

Cementitious materials in biogas systems: Biodeterioration mechanisms and kinetics in CEM I and CAC based materials

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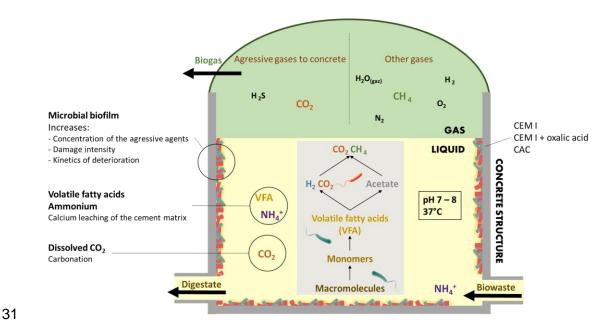
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- 1 Cementitious materials in biogas systems: biodeterioration mechanisms
- 2 and kinetics in CEM I and CAC based materials
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- 13 ABSTRACT
- 14 In anaerobic digestion plants, biological activities transform biowaste into a
- 15 renewable energy: the biogas. This process produces aggressive and complex
- 16 media in direct contact with the structure. Few studies have focused on the
- 17 deterioration of concrete in such environments. In order to support the
- 18 sustainable development of biogas production growing sector, this study intends
- 19 to provide thorough information on the biowaste chemical composition and on
- 20 materials biodeterioration mechanisms occurring in biogas plants. Cementitious
- 21 materials with various surface compositions (pastes of CEM I, CAC and CEM I
- treated with oxalic acid) were immersed in the liquid phase of laboratory
- 23 bioreactors reproducing anaerobic digestion conditions. Altered depths
- 24 measurements in time highlighted the better resistance of the CAC material. The

microbial biofilm coverage on the material's surface was delayed by the surface pretreatment with oxalic acid and was less dense for the CAC paste. Chemical and mineralogical analyses mainly showed leaching and carbonation phenomena.

Keywords: degradation, cement paste, durability, SEM, calcium aluminatecement



1. INTRODUCTION

The microbial fermentation of organic matter in the absence of oxygen, called anaerobic digestion, leads to the production of biogas, a sustainable fuel gas energy mostly composed of methane (65% CH₄) but also containing a significant amount of carbon dioxide (35% CO₂), and leaves a solid residue, the digestate.

Anaerobic digestion ensures the treatment of biowaste directly at the production source (thus reducing the final quantity of waste). Biogas can be used to produce heat with a good energy efficiency and can also be used as a fuel to produce electricity, with a lower yield. The most common energy recovery method for

biogas is cogeneration, which consists of combining the production of heat and of electricity by recovering the heat of combustion to transform it into electricity [1]. The digestate, made up of non-biodegradable organic matter (mostly lignin is the case of vegetal waste), mineral matter and water, is used as a soil fertilizer and can be a source of additional revenue [2]. In addition, anaerobic digestion permits greenhouse gas emissions to be reduced and helps to increase the incomes of the biogas plants operators. This technology of sustainable energy production has been evaluated as one of the most energy-efficient and environmentally friendly [3]. In view of these multiple advantages, biogas production by anaerobic digestion is strongly encouraged by European policies and its development has become global, using many sources of organic waste. Since 2009, the number of biogas plants in Europe has steadily increased from 6227 to 17662 anaerobic digestion structures in 2016 [4]. As for agricultural and agro-industrial buildings, concrete is the material the most commonly used for the construction of biogas plants. This is related not only to its low cost and ease of implementation but also to its air- and water-tightness and its good thermal inertia [5,6]. However, in digesters, structural concrete is directly in contact with both the liquid and gas phases of the digesting biowaste, which are aggressive to concrete [7,8]. Biogas is the final product of four consecutive degradation reactions that make up the anaerobic process: hydrolysis, acidogenesis, acetogenesis and methanogenesis [9,10]. In most plants, the digestion process is performed under mesophilic conditions, with an optimal operating temperature of around 37°C which requires low self-consumption of energy for the heating [11]. In the first

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step of the process, hydrolysis decomposes complex organic polymers (proteins, lipids and carbohydrates) into simple soluble molecules: amino acids, long-chain fatty acids and sugars. These compounds are then reduced into short chain volatile fatty acids (VFA) during the acidogenesis step. Acetogenic bacteria degrade the organic acids into acetate, hydrogen and carbon dioxide. These three compounds are finally consumed by the methanogenic archaea to produce methane, and also CO₂ [12–14]. The degradation of nitrogenous compounds such as proteins and urea leads to the production of ammonia, mainly present in the form of ammonium ions under the pH conditions of the anaerobic digestion [15,16].

Consequently, by being in contact with the biowaste at every step of the anaerobic digestion, concrete may undergo deterioration due to both the chemical and biological agents. Several compounds in the medium [6,7] are aggressive to concrete: volatile fatty acids, excreted by the acidogenic bacteria, ammonium and dissolved CO₂:

- Volatile fatty acids metabolized during anaerobic digestion are organic
 acids whose calcium salts are soluble in water. Even if the acidity of
 organic acids is lower than mineral acids, the buffer effects and the
 solubility of the organic salts make them not less aggressive to concrete.
 Their attack results in a progressive dissolution of all the initial hydrated
 phases of the cement paste [17–19].
- The ammonium cation NH₄⁺ is the conjugated acid of ammonia NH₃, produced during the anaerobic digestion. The ammonium ion react with the cementitious matrix supposedly according to an exchange mechanism 2 NH₄⁺ → Ca²⁺ [20,21]. This leads to the dissolution of portlandite and C-S-H and the decalcification of the cementitious matrix [22,23].

• The presence of carbon dioxide in the medium leads to the carbonation of the cement paste: portlandite and C-S-H are dissolved and calcium carbonates are formed [24–26]. Even if most of the time, carbonation can be beneficial for the concrete durability by clogging the porosity [24,25], in submerged systems such as anaerobic digestion medium, dissolved CO₂ can lead to the dissolution of calcium bearing phases, including the calcium carbonates [21].

The microorganisms involved may organize themselves as a biofilm on the surface of the material [27] and induce very high local concentrations of aggressive agents, accentuating the deterioration [7,28]. In this context, the concrete undergoes deterioration resulting in increased porosity and reduced impermeability, leading to a greater risk of reinforcement corrosion [6,29].

Damage to the structures could lead to both important economic and environmental impacts such recovery of a smaller amount of biogas, pollutant flows nearby the plant, and the need for repairs.

The European standard EN 206 [30] and the French information document FD P 18-011 [31] classify environments that are chemically aggressive for concrete by defining three classes of increasing aggressiveness, XA1 to XA3. Among the aggressive species present in the liquid phase of fermenting biowaste, the texts consider the presence and the concentration of carbon dioxide (aggressive CO₂), sulphate (SO₄²⁻), magnesium (Mg²⁺), and ammonium (NH₄⁺), together with the pH and the water hardness. However, neither document considers the action of organic acids or of microorganisms [17,29], nor the temperature effect, which could accelerate the degradation reactions [32]. Moreover, the pH is not suitable to characterize the aggressiveness of some liquid media, including those containing organic acids [5,33]. It therefore appears that the normative texts are

118 incomplete as far classifying the aggressiveness of agricultural environments, 119 and especially the anaerobic digestion environment, is concerned. In order to 120 facilitate their classification in the standards, better knowledge of these complex environments, in terms of composition and aggressiveness toward concrete, is 122 needed. 123 This study aims to (i) widely characterize the medium in terms of chemical 124 composition; (ii) identify and understand the biodeterioration mechanisms and (iii) 125 compare the performances of different cementitious materials (CEM I, CAC and 126 CEM I surface-treated with oxalic acid). The influence of the oxalic acid 127 treatment, the type of binder and the biofilm coverage on the material durability

129 2. MATERIALS AND METHODS

130 2.1 **Materials**

are discussed.

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131 2.1.1 Cement paste manufacturing

Three types of materials were used for this study: (i) cement pastes of ordinary Portland cement, CEM I 52.5 R (denominated CEM I); (ii) cement pastes made of calcium aluminate cement, Secar 51 RG (CAC) (chemical oxide composition and Blaine specific surface of both the previous cements are given in Table 1) and (iii) CEM I pastes the surface of which had been treated with a 0.28M oxalic acid solution (denominated CEM I-Ox) [34]. CEM I paste was considered as the reference material and CAC and CEM I-Ox cements were chosen because of their already proven higher durability in an acid environment [33-36].

140 Table 1. Oxide compositions of the cements in mass percentages and Blaine's specific surface area

	CaO	SiO ₂	Al ₂ O ₃	Fe ₂ O ₃	MgO	SO ₃	Na ₂ O	K ₂ O	Blaine's specific surface (cm²/g)
CEM I	64.5	20.1	4.83	2.47	1	3.49	0.14	0.17	4970
CAC	44	5.6	52.9	1.47	0.2	<0.2	0.23	0.37	3870

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According to several authors [33,35,36], calcium aluminate cements show high durability in acidic media, with a good chemical stability. This could be explained by the alkalinity and the stability of the initial (e.g. katoite C₃AH₆, aluminium hydroxide AH₃) phases and that newly formed through acidic attack, aluminium hydroxide AH₃, which has a high thermodynamic stability [37,38]. Moreover, the alkalinity of CAC binders vs. OPC ones is detailed in the study of Buvignier et al. [39] by comparing the neutralization capacity of an OPC with the one of CACbased materials on the basis of the calculation by Letourneux and Scrivener [40]. The higher neutralization capacity of the CAC was highlighted and it was assumed that it could contribute to the higher resistance of CAC materials to microorganisms-bearing environments by hindering or inhibiting microbial activity on their surface [39].

Chemical action of the oxalate anion has already been studied and used to provide a surface protection for marble [41,42]; electron microscope observations showed a structural correspondence between the calcium oxalate and the calcite crystals of the marble [43]. The chemical reaction is thought to be based on the transformation of calcium carbonate from the marble into calcium oxalate [44]. This surface layer protects the marble without clogging the pores of the stone [42]. This type of treatment has also been explored on cementitious materials in

laboratory studies, which have shown that, despite the very low pH of the oxalic solution used (pH=1), ordinary Portland paste does not lose mass and is not degraded in depth [33,34]. Calcium oxalate forms a layer on the surface of the material, filling the peripheral porosity. The salt formation is believed to be due to the reaction of the oxalate solution with the Portland cement matrix [17], where the sacrificial dissolution of the portlandite allows the formation of calcium oxalate in the outer layer of the paste specimen. According to Larreur-Cayol et al. [33], the protective effect of the calcium oxalate salt could come from its molar volume which allows the salt to fill the volume released by the dissolution of portlandite and also some of the capillary porosity. Voegel et al. [34] also showed that the oxalic acid treatment on a CEM I paste could delay the paste alteration by an acetic acid solution.

The CEM I and CAC paste specimens were made with a water/binder ratio of 0.4. This ratio optimizes the effects of the acid oxalic treatment and a study by Voegel et al. [34] showed that the water/binder ratio of 0.4 led to a positive mass variation, whereas no mass change was detected for a ratio of 0.27. Thus, the higher porosity of the matrix with a water/binder ratio of 0.4 could have led to the precipitation of a larger amount of calcium oxalate [34]. The pastes were mixed according to the French standard NF EN 196-3 [45] and cast in cylindrical moulds 75 mm high and 25 mm in diameter. The pastes were cast in three consecutive layers, each vibrated for 120 seconds. They were removed from their moulds 24h after pouring. Then, each specimen underwent an exogenous cure in a small amount of water at 20 °C for 27 days, which allowed the hydration to continue while limiting leaching. After the curing period, 5 mm thick slices were sawn from paste cylinder specimen, to be used for the biofilm growth testing. Also, some CEM I paste cylinders and slices were immersed in a 0.28 M oxalic acid solution

 $(C_2H_2O_4)$ (pH=0.76) for 7 days, the other specimens being kept in sealed bags. The initial porosities of the paste specimens were measured by water intrusion porosimetry adapting the French standard NF P18-459 [46] for cement pastes specimens. The results were 40±1 % for CEM I and 27±1 % for CAC.

2.1.2 Characterization of cement paste surfaces

Just before immersion in the biowaste, the surface pH of the specimens was measured with pH paper: it was between 12 and 13 for the CEM I and CAC pastes (sound cementitious materials) whereas it was about 1 for the CEM I-Ox paste (because of the pH 0.76 oxalic acid treatment) [27].

The main mineralogical phases of the cement pastes surfaces before the immersion were determined by X-Ray Diffraction (XRD) analyses. The diffractograms showed the phases habitually encountered in sound CEM I and CAC cement pastes. The CEM I paste surface was composed of C-S-H, portlandite (Ca(OH)₂), ettringite, C_xAH_y , C_3S and C_2S . The CAC paste surface showed C_3AH_6 , gibbsite (AH₃), gehlenite (C_2AS), mayenite ($C_{12}A_7$), C_2AH_8 , brownmillerite (C_4AF) and calcium aluminate oxide (CA). Microscopic observations (performed with a Scanning Electron Microscope (SEM) in back scattered electron mode (BSE)) and mineralogical analysis (by XRD) were made on the surface of the CEM I specimens treated with oxalic acid. They showed an adherent, continuous layer of precipitate of whewellite (calcium oxalate monohydrate) covering the whole specimen (Figure 1). This layer was formed over a thickness of 100 to 200 μ m and clogged the porosity locally [33,34]. More details about the method are given in §2.5.

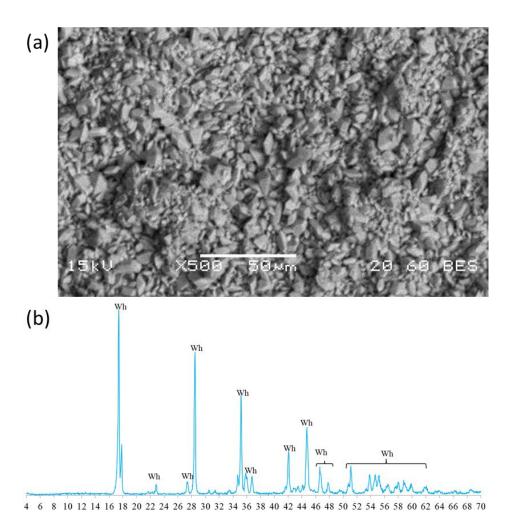


Figure 1. (a) SEM observation and (b) mineralogical analysis of the CEM I-Ox paste surface after the 7 days oxalic acid treatment (Wh: whewellite (calcium oxalate monohydrate))

20 (Co)

2.2 Immersion of cement pastes in the synthetic biowaste

The substrate for biogas production by anaerobic digestion used in this study was a laboratory synthetic biowaste representative of the organic domestic waste currently produced in France according to the CIRSEE (International Research Center On Water and Environment). Its composition was provided by IRSTEA of Antony (France): a mix of different food products. All the details on composition and production are given in Voegel et al. [7]. Bacterial inoculation was made from a sludge sampled in a municipal wastewater treatment in Toulouse (France) in

order to induce the anaerobic digestion process [47]. The inoculation was operated with a V_{inoculum}/V_{substrate} ratio of 2.5, which corresponded to an optimal ratio of 1 g COD_{inoculum}/ g COD_{substrate}, (with COD the chemical oxygen demand) as already established in previous works [7,48]. Cement pastes were totally immersed in the liquid phase of the biowaste in anaerobic bioreactors for 5 weeks. The surface area/volume ratio (cement paste surface area/inoculated biowaste volume) was 224 cm² L⁻¹ for the cylindrical samples and 34 cm² L⁻¹ for the slices used for biofilm development monitoring. These two ratios are guite high in comparison with real situations of industrial digester (4 cm² L⁻¹ [48]) but allow to amplify the phenomena occurring close to the material. The bioreactors (two reactors per type of material, experiments run in duplicate) were kept in an oven at 37° C throughout the experiment. The reaction medium, with an initial volume of 525 mL, was continuously stirred by a magnetic stirring system. Biowaste was renewed at the end of each 5-week digestion cycle. This period corresponds to the average time required for the complete digestion of a substrate [49] and is commonly used as a typical duration for laboratory tests [50,51]. Two cycles were carried out.

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2.3 Analysis of the liquid phase of the digesting biowaste

The liquid phase composition of the digesting biowaste was monitored over time in order to analyse the concentration of organic acids, ammonium and CO₂, and the pH [7]. Regular sampling of 1.5 mL of the liquid phase was carried out and concentrations of the organic acids were measured using High Performance Liquid Chromatography analyses (Thermo Fisher U3000; column: Aminex HPX-87H BIORAD; eluant: H₂SO₄; flow rate: 0.5 mL.min⁻¹) [7,48]. Ammonium concentrations were measured using Ionic Chromatography (Thermo Electron ICS 3000, column: CS16; pre-column; cartridge holder; eluant: 30 mM

methanesulfonic acid; flow rate: 1.0 mL.min⁻¹). The soluble CO₂ produced from the microbial activity was occasionally determined by analysis of total inorganic carbon (TOC-SCHIMADZU) [7].

2.4 Observation of biofilm on cement pastes

Cement paste slices were sawn carefully in order to preserve the biofilm on their surfaces. Before the observations, the biofilms were first chemically fixed and then dehydrated according to the method of Voegel et al. [7,48]. The biofilm was observed by SEM-FEG (JEOL 7100F TTLS combined with EDX XMax, 5kW) after 5 weeks (CEM I, CEM I-Ox, CAC) and 10 weeks (CEM I-Ox).

2.5 Microscopic observations and analysis of chemical and mineralogical changes in cementitious materials

The cylindrical cement paste specimens were removed from the biowaste after each digestion cycle (i.e. after 5 and 10 weeks of immersion) and 5 mm cement paste slices were collected by sawing the cylindrical specimens perpendicularly to their longitudinal axis with a diamond saw. A part of a slice was then embedded in an epoxy resin (Mecaprex MA2+ by Presi) and dry-polished using silicon carbide polishing disks (Presi) according to the procedure described by Bertron et al. [52]. The polished sections were coated with carbon before being analysed. Microstructural observations of cement pastes were made by Scanning Electron Microscopy (SEM, JEOL JSM 6380LV) using back scattered electron (BSE) mode. Chemical analyses were performed on the same specimens using energy dispersive X-ray spectrometry (EDS, Brucker Xflash 6/30 - SDD) and electron-probe micro-analysis (EPMA, Cameca SXFive, 15 kV, 20 nA, scanning area of the beam 2 x 2 μ m²). Punctual analyses were carried out by positioning a hundred points manually, carefully avoiding anhydrous grains, from the degraded

surface to the sound core [7]. A microstructural and chemical zonation of the deteriorated specimens according to the distance to the surface was identified from the correlated results of SEM+EDS and EPMA: the zonation is primarily set through chemical composition profiles obtained from EPMA and then X-ray diffraction (XRD) analyses are performed to evaluate the mineralogical composition in each zone. Mineralogical analyses in the different zones identified were carried out by XRD (Siemens D5000, Co cathode, 40 kV, 30 nA) using the procedure presented in Bertron et al. [53].

3. RESULTS

3.1 Composition of the liquid fraction of the digesting biowaste

Table 2 summarizes the composition of the liquid fraction of the bioreactors through several cycles. The values in the table are averages over the six bioreactors run at the same time (two bioreactors per material type). Figure 2 shows the typical evolutions of the pH and the acid concentrations during a cycle of anaerobic digestion [7].

From Table 2, no major difference in the composition of the liquid fraction is observed between the two cycles (5 and 10 weeks). The acetic acid concentration reaches 23 mmol.L⁻¹ on average, whereas the propionic and butyric acids concentrations reach 9 mmol.L⁻¹. Table 2 and Figure 2 show that the pH decreases in the first few days, to values around 4.5. Within this period, hydrolysis and acidogenesis steps correlated with the production of organic acids. After that, the pH reaches 7.0 on the 10th day and stabilizes around 7.8 thereafter due to a progressive and then total consumption of the VFA during the acetogenesis and methanogenesis steps. Koenig and Dehn [8] found similar pH values (pH of approximately 7) in the liquid phase of their laboratory fermenter.

The average ammonium concentrations reach 55 mmol.L⁻¹. This value is consistent with the studies by Voegel et al. [7] and Giroudon et al. [54], where the evolutions of the ammonium concentration were similar and reached about 800 mg.L⁻¹ (40 mmol.L⁻¹) and with McCarty [49] who states that ammonium concentrations can reach 1000 mg.L⁻¹ for a digester in good function.

Punctual analyses of dissolved CO_2 concentrations showed that, after 3 weeks of biowaste anaerobic digestion, the CO_2 concentration was 2.3 mmol $L^{-1} \pm 0.2$ mg. L^{-1} . The digesting biowaste contained little or no sulfur bearing compounds.

Table 2. Composition of the liquid fraction of the digesting biowaste in terms of VFA concentration, ammonium concentration and pH: minimum value—maximum value (standard deviations).

	VFA con	centration (mm	Ammonium		
	Acetic acid	Propionic acid	Butyric acid	concentration (mmol.L ⁻¹)	рН
1 st cycle (week 1 to 5)	0.14-22.99 (0.22-2.14)	0.26-10.79 (0.12-2.54)	0.00-8.31 (0-1.17)	0.85-54.90 (0.11-1.00)	4.33-7.81 (0.31- 0.18)
2 nd cycle (week 6 to 10)	1.14-24.42 (0.10-2.87)	0.14-9.84 (0.09-2.55)	0.00-9.83 (0-2.32)	0.83-52.24 (0.09-2.11)	4.47-7.52 (0.23- 0.15)

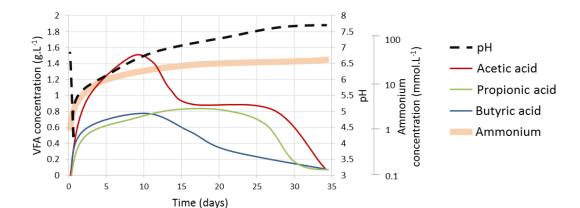


Figure 2. Typical evolution of pH, VFA concentrations and ammonium concentration for a single anaerobic digestion cycle, adapted from Voegel et al. [7]

3.2 Altered depths of the cement pastes

Figure 3 shows the altered depths of the cement pastes after 5 weeks (one digestion cycle) and 10 weeks (two digestion cycles) of immersion in the digesting biowaste. Altered depths were obtained by analyses of flat polished sections using BSE-mode SEM coupled to EDS and EPMA. Altered depths correspond to limits between the sound zone (zone 1) and the beginning of the altered zone (zone 2) on Figures 5, 7 and 9.

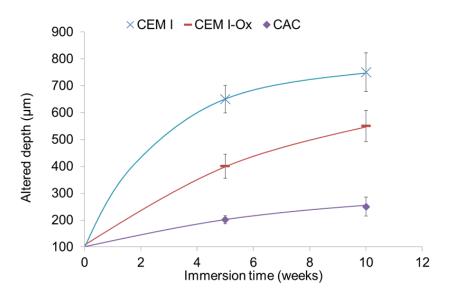


Figure 3. Altered depths of the cement pastes immersed in the digesting biowaste

After 5 weeks of exposure to the digesting biowaste, the CAC samples had the smallest altered depth (200 μ m) whereas CEM I-Ox and CEM I pastes had altered depths of 400 and 650 μ m respectively. After 10 weeks, the degraded depth of the CAC pastes had increased slightly, reaching a value of 250 μ m, whereas CEM I-Ox and CEM I reached degraded depths of 550 and 750 μ m.

seemed to be efficient as the CEM I-Ox degraded depth was 1.6 times less than that of CEM I pastes. During the second cycle (between 5 and 10 weeks), the

increase of the degraded depth of the CEM I-Ox paste was 150 μ m which was greater than the increase of the degraded depth of the non-treated CEM I paste (of 100 μ m). Thus, it seems that the calcium oxalate treatment was no longer effective at this point.

The CAC pastes showed positive achievement throughout the experiment as their degraded depths after 10 weeks were only 250 µm which was 2 or 3 times less than the CEM I-Ox and CEM I degraded depths after 5 weeks.

3.3 Microscopic observation of biofilms

Figure 4 presents the SEM observation of the surface of CEM I, CAC and CEM I-Ox pastes after 5 weeks of immersion and 10 weeks of immersion for the CEM I-Ox sample. At both sampling dates (i.e. 5 and 10 weeks), the CEM I pastes surfaces showed a uniform, dense biofilm whereas the biofilm observed on the surface of the CAC was less dense, with few microorganisms. In contrast, microbial colonization was not observed on the surface of the CEM I-Ox slice after 5 weeks of anaerobic digestion and calcium oxalate precipitates, similar to those observed on the cement paste immediately after the oxalic acid treatment (Figure 1), covered the surface. After 10 weeks of immersion, the entire surface of the CEM I-Ox paste was covered by a dense biofilm and calcium oxalate precipitates were no longer observed. It can be assumed that between 5 and 10 weeks of exposure the surface conditions enabled a biofilm to form on the surface. The biofilm may have produced local conditions that finally dissolved the oxalate salt.

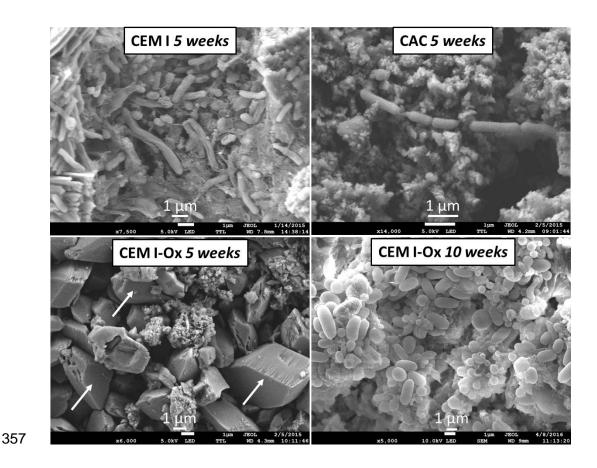


Figure 4. SEM observations of biofilms on cement pastes after 5 weeks (CEM I, CAC, CEM I-Ox) and 10 weeks (CEM I-Ox) of immersion in the digesting biowaste. Calcium oxalate crystals are identified by white arrows.

3.4 Chemical and mineralogical analysis of the cementitious specimens

3.4.1 CEM I paste

The chemical composition profiles of the CEM I paste sample at 10 weeks of immersion, as a function of the distance to the surface, and the image of the same specimen by SEM in back-scattered electron (BSE) mode are presented on the right side of Figure 5. The outer layer of the specimen was partly dissolved when it was removed from the biowaste. The thickness of the material lost is reported on the figure (about 150 μ m) and was measured by measuring the difference between the initial diameter of the sample and its final dimensions. The left side of Figure 5 gives the chemical composition profile of the outer layers of the CEM I paste immersed for 5 weeks in the fermenting biowaste (adapted from

Voegel et al. [7]). A chemical zonation of the specimens was identified and is represented on the figure, from zone 1, corresponding the intact core, to zone 5, corresponding to the outer layer in contact with the biofilm and the biowaste. The mineralogical characterization by XRD in each zone defined in Figure 5 is presented in Figure 6. From the core to the outer layer, the chemical and mineralogical evolution of the cementitious matrix is as follows:

- Zone 1 corresponds to the sound zone. The mineralogical analyses showed a composition similar to that of an unaltered specimen, mainly made of hydrated phases (portlandite, ettringite, and C-S-H) and anhydrous grains (C₃S, C₂S, brownmillerite C₄AF). The SEM observations show the high density of the anhydrous residual cement grains (in lighter grey on the SEM image).
- Zone 2 is 350 µm thick and shows the same two main phenomena in each cycle: (i) a slight decalcification corresponding to the dissolution of portlandite, and (ii) an enrichment in sulfur, which can be correlated with a slight intensification of the ettringite peaks in the mineralogical pattern (Figure 6). This secondary ettringite comes from the diffusion of sulfates from the altered zone, and is classically observed in transition zone of specimens exposed to leaching [53,55]. It is probably not expansive as no microcracks are observed by SEM in this zone. The density of the anhydrous grains remains high in this zone, as can be observed in the SEM picture.
- After each cycle, zone 3 is mainly characterized by the dissolution of the anhydrous grains shown in the SEM picture and a lower density of the hydrated matrix (darker colour of the cement paste). The mineralogical pattern (Figure 6) shows that calcite is the main crystallized phase in this

zone and that ettringite, C_3S , C_2S and brownmillerite phases are dissolved. The chemical composition (Figure 5) shows the decrease in the sulfur content, correlated with the disappearance of the ettringite peaks and the decrease of the total amount of oxides. This zone is 180 μ m thick.

- Zone 4 is mainly composed of the two polymorphs of calcium carbonate (CaCO₃), calcite and vaterite, which form for C/S ratio ≥ 0.67 in the C-S-H [56]. The chemical composition (Figure 5) shows the partial decalcification of the sample and a peak of phosphorous, probably brought by the biowaste. No peak of a crystallized phase containing phosphorus is visible on the diffraction pattern, at either 5 or 10 weeks, suggesting that an amorphous P-phase has precipitated. This zone is 30 µm and 80 µm thick respectively after the 5th and the 10th weeks, respectively, and shows a low density in the SEM image.
- Zone 5 is the outer layer, mainly visible after 5 weeks (70 µm thick). This zone is mainly amorphous (Figure 6) and is likely to be composed of a Si-Al gel, containing phosphorus in its outer part. At 10 weeks, it can be assumed that this external zone was partially dissolved or detached from the matrix. The missing zone was determined to be around 150 µm thick according to image analysis measurements performed on the cross section of the specimen.

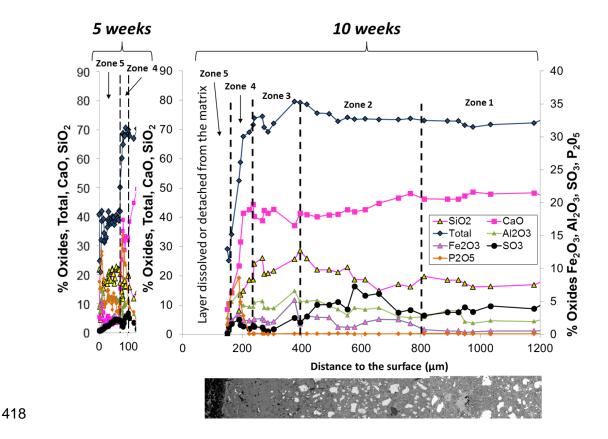


Figure 5. Chemical composition after 10 weeks of immersion and chemical composition of the outer layers of the CEM I paste after 5 weeks [7] according to the distance to the surface (EPMA) (top) and SEM observations of the polished section in BSE mode (bottom)

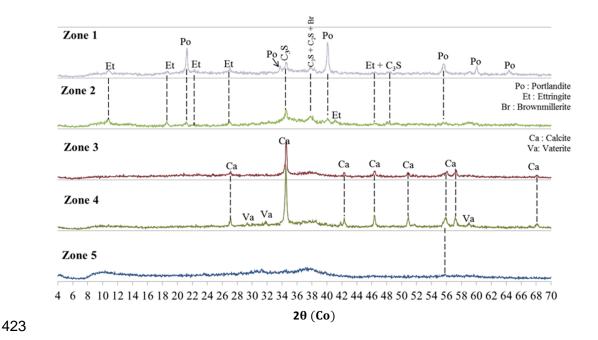


Figure 6. Mineralogical analysis by XRD of CEM I paste immersed for 5 and 10 weeks

3.4.2 CEM I-Ox paste

The chemical composition profiles, according to the distance to the surface, of the CEM I-Ox paste sample at 10 weeks of immersion, and the image of the same specimen by SEM in BSE mode are presented in Figure 7, right side. The left side of the figure gives the chemical composition profile of the outer layers of the CEM I-Ox paste immersed for 5 weeks in the fermenting biowaste. The mineralogical characterization by XRD of each zone is presented in Figure 8. In light of the chemical and mineralogical analyses, four zones can be distinguished after 5 and 10 weeks, with differences in the composition of the outer layer between the two exposure times. The zonation is as follows:

- Zone 1 corresponds to the unaltered zone. Its mineralogical and chemical composition is similar to that of zone 1 of the CEM I paste.
- As for the CEM I paste specimen, Zone 2 shows a slight decalcification
 with an increase of the sulfur oxide content. From a mineralogical point

of view, portlandite peaks are reduced whereas ettringite peaks are intensified, which could be correlated with the increase of sulfur oxides content. This zone is 400 μ m thick and the SEM image shows a less dense structure here than in zone 1.

- Zone 3, 180 µm thick, contains mainly calcium in the form of calcite and vaterite. Anhydrous grains are no longer observed in the SEM image and the paste density has decreased.
- For zone 4, two different patterns are observed at 5 and 10 weeks of exposure to the biowaste. At 5 weeks, this zone is mainly composed of calcium and phosphorus. The CaO content is related to the precipitation of the calcium oxalate salt (whewellite), preserved on the surface, the XRD pattern being very similar to that of the surface before the immersion (Figure 1). The phosphorus enrichment is probably due to diffusion and fixation of this element from the biowaste. At 10 weeks, the calcium content tends to zero in the outer part of this zone. Whewellite has been dissolved and the XRD diffractogram shows an amorphous pattern of the remaining cement matrix.

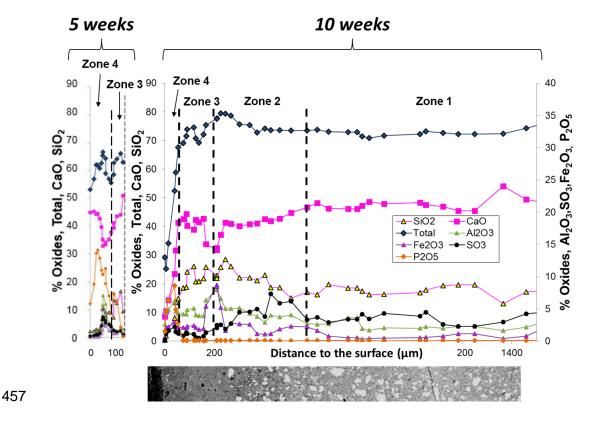


Figure 7. Chemical composition of the CEM I-Ox paste after 10 weeks of immersion and chemical composition of the outer layers of this paste after 5 weeks of immersion according to the distance to the surface (EPMA) (top), and SEM observations of the polished section in BSE mode (bottom)

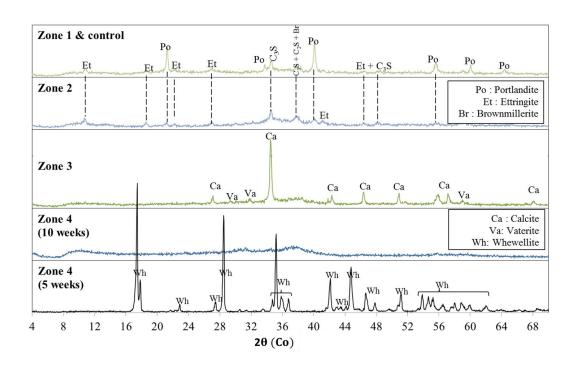


Figure 8. Mineralogical analysis by XRD of the CEM I-Ox paste immersed for 10 weeks and 5 weeks (Zone 4)

3.4.3 CAC paste

The chemical composition of the CAC paste after 10 weeks of immersion is presented in Figure 9, together with a BSE-mode SEM image of the sample. Chemical compositions are similar after 5 and 10 weeks. The mineralogical analyses from the core to the surface are presented in Figure 10. Three zones are distinguished:

Zone 1 has similar composition to that of a sound specimen, i.e. it is mainly made of aluminium and calcium, and contains crystalline anhydrous (calcium aluminium oxides (CA), mayenite (C₁₂A₇), brownmillerite (C₄AF), gehlenite (C₂AS)) and hydrated phases (C₃AH₆, gibbsite (AH₃), C₂AH₈), usually found in aluminate cement matrices. It shows a high density (lighter grey zone on the SEM images).

• Zone 2, 180 µm thick, shows a slight decalcification, and therefore a slight relative enrichment of Al. Mineralogical analyses show the same phases as in zone 1 overall. However, some slight differences can be noted: the peaks of the anhydrous phases, CA and gehlenite are less intense closer to the surface; in contrast, gibbsite peaks (AH₃) increased. Aragonite is detected, probably due to the carbonation of katoite C₃AH₆ [57]. Moreover, the density of the paste decreases (darker background on the SEM image).

Zone 3 shows an enrichment in phosphorus with an intense
decalcification and a decrease of the aluminium content. However, no
peak of a crystallized P-bearing phase is observed on the diffraction
pattern. The total oxides content decreases from 65 to 50%. The
mineralogical analysis shows intense peaks of AH₃, the broad bases of
which characterize the poorly crystallized character of this phase. The
density of the peripheral zone remains relatively high.

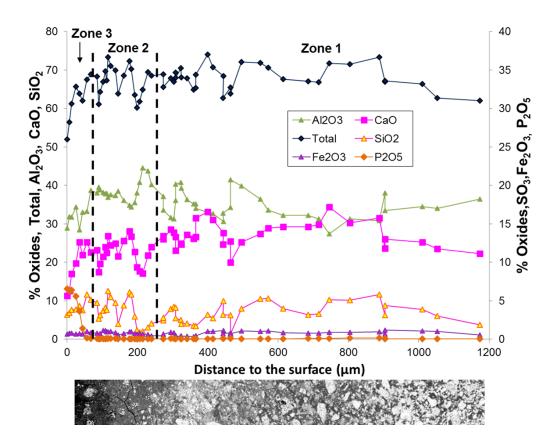


Figure 9. Chemical composition of the CAC paste after 10 weeks of immersion according to the distance to the surface (EPMA) (top), and SEM observations of the polished section in BSE mode (bottom)

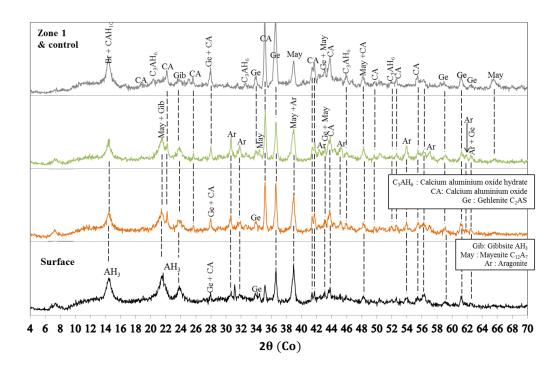


Figure 10. Mineralogical analysis by XRD of the CAC paste immersed for 5 weeks in digesting biowaste

4. DISCUSSION

4.1 Chemical and microbial composition of the liquid fraction of the digesting biowaste

The liquid fraction contained in lab-scale anaerobic bioreactor has properly undergone the consecutive reactions of the anaerobic digestion process. First, the hydrolysis and acetogenesis steps led to a decrease of the pH and an increase of the VFAs concentration: the total VFAs concentration reached about 75 mmol.L⁻¹ and the pH dropped to 4.3. The VFAs metabolized in the digesting biowaste were acetic, propionic and butyric acids, which are acids with soluble calcium salts. Then, the acetogenesis and methanogenesis steps gave rise to consumption of the VFAs and an increase to the neutrality area of the pH, which reached 7.8 at the end of the digestion cycle. The ammonium concentration

increased quickly during the first few days of digestion and then stabilized at values around 55 mmol.L⁻¹. Both cycles showed similar aggressive conditions in terms of concentrations of aggressive metabolites (VFA, ammonium and dissolved CO₂) and pH values. This means that concrete structures in which anaerobic digestion takes place will undergo repeated aggression when they are replenished in fed-batch, or a continuous aggression in the case of continuously-fed digesting installation.

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These concentrations of aggressive metabolites in the liquid phase of the bioreactors give information on the aggressiveness of the environment for concrete. Table 3 summarizes the limit values for solutions according to the exposure classes as given in FD P 18-011 [31] for the concentrations of aggressive CO₂ and the ammonium, together with the pH. Considering that, for each cycle, the pH increased to values up to 6.5 after 7 days, the pH of the anaerobic digestion in normal operation should not lead to aggressive conditions for concrete according to the French information document FD P 18-011 [31] where the upper limit of pH of the XA1 exposure class is 6.5. The standard documents [30,31] also take the presence and the concentration of ammonium and dissolved CO2 into account. The maximum concentration values measured during the anaerobic digestion (55 mmol.L⁻¹ for ammonium and 2.3 mmol.L⁻¹ for dissolved CO₂) made the liquid phase of the anaerobic digestion a highly aggressive medium for cement based materials (class XA3 according to Table 3): the maximum concentration of ammonium measured was more than 7 times the upper limit considered in the XA3 range (3.33 – 5.55 mmol.L⁻¹) and the dissolved CO₂ concentration was above the lower limit of the XA3 class (1.67 mmol.L⁻¹). It also can be noticed that ammonium and bicarbonate concentrations of several grams per liter have been found in fully functioning industrial digesters [58-60].

Moreover, it can be highlighted that neither document considers the presence and concentration of VFA, even though several authors [17,33] have shown that the acid concentration is a much more significant parameter than the pH in the evaluation of the aggressiveness of certain types of acids, including organic acids.

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account for the exposure classes.

The presence and impact of microorganisms, in the form of a biofilm or not, are not considered either by the FD P18-011 [31] regulation. However, the establishment of a microbial biofilm between the 5th and the 10th week was clearly a factor increasing the deterioration kinetics of the CEM I-Ox paste. In contrast, the CAC paste was particularly resilient to the biofilm proliferation and was the material that showed the strongest resistance against the biodeterioration. Several studies agree that microorganisms are acting as accelerating agents on the deterioration of concrete, aggravating its biodeterioration [7,8,28,29,61]. More particularly, a study by Magniont et al. [28] has highlighted the clear influence of a biofilm formation of *E. coli* intensifying the CEM I degradation. In addition, it was also demonstrated that the ability of bacteria to create a biofilm could be very harmful for concrete structures [5,28]. According to the non-aggressive pHconditions, the dissolution of the external zone (Si-Al gel) of the CEM I paste presently observed after 10 weeks of exposure can be explained by particularly severe local conditions brought by the establishment of a biofilm at the surface. Lastly, the standard considers temperatures between 5 and 25 °C while the anaerobic digestion process occurs mostly at mesophilic conditions (around 37 °C) in the industrial sector [11]. Even if a rise in temperature increases reaction rates and finally the concrete degradation kinetics, this factor is not taken in

Aggressive agents		ness class a NF EN 206 [30	In the biowaste studied	
	XA1	XA2	XA3	
Aggressive CO ₂ (mg.L ⁻¹)	≥ 15 and ≤ 20	> 40 and ≤ 100	> 100	101
NH₄⁺ (mg.L⁻¹)	≥ 15 and ≤ 30	> 30 and ≤ 60	> 60 and ≤ 100	990
рН	≤ 6,5 and ≥ 5,5	< 5,5 and ≥ 4,5	< 4,5 and ≥ 4,0	7.5

4.2 Performances of tested materials in relation with biofilm-materials interaction phenomena

The CEM I paste showed the highest deterioration kinetics, with the greatest degraded depths after 5 and 10 weeks and complete biofilm coverage observed at 5 weeks of exposure. The early microbial colonization of the CEM I paste could be explained by the topology of the surface of CEM I, especially the abundance of micrometric holes, which Voegel et al. [27] found to be favourable to adhesion and microbial proliferation.

The oxalic acid treatment has resulted after 5 weeks in a less intense degraded depth than in the CEM I specimen, a chemical composition close to that of a sound sample, and no biofilm coverage. This resistance against the microbial biofilm growth could come from the surface texture created on the CEM I treated by oxalic acid: according to Voegel et al. [27], considering the characteristics of its surface height distribution, the CEM I-Ox paste covered by calcium oxalate was less favourable to microbial growth. However, after 10 weeks of immersion, the salt was no longer observed and the sample was covered by biofilm. The CEM I-Ox paste then showed the same degradation mechanisms as the CEM I

paste, with a large degraded depth. Thus, the treatment seemed to have a protective effect for a limited duration only: it allowed degradation to be delayed by delaying the biofilm formation on the surface, which underlines the worsening effect of this formation. The oxalic acid surface treatment probably provided two types of protection: the delay of biofilm formation and the presence of a layer of calcium oxalate, which was initially stable against acid attacks, thus protecting the matrix, but dissolved in the short term, revealing a surface composition close to that of the non-treated CEM I favourable to microbial proliferation.

Although the CAC pastes presented surfaces favourable to proliferation [27], this

material showed the highest durability, with the smallest degraded depths after 10 weeks and only a non-uniform weak surface coverage by a microbial biofilm. The anhydrous phases of the CAC paste were the most preserved and the aluminate cement showed some peaks of crystallized AH₃. However, a halo baseline characteristics of amorphous compounds is also observed on the surface layer.

4.3 Biodeterioration mechanisms

Figure 11 summarizes the main chemical and mineralogical changes in the CEM I and CAC pastes exposed to the anaerobic digesting biowaste for two cycles, together with exposure conditions and degraded depths.

CEM I CAC

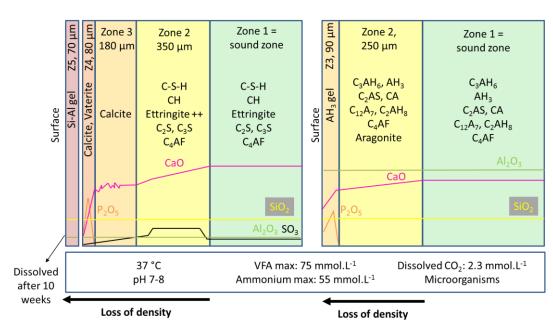


Figure 11. Chemical and mineralogical changes and degraded depths of CEM I and CAC pastes exposed to the liquid-solid phase of an anaerobic digestion bioreactor after 10 weeks of immersion and their conditions of exposure.

4.3.1 CEM I paste

In accordance with Voegel et al. [7], the analyses for the ordinary Portland cement showed alteration mechanisms corresponding to a combination of (i) leaching resulting from the exposure of cementitious materials to VFA [62] and ammonium [20], and (ii) carbonation due to the presence of aqueous CO₂ [21]. As the biowaste contained little or no sulfur, it is likely that the relative enrichment in sulfur at depth, expressed by the formation of secondary non-expansive ettringite, was due to the movement of sulfur from the outer layer, where S-bearing phases were dissolved, into zone 2. The peak of phosphorus in zone 4 was associated with the presence of phosphorus in the substrate [7]. Unlike the CAC paste, the CEM I paste was significantly affected by biofilm formation, leading to an external zone that was totally decalcified: anhydrous grains were completely dissolved after only 5 weeks of immersion, from zone 5 to zone 3.

4.3.2 CAC paste

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In contact with biowaste, the CAC paste underwent chemical and mineralogical changes. Several phenomena were identified, such as surface decalcification, (incomplete) dissolution of the anhydrous grains and the hydrates on the surface, precipitation of calcium carbonates at depth and a phosphorous peak in the intermediate zone. It could also be noted that the peripheral zone maintained a relatively high density. The fact that the CAC paste performs best could be linked to lower colonization of the CAC pastes compared to other binders [63,64]. Herisson et al. [65] explained this by the bacteriostatic effect of aluminium, present in large amounts in the CAC cements. It should be noted that, when considering very aggressive sulfur oxidizing conditions comparable to those in sewer networks, Buvignier et al. [61] showed no direct effect of the aluminium on the bacteria responsible for the acid attack, which the authors attributed to the adaptation of the microbial biofilm to high Al concentrations. The particular effect of CAC on microbial biofilm development was also observed in different conditions by Peyre Lavigne et al. [66] (sulfur-oxidizing biofilms) and Dalod et al. [63] (photosynthetic biofilms). Results of both studies suggested that the biofilm development and/or activity was slown down on the CAC substrates compared to the one grown on ordinary binder based materials. Moreover, Peyre Lavigne et al. [66] showed that the biofilm on CAC was less dense, and drier, and with lower microbial diversity compared to biofilm grown on other binders. Moreover, according to the literature, the formation of an aluminium gel (AH₃) on the surface leads to better durability of calcium aluminate cement matrices, since this gel is known to be stable at moderately acidic pH [37].

5. CONCLUSION

In this study, cement paste specimens were exposed to the anaerobic digestion liquid phase of laboratory reactors for 10 weeks. At the end of the experiment, all the cementitious materials tested: made of CEM I (ordinary Portland cement), CAC (calcium aluminate cement) or CEM I-Ox (CEM I treated with oxalic acid) were covered by biofilm and presented biodeterioration. The pastes were affected by alteration mechanisms including leaching, carbonation and phosphorus enrichment. However, it should be noted that differences of behaviour were highlighted according to the material. The treatment with oxalic acid seemed to protect the specimen from biodeterioration by preventing the formation of biofilm during the first 5 weeks. Calcium oxalate therefore seems to have a proliferation inhibiting effect, at least in the short term. After this period, CEM I and CEM I-Ox followed the same degradation kinetics and mechanisms. with the formation of secondary non-expansive ettringite at depth. CAC paste showed the best durability performances and smallest degraded depth in the long term. Moreover, the anhydrous phases of the CAC paste were the best preserved and some of them were still present in the surface layer. This good durability could be linked to a less dense coverage by the biofilm but probably also to the CAC mineralogy and to the formation of a peripheral gel of AH₃, stable at moderately acidic pH.

6. ACKNOWLEDGEMENTS

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