

# Theoretical and experimental adhesion of yeast strains with high chromium removal potential

Meryem Asri, Alae Elabed, Naïma El Ghachtouli, Saad Ibnsouda Koraichi, Wifak Bahafid, Soumya Elabed

#### ▶ To cite this version:

Meryem Asri, Alae Elabed, Naïma El Ghachtouli, Saad Ibnsouda Koraichi, Wifak Bahafid, et al.. Theoretical and experimental adhesion of yeast strains with high chromium removal potential. Environmental Engineering Science, 2017, 34 (10), pp.693-702. 10.1089/ees.2016.0515 . hal-02134817

### HAL Id: hal-02134817 https://hal.science/hal-02134817v1

Submitted on 20 May 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



## Open Archive Toulouse Archive Ouverte (OATAO)

OATAO is an open access repository that collects the work of some Toulouse researchers and makes it freely available over the web where possible.

This is an author's version published in: http://oatao.univ-toulouse.fr/20354

Official URL: https://doi.org/10.1089/ees.2016.0515

#### To cite this version:

Asri, Meryem and Elabed, Alae and El Ghachtouli, Naïma and Koraichi, Saad Ibnsouda and Bahafid, Wifak and Elabed, Soumya Theoretical and experimental adhesion of yeast strains with high chromium removal potential. (2017) Environmental Engineering Science, 34 (10). 693-702. ISSN 1557-9018

Any correspondence concerning this service should be sent to the repository administrator: tech-oatao@listes-diff.inp-toulouse.fr

## Theoretical and Experimental Adhesion of Yeast Strains with High Chromium Removal Potential

Meryem Asri, Alae Elabed, Naïma El Ghachtouli, Saad Ibnsouda Koraichi, Wifak Bahafid, and Soumya Elabed, tabled

<sup>1</sup>Laboratoire de Biotechnologie Microbienne, Faculté des Sciences et Techniques, Université Sidi Mohamed Ben Abdellah, Fès, Maroc.

<sup>2</sup>Laboratoire de Génie Chimique, CNRS-Université de Toulouse (INPT), Toulouse, France.

<sup>3</sup>Centre Universitaire Régional d'Interface, Université Sidi Mohamed Ben Abdellah, Fès, Maroc.

#### **Abstract**

Biofilm-based bioprocesses are increasingly used in wastewater treatment. Microbial adhesion constitutes the key step in stability of these depollution systems. For adhesion studies, physicochemical characterization of microbial cells and supports has proved to be of extreme importance. In this work, estimation of interaction between five yeast strains with a high potential for Cr (VI) removal using extended Derjaguin–Landau–Verwey and Overbeek (XDLVO) theory as a powerful predictive tool of adhesion was investigated. Predictions showed that wood husk could be a good support for the formation of tested yeast biofilm, beech and oak exhibit better properties than other wood species studied with 100% of potential for adhesion. From a thermodynamic point of view, pine and teak woods are not suitable for biofilm formation for all tested yeast strains, presenting positive values of free energy adhesion ( $\Delta G^{XDLVO}$ ). Environmental scanning electronic microscopy (ESEM) and Matlab® image analysis confirmed that all tested yeast strains were able to adhere to pine wood and, except for *Wickerhamomyces anomalus* they were unable to adhere to oak wood. Adhesion experiments were found to be well related to the theoretical prediction. To our knowledge, this is the first study dealing with biofilm-mediated depollution from an adhesion point of view aiming to optimize the stability of the system. It allows expanding knowledge about adhesion phenomena of yeast strains on wooden surface and contributes to select the best biofilm-support combination that would be used in a performant biological system for chromium removal.

**Keywords:** adhesion; wastewater; wood husk; XDLVO theory; yeast strains

#### Introduction

O VER THE LAST few decades, biofilm-based bioprocesses have gained increasing attention in various applications. Several studies on the potential of biofilm communities for bioremediation processes have been realized (Silva *et al.*, 2008; Quintelas *et al.*, 2009). Due to the close, mutually beneficial physical and physiological interactions in a biofilm community, biofilm-mediated bioremediation appears to be a proficient alternative to bioremediation with planktonic microorganisms (Das *et al.*, 2013).

The stability of the biofilm during its functional lifetime is a key factor for its application in a wastewater treatment process. It is essentially related to initial microbial attachment on substrates. Thus, an optimal maintain of biofilm adhesion may provide efficient degradation of hazardous substances in a wastewater treatment process. This step of biofilm formation is a complicated process, enhanced by multifactorial parameters especially physicochemical properties of both support material and microbial surfaces (Elabed *et al.*, 2011).

In biofilm-based systems for wastewater treatment, many chemical materials have been used as support. The most frequently employed are granulated activated carbon (Quintelas *et al.*, 2008), zeolite (Silva *et al.*, 2008) and kaolin (Quintelas *et al.*, 2009). These materials are especially employed for their depolluting activity. However, they represent the disadvantage of being expensive. Therefore, natural materials are gaining increasing attention. Wood husk is a natural support increasingly employed, because it is considered as safe, cheap, and eco-friendly. It also represents a natural source of cellulose for cell growth (Zainul Akmar *et al.*, 2007). Very few works have dealt with interactions between fungal and bacterial adhesion on wood species and the different parameters enhancing the strength of cell-wood

<sup>\*</sup>Corresponding author: Laboratoire de Biotechnologie Microbienne, Faculté des Sciences et Techniques, Université Sidi Mohamed Ben Abdellah, Route Immouzer, PO Box 2202, Fez 30000, Morocco. *Phone:* +212 663 45 06 48; *Fax:* +212 (0) 535 60 82 14; *E-mail:* soumya.elabed@usmba.ac.ma

interactions (Elabed *et al.*, 2011, 2013). On the other hand, the majority of studies on biofilm-mediated bioremediation are focusing on bacterial strains (Quintelas *et al.*, 2008; Pan *et al.*, 2014), nevertheless, yeast strains present a good efficiency in accumulation and/or biosorption of pollutants including heavy metals like chromium (Ksheminska *et al.*, 2005; Bahafid *et al.*, 2013). Thus, there is a clear need to study yeast attachment on wood surfaces to form a potential biofilm that can be used in bioprocesses for treating wastewater rich in chromium.

A fundamental understanding of interactions between yeast and substratum surface is of extreme importance for characterizing cell attachment to wood husk. Interactions related to cell attachment to a given surface can be classified into Lifshitz-van der Waals interactions, electrostatic interactions, and polar or Lewis acid–base interactions (electrondonor and electron-acceptor) (Gannon *et al.*, 1991; Vernhet and Bellon, 1995).

Relevant works have shown that the interaction between microorganisms and the abiotic surfaces is of paramount importance in environmental systems (Grasso *et al.*, 2002). In microbially mediated *in situ* pollutant degradation, these interactions are the key event that determines the microorganisms' migration in geological formation and consequently the efficacy of the pollutant removal (Grasso *et al.*, 1996; Grasso and Smets, 1998).

Efficiency of *in situ* bioremediation of contaminated aquifers is often dependent on the ability of the effective transport of microorganisms to the contaminant site, where the propensity of cells adhesion to porous media surfaces is one of the affecting phenomena.

Physicochemical interactions between these two components have been reported to govern the adhesion phenomenon (Smets *et al.*, 1999).

Biofilm systems are relatively recent; the most prominent question that researchers face is the determination of a stable state of the formed biofilm. For this reason many researchers' works have employed theoretical frameworks to describe and predict this stability and biofilm formation ability. Several theoretical models have been employed to predict microbialsurface interactions (Bos et al., 1999): (i) the thermodynamic approach considering the balance of interfacial free energies between the interacting surfaces (Güleç et al., 2006); (ii) the Derjaguin–Landau–Verwey and Overbeek (DLVO) theory was used to evaluate the total interaction energy between a spherical colloid (microbial cells) and the solid surface taking into account electrostatic repulsion and the van der Waals force (Bos et al., 1999); and (iii) the extended DLVO theory (XDLVO), as the acid-base (AB) interactions have been shown to be a major contributor in adhesion processes this theory has been developed by the addition of AB interactions, including both the electron-donating and electron-accepting energies.

Classic DLVO model has been extensively reported, however, it was found unable to fully describe biotic and abiotic colloidal behavior in aqueous media (Grasso *et al.*, 2002). The XDLVO theory was the most substantial theory to explain the experimental findings of microbial adhesion (Elabed *et al.*, 2011, 2013; Sadiki *et al.*, 2015). In addition, this predictive tool takes into account the distance between cell and substratum surfaces (Bos *et al.*, 1999). Currently, these models are employed to describe the attachment of

microbial cells to surfaces in various environments. The application of these theoretical tools with experimental studies may give valuable insights into the adhesion step and the stability of the biofilm-mediated system (Van wey *et al.*, 2011).

The prediction of microbial adhesion, using XDLVO, on different materials such as stainless steel (Nguyen *et al.*, 2011), glass (Azeredo *et al.*, 1999), and wood surfaces (Elabed *et al.*, 2013) was previously documented. To our knowledge, yeast adhesion on wood surface has not been investigated.

A scrutiny of literature indicates a paucity of theoretical predictions based on XDLVO theory, aiming to choose the most suitable support for biofilm formation to be used in wastewater treatment process. This study is of extreme importance for the optimization of microbial adhesion, biofilm stability, and consequently wastewater treatment conditions.

Thus, this work is aimed to study theoretical and experimental ability of yeast cells to adhere to wood surface for the biofilm-mediated system using XDLVO theory. These yeast strains were reported to have a high chromium removal potential (Bahafid *et al.*, 2013). This must have significance in the development of strategies to improve their adherence to this type of substratum to create an efficient biofilm with high performance in wastewater treatment processes.

#### **Materials and Methods**

#### Yeast strains and growing conditions

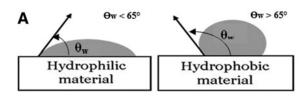
Five yeast strains *Cyberlindnera fabianii*, *Wickerhamomyces anomalus*, *Candida tropicalis*, *Pichia fermentans*, and *Galactomyces geotrichum* were used in this study. These strains were isolated from soil and wastewater samples heavily contaminated with chemical industrial effluents in Fez and were selected on the basis of their chromium resistance as reported by Bahafid *et al.* (2011). All studied yeast strains showed a high ability of removing chromium and were considered as excellent biosorbents. *C. fabianii*, *W. anomalus*, and *C. tropicalis* strains were reported to have a high chromium biosorption capacity (Bahafid *et al.*, 2013). It is also the case for *P. fermentans* and *G. geotrichum* (not published data).

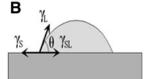
Yeast strains were seeded on yeast medium agar (1% peptone, 1% yeast extract, 2% glucose, and 1.5% agar) plates and incubated for 48 h at 30°C.

#### Cells preparation

Preparation of yeast strains suspension for cell surface contact angle measurements (CAM) was carried out following the protocol of Mohd-Al-Faisal *et al.* (2013) with slight modifications: 2 g of yeast strains was inoculated in yeast medium (YPG) (1% peptone, 1% yeast extract, and 2% glucose) and incubated at 30°C for 48 h where the log phase was attained.

Then, cells were harvested by centrifugation at  $8,000 \times g$ . Cell pellets were washed twice with KNO<sub>3</sub> (0.1 M) and resuspended in the same buffer. At 550 nm, the cell density was adjusted at an absorbance of 0.450 that is equivalent to  $1 \times 10^7$  cells/mL (Mohd-Al-Faisal *et al.*, 2013). Microscopic examination at log phase allowed the visualization of cells presenting characteristics of yeast.





**FIG. 1.** Schematic representation of contact angles measurement; **(A)** according to Vogler's approach; **(B)** according to Van Oss.

#### Contact angle measurements

Hydrophobicity. To prepare microbial lawns suitable for CAM, microbial cells suspended in KNO<sub>3</sub> (0.1 M) sterile solution were deposited on a cellulose acetate membrane filter (0.45  $\mu$ m) by filtration of the suspension using negative pressure. Filters containing microorganisms (yeast cells  $\sim 1 \times 10^7$  cell/mm<sup>2</sup>) were placed to air dry for 30–60 min to obtain stable lawns for contact angles measurements. Contact angles were measured in triplicate with separately cultured microbes (Elabed *et al.*, 2011).

According to Vogler's approach (Vogler, 1998), the value of water contact angle  $\theta_{\rm W}$  permits to evaluate the hydrophobicity of a surface qualitatively, a value higher than 65° indicate a hydrophobic surface, conversely a value lower than 65° allows classifying a surface as being hydrophilic qualitatively (Fig. 1A).

By contrast with this approach, Van Oss'approach appears to be more precise, it allows the determination of the absolute degree of hydrophobicity of a surface, which is calculated using formula Equation (1) (Van Oss *et al.*, 1986).

In this approach, the degree of hydrophobicity of a given material (i) is expressed as the free energy of interaction between two identical surfaces immersed in water ( $\Delta G$ iwi). If the interaction between two surfaces is stronger than the interaction between each surface with water ( $\Delta G$ iwi <0), the surface is considered hydrophobic and conversely ( $\Delta G$ iwi >0), the surface is considered hydrophilic.

$$\Delta Giwi = -2\gamma_{iw} = -2\left[\left(\left(\gamma_i^{LW}\right)^{1/2} - \left(\gamma_W^{LW}\right)^{1/2}\right)^2 + 2\left(\left(\gamma_i^+ \gamma_i^-\right)^{1/2} + \left(\gamma_w^+ \gamma_w^-\right)^{1/2} - \left(\gamma_i^+ \gamma_w^+\right)^{1/2}\right) - \left(\gamma_i^+ \gamma_w^+\right)^{1/2}\right], \quad ($$

where  ${\gamma_i}^{LW}$  is Lifshitz-van der Waals component,  ${\gamma_w}^{LW}$  is Lifshitz-van der Waals component of water,  ${\gamma}i^+$  electron acceptor of a given material (i),  ${\gamma}i^-$  electron donor of a given material (i),  ${\gamma}_w^+$  electron acceptor of water, and  ${\gamma}_w^-$  electron donor of water.

Surface tension components. Once the contact angles were measured, the Lifshitz-van der Waals  $(\gamma^{LW})$  and acidbase  $(\gamma^{AB})$  surface tension components were obtained by the three equation system from the application of the Young-Dupré equation to each probe liquid (Van Oss, 1996). By using three different liquids with known surface parameter values  $\gamma_{l}^{LW}$ ,  $\gamma_{l}^{+}$ , and  $\gamma_{l}^{-}$ , for example, water, formamide, and diiodomethane. The unknown surface tension components of a solid surface  $(\gamma_{s}^{LW}, \gamma_{s}^{+},$  and  $\gamma_{s}^{-})$  or microbial surface  $(\gamma_{s}^{LW}, \gamma_{s}^{+},$  and  $\gamma_{s}^{-})$  or microbial surface  $(\gamma_{s}^{LW}, \gamma_{s}^{+},$  and  $\gamma_{s}^{-})$  can be estimated.

$$\gamma_L(\cos\theta + 1) = 2\left[ (\gamma_s^{LW} \gamma_L^{LW})^{1/2} + (\gamma_s^+ \gamma_L^-)^{1/2} + (\gamma_s^- \gamma_L^+)^{1/2} \right], \tag{2}$$

where  $\theta$  is the measured contact angle and the subscripts (S) and (L) are solid surface and liquid phases, respectively.  $\gamma^{LW}$  is the Lifshitz-van der Waals component of the surface free energy,  $\gamma^+$  and  $\gamma^-$  are the electron acceptor and electron donor parameters, respectively, of the Lewis acid-base component ( $\gamma^{AB}$ ). The surface free energy is expressed as  $\gamma_S = \gamma_S^{LW} + \gamma_S^{AB}$  where  $\gamma_S^{AB} = 2 \left(\gamma_S^- \gamma_S^+\right)^{1/2}$  the acid-base free energy component (Fig. 1B).

Wood species and surface characteristics. Surface free energy data concerning nine wood species (Alder, Hazel, Maple, beech, ash, oak, cedar, pine, and teak) were obtained from the literature. Table 1 presents the water, formamide,

Table 1. Contact Angle Values Using Water  $\theta_{\rm w}$ , Formamide  $\theta_{\rm F}$ , and Diiodométhane  $\theta_{\rm D}$ , Lifshitz-van der Waals  $(\gamma^{\rm LW})$ , Electron-Donor  $(\gamma^-)$  and Electron-Acceptor  $(\gamma^+)$  Parameters and Surface Energies ( $\Delta G$ iwi) of Wood Species

Wood species	Contact angles (°)			Surface tension: components and parameters (mJ/m²)			Surface energies			
	$\theta_w$	$\theta_F = \theta_D = \gamma^{LW} = \gamma^+$		γ_	$\Delta Giwi \ (mJ/m^2)$	References				
Cedar	82.5	61.8	44.5	37.3	0	5.5	-58.82	Elabed et al. (2011)		
Beech	54.5	16.2	13.1	49.5	9.1	1.5	-42.4	Gérardin P et al. (2007)		
Pine	55.4	27.4	27.1	47.6	1.2	24.2	-12.0	Gérardin et al. (2007)		
Ash	68	nd	nd	45.2	2	6.1	-37.8	Mohammed-Ziegler et al. (2004)		
Oak	81	nd	nd	42.9	3.5	0.3	-64.5	Mohammed-Ziegler et al. (2004)		
Teak	18	24	34	42.6	0.2	59.3	-41.9	Mohammed-Ziegler et al. (2004)		
Alder	70	28	27	45.3	2.43	3.75	32.09	Mohammed-Ziegler et al. (2004)		
Hazel	62	32	19	47.9	0.75	11.38	-15.48	Mohammed-Ziegler et al. (2004)		
Maple	75	39	36	41.47	1.97	3.10	31.18	Mohammed-Ziegler et al. (2004)		

and diiodomethane contact angles on wood species. Except beech wood ( $\gamma^-=1.5 \text{ mJ/m}^2$ ,  $\gamma^+=9.1 \text{ mJ/m}^2$ ) and oak wood ( $\gamma^-=0.3 \text{ mJ/m}^2$ ,  $\gamma^+=3.5 \text{ mJ/m}^2$ ), all wood species are predominantly electron donor (high value of  $\gamma^-$ ) ranging from 3.10 to 59.3 mJ/m<sup>2</sup> and presenting weak electron acceptors ranging from 0 to 2.43 mJ/m<sup>2</sup>.

#### Calculation of free energy adhesion of the yeast strains to wood species by extended DLVO theory

Classical DLVO theory (Hermansson, 1999) was the first successful effort to quantitatively describe colloidal stability interactions (Sharma and Hanumantha Rao, 2003).

In this theory, the interaction energy ( $\Delta G^{DLVO}$ ) requires to bring a microorganism (m) into contact with a flat substratum surface (s) immersed in aqueous medium (l).

The net interaction energy was developed by balancing repulsive or attractive electrostatic energy  $(G^{EL})$  and Lifshitz-van der Waals attractive forces  $(G^{LW})$ .

The total interaction or adhesion energy as a function of the separation distance (d) between a bacterium (sphere) and a substratum (flat plane) surface; it can be expressed as follows:

$$\Delta G^{DLVO}\left(d\right) = \Delta G^{LW}(d) + \Delta G^{EL}(d) \tag{3}$$

An extension of the DLVO theory (XDLVO) was suggested (Van Oss, 1996), in which the acid–base interaction ( $\Delta G_{AB}$ ) was added to formulate the extended DLVO theory. The total interaction energy in this approach between microbial cell (m) and substratum (s) through water (w) is described as a balance between attractive Lifshitz-van der Waals force, repulsive or attractive electrostatic force, and acid–base interaction forces. It can be written as follows:

$$\Delta G^{XDLVO}(d) = \Delta G^{LW}(d) + \Delta G^{EL}(d) + \Delta G^{AB}(d), \quad (4)$$

where

$$\begin{split} \Delta G^{\text{LW}} &= \left( \left( \gamma_M^{LW} \right)^{\frac{1}{2}} - \left( \gamma_S^{LW} \right)^{\frac{1}{2}} \right)^2 - \left( \left( \gamma_M^{LW} \right)^{\frac{1}{2}} - \left( \gamma_L^{LW} \right)^{\frac{1}{2}} \right)^2 \\ &- \left( \left( \gamma_S^{LW} \right)^{\frac{1}{2}} - \left( \gamma_L^{LW} \right)^{\frac{1}{2}} \right)^2 \;, \end{split} \tag{4a}$$

and

$$\begin{split} \Delta G^{\text{AB}} &= 2 \left[ \left( \gamma_L^+ \right)^{\frac{1}{2}} \left( \left( \gamma_M^- \right)^{\frac{1}{2}} + \left( \gamma_S^- \right)^{\frac{1}{2}} - \left( \gamma_L^- \right)^{\frac{1}{2}} \right) \right. \\ &+ \left. \left( \gamma_L^- \right)^{\frac{1}{2}} \left( \left( \left( \gamma_M^+ \right)^{\frac{1}{2}} + \left( \gamma_S^+ \right)^{\frac{1}{2}} - \left( \gamma_L^+ \right)^{\frac{1}{2}} \right) \right. \\ &\left. - \left( \gamma_L^- \gamma_S^+ \right)^{\frac{1}{2}} \right) - \left( \gamma_L^+ \gamma_S^- \right)^{\frac{1}{2}} \right] \end{split} \tag{4b}$$

XDLVO predictions of the free energy of adhesion between yeast strains and wood species were investigated. The total free energy of interaction defined by the extended DLVO theory is the sum of the Lifshitz-van der Waals, electrostatic and acid–basic interactions as calculated from Equation (4). As reported by Gallardo-moreno *et al.* (2002) and Rijnaarts *et al.* (1999), the employment of a suspending liquid with high ionic strength (KNO<sub>3</sub> 0.1 M) allows the negligence of electrostatic interaction free energy  $\Delta G^{\rm EL}$ .

#### Adhesion experiments

Adhesion experiments were realized following the protocol described by Elabed *et al.* (2013) with slight modifications. Ten cubic millimeters of yeast cells suspension containing 10<sup>7</sup>–10<sup>8</sup> CFU/mL was incubated, for 48 h at 30°C, in a Petri dish containing wood species coupons singly. The wood coupons were then rinsed thrice with sterile distilled water to eliminate nonadherent cells and moved in a small Petri dish for scanning electron microscopy analysis.

#### Environmental scanning electron microscopy analysis

After the contact period, wood coupons were imaged by using environmental scanning electron microscopy (ESEM Quanta 200). All SEM images were then processed with Matlab® program to determine the adhesion percentage of microbial cells (Hamadi *et al.*, 2005). Matlab software quantifies the experimental adhesion by calculating the ratio of the surface covered by microbial cells and the wood total surface. It is a useful software widely used for this adhesion quantification purpose (Elabed *et al.*, 2012; Sadiki *et al.*, 2015).

#### **Results and Discussion**

#### Cell surface characteristics

Cell surface hydrophobicity. There are different ways to obtain information about cell surface properties. The most commonly used methods are the microbial adhesion to hydrocarbon (MATH) method (Zhang *et al.*, 2010) and CAM (Elabed *et al.*, 2011). In this work, physicochemical characterization of the surfaces of five yeast strains was carried out by the measurement of contact angles of test liquids, using diiodométhane ( $\theta_D$ ), formamide ( $\theta_F$ ), and water ( $\theta_W$ ). Contact angle values  $\theta_W$ ,  $\theta_F$ ,  $\theta_D$ , and the free energy of interaction between two surfaces immersed in water are listed in Table 2.

The results of hydrophobicity obtained are expressed qualitatively  $(\theta_w)$  and quantitatively (in terms of  $\Delta G$ iwi) (Table 2). Among the five yeast strains tested, four have positive values of the free energy of interaction  $\Delta G$ iwi and thus classified as hydrophilic strains. *C. fabianii* is the only yeast strain showing a negative value of free energy of interaction, indicating that this strain is hydrophobic.

These findings agree with previous results of Mercier-Bonin *et al.* (2004) which describe hydrophilic behavior of *Saccharomyces cerevisiae* cells. It is also in agreement with results reported for other genera (Amaral and Lehocky, 2006).

Many studies have aimed to understand microbial cell surface properties. Correlation between yeast surface hydrophobicity and surface protein concentration have been previously suggested (Smit *et al.*, 1992; Suzzi *et al.*, 1994; Vichi *et al.*, 2010), while no correlation was found with hydrocarbon compounds or polysaccharide concentrations (Dengis and Rouxhet, 1997).

It is noteworthy that the interest in yeasts cell surface hydrophobicity has been mainly focused on *Candida* species (Gallardo-moreno *et al.*, 2004) and to a minor degree on *S. cerevisiae* (Vichi *et al.*, 2010) due to their importance in pathology and biotechnology respectively. However, there is extreme evidence that cell surface hydrophobicity plays an important role in the environmental bioremediation applications.

Table 2. Contact Angle Values Using Water  $\theta_{\rm w}$ , Formamide  $\theta_{\rm F}$ , and Diiodométhane  $\theta_{\rm D}$ , Lifshitz-van der Waals  $(\gamma^{\rm LW})$ , Electron-Donor  $(\gamma^-)$  and Electron-Acceptor  $(\gamma^+)$  Parameters and Surface Energies ( $\Delta G$ iwi) of Yeast Cells

	Contact angles (°)			S	Surface energies			
Yeast strains	$\theta_w$	$\theta_F$	$\theta_D$	$\gamma^{LW}$	$\gamma^+$	γ_	$\gamma^{AB}$	∆Giwi
Pichia fermentans Galactomyces geotrichum				$29.38 \pm 0.36$ $21.68 \pm 0.17$			$-6.89 \pm 2.77$ $-10.88 \pm 0.55$	12.63 ± 5.63 21.76 ± 1.11
Cyberlindnera fabianii	$51.0 \pm 1.21$	$35.6 \pm 1.90$	$81.1 \pm 1.38$	$16.87 \pm 0.69$	$9.85 \pm 0.75$	$23.51 \pm 1.30$	$0.75 \pm 0.45$	$-2.33 \pm 1.06$
Candida tropicalis Wickerhamomyces anomalus	$48.4 \pm 0.74$ $33.5 \pm 0.84$	$50.4 \pm 2.14$ $49.3 \pm 1.16$	$74.2 \pm 0.43$ $82.5 \pm 1.03$	$20.48 \pm 0.24$ $16.18 \pm 0.51$	$2.49 \pm 0.15$ $3.19 \pm 0.34$	$39.51 \pm 2.85$ $60.14 \pm 1.30$	$-8.64 \pm 2.02$ $-17.67 \pm 1.07$	$17.25 \pm 4.05$ $34.58 \pm 2.24$

Surface tension components. The Lifshitz-van der Waals  $(\gamma^{LW})$  component, electron-donor  $(\gamma^-)$  and electron-acceptor  $(\gamma^+)$  parameters are presented in Table 2. Surfaces of all yeast strain appear to behave predominantly as electron donors/ Lewis bases with high values of  $\gamma^-$  ranging from  $23.51\pm1.30$  to  $60.14\pm1.30$  mJ/m². The results show also that all strains are weak electron acceptors ranging from  $0.55\pm0.18$  to  $9.58\pm0.75$  mJ/m². Our results are in agreements with a previous study presented by Van Der Mei *et al.* (1998) showing that microbial cell surfaces are electron-donating, while sometimes rare electron-accepting cell surfaces can be found (Van Der Mei *et al.*, 1998). This was also in agreement with results reported by Mercier-Bonin *et al.* (2004) and Vichi *et al.* (2010) showing that *S. cerevisiae* cells are exhibiting a Lewis basic character.

The nature of chemical groups on the surface of microbial strains may explain the predominance of their electron-donor character. It was reported that this character can be attributed to the presence of the chemical groups negatively charged or neutral exposed on the surface. These groups are essentially carboxylate groups (COO<sup>-</sup>), amino groups (NH<sub>2</sub>), phosphate (PO<sub>4</sub>) groups from phospholipids, lipoproteins, and lipopolysaccharides, and (-SO<sub>3</sub>) groups from sulfur clusters (Briandet et al., 1999; Hamadi et al., 2004; Hamadi et al., 2008). For yeast strains, it was reported that yeast cells exhibit negatively charged surfaces due to the presence of phosphate groups in their cell wall (Vichi et al., 2010) as suggested earlier (Amory and Rouxet (1988). Previous Fourier transform infrared spectroscopy (FTIR) analysis of W. anomalus confirms the presence of phosphate groups on its cell surface (Bahafid et al., 2011).

#### Prediction of adhesion of the yeast strains to wood species by XDLVO theory

Very few studies on bacterial and fungal adhesion phenomena on wooden substratum have been reported (Elabed et al., 2012). To date, there is no information on yeast strains attachment on this type of substratum. Theoretical physicochemical models can successfully explain biofilm formation phenomena, since the main factors affecting microbial adhesion are physicochemical properties of microbial cell and support surfaces. Indeed, many works correlating the experimental adhesion of microbial cells visualized by ESEM with surface physicochemical characteristics were reported (Sadiki et al., 2015).

Application of these models to predict microbial adhesion to different materials has been reported. Nevertheless, very few works have proposed to employ XDLVO approach to study the interaction between the two components of the biofilm, to develop a new bioprocess for the treatment of wastewater.

XDLVO predictions of the free energy of adhesion between yeast strains and wood species are presented in Table 3. The positive values of the  $\Delta G^{\rm XDLVO}$  indicate unfavorable adhesion or repulsion. On the opposite, attraction between two interacting surfaces occurs when the total energy values  $\Delta G^{\rm XDLVO}$  are negative. In the present case, XDLVO predictions show that the adhesion is unfavorable between pine, teak, and all yeast strains tested with positive values of total free energy of adhesion. For pine wood,  $\Delta G^{\rm XDLVO}$  values ranged from 0.42 to 23.43 mJ/m² and for

Table 3. Total Interaction Free Energy ( $\Delta G^{\text{XDLVO}}$  (mJ/m<sup>2</sup>)) of Yeast Adhesion on Wood Species

			Inte	raction free	energy ∆G	XDLVO (mJ/i	$m^2$ )		
	Wood species								
Yeast strains	Alder	Hazel	Maple	Beech	Pine	Ash	Oak	Teak	Cedar
P. fermentans G. geotrichum C. fabianii C. tropicalis W. anomalus	-24.35 -5.24 -10.99 -12.40 1.23	-11.15 5.76 -5.56 -0.65 14.61	-25.18 -5.70 -12.05 -13.32 0.55	-33.27 -14.81 -12.78 -20.85 -10.91	1.83 13.97 0.42 9.51 23.43	-19.52 -1.58 -9.02 -8.34 5.49	-36.55 -14.55 -16.38 -22.86 -9.76	27.39 32.67 10.37 30.32 44.61	-17.41 2.92 -10.76 -5.89 11.53

100 90 80 70 Adhesion percentage (%) 60 50 40 30 20 10 Cedar Beech Pine Ash Oak Alder Hazel Maple Teak Wood species

**FIG. 2.** Potentiality of theoretical adhesion of yeast strains on wood species.

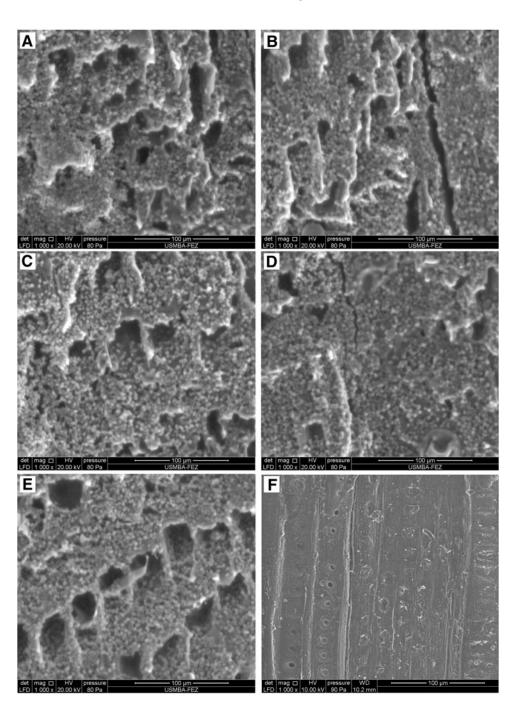


FIG. 3. Images of yeast strain cells adhered onto oak wood coupons, visualized by environmental scanning electron microscopy, after 48 h of contact period at 30°C. (A) Pichia fermentans, (B) Galactomyces geotrichum, (C) Cyberlindenera fibianii, (D) Candida tropicalis, (E) Wickerhamomyces anomalus, and (F) control wood.

#### ADHESION OF YEAST WITH HIGH CHROMIUM REMOVAL

teak, values of  $\Delta G^{\rm XDLVO}$  ranged from 10.37 to 44.61 mJ/m². These results show that pine and teak wood should not be used as a support for biofilm formation for all yeast strains tested. On the opposite, alder, hazel, maple, oak, beech, ash, and cedar wood show favorable adhesion with different percentage of potentiality of adhesion. Percentage of adhesion of alder, hazel, maple, beech, ash, oak, and cedar wood are 80%, 60%, 80%, 100%, 80%, 100%, and 60% respectively (Fig. 2). Results show that beech and oak exhibit better properties than the other wood species studied. These wood species may be the most suitable to be used in a wastewater treatment process as a support for biofilm formation of the tested yeast strains. The combination beech woodyeast strains exhibit negative  $\Delta G^{\rm XDLVO}$  values, ranged from

–10.91 to –33.27 mJ/m². For oak wood, negative  $\Delta G^{\rm XDLVO}$  values ranged from –9.76 to –36.55 mJ/m².

## Experimental adhesion of yeast species on wood species

To verify the applicability of XDLVO theory as a predictive physicochemical approach to study yeast cells adhesion to wood surface, ESEM was used to visualize the experiment adhesion after 48 h of contact period.

It is noteworthy that wood surface roughness parameter has not been fixed in this investigation. Indeed, the developing process is involving wood husk as biofilm support, which may present a wide variation in roughness.

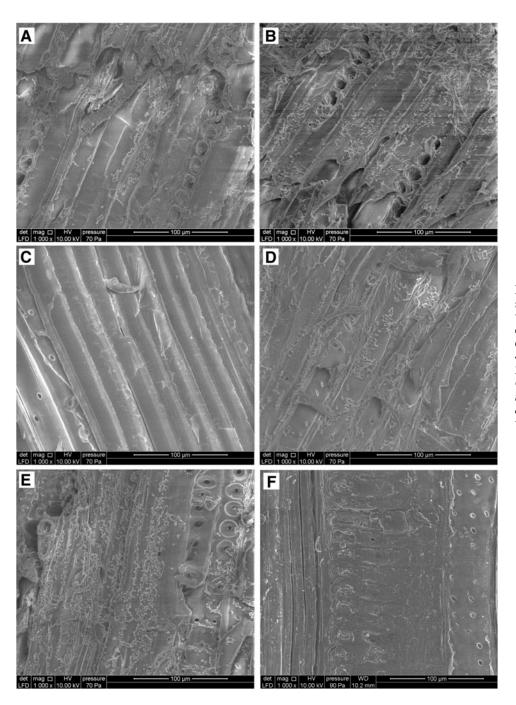


FIG. 4. Images of yeast strain cells adhered onto pine wood coupons, visualized by environmental scanning electron microscopy, after 48 h of contact period at 30°C. (A) P. fermentans, (B) G. geotrichum, (C) C. fibianii, (D) C. tropicalis, (E) W. anomalus, and (F) control wood.

Table 4. Percentage of Experimental ADHESION BY YEAST CELLS TO OAKAND PINE WOOD BASED ON IMAGE ANALYSIS BY MATLAB

Percentage of adhesion			
Oak	Pine		
12.81	85.35		
22.58	62.65		
3.42	81.84		
21.45	79.89		
32.06	65.7		
	Oak  12.81 22.58 3.42 21.45		

Furthermore, it was reported that roughness appears to be a minor factor influencing initial biofilm adhesion (Boulangé-Petermann et al., 1997; Bos et al., 1999).

Environmental scanning electron microscopy images show the experimental adhesion behavior of the yeast cells on oak wood (Fig. 3) presenting attractive interactions with all tested strains according to the XDLVO theory predictions by contrast to pine wood (Fig. 4), which was predicted to be repulsive for all tested yeast strains as indicated by positive values of  $\Delta G^{\rm XDLVO}$  (Table 3).

ESEM observations reveal the porous nature of wood substrata, with depressions and grooves on the surface conferring to wood its microbial attachment property (Zainul Akmar et al., 2007) and a large number of binding sites for chromium ions fixation (Figs. 3 and 4).

Obtained scanning electron microscopy (SEM) images show the yeast cells immobilized on the oak wood surfaces within a compact microbial layer where the adhering cells were distributed relatively uniformly across the entire wood surface (Fig. 3). Figure 3A and C shows that the adhering cells were deposited in aggregate forms within clumps. Microbial clumps are characteristic of hydrophobic substrata (Van Pelt et al., 1985).

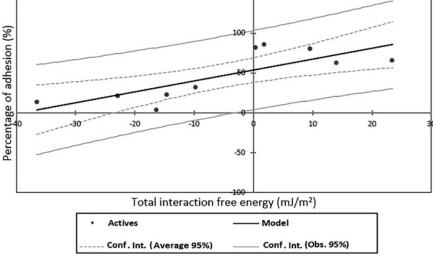
Indeed, Matlab software showed high percentage values of adhesion ranging from 62.65% to 85.35%. For ESEM observation on pine wood, yeast strains were observed to be randomly dispersed on wood surface attached as single cells or pairs of cells (Fig. 4). Even though W. anomalus represented a relative important adhesion percentage compared to the other yeast strains tested, it can be seen that the microbial layer does not completely cover the wood surface (Fig. 4E).

Results of yeast experimental adhesion to pine surface according to the image analysis by the Matlab software program indicates low adhesion percentages ranging from 3.42% to 22.58% (Table 4). An exception was obtained with W. anomalus (Fig. 4E), which represents a relatively important percentage of experimental adhesion (32.02%) (Table 4) according to Matlab software analysis, despite the unfavorable theoretical adhesion obtained by XDLVO prediction tool.

In this investigation, a good relation between theoretical prediction using XDLVO theory and experimental yeast cells adhesion on wood surfaces was obtained (Fig. 5). Thus, a high degree of agreement was noted between theoretical and experimental results (r=0.7901). Indeed, the experimental assay proves that all tested strains are able to adhere to oak wood with  $\Delta G^{XDLVO}$  < 0 and except *W. anomalus*, all tested strains are not able to adhere to pine wood ( $\Delta G^{XDLVO} > 0$ ). Our findings corroborate with previous results (Hamadi et al., 2009; Elabed et al., 2013; Sadiki et al., 2015), which reported a good correlation between XDLVO results and experimental adhesion.

In many works physicochemical models has succeeded to explain microbial adhesion visualized using ESEM results (Elabed et al., 2013; Sadiki et al., 2015). In fact, XDLVO approach proved to be more effective than other predictive tools (Sharma and Hanumantha Rao, 2003). However, microbial adhesion to surfaces is a very complex phenomenon due to the complex and heterogeneous surface composition, which may in some cases limit the applicability of surface theoretical predictive tools. Furthermore, it was reported that discrepancies between the XDLVO theoretical predictions and the experimental adhesion results may be attributed to additional interactions, non-DLVO forces, or parameters such as surface roughness, the complex chemical and

FIG. 5. Relation between total interaction free energy and adhesion percentage of yeast strains.



Percentage of adhesion (%) = 53,69404+1,35827\*x; 0.95 Conf.Int. r2=0,6244; r=0,7901

morphologic structure of surfaces (Brant and Childress, 2002; Chin et al., 2002; Sharma and Hanumantha Rao, 2003).

#### Conclusion

The potential adhesion of five yeast strains on nine wood species was investigated employing extended DLVO approach. The results revealed that microbial adhesion on wooden substrata was essentially dependent on two factors, wood species and tested microorganisms. The results presented here demonstrate that the use of Matlab program to analyze the images obtained by SEM allows confirming the theoretical prediction of adhesion obtained using XDLVO tool. Results suggest that pine and teak wood are not suitable supports for tested yeast strains presenting positive values of  $\Delta G^{XDLVO}$  toward all tested strains and 0% of adhesion percentage toward all studied yeast strains. Beech and oak wood have better proprieties to be used as a support for tested yeast strains with 100% of adhesion percentage and negative  $\Delta G^{\mathrm{XDLVO}}$  values. These combinations could form an efficient biofilm for chromium removal. These findings have important potential in the stability of the biofilm during its functional lifetime and its applicability in a wastewater treatment process.

#### Acknowledgments

The authors thankfully acknowledge the financial and scientific support of the Microbial Biotechnology Laboratory, Faculty of Sciences and Technology, SMBA University, Fez, Morocco.

#### **Author Disclosure Statement**

No competing financial interests exist.

#### References

- Amaral, P.F.F., and Lehocky, M. (2006). Cell surface characterization of *Yarrowia lipolytica*. Wiley Intersci. 23, 867.
- Amory, E.D., and Rouxet, .PG. (1988). Flocculence of brewery yeasts and their surface properties: Chemical composition, electrostatic charge and hydrophobicity. *J. Inst. Brew.* 94, 79.
- Azeredo, J., Visser, J., and Oliveira, R. (1999). Exopolymers in bacterial adhesion: Interpretation in terms of DLVO and XDLVO theories. *Colloid Surf. B.* 14, 141.
- Bahafid, W., Sayel, H., Tahri Joutey, N., and El Ghachtouli, N. (2011). Removal mechanism of hexavalent chromium by a novel strain of Pichia anomala isolated from industrial effluents of Fez (Morocco). *J. Environ. Sci. Eng.* 5, 980.
- Bahafid, W., Tahri Joutey, N., Sayel, H., Iraqui-Houssaini, M., and El Ghachtouli, N. (2013). Chromium adsorption by three yeast strains isolated from sediments in Morocco. *Geomicrobial. J.* 30, 422.
- Bos, R., Mei, H.C., and Busscher, H.J. (1999). Physicochemistry of initial microbial adhesive interactions-its mechanisms and methods for study. FEMS Microbiol. Rev. 23, 179.
- Boulangé-Petermann, L., Rault, J., and Bellon-Fontaine, M.N. (1997). Adhesion of *Streptococcus thermophiles* to stainless steel with different surface topography and roughness. *Biofouling*. 11, 201.
- Brant, J.A., and Childress, A.E. (2002). Membrane-colloid interactions: Comparison of extended DLVO predictions with AFM force measurements. *Environ. Eng. Sci.* 19, 413.

- Briandet, R., Meylheuc, T., Maher, C., and Bellon-fontaine, M.N. (1999). *Listeria monocytogenes* Scott A: Cell surface charge, hydrophobicity, electron donor and acceptor characteristics under different environmental growth conditions. *Appl. Envion. Microbiol.* 65, 5328.
- Chin, C.J., Yiacoumi, S., and Tsouris, C. (2002). Influence of metal ion sorption on colloidal surface forces measured by atomic force microscopy. *Environ. Sci. Technol.* 36, 343.
- Das, N., Basak, L.V.G., Salam, JA., and Abigail, E.A.M. (2013). Application of biofilms on remediation of pollutants—An overview. J. Microbial. Biotech. Res. 2, 783.
- Dengis, P.B., and Rouxhet, P.G. (1997). Surface properties of top- and bottom-fermenting yeast. *Yeast.* 13, 931.
- Elabed, S., Moustakim, M., Berguadi, F., Latrache, H., Houari, A., Hamadi, F., and Ibnsouda Koraichi, S. (2011). Study of microbial adhesion on some wood species: Theoretical prediction. *Microbiology*. 8, 43.
- Elabed, S., Ibnsouda Koraichi, S., Latrache, H., Meftah, H., Tahri Joutey, N., and Hamadi, F. (2012). Environmental Scanning Electron Microscopy characterization of the adhesion of conidia from *Penicillium expansum* to cedar wood substrata at different pH values. *World J. Microb. Biotechnol.* 28, 1707.
- Elabed, S., Ibnsouda Koraichi, S., Houari, A., and Latrache, H. (2013). Experimental and theoretical investigations of the adhesion time of *Penicillium* spores to cedar wood surface. *Mater. Sci. Eng. C.* 33, 1276.
- Gallardo-moreno, A.M., Gonzalez-Martin, M.L., Pérez-giraldo, C., Bruque, J.M., and Gomez-Garcia, A.C. (2002). Serum as a factor influencing adhesion of *Enterococcus faecalis* to glass and silicone. *Appl. Environ. Microbiol.* 68, 5784.
- Gallardo-moreno, A.M., González-Martín, M.L., Pérez-Giraldo, C., Bruque, J.M., and Gómez-García, A.C. (2004). The measurement temperature: An important factor relating physicochemical and adhesive properties of yeast cells to biomaterials. J. Colloid. Interface Sci. 271, 351.
- Gannon, J.T., Manilal, V.B., and Alexander, M. (1991). Relationship between cell surface properties and transport of bacteria through soil. Appl. Environ. Microbiol. 57, 190.
- Gérardin, P., Petric, M., Petrissans, M., Lambert, J., and Ehrhradt, J.J. (2007). Evolution of wood surface free energy after heat treatment. *Polym. Degrad. Stabil.* 92, 653.
- Grasso, D., Smets, B.F., Strevett, K.A., Machinist, B.D., Van Oss, C.J., Giese, R.F., and Wu, W. (1996). Impact of physiological state on surface thermodynamics and adhesion of *Pseudomonas Aeruginosa*. Environ. Sci. Technol. 30, 3604.
- Grasso, D., and Smets B.F. (1998). Equilibrium modeling of *Pseudomonas* aggregation and partitioning to dolomite. *J. Disper. Sci. Technol.* 19, 1081.
- Grasso, D., Subramaniam, M., Butkus, M., Strevett, K., and Bergengahl, J. (2002). A review of non-DLVO interactions in environmental colloidal systems. *Rev. Environ. Sci. Bio*technol. 1, 17.
- Güleç, H.A., Sarioğlu, K., and Mutlu, M. (2006). Modification of food contacting surfaces by plasma polymerisation technique. Part I: Determination of hydrophilicity, hydrophobicity and surface free energy by contact angle method. *J. Food Eng.* 2, 187.
- Hamadi, F., Latrache, H., El Ghmari, A., and Ellouali, M. (2004). Effect of pH and ionic strength on hydrophobicity and electron donor and acceptor characteristics of *Escherichia coli* and *Staphylococcus aureus*. *Ann. Microbial*. 54, 213.
- Hamadi, F., Latrache, H., Mabrrouki, M., Elghmadi, A., Outzourhit, A., Ellouali, M., and Chtaini, M. (2005). Effect of pH

- on distribution and adhesion of *Staphylococcus aureus* to glass. *J. Adhes. Sci. Technol.* 19, 73.
- Hamadi, F., Latrache, H., Zahir, H., Elghmari, A., Timinouni, M., and Ellouali, M. (2008). The relation between *Escherichia coli* surface functional groups' composition and their physicochemical properties. *Braz. J. Microbiol.* 39, 10.
- Hamadi, F., Latrache, H., Zekraoui, M., Ellouali, M., and Bengourram, J. (2009). Effect of pH on surface energy of glass and Teflon and theoretical prediction of *Staphylococcus* aureus adhesion. *Mater. Sci. Eng. C.* 29, 1302.
- Hermansson, M. (1999). The DLVO theory in microbial adhesion. *Colloid Surf. B.* 14, 105.
- Ksheminska, H., Fedorovych, D., Babyak, L., Yanovych, D., Kaszycki, P., and Koloczek, H. (2005). Chromium (III) and (VI) tolerance and bioaccumulation in yeast: A survey of cellular chromium content in selected strains of representative genera. *Process Biochem.* 40, 1565.
- Mercier-Bonin, M., Ouazzani, K., Schmitz, P., and Lorthois, S. (2004). Study of bioadhesion on a flat plate with a yeast/glass model system. *J. Colloid Interface. Sci.* 271, 342.
- Mohammed-Ziegler, I., Oszlánczi, Á., Somfai, B., Hórvölgyi, Z., Pászli, I., Holmgren A., and Forsling, W. (2004). Surface free energy of natural and surface-modified tropical and European wood species. J. Adhesion Sci. Technol. 18, 687.
- Mohd-Al-Faisal, N., Wan Harun, W.H.A., and Abdul Razak, F. (2013). An in vitro study on the anti-adherence effect of *Brucea javanica* and *Piper betle* extracts towards oral Candida. *Arch. Oral Biol.* 58, 1335.
- Nguyen, V.T., Chia, T.W.R., Turner, M.S., Fegan, N., and Dykes, G.A. (2011). Quantification of acid-base interactions based on contact angle measurement allows XDLVO predictions to attachment of *Campylobacter jejuni* but not *Salmonella*. *J. Microbiol. Methods*. 86, 89.
- Pan, X., Liu, Z., Chen, Z., Cheng, Y., Pan, D., Shao, J., Lin, Z., and Guan, X. (2014). Investigation of Cr(VI) reduction and Cr(III) immobilization mechanism by planctonik cells and biofilm of *Bacillus subtilis* ATCC-6633. *Water Res.* 55, 21.
- Quintelas, C., Fernandes, B., Castro, J., Figueiredo, H., and Tavares, T. (2008). Biosorption of Cr (VI) by a *Bacillus coagulans* biofilm supported on granular activated carbon (GAC). *J. Hazard. Mater.* 153, 799.
- Quintelas, C., Rocha, Z., Silva, B., Fonseca, B., Figueiredo, H., Tavares, T. (2009). Removal of Cd(II), Cr(VI), Fe(III) and Ni(II) from aqueous solutions by an *E.coli* biofilm supported on kaolin. *Chem. Eng. J.* 149, 319.
- Rijnaarts, H.H.M., Norde, W., Lyklema, J., and Zehnder, A.J.B. (1999). DLVO and steric contributions to bacterial deposition in media of different ionic strenghts. *Colloid Surf.* B. 14, 179.
- Sadiki, M., Elabed, S., Barkai, H., Laachari, F., and Ibnsouda Koraichi, S. (2015). The impact of *Thymus vulgaris* extractives on cedar wood surface energy: Theoretical and experimental of *Penicillium* spores adhesion. *Ind. Crop. Prod.* 77, 1020.

- Sharma, P.K., and Hanumantha Rao, K. (2003). Adhesion of Paenibacillus polymyxa of chalcopyrite and pyrite: Surface thermodynamics and extended DLVO theory. Colloid Surface B. 29, 21.
- Silva, B., Figueiredo, H., Quintelas, C., Neves, I.C., and Tavares, T. (2008). Zeolithes as supports for the biorecovery of hexavalent and trivalent chromium. *Microporous Mesoporous Mater*. 116, 555.
- Smets, B.F., Grasso, D., Engwall M.A., and Machinist B.J.(1999). Surface physicochemical properties of *Pseudo-monas Fluorescencs* and impact on adhesion and transport. *Colloid Surface B.* 14, 121.
- Smit, G., Straver, M.H., Lugtenberg, B.J.J., and Kijne, J.W. (1992). Flocculence of *Saccharomyces cerevisiae* cells is induced by nutriment limitation, with cell surface hydrophobicity as a major determinant. *Appl. Environ. Microbiol.* 58, 3709.
- Suzzi, G., Romano, P., and Vannini, L. (1994). Cell surface hydrophobicity and floculence in *Saccharomyces cerevisiae* wine yeasts. *Colloid Surface B.* 2, 505.
- Van Der Mei, H.C., Bos, R., and Busscher, H.J. (1998). A reference guide to microbial cell surface hydrophobicity based of contact angles. *Colloid Surface B.* 11, 213.
- Van Oss, C.J. (1996). *Inerfacial Forces in Aqueous Media*. New York: Dekker.
- Van Oss, C.J., Good, R.J., and Chaudhury, M.K. (1986). The role of Van Der Waals forces and hydrogen bonds in hydrophobic interactions' between biopolymers and low energy surfaces. J. Colloid Interface Sci. 2, 378.
- Van Pelt, A.W.J., Weerkamp, A.H., Uyen, M.H.W.J.C., Busscher, H.J., De Jong, H.P, and Arends, J. (1985). Adhesion of Streptococcus sanguis CH3 to polymers with different surface free energies. *Appl. Environ. Microb.* 5, 1270.
- Van wey, A.S., Cookson, A.L., Roy, N.C., McNabb, W.C., Soboleva, T.K., and Shorten P.R. (2011). Bacterial biofilms associated with food particles in the human large bowel. *Mol. Nutr. Food Res.* 7, 969.
- Vernhet, A., and Bellon, N. (1995). Role of bentonites in the prevention of *Saccharomyces cerevisiae*. *Colloid Surface B*. 3, 255.
- Vichi, S., Gallardo-Chacon, J.J., Pradelles, R., Chassagne, D., Lopez-Tamames, E., and Buxaderas, S. (2010). Surface properties of Saccharomyces cerevisiae lees during sparkling wine ageing and their effect on flocculation. Int. J. Food Microbiol. 140, 125.
- Vogler, E.A. (1998). Structure and reactivity of water at biomaterial surfaces. *Adv. Colloid Interface Sci.* 74, 69.
- Zainul Akmar, Z., Zainoha, Z., Salmijah, S., and Wan Azlina, A. (2007). Biological detoxification on Cr(VI) using woodhusk immobilized *Acinetobacter haemolyticus*. *J. Hazard. Mater.* 148, 164.
- Zhang, C., Jia, L., Wang, S., Qu, J., Li, K., Xu, L., Shi, Y., and Yan, Y. (2010). Biodegradation of beta-cypermethrin by two *Serratia spp.* with different cell surface hydrophobicity. *Bioresour. Technol.* 101, 3423.