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# Self-assembly of PEG-*b*-PTMC copolymers: micelles and polymersomes size control

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 [Supporting Information](#)

## ABSTRACT

Poly(ethylene glycol)<sub>45</sub>-*b*-poly(trimethylene carbonate)<sub>n</sub> PEG<sub>45</sub>-*b*-PTMC<sub>n</sub> diblock copolymers were synthesized with five different PTMC degrees of polymerization ( $n = 38, 96, 144, 170, 332$ ) and their self-assembly properties in water were studied using a manual nanoprecipitation procedure. We confirmed that the copolymer's hydrophilic weight fraction ( $f_{\text{PEG}}$ ) is controlling nanoparticles morphology. We determined that the PEG<sub>45</sub>-*b*-PTMC<sub>96</sub> with  $f_{\text{PEG}} \approx 17\%$  is the optimal hydrophilic fraction for the stabilization of well-defined unilamellar vesicles with a membrane thickness of  $\delta \approx 14.6$  nm. Maintaining this fraction constant and modulating the overall molar mass of the block copolymers allowed the establishment of a power law of  $\delta \propto \overline{M}_n^{0.56}$  which provides a robust correlation between the molar mass of PTMC and vesicles' membrane thickness. Finally, we proved that controlling nanoprecipitation's conditions by microfluidics allowed fine-tuning and control of the nanoparticles size and polydispersity index while maintaining their shape with a perfect batch-to-batch reproducibility.

## INTRODUCTION

In recent years, polymeric nanoparticles have been widely investigated as drug delivery systems with some currently being under clinical trials.<sup>1-4</sup> Among the different nanocarriers being developed, a significant part are resulting from the self-assembly of amphiphilic block copolymers in aqueous solution in different morphologies such as micelles, vesicles or rods.<sup>5-7</sup>

Polymeric vesicles – also called polymersomes (Ps) – are of great interest for imaging purposes and as drug delivery systems in various types of therapy, the most explored being cancer treatment.<sup>8-12</sup>

Compared to their low molar mass lipidic analogs,<sup>13</sup> polymersomes have a thicker hydrophobic membrane, providing significant storage capacity for water-insoluble substances. They are also enclosing a large hydrophilic reservoir which can be loaded with water-soluble substances. Their surface can be (multi)functionalized with imaging probes and/or targeting moieties. Their membrane thickness dictates several physico-chemical properties such as colloidal stability, fluidity and permeability which can be easily modulated by tuning the hydrophobic block molar mass.<sup>14-18</sup> Membrane permeability can also be tuned by changing the nature of the hydrophobic block or by chemical post-polymerization modifications.<sup>19-21</sup>

The formation of a majority of vesicles over other morphologies can be predicted by the use of a copolymer with a specific hydrophilic weight fraction ( $f$ ).<sup>5</sup> However, this tool can only be employed for objects obtained at or close to thermodynamic equilibrium. Indeed, it was shown that the same amphiphilic copolymer can form a variety of kinetically-trapped morphologies according to the formulation process.<sup>5,16,22</sup> Generally speaking, one can anticipate that when increasing  $f$ , self-assembled morphologies pass from vesicles to rods to micelles.<sup>23</sup>

Ps can be obtained in different sizes through different formulation techniques such as film hydration or solvent-displacement method (also called nanoprecipitation).<sup>22,24</sup> Microfluidic systems are particularly interesting for the formation of monodisperse, size-controlled Ps since they allow the mixture between solutions to be controlled and easily tuned in a repeatable manner (e.g. flow ratios between solutions and overall total flow rate).<sup>22,25–28</sup> This permits reproducibility which is absolutely necessary for the development of Ps as drug delivery systems.

The copolymer's chemical nature is also important in order to access Ps with specific properties such as biodegradability or stimuli-responsiveness.<sup>29–32</sup> Poly(ethylene glycol) (PEG) is often used as a hydrophilic block since it was found to delay Ps clearance from the blood.<sup>33,34</sup> Biodegradable polymers, especially aliphatic polyesters and polycarbonates, such as poly(lactic acid) (PLA), polycaprolactone (PCL) and poly(trimethylene carbonate) (PTMC), are well studied in pharmaceuticals due to their low systemic toxicity.<sup>35</sup> For instance, after studying PEG-*b*-PLA and PEG-*b*-PCL self-assembly and stability, Discher *et al.* showed they could form polymersomes which biodegrade upon hydrolysis.<sup>36</sup> Recently, PTMC-based materials were investigated since PTMC has a unique degradation which is mostly sensitive to enzymatic hydrolysis with a surface erosion mechanism generating non-acidic products.<sup>37,38</sup> Our group demonstrated the stability in aqueous conditions and enzymatic degradation of vesicles made from PGA-*b*-PTMC copolymers presenting an anionic poly(L-glutamic acid) (PGA) as the hydrophilic block.<sup>22</sup> In addition, we determined that the PTMC semi-crystalline nature impacted the PTMC conformation in the membrane.<sup>39</sup>

Because PEG is routinely employed as a hydrophilic block for its stealthiness property, the self-assembly of PEG-*b*-PTMC copolymers was also studied. Van Hest and co-workers reported the formation of PEG-*b*-PTMC multilamellar vectors<sup>40</sup> or micelles<sup>41</sup>, while Fang and co-workers.<sup>42</sup> formed spherical nanoparticles without elucidating their shape in deep details.

To our knowledge, only two articles showed the possible formation of PEG-*b*-PTMC vesicles: Feijen's team mentioned that a PEG<sub>5.8k</sub>-*b*-PTMC<sub>24k</sub> ( $f_{\text{PEG}} = 19.5\%$ ) could form polymersomes<sup>43</sup> and Zhong and co-workers confirmed that this hydrophilic fraction was suitable to preferentially form vesicles with -COOH or -NH<sub>2</sub> terminated PEG<sub>5k</sub>-*b*-PTMC<sub>20.3k</sub> ( $f_{\text{PEG}} = 19.8\%$ ).<sup>44</sup> This value of  $f_{\text{PEG}}$  was not following the empirical rule of Discher & Eisenberg of  $25\% < f_{\text{PEG}} < 45\%$  that were established for polydiene based copolymers.<sup>5</sup> This is not surprising since diblock copolymers made of PEG and polar polymers such as PLA or PCL were shown to form vesicles for  $f_{\text{PEG}} < 20\%$ .<sup>23</sup>

The purpose of our study was to obtain monodispersed and size-controlled PEG-*b*-PTMC polymersomes with different PTMC membrane thicknesses while keeping the length of PEG constant at 2000 g/mol, this molar mass being often reported to provide interesting stealth behavior.<sup>34</sup> To reach this goal, we first synthesized and characterized a series of amphiphilic block copolymers, PEG<sub>45</sub>-*b*-PTMC<sub>*n*</sub>, with different PTMC block length. We then studied the influence of the copolymer's hydrophilic weight fraction ( $f_{\text{PEG}}$ ) on nanoparticles morphology obtained upon self-assembly in water by manual nanoprecipitation. Finally, we studied the impact of nanoprecipitation conditions by microfluidics on nanoparticles' size and polydispersity index.

## EXPERIMENTAL SECTION

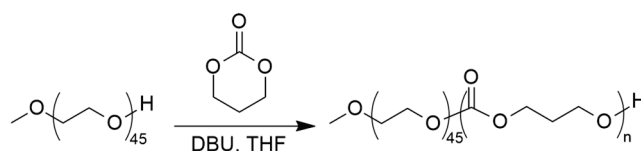
### Materials

Solvents: tetrahydrofuran (THF), dimethyl sulfoxide (DMSO), ethyl acetate, toluene, acetic acid, methanol) were purchased from Sigma-Aldrich. THF was dried by a solvent purification system. Trimethylene carbonate (1,3-dioxane-2-one; TMC) was purchased from TCI Europe and purified by three successive recrystallizations in dry ethyl acetate. Poly(ethylene glycol)methyl ether (MeO-PEG<sub>45</sub>-OH;  $\overline{M}_n = 2000$  g/mol) was purchased from Iris Biotech

GMBH and dried by azeotropic distillation in toluene. DBU (1,8-diazabicyclo[5.4.0]undec-7-ene) was purchased from Sigma-Aldrich and distilled before use.

### Synthesis of PEG-*b*-PTMC block copolymers

The synthesis of the PEG<sub>45</sub>-*b*-PTMC<sub>*n*</sub> block copolymers was adapted from a previously described method of ring-opening polymerization (ROP) of TMC using MeO-PEG-OH as a macro-initiator (Scheme 1).<sup>45</sup> All the compounds and solvents must be anhydrous or distilled to avoid secondary reactions with water. All polymerization reactions were performed in flame-dried glassware employing standard high vacuum techniques and carried out in a glovebox under inert atmosphere (argon, [O<sub>2</sub>] < 0.5 ppm, [H<sub>2</sub>O] < 5 ppm).



**Scheme 1.** Reaction scheme of the synthesis of PEG<sub>45</sub>-*b*-PTMC<sub>*n*</sub> by ring-opening polymerization (ROP) of TMC on MeO-PEG<sub>45</sub>-OH in THF and initiated by DBU.

The procedure described below was used for a targeted PTMC degree of polymerization (DP) of 40 and adapted for DPs 120, 190 and 400. In a glovebox, dry THF (2 mL) was added in a Schlenk containing dried MeO-PEG<sub>45</sub>-OH (2.2 g, 1.1 mmol) and let to stir until total dissolution. Separately, TMC (previously recrystallized, 4 g, 39 mmol) was dissolved in dry THF (6 mL). The TMC solution was added to the PEG solution. DBU (previously distilled, 161  $\mu$ L, 1.1 mmol) was introduced and the mixture was left to stir at room temperature. The reaction progress was checked by <sup>1</sup>H NMR of the reaction mixture. When the PTMC DP was reached (95% of conversion), the reaction was quenched with an excess of acetic acid. The resulting copolymer was precipitated in cold methanol, centrifuged to remove the supernatant,

washed 3 more times with cold methanol and dried under vacuum overnight. The pure copolymer was obtained as a white powder.

$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ,  $\delta$  ppm): 4.29 (2H,  $-\text{PTMC}_{n-1}-\text{CO}-\text{O}-\text{CH}_2-\text{CH}_2-\text{CH}_2-\text{OH}$ ); 4.23 (t,  $J = 8.34$  Hz,  $4*(n-1)\text{H}$ ,  $-(\text{CO}-\text{O}-\text{CH}_2-\text{CH}_2-\text{CH}_2-\text{O})_{n-1}-$ ); 3.73 (2H,  $-\text{PTMC}_{n-1}-\text{CO}-\text{O}-\text{CH}_2-\text{CH}_2-\text{CH}_2-\text{OH}$ ); 3.63 (s,  $(4*45)\text{H}$ ,  $-(\text{CH}_2-\text{CH}_2-\text{O})_{45}-$ ); 3.37 (s, 3H,  $\text{H}_3\text{C}-\text{O}-\text{PEG}-$ ); 2.04 (quint,  $J = 8.34$  Hz,  $2*(n-1)\text{H}$ ,  $-(\text{CO}-\text{O}-\text{CH}_2-\text{CH}_2-\text{CH}_2-\text{O})_{n-1}-$ ); 1.91 (quint,  $J = 8.12$  Hz, 2H,  $-\text{PTMC}_{n-1}-\text{CO}-\text{O}-\text{CH}_2-\text{CH}_2-\text{CH}_2-\text{OH}$ ).

### Formulation of nanoparticles

**Manual nanoprecipitation (mNP).** Ultrapure water and copolymer solution in DMSO (10 g/L) were respectively filtered off with cellulose acetate 0.22  $\mu\text{m}$  and polytetrafluoroethylene (PTFE) 0.45  $\mu\text{m}$  syringe filters. Filtered water (2.7 mL) was quickly added in the filtered copolymer solution under stirring (400 rpm, 25  $^\circ\text{C}$ , 300  $\mu\text{L}$  of a 10 g/L solution) to obtain a suspension at 1 g/L of copolymer and 90 vol% of water. To remove the organic solvent, the suspension was dialyzed against deionized water using a 3.5 kD cut-off dialysis membrane (Spectra/Por<sup>®</sup> RC, 5 L of water, 2 changes). With the  $\text{PEG}_{45}-b\text{-PTMC}_{96}$  copolymer, the reverse addition (mNP-ra) was also performed at 90 vol% of water by quickly adding under stirring the filtered copolymer solution (400 rpm, 25  $^\circ\text{C}$ , 300  $\mu\text{L}$  of a 10 g/L solution) into filtered ultrapure water (2.7 mL).

**Controlled nanoprecipitation using a microfluidic system (cNP).** The Dolomite Microfluidics<sup>®</sup> system used was constituted of: two pressure pumps (3200175), two flow rate sensors (3200097) and a micromixer chip (3200401) all connected with FEP tubing (1/16" x 0.25 mm, 3200063). Under nitrogen pressure, flow rates were controlled by the Mitos Flow Control Center 2.5.17 software. A scheme of the set-up is available in Figure S7. Ultrapure water and copolymer solution in DMSO (1 g/L) were respectively filtered off with cellulose acetate 0.22  $\mu\text{m}$  and PTFE 0.45  $\mu\text{m}$  syringe filters. One pump was filled with the filtered



copolymer solution in DMSO (P-DMSO) and connected to the chip through the first and third inputs using a T-connector. The other pump was filled with filtered ultrapure water and directly linked to the chip through the second input. A camera was used to verify that the chip was dust free and air bubbles free. Flow rates were set at 233  $\mu\text{L}/\text{min}$  of P-DMSO for 100  $\mu\text{L}/\text{min}$  of  $\text{H}_2\text{O}$ . Suspension (333  $\mu\text{L}/\text{min}$  total flow rate; 30 vol% water content) was collected into a glass vial from the chip's output after 12 mixing stages. The organic solvent was removed by dialysis against deionized water using a 3.5 kD cut-off dialysis membrane (Spectra/Por<sup>®</sup> RC, 5 L of water, 2 changes). With the PEG<sub>45</sub>-*b*-PTMC<sub>96</sub> copolymer, different conditions were tested to apprehend the system (Figure S8 and Table S3). All suspensions were prepared at room temperature.

### Characterization techniques

**NMR.** Nuclear magnetic resonance (NMR) experiments were performed at 298 K on a Bruker Avance I NMR spectrometer operating at 400 MHz, using TMS as standard, and equipped with a Bruker multinuclear z gradient direct probe head capable of producing gradients in the z direction with 53.5  $\text{G}\cdot\text{cm}^{-1}$  strength. <sup>1</sup>H NMR spectra were recorded with a D1 of 5 s and 32 scans. To describe the multiplicities of the signals, the following abbreviations were used: s: singlet, t: triplet and quint: quintuplet. The coupling constants, abbreviated J, were expressed in Hz.

**SEC.** Size exclusion chromatography (SEC) analyses were performed on a Varian apparatus equipped with a refractive index detector and a three-column set of TSK gel TOSOH (G2000HXL, G3000HXL, G4000HXL with pore sizes respectively of 20, 75, and 200 Å). Analyses were carried out on polymer samples (10 mg/mL) at 40 °C using THF as eluent (1 mL/min) and trichlorobenzene as a flow marker. Values of the refractive index increment ( $\text{dn}/\text{dc}$ ) of each copolymer were determined by measuring the differential refractive index between copolymer solutions at different concentrations (1, 5, 10 and 15 g/L) and pure

THF using the refractive index detector of the Varian apparatus. The  $dn/dc$  corresponds to the slope of the linear regression of the differential refractive index values against the copolymer concentration. The  $dn/dc$  values were then used to determine the true copolymer's average molar mass ( $\overline{M}_n$ ) and dispersity ( $\mathcal{D}$ ).

**TGA.** Thermogravimetric analyses (TGA) were performed on a TA Instruments Q500 under a continuous nitrogen flow using approximatively 8 mg of samples in a platinum pan. The heating run was recorded at 10 °C/min from room temperature to 500 °C.

**DSC.** Differential scanning calorimetry (DSC) analyses were carried out on a TA Instruments Q100 under a continuous flow of nitrogen and helium (25 mL/min), using approximatively 5 mg of samples in sealed aluminum pans. Thermograms of three successive runs were recorded: (i) first heating run from –60 to 120 °C at 10 °C/min; (ii) cooling run from 120 to –60 °C at 10 °C/min; (iii) second heating run from –60 to 80 °C at 10 °C/min. As the crystallization process of the copolymers was slow, the melting temperature ( $T_m$ ) and the glass transition temperature ( $T_g$ ) were measured only on the first heating run.

**DLS 90°.** Dynamic light scattering measurements (DLS 90°) were carried out on a Malvern Zetasizer Nano ZS equipped with a He-Ne laser (633 nm), at 25 °C and a scattering angle of 90°. Values of viscosity were corrected according to the mixture of DMSO/H<sub>2</sub>O used. Hydrodynamic radius ( $R_h$ ) of the nanoparticles and polydispersity index (PDI) were calculated from autocorrelation functions using the cumulant method.

**MALS.** Multi-angle dynamic and static light scattering (MALS) measurements were performed using an ALV-5000 goniometer with a He-Ne laser (633 nm) and an ALV-5000/EPP multiple tau digital correlator (initial sampling time of 125 ns). Suspensions at 0.1 g/L of copolymer were placed in 10 mm outer diameter quartz tubes. Temperature of the toluene bath was set at 25 °C. Data were acquired on the ALV correlator software at several angles from 30° to 150° by step of 10° (counting time of 10 s). The hydrodynamic radii ( $R_h$ )

and PDI were determined using the cumulant method. The radii of gyration ( $R_g$ ) were calculated using Guinier's approximation.

**Cryo-TEM.** Cryo-transmission electron microscopy (cryo-TEM) images were recorded at the Institut de Minéralogie, de Physique des Matériaux et de Cosmochimie (Paris) on a LaB6 JEOL 2100 (JEOL, Japan) working at 200 kV. Samples were prepared by placing a drop of a suspension of nanoparticles onto a holey carbon grid (Quantifoil® Micro Tools GmbH, Germany). Excess of solution was blotted by a filter paper. The grid was rapidly plunged into liquid ethane, placed onto a Gatan 626 cryo-holder and transferred into the microscope. The grids were observed at low temperature ( $-180\text{ }^{\circ}\text{C}$ ) and under low dose conditions (JEOL minimum dose system). Pictures were taken with an Ultrascan 2k CDD camera (Gatan, USA) and analyzed with the ImageJ software.

**SANS.** Small angle neutron scattering experiments (SANS) were performed at the Laboratoire Léon Brillouin (CEA-CNRS, Saclay) on the PACE and PA20 spectrometers. Four configurations were chosen by varying the wavelength  $\lambda$  and the sample-to-detector distance  $D$ , to cover a large range of scattering vectors  $q$  ( $q = \frac{4\pi}{\lambda} \sin(\frac{\theta}{2})$ ) from  $1.1 \times 10^{-3}$  to  $1.1 \times 10^{-1} \text{ \AA}^{-1}$ : i)  $\lambda = 6 \text{ \AA}$ ,  $D = 3 \text{ m}$ ; ii)  $\lambda = 6 \text{ \AA}$ ,  $D = 5 \text{ m}$ ; iii)  $\lambda = 13 \text{ \AA}$ ,  $D = 5 \text{ m}$  (on PACE) and iv)  $\lambda = 13 \text{ \AA}$ ,  $D = 16 \text{ m}$  (on PA20). Analyses were done on samples in  $\text{D}_2\text{O}$  at approximately 1 g/L of copolymer.

Samples were first prepared in  $\text{H}_2\text{O}$  and the change of solvent to  $\text{D}_2\text{O}$  was done using an ultrafiltration system (10 mL Amicon Millipore stirred cell, Ultracel® disc of 100 kD cut-off). Suspensions in  $\text{H}_2\text{O}$  were diluted 5 times with  $\text{D}_2\text{O}$  (Euriso-Top®), concentrated, washed a second time and concentrated again.

Samples were placed into Thuet® quartz cells of 5 mm optical path length. Absolute values of the scattering intensity,  $I(q)$ , (in  $\text{cm}^{-1}$ ) were obtained from the direct determination of the incident beam flux.<sup>46</sup>

Scattering intensity from  $N$  identical particles of spherical symmetry with a volume  $V_p$  placed in a total volume  $V$  can be expressed as follows:

$$I(q) = \Phi(\Delta\rho)^2 V_p P(q) S(q) \quad (1)$$

where  $P(q)$  is the form factor of the particles,  $S(q)$  the structure factor,  $\Phi = \frac{NV_p}{V}$  the volume fraction and  $(\Delta\rho)^2$  is the contrast, with  $\Delta\rho$  the difference of scattering length densities between particle and solvent. Assuming that PEG chains were highly hydrated by D<sub>2</sub>O, we considered  $\Delta\rho$  only between PTMC and D<sub>2</sub>O ( $\Delta\rho = 4.70 * 10^{-6} \text{ \AA}^{-2}$ ).

Scattering curves were analyzed by fitting them with theoretical curves using SasView software (<http://www.sasview.org>), considering the background, volume fraction, scattering length densities of polymer and solvent, object polydispersity and measurement uncertainties ( $dq$  and  $dI$ ).

Different form factors were used to fit the data: sphere, vesicle and vesicular aggregate.

**Sphere model** is the historical model used by Guinier where  $R$  is the sphere radius.

$$P(q) = \left[ 3 \frac{\sin(qR) - qR \cos(qR)}{(qR)^3} \right]^2 \quad (2)$$

**Vesicle model** is a shell model where particles are defined by spherical shells of PTMC:

$$P(q) = \frac{1}{(V_e - V_c)^2} \left[ 3V_e \frac{\sin(qR_e) - qR_e \cos(qR_e)}{(qR_e)^3} - 3V_c \frac{\sin(qR_c) - qR_c \cos(qR_c)}{(qR_c)^3} \right]^2 \quad (3)$$

where  $(R_e, R_c)$  and  $(V_e, V_c)$  are respectively the external and core radii and volumes and  $\delta = R_e - R_c$  the hydrophobic membrane thickness.

**Vesicular aggregate model** is a fractal aggregate model of core-shell particles.<sup>47</sup> Here, for:

$$I(q) = \phi P(q) S(q) \quad (4)$$

$$P(q) = \frac{1}{V_e} \left[ 3V_c(\rho_c - \rho_s) \frac{\sin(qR_c) - qR_c \cos(qR_c)}{(qR_c)^3} + 3V_e(\rho_s - \rho_{D_2O}) \frac{\sin(qR_e) - qR_e \cos(qR_e)}{(qR_e)^3} \right]^2 \quad (5)$$

and:

$$S(q) = 1 + \frac{D_f \Gamma(D_f - 1)}{[1 + 1/(q\xi)^2]^{(D_f-1)/2}} \sin[(D_f - 1) \tan^{-1}(q\xi)] \quad (6)$$

where  $(R_e, R_c, R_s)$ ,  $(V_e, V_c, V_s)$  ( $\rho_e, \rho_c, \rho_s$ ) are respectively the radii, the volumes and scattering length densities of the whole particle, core and shell.  $D_f$  is the fractal dimension and  $\xi$  the correlation length of the fractal aggregates.

**For all models**, the radius polydispersity  $\sigma_R$  was estimated as a standard deviation of a lognormal distribution whereas the thickness polydispersity  $\sigma_\delta$  was done on a gaussian distribution. Standard deviations ( $SD_R$ ,  $SD_\delta$ ) were calculated for the radius and the thickness by multiplying the respective polydispersity ( $\sigma_R$ ,  $\sigma_\delta$ ) with either the value of the radius ( $R$ ) or of the thickness ( $\delta$ ).

## RESULTS AND DISCUSSION

### Synthesis of copolymers and characterization

A series of poly(ethylene glycol)-*b*-poly(trimethylene carbonate) PEG<sub>45</sub>-*b*-PTMC<sub>n</sub> block copolymers was obtained by ring-opening polymerization (ROP) using MeO-PEG<sub>45</sub>-OH ( $\overline{M}_n$  = 2000 g/mol) as macro-initiator in the presence of DBU as catalyst (Scheme 1).

The DP ( $n$  value) of the PTMC block was easily calculated by proton NMR in CDCl<sub>3</sub> from the integration of the resonance peaks at 2.04 and 4.23 ppm corresponding to PTMC protons. The calibration was performed based on the PEG block with peaks at 3.63 ppm (184H) and 3.37 ppm (3H) (Figure S1). The DP of PTMC was also determined by steric exclusion chromatography (SEC) in THF using each copolymer's actual refractive index increment (dn/dc). Both methods provided consistent and similar results.

Five block copolymers PEG<sub>45</sub>-*b*-PTMC<sub>n</sub> with  $n$  = 38, 96, 144, 177 and 332 were synthesized with a hydrophilic weight fraction  $f_{\text{PEG}}$  ranging from 34% to 6%. Their

characteristics found by NMR, SEC and differential scanning calorimetry (DSC) are summarized in Table 1.

SEC analyses in THF revealed a narrow molar mass distribution of the copolymers (dispersity  $\bar{D}$  ranging from 1.05 to 1.17) (Figure S2). Thermogravimetric analyses (TGA) provided information on the copolymers degradation and purity (Figure S3). The first block to be degraded was the PTMC around 200 °C followed by the one of PEG around 320 °C. The larger the PTMC block, the higher its degradation temperature. There were no carbon residues at 500 °C which corroborates the excellent purity of the copolymers evidenced by NMR and SEC.

**Table 1.** Characteristics of PEG<sub>45</sub>-*b*-PTMC<sub>n</sub> block copolymers obtained by <sup>1</sup>H NMR, SEC and DSC.

Block copolymer	n, DP (PTMC)		$\bar{M}_n$ (PTMC <sub>n</sub> )		$\bar{M}_n$ (PEG <sub>45</sub> - <i>b</i> -PTMC <sub>n</sub> ) <sup>a</sup>	$f_{\text{PEG}}$ <sup>a</sup>	$\bar{D}$ <sup>a</sup>	$T_g$ <sup>b</sup>	$T_m$ <sup>b</sup>
	<sup>1</sup> H NMR	SEC <sup>a</sup>	<sup>1</sup> H NMR	SEC <sup>a</sup>					
			(g/mol)	(g/mol)	(g/mol)	(%)	(°C)	(°C)	
PEG <sub>45</sub> - <i>b</i> -PTMC <sub>38</sub>	38	38	3900	3900	5900	34	1.05	−40	41
PEG <sub>45</sub> - <i>b</i> -PTMC <sub>96</sub>	90	96	9200	9800	11800	17	1.05	−32	39
PEG <sub>45</sub> - <i>b</i> -PTMC <sub>144</sub>	144	144	14700	14700	16700	12	1.06	−27	40
PEG <sub>45</sub> - <i>b</i> -PTMC <sub>170</sub>	167	170	17000	17400	19400	10	1.11	−26	40
PEG <sub>45</sub> - <i>b</i> -PTMC <sub>332</sub>	332	332	33900	33900	35900	6	1.17	−21	42

<sup>a</sup> Results from SEC in THF ( $\bar{D}$ : dispersity)

<sup>b</sup> Results from DSC

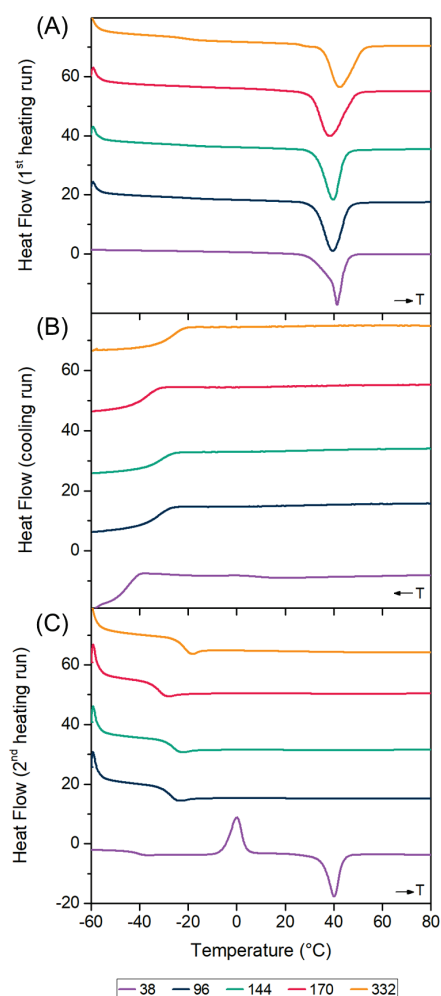
n: degree of polymerization (DP) of PTMC

$f_{\text{PEG}}$ : ratio of the PEG molar mass on the copolymer molar mass

$T_g$ : glass transition temperature

$T_m$ : melting temperature (DSC)

PEG<sub>45</sub>-*b*-PTMC<sub>n</sub> copolymers were also analyzed by DSC between −60 °C and 120 °C (Figure 1). During the analysis, copolymers underwent a first heating run (Figure 1A), a cooling run (Figure 1B) and a second heating run (Figure 1C). The five copolymers had a glass transition temperature  $T_g$  (Figure 1C) between −40 °C and −21 °C (the  $T_g$  increased when the DP of PTMC increased), as well as a melting temperature  $T_m$  around 40 °C (Figure 1A, endothermic peak) which reflects the semi-crystalline nature of the PEG<sub>45</sub>-*b*-PTMC<sub>n</sub>. However, there was no exothermic peak corresponding to the copolymer recrystallization during the cooling run (Figure 1B), and therefore no endothermic peak during the second heating run, except for PEG<sub>45</sub>-*b*-PTMC<sub>38</sub>, with the smallest DP of PTMC which indicates a probable very slow recrystallization kinetics. The thermal behavior of PEG<sub>45</sub>-*b*-PTMC<sub>38</sub> is actually different from the other copolymers, possibly because the PTMC is the shortest and its ratio in the copolymer is the lowest, both favoring its crystallization. Indeed, during the cooling run, the copolymer began to recrystallize around 0 °C (Figure 1B) and continued during the second heating run (Figure 1C). Recrystallization kinetics at 21 °C were performed by DSC analyses for PEG<sub>45</sub>-*b*-PTMC<sub>n</sub>  $n = 96, 144, 170$  and  $332$  in order to estimate the required time for the beginning of recrystallization (Figure S4 and Table S1). Results showed that the higher the ratio of PTMC in the copolymer, the slower the recrystallization (between a few minutes to several weeks), in good agreement with the work of Zhu *et al.*<sup>48</sup> This parameter could be important to consider concerning the self-assembly properties of the copolymers, especially when using a direct dissolution method. Oppositely, if nanoprecipitation is used, the process is so fast that this phenomenon is negligible, especially when small nanoparticles are obtained, leading to a strong confinement of PTMC chains.<sup>39</sup>



**Figure 1.** DSC thermograms of PEG<sub>45</sub>-b-PTMC<sub>n</sub> diblock copolymers ( $n = 38$  (purple), 96 (blue), 144 (green), 170 (red), 332 (orange)). (A) First heating run; (B) Cooling run; (C) Second heating run.



## **Self-assembly**

Nanoparticles were prepared by self-assembly of the different PEG<sub>45</sub>-*b*-PTMC<sub>n</sub> diblock copolymers. In order to evaluate the influence of the copolymer's hydrophilic weight fraction on objects' size and morphology, the same formulation technique was applied to all copolymers.

## **Direct dissolution**

We firstly investigated the direct dissolution method. Copolymers dissolution in water was performed for PEG<sub>45</sub>-*b*-PTMC<sub>96</sub> and PEG<sub>45</sub>-*b*-PTMC<sub>144</sub> at two different temperatures: below (25 °C) and above (40 °C) the copolymers' melting temperature. The objects hydrodynamic diameter ( $D_h$ ) and polydispersity index (PDI) were followed in time by dynamic light scattering (DLS) at 90° (Figure S5). It took at least 1 week for the different systems to reach a state close to equilibrium, meaning to form only one population of sub-micron particles. In each case, the higher temperature (40 °C) allowed objects to be smaller. However, the impact of temperature on the polydispersity indexes (PDI) remains uncertain. Particles of PEG<sub>45</sub>-*b*-PTMC<sub>96</sub> had a smaller PDI when formed at 25 °C rather than 40 °C, while the opposite situation was observed with PEG<sub>45</sub>-*b*-PTMC<sub>144</sub> particles. Overall, the PDI improved with time but were still high (> 0.2 after 37 days of dissolution). Therefore, we decided to use another method to form monodisperse nanoparticles, namely a solvent-displacement method, also referred to as nanoprecipitation.

## **Nanoprecipitation**

### **Influence of the hydrophilic fraction**

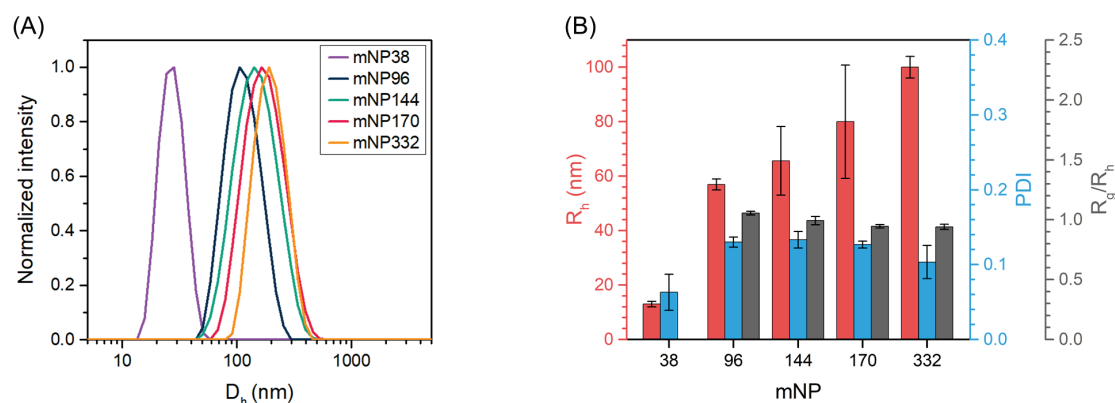
We studied the self-assembly of the whole series of block copolymers by manual nanoprecipitation (mNP). In this process, water was manually added to each copolymer

solution in DMSO to trigger the self-assembly and the organic solvent was then removed by dialysis against water.

mNP samples were produced in triplicate and each sample was analyzed by DLS at 90° and both static and dynamic multi-angle light scattering (MALS). Size distributions obtained by DLS 90° proved that NPs were spherical and monodisperse. Indeed, examples of DLS 90° curves for one sample per type of NP (Figure 2A) showed narrow log-normal distributions without aggregation. Radius of gyration ( $R_g$ ) was estimated by plotting the scattered intensity from SLS measurement with a Guinier plot while hydrodynamic radius ( $R_h$ ) was calculated from DLS measurement using the Stokes-Einstein equation once the diffusion coefficient was determined using Fick's law of diffusion (Figure S6 and Table S2). All the NPs were found to have a hydrodynamic radius below 100 nm with a low dispersity ( $PDI < 0.15$ ) (Figure 2B). It could seem that the longer the PTMC in the copolymer, the larger the NPs. However, the manual nanoprecipitation procedure did not allow a perfect reproducibility of the size of the NPs (see standard deviations (SD) in Table S2). Repeatability was proven for mNP38, 96 and 332 since their size' SD was lower than 4 nm and their PDI' SD lower than 0.02. It is thus possible to compare their sizes accurately and draw some conclusions: mNP38 were the smallest ( $R_h = 13 \pm 1$  nm), mNP332 were the largest ( $R_h = 100 \pm 4$  nm) and mNP96 were in between ( $R_h = 57 \pm 2$  nm). However, repeatability was not achieved for mNP144 and 170 with respective size' SD of 13 and 21 nm meaning that their sizes could vary more than 20% of the mean value, even if at this stage of the characterization, it was not possible to explain the lack of repeatability of the manual nanoprecipitation only for these two copolymers.

The  $R_g/R_h$  ratio was also calculated since it is representative of the objects morphology.<sup>49</sup> Indeed, a vesicular shape should give  $R_g/R_h \approx 1$  whereas  $R_g/R_h \approx 0.67$  is expected for micellar structures.

The PEG<sub>45</sub>-*b*-PTMC<sub>38</sub> nanoparticles (mNP38) were the smallest, with  $R_h = 13 \pm 1$  nm, which was too low to be able to measure  $R_g$  by static MALS. Even if we were not able to obtain the mNP38  $R_g/R_h$  ratio, the NPs' size strongly suggested spherical micelle morphology. Ratios  $R_g/R_h$  of mNP96, 144, 170 and 332 were all found to be around 1, meaning that the objects were probably vesicles.



**Figure 2.** A) Example of size distributions obtained by DLS 90° on the different types of nanoparticles produced by manual nanoprecipitation (mNP) using the series of PEG<sub>45</sub>-*b*-PTMC<sub>n</sub> diblock copolymers ( $n = 38$  (purple), 96 (blue), 144 (green), 170 (red), 332 (orange)). B) Average MALS data obtained for each sample of NPs produced in triplicate by mNP:  $R_h$  (red), PDI (blue) and  $R_g/R_h$  ratio (grey).  $R_g$  for mNP38 could not be calculated (see example of Guinier plot in Figure S6A). Details of the data obtained for each sample in Table S2.

To further characterize the objects, cryo-transmission electron microscopy (cryo-TEM) experiments were performed for each type of nanoparticles obtained from mNP (Figure 3A). Different morphologies were observed: micelles for mNP38, unilamellar vesicles for mNP96, vesicular aggregates for mNP144 and 170 and a mixture of vesicular aggregates and spheres for mNP332.

Small angle neutron scattering (SANS) experiments were also performed to verify the supposed morphologies. All the samples were studied in D<sub>2</sub>O to have a better contrast

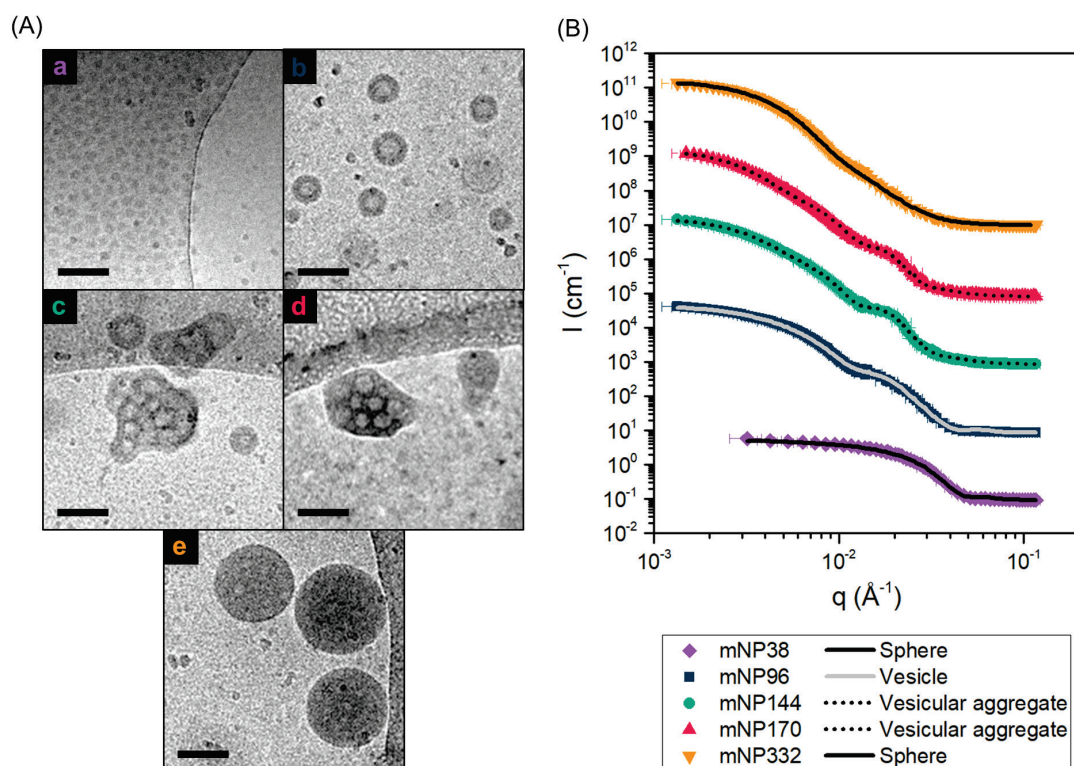
between PTMC and solvent. Scattering curves were fitted with an appropriate model for the form factor which was chosen according to cryo-TEM pictures (Figure 3B). Data plots for mNP38 and 332 were well adjusted with a sphere model, mNP96 with a unilamellar vesicle model and mNP144 and 170 with a specific model of a fractal core-shell to mimic a vesicular aggregate.

All the characteristics obtained from DLS 90°, MALS, cryo-TEM and SANS are summarized in Table 2. They provided a detailed internal structure of the different types of nanoparticles.

The radius from DLS was found to be higher than the radii found by cryo-TEM and SANS. For these two techniques, both the instrumental limitation and the low contrast of PEG lead to an underestimation of the radius whereas DLS is more sensitive to larger objects and give a hydrodynamic radius comprising the PEG corona and the solvation layer. This explains the difference of size determined from these different methods. Thus, the size of the NPs was only discussed with the hydrodynamic radius from DLS while cryo-TEM and SANS were both employed to assess the hydrophobic membrane thickness ( $\delta$ ) of the vesicular structures obtained for mNP96, 144 and 170.

Values of  $\delta$  were similar between cryo-TEM and SANS (Table 2). Standard deviations on  $\delta$  found with both techniques were also comparable, meaning that  $\delta$  measurements were equally precise. However, since SANS is statistically more relevant than cryo-TEM, we chose to discuss on  $\delta_{SANS}$  values, which were the following:  $14.6 \pm 1.5$  nm for mNP96,  $18.1 \pm 3.3$  nm for mNP144 and  $20.2 \pm 4.0$  nm for mNP170.

The higher values of standard deviations for both mNP144 and 170 compared to mNP96 can be explained by the fact that they both are vesicular aggregates and not well-defined unilamellar vesicles like mNP96.



**Figure 3.** Cryo-TEM images (A) and SANS curves (B) of the different nanoparticles obtained by manual nanoprecipitation (mNP) using the series of PEG<sub>45</sub>-*b*-PTMC<sub>n</sub> diblock copolymers ( $n = 38$  (purple, a), 96 (blue, b), 144 (green, c), 170 (red, d), 332 (orange, e)). A) Scale bar: 100 nm. B) Neutron scattering curves were fitted with theoretical plots for different morphology models: sphere (black line), unilamellar vesicle (grey line) or vesicular aggregate (black dotted line) with  $10^2 \text{ cm}^{-1}$  of offset between curves for clarity.

**Table 2.** DLS 90°, MALS, cryo-TEM and SANS data obtained on the same nanoparticles formed by mNP and analyzed in H<sub>2</sub>O for DLS 90° and MALS and D<sub>2</sub>O for cryo-TEM and SANS.

mNP	DLS 90°		MALS			Cryo-TEM					SANS					
	R <sub>h</sub>	PDI	R <sub>g</sub>	R <sub>h</sub>	R <sub>g</sub> /R <sub>h</sub>	m	R	SD <sub>R</sub>	δ	SD <sub>δ</sub>	m	φ	R	SD <sub>R</sub>	δ	SD <sub>δ</sub>
	(nm)		(nm)	(nm)			(nm)	(nm)	(nm)	(nm)		(10 <sup>-4</sup> g/mL)	(nm)	(nm)	(nm)	(nm)
38	15	0.09	-	14	-	s	7.8	1.2	-	-	s	5.6	8.8	1.1	-	-
96	55	0.13	53	51	1.04	v	37.3	15.0	14.6	1.6	v	8.8	30.1	10.5	14.6	1.5
144	85	0.14	69	73	0.95	va	67.5	24.7	17.5	2.9	va	10.1	29.6	10.4	18.1	3.3
170	90	0.14	71	81	0.88	va	72.1	29.0	19.2	3.9	va	6.4	29.7	12.5	20.2	4.0
332	70	0.06	55	68	0.81	s	47.1	13.3	-	-	s	9.5	39.4	11.0	-	-

R<sub>h</sub>: hydrodynamic radius

R<sub>g</sub>: radius of gyration

R<sub>g</sub>/R<sub>h</sub>: ratio of R<sub>g</sub> over R<sub>h</sub>

m: morphology observed by cryo-TEM and model chosen for SANS data fitting: s for sphere, v for vesicle and va for vesicular aggregate

R: radius measured on cryo-TEM images and radius estimated with SANS data fitting: s) core radius, v and va) external radius (radius underestimated compared to DLS due to low contrast of PEG for both cryo-TEM and SANS)

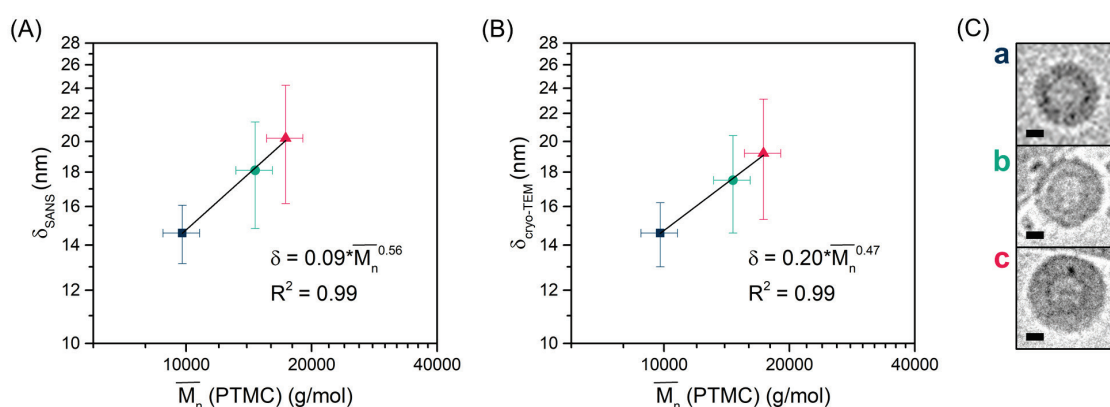
δ: hydrophobic membrane thickness measured on cryo-TEM images and estimated with SANS data fitting

SD<sub>R</sub> and SD<sub>δ</sub>: standard deviations respectively on R and δ from cryo-TEM images and SANS data fitting

φ: volume fraction: PTMC concentration in particles

The vesicles hydrophobic membrane thickness was plotted as function of the hydrophobic block molar mass (Figure 4). A typical scaling law with a power-dependency was determined with both SANS and cryo-TEM data:  $\delta_{SANS} \propto \overline{M}_n^{0.56}$  (Figure 4A) and  $\delta_{cryo-TEM} \propto \overline{M}_n^{0.46}$  (Figure 4B, 4C). This scaling law can be explained considering that a polymer chain in a  $\theta$  solvent has a radius of gyration proportional to its molar mass power 0.5 ( $R_g \propto \overline{M}_n^{0.5}$ ).<sup>50</sup> The

exponent for PEG<sub>45</sub>-*b*-PTMC<sub>n</sub> block copolymers found to be slightly above 0.5 shows that the polymer chains might be slightly stretched in the membrane due to the segregation between PEG and PTMC domains at the vesicle membrane interface. The establishment of such scaling law is important to predict which PTMC block molar mass is needed to obtain a specific hydrophobic membrane thickness, while keeping the  $f_{\text{PEG}}$  close to 17% in order to guarantee the formation of unilamellar vesicles using the manual nanoprecipitation protocol.



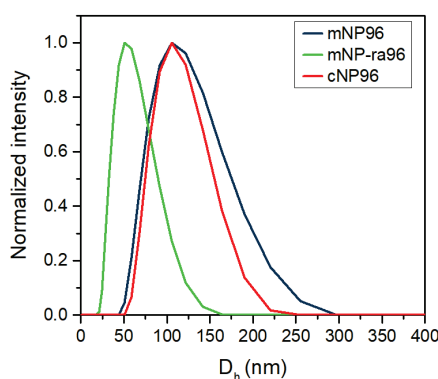
**Figure 4.** Hydrophobic membrane thickness ( $\delta$ ) of vesicular objects as a function of the hydrophobic block molar mass ( $\bar{M}_n$  (PTMC)) for three PEG<sub>45</sub>-*b*-PTMC<sub>n</sub> copolymers with  $n = 96$  (blue), 144 (green), 170 (red). Membrane thicknesses were: A) estimated by fitting SANS data ( $\delta_{\text{SANS}}$ ) and B) measured on cryo-TEM images ( $\delta_{\text{cryo-TEM}}$ ). Fits show a power dependency of  $\delta$  with  $\bar{M}_n$  by an exponent of 0.56 for  $\delta_{\text{SANS}}$  (A) and 0.47 for  $\delta_{\text{cryo-TEM}}$  (B). C) Cryo-TEM images focusing on one polymersome of mNP96 (a, blue), mNP144 (b, green) and mNP170 (c, red). Scale bar: 20 nm.

The results of this self-assembly study using the same formulation protocol showed how the hydrophilic fraction  $f_{\text{PEG}}$  of PEG<sub>45</sub>-*b*-PTMC<sub>n</sub> is dictating the nanoparticles morphology.

In a second time, we studied the influence of the nanoprecipitation process, including the use of a microfluidic device, on the size and morphology of the self-assembled nanoparticles, focusing on PEG<sub>45</sub>-*b*-PTMC<sub>96</sub> block copolymer.

### Influence of the nanoprecipitation process

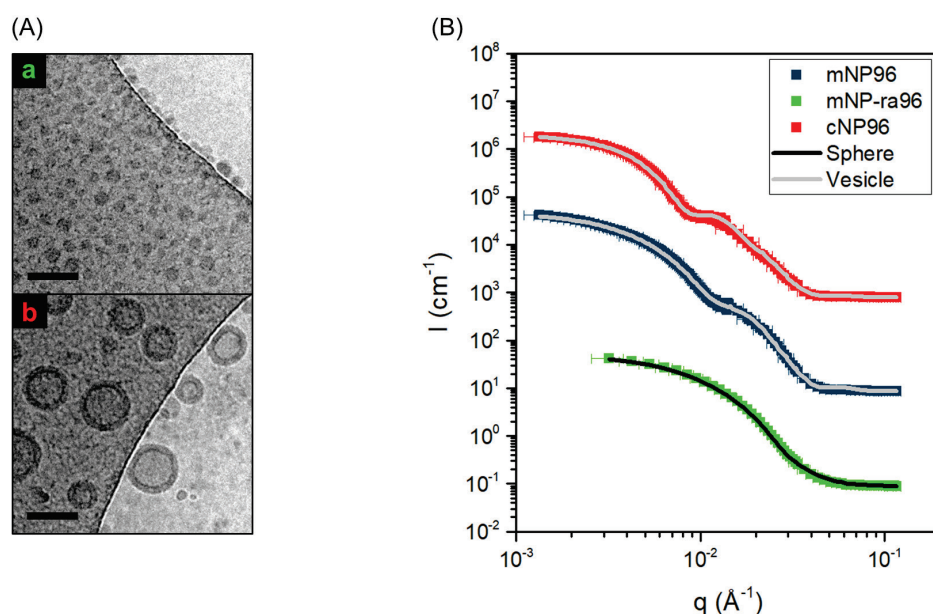
Manual nanoprecipitation of PEG<sub>45</sub>-*b*-PTMC<sub>96</sub> was first performed by a reverse addition (mNP-ra96) of the copolymer solution in DMSO into water and subsequent dialysis against water. DLS 90° curve showed smaller objects for mNP-ra96 than for mNP96 with  $R_h \approx 30$  nm versus 55 nm (Figure 5).



**Figure 5.** DLS 90° curves of the different PEG<sub>45</sub>-*b*-PTMC<sub>96</sub> nanoparticles obtained by different processes: manual nanoprecipitation upon addition of water in the copolymer solution (mNP96, blue) or upon reverse addition (mNP-ra96, green) and by controlled nanoprecipitation using microfluidics (cNP96, red).

Combined characteristics from MALS, cryo-TEM and SANS (Figure 6, Table 3) confirmed for mNP-ra96 a micellar structure of the objects with a core size close to 11 nm. This change in morphology correlated to a modification in the order of solvent addition can be expected since it may increase the rate at which the copolymer is precipitated, the overall system being thermodynamically out of equilibrium (in a “frozen” state).





**Figure 6.** Cryo-TEM images (A) and SANS curves (B) of the different PEG<sub>45</sub>-*b*-PTMC<sub>96</sub> nanoparticles obtained by different processes: manual nanoprecipitation upon addition of water in the copolymer solution (mNP96, blue) or upon reverse addition (mNP-ra96, green, a) and by controlled nanoprecipitation using microfluidics (cNP96, red, b). A) Scale bar: 100 nm. For comparison, cryo-TEM image of mNP96 can be found in Figure 3. B) Neutron scattering curves were fitted with theoretical plots for different morphology models: sphere (black line) or unilamellar vesicle (grey line) with  $10^2 \text{ cm}^{-1}$  of offset between curves. Curves of mNP96 (blue) is reminded for easier comparison.

**Table 3.** DLS 90°, MALS, cryo-TEM and SANS data of the different PEG<sub>45</sub>-*b*-PTMC<sub>96</sub> nanoparticles obtained by different processes: manual nanoprecipitation upon addition of water in the copolymer solution (mNP96) or upon reverse addition (mNP-ra96) and by controlled nanoprecipitation using microfluidics (cNP96).

	DLS 90°		MALS			Cryo-TEM					SANS						
Sample	R <sub>h</sub>	PDI	R <sub>g</sub>	R <sub>h</sub>	R <sub>g</sub> /R <sub>h</sub>	m	R	SD <sub>R</sub>	δ	SD <sub>δ</sub>	m	φ	R	SD <sub>R</sub>	δ	SD <sub>δ</sub>	
	(nm)		(nm)	(nm)			(nm)	(nm)	(nm)	(nm)		(10 <sup>-4</sup> g/mL)	(nm)	(nm)	(nm)	(nm)	
mNP96	55	0.13	53	51	1.04	v	37.3	15.0	14.6	1.6	v	8.8	15.5	5.4	14.6	1.5	
mNP-ra96	30	0.17	34	28	1.21	s	11.3	4.6	-	-	s	7.2	11.3	3.5	-	-	
cNP96	55	0.04	49	51	0.96	v	40.6	7.4	14.7	1.6	v	3.4	25.0	6.5	14.2	1.8	

R<sub>h</sub>: hydrodynamic radius

R<sub>g</sub>: radius of gyration

R<sub>g</sub>/R<sub>h</sub>: ratio of R<sub>g</sub> over R<sub>h</sub>

m: morphology observed by cryo-TEM and model chosen for SANS data fitting: s for sphere and v for vesicle

R: radius measured on cryo-TEM images and radius estimated with SANS data fitting: s) core radius and v) external radius (radius underestimated compared to DLS due to low contrast of PEG for both cryo-TEM and SANS)

δ: hydrophobic membrane thickness measured on cryo-TEM images and estimated with SANS data fitting

SD<sub>R</sub> and SD<sub>δ</sub>: standard deviations respectively on R and δ from cryo-TEM images and SANS data fitting

φ: volume fraction: PTMC concentration in particles

The third preparation process used was controlled nanoprecipitation (cNP), carried out using a microfluidic system depicted in Figure S7. A screening of different conditions with such a versatile system was performed again with PEG<sub>45</sub>-*b*-PTMC<sub>96</sub>. Different flow rate, water content and initial copolymer concentration were tested in three different microfluidic set-ups (Figure S8). Nanoparticles obtained from the H<sub>2</sub>O/DMSO mixture (before removal of DMSO by dialysis) were characterized by DLS 90° with the adequate solvent viscosity.

Results indicated in Table S3 revealed that nanoparticles' size was increased upon an increase of the copolymer concentration in the final H<sub>2</sub>O/DMSO mixture. This trend was observed when the water content was decreased while keeping the initial copolymer concentration constant. The same trend was also found when the initial copolymer concentration was increased while keeping the water content constant. Secondly, the decrease of the total flow rate gave slightly bigger objects for water contents of 50 and 70 vol%. In addition, for 30 vol%, no real impact on the size was observed. Finally, a negligible effect was observed on the objects size formed with the three different set-ups that were tested.

With all the different conditions carried out, we managed to control the objects size with  $R_h$  between 22 and 85 nm and a PDI repeatedly lower than 0.1. Only size is mentioned here since no other characterization than DLS 90° was performed to determine the objects morphology. However, since mNP-ra96 was demonstrated to form micelles of  $R_h \approx 30$  nm with a fast mixing process, one can expect micelles for the smaller objects formed by microfluidics in rather similar conditions, and vesicles for objects with  $R_h \geq 50$  nm since mNP96 was shown to form vesicles when close to equilibrium. As such, one specific sample giving  $R_h = 58$  nm before dialysis (cNP96) was studied thoroughly to confirm the vesicular structure. The conditions used in the microfluidics system were as follows: a flow rate of 100  $\mu$ L/min for H<sub>2</sub>O, 233  $\mu$ L/min for the copolymer solution in DMSO at 1 g/L, meaning a water content of 30 vol%. First, DLS 90° was done after dialysis (Figure 5) and showed  $R_h = 55$  nm with PDI = 0.04 (Table 3) which was much lower than the one of mNP96 with 0.13. Further data from MALS, cryo-TEM and SANS confirmed a vesicle morphology of cNP96 (Figure 6, Table 3) with the same membrane thickness ( $\delta \approx 14.6$  nm) as for mNP96 vesicles, since it is an intrinsic characteristic of the PEG<sub>45</sub>-*b*-PTMC<sub>96</sub> copolymer.

However, for cNP96, smaller size dispersity was observed by cryo-TEM and the oscillation characteristic of the bilayer thickness observed by SANS was well marked, confirming an

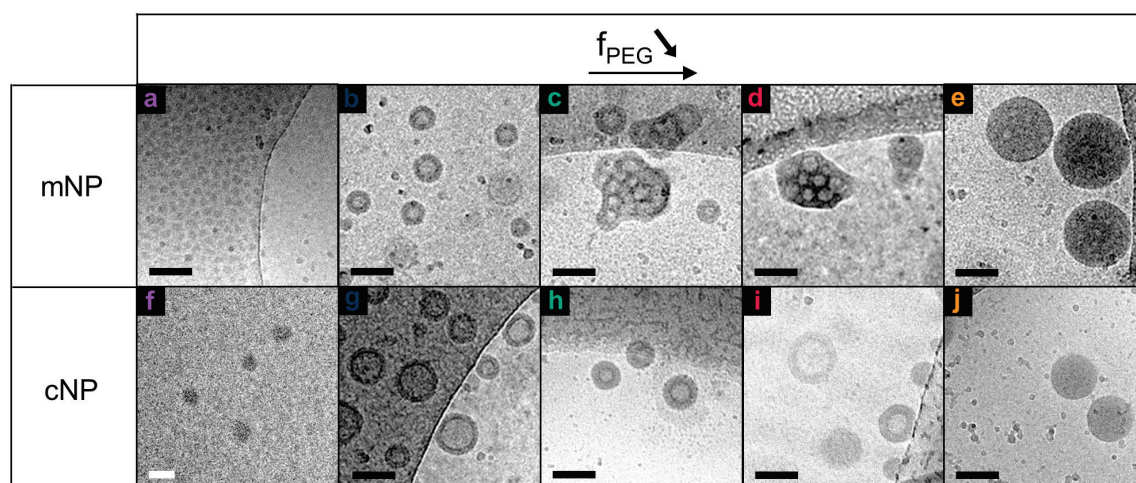
overall well-defined size of vesicles. To conclude, microfluidics was found to be a successful technique with this particular condition to obtain more monodisperse vesicles than the manual nanoprecipitation procedure.

Controlled nanoprecipitation (cNP) with the same microfluidic conditions was then applied for all the other block copolymers of the series, and the obtained nanoparticles were characterized by DLS 90°, MALS and cryo-TEM (Figure 7f-j, Table S4). cNP38 gave micelles with the same core radius than for mNP38 of 8 nm but aggregates were obtained too, leading to a poor PDI of 0.7. cNP144 and 170 generated a mixture of vesicles of 40 nm and spheres of 20 to 30 nm in radius, as determined by cryo-TEM. Hence, vesicular aggregates were avoided *via* microfluidics but the hydrophilic weight fraction of these two block copolymers are probably too low to find a good condition to obtain exclusively a vesicular shape. Values of membrane thicknesses were a bit higher than the ones obtained for mNP144 and 170 (Table 2) but still within the standard deviations. Finally, cNP332 produced spheres of  $R_h \approx 50$  nm, smaller than for mNP332 with a good PDI of 0.14. These results showed that applying the same microfluidic condition on four different block copolymers of our series with  $f_{\text{PEG}} \leq 17\%$  (PEG<sub>45</sub>-*b*-PTMC<sub>n</sub> n = 96, 144, 170, 332) confirmed the power of microfluidics to control nanoparticles size with low dispersity for morphologies between vesicles and spheres. An optimization of the different microfluidics conditions could be investigated in order to perfect the resulting nanoparticles, especially on PEG<sub>45</sub>-*b*-PTMC<sub>38</sub> with  $f_{\text{PEG}} \approx 34\%$  in order to obtain monodisperse micelles.

## CONCLUSIONS

In this work, the synthesis by ROP of five PEG<sub>45</sub>-*b*-PTMC<sub>n</sub> block copolymers was performed to obtain different hydrophilic weight fraction  $f_{\text{PEG}}$  between 34% and 6% by

changing the PTMC chain length ( $n = 38, 96, 140, 170, 332$ ). These copolymers are of particular interest for the design of biomaterials due to the known biocompatibility of PEG and the low systemic toxicity of PTMC.<sup>35</sup> In addition, the PTMC block has also been interestingly reported to be biodegradable upon enzymatic hydrolysis.<sup>37,38,22</sup> This series was self-assembled using different processes to form nanoparticles with different sizes and morphologies (Figure 7).



**Figure 7.** Summary of cryo-TEM images of the different nanoparticles obtained by manual nanoprecipitation (mNP, a-e) and controlled nanoprecipitation (cNP, f-j) using the series of PEG<sub>45</sub>-b-PTMC<sub>n</sub> diblock copolymers with  $n = 38$  (purple, a/f), 96 (blue, b/g), 144 (green, c/h), 170 (red, d/i), 332 (orange, e/j). Black scale bar: 100 nm; white scale bar (f): 20 nm.

First, by applying the same manual nanoprecipitation protocol for all copolymers, different morphologies were obtained according to  $f_{\text{PEG}}$ : micelles for  $f_{\text{PEG}} \approx 34\%$ , vesicles for  $17\% < f_{\text{PEG}} < 10\%$  and spheres for  $f_{\text{PEG}} \approx 6\%$  (Figure 7, mNP). Then, by controlled nanoprecipitation using a microfluidic system, it was possible to control nanoparticles size while keeping the same morphologies (Figure 7, cNP), indicating that the shape of the self-assembled structures is truly dictated by the macromolecular parameters ( $f_{\text{PEG}}$ ), when formed at equilibrium.

Indeed, when mixing rate is fast enough to be thermodynamically out of equilibrium and in a “frozen” state, other morphologies can also be obtained. This was illustrated by the observation of vesicles formed by manual nanoprecipitation of PEG<sub>45</sub>-*b*-PTMC<sub>96</sub> upon addition of water in the copolymer solution while “frozen” micelles were obtained when the addition was performed reversely, leading to a rapid nanoprecipitation process.

Furthermore, a microfluidic study on the best copolymer candidate to form vesicles, PEG<sub>45</sub>-*b*-PTMC<sub>96</sub> ( $f_{\text{PEG}} \approx 17\%$ ), showed that vesicles size can be tuned and controlled by changing different parameters such as flow rates, water content and copolymer concentration. One condition was found to match the same vesicle size as the one obtained by manual nanoprecipitation ( $D_h \approx 100$  nm) with excellent monodispersity ( $\text{PDI} < 0.1$ ). Vesicles had the same membrane thickness ( $\delta \approx 14.6$  nm), illustrating that  $\delta$  is only imposed by the copolymer and not dependent on the formulation process.

Finally, a scaling law was drawn:  $\delta \propto \overline{M}_n^{0.56}$ , the hydrophobic membrane thickness  $\delta$  is proportional to the molar mass of the hydrophobic block  $\overline{M}_n$  to a power 0.56. This simple but influential law will provide anyone working on similar copolymers to predict the membrane thickness of vesicles and engineer the appropriate copolymer, providing that  $f_{\text{PEG}}$  remains close to 17% to maintain a vesicular structure.

As a general outlook, we now have all the tools needed to form PEG-*b*-PTMC vesicles with different membrane thicknesses and PEG lengths in order to tune certain physicochemical membrane properties (stability, permeability, fluidity...) which are critical parameters for future applications in biomaterial science and drug delivery. In future work, this will be especially interesting to study the effect of PEG length on colloidal stability and stealth properties, and the effect of the polymersome membrane thickness on mechanical and diffusion properties. In addition, the ability to fine-tune the polymersome size using a microfluidic assisted nanoprecipitation method, with very low dispersity and high

repeatability is crucial for clinical and industrial development of nanoparticles, especially polymersomes.

## ASSOCIATED CONTENT

Supporting Information is available.  $^1\text{H}$  NMR spectra, SEC chromatograms, TGA thermograms, additional DSC, DLS, MALS, cryo-TEM data and scheme of the microfluidic system.

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## CONFLICTS OF INTEREST

The authors declare no conflicts of interest.



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