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Nonlinear plasmonic nanohybrids as probes for multimodal cell imaging and potential phototherapeutic agents

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Abstract

Silicon carbide (SiC) nanoparticles were coated with gold nanorods to prepare nonlinear plasmonic nanohybrids. Their structural properties were evaluated by Transmission Electron Microscopy. Nonlinear optical properties of the two components (second harmonic generation for SiC nanoparticles, two-photon luminescence for gold nanorods) were simultaneously found into nanohybrids and spatially colocalized. Photothermal effect was observed for nanohybrids. The cytotoxicity of nanoparticles, nanorods and nanohybrids towards healthy and cancer cell lines was then evaluated. SiC nanoparticles and nanohybrids showed no cytotoxicity for healthy

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3 cells while gold nanorods were found to exhibit an increasing toxicity with increasing
4 concentrations, which was related to the presence of CTAB surfactant on their surface. Finally,
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6 nanohybrids exhibit toxicity for cancer cells and this effect was increased under laser irradiation
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8 at 808 nm, which matches the plasmon resonance of gold nanorods.
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12 13 **1. Introduction**

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15 The World Health Organization recently reported that cancer is the second leading cause of
16 death worldwide.^[1] Therefore, tremendous efforts have been devoted to novel cancer diagnostic
17 and therapeutic approaches. Among other, tumor cells apoptosis can be triggered by the local
18 increase of the temperature. Such local heating can be achieved by photothermia, the conversion
19 of light into heat. Gold nanoparticles have attracted attention as they can be easily synthesized,
20 their surface can be engineered for tumor cell targeting, and their optical properties can be tuned
21 by controlling their size and shape.^[2] Light absorption near the plasmon band resonance can be
22 up to 4-6 times higher than organic dyes.^[3,4] Moreover, upon light absorption, the incoming
23 energy is transformed into heat and bubbles are formed leading to cellular apoptosis due to
24 thermal and mechanical stress.^[5] However, gold nanospheres suffer from an absorption band
25 located at 520 nm, far from the biological transparency window which is located in the near
26 infrared (NIR) spectral region (650-1050 nm), low heating efficiency and low cavitation
27 bubbles formation threshold.^[6] To overcome these shortcomings, efforts have been made to
28 develop gold nanorods, nanoshells or nanoclusters of different shapes (nanostars, nanocages)
29 for which the plasmon resonance can be shifted toward the NIR with a narrow and intense
30 absorption band. In particular, for gold nanorods, the variation of aspect ratio (length to
31 diameter ratio) allows a fine tuning of plasmon resonance in this spectral range. Besides, El
32 Fayed *et al* demonstrated using Mie based calculations that gold nanorods absorption
33 coefficient in the NIR region is higher than for gold nanoshells.^[7] Finally, the gold nanorods
34 cytotoxicity seems to be related to the presence of hexadecyltrimethylammonium bromide
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3 (CTAB) which is commonly used for their synthesis.^[2] Alternative surfactant or capping of the
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5 nanorods with a polymeric layer has been proposed as possible way to overcome this toxicity.^[8]
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8 Another advantage of gold nanostructures such as gold nanorods is that they can exhibit two-
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10 photon luminescence (TPL), which allows cell imaging under excitation in the NIR region.^[9]
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12 However, the signal is generally broad and not specific as many other biological components,
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14 such as proteins and co-factors possess overlapping emission bands.^[10] Thus, sensitive
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16 detection and high contrast imaging can be reached only if gold nanorods are coupled to other
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18 nanostructures. Nanoparticles based on non-centrosymmetric crystalline materials (also called
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20 harmonic nanoparticles) exhibit an intense and sharp second harmonic generation (SHG) signal
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22 in a wide range of excitation wavelengths, that can be used for cell imaging and tracking^[11,12].
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24 Moreover, these materials generally exhibit low cytotoxicity at the nanoscale^[13]. In particular,
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26 silicon carbide (SiC) nanoparticles were recently used for cancer cells targeting^[14].
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32 Coupling gold nanorods to other nanostructures was mainly achieved through Coulombic
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34 interactions. Recently, gold nanorods were immobilized on silica-coated iron oxide
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36 nanoparticles by polyethylenimine/poly(styrenesulfonate)^[15] and CTAB/silica^[16] interactions
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38 allowing for the fabrication of multifunctional nanohybrids featuring photothermal properties
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40 and magnetic properties. Direct assembly between iron oxide nanoparticles and gold nanorods
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42 was also achieved through polydopamine/disulphide-modified poly(2-dimethyl amino)ethyl
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44 methacrylate interaction^[17]. In this case, the objective was to combine photothermal/gene
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46 therapy using plasmid DNA. Gold nanorods were also coupled to upconversion nanoparticles
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48 via poly-L-lysine/silica^[18] and chitosan/aminosilane^[19] interactions for phototherapeutic
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50 applications. Polyethyleneglycol (PEG)-based linkers were also used to immobilize gold
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52 nanorods onto various kinds of nanoparticles. For instance, gold nanorods were linked to
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54 mesoporous silica nanocarriers for light triggered photothermal and photodynamic therapy^[20].
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59 In another report, Prussian-blue-functionalized lanthanide-ion-doped inorganic nanoparticles
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3 were coated with gold nanorods using a PEG-thiol linker for targeted photothermal therapy on
4 breast cancer cells [21]. Hu *et al* prepared cellulose-gold nanorods nanohybrids with lipoic acid
5 as a linker and used them for photoacoustic imaging and combined photothermal/gene therapy
6 [22].
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12 This paper is focused on the synthesis of nanohybrids composed of gold nanorods immobilized
13 on a non-centrosymmetric core (SiC) exhibiting SHG signal. Positively-charged gold nanorods
14 were synthesized using the standard CTAB-based method and then simply immobilized by
15 electrostatic self-assembly on the negatively-charged SiC cores. The nonlinear optical
16 properties (TPL and SHG) and photothermal effect of nanohybrids, gold nanorods and non-
17 centrosymmetric cores were compared. Then, nanoparticles incubated with healthy cells were
18 observed using multiphoton microscopy and their cytotoxicity was investigated. Finally, the
19 phototherapeutic properties of nanoparticles incubated with cancer cells were studied under
20 laser irradiation at 808 nm.
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33 34 35 **2. Experimental**

36 37 **2.1. Materials**

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39 SiC-3C nanoparticles were purchased from KM Labs (Ukraine). They were produced by
40 carbothermal reduction of amorphous silica in presence of sucrose and citric acid at 240°C.
41 Then the obtained powder was heated at 1420°C under argon. All other chemicals were
42 obtained from Sigma-Aldrich and used as received. Milli-Q water (18.2 MΩ cm) was used in
43 all the preparations.
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50 51 **2.2. Initial stabilization of SiC nanoparticles**

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53 The initial powder of SiC nanoparticles was heated at 600°C, in air, during 1 h. After cooling,
54 the powder was dispersed at a concentration of 80 mg mL⁻¹ in an aqueous KOH 9 M solution
55 and the dispersion was sonicated during 1 h. Then, nanoparticles were washed three times by
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3 centrifugation (1000g, 1-5 min) and redispersed in water. After redispersion, nanoparticles were
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5 let for sedimentation during 24 h. The resulting precipitate was discarded and the supernatant,
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7 containing a concentration of around 8 mg mL⁻¹ in SiC nanoparticles, was kept for
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9 functionalization. The resulting nanoparticles suspensions were stable for several days.
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13 2.3. Synthesis of Au nanorods

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15 Nanorods were prepared using a previously reported protocol [23]. Gold seeds were synthesized
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17 by mixing 629 μL of aqueous CTAB solution (150 mM), 216 μL of deionized water and 55.1
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19 μL of aqueous HAuCl₄ solution (4.28 mM). Then, 100 μL of NaBH₄ aqueous solution was
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21 added and the obtained solution was left to react during 3 h at 25°C. Next, the growth solution
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23 was prepared by mixing 9.87 mL of aqueous CTAB solution (150 mM), 2.2 mL of deionized
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25 water and 1.73 mL of aqueous HAuCl₄ solution (4.28 mM). Then, 200 μL of an aqueous AgNO₃
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27 solution (7.4 mM) were added, to obtain nanorods with an aspect ratio of 4.4. The obtained
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29 solution was stirred slowly during 2 min and 1 mL of aqueous solution of ascorbic acid (8.18
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31 mM) was then added. After a slow stirring of 1 min, 25 μL of the previously prepared gold seed
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33 suspension was added. The solution was stirred for 1 min and let to react, at 30°C, overnight.
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35 Centrifugation at 6700g during 5 min was used to stop the reaction and remove reactants and
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37 by-products from nanorod suspension after reaction. Finally, a shape-selective purification was
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39 applied to eliminate nanospheres from nanorods suspensions according to a previously reported
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41 procedure.^[24]
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48 2.4. Elaboration of SiC@Au nanohybrids

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50 500 μL of Au nanorods suspension (100 mg/L in water) were centrifuged at 6700g during 6
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52 min for three times to eliminate the excess of CTAB. They were redispersed in 500 μL of water.
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54 Then 30 μL or 120 μL of SiC nanoparticles suspension (2.67 g/L in water) were added to the
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56 Au nanorods suspension. The mixture was stirred at 500 rpm overnight. Finally, the
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58 nanohybrids suspension was centrifuged at 6700g during 2 min and redispersed in 500 μL of
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3 water. This short centrifugation time was necessary to keep free nanorods in the supernatant.
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5 Then, two populations of nanohybrids were identified: a first population that sedimented within
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7 a few minutes and a second one that remained in suspension for several days. Only the second
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9 population (in the supernatant) was kept for further measurements after natural sedimentation
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11 of the first one.
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14 15 2.5. Cell incubation and irradiation with nanoparticles and nanohybrids

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17 Cell cultures of 3T3-L1 mouse fibroblasts (American Type Culture Collection, Manassas, VA,
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19 USA) and human hepatocarcinoma-derived HuH7 cells (European Collection of Cell Cultures,
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21 Salisbury, UK) were initially grown in Dulbecco's modified Eagle's medium (DMEM)
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23 containing 4.5 g L⁻¹ glucose supplemented with 10% newborn calf serum (American Type
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25 Culture Collection, Manassas, VA, USA), 100 IU penicillin, and 100 µg streptomycin at 37°C
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27 in a water-saturated atmosphere with 5% CO₂, in a Heraeus incubator. A 96-well microplate
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29 with wells of 8 mm in diameter was used for incubation. Cells were seeded at 2500 cells per
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31 well. Nanoparticles were introduced at concentrations ranging from 1.25 to 20 µg mL⁻¹. After
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33 2 h of incubation with HuH7 cells, half of the wells were irradiated with a continuous-wave
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35 Newport laser emitting at a wavelength of 808 nm during 12 min using an optical fiber of 200
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37 µm in diameter and a numerical aperture of 0.22. The measured surface power at the apex of
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39 the fiber was 40 mW cm⁻². An empty microplate was put on the top of the microplate containing
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41 cells incubated with nanoparticles. Then the optical fiber of the laser beam was put inside the
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43 wells of the empty microplate for irradiation. During irradiation, the microplate was kept under
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45 a temperature of 37°C.
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52 53 2.6. Photothermal properties evaluation

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55 Drops of 3 µL of nanoparticles suspensions were deposited on a microstructured hydrophobic
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57 glass slide. Then the drops were successively irradiated with the 808 nm continuous-wave laser
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3 via the optical fiber. The heat generation was recorded with a ThermoVision A20M infrared
4 camera from Flir Systems in a top view of the glass slide.
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7 8 2.7. Toxicity assay 9

10 Cell proliferation and/or survival was monitored with the xCELLigence Real-time Cell
11 Analyser (RTCA) System (ACEA Biosciences, Inc., San Diego, USA), which allows label-free
12 and non-destructive monitoring changes of cell number, viability, morphology and quality of
13 cell attachment by measurement of cell to electrode responses of cells seeded in E96-well plates
14 manufactured with integrated microelectronic sensor arrays. RTCA system measures real-time
15 impedance variations that are then converted in cell surface occupancy, i.e. cell index, taking
16 into account cell number, cell size, and adhesion force. When cell index reaches a sufficient
17 value (typically between 1 and 2), nanoparticles are introduced and incubated with cells during
18 24 h using the protocol detailed in the previous section. During this 24 h duration, the colloidal
19 stability of nanoparticles suspensions was ensured. Cell indexes are normalized at the
20 introduction of nanoparticles in order to facilitate the comparison between the different
21 conditions. Then nanoparticles are removed by replacing the cell culture medium by a fresh
22 one. The impedance measurements are maintained during 24 h in order to study the influence
23 of washing.
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43 2.8. Multiphoton imaging 44

45 Nanoparticles drop casted on a substrate or in fixed cells were observed with a Nikon
46 multiphoton inverted microscope (A1R-MP) coupled to a Mai-Tai tunable Ti:sapphire
47 oscillator (Spectra-Physics; 100 fs, 80 MHz, 700–1000 nm). A plan apochromat 20× water-
48 immersion objective with a 0.75 numerical aperture was used to focus the excitation laser at
49 790 nm and epi-collect the two-photon luminescence (TPL) and second harmonic generation
50 (SHG) emission. The collected signals were processed by non-descanned detectors with using
51 band-pass filters (SemRock) of 395/11 nm for SHG and 607/70 nm for two-photon
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luminescence. Alternatively the epi-collected signal were sent through a fiber to a spectral detection unit allowing the simultaneous detection of up to 32 independent channels.

2.9. Other characterization techniques

Zeta potential was evaluated using a Zetasizer apparatus (Malvern Instruments). UV-Vis absorption spectra were collected in a quartz cell with 1.0 cm path length using a SAFAS-UV mc2 double-beam spectrophotometer. TEM images were realized with a JEOL 2100HT microscope working at 200 kV. For TEM studies, 2 μ L of the diluted dispersion of nanoparticles was deposited onto a carbon holey grid (Ted Pella, Inc.). Images were analyzed with ImageJ 1.46r software.

3. Results and discussion

3.1. Structural properties of SiC@Au nanohybrids

Gold nanorods were first characterized using UV-Vis absorption spectroscopy and TEM (Figure 1).

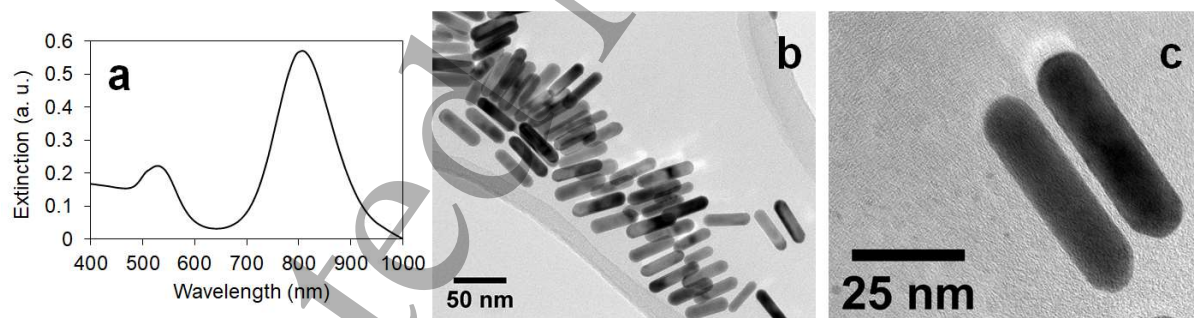
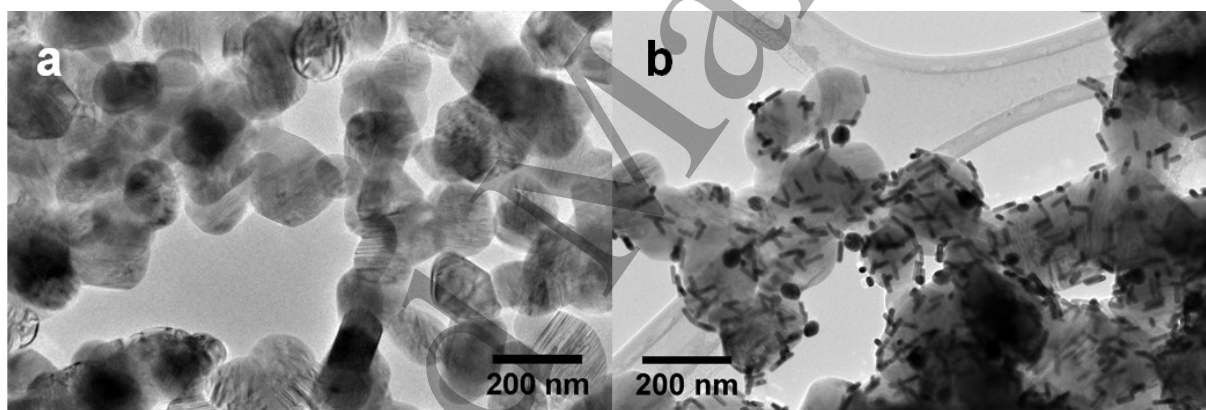


Figure 1. Extinction spectrum (a) and TEM images (b and c) of Au nanorods.

Au nanorods exhibited a high intensity absorption band centered at 808 nm and a low intensity absorption band around 529 nm (Figure 1a). These two bands can be attributed to the longitudinal plasmon resonance mode of nanorods and to a combination of transverse plasmon resonance mode of nanorods and plasmon resonance of remaining nanospheres (less than 7%

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3 after shape-selective purification^[24]), respectively. This was confirmed by TEM images
4 (Figures 1b and 1c) in which a mean length of 47.6 ± 3.9 nm and a mean diameter of 12.3 ± 1.2
5 nm were determined (aspect ratio: 3.9). This aspect ratio is in agreement with the position of
6 the measured longitudinal plasmon resonance band of gold nanorods according to previous
7 works ^[25,26]. As shown in Supporting Information (Figure S2), SiC nanoparticles exhibited a
8 zeta potential around -50 mV. After their synthesis, Au nanorods were coated with CTAB, a
9 positively charged tensio-active molecule, thus they naturally exhibited a positive zeta potential
10 of +55 mV. Due to the highly negative value of zeta potential for SiC nanoparticles, SiC@Au
11 nano hybrids were elaborated through the ionic interaction between the oppositely charged
12 surfaces of SiC and as-synthesized Au nanorods coated with CTAB. Figure 2 showed TEM
13 images of initial SiC nanoparticles (Figure 2a) compared to SiC@Au nano hybrids (Figure 2b).
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43 **Figure 2.** TEM images of SiC nanoparticles (a) and SiC@Au nano hybrids (b).
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46 As shown in Figure 2a, SiC nanoparticles exhibited a diameter, ranging from 80 to 220 nm,
47 centered around 150 nm. The success of Au nanorods immobilization onto SiC nanoparticles
48 was evidenced by dark spots (Figure 2b) corresponding to Au nanorods onto SiC@Au
49 nano hybrids. No free Au nanorods were observed. The clustering effect observed on TEM
50 images was mainly due to the drying of aqueous suspensions of nanoparticles onto hydrophobic
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TEM grids (coated with a carbon membrane). It did not reflect the behaviour of nanoparticles in suspension, as colloidal stability was ensured for several days.

3.2. Nonlinear optical properties of SiC@Au

Then, the nonlinear optical properties of Au nanorods, SiC nanoparticles and SiC@Au nano hybrids were studied. Figure 3 shows the nonlinear optical responses from Au nanorods, SiC nanoparticles and SiC@Au nano hybrids under 820 nm excitation.

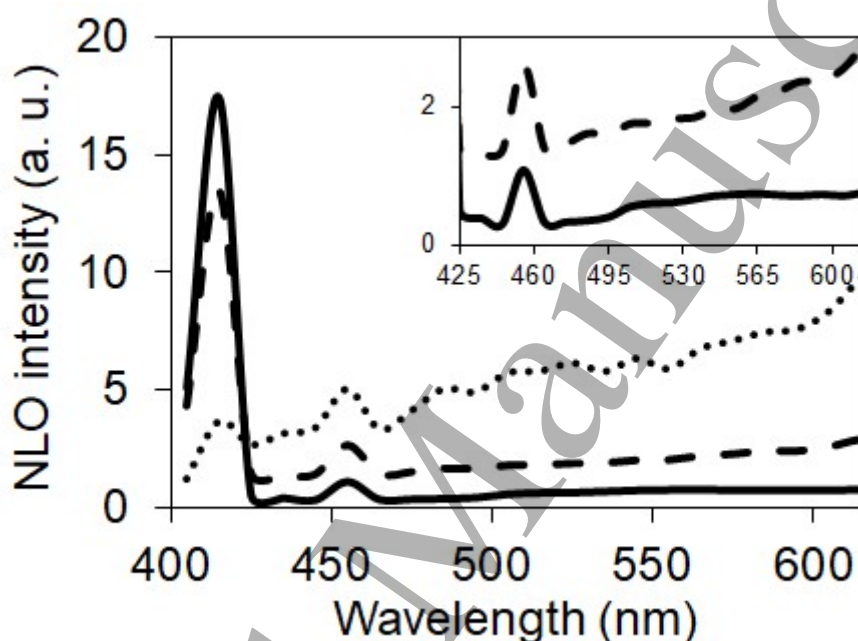
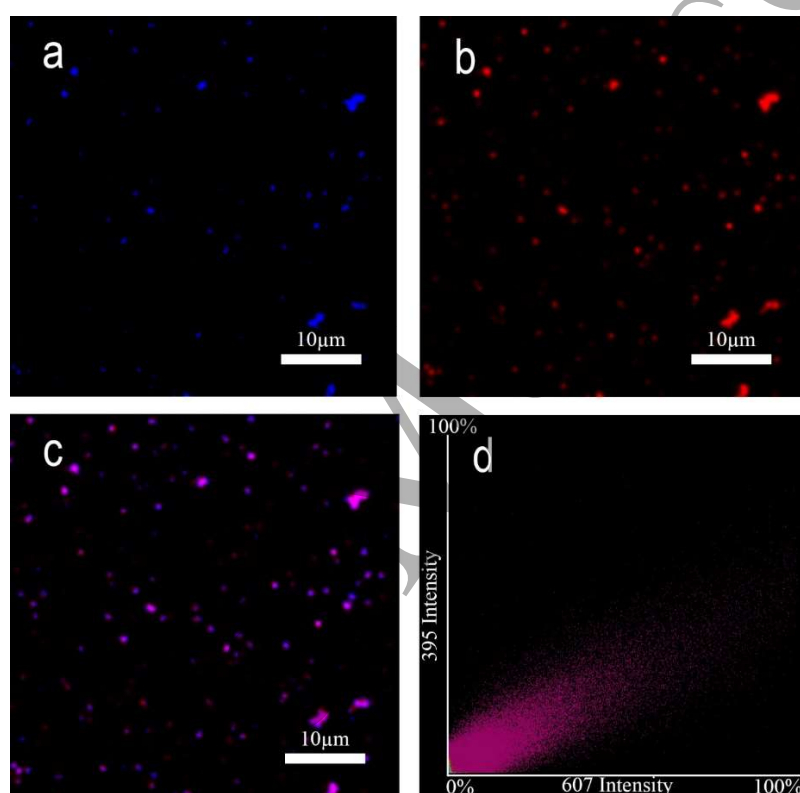


Figure 3. Nonlinear optical (NLO) response from Au nanorods (dot line), SiC nanoparticles (solid line) and SiC@Au nano hybrids (dashed line) under 820 nm excitation. Insert: magnification on the 425-615 nm region.

On one hand, it reveals a strong SHG response at 410 nm for SiC nanoparticles and SiC@Au nano hybrids while this peak is absent for Au nanorods. The lower SHG peak observed for SiC@Au nano hybrids is due to our preparation process. As presented in Section 2.4, for the as-prepared SiC@Au nano hybrids, two populations were identified: a first population that sedimented in few minutes and a second one that stayed in suspension for several days. Only the second population (in the supernatant) was kept for nonlinear optical measurements after

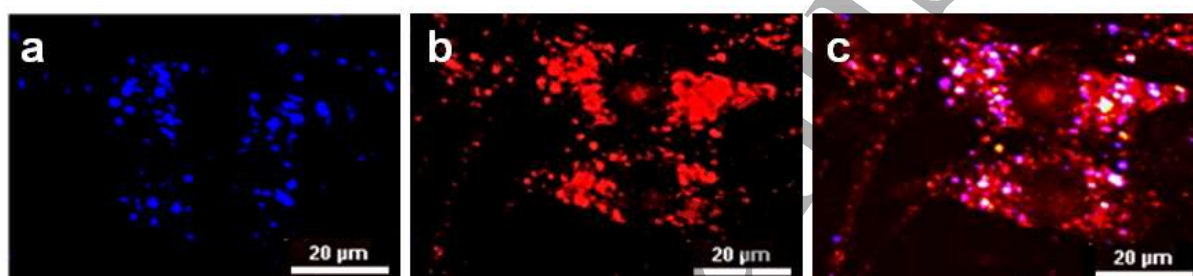
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3 natural sedimentation of the first one. Thus the concentration of SiC in SiC@Au nanohybrids
4 is lower than the initial concentration in SiC suspension. This could explain the lower SHG
5 signal. On the other hand, Au nanorods exhibit a non negligible signal between 420 and 610
6 nm, such as SiC@Au nanohybrids (Figure 3, insert). This signal increases with increasing
7 detection wavelength, which proves that it is due to the TPL emission of Au nanorods. The low
8 intensity peak detected at 455 nm for all samples is due to an artefact of the instrument. SiC@Au
9 nanohybrids were then observed under multiphoton imaging (Figure 4).
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46 **Figure 4.** SiC@Au nanohybrids observed under multiphoton imaging (excitation wavelength:
47 790 nm): detection at 395 nm (a), detection at 607 nm (b) and all detection channels overlaid
48 (c). Colocalization plot between 395 nm and 607 nm detection channels (d).
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53 As shown in Figures 4a and 4b, nanohybrids can be detected at both 395 nm and 607 nm. The
54 detection at 395 nm corresponds to the half of excitation wavelength thus to SHG signal from
55 SiC nanoparticles while at 607 nm, the TPL signal of Au nanorods is observed. These two
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3 signals appear to be co-localized in the same region, as shown in Figure 4c. This co-localization
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5 was quantified using an algorithm that counts the number of pixel intensities in the different
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7 channels and makes a correlation plot (Figure 4d). A Pearson's coefficient of 0.88 was obtained,
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9 which proves the strong cohesion of SiC with Au nanorods inside nanohybrids. By counting a
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11 total number of observed SiC nanoparticles and SiC nanoparticles colocalized with 607 nm
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13 channel we estimated that up to 90% of SiC nanoparticles were functionalized with gold
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15 nanorods. Next healthy cells were then incubated with SiC@Au nanohybrids and observed
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17 under multiphoton imaging (Figure 5).
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32 **Figure 5.** 3T3-L1 healthy cells labelled with SiC@Au nanohybrids observed under multiphoton
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34 imaging (excitation wavelength: 790 nm): detection at 395 nm (a), detection at 607 nm (b) and
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36 all detection channels overlayed (c).
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42 The signal detected at 395 nm (Figure 5a) corresponds to the SHG from SiC nanoparticles while
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44 the signal detected at 607 nm (Figure 5b) corresponds to the TPL of Au nanorods. To avoid the
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46 observation of cells autofluorescence, a low laser power was used. In this case, the
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48 autofluorescence signal is highly reduced and cannot be confused with the bright spots coming
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50 from nanoparticles and nanohybrids.
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56 3.3. Photothermal properties of SiC@Au nanohybrids

57 Drops of nanoparticles and nanohybrids suspensions were directly irradiated using a 808 nm
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59 continuous-wave laser through an optical fiber. The power per surface unit emitted at the exit
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of the optical fiber was measured to be 40 mW cm^{-2} . Instantaneously, heat generation was recorded with an infrared camera. The corresponding thermal images and temperature measurements are presented in Figure 6.

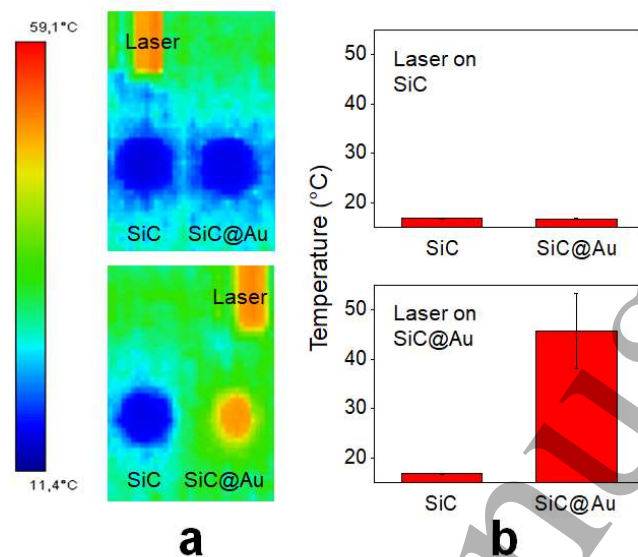


Figure 6. Thermal images (a) and corresponding temperatures (b) of drops of nanoparticles suspensions under 808 nm continuous-wave laser irradiation: SiC nanoparticles (left drop), and SiC@Au nanohybrids (right drop). The drop that is irradiated can be identified by the position of the laser optical fiber on each thermal image.

As shown in Figure 6, SiC nanoparticles irradiation did not lead to temperature increase while for SiC@Au nanohybrids, an instantaneous temperature elevation from 17°C to 45°C was obtained upon laser irradiation. A high temperature increase was also observed for Au nanorods dispersions (Figure S5, Supporting Information). Therefore, photothermal effect may be attributed to presence of Au nanorods into nanohybrids. As discussed in previous reviews, heat is mainly due to Joule effect generated by the current from oscillating electrons in the Au nanorod volume.^[27,28] However, surface effects can also appear due to the immersion of nanoparticles in a liquid, the presence of molecules (CTAB) at the surface of Au nanorods and their immobilization at the surface of a larger nanostructure (SiC nanoparticles). As a

consequence, surface thermal resistivity can occur and influence the heat release. Therefore, different Au nanorods coating may lead to different photothermal effects.

3.4. Toxicity assay towards healthy cells

Then, the toxicity of SiC nanoparticles and SiC@Au nanohybrids towards 3T3-L1 healthy cells was evaluated. The in-time evolution of cell index (proportional to the percentage of remaining living cells) was recorded using a non-destructive impedance-based method with an xCELLigence device (Figure 7). For a better comparison of the different samples, cell indices were normalized at the injection time t_0 of nanoparticles (48 h after the beginning of cell culture).

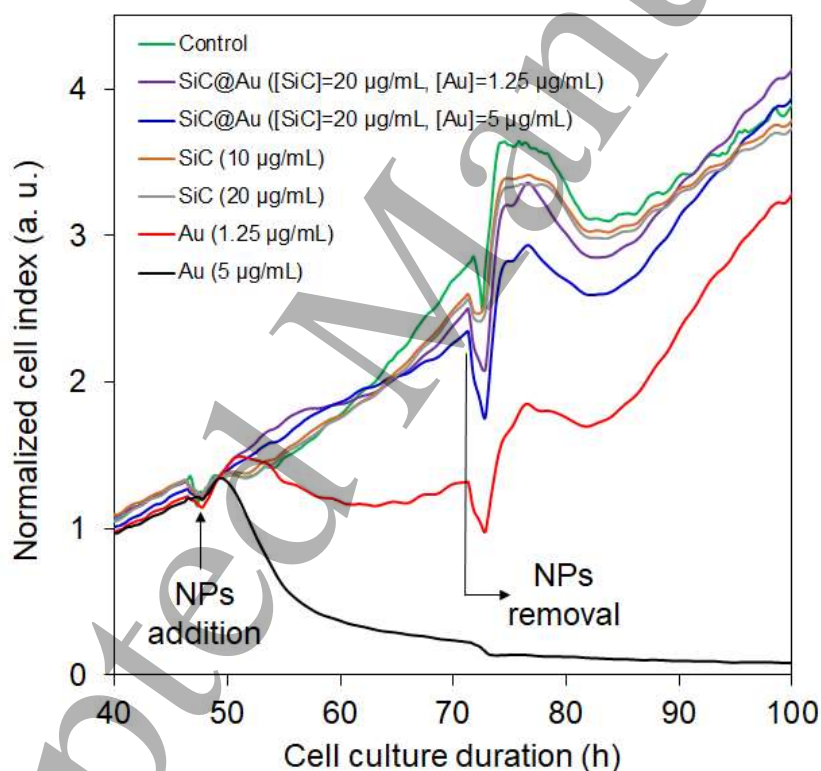


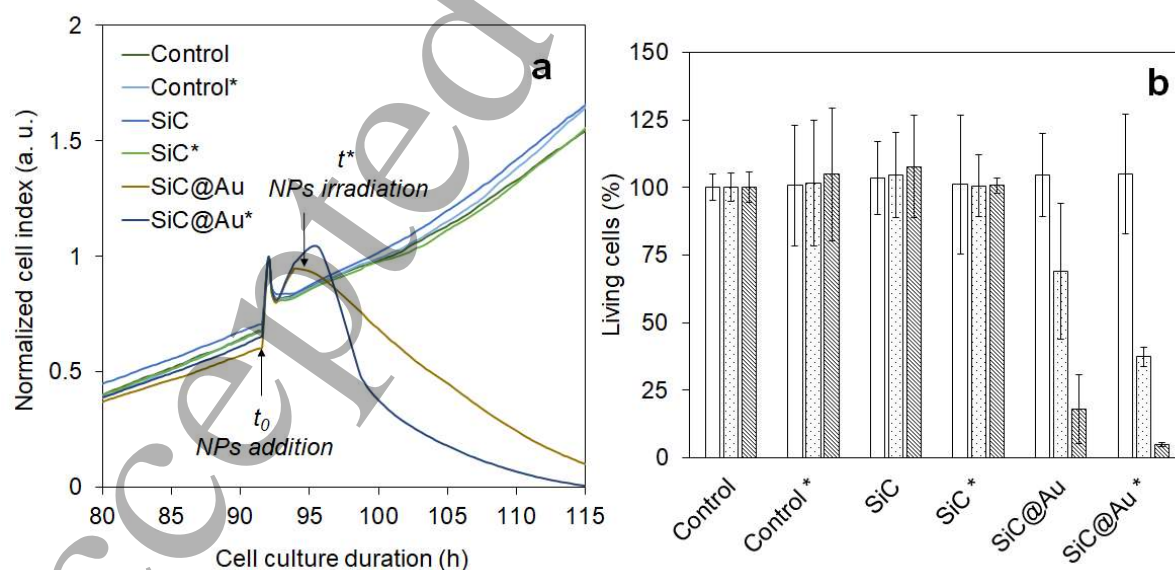
Figure 7. Normalized cell index measured by xCELLigence system as a function of cell culture duration for 3T3-L1 healthy cells in presence of SiC, SiC@Au and Au nanorods at various concentrations. Control samples correspond to 3T3-L1 healthy cells only. Nanoparticles were added at $t_0 = 48$ h after the beginning of cell culture and removed at 72 h ($t_0 + 24$ h).

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3 As shown in Figure 7, SiC nanoparticles of concentrations ranging from 10 to 20 $\mu\text{g/mL}$ did
4 not interfere with cell proliferation. This is in agreement with previous work ^[29] in which SiC
5 nanoparticles with a diameter larger than 15 nm did not exhibit cytotoxic effect. Besides, their
6 highly negative zeta potential (lower than -40 mV, Figure S2) did not favor the uptake of these
7 large diameter nanoparticles by cells ^[30–32]. On the contrary, Au nanorods exhibited a high
8 toxicity for 3T3-L1 healthy cells, with a cytostatic effect for a concentration of 1.25 $\mu\text{g/mL}$ and
9 a clear cytotoxic effect for a concentration of 5 $\mu\text{g/mL}$ although Au nanorods are known to be
10 biocompatible in a broad range of aspect ratios from 2.2 to 5.9.^[33] However, their surface
11 chemistry also have a great importance on their cytotoxicity.^[34] In particular, the presence of
12 CTAB onto Au nanorods surface may be related to their observed high toxicity.^[35] Indeed,
13 CTAB induces a positive surface charge (Figure S2, Supporting Information) resulting to an
14 increased cellular uptake, which can lead to intracellular toxicity. A previous study also showed
15 that toxicity of Au nanorods was mainly due to free CTAB residues issued from desorption ^[36].
16 SiC@Au nano hybrids were prepared according to the protocol detailed in Section 2.4 using the
17 lowest (1.25 $\mu\text{g/mL}$) and the highest (5 $\mu\text{g/mL}$) concentrations of Au nanorods. Although it was
18 difficult to determine the amount of Au nanorods really immobilized onto SiC nanoparticles
19 surface, the two kinds of SiC@Au nano hybrids were found to be non toxic. Thus the
20 concentration of free Au nanorods after washing of SiC@Au nano hybrids was supposed to be
21 negligible, even when using the highest Au nanorods concentration for their preparation. The
22 toxic effect of CTAB might be reduced in SiC@Au nano hybrids compared to Au nanorods
23 because of the screening effect of SiC nanoparticles. Attempts to exchange CTAB by other
24 ligands were carried out but lead to poor colloidal stability. Besides, Au nanorods linked to
25 larger diameter SiC nanoparticles (150 nm) may behave like high diameter spherical
26 nanoparticles that are too big to be internalized by endocytosis. It was shown that above 50 nm
27 in diameter, the capacity of a nanoparticle to be submitted to endocytosis decreases and
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becomes negligible above 100 nm.^[37] Such results are in agreement with the observation of 3T3-L1 cells labelled with SiC nanoparticles and Au nanorods (Figure S3, Supporting Information) showing that SiC nanoparticles were more located on the periphery of cells while Au nanorods are more homogeneously distributed onto cells and are able to penetrate inside.

3.5. Phototherapeutic properties

Finally, nanoparticles and nanohybrids were incubated with cancer cells and their toxicity was compared with and without continuous-wave laser irradiation at 808 nm (40 mW cm^{-2}), which corresponds to the spectral region where Au nanorods strongly absorbs light and thus can lead to photothermal effect. In order to evaluate this effect, the in-time evolution of cell index (proportional to the percentage of remaining living cells) was recorded. For a better comparison of the different samples, cell indices were normalized at the injection time t_0 of nanoparticles (92 h after the beginning of cell culture). The evolution curves for SiC nanoparticles and SiC@Au nanoparticles and nanohybrids are shown in Figure 8a. The percentage of remaining living cells at different stages of the experiment extracted from these curves is presented in Figure 8b.



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3 **Figure 8.** (a) Normalized cell index measured by xCelligence system as a function of cell
4 culture duration for HuH7 cancer cells in presence of SiC nanoparticles and SiC@Au
5 nano hybrids. Control samples correspond to HuH7 cells only. Nanoparticles were added at $t_0 =$
6 92 h after the beginning of cell culture. The irradiation (samples labelled with a *) was
7 performed at $t^* = 94$ h for 6 min. (b) Percentage of remaining living HuH7 cancer cells at $t_0 +$
8 1 h (white bars), $t^* + 6$ h (dot bars) and $t^* + 16$ h (stripe bars).

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19 As shown in Figure 8a, HuH7 cells only or HuH7 cells in presence of SiC nanoparticles were
20 still proliferating with or without laser irradiation. However, the presence of nano hybrids
21 produced a significant decrease of cancer cells proliferation, even without laser irradiation. This
22 effect was more pronounced for SiC@Au under laser irradiation: 6 hours after nanoparticles
23 irradiation, the rate of living cells was around 70% for non-irradiated SiC@Au while it was of
24 37% for irradiated SiC@Au (Figure 8b). These rates fell to 18% for non-irradiated SiC@Au
25 and 5% for irradiated SiC@Au 16 h after nanoparticles irradiation. Besides, as shown in Figure
26 8a, nano hybrids injection into cancer cells was accompanied by an increase of normalized cell
27 index from $t = 92.5$ h to $t = 95$ h, which was not observed for control sample, SiC nanoparticles.
28 This could be attributed to an increase of cell volume, which may be due to cellular necrosis
29 that is often preceded with a phase of cellular swelling prior to cellular lysis. As discussed in
30 Section 3.4, such a behavior implies the probable presence of free Au nanorods in the samples.
31 Although SiC@Au nano hybrids were found to be non toxic for 3T3-L1 healthy cells (Figure
32 7), a different behavior was observed in the presence of HuH7 cancer cells. Indeed, it was
33 demonstrated that the exposure of different cell types to the same kind of nanoparticles may
34 influence cell morphology, ATP and ROS production and their capacity of cellular uptake [38-
35 41]. Besides, concerning *in vivo* studies, it is also well known that nanoparticles may be more
36 efficiently accumulated by tumor tissue than by healthy one due to enhanced penetration and
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3 retention (EPR) effect ^[42]. This can potentially induces the penetration of some free Au
4 nanorods inside HuH7 cells, leading to toxicity. However, the higher toxicity observed after of
5 irradiation may due to the photothermal effect of nanohybrids, without excluding the possible
6 presence of free Au nanorods inside the cells, as discussed before.
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14 **4. Conclusions**

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16 In this study, nanohybrids were prepared from harmonic nanoparticles coated with gold
17 nanorods. Their structural and nonlinear optical properties were investigated, showing that the
18 two kinds of nanoparticles were colocalized. Photothermal effect was measured. Harmonic
19 nanoparticles and nanohybrids showed no effect on cell proliferation as assessed by impedance
20 measurements cytotoxicity for healthy cells while gold nanorods were found to exhibit a dose
21 dependent toxicity. However, this effect may not be related to gold nanorods by themselves but
22 to CTAB coated on their surface. Finally, the phototherapeutic properties of nanohybrids upon
23 cancer cells were studied. Without laser irradiation, nanohybrids exhibit toxicity for cancer
24 cells. This effect is increased under laser irradiation. This work can provide several
25 opportunities in the field of cell imaging and cancer therapy. The measurement of two distinct
26 nonlinear optical signals for the two components (SHG for harmonic nanoparticles and TPL for
27 gold nanorods) offers the possibility of multiplexed detection. This could bring more accuracy
28 for the detection of cancer cells. Furthermore, thanks to the NIR transparency window and the
29 spatial resolution of SHG, one can expect to be able to gain 3D information on the morphology
30 of tumoral tissue. The elaborated nanohybrids also have the potential to affect cancer cells while
31 preserving healthy cells. Although the photothermal effect is probably not the only process
32 involved, this represents interesting perspectives in the cancer nanotherapy research field.
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