa Speyer (Germany) were chosen with different
characteristics (Table 1): texture ranging from sand to clayey loam, organic carbon content from 0 to
2.09%, cation exchange capacity (CEC) from 0 to 10 meq/100 g and pH from 5.1 to 7.2.

	Sand	Silty sand	Loamy sand	Clayey loam		
Particle size (%)						
<0.002 mm	0.0 ± 0.0	2.9 ± 0.8	6.4 ± 0.9	41.7 ± 1.3		
0.002-0.05 mm	0.0 ± 0.0	9.1 ± 1.4	11.6 ± 0.7	36.1 ± 2.2		
0.05 – 2 mm	100.0 ± 0.0	88.0 ± 1.0	82.0 ± 0.7	22.2 ± 1.5		
Water holding capacity						
g/100 g	25.3 ± 2.8	31.8 ± 3.0	46.5 ± 6.0	39.6 ± 5.0		
Weight per Volume						
g/1000 mL	1756 ± 6	1430 ± 57	1220 ± 78	1330 ± 96		
Organic Carbon						
%	0.0 ± 0.0	0.74 ± 0.14	2.09 ± 0.40	1.69 ± 0.17		

		pH value		
0.01M CaCl ₂	6.3 ± 0.0	5.1 ± 0.5	5.5 ± 0.1	7.2 ± 0.1
	Cat	tion Exchange Capa	city	
Meq/100g	<2	4.0 ± 1.0	10.0 ± 0.5	22.0 ± 6.0
		Leachates		
mL	10.6 ± 0.1^{c}	9.1 ± 0.2^{bc}	8.0 ± 0.3^{ab}	6.3 ± 0.6^{a}

Table 1. Soil characteristics (soil provided by Lufa Speyer - Germany: silty sand: soil 2.1, loamy sand:
 soil 2.2, clayey loam: soil 6S). Characteristics were provided by the supplier. For leachates, different
 lowercase letters indicate significant differences (p<0.05) among all treatments.

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105 Exposure concentrations of TiO₂-NPs represented a realistic condition (100 mg.kg⁻¹) and a higher and 106 plausible concentration after several years of sewage sludge spreading (500 mg.kg⁻¹). TiO₂-NP 107 suspensions (100 or 500 mg.L⁻¹ and ultrapure water as control) were mixed with soil with a ratio of 1:1 (w:w) for 12 hours on a shaker (200 rpm) in containers wrapped with aluminum foil. This mixing 108 109 strategy was chosen to ensure the most homogeneous matrix as possible for plant exposure. After this 110 step, the mixture was poured in a folded standard filter paper (Fisherbrand, Dia/Size 330 mm) and 111 separated in soil leachates and soil (after 8h of settling). After weighting, soil leachates were 112 evaporated and prepared for Ti quantification to determine the Ti mobility.

6 individual replicates for each type of soils (4) and each level of contamination (3) were set-up with each a wheat seed (*Triticum aestivum* L., cv. Courtaud). All experiments were carried out in a growth chamber under controlled conditions: day/night photoperiod (16/8 h), 100 µmol/m²/s, day/night temperature, (24/20±1 °C) and day/night relative humidity (70/75%). Leaf height and chlorophyll content (determined using a SPAD-502 chlorophyll meter, Minolta Camera Co., Japan) were assessed every day for 3 weeks until harvest. At the end of the experiment fresh leaf weight and dry leaf weight after 48h in an oven at 70°C were recorded.

121 **2.2. Bulk Ti quantification by ICP-AES**

122 Different tests have been performed to ensure a proper acidic digestion of TiO₂-NPs which represents 123 a critical step to avoid underestimation of Ti concentration in the samples. Several digestion protocols 124 adapted from the literature have been compared (Figure S2A) with different acid cocktails, temperatures and evaporation steps. The best digestion yield was obtained with a protocol adapted 125 126 from Mariet et al. (2008) with a mix of HNO₃, NH₄F, HClO₄ and H₂O₂ with several evaporation steps at 100 °C in a DigiPrep[®] system (see Supporting Information for more details)²⁸. The yield was about 88% 127 128 and reproducible. This test demonstrates that the digestion protocol needs to be re-optimized for each 129 TiO₂-NP. Indeed, in one of our previous experiments using 4 nm anatase TiO₂-NPs, the protocol from 130 Mariet et al. was not efficient²⁹. The linearity of the digestion was also assessed with increasing TiO₂-NP concentrations (Figure S2B). Finally the stability of the solution after digestion over a month was 131 132 monitored (Figure S2C). After digestion, samples (soils, leachates, plant shoots) were analyzed using 133 Inductively Coupled Plasma – Mass Spectrometry (ICP-AES IRIS, Thermo Scientific). Quality of sample 134 preparation was checked with blank samples containing only chemicals and with reference samples 135 (tomato leaves, NIST RM 1573a) and permitted to validate sample preparation and ICP analysis (Figure 136 S2bis, Table S1).

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138 **2.3. Spatially resolved Ti semi-quantification by μPIXE/RBS**

A combination of micro-particle induced X-ray emission (μPIXE) and Rutherford Backscattered
 spectroscopy (RBS) has been used for Ti semi-quantification. μPIXE/RBS quantification of Ti was chosen
 as a spatially resolved technique to avoid mixing the Ti signal coming from NPs adsorbed at the surface
 of roots with Ti really absorbed inside the root tissues.

Wheat roots were thoroughly washed with deionized water to take off NPs lightly bound to the surface
and immersed in a droplet of resin (Tissue Teck Sakura[®]) to be immediately cryo-fixed by plunging the

sample in isopentane cooled with liquid nitrogen. Samples were then cut in thin cross-sections (50 μm)
using a cryo-microtome (Leica) and finally freeze-dried (48h, -52°C, 0.01 mbars).

Freeze-dried sections were analyzed at the nuclear microprobe available at the Atomic Energy Commission (CEA) Center of Saclay (France) with a proton source of 3 MeV, a beam focused to 2.5 μm and a current intensity of 500 pA. Data processing was performed using Rismin³⁰ software to define regions of interest and extract spectra, and SIMNRA³¹ and GUPIX³² codes to reduce RBS and PIXE data, respectively. The Ti/Ca ratio was used as a Ti enrichment indicator as Ca is an abundant endogenous element.

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154 **2.4.** Ti distribution and speciation by μXRF and μXANES

Samples were prepared as described previously for nuclear microprobe experiments (section 2.3.).
 Synchrotron experiments were carried out on LUCIA beamline at SOLEIL (Saint Aubain, France)³³. Micro
 X-ray fluorescence (μXRF) maps were recorded above the Ti-K edge (5.1 keV) with a step size of 5 μm
 and a dwell time of 2000 ms. μXRF data were fitted using PyMCA software ³⁴.

Ti-K edge (4.5 to 5.5 keV energy range, 0.5 eV step) micro X-ray absorption near edge spectroscopy (µXANES) spectra were recorded in regions of interest of the fluorescence maps. XANES spectra on reference compounds were recorded during previous experiments ^{29,35}. For the fitting, 3 groups of references were created: anatase, rutile and others (containing Ti-oxysulfate, amorphous Ti oxide and Ti acetate). XANES data treatment was performed using Athena software ³⁶ as previously ^{29,35}.

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165 **2.5. Statistical analysis**

Data were checked for normality (Shapiro test) and homoscedasticity (Bartlett or Levene's tests). They were compared using two-way ANOVA, with soil type and TiO₂-NP concentration as main effects, and the interaction (soil type x NP concentration). If the whole set of data did not follow a normal distribution even after transformation, Student T test, Wilcoxon test, one-way ANOVA or Kruskal– Wallis test were performed on subset of data (according to soil type or NP concentration). Post hoc tests (Tukey's HSD or Dunn's test) were applied to detect differences between groups of samples.

Differences were considered significant when *P*< 0.05. All statistical analyses are gathered in Table
S21.

To study the global influence of TiO₂-NPs on plant elemental content, MANOVA based on Pillai's trace
test was used on the whole set of micro- and macronutrients measured by ICP-AES.

All statistical analyses were performed using the R statistical software (version 3.1.3) ³⁷, with car ³⁸,
 multcompView ³⁹, pgirmess ⁴⁰ and Ismeans ⁴¹ packages.

- 178
- 179

180 3. Results and discussion

181 **3.1.** Ti partitioning between soil and soil leachates

182 As stated earlier, Ti is a geogenic element of the earth crust. As such, the 4 natural soils (control condition) contained Ti concentrations ranging from 43 to 1671 mg.kg⁻¹ with sand having the lowest 183 184 concentration followed by the group of silty and loamy sands and finally the clayey loam being the 185 richest in Ti (P<0.0001, Figure 1A). Ti concentration in natural soils was positively correlated with the fraction of soil particles in the 0.002 to 0.005 mm size range (r²= 0.9997, Table 1) and to a lesser extent 186 187 to the fraction <0.002 mm (r²=0.9618). After addition of 100 mg.kg⁻¹ of TiO₂-NPs, the Ti concentration in soils did not significantly increase in any soil. After addition of 500 mg.kg⁻¹ of TiO₂-NPs, the increase 188 189 in Ti was only significant for sand and silty sand (P<0.0001), i.e. soils with the lowest clay fraction.

Those results can be confronted with Ti concentrations in soil leachates (Figure 1B). In natural soils, Ti concentrations in the leachates after 12h mixing were respectively 0.03, 0.18, 0.07 and 0.23 mg.L⁻¹ for sand, silty sand, loamy sand and clayey loam. After addition of 100 mg.kg⁻¹ of TiO₂-NPs, only the sand leachate had a significantly higher Ti concentration (13.18 mg.L⁻¹, P=0.0379) with a more than 400 fold increase in comparison with the control sand. This is also the soil in which the leachate volume recovered was the highest (Table 1): 71% of the 15 mL of the contamination suspension was collected in the leaching compartment. This fraction was decreased to 42% in the case of the clayey loam soil. 197 Crossing this result with Ti concentration in soil, one can conclude that most of the TiO_2 -NP suspension 198 added in the 100 mg.kg⁻¹ condition went through the sand column and ended up in the soil leachate. 199 After addition of 500 mg.kg⁻¹ of TiO_2 -NPs, Ti concentration in soil leachate was significantly increased 200 as compared to the control for all soils except for the clayey loam soil (*P*=0.0019 for silty sand and *P*= 201 0.0031 for loamy sand).



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Figure 1. Ti concentration in the soil system: soil (A) and soil leachate (B) determined by ICP-AES.
Different lowercase letters indicate significant differences (p<0.05) among all treatments in A or among
treatment within a soil type in B. Capital letters in A highlight significant differences among soil types.
(n=6).

208 If a contamination occurs on a sand, most TiO₂-NPs will likely go through the soil and reach the aquifers. 209 Only a small fraction will remain in the soil if a high dose (500 mg.kg⁻¹) of NPs is applied. In the silty 210 sand, Ti increase in both soil and soil leachate was only detectable for the highest concentration added. For the loamy sand, the high Ti natural background (> 500 mg.kg⁻¹) shadowed the addition of NPs in 211 212 the soil and Ti increase was visible only in the leachate at the highest NP concentration added. Finally, 213 in the clayey loam, all added Ti was retained in the soil but did not increase significantly Ti 214 concentration also because of the very high geogenic Ti level. Depth column studies would be needed 215 to confirm these results about NP mobility and gain a deeper understanding.

216 Soils with coarser texture, lower organic carbon content as well as lower CEC (sand and silty sand) were 217 the ones in which NPs displayed the highest availability. This result is in agreement with previous literature ^{42–45}. Authors suggested that soils with a higher clay content have also smaller pore size and 218 219 larger surface area which increase both electrostatic interactions and physical straining explaining the 220 lower transport of NPs to soil leachates. Likewise, Cu-NP translocation was observed to be low in soils enriched in organic matter and high in clay ⁴⁶. Another important point that could be considered is the 221 222 interaction of NPs with other potential contaminants of the soil. Indeed, it has been shown in the 223 literature that addition of TiO₂-NPs in sandy soils can improve Cu and Pb mobility in soils ^{43,47}. Sewage sludge also often contains a certain amount of heavy metals ¹⁵; NP contamination of sewage sludge 224 225 amended soils could thus represent a threat to both terrestrial and aquatic ecosystems through higher 226 heavy metal bioavailability and leaching towards aquifers. Finally, different microbial communities 227 likely occur in different soils, and might have a role on TiO₂ fate and speciation over long term 228 exposure.

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230 3.2. Ti uptake in wheat roots

Spatially resolved techniques were employed to investigate Ti uptake in wheat roots exposed in control
 soil and soil contaminated with 100 mg.kg⁻¹ of TiO₂-NPs (the most environmentally relevant condition).

233 Figure 2A-D displays Ti distribution in roots of wheat seedlings grown in the 4 different natural soils. Ti 234 was mainly detected adsorbed at the surface of the roots but also in the parenchyma with maximum 235 fluorescence intensity of 42 counts in average for sand, loamy sand and clayey loam and up to 133 236 counts in silty sand roots. In roots grown in contaminated soils (Figure 2E-H), the same distribution 237 pattern appeared, with Ti mainly detectable on root surface and less Ti observed adsorbed in the clayey 238 loam soil in comparison with the 3 other ones. However, the maximum fluorescence intensities were 239 increased in root inner tissues (parenchyma and vascular cylinder) especially for roots grown in sand 240 and silty sand. In first approach, these results seem to correlate with those obtained in the Ti 241 partitioning experiment: Ti added to sandy soil is more available and thus internalized in roots, while 242 Ti added in soils with more clay and organic matter is more retained by the soil matrix and lead only 243 to a slight Ti fluorescence signal increase in wheat roots.

244 ICP-AES measurements of Ti content in roots would not be reliable because Ti can be heavily adsorbed 245 on their surface, leading to over-estimation of Ti absorption. Thus, to have a proper quantitative 246 assessment of Ti accumulation in roots, thin sections were analyzed using μ PIXE coupled to μ RBS, 247 which permitted the quantification of all the chemical elements of the periodic table, light elements 248 (C, H, O, N) included (Figure 2I). The main difference evidenced by the analysis was the one related to 249 soil type (P = 0.0003): roots from clayey soils contained significantly less Ti than the roots grown in 250 loamy sand and silty sand. Wheat roots accumulated significantly more Ti after addition of TiO₂-NPs in silty sand only (P = 0.0008). In this condition, Ti content was multiplied by a factor of 7. Surprisingly, in 251 252 contrast with µXRF results, Ti content in roots grown on sand did not show any significant increase (P 253 = 0.4699). However, this is in agreement with the fact that Ti content in sand after addition of 100 254 mg.kg⁻¹ TiO₂-NPs did not increase significantly. One possible explanation for this discrepancy is that µXRF technique has a lower limit of detection than µPIXE/RBS⁴⁸. Overall, no correlation was detectable 255 256 between Ti concentration in soil and Ti/Ca content in roots (r=0.111, P=0.742).



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Figure 2. Ti in wheat roots after a 3 week exposure: elemental distribution by μ XRF of Ti (in red) and Ca (in green) in root sections of wheat grown in control soils (A-D) or in contaminated soils (E-F). Numbers in red (fluorescence counts) and arrows indicate the highest fluorescence intensity detected inside the root . ep. epidermis, p. parenchyma, v.c. vascular cylinder. Scale bar = 50 μ m. Semi-quantitative analysis of Ti by μ PIXE/RBS in root sections of wheat grown in natural soils and soils contaminated with 100 mg.kg⁻¹ TiO₂-NPs (I). Capital letters indicate significant differences among soil types and * indicate significant differences between exposed plants and their corresponding control (p<0.05) (n=3).

Ti speciation was then analyzed by μ XANES on high intensity spots identified by μ XRF (Figure 3). This analysis was performed in roots grown in the 4 different types of soils and in both natural and contaminated (100 mg.kg⁻¹ added NPs) conditions. Different chemical forms of Ti were detected in wheat grown in natural soils from mainly rutile in clayey loam and sand (with respectively 73 and 77% 270 of rutile crystalline form) to mainly anatase in loamy sand (63% anatase). Anatase and rutile forms of TiO₂ correspond to an octahedral arrangements of the O atoms around Ti. In the natural silty sand, 271 272 there is a shift of the A3 peak towards the lower energies (about 0.5 eV) with an increase of the peak height and area which suggests a different conformation of the TiO unit going from octahedral towards 273 274 trigonal bipyramidal (5 coordinations) or tetrahedral (4 coordinations) conformations ⁴⁹. After 275 exposure to TiO₂-NPs (89% anatase, 11% rutile, Figure S1), the ratio between anatase and rutile in 276 comparison with the same ratio in the control roots increased in the roots of all 4 conditions 277 highlighting the contribution of the mainly anatase added NPs. The increase was more important in 278 the case of plants grown in the soils with coarser texture (sand and silty sand).

There are not many studies evaluating NP fate in plants after soil exposure even though this number 279 280 has been increasing over the last few years. But there are even fewer articles reporting the fate of NPs in roots (Majumdar et al. 2016 on CeO₂-NPs ⁵⁰ or Gao et al., 2018 on CuO-NPs⁵¹). Indeed it is very 281 282 challenging to obtain this type of data and only spatially resolved techniques are able to provide 283 reliable data getting rid of the geogenic signal surrounding the roots. Moreover, if one wants to 284 perform a comparative study, the technique also needs to be quantitative which leaves the scientific 285 community with few tools mainly based on spectroscopy. In this study by combining 4 different 286 techniques namely µXRF, µPIXE/RBS and µXANES, we were able to map Ti distribution in wheat roots 287 with a good sensitivity, quantify it and investigate its speciation in vivo. Our results demonstrate that 288 the added TiO₂-NPs can be taken up by plant roots when exposure occurs in soil and in particular in 289 sand and silty sand. Considering the concentration of NPs added to natural soils (no significant increase 290 of Ti concentration in soils after addition of 100 mg.kg⁻¹, Figure 1A), our results also suggest that Ti 291 from NPs was taken up by roots preferentially than geogenic Ti (no significant Ti concentration increase 292 in soil vs. increase in roots). This highlights that the added Ti had a different behavior than the natural 293 one and a different bioavailability which justifies the fact to study the impact of a TiO₂-NP 294 contamination in the environment even though plants have evolved with high Ti concentrations in soils 295 for ages.



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Figure 3. Ti speciation assessed by μ XANES in wheat roots grown in control soils (Ctl) or soils contaminated with 100 mg.kg⁻¹ TiO₂-NPs (+TiO₂) after a 3 week exposure. A. anatase, R. rutile, other gathers Ti-acetate, Ti-oxysulfate and amorphous Ti oxyde. In colors: experimental spectra, in dashed black: fits. Red dashed vertical line shows the position of A3 peak in [Ti]₆ spectra. The fit of Ctl silty sand is not displayed since a major reference compound was not available (trigonal bipyramidal and/or tetrahedral form(s)) for a proper fitting.

Moreover, the results obtained in this work are also in agreement with several similar articles of the literature. For instance, in a study carried out with the same NPs (but much higher concentration) and a soil similar to our silty sand, Ti based NPs have been detected (TEM-EDX) in the roots of red clover exposed to 1000 mg.kg⁻¹ after 14 weeks of exposure⁵². Previously in 2011, Du *et al.*, also detected by TEM, TiO₂-NPs in the roots of wheat grown for 2 weeks in an artificially contaminated loamy clay soil⁵³. Finally those results are also in line with those we previously demonstrated for wheat seedlings

exposed for 7 days to the same nanomaterial in hydroponics: anatase TiO₂-NPs were also evidenced in
 the roots by various techniques ^{54,55}.

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313 **3.3. Ti translocation to wheat leaves**

Ti accumulation in aerial parts was quantified by ICP-AES (Figure 4A). Wheat leaves had a basal Ti concentration of 9 mg.kg⁻¹ in average in control soils. The only significant Ti concentration increase in leaves after exposure was detected for seedlings grown in sand after addition of 500 mg.kg⁻¹ TiO₂-NPs (*P*<0.0001) with a concentration reaching more than 100 mg Ti.kg⁻¹ DW. The same trend was also noticeable in silty sand but not significant (from 7 in natural soil to 19 mg.kg⁻¹ in the most contaminated condition). Plants grown on clayey loam had the lowest Ti concentration in all conditions (4.4 mg.kg⁻¹ on average) even though this was the soil with the highest Ti concentration.

321 Another factor of interest after contamination of an agro-ecosystem is the translocation factor which 322 corresponds to the fraction of Ti the plant is able to transfer from the soil to its leaves (Figure 4B). Even 323 though the clayey loam soil was the richest in Ti - mainly geogenic Ti as demonstrated earlier - it was 324 also the one in which Ti translocation factors were significantly lower than in other soils with an 325 average of 0.4% (P < 0.0001). The addition of TiO₂-NPs to silty sand, loamy sand and clayey loam had 326 no influence on Ti translocation factors whatever the concentration added. Translocation factor was 327 only increased after addition of NPs in sand (P= 0.0108). Sand was the soil with the lowest Ti content, but with the highest translocation factors ranging from 23.4 to 61.9% meaning that Ti was more 328 bioavailable in sand soil. All conditions considered, a negative significant correlation was evidenced 329 330 between Ti in soil and Ti in leaves (r= -0.374, P=0.0130).



Figure 4. A. Ti quantification in leaves and B. Translocation factors ($[Ti]_{leaf}$ / $[Ti]_{soil}$) determined by ICP-AES in seedlings grown in control soils and soils contaminated with 100 or 500 mg.kg⁻¹ TiO₂-NPs, after

a 3 week exposure. Lowercase letters indicate significant differences (p<0.05) among NP
 concentrations (control, 100 or 500 mg.kg⁻¹) within a soil type in A or among all treatments in B. Capital
 letters indicate significant differences among soil types. (n=6)

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338 Results in the literature also report very little to no Ti translocation to leaves when plants were exposed in soils. For instance, tomato plants exposed to 250 mg.kg⁻¹ 25 nm TiO₂-NPs in calcinated fuller earth 339 for 66 days accumulated between 25 to 83 mg Ti.kg⁻¹ approximately in their aerial parts with no 340 341 significant difference with the control condition⁵⁶. Likewise, Gogos et al. did not evidenced any 342 significant increase in red clover shoot and wheat grain after exposure to the same TiO₂-NPs as ours in a silty sand contaminated with concentrations ranging from 1 to 1000 mg.kg^{-1 52}. TiO₂-NPs were also 343 344 not detected in the leaves of wheat plants grown on a contaminated loamy clay (clay: 32.6%, OM content: 4.6%, added NPs: 90 mg.kg⁻¹) for 2 weeks ⁵³. Ti was detected up to the fruits of cucumber 345 346 plants after exposure to TiO₂-NPs in a sandy loam soil contaminated with 750 mg.kg⁻¹ for 150 days but with no quantitative data or comparison to control ⁵⁷. Results were also similar for other NPs and plant 347 species (CuO and lettuce⁵⁸, CeO₂ and radish⁵⁹), unless very high concentrations were introduced (>1000 348 $mg.kg^{-1}$). 349

350 Others in the literature also investigated soil characteristic influence on NP translocation, in particular the impact of organic matter. Interestingly most of the available studies concluded that increasing 351 352 organic matter leads to higher metal concentration in plant shoots (Ce in kidney bean ⁵⁰, Ce in fescue ⁶⁰, Zn in cucumber ⁶¹). However, in another study, about 10 times more Zn was found in the shoot of 353 354 wheat grown on "acid soil" (with 1.6% OM and 8% clay) than in "calcareous alkaline soil" (with 4.1% 355 OM and 28% clay) after exposure to ZnO NPs. This was related to a higher proportion of soluble Zn in the acidic soil (200 times more) ⁶². Authors tried to confirm the relationship between OM and metal 356 357 translocation by comparing Zn internalization in plants grown on sand with increasing concentrations 358 of humic acid, but they were not able to reproduce the results obtained in natural soil: adding humic 359 acid did not influenced Zn internalization. Finally, corn plants exposed to CeO₂ NPs in untreated soil 360 contained significantly less Ce in roots but more Ce in shoot than plants exposed in the same soil 361 enriched in organic matter. In parallel, ICP measurements demonstrated that the organic soil 362 contained more Ce in the soil solution than the untreated soil (375 vs. 18 mg/100 mL soil solution) ⁶³ 363 which is in contradiction with the previous study. Thus it seems that the correlation between OM 364 content and NP/metal uptake is still not clear. OM could also have an indirect effect on plant 365 development, possibly leading to an increased foliar area and thus plant evapotranspiration which 366 could in turn increase NP uptake by plants. Those discrepancies might also be related to plant species and their different strategies for metal uptake (monocot vs. dicot ⁶⁴) or related to NP surface charge 367 368 or biotransformation. Indeed, XANES studies have demonstrated that Zn is mainly detected in soil and plant shoot as dissolved ions recomplexed to either phosphate, phytate, citrate or histidine ^{65,66} while 369 370 Ce has been detected as Ce(IV)O₂-NPs in roots and Ce(III) carboxyl in the stem and leaves, suggesting 371 that the dissolution of CeO₂-NPs took place in the plants rather than in the soil. Ce would then be 372 present in the soil as NPs while Zn would be under ionic form which implies different interactions with 373 the soil particles and thus different uptake dynamics. Ti XANES spectra in plant leaves, in this study 374 and in our previous work, were dominated by the anatase NP crystalline form which indicates that TiO₂-NPs are more inert than ZnO-NPs and would behave in the soil more like CeO₂-NPs. Obviously
 more studies are needed to really understand how OM impacts different types of NP uptake by plants.

Soil texture, and in particular the amount of clay, is another parameter of interest to determine the fate of NPs. Our results showed that an increased percentage of clay in the soil lead to a decrease of Ti uptake in plant shoots. They are in agreement with a study from Zhang *et al.*, in which radish roots and shoots tend to accumulate more Ce after exposure to CeO₂-NPs in the loamy sand (76% sand) than in the silty loam (16% sand) ⁵⁹. Likewise, Layet *et al.*, concluded that increasing clay fraction enhanced the retention of CeO₂-NPs in soil and thus decreased its uptake by fescue ⁶⁰.

383

384 3.4. TiO₂-NP phytotoxicity

TiO₂-NP toxicity was evaluated during the time course of the exposure via seedling shoot length and chlorophyll content. The main noticeable difference was linked to soil type with plants growing in sand being the smallest followed by plants grown in silty sand, loamy sand and clayey loam (for instance at day 16, P < 0.0001) (Figure S3) but no significant difference was highlighted between NP treatment (for instance at day 16, P = 0.528). Results for chlorophyll content were similar (data not shown): the only difference was related to soil type (P= 0.0080) but not to NP addition (P=0.9290).

391 After 3 weeks, leaf fresh and dry biomasses were recorded (Figure 5). Fresh leaf biomass was only 392 influenced by soil type (P<0.0001) with 3 groups of individuals: sand < silty sand and loamy sand < 393 clayey loam (Figure 5A). Dry leaf biomass results were similar (P=0.0008) with plants grown in sand 394 having the lowest biomass with 20 mg on average, followed by plants grown in silty sand, loamy sand 395 and clayey loam with respectively 36 mg, 41 mg and 43 mg (Figure 5B). The ratio between fresh matter 396 and dry matter seems to be a more sensitive parameter which evidenced a soil effect (P=0.0080): 397 plants grown on clayey loam had 19% of dry matter vs. plants grown in loamy sand with 25% (Figure 398 5C). Moreover, this ratio also highlighted a NP concentration effect (P=0.0410) with plants exposed in soil with 500 mg.kg⁻¹ TiO₂-NPs having a lower content in dry matter as compared with plants grown in control soils. It has already been shown previously that NPs are able to interact with water uptake processes 67 .



402

Figure 5. TiO₂-NP phytotoxicity on wheat biomass (fresh: A, dry: B, ratio between fresh and dry: C) after
a three week exposure to different concentrations of NPs (0, 100, 500 mg.kg⁻¹) in different types of soils.
Capital letters indicate significant differences (p<0.05) among soil types (n=6).

406

407 TiO₂-NP impact on plant hydromineral nutrition was also investigated at harvest after ICP-AES 408 measurements of plant macro- (Ca, K, P, S) and micro- (Al, B, Ba, Cu, Fe, Mg, Mn, Na, Si, Zn) nutrient 409 concentrations. Again, the main effect detected was a soil effect (Manova with Pillai's trace, $F_{3,32}$ = 10.4, 410 P < 0.0001) but no NP specific effect was highlighted (Manova with Pillai's trace, $F_{2,33}$ = 1.5, P = 0.1210). In the literature, no major acute toxicity has been reported after plant exposure to TiO₂-NPs in soil. For instance, there was no significant difference in catalase and ascorbate peroxidase activities in cucumber exposed to up to 750 mg.kg⁻¹ TiO₂-NPs. Likewise, no effect on chlorophyll content was detected until 500 mg.kg⁻¹ NPs (but an increase was seen at 750 mg.kg⁻¹) ⁵⁷. The same results were also obtained previously in hydroponics for short term exposure ^{29,54,55}. It would be interesting to confirm this lack of toxicity through multigenerational tests and through more in-depth endpoints (-omics approaches).

418

419 Conclusion

The results obtained in this study demonstrated that TiO₂-NPs can be significantly taken up by wheat 420 seedlings after a 3 week exposure in a sandy soil in which 500 mg.kg⁻¹ of engineered NPs have been 421 422 added. This is also in this soil type that NPs were able to reach in higher quantities the leachates. Thus 423 if a contamination occurs in a sand, NPs will be very mobile (towards aquifers) and bioavailable to crop 424 plants representing a risk for trophic transfer. However, sand is not a relevant medium for agriculture. 425 The same trend has been highlighted in silty sand but in smaller proportions. In contrast, in loamy sand 426 and clayey loam no difference has been evidenced after addition of NPs to the system: no Ti 427 concentration increase in the soil, partly due to the high Ti geogenic background, no increase in plant roots and leaves and little to no increase in the soil leachates. This difference in TiO2-NP behavior can 428 429 be related to 3 characteristics of the soil used: organic carbon content, clay content and CEC, but the 430 experimental plan do not allow to go further. In all conditions, no acute sign of phytotoxicity was detected in the endpoints analyzed. The overall conclusion for agro-ecosystems is that TiO₂-NP 431 432 contamination (after a short term exposure: 3 weeks) seems to have little impact on plant health and 433 low chance of uptake as well as low risk of leaching to the aquifers. The soil displaying the highest risk 434 for food safety would be the silty sand but this soil has little nutrients for plant growth and is not the 435 best featured for agriculture. However, those results also imply that TiO₂-NPs will remain in the soil 436 with possible implications for soil micro and macro-organisms. It is thus important to understand NP

437	fate in soils upon weathering on a long term experiment. Finally, an experiment covering the full life
438	cycle of different types of crop plants (fruit crop, leaf crop, root crop) is also needed to confirm data
439	obtained here and further investigate the potential risk on silty sand soil.

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453 Supporting Information is available, it contains 4 figures and 2 tables.

454 References

- 455 1. www.nanotechproject.org. The project on emerging nanotechnologies. (2015).
- 456 2. consumer products. *the nanodatabase* Available at: http://nanodb.dk/en/analysis/consumer-

457 products/.

- 458 3. Piccinno, F., Gottschalk, F., Seeger, S. & Nowack, B. Industrial production quantities and uses of
- 459 ten engineered nanomaterials in Europe and the world. J. Nanoparticle Res. **14**, 1109 (2012).
- 460 4. Jovanovic, B. Critical Review of Public Health Regulations of Titanium Dioxide, a Human Food
 461 Additive. *Integr. Environ. Assess. Manag.* 11, 10–20 (2015).
- 462 5. Weir, A., Westerhoff, P. K., Fabricius, L. & von Goetz, N. Titanium Dioxide Nanoparticles in Food
 463 and Personal Care Products. *Env. Sci Technol* 46, 2242–2250 (2012).
- 464 6. Robichaud, C. O., Uyar, A. E., Darby, M. R., Zucker, L. G. & Wiesner, M. R. Estimates of Upper
- Bounds and Trends in Nano-TiO2 Production As a Basis for Exposure Assessment. *Env. Sci Technol* 43, 4227–4233 (2009).
- 467 7. Windler, L. *et al.* Release of Titanium Dioxide from Textiles during Washing. *Environ. Sci. Technol.*468 46, 8181–8188 (2012).
- 469 8. Mackevica, A. & Foss Hansen, S. Release of nanomaterials from solid nanocomposites and
- 470 consumer exposure assessment a forward-looking review. *Nanotoxicology* **10**, 641–653 (2016).
- 471 9. Zhang, Y., Leu, Y.-R., Aitken, R. J. & Riediker, M. Inventory of Engineered Nanoparticle-Containing
- 472 Consumer Products Available in the Singapore Retail Market and Likelihood of Release into the
 473 Aquatic Environment. *Int. J. Environ. Res. Public. Health* **12**, 8717–8743 (2015).
- 474 10. Mitrano, D. M., Motellier, S., Clavaguera, S. & Nowack, B. Review of nanomaterial aging and
 475 transformations through the life cycle of nano-enhanced products. *Environ. Int.* 77, 132–147
 476 (2015).
- 477 11. Sun, T. Y., Bornhöft, N. A., Hungerbülher, K. & Nowack, B. Dynamic probabilistic modeling of
- 478 environmental emissions of engineered nanomaterials. *Env. Sci Technol* 4701–4711 (2016).

- 479 12. Gottschalk, F., Sonderer, T., Scholz, R. W. & Nowack, B. Modeled Environmental Concentrations
- 480 of Engineered Nanomaterials (TiO(2), ZnO, Ag, CNT, Fullerenes) for Different Regions. *Environ.*481 *Sci. Technol.* 43, 9216–9222 (2009).
- 482 13. Kim, B., Murayama, M., Colman, B. P. & Hochella, M. F. Characterization and environmental
- 483 implications of nano- and larger TiO2 particles in sewage sludge, and soils amended with sewage
- 484 sludge. J. Environ. Monit. **14**, 1128–1136 (2012).
- 485 14. Gottschalk, F., Sun, T. & Nowack, B. Environmental concentrations of engineered nanomaterials:
 486 review of modeling and analytical studies. *Env. Pollut* 181, 287–300 (2013).
- 487 15. Sharma, B., Sarkar, A., Singh, P. & Singh, R. P. Agricultural utilization of biosolids: a review on
- 488 potential effects on soil and plant grown. *Waste Manag.* **64,** 117–132 (2017).
- 489 16. Zimbone, M., Cacciato, G., Boutinguiza, M., Privitera, V. & Grimaldi, M. G. Laser irradiation in
- 490 water for the novel, scalable synthesis of black TiOx photocatalyst for environmental
- 491 remediation. *Beilstein J. Nanotechnol.* **8**, 196–202 (2017).
- 492 17. Bessa da Silva, M., Abrantes, N., Nogueira, V., Gonçalves, F. & Pereira, R. TiO2 nanoparticles for
- 493 the remediation of eutrophic shallow freshwater systems: Efficiency and impacts on aquatic
- 494 biota under a microcosm experiment. *Aquat. Toxicol. Amst. Neth.* **178**, 58–71 (2016).
- 495 18. Wu, M. *et al.* Simultaneous biological-photocatalytic treatment with strain CDS-8 and TiO2 for
- 496 chlorothalonil removal from liquid and soil. J. Hazard. Mater. **320**, 612–619 (2016).
- 497 19. Sanz Lobón, G. *et al.* Efficient electrochemical remediation of microcystin-LR in tap water using
 498 designer TiO2@carbon electrodes. *Sci. Rep.* 7, (2017).
- 20. Bergeson, L. L. Nanosilver pesticide products: What does the future hold? *Environ. Qual. Manag.*500 **19**, 73–82 (2010).
- 501 21. Sastry, K., Rashmi, H. & Rao, N. Nanotechnology patents as R&D indicators for disease
- 502 management strategies in agriculture. *J. Intellect. Porperty Rights* **15**, 197–205 (2010).
- 503 22. Khot, L. R., Sankaran, S., Maja, J. M., Ehsani, R. & Schuster, E. W. Applications of nanomaterials in
- agricultural production and crop protection: A review. *Crop Prot.* **35,** 64–70 (2012).

- 505 23. Ghormade, V., Deshpande, M. V. & Paknikar, K. M. Perspectives for nano-biotechnology enabled
 506 protection and nutrition of plants. *Biotechnol Adv* 29, 792–803 (2011).
- 507 24. Gogos, A., Knauer, K. & Bucheli, T. D. Nanomaterials in plant protection and fertilization: current
- 508 state, foreseen applications, and research priorities. *J. Agric. Food Chem.* **60**, 9781–9792 (2012).
- 509 25. Nair, R. et al. Nanoparticulate material delivery to plants. Plant Sci. 179, 154–163 (2010).
- 510 26. Cox, A., Venkatachalam, P., Sahi, S. & Sharma, N. Silver and titanium dioxide nanoparticle toxicity
- 511 in plants: A review of current research. *Plant Physiol. Biochem.* **107**, 147–163 (2016).
- 512 27. Laxma Reddy, P. V., Hernandez-Viezcas, J.-A., Peralta-Videa, J. R. & Gardea-Torresdey, J. L.
- 513 Lessons learned: Are engineered nanomaterials toxic to terrestrial plants? Sci. Total Environ. 568,
- 514 470–479 (2016).
- 515 28. Mariet, C., Belhadj, O., Leroy, S., Carrot, F. & Metrich, N. Relevance of NH4F in acid digestion
 516 before ICP-MS analysis. *Talanta* 77, 445–450 (2008).
- 517 29. Larue, C. *et al.* Fate of pristine TiO2 nanoparticles and aged paint-containing TiO2 nanoparticles
 518 in lettuce crop after foliar exposure. *J Hazard Mater* 273, 17–26 (2014).
- 519 30. Daudin, L., Khodja, H. & Gallien, J. P. Development of 'position-charge-time' tagged spectrometry
- for ion beam microanalysis. *Nucl. Instrum. Methods Phys. Res. Sect. B-Beam Interact. Mater. At.*210, 153–158 (2003).
- 522 31. Mayer, M. SIMNRA, a simulation program for the analysis of NRA, RBS and ERDA. in *Application*
- 523 of Accelerators in Research and Industry, Pts 1 and 2 (eds. Duggan, J. L. & Morgan, I. L.) **475,** 541–

524 544 (Amer Inst Physics, 1999).

- 525 32. Campbell, J. L., Hopman, T. L., Maxwell, J. A. & Nejedly, Z. The Guelph PIXE software package III:
- 526 Alternative proton database. Nucl. Instrum. Methods Phys. Res. Sect. B-Beam Interact. Mater. At.
- 527 **170,** 193–204 (2000).
- 528 33. Vantelon, D. *et al.* The LUCIA beamline at SOLEIL. *J Synchrotron Radiat* **23**, 635–640 (2016).

- 529 34. Sole, V. A., Papillon, E., Cotte, M., Walter, P. & Susini, J. A multiplatform code for the analysis of
- energy-dispersive X-ray fluorescence spectra. *Spectrochim. Acta Part B-At. Spectrosc.* 62, 63–68
 (2007).
- 532 35. Larue, C. et al. Foliar exposure of the crop Lactuca sativa to silver nanoparticles: Evidence for
- 533 internalization and changes in Ag speciation. J. Hazard. Mater. **264**, 98–106 (2014).
- 534 36. Ravel & Newville. ATHENA, ARTEMIS, HEPHAESTUS: data analysis for X-ray absorption
- 535 spectroscopy using IFEFFIT. J. Synchrotron Radiat. **12**, 537–541 (2005).
- 536 37. R Core Team. R: A language and environment for statistical computing. (2015).
- 537 38. Fox, J. & Weisberg, S. *An* {*R*} *Companion to Applied Regression*. (Thousand Oaks CA: Sage, 2011).
- 538 39. Graves, S., Piepho, H.-P. & Selzer, L. Visualizations of Paired Comparisons. (2015).
- 539 40. Giraudoux, P. pgirmess: Data Analysis in Ecology. (2017).
- 540 41. Lenth, R. V. Least-Squares Means: The R Package Ismeans. J. Stat. Softw. 69, 1–33 (2016).
- 42. Fang, J., Shan, X. Q., Wen, B., Lin, J. M. & Owens, G. Stability of titania nanoparticles in soil
- 542 suspensions and transport in saturated homogeneous soil columns. *Environ. Pollut.* 157, 1101–
- 543 1109 (2009).
- 43. Fang, J. *et al.* Transport of copper as affected by titania nanoparticles in soil columns. *Environ.*
- 545 *Pollut. Barking Essex 1987* **159,** 1248–1256 (2011).
- 44. Pachapur, V. *et al.* Behavior and characterization of titanium dioxide and silver nanoparticles in
 soils. *Science of the Total Environment* 933–943 (2016).
- 45. Wang, M., Gao, B. & Tang, D. Review of key factors controlling engineered nanoparticle transport
- in porous media. *Journal of Hazardous Materials* 233–246 (2016).
- 46. Shah, V. *et al.* Fate and impact of zero-valent copper nanoparticles on geographically-distinct
- soils. *Science of the Total Environment* 661–670 (2016).
- 47. Fang, J. *et al.* Co-transport of Pb(2+) and TiO2 nanoparticles in repacked homogeneous soil
- 553 columns under saturation condition: Effect of ionic strength and fulvic acid. *Sci. Total Environ.*
- **554 571,** 471–478 (2016).

- 48. Larue, C. *et al.* Innovative combination of spectroscopic techniques to reveal nanoparticle fate in
- a crop plant. *Spectrochim. Acta Part B At. Spectrosc.* **119**, 17–24 (2016).

49. Farges, F., Brown, G. E. & Rehr, J. J. Ti K-edge XANES studies of Ti coordination and disorder in

- 558 oxide compounds: Comparison between theory and experiment. *Phys. Rev. B* 56, 1809–1819
- 559 (1997).
- 560 50. Majumdar, S. *et al.* Soil organic matter influences cerium translocation and physiological
- 561 processes in kidney bean plants exposed to cerium oxide nanoparticles. Sci. Total Environ. 569–
- **562 570,** 201–211 (2016).
- 563 51. Gao, X. et al. CuO Nanoparticle Dissolution and Toxicity to Wheat (Triticum aestivum) in
- 564 Rhizosphere Soil. *Env. Sci Technol* (2018). doi:DOI: 10.1021/acs.est.7b05816
- 565 52. Gogos, A. *et al.* Vertical transport and plant uptake of nanoparticles in a soil mesocosm
- 566 experiment. *Journal of Nanobiotechnology* (2016).
- 567 53. Du, W. C. *et al.* TiO(2) and ZnO nanoparticles negatively affect wheat growth and soil enzyme
 568 activities in agricultural soil. *J. Environ. Monit.* 13, 822–828 (2011).
- 569 54. Larue, C. et al. Accumulation, translocation and impact of TiO2 nanoparticles in wheat (Triticum
- 570 aestivum spp.): influence of diameter and crystal phase. *Sci Total Env.* **431**, 197–208 (2012).
- 571 55. Larue, C. *et al.* Comparative Uptake and Impact of TiO(2) Nanoparticles in Wheat and Rapeseed.
- 572 *J. Toxicol. Environ. Health A* **75,** 722–34 (2012).
- 573 56. Raliya, R., Nair, R., Chavalmane, S., Wang, W.-N. & Biswas, P. Mechanistic evaluation of
- translocation and physiological impact of titanium dioxide and zinc oxide nanoparticles on the
- tomato (Solanum lycopersicum L.) plant. *Metallomics* 1584–1594 (2015).
- 576 57. Servin, A. D. *et al.* Synchrotron verification of TiO2 accumulation in cucumber fruit: a possible
- pathway of TiO2 nanoparticle transfer from soil into the food chain. *Env. Sci Technol* 47, 11592–
 11598 (2013).
- 579 58. Servin, A. D. et al. Weathering in soil increases nanoparticle CuO bioaccumulation within a
- 580 terrestrial food chain. *Nanotoxicology* 98–111 (2017).
 - 27

- 581 59. Zhang, W. *et al.* Bioavailability of cerium oxide nanoparticles to Raphanus sativus L. in two soils.
- 582 Plant physiology and biochemistry: PPB 185–193 (2017).
- 583 60. Layet, C. et al. Evidence that Soil Properties and Organic Coating Drive the Phytoavailability of
- 584 Cerium Oxide Nanoparticles. *Env. Sci Technol* **51,** 9756–9764
- 585 61. Moghaddasi, S. et al. Bioavailability of coated and uncoated ZnO nanoparticles to cucumber in
- 586 soil with or without organic matter. *Ecotoxicology and Environmental Safety* 543–551 (2017).
- 587 62. Watson, J.-L. *et al.* The phytotoxicity of ZnO nanoparticles on wheat varies with soil properties.
- 588 Biometals **28**, 101–112 (2015).
- 589 63. Zhao, L. et al. Effect of surface coating and organic matter on the uptake of CeO2 NPs by corn
- plants grown in soil: Insight into the uptake mechanism. *J. Hazard. Mater.* **225–226,** 131–138
- 591 (2012).
- 592 64. Palmer, C. M. & Guerinot, M. L. Facing the challenges of Cu, Fe and Zn homeostasis in plants.
 593 *Nat. Chem. Biol.* 5, 333–340 (2009).
- 594 65. Dimkpa, C. O. D. C. O. *et al.* Fate of CuO and ZnO Nano- and Microparticles in the Plant
- 595 Environment. *Env. Sci Technol* **47,** 4734–4742 (2013).
- 596 66. Wang, P. et al. Fate of ZnO Nanoparticles in Soils and Cowpea (Vigna unguiculata). Env. Sci
- 597 *Technol* (2013). doi:10.1021/es403466p
- 598 67. Khodakovskaya, M. et al. Carbon nanotubes are able to penetrate plant seed coat and
- 599 dramatically affect seed germination and plant growth. *Acs Nano* **3**, 3221–3227 (2009).
- 600
- 601