



HAL
open science

AFM dendritips functionalized with molecular probes specific to cell wall polysaccharides as a tool to investigate cell surface structure and organization

Marion Schiavone, Nathalie Sieczkowski, Mathieu Castex, Emmanuelle
Trévisiol, Etienne Dague, Jean Marie François

► To cite this version:

Marion Schiavone, Nathalie Sieczkowski, Mathieu Castex, Emmanuelle Trévisiol, Etienne Dague, et al.. AFM dendritips functionalized with molecular probes specific to cell wall polysaccharides as a tool to investigate cell surface structure and organization. *The Cell Surface*, 2019, 5, pp.100027. 10.1016/j.tcs.2019.100027 . hal-02295500

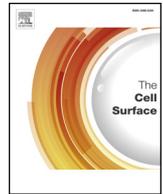
HAL Id: hal-02295500

<https://hal.science/hal-02295500v1>

Submitted on 24 Sep 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



AFM dendritips functionalized with molecular probes specific to cell wall polysaccharides as a tool to investigate cell surface structure and organization



Marion Schiavone^{a,b}, Nathalie Sieczkowski^b, Mathieu Castex^b, Emmanuelle Trevisiol^c, Etienne Dague^{c,*}, Jean Marie François^{c,*}

^a LISBP, UMR INSA-CNRS 5504 & INRA 792, F-31077 Toulouse, France

^b Lallemand SAS, 19, rue des briquetiers, 31702 Blagnac, France

^c CNRS, LAAS, 7 avenue du colonel Roche, F-31400 Toulouse, France

ARTICLE INFO

Keywords:

Cell surface
Cell wall
 β -Glucan
Chitin
AFM dendritips
Anti β -1,3-glucan antibodies
Molecular interaction

ABSTRACT

The yeast cell wall is composed of mannoproteins, β -1,3/ β -1,6-glucans and chitin. Each of these components has technological properties that are relevant for industrial and medical applications. To address issues related to cell wall structure and alteration in response to stress or conditioning processes, AFM dendritips were functionalized with biomolecules that are specific for each of the wall components, which was wheat germ agglutinin (WGA) for chitin, concanavalin A (ConA) for mannans and anti- β -1,3/anti- β -1,6-glucan antibodies for β -1,3/ β -1,6-glucans. Binding specificity of these biomolecules were validated using penta-*N*-acetylchitopentaose, α -mannans, laminarin (short β -1,3-glucan chain) and gentiobiose (2 glucose units linked in β 1 \rightarrow 6) immobilized on epoxy glass slides. Dynamic force spectroscopy was employed to obtain kinetic and thermodynamic information on the intermolecular interaction of the binary complexes using the model of Friddle-Noy-de Yoreo. Using this model, transition state distance x_t , dissociate rate k_{off} and the lowest force (f_{eq}) required to break the intermolecular bond of the complexes were approximated. These functionalized dendritips were then used to probe the yeast cell surface treated with a bacterial protease. As expected, this treatment, which removed the outer layer of the cell wall, gave accessibility to the inner layer composed of β -glucans. Likewise, bud scars were nicely localized using AFM dendritip bearing the WGA probe. To conclude, these functionalized AFM dendritips constitute a new toolbox that can be used to investigate cell surface structure and organization in response to a wide arrays of cultures and process conditions.

1. Introduction

The yeast cell wall of *Saccharomyces cerevisiae* is a solid armour of about 150 nm thick, which represents 20–25% of the cell dry mass (Aguilar-Uscanga and Francois, 2003). This cell wall consists of mannoproteins (40–45% of wall dry mass), β -1,3 and β -1,6-glucans (45–55% of wall dry mass) and a tiny amount of chitin (less than 5% of wall dry mass), which are cross-linked in various ways to form higher-order complexes (Klis et al., 2006). Under transmission electron microscopy, the cell wall appears as the juxtaposition of two layers, an outer layer of about 30–40 nm thick that is merely composed of mannoproteins and an inner layer of about 70–100 nm mainly composed of β -glucans (Osumi et al., 1984). These two layers are however

interconnected through different covalent linkages of mannoproteins to β -glucans (see Orlean (2012), for a review on the subject). Although the chemical composition of the yeast cell wall is well known, its molecular and spatial organization is still challenging to draw. Moreover, the amount, the physical structure and the interlinkages between cell wall components are subject to dramatic changes in response to various stress, culture conditions, fermentation processes and downstream processing (Aguilar-Uscanga and Francois, 2003; Pillet et al., 2014; Schiavone et al., 2016, 2015). The cell wall composition was also reported to be substantially different among yeast species (Nguyen et al., 1998).

Each of the cell wall components is endowed with technological properties that are pertinent for industrial and medical applications. A

* Corresponding authors.

E-mail addresses: schiavon@insa-toulouse.fr (M. Schiavone), nsieczkowski@lallemand.com (N. Sieczkowski), mcastex@lallemand.com (M. Castex), etrevisi@laas.fr (E. Trevisiol), edague@laas.fr (E. Dague), fran_jm@insa-toulouse.fr (J.M. François).

<https://doi.org/10.1016/j.tcs.2019.100027>

Received 8 April 2019; Received in revised form 13 May 2019; Accepted 14 June 2019

Available online 05 August 2019

2468-2330/© 2019 The Author(s). Published by Elsevier B.V. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

Table 1
Structure of the polysaccharides used in this study.

Name	MW (kDa)	Formula	Chemical representation
Laminarin	~5	β -1,3-Glucan with ramifications of β -1,6-glucan	
Gentiobiose	0.342	2 units of glucose linked in β (1 \rightarrow 6)	
Mannan	54	Mannose units linked in α (1 \rightarrow 4); α (1 \rightarrow 2); α (1 \rightarrow 6)	
Penta-N-acetylchitopentaose	~1.0	5 units of N-acetyl-D-glucosamine linked in β (1 \rightarrow 4)	

well-recognized property of the yeast cell wall mannoproteins resides in the adhesion to inert surface and in the consequent formation of biofilms (Blankenship and Mitchell, 2006; Bojsen et al., 2012), as well as in the capacity to interact with aromatic and phenolic compounds during winemaking process (Chalier et al., 2007; Mekoue Nguela et al., 2016; Pradelles et al., 2008). The *manno*-oligosaccharides that decorate cell wall proteins are also reported to exhibit prebiotic properties as illustrated by the stimulation of gut microbiota of chickens (Pourabedin et al., 2017; Pourabedin and Zhao, 2015) and by preventing expansion of pathogenic bacteria via direct connexion to specific fimbriae present at the bacteria surface (Ganner and Schatzmayr, 2012). Moreover, yeast mannans have been reported to improve immune responses and growth efficiency of pigs infected with virus (Che et al., 2012). On the other hand, immuno-stimulatory, anti-tumor and anti-diabetes properties have been documented for β -glucans, which is due to their interactions with receptors among which dectin-1 and complement receptor CR3 are the most characterized (Aleem, 2013; Clemons et al., 2014; Du et al., 2014; Legentil et al., 2015). However, the effects of β -glucans are dependent on the degree of branching and on the enchainment of the glucosyl units (1 \rightarrow 3, 1 \rightarrow 4, 1 \rightarrow 6) (Bohn and Bemiller, 1995; Raa,

2015), what can explain some discrepancies in the reported effects of the polysaccharides originated from various biological sources or isolated by different procedures (Cleary et al., 1999; Kulicke et al., 1997). Chitin also shows immunostimulating properties in mammals (Lee et al., 2008), which are strongly influenced by the size of chitin particles (Cuesta et al., 2003). This polysaccharide was also reported to induce cytokine production for leukocyte recruitment and activation of macrophages (Da Silva et al., 2009; Esteban et al., 2001).

The rapid development of Atomic Force Microscopy (AFM) in liquid has allowed to address several problems on the structural organization and interaction of cell surface with its environment, which could not be attempted by other techniques. The remarkable properties of AFM is to be a non-destructive technique that requires a minimal sample preparation. It enables imaging individual cells at high resolution and measuring interaction forces in the range of piconewtons (Dufrene, 2008; Muller and Dufrene, 2008). Using this tool, quantitative data regarding nanomechanical properties (*i.e.* stiffness, elasticity) of microbial cells have been obtained (Dague et al., 2010, 2007). More remarkably, the functionalization of AFM tips with specific biomolecules or cells opened a new angle of investigating the interaction at the single

molecular level (single-molecule force spectroscopy or SMFS) (Alsteens et al., 2011; Hinterdorfer and Dufrene, 2006) or single cell level (single-cell force spectroscopy or SCFS) (Alsteens et al., 2013; Benoit et al., 2000; El-Kirat-Chatel and Beaussart, 2018). Dynamic-force spectroscopy (DFS) can then be employed to measure binding properties of biomolecules in a dynamic manner, which means the force it takes to manipulate a biomolecule or a biomolecular complex (Merkel et al., 1999; Takeuchi et al., 2006). It requires the application of an external force that increases linearly with time so that the molecule or complex is exposed to a constant loading rate. Evans and Ritchie (1997) found that the bond strength, which corresponds to the rupture force between two interacting molecules, varies as the logarithm of the loading rate. However, force spectra are rarely linear, and biological entities such as pili or polymers have shown a nonlinear force response (Bjornham and Andersson, 2017). The nonlinear trend of the force spectra was investigated by Friddle et al. (2012). They proposed a new DFS model based on the existence of two regimes that account for this non-linearity, namely (i) a near equilibrium regime characterized by a finite force f_{eq} that corresponds to the lowest force required to break the bond and (ii) a kinetic regime characterized by a dependence of the force to the logarithm of the loading rate. Application of this model provides the energy landscape of molecular interaction giving rise to force, dissociate rate and energy barrier of these interactions.

The purpose of this work was to investigate the dynamics of interaction between AFM tips functionalized with biomolecules that shall specifically interact with the four different components of the yeast cell wall. To this end, dendrimer-activated tips (so-called dendritip) were functionalized with either wheat germ agglutinin (WGA), concanavalin A (Con A), anti- β -1,3-glucan or anti- β -1,6-glucan antibodies. We validated the specificity of these functionalized dendritips by probing epoxy-activated glass slides coated with laminarin (a β -1,3-glucan oligosaccharide), gentiobiose (a disaccharide linked in β 1 \rightarrow 6), penta-*N*-acetylchitopentaose (5 units of *N*-acetylglucosamine linked in β 1 \rightarrow 4) (see Table 1) and α -mannans as sugar substrates that are representative of the yeast cell wall polysaccharides. Moreover, kinetic and thermodynamic information about these interactions were extracted using the Friddle-Noy-de Yoero model (Friddle et al., 2012). Finally, we employed these functionalized AFM dendritips to explore the yeast cell surface after a protease treatment as well as to map chitin at surface of the yeast cell concomitantly with bud scars.

2. Materials and methods

2.1. Chemicals and biochemicals

Ethanolamine hydrochloride, tetrahydrofuran (THF), dichloromethane, dimethylsulfoxide (DMSO), sodium cyanoborohydride, acetone, ethanol, bovine serum albumin (BSA), *o*-phenylenediamine (OPD), laminarin from *Laminaria digitata*, α -mannan from *Saccharomyces cerevisiae* (M75014), D-glucose, D-mannose, *N*-acetyl-D-glucosamine were purchased from Sigma-Aldrich and used without further purification. Pustulan (β -1,6-glucan) and β -1,6-gentiobiose were obtained from Calbiochem and Carbosynth, respectively. β -1,3-laminaribiose and penta-*N*-acetylchitopentaose were provided by Megazyme. PBS buffer (10 mM Na_2HPO_4 , 138 mM NaCl, 2.7 mM KCl, pH 7.4) was purchased in Euromedex. Epoxide-activated glass slides were from Schott Nexterion (Nexterion® Slide E). The MLCT AFM tips were obtained from Bruker. Wheat Germ Agglutinin (WGA) and concanavalin A (ConA) from Sigma-Aldrich. Anti- β -1,3-glucan, a mouse monoclonal IgG was provided by Biosupplies (<http://www.biosupplies.com.au/>, Australia). The anti- β -1,6-glucan antibody, a rabbit polyclonal antibody IgG, was kindly provided by Prof. Frans Klis (Swammerdam University of Amsterdam). A goat-anti-rabbit and a rabbit anti-mouse IgG-peroxidase conjugates were purchased from ThermoFisher Scientific.

2.2. Yeast, culture condition and protease treatment

The yeast *Saccharomyces cerevisiae* wild-type BY4741 (MATa his3 Δ 1; leu2 Δ 0; met15 Δ 0; ura3 Δ 0) stored at -80°C was revived on YPD agar plate and then cultivated in YPD medium (1% w/v yeast extract, 2% w/v peptone, 2% w/v glucose) at 30°C with shaking at 200 rpm until $\text{OD}_{600} = 1$ ($\sim 1.4 \times 10^7$ cells/ml). Unless otherwise stated, cells were harvested by centrifugation (3000 rpm \times 3 min), washed with PBS or acetate buffer (18 mM, pH 5.2) and re-suspended to obtain around 5 OD unit at 600 nm. Yeast cells were immobilized by mechanical trapping into polydimethylsiloxane (PDMS) stamps (Formosa et al., 2015a). Firstly, a bare AFM tip was used to visualise and image a single cell before to change it with a functionalized dendritip. For protease treatment, YPD-cultivated yeast cells were treated with 2 mg/ml of *Streptomyces griseus* protease (P6911, Sigma-Aldrich) at 37°C for 6 h at pH 7.5, then collected by centrifugation and processed as above. Eight cells from 3 independent experiments were analysed for each treatment and a total of 8192 force curves were analysed.

2.3. Characterization of antibodies specific for β -1,6-glucan and β -1,3-glucan

The binding specificity of the antibodies was determined by indirect ELISA assay against β -1,3-glucan-BSA and β -1,6-glucan-BSA as described by Montijn et al. (1994). β -1,3-laminaribiose and β -1,6-gentiobiose were conjugated to BSA by reductive amination described by Roy et al. (1984). BSA (6.8 mg, 0.1 μmol), oligosaccharides (10 mg, 29.2 μmol) and NaBH_3CN (18 mg, 0.159 mmol) were dissolved in 2.5 ml of sodium borate buffer (0.2 M, pH 9.0). The reaction was carried out at room temperature for 96 h with continuous stirring. Conjugates were purified from reaction mixtures using a spin column with a 10-kDa cut off (Amicon, Millipore), lyophilized and resuspended in PBS to a final concentration of 1 mg.ml $^{-1}$. For the indirect ELISA assays, 96-wells plates were coated with either β -1, 3-glucan-BSA or β -1, 6-glucan-BSA (100 μl of 10 $\mu\text{g.ml}^{-1}$ in PBS, pH 7.2). The coated plates were washed four times with PBS (200 μl per well) and incubated for 1 h at 37°C with serial dilutions (1:5000 to 1:20,000) of purified antibodies diluted in PBS containing 3% (w/v) BSA. The plates were washed four times with PBS, and then incubated with either a goat anti-rabbit or a rabbit anti-mouse IgG-peroxidase conjugate (200 μl per well of dilutions 1:1000 and 1:2000) during 1 h at 37°C . After incubation, the plates were washed 4 times with 100 μl of PBS. Then, 100 μl of a 0.5 mg.ml $^{-1}$ *O*-phenylenediamine solution in peroxidase buffer was added per well as chromogenic substrate. The plates were incubated at 37°C for 30 min and then read at 450 nm. Control assay was performed by the same procedure with 100 μl of PBS buffer instead of BSA conjugates.

2.4. Functionalization of AFM dendritips

The Si_3N_4 AFM tips were first functionalized with dendrimers, bearing 96 aldehydes groups according to the procedure described in Jauvert et al. (2012) to give rise to AFM dendritips. Then these tips were incubated for 1 h at room temperature in 100 μl of 0.2 mg.ml $^{-1}$ of lectin solution (ConA or WGA) in 100 mM carbonate sodium buffer pH 8.4 to which 2 μl of 1 M NaBH_3CN in 100 mM sodium hydroxide was added. For coupling with antibodies, AFM dendritips were incubated overnight at 4°C in 100 μl of 0.1 mg.ml $^{-1}$ of antibodies solution in 100 mM carbonate buffer with 2 μl of 1 M NaBH_3CN . After incubation, the AFM dendritips were washed two times in PBS-Tween (PBS containing 0.05% Tween® 20). They were immediately used after functionalization.

2.5. Immobilization of polysaccharides on epoxy-activated glass slides

To epoxy-activated glass slides (Nexterion® Slide E from Schott, Germany), 100 μl of 10 mg.ml $^{-1}$ of the different substrates (Table 1)

prepared, unless otherwise stated, in PBS buffer was deposited and incubated for 16 h at room temperature. As the coating is sensible to light, the slides were kept in dark. After incubation, the slides were washed two times with 100 μ l PBS. For ConA experiment, substrates were prepared in 18 mM sodium acetate buffer, pH 5.2 containing 1 mM MnCl₂ and 1 mM CaCl₂ and the slides were washed with the same buffer.

2.6. Single molecular atomic force spectroscopy measurements

The force spectroscopy experiments were carried out with a Nanowizard III atomic force microscope (Bruker-JPK Instruments). The spring constant of each dendritip was determined by the thermal noise method (Hutter and Bechhoefer, 1993) and were found to be in the range of 10–20 pN/nm. All experiments with antibodies and WGA were performed in PBS buffer. Experiments using ConA-AFM tips were done in sodium acetate buffer (18 mM, pH 5.2) containing 1 mM CaCl₂ and 1 mM MnCl₂. AFM images were recorded in the Quantitative Imaging mode (Chopin et al., 2013), at 20 °C in buffer solution. Force–distance curves were recorded in buffer solution at different retraction speeds ranging from 1 to 6 μ m.s⁻¹ and at a constant applied force of 500 pN. Adhesion force maps were obtained by recording 32 × 32 force–distance curves. For specific blocking of the interaction, 100 μ l of 0.2 M solution of D-glucose, D-mannose or N-acetyl-D-glucosamine was added to the embedded cells in the PDMS chamber before AFM measurements

2.7. Dynamic force spectroscopy

To determine the parameters of energy landscapes for each couple of interacting molecules the values of rupture forces were determined from the force–distance curves recorded at different loading rates. Thousands of approach–retract cycles were carried out to provide sufficient data to fit to the theoretical model. The intermolecular interactions of biomolecules were described as free energy landscapes using the Friddle–Noy–de Yoreo model (Friddle et al., 2012). In this model, the rupture force F that varies as a function of the loading rate r can be approximated by the following equation:

$$F(r) \cong f_{eq} + f_{\beta} * \ln \left(1 + \frac{re^{-\gamma}}{f_{\beta} * k_{off}(f_{eq})} \right) \quad (1)$$

where f_{eq} is the equilibrium force, f_{β} is the thermal force, γ is the Euler's constant ($\gamma = 0.577$) and K_{off} is the dissociate rate constant at f_{eq} . The loading rate r is defined as the change in force with time, therefore it influences the rupture force and bond survival.

The thermal force f_{β} is defined as:

$$f_{\beta} = \frac{k_B T}{x_t} \quad (2)$$

where k_B is the Boltzmann's constant, T is the temperature and x_t is the width of the energy barrier.

The equilibrium force f_{eq} is defined as the force at which the dissociation and association rate intersect and is given by the equation:

$$f_{eq} = \sqrt{2k_c \Delta G} \quad (3)$$

where k_c is the spring constant of the cantilever and ΔG is the Gibbs free energy of binding, representing the height of the activation barrier.

The bond lifetime is calculated as

$$\tau = 1/k_{off} \quad (4)$$

Origin 8.0 software (OriginLab) was used to fit Eq. (1) to experimental DFS plot with a three-parameters nonlinear fit, along with 99% confidence intervals and 99% prediction intervals of the rupture forces. Fitting convergence criteria were approximated with a nonlinear iterative Levenberg–Marquardt algorithm. The maximum of iterations were fixed at 500, the chi-square (χ^2) tolerance between each iteration

was fixed at 10^{-15} . The parameters f_{eq} , k_{off} , f_{β} were extracted from fitting and parameters x_{β} and ΔG were calculated using Eqs. (2) and (3).

2.8. Data analyses

AFM images and force–distance curves were analysed with the JPK Data Processing software (Bruker-JPK Instruments).

3. Results and discussion

3.1. Functionalization of AFM dendritips with wheat germ agglutinin, concanavalin A, anti- β -1,3 and anti β -1,6-glucan antibodies

To address the probing of the different polysaccharides that compose the yeast cell wall at a nanoscale level, we functionalized AFM tips with a biomolecule or ligand that should unambiguously recognize these cell wall macromolecular components. Until now, AFM tips bearing concanavalin A (ConA) that specifically interacts with mannose units of *manno*-oligosaccharides have been used to map mannoproteins at the yeast surface (Alsteens et al., 2008; Beaussart et al., 2012; Formosa et al., 2015b; Francius et al., 2009; Schiavone et al., 2015). El-Kirat-Chatel et al. reported the functionalization of AFM tips with wheat germ agglutinin (WGA) and monoclonal anti- β -1,3-glucan antibody to probe chitin and β -glucans, respectively, at the cell surface of different yeast species and after a heat shock treatment (El-Kirat-Chatel et al., 2013). The use of WGA to detect chitin was justified by early works showing that this lectin possesses eight adjacent sugar-binding sites (Schwefel et al., 2010), which binds to GlcNAc or β -(1,4)-linked oligomer derivatives, as determined by extensive studies using NMR, fluorescence and X-Ray diffraction experiments (Kristiansen et al., 1999; Lotan and Sharon, 1973; Wright et al., 1984). In this work, we wished to revisit the specificity and accuracy of the lectins and antibodies that interact with cell wall polysaccharides, starting from the process of chemical functionalization of silicon-nitride (Si₃N₄) AFM cantilevers with dendrimers to the immobilization of polysaccharide substrates on epoxy-activated glass slides.

We first verified the specificity of anti β -1,3- and anti β -1,6-glucans antibodies by performing ELISA assays against BSA- β -(1,3)-glucan and BSA- β -(1,6)-glucan conjugates according to the procedure of Montijn et al. (1994). Indirect ELISA titration of polyclonal anti- β -1,6-glucan antibodies against BSA-gentiobiose, a two units of glucose linked by a β 1 → 6 bond (Table 1), showed a half saturation value of 206 ng.ml⁻¹ with no interference with BSA- β -1,3-laminarin. This result shows the high specificity of this antibody to β -1,6-glucan since laminarin from *laminari digitata* used in this work is reported to be a short linear β 1,3-glucan chain cross-linked with β -1,6-glucan at a ratio of 7 (Hrmova and Fincher, 1993). On the other hand, a value of 30 ng.ml⁻¹ was obtained for the anti- β -1,3-glucan antibody against BSA-laminarin with no cross-reactivity with gentiobiose, in accordance with the data from the commercial supplier (data not shown). In a second step, we fabricated dendritips by amination of the silicon nitride AFM tips with ethanolamine followed by incubation of these amino-derived tips with G4-aldehyde dendrimers as described in Jauvert et al. (2012). These AFM dendritips were afterwards incubated with either WGA, ConA, anti- β -1,3-glucan or anti- β -1,6-glucan antibodies. This incubation was followed by reduction of the imine bond formed between the dendrimers and the silicon nitride tip and between dendrimers and the epsilon amino group of lysine of the protein molecules into amine in the presence of sodium cyanoborohydride. The biofunctionalized AFM dendritip were then be used for single-molecule force spectroscopy analyses.

The third step was to qualify the interaction of these functionalized AFM dendritips with their cell wall substrates. To this end, we sought to use epoxy-activated glass slides coated with soluble polysaccharides that are representative of β -1,3-glucan, β -1,6-glucan, chitin and α -mannans. The hydroxyl groups present on the sugar polysaccharides

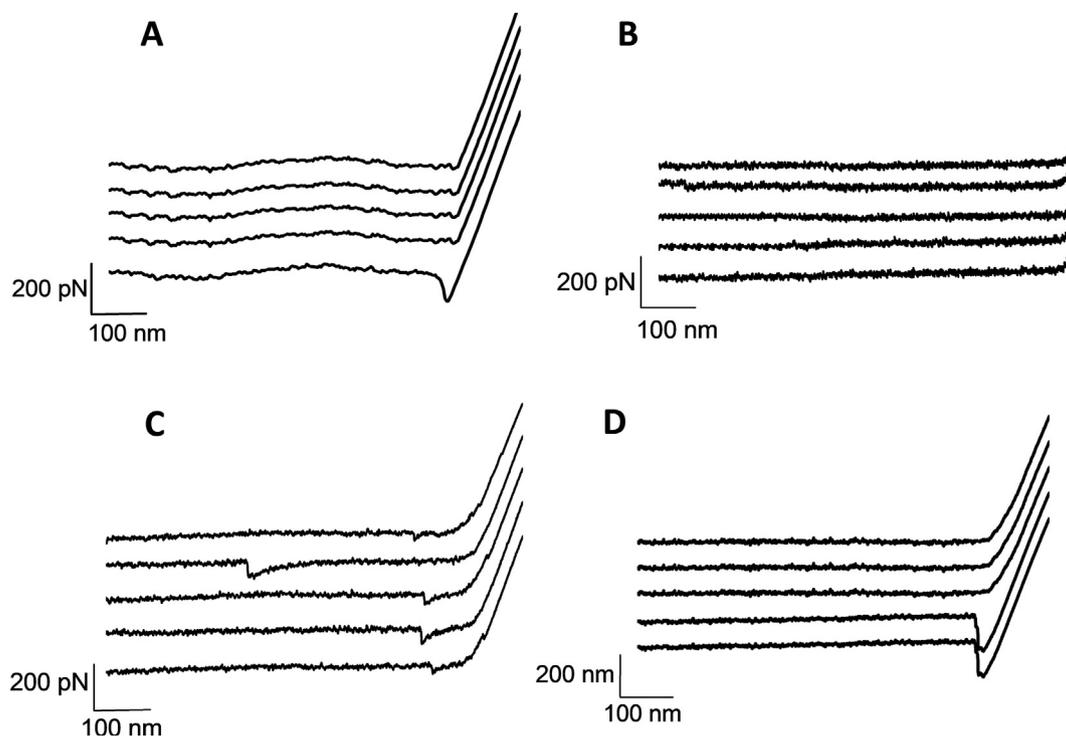


Fig. 1. Specificity of the interaction of AFM dendritips functionalized with anti- β -1,3-glucan antibodies on epoxide-activated glass slide alone (A) or coated with laminarin (C) and gentiobiose (D). The interaction of AFM tips is reported as force-distance curve, at a retraction rate of $1 \mu\text{m}\cdot\text{s}^{-1}$. In (B) is shown the result with a naked AFM dendritip on laminarin-coated slide.

should react with the epoxide function, leading to a covalent linkage between the sugar and the glass slide. We validated this coating strategy by spotting laminarin – a β -1,3-glucan substrate, penta-*N*-acetylchitopentaose, a substrate derived from chitin, and purified yeast α -mannans on epoxy-activated glass slides (Table 1). The coating of the first two substrates, which were biotinylated prior spotting on the glass slide was verified with streptavidin–Alexa fluor[®]647, whereas ConA labelled with this fluorophore was used to show that α -mannans were immobilized on the epoxy-activated glass slide. In addition, we showed that only WGA–Alexa fluor[®]647 binds to the immobilized chitopentaose (see Figs. S1 and S2 in Supplementary data).

3.2. The functionalized AFM dendritips specifically recognize their cognate substrates immobilized on epoxy-activated slides.

When probing the surface of a pristine glass slide with a AFM dendritip bearing anti- β -1,3-glucan antibodies, a deflection peak at the contact point with the surface was obtained (Fig. 1A). Similar data were obtained with AFM dendritips functionalized with the other biomolecules, such as with WGA (Fig. S3). Also, we did not find any interaction between AFM dendritip functionalized with anti- β -1,3-glucan antibodies and epoxy-activated glass slide coated with gentiobiose (Fig. 1D). Likewise, no interaction of WGA-tip to pristine glass slides or coated with laminarin was found (Fig. S3). In contrast, deflections at distance ranging from 40 to 500 nm were recorded from the retraction of an AFM dendritip functionalized with anti- β -1,3-glucan antibody on a laminarin-coated glass slide (Fig. 1C), demonstrating the specificity of the interaction of this antibody to its substrate. To further validate the specificity and affinity of the functionalized AFM dendritips towards their cognate polysaccharide substrates, we carried out time course interaction experiments at a constant applied force of 500 pN. Results of this experiment showed that the kinetic of the interaction frequency for each of the functionalized AFM dendritips to their polysaccharide target followed a seemingly hyperbolic curve that can be fitted to a Michaelis-Menten model (Fig. 2). Plotting the values according to a double

reciprocal plot, *i.e.* $1/\text{frequency}$ of interaction vs $1/\text{contact time}$ allowed us to estimate a half saturation value ($k_{0.5}$) for each couple of interacting complex. This $k_{0.5}$ could express the time needed to reach 50% of the maximal interaction frequency between the functionalized AFM dendritip and its immobilized polysaccharide target, under the conditions specified in Methods. As shown in Fig. 2, the interaction of the AFM dendritip bearing anti- β -1,3-glucan antibodies and laminarin was the most faster, giving rise to a $k_{0.5}$ 10–20 fold lower than that of interaction between dendritip functionalized with WGA, con A or anti- β -1,6-glucan and its corresponding polysaccharides target (Table S1).

3.3. Force spectroscopy analyses for molecular interaction and determination of unbinding force

By collecting thousands of force-distance curves at a retraction speed of $1 \mu\text{m}\cdot\text{s}^{-1}$ between the immobilized polysaccharide substrates (target) and the AFM dendritips functionalized with their specific probes *i.e.* lectins or antibodies, we could draw a distribution of the adhesion force versus the frequency of the interaction for each probe/target couple (Fig. 3). The representation of these force-distance curves revealed that essentially single adhesion events were recorded, which was spread over a rupture distance ranging from 20 to 200 nm. Also, the adhesion frequency of WGA to the penta-*N*-acetylchitopentaose was significantly higher than that with the other probes to their target (from 30 to 51% vs 5 to 15%), arguing that the association constant (k_{on}) of penta-*N*-acetylchitopentaose substrate to WGA probe is stronger than that of the other probes-polysaccharides substrates. By fitting these distributions to a Gaussian curve, a mean adhesion (or unbinding) force was estimated at 36 ± 19 pN for the interaction between anti- β -1,3-glucan and laminarin, and 83 ± 25 pN for WGA/chitopentaose at a pulling rate of $1 \mu\text{m}\cdot\text{s}^{-1}$. These data were comparable to those reported by El-Kirat-Chatel et al. (2013) using heat-treated yeast cell. Indeed, these authors identified two maximum forces at 42 ± 7 pN and 84 ± 24 pN when probing the surface of the heat-treated cell with an AFM tip bearing a monoclonal anti- β -1,3-glucan antibody. While the

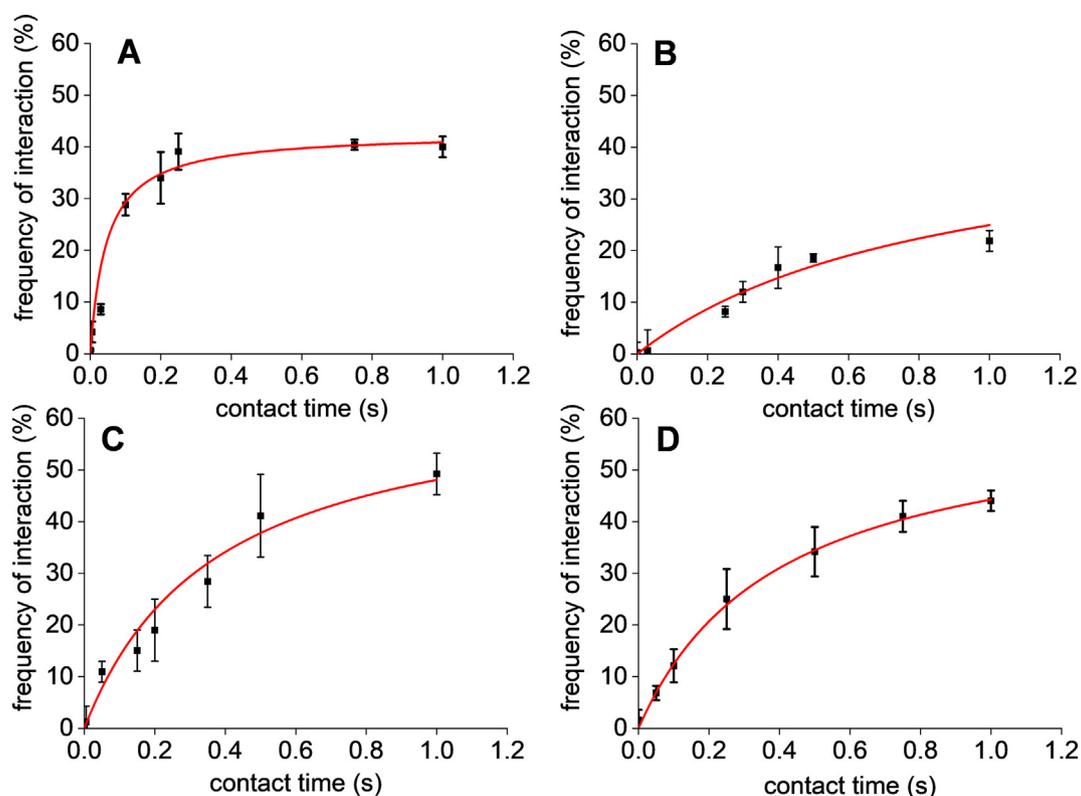


Fig. 2. Effect of the contact time on the interaction between functionalized AFM dendritips and their cognate substrates immobilized on epoxy-activated glass slide. (A) interaction of monoclonal anti- β -1,3-glucan antibody to laminarin-coated slide; (B) interaction of polyclonal anti- β -1,6-glucan antibody to gentiobiose-coated slide, (C) interaction of WGA to penta-*N*-acetylchitopentaose-coated slide and (D) interaction of conA to α -mannan-coated slide.

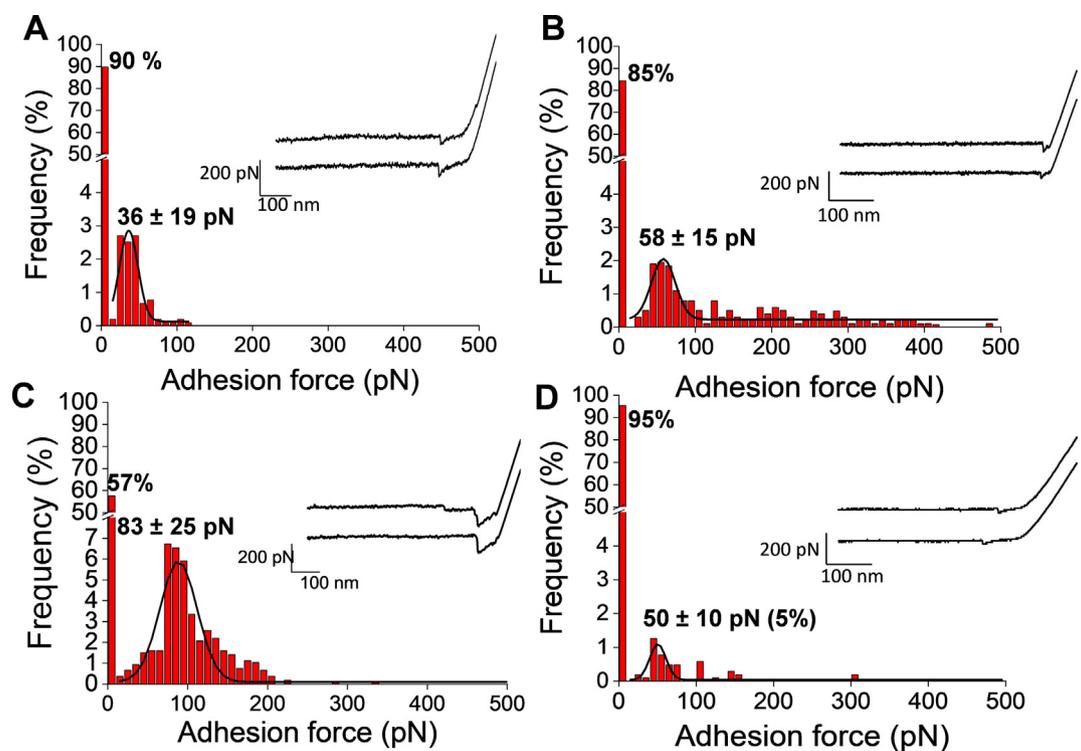


Fig. 3. Determination of unbinding force and frequency of adhesion of functionalized AFM dendritips to their specific substrate coated on epoxy-activated glass slide coated. The frequency, the estimated unbinding force and force-curve distance are shown with AFM tip bearing in (A) monoclonal anti- β -1,3-glucan antibody versus laminarin, in (B) polyclonal anti- β -1,6-glucan antibody vs gentiobiose, in (C) WGA vs penta-*N*-acetylchitopentaose and in (D) conA vs mannan.

value of the first peak is similar to ours, the second peak at 84 pN may be explained by the interaction with two β -glucan chains. The same authors also reported an adhesion force of 54 pN for the interaction of chitin with WGA while a force of 83 pN was estimated under our condition (Fig. 3). This difference may be attributed to the number of GlcNAc units of the chitin chains that can interact with WGA, since it has been reported that the strength of the WGA-GlcNAc interaction is proportional to the number of GlcNAc units (Bains et al., 1992). We also found that the adhesion force of 50 ± 10 pN we determined between ConA-functionalized AFM tip and α -mannan immobilized on an epoxy-activated glass slide was comparable to that reported by Francius et al. (2009) (i.e. 57 ± 19 pN) for a single ConA-mannose interaction. Finally, we report for the first time an unbinding force of 58 ± 15 pN that characterized the interaction of anti- β -1,6-glucan antibodies functionalized AFM dendritip to gentiobiose immobilized on epoxy-glass slide.

3.4. Energy landscape of the intermolecular interactions between cell wall polysaccharides and their specific probe determined by dynamic force spectroscopy

We next exploited DFS to explore the energy landscape of the intermolecular bonds that characterize the interactions between each of the biomolecules covalently fixed on the AFM dendritips and their corresponding polysaccharides target immobilized on epoxy-activated glass slide. According to the theory of Evans (2001), the probability to rupture a molecular bond increases with increasing force at a constant loading rate. However, and in contrast to Evans' law, a non-linear behavior in the force versus loading rate spectra was obtained on a range of loading rate from 100 to 20,000 pN/s. This behavior is illustrated in Fig. 4 where each dot corresponds to the critical loading rate, which is

the loading rate at the exact rupture of the interactions (see also Figs. S4–S7, which illustrate the frequency of interaction and the peak force calculated at different loading rate for each couple of probe-polysaccharides substrates). These nonlinear trends are typical of intermolecular interactions between biological entities and have been modelled by Friddle et al. (2012). Indeed, to account for this non-linearity, the model proposed that the force spectrum reflects two fundamental regimes of bond rupture: (i) a near equilibrium regime between rupture and reforming bond, independently of the loading rate, and (ii) a kinetic regime characterized by a dependence of the rupture force to the log- loading rate ($f \sim \ln r$). Fitting the analytical approximation of the Friddle equation to the dynamic force vs loading rate spectra for each couple of interacting biomolecules within a confidence interval of 99% (Fig. 4), kinetic and thermodynamic parameters of the non-covalent intermolecular bonds between the polysaccharide and their specific probe could be approximated (Table 2). The f_{eq} , corresponding to the lowest force required to break the bond ranged from 35 to 88 pN, which agrees with those commonly obtained for single-bond interaction (Evans, 2001; Friddle et al., 2012). These f_{eq} values were actually close to the adhesion force reported in Fig. 3. With these data and using Eq. (3) (defined in Material and methods), the free energy barrier ΔG needed to create the bonding complex could be calculated. This ΔG for the WGA/chitopentaose was 10 times higher than that for the binding energy of this complex obtained by isothermal titration calorimetry (Bains et al., 1992), whereas it was 2 times lower than that determined by QCM (Coulibaly and Youan, 2014) for the ConA/ α -mannans complex. These discrepancies can be due to difference in pH, temperature and solvation between the assay conditions, which are known to affect the free energy of binding of these ligands. The model can give access to the transition state distance x_t , which corresponds to the width of the energy barrier. The calculated values

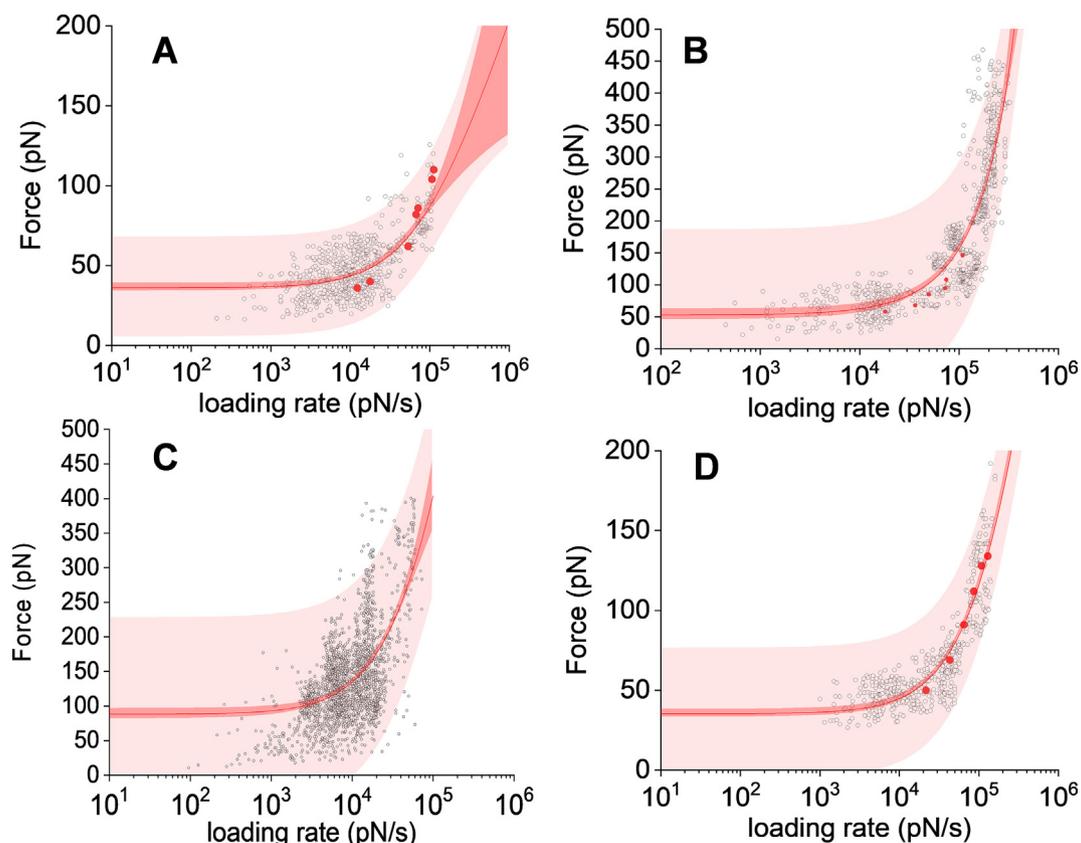


Fig. 4. Dynamic force spectra of functionalized AFM dendritips towards their specific substrates immobilized on epoxide-activated glass slides. The figure reports the rupture force vs the logarithm of the loading rate of an AFM dendritip functionalized with (A) anti- β -1,3-glucan antibody probing a laminarin-coated slide, (B) anti- β -1,6-glucan antibody probing gentiobiose-coated slide, (C) WGA probing a penta-*N*-acetylchitopentaose-coated slide, and (D) conA probing α -mannan-coated slide.

Table 2

Fitted parameters of the energy landscape for the interaction of functionalized AFM dendritips with specific probe to cell wall polysaccharides. The Friddle-Noy-de Yoreo model (Friddle et al., 2012) has been used to determine the energy landscape of the intermolecular interaction between each couple of polysaccharide with its specific probes covalently attached on a AFM dendritip. Details about the nature of each of the parameters is described in the Results section.

Functionalized AFM dendritip	Polysaccharides coated on epoxide-glass slide	x_t (Å)	f_{eq} (pN)	k_{off} (s^{-1})	τ_0 (ms)	ΔG (kcal.mol $^{-1}$)
Anti- β -1,3-glucan antibodies	β -1,3-Laminarin	0.6 ± 0.2	36.2 ± 1.0	732.4 ± 10.4	1.37	-5.3
Anti- β -1,6-glucan antibodies	β -1,6-Gentiobiose	0.3 ± 0.1	52.7 ± 4.1	1046.7 ± 197.6	0.96	-11.0
Concanavalin A	α -Mannan	0.2 ± 0.1	35.3 ± 1.0	569.9 ± 15	1.75	-4.2
Wheat germ agglutinin	β -1,4-Penta- <i>N</i> -acetylchitopentaose	0.1 ± 0.04	88.1 ± 2.9	113.6 ± 10.1	8.81	-55.1

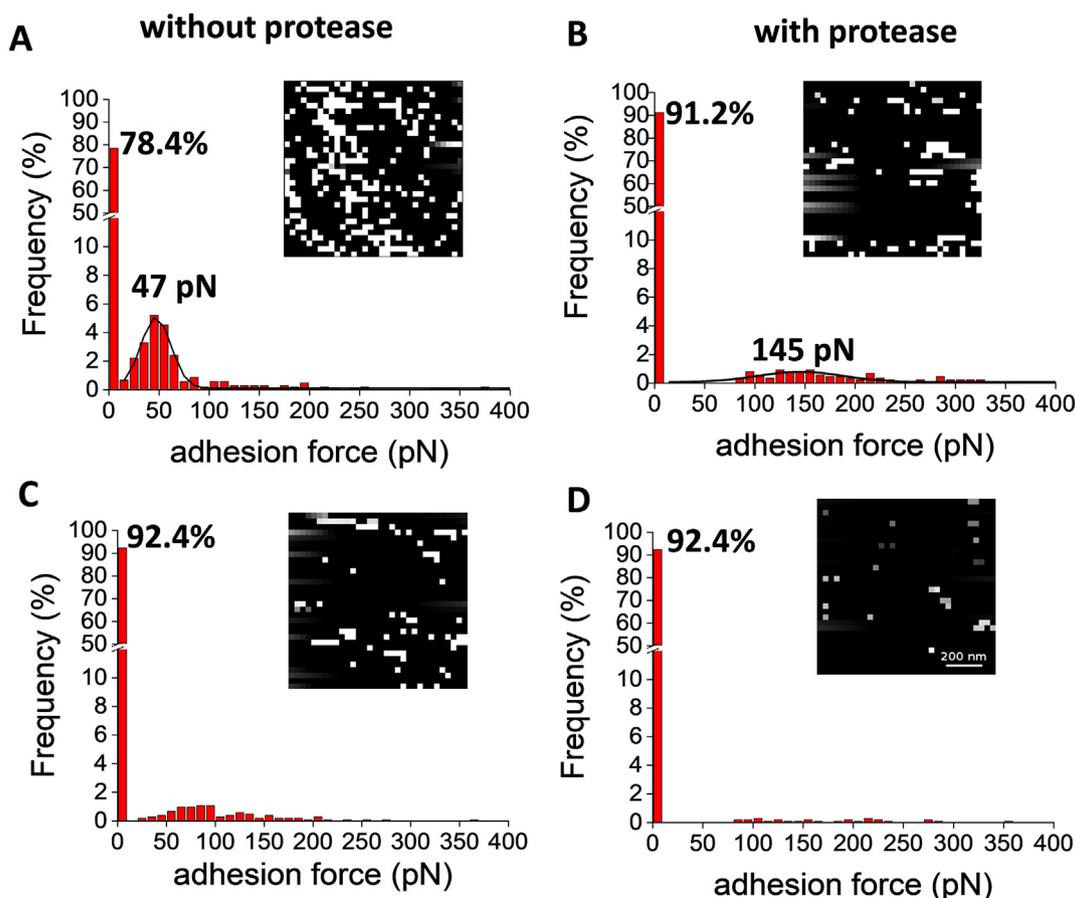


Fig. 5. Molecular mapping of a yeast cell surface using AFM dendritip functionalized with ConA before (A, C) and after (B, D) protease treatment. Inserts are represented the adhesion force map, where white pixels correspond to adhesion events whereas black pixels represent absence of interaction. In (C, D) are shown adhesion force after addition of excess of mannose to the immobilized cells.

were lower than 1 \AA , which is in the range of most of the biomolecular complex interactions (Friddle et al., 2012; Lv et al., 2012). This low distance also indicated that the bond cannot withstand a large deformation when a high force is exerted. We also determined the k_{off} that characterizes the dissociate rate constant of the intermolecular bonds of the complex according to Eq. (1) (In Material and methods section). This value also gave access to the lifetime of the bonds ($\tau_0 = 1/k_{off}$). As indicated in Table 2, the k_{off} of WGA/*N*-acetylchitopentaose complex was 5–8 times lower than that estimated for the 3 other biological complexes (anti- β -1,3-glucan antibodies/laminarin, anti- β -1,6-antibody/gentiobiose and conA/ α -mannan). These data indicated that the intermolecular bonds between WGA and chitin substrate are stronger than those of the other biological complexes, which is further supported by a ΔG that is also higher for the formation of the WGA/penta-*N*-acetylchitopentaose complex (Table 2). To conclude, our dynamic force spectroscopy analysis showed that the intermolecular interactions of cell wall polysaccharides with their specific probes fall into a single-barrier (single bond) model from which the transition state distance x_t ,

unbinding rate k_{off} and equilibrium f_{eq} could be determined.

3.5. Applications of biofunctionalized AFM dendritips to probe surface of living yeast cells after a proteolytic treatment and during budding.

We used our functionalized AFM dendritips to probe the surface of yeast cells prior and after a treatment with a bacterial protease. The rationale of this proteolytic treatment was to remove the outer layer of the cell wall made of mannoproteins to get access to the inner layer composed of β -glucans, as originally reported by Zlotnik et al. (1984). As shown in Fig. 5, the adhesion events recorded by probing the untreated yeast cell with a ConA-functionalized AFM dendritip dropped to less than 8% after addition of 200 mM *D*-mannose, suggesting that the remaining interactions were likely nonspecific. In addition, the mean adhesion force (47 pN) measured for this interaction was comparable to the force of a single ConA-mannose interactions (Francius et al., 2009). Upon digestion of the yeast cell wall with the protease, the frequency of adhesion events dropped to a level similar to that after addition of

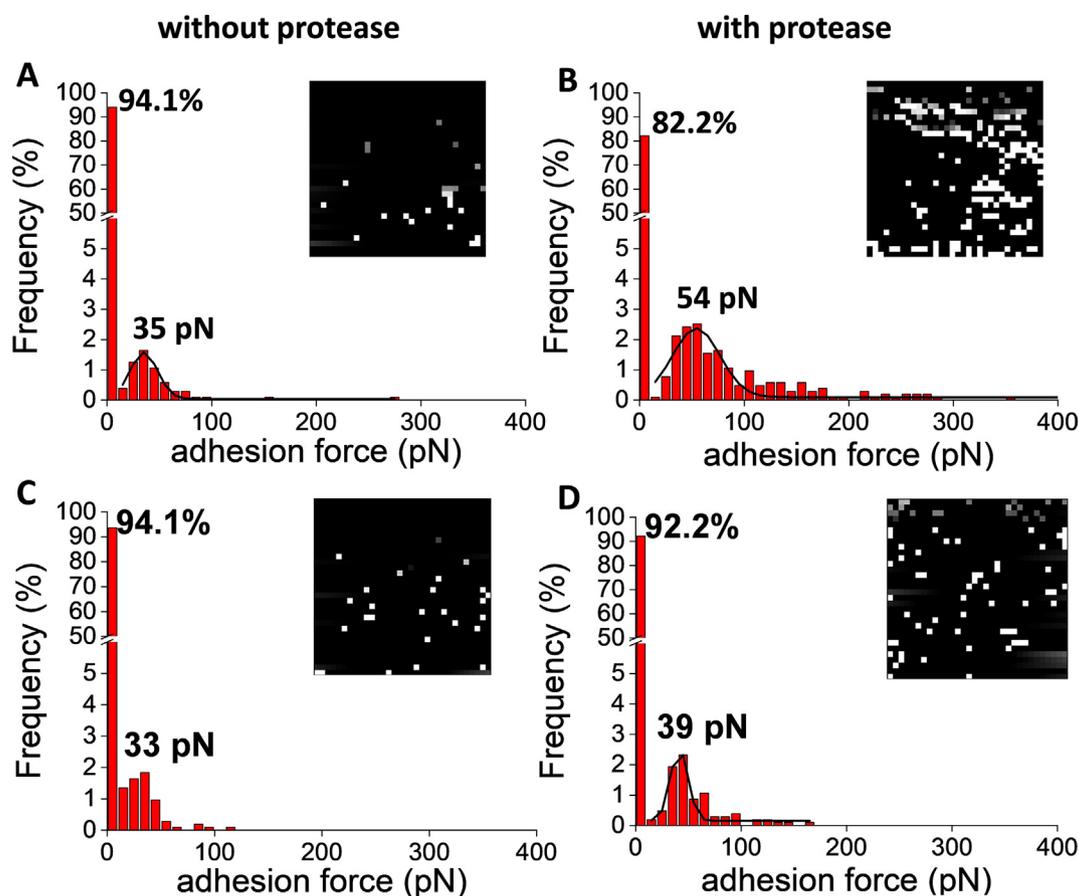


Fig. 6. Molecular mapping of yeast cell surface using an AFM dendritip functionalized with anti- β -1,3-glucan (A, B) and anti- β -1,6-glucan antibodies (C, D) before (A, C) and after (B, D) protease treatment. AFM tip functionalized with either anti- β -1,6-glucan antibody on a yeast cell before (C) and after protease treatment (D). Force maps and histograms of the frequency of adhesion of the tip functionalized on the yeast cell surface in function of the adhesion force detected.

mannose to untreated yeast cell, arguing that the mannoproteins coat of the cell wall has been largely removed by this treatment (Fig. 5B). Addition of excess of mannose prior to AFM interaction resulted in the loss of minor binding events exhibiting adhesion forces in the range of 100–300 pN (Fig. 5D). However, about 8% of unspecific interactions as observed with untreated cells remained.

We next used an AFM dendritip functionalized with anti- β -1,3-glucan antibodies to probe the cell surface before and after the protease treatment (Fig. 6). As expected, the interaction events of an untreated yeast cell with this functionalized dendritip were almost inexistent as shown also on the adhesion force map (< 6%, Fig. 6A), even though there were some minor and most likely unspecific interaction events that were spread in the range of 10 to 100 pN of adhesion force. In contrast, the frequency of adhesion events significantly increased by almost 20% by probing the protease-treated cell with this functionalized dendritip (Fig. 6B). This result confirmed that the cell wall outer layer has been removed and has given access to the inner layer composed of β -1,3-glucan. However, the mean adhesion force peaked at \sim 54 pN instead of 36 pN as determined earlier using laminarin-coated glass slide. As already noticed above, the binding affinity of anti β -1,3-antibody to β -glucan has been reported to be dependent on its molecular mass, degree of branching and structure (Mueller et al., 2000; Strandh et al., 1998). Clearly the molecular size and structure of yeast β -1,3-glucan chains are different from the laminarin which is a short β -glucan chain containing glucose mostly linked in β 1 \rightarrow 3 (Table 1). When probing the cell surface with AFM dendritip functionalized with an anti- β -1,6-glucan antibody, again very few detection events were recorded on the untreated yeast cell (Fig. 6C). However, the frequency of the interaction slightly increased with the protease-treated yeast cells

(Fig. 6D). Since this weak increase of interaction frequency with a mean adhesion force peaking at 39 pN was abolished when excess of glucose was added prior to AFM experiment (data not shown), we could conclude that β -1,6-glucans chains were recognized on the yeast surface after treatment with the protease.

We finally evaluated the pertinence of our AFM dendritip functionalized with WGA by searching the cell surface for bud scars, as they are reported to be highly enriched in chitin (Cabib and Arroyo, 2013). We first employed a simple AFM tip to image a yeast cell, which enabled to localize three bud scars on the surface (Fig. 7B). Using the AFM dendritip bearing WGA as the probe, we mapped the same region and recorded 1024 force -distance curves. We showed that the adhesion force maps overlapped with the localization of the bud scars, which is consistent with the high concentration of chitin in this region (Fig. 7C). In addition, the distribution of the adhesion force ranged from 20 to 200 pN with a maximum at 112 pN. This value is higher than that previously reported (54 pN in El-Kirat-Chatel et al. (2013) and 83 pN, see Fig. 3), which can be explained by the number of GlcNAc units interacting with WGA. Finally, we confirmed that the WGA-functionalized AFM dendritip specifically interacted with *N*-acetylglucosamine moieties of chitin since this interaction was lost upon addition of excess (200 mM) of *N*-acetylglucosamine prior to another force mapping measurements (Fig. 7E & F). Taking these values, we can approximate that the binding of WGA with one GlcNAc unit is approximately 27 pN.

4. Conclusions

In this paper, we report on the development and use of AFM

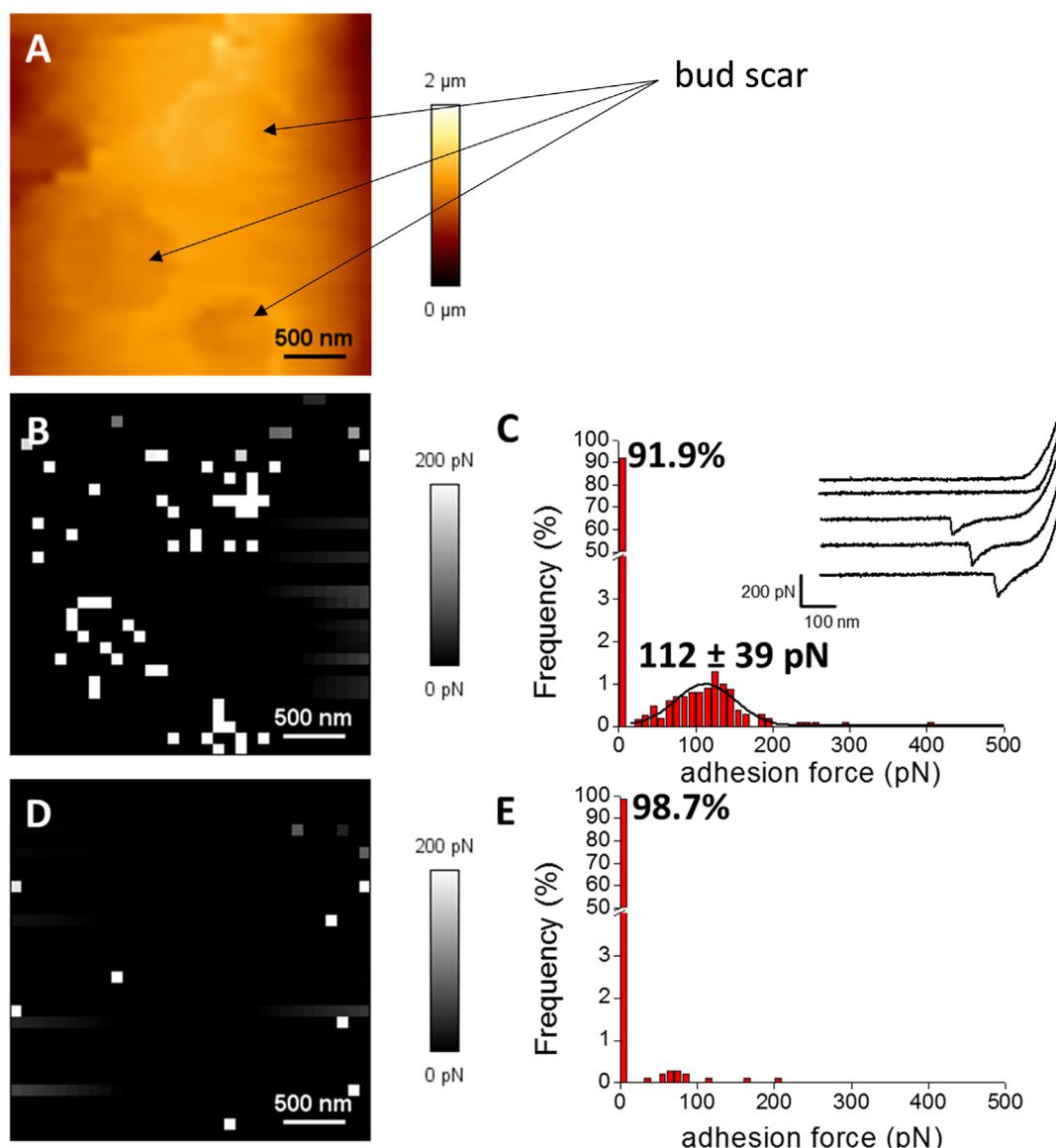


Fig. 7. Molecular mapping of cell surface using an AFM tip functionalized with wheat germ agglutinin. In A is shown the AFM image at high resolution from a single cell of the yeast *S. cerevisiae*, illustrating the presence of 3 bud scars. Panel B & C correspond to the adhesion force maps recorded in PBS buffer between the AFM dendritip functionalized with WGA and the yeast cell surface in the absence (B) or after addition of 200 mM *N*-acetyl glucosamine (C). White pixels indicate specific adhesion events, whereas black pixels represent non-adhesive events. Corresponding adhesion force histograms (for 1024 force-distance curves) of C and E are shown in panel D and F.

dendritips functionalized with anti- β -1,3-glucan and anti- β -1,6-glucan antibodies, wheat germ agglutinin and concanavalin A to probe β -1,3-glucans, β -1,6-glucans, α -mannans and chitin that compose the yeast cell wall. We qualified these functionalized AFM dendritips by determining their specificity towards their cognate cell wall polysaccharides previously immobilized on epoxy glass slides. We then demonstrated that intermolecular interactions between cell wall polysaccharides and their specific molecular probes fall into the single-barrier or single-bond model proposed by Friddle et al. (2012). Using this model, we could extrapolated the transition state distance x_t , the dissociate rate k_{off} and the lowest force (f_{eq}) that is required to break the intermolecular bond of each of the complexes. We then applied these functionalized AFM dendritips to probe the cell surface of yeast before and after a protease treatment, which was aimed at removing the outer-mannoproteins-layer. Accordingly, the frequency of interaction of the surface of the treated cells with an AFM tip bearing ConA dropped to near zero while the adhesion events with an AFM dendritip functionalized with anti- β -1,3-glucan significantly increased. Finally, using the

AFM dendritip coated with WGA, we showed that the adhesion force maps nicely overlapped with bud scars, consistent with abundant level of chitin in these specific structures of the cell wall. Altogether, we report in this paper a new AFM-associated toolbox that can be applied to explore cell surface of any fungal cell made of mannoproteins, glucans and chitin.

Funding

This work was supported in part by a grant from Lallemand Inc. (project Lallwall, n°SAIC2016/048 & SAIC/2018/010).

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

We are grateful Prof Frans Klis from Department of Molecular Biology and Microbial Food Safety, Swammerdam Institute for Life Sciences in University of Amsterdam for providing the anti- β -1,6-glucan antibodies.

Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.tcsuw.2019.100027>.

References

- Aguilar-Uscanga, B., Francois, J.M., 2003. A study of the yeast cell wall composition and structure in response to growth conditions and mode of cultivation. *Lett. Appl. Microbiol.* 37 (3), 268–274.
- Aleem, E., 2013. β -Glucans and their applications in cancer therapy: focus on human studies. *Anticancer Agents Med. Chem.* 13 (5), 709–719.
- Alsteens, D., Beaussart, A., Derclaye, S., El-Kirat-Chatel, S., Park, H.R., Lipke, P.N., Dufrene, Y.F., 2013. Single-cell force spectroscopy of Als-mediated fungal adhesion. *Anal. Methods* 5 (15), 3657–3662. <https://doi.org/10.1039/C3AY40473K>.
- Alsteens, D., Dupres, V., Andre, G., Dufrene, Y.F., 2011. Frontiers in microbial nanoscopy. *Nanomedicine (London)* 6 (2), 395–403. <https://doi.org/10.2217/nmm.10.151>.
- Alsteens, D., Dupres, V., Mc, E.K., Wildling, L., Gruber, H.J., Dufrene, Y.F., 2008. Structure, cell wall elasticity and polysaccharide properties of living yeast cells, as probed by AFM. *Nanotechnology* 19 (38), 384005. <https://doi.org/10.1088/0957-4484/19/38/384005>. S0957-4484(08)72159-7 [pii].
- Bains, G., Lee, R.T., Lee, Y.C., Freire, E., 1992. Microcalorimetric study of wheat germ agglutinin binding to *N*-acetylglucosamine and its oligomers. *Biochemistry* 31 (50), 12624–12628.
- Beaussart, A., Alsteens, D., El-Kirat-Chatel, S., Lipke, P.N., Kucharikova, S., Van, D.P., Dufrene, Y.F., 2012. Single-molecule imaging and functional analysis of Als adhesins and mannans during *Candida albicans* morphogenesis. *ACS Nano* 6 (12), 10950–10964. <https://doi.org/10.1021/nn304505s>.
- Benoit, M., Gabriel, D., Gerisch, G., Gaub, H.E., 2000. Discrete interactions in cell adhesion measured by single-molecule force spectroscopy. *Nat. Cell Biol.* 2 (6), 313–317. <https://doi.org/10.1038/35014000>.
- Bjornham, O., Andersson, M., 2017. Theory for nonlinear dynamic force spectroscopy. *Eur. Biophys. J.* 46 (3), 225–233. <https://doi.org/10.1007/s00249-016-1158-6>.
- Blankenship, J.R., Mitchell, A.P., 2006. How to build a biofilm: a fungal perspective. *Curr. Opin. Microbiol.* 9 (6), 588–594.
- Bohn, J.A., Bemiller, J.N., 1995. (1 \rightarrow 3)- β -D-Glucans as biological response modifiers: a review of structure-functional activity relationships. *Carbohydr. Polym.* 28, 3.
- Bojsen, R.K., Andersen, K.S., Regenb, B., 2012. *Saccharomyces cerevisiae* – a model to uncover molecular mechanisms for yeast biofilm biology. *FEMS Immunol. Med. Microbiol.* 65 (2), 169–182. <https://doi.org/10.1111/j.1574-695X.2012.00943.x>.
- Cabib, E., Arroyo, J., 2013. How carbohydrates sculpt cells: chemical control of morphogenesis in the yeast cell wall. *Nat. Rev. Microbiol.* 11 (9), 648–655.
- Chalier, P., Angot, B., Delteil, D., Doco, T., Gunata, Z., 2007. Interactions between aroma compounds and whole mannoprotein isolated from *Saccharomyces cerevisiae* strains. *Food Chem.* 100, 22–30.
- Che, T.M., Johnson, R.W., Kelley, K.W., Dawson, K.A., Moran, C.A., Pettigrew, J.E., 2012. Effects of mannan oligosaccharide on cytokine secretions by porcine alveolar macrophages and serum cytokine concentrations in nursery pigs. *J. Anim. Sci.* 90 (2), 657–668. <https://doi.org/10.2527/jas.2011-4310>.
- Chopin, L., Formosa, C., Rols, M.P., Duval, R.E., Dague, E., 2013. Imaging living cells surface and quantifying its properties at high resolution using AFM in QI mode. *Micron* 48, 26–33.
- Cleary, J.A., Kelly, G.E., Husband, A.J., 1999. The effect of molecular weight and β -1,6-linkages on priming of macrophage function in mice by (1,3)- β -D-glucan. *Immunol. Cell Biol.* 77 (5), 395–403. <https://doi.org/10.1046/j.1440-1711.1999.00848.x>.
- Clemons, K.V., Danielson, M.E., Michel, K.S., Liu, M., Ottoson, N.C., Leonardo, S.M., Martinez, M., Chen, V., Antonysamy, M.A., Stevens, D.A., 2014. Whole glucan particles as a vaccine against murine aspergillosis. *J. Med. Microbiol.* 63 (Pt 12), 1750–1759. <https://doi.org/10.1099/jmm.0.079681-0>. jmm.0.079681-0 [pii].
- Coulibaly, F.S., Youan, B.B., 2014. Concanavalin A-polysaccharides binding affinity analysis using a quartz crystal microbalance. *Biosens. Bioelectron.* 59, 404–411. <https://doi.org/10.1016/j.bios.2014.03.040>.
- Cuesta, A., Esteban, M.A., Meseguer, J., 2003. In vitro effect of chitin particles on the innate cellular immune system of gilthead seabream (*Sparus aurata* L.). *Fish Shellfish Immunol.* 15 (1), 1–11.
- Da Silva, C.A., Chalouni, C., Williams, A., Hartl, D., Lee, C.G., Elias, J.A., 2009. Chitin is a size-dependent regulator of macrophage TNF and IL-10 production. *J. Immunol.* 182 (6), 3573–3582. <https://doi.org/10.4049/jimmunol.0802113>.
- Dague, E., Bitar, R., Ranchon, H., Durand, F., Yken, H.M., Francois, J.M., 2010. An atomic force microscopy analysis of yeast mutants defective in cell wall architecture. *Yeast* 27 (8), 673–684.
- Dague, E., Gilbert, Y., Verbelen, C., Andre, G., Alsteens, D., Dufrene, Y.F., 2007. Towards a nanoscale view of fungal surfaces. *Yeast* 24 (4), 229–237. <https://doi.org/10.1002/yea.1445>.
- Du, B., Bian, Z., Xu, B., 2014. Skin health promotion effects of natural beta-glucan derived from cereals and microorganisms: a review. *Phytother. Res.* 28 (2), 159–166. <https://doi.org/10.1002/ptr.4963>.
- Dufrene, Y.F., 2008. Towards nanomicrobiology using atomic force microscopy. *Nat. Rev. Microbiol.* 6 (9), 674–680. <https://doi.org/10.1038/nrmicro1948>.
- El-Kirat-Chatel, S., Beaussart, A., 2018. Probing bacterial adhesion at the single-molecule and single-cell levels by AFM-based force spectroscopy. *Methods Mol. Biol.* 1814, 403–414. https://doi.org/10.1007/978-1-4939-8591-3_24.
- El-Kirat-Chatel, S., Beaussart, A., Alsteens, D., Sarazin, A., Jouault, T., Dufrene, Y.F., 2013. Single-molecule analysis of the major glycopolymers of pathogenic and non-pathogenic yeast cells. *Nanoscale* 5 (11), 4855–4863. <https://doi.org/10.1039/c3nr00813d>.
- Esteban, M.A., Cuesta, A., Ortuno, J., Meseguer, J., 2001. Immunomodulatory effects of dietary intake of chitin on gilthead seabream (*Sparus aurata* L.) innate immune system. *Fish Shellfish Immunol.* 11 (4), 303–315. <https://doi.org/10.1006/fsim.2000.0315>.
- Evans, E., 2001. Probing the relation between force-lifetime-and chemistry in single molecular bonds. *Annu. Rev. Biophys. Biomol. Struct.* 30, 105–128. <https://doi.org/10.1146/annurev.biophys.30.1.105>.
- Evans, E., Ritchie, K., 1997. Dynamic strength of molecular adhesion bonds. *Biophys. J.* 72 (4), 1541–1555. [https://doi.org/10.1016/S0006-3495\(97\)78802-7](https://doi.org/10.1016/S0006-3495(97)78802-7).
- Formosa, C., Pillet, F., Schiavone, M., Duval, R.E., Ressler, L., Dague, E., 2015a. Generation of living cell arrays for atomic force microscopy studies. *Nat. Protoc.* 10 (1), 199–204. <https://doi.org/10.1038/nprot.2015.004>.
- Formosa, C., Schiavone, M., Boisrame, A., Richard, M.L., Duval, R.E., Dague, E., 2015b. Multiparametric imaging of adhesive nanodomains at the surface of *Candida albicans* by atomic force microscopy. *Nanomedicine* 11 (1), 57–65. <https://doi.org/10.1016/j.nano.2014.07.008>. S1549-9634(14)00418-3 [pii].
- Francius, G., Alsteens, D., Dupres, V., Leber, S., De, K.S., Vanderleyden, J., Gruber, H.J., Dufrene, Y.F., 2009. Stretching polysaccharides on live cells using single molecule force spectroscopy. *Nat. Protoc.* 4 (6), 939–946. <https://doi.org/10.1038/nprot.2009.65>. nprot.2009.65 [pii].
- Fridde, R.W., Noy, A., De Yoreo, J.J., 2012. Interpreting the widespread nonlinear force spectra of intermolecular bonds. *Proc. Natl. Acad. Sci. U.S.A.* 109 (34), 13573–13578. <https://doi.org/10.1073/pnas.1202946109>.
- Ganner, A., Schatzmayr, G., 2012. Capability of yeast derivatives to adhere enteropathogenic bacteria and to modulate cells of the innate immune system. *Appl. Microbiol. Biotechnol.* 95 (2), 289–297. <https://doi.org/10.1007/s00253-012-4140-y>.
- Hinterdorfer, P., Dufrene, Y.F., 2006. Detection and localization of single molecular recognition events using atomic force microscopy. *Nat. Methods* 3 (5), 347–355.
- Hrmova, M., Fincher, G.B., 1993. Purification and properties of three (1 \rightarrow 3)- β -D-glucanase isoenzymes from young leaves of barley (*Hordeum vulgare*). *Biochem. J.* 289 (Pt 2), 453–461. <https://doi.org/10.1042/bj2890453>.
- Hutter, J.L., Bechhoefer, J., 1993. Calibration of atomic force microscope tips. *Rev. Sci. Instrum.* 64 (7), 1868–1873.
- Jauvert, E., Dague, E., Severac, M., Caminade, A., Ressler, L., Majoral, J.E.T., 2012. Probing single molecule interactions by AFM using biofunctionalized dendritips. *Sens. Actuators B: Chem.* 168, 436–441. <https://doi.org/10.1016/j.snb.2012.04.048>.
- Klis, F.M., Boorsma, A., de Groot, P.W., 2006. Cell wall construction in *Saccharomyces cerevisiae*. *Yeast* 23 (3), 185–202.
- Kristiansen, A., Nysaeter, A., Grasdalen, H., Varum, K.M., 1999. Quantitative studies of the binding of wheat germ agglutinin (WGA) to chitin-oligosaccharides and partially *N*-acetylated chitosans suggest inequivalence of binding sites. *Carbohydr. Polym.* 38 (1), 23. [https://doi.org/10.1016/S0144-8617\(98\)00106-4](https://doi.org/10.1016/S0144-8617(98)00106-4).
- Kulicke, W.M., Lettau, A.L., Thielking, H., 1997. Correlation between immunological activity, molar mass, and molecular structure of different (1 \rightarrow 3)- β -D-glucans. *Carbohydr. Res.* 297 (2), 135–143.
- Lee, C.G., Da Silva, C.A., Lee, J.Y., Hartl, D., Elias, J.A., 2008. Chitin regulation of immune responses: an old molecule with new roles. *Curr. Opin. Immunol.* 20 (6), 684–689. <https://doi.org/10.1016/j.coi.2008.10.002>.
- Legentil, L., Paris, F., Ballet, C., Trouvelot, S., Daire, X., Vetricka, V., Ferrieres, V., 2015. Molecular interactions of β -(1 \rightarrow 3)-glucans with their receptors. *Molecules* 20 (6), 9745–9766. <https://doi.org/10.3390/molecules20069745>.
- Lotan, R., Sharon, N., 1973. The fluorescence of wheat germ agglutinin and of its complexes with saccharides. *Biochem. Biophys. Res. Commun.* 55 (4), 1340–1346.
- Lv, Z., Wang, J., Chen, G., 2012. Exploring the energy profile of human IgG/rat anti-human IgG interactions by dynamic force spectroscopy. *Protein J.* 31 (5), 425–431. <https://doi.org/10.1007/s10930-012-9419-4>.
- Mekoue Nguela, J., Poncet-Legrand, C., Siczekowski, N., Vernhet, A., 2016. Interactions of grape tannins and wine polyphenols with a yeast protein extract, mannoproteins and β -glucan. *Food Chem.* 210, 671–682. <https://doi.org/10.1016/j.foodchem.2016.04.050>.
- Merkel, R., Nassoy, P., Leung, A., Ritchie, K., Evans, E., 1999. Energy landscapes of receptor-ligand bonds explored with dynamic force spectroscopy. *Nature* 397 (6714), 50–53. <https://doi.org/10.1038/16219>.
- Montijn, R.C., van Rinsum, J., van Schagen, F.A., Klis, F.M., 1994. Glucmannoproteins in the cell wall of *Saccharomyces cerevisiae* contain a novel type of carbohydrate side chain. *J. Biol. Chem.* 269 (30), 19338–19342.
- Mueller, A., Raptis, J., Rice, P.J., Kalbfleisch, J.H., Stout, R.D., Ensley, H.E., Browder, W., Williams, D.L., 2000. The influence of glucan polymer structure and solution conformation on binding to (1 \rightarrow 3)- β -D-glucan receptors in a human monocyte-like cell line. *Glycobiology* 10 (4), 339–346.
- Muller, D.J., Dufrene, Y.F., 2008. Atomic force microscopy as a multifunctional molecular toolbox in nanobiotechnology. *Nat. Nanotechnol.* 3 (5), 261–269. <https://doi.org/10.1038/nnnano.2008.100>.

- 1038/nano.2008.100.
- Nguyen, T.H., Fleet, G.H., Rogers, P.L., 1998. Composition of the cell walls of several yeast species. *Appl. Microbiol. Biotechnol.* 50 (2), 206–212.
- Orlean, P., 2012. Architecture and biosynthesis of the *Saccharomyces cerevisiae* cell wall. *Genetics* 192 (3), 775–818.
- Osumi, M., Baba, M., Yamaguchi, H., 1984. *Electron Microscopical Study on the Biosynthesis and Assembly Cell Wall Components, Cell wall Synthesis and Autolysis*. Elsevier, Amsterdam, pp. 137–142.
- Pillet, F., Lemonier, S., Schiavone, M., Formosa, C., Martin-Yken, H., Francois, J.M., Dague, E., 2014. Uncovering by atomic force microscopy of an original circular structure at the yeast cell surface in response to heat shock. *BMC Biol.* 12 (6). <https://doi.org/10.1186/1741-7007-12-6>. 1741-7007-12-6 [pii].
- Pourabedin, M., Chen, Q., Yang, M., Zhao, X., 2017. Mannan- and xylooligosaccharides modulate caecal microbiota and expression of inflammatory-related cytokines and reduce caecal *Salmonella* Enteritidis colonisation in young chickens. *FEMS Microbiol. Ecol.* 93 (1). <https://doi.org/10.1093/femsec/fiw226>.
- Pourabedin, M., Zhao, X., 2015. Prebiotics and gut microbiota in chickens. *FEMS Microbiol. Lett.* 362 (15), fnv122. <https://doi.org/10.1093/femsle/fnv122>.
- Pradelles, R., Alexandre, H., Ortiz-Julien, A., Chassagne, D., 2008. Effects of yeast cell-wall characteristics on 4-ethylphenol sorption capacity in model wine. *J. Agric. Food Chem.* 56 (24), 11854–11861. <https://doi.org/10.1021/jf802170p>.
- Raa, J., 2015. Immune modulation by non-digestible and non-absorbable beta-1,3/1,6-glucan. *Microb. Ecol. Health Dis.* 26, 27824. <https://doi.org/10.3402/mehd.v26.27824>.
- Roy, R., Katzenellenboen, E., Jennings, H.J., 1984. Improved procedures for the conjugation of oligosaccharides to protein by reductive amination. *Can. J. Biochem. Cell Biol.* 62 (5), 270.
- Schiavone, M., Formosa-Dague, C., Elstein, C., Teste, M.A., Martin-Yken, H., de Morais, M.A.J., Dague, E., Francois, J.M., 2016. Evidence for a role for the plasma membrane in the nanomechanical properties of the cell wall as revealed by an atomic force microscopy study of the response of *Saccharomyces cerevisiae* to ethanol stress. *Appl. Environ. Microbiol.* 82 (15), 4789–4801. <https://doi.org/10.1128/AEM.01213-16>. AEM.01213-16 [pii].
- Schiavone, M., Siczkowski, N., Castex, M., Dague, E., Marie Francois, J., 2015. Effects of the strain background and autolysis process on the composition and biophysical properties of the cell wall from two different industrial yeasts. *FEMS Yeast Res.* 15 (2). <https://doi.org/10.1093/femsyr/fou012>.
- Schwefel, D., Maierhofer, C., Beck, J.G., Seeberger, S., Diederichs, K., Moller, H.M., Welte, W., Wittmann, V., 2010. Structural basis of multivalent binding to wheat germ agglutinin. *J. Am. Chem. Soc.* 132 (25), 8704–8719. <https://doi.org/10.1021/ja101646k>.
- Strandh, M., Persson, B., Roos, H., Ohlson, S., 1998. Studies of interactions with weak affinities and low-molecular-weight compounds using surface plasmon resonance technology. *J. Mol. Recognit.* 11 (1–6), 188–190. [https://doi.org/10.1002/\(SICI\)1099-1352\(199812\)11:1/6<188::AID-JMR420>3.0.CO;2-O](https://doi.org/10.1002/(SICI)1099-1352(199812)11:1/6<188::AID-JMR420>3.0.CO;2-O).
- Takeuchi, O.M.T., Taninaka, A., Tanaka, K., Cho, D., Fujita, M., Yasuda, S., Jarvis, S.P., Shigekawa, H., 2006. Dynamic-force spectroscopy measurement with precise force control using atomic-force microscopy probe. *J. Appl. Phys.* 100 (7) 074315-074311.
- Wright, C.S., Gavilanes, F., Peterson, D.L., 1984. Primary structure of wheat germ agglutinin isolectin 2. Peptide order deduced from X-ray structure. *Biochemistry* 23 (2), 280–287.
- Zlotnik, H., Fernandez, M.P., Bowers, B., Cabib, E., 1984. *Saccharomyces cerevisiae* mannanoproteins form an external cell wall layer that determines wall porosity. *J. Bacteriol.* 159 (3), 1018–1026.