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Chiral Imines on the Wave: Reactivity of *tert*-Butyl Acrylate and Stereoselectivity Determination Using NMR in Liquid Crystals

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Abstract: In connection with synthetic applications, we have foreseen to study the reactivity of the poorly reactive *tert*-butyl acrylate electrophile with chiral imines of unsymmetrical ketones, using conventional or microwave flash heating under carefully controlled reaction conditions in order to develop a selective and efficient access to the corresponding Michael adducts in which a created stereogenic quaternary carbon center was fully stereocontrolled. Depending on the conditions, either a keto ester or a lactam were obtained. A good correlation was obtained between experiment and theoretical calculations. The stereoselectivity of the process ($\epsilon > 95\%$) was determined using natural abundance ^{13}C - $\{^1\text{H}\}$ NMR in a chiral polypeptide liquid crystal. The scope of the reaction was screened using a set of electrophilic alkenes giving Michael adducts in good yield and similar high enantiocontrol.

Keywords: Aza-ene process, Asymmetric Michael reaction, Enamine activation, Unsymmetrical ketones, *tert*-Butyl acrylate, Quaternary carbon center, chiral polypeptide, liquid crystal, ^{13}C - $\{^1\text{H}\}$ NMR, MW assisted synthesis.

1. Introduction

Although a broad range of methods able to generate new carbon-carbon bonds exists, the establishment of quaternary carbon centres in the proper configuration is among the most restrictive in organic synthesis.¹ Since forty years, the asymmetric Michael addition of chiral imines (AMACI) under neutral conditions has attracted widespread attention as a versatile carbon-carbon bond-forming method, leading to Michael adducts with high levels of regio- and stereocontrol.² Such Michael adducts featuring a stereogenic tetrasubstituted carbon center are useful synthons for the asymmetric synthesis of a variety of bioactive compounds.³ Besides its remarkable efficiency due to a concerted *aza-ene* type mechanism,⁴ the reaction tolerates a large

2. Results and Discussion

As a part of our program directed at exploring the scope of the AMACI, and in connection with synthetic applications for which an orthogonal ester protection cleavable in acidic medium was desired in the Michael adduct, we have foreseen to study the reactivity of *tert*-butyl acrylate **3** with chiral imine **2a** derived from 2-methylcyclopentanone **1a**. Although considerable attention has recently been focused on organocatalytic asymmetric transformations as efficient and convenient methodologies owing to their environmentally friendly characteristics, none of them address the reactivity of bulky electrophiles in this reaction.⁹ Prior to engage in such studies, and because the chiral inductor is available at low price and easily recovered without loss of optical activity at the end of the process, we first analyze the stoichiometric transformation. Due to the steric hindrance of the

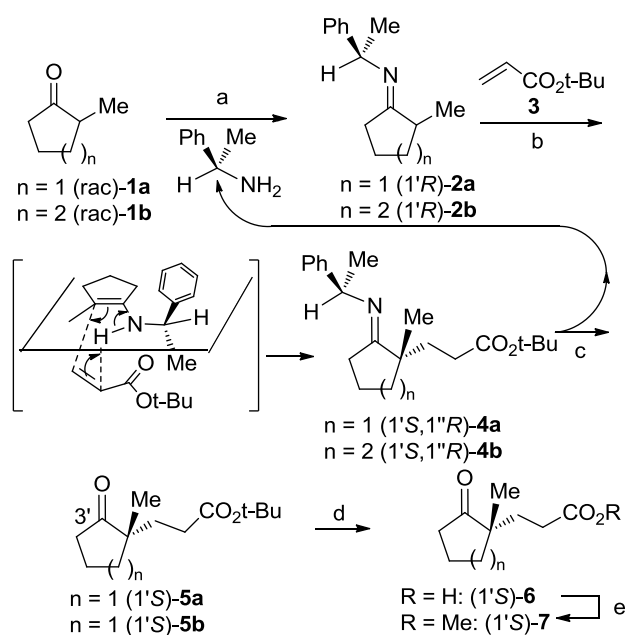
variability in both carbonyl compounds and Michael acceptors, with some limitations for hindered systems for which high pressure activation is required.⁵ Moreover, in the context of improved reaction efficiency, clean processes and short reaction times are desired. In this respect, μW is an efficient way of promoting organic transformations, mainly in solvent-free systems. Thus, interest in μW assisted organic reactions has recently considerably increased.⁶ Nevertheless, little attention has been devoted to the μW effects on selectivity,⁷ particularly for asymmetric Michael reactions.⁸ Herein, we describe our investigation of the reactivity of chiral imines with hindered *tert*-butyl acrylate and related electrophiles under microwave activation.

acceptor, we have turned to use microwave irradiation (μW) under carefully controlled reaction conditions, in order to develop a simple and efficient access to Michael adduct **5a** and the derived keto-acid **6** and compared this method to conventional heating (Δ) (Scheme 1).

In general, the AMACI is carried out from the intermediary chiral imine and the electrophile, in the absence of solvent, at room temperature or using moderate Δ (up to 80 °C) as the regio- and stereocontrol of this reaction are only slightly sensitive to heat.² When the reaction was performed at 25 °C for 7 days, the conversion was not complete and 15% yield of Michael adduct **5a** was obtained upon hydrolysis (Table 1, entry 1). Thus, due to its bulkiness, *tert*-butyl acrylate **3** reacts much slower than its methyl counterpart.¹⁰ At 60 °C, all the imine **2a** was consumed in one day; however, side

polymerization reactions resulted in a modest 58% yield of adduct **5** (Table 1, entry 2).

20 °C (d) HCO₂H, 20 °C, 89%; (e) CH₂N₂, Et₂O, 0 to 20 °C, 2 h, 95%.



Scheme 1 Reagents and conditions: (a) 1.01 eq 1-phenylethylamine, cyclohexane, reflux, Dean-Stark, overnight, 89% (b) 2 equiv. **2**, 25-200 °C (see Tables 1 and 2); (c) 20% aq. AcOH, THF,

When the reaction was performed at 100 °C for 4 h, alkylated imine **4a**¹¹ was obtained as the sole product leading upon hydrolysis to keto ester **5** in 79% yield. On the basis of its ¹³C NMR spectrum, crude imine **4a** exists as a single stereoisomer (de >95%),¹¹ as the result of a highly stereo-controlled process, giving rise to Michael adduct **5a** with a >95% enantiomeric excess (ee). The reaction duration was markedly reduced to 30 min at 150 °C and ketoester **5** was obtained in an optimum 92% yield upon hydrolysis. However, the stereoselectivity proved to be lower (ee 80%) than at 100 °C (Table 1, entry 6). Finally, both the efficiency and the stereoselectivity dropped at 200 °C (Table 1, entry 7).

Table 1. Effect of temperature on the synthesis of ketoester **4** by condensation of chiral imine **1** with *tert*-butyl acrylate **2**.^a

Entry	Temperature (°C)	Time	5	
			yield % ^b	ee% ^d
1	25	7 d	15	nd ^c
2	60	1 d	58	> 95
3 ^b	100	4 h	79	> 95
4	150	5 min	14	nd ^c
5	150	15 min	59	nd ^c
6	150	30 min	92	80
7	200	30 min	57	80

^a: 2 Equivalents; ^b: Isolated yield of purified keto-ester **5**; ^c: Not determined

^d: Determined by ¹³C-¹H NMR in chiral liquid crystals.

Concerning the stereoselectivity of the process, attempts to measure accurately the ee in adduct **5a** or in the related keto-acid **6** using either chiral HPLC or ¹H NMR spectroscopy in the presence of chiral shift reagent [Eu(hfc)₃] were

unsuccessful. However, screening of the selectivity was made possible on the examination of ¹³C NMR spectra of crude imine **4a** and gave satisfactory results. In order to ascertain that no epimerization of Michael adduct **5a** occurred

during hydrolysis of imine **4a**, we turned our attention to NMR spectroscopy in polypeptide chiral liquid crystals (CLC) that generally provides an efficient alternative to classical methods when these latter failed or gave poor results.¹² We used here $^{13}\text{C}\{-^1\text{H}\}$.¹³ Spectral enantio-discriminations using $^{13}\text{C}\{-^1\text{H}\}$ NMR in a CLC are based on ^{13}C chemical shift anisotropy (CSA) differences. In practice, when the enantiomers are oriented differently inside the CLC, we can expect to observe two distinct resonances for each non-equivalent carbon atom discriminated. A priori, each carbon atom is a potential spy, thus increasing the possibility to visualize enantiomers.¹⁴ Various carbon sites show enantiodiscrimination, but the best spectral separation was obtained for C3' in compound **5**. Considering the S/N ratio, the error on the ee has

been estimated around 5% of the true value. Figure 1 shows the evolution of two $^{13}\text{C}\{-^1\text{H}\}$ signals associated to C-3' (Scheme 2) both in racemic (a) and enantio-enriched forms [Table 1, entry 6 (b) and Table 2, entry 4 (c)] oriented in a PBLG/ dichloromethane phase. The differences in peak intensity reveal the evolution of the ee. Although the separation observed at 100 MHz is rather small (< 3 Hz), a suitable evaluation of the ee is possible using deconvolution process. The absence of peak for the minor enantiomer in Fig. 1c indicates that the ee is >95%. Accuracy of the method was ascertained by measuring gradual mixtures of the racemic ketoester **5a** (prepared from racemic 1-phenylmethanimine using the same conditions) with the pure enantiomer.

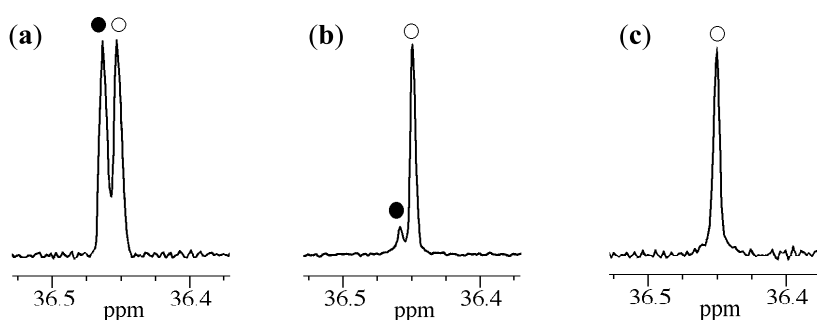


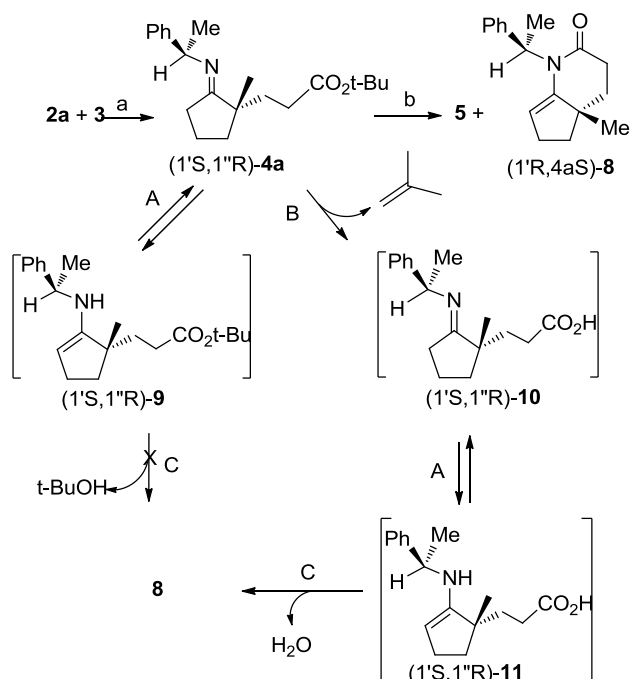
Figure 1. $^{13}\text{C}\{-^1\text{H}\}$ signals of carbon atom C-3' in ketoester **5** prepared in racemic form (a) and (S)-enriched one (b, c).

Analysis of NMR results obtained for synthesis of adduct **5a** using Δ clearly indicates a sharp decrease in the stereoselectivity (>95 to 80% ee) when the temperature was increased from 100 °C to 200 °C (Table 1, entries 3-4 and 6-7). This phenomenon can tentatively be explained assuming a possible competitive retro-Michael addition leading to a partial racemization of the Michael adduct under rather drastic conditions.¹⁵

Having secured an access to Michael adduct **5a**, a chemical correlation of its corresponding methyl keto ester **6**¹⁰ was undertaken to ascertain the absolute configuration in **5a** (Scheme 1). Thus, keto acid **6** was easily obtained upon formic acid treatment of ketoester **5a** (Table 1, entry 3) in good yield and directly subjected to diazomethane esterification leading to (-)-(S)-**7**. As expected, the sense of induction is in

accordance to the empirical rule defined for this reaction,^{2,3} with the same sense of asymmetric induction as those obtained using methyl acrylate as the electrophile. This stereoselectivity originates from the *aza-ene* type mechanism in which internal transfer of the proton born by the nitrogen atom of the more substituted tautomeric enamine is concerted with the creation of the C-C bond (Scheme 1).

We then turned our focus to the study of this Michael reaction, carrying out the experiments under μ W. As can be seen from Table 2 and Scheme 3, μ W affects the reactivity, the stereo- and unexpectedly the chemoselectivity (Table 2). The closed vessel system was chosen in order to contain the toxic and volatile Michael acceptors in the reaction vessel, and to monitor the possible extent of pressure elevation during the microwave irradiation. The possibility of running reactions in an inert gas atmosphere is another distinct advantage with the sealed reaction vessel strategy. Despite sensitivity of chiral imines **2** toward water, this was not necessary in this case. The first observation was the role of stirring upon efficiency of the reaction. This parameter was found to be critical to the success of the reaction (compare entries 1,3,5 with entries 2,4,6 and 7).¹⁶



Scheme 2 Reaction pathway to lactam **8** under μ W. *Reagents and conditions:* (a) 2 equiv. **3**, μ W, 200 °C, 30 min; (b) 20% aq. AcOH, THF, 20 °C.

When mixtures of imine **2a** and alkene **3** were submitted to μ W for 30 min at 100 °C with an optimal power of 30 W (Table 2, entry 2), the only reaction product was the expected chiral imine **4a**, leading to keto-ester **5a** upon hydrolytic workup. This result is a noteworthy improvement over results obtained from conventional heating at the same temperature for 4 hours (Table 1, entry 3) in terms of yields and reaction time, the enantioselectivity remaining the same.

At 150 °C, within 30 min, whereas yields were nearly comparable (Table 1, entry 6 and Table 2, entry 4), the enantioselectivity was largely improved under μ W when compared to Δ (see Figure 1) with similar set of conditions (temperature, pressure, profiles of heating rates).

Table 2. Effect of μ W irradiation and stirring upon condensation of imine **2a** (entries 1-7) and **2b** (entries 8-9) to *tert*-butyl acrylate **3**^a.

Entry	P (W)	T (°C)	Stirring	ΔP (bar)	5a/5b yield (ee) %	8
1	30	100	no	0.1	85	0
2	30	100	yes	0.1	100 (> 95 ^d)	0
3	80	150	no	0.7	67	0
4	80	150	yes	0.3	89 (> 95 ^d)	0
5	80	200	no	3.5	11	25
6	80	200	yes	1.4	41	14
7	100	200	yes	13.0	0	53
8	80	100	yes	0.8	92 (> 95 ^d)	0
9	100	200	yes	14.0	0	68

^a: 2 Equivalents; ^b: Isolated yield of purified keto-ester **5a** after 30 min μW irradiation and subsequent hydrolysis; ^c: Not determined; ^d: Determined by ¹³C-¹H NMR in chiral liquid crystals.

The most intriguing feature concerned the production of lactam **8** under forcing μW conditions (Scheme 3; Table 2, entries 5-7) instead of keto ester **5a**, since this lactam **8** was never observed in the conventional thermal process (Δ). This very important specific μW effect appeared when the reaction was performed at 200 °C. In contrast with Δ where the expected Michael adduct **5a** was obtained (Table 1, entry 7), within 30 min under μW irradiation at 200 °C, quite surprisingly, the lactam **8** was formed as the sole product (Scheme 2; Table 2, entry 7).

This noticeable finding on chemoselectivity can be justified by considering the possibility for μW to favor a very polar mechanism consisting in the nucleophilic addition (Scheme 2, routes C) of the enamines **9** or **11** to the carbonyl group of either a *tert*-butyl ester (**9** \rightarrow **8**) with the release of *tert*-butanol, or an acid (**11** \rightarrow **8**) with elimination of a water molecule. Secondary enamines **9** or **11** are in tautomeric equilibrium (Scheme 2, routes A) with imines **4a** or **10** respectively, the latter being issued from the thermolysis of the *tert*-butyl ester group in imine

4 with concomitant generation of isobutene (Scheme 2, route B, **4** \rightarrow **10**).

In order to elucidate the pathway to lactam **8**, a GC-mass analysis of the headspace of the reaction mixture was undertaken, and compared to those of the starting chiral imine **2**, the acrylate **3** and *tert*-butanol having been separately irradiated under the same conditions (200 °C, 100 W, 30 min in closed reaction vessels). While isobutene was detected in the headspace of the reaction mixture and the *tert*-butyl acrylate **3** sample, it was not present in the *tert*-butanol or starting imine **1** ones.¹⁷ This indicates that, under μW at 200 °C, the formation of lactam **8** occurred via a tandem Michael addition/deprotection/aza-annulation sequence implying the thermolysis of the *tert*-butyl ester group in imine **4** (path B/C).

Dealing with μW effect on enantioselectivity, the superiority of μW reveals the intervention of non-purely thermal μW specific effects. Although the effects observed in microwave-irradiated chemical transformations can in most cases be rationalized by purely bulk thermal

phenomena associated with rapid heating to elevated temperatures,¹⁸ we have conducted all experiments in the same conditions (closed vessels, same scale, same magnetic barrel) either in the microwave chamber or by immersion in a preheated oil bath in order to avoid as possible any difference in the temperature profiles. They can be justified by considering the reaction mechanism,⁴ expecting μW effects when the polarity of the system increases during the reaction progress. It will be the case when the transition state (TS) of a reaction is more polar than its ground state, thus leading to a decrease in the activation energy.¹⁹ Data obtained for the transition states for the *Re* and *Si*-approaches of imine **2** and *tert*-butyl acrylate **3** are consistent with the previous studies:^{4b} *Re* approach: forming CC bond: 1.87 Å and forming CH bond: 2.56 Å; *Si* approach: forming CC bond: 1.89 Å and forming CH bond: 2.50 Å (Figure 2). As it was shown that this Michael addition proceeds through a concerted *asynchronous* 'aza-ene like' mechanism,⁴ the TS has thus a certain polarity, a

situation for which μW effects are expected. This fact explains the intervention of μW effects upon exaltation of reactivity (comparing yields at 100 °C). To support this assumption, taking into account previous related AM1 computational investigations,²⁰ we calculate the corresponding approaches between the enamine tautomer of imine **2** with *tert*-butyl acrylate **3**. The *Re*-approach (the favored one) leads to a slightly more polar as well as more asynchronous than the *Si*-approach TS (Figure 2).²¹ Consequently, this TS will be slightly favored due to a better dipole-dipole stabilization by μW . Therefore, under μW , the selectivity in favor of the *Re*-approach will be even more improved (exp. from 80 to > 95% ee). Reduced reversibility of the Michael reaction under microwave conditions could also contribute to the superior enantioselectivity, both phenomenon leading to an increased stereoselectivity in such conditions.

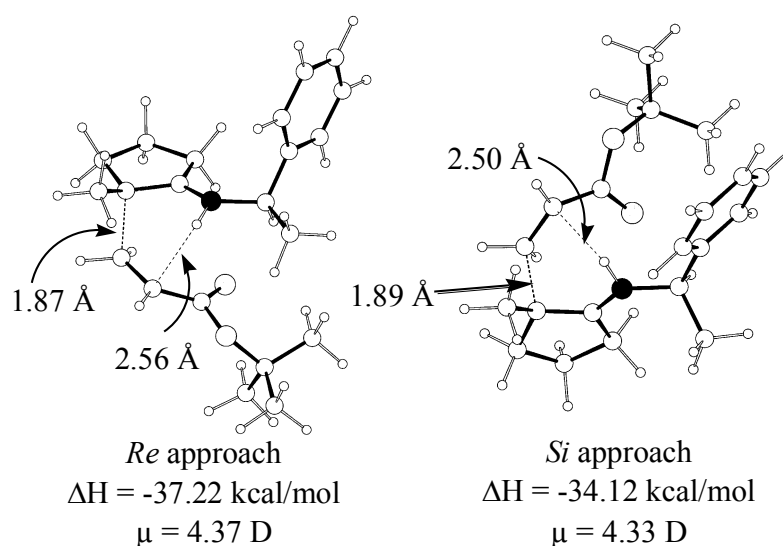


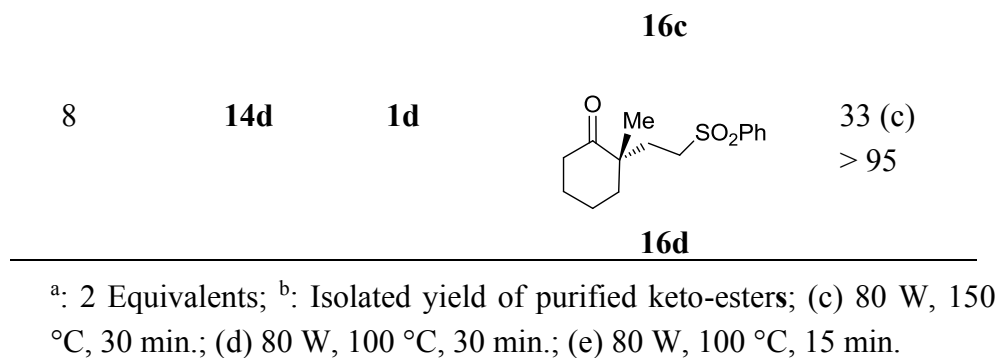
Figure 2. Transition structures at the RHF AM1 level for *Re* (left) and *Si* (right) approaches of *tert*-butyl acrylate **3** to enamine imine **2**, their respective enthalpies of formation and dipole moment.²⁰

The same trend was observed with chiral imine **2b** derived from 2-methylcyclohexanone **1b**. Lactame **8b** was obtained when the reaction was performed at 200 °C upon irradiation at 100 W for 30 min (Table 2, entry 9) in a 68% yield while the expected Michael adduct **5b** was

obtained in the optimized conditions (100 °C, 80 W, Table 2, entry 8) in 92% yield with >95% ee. We next examine the reactivity of these imine in this μ W promoted AMACI (Table 3).

Table 3. Effect of temperature on the synthesis of ketoester **4** by condensation of chiral imine **1** with electrophilic alkenes **12a-d**.^a

Entry	Electrophile	Ketone	Michael adduct	Yield % ^b ee%
1		1a	 7a	87 ^(d) > 95
2		1a	 15b	76 ^(d) > 95
3 ^b		1a	 15c	47 ^(e) > 95
4		1a	 15d	95 ^(b) >95
5	14a	1b	 7b	77 ^(d) > 95
6	14b	1b	 16b	60 ^(c) > 95
7	14c	1c		74 ^(c) > 95



The reaction performed well, giving the expected Michael adducts in adduct in 33-95% yield, in a first series of experiments. Ee of the adducts were measured at the level of the crude imines as >95% (no diastereoisomers detected).

With these satisfactory results in hand, we will turn to the study of a catalytic version of this

3. Materials and Methods

3a. Chemistry: General: All reactions not involving aqueous media were carried out under a nitrogen atmosphere in a flame-dried glassware. Commercial reagents were used without further purification. Reactions were followed by ¹H NMR in CDCl₃ or using thin-layer chromatography, carried out on silica gel plates, which were viewed by UV irradiation at 254 nm and/or by staining with phosphomolybdic acid or *p*-anisaldehyde. Flash column chromatography was performed with 230-400 mesh silica gel. Melting points were recorded on a digital melting point apparatus. IR spectra were recorded with a Fourier transform spectrometer Bruker VECTOR 22. NMR spectra of the crude reaction mixtures were recorded in CDCl₃ containing a pinch of sodium carbonate in order to prevent hydrolysis of the imines. Imines proved to be stable for days in such conditions. ¹H NMR spectra were recorded at 300 K, at 200 or 400 MHz on a Bruker AC 200 or Bruker Avance 400 spectrometer, with CHCl₃ as internal

reaction, in the context of a greener generation of quaternary carbon centers in a simple manner. Work is in progress to extend this μW activation mode to engage substituted acceptors in the AMACI.

standard ($\delta_{\text{H}} = 7.26$ ppm). ¹³C NMR spectra were recorded at 300 K, at 50 or 100 MHz, with the central peak of CHCl₃ as internal standard ($\delta_{\text{C}} = 77.0$ ppm, central line). Recognition of methyl, methylene, methine and quaternary carbon nuclei in ¹³C NMR spectra rests on the *J*-modulated spin-echo sequence. 2D NMR experiments (COSY, HMQC, HMBC and NOESY) were used for the assignments of signals in the ¹H and ¹³C NMR spectra. For NMR measurements, ¹³C 1D NMR experiments in polypeptidic oriented solvents were performed on a Bruker DRX-400 equipped with a BBO probe, and hence no additional hardware equipment is basically required. All proton-decoupled ¹³C NMR experiments were recorded by applying the WALTZ-16 composite pulse sequence to decouple protons and benefit from NOE effect. For unambiguous assignment of enantiomers in chiral NMR, comparison was made in all cases with the corresponding racemates. Optical rotations were measured at 589 nm in a 1 dm-cell

using an Optical Activity Limited AA-10R apparatus and are expressed in g/100mL. Elemental analyses were performed by the Service de microanalyse, BioCIS, Châtenay-Malabry, France, with a Perkin-Elmer 2400 analyzer. A CEM Discover monomode reactor with an accurate control of temperature and pressure by modulation of emitted W was used for the microwave experiments. **Caution:** It is essential that great precaution be taken when carrying out organic reactions in sealed vessels. In particular, safety devices are to be used including appropriate septa as a pressure relief system and an automatic cut off of the microwave irradiation before the pressure limit of the vessels has been reached. See: Raner, K. D.; Strauss, C. R.; Trainor, R. W.; Thorn, S. J. *J. Org. Chem.* **1995**, *60*, 2456 and references cited therein. In this study, the maximum developed pressure was 14 bar at 100 W and 200 °C, far below the pressure limit (20 bar).

Preparation of chiral imines **2** exemplified for **2a**: In a 100 mL round bottom flask equipped with a Dean-Stark apparatus, 21.6 mL (0.2 mol) 2-methylcyclopentanone, 27.3 mL (0.21 mol, 1.05 equiv.) (*R*)-1-phenylethylamine (ee = 99%) are successively added to cyclohexane (50 mL). The resulting mixture was stirred for 18 h under nitrogen at 110 °C (oil bath) with azeotropic removal of water. Cyclohexane was then distilled and fractional distillation of the crude under reduced pressure afforded the desired chiral imine **1** as a colourless oil.

(1'R)-(2-Methylcyclopentylidene)-(1'-phenylethyl)-amine (2a). Colorless oil (89%); B.p. = 80 °C (0.01 Torr); IR (neat, ν cm⁻¹): 2958, 1673; ¹H NMR (200 MHz, CDCl₃) δ ppm: 7.33-7.05 (m, 5H), 4.43-4.28 (m, 1H), 2.45-1.15 (m, 7H), 1.43 and 1.41 (d, *J* = 6.1 Hz, 3H), 1.11 and 1.10 (d, *J* = 6.7 Hz, 3H); ¹³C NMR (50 MHz, CDCl₃)

δ ppm: 181.0 and 180.6 (C), 146.2 and 145.9 (C), 128.1 (2CH), 126.5 (CH), 126.3 (2CH), 61.5 and 61.2 (CH), 41.2 and 41.1 (CH), 32.8 and 32.7 (CH₂), 28.7 and 28.5 (CH₂), 24.8 and 24.5 (CH₃), 22.5 and 22.4 (CH₂), 17.5 (CH₃).

General procedure for the asymmetric Michael reactions using microwaves: Mixtures of imines **2** (1 to 6 mmol) and 2 equivalents of electrophilic alkene were placed in sealed,³³ 10 mL heavy-walled pyrex tubes.³⁴ The tubes were introduced in the cavity of a single-mode³⁵ device allowing control of irradiation power (up to 300 W), time, temperature and pressure (see article, Tables 1 and 2).³⁶ The tube was opened after the reaction mixture was rapidly cooled down to room temperature, and excess *tert*-butyl acrylate was removed in vacuo. An aliquot of crude reaction mixture was analyzed by ¹H and ¹³C NMR. For the crude reaction mixtures containing alkylated imine, after being vigorously stirred with 20% aqueous acetic acid (2 mL/mmol) and THF (2 mL/mmol) for 17 hours, the reaction mixture was concentrated, then thoroughly extracted with Et₂O (3 x 10 mL). The combined organic phase was washed successively with saturated NaHCO₃ and NaCl solutions, dried (MgSO₄), filtered over Celite[®] and concentrated in vacuo. Chromatographic purification on silica gel (cyclohexane:ethyl acetate, 9:1) afforded keto derivatives **5** and/or lactam **8** as colorless oils.

tert-Butyl (1'S,1'R)-3-[1'-Methyl-2-(1''-phenyl-ethyl-imino)-cyclopentyl]-propionate 4a. IR (neat, ν cm⁻¹): 2966, 2868, 1726, 1673; ¹H NMR (200 MHz, CDCl₃) δ ppm: 7.38-7.07 (m, 5H), 4.35 (q, 1H, *J* = 6.6 Hz), 2.37-2.04 (m, 4H), 1.86-1.41 (m, 6H), 1.38 (s, 9H), 1.36 (d, 3H, *J* = 6.6 Hz), 0.98 (s, 3H); ¹³C NMR (50 MHz, CDCl₃) δ ppm: 180.7 (C), 173.5 (C), 146.2 (C), 128.0 (2CH), 126.3 (2CH), 126.1 (CH), 79.7 (C), 60.8 (CH), 45.5 (C), 37.0 (CH₂), 33.4 (CH₂), 30.9

(CH₂), 28.4 (CH₂), 28.0 (3 CH₃), 24.8 (CH₃), 23.9 (CH₃), 20.5 (CH₂).

tert-Butyl (1'S,1'R)-3-[1'-Methyl-2-(1''-phenyl-ethyl-imino)-cyclohexyl]-propionate

4b. IR (neat, ν cm⁻¹): 3061, 3063, 2966, 2926, 1658, 1493, 1447; ¹H NMR (300 MHz, CDCl₃) δ ppm: 7.41-7.12 (m, 5H), 4.69 (q, 3H, $J = 6.4$ Hz), 2.40-2.21 (m, 1H), 1.57-1.30 (m, 8H), 1.17-1.11 (2d, 3H, $J = 4.3$ Hz), 1.01-0.99 (d, 3H, $J = 4.9$ Hz); RMN ¹³C (75 MHz, CDCl₃) δ ppm: 173.1 (C), 146.7 (C), 127.9 (CH), 126.3 (CH), 126.2 (CH), 125.9 (CH), 125.8 (CH), 57.3 (CH), 42.0 (CH₂), 35.6 (CH), 27.5 (CH₂), 25.4 (CH₃), 25.3 (CH₂).

tert-Butyl (1'S)-3-(1'-Methyl-2'-oxocyclopentyl)-propionate 5a. IR (neat, ν cm⁻¹): 2968, 2934, 2872, 1727; ¹H NMR (200 MHz, CDCl₃) δ ppm: 2.38-2.06 (m, 4H), 2.0-1.54 (m, 6H), 1.36 (s, 9H), 0.98 (s, 3H); ¹³C NMR (50 MHz, CDCl₃) δ ppm: 222.3 (C), 172.6 (C), 80.1 (C), 47.4 (C), 37.3 (CH₂), 35.9 (CH₂), 31.3 (CH₂), 30.5 (CH₂), 27.9 (3CH₃), 21.3 (CH₃), 18.4 (CH₂); $[\alpha]_D^{24} = -27.8$ ($c = 4.5$, EtOH_{abs}); Anal. calcd for C₁₃H₂₂N: C, 68.99; H, 9.80. Found: C, 68.84; H, 9.75%.

tert-Butyl (1'S)-3-(1'-Methyl-2'-oxocyclohexyl)-propionate 5b. IR (neat, ν cm⁻¹): 2933-2866, 1727, 1704; RMN ¹H (200 MHz, CDCl₃) δ ppm: 2.49-2.33 (2H), 2.28-2.12 (1H), 2.10-1.94 (3H), 1.92-1.52 (8H), 1.42 (9H), 1.02 (3H); RMN ¹³C (75 MHz, CDCl₃): δ ppm: 215.7 (C), 173.1 (C), 82.3 (C), 47.9 (C), 39.3 (CH₂), 38.7 (CH₂), 32.4 (CH₂), 30.3 (CH₂), 28.7 (3CH₂), 27.4 (CH₂), 22.4 (CH₃), 21.0 (CH₂).

(4aS,1'R)-4a-Methyl-1-(1'-phenylethyl)-1,3,4,4a, 5,6-hexahydro-[1]pyrindin-2-one 8. IR (neat, ν cm⁻¹): 1665, 1629; ¹H NMR (CDCl₃, 200 MHz) δ ppm: 7.24-7.06 (m, 5H), 6.14 (q, 1H, $J = 7.1$ Hz), 4.31 (t, 1H, $J = 2.5$ Hz), 2.62-2.53 (m, 1H), 2.57 (dd, 1H, $J = 8.3, 4.0$ Hz), 2.30 (m, 1H), 2.05 (ddd, 1H, $J = 15.6, 9.0, 3.1$ Hz), 1.76-1.43 (m, 4H), 1.55 (d, 3H, $J = 7.1$ Hz), 1.05

(s, 3H); ¹³C NMR (CDCl₃, 50 MHz) δ ppm: 169.3 (C), 143.7 (C), 141.2 (C), 128.3 (2CH), 126.5 (CH), 126.1 (2CH), 105.7 (CH), 49.9 (CH), 43.5 (C), 38.3 (CH₂), 33.4 (CH₂), 29.9 (CH₂), 28.5 (CH₂), 21.1 (CH₃), 14.8 (CH₃); Anal. calcd for C₁₇H₂₁NO: C, 79.96; H, 8.29; N, 5.49. Found: C, 80.04; H, 8.56; N, 5.15%.

3-[1-Methyl-2-(1-phenyl-ethylimino)-cyclopentyl]-propionitrile 17c. IR (neat, ν cm⁻¹): 3027, 2962, 2867, 2245, 1672; ¹H NMR (CDCl₃, 300 MHz) δ : 7.35-7.17 (m, 5H), 4.41 (q, 1H, $J = 6.6$ Hz), 2.65-2.27 (m, 4H), 1.94-1.57 (m, 6H), 1.41 (d, 3H, $J = 6.6$, Hz), 1.06 (s, 3H); ¹³C NMR (CDCl₃, 75 MHz) δ ppm: 179.9 (C), 145.8 (C), 128.2 (2CH), 126.4 (CH), 126.3 (2CH), 120.7 (C), 61.2 (CH), 45.4 (C), 37.1 (CH₂), 35.8 (CH₂), 28.5 (CH₂), 24.9 (CH₃), 23.8 (CH₃), 20.5 (CH₂), 12.4 (CH₂).

3-[1-Methyl-2-(1-phenyl-ethylimino)-cyclohexyl]-propionitrile 18c. IR (neat, ν cm⁻¹): 2969, 2931, 2866, 2247, 1705, 1650; ¹H NMR (CDCl₃, 300 MHz) δ ppm: 7.39-7.20 (m, 5H), 4.70 (q, 1H, $J = 6.6$ Hz), 2.53-2.14 (m, 5H), 2.13-1.81 (m, 1H), 1.74-1.27 (m, 6H), 1.36 (d, 3H, $J = 6.6$, Hz), 1.04 (s, 3H); ¹³C NMR (CDCl₃, 75 MHz) δ ppm: 172.1 (C), 146.6 (C), 128.2 (2CH), 126.4 (3CH), 121.1 (C), 57.8 (CH), 43.2 (C), 39.0 (CH₂), 35.2 (CH₂), 27.1 (CH₂), 25.8 (CH₃), 24.9 (CH₂), 23.9 (CH₃), 21.2 (CH₂), 12.4 (CH₂).

Methyl (1'S)-3-(1'-Methyl-2'-oxocyclohexyl)-propionate 7b. IR (neat, ν cm⁻¹): 2960, 1731, 1437; ¹H NMR (200 MHz, CDCl₃) δ ppm: 3.33 (s, 3H), 2.74-2.18 (m, 4H), 1.98-1.62 (m, 6H), 1.01 (s, 3H); ¹³C NMR (50 MHz, CDCl₃) δ ppm: 222.3 (C), 173.8 (C), 51.5 (CH₃), 47.4 (C), 37.3 (CH₂), 35.9 (CH₂), 32.0 (CH₂), 29.2 (CH₂), 21.2 (CH₃), 18.4 (CH₂); Anal. for C₁₀H₁₆O₃, calcd. C, 65.19; H, 8.75; found C, 65.08; H, 8.79.; **12a** $[\alpha]_D^{20} -34.5$ ($c = 2$, EtOH_{abs}); lit.³⁸ *ent-12a* $[\alpha]_D^{20} +35.7$ ($c = 2$, EtOH_{abs}).

3-(1-Methyl-2-oxo-cyclopentyl)-propionitrile

15c. IR (neat, ν cm^{-1}): 2965, 2927, 2872, 2247, 1731; ^1H NMR (300 MHz, CDCl_3) δ ppm: 2.46-2.14 (m, 4H), 1.97-1.69 (m, 6H), 1.02 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ ppm: 221.2 (C), 119.7 (C), 47.2 (C), 37.2 (CH_2), 35.7 (CH_2), 32.0 (CH_2), 20.9 (CH_3), 18.4 (CH_2), 12.4 (CH_2); $[\alpha]_{\text{D}}^{20} = -34.6^\circ$ ($c = 0.003$, EtOH_{abs}); Anal. Calcd for $\text{C}_9\text{H}_{13}\text{NO}$: C, 71.49; H, 8.67; N, 9.26; O, 10.58. Found: C, 71.06; H, 8.12; N, 9.07.

3-(1-Methyl-2-oxo-cyclohexyl)-propionitrile

16c. IR (neat, ν cm^{-1}): 2937, 2867, 2246, 1700, 1451; ^1H NMR (300 MHz, CDCl_3) δ ppm: 2.48-2.37 (m, 1H), 2.33-2.26 (m, 3H), 1.97-1.64 (m, 8H), 1.12 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ ppm: 214.3 (C), 120.0 (C), 47.6 (C), 38.5 (2 CH_2), 33.6 (CH_2), 27.1 (CH_2), 22.2 (CH_3), 20.8 (CH_2), 12.3 (CH_2); Anal. for $\text{C}_{10}\text{H}_{15}\text{NO}$, calcd. C, 72.69; H, 9.15; found C, 72.23; H, 8.49; MS (ESI): 166 ($\text{M}+1$) **16c** $[\alpha]_{\text{D}}^{24} 9.7$ ($c = 0.02$, EtOH_{abs}).

[2-(2-Benzenesulfonyl-ethyl)-2-methyl-cyclopentylidene]-(1-phenyl-ethyl)-amine 17d IR (neat, ν cm^{-1}): 2962, 1671, 1447, 1305, 1145; ^1H NMR (300 MHz, CDCl_3) δ ppm: 7.70-7.45 (m, 10H), 4.34 (q, 1H, $J = 6.6$ Hz), 3.37 (mc, 2H), 2.24 (bt, 2H, $J = 5.9$ Hz), 1.80-1.40 (m, 6H), 1.31 (d, 3H, $J = 6.6$ Hz), 1.00 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ ppm: 179.6 (C), 146.0 (C), 138.3 (CH), 133.6 (CH), 129.2 (2CH), 129.1 (2CH), 128.1 (CH), 127.7 (CH), 126.2 (CH), 67.8 (CH_2), 61.1 (CH), 52.2 (C), 37.6 (CH_2), 28.3 (CH_2), 25.5 (CH_2), 25.0 (CH_3), 23.8 (CH_3), 20.4 (CH_2).

[2-(2-Benzenesulfonyl-ethyl)-2-methyl-cyclohexylidene]-(1-phenyl-ethyl)-amine 18d IR (neat, ν cm^{-1}): 2928, 1707, 1650; ^1H NMR (300 MHz, CDCl_3) δ ppm: 8.00-7.87 (m, 4H), 7.66-7.49 (m, 6H), 4.61 (q, 1H, $J = 6.6$ Hz), 3.35-3.20 (m, 2H), 2.42-2.29 (m, 2H), 2.18-2.00 (m, 2H), 1.90-1.38 (m, 6H), 1.24 (d, 3H, $J = 6.6$ Hz), 1.01 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ ppm: 172.2 (C), 146.6 (C), 139.3 (C), 133.4 (CH),

129.1 (CH), 128.2 (CH), 126.4 (CH), 57.7 (CH), 52.2 (CH_2), 43.0 (C), 39.3 (CH_2), 31.9 (CH_2), 27.0 (CH_2), 25.7 (CH_3), 24.7 (CH_2), 24.2 (CH_3), 21.2 (CH_2).

2-(2-Benzenesulfonyl-ethyl)-2-methyl-cyclopentanone 15d Mp = 69 °C; IR (neat, ν cm^{-1}): 2965, 2870, 1729; ^1H NMR (300 MHz, CDCl_3) δ ppm: 7.87 (bd, 2H, $J = 8.3$ Hz), 7.68-7.45 (m, 3H), 3.14 (ddd, 1H, $J = 18.4, 12.0, 4.7$ Hz), 2.96 (ddd, 1H, $J = 18.4, 12.2, 4.9$ Hz), 2.35-2.07 (m, 2H), 1.91-1.72 (m, 6H), 0.94 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ ppm: 221.4 (C), 138.7 (C), 133.7 (CH), 129.2 (2CH), 127.9 (2CH), 51.8 (CH_2), 46.7 (C), 37.2 (CH_2), 36.2 (CH_2), 28.9 (CH_2), 21.0 (CH_3), 18.4 (CH_2); $[\alpha]_{\text{D}}^{23} = -19.4^\circ$ ($c = 0.0075$, EtOH_{abs}); MS (APCI): m/z 267 (100%) $[\text{M} + \text{H}]^+$; Anal. Calcd for $\text{C}_{14}\text{H}_{18}\text{O}_3\text{S}$: C, 63.13; H, 6.81; O, 18.02; S, 12.04. Found: C, 62.65; H, 6.80.

2-(2-Benzenesulfonyl-ethyl)-2-methyl-cyclohexanone 16d Mp = 74-77 °C; IR (neat, ν cm^{-1}): 2934, 2868, 1700; ^1H NMR (300 MHz, CDCl_3) δ ppm: 7.86 (bd, 2H, $J = 8.3$ Hz), 7.65-7.49 (m, 3H), 3.06 (ddd, 1H, $J = 13.7, 11.9, 5.0$ Hz), 2.98 (ddd, 1H, $J = 13.7, 11.7, 5.0$ Hz), 2.39-2.13 (m, 2H), 2.00-1.88 (m, 1H), 1.79-1.55 (m, 7H), 1.01 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ ppm: 214.3 (C), 138.9 (C), 133.6 (CH), 129.2 (2CH), 127.9 (2CH), 51.8 (CH_2), 47.4 (C), 38.9 (CH_2), 38.4 (CH_2), 30.2 (CH_2), 27.1 (CH_2), 22.3 (CH_3), 20.8 (CH_2); $[\alpha]_{\text{D}}^{24} = +2.9^\circ$ ($c = 0.01$, EtOH_{abs}); MS (APCI): m/z 281 (100%) $[\text{M} + \text{H}]^+$; Anal. Calcd for $\text{C}_{15}\text{H}_{20}\text{O}_3\text{S}$: C, 64.26; H, 7.19; O, 17.12; S, 11.44. Found: C, 64.40; H, 7.03.

General procedure for the synthesis of ketoacid 6

A mixture of adduct **6a** (452 mg, 2 mmol) and formic acid (2 mL) was stirred at 20 °C for 2 h. Formic acid was distilled, the crude was taken up in Et_2O (10 mL), washed with saturated NaHCO_3

solution (2 x 10 mL). The aqueous layer was acidified at 0 °C (6N HCl) and thoroughly extracted (4 x 10 mL Et₂O). The combined organic phase was dried (MgSO₄) and filtered (Celite[®]) and the crude concentrated in vacuo to give keto acid **6a** (302 mg, 89%) as a colorless oil. This material was used without further purification in the next step.

(1'S)-3-(1'-Methyl-2-oxocyclopentyl)-

propionic acid 6a. IR (neat, ν cm⁻¹): 3512, 3090, 2963, 2873, 2663, 1729, 1706; ¹H NMR (200 MHz, CDCl₃) δ ppm: 11.0 (bs, 1H), 1.67-1.33 (m, 4H), 1.19-0.81 (m, 6H), 0.20 (s, 3H); ¹³C NMR (50 MHz, CDCl₃) δ ppm: 222.8 (C), 178.3 (C), 47.0 (C), 36.8 (CH₂), 35.4 (CH₂), 30.6 (CH₂), 28.7 (CH₂), 20.7 (CH₃), 18.0 (CH₂); Anal. for C₉H₁₄O₃, calcd. C, 63.51; H, 8.29; found C, 63.28; H, 8.39; $[\alpha]^{25}_D$ -40.6 (c = 1.6, EtOH_{abs}).

(1'S)-3-(1'-Methyl-2-oxocyclohexyl)-

propionic acid 6b. IR (neat, ν cm⁻¹): 3502, 2935, 2866, 1700; ¹H NMR (300 MHz, CDCl₃) δ ppm: 9.4 (bs, 1H), 2.38-2.29 (m, 3H), 2.23-2.12 (m, 1H), 2.03-1.93 (m, 1H), 1.84-1.68 (m, 6H), 1.62-1.58 (m, 1H), 1.05 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ ppm: 215.5 (C), 179.3 (C), 47.7 (C), 39.0 (CH₂), 38.5 (CH₂), 32.2 (CH₂), 28.9 (CH₂), 27.3 (CH₂), 22.3 (CH₃), 20.8 (CH₂); MS (ESI) 207 (M+23) 391 (2M+23); $[\alpha]^{28}_D$ -85 (c = 0.13, EtOH_{abs}).

Chemical correlation to Methyl (1'S)-3-(1-Methyl-2-oxo-cyclopentyl)-propionate 7a: A 0.5 M solution of diazomethane in Et₂O (5 mL) was added to a solution of the acid **6** (97 mg, 0.57 mmol) in dry Et₂O (20 mL) at 0 °C. The resulting mixture was stirred at room temperature for 2 hours and the excess of diazomethane was destroyed with acetic acid. The reaction mixture was washed with brine (2x5 mL), dried (MgSO₄), and concentrated in vacuo. The residue was purified by column chromatography (cyclohexane/AcOEt, 9:1) to give methyl ester

7a as a colourless oil (100 mg, 95%); $[\alpha]^{20}_D$ -32.5 (c = 2; EtOH_{abs}); lit.¹⁰ *ent*-**12a** $[\alpha]^{20}_D$ +35.7 (c = 2, EtOH_{abs}).

3b. Enantiomeric excess determination: For ¹³C-¹H} 1D NMR experiments in polypeptidic oriented solvents, the sample preparation consisted of directly weighting 25-30 mg of solute, around 140 mg of poly- γ -benzyl-L-glutamate (PBLG, DP= 463, commercially available) and adding about 350 mg of dichloromethane into a 5 mm NMR tube. Under these conditions, the total volume of the sample is optimal compared to the length of the coil of a 5 mm diameter probe-head. Compared to previous work, we have used a larger amount of PBLG than usual (100 mg) in order to obtain a clean liquid crystalline phase. This is a consequence of the relatively low degree of polymerization (DP= 463, i.e. MW \approx 101000 g.mol⁻¹). The exact composition of each NMR sample is given in Table 3. To avoid the evaporation of dichloromethane during long NMR experimental time, we have sealed the samples. Note here that a mixture of protonated and deuterated dichloromethane was used. This solution allows to easily shim the magnet on the proton FID as well as to minimize the digitization problems associated with the dynamic range of the Analogue-to-Digital Converter (ADC) induced by the difference of ¹³C signal intensity between dichloromethane and solute. In other hands, the shape and the line width of ¹³C resonances of dichloromethane provide a serious control of the magnet stability as well as possible time-evolution of the sample homogeneity during the experiments. The sample is then centrifuged during few seconds, then inverted and centrifuged again. This process is repeated until an optically homogeneous birefringent phase is obtained.

Table 3. Composition of liquid-crystalline representative NMR samples investigated

Sample	Solute	Solute / mg ^a	Co-solvent / mg ^a	Polymer % in weight
1	(<i>rac</i>)- 6	27	75/275	27.1
2	(<i>S</i>)- 6	28	75/277	26.9
3	(<i>S</i>)- 6	28	75/277	27.1

Conditions: Polymeric solvent: PBLG; Degree of polymerization of polypeptide used: 463; Co-solvent: CH₂Cl₂ / CD₂Cl₂; *Polymer/mg:* 140; ^aThe accuracy on the weighing is 1 mg.

The NMR tube was not spun in the magnet and its temperature was regulated carefully at 299 K using the standard variable temperature control unit (BVT 3000). ¹³C spectra were recorded by adding 1000 to 3000 scans. Gaussian filtering was applied to improve the spectral separation of resonances. The area measurement was performed using a curve fitting algorithm based on complex least squares treatment of the ¹³C NMR signals with and without filtering. Note that the experiments and the area measurements were repeated several times to estimate accurately the error on the enantiomeric excess of the mixture.

3c. GC-MS mechanistic studies: An HP 5989A GC-MS system (Hewlett-Packard, Palo Alto, CA, USA) was used. The chromatographic separation was performed with an Omega delta-3 capillary column (length: 25 m; I.D. 0.20 mm; film thickness: 0.2 μm) (Macherey-Nagel, Düren, Germany). In view of comparison, samples consisting of either the asymmetric Michael addition reaction mixture [1 equiv. chiral imine (**2a**) and 1.1 equiv. *tert*-butyl acrylate (**3**)] or each of the individual components (*tert*-butyl acrylate (**3**), chiral imine (**2a**), 2-methyl cyclopentanone **1a**, 1-phenylethylamine, 2-methyl-2-propanol) were

separately irradiated at 100 W and 200 °C for 30 min in 10 mL teflon sealed glass vials and cooled to r.t. prior to GC-mass analysis. The teflon sealed glass vials filled with the reaction medium were maintained at 70°C during 15 minutes prior analysis and immediately processed. The head space (5 μL) was sampled using an airtight syringe and injected in splitless mode. Helium pressure was 50 kPa. The injector temperature was 250 °C and the initial oven temperature was 35 °C. This temperature was maintained for 1 min, the temperature was then programmed as follows: 4 °C/min up to 50 °C then 6 °C/min up to 100 °C followed by a 5 min hold. The transfer line temperature was set to 280 °C. Analysis was performed by electronic impact ionisation. The ion source and quadrupole temperature was set to 200 °C and 100 °C respectively. The electron energy was 70 eV. Acquisition was performed in scan mode over the range of 20 to 150 at a scan rate of 0.9 scan/sec (4 samplings per scan). Analysis of the results obtained for the head space of the asymmetric Michael reaction showed that the peak observed at 1.605 min correspond to 2-methyl-2-butene (MW = 56 g.mol⁻¹, identical fragmentation and comparable abundances)¹⁷ A similar peak was not detected from the head space of the other samples, except for the *tert*-butyl acrylate (MW = 128 g.mol⁻¹) one. However, ions at m/z 55, 57 and 59 are not present in the same ratio for the 2-methyl-2-butene mass spectrum. Moreover, analysis of the *tert*-butanol (MW = 74 g.mol⁻¹) sample indicates that in the reaction conditions, *tert* butanol did not led to 2-methyl-2-butene. This set of results gave evidence that 2-methyl-2-butene and not *tert*-butanol was released during the lactamization process of Michael adduct (**5**) (see article, Scheme 3).

3d. Theoretical calculations: Geometries for the reactants were optimized by means of

gradient technique at RHF AM1 level²⁰ by using the semi-empirical molecular orbital program MOPAC.²¹ All the RHF AM1 transition structures were located using the procedures implemented in MOPAC (Version 5.0). All variables were optimized by minimizing the sum of the squared scalar gradients (NLLSQ and

SIGMA).^{22,23} Force calculations were carried out to ensure that the transition structures located had one imaginary frequency. Final values of the gradient norms were <1 kcal/Å and each transition structure had one negative eigenvalue in the Hessian matrix as required.

4. Conclusions

In conclusion, we have demonstrated that the reaction of hindered *tert*-butyl acrylate **2** in the AMACI was efficiently promoted under μ W activation, compared to conventional heating. As expected, regardless the activation mode, the control of the stereochemistry of the newly created quaternary carbon center in such Michael adducts is always dictated by the configuration of the chiral inductor. The stereoselectivity of the process was determined using natural abundance ¹³C-¹H NMR in a chiral polypeptide liquid crystal. Moreover, the temperature profiles achieved under microwave irradiation are not accessible in conventional heating and can allow a differentiation in the reaction pathways. A direct and stereoselective access to lactams **8** was thus achieved only under μ W, although in moderate yield. A highly stereoselective process (ee > 95 %) was obtained either at 100 °C for 4 h (Δ) or for 30 min (μ W, 100 W). A good correlation was obtained between experiment and theoretical calculations. Both the more polar and asynchronous transition state led to the expected Michael adduct (*S*)-**4**, and are favored under μ W activation, allowing the reaction to proceed efficiently in minutes. Finally, Michael adducts from methyl acrylate, benzyl acrylate, vinylsulfone and acrylonitrile are regio- and stereoselectively obtained in high yield and short time using the microwave process.

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Author Contributions

Françoise Dumas ensure the conception and design of the chemistry and Philippe Lesot the the enantiomeric purity determination using natural abundance ¹³C-¹H NMR. Lucie Vandromme, Li Chen and Lai Wei performed the chemical experiments and analyzed the data, Franck Le Bideau, and Françoise Dumas wrote the manuscript, André Loupy helped and advised us for the microwave chemistry, Olivier Lafon and Philippe Lesot were in charge of the NMR determination of selectivity in chiral liquid phase, Elise Tran Huu Dau performed the theoretical calculations and Pierre Chaminade the CPV analysis.

Conflicts of Interest

The authors declare no conflict of interest.

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