

# Paleoecological insights from fossil freshwater mollusks of the Kanapoi Formation (Omo-Turkana Basin, Kenya)

Bert Van Bocxlaer

## ▶ To cite this version:

Bert Van Bocxlaer. Paleoecological insights from fossil freshwater mollusks of the Kanapoi Formation (Omo-Turkana Basin, Kenya). Journal of Human Evolution, 2020, 140, pp.102341. 10.1016/j.jhevol.2017.05.008. hal-02323652

HAL Id: hal-02323652

https://hal.science/hal-02323652

Submitted on 18 Aug 2020

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

- 1 Paleoecological insights from fossil freshwater mollusks of the Kanapoi
- 2 Formation (Omo-Turkana Basin, Kenya)

4 Bert Van Bocxlaer<sup>a,b,\*</sup>

3

5

- 6 Limnology Unit, Department of Biology, Ghent University, B-9000 Ghent, Belgium.
- <sup>b</sup>CNRS, Univ. Lille, UMR 8198 Evolution, Ecology, Paleontology, F-59000 Lille, France
- 8 \*Corresponding author. Address: UMR 8198 Evolution, Ecology, Paleontology; Lille University –
- 9 Science and Technology, Cité Scientifique Bât. SN5, F-59655 Villeneuve d'Ascq cedex, France; E-
- mail: bert.van-bocxlaer@univ-lille1.fr; tel: +33 3 20 43 40 15.
- 12 Running Title: Fossil freshwater mollusks from Kanapoi



#### Abstract

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

The Early Pliocene Kanapoi Formation of the Omo-Turkana Basin consists of two fluvial/deltaic sedimentary sequences with an intermediate lacustrine sequence that was deposited in Paleolake Lonyumun, the earliest large lake in the basin. Overall, the geology and vertebrate paleontology of the Kanapoi Formation are well studied, but its freshwater mollusks, despite being a major component of the benthic ecosystem, have not been subjected to in-depth study. Here I present the first treatment of these mollusks, which have been retrieved mainly from the lacustrine but also from the upper fluvial sediments, with a focus on paleoecological implications. Overall, the freshwater mollusk fauna is reasonably diverse and contains the gastropods Bellamya (Viviparidae), Melanoides (Thiaridae), Cleopatra (Paludomidae) and Gabbiella (Bithyniidae), as well as the unionoid bivalves Coelatura, Pseudobovaria (Unionidae), Aspatharia, Iridina (Iridinidae) and Etheria (Etheriidae). Material is typically recrystallized, lithified and its taphonomy suggests deposition in a system with intermediate energy, such as a beach, with post-depositional deformation and abrasion. The mollusk assemblage is indicative of perennial, fresh and well-oxygenated waters in the Kanapoi region. It suggests that Paleolake Lonyumun had largely open shores with limited vegetation and that swampy or ephemeral backwaters were rare. Overall, these findings support earlier paleoecological interpretations based on the fish assemblage of Paleolake Lonyumun at Kanapoi. Moreover, mollusk assemblages from this lake are very similar across the Omo-Turkana Basin (Nachukui, Usno, Mursi and Koobi Fora Formations) suggesting that the lacustrine paleoecological conditions found in the Kanapoi Formation existed throughout the basin.

33

34 Keywords: freshwater gastropods; freshwater bivalves; taphonomy; paleoecology; East African Rift

35 System



#### Introduction

36

The Kanapoi Formation provides a unique insight into Early Pliocene environments in the southern 37 Omo-Turkana Basin, including into the terrestrial and aquatic faunas of the region at that time (e.g., 38 Harris et al., 2003; Stewart, 2003; Werdelin, 2003). Here I provide an overview of freshwater 39 mollusks from the Kanapoi Formation, with a focus on their paleoenvironmental implications. 40 41 Treatment of these mollusks is warranted here because unlike the geological setting and vertebrate 42 remains of the Kanapoi Formation (e.g., Harris and Leakey, 2003), they have not been subjected to detailed studies before. Moreover, they arguably are an important component of the aquatic 43 ecosystem, as evidenced by various shell assemblages, Etheria bioherms of up to 1 m in thickness 44 (Feibel, 2003a), and the associated rich fauna of medium to large molluscivorous fishes (Stewart, 45 46 2003). Sediments of the Kanapoi Formation have a composite thickness of ~60 m, and they have been 47 deposited during two fluvial/deltaic phases of the Kerio River system, with an intervening lacustrine 48 phase (Fig. 1) (Feibel, 2003a). Three tuffs occur in the formation, i.e., the lower and upper 49 pumiceous airfall tuffs and the vitric Kanapoi tuff. These tuffs have been dated to  $4.195 \pm 0.033$  Ma, 50  $4.147 \pm 0.019$  Ma and  $4.108 \pm 0.029$  Ma, respectively (McDougall and Brown, 2008). The Kanapoi 51 52 Formation is capped by the Kalokwanya Basalt (Feibel, 2003a), which has been dated to ~3.4 Ma (Leakey et al., 1995), although its reversed magnetic polarity (Powers, 1980) is inconsistent with this 53 age (McDougall and Brown, 2008). 54 Most of the fossil remains of the Kanapoi Formation have been obtained from deposits between the 55 56 lower pumiceous tuff and the Kanapoi tuff, including remains of Australopithecus anamensis Leakey, 1995 (Leakey et al., 1995; Leakey et al., 1998; Ward et al., 2013). Freshwater mollusks have only been 57 obtained from sediments above the upper pumiceous tuff: some mollusk material derives from the 58



upper fluvial phase, but most is obtained from strata that were deposited during the lacustrine phase, i.e., in stratigraphic proximity to the Kanapoi tuff. These lacustrine sediments also contain ostracodes and plant fragments, and were deposited in Paleolake Lonyumun, the earliest extensive lacustrine interval in the Omo-Turkana Basin (Fig. 1) (Feibel, 1997, 2003a, 2011). Evidence of this paleolake is recorded throughout the whole Omo-Turkana Basin: from the Usno and Mursi Formations along the Omo River in the North (de Heinzelin, 1983), from the Lonyumun Member of the Nachukui Formation in West Turkana (Harris et al., 1988), from the Murungori Member of the same formation at Lothagam (Feibel, 2003b), from the Kanapoi Formation in Southwest Turkana (Feibel, 2003a), and from the Lonyumun Member of the Koobi Fora Formation in East Turkana (Brown and Feibel, 1991; Feibel et al., 1991). The Omo, Kerio and Turkwell Rivers were the main sources of inflow for Paleolake Lonyumun, and the lake is presumed to have been hydrologically open, with the Turkana River in the Southeast as outflow (Feibel, 2011). Paleolake Lonyumun appears to have been extensive but shallow and probably persisted for about 100 ka (Feibel, 2011). Below, I first introduce the mollusk fauna of the Kanapoi Formation, and subsequently I discuss its paleoenvironmental implications.

75 Material and methods.

59

60

61

62

63

64

65

66

67

68

69

70

71

72

73

- 76 Remains of freshwater mollusks from the Kanapoi Formation have been collected during the
- 77 Harvard University expeditions led by B. Patterson from 1964 to 1968, during expeditions by D. W.
- Powers in 1972, by C. S. Feibel in 1995 and 1996, and during fieldwork in 2014 led by C. W. Ward
- and F. K. Manthi. I here report on all of this material, ~925 specimens, however, as most material is
- 80 preserved in lithified blocks of sediment future disaggregation may yield more specimens. Material is
- 81 reported with original processing numbers for the first two expeditions (e.g., DVD KP6501L, with



82 DVD indicating Dirk Van Damme, who assigned the processing number, KP65 indicating the Kanapoi 1965 collecting event, 01 the sample number in that collecting event, and L the stratigraphic 83 position. For this material two stratigraphic positions, i.e., lower [L] and upper [U] are possible, 84 representing lacustrine and fluvial/deltaic environments, respectively. Other material is indicated 85 with fieldcodes, e.g., CSF K96-5643 with CSF indicating Craig S. Feibel [collector], K96 representing 86 87 Kenya 1996 and 5643 the sample number; KAP14-001 represents sample 001 from fieldwork at 88 Kanapoi in 2014. Material will be deposited in museum collections after formal taxonomic treatment. Furthermore, some scattered Late Pleistocene-Holocene mollusk-bearing sediments occur at 89 Kanapoi, but as these do not belong to the Kanapoi Formation this material is not treated here— 90 they contain Melanoides cf. tuberculata (Müller, 1774) and Iridina turkanica Van Bocxlaer and Van 91 92 Damme, 2009. Overall, the preservation of mollusks at Kanapoi is moderate to poor, certainly in comparison to 93 those of the Nachukui, Koobi Fora and Shungura Formations, which relates to corrosion of 94 specimens, recrystallization and abrasion. Much of the abrasion has occurred after deposition of 95 material as part of the processes that have resulted in lithified shell beds: bivalves regularly are 96 fragmented but the fragments are usually still in place, whereas flattened gastropod shells also 97 98 evidence the great pressures associated with sediment compaction and potentially tectonic processes to which assemblages have been exposed. Additionally, shells may be dissolved, leaving mainly 99 imprints of the external shell and internal molds. Similar processes are observed in other shell beds in 100 the Turkana Basin of the same age, however, to varying degrees. For example, several shell beds in 101 102 the Nachukui Formation have not been exposed to the same pressures as those experienced at Kanapoi, resulting in better preservation and less aggregation, so that in situ collecting is easier. As a 103



result of its preservation, mollusk material from the Kanapoi Formation is typically collected as shellenclosing sediment blocks, or as weathered, surface-picked specimens. Here, I describe the freshwater mollusk fauna from the Kanapoi Formation up to genus level and provide comments on specific attributes in the context of the current literature. Some of the included species are likely new to science, but I refrain from formal species-level description here for several reasons: First, my main focus here is on paleoenvironmental implications, which can be inferred at genus level, rather than on mollusk taxonomy. Second, descriptions ideally are complemented with objective morphometric comparisons and detailed diagnoses that allow distinguishing between closely related species that have been obtained from other regions in the Omo-Turkana Basin and other periods, which would detract from a strong focus on the Kanapoi Formation. Finally, for some taxa more typical and better preserved material is found elsewhere, and I consider it warranted that the type material is as complete as possible regarding diagnostic features. Nevertheless, as the mollusk fauna of the Kanapoi Formation is poorly known, I provide concise treatments and schematic reconstructions to allow identification of material in the field and to make paleoenvironmental interpretations more traceable and reproducible. However, I first investigate the effect of sample size upon the sampled genus-level diversity as a measure of how reliable we can expect paleoenvironmental interpretations to be. I performed individual-based rarefaction at the genus level with all abundance data pooled into a single reference sample. Assumptions and requirements for rarefaction are indicated in the discussion. Rarefaction was performed in R version 3.2.1 (R Core Team, 2015) using the package iNEXT (Chao et al., 2014; Hsieh et al., 2016) and in EstimateS version 9.1.0 (Colwell et al., 2012; Colwell, 2013). Calculations were performed with variance estimators that are conditional or unconditional on the sampling data to calculate 95 % confidence intervals (CIs). Use of the conditional variance estimator allows examining expectancies



104

105

106

107

108

109

110

111

112

113

114

115

116

117

118

119

120

121

122

123

124

125

for samples that are considerably smaller than the observed 'reference' sample, but it results in increasingly narrow confidence intervals when the size of the estimated sample approaches that of the reference sample. As the reference sample represents a random sample from an unknown ecological community/assemblage, we can use the unconditional variance estimator to examine how reliable the sample represents the total assemblage, and to calculate extrapolated species richness estimates (Colwell et al. 2012). Finally, I calculated Chao1 as a non-parametric estimator of asymptotic richness.

134

135

127

128

129

130

131

132

133

#### Results

- 136 Systematic Paleontology
- 137 Viviparidae Gray, 1847
- 138 Bellamyinae Rohrbach, 1937
- 139 Bellamya Jousseaume, 1886

**Remarks.** A single species of *Bellamya* appears to occur in the Kanapoi Formation (Fig. 2A), and it is 140 morphologically very similar to material that has been referred to by Fuchs (1939: 269) as Bellamya 141 unicolor var. rudolfianus from deposits at Gaza waterhole (Alia Bay area, East Turkana), the Losidok 142 (Lothidok) Hills (West Turkana) and from northwest of Labur (Northwest Turkana). However, the 143 currently accepted concept of Bellamya unicolor (Olivier, 1804), described from extant material from 144 145 Alexandria, Egypt (Brown, 1994) appears to contain multiple phylogenetic lineages, like is the case for B. capillata (Frauenfeld, 1865) (Schultheiß et al., 2014; Sengupta et al., 2009), and as such it is in 146 147 need of revision. The specimens of the Kanapoi Formation attain a size up to 30 mm and display a basal ridge surrounding the umbilicus. They likely belong to a species that is new to science, and that 148 149 was abundant in Paleolake Lonyumun.



151	Material examined: CSF K95-5487 (23 specimens); CSF K95-5557 (12 specimens); CSF K96-5643
152	(4 specimens, 3 fragments); DVD KP6502L (10 fragmentary molds in a block); DVD KP6504L,
153	KP6506L (3 internal molds); DVD KP6601L (15 internal molds, 1 recrystallized juvenile shell and 9
154	fragmentary shells); DVD KP6602L (19 internal molds, 12 fragmentary internal molds); DVD
155	KP6601L, 6602L, 6604L, 7202L (4 specimens, somewhat deformed, 1 external mold, 1 fragment);
156	DVD KP7201L (4 deformed fragments); DVD KP7203L (4 fragments, 9 internal molds, 1 external
157	mold); DVD KP7205L (28 internal molds, 1 external mold, 3 fragments); DVD KP7206L (2
158	fragments); KAP14-001 (1 specimen); KAP14-002 (1 specimen); KAP14-003 (1 specimen); KAP14-
159	004 (1 deformed specimen); KAP14-005 (3 specimen, 2 internal molds, 1 fragment). Other material
160	of the same species is known from the Mursi Formation, the Nachukui Formation at Lothagam, and
161	the Koobi Fora Formation.
162	

163

166

167

168

169

170

171

- Paludomidae Stoliczka, 1868
- 164 Cleopatrinae Pilsbry & Bequaert, 1927
- Cleopatra Troschel, 1856 165
  - **Remarks.** A single species of *Cleopatra* appears to occur in the Kanapoi Formation (Fig. 2B), although dimorphism seems to exist in some samples, with a rare, small and often poorly preserved morph besides the typical form. I tentatively consider this small morph to be a variant of the same species. The typical form is very similar to the Cleopatra species described by Roger (1944) from the Omo Valley (between Kalam and Bourillé) as C. arambourgi, and also to material from Kubi Algi (the Lonyumun Member of the Koobi Fora Formation) that Williamson (1979, 1981), perhaps following Fuchs (1939), referred to the extant species Cleopatra ferruginea (Lea and Lea, 1850). However, as the



type locality of *C. ferruginea* is Zanzibar (Brown, 1994), substantial differences exist in geographical and temporal occurrence with material from the Kanapoi Formation. Moreover, growth patterns differ somewhat between both species, with *C. arambourgi* growing more in width early on, and later in length, resulting in a wider apical angle in juveniles than adults, whereas the apical angle in *C. ferruginea* remains more constant through life. As such, I consider it unlikely that *C. arambourgi* is conspecific with *C. ferruginea*. *Cleopatra* sp. A of Van Damme and Gautier (1972) from the Omo Valley is also conspecific with *C. arambourgi*.

180

181

182

183

184

185

186

187

173

174

175

176

177

178

179

Material examined: CSF K96-5643 (~70 shells); DVD KP6502L (14 specimens, 17 fragments);

DVD KP6505U (16 specimens in blocks); DVD KP6601L (15 specimens, 1 fragment, 1 external

mold); DVD KP6601L, 6602L, 6604L, 7202L (2 specimens); DVD KP6605L (3 specimens); DVD

KP7201L (3 internal molds); DVD KP7203L (16 internal molds, 1 external mold); DVD KP7205L

(~95 internal molds in blocks); DVD KP7206L (7 fragments); KAP14-004 (1 fragment); KAP14-005

(33 specimens, 1 fragment). Material is also known from East Turkana, the Omo Valley and the

Lothidok area.

188

189

191

192

193

194

Thiaridae Gill, 1871

190 Melanoides Olivier, 1804

Williamson (1979, 1981) considered all Melanoides specimens from the lower part of the Koobi Fora

Formation and Holocene Beds in the Turkana Basin to be Melanoides tuberculata, and he specifically

referred material from the Kanapoi Formation to belong to Melanoides tuberculata as well. Earlier, Van

Damme and Gautier (1972) considered Melanoides from the Mursi Formation of the Omo-Turkana

Basin to be a new subspecies of Melanoides tuberculata. A difficulty with the identification of Melanoides



is that several species appear to be morphologically very variable, perhaps because of parthenogenetic reproduction, and, furthermore, that polyploid lineages exist (Jacob, 1957; Samadi et al., 1999, 2000). Moreover, recent molecular work indicates that a number of different species with similar shell morphological features are likely aggregated under *M. tuberculata*, including a native African as well as an invasive oriental lineage (Genner et al., 2004; Van Bocxlaer et al., 2015). The material from the Kanapoi Formation is morphologically variable, but most specimens are ornamented with strong, often nodular ribs, a subsutural cord and other spiral cords at the base, as well as fainter spiral lines in between (Fig. 2C). The ribs are more widely spaced than those of *M. recticosta* (von Martens, 1882), and the set of ornamentation displayed by *Melanoides* from the Kanapoi Formation is not observed in extant populations of *M. tuberculata* (Van Bocxlaer et al., 2015). As such, I consider it more likely that the *Melanoides* species of the Kanapoi Formation represents a separate species, but more in-depth morphometric work is required to confirm this issue.

**Material examined:** CSF K96-5643 (~60 relatively well preserved specimens); DVD KP6501L (48 specimens in blocks); DVD KP6503L (1 specimen); DVD KP6601L (1 specimen); DVD KP6605L (~105 specimens in blocks); KAP14-005 (53 isolated specimens and more specimens in blocks of sediment). In-depth work on other faunas is required to establish the geographical extent of this taxon.

- Bithyniidae Gray, 1857
- 216 Gabbiella Mandahl-Barth, 1968
- Remarks. Some evidence of the presence of Bithyniidae has been found in the Kanapoi Formation,consisting of one shell, an inner core, and several opercula. As the preservation is poor, identification



at the genus level is somewhat tentative, but supported by the strongly calcified opercula and a specimen resembling *Gabbiella* in contemporaneous deposits of the Mursi Formation (DVD MU6801). It is somewhat surprising that this taxon is mainly known from opercula in the Kanapoi Formation, because *Gabbiella* typically has a relatively thick shell, so that preservation bias is not a likely cause for its absence. It is possible that the taxon was rare in general, or largely restricted to specific microhabitats, e.g., associated with aquatic plants, that may have been underrepresented in the Kanapoi Formation.

226

227

228

229

219

220

221

222

223

224

225

Material examined: CSF K96-5640 (~5 opercula); DVD KP6502L (1 specimen, identification tentative); DVD KP6601L (1 inner core and 2 opercula). As mentioned, the taxon also occurs in the Mursi Formation.

- 231 Unionidae Rafinesque, 1820
- 232 Coelaturinae Modell, 1942
- 233 Coelatura Conrad, 1853
- Remarks. I reported earlier that two *Coelatura* species appear to be present in the Kanapoi
- Formation, i.e., C. aegyptiaca (Cailliaud, 1827) and C. bakeri (Adams, 1866), although these
- identifications were reported as tentative (Van Bocxlaer, 2011). Both taxa are very variable in shell
- shape in the extant fauna (Mandahl-Barth, 1954, 1988), and in fossil deposits from the Turkana
- Basin, as evidenced from shell shape data based on elliptic Fourier analysis (Van Bocxlaer, 2011).
- However, C. aegyptiaca specimens have a posterior wing and more prominent angulations at the
- anterior and posterior sides of the dorsal margin than C. bakeri, which typically is more elongate
- 241 (Mandahl-Barth, 1954, 1988). The morphotype resembling *C. aegyptiaca* is represented in Fig. 2E.



Typical *C. bakeri* specimens display zigzag sculpture on the umbo, whereas *C. aegyptiaca* bears tubercles (Van Bocxlaer, 2011). Two issues affect identification of *Coelatura* specimens from the Kanapoi Formation: most specimens are preserved as fragments and, second, umbonal sculpture is difficult to discern on most specimens of the Kanapoi Formation, perhaps because of recrystallization of the material. Nevertheless, specimens with features of each taxon, including umbonal sculpture, have been obtained from the Kanapoi Formation, suggesting that both were present. Additional, better-preserved material from the Kanapoi Formation and molecular work on extant representatives of both species may elucidate the current ambiguity.

Material examined: (identifications are tentative) *C. aegyptiaca*: CSF K96-5643 (5 doublets, some more valves and fragments); DVD-KP6606L (mold of 1 doublet, 10 internal molds of valves, 6 fragmentary valves); KAP14-005 (1 valve); *C. bakeri*: DVD KP6501L (2 doublets, 1 valve); DVD KP6503L (5 fragmentary valves); DVD-KP6601L/6602L/KP6604L/7202L (1 doublet, 3 imprints); DVD-KP6605L (5 doublets, 19 fragmentary valves); KAP14-005 (1 doublet); *Coelatura* sp. indet.: DVD KP6501L (2 fragmentary valves, 2 external molds); DVD-KP6503L (1 valve, 2 external molds); DVD KP6602L (2 fragmentary valves); DVD-KP6605L (37 fragmentary valves); DVD KP6606L (2 valves); KAP14-005 (18 fragmentary valves). *Coelatura aegyptiaca* and *C. bakeri* occur in various regions of the Turkana Basin, including in the Nachukui Formation at Lothagam and West Turkana, the Mursi Formation, the Koobi Fora Formation and the Shungura Formation, and in various periods, notably in Paleolakes Lonyumun and Lorenyang (Van Bocxlaer, 2011).

Unionidae with uncertain subfamily position

264 Pseudobovaria Adam, 1957



**Remarks.** Several features, including the hinge, indicate that the extinct genus *Pseudobovaria* belongs to Unionidae, however, its attribution at the subfamily level is uncertain. Morphologically, representatives of the genus compare well to the North American genera Obovaria Rafinesque, 1819, and Pleurobema Rafinesque, 1819, which belong to the subfamily Ambleminae Rafinesque, 1820. Extant representatives of this subfamily are deeply split molecularly, and geographically distant from African taxa (Whelan et al., 2011), suggesting that remarkable convergence is the most likely cause of similarity. Also Gonideinae Ortmann, 1916, to which Van Damme and Pickford (2010) attributed Pseudobovaria, is deeply split from Coelaturinae Modell, 1942 (Whelan et al., 2011), and given that good arguments for any subfamily attribution are lacking, I consider the genus' position within Unionidae currently uncertain. In any case, the few Pseudobovaria valves that have been obtained from the Kanapoi Formation differ morphologically from previously described Pseudobovaria species in that they are more elongate, with valves and hinges that are less strongly calcified (Fig. 2D). As such they likely represent a new species. Better preserved material has been obtained from the Nachukui Formation near Kalokol. Van Damme and Gautier (1972) also obtained material of this taxon from the Mursi and Usno Formations, but at the time they considered it to belong to Nyassunio Haas, 1936.

281

282

283

265

266

267

268

269

270

271

272

273

274

275

276

277

278

279

280

**Material examined:** DVD KP6502L (1 specimen); DVD KP6601L (17 specimens in blocks, some being fragmentary). Finds at other localities are indicated in the remarks.

284

- Iridinidae Swainson, 1840
- 286 Aspathariinae Modell, 1942
- 287 Aspatharia Bourguignat, 1885



Remarks. Some material that can be attributed to *Aspatharia* has been obtained from the Kanapoi Formation, but it is not well preserved. However, at least part of the fragments that have been obtained display conspicuous radial ridges on the posterior-dorsal surface of the shell, linking the material with better preserved specimens from the Mursi Formation. This material has been assigned to *A. cf. rugifera* (Dunker, 1858) (Van Bocxlaer and Van Damme, 2009). More and better preserved material, as well as insights into the phylogenetic relationships between extant *Aspatharia* species, may elucidate the affinities of the material better in the future. In any case, *A. rugifera* is currently confined to West African rivers (Mandahl-Barth, 1988).

**Material examined:** DVD KP 6603L (1 internal mold, fragments of 6 doublets, 1 fragmentary valve with the typical ornamentation on the posterior side): KP 6504L (1 fragment of a doublet); KP6501L (fragments of ? *Aspatharia*). More material of this taxon has been obtained from the Mursi Formation.

Iridininae Swainson, 1840

302 Iridina Lamarck, 1819

**Remarks.** Much material from the Kanapoi Formation belonging to Iridininae is fragmentary, but at least part of the material can be clearly assigned to *Iridina subelongata* Gautier, 1965 (Van Bocxlaer and Van Damme, 2009). Some other fragments do not display a denticulate hinge, which may have resulted from corrosion or abrasion. As Van Bocxlaer and Van Damme (2009) indicated, it is possible that a second iridinine species occurs in the Kanapoi Formation, but no sufficiently well-preserved material exists to confirm this hypothesis.



Material examined: CSF K96-5643 (3 doublets, fragmentary valves of *Iridina* sp. indet); CSF K95-5487 (fragments of *Iridina* sp. indet); DVD-KP 6503L (1 doublet); 6605L (fragments of 4 doublets and 1 single valve); 6606L (5 fragmentary external molds); 7204L (fragmentary molds); all of these samples, except for 6605L contain additional unidentifiable *Iridina* fragments and/or internal molds; KAP14-005 (fragments of *Iridina*, possibly of *I. subelongata*). More material has been obtained from the Mursi, Usno, Nachukui, Koobi Fora and Shungura Formations.

Etheriidae Deshayes, 1832

318 Etheria Lamarck, 1807

Remarks. Etheria has been considered to be monospecific in the latest systematic treatments of extant African freshwater mollusks (Mandahl-Barth, 1988; Daget, 1998; Graf and Cummings, 2007), and in this context Van Damme and I attributed all Etheria specimens obtained from the Turkana Basin to Etheria elliptica Lamarck, 1807 (Van Bocxlaer and Van Damme, 2009). Recent work (Elderkin et al., 2016), however, indicates deep phylogenetic splits between populations of Etheria, suggesting that some of the other described species, e.g., by Lamarck (1807), may actually be valid. As Etheria has very limited diagnostic shell-morphological features at the species level (Mandahl-Barth, 1988) it will be a challenge to clear up the existing taxonomic confusion. As such, the Turkana Basin may have, over time, been home to several Etheria species, but for now, I am unable to revise our previous assignment as knowledge on the extant Etheria fauna remains too fragmentary. I have not had an opportunity to investigate Etheria specimens from Kanapoi, but Feibel (2003a) identified a reef of Etheria in his section CSF 96-12, directly on top of the Kanapoi Tuff. The lack of samples may relate to the practical difficulties to sample a bioherm. The reconstruction provided here (Fig. 2H) is based on a contemporary specimen from the Lothagam area of the Nachukui Formation.



Rarefaction

As mentioned, all the examined material was pooled into a single sample for individual-based rarefaction. For *Etheria* five specimens were included to cover the existing reports on its presence in the Kanapoi Formation, resulting in a total sample size of n = 931. The accumulation curve reaches an asymptote well before the total number of individuals sampled, and the associated 95 % CIs are narrow, even upon using the unconditional variance estimator and at sample sizes approximating the total size of the reference sample (Fig. 3). Non-parametric asymptotic richness (Chao1; mean and 95 % CI) converges to the generic richness in our reference sample (n = 9).

## Discussion

Rarefaction has a number of requirements that may be difficult to meet in a paleontological context (e.g., standardized sampling of lithified shell assemblages), and it makes a number of assumptions (e.g., random distribution of individuals in environments), which may be affected by taphonomic processes (see e.g., Foote, 1992; Gotelli and Colwell, 2001). As such, results are to be interpreted with care. Nevertheless, chances that the number of genera sampled underestimates the true but unknown diversity significantly are small if our collective, historic sampling of freshwater mollusks is representative of all existing shell beds in the Kanapoi Formation. Note that the seeming sufficiency of the collected material for reliable paleoecological interpretations does not contradict the need of more and better preserved material for species-level taxonomy.

I have made some comments on the preservation of mollusk specimens already, but discuss taphonomic aspects related to depositional environments in more detail here. Shells from the Kanapoi Formation have typically been rolled somewhat before they were deposited, resulting



regularly in the abrasion of early whorls and the aperture in gastropods, and sometimes in the separation of both valves in bivalves. However, the abrasion taking place before deposition remains overall limited and doublets are still commonly obtained in bivalves. Overall this indicates a depositional environment with intermediate water energy, much like is the case for beach deposits in other African Great Lakes. I have not observed indications that would suggest long distance transportation, except perhaps the difference in abundance between bithyniid opercula and shells. The taxa recovered from the Kanapoi Formation are medium to large (longest axis ~1.0 cm or larger) and rather strongly calcified, indicating that smaller and more fragile taxa, such as some heterobranchs, may have been present, especially in fluvial deposits, but not preserved due to preservation bias. Perhaps the rarity of bithyniid shells is caused by this factor, but Gabbiella is typically rather thick-walled. Furthermore, the presence of even smaller, but chemically more resistant calcareous bithyniid opercula indicates that size alone is no major driver of any preservation bias. Aragonite dissolution does not provide a straightforward explanation for the absence of certain taxa neither. Overall the effects of dissolution vary between sampling sites of Paleolake Lonyumun, but even in the Mursi Formation, which is heavily affected by dissolution, a Gabbiella shell (our results) and two genera of Sphaeriidae Deshayes, 1855 with very fragile shells were preserved (Van Damme and Gautier, 1972). Furthermore, most assemblages of the Kanapoi Formation show limited evidence for dissolution, e.g., the fragile, aragonitic early teleoconchs of *Melanoides* are occasionally preserved. Variation in preservation potential may vary between habitats (e.g., sites with more vegetation may create more acidic depositional environments due to decaying plant material), but a specimen of the heterobranch Bulinus Müller, 1781 is known from the Nachukui Formation at Lothagam, indicating, together with the observations in the Mursi Formation, that there are no basin-wide factors that have eliminated the chances for the preservation of fragile specimens in



356

357

358

359

360

361

362

363

364

365

366

367

368

369

370

371

372

373

374

375

376

377

deposits of Paleolake Lonyumun. Most thin-shelled heterobranchs typically adopt a cryptic lifestyle within vegetation, where they are better protected against predators (Brown, 1994). None of the freshwater mollusk taxa present in the Kanapoi Formation prefers this type of habitat, which may indicate the overall limited occurrence of vegetation-rich habitats in Paleolake Lonyumun at Kanapoi. This finding is corroborated by the fish assemblages of the Kanapoi Formation, which are rare in herbivores (Stewart, 2003). Note that the remains of such herbivorous fish would not be affected by any of the abovementioned potential sources of preservation bias for mollusks. In general, the freshwater mollusk fauna of the Kanapoi Formation is with ~10 species modestly to reasonably diverse. In genus-level composition the fauna is very similar to the modern mollusk fauna of the southern waters of Lake Chad, apart from the fact that a diversity of (small, fragile) heterobranchs have been recorded from the surroundings of this lake (Lévêque and Saint-Jean, 1979; Brown, 1994). Lake Chad lacks the extinct *Pseudobovaria*, but contains the similar-sized corbiculid bivalve Corbicula von Mühlfeld, 1811. Corbicula appears to have been absent from Paleolake Lonyumun, but it is present in later paleolakes in the Turkana Basin (e.g., Van Bocxlaer et al., 2008; Williamson, 1981). Not unlike in Lake Chad, mollusks reached great biomass in Paleolake Lonyumun, as evidenced from the deposition in a mollusk packstone of up to 1 m in thickness. The gastropods Cleopatra, Melanoides and Bellamya are the most abundant taxa, representing ~32, 29 and 19 % of the recovered specimens, respectively. The substantial molluscan biomass in Paleolake Lonyumun is also corroborated by the presence of a diverse community of medium to large molluscivore fishes (Stewart, 2003). Substantial diversity in unionoids, which have a fish-parasitizing



larval stage, is furthermore congruent with substantial fish diversity.

379

380

381

382

383

384

385

386

387

388

389

390

391

392

393

394

395

396

397

398

399

Accepted Manuscript, Journal of Human Evolution (online Sept 2017, in print Jan 2020) Published version: http://www.sciencedirect.com/science/article/pii/S0047248417302245 DOI: http://dx.doi.org/10.1016/j.jhevol.2017.05.008

The mollusk assemblage indicates the presence of year-round waters, with open sandy shorelines, and some aquatic macrophytes. Reed fringes and swampy areas, as regularly associated with deltaic conditions, were likely restricted at Kanapoi during the lacustrine phase—several of the occurring mollusk taxa do not tolerate desiccation or oxygen stress, avoid marginal reed mats, and most of the occurring taxa are typically restricted to perennial waterbodies (Bellamya, Gabiella, and unionoids) (Brown, 1994; Van Bocxlaer et al., 2011). Moreover, some of these taxa, e.g., Bellamya, have a limited tolerance for brackish/saline conditions (Lévêque, 1972) indicating that Paleolake Lonyumun was substantially less saline than the current Lake Turkana. The freshwater mollusks of the Kanapoi Formation therefore indicate well-oxygenated, freshwater conditions, which is also suggested by the associated fish assemblage (Stewart, 2003). Furthermore, amphibious snails that are adapted to swamp-like and seasonally desiccating habitats, e.g., the ampullariids Pila Röding, 1798 and (to somewhat lesser extent) Lanistes Montfort, 1810, are absent, not only from the Kanapoi Formation, but also from most other contemporaneous deposits of Paleolake Lonyumun. The general absence of ampullariids indicates that the lake, despite presumably being shallow (Feibel, 1997), had substantial stretches of open water, probably not unlike Lake Victoria today, and that shallow, densely vegetated and potentially seasonally desiccating lagoons were rare. Some Lanistes and Bulinus specimens have been obtained from the Nachukui Formation at Lothagam, but their exact age is somewhat uncertain. Nevertheless, the occurrence of amphibious snails at Lothagam is again congruent with the fish community, which unlike that at Kanapoi, includes species that are regularly associated with vegetated, shallow backwaters or bays, i.e., Protopterus Owen, 1839, Polypterus Geoffroy Saint-Hilaire, 1802 and Heterotis Ruppell, 1829 (Stewart, 2003). As such, both the fish and mollusk communities suggest that lacustrine environments at Kanapoi were substantially different from those at Lothagam. The contrast in habitats is perhaps most eloquently indicated by comparing



401

402

403

404

405

406

407

408

409

410

411

412

413

414

415

416

417

418

419

420

421

422

the mollusk fauna obtained from lacustrine and fluvial/deltaic deposits of the Kanapoi Formation (Table 1), although in all fairness, fluvial mollusk-bearing deposits are much rarer and likely still underrepresented in current samples of the Kanapoi Formation. In any case, it is probably not a coincidence that only *Cleopatra*, which is more tolerant to a number of stressors than *Bellamya* and unionoid bivalves (Brown, 1994; Van Bocxlaer et al., 2011), was obtained from fluvial/deltaic deposits. Although the sediments of the Kanapoi Formation provide the oldest record of fluvial sediments deposited by the Kerio River, and although most of the terrestrial vertebrates may have been deposited in the paleoenvironment of a deltaic tongue that extended into the subsequently expanding Palaeolake Lonyumun (Feibel, 2003a), most of the freshwater mollusks have been deposited in a fully lacustrine context rather than in a fluvial/deltaic environment.

Exploratory comparisons of the freshwater mollusk fauna of the Kanapoi Formation to mollusk faunas that have been deposited in other parts of the Turkana Basin during the existence of Paleolake Lonyumun indicate overall great faunal resemblance across regions (see also systematic paleontology). This resemblance indicates that the lake was indeed large, but that faunal connectivity between its various regions was nevertheless good. If this finding remains standing after more indepth morphometric comparisons, it suggests that the lake was too short-lived for intra-lacustrine speciation and the emergence of regional endemism. Our understanding of the Early Pliocene freshwater mollusk faunas of Africa is too limited to make strong predictions as to the biogeographic origin of the molluscan assemblage of the Kanapoi Formation. However, some similarities exist with the fauna of the Albertine Basin, where *Iridina subelongata* and *Pseudobovaria* occurred since the Miocene (Van Damme and Pickford, 2010). Moreover, the hydrographic catchment of the Albertine Basin extended substantially further eastward in the Early Pliocene than it does currently (Salzburger



et al., 2014). The occurrence of A. cf. rugifera in the Turkana Basin may also highlight a stronger connectivity between East and West African faunas in the Early Pliocene. Care needs to be taken with this interpretation as the Early Pliocene freshwater mollusk fauna of North Africa, and the hydrographic connections of the Omo River, are poorly known. Nevertheless, progressive rift formation and the associated emergence of long-lasting waters in the Early Pliocene Turkana Basin have attracted diverse terrestrial and aquatic faunas. The Kanapoi Formation provides, among the first places, testimony to the start of a strongly interlinked succession of terrestrial and aquatic faunal diversity in the Omo-Turkana Basin as climate and environments changed through time.

## Acknowledgements

Kanapoi fieldwork in 2014 was supported by NSF BCS-1231749 (to C. V. Ward and F. K. Manthi), by NSF BCS-1231675 (to J. M. Plavcan and P. S. Ungar), the Wenner Gren Foundation and the University of Missouri Research Board. I thank C. V. Ward and F. K. Manthi for their invitation to participate to this research, and their support along the way, F. H. Brown, C. S. Feibel and F. K. Manthi for samples from Kanapoi and other regions in the Turkana Basin, and F. H. Brown for insights in the field. Research was funded by a postdoctoral Fellowship of the Flanders Research Foundation. I am not aware of any conflict of interest as to this paper.

#### References

Amante, C., Eakins, B.W., 2009. ETOPO1 1 Arc-Minute Global Relief Model: Procedures, Data Sources and Analysis. NOAA Technical Memorandum NESDIS NGDC-24. 19 pp.

Brown, D.S., 1994. Freshwater snails of Africa and their medical importance. Taylor & Francis, London.

- Brown, F.H., Feibel, C.S., 1991. Stratigraphy, depositional environments, and palaeogeography of the
- Koobi Fora Formation, in: Harris, J.M. (Ed.), Koobi Fora Research Project. Volume 3—The
- fossil ungulates: geology, fossil artiodactyls and palaeoenvironments. Clarendon Press, Oxford,
- 473 pp. 1-30.
- Chao, A., Gotelli, N.J., Hsieh, T.C., Sander, E.L., Ma, K.H., Colwell, R.K., Ellison, A.M., 2014.
- Rarefaction and extrapolation with Hill numbers: a framework for sampling and estimation in
- species diversity studies. Ecological Monographs 84, 45-67.
- 477 Colwell, R.K., 2013. EstimateS: Statistical estimation of species richness and shared species from
- samples. Version 9.1.0; accessed via: purl.oclc.org/estimates.
- 479 Colwell, R.K., Chao, A., Gotelli, N.J., Lin, S.-Y., Mao, C.X., Chazdon, R.L., Longino, J.T., 2012.
- 480 Models and estimators linking individual-based and sample-based rarefaction, extrapolation and
- comparison of assemblages. Journal of Plant Ecology 5, 3-21.
- 482 Daget, J., 1998. Catalogue raisonnée des mollusques bivalves d'eau douce africains. Backhuys
- 483 Publishers and Orstom, Leiden and Paris.
- de Heinzelin, J., 1983. The Omo group. Annales du Musée Royal de l'Afrique Centrale, Tervuren -
- 485 Série 8 Sciences géologiques 85, 1-365.
- Elderkin, C.L., Clewing, C., Wembo Ndeo, O., Albrecht, C., 2016. Molecular phylogeny and DNA
- barcoding confirm cryptic species in the African freshwater oyster Etheria elliptica Lamarck, 1807
- 488 (Bivalvia: Etheriidae). Biol. J. Linn. Soc. 118, 369-381.
- 489 Feibel, C.S., 1997. A terrestrial auxiliary stratotype point and section for the Plio-Pleistocene
- Boundary in the Turkana Basin, East Africa. Quat. Int. 40, 73-79.
- 491 Feibel, C.S., 2003a. Stratigraphy and depositional setting of the Pliocene Kanapoi Formation, Lower
- Kerio Valley, Kenya, in: Harris, J.M., Leakey, M.G. (Eds.), Geology and vertebrate paleontology



- of the Early Pliocene site of Kanapoi, Northern Kenya. Natural History Museum of Los Angeles
- 494 County, Los Angeles, pp. 9-20.
- 495 Feibel, C.S., 2003b. Stratigraphy and depositional history of the Lothagam sequence, in: Leakey,
- 496 M.G., Harris, J.M. (Eds.), Lothagam: The dawn of humanity in Eastern Africa. Columbia
- 497 University Press, New York, pp. 17-29.
- 498 Feibel, C.S., 2011. A geological history of the Turkana Basin. Evol. Anthropol. 20, 206-216.
- 499 Feibel, C.S., Harris, J.M., Brown, F.H., 1991. Palaeoenvironmental context for the late Neogene of
- the Turkana Basin, in: Harris, J.M. (Ed.), Koobi Fora Research Project. Volume 3-The fossil
- ungulates: geology, fossil artiodactyls and palaeoenvironments. Clarendon Press, Oxford, pp.
- **502** 321-370.
- Foote, M., 1992. Rarefaction analysis of morphological and taxonomic diversity. Paleobiology 18, 1-
- 504 16.
- 505 Fuchs, V.E., 1939. The geological history of the Lake Rudolf Basin, Kenya colony. Philos. Trans. R.
- 506 Soc. Lond. B. Biol. Sci. 229, 219-274 (+ 9 plates).
- 507 Genner, M.J., Michel, E., Erpenbeck, D., De Voogd, N., Witte, F., Pointier, J.-P., 2004. Camouflaged
- 508 invasion of Lake Malawi by an Oriental gastropod. Mol. Ecol. 13, 2135-2141.
- Gotelli, N.J., Colwell, R.K., 2001. Quantifying biodiversity: procedures and pitfalls in the
- measurement and comparison of species richness. Ecol. Lett. 4, 379-391.
- 511 Graf, D.L., Cummings, K.S., 2007. Review of the systematics and global diversity of freshwater
- mussel species (Bivalvia: Unionoida). J. Molluscan Stud. 73, 291-314.
- Harris, J.M., Brown, F.H., Leakey, M.G., 1988. Stratigraphy and paleontology of Pliocene and
- Pleistocene localities west of Lake Turkana, Kenya. Natural History Museum of Los Angeles
- 515 County, Los Angeles.



516 Harris, J.M., Leakey, M.G., 2003. Geology and vertebrate paleontology of the Early Pliocene site of Kanapoi, Northern Kenya. Natural History Museum of Los Angeles County, Los Angeles. 517 Harris, J.M., Leakey, M.G., Cerling, T.E., Winkler, A.J., 2003. Early Pliocene tetrapod remains from 518 519 Kanapoi, Lake Turkana Basin, Kenya, in: Harris, J.M., Leakey, M.G. (Eds.), Geology and vertebrate paleontology of the Early Pliocene site of Kanapoi, Northern Kenya. Natural History 520 521 Museum of Los Angeles County, Los Angeles, pp. 39-113. 522 Hsieh, T.C., Ma, K.H., Chao, A., 2016. iNEXT: Interpolation and extrapolation for species diversity. 523 R package, version 2.0.12. Jacob, J., 1957. Cytological studies of Melaniidae (Mollusca) with special reference to 524 525 parthenogenesis and polyploidy. I. Oögenesis of the parthenogenetic species of Melanoides 526 (Prosobranchia-Gastropoda). Trans. R. Soc. Edinb. 63, 341-352. Lamarck, J.B., 1807. Sur l'Aethérie, nouveau genre de coquille bivalve de la famille des Camacés. 527 Annls Mus. Hist. nat. Paris 10, 398-408. 528 Leakey, M.G., Feibel, C.S., McDougall, I., Walker, A., 1995. New four-million-year-old hominid 529 530 species from Kanapoi and Allia Bay, Kenya. Nature 376, 565-571. 531 Leakey, M.G., Feibel, C.S., McDougall, I., Ward, C., Walker, A., 1998. New specimens and 532 confirmation of an early age for Australopithecus anamensis. Nature 393, 62-66. Lévêque, C., 1972. Mollusques benthiques du lac Tchad: écologie, étude des peuplements et 533 estimation des biomasses. Cah. ORSTOM Sér. Hydrobiol. 6, 3-45. 534 535 Lévêque, C., Saint-Jean, L., 1979. Production secondaire (Zooplancton - Benthos) dans le lac Tchad, 536 Réunion de travail sur la limnologie africaine. Nairobi (KEN), 1979/12/16-23. ORSTOM: Paris,

pp. 1-40.

- Mandahl-Barth, G., 1954. The freshwater mollusks of Uganda and adjacent territories. Annales du
- Musée Royal du Congo Belge, Sciences Zoologiques 32, 1-206.
- Mandahl-Barth, G., 1988. Studies on African freshwater bivalves. Danish Bilharziasis Laboratory,
- 541 Charlottenlund.
- McDougall, I., Brown, F.H., 2008. Geochronology of the pre-KBS Tuff sequence, Omo Group,
- 543 Turkana Basin. J. Geol. Soc. (Lond.) 165, 549-562.
- Powers, D.W., 1980. Geology of Mio-Pliocene sediments of the lower Kerio River Valley, Kenya,
- PhD dissertation. Princeton University, Princeton, NJ.
- R Core Team, 2015. R: A language and environment for statistical computing. Version 3.2.1; R
- foundation for statistical computing, Vienna, Austria.
- Roger, J., 1944. Mollusques fossiles et subfossiles du bassin du lac Rudolphe, in: Arambourg, C.
- 549 (Ed.), Mission Scientifique de l'Omo (1932-1933). Museum national d'Histoire naturelle, Paris,
- 550 pp. 119-155 (+ 2 Pl.).
- 551 Salzburger, W., Van Bocxlaer, B., Cohen, A.S., 2014. Ecology and evolution of the African Great
- Lakes and their faunas. Annu. Rev. Ecol. Evol. Syst. 45, 519-545.
- 553 Samadi, S., David, P., Jarne, P., 2000. Variation of shell shape in the clonal snail *Melanoides tuberculata*
- and its consequences for the interpretation of fossil series. Evolution 54, 492-502.
- Samadi, S., Mavárez, J., Pointier, J.-P., Delay, B., Jarne, P., 1999. Microsatellite and morphological
- analysis of population structure in the parthenogenetic freshwater snail Melanoides tuberculata:
- insights into the creation of clonal variability. Mol. Ecol. 8, 1141-1153.
- 558 Schultheiß, R., Van Bocxlaer, B., Riedel, F., von Rintelen, T., Albrecht, C., 2014. Disjunct
- distributions of freshwater snails testify to a central role of the Congo system in shaping
- biogeographical patterns in Africa. BMC Evol. Biol. 14, e42 (12p.).



561 Sengupta, M.E., Kristensen, T.K., Madsen, H., Jørgensen, A., 2009. Molecular phylogenetic investigations of the Viviparidae (Gastropoda: Caenogastropoda) in the lakes of the Rift Valley 562 area of Africa. Mol. Phylogenet. Evol. 52, 797-805. 563 Stewart, K., 2003. Fossil fish remains from the Pliocene Kanapoi site, Kenya, in: Harris, J.M., Leakey, 564 M.G. (Eds.), Geology and vertebrate paleontology of the Early Pliocene site of Kanapoi, 565 Northern Kenya. Natural History Museum of Los Angeles County, Los Angeles, pp. 21-38. 566 567 Van Bocxlaer, B., 2011. Palaeobiology and evolution of the late Cenozoic freshwater molluscs of the Turkana Basin: Unionidae Rafinesque, 1820, partim Coelatura (Bivalvia: Unionoidea). J. Syst. 568 Palaeontol. 9, 523-550. 569 570 Van Bocxlaer, B., Clewing, C., Mongindo Etimosundja, J.-P., Kankonda, A., Wembo Ndeo, O., 571 Albrecht, C., 2015. Recurrent camouflaged invasions and dispersal of an Asian freshwater gastropod in tropical Africa. BMC Evol. Biol. 15, e33 (18 p.). 572 Van Bocxlaer, B., Van Damme, D., 2009. Palaeobiology and evolution of the late Cenozoic 573 freshwater molluscs of the Turkana Basin: Iridinidae Swainson, 1840 and Etheriidae Deshayes, 574 575 1830 (Bivalvia: Etheroidea). J. Syst. Palaeontol. 7, 129-161. Van Bocxlaer, B., Van Damme, D., Feibel, C.S., 2008. Gradual versus punctuated equilibrium 576 577 evolution in the Turkana Basin molluscs: evolutionary events or biological invasions? Evolution 578 62, 511-520. Van Bocxlaer, B., Verschuren, D., Schettler, G., Kröpelin, S., 2011. Modern and early Holocene 579 mollusc fauna of the Ounianga lakes (northern Chad): implications for the palaeohydrology of 580

the central Sahara. J. Quat. Sci. 26, 433-447.

Van Damme, D., Gautier, A., 1972. Molluscan assemblages from the Late Cenozoic of the Lower

Omo Basin, Ethiopia. Quat. Res. 2, 25-37.



581

582

584 Van Damme, D., Pickford, M., 2010. The Late Cenozoic bivalves of the Albertine Basin (Uganda-Congo). Geo-Pal Uganda 2, 1-121. 585 Ward, C.V., Manthi, F.K., Plavcan, J.M., 2013. New fossils of Australopithecus anamensis from 586 Kanapoi, West Turkana, Kenya (2003-2008). J. Hum. Evol. 65, 501-524. 587 Werdelin, L., 2003. Carnivora from the Kanapoi hominid site, Turkana Basin, Northern Kenya, in: 588 Harris, J.M., Leakey, M.G. (Eds.), Geology and vertebrate paleontology of the Early Pliocene site 589 590 of Kanapoi, Northern Kenya. Natural History Museum of Los Angeles County, Los Angeles, pp. 114-132. 591 Whelan, N.V., Geneva, A.J., Graf, D.L., 2011. Molecular phylogenetic analysis of tropical freshwater 592 mussels (Mollusca: Bivalvia: Unionoida) resolves the position of Coelatura and supports a 593 594 monophyletic Unionidae. Mol. Phylogenet. Evol. 61, 504-514. Williamson, P.G., 1979. Evolutionary implications of Late Cenozoic mollusc assemblages from the 595 596 Turkana Basin, North Kenya. University of Bristol, Bristol, UK, p. 376 (+ appendices). Williamson, P.G., 1981. Palaeontological documentation of speciation in Cenozoic molluscs from 597 598 Turkana Basin. Nature 293, 437-443.

**Table 1.** Comparison between the mollusk fauna derived from lacustrine and fluviatile/deltaic deposits of the Kanapoi Formation. Note however that fluviatile/deltaic mollusk-bearing deposits are substantially less abundant than lacustrine ones, and therefore underrepresented. A = absent; P = present.

Taxa	Lac.	Fluv./Delt.
Gastropoda		
Bellamya	P	A
Cleopatra	P	P
Melanoides	P	A
Gabbiella	P	A
Bivalvia		
Coelatura	P	A
Pseudobovaria	P	A
Aspatharia	P	A
Iridina	P	A
Etheria	P	A

599

600

601

## Figures.

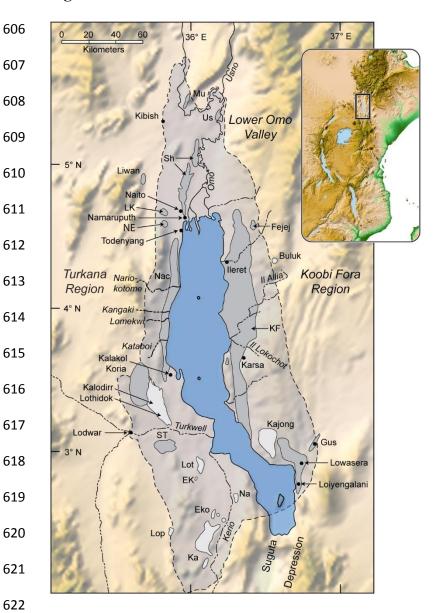
605

623

624

625

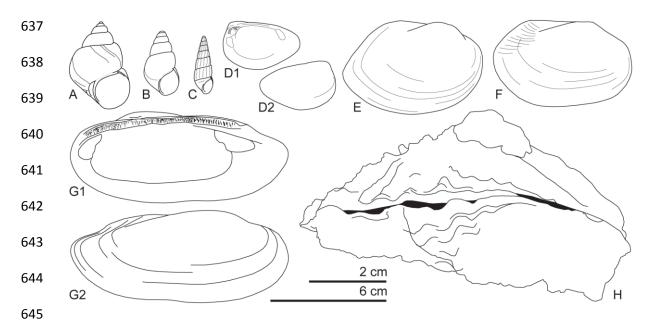
626



**Figure 1.** Topographic representation of the East African Rift System and the Omo-Turkana Basin, with major areas of exposure of Neogene sediments indicated in gray (light gray areas in the South are Mio-Pliocene localities, darker grey areas in the north indicate exposures belonging to the Omo Group). Lake Turkana is indicated in blue and the approximate extent of Paleolake Lonyumun at ~4.1 Ma ago, in which the lacustrine deposits of the Kanapoi Formation (southwestern Omo-



628	Turkana Basin) were deposited, with a dashed line. Although the Kanapoi Formation is
629	geographically restricted and although it captures a relatively short period in the history of the basin,
630	studies of its deposits have been exceptionally informative as to the geology of the basin as well as
631	biological and hominin cultural evolution. Abbreviations: KF = Koobi Fora Formation (Fm.); LK =
632	Loruth Kaado Fm.; Lop = deposits at Loperot; Lot = Lothagam area of Nachukui Fm.; Mu =
633	Mursi Fm.; Na = deposits at Nakoret; Nac = Nachukui Fm., NE = Nayiena Epul Fm.; Sh =
634	Shungura Fm.; ST = deposits at South Turkwell; US = Usno Fm. Map compiled from topographic
635	data of Amante and Eakins (2009) with data from Brown and Feibel (1991) and Feibel (2011).



**Figure 2.** Freshwater mollusks of the Kanapoi Formation. A) *Bellamya*, B) *Cleopatra*, C) *Melanoides*, D) *Pseudobovaria*, E) *Coelatura*, F) *Aspatharia*, G) *Iridina*, H) *Etheria* (ventral view). Scale bar of 2 cm for A-E, that of 6 cm for F-H.

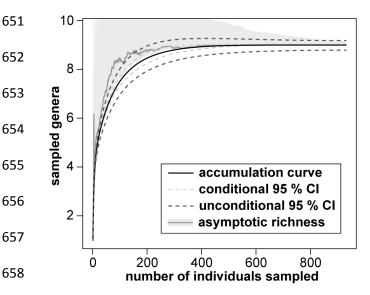


646

647

648

649



**Figure 3.** Individual-based rarefaction at the genus level with all fossil freshwater mollusk finds of the Kanapoi Formation pooled into a single reference sample. The rarefaction curve reaches a plateau well before the total number of individuals sampled (n = 931), and assuming that the reference sample is representative of the total diversity of shell assemblages in the Kanapoi Formation, our material likely approaches the real but unknown genus-level diversity of freshwater mollusks in the formation. This interpretation is supported by the narrow confidence interval based on the unconditional variance at the total sample size and the convergence of the asymptotic richness to the accumulation curve for large samples. 95 % confidence intervals on the rarefaction curve based on conditional or unconditional variance estimators are indicated with dashed lines, the 95 % confidence interval on asymptotic richness as a grayscale polygone.