

Paleogenomic insights into the origins of French grapevine diversity

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▶ To cite this version:

Jasmin Ramos-Madrigal, Anne Kathrine Wiborg Runge, Laurent Bouby, Thierry Lacombe, José Alfredo Samaniego Castruita, et al.. Paleogenomic insights into the origins of French grapevine diversity. Nature Plants, 2019, 5, pp.595-603. 10.1038/s41477-019-0437-5 . hal-02187462

HAL Id: hal-02187462 https://hal.science/hal-02187462

Submitted on 18 May 2022 $\,$

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Article:

Ramos Madrigal, Jazmín, Runge, Anne Kathrine Wiborg, Bouby, Laurent et al. (14 more authors) (2019) Palaeogenomic insights into the origins of French grapevine diversity. Nature Plants. ISSN 2055-026X

https://doi.org/10.1038/s41477-019-0437-5

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1 Palaeogenomic insights into the origins of French grapevine

2 diversity

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30

31	The Eurasian grapevine (<i>Vitis vinifera</i>) has long been important for wine
32	production and a food source. Despite being clonally propagated, modern
33	cultivars exhibit great morphological and genetic diversity, with thousands
34	of varieties described in historic and contemporaneous records. Through
35	historical accounts, some varieties can be traced to the Middle Ages, but the
36	genetic relationships between ancient and modern vines remain unknown.
37	We present target-enriched genome-wide sequencing data from 28
38	archaeological grape seeds dating to the Iron Age, Roman era, and
39	medieval period. When compared to domesticated and wild accessions, we
40	found the archaeological samples were closely related to Western
41	European cultivars used for winemaking today. We identified seeds with
42	identical genetic signatures present at different Roman sites, as well as
43	seeds sharing parent-offspring relationships with varieties grown today.
44	Furthermore, we discovered one seed dated to ~1100 CE was a genetic
45	match to 'Savagnin Blanc', providing evidence for 900 years of
46	uninterrupted vegetative propagation.
47	

Since its domestication in Southwestern Asia more than 6000 years ago ¹⁻³, the Eurasian grapevine (*Vitis vinifera* L.) has become one of the world's most widely produced and economically valuable fruit crops. Although grapevine products are widely consumed as table grapes, dried raisins, fruit preserves, and cooked leaves, both archaeological and historical evidence indicates that wine has been

53	its primary use ^{4,5} . A key unresolved question in ancient viniculture is the origin
54	and proliferation of vegetative propagation ⁶ . Like many other fruit crops,
55	grapevine is grown almost exclusively as clonal lineages, wherein favored
56	varieties are maintained through horticultural techniques like grafting, layering,
57	and planting of shoots ^{7,8} . These methods take advantage of its natural ability to
58	reproduce asexually under certain conditions, and ultimately enable the
59	establishment of genetic clones of valuable cultivars. With vegetative
60	propagation, viniculturists can consistently harvest berries with a desired flavor
61	profile, and with relatively limited effort, have the potential to expand cultivars
62	to new vineyards and distant regions. The alternative approach of sowing seeds
63	is unreliable because grapevine genomes are highly heterozygous and
64	individuals grown from seed are highly diverse in quality, yield, phenotype, and
65	phenology ⁸ . Moreover, winemakers have to wait from three to five years until
66	vines reach maturity ⁹ , before it is possible to assess berry quality and yield.
67	Thus, clonal lineages of high-quality vines have become indispensable in modern
68	viniculture. Discovering the antiquity of vegetative propagation technologies and
69	the unique histories of individual grapevine varieties will mark a major
70	advancement in our understanding of ancient viniculture, provide a means to
71	investigate longstanding local agricultural traditions, and generate pertinent
72	information for future development of breeding schemes (<i>e.g.</i> through better
73	understanding why some varieties have been more successful than others, or
74	adding historical value to present-day cultivars).
75	

The history of winemaking in France provides a useful model to explore howvegetative propagation helped establish ancient vineyards, and how those

78 actions ultimately shaped the economy and landscape of one of the world's most esteemed winegrowing countries. Written sources and archaeological 79 80 records indicate vineyards were first planted at the Greek colony of *Massalia*, 81 present day Marseille, during the 6th century BCE ^{10,11}. Winemaking subsequently 82 spread along the Mediterranean coast ¹², but it was not until end of the first 83 century BCE that Romans greatly increased wine production across southern 84 France ¹⁰. Roman authors, including Pliny the Elder in the first century CE (¹³: 85 Book XIV), discussed grafting and grapevine varieties, thereby demonstrating 86 their proficiency in vegetative propagation techniques. While Pliny describes 91 87 varieties, it is currently impossible to link Roman names to modern grapevines; 88 however, it is frequently speculated that some living varieties were grown by the 89 Romans, and that those genetic clones have been maintained for two millennia 9. 90 After the fall of the Roman Empire, winemaking traditions continued in France, 91 and by the Middle Ages, contemporary variety names appear in written records 92 ¹⁴. Even though historic names are still used today, it remains unknown whether 93 the same genetic clone has been maintained, or if names have been assigned to 94 other lineages.

95

Archaeobotanical remains, in particular seeds, have the potential to shed new
light on the legacy of French grapevine varieties, and more generally on the
history of viniculture. Using morphometric analyses of seed shape, researchers
have shown seeds from most domesticated grapevines (*V. vinifera* subsp. *vinifera*) can be distinguished from those produced by wild vines (*V. vinifera*subsp. *sylvestris*) ^{15,16}. With this approach, Bouby *et al.* ¹⁰ determined that early
Roman sites in Southern France (50 BCE–225 CE) contained greater numbers of

103	morphologically wild seeds than the following period (225–600 CE), raising the
104	question of whether Romans collected and cultivated wild berries for
105	winemaking. Through this time series, seed shapes tended toward domesticated
106	morphotypes, a finding the authors hypothesize represents a combination of
107	continued selective pressures with a sporadic incorporation of native varieties
108	through sexual reproduction. While these interpretations are thought provoking,
109	the authors also recognize critiques that some wild and domesticated vines
110	produce morphologically indistinguishable seeds.
111	
112	One of the most promising avenues of research for ancient viniculture is
113	palaeogenomic (or ancient DNA, aDNA) analysis of well-preserved
114	archaeological pips ^{17–19} . For example, Wales <i>et al</i> . ²⁰ demonstrated that many
115	waterlogged grape seeds contain high proportions of endogenous DNA that
116	could be interrogated with state-of-the-art, high-throughput aDNA sequencing.
117	With the establishment of genomic databases for hundreds of modern cultivars
118	and wild grapevines ²¹ , we sought to examine how DNA recovered from
119	archaeological samples could sidestep some of the challenges of conventional
120	archaeobotanical methods and reveal relationships between ancient samples
121	and modern varieties, thereby providing otherwise unachievable insights on past
122	implementation of vegetative propagation and the antiquity of some of the
123	world's most produced grapevine varieties.
124	
125	Results and discussion
126	

127 Successful enrichment of SNP loci in archaeological pips

128	We performed targeted enrichment and shotgun sequencing of 10,000 SNP loci
129	in 28 archaeological grape seeds. The pips were recently excavated from
130	waterlogged features (wells, latrines, ditches, and pits) at 9 French sites
131	(Supplementary Fig. 1), and based on archaeological context, date as early as the
132	Iron Age (510–475 BCE) and as late as the medieval period (1050–1200 CE)
133	(Table 1). SNP loci were selected from the GrapeReSeq panel, a DNA microarray
134	that was developed to authenticate varieties for breeding and germplasm
135	management ²¹ . This reference panel provides data for 783 domesticated
136	varieties (V. vinifera subsp. vinifera), 112 wild (V. vinifera subsp. sylvestris)
137	accessions collected from Eurasia and North Africa, and 11 other Vitis species.
138	We obtained a 4- to 400-fold enrichment at the targeted SNP sites, leading to an
139	on-target depth of coverage of 1–25.7 $ imes$ (Supplementary Table 1 and
140	Supplementary Fig. 2). Nucleotide misincorporation patterns observed in the
141	sequencing data and read length distributions were consistent with those
142	expected for degraded DNA 22 (Supplementary Figs. 3, 4 and 5a) .
143	
144	Archaeological seeds related to European winemaking lineages
145	We employed multidimensional scaling (MDS) to investigate whether
146	archaeological samples were more closely related to wild accessions or
147	domesticated varieties. Samples were compared to the GrapeReSeq panel
148	following the random allele sampling strategy described in <i>bammds</i> ²³ , to
149	account for varying depth of coverage in the archaeological samples.
150	Additionally, we expanded our reference dataset with publicly-available whole-
151	genome sequencing data from 27 wild and domesticated grape accessions $^{24-26}$
152	(Supplementary Table 2). The MDS plots showed all 28 archaeological samples

153	fall within the variability of domesticated grapevines, suggesting none of the
154	seeds originated from truly wild vines (Fig. 1a). While it is plausible that samples
155	near the boundary of the domesticated and wild clusters could represent F_1
156	hybrids between domesticated varieties and wild grapevines (e.g. specimen R-
157	LLE_09), we find no evidence for large-scale collection of wild berries by Romans
158	or medieval people at these sites. Likewise, the oldest sample, from the Iron Age
159	site of La Cougourlude dating to $510-475$ BCE, also falls within the MDS space
160	composed of cultivated grapevines. These findings support Bouby <i>et al.</i> 's 10
161	hypothesis that even though many pips from Roman and medieval sites exhibit
162	wild morphotypes they in fact originate from domesticated varieties.
163	
164	Once we determined that archaeological seeds likely originated from
165	domesticated grapevines, we repeated the MDS analysis without wild accessions
166	to achieve a more refined picture of the relationships to regional varieties and
167	types of berries (<i>i.e.</i> , predominantly used in winemaking or as table grapes). The
168	majority of the archaeological pips were most closely related to wine cultivars
169	from West and Central Europe (Fig. 1b), although the three Early Roman samples
170	from the Mas de Vignoles XIV site had a closer affinity to wine grapes from the
171	Balkans and the Iberian Peninsula. Overall, this analysis shows that the
172	archaeological seeds are predominantly related to Western European varieties
173	that are used for winemaking, and not grapevines that are today grown further
174	east for wine or table grapes. These data suggest that 2000 years ago cultivated
175	vines in the modern territory of France were distinct from their Near Eastern
176	ancestors and well on their way to founding the germplasm of modern varieties
177	used in Western European winemaking. We also verified that the patterns

observed in the MDS analysis using the GrapeReSeq panel were consistent with
those obtained from a whole-genome (WG) reference panel (Supplementary
Table 2 and Supplementary Fig. 6).

181

We further explored the genetic structure of the archaeological seeds with a twostep model-based clustering analysis. First, ADMIXTURE ²⁷ was used to infer the
ancestry proportions within the samples in the reference panel, and then
FastNGSAdmix ²⁸ was used to estimate the ancestry proportions in the
archaeological samples (Fig. 1c and Supplementary Fig. 7). The results were
consistent with the MDS analysis, showing that most archaeological seeds were
related to wine grapes from Western Europe.

189

190 As there is evidence for gene flow with local wild grapevines in Western Europe 191 ¹, we explored the wild ancestry components identified through the clustering 192 analysis. Since these proportions are estimated on the GrapeReSeq SNPs they do 193 not necessarily represent whole-genome ancestry proportions. However, this 194 allowed us to: 1) compare the proportions between present-day varieties and 195 the archaeological seeds at these diagnostic sites, and 2) identify the potential 196 source of the wild grape ancestry in the archaeological seeds. Wild grapevines 197 carry four main ancestry components when assuming 8 clusters (Fig. 1c). While 198 American and Asian Vitis species (yellow) and Eurasian wild grapes from the 199 Caucasus and Turkey (light blue) separate into individual clusters that do not 200 contribute significant ancestry to any other group, wild grapes from the African 201 and Western European populations display two ancestry components (dark and 202 light green) that are found in some domesticated grapes. All archaeological

203	samples except for the most recent (M-LM_22) show evidence of genetic
204	contributions from wild grapevines (Fig. 1c and Supplementary Fig. 7), and these
205	wild ancestries are primarily associated with Western and Central European
206	vines. While these data provide the first clues on the timing of genetic
207	introgression from local vines into domesticated lineages, the amount of wild
208	ancestry does not follow a consistent pattern related to sample age. For example,
209	the oldest sample (La Cougourlude, 510–475 BCE) shows some of the highest
210	levels of wild ancestry (\sim 45%), while other early samples from Mas de Vignoles
211	XIV ($2^{nd}-1^{st}$ century BCE) have marginal amounts of wild ancestry (3.5-4.5%),
212	and five samples from La Lesse-Espagnac (175–225 CE) range from \sim 10–38%. In
213	fact, these proportions of wild ancestry are similar to those observed in modern
214	French varieties, suggesting that the admixture with wild grapevines took place
215	at the earliest stages of viniculture in France, and potentially before other
216	cultivated lineages were introduced to France (<i>i.e.</i> , from Greece or the Italian
217	Peninsula). Together, these results suggest that the local wild gene pool played
218	an early role for domesticated varieties, with the gene flow between wild
219	grapevines and domesticated cultivars occurring at least 2500 years ago.
220	

221 Ancient use of vegetative propagation

The availability of genotype data for hundreds of cultivars in the GrapeReSeq panel, allowed us to explore relationships between archaeological pips excavated from individual sites and across different regions of France. We estimated kinship coefficients among pairs of samples using *KING*²⁹ and *NgsRelate*³⁰. Pairs of samples were classified based on the kinship coefficients and the proportion of sites with 'zero alleles Identical by State' (IBS0)²⁹, into the following

228	categories: identical clones, parent-offspring, highly-related/full-siblings or
229	unrelated ²¹ (Supplementary Table 3). We found six instances of genetically
230	identical pairs or groups of seeds (Fig. 2a). Additionally, we identified first-
231	degree relationships (parent-offspring and highly-related /full-siblings) and
232	unrelated varieties (Fig 2b). However, since grape seeds that have been cross-
233	fertilized contain paternal derived DNA ³¹ which could affect the relatedness
234	analyses, we explored whether the archaeological seeds contained maternal DNA
235	only (as expected from empty seeds), or both paternal and maternal DNA. To do
236	so, we generated sequencing data from three seeds and a wood sample of the
237	same plant and conducted a simulation study, in an attempt to estimate the
238	parental contribution in the archaeological samples (Supplementary Fig. 8;
239	Supplementary Section 16). We found that data from all archaeological seeds,
240	except R-TDM_06, R-TDM_08, R-HW71_03 and M-MDV12_09, were consistent
241	with a paternal DNA contribution of $\leq 10\%$ (Supplementary Figs. 8 to 11).
242	Moreover, we studied the dependence of the relatedness analyses on such
243	contribution and found that $\leq 10\%$ paternal DNA does not significantly affect the
244	results (Supplementary Fig. 12). Therefore, we consider that clonal and parent-
245	offspring relationships are not affected in most samples. On the other hand, full-
246	sibling relationships could derive from multiple scenarios if the samples
247	involved contain paternal DNA (Supplementary Fig. 12c), and thus we classified
248	pairs of samples with this type of relationship as 'highly related'.
249	Grape seeds have been found to follow a degradation process of the two tissues
250	that contain paternal DNA, the endosperm and embryo, resulting in empty seeds
251	(<i>e.g.</i> in up to 30% of the cases for 'Chardonnay' variety 32,33). Our results
252	suggest that the observed clonal clusters among archaeological samples

253	represent empty seeds with only maternal tissue, either produced by the same
254	plant, such as might occur at one archaeological site, or by one grapevine variety
255	spread through vegetative propagation (Fig. 2 and Supplementary Table 3). Five
256	of these clonal clusters consist of two or three seeds from a single stratigraphic
257	context: an Early Roman ditch at Mas de Vignoles XIV near Nîmes city ($2^{nd}-1^{st}$
258	century BCE), a Roman well at Mont Ferrier, Tourbes (1st century CE), a Roman
259	well at La Lesse-Espagnac (ca. 200 CE), a Roman well at Terrasses de Montfau,
260	Magalas (4th century CE), and an early medieval well at Mas de Vignoles XIV (ca.
261	800 CE). Given that bunches of grapes might have been pressed for juice and
262	discarded en masse, these genetically identical specimens may well represent
263	seeds from single plants. The other genetic cluster consists of three seeds from
264	Horbourg-Wihr in Alsace and one seed from La Lesse-Espagnac in
265	Mediterranean France (Fig 2b); while all four samples date to the 2^{nd} century CE,
266	these genetic clones suggest that Romans transported grapevine across long
267	distances (>600 km), most likely as cuttings.
268	Five archaeological sites in Southern France demonstrated the presence of
269	multiple genotypes within a single temporal stratum, providing genetic evidence
270	that multiple lineages or varieties were maintained at individual vineyards. For
271	example, we identified six different genotypes at Mas de Vignoles XIV near
272	Nîmes, three of which shared first-degree relationships and three of which were
273	unrelated (Fig. 2b). Overall, these relationship data indicate that vegetative
274	propagation, long-distance transportation of varieties, and multivarietal
275	cultivation have been practiced in France since the Roman era, consistent with
276	historic accounts ⁴ .
277	

278 The antiquity of modern French varieties

279	We lastly investigated the relatedness between archaeological and modern
280	varieties, by computing kinship coefficients and the proportion of IBSO sites
281	between pairs of archaeological samples and samples present in the GrapeReSeq
282	panel using <i>KING</i> ²⁹ (Fig. 3 and Supplementary Tables 4 and 5). Our results
283	confirm long-held beliefs that Roman and medieval viticulturists maintained
284	ancient lineages using vegetative propagation ¹³ , and that modern French
285	viniculture is in large part a product of these traditions. One archaeological
286	sample from La Madeleine (Orléans), dating to 1050–1200 CE, was an identical
287	clone of 'Savagnin Blanc' (VIVC17636), a variety today cultivated for wine
288	production in Northeastern France and other countries from Central Europe
289	(<i>kinship coeff.</i> =0.496; <i>IBS0</i> ~0.0001; Identity of 99.7% and 99.9% for the
290	GrapeReSeq and WG panels, respectively) (Supplementary Tables 4 and 5).
291	Several researchers previously identified 'Savagnin Blanc', also known as '
292	Traminer Weiss', as a recurrent parent of many commercially important
293	European varieties ^{1,34,35} , and written accounts document the appellation as early
294	as 1539 CE ³⁶ . Our findings extend the presence of this variety in France by
295	hundreds of years and furthermore, suggest that either 'Savagnin Blanc' or its
296	direct relatives have been cultivated in France since the 1^{st} century CE, since
297	archaeological seeds from Mont Ferrier, Tourbes have a parent-offspring
298	relationship with 'Savagnin Blanc' (Figs. 2b and 3).
299	
300	Several archaeological seeds were closely related to 'Mondeuse Blanche'
301	(VIVC7919), a French variety characteristic of the Northern French Alps that has

been suggested to have acted as a key progenitor ^{35,37} . We found that four
genetically identical 2^{nd} century CE seeds from Horbourg-Wihr and La Lesse-
Espagnac have a parent-offspring relationship with 'Mondeuse Blanche',
indicating that just one reproductive cycle has taken place in this lineage in the
past 1800 years (Fig. 3). This finding presents an exciting consilience of genetic
and archaeobotanical data; using morphometric analysis, Terral <i>et al.</i> ¹⁶ also
found evidence for 'Mondeuse Blanche' among 1^{st} – 2^{nd} century CE pips from the
Rec-de-Ligno site, which lies less than 10 km from La Lesse-Espagnac. We also
observed that 'Mondeuse Blanche' is highly related (full-sibling or similar
relationship) to an archaeological seed from Colletiere, dating to circa 1000 CE,
close to the region where 'Mondeuse Blanche' is still grown today (Savoie, Ain)
(Fig. 3). Interestingly, the medieval seed is also highly related to 'Tressot'
(12640) (cited since 1396 in France ³⁸) and 'Servanin' (VIVC11526), both French
varieties that are rarely cultivated today.
In addition to 'Mondeuse Blanche', four other Roman seeds from Southern
France provided parent-offspring relationships to modern Alpine varieties: three
1^{st} century CE seeds from Mont Ferrier are highly related to 'Arvine'
(VIVC664) and 'Amigne' (VIVC425) and one 1^{st} - 3^{rd} century CE seed from
Roumeges is a first-degree relative to 'Humagne Blanc' (VIVC5450) (Fig. 3).
All three are Swiss varieties used for white wine, and the former two are
recorded in Switzerland by the 17^{th} century CE 39 . Tradition holds that the
Romans brought 'Amigne' to Switzerland as a variety they referred to as

326 etymological, with the retained usage of the Latin word *amoenus* for "delicious" ⁴⁰. Our findings suggest there indeed is a close genetic link between the varieties 327 328 grown by the Romans and some modern Swiss cultivars, including 'Amigne'. 329 Moreover, these data suggest that modern Alpine varieties may have been 330 cultivated in a more widespread geographic region during the Roman period, 331 thus posing an important question on their origin and the adaptation of modern 332 grapes. The approaches established here can be applied to other archaeological 333 pip assemblages with the aim of detecting when regional and economically 334 important lineages first appeared and how they were maintained.

335

336 Impact of cultural changes in the viniculture of France

337 Specimens from the Mas de Vignoles XIV site in Nîmes provide one final 338 observation on the changing nature of viniculture in France. This site allowed us 339 to investigate a transect of three time periods: $2^{nd} - 1^{st}$ century BCE in the early 340 Roman period, 417–515 CE in the Late Roman period when viticulture was fully 341 established in the region, and 731–851 CE in the early medieval period. While 342 cultivars from the most recent period were found to share first-degree 343 relationships with modern French varieties, no relationship was found between 344 cultivars from the Roman period and the modern varieties (Fig. 3). Our results 345 from Mas de Vignoles XIV suggest a change in grapevine diversity from Roman to 346 Medieval times. This transition can also be observed in the MDS analyses (Fig. 347 1b); the three seeds from the early Roman period (R-MDV14_04/07/09) are 348 placed closer to East European and Iberian grape varieties, while Late Roman 349 and early medieval seeds are more similar to West Europe varieties. These 350 results show the relatively high diversity of grapes cultivated in this region

during this period, as well as replacement and incorporation of new varietiesthrough time.

353

354 **Concluding remarks**

355

356 Palaeogenomic analysis of archaeobotanical remains has helped reveal the 357 evolutionary histories of annual crops like barley ⁴¹ and maize ^{42,43}, but this 358 project represents the first nuclear aDNA study of a vegetatively propagated fruit 359 crop. Our results highlight the utility of state-of-the-art palaeogenomic methods 360 in the study of ancient viniculture through space and time. While previous 361 studies on ancient chloroplast DNA ²⁰, microsatellites ^{18,19,44}, and proteins ¹⁸ have 362 provided insights into the history of grapevine cultivation, their resolution is 363 limited. With the availability of a nuclear DNA diversity panel, we interrogated 364 genome-wide data from archaeological grape seeds, identified relationships 365 between ancient pips and modern varieties, observed connections between 366 distant sites, and traced the history of vegetative propagation in France. Future 367 palaeogenomic research on archaeological grape seeds holds great potential in 368 identifying the links between past and present grape varieties, and especially for 369 refining our knowledge of the pace of domestication and improvement under 370 vegetative propagation ⁴⁵. 371

372 Materials and methods

373

374 Archaeological sample description

375 Grape seeds were collected from nine archaeological sites in France during 376 excavations of wells, latrines, pits, and ditches (Supplementary Fig. 1; see 377 Supplementary Section 1 for a description of the archaeological sites). Sediment 378 samples were systematically collected and immediately isolated to prevent 379 contamination and stored in cool conditions (4° C). The sediment samples were 380 processed at the Institut des Sciences de l' Evolution (ISEM) in Montpellier, 381 France. To prevent contamination with modern material, seeds were isolated in 382 a clean room separate from the archaeobotanical laboratory. Additionally, 383 surfaces and tools were cleaned with bleach prior to handling. Most of the 384 samples included in this study were photographed inside the clean room, with 385 specific equipment in order to carry out morphological analyses. Archaeological 386 samples were dated either by association with archaeological artifacts found in 387 the same stratigraphic units, dendrochronology, or radiocarbon dating. The age 388 of the samples ranged from the Iron Age (510–475 BCE) to the medieval period 389 (1050–1200 CE) (Supplementary Fig. 1 and Table 1).

390

391 Archaeological samples processing

392 Archaeological samples were processed in dedicated aDNA facilities at the

393 University of Copenhagen following standard measures to prevent

394 contamination. Individual seeds were decontaminated with 10% bleach, rinsed

395 with molecular biology grade water, and pulverized. DNA was extracted from the

396 resulting powder following standard protocols standardized for

397 archaeobotanical remains ⁴⁶. DNA extracts were converted into double-stranded

- 398 Illumina libraries using the NEBnext DNA Library Prep Mast Mix Set 2 (E6070L,
- 399 New England BioLabs) with modifications described in Wales *et al.* ⁴⁷ (see

400	Supplementary Section 4 for a description of the protocol). Resulting Illumina
401	libraries were enriched for a set of genomic loci present in the GrapeReSeq
402	reference panel ²¹ (Supplementary Section 5). This panel covers genomic sites
403	known to be informative for identification of grape cultivars. Libraries were
404	captured following the MYbaits protocol as described in Supplementary Section
405	6. Finally, pre- and post-capture libraries were sequenced on an Illumina 2500
406	HiSeq platform in SR100 mode. Sequencing reads obtained from the pre-
407	captured libraries were used to assess the capture efficiency only.
408	

409 Sequencing data processing

*AdapterRemoval2.0*⁴⁸ was used to remove Illumina adapter sequences, low 410 411 quality stretches and ambiguous bases from the read ends. Resulting reads \geq 30 412 base pairs were mapped to the grape nuclear reference genome 12X.2⁴⁹, 413 chloroplast ⁵⁰ and mitochondrial ⁵¹ genomes using *bwa aln* (0.7.5a) ⁵²; seeding 414 was disabled (-l was set to 1000) to improve the mapping sensitivity of aDNA 415 reads ⁵³. Reads with mapping quality below 30 or ambiguously mapping were 416 discarded, PCR duplicates were removed using *MarkDuplicates* 417 (http://picard.sourceforge.net), reads were realigned around indels using GATK ⁵⁴ and the MD-tag was recalculated using *samtools* 1.2 ⁵⁵. Finally, we excluded 5 418 419 bases from the 5' and 3' ends of each read from subsequent analyses to reduce 420 the proportion of bases with aDNA damage. Genotype calling was performed in 421 the resulting alignments using a combination of the *HaplotypeCaller* and 422 *UnifiedGenotyper* algorithms from *GATK*⁵⁴ on sites with a minimum coverage of 423 10× as described in Supplementary Section 12. To evaluate the genotyping 424 pipeline, we generated sequencing data for two modern grape cultivars using the

425 SNP capture protocol. These two varieties are present in the GrapeReSeq panel,

426 thus provide a direct comparison between our method and the GrapeReSeq

427 microarray. We found a concordance of 99.4% and 99.3%, between the called

428 genotypes and their corresponding genotypes in the GrapeReSeq panel.

429

430 Ancient DNA authentication

431 The authenticity of the aDNA data was assessed on the basis of the length

432 distribution and the nucleotide misincorporation patterns observed in the

- 433 sequencing data. We used *bamdamage*²³ to estimate per base nucleotide
- 434 substitutions in the mapped reads. Reads with mapping quality lower than 30
- 435 and base quality lower than 20 were discarded. Archaeological samples

436 displayed increased C-to-T and G-to-A substitutions as well as short reads

437 (Supplementary Figs. 3 and 4), consistent with aDNA data ²².

438

439 **Reference datasets**

440 We used two reference datasets to compare the archaeological grape seeds to 441 present-day grape varieties (see Supplementary Section 11 for a detailed 442 description of the reference panels used). 1) The GrapeReSeq panel consists of 443 783 modern grape cultivars (V. vinifera subsp. vinifera) and 112 wild grape 444 individuals (*V. vinifera* subsp. sylvestris) representative of the genetic diversity 445 found in Europe (81 accessions), as well as from North Africa (18 accessions) 446 and the Caucasus (13 accessions) genotyped for 10,000 diagnostic SNPs²¹. 2) We 447 assembled a whole-genome (WG) reference panel incorporating sequencing data 448 from 27 publicly available wild and domesticated grape accessions ^{24–26}. Raw 449 reads were obtained from the NCBI SRA, mapped and processed using similar

450 parameters as the archaeological samples. To avoid ambiguities due to

451 synonymy the VIVC number ⁵⁶ is assigned to cultivars as indicated in

452 Supplementary Table 4.

453

454 **Genetic structure analyses**

455 We explored the genetic relationships between the archaeological grape seeds 456 and the samples in the two assembled reference panels using multidimensional 457 scaling as implemented in *bammds*²³ (Figs. 1a and 1b, and Supplementary Fig. 458 6). Samples with an on-target depth of coverage $\geq 3 \times$ were included to the 459 reference panel by sampling a random allele from the called genotypes; while the 460 six low coverage samples were incorporated from a majority count consensus 461 sequence (Supplementary Table 1). After filtering low-quality SNPs, the final 462 datasets consisted of 9,896 and 3,076,549 sites for the GrapeReSeg and WG 463 panels, respectively. Note that, for analyses using the GrapeReSeq panel we did 464 not exclude transition sites. However, data from the genotype calls and majority 465 count consensus sequences obtained for the archaeological samples showed 466 error rates comparable to those of modern grape samples (Supplementary Fig. 467 5), suggesting that the aDNA derived error is unlikely to have a substantial effect 468 in the analyses. 469 We used the model-based clustering approaches implemented in *fastNGSadmix* 470 ²⁸ and *ADMIXTURE* ²⁷ to estimate ancestry proportions in the archaeological

471 samples (Fig. 1c and Supplementary Fig. 7). First, ADMIXTURE was run on the

472 GrapeReSeq panel assuming 2-8 populations/clusters (*K*=2-8). We obtained

473 1,000 independent replicates for each value of *K* and kept the one with the best

474 likelihood. Then, we estimated genotype likelihoods for the archaeological

samples using the *samtools* model implemented in *ANGSD* v1.9 ⁵⁷ at the sites
included in the GrapeReSeq panel. Finally, we obtained maximum likelihood
estimates of the ancestry proportions for the archaeological samples using the
genotype likelihoods and the *ADMIXTURE*-inferred allele frequencies for each
value of *K* using *fastNGSadmix*. Figure 1c shows the results for the *K*=8, which
resulted among the lowest cross-validation errors (Supplementary Fig. 7b).

482 **Relatedness analyses**

483 To explore the relationships among pairs of archaeological samples and between

the archaeological samples and samples in the reference panels, we estimated

485 kinship coefficients using two approaches: the called genotype-based approach

486 implemented in *KING*²⁹ and the genotype likelihood-based approach

487 implemented in *NgsRelate* ³⁰ (Supplementary Tables 3 to 5).

488 *KING* was run assuming non-homogeneous population structure for the two

489 reference panels and using called genotypes for the archaeological samples. Pairs

490 of samples were classified based on the kinship coefficients and the proportion

491 of sites with 'zero alleles Identical by State' (IBS0), as suggested in

492 Manichaikul *et al.* ²⁹, in the following categories: identical clones ($K \ge 0.49$ and

493 IBS0≤0.001), parent-offspring (0.177<K<0.354 and IBS0≤0.001), highly

494 related/sibling (0.177<K<0.354 and IBS0≤0.25) or unrelated (Supplementary

495 Tables 3 to 5). These values have been shown to be reliable in discerning known

496 first-degree relationships among grape cultivars ²¹.

497 *NgsRelate* was used as a complementary method to validate the results obtained

498 using *KING* and to include low coverage samples for which it was not possible to

499 call genotypes. To run *NgsRelate*, we first estimated genotype likelihoods for the

500	archaeological samples using the <i>samtools</i> model (-gl 1) implemented in ANGSD
501	v1.9 57 . Reads with mapping quality lower than 30 and bases with quality lower
502	than 20 were discarded. We then estimated allele frequencies for the two
503	reference panels using <i>PLINK 1.9</i> ⁵⁸ . These frequencies together with genotype-
504	likelihoods were used to obtain kinship coefficients and the proportion of sites
505	sharing 0, 1 or 2 alleles identical by descent (IBD) between pairs of samples
506	(Supplementary Table 3). These results were evaluated together with the
507	obtained from the genotype-based approach to assign relationships between
508	pairs of archaeological samples.
509	In Supplementary Section 16, we explore the possibility of paternal DNA present
510	in the archaeological seeds through a simulation study and comparing the
511	archaeological seeds data with that obtained from fresh seeds (Supplementary
512	Figs. 8 to 11 and Supplementary Table 6). While most of the archaeological seeds
513	were found to be consistent with data derived from a single individual, our
514	analyses indicate four seeds contain $\geq 10\%$ of paternal DNA (Supplementary Fig.
515	11). Additionally, we evaluate potential effects of paternal DNA in the
516	relatedness analyses and found that: 1) clonal relationships can only be detected
517	from true identical individuals even in the presence of paternal DNA, 2) parent-
518	offspring relationships are only ambiguous when the sample contains >10% $$
519	paternal DNA, and 3) apparent full-sibling relationships can result from multiple
520	scenarios, thus pairs of samples with this type of relationship were classified as
521	'Highly related pairs' (Supplementary Fig. 12). Relationships between
522	archaeological seeds and modern grapes were evaluated based on the
523	conclusions from Supplementary Section 16.

524	We further explored the effect of sequencing depth and panel ascertainment in
525	the robustness of the relatedness inferences (Supplementary Section 17). The
526	results indicate that the metrics used to identify relationships between the
527	samples are reliable for samples with an on-target depth of coverage of \geq 2 $ imes$
528	when using genotypes, and $1 imes$ when using genotype likelihoods
529	(Supplementary Table 7). Additionally, we confirmed that samples identified as
530	identical clones display an IBS distance < 0.0001 both in the sites overlapping the
531	GrapeReSeq panel and in off-target sites (Supplementary Fig. 13; Supplementary
532	Section 18).

1.1

533

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- 684

685 Acknowledgments

- 686 We thank the Danish National High-throughput Sequencing Centre for assistance
- 687 in generating the sequencing data. This project was funded by the Danish Council
- 688 for Independent Research (10-081390) and the Danish National Research
- 689 Foundation (DNRF94). L.B. and R.B. were supported by the French National
- Agency of Research (VINICULTURE project ANR-16-CE27-0013). We would like

691	to thank the following scientific and technical directors and corresponding
692	institutions for providing the archaeological material used in this project as well
693	as contextual information: P. Blanchard (Inrap, site: La Madeleine), E. Verdel
694	(Isère Patrimoine, site: Colletière), H. Pomarèdes (Inrap, sites: Mas de Vignoles
695	XIV and La Lesse-Espagnac), O. Ginouvez (Inrap, site: Terrasses de Montfau), R.
696	Bourgaut (Communauté d'Agglomération du Bassin de Thau, site: Roumèges), P.
697	Flotte (Archéologie Alsace, site: Horbourg-Wihr), M. Compan (Inrap, site : Mont
698	Ferrier), I. Daveau (Inrap, site: Cougourlude) and C. Tardy (Inrap). We are also
699	grateful to the GrapeReSeq consortium for early access to the genotype data.
700	Finally, we would like to thank J. Víctor Moreno-Mayar, Shyam Gopalakrishnan,
701	Filipe G. Vieira, David Maghradze, and Angela Schlumbaum for their helpful
702	discussion.
703	

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706

707 Author contributions

- The project was conceived by N.W., M.T.P.G., R.B., and L.B., and headed by N.W.
- and M.T.P.G.; J.A.S.C., A.K.W.R, R.B. and N.W. designed experimental enrichment
- 710 methodology with input from J.M.M.Z., R.T. and A.F.A.B.; A.K.W.R. processed
- ancient DNA with input from N.W.; J.R.M., A.K.W.R., R.B. and N.W. designed
- analysis strategy; J.R.M. performed bioinformatic analysis with assistance from
- 713 B.P. and T.S.P. and input from N.W, M.T.P.G. and R.B.; J.R.M., N.W., M.T.P.G., R.B.,
- 714 L.B., T.L. and P.T. interpreted the results; L.B., I.F., C.S. and C.H., curated

- archaeological material; J.R.M., N.W. and M.T.P.G. wrote the manuscript with
- 716 input from R.B., L.B. and T.L. and the other authors. All authors revised, edited
- and accepted the manuscript. Primary funding acquired by M.T.P.G.

719 **Competing interest**

720 The authors declare no competing interests.

721

722 Data availability

- 723 Sequencing data produced in this study are available at the NCBI SRA under the
- reference PRJNA489970. Genotype data are available in the *figshare* repository
- under the following DOI: 10.6084/m9.figshare.7610987.

726

727 Figure legends

728 Figure 1. Genetic affinities between archaeological grape seeds and

729 modern *Vitis vinifera* accessions. a. Multidimensional scaling plot (MDS)

- including archaeological samples, wild V. vinifera subsp. sylvestris accessions, and
- 731 domesticated varieties. b. MDS plot restricted to archaeological samples and
- domesticated varieties. Colors correspond to the main ancestry clusters
- 733 identified in Laucou *et al.*²¹. *Archaeological samples that were incorporated to
- the dataset by sampling a random allele from a majority count consensus
- sequence instead of called genotypes c. Model-based clustering analysis of the
- 736 GrapeReSeq panel assuming K=8 clusters. Vertical bars represent individual
- accessions, colors represent the inferred ancestry components, and the fraction
- of each color corresponds to the estimated ancestry proportion. Archaeological

samples are sorted by age, and by sample identification within a stratigraphic
context. Samples that were identified as identical clones are grouped with black
lines and capital letters (A-F) at the bottom.

Figure 2. Geographic distribution and relationships between the distinct 742 743 genetic types of archaeological samples. a. Relatedness among pairs of 744 archaeological samples. Kinship coefficients were estimated using *NgsRelate* 745 between pairs of samples for SNP loci present in the GrapeReSeq panel. Capital 746 letters (A-F) on the left indicate genetically identical clones, *i.e.*, putative ancient 747 and historical varieties. *Archaeological seeds that were found consistent with 748 carrying >10% paternal DNA. b. Map displaying the distribution of genetic types 749 (circles) in each archaeological site. Capital letters (A-F) on the circles indicate 750 clusters of genetically identical seeds represented by more than one seed. 751 Shading of the circles indicates sample age. In red is shown the genetic type that 752 was found in more than one archaeological site. Lines connect pairs of samples 753 that are related as parent-offspring (solid lines) or highly-related/full-sibling 754 (dotted lines). Note that, since in the presence of paternal DNA full-sibling 755 relationships could derive from multiple scenarios, we classified samples 756 consistent with full-sibling relationships as 'highly-related' (see Supplementary 757 Section 16). 758 Figure 3. Genetic origins of ancient and historic French grapevine varieties. 759 Relationships identified between archaeological samples and modern cultivars 760 included in the GrapeReSeq panel. Solid lines represent parent-offspring 761 relationships and dotted lines represent pairs of highly related (full-sibling or 762 similar) samples. Sibling relationships involving pairs of modern cultivar are not

763 displayed for simplicity. *Archaeological seeds that were found consistent with

- carrying >10% paternal DNA. The VIVC (http://www.vivc.de) and GrapeReSeq
- identifiers for the modern cultivars can be found in Supplementary Table 4.

767

Table 1. Description of the archaeological grape seeds used in the study.

#	Sample ID	Geographic coordinates	Archaeologica l site	Stratigraphi c unit	Structure	Age	Dating method	Period	GC †
1	IA-LC_01	43.573639, 3.914750	La Cougourlude, Lattes	US 31084	Ditch FO 30277	510-475 BCE/2480 ± 30 BP (769- 417 cal BCE)	Archaeological artifacts/C14	Iron Age	
2	R-MDV14_04	43.808222, 4.368222	Mas de Vignoles XIV, Nîmes	US 14152	Rural ditch FO 14194	2nd-1st c BCE	Archaeological artifacts	Early Roman	А
3	R-MDV14_07	43.808222, 4.368222	Mas de Vignoles XIV, Nîmes	US 14152	Rural ditch FO 14194	2nd-1st c BCE	Archaeological artifacts	Early Roman	A
4	R-MDV14_09	43.808222, 4.368222	Mas de Vignoles XIV, Nîmes	US 14152	Rural ditch FO 14194	2nd-1st c BCE	Archaeological artifacts	Early Roman	A
5	R-MF_21	43.432556, 3.394222	Mont Ferrier, Tourbes	US 2076	Well PT 2052	1st c CE	Archaeological artifacts	Roman	В
6	R-MF_23	43.432556, 3.394222	Mont Ferrier, Tourbes	US 2076	Well PT 2052	1st c CE	Archaeological artifacts	Roman	
7	R-MF_25	43.432556, 3.394222	Mont Ferrier, Tourbes	US 2076	Well PT 2052	1st c CE	Archaeological artifacts	Roman	В
8	R-HW70_18	48.080500, 7.399194	Horbourg-Wihr	N.D.	Pit ST7054	2nd c CE	Dendrochronology / Archaeological artifacts	Roman	С
9	R-HW71_03	48.080500, 7.399194	Horbourg-Wihr	N.D.	Pit ST7172	2nd c CE	Archaeological artifacts	Roman	С
10	R-HW71_17	48.080500, 7.399194	Horbourg-Wihr	N.D.	Pit ST7172	2nd c CE	Archaeological artifacts	Roman	С
11	R-R_09	43.471306, 3.670139	Roumeges, Poussan	US 5007(12/13)	Well PT 5001	1st-3rd c CE	Archaeological artifacts	Roman	
12	R-R_14	43.471306, 3.670139	Roumeges, Poussan	US 5007(12/13)	Well PT 5001	1st-3rd c CE	Archaeological artifacts	Roman	
13	R-LLE_02	43.300806, 3.239917	La Lesse- Espagnac, Sauvian	US 3019	Well PT 3005	175-225 CE	Archaeological artifacts	Roman	
14	R-LLE_08	43.300806, 3.239917	La Lesse- Espagnac, Sauvian	US 3019	Well PT 3005	175-225 CE	Archaeological artifacts	Roman	С
15	R-LLE_09	43.300806, 3.239917	La Lesse- Espagnac, Sauvian	US 3019	Well PT 3005	175-225 CE	Archaeological artifacts	Roman	
16	R-LLE_13	43.300806, 3.239917	La Lesse- Espagnac, Sauvian	US 3019	Well PT 3005	175-225 CE	Archaeological artifacts	Roman	D
17	R-LLE_14	43.300806, 3.239917	La Lesse- Espagnac, Sauvian	US 3019	Well PT 3005	175-225 CE	Archaeological artifacts	Roman	D
18	R-TDM_06	43.472806, 3.223000	Terrasses de Montfau, Magalas	US 4015	Well PT 4000	4th c CE	Archaeological artifacts	Roman	Е
19	R-TDM_08	43.472806, 3.223000	Terrasses de Montfau, Magalas	US 4015	Well PT 4000	4th c CE	Archaeological artifacts	Roman	Е
20	R-TDM_10	43.472806, 3.223000	Terrasses de Montfau, Magalas	US 4015	Well PT 4000	4th c CE	Archaeological artifacts	Roman	
21	M-MDV13_07	43.808222, 4.368222	Mas de Vignoles XIV, Nîmes	US 13525	Well PT 13319	1605 ± 35 BP (417-515 CE)	C14	Late Roman/ Medieval	

22	M-MDV12_02	43.808222, 4.368222	Mas de Vignoles XIV,	US 12111	Well PT 12024	1220 ± 30 BP (731-851 CE)	C14	Early Medieval	
23	M-MDV12_04	43.808222, 4.368222	Nîmes Mas de Vignoles XIV, Nîmes	US 12111	Well PT 12024	1220 ± 30 BP (731-851 CE)	C14	Early Medieval	F
24	M-MDV12_05	43.808222, 4.368222	Mas de Vignoles XIV, Nîmes	US 12111	Well PT 12024	1220 ± 30 BP (731-851 CE)	C14	Early Medieval	
25	M-MDV12_07	43.808222, 4.368222	Mas de Vignoles XIV, Nîmes	US 12111	Well PT 12024	1220 ± 30 BP (731-851 CE)	C14	Early Medieval	
26	M-MDV12_09	43.808222, 4.368222	Mas de Vignoles XIV, Nîmes	US 12111	Well PT 12024	1220 ± 30 BP (731-851 CE)	C14	Early Medieval	F
27	M-C_27	45.436417, 5.520306	Colletiere, Charavines	N.D.	Cultural layer, rubbish deposits	1006-1040 CE	Dendrochronology	Medieval	
28	M-LM_22	47.900472, 1.884333	La Madeleine, Orléans	US 15126	Cesspit F 1517	1050-1200 CE	Archaeological artifacts	Medieval	

[†] Genetic clusters composed of identical clones. The genetic cluster was assigned according to the relatedness analyses described in the results section.





