

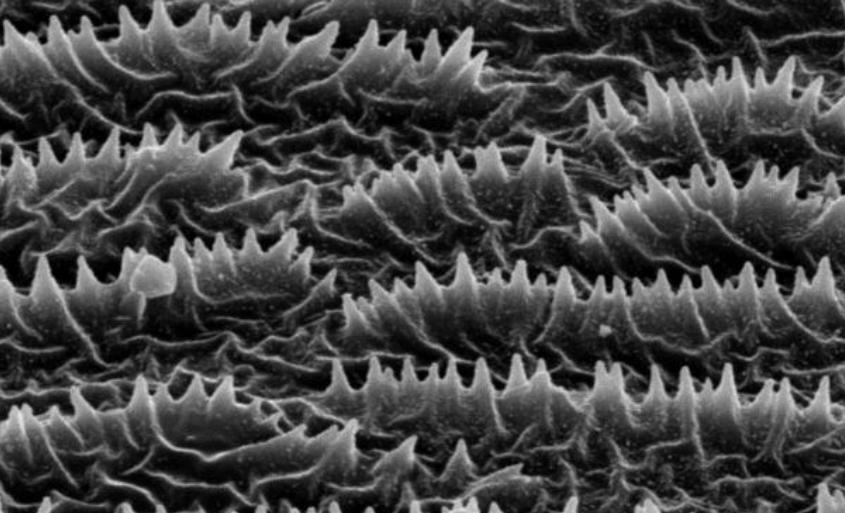


**INRA**  
SCIENCE & IMPACT



Yvan Rahb  

abdomen



# Des stylet(s) à leur cuticule: transcriptomique et genomique des prot  ines cuticulaires de puceron

10 µm

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CBI 2019, ALBI Campus Champollion  
26-28 Juin 2019



# Plan

1

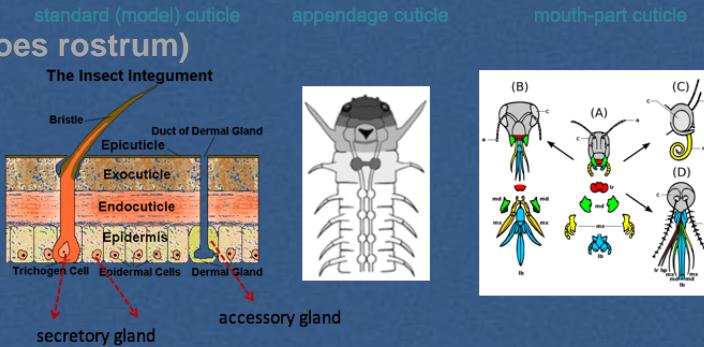
- ❖ Introduction: the biological issue, or why do we need stylet « analytics » ?
  - ❖ What is really « stylet transcriptomics » ?: Retort organ revisited.
  - ❖ Two (x2) stylets for one bundle = maxillary vs mandibular.
- 2
- ❖ What is a cuticular protein ? Domain conservation is chitin-binding
  - ❖ Cuticular protein profiles and CuP (=CP) compositional complexity: Shannon again.
- 3
- ❖ Compositional bias in cuticular proteins: Structure, composition, function.
  - ❖ Parsing genome for compositional bias: A brand new world ? (in retort organs)
  - ❖ Conclusions: Homology and orthology in cuticular proteins

# The biological issue

- ❖ Hemipteran stylet and mouth cuticle territories do bind microbes:
  - viruses (*non-circulative* , CaMV... -aphids-)
  - bacteria (*Xylella* -sharpshooters-)
- ❖ What is mouthpart cuticle made of ? Structure ? Components ?
  - cuticular proteins ?
  - lipids ?
- ❖ How to achieve such characterization ?

# Analysis of mouthpart cuticular proteins

Very scarce data at this time (a single analysis on mosquitoes rostrum)



- ❖ Combined analysis with proteomics, transcriptomics & genomics (structural, annotational, comparative).
- ❖ The advantage of transcriptomics (in identifying the main -gene- players)
  - Sensitivity
  - Insensitivity to post-translational modifications
  - Expression (levels), quantitative: therefore a partial link to function

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# Stylet transcriptomics

Retort organ revisited



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# The retort organ in space

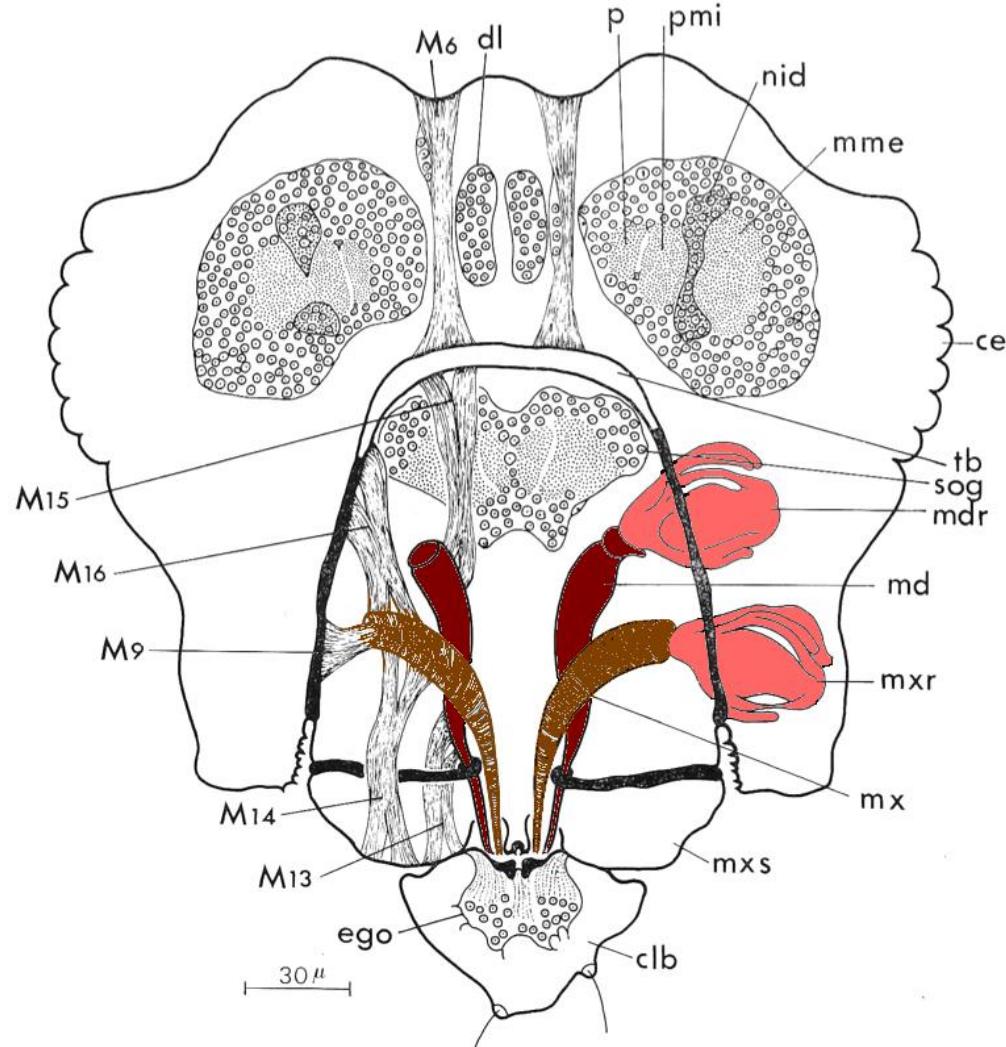
A couple of glands to re-discover

❖ Ponsen (1972)

- For *Myzus persicae*
- Partially for other aphids

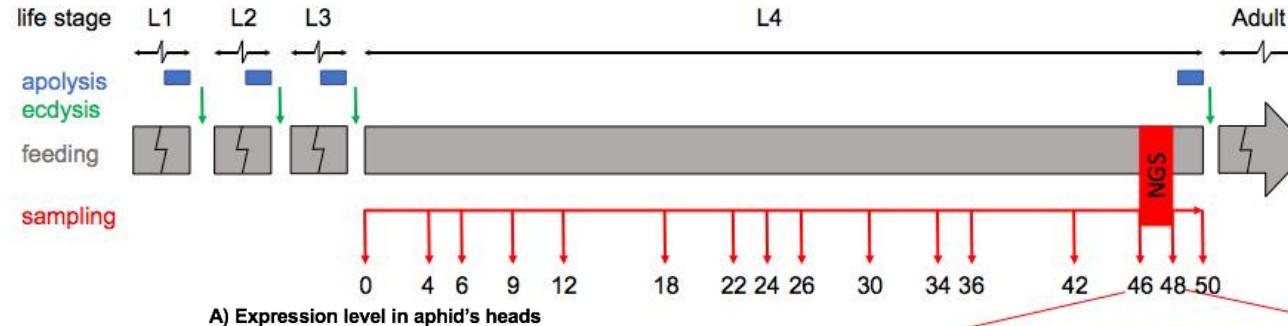
❖ Cicero (2016)

- For a psyllid
- Cicero JM: Stylet biogenesis in *Bactericera cockerelli* (Hemiptera: Triozidae). Arthropod Structure & Development 2016.

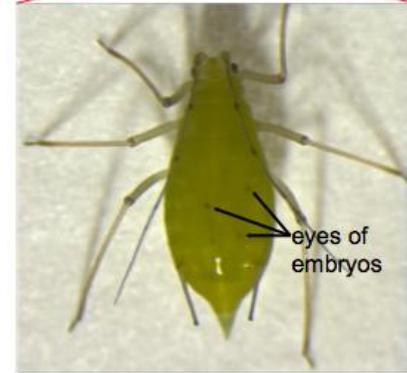
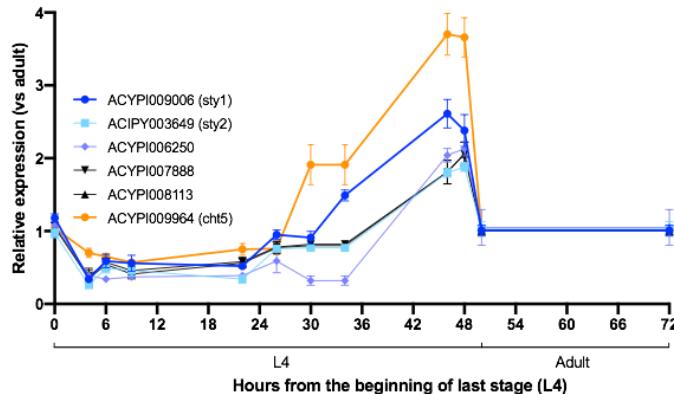


# The retort organ

in time



A) Expression level in aphid's heads



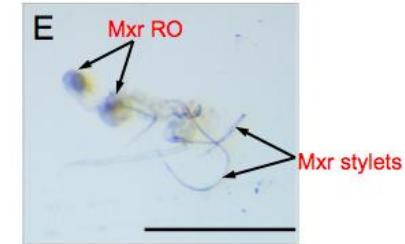
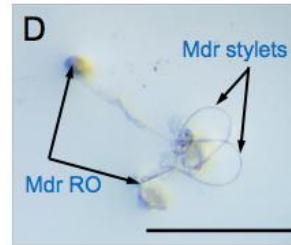
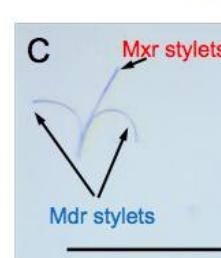
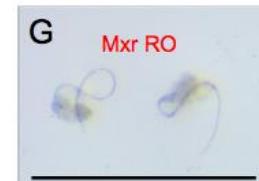
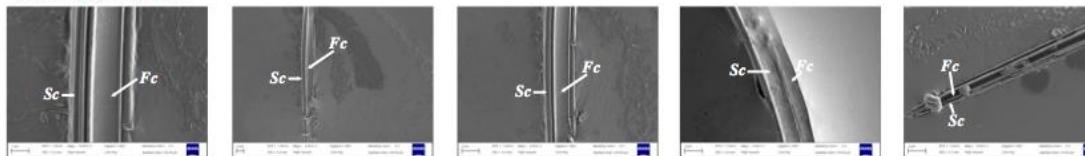
# The retort organ(s)

*in vitro: dissection of maxillary and mandibular stylet glands*

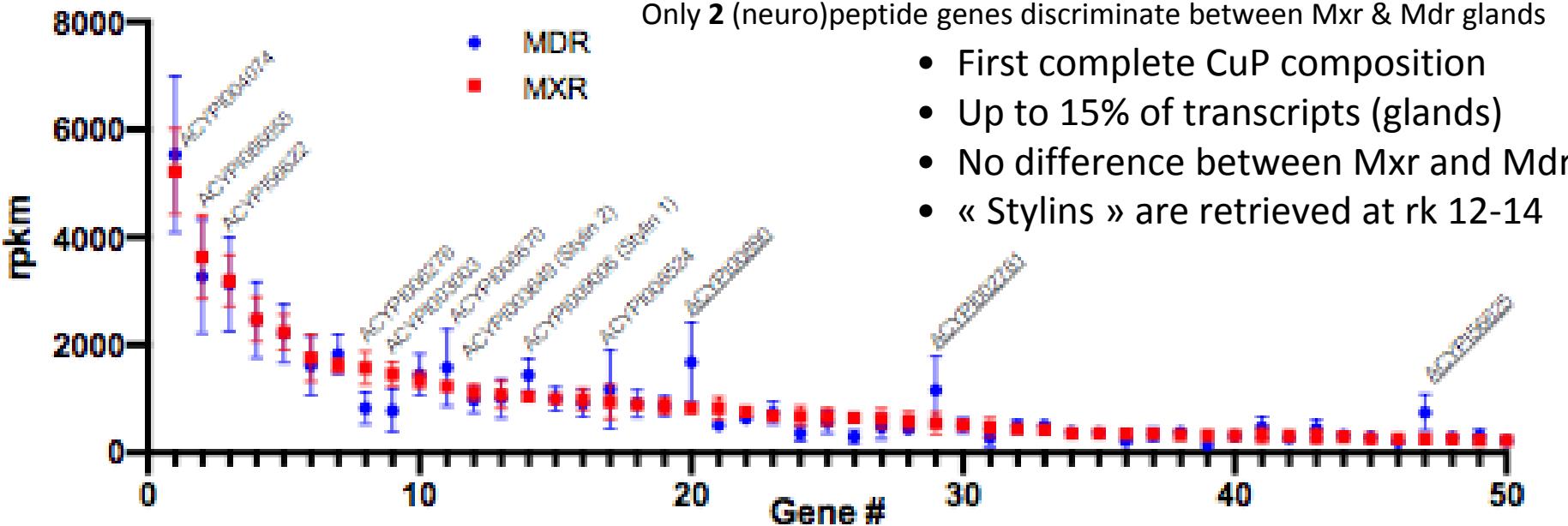
Mandibular stylets:



Maxillary stylets:

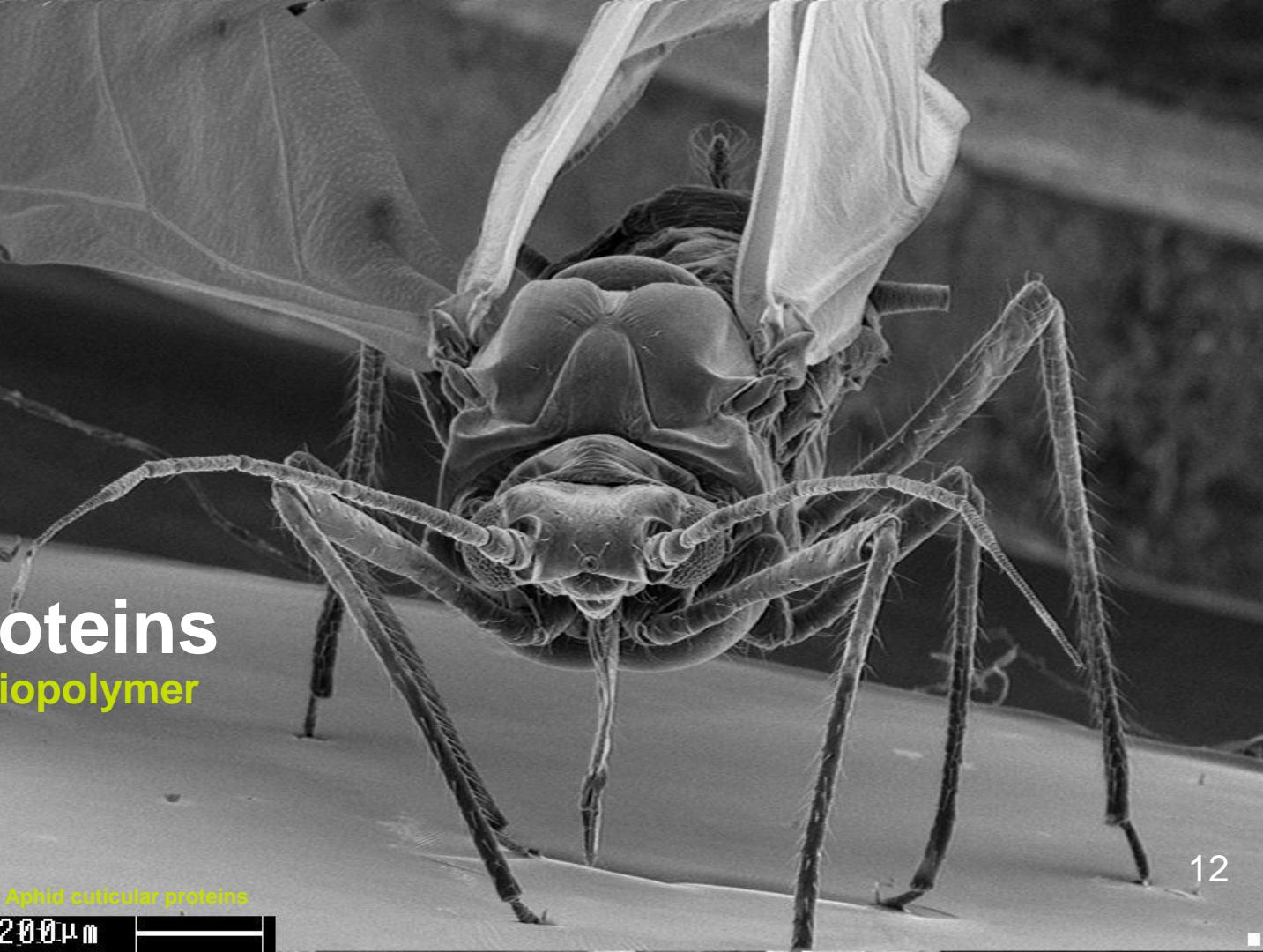


# The retort organ in (cuticular) proteins

**C**

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## Cuticular proteins and Cuticle Biopolymer



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# What are cuticular proteins ?

## in aphids

Basically: Proteins with a (set of specific) chitin-binding domain(s)

Table 5

Proteins<sup>a</sup> identified from 14 arthropod proteomes using CutProtFam-Pred (see Section 2.6).

Family	<i>Drosophila melanogaster</i>	<i>Glossina morsitans</i>	<i>Culex quinquefasciatus</i>	<i>Aedes aegypti</i>	<i>Anopheles gambiae</i>	<i>Bombyx mori</i>	<i>Danaus plexippus</i>	<i>Tribolium castaneum</i>	<i>Apis mellifera</i>	<i>Nasonia vitripennis</i>	<i>Acyrthosiphon pisum</i>	<i>Pediculus humanus corporis</i>	<i>Daphnia pulex</i>	<i>Tetranychus urticae</i>
CPR_RR-1 <sup>b</sup>	61	33 <sup>c</sup>	49	66	43	47	47	34	13	19	9	9	101 <sup>c</sup>	0
CPR_RR-2 <sup>b</sup>	42	27 <sup>c</sup>	97	150	103	78	57	55	15	32	84	15	36	7
CPR_UncL <sup>b</sup>	34 <sup>a</sup>	17	30	28 <sup>c</sup>	21 <sup>c</sup>	19	18 <sup>c</sup>	21 <sup>c</sup>	10	18 <sup>c</sup>	20 <sup>c</sup>	17	152 <sup>c</sup>	31 <sup>c</sup>
CPAP1	29	11	10	14	13	13	16	13	15	16	10	12	20	14
CPAP3	10	6	8	9	10	6	10	7	7	6	8	6	12	5
CPCFC	1	1	1	1	1	1	1	2	0	0	1	1	0	0
CPF	5	1	5	3	4	1	1	5	4	5	2	1	0	0
CPLCA	13	9	3	3	3	2	1	1	0	0	0	0	0	0
CPLCG	4	4	41	18	25	0	1	1	0	0	0	0	0	0
CPLCW	0	0	0	7	10	0	0	0	0	0	0	0	0	0
Tweedle	29	9	9	6	12	4	5	3	2	2	3	2	0	0
Total	228	118	253	305	245	171	157	142	66	98	137	63	321	57

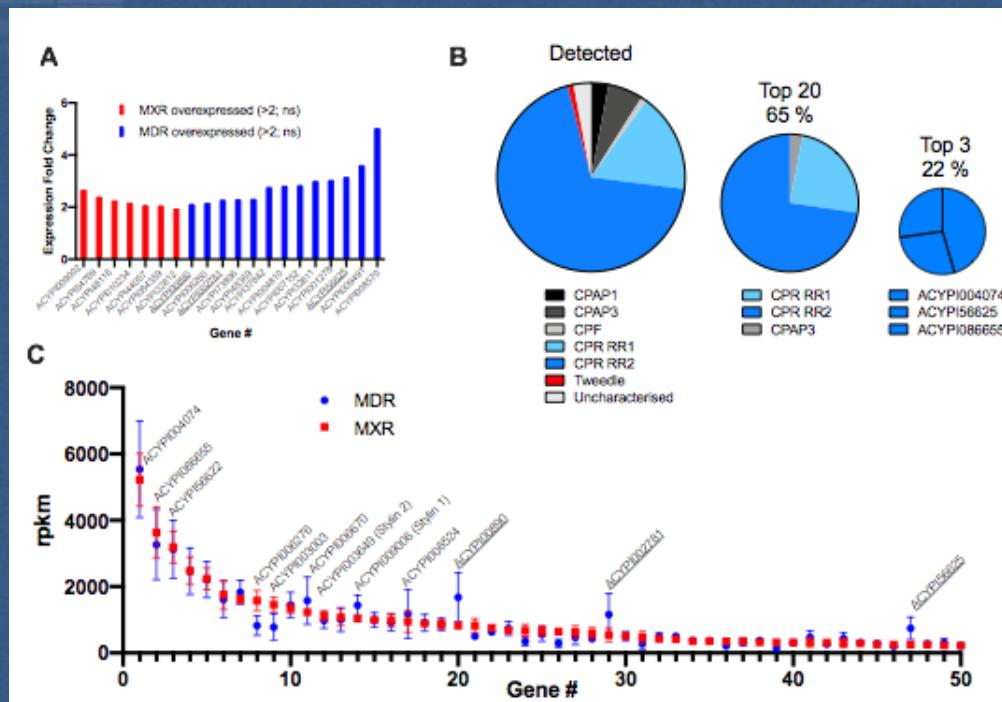
<sup>a</sup> Numbers include all splice variants. See Supplementary File 2 and Supplementary File 3 for values based on gene numbers.

<sup>b</sup> Sequences that score equal or above the assigned score cutoffs for the CPR\_RR-1 and CPR\_RR-2 models are classified in the corresponding family, while sequences that score below the assigned score cutoffs and above 0 for

# Cuticular protein compositional complexity in aphid stylets

Shannon's diversity index

$$H' = - \sum_{i=1}^R p_i \ln p_i$$

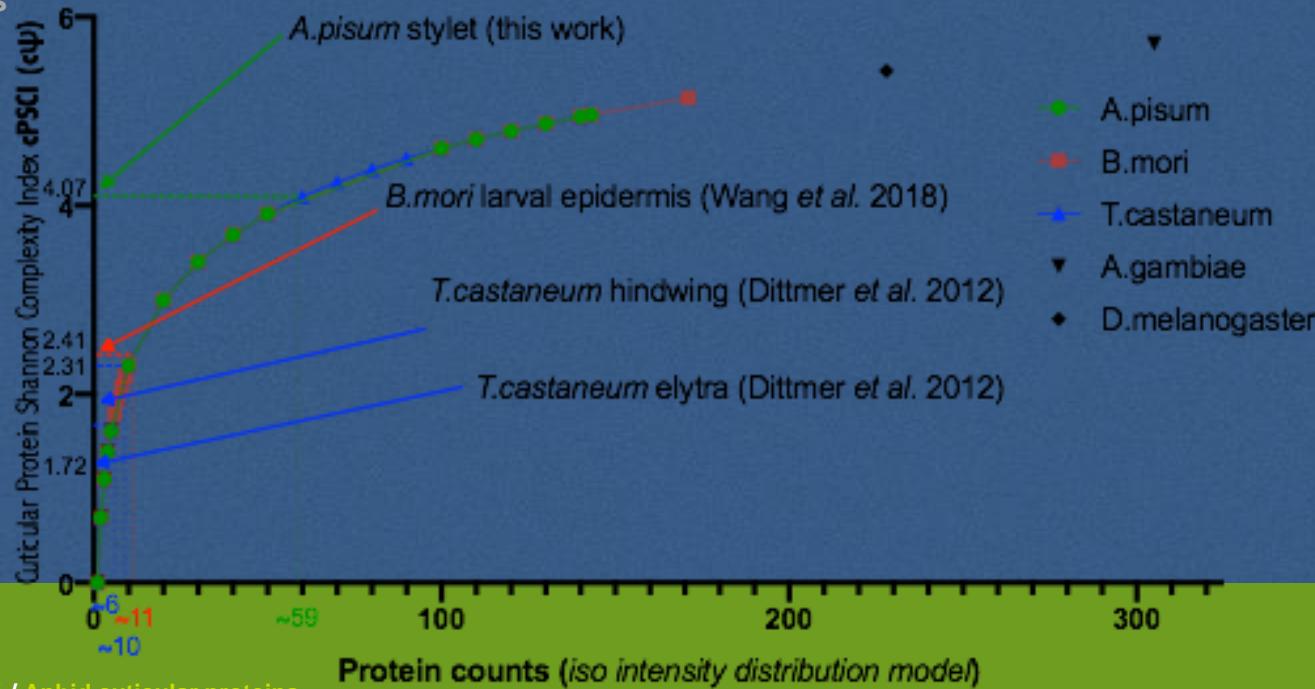


# Cuticular protein compositional complexity

## in insect cuticles

Figure 11: cψ Plots

Compared Complexity  
of insect cuticles  
analyzed for  
whole genome transcripts



# \_03

## Compositional bias vs homology domains

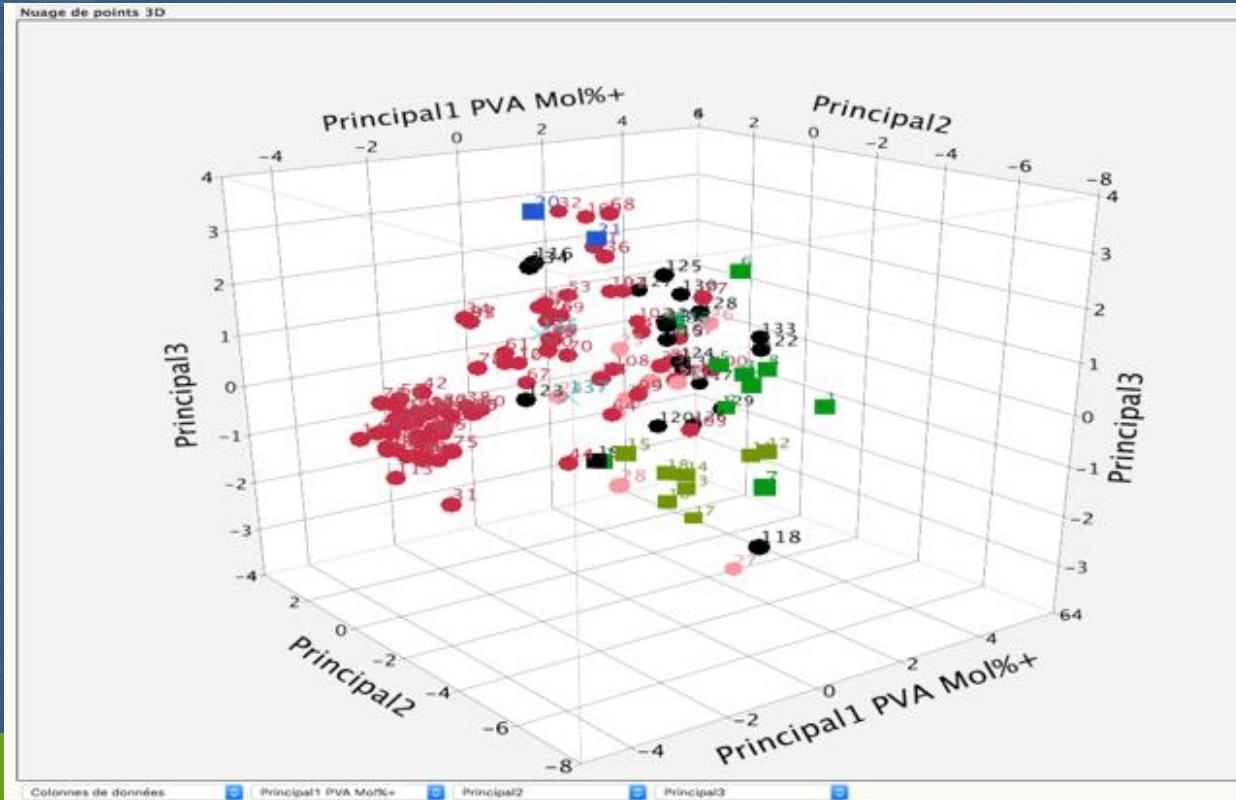


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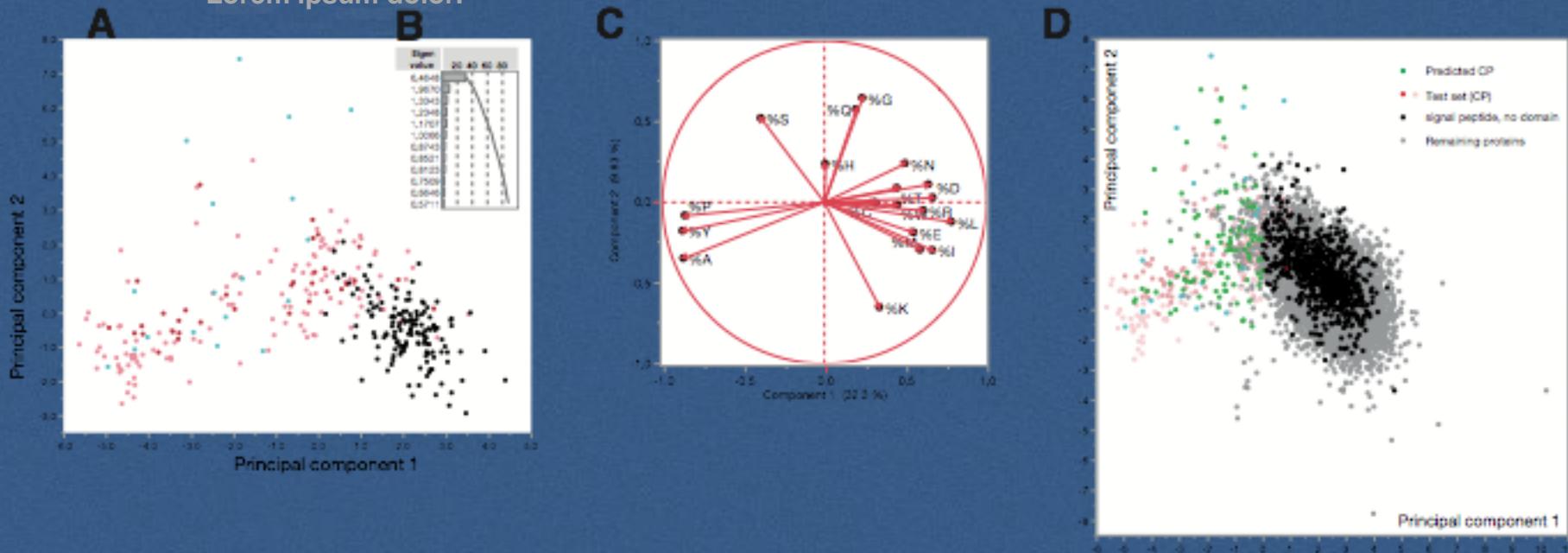
# Compositional bias in cuticular proteins

## in « standard » (recognized) cuticular proteins



# Compositional bias in (cuticular?) proteins in aphid retort organ and aphid whole genome

Lorem ipsum dolor.





## sample sequences

>ACYPI087730 grydary

EEAKKETKVVEKRSDKSDLTPAATGRSFLGGYNSYPTGYSSGYNSGYNSYPSTYSGGYSSYPSYSSYPSSGY

>ACYPI082258 «paysa»ine

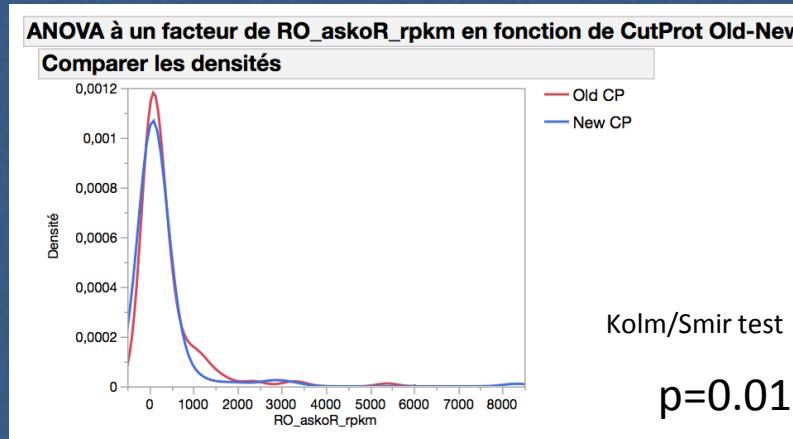
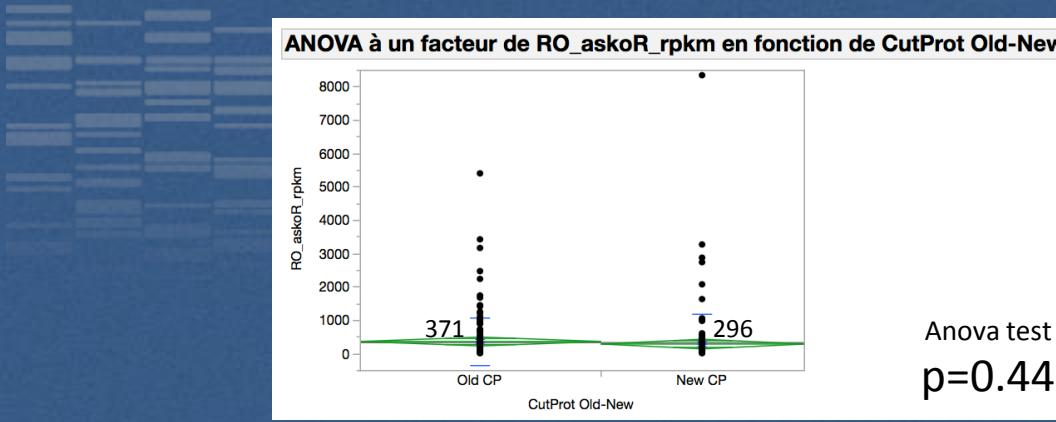
QEAPKAEEQVGAEAARVKKHAYIAAPALAYSSPAYPYAYSAYPSAYPSYSYPAAYPTAYASYPSYAYAHDDGKY

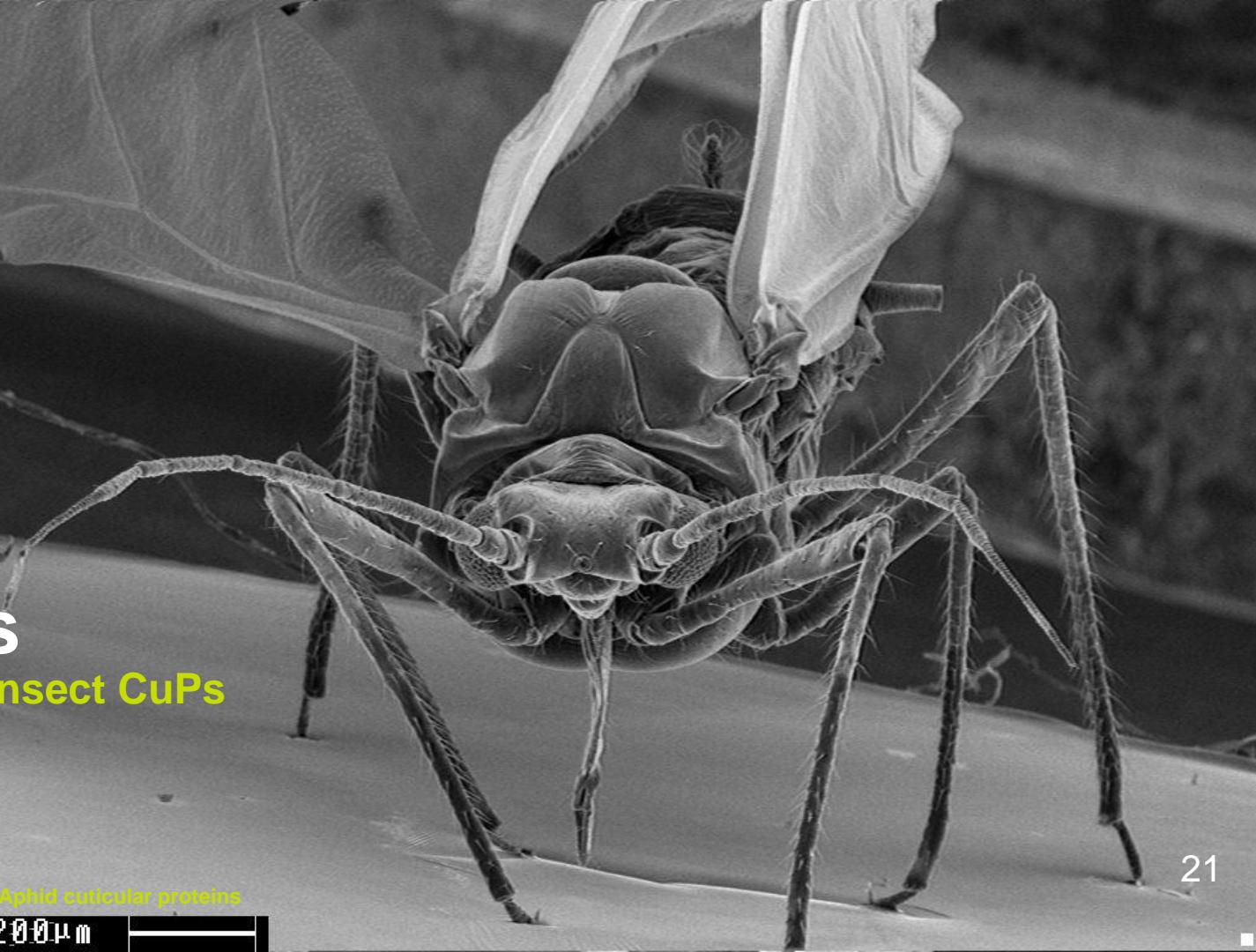
>ACYPI001038 hyperstylin

APSHGFISPVLPKVHLASPDVTVRQPYVVQQTEPVFRHVYYAGDSVPYVSHVQAYHQHPAVVAAYQTAPA

>J9JUP8\_ACYPI

SVHSINFSVYSISTVHNTLTKMAAKLIIFTVCVASAFAQYSAPAYKPAYSSPAYAAPAYA  
APKAYAPEPAYAPAPYNEFYSVNDPHTYDVKSQSEYSDGNGYVKGSYSLLLEADGST  
YTADDYNGFNAVVKNEGGYKAPAYAAPAYAAPAYSAPAYKPAYSAPAYSAPAYKPAY  
AYSAPAYSAPAYKPAYKPAY



A detailed scanning electron micrograph (SEM) of an aphid, showing its segmented body, long antennae, and six legs. The aphid is positioned against a dark background, highlighting its intricate surface textures and appendages.

# \_the end

## Conclusions

homology in insect CuPs

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# Yvan Rahbé, Natalia Guschinskaya and Marilyne Uzest



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Natalia Guschinskaya, Denis Ressnikoff, Karim Arafah, Sébastien Voisin, Philippe Bulet, Marilyne Uzest, Yvan Rahbé. Insect mouthpart transcriptome unveils extension of cuticular protein repertoire and complex organization, *iScience*, in revision.

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Yvan Rahbé / **Aphid cuticular proteins**

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