



HAL
open science

Functional Mechanism of the Efflux Pumps Transcription Regulators From *Pseudomonas aeruginosa* Based on 3D Structures

Karim Housseini B Issa, Gilles Phan, Isabelle Broutin

► **To cite this version:**

Karim Housseini B Issa, Gilles Phan, Isabelle Broutin. Functional Mechanism of the Efflux Pumps Transcription Regulators From *Pseudomonas aeruginosa* Based on 3D Structures. *Frontiers in Molecular Biosciences*, 2018, 5, 10.3389/fmolb.2018.00057 . hal-02150022

HAL Id: hal-02150022

<https://hal.science/hal-02150022v1>

Submitted on 6 Jun 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Functional Mechanism of the Efflux Pumps Transcription Regulators From *Pseudomonas aeruginosa* Based on 3D Structures

Karim Housseini B Issa, Gilles Phan and Isabelle Broutin*

Laboratoire de Cristallographie et RMN Biologiques (UMR 8015), Centre National de la Recherche Scientifique, Faculté de Pharmacie, Université Paris Descartes, Université Sorbonne Paris Cité, Paris, France

OPEN ACCESS

Edited by:

Vassily Bavro,
University of Essex, United Kingdom

Reviewed by:

Zhao Wang,
Baylor College of Medicine,
United States
Anastassios C. Papageorgiou,
University of Turku, Finland

*Correspondence:

Isabelle Broutin
isabelle.broutin@parisdescartes.fr

Specialty section:

This article was submitted to
Structural Biology,
a section of the journal
Frontiers in Molecular Biosciences

Received: 28 February 2018

Accepted: 31 May 2018

Published: 19 June 2018

Citation:

Housseini B Issa K, Phan G and
Broutin I (2018) Functional Mechanism
of the Efflux Pumps Transcription
Regulators From *Pseudomonas*
aeruginosa Based on 3D Structures.
Front. Mol. Biosci. 5:57.
doi: 10.3389/fmolb.2018.00057

Bacterial antibiotic resistance is a worldwide health problem that deserves important research attention in order to develop new therapeutic strategies. Recently, the World Health Organization (WHO) classified *Pseudomonas aeruginosa* as one of the priority bacteria for which new antibiotics are urgently needed. In this opportunistic pathogen, antibiotics efflux is one of the most prevalent mechanisms where the drug is efficiently expelled through the cell-wall. This resistance mechanism is highly correlated to the expression level of efflux pumps of the resistance-nodulation-cell division (RND) family, which is finely tuned by gene regulators. Thus, it is worthwhile considering the efflux pump regulators of *P. aeruginosa* as promising therapeutic targets alternative. Several families of regulators have been identified, including activators and repressors that control the genetic expression of the pumps in response to an extracellular signal, such as the presence of the antibiotic or other environmental modifications. In this review, based on different crystallographic structures solved from archetypal bacteria, we will first focus on the molecular mechanism of the regulator families involved in the RND efflux pump expression in *P. aeruginosa*, which are TetR, LysR, MarR, AraC, and the two-components system (TCS). Finally, the regulators of known structure from *P. aeruginosa* will be presented.

Keywords: multidrug resistance, efflux pumps regulators, activator, repressor, X-ray structures

INTRODUCTION

Just after the introduction of antibiotics on the market in the mid-twentieth century, bacterial resistance was recognized as a natural but worrisome phenomenon (McDermott et al., 2003; Hede, 2014). More than 60 years later, the resistance is still a worldwide health concern (Frieri et al., 2017), threatening the effectiveness of antibacterial therapy, and challenging the efforts of developing novel antibiotics (Li et al., 2015), but fortunately some studies tend to give hope in this research area (D'Costa et al., 2011; Rolain et al., 2016).

To survive, bacteria have developed an inexhaustible range of antibiotic resistance mechanisms (Coates et al., 2002; Levy and Marshall, 2004). One of them involves the efflux of toxic compounds through bacterial cell-wall by membrane-bound protein transporters called multidrug efflux pumps (Poole and Srikumar, 2001; Rahman et al., 2017). These multidrug efflux systems (MES) existed in

bacterial genomes long before the use of antibiotics by human to cure infection (Davies and Davies, 2010). MES are essential in bacterial physiology and natural defenses (Poole, 2008; Li and Nikaido, 2009; Alvarez-Ortega et al., 2013; Blanco et al., 2016), including export of organic solvent, detergents, fatty acids, toxic lipids and quorum sensing molecules. Because many structurally unrelated compounds are extruded by the same system, MES are also responsible for the multidrug resistance (MDR) phenotype (Nikaido, 2009). Efflux pumps have been categorized into five different families (Li and Nikaido, 2009), based on three criteria: the amino acid sequence identity, the energy source required to drive export and the substrate specificities (Li et al., 2015). The five major known families are the ATP-binding cassette (ABC) (Szakács et al., 2008; Locher, 2009), the small multidrug resistance (SMR) (Schuldiner, 2009), the major facilitator superfamily (MFS) (Kumar et al., 2013; Yan, 2015), the resistance-nodulation-cell division (RND) (Du et al., 2014; Yamaguchi et al., 2015; Daury et al., 2016; Vargiu et al., 2016) and the multidrug and toxic compound extrusion (MATE) (Hvorup et al., 2003; Kuroda and Tsuchiya, 2009). ABC superfamily belongs to the primary active transporters class which function depends on ATP hydrolysis, whereas the other pumps are secondary active transporters (symporters, antiporters, and uniporters) using energy from proton and/or sodium gradient (Mousa and Bruner, 2016).

Multidrug efflux transporters overexpression is tightly regulated by transcriptional activators and/or repressors upon the presence of toxic compounds (Sun et al., 2014). Interestingly, the regulators themselves are potentially triggered by the substrate that will be transported in turn by the regulated pumps (Schumacher and Brennan, 2002). A very specific and imbricated regulation system seems to link the transcriptional regulators to the cognate efflux pumps expression. In order to combat antibiotic resistance, all the different resistance mechanisms must be targeted, and despite recent encouraging results (Fair and Tor, 2014; Khameneh et al., 2016; Cheesman et al., 2017), multidrug transporters remain largely responsible for antibiotherapy failures (Sun et al., 2014). Because the last discovered antibiotic is specific to Gram positive (Gram+) bacteria (Ling et al., 2015), it is urgent to find new drugs targeting the Gram negative (Gram-). This is also supported by the fact that most of the main problematic multiresistant pathogens in hospitals or “ESKAPE”

bacteria are Gram- (Tacconelli et al., 2017). This acronym comes from the initials of six superbugs which are *Enterococcus faecium* and *Staphylococcus aureus*, both Gram+, and the Gram- *Klebsiella pneumoniae*, *Acinetobacter baumannii*, *Pseudomonas aeruginosa* and *Enterobacter*. The particularity of Gram- bacteria is the presence of two membranes and a soluble space in between called the periplasm. Consequently, the efflux pumps must cross two lipidic barriers and the periplasm to transport the molecules out of the cell. This can be achieved by RND transporters that represent the major drug efflux pumps in Gram- bacteria, and more specifically by the hydrophobe/amphiphile efflux 1 (HAE1) sub-family (Nikaido, 2018) in which the transporter is a homotrimer that belongs to a tripartite complex. These efflux pumps are constituted of three different proteins forming an elongated nanomachine. The transporter itself, called RND, is localized in the inner membrane. It is the motor of the pump activated by the proton motive force. Another partner protein called Outer Membrane Factor (OMF) is embedded in the outer membrane. The third protein called Membrane Fusion Protein (MFP) is localized at the periplasm with a lipidic anchor inserted in the inner membrane. The 3D structure of the whole assembly has been solved recently by cryo-EM (Du et al., 2014; Daury et al., 2016; Wang Z. et al., 2017) highlighting how the different protein partners interact. Similar assembly architecture is also observed in the ABC family despite a different oligomeric organization of the transporter (Fitzpatrick et al., 2017). The fact that the HAE1-RND efflux system is only found in Gram- makes them interesting and specific target. Nevertheless, more than 10 years of research on this transporter family has not provided active and non-toxic drug yet. Thus, targeting the efflux pump expression regulation appears as an attractive alternative. Prokaryotic transcriptional regulators are classified into two groups: the one-component system and the two-components system. The gene expression regulation of the same HAE1-RND could involve both systems. A better comprehension of the molecular basis of efflux pumps genes expression is highly needed to pave the way for the design of new drugs toward multidrug resistance by efflux pump.

This review will focus on the regulators of the HAE1-RND efflux pumps (later called RND for simplification) involved in drug-resistance, with a particular focus on the regulator families controlling the pumps of *P. aeruginosa*, one of the most difficult bacteria to treat in clinic. For each family of the two-component and one-component regulators, a description of the molecular mechanism will be given based on structural knowledge obtained from archetypal organisms.

THE REGULATION OUTLINE OF *P. AERUGINOSA* RND EFFLUX PUMPS

Pseudomonas aeruginosa is an opportunistic bacterium that has the ability to rapidly grow in diverse environmental niches, from different soils to human respiratory tract. It is involved in severe human diseases like meningitis, septicaemia or cystic fibrosis and is also a major cause of nosocomial infections due to its high capacity to develop resistances (Poole, 2011). One of

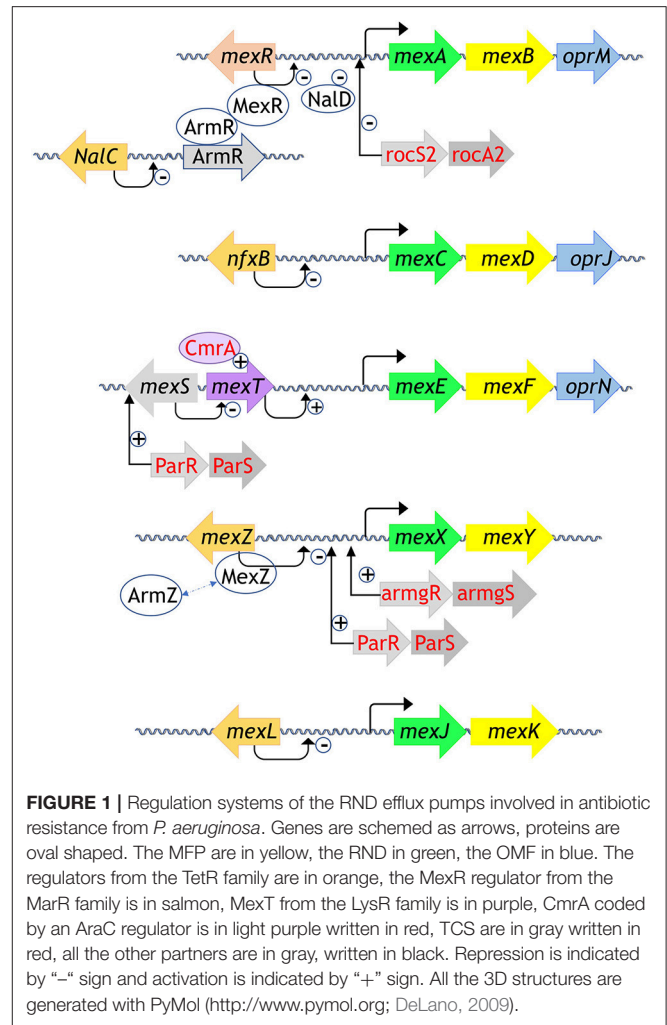
Abbreviations: ABS, Activation binding site; ABC, ATP-binding cassette; CA, Catalytic and ATP binding domain; DBD, DNA-Binding Domain; DHp, Dimerization and histidine phosphotransfer domain; GAF, cGMP-specific phosphodiesterases, Adenyl cyclases, FhlA domain; HAE1, hydrophobe/amphiphile efflux 1; HAMP, Histidine kinases, Adenyl cyclases, Methylaccepting proteins, Phosphatases domains; HK, Histidine kinase domain; HTH, Helix-turn-helix; LBD, Ligand binding domain; LTTR, LysR-type transcription regulators; MATE, Multidrug and toxic compound extrusion; MDR, Multidrug resistance; MES, Multidrug efflux systems; MFS, Major facilitator superfamily; PAS, Per arnt sim domain; PDB, Protein data bank; RBS, Regulatory binding site; RD, Regulatory domain; REC, N-terminal receiver domain; RND, Resistance-nodulation-cell division; ROS, Reactive oxygen species; RR, Response regulator; SD, Sensor domain; SK, Sensor kinase receptor; SMR, Small multidrug resistance; TCS, Two-component system; TetR, Tetracycline repressor; TF, Transcription factor; TFBS, Transcription factor binding site; TG, Target gene; TM, Trans-membrane domain; TSA, paratoluene sulfonate.

the most efficient resistance mechanisms is the overexpression of the tripartite RND-MFP-OMF efflux pumps. Up to twelve genes coding for the efflux pumps were identified in PAO1 genome, each of them showing substrates specificity (Stover et al., 2000). Nevertheless, only five of the efflux pumps are involved in resistance in clinical strains, i.e., MexA^{MFP}-MexB^{RND}-OprM^{OMF}, MexX^{MFP}-MexY^{RND}-OprM^{OMF}, MexC^{MFP}-MexD^{RND}-OprJ^{OMF}, MexE^{MFP}-MexF^{RND}-OprN^{OMF}, and MexJ^{MFP}-MexK^{RND}-OprM^{OMF} (Lister et al., 2009; Li et al., 2015). These tripartite pumps are encoded in operon, but some of them do not bear their own OMF gene, such as MexXY, MexJK, MexVW, and MexMN. In that case, they usually assemble with OprM, the versatile OMF of *P. aeruginosa*, which structure has been extensively studied (Phan et al., 2010, 2015; Monlezun et al., 2015). MexAB-OprM is the only pump that is constitutively expressed and is able to transport most of the antibiotic families whereas the others are more selective and are induced under specific conditions. Dedicated activators or repressors regulate the efflux pumps expression, but complementary regulators, including the two-components system, could intervene to finely orchestrate the operons transcription. A blast analysis (<https://blast.ncbi.nlm.nih.gov/>) of the regulators based on *P. aeruginosa* amino acid sequences and known 3D structure homologs allows us to identify the main families of regulators in *P. aeruginosa* as illustrated in **Figure 1**. A majority of them belongs to the TetR family. We will first describe the regulators function of each family from archetypal structures. We will start with the two-component system (TCS) and then the one-component systems, i.e., TetR, LysR, MarR, and AraC. Finally, we will present MexR, NalD and MexZ, the only 3D structures solved from *P. aeruginosa* one-component regulators so far.

THE TWO-COMPONENT SYSTEM REGULATORS IN *P. AERUGINOSA*

In bacteria, efficient adaptation to environmental changes is very often orchestrated by the two-component systems (TCS) (Stock et al., 2000). As such, TCS is one of the most abundant bacterial molecular devices to cope the variety of environmental signals (Krell et al., 2010; Capra and Laub, 2012; Jung et al., 2012). In particular, according to the whole genome prediction, *P. aeruginosa* owns around 130 different TCS (Rodrigue et al., 2000) and uses more than 60 TCS to regulate virulence and antibiotic resistance (Gooderham and Hancock, 2009; Muller et al., 2011; Li et al., 2015). All the TCSs regulating *P. aeruginosa* efflux pumps belong to the prototypical system, which mechanism will be described here.

Regarding the specific genes regulation of the RND efflux pumps, five TCSs were identified so far (see **Figure 1** for those involved in resistance in clinical strains) (Li et al., 2015). The RocS2-RocA2 system was shown to downregulate the expression of the constitutive efflux pump MexAB-OprM in order to favor biofilm set up (Sivaneson et al., 2011). The TCSs ParR-ParS and AmgR-AmgS switch on the expression of the efflux pump MexXY following bacterial envelope stress and membrane perturbation by either colistin or polymyxin B (Fernández et al., 2010;



Muller et al., 2011; Lau et al., 2015). Besides, ParR-ParS also upregulates the efflux pump MexEF-OprN operon by enhancing the expression of the activator MexS (Wang D. et al., 2013). Finally, both systems CzcR-CzcS and CopR-CopS stimulate the expression of the heavy-metal efflux pump CzcABC (Perron et al., 2004; Caille et al., 2007). This pump is not involved in antibiotic resistance but it is mentioned here as the only TCS crystal structure solved so far in *P. aeruginosa*. Indeed, the sensor domain of the zinc-responsive histidine kinase CzcS shows the typical mixed α/β -fold of the PhoQ family (Wang D. et al., 2017).

Most of these TCS targeting the efflux pump genes of *P. aeruginosa* belong to the OmpR/PhoB family, except RocS2-RocA2, which is part of the CheY family. The architecture of the OmpR/PhoB and CheY families corresponds to the typical TCS which is a duet of phosphor-relay proteins (**Figure 2**): (1) a receptor Histidine-Kinase (HK), also named Sensor Kinase (SK), receives the extra-cytoplasmic (or periplasmic) signal and then activates (3) a cognate intracellular response regulator (RR) through a concerted trans-phosphorylation process (2). Subsequently, activated RR displays generally a DBD domain that targets a repeated sequence (4) upstream or within the promoter,

in order to up-regulate or to repress the expression of specific genes (Krell et al., 2010; Capra and Laub, 2012; Zschiedrich et al., 2016).

Despite the lack of the full-length structure of HK, more than 100 structures of the different domains and fragments from different bacterial species shed light on the transduction mechanism upon ligand binding (Bhate et al., 2015; Zschiedrich et al., 2016). HK are membrane proteins and function as homodimeric receptors. The canonical monomer is made of 4 domains: (i) a sensor domain (SD) that recognizes various signals (gas, ions, osmotic change, temperature, light or variable organic compounds including antibiotics (Krell et al., 2010), (ii) a transmembrane domain (TM), (iii) one or several signal transduction domains (i.e., the domains HAMP - *histidine kinases*, *adenyl cyclases*, *methylaccepting proteins*, *phosphatases*, PAS—*per arnt sim* or GAF—*cGMP-specific phosphodiesterases*, *adenylyl cyclases*, *FhlA*) and (iv) an autokinase domain made of two subdomains DHP (*dimerization and histidine phosphotransfer*) and CA (*catalytic and ATP binding*). Thus, HK are multi-domains receptors with variable and complex architectures, but the structure of the transmembrane and the cytosolic domains tend to be more conserved than the sensor domains across the receptor family (Krell et al., 2010; Bhate et al., 2015; Zschiedrich et al., 2016).

The different domains of HK show diverse folds, mainly from α/β to all- α classes. Behind the variety of stimuli and domain structures, signal transduction mechanism tends to be well conserved, driven by key α -helices connecting the different domains along the receptor. Signal transduction mechanism starts at the sensor domain by the p-helices (*periplasmic helices*) which are localized at the dimer interface of the sensor domain. Upon binding of the ligand, helices rearrangements, described as a piston-like shift, are transmitted to the TM domain across a bundle of two pairs of anti-parallel α -helices connected to the cytosolic domain, mainly HAMP, PAS, or GAF domains. As for the sensor domain, cytosolic domain folds are variable but the signal transduction is again driven by specific α -helices, in particular a tandem of input and output helices. Through dynamic scissoring and rotation of the α -helices bundle, a major structural event is the transition from a symmetrical apo-conformation (ligand-free) to an asymmetric holo-conformation (ligand-bound) and then active state. To summarize the complex signal transduction within the receptor structure, a global α -helices coiled-coil disturbance mediated by the key helices stretching and rotation leads to an asymmetric kinase-competent state (Wang C. et al., 2013; Mechaly et al., 2014; Molnar et al., 2014; Gushchin et al., 2017).

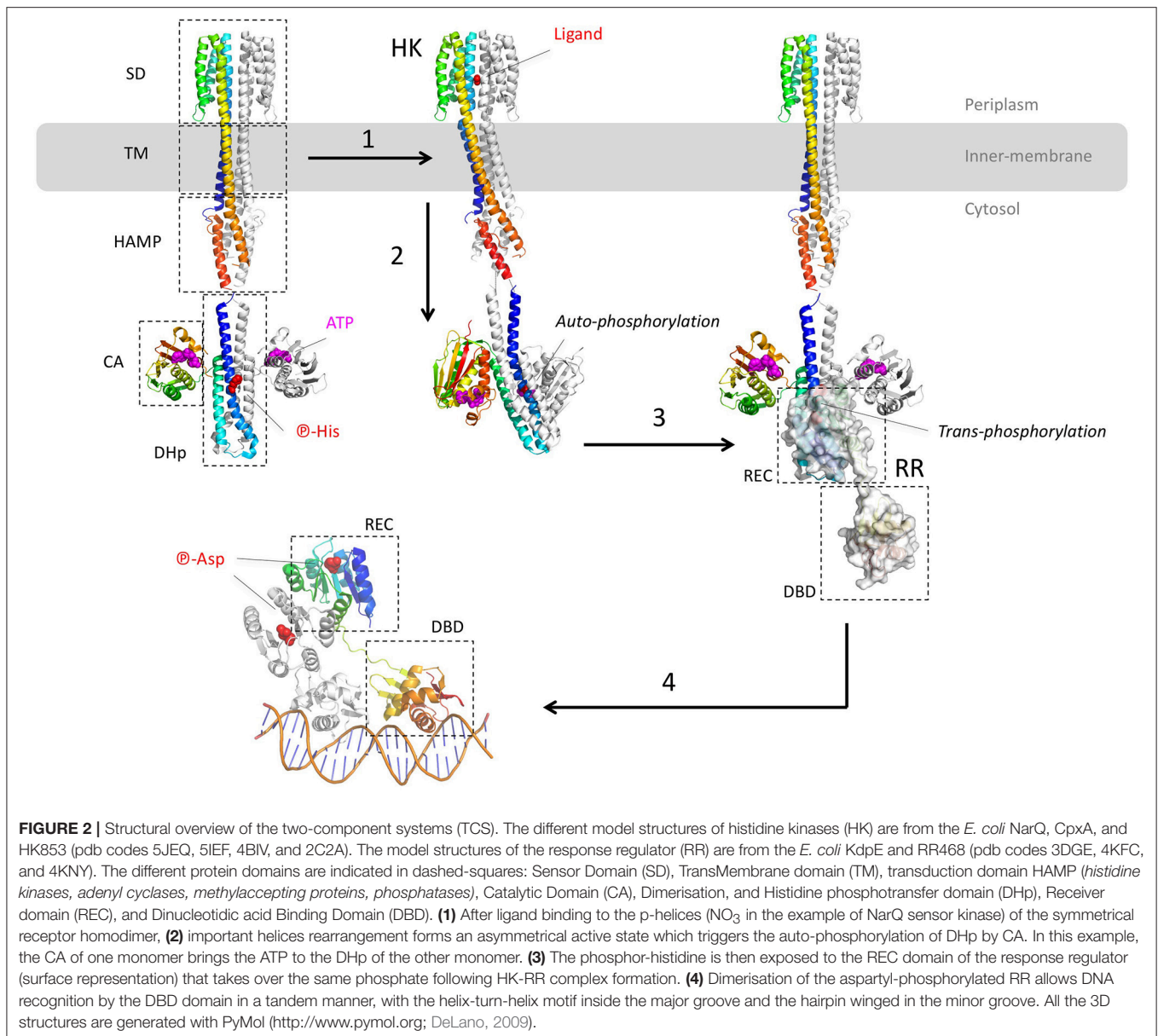
The catalytic event of HK takes place at the DHP and CA domains. It starts with the auto-phosphorylation of the conserved histidine of DHP domain which is a symmetrical dimer of helix-turn-helix. The CA domain binds to the upper region of the DHP and captures the phosphoryl group from one ATP to phosphorylate the histidine of DHP. Depending of the orientation of the helices, histidine phosphorylation could be within the same protomer (*cis*-phosphorylation) or between (*trans*-phosphorylation) the subunits of the homodimer receptor (Casino et al., 2009, 2010). The lower part of DHP receives the

response regulator RR. Again, a critical switch to an asymmetric conformation of the DHP correlates with the active kinase state. The formed phosphor-histidine is then available for the trans-phosphorylation to the RR.

Actually, the RR protein is a kinase itself and catalyzes its own phosphorylation on a conserved aspartate. The prototypical RR of the OmpR/PhoB superfamily presents two domains: a conserved N-terminal *receiver* (REC) domain linked to a more variable C-terminal domain, mainly a DBD. The typical REC domain consists of a typical α/β fold with five parallel β -strands surrounded by five α -helices. The REC domain docks onto the lower part of the DHP domain and catalyzes its own aspartyl-phosphorylation from the phospho-histidine donor. This phosphor-relay disturbs the molecular surface of REC and triggered the switch of two conserved residues T/S and F/Y localized between the strands β 4- β 5 of REC, nearby the phosphor-aspartate. This event induces the symmetric dimerization of the REC domain (Gao and Stock, 2009) and brings in close proximity the C-terminal DBDs each other. Thus, the DBD tandem is able to recognize and to interact with the DNA repeat sequence. Remarkably both REC domains of the RR dimer form a symmetrical head-to-head complex, whereas the associated DBD are poised asymmetrically on the cognate DNA, in a head-to-tail manner (Narayanan et al., 2014; Lou et al., 2015; He et al., 2016). Structural determination of the DNA-RR complexes revealed that the DBD-DNA interface is conserved, described as a winged-helix fold where the recognition α 3 penetrates the major groove whereas the α - β hairpin wings interacts with the minor groove, motif found in the LysR and MarR of the one-component system (see below) (Blanco et al., 2002; Narayanan et al., 2014; Lou et al., 2015; He et al., 2016). The surface contact of the DBD-DNA complex covers around 1,800 Å² with mainly van der Waals interactions toward the ribose groups and electrostatic attractions to the phosphates backbone.

THE ONE-COMPONENT SYSTEM REGULATORS

The one-component regulator system comprises both activators and repressors depending on the location of their binding site (TFBS) with the one of the RNA polymerase (RNAP). If the TFBS interferes with RNAP binding, the transcription factor will act as a repressor. When located upstream, the transcription factor helps RNAP recruitment as an activator or by competing with a repressor. One-component regulators can either act locally, interfering directly with the regulated operon, or remotely through general signaling events. These regulatory proteins are composed of two domains, one DNA-binding domain (DBD) comprising a Helix-Turn-Helix (HTH) motif, and one sensory domain involved in the oligomerization of the protein, often triggered by the binding of the sensor molecule. The one-component regulators are implicated in most of the essential signaling events in prokaryotic cells that is why a majority of bacterial regulators belong to this system. They are classified into more than 20 families (Cuthbertson and Nodwell, 2013) mainly based on sequence similarity of the DBD (Grkovic et al.,



2002), and most of them belong to six major families: TetR, MarR, LacI, LysR, AraC and MerR (Spengler et al., 2017). Four of them are involved in *P. aeruginosa* RND efflux pumps regulation (TetR, MarR, LysR, AraC) and they will be described here.

The TetR Family

The tremendous amount of sequences deposited in the UniProtKB databank (>2,300 verified TetR family assigned sequences in 2005; Ramos et al., 2005) supports the fact that most of bacterial genomes carries several TetR regulators to control vital and diverse functions. Interestingly, more than 15% of them regulate membrane-associated proteins, which are transporters in majority. As expected, the number of solved structures deposited in the protein data bank (PDB) is still low

with hardly more than 280 entries (December 2017). The first solved 3D structure of a member of this family corresponds to the tetracycline repressor (TetR) from *E. coli* [2TCT (Kisker et al., 1995); 2TRT (Hinrichs et al., 1994)] repressing the expression of TetA, a MFS efflux pump expelling the tetracycline antibiotic. Surprisingly, structural alignment of the known structures of this family did not provide conserved motif because of a low sequence identity (as low as 7%), despite the well conserved fold of the N-terminal DBD domain (≈ 50 amino acids). Nevertheless, this analysis highlights the fact that most of the studied regulators are frequently involved in antibiotic resistance or virulence of pathogenic bacteria. The larger group of solved structures corresponds to EthR from *Mycobacterium tuberculosis* with 57 entries (Carette et al., 2012; Blondiaux et al., 2017; Nikiforov et al., 2017). It negatively regulates the expression of EthA

monooxygenase implicated in the inactivation of the anti-tuberculosis drug ethionamide.

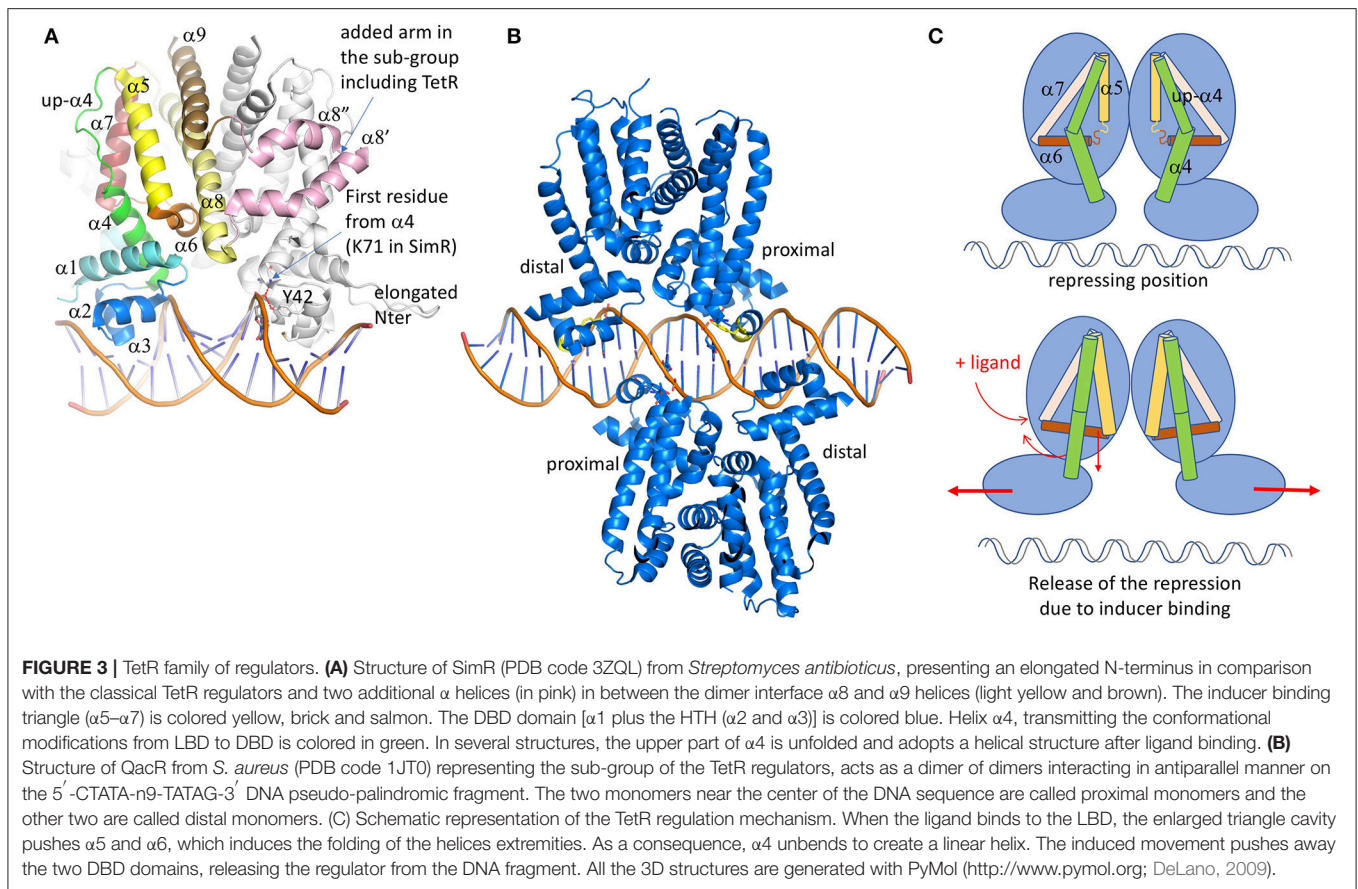
In spite of low sequence identity, the 3D structure of these regulators is conserved (**Figure 3A**). It is formed of 9 α -helices, with $\alpha 1$ - $\alpha 3$ corresponding to the DBD and $\alpha 4$ - $\alpha 9$ to the ligand-binding domain (LBD) (Deng et al., 2013). The contacts between the two domains involve $\alpha 1$ with $\alpha 4$ and $\alpha 6$, the latter being perpendicular to the dimer interface formed by $\alpha 8$ and $\alpha 9$ of each subunit (**Figure 3**). These inter-domain helices play a key role in the regulator activation through conformational modifications induced by the ligand binding. Among the 280 structures of TetR deposited in the PDB, more than 80 are declared to be associated with ligands. The chemical nature of these ligands is diverse, from a simple benzenediol such as resorcinol interacting with RolR, regulator of the aromatic catabolism from *Corynebacterium glutamicum* (3AQT; Li et al., 2011), to a more complex nucleotide derivative compound like isovaleryl coenzyme A (IV-CoA), an important building block in the formation of iso-fatty acids, interacting with AibR from *Myxococcus xanthus* (5K7H, 5K7Z; Bock et al., 2017). Most of the regulators classically repress the transcription in their apo-form, except for the AibR which is a repressor in a ligand-bound form of a whole operon bearing five genes involved in the biosynthesis process of IV-CoA, once bound to the same ligand. Besides, protein partners instead of chemicals also modulate some regulators. For instance, the SlmA regulator involved in cell-division of *E. coli* (Schumacher and Zeng, 2016) or AmtR, the nitrogen regulator of *Corynebacterium glutamicum* (Palanca and Rubio, 2016), both have to bind to DNA and another cognate protein at the same time to act. The diversity of the TetR regulator/ligand complexes is not limited to the ligand nature, but also depends on the localization of the binding regions which could be at the protein surface, close to the dimer or the domain interfaces, or in a deeply buried cavity that could even cross the entire protein like ActR (see Figure 8 from Cuthbertson and Nodwell, 2013 for a graphical representation). It differs also by the stoichiometry of the binding molecules, perfectly exemplified with TtgR (Alguel et al., 2007), a repressor of the key efflux pump TtgABC in *Pseudomonas putida*. Unlike TetR, which is only activated by tetracycline, TtgR can accommodate different molecules in the same ligand-binding site. Five structures have been solved of TtgR in complex with different antimicrobial molecules (2UXH, 2UXI, 2UXO, 2UXP, and 2UXU). In three different structures, the ligand occupies a large cavity formed by helices $\alpha 5$ - $\alpha 8$, within each monomer (quercetin in 2UXH, chloramphenicol in 2UXP and naringenin in 2UXU). In the complex with tetracycline (2UXO), a single monomer site is occupied despite the structural similarity with quercetin or naringenin. On the contrary, the structure of TtgR with phloretin shows both cavities occupied with an additional molecule close to $\alpha 6$ but in one monomer only. Using isothermal calorimetry to measure the binding of the different molecules, it has been shown that the affinity of phloretin for the different binding sites differs by two orders of magnitude, suggesting the existence of a positive cooperativity between the two sites.

Because of the important diversity of the ligands and the binding sites, it will not be possible to bring out a prototypical interaction mechanism. Nevertheless, the global structural

analysis of the LBD shows a conserved helical architecture built around helices $\alpha 5$ - $\alpha 7$ forming the so-called triangle that can be superimposed easily despite a root-mean-square deviation (rmsd) of more than 15 Å. This long atomic distance is due to differences in helices length and curvature. $\alpha 8$ is parallel to $\alpha 5$, stabilizing the triangle. Even if most of LBD domains are formed by 6 helices ($\alpha 4$ to $\alpha 9$), TetR belongs to a subclass itself, together with some minority proteins like the ActR regulator of actinorhodin efflux pump from *Streptomyces coelicolor* (Willems et al., 2008). We notice that the family name TetR was historically given because it was the first structure solved, but it is actually not representative since the folds are quite variable. For instance, in a sub-class of the TetR family, two additional helices are inserted between the classical $\alpha 8$ and $\alpha 9$, keeping the C-terminal helix at the conserved dimer interface. The two extra helices adopt a coiled-coil structure forming an arm that shell the second monomer, thus stabilizing the dimer. This is not the only difference in this sub-family. They also present a shorter helix $\alpha 4$ at the interface between the LBD and the DBD, the upper part of the traditionally curved $\alpha 4$ being unstructured but still parallel to $\alpha 5$.

In all the TetR family regulators, the dimer interface is formed by two antiparallel coiled-coil helices from each monomer ($\alpha 8$ and $\alpha 9$), forming a symmetrical four-helices bundle with the other monomer. The dimer is not always symmetrical, that is why ligands are not always present in both cavities of the complex structures. This is the case of QacR (Schumacher et al., 2004), which presents a much smaller cavity in one of the two monomers due to the flexibility of the last turn of $\alpha 5$, giving more freedom to $\alpha 6$ that can move upper then reducing the triangle cavity. The binding of the ligand in only one cavity is sufficient to release the transcription repression, but two dimers of QacR must interact with DNA (Schumacher et al., 2002), so at the end two ligand-bound cavities are necessary for the DNA recognition. In QacR the enlarged cavity can accept two different molecules (ethidium and proflavin), even if proflavin displaces ethidium from the binding site. Ligand binding pushes $\alpha 6$ in an allosteric way, which is transmitted to $\alpha 4$ that adopts a pendulum-like movement as described in Resch et al. (2008), driving away the two DNA binding sites (**Figure 3C**). It has also been described in HrtR, the heme homeostasis regulator in *Lactococcus lactis* (Sawai et al., 2012), an induced structural modification of this $\alpha 4$ from a partial random-coil to stable α -helical structure once associated to the cognate DNA. In the case of QacR, the distance between the two sites increases from 37 to 45 Å, causing the detachment of the regulator from the DNA fragment (Schumacher et al., 2001). It has been suggested by Reichheld et al. (2009) that the rigidification of the structure drives the DNA release. This hypothesis is based on far-UV circular dichroism experiments on TetR wild-type and mutants in the presence of increasing concentration of urea, with and without ligands. In the absence of the ligand, the DBD unfolds first followed by the LBD. In presence of the ligand, the two domains unfold at the same time, suggesting cooperativity. The instability of DBD domain without the ligand would be a clue for its DNA adaptation.

The DBD, generally localized at the N-terminus of the protein, is composed of $\alpha 1$, HTH domain, and the beginning of $\alpha 4$.



Among the 280 deposited TetR structures, only 19 are associated with the cognate DNA fragments, highlighting the difficulty to stabilize these protein/DNA complexes. These regulator-DNA complex structures were crucial to understand the functional mechanism of this regulator family. There are two sub-classes of DNA-binding mode: one represents DNA promoters interacting with a dimer (1QPI: TetR; 3ZQL: SimR; 3LSP and 3LSR: DesT; 3VOK: HrtR; 5UA2 and 5UA1: KstR; 5DY0: AmtR; 4I6Z: Tm1030; 5K7Z: Aibr). The second involves two dimers interaction (1JT0: QacR; 5HAW, 5HBU and 5k58: SImA; 4PXI and 5H58: CprB; 2YVH: CgmR; 4JL3: Ms6564; 5GPC: FadR) (**Figure 3B**). The first DNA complex was determined 5 years after the first structures of TetR by the same research group (1QPI; Orth et al., 2000). The main interacting region corresponds to the HTH domain formed by $\alpha 2$ and $\alpha 3$, which is also the most conserved sequence region used to create an identification profile of the TetR family (Ramos et al., 2005). The two helices deeply enter the DNA major groove but most of the contacts involve $\alpha 3$ only. In DesT, a regulator that controls the fatty acid saturation ratio in membrane lipid biogenesis, additional interactions were described with the DNA minor groove involving the elongated N-terminus of the protein (Miller et al., 2010). This is also the case for SimR, an exporter of a potent DNA gyrase inhibitor from *Streptomyces antibioticus*, which presents an even longer N-terminus that turns back to the added “arm” between $\alpha 8$ and $\alpha 9$

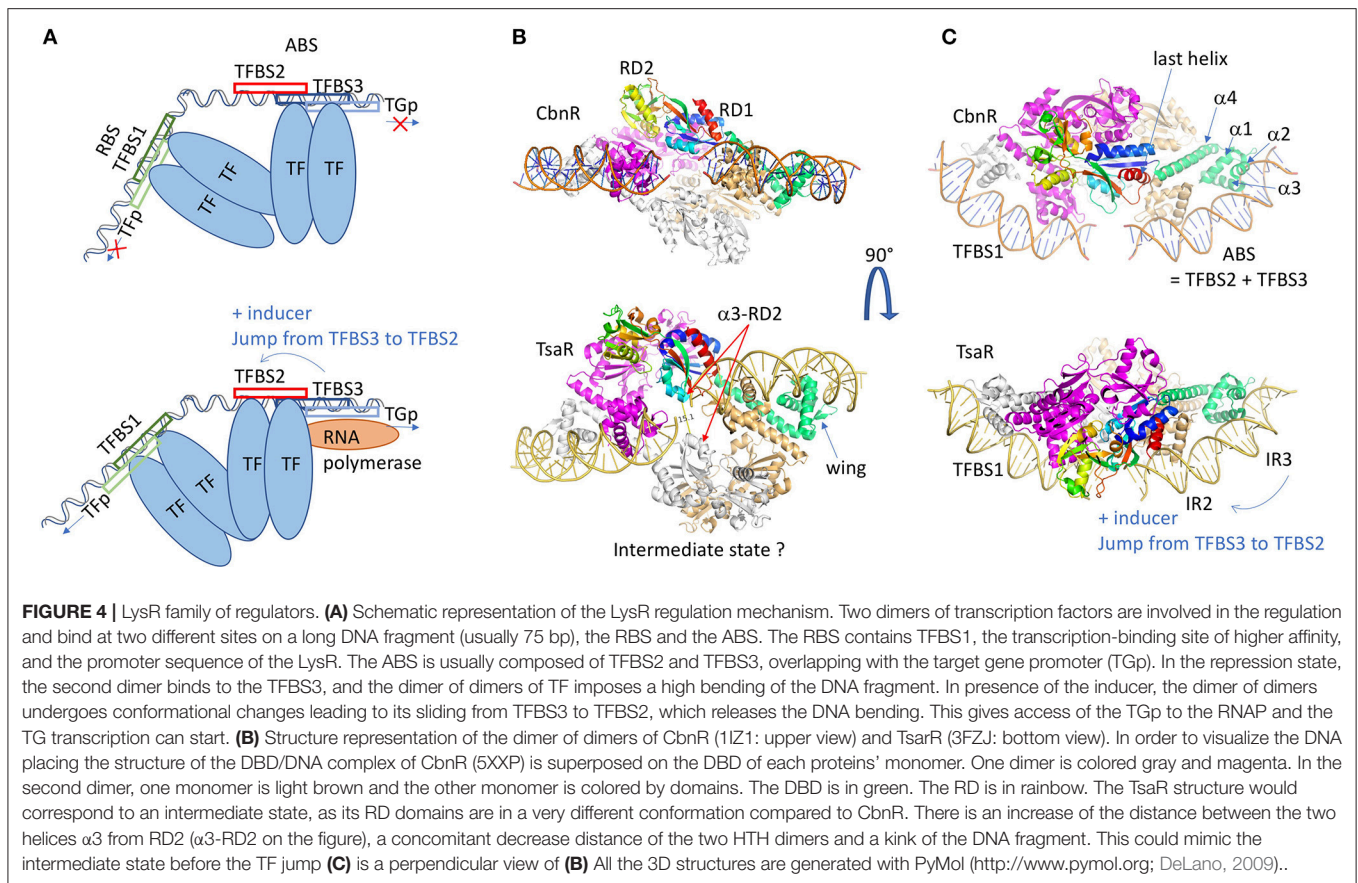
of the second monomer (Le et al., 2011). CprB from *S. coelicolor*, a receptor of c-butyrolactones, a class of quorum sensing molecules (Bhukya et al., 2017), also takes part of this N-terminal extended sub-family. AmtR, the global nitrogen regulator of *C. glutamicum* that is activated by a protein instead of a small molecule (Palanca and Rubio, 2016), also shows an additional C-terminal helix of unclear function. The recognized DNA operator is often formed by one central base pair (bp) surrounded by a palindromic sequence of a minimum of 6 bp in each opposite direction from the center (Yu et al., 2010). So, the complex is formed by a symmetrical protein dimer bound to a symmetric DNA fragment. Among the sub-class acting as a dimer of dimers, the regulators bind two overlapping DNA palindromic sequences instead of one in a cooperative manner. It is the case of QacR (Grkovic et al., 2001; Schumacher et al., 2002), which represses the expression of the MFS efflux pump QacA, transporting toxic organic compounds like the quaternary ammonium compounds. In this case, the palindromic DNA fragment is elongated by 28 bp with a longer central non-palindromic sequence (6 instead of 1) in order to accommodate the two dimers. For this class of regulators, the following nomenclature has been adopted: depending on the distance of the dimer from the central DNA sequence, there are the proximal and the distal subunits. Both dimers bind on opposite side of the promoter sequence, the two proximal monomers being very close to each other, sharing

several DNA base pairs in their binding site. With the opposite binding mode, the two dimers axis form an angle of $<180^\circ$ except Ms6564 (Yang et al., 2013), a broad regulator in *Mycobacterium smegmatis* that presents a smooth interaction involving water molecules which allow a sliding motion of the regulator along the genome to target several genes. For instance, the measured angles are 130° for both QacR and SlmA (the smallest ones), 142° for CprB and 145° for CgmR (Itou et al., 2010; Bhukya et al., 2014). In spite of this particularity, the interaction of two dimers with the DNA is very similar to that of one dimer. The main difference is a larger spacing between the two DNA binding sites within a dimer (3 bp for TetR-like regulators and 4 bp for QacR-like), reducing the induced bending of the DNA fragment ($\approx 3^\circ$ compared to $\approx 16^\circ$ for the one-dimer class) and increasing the $\alpha 3$ - $\alpha 3'$ distance from ≈ 34 to ≈ 37 Å. This distance is comparable to one turn of DNA-B form and was questionable for a long time. The numerous unbound and ligand-bound structures show very variable distances between these two $\alpha 3$ helices, ranging from 37.4 to 48.1 Å (Yu et al., 2010). This was confusing for scientists in their interpretation of the regulation mechanism, as discussed by Frénois et al. (2004) when they compared the structure of EthR solved with hexadecyl octanoate in LBD, and that of QacR solved in different forms (free, complexed with a ligand and complexed with DNA).

When superposing the DBD from the solved TetR structures in complex with DNA, it clearly appears that the HTH and the N-terminus of $\alpha 4$ of the DNA binding site match perfectly without any amino acid insertion. There are eight important residues for the binding: the two first correspond to the first residues of $\alpha 2$ and $\alpha 4$ that are not conserved since the interaction involve the protein backbone only with the phosphates of the DNA. The six other residues correspond to the whole $\alpha 3$ helix except the central residue oriented toward $\alpha 2$ (T41 in TetR). The nature of these amino acids varies with the cognate DNA sequence, even though most of the protein/DNA interactions involve the DNA backbone and not the nucleic base. This is the case for AibR presenting seven residues interacting with phosphates backbone and only two involving specific contacts with the base (Bock et al., 2017). Nevertheless, one residue is particularly well conserved, a tyrosine (Y42 in TetR and Y40 in QacR) that interacts with the same DNA phosphate as the amine group of the N-terminal residue of $\alpha 4$, reinforcing the DNA binding. This phosphate corresponds to the center of the DNA palindromic sequence. The amino acid the most deeply buried into the DNA major groove is positioned four residues before the conserved tyrosine (Y-4), in the turn preceding helix $\alpha 3$. This amino acid is often a short one (Ramos et al., 2005). In the case of FadR, a regulator involved in the fatty-acid degradation and synthesis pathways, mutation of each of these two conserved residues led to a significant reduction of DNA binding as proved by electrophoretic mobility shift assays (Yeo et al., 2017). It seems that the regulator could slide on the DNA until the $\alpha 3$ helix recognizes the cognate sequence, using a short residue at position Y-4 to enter more deeply into the groove. Then the clamp formed by the tyrosine and the first residue of $\alpha 4$ lock the interaction.

The LysR Family

The transcription factors (TF) belonging to the LysR family (LysR-Type Transcription Regulators: LTTR) are the most abundant in prokaryotes. This is due to the fact they regulate the expression of genes coding for proteins involved in very diverse functions like β -lactamase, transporter, amino acids biosynthesis, metabolic signaling, secretion, oxidative-stress response, cell division, quorum sensing, virulence, motility, detoxification, attachment (Schell, 1993; Maddocks and Oyston, 2008; Jiang et al., 2018). The family was named after the extensively studied transcription regulator of *lysA* implicated in lysine biosynthesis (Stragier et al., 1983) and is composed of both activators and repressors (Maddocks and Oyston, 2008) depending on the location of the transcription factor binding site (TFBS). The genetic organization of LTTRs targeted promoters and TFBSs has been studied by a computational protocol termed Phylogenetic Profile of Consensus Motifs issued by the analysis of Phylogenetic Footprinting technics (Oliver et al., 2016). In the LysR family, the gene coding for the TF is divergently oriented from its target gene (TG) and located <100 nucleotides upstream the beginning of the TG, and sometimes up to 500 bp from the initiation codon (Heroven and Dersch, 2006). Two to three different TFBSs are found in the intergenic region, the inter-motif length between the two first is generally seven nucleotides, except for the LyrR which is six. When the third motif exists, it very often overlaps with the second one, the global site being called the ABS (activation binding site) (Figure 4A). LysR also differs for this rule, because it presents 19 bp intermotif length between TFBSs 2 and 3. In each case, the transcription activation involves the binding, with different affinities, of two activated TF dimers in a cooperative manner, triggered by one or several inducers. An overlap of TF promoter and TFBSs causes an auto-repression by the TF when bound to the TFBS on the opposite strand of the DNA. The global site is called the RBS (regulatory binding site). Depending on the affinity of the TF for one of the different TFBSs, the TF will be an activator or a repressor. The mean length of the TFBSs is 15 bp and the consensus sequence, originally described as 5'-T-n11-A-3' (Goethals et al., 1992), has been extended to 5'-CTATA-n9-TATAG-3' (Oliver et al., 2016). Based on DNaseI protection assays combined with structural analysis (Wang and Winans, 1995; Muraoka et al., 2003; Picossi et al., 2007), a model has been proposed for the molecular mechanism of the LysR-type transcription regulators when three TFBSs are present, which is the majority of the LysR-type intergenic organization (Figure 4A). The affinity of the TF for the first and last TFBSs is greater than for the second one. Then in absence of the co-inducer, the formation of the dimer imposes a large bending of the DNA from 50 to 100° . With the inducer, the dimer interacts with the second TFBS which releases the third TFBS and unbends the DNA of 9° up to 50° . From this conformation, the TF will interact with the α -subunit of the RNAP so that the transcription of the TG can start. This mechanism model was reinforced by DNA-binding studies performed on modified DNA sequences, and is known as the "sliding dimer" (Porrúa et al., 2007). To get insights into the molecular details of this model, several 3D



structures were necessary and several examples will be described below.

The LysR-type genes code for proteins of around 330 amino acids. When searching for LysR transcription regulator in the PDB, it issues 87 entries corresponding to 27 different proteins, the majority corresponds to BenM from *Acinetobacter baylyi* or *sp.*, DntR from *Burkholderia sp.* or *cepacia*, OxyR from *Vibrio vulnificus*, CysB from *Salmonella typhimurium*, TsaR from *Comamonas testosteroni* and AphB from *Vibrio vulnificus*. Their structure shows a HTH motif in the DBD at the N-terminus like the TetR family, and a regulatory domain (RD) receiving an inducer (Henikoff et al., 1988). A long helical linker separated the two domains. The HTH is the most conserved region and is used to identify the members of this family in genome analysis (Schell, 1993). In contrary to the TetR HTH motif, there are two additional β -strands between α_3 and α_4 , a particular topology called winged-HTH. It also differs by the relative orientation of the three helices forming the DBD domain. Unlike the TetR family, the N-terminus of LTTR is localized at the dimer interface. As a consequence, all the helix axes are reverted as we can see by comparing **Figure 3A** with the DBDs on the ABS site of **Figure 4C**. The rest of the sequence is not very conserved except the C-terminal fragment of about 15 residues. Mutational analysis indicated implication in DNA binding or oligomerization (Schell et al., 1990; Bartowsky and Normark, 1991) which was partially confirmed with the

first crystal structure of the RD of CysB from *Salmonella typhimurium*, the regulator of the cysteine regulon expression (1AL3, Tyrrell et al., 1997). The co-inducer binding domain is composed of two α/β Rossmann fold-like subdomains RD1 and RD2 (**Figure 4B**) with a long β -sheet in between. The RD architecture forms a bend where the co-inducer interacts. Punctual mutations introduced in the co-inducer binding site of CysB led to an uncontrolled activation phenotype in spite of a proper interaction with the TFBS according to gel-shift assays (Colyer and Kredich, 1996). The nature of the replacing residue was important since the mutant T149M is comparable to the wild type, whereas T149P shows only a 10% activity, which suggests that the conformational flexibility of the protein is required for the co-activator effect.

Several structures of the isolated RD domain were solved with a monomer or a dimer in the asymmetric unit of the crystal, but none of them reveals the functional mechanism by a tetramer. The first structure of a full-length LTTR corresponds to the one of CbnR from *Cupriavidus necator*, involved in the degradation of chlorocatechol converted from 3-chlorobenzoate, using *cis,cis*-muconate as inducer (1IXC and 1IZL: Muraoka et al., 2003). The structure shows a tetramer that can be considered as a dimer of dimer, composed of two types of subunits with different conformations (**Figures 4B,C** upper panel), either compact or extended forms. Within one dimer, the main interacting region corresponds to the helical linker α_4 localized between the DBD

and the RD. The two $\alpha 4$ helices bind in an anti-parallel manner, imposing the head-to-head orientation of both DBD interacting by their N-termini. In this architecture, the distance between the two $\alpha 3$ helices is compatible with an interaction with the major groove of the B-form DNA. The angle between $\alpha 4$ and the RD axis is about 50° in the compact form and 130° in the extended form. Contacts between two RD domains appear through the interaction of the tetramer, i.e., two LTTRs dimers. It has to be noticed that the so-formed RD dimer interface is similar to those described in the structures of the isolated domain from several LTTRs, such as BenM from *Acinetobacter baylyi* regulating aromatic compound degradation (2F8D; Ezezika et al., 2007a). The two dimers are properly superimposed without showing any hinge movement between RD1 and RD2, with the exception of the C-terminal α/β domain of RD1 that is more divergent. Because the last helix is localized in the continuity of the linker $\alpha 4$ in the extended form of CbnR (see **Figure 4C** upper panel), this swapping domain could be involved in the conformational changes necessary for proper function of this family of regulators. The global quaternary structure of CbnR is compatible with an interaction with two DNA binding sites on a bended DNA fragment, supporting that the crystal structure is biologically relevant. Among the different full-length structures solved later, the one of TsaR (3FXU and 3FZJ; Monferrer et al., 2010) from the soil bacteria *Comamonas testosteroni* brought interesting insights in the regulation mechanism of the LTTRs. TsaR regulates the degradation of paratoluenesulfonate (TSA), a commonly found industrial pollutant, that also induces the regulator transcription itself. The tetrameric structure solved in complex with TSA is flatter than CbnR and presents less contacts between the different RD domains as they swing almost perpendicular to the tetramer plane, which yields a different interface (**Figures 4B,C** lower panel). In this conformation, the hinge between the DBD and the RD reaches 153° whereas 130° was measured for CbnR. On the contrary, for the compact form, the angle of 50° is conserved. The distance between the two pairs of $\alpha 3$ helices of the DBD domain varies largely because of the surface convexity between two pairs of DBDs. It has been suggested that the open form structure of TsaR could represent the active tetramer whereas the more compact form of CbnR tetramer, through contacts between the third helix from the RD2 domains ($\alpha 3$ -RD2) (see **Figure 4B** lower panel), would represent the inactive form. The transit to the active form could be induced by the binding of the TSA inducer in the cleft formed by two RD domains: in the structure of TsaR, the crossing β -sheet is broken in the middle when compared to CbnR. The hypothesis of a switch from a compact to an extended conformation of the tetramer once activated by the inducer in the RD cleft was confirmed by several LTTRs structures, i.e., ArgP regulating chromosome replication in *Mycobacterium tuberculosis* (Zhou et al., 2010), NdhR from *Synechocystis* involved in the control of carbon metabolism (Jiang et al., 2018), many of BenM (Ezezika et al., 2007b), and the SAXS experiments performed on DntR (Lerche et al., 2016). Nevertheless, the activation does not always depend on an inducer binding. For instance, a redox switch activates the oxidative stress regulator OxyR (Jo et al., 2015, 2017). Two cysteines (C199 and C208) from the helix $\alpha 3$ -RD2

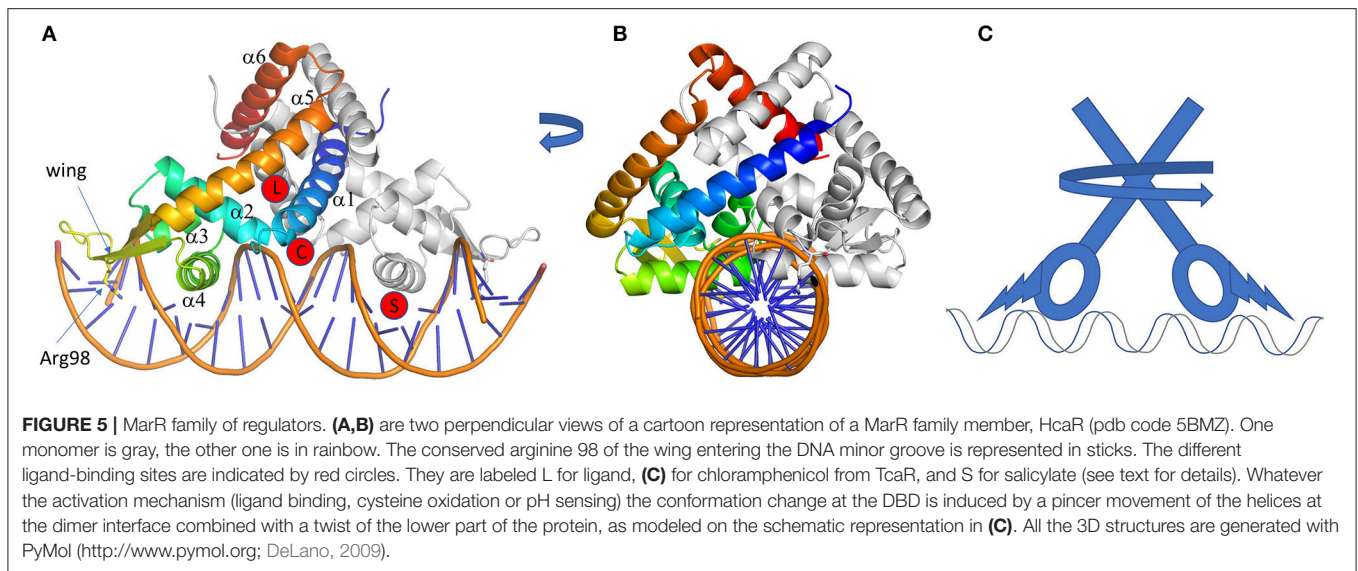
form a disulfide bond in the presence of H_2O_2 , which results in the unfolding of the helix and subsequent conformation modifications.

Only three structures of LTTR were solved in complex with DNA [two of BenM (4IHT, 4IHS) and one of CbnR (5XXP)] and all of them concern only the N-terminus DBD, including the HTH and the $\alpha 4$ in order to stabilize the dimer (Alanazi et al., 2013; Koentjoro et al., 2018). As already mentioned, the antiparallel $\alpha 4$ helices coiled-coil and the N-terminus are at the proximity of the pseudo-palindromic DNA center. Helix $\alpha 3$ enters deeply in the DNA major groove and brings most of the specific contacts with the DNA bases. The winged β -strand between the HTH and $\alpha 4$ makes contacts with the DNA minor groove, mainly through the phosphates, and one residue (R53) makes selective contact in BenM, which is absent in CbnR. Nevertheless, most of the selective residues are located in $\alpha 3$. Among these residues (A28, Q29, P30, P31, and R34 in CbnR), the remarkable mutation Q29A did not modify the interaction with DNA as proved by EMSA. Nevertheless, the mutant does not activate the transcription of the TG. This supports the importance of residues of the ABS site instead of the RBS. A plausible hypothesis is the involvement of Q29 in the RNA polymerase recruitment, since it is highly conserved among the LTTR family.

The MarR Family

The members of the Multiple Antibiotic Resistance Regulator (MarR) family are usually repressors found in bacteria and Archaea genomes (Wilkinson and Grove, 2006). They are mainly activated by sensors of environmental changes, like the presence of nutrients or toxins. Logically they often regulate genes coding for exporters of antibiotics, but they are also implicated in virulence, degradation processes, stress response and metabolic pathway (Alekhshun and Levy, 1997; Perera and Grove, 2010). MarR family are also involved in aromatic compounds metabolism which is one of the attractive field in industrial research of renewable energy (Davis and Sello, 2010; Fuchs et al., 2011; Bugg and Rahmanpour, 2015; Kallscheuer et al., 2016; Grove, 2017). Due to the essential role, MarR family is widely spread in bacterial genomes, up to 24 in *Bacillus subtilis* according to UniProt data bank.

The MarR transcription regulators are small proteins of <150 residues containing a winged HTH domain at the N-terminus similar to the LTTR family but acting in a dimeric form like TetR family. About 120 structure entries are in the PDB and the first one (1JGS) corresponds to MarR regulator from *E. coli* (Alekhshun et al., 2001). Compare to the other regulator families, the structure of MarR is quite simple, made of a DBD domain with an extension α -helix at each extremity (**Figure 5**). Those additional helices ($\alpha 1$ and $\alpha 6$) are the dimer interface. Besides, some MarR regulators present additional elements, for instance the regulator PcaV, involved in protocatechuate metabolism, an intermediary product of lignin degradation from *Streptomyces coelicolor*, possesses an additional β -strand between $\alpha 2$ and $\alpha 3$, forming a β -sheet wing (strands $\beta 1$ and $\beta 2$) (Davis et al., 2013). Another additional secondary structure is found in HucR from *Deinococcus radiodurans* (2fbk, Bordelon et al., 2006) involved



in oxidative stress response to uric acid. HucR possesses an additional helix at its N-terminus that stabilizes the dimer in the absence of DNA by pinching the helices of the dimer interface.

TF from the MarR family recognizes one or two types of intergenic region among the different regulated genes. The TFBSs are 16 to 20 bp long and are not always perfectly palindromic (Martin and Rosner, 1995; Perera and Grove, 2010). Like the LysR family, the HTH motif (here $\alpha 3$ and $\alpha 4$) enters in the DNA major groove and the supplementary β -strand wing interacts with the minor groove. This wing extension is essential for DNA binding, especially the arginine of the loop connecting the β -strands (Kumarevel, 2012), which deeply enters the DNA minor groove (**Figure 5A**). The importance of this basic amino acid for the regulation mechanism was analyzed for the ST1710 regulator from *Sulfolobus tokodaii* involved in antibiotic resistance. Several basic residues from this loop were mutated into alanine (R90A), showing a decrease of binding affinity for the cognate DNA sequence by gel-mobility shift assays (Kumarevel et al., 2009). When two TFBSs are necessary for gene regulation, the binding position of two TFs is either on opposite side or adjacent on DNA depending on the intergenic length and of the size of the β -strands wing, even though a dissociation of the wing was reported in the structure of Rv2887 regulator from *Mycobacterium tuberculosis* in complex with two TFBSs of 30 bp (Gao et al., 2017).

Concerning the LBD, it is reduced to a smaller fold so that the “triangle cavity” described previously in the TetR family does not really exist, even if $\alpha 5$ and $\alpha 6$ seem to adopt a triangle-like shape (**Figure 5**). This structure is sufficient to create a cavity surrounded by helix $\alpha 1$ and sometimes a long loop between $\alpha 1$ and $\alpha 2$ like in ZitR, a zinc metalloregulator (Zhu et al., 2017). This cavity is a receiving platform for several kinds of molecules, like coumaric acid, ferulic acid, vanillin and 3,4-dihydroxybenzoic acid in the case of HcaR from *Acinetobacter*, a regulator of the hydroxycinnamate degradation pathway (Kim et al., 2016). The binding of a molecule into the cavity will rearrange the dimer

interface conformation by modulating the distance between the two HTH domains and subsequently controlling the regulator release from the DNA. Nevertheless, structural modification of the regulator could be small for a large group of MarR regulators, like in SlyA, a virulence regulator from *Salmonella* (Dolan et al., 2011). The apo-form and ligand-bound structures are already in a conformation favorable to B-DNA interaction, with a distance of around 30 Å between the two DNA recognition helices. In this case, the presence of the inducer simply add stability to the regulator-DNA complex as demonstrated on MexR by thermal unfolding experiment and surface plasmon resonance (Andrésen et al., 2010). This is also supported by the structure of PcaV (4G9Y and 4FHT; Davis et al., 2013) showing the importance of an arginine (R15) in the functional mechanism (**Figure 5A**). By comparing the structure of the apo-form and the regulator complex with the natural ligand 3,4-dihydroxybenzoic acid, the R15 occupies the binding site in the absence of ligand. It is then pushed away by the ligand and forms hydrogen bonds with residues localized in $\alpha 2$, $\alpha 3$, $\alpha 3$ - $\alpha 4$ loop, $\alpha 1$ of the other monomer and the ligand itself. This pulls $\alpha 1$ by 10° toward the second monomer with an allosteric effect on the DBD orientation and a large movement of the β -sheet wing. To sum up, R15 and the ligand make a stable bridge between the DBD and the dimer interface by many hydrogen bonds. This specific mechanism was also described for NadR, the Neisserial Adhesin NadA repressor from *Neisseria meningitidis* studied by HDX-MS and molecular dynamic (Brier et al., 2012). Other binding sites were suggested for this family. In the case of the monomeric MarR (1JGS), two molecules of sodium salicylate bind on each side of the HTH- $\alpha 4$ of the DBD domain (**Figure 5A**). This compound is an inhibitor of MarR that will activate *marA* gene transcription (Cohen et al., 1993). It suggests the possibility of a regulation mechanism directly at the DBD site. Several other possible interacting sites were described, based on the structures of TcaR solved with different compounds (4EJT, 4EJU, 4EJV, 4EJW, Chang et al., 2010, 2013). For instance, kanamycin binds to similar sites as the ligand

of PcaV, but chloramphenicol binds to an unusual large cavity below the dimer interface (see C in **Figure 5A**), as well as a second site close to the $\alpha 4$. The latter is similar to that of salicylate in MarR or MTH313, a MarR homolog from *Methanobacterium thermoautotrophicum* (Saridakis et al., 2008).

Nevertheless, ligand binding is not the only activation mode of MarR family regulators. Some of them are sensitive to oxidative stress like AbfR from *Staphylococcus epidermidis* (Liu et al., 2017). The monomer possesses two cysteines, each one in the terminal helices ($\alpha 1$ and $\alpha 6$). The regulator binds to DNA in a reduced state. Under oxidative environment, sulfenic acid intermediates catalyze disulfide-bridge formation of the cysteines, which results in a large movement of the two monomers and destabilizes DNA interaction. The same regulation mechanism is also found in OhrR, the regulator of a peroxidase that reduces organic hydroperoxides to alcohols (Newberry et al., 2007). Upon oxidation by hydroperoxides, helix $\alpha 5$ of OhrR is locally unfolded which brings in close proximity the two cysteines to form the disulfide-bond responsible for a rigid-body rotation of the winged-HTH and then DNA release. Another regulation mechanism that depends on the pH was described for HucR regulator (Deochand et al., 2016). As shown by circular dichroism study at different pH, the N-terminal helices from each monomer interact by H51 stacking which protonation results in a molten globule intermediate with a low affinity for DNA. This explains the necessity for an additional helix at N-terminus to maintain the 3D structure of HucR during the conformation changes.

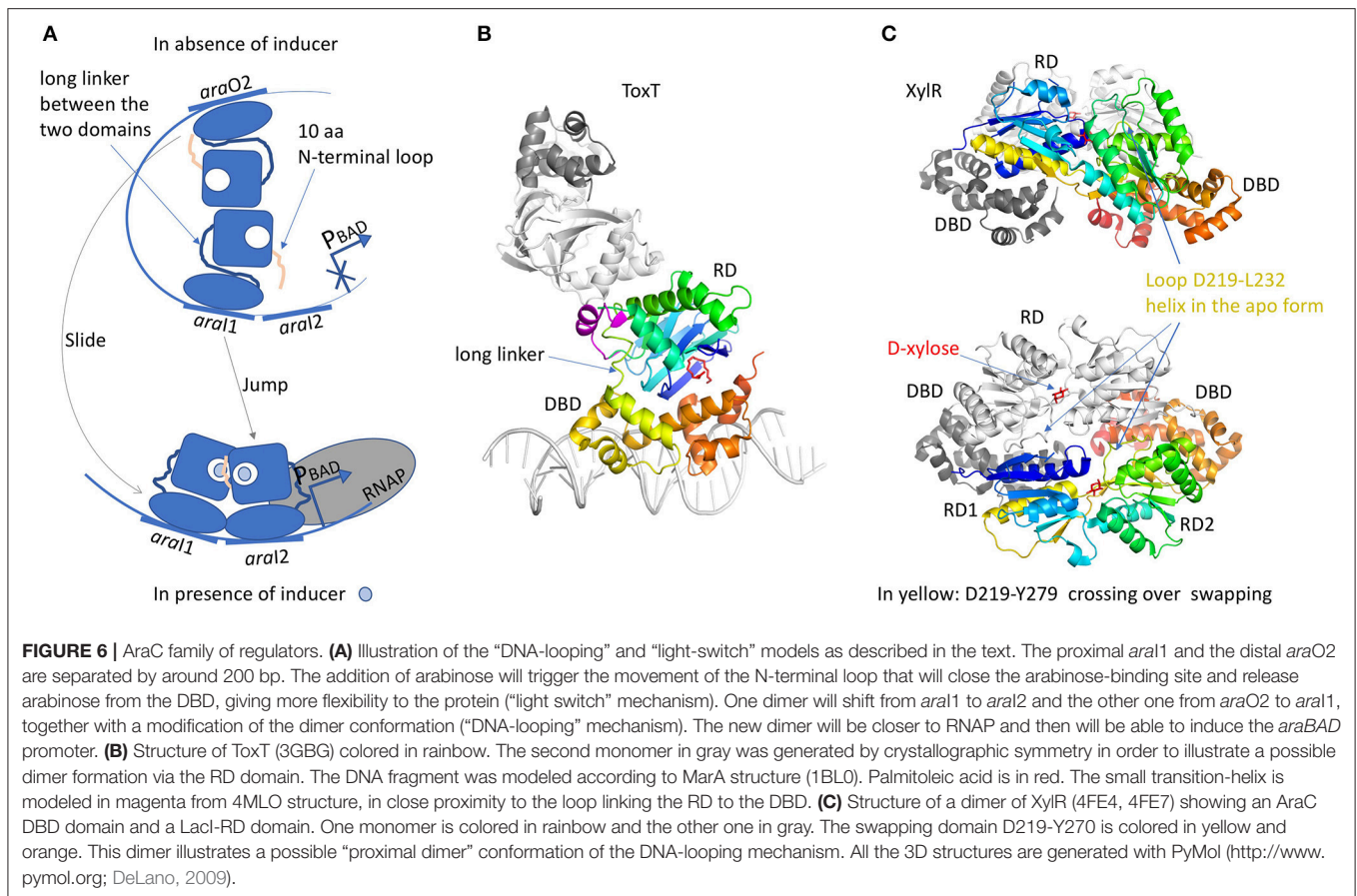
The AraC/XylS Family

The transcription regulators of AraC family (AFTRs) are generally activators found in all bacterial genomes, except in archaeobacteria (Gallegos et al., 1997). Like most of activators, the main TFBS is located around the -35 region of the promoter, which eases direct interaction with the RNAP (Ptashne and Gann, 1997). The members of the AFTR group are involved in pathogenesis, virulence and environment sensing, responding to oxidative stress, pH, temperature and antibiotics. They are also highly implicated in the regulation of essential metabolism pathways of the carbons, such as sugars, amino acids, alcohols or herbicides degradation (Gallegos et al., 1997; Egan, 2002; Ibarra et al., 2008).

The regulators of this family are generally 300 residues long, with some exceptions such as MarA or SoxS from *E. coli* which are reduced to the sole DBD domain (Rhee et al., 1998). When searching “AraC transcription” in the PDB, only 22 entries are listed which is low compared to the other families. An explanation was published in an essay of Schleif and his group who investigated the AraC regulation mechanism (Schleif, 2003). Because AraC interacts with more than 40 bp, it must be partially unfolded in the absence of DNA to reduce the binding energy for a reversible interaction. Thus, the intrinsically disordered state of this regulator family makes them difficult to handle (Schleif, 2010). Nevertheless, with the constant progress in protein expression and purification methods, several structures were obtained for more than ten different members of the family. Most of the ATFRs are made of two domains: a response domain (RD) involved in protein dimerization and a conserved DBD of

around 100 residues. The HTH motif interacts with the DNA in a similar way to the TetR family with one helix entering the DNA major groove. But unlike TetR, there is no wing interacting with the minor groove and one monomer carries two HTH separated by a long linking-helix. When both HTH interact with two contiguous DNA major grooves (Rhee et al., 1998), the DNA is bended by $\approx 35^\circ$ (Martin and Rosner, 2001). Besides, some proteins, such as the transcription factor Rob involved in antibiotic resistance and organic solvent tolerance in *E. coli*, makes direct contacts with only one DNA major groove with the first HTH, whereas the second interacts with the RNAP (Bhende and Egan, 2000). The tandem HTHs are generally at the C-terminus but in Rob regulator (PDB code 1D5Y), the HTH motifs are localized at the N-terminus. Even though the interacting configuration of the AraC family is quite specific, HTH motifs are poised from either side of the DNA fragment (Kwon et al., 2000) in a comparable manner to the QacR sub-group of TetR. Affinity measurements and DNA foot-printing experiments show that AraC recognizes DNA sequences as a tandem or inverted repeat orientation with different affinities (Carra and Schleif, 1993; Reeder and Schleif, 1993).

The RD domain is not conserved across the AraC family and it has very different functions. Both RD and DBD domains are independently associated by a long linker. A chimeric construct of the AraC-RD with the LexA-DBD from the LexA repressor (Bustos and Schleif, 1993) results in a protein that was able to dimerize and to repress *lexA* operator in response to L-arabinose. In spite of more than 20 years of work, the full-length structure of AraC is still unsolved. Nevertheless, crystal structures of the RD domain were solved with and without the arabinose ligand (Soisson et al., 1997; Weldon et al., 2007). The domain is a jellyroll that ended by a helix coiled-coil. RD dimer is found in the asymmetric unit only in the presence of arabinose. The dimer interface involves hydrophobic residues from the coiled-coil motif and an additional helix from the region between the jellyroll and the coiled-coil. The arabinose binds in a cavity formed by the jellyroll and locked by N-terminal loop of 10 residues. Without arabinose, this loop was not visible in the electron density. Genetic, biochemical and biophysical characterizations of AraC brought hypothesis on the regulation mechanism by the N-terminal loop (Carra and Schleif, 1993). In *E. coli*, L-arabinose is involved in the regulation of four operons, *araBAD*, *araE*, *araFGH* and *araC*. AraC represses the expression of *araBAD* and *araC* promoters by binding to two different DNA half sites separated by around 200 bp, the proximal site *araI1* and the distal site *araO2*, leading to the formation of a DNA loop (**Figure 6A**). In this configuration, the N-terminal loop makes contacts with the DBD, constraining the monomer in a compact structure. Arabinose binding results in the release of this loop by a “light switch” mechanism, allowing the RDs to form a totally different dimer. Then the DBD shifts from *araO2/araI1* to *araI1/araI2* sites, closer to the RNAP and the induction of the *araBAD* promoter. This mechanism is found in several regulators like ToxT from *Vibrio cholerae* (Lowden et al., 2010) or RegA from *Citrobacter rodentium* (Yang et al., 2009, 2011) respectively regulated by fatty acids or bicarbonate. The structure of the full-length protein ToxT from *Vibrio cholerae*



was solved in the absence of DNA (3GBG; Lowden et al., 2010; **Figure 6B**). The RD domain is very similar to that of the AraC but without the N-terminal loop. Instead, Li et al. (2016) revealed a small α -helix (D101 to E110) in proximity of the RD to DBD linker. Mutational analysis in ToxT of Met103, Arg105 or Asn106 showed a threefold activation increase of the *ctxA* promoter, pointing out the important role of this small α -helix (Childers et al., 2007). It could play the same role as the N-terminus loop of AraC by controlling protein flexibility. Another regulator of the AraC family was solved full-length: XylR (4FE4, 4FE7; Ni et al., 2013; **Figure 6C**). It is activated in *E. coli* in the absence of glucose, in order to use D-xylose as an alternative carbon source (Brückner and Titgemeyer, 2002). From structural comparison, the C-terminal DBD domain is very similar to that of AraC, but not the N-terminal RD domains. The XylR-RD domain looks like the periplasmic binding-protein of PurR from the LacI/GalR regulator family. It is composed of two α/β sub-domains linked by a small loop. The ligand-binding cavity is localized at the interface between the two α/β sub-domains. At the beginning of the swapping region between the RD and DBD domains, a flexible loop (D219-L232) in the complex structure with D-xylose changes into α -helix in the apo-form, which modulates the binding affinity of the DBD toward the cognate DNA operator. XylR forms

antiparallel dimers orienting the DBD domains in a head-to-head manner. Because XylR has to interact with two distant operator sites, a DNA loop must be formed by the DBD dimer, as demonstrated by AFM studies performed on XylR dimer in complex with a 500 bp DNA fragment. Note that unlike AraC, the DNA loop is formed in presence of the inducer ligand D-xylose. Nevertheless, there is a cross talk between the two regulatory mechanisms since AraC binds a DNA region containing the XylR promoter. It gives the possibility to activate one or the other sugar-producing pathway according to the available source of carbon.

All of the AFTRs does not respond to this simple mechanism. For example, InvF from *S. enterica serovar Typhimurium* is not able to activate virulence genes without the chaperone protein SicA (Darwin and Miller, 2000). It is suggested that InvF would function as a monomer but associated with SicA. MarA (Rhee et al., 1998; Gillette et al., 2000) and SoxS (Griffith and Wolf, 2002) involved in response to oxidative stress are small proteins of around 100 amino acids, formed by HTH domain only, thus devoid of responsive domain for a signaling ligand. They act as monomers and their regulation depends on the position of the TFBS toward the RNAP.

The tight regulation mechanism of AraC has inspired biotechnology developments and it is largely used as bacterial

expression systems for recombinant protein expression (Brautaset et al., 2009).

The One-Component Regulators of *P. aeruginosa* RND Efflux Pumps

As mentioned in the introduction, very few structural information is available on *P. aeruginosa* regulators, with only three solved structures to date: MexR, NalD and MexZ. Two of them are involved in the regulation of the MexAB-OprM pump which is considered as constitutive, although mutations in either *mexR*, *nalC* or *nalD* cause an over-expression of the pump (Boutoille et al., 2004; Sobel et al., 2005). MexR is the primary regulator of MexAB-OprM; it belongs to the MarR family and binds to its own promoter and that of the *mexAB-oprM* operon. One MexR regulatory pathway involves the binding of ArmR, a polypeptide of 53 amino acids, which expression is controlled by NalC, another TetR repressor. The structure of NalC has not been solved yet but it shows 31% of similarity with the N-terminal half of MLR_4833 from *Mesorhizobium loti* (3BHQ), corresponding to a canonical DBD of the TetR family. The LBD structure is not known but pentachlorophenol and other chlorinated phenol molecules have been identified as NalC signaling inducer (Muller et al., 2007; Ghosh et al., 2011). Several structures of MexR were solved: in the apo-form (1LNW, Lim et al., 2002), in complex with the C-terminal part of ArmR (3ECH, Wilke et al., 2008) and a clinical mutant (R21W) that induces overexpression of the pump (4ZZL, Anandapadamanaban et al., 2016). In addition, another apo-form structure was solved upon oxidation of the cysteines (3MEX, Chen et al., 2008), highlighting a new mechanism of regulation by inter-monomer disulfide bond (Chen et al., 2010). The comparison between the four different structures gives some information on the induced conformational changes necessary for the regulation. In the MexR/ArmR complex, ArmR binds into the classical MarR cavity with an oligomeric ratio of 1:2. The C-terminus enters deeply in the protein toward the cavity of the second monomer, labeled as “C” on **Figure 5**. In the mutant R21W structure, the dimer is more packed with a more constricted ligand cavity: the tryptophan mutant is stacked in between four prolines (P37 and P38 from each monomer) at the position normally occupied by G49 of ArmR (**Figure 5C**). This will cause the closure of the pincer formed by helices $\alpha 1$ and $\alpha 6$, and consequently will increase the distance between the two β -wings. The dimer does not fit anymore to a B-DNA conformation, which releases the repression and promotes *Pseudomonas* antibiotic resistance by MexAB-OprM efflux. The structure of the oxidized MexR is comparable to that of mutant R21W despite the 16 Å displacement of the $\alpha 3$ - $\alpha 4$ loop resulting in the disulfide bond formation between C62 and C30 from each monomer. The distance between the two helices $\alpha 4$ of the wild-type dimer (29 Å) is more suitable for tandem DNA major grooves interaction (34 Å) compared to the mutant (23.5 Å). These structures have brought important information to understand the regulation mechanisms of MexR.

Another regulator of known structure is NalD, a secondary regulator of MexAB-OprM transcription (Morita et al., 2006).

NalD is a TetR repressor that recognizes a TFBS upstream the operon *mexAB-OprM*. The NalD structure was solved in its apo-form (Chen et al., 2016) and is very similar to that of TtgR, the regulator of TtgABC efflux in *Pseudomonas putida*, which has a less folded $\alpha 4$ helix. It was proved that NalD is able to bind to novobiocin in a similar pocket to that of TtgR, resulting in resistance increase of *P. aeruginosa* strains.

The repressor NfxB of the MexCD-OprJ efflux pump was first classified in the LacI/GalR family, but the closest sequence homology turns to be the TetR-like regulator LFRR from *Mycobacterium smegmatis*, especially the DBD domain. Besides, MexL also belongs to the TetR family, with sequence identity alignment coverage of 93%. It shares 46.5% sequence identity with the DBD domain of NalD, which suggests a possible competition for the same TFBS. Another TetR repressor is the primary repressor MexZ of the MexXY-OprM efflux pump, which selectively transports aminoglycosides. The structure of MexZ was solved in its apo-form (Alguet et al., 2010). It presents a classical TetR structure, with a partially unfolded $\alpha 4$ helix. At present, the inducer molecule of MexZ is not known. A novel protein partner ArmZ, classified as a RNA-ligase, is suggested to sequester MexZ which releases the repression of *mexXY* (Hay et al., 2013). The gene of MexZ is the most frequently mutated in *P. aeruginosa* strains from cystic fibrosis patients (Smith et al., 2006). Most of the mutations are found in the DBD or strategic positions such as the dimer interface. One mutation has been reported on the surface of the helix $\alpha 7$ (L128M, see **Figure 3**; Guénard et al., 2014), which suggests $\alpha 7$ as part of the recognition site of ArmZ.

Finally, an activator regulates the efflux pump MexEF-OprN this time: MexT, which belongs to the LysR family (Fetar et al., 2011). MexT acts as a primary regulator of the pump together with the repressor of *oprD* porin. A redox mechanism seems to regulate MexT through *mexS* gene which codes for an oxydoreductase that upregulates MexEF-OprN (Morita et al., 2015; Richardot et al., 2016). We modeled the structure of MexT based on that of the DntR regulator from *Burkholderia* and localized in the RD domain the only cysteine that could possibly react to the ROS (Reactive Oxygen Species). Nevertheless, this cysteine can hardly form a disulfide-bond within a dimer of MexT according to the homology model structure. Recently a secondary activator of MexEF-OprN was described to upregulate MexEF-OprN through MexS and MexT (Juarez et al., 2018): it is named CmrA for Chloramphenicol Resistance Activator (Juarez et al., 2017). This regulator belongs to the AraC family and presents several cysteines, which is interesting in the context of the redox regulation mechanism.

CONCLUSION

As a multidrug-resistant pathogen, *Pseudomonas aeruginosa* possesses many RND efflux pumps. But some of them are functionally redundant, which is *a priori* not needful in term of biological evolution. Surprisingly, efflux pumps that transport similar molecules are not regulated by the same transcriptional systems. This certainly reflects the need to a prompt reactivity of the bacteria upon environment modification. All the 3D

structures of the different regulators solved from different bacteria brought complementary informations to genetic, biochemical and biophysical data, in particular the crystal structures gave important insights in the comprehension of the main regulatory mechanisms. Nevertheless, with the intention of doing specific drug-design, high-resolution structures of regulators from *P. aeruginosa* are still necessary. Recent structures of the virulence factor regulator MvfR, member of the LysR family from *P. aeruginosa*, solved in complex with inducer and inhibitor, illustrate the interest of the structural approach (Kitao et al., 2018). Both molecules bind within the same cavity with subtle interaction differences that are the keystone of the regulation mechanism. Thus, the knowledge of the 3D structures of each specific regulator is mandatory to develop new and specific drugs. From the complex signaling regulation of RND pumps expression illustrated on **Figure 1**, it is clear that

there is a real lack of information concerning the structures of *P. aeruginosa* regulators. As this bacterium belongs to the group of the most problematic clinical pathogens, structural study of the regulators of *P. aeruginosa* are urgently needed.

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

FUNDING

Association Vaincre la Mucoviscidose (RF20150501358) and Association Grégory Lemarchal. CNRS. University Paris Descartes. KH was supported by a grant of Vaincre la Mucoviscidose and the Association Grégory Lemarchal.

REFERENCES

- Alanazi, A. M., Neidle, E. L., and Momany, C. (2013). The DNA-binding domain of BenM reveals the structural basis for the recognition of a T-N11-A sequence motif by LysR-type transcriptional regulators. *Acta Crystallogr. D Biol. Crystallogr.* 69(Pt 10), 1995–2007. doi: 10.1107/S0907444913017320
- Alekshun, M. N., and Levy, S. B. (1997). Regulation of chromosomally mediated multiple antibiotic resistance: the mar regulon. *Antimicrob. Agents Chemother.* 41, 2067–2075.
- Alekshun, M. N., Levy, S. B., Mealy, T. R., Seaton, B. A., and Head, J. F. (2001). The crystal structure of MarR, a regulator of multiple antibiotic resistance, at 2.3 Å resolution. *Nat. Struct. Biol.* 8, 710–714. doi: 10.1038/90429
- Alguel, Y., Lu, D., Quade, N., Sauter, S., and Zhang, X. (2010). Crystal structure of MexZ, a key repressor responsible for antibiotic resistance in *Pseudomonas aeruginosa*. *J. Struct. Biol.* 172, 305–310. doi: 10.1016/j.jsb.2010.07.012
- Alguel, Y., Meng, C., Terán, W., Krell, T., Ramos, J. L., Gallegos, M. T., et al. (2007). Crystal structures of multidrug binding protein TtgR in complex with antibiotics and plant antimicrobials. *J. Mol. Biol.* 369, 829–840. doi: 10.1016/j.jmb.2007.03.062
- Alvarez-Ortega, C., Olivares, J., and Martínez, J. L. (2013). RND multidrug efflux pumps: what are they good for? *Front. Microbiol.* 4:7. doi: 10.3389/fmicb.2013.00007
- Anandapadamanaban, M., Pilstål, R., Andresen, C., Trehwella, J., Moche, M., Wallner, B., et al. (2016). Mutation-induced population shift in the MexR conformational ensemble disengages DNA binding: a novel mechanism for MarR family derepression. *Structure* 24, 1311–1321. doi: 10.1016/j.str.2016.06.008
- Andrésen, C., Jalal, S., Aili, D., Wang, Y., Islam, S., Jarl, A., et al. (2010). Critical biophysical properties in the *Pseudomonas aeruginosa* efflux gene regulator MexR are targeted by mutations conferring multidrug resistance. *Protein Sci.* 19, 680–692. doi: 10.1002/pro.343
- Bartowsky, E., and Normark, S. (1991). Purification and mutant analysis of *Citrobacter freundii* AmpR, the regulator for chromosomal AmpC beta-lactamase. *Mol. Microbiol.* 5, 1715–1725. doi: 10.1111/j.1365-2958.1991.tb01920.x
- Bhate, M. P., Molnar, K. S., Goulian, M., and DeGrado, W. F. (2015). Signal transduction in histidine kinases: insights from new structures. *Structure* 23, 981–994. doi: 10.1016/j.str.2015.04.002
- Bhende, P. M., and Egan, S. M. (2000). Genetic evidence that transcription activation by RhaS involves specific amino acid contacts with sigma 70. *J. Bacteriol.* 182, 4959–4969. doi: 10.1128/JB.182.17.4959-4969.2000
- Bhukya, H., Bhujbalrao, R., Bitra, A., and Anand, R. (2014). Structural and functional basis of transcriptional regulation by TetR family protein CprB from *S. coelicolor* A3(2). *Nucleic Acids Res.* 42, 10122–10133. doi: 10.1093/nar/gku587
- Bhukya, H., Jana, A. K., Sengupta, N., and Anand, R. (2017). Structural and dynamics studies of the TetR family protein, CprB from *Streptomyces coelicolor* in complex with its biological operator sequence. *J. Struct. Biol.* 198, 134–146. doi: 10.1016/j.jsb.2017.03.006
- Blanco, A. G., Sola, M., Gomis-Rüth, F. X., and Coll, M. (2002). Tandem DNA recognition by PhoB, a two-component signal transduction transcriptional activator. *Structure* 10, 701–713. doi: 10.1016/S0969-2126(02)00761-X
- Blanco, P., Hernando-Amado, S., Reales-Calderon, J. A., Corona, F., Lira, F., Alcalde-Rico, M., et al. (2016). Bacterial multidrug efflux pumps: much more than antibiotic resistance determinants. *Microorganisms* 4:E14. doi: 10.3390/microorganisms4010014
- Blondiaux, N., Moune, M., Desroses, M., Frita, R., Flipo, M., Mathys, V., et al. (2017). Reversion of antibiotic resistance in *Mycobacterium tuberculosis* by spiroisoxazoline SMART-420. *Science* 355, 1206–1211. doi: 10.1126/science.aag1006
- Bock, T., Volz, C., Hering, V., Scrima, A., Müller, R., and Blankenfeldt, W. (2017). The AibR-isovaleryl coenzyme A regulator and its DNA binding site - a model for the regulation of alternative de novo isovaleryl coenzyme A biosynthesis in *Myxococcus xanthus*. *Nucleic Acids Res.* 45, 2166–2178. doi: 10.1093/nar/gkw1238
- Bordelon, T., Wilkinson, S. P., Grove, A., and Newcomer, M. E. (2006). The crystal structure of the transcriptional regulator HucR from *Deinococcus radiodurans* reveals a repressor preconfigured for DNA binding. *J. Mol. Biol.* 360, 168–177. doi: 10.1016/j.jmb.2006.05.005
- Boutoille, D., Corvec, S., Caroff, N., Giraudeau, C., Espaze, E., Caillon, J., et al. (2004). Detection of an IS21 insertion sequence in the mexR gene of *Pseudomonas aeruginosa* increasing beta-lactam resistance. *FEMS Microbiol. Lett.* 230, 143–146. doi: 10.1016/S0378-1097(03)00882-6
- Brautaset, T., Lale, R., and Valla, S. (2009). Positively regulated bacterial expression systems. *Microb. Biotechnol.* 2, 15–30. doi: 10.1111/j.1751-7915.2008.00048.x
- Brier, S., Fagnocchi, L., Donnarumma, D., Scarselli, M., Rappuoli, R., Nissum, M., et al. (2012). Structural insight into the mechanism of DNA-binding attenuation of the Neisserial adhesin repressor NadR by the small natural ligand 4-hydroxyphenylacetic acid. *Biochemistry* 51, 6738–6752. doi: 10.1021/bi300656w
- Brückner, R., and Titgemeyer, F. (2002). Carbon catabolite repression in bacteria: choice of the carbon source and autoregulatory limitation of sugar utilization. *FEMS Microbiol. Lett.* 209, 141–148. doi: 10.1016/S0378-1097(02)00559-1
- Bugg, T. D., and Rahmanpour, R. (2015). Enzymatic conversion of lignin into renewable chemicals. *Curr. Opin. Chem. Biol.* 29, 10–17. doi: 10.1016/j.cbpa.2015.06.009
- Bustos, S. A., and Schleif, R. F. (1993). Functional domains of the AraC protein. *Proc. Natl. Acad. Sci. U.S.A.* 90, 5638–5642. doi: 10.1073/pnas.90.12.5638
- Caille, O., Rossier, C., and Perron, K. (2007). A copper-activated two-component system interacts with zinc and imipenem resistance in *Pseudomonas aeruginosa*. *J. Bacteriol.* 189, 4561–4568. doi: 10.1128/JB.00095-07

- Capra, E. J., and Laub, M. T. (2012). Evolution of two-component signal transduction systems. *Annu. Rev. Microbiol.* 66, 325–347. doi: 10.1146/annurev-micro-092611-150039
- Carette, X., Blondiaux, N., Willery, E., Hoos, S., Lecat-Guillet, N., Lens, Z., et al. (2012). Structural activation of the transcriptional repressor EthR from *Mycobacterium tuberculosis* by single amino acid change mimicking natural and synthetic ligands. *Nucleic Acids Res.* 40, 3018–3030. doi: 10.1093/nar/gkr1113
- Carra, J. H., and Schleif, R. F. (1993). Variation of half-site organization and DNA looping by AraC protein. *EMBO J.* 12, 35–44.
- Casino, P., Rubio, V., and Marina, A. (2009). Structural insight into partner specificity and phosphoryl transfer in two-component signal transduction. *Cell* 139, 325–336. doi: 10.1016/j.cell.2009.08.032
- Casino, P., Rubio, V., and Marina, A. (2010). The mechanism of signal transduction by two-component systems. *Curr. Opin. Struct. Biol.* 20, 763–771. doi: 10.1016/j.sbi.2010.09.010
- Chang, Y. M., Chen, C. K., Ko, T. P., Chang-Chien, M. W., and Wang, A. H. (2013). Structural analysis of the antibiotic-recognition mechanism of MarR proteins. *Acta Crystallogr. D Biol. Crystallogr.* 69(Pt 6), 1138–1149. doi: 10.1107/S0907444913007117
- Chang, Y. M., Jeng, W. Y., Ko, T. P., Yeh, Y. J., Chen, C. K., and Wang, A. H. (2010). Structural study of TcaR and its complexes with multiple antibiotics from *Staphylococcus epidermidis*. *Proc. Natl. Acad. Sci. U.S.A.* 107, 8617–8622. doi: 10.1073/pnas.0913302107
- Cheesman, M. J., Ilanko, A., Blonk, B., and Cock, I. E. (2017). Developing new antimicrobial therapies: are synergistic combinations of plant extracts/compounds with conventional antibiotics the solution? *Pharmacogn. Rev.* 11, 57–72. doi: 10.4103/phrev.phrev_21_17
- Chen, H., Hu, J., Chen, P. R., Lan, L., Li, Z., Hicks, L. M., et al. (2008). The *Pseudomonas aeruginosa* multidrug efflux regulator MexR uses an oxidation-sensing mechanism. *Proc. Natl. Acad. Sci. U.S.A.* 105, 13586–13591. doi: 10.1073/pnas.0803391105
- Chen, H., Yi, C., Zhang, J., Zhang, W., Ge, Z., Yang, C. G., et al. (2010). Structural insight into the oxidation-sensing mechanism of the antibiotic resistance of regulator MexR. *EMBO Rep.* 11, 685–690. doi: 10.1038/embor.2010.96
- Chen, W., Wang, D., Zhou, W., Sang, H., Liu, X., Ge, Z., et al. (2016). Novobiocin binding to NalD induces the expression of the MexAB-OprM pump in *Pseudomonas aeruginosa*. *Mol. Microbiol.* 100, 749–758. doi: 10.1111/mmi.13346
- Childers, B. M., Weber, G. G., Prouty, M. G., Castaneda, M. M., Peng, F., and Klose, K. E. (2007). Identification of residues critical for the function of the *Vibrio cholerae* virulence regulator ToxT by scanning alanine mutagenesis. *J. Mol. Biol.* 367, 1413–1430. doi: 10.1016/j.jmb.2007.01.061
- Coates, A., Hu, Y., Bax, R., and Page, C. (2002). The future challenges facing the development of new antimicrobial drugs. *Nat. Rev. Drug Discov.* 1, 895–910. doi: 10.1038/nrd940
- Cohen, S. P., Hächler, H., and Levy, S. B. (1993). Genetic and functional analysis of the multiple antibiotic resistance (mar) locus in *Escherichia coli*. *J. Bacteriol.* 175, 1484–1492. doi: 10.1128/jb.175.5.1484-1492.1993
- Colyer, T. E., and Kredich, N. M. (1996). *In vitro* characterization of constitutive CysB proteins from *Salmonella typhimurium*. *Mol. Microbiol.* 21, 247–256. doi: 10.1046/j.1365-2958.1996.6301347.x
- Cuthbertson, L., and Nodwell, J. R. (2013). The TetR family of regulators. *Microbiol. Mol. Biol. Rev.* 77, 440–475. doi: 10.1128/MMBR.00018-13
- Darwin, K. H., and Miller, V. L. (2000). The putative invasion protein chaperone SicA acts together with InvF to activate the expression of *Salmonella typhimurium* virulence genes. *Mol. Microbiol.* 35, 949–960. doi: 10.1046/j.1365-2958.2000.01772.x
- Daurly, L., Orange, F., Taveau, J. C., Verchère, A., Monlezun, L., Gounou, C., et al. (2016). Tripartite assembly of RND multidrug efflux pumps. *Nat. Commun.* 7:10731. doi: 10.1038/ncomms10731
- Davies, J., and Davies, D. (2010). Origins and evolution of antibiotic resistance. *Microbiol. Mol. Biol. Rev.* 74, 417–433. doi: 10.1128/MMBR.00016-10
- Davis, J. R., Brown, B. L., Page, R., and Sello, J. K. (2013). Study of PcaV from *Streptomyces coelicolor* yields new insights into ligand-responsive MarR family transcription factors. *Nucleic Acids Res.* 41, 3888–3900. doi: 10.1093/nar/gkt009
- Davis, J. R., and Sello, J. K. (2010). Regulation of genes in *Streptomyces* bacteria required for catabolism of lignin-derived aromatic compounds. *Appl. Microbiol. Biotechnol.* 86, 921–929. doi: 10.1007/s00253-009-2358-0
- D’Costa, V. M., King, C. E., Kalan, L., Morar, M., Sung, W. W., Schwarz, C., et al. (2011). Antibiotic resistance is ancient. *Nature* 477, 457–461. doi: 10.1038/nature10388
- DeLano, W. L. (2009). PyMOL molecular viewer: updates and refinements. *Abst. Papers Am. Chem. Soc.* 238.
- Deng, W., Li, C., and Xie, J. (2013). The underlying mechanism of bacterial TetR/AcrR family transcriptional repressors. *Cell. Signal.* 25, 1608–1613. doi: 10.1016/j.cellsig.2013.04.003
- Deochand, D. K., Perera, I. C., Crochet, R. B., Gilbert, N. C., Newcomer, M. E., and A., G. (2016). Histidine switch controlling pH-dependent protein folding and DNA binding in a transcription factor at the core of synthetic network devices. *Mol. Biosyst.* 12, 2417–2426. doi: 10.1039/C6MB00304D
- Dolan, K. T., Duguid, E. M., and He, C. (2011). Crystal structures of SlyA protein, a master virulence regulator of *Salmonella*, in free and DNA-bound states. *J. Biol. Chem.* 286, 22178–22185. doi: 10.1074/jbc.M111.242528
- Du, D., Wang, Z., James, N. R., Voss, J. E., Klimont, E., Ohene-Agyei, T., et al. (2014). Structure of the AcrAB-TolC multidrug efflux pump. *Nature* 509, 512–515. doi: 10.1038/nature13205
- Egan, S. M. (2002). Growing repertoire of AraC/XylS activators. *J. Bacteriol.* 184, 5529–5532. doi: 10.1128/JB.184.20.5529-5532.2002
- Ezeizika, O. C., Haddad, S., Neidle, E. L., and Momany, C. (2007a). Oligomerization of BenM, a LysR-type transcriptional regulator: structural basis for the aggregation of proteins in this family. *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.* 63(Pt 5), 361–368. doi: 10.1107/S1744309107019185
- Ezeizika, O. C., Haddad, S., Clark, T. J., Neidle, E. L., and Momany, C. (2007b). Distinct effector-binding sites enable synergistic transcriptional activation by BenM, a LysR-type regulator. *J. Mol. Biol.* 367, 616–629. doi: 10.1016/j.jmb.2006.09.090
- Fair, R. J., and Tor, Y. (2014). Antibiotics and bacterial resistance in the 21st century. *Perspect. Medicin. Chem.* 6, 25–64. doi: 10.4137/PMC.S14459
- Fernández, L., Gooderham, W. J., Bains, M., McPhee, J. B., Wiegand, I., and Hancock, R. E. (2010). Adaptive resistance to the “last hope” antibiotics polymyxin b and colistin in *Pseudomonas aeruginosa* is mediated by the novel two-component regulatory system ParR-ParS. *Antimicrob. Agents Chemother.* 54, 3372–3382. doi: 10.1128/AAC.00242-10
- Fetar, H., Gilmour, C., Klinoski, R., Daigle, D. M., Dean, C. R., and Poole, K. (2011). mexEF-oprN multidrug efflux operon of *Pseudomonas aeruginosa*: regulation by the MexT activator in response to nitrosative stress and chloramphenicol. *Antimicrob. Agents Chemother.* 55, 508–514. doi: 10.1128/AAC.00830-10
- Fitzpatrick, A. W. P., Llabrés, S., Neuberger, A., Blaza, J. N., Bai, X. C., Okada, U., et al. (2017). Structure of the MacAB-TolC ABC-type tripartite multidrug efflux pump. *Nat. Microbiol.* 2:17070. doi: 10.1038/nmicrobiol.2017.70
- Frénois, F., Engohang-Ndong, J., Loch, C., Baulard, A. R., and Villeret, V. (2004). Structure of EthR in a ligand bound conformation reveals therapeutic perspectives against tuberculosis. *Mol. Cell* 16, 301–307. doi: 10.1016/j.molcel.2004.09.020
- Frieri, M., Kumar, K., and Boutin, A. (2017). Antibiotic resistance. *J. Infect. Public Health* 10, 369–378. doi: 10.1016/j.jiph.2016.08.007
- Fuchs, G., Boll, M., and Heider, J. (2011). Microbial degradation of aromatic compounds - from one strategy to four. *Nat. Rev. Microbiol.* 9, 803–816. doi: 10.1038/nrmicro2652
- Gallegos, M. T., Schleif, R., Bairoch, A., Hofmann, K., and Ramos, J. L. (1997). Arac/XylS family of transcriptional regulators. *Microbiol. Mol. Biol. Rev.* 61, 393–410.
- Gao, R., and Stock, A. M. (2009). Biological insights from structures of two-component proteins. *Annu. Rev. Microbiol.* 63, 133–154. doi: 10.1146/annurev.micro.091208.073214
- Gao, Y. R., Li, D. F., Fleming, J., Zhou, Y. F., Liu, Y., Deng, J. Y., et al. (2017). Structural analysis of the regulatory mechanism of MarR protein Rv2887 in *M. tuberculosis*. *Sci Rep* 7:6471. doi: 10.1038/s41598-017-01705-4
- Ghosh, S., Cremers, C. M., Jakob, U., and Love, N. G. (2011). Chlorinated phenols control the expression of the multidrug resistance efflux pump MexAB-OprM in *Pseudomonas aeruginosa* by interacting with NalC. *Mol. Microbiol.* 79, 1547–1556. doi: 10.1111/j.1365-2958.2011.07544.x

- Gillette, W. K., Martin, R. G., and Rosner, J. L. (2000). Probing the *Escherichia coli* transcriptional activator MarA using alanine-scanning mutagenesis: residues important for DNA binding and activation. *J. Mol. Biol.* 299, 1245–1255. doi: 10.1006/jmbi.2000.3827
- Goethals, K., Van Montagu, M., and Holsters, M. (1992). Conserved motifs in a divergent nod box of *Azorhizobium caulinodans* ORS571 reveal a common structure in promoters regulated by LysR-type proteins. *Proc. Natl. Acad. Sci. U.S.A.* 89, 1646–1650. doi: 10.1073/pnas.89.5.1646
- Gooderham, W. J., and Hancock, R. E. (2009). Regulation of virulence and antibiotic resistance by two-component regulatory systems in *Pseudomonas aeruginosa*. *FEMS Microbiol. Rev.* 33, 279–294. doi: 10.1111/j.1574-6976.2008.00135.x
- Griffith, K. L., and Wolf, R. E. Jr. (2002). A comprehensive alanine scanning mutagenesis of the *Escherichia coli* transcriptional activator SoxS: identifying amino acids important for DNA binding and transcription activation. *J. Mol. Biol.* 322, 237–257. doi: 10.1016/S0022-2836(02)00782-9
- Grkovic, S., Brown, M. H., Schumacher, M. A., Brennan, R. G., and Skurray, R. A. (2001). The staphylococcal QacR multidrug regulator binds a correctly spaced operator as a pair of dimers. *J. Bacteriol.* 183, 7102–7109. doi: 10.1128/JB.183.24.7102-7109.2001
- Grkovic, S., Brown, M. H., and Skurray, R. A. (2002). Regulation of bacterial drug export systems. *Microbiol. Mol. Biol. Rev.* 66, 671–701, table of contents. doi: 10.1128/MMBR.66.4.671-701.2002
- Grove, A. (2017). Regulation of metabolic pathways by MarR family transcription factors. *Comput. Struct. Biotechnol. J.* 15, 366–371. doi: 10.1016/j.csbj.2017.06.001
- Guénard, S., Muller, C., Monlezun, L., Benas, P., Broutin, I., Jeannot, K., et al. (2014). Multiple mutations lead to MexXY-OprM-dependent aminoglycoside resistance in clinical strains of *Pseudomonas aeruginosa*. *Antimicrob. Agents Chemother.* 58, 221–228. doi: 10.1128/AAC.01252-13
- Gushchin, I., Melnikov, I., Polovinkin, V., Ishchenko, A., Yuzhakova, A., Buslaev, P., et al. (2017). Mechanism of transmembrane signaling by sensor histidine kinases. *Science* 356:eaah6345. doi: 10.1126/science.aah6345
- Hay, T., Fraud, S., Lau, C. H., Gilmour, C., and Poole, K. (2013). Antibiotic inducibility of the mexXY multidrug efflux operon of *Pseudomonas aeruginosa*: involvement of the MexZ anti-repressor ArmZ. *PLoS ONE* 8:e56858. doi: 10.1371/journal.pone.0056858
- He, X., Wang, L., and Wang, S. (2016). Structural basis of DNA sequence recognition by the response regulator PhoP in *Mycobacterium tuberculosis*. *Sci. Rep.* 6:24442. doi: 10.1038/srep24442
- Hede, K. (2014). Antibiotic resistance: an infectious arms race. *Nature* 509, S2–S3. doi: 10.1038/509S2a
- Henikoff, S., Haughn, G. W., Calvo, J. M., and Wallace, J. C. (1988). A large family of bacterial activator proteins. *Proc. Natl. Acad. Sci. U.S.A.* 85, 6602–6606. doi: 10.1073/pnas.85.18.6602
- Heroven, A. K., and Dersch, P. (2006). RovM, a novel LysR-type regulator of the virulence activator gene rovA, controls cell invasion, virulence and motility of *Yersinia pseudotuberculosis*. *Mol. Microbiol.* 62, 1469–1483. doi: 10.1111/j.1365-2958.2006.05458.x
- Hinrichs, W., Kisker, C., Düvel, M., Müller, A., Tovar, K., Hillen, W., et al. (1994). Structure of the Tet repressor-tetracycline complex and regulation of antibiotic resistance. *Science* 264, 418–420. doi: 10.1126/science.8153629
- Hvorup, R. N., Winnen, B., Chang, A. B., Jiang, Y., Zhou, X. F., and Saier, M. H. Jr. (2003). The multidrug/oligosaccharidyl-lipid/polysaccharide (MOP) exporter superfamily. *Eur. J. Biochem.* 270, 799–813. doi: 10.1046/j.1432-1033.2003.03418.x
- Ibarra, J. A., Pérez-Rueda, E., Segovia, L., and Puente, J. L. (2008). The DNA-binding domain as a functional indicator: the case of the AraC/XylS family of transcription factors. *Genetica* 133, 65–76. doi: 10.1007/s10709-007-9185-y
- Itou, H., Watanabe, N., Yao, M., Shirakihara, Y., and Tanaka, I. (2010). Crystal structures of the multidrug binding repressor *Corynebacterium glutamicum* CgmR in complex with inducers and with an operator. *J. Mol. Biol.* 403, 174–184. doi: 10.1016/j.jmb.2010.07.042
- Jiang, Y. L., Wang, X. P., Sun, H., Han, S. J., Li, W. F., Cui, N., et al. (2018). Coordinating carbon and nitrogen metabolic signaling through the cyanobacterial global repressor NdhR. *Proc. Natl. Acad. Sci. U.S.A.* 115, 403–408. doi: 10.1073/pnas.1716062115
- Jo, I., Chung, I. Y., Bae, H. W., Kim, J. S., Song, S., Cho, Y. H., et al. (2015). Structural details of the OxyR peroxide-sensing mechanism. *Proc. Natl. Acad. Sci. U.S.A.* 112, 6443–6448. doi: 10.1073/pnas.1424495112
- Jo, I., Kim, D., Bang, Y. J., Ahn, J., Choi, S. H., and Ha, N. C. (2017). The hydrogen peroxide hypersensitivity of OxyR2 in *Vibrio vulnificus* depends on conformational constraints. *J. Biol. Chem.* 292, 7223–7232. doi: 10.1074/jbc.M116.743765
- Juarez, P., Broutin, I., Bordi, C., Plésiat, P., and Llanes, C. (2018). Constitutive activation of MexT by amino acid substitutions results in MexEF-OprN overproduction in clinical isolates of *Pseudomonas aeruginosa*. *Antimicrob. Agents Chemother.* 62:e02445–17. doi: 10.1128/AAC.02445-17
- Juarez, P., Jeannot, K., Plésiat, P., and Llanes, C. (2017). Toxic electrophiles induce expression of the multidrug efflux pump MexEF-OprN in *Pseudomonas aeruginosa* through a novel transcriptional regulator, CmrA. *Antimicrob. Agents Chemother.* 61. doi: 10.1128/AAC.00585-17
- Jung, K., Fried, L., Behr, S., and Heermann, R. (2012). Histidine kinases and response regulators in networks. *Curr. Opin. Microbiol.* 15, 118–124. doi: 10.1016/j.mib.2011.11.009
- Kallscheuer, N., Vogt, M., Kappelman, J., Krumbach, K., Noack, S., Bott, M., et al. (2016). Identification of the phd gene cluster responsible for phenylpropanoid utilization in *Corynebacterium glutamicum*. *Appl. Microbiol. Biotechnol.* 100, 1871–1881. doi: 10.1007/s00253-015-7165-1
- Khameneh, B., Diab, R., Ghazvini, K., and Fazly Bazzaz, B. S. (2016). Breakthroughs in bacterial resistance mechanisms and the potential ways to combat them. *Microb. Pathog.* 95, 32–42. doi: 10.1016/j.micpath.2016.02.009
- Kim, Y., Joachimiak, G., Bigelow, L., Babnigg, G., and Joachimiak, A. (2016). How aromatic compounds block DNA binding of HcaR catabolite regulator. *J. Biol. Chem.* 291, 13243–13256. doi: 10.1074/jbc.M115.712067
- Kisker, C., Hinrichs, W., Tovar, K., Hillen, W., and Saenger, W. (1995). The complex formed between Tet repressor and tetracycline-Mg²⁺ reveals mechanism of antibiotic resistance. *J. Mol. Biol.* 247, 260–280. doi: 10.1006/jmbi.1994.0138
- Kitao, T., Lepine, F., Babloui, S., Walte, F., Steinbacher, S., Maskos, K., et al. (2018). Molecular insights into function and competitive inhibition of *Pseudomonas aeruginosa* multiple virulence factor regulator. *mBio* 9:e02158-17. doi: 10.1128/mBio.02158-17
- Koentjoro, M. P., Adachi, N., Senda, M., Ogawa, N., and Senda, T. (2018). Crystal structure of the DNA-binding domain of the LysR-type transcriptional regulator CbnR in complex with a DNA fragment of the recognition-binding site in the promoter region. *FEBS J.* 285, 977–989. doi: 10.1111/febs.14380
- Krell, T., Lacal, J., Busch, A., Silva-Jiménez, H., Guazzaroni, M. E., and Ramos, J. L. (2010). Bacterial sensor kinases: diversity in the recognition of environmental signals. *Annu. Rev. Microbiol.* 64, 539–559. doi: 10.1146/annurev.micro.112408.134054
- Kumar, S., Mukherjee, M. M., and Varela, M. F. (2013). Modulation of bacterial multidrug resistance efflux pumps of the major facilitator superfamily. *Int. J. Bacteriol.* 2013:204141. doi: 10.1155/2013/204141
- Kumarevel, T. (2012). “The MarR Family of Transcriptional Regulators – A Structural Perspective,” in *Antibiotic Resistant Bacteria – A Continuous Challenge in the New Millennium*, ed M. Pana (London, UK: InTech), 403–418. doi: 10.5772/1058
- Kumarevel, T., Tanaka, T., Umehara, T., and Yokoyama, S. (2009). ST1710-DNA complex crystal structure reveals the DNA binding mechanism of the MarR family of regulators. *Nucleic Acids Res.* 37, 4723–4735. doi: 10.1093/nar/gkp496
- Kuroda, T., and Tsuchiya, T. (2009). Multidrug efflux transporters in the MATE family. *Biochim. Biophys. Acta* 1794, 763–768. doi: 10.1016/j.bbapap.2008.11.012
- Kwon, H. J., Bennis, M. H., Demple, B., and Ellenberger, T. (2000). Crystal structure of the *Escherichia coli* Rob transcription factor in complex with DNA. *Nat. Struct. Biol.* 7, 424–430. doi: 10.1038/75213
- Lau, C. H., Krahn, T., Gilmour, C., Mullen, E., and Poole, K. (2015). AmgRS-mediated envelope stress-inducible expression of the mexXY multidrug efflux operon of *Pseudomonas aeruginosa*. *Microbiologyopen* 4, 121–135. doi: 10.1002/mbo3.226
- Le, T. B., Schumacher, M. A., Lawson, D. M., Brennan, R. G., and Buttner, M. J. (2011). The crystal structure of the TetR family transcriptional repressor SimR bound to DNA and the role of a flexible N-terminal extension in minor groove binding. *Nucleic Acids Res.* 39, 9433–9447. doi: 10.1093/nar/gkr640

- Lerche, M., Dian, C., Round, A., Lönneborg, R., Brzezinski, P., and Leonard, G. A. (2016). The solution configurations of inactive and activated DntR have implications for the sliding dimer mechanism of LysR transcription factors. *Sci. Rep.* 6:19988. doi: 10.1038/srep19988
- Levy, S. B., and Marshall, B. (2004). Antibacterial resistance worldwide: causes, challenges and responses. *Nat. Med.* 10, S122–S129. doi: 10.1038/nm1145
- Li, D. F., Zhang, N., Hou, Y. J., Huang, Y., Hu, Y., Zhang, Y., et al. (2011). Crystal structures of the transcriptional repressor RolR reveals a novel recognition mechanism between inducer and regulator. *PLoS ONE* 6:e19529. doi: 10.1371/journal.pone.0019529
- Li, J., Wehmeyer, G., Lovell, S., Battaile, K. P., and Egan, S. M. (2016). 1.65 Å resolution structure of the AraC-family transcriptional activator ToxT from *Vibrio cholerae*. *Acta Crystallogr. F Struct. Biol. Commun.* 72(Pt 9), 726–731. doi: 10.1107/S2053230X1601298X
- Li, X. Z., and Nikaido, H. (2009). Efflux-mediated drug resistance in bacteria: an update. *Drugs* 69, 1555–1623. doi: 10.2165/11317030-000000000-00000
- Li, X. Z., Plésiat, P., and Nikaido, H. (2015). The challenge of efflux-mediated antibiotic resistance in Gram-negative bacteria. *Clin. Microbiol. Rev.* 28, 337–418. doi: 10.1128/CMR.00117-14
- Lim, D., Poole, K., and Strynadka, N. C. (2002). Crystal structure of the MexR repressor of the mexRAB-oprM multidrug efflux operon of *Pseudomonas aeruginosa*. *J. Biol. Chem.* 277, 29253–29259. doi: 10.1074/jbc.M111381200
- Ling, L. L., Schneider, T., Peoples, A. J., Spoering, A. L., Engels, I., Conlon, B. P., et al. (2015). A new antibiotic kills pathogens without detectable resistance. *Nature* 517, 455–459. doi: 10.1038/nature14098
- Lister, P. D., Wolter, D. J., and Hanson, N. D. (2009). Antibacterial-resistant *Pseudomonas aeruginosa*: clinical impact and complex regulation of chromosomally encoded resistance mechanisms. *Clin. Microbiol. Rev.* 22, 582–610. doi: 10.1128/CMR.00040-09
- Liu, G., Liu, X., Xu, H., Liu, X., Zhou, H., Huang, Z., et al. (2017). Structural insights into the redox-sensing mechanism of MarR-type regulator AbfR. *J. Am. Chem. Soc.* 139, 1598–1608. doi: 10.1021/jacs.6b11438
- Locher, K. P. (2009). Review. Structure and mechanism of ATP-binding cassette transporters. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 364, 239–245. doi: 10.1098/rstb.2008.0125
- Lou, Y. C., Weng, T. H., Li, Y. C., Kao, Y. F., Lin, W. F., Peng, H. L., et al. (2015). Structure and dynamics of polymyxin-resistance-associated response regulator PmrA in complex with promoter DNA. *Nat. Commun.* 6:8838. doi: 10.1038/ncomms9838
- Lowden, M. J., Skorupski, K., Pellegrini, M., Chiorazzo, M. G., Taylor, R. K., and Kull, F. J. (2010). Structure of *Vibrio cholerae* ToxT reveals a mechanism for fatty acid regulation of virulence genes. *Proc. Natl. Acad. Sci. U.S.A.* 107, 2860–2865. doi: 10.1073/pnas.0915021107
- Maddocks, S. E., and Oyston, P. C. (2008). Structure and function of the LysR-type transcriptional regulator (LTTR) family proteins. *Microbiology* 154(Pt 12), 3609–3623. doi: 10.1099/mic.0.2008/022772-0
- Martin, R. G., and Rosner, J. L. (1995). Binding of purified multiple antibiotic-resistance repressor protein (MarR) to mar operator sequences. *Proc. Natl. Acad. Sci. U.S.A.* 92, 5456–5460. doi: 10.1073/pnas.92.12.5456
- McDermott, P. F., Walker, R. D., and White, D. G. (2003). Antimicrobials: modes of action and mechanisms of resistance. *Int. J. Toxicol.* 22, 135–143. doi: 10.1080/10915810305089
- Martin, R. G., and Rosner, J. L. (2001). The AraC transcriptional activators. *Curr. Opin. Microbiol.* 4, 132–137. doi: 10.1016/S1369-5274(00)00178-8
- Mechaly, A. E., Sassoon, N., Betton, J. M., and Alzari, P. M. (2014). Segmental helical motions and dynamical asymmetry modulate histidine kinase autophosphorylation. *PLoS Biol.* 12:e1001776. doi: 10.1371/journal.pbio.1001776
- Miller, D. J., Zhang, Y. M., Subramanian, C., Rock, C. O., and White, S. W. (2010). Structural basis for the transcriptional regulation of membrane lipid homeostasis. *Nat. Struct. Mol. Biol.* 17, 971–975. doi: 10.1038/nsmb.1847
- Molnar, K. S., Bonomi, M., Pellarin, R., Clinthorne, G. D., Gonzalez, G., Goldberg, S. D., et al. (2014). Cys-scanning disulfide crosslinking and bayesian modeling probe the transmembrane signaling mechanism of the histidine kinase, PhoQ. *Structure* 22, 1239–1251. doi: 10.1016/j.str.2014.04.019
- Monferrer, D., Tralau, T., Kertesz, M. A., Dix, I., Solá, M., and Usón, I. (2010). Structural studies on the full-length LysR-type regulator Tsar from *Comamonas testosteroni* T-2 reveal a novel open conformation of the tetrameric LTTR fold. *Mol. Microbiol.* 75, 1199–1214. doi: 10.1111/j.1365-2958.2010.07043.x
- Monlezun, L., Phan, G., Benabdelhak, H., Lascombe, M. B., Enguéné, V. Y., Picard, M., et al. (2015). New OprM structure highlighting the nature of the N-terminal anchor. *Front. Microbiol.* 6:667. doi: 10.3389/fmicb.2015.00667
- Morita, Y., Cao, L., Gould, V. C., Avison, M. B., and Poole, K. (2006). nalD encodes a second repressor of the mexAB-oprM multidrug efflux operon of *Pseudomonas aeruginosa*. *J. Bacteriol.* 188, 8649–8654. doi: 10.1128/JB.01342-06
- Morita, Y., Tomida, J., and Kawamura, Y. (2015). Efflux-mediated fluoroquinolone resistance in the multidrug-resistant *Pseudomonas aeruginosa* clinical isolate PA7: identification of a novel MexS variant involved in upregulation of the mexEF-oprN multidrug efflux operon. *Front. Microbiol.* 6:8. doi: 10.3389/fmicb.2015.00008
- Mousa, J. J., and Bruner, S. D. (2016). Structural and mechanistic diversity of multidrug transporters. *Nat. Prod. Rep.* 33, 1255–1267. doi: 10.1039/C6NP00006A
- Muller, C., Plésiat, P., and Jeannot, K. (2011). A two-component regulatory system interconnects resistance to polymyxins, aminoglycosides, fluoroquinolones, and beta-lactams in *Pseudomonas aeruginosa*. *Antimicrob. Agents Chemother.* 55, 1211–1221. doi: 10.1128/AAC.01252-10
- Muller, J. F., Stevens, A. M., Craig, J., and Love, N. G. (2007). Transcriptome analysis reveals that multidrug efflux genes are upregulated to protect *Pseudomonas aeruginosa* from pentachlorophenol stress. *Appl. Environ. Microbiol.* 73, 4550–4558. doi: 10.1128/AEM.00169-07
- Muraoka, S., Okumura, R., Ogawa, N., Nonaka, T., Miyashita, K., and Senda, T. (2003). Crystal structure of a full-length LysR-type transcriptional regulator, CbnR: unusual combination of two subunit forms and molecular bases for causing and changing DNA bend. *J. Mol. Biol.* 328, 555–566. doi: 10.1016/S0022-2836(03)00312-7
- Narayanan, A., Kumar, S., Evrard, A. N., Paul, L. N., and Yernool, D. A. (2014). An asymmetric heterodomain interface stabilizes a response regulator-DNA complex. *Nat. Commun.* 5:3282. doi: 10.1038/ncomms4282
- Newberry, K. J., Fuangthong, M., Panmanee, W., Mongkolsuk, S., and Brennan, R. G. (2007). Structural mechanism of organic hydroperoxide induction of the transcription regulator OhrR. *Mol. Cell* 28, 652–664. doi: 10.1016/j.molcel.2007.09.016
- Ni, L., Tonthat, N. K., Chinnam, N., and Schumacher, M. A. (2013). Structures of the *Escherichia coli* transcription activator and regulator of diauxie, XylR: an AraC DNA-binding family member with a LacI/GalR ligand-binding domain. *Nucleic Acids Res.* 41, 1998–2008. doi: 10.1093/nar/gks1207
- Nikaido, H. (2009). Multidrug resistance in bacteria. *Annu. Rev. Biochem.* 78, 119–146. doi: 10.1146/annurev.biochem.78.082907.145923
- Nikaido, H. (2018). RND transporters in the living world. *Res. Microbiol.* doi: 10.1016/j.resmic.2018.03.001. [Epub ahead of print].
- Nikiforov, P. O., Blaszczyk, M., Surade, S., Boshoff, H. I., Sajid, A., Delorme, V., et al. (2017). Fragment-sized EthR inhibitors exhibit exceptionally strong ethionamide boosting effect in whole-cell mycobacterium tuberculosis assays. *ACS Chem. Biol.* 12, 1390–1396. doi: 10.1021/acschembio.7b00091
- Oliver, P., Peralta-Gil, M., Tabche, M. L., and Merino, E. (2016). Molecular and structural considerations of TF-DNA binding for the generation of biologically meaningful and accurate phylogenetic footprinting analysis: the LysR-type transcriptional regulator family as a study model. *BMC Genomics* 17:686. doi: 10.1186/s12864-016-3025-3
- Orth, P., Schnappinger, D., Hillen, W., Saenger, W., and Hinrichs, W. (2000). Structural basis of gene regulation by the tetracycline inducible Tet repressor-operator system. *Nat. Struct. Mol. Biol.* 7, 215–219. doi: 10.1038/73324
- Palanca, C., and Rubio, V. (2016). Structure of AmtR, the global nitrogen regulator of *Corynebacterium glutamicum*, in free and DNA-bound forms. *FEBS J.* 283, 1039–1059. doi: 10.1111/febs.13643
- Perera, I. C., and Grove, A. (2010). Molecular mechanisms of ligand-mediated attenuation of DNA binding by MarR family transcriptional regulators. *J. Mol. Cell Biol.* 2, 243–254. doi: 10.1093/jmcb/mjq021
- Perron, K., Caille, O., Rossier, C., Van Delden, C., Dumas, J. L., and Köhler, T. (2004). Czcr-CzcS, a two-component system involved in heavy metal and carbapenem resistance in *Pseudomonas aeruginosa*. *J. Biol. Chem.* 279, 8761–8768. doi: 10.1074/jbc.M312080200

- Phan, G., Benabdelhak, H., Lascombe, M. B., Benas, P., Rety, S., Picard, M., et al. (2010). Structural and dynamical insights into the opening mechanism of *P. aeruginosa* OprM channel. *Structure* 18, 507–517. doi: 10.1016/j.str.2010.01.018
- Phan, G., Picard, M., and Broutin, I. (2015). Focus on the outer membrane factor OprM, the forgotten player from efflux pumps assemblies. *Antibiotics* 4, 544–566. doi: 10.3390/antibiotics4040544
- Picossi, S., Belitsky, B. R., and Sonenshein, A. L. (2007). Molecular mechanism of the regulation of *Bacillus subtilis* gltAB expression by GltC. *J. Mol. Biol.* 365, 1298–1313. doi: 10.1016/j.jmb.2006.10.100
- Poole, K. (2008). Bacterial multidrug efflux pumps serve other functions. *Microbe* 3, 179–185. doi: 10.1128/microbe.3.179.1
- Poole, K. (2011). *Pseudomonas aeruginosa*: resistance to the max. *Front. Microbiol.* 2:65. doi: 10.3389/fmicb.2011.00065
- Poole, K., and Srikanth, R. (2001). Multidrug efflux in *Pseudomonas aeruginosa*: components, mechanisms and clinical significance. *Curr. Top. Med. Chem.* 1, 59–71. doi: 10.2174/1568026013395605
- Porriá, O., García-Jaramillo, M., Santero, E., and Govantes, F. (2007). The LysR-type regulator AtzR binding site: DNA sequences involved in activation, repression and cyanuric acid-dependent repositioning. *Mol. Microbiol.* 66, 410–427. doi: 10.1111/j.1365-2958.2007.05927.x
- Ptashne, M., and Gann, A. (1997). Transcriptional activation by recruitment. *Nature* 386, 569–577. doi: 10.1038/386569a0
- Rahman, T., Yarnall, B., and Doyle, D. A. (2017). Efflux drug transporters at the forefront of antimicrobial resistance. *Eur. Biophys. J.* 46, 647–653. doi: 10.1007/s00249-017-1238-2
- Ramos, J. L., Martínez-Bueno, M., Molina-Henares, A. J., Terán, W., Watanabe, K., Zhang, X., et al. (2005). The TetR family of transcriptional repressors. *Microbiol. Mol. Biol. Rev.* 69, 326–356. doi: 10.1128/MMBR.69.2.326-356.2005
- Reeder, T., and Schleif, R. (1993). AraC protein can activate transcription from only one position and when pointed in only one direction. *J. Mol. Biol.* 231, 205–218. doi: 10.1006/jmbi.1993.1276
- Reichheld, S. E., Yu, Z., and Davidson, A. R. (2009). The induction of folding cooperativity by ligand binding drives the allosteric response of tetracycline repressor. *Proc. Natl. Acad. Sci. U.S.A.* 106, 22263–22268. doi: 10.1073/pnas.0911566106
- Resch, M., Striegl, H., Hensler, E. M., Sevana, M., Egerer-Sieber, C., Schiltz, E., et al. (2008). A protein functional leap: how a single mutation reverses the function of the transcription regulator TetR. *Nucleic Acids Res.* 36, 4390–4401. doi: 10.1093/nar/gkn400
- Rhee, S., Martin, R. G., Rosner, J. L., and Davies, D. R. (1998). A novel DNA-binding motif in MarA: the first structure for an AraC family transcriptional activator. *Proc. Natl. Acad. Sci. U.S.A.* 95, 10413–10418. doi: 10.1073/pnas.95.18.10413
- Richardot, C., Juarez, P., Jeannot, K., Patry, I., Plésiat, P., and Llanes, C. (2016). Amino acid substitutions account for most MexS alterations in clinical *nfxC* mutants of *Pseudomonas aeruginosa*. *Antimicrob. Agents Chemother.* 60, 2302–2310. doi: 10.1128/AAC.02622-15
- Rodrigue, A., Quentin, Y., Lazdunski, A., Méjean, V., and Foglino, M. (2000). Two-component systems in *Pseudomonas aeruginosa*: why so many? *Trends Microbiol.* 8, 498–504. doi: 10.1016/S0966-842X(00)01833-3
- Rolain, J. M., Abat, C., Jimeno, M. T., Fournier, P. E., and Raoult, D. (2016). Do we need new antibiotics? *Clin. Microbiol. Infect.* 22, 408–415. doi: 10.1016/j.cmi.2016.03.012
- Saridakis, V., Shahinas, D., Xu, X., and Christendat, D. (2008). Structural insight on the mechanism of regulation of the MarR family of proteins: high-resolution crystal structure of a transcriptional repressor from *Methanobacterium thermoautotrophicum*. *J. Mol. Biol.* 377, 655–667. doi: 10.1016/j.jmb.2008.01.001
- Sawai, H., Yamanaka, M., Sugimoto, H., Shiro, Y., and Aono, S. (2012). Structural basis for the transcriptional regulation of heme homeostasis in *Lactococcus lactis*. *J. Biol. Chem.* 287, 30755–30768. doi: 10.1074/jbc.M112.370916
- Schell, M. A. (1993). Molecular biology of the LysR family of transcriptional regulators. *Annu. Rev. Microbiol.* 47, 597–626. doi: 10.1146/annurev.mi.47.100193.003121
- Schell, M. A., Brown, P. H., and Raju, S. (1990). Use of saturation mutagenesis to localize probable functional domains in the NahR protein, a LysR-type transcription activator. *J. Biol. Chem.* 265, 3844–3850.
- Schleif, R. (2003). AraC protein: a love-hate relationship. *Bioessays* 25, 274–282. doi: 10.1002/bies.10237
- Schleif, R. (2010). AraC protein, regulation of the l-arabinose operon in *Escherichia coli*, and the light switch mechanism of AraC action. *FEMS Microbiol. Rev.* 34, 779–796. doi: 10.1111/j.1574-6976.2010.00226.x
- Schuldiner, S. (2009). EmrE, a model for studying evolution and mechanism of ion-coupled transporters. *Biochim. Biophys. Acta* 1794, 748–762. doi: 10.1016/j.bbapap.2008.12.018
- Schumacher, M. A., and Brennan, R. G. (2002). Structural mechanisms of multidrug recognition and regulation by bacterial multidrug transcription factors. *Mol. Microbiol.* 45, 885–893. doi: 10.1046/j.1365-2958.2002.03039.x
- Schumacher, M. A., Miller, M. C., and Brennan, R. G. (2004). Structural mechanism of the simultaneous binding of two drugs to a multidrug-binding protein. *EMBO J.* 23, 2923–2930. doi: 10.1038/sj.emboj.7600288
- Schumacher, M. A., Miller, M. C., Grkovic, S., Brown, M. H., Skurray, R. A., and Brennan, R. G. (2001). Structural mechanisms of QacR induction and multidrug recognition. *Science* 294, 2158–2163. doi: 10.1126/science.1066020
- Schumacher, M. A., Miller, M. C., Grkovic, S., Brown, M. H., Skurray, R. A., and Brennan, R. G. (2002). Structural basis for cooperative DNA binding by two dimers of the multidrug-binding protein QacR. *EMBO J.* 21, 1210–1218. doi: 10.1093/emboj/21.5.1210
- Schumacher, M. A., and Zeng, W. (2016). Structures of the nucleoid occlusion protein SlmA bound to DNA and the C-terminal domain of the cytoskeletal protein FtsZ. *Proc. Natl. Acad. Sci. U.S.A.* 113, 4988–4993. doi: 10.1073/pnas.1602327113
- Sivaneson, M., Mikkelsen, H., Ventre, I., Bordi, C., and Filloux, A. (2011). Two-component regulatory systems in *Pseudomonas aeruginosa*: an intricate network mediating fimbrial and efflux pump gene expression. *Mol. Microbiol.* 79, 1353–1366. doi: 10.1111/j.1365-2958.2010.07527.x
- Smith, E. E., Buckley, D. G., Wu, Z., Saenphimmachak, C., Hoffman, L. R., D'Argenio, D. A., et al. (2006). Genetic adaptation by *Pseudomonas aeruginosa* to the airways of cystic fibrosis patients. *Proc. Natl. Acad. Sci. U.S.A.* 103, 8487–8492. doi: 10.1073/pnas.0602138103
- Sobel, M. L., Hocquet, D., Cao, L., Plesiat, P., and Poole, K. (2005). Mutations in PA3574 (*nalD*) lead to increased MexAB-OprM expression and multidrug resistance in laboratory and clinical isolates of *Pseudomonas aeruginosa*. *Antimicrob. Agents Chemother.* 49, 1782–1786. doi: 10.1128/AAC.49.5.1782-1786.2005
- Soisson, S. M., MacDougall-Shackleton, B., Schleif, R., and Wolberger, C. (1997). Structural basis for ligand-regulated oligomerization of AraC. *Science* 276, 421–425. doi: 10.1126/science.276.5311.421
- Spengler, G., Kincses, A., Gajdacs, M., and Amaral, L. (2017). New roads leading to old destinations: efflux pumps as targets to reverse multidrug resistance in bacteria. *Molecules* 22:E468. doi: 10.3390/molecules22030468
- Stock, A. M., Robinson, V. L., and Goudreau, P. N. (2000). Two-component signal transduction. *Annu. Rev. Biochem.* 69, 183–215. doi: 10.1146/annurev.biochem.69.1.183
- Stover, C. K., Pham, X. Q., Erwin, A. L., Mizoguchi, S. D., Warriner, P., Hickey, M. J., et al. (2000). Complete genome sequence of *Pseudomonas aeruginosa* PAO1, an opportunistic pathogen. *Nature* 406, 959–964. doi: 10.1038/35023079
- Stragier, P., Richaud, F., Borne, F., and Patte, J. C. (1983). Regulation of diaminopimelate decarboxylase synthesis in *Escherichia coli*. I. Identification of a lysR gene encoding an activator of the *lysA* gene. *J. Mol. Biol.* 168, 307–320. doi: 10.1016/S0022-2836(83)80020-5
- Sun, J., Deng, Z., and Yan, A. (2014). Bacterial multidrug efflux pumps: mechanisms, physiology and pharmacological exploitations. *Biochem. Biophys. Res. Commun.* 453, 254–267. doi: 10.1016/j.bbrc.2014.05.090
- Szakács, G., Varadi, A., Ozvegy-Laczka, C., and Sarkadi, B. (2008). The role of ABC transporters in drug absorption, distribution, metabolism, excretion and toxicity (ADME-Tox). *Drug Discov. Today* 13, 379–393. doi: 10.1016/j.drudis.2007.12.010
- Tacconelli, E., Carrara, E., Savoldi, A., Harbarth, S., Mendelson, M., Monnet, D. L., et al. (2017). Discovery, research, and development of new antibiotics: the WHO priority list of antibiotic-resistant bacteria and tuberculosis. *Lancet Infect. Dis.* 18, 318–327. doi: 10.1016/S1473-3099(17)30753-3

- Tyrrell, R., Verschuere, K. H., Dodson, E. J., Murshudov, G. N., Addy, C., and Wilkinson, A. J. (1997). The structure of the cofactor-binding fragment of the LysR family member, CysB: a familiar fold with a surprising subunit arrangement. *Structure* 5, 1017–1032. doi: 10.1016/S0969-2126(97)00254-2
- Vargiu, A. V., Pos, K. M., Poole, K., and Nikaido, H. (2016). Editorial: bad bugs in the XXIst century: resistance mediated by multi-drug efflux pumps in gram-negative bacteria. *Front. Microbiol.* 7:833. doi: 10.3389/fmicb.2016.00833
- Wang, C., Sang, J., Wang, J., Su, M., Downey, J. S., Wu, Q., et al. (2013). Mechanistic insights revealed by the crystal structure of a histidine kinase with signal transducer and sensor domains. *PLoS Biol.* 11:e1001493. doi: 10.1371/journal.pbio.1001493
- Wang, D., Chen, W., Huang, S., He, Y., Liu, X., Hu, Q., et al. (2017). Structural basis of Zn(II) induced metal detoxification and antibiotic resistance by histidine kinase CzcS in *Pseudomonas aeruginosa*. *PLoS Pathog.* 13:e1006533. doi: 10.1371/journal.ppat.1006533
- Wang, D., Seeve, C., Pierson, L. S., and Pierson, E. A. (2013). Transcriptome profiling reveals links between ParS/ParR, MexEF-OprN, and quorum sensing in the regulation of adaptation and virulence in *Pseudomonas aeruginosa*. *BMC Genomics* 14:618. doi: 10.1186/1471-2164-14-618
- Wang, L., and Winans, S. C. (1995). High angle and ligand-induced low angle DNA bends incited by OccR lie in the same plane with OccR bound to the interior angle. *J. Mol. Biol.* 253, 32–38. doi: 10.1006/jmbi.1995.0533
- Wang, Z., Fan, G., Hryc, C. F., Blaza, J. N., Serysheva, I. I., Schmid, M. F., et al. (2017). An allosteric transport mechanism for the AcrAB-TolC multidrug efflux pump. *Elife* 6:e24905. doi: 10.7554/eLife.24905
- Weldon, J. E., Rodgers, M. E., Larkin, C., and Schleif, R. F. (2007). Structure and properties of a truly apo form of AraC dimerization domain. *Proteins* 66, 646–654. doi: 10.1002/prot.21267
- Wilke, M. S., Heller, M., Creagh, A. L., Haynes, C. A., McIntosh, L. P., Poole, K., et al. (2008). The crystal structure of MexR from *Pseudomonas aeruginosa* in complex with its antirepressor ArmR. *Proc. Natl. Acad. Sci. U.S.A.* 105, 14832–14837. doi: 10.1073/pnas.0805489105
- Wilkinson, S. P., and Grove, A. (2006). Ligand-responsive transcriptional regulation by members of the MarR family of winged helix proteins. *Curr. Issues Mol. Biol.* 8, 51–62. Available online at: <http://www.caister.com/cimb/v/v8/51.pdf>
- Willems, A. R., Tahlan, K., Taguchi, T., Zhang, K., Lee, Z. Z., Ichinose, K., et al. (2008). Crystal structures of the *Streptomyces coelicolor* TetR-like protein ActR alone and in complex with actinorhodin or the actinorhodin biosynthetic precursor (S)-DNPA. *J. Mol. Biol.* 376, 1377–1387. doi: 10.1016/j.jmb.2007.12.061
- Yamaguchi, A., Nakashima, R., and Sakurai, K. (2015). Structural basis of RND-type multidrug exporters. *Front. Microbiol.* 6:327. doi: 10.3389/fmicb.2015.00327
- Yan, N. (2015). Structural Biology of the Major Facilitator Superfamily Transporters. *Annu. Rev. Biophys.* 44, 257–283. doi: 10.1146/annurev-biophys-060414-033901
- Yang, J., Dogovski, C., Hocking, D., Tauschek, M., Perugini, M., and Robins-Browne, R. M. (2009). Bicarbonate-mediated stimulation of RegA, the global virulence regulator from *Citrobacter rodentium*. *J. Mol. Biol.* 394, 591–599. doi: 10.1016/j.jmb.2009.10.033
- Yang, J., Tauschek, M., and Robins-Browne, R. M. (2011). Control of bacterial virulence by AraC-like regulators that respond to chemical signals. *Trends Microbiol.* 19, 128–135. doi: 10.1016/j.tim.2010.12.001
- Yang, S., Gao, Z., Li, T., Yang, M., Zhang, T., Dong, Y., et al. (2013). Structural basis for interaction between *Mycobacterium smegmatis* Ms6564, a TetR family master regulator, and its target DNA. *J. Biol. Chem.* 288, 23687–23695. doi: 10.1074/jbc.M113.468694
- Yeo, H. K., Park, Y. W., and Lee, J. Y. (2017). Structural basis of operator sites recognition and effector binding in the TetR family transcription regulator FadR. *Nucleic Acids Res.* 45, 4244–4254. doi: 10.1093/nar/gkx009
- Yu, Z., Reichheld, S. E., Savchenko, A., Parkinson, J., and Davidson, A. R. (2010). A comprehensive analysis of structural and sequence conservation in the TetR family transcriptional regulators. *J. Mol. Biol.* 400, 847–864. doi: 10.1016/j.jmb.2010.05.062
- Zhou, X., Lou, Z., Fu, S., Yang, A., Shen, H., Li, Z., et al. (2010). Crystal structure of ArgP from *Mycobacterium tuberculosis* confirms two distinct conformations of full-length LysR transcriptional regulators and reveals its function in DNA binding and transcriptional regulation. *J. Mol. Biol.* 396, 1012–1024. doi: 10.1016/j.jmb.2009.12.033
- Zhu, R., Song, Y., Liu, H., Yang, Y., Wang, S., Yi, C., et al. (2017). Allosteric histidine switch for regulation of intracellular zinc(II) fluctuation. *Proc. Natl. Acad. Sci. U.S.A.* 114, 13661–13666. doi: 10.1073/pnas.1708563115
- Zschiedrich, C. P., Keidel, V., and Szurmant, H. (2016). Molecular mechanisms of two-component signal transduction. *J. Mol. Biol.* 428, 3752–3775. doi: 10.1016/j.jmb.2016.08.003

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Housseini B Issa, Phan and Broutin. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.