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Assessing the Potential of Biochars Prepared by Steam-Assisted Slow Pyrolysis for CO₂ Adsorption and Separation

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Supporting Information

ABSTRACT: The potentialities in the use of biochars prepared by steam-assisted slow pyrolysis as adsorbents of gases of strategic interest (N₂, CO₂, and CH₄) and their mixtures were explored. The biochars prepared from *Populus nigra* wood and cellulose fibers exhibited a narrow microporosity, with average pore sizes ranging between 0.55 and 0.6 nm. The micropore volume increased with the pyrolysis temperature, allowing CO₂ and CH₄ uptakes at room temperature between 1.5 and 2.5 mmol/g and between 0.1 and 0.5 mmol/g, respectively. These values are in line with those from the literature on biomass-derived carbon-based materials, exhibiting much higher porous features than those reported herein. As for the separation of CO₂/N₂ and CO₂/CH₄ gas mixtures, data showed that the prepared biochars exhibited good selectivities for CO₂ over both N₂ and CH₄: between ca. 34 and 119 for a CO₂/N₂ mixture in typical post-combustion conditions (15:85, v/v) and between 14 and 34 for a CO₂/CH₄ mixture typical of natural gas upgrading (30:70, v/v).

INTRODUCTION

A biochar is a carbon-based solid product obtained through thermochemical conversions, such as pyrolysis, gasification, torrefaction, and hydrothermal carbonization of biomass.^{1,2} The number and type of potential feedstocks for biochar production are huge, e.g., wood and agricultural wastes, rice husks and straw, leaves, food waste, paper sludge, bagasse, and many others.^{1,2} Under certain operating conditions, biochar production is an energy self-sufficient process and can produce biofuel for use in various energy conversion processes, including transportation and electricity production.³ This means that agricultural and animal waste disposal can be reduced by recycling the waste, using pyrolysis, into biochar, energy, and value-added products.

Biochars consist of fixed carbon, ash components, moisture, and, on the basis of production conditions, labile carbon and other volatile compounds.^{4–6} These materials have recently attracted much attention as inexpensive, environmentally friendly, carbon-rich materials with potential applications in a variety of fields, such as soil remediation, waste management, greenhouse gas reduction, and energy production.^{4–9} New applications using biochar as material for electronics and bioenergy (fuel cells and supercapacitors), as a catalyst for syngas cleaning and conversion of syngas to liquid hydrocarbon, and as a sorbent for contaminant reduction in gaseous streams are emerging.^{1,10–13} Because there is a limited literature on the applications of the biochar as a gas adsorbent and/or catalyst, research efforts in this field are of great importance.

On the other hand, adsorption on a solid matrix represents a well-assessed method for the capture and separation of CO₂ from exhaust flue gases.^{14,15} An ideal CO₂ adsorbent should (i) exhibit high selectivity toward CO₂ over N₂ and other exhaust

components (CO, NH₃, and light hydrocarbons, such as CH₄), (ii) be produced via inexpensive and low-energy consumption processes, using renewable resources as precursors, (iii) exhibit flexible morphologies, pore structures, and functionalities, and (iv) present good mechanical properties to undergo repeated adsorption–desorption cycles.^{15,16} In this framework, the biochar is gaining a lot of interest because it could represent a valid low-cost substitute of the more expensive activated carbons.

With these premises, the aim of this work was to assess the potential of biochars prepared from steam-assisted slow pyrolysis of biomass feedstocks (*Populus nigra* wood and cellulose fibers) in carbon capture and storage applications. For this, a series of biochars with different characteristics were prepared by steam-assisted slow pyrolysis and tested to adsorb and separate CO₂ from N₂ and CH₄ in a typical scenario of flue gases. Although slow pyrolysis is typically carried out in an inert atmosphere, the presence of oxidant agents, such as steam, has been reported to improve the porosity of the chars as a result of its ability to efficiently penetrate on the precursor during the slow pyrolysis, enhancing devolatilization, and, if the temperature is high enough, to promote gasification reactions.^{17–21} The gas adsorption and selectivity were measured by means of equilibrium adsorption isotherms for pure gases at ambient conditions. In particular, gas adsorption data were used to predict the adsorption of binary mixtures

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and determine CO₂/N₂ and CO₂/CH₄ selectivities by applying the ideal adsorbed solution theory (IAST).

■ EXPERIMENTAL SECTION

Biochar Precursors. *P. nigra* wood and cellulose fibers were used as biomass feedstocks for the preparation of the biochars. *P. nigra* wood was selected as a precursor because it allows for the production of a biochar with a higher specific surface area compared to other lignocellulosic biomasses treated in the same pyrolytic conditions.¹⁷ Moreover, *P. nigra* is among the faster growing trees for short rotation coppice, with an annual dry matter production of 17.8 Mg ha⁻¹ year⁻¹.¹⁷ Cellulose fibers were selected as model feedstock; cellulose is one of the three main biomass components, and the most abundant component in *P. nigra* wood (55.4 wt % cellulose, 11.6 wt % lignin, and 26.8 wt % hemicellulose).¹⁷

Synthesis of the Biochars. Biochars were produced from *P. nigra* wood and cellulose fibers (Sigma-Aldrich C6663) through a steam-assisted slow pyrolysis process as described in previous works.^{17,18} Briefly, the biomass (400–600 μm size range) was pre-dried in a thermoventilated oven at 105 °C for 2 h to remove the residual moisture before the pyrolysis experiments. Then, about 6 g of biomass were spread as thin layers (approximately 1 mm thick) over four trays in the pyrolytic reactor¹⁸ and exposed to steam flow (ca. 0.25 g/s, with an average residence time of 2 s) up to the desired temperature (480–700 °C). The heating rate was fixed to ca. 5 °C/min and pressure to ca. 5 × 10⁵ Pa. Gaseous stream driven by the flux of steam exited the reaction unit and passed through a condensation device, and the permanent gases were sent to the exhaust. Biochar was recovered from the trays at the end of each experimental test, and the yield was determined gravimetrically with respect to the feed sample. The reliability of the experimental apparatus has been evaluated on two replicates of the same experimental run, and an absolute error lower than 5% of the measured values has been observed for the yields of solid. Biochars from cellulose fibers were prepared at three pyrolysis temperatures of 530, 650, and 700 °C, and biochars from *P. nigra* wood were obtained at 480 and 600 °C. As reported in previous works,^{17,18} these temperatures have been selected on the basis of the thermal degradation profiles of the biochar precursors (Figure S1 of the Supporting Information). Moreover, in accordance with the results reported in previous works, to obtain chars with a well-developed porosity, pyrolysis temperatures below 480 °C were discarded as a result of the incomplete thermal degradation of the biomass precursors.^{17,18,22} The nomenclature of the biochars will be CEL-T and PN-T for those obtained from cellulose fibers and *P. nigra* wood precursors, respectively, where T refers to the final temperature of the steam-assisted slow pyrolysis process applied for the production of the sample.

Characterization Methods. The carbon, hydrogen, and nitrogen contents of the samples were measured by a CHN 628 LECO elemental analyzer according to the ASTM E870 procedure using ethylenediaminetetraacetic acid (EDTA) as the standard. For each sample, two replicates were performed and the average values are reported (maximum relative error of around 0.7%). The ash content was evaluated by thermogravimetric analysis in a TGA 701 LECO analyzer. Fourier transform infrared spectroscopy (FTIR) analysis of the biochars was performed on solid sample dispersions prepared by mixing and grinding the powdered materials (0.5–0.8 wt %) with KBr. Pellets of the biochar and KBr mixtures were obtained upon compression at 10 tons for 10 min. FTIR spectra in the 3400–600 cm⁻¹ range were acquired in the transmittance mode using a 5700 Nicolet spectrophotometer. Each spectrum was obtained by collecting 32 interferograms with 0.02 cm⁻¹ resolution. The porosity of the biochars was characterized by gas adsorption experiments. High-resolution equilibrium nitrogen adsorption/desorption isotherms at -196 °C were collected in a volumetric analyzer (Triflex, Micromeritics) provided with three pressure transducers to allow for high resolution in the low pressure range. The samples were previously outgassed under dynamic vacuum (ca. 10⁻⁵ Torr) at 120 °C overnight. The Brunauer–Emmett–Teller (BET) theory was used to

calculate the specific surface areas and total pore volumes. Additionally, the narrow microporosity was assessed by CO₂ equilibrium adsorption/desorption isotherms at 0 °C (Tristar II 3020, Micromeritics). The micropore volumes were evaluated using the Dubinin–Radushkevich (DR) formalism applied to nitrogen gas adsorption data. The narrow microporosity was further evaluated from the CO₂ adsorption/desorption isotherms at 0 °C, using the DR equation and the two dimensional non-local density functional theory (2D-NLDFT) method.²³ Ultrapure gases were used for all of the measurements. The reproducibility of the adsorption data was assessed by measuring the adsorption isotherms on at least two fresh aliquots of selected samples.

Gas Adsorption Tests. Gas adsorption/desorption isotherms of selected gases (i.e., CO₂, CH₄, and N₂) at temperatures near ambient (i.e., 0, 10, and 25 °C) were collected in a volumetric instrument (Tristar II 3020, Micromeritics) in the pressure range of 0.1–900 Torr. Gas adsorption data near room temperature were recorded with over 60 points in the whole range of relative pressures, with readings at least every ca. 1.5 cm³/g to obtain accurate data. The analyzer is equipped with a pressure transducer of 1000 mmHg capacity (accuracy within 0.15% of reading). The outgassing conditions before the adsorption measurements were as indicated above. The temperature of the analysis was maintained using a water circulating bath. The CO₂ isosteric heats of adsorption were calculated using the Clausius–Clapeyron equation with the specific tool of Microactive software (Micromeritics) from the set of equilibrium isotherms acquired at near ambient conditions (0, 10, and 25 °C). The isosteric enthalpies of adsorption were obtained from the slopes of the lines in the plots of ln P versus 1/T for a given amount adsorbed in the isotherms at different temperatures. An average Q_{st} value was calculated over the whole range of CO₂ uptakes. The IAST was used to predict binary mixture adsorption from the experimental pure gas isotherms.²⁴ The integration required by IAST was achieved by fitting the single-component gas isotherms to the Jensen equation.²⁵ Typical conditions of flue gases in post-combustion processes (e.g., CO₂/N₂ mixtures of 15:85, v/v) and natural gas fields (e.g., CO₂/CH₄ mixtures of 30:70, v/v) were used in the binary mixture predictions. Selectivity for gas component *i* over *j* (*S*_{*ij*}) was calculated at 25 °C and 1 atm as *S*_{*ij*} = *C*_{*i*}/*x*_{*i*}/*C*_{*j*}/*x*_{*j*}, where *C*_{*i*} and *C*_{*j*} are the modeled adsorbed amounts of components *i* and *j* in the mixture and *x*_{*i*} and *x*_{*j*} are their molar fractions in the mixture.

■ RESULTS AND DISCUSSION

Biochar Compositional Properties. The chemical composition of the synthesized biochars is listed in Table 1.

Table 1. Chemical Composition of the Biochars (Dry and Ash-Free Basis)

	CEL-530	CEL-650	CEL-700	PN-480	PN-600
C (wt %)	85.40	88.40	83.70	80.70	89.60
H (wt %)	2.40	2.20	0.60	3.50	2.60
N (wt %)	0	0	0	0.70	1.00
O (wt %) ^a	12.20	9.40	15.70	15.10	6.80
ash (wt %)	0	0	0	8.70	12.80
H/C	0.34	0.30	0.09	0.52	0.35
O/C	0.11	0.08	0.14	0.14	0.06
yield (wt %)	18	16	16	27	21

^aCalculated by difference

H/C and O/C atomic ratios are also reported, because they correlate with the degree of aromaticity and polarity of the biochars, respectively.²⁶ The ash content of the biochars obtained from cellulose fibers was negligible, in agreement with previous results¹⁸ and as opposed to those obtained from *P. nigra* wood feedstock.¹⁷ The biochar yields ranged from 16 to 27 wt %, being higher in the biochars obtained from *P. nigra*

and following the expected decreasing trend with the pyrolysis temperature for a given precursor (Table 1).

With regard to chemical composition, all of the prepared biochars are rich-carbon materials. As a general rule, the carbon content increased with the pyrolysis temperature, with the exception of CEL-700. This is attributed to a higher extent in the devolatilization reactions of the precursor under a steam/nitrogen flow; for the case of CEL-700, the temperature might be too high, leading to the consumption of carbon.¹⁸ The decrease of the H/C values with the pyrolysis temperature indicates the higher aromatic character of the chars obtained at higher temperatures. A similar trend was reported for biochars from other biomass precursors.^{17,26} On the other hand, the polarity of the biochars, evaluated on the basis of the O/C values, is quite similar (Table 1).

The analysis of infrared spectra confirmed the presence of an aromatic carbonaceous network in all of the biochars. As seen in Figure 1, all of the samples were characterized by broad-

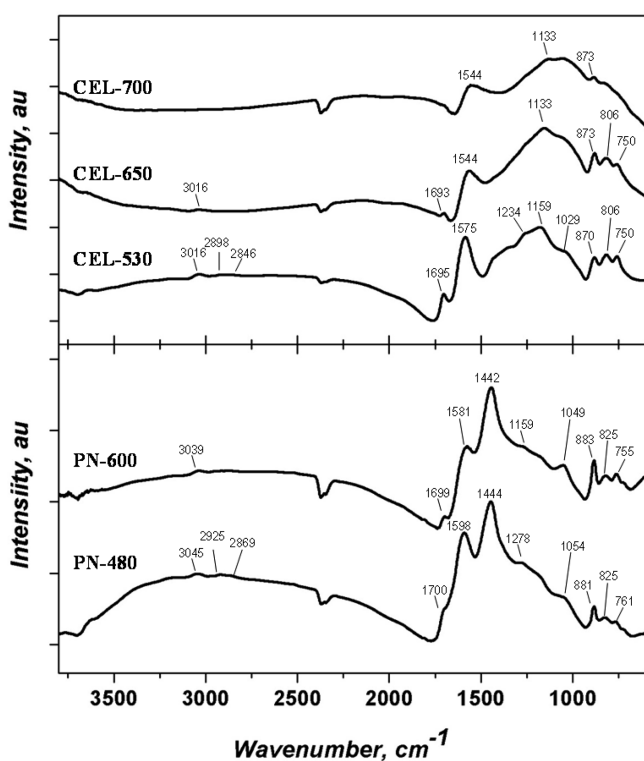


Figure 1. FTIR spectra of biochars synthesized from (top) cellulose fibers and (bottom) *P. nigra* wood.

shaped spectra, with signals in spectral regions typical of ring vibrations in large condensed aromatic carbon skeletons. Around 3000 cm^{-1} , the low-intense bands as a result of the stretching vibrations of aliphatic and aromatic C–H bonds were detected; in the medium-frequency range (between 1700 and 1000 cm^{-1}), a broad combination of peaks generated by the overlapping of carbon skeleton adsorption bands (C=O, C=C, C–C, and C–H–C–O stretching and bending modes) was found. A third group of bands characteristic of aromatic C–H out-of-plane (OPLA) bending modes was also detected between 1000 and 600 cm^{-1} .^{27–31} With the increase in the pyrolysis temperature, the number of peaks in the infrared spectra is reduced; this is likely due to charring reactions and the removal of the labile functional groups (most likely oxygen-

containing groups) that are favored at such high pyrolysis temperatures.³²

For instance, the spectrum of sample CEL-530 is characterized by low-intense bands at 3016, 2988, and 2846 cm^{-1} , ascribed to aromatic and aliphatic C–H stretching modes in amorphous carbons.^{28–31} The peak at 1695 cm^{-1} is associated with stretching modes of C=O moieties in conjugated systems, while the intense peak at 1575 cm^{-1} arises from C=C stretching modes in aromatic networks.^{30,31} In the region between 900 and 1500 cm^{-1} , overlapped signals as a result of aromatic skeleton vibrations in carbon-defective networks (between 1100 and 1500 cm^{-1}), oxygen-containing groups (C–O–C glycosidic bonds around 1150 cm^{-1}), and C–H and C–OH ring deformations of carbohydrate moieties (1040 cm^{-1}) were detected.³² In the 900–700 cm^{-1} region, three main peaks assigned to solo (870 cm^{-1}), duo (806 cm^{-1}), and trio (750 cm^{-1}) configurations (e.g., OPLA bending modes of aromatic C–H bonds)^{28–31} were observed. The intensity of the three peaks is almost the same, indicating a high substitution degree in the aromatic network of this sample.^{30,31}

The infrared spectrum of sample CEL-650 displayed features similar to CEL-530, with bands showing a less structured shape and an overall decrease in the intensity in most of them. In the case of sample CEL-700, the bands in the high-frequency region are almost not detectable and the peaks ascribable to C=O stretching modes and aromatic C–H OPLA disappeared. Only the peak at 1548 cm^{-1} as a result of C=C stretching modes and the overlapped peaks between 900 and 1500 cm^{-1} as a result of skeleton vibrations are still present.

Comparatively, the infrared spectra of the *P. nigra*-derived biochars were quite different compared to those of cellulose-derived materials. Indeed, in the region between 900 and 1700 cm^{-1} , peaks ascribable to the aromatic network of lignin are identified for PN-480 and PN-600. The spectra of the latter also featured low-intense bands as a result of aromatic and aliphatic C–H stretching in amorphous carbons (ca. 3045, 2925, and 2869 cm^{-1}). The bands associated with stretching modes of C=O in ketones or carboxylic moieties in conjugated systems (ca. 1700 cm^{-1}), C=C stretching modes in aromatic networks (ca. 1598 cm^{-1}), lignin-derived moieties (ca. 1444 and 1278 cm^{-1}), and C–OH stretching of pyranose rings from the holocellulosic fraction of *P. nigra* wood were also observed.³²

In the region below 900 cm^{-1} , bending modes of solo, duo, and trio configurations with varied intensities were detected too; as opposed to the cellulose-derived biochars, the intensity of the solo signal was the highest, indicating a different but also high degree of condensation and substitution in the *P. nigra* wood-derived biochars. As for the impact of the pyrolysis temperature, samples PN-480 and PN-600 displayed similar spectral features, with a slight decrease in the intensity of the peak at 1590 cm^{-1} .

Textural Properties. It is well-known that chars usually display a constricted pore network comprised of nanopores of small dimensions accessible through pore mouths or necks of narrow dimensions.³³ As a result, gas adsorption at cryogenic temperatures (typically argon and nitrogen at -186 and -196 °C) may present kinetic restrictions and, thus, provide a limited value for the characterization of narrow micropores. Following International Union of Pure and Applied Chemistry (IUPAC) recommendations,³⁴ a combined approach based on the use of N_2 at -196 °C and CO_2 at 0 °C has been used to

better describe the porosity of prepared biochars herein.³⁵ This allows for the detection of any kinetic limitations at cryogenic temperatures while enabling the evaluation of the narrow microporosity from the analysis of the CO₂ adsorption isotherms at 0 °C.

The N₂ adsorption/desorption isotherms at −196 °C of the prepared samples are shown in Figure 2. With the exception of

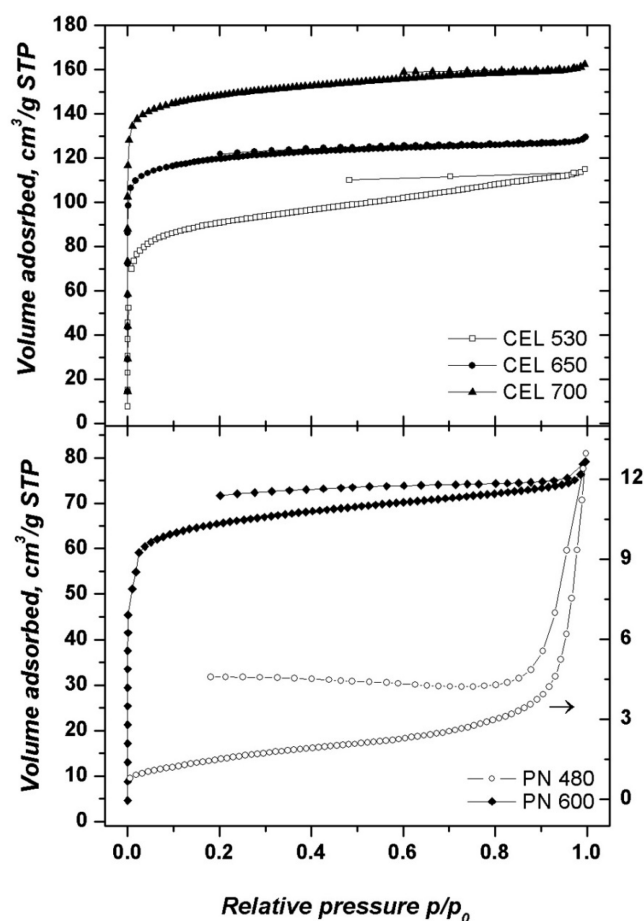


Figure 2. High-resolution N₂ adsorption/desorption isotherms at −196 °C of the synthesized biochars.

PN-480, all of the biochars presented N₂ adsorption isotherms belonging to type I in the IUPAC classification,³⁴ characteristic of microporous materials. Also, all of the isotherms present a non-reversible desorption branch, which is indicative of materials displaying a constricted microporous structure, often reported for carbonaceous chars (as mentioned above).^{33,35} This feature was less pronounced but still detected in the case of samples CEL-700 and CEL-650 (see magnification of the desorption branch in Figure S2 of the Supporting Information).

The nitrogen uptake is not very high for any of the obtained materials produced by steam-assisted slow pyrolysis, as expected for materials prepared upon carbonization at low temperature. Indeed, slow pyrolysis is typically carried out in an inert atmosphere, although the presence of oxidant agents, such as steam, has been reported to improve the porosity of the resulting chars as a result of the enhanced devolatilization favored by the efficient diffusion of steam in the precursor during pyrolysis, and, if the temperature is high enough, the capacity of steam to promote gasification reactions.^{17–21} The

temperatures selected herein for the steam-assisted slow pyrolysis tests were sufficiently low to limit the gasification of the feedstock, as evidenced in our previous work based on the analysis of the pyrolysis gas exhausts.^{17,18,36–38} Thus, the porosity of the obtained chars is mainly due to the enhanced devolatilization of the raw precursor in the presence of steam.

The low nitrogen uptake is more pronounced in the case of PN-480; this feature can be attributed to either a poor textural development or the presence of a constricted pore structure, through which the diffusion of nitrogen at cryogenic temperatures is restricted.³⁹ To further discriminate this effect, the CO₂ adsorption isotherms at 0 °C were recorded (Figure 3) and analyzed using various methods. This allowed us to

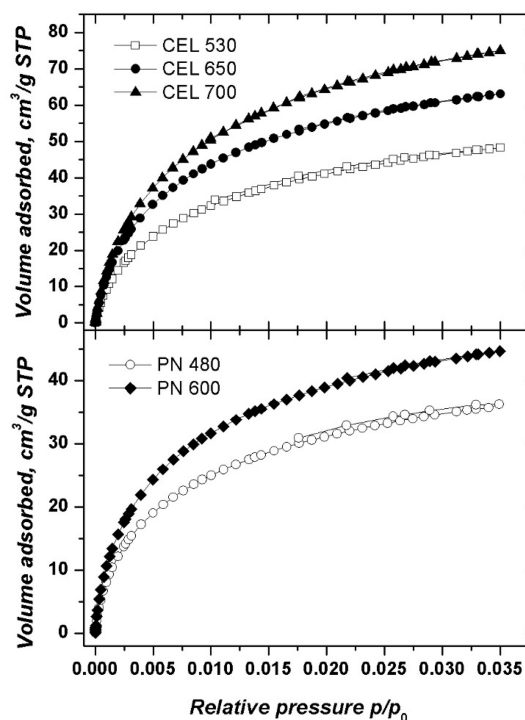


Figure 3. CO₂ adsorption/desorption isotherms at 0 °C of the synthesized biochars.

evaluate the so-called narrow microporosity (ultramicropores) of the biochars.⁴⁰ Data shown in Figure 3 and Table 2 confirmed that all of the samples displayed a well-developed narrow microporosity, including biochar PN-480. Interestingly, for all of the samples, the volume of total micropores (evaluated from N₂ data) is lower than the volume of narrow micropores evaluated from CO₂ data (Table 2); this confirmed the presence of nanopores of small dimensions in all of the biochars, which are not accessible to nitrogen as a result of diffusional limitations at −196 °C. Thus, sample PN-480 is a microporous biochar, despite the low BET surface area value.^{40,41} The uptake of CO₂ at 0 °C increased with the pyrolysis temperature, indicating that the development of the narrow microporosity is favored at higher temperatures.

The average narrow micropore size (L) was estimated from the CO₂ adsorption isotherms at 0 °C using the empirical correlation proposed by Stoeckli–Ballarini,⁴² valid for pore sizes between 0.35 and 1.3 nm.⁴³ As seen in Table 2, average values of ca. 0.6 and 0.55 nm were obtained for the cellulose- and *P. nigra*-derived biochars, respectively. These values are in

Table 2. Main Textural Features of the Synthesized Biochars Obtained from the N₂ and CO₂ Adsorption/Desorption Isotherms at −196 and 0 °C, Respectively

	S_{BET} (m ² /g)	V_{total}^a (cm ³ /g)	$V_{\text{micro}}^{(\text{DR},\text{N}_2)^b}$ (cm ³ /g)	$V_{\text{micro}}^{(\text{DR},\text{CO}_2)^c}$ (cm ³ /g)	$V_{\text{micro}}^{(\text{NLDFT},\text{CO}_2)^d}$ (cm ³ /g)	L^e (nm)
CEL-530	351	0.177	0.141	0.160	0.138	0.61
CEL-650	473	0.199	0.183	0.220	0.169	0.60
CEL-700	593	0.250	0.220	0.250	0.205	0.60
PN-480	6	0.018	0.002	0.120	0.097	0.56
PN-600	217	0.121	0.093	0.140	0.129	0.54

^aTotal pore volume measured at $p/p_0 \sim 0.99$. ^bTotal micropore volume evaluated by the DR method applied to N₂ adsorption data. ^cNarrow micropore volume evaluated by the DR method applied to CO₂ adsorption data. ^dNarrow micropore volume evaluated by NLDFT applied to CO₂ adsorption data. ^eAverage narrow micropore size evaluated by the Stoekli–Ballerini equation applied to CO₂ adsorption data.

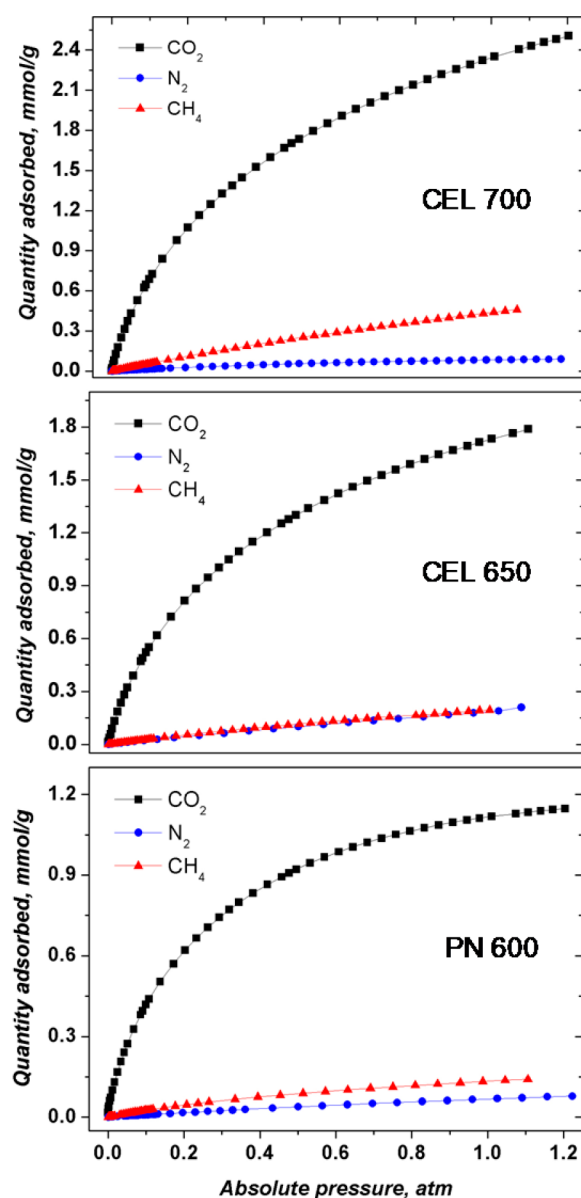
line with the average micropore sizes reported for carbon-based materials with good CO₂ adsorption capacities.⁴⁴

The BET surface area values ranged between 250 and 600 m²/g, increasing as the pyrolysis temperature is raised (Table 2). The biochars synthesized from *P. nigra* wood presented lower surface area values than those from cellulose fibers, which may be attributed to the different composition of the biomass precursors. Indeed, *P. nigra* wood has two major components (e.g., lignin and hemicellulose) besides cellulose, which would prevent the development of an incipient porosity during the pyrolysis (likely the cellulose fraction is entrapped in a compact matrix).³⁶ On the other hand, the contribution of the ashes (Table 1) to the lower surface area should not be neglected, because this magnitude is expressed per gram of material.³⁷ To further evaluate this issue, biochar PN-600 was washed in a 1 M NaOH solution to remove the alkali-soluble inorganic species. The washed material displayed a negligible ash content, and despite the surface area and total pore volume increased by ca. 30% (Table S1 and Figure S3 of the Supporting Information), it still presented lower values than the biochars obtained from cellulose fibers.

CO₂ Uptake and Separation Capacity. The biochars showing the highest micropore volumes (namely, CEL-650, CEL-700, and PN-600) were selected for evaluating their ability to store and separate CO₂ at room temperature. The CO₂ isotherms at 25 °C for the selected samples are reported in Figure 4, along with the uptakes of N₂ and CH₄.

The highest CO₂ uptake was obtained for the cellulose-derived biochars (ca. 2.33 and 1.72 mmol/g for CEL-700 and CEL-650, respectively), with values almost twice as large as those of PN-600 (ca. 1.12 mmol/g). These values have been compared in Table 3 to the CO₂ uptakes of other biomass-derived carbon-based materials available in the literature. In Table 3, the CO₂ uptakes at 1, 0.15, and 0.3 atm were reported. The CO₂ uptakes at low pressures, namely, typical pressures for CO₂ capture in post-combustion purposes (0.15 atm) and biogas upgrading (0.30 atm), were included because, depending upon the pore size distribution, the fraction of the total CO₂ uptake at relatively low partial pressures could vary a lot for different carbon-based adsorbents.

Data in Table 3 provide evidence of a large variability of CO₂ uptake for biomass-derived carbons (covering biochars and biomass-derived activated carbons), likely as a result of the differences in the properties of the material associated with the nature of the starting feedstock and the synthesis conditions. Available data in the literature on CO₂ uptake of biochars are limited; however, it can be observed that the adsorption capacities of prepared biochars herein are comparable to those reported in the literature for other biomass-derived adsorbents obtained by slow pyrolysis or carbonization. For instance, the

**Figure 4.** Equilibrium CO₂, N₂, and CH₄ uptakes at 25 °C of selected biochars.

CO₂ uptake of CEL-700 (2.33 mmol/g) is similar to that reported for cellulose fibers carbonized at 700 °C (2.2 mmol/g).⁴⁵ On the other hand, the adsorption capacities are lower than those of activated carbons prepared by physical and/or chemical activation of biomass. This is reasonable considering that the porosity of reported biochars herein is much lower

Table 3. CO₂ Uptake at Room Temperature of the Synthesized Biochars^a

feedstock	type	synthesis	S _{BET} (m ² /g)	V _{micro} (cm ³ /g)	CO ₂ uptake (mmol/g)			reference
					1 atm	0.15 atm	0.30 atm	
cellulose fibers	biochar	steam-assisted slow pyrolysis at 650 °C	473	0.183	1.72	0.7	1	this work
cellulose fibers	biochar	steam-assisted slow pyrolysis at 700 °C	593	0.220	2.33	0.9	1.4	this work
<i>P. nigra</i> wood	biochar	steam-assisted slow pyrolysis at 600 °C	217	0.093	1.12	0.5	0.75	this work
hickory wood	biochar	slow pyrolysis at 600 °C	401		1.39			46
sugar cane bagasse	biochar	slow pyrolysis at 600 °C	388		1.67			46
sawdust	biochar	gasification at 850 °C	182	0.0036	1.08			47
almond shell	biochar	single-step oxidation (3% O ₂)	557	0.21 ^b	2.11	~1.05	~1.5	48
olive stone	biochar	single-step oxidation (3% O ₂)	697	0.27 ^b	2.02	~0.8	~1.3	48
cellulose fibers	activated carbon	carbonization at 700 °C	499	0.193	2.21	~0.7	~1.2	45
cellulose fibers	activated carbon	carbonization at 700 °C and physical activation	599	0.229	2.61	~1	~1.6	45
cellulose fibers	activated carbon	carbonization at 800 °C and physical activation	863	0.334	3.78	~1.15	~2	45
pine nut shell	activated carbon	carbonization and chemical activation	1486	0.64	5.0	2	~2.9	44
olive stone	activated carbon	physical activation	1479	0.594	3.05	0.80	~1.4	49
lignin	activated carbon	chemical activation	2246	0.753	2.38	0.52	~0.9	49
lignin	activated carbon	carbonization	71	0.033	2.20	0.92	~1.4	49
eucalyptus wood	activated carbon	chemical activation	1889	1.063	2.98			50
bamboo	activated carbon	chemical activation	1846	0.36 ^b	4.5	~1.2	~2.1	51
coconut shell	activated carbon	carbonization and physical activation	1327	0.55	3.9	~1.5	~1.9	52
African palm shells	activated carbon	carbonization	365	0.16	1.9	~0.9	~1.9	53
African palm shells	activated carbon	chemical activation	1250	0.55	4.4	~1.5	~1.9	53
vine shoots	activated carbon	physical activation	767	0.245	3.1	~1.2	~1.8	54
vine shoots	activated carbon	chemical activation	1439	0.493	4	~1.2	~2	54

^aData from selected best performing biomass-derived carbons reported in the literature are included for comparison purposes, along with selected characteristics and textural parameters (unless otherwise stated, micropore volumes were estimated from N₂ adsorption data and CO₂ uptakes were estimated at 25/30 °C and 1 bar). ^bEstimated from CO₂ adsorption data at 0 °C.

than that of activated carbons (steam-assisted slow pyrolysis conditions enhanced the surface area and overall microporosity of the prepared biochars).

It is well-known that the CO₂ adsorption capacity is strongly influenced by the adsorbent structural features and the operating conditions and that various mechanisms may be involved. In carbon-based adsorbents, the main parameters governing both the CO₂ uptake capacity and selectivity at low partial pressures and ambient temperature are the microporosity (volume and size) and functionalization of the pores.^{16,49} At low pressure, the adsorption is favored in narrow micropores and the mechanism is based on short-range non-specific interactions between the gas and adsorbent. At higher pressures, the surface coverage is the predominant mechanism and, therefore, wider micropores become more relevant (see ref 55 and references therein).

Considering this and aiming to better understand the CO₂ uptake of the prepared biochars under typical post-combustion conditions, we have evaluated the isosteric heat of CO₂ adsorption and discussed the adsorption capacity in terms of the physicochemical features of the biochars. The values of isosteric heat of CO₂ adsorption (determined using the Clausius–Clapeyron equation) were ca. 35, 22, and 34 kJ/mol for CEL-650, CEL-700, and PN-600, respectively (Figure S4 of the Supporting Information). The enthalpy values below 40 kJ/mol confirmed that physisorption is the dominant adsorption mechanism in these samples,⁵⁶ with the narrow micropores being the main adsorption sites. Indeed, the heat of adsorption of biochar CEL-700 was significantly lower than those of CEL-650 and PN-600, which is attributed to its wider distribution of micropores (Table 2). Additionally, the isosteric

heats of adsorption were quite constant with the CO₂ coverage (Figure S3 of the Supporting Information), indicating energetically homogeneous adsorption sites. Overall, the CO₂ isosteric heats of adsorption are in line with those reported for other biochars obtained by slow pyrolysis.⁴⁶ These values are also within those recommended for post-combustion capture from an engineering point of view (between 30 and 60 kJ/mol).^{55,56}

The ability of the biochars to adsorb N₂ and CH₄ at room temperature was also explored. As seen in Figure 3, the uptake of both gases is much lower than that of CO₂, indicating that the biochars are good materials for the separation of these gases. It is well-known that carbon adsorbents display a much lower affinity for nitrogen than carbon dioxide; in the case of methane, the lower uptake is attributed to the pore structure of the biochars, because methane is slightly bulkier than CO₂. CEL-700 exhibited the highest CH₄ uptake (ca. 0.44 mmol/g), and PN-600 exhibited the lowest CH₄ uptake (ca. 0.14 mmol/g). These values are within those reported for other biochars.⁵⁷

As opposed to CO₂, where the uptake correlates with the volume of narrow micropores (Figure S5 of the Supporting Information), the uptake of CH₄ correlates well with the total micropore volume evaluated from the N₂ adsorption data (Figures S6 and S7 of the Supporting Information). This is due to the different sizes of the probe molecules and confirmed that materials with a narrow distribution of micropores are suitable for CO₂ storage and separation of CO₂/CH₄ mixtures. On the contrary, wider micropores would favor the uptake of methane,^{55,58,59} as in the case of sample CEL-700.

Gas Selectivity (CO₂/N₂ and CO₂/CH₄). On the basis of their adsorption features and potential adequateness to

separate CO₂ from N₂ and/or CH₄, we further investigated the selectivity of these biochars by the application of the IAST to the single-component experimental gas adsorption isotherms. IAST has been used for the prediction of multi-component adsorption mixtures on a wide range of porous materials having homogeneous energetic adsorption sites.^{60,61} We have herein applied IAST to CO₂/N₂ and CO₂/CH₄ mixtures, representative of synthetic flue gases in post-combustion processes (with a v/v gas composition of 15:85) and natural gas upgrading processes (e.g., with v/v gas compositions of 30:70), respectively. The modeled isotherms are shown in Figure S8 of the Supporting Information, while IAST selectivities are compiled in Table 4. As seen, CEL-700 and PN-600 displayed higher CO₂/N₂ selectivities compared to CEL-650. However, a different trend was obtained for the

separation of CO₂/CH₄, where biochar CEL-700 showed a lower selectivity value. This is in agreement with the wider micropore size distribution of this biochar (Table 2); thus, the micropores are accessible to methane. Furthermore, the CO₂/N₂ and CO₂/CH₄ selectivities show a good correlation with textural parameters of the biochars in terms of the pore volume and average micropore size, respectively (Figure S9 of the Supporting Information).

Table 4 also compiles CO₂/N₂ selectivity values reported in the literature for other adsorbents evaluated for typical post-combustion flue gas mixtures (i.e., CO₂/N₂, 15:85, v/v). The selectivities of our biochars are within the range of those reported for some activated carbons with much higher porosity but lower than those of synthetic polymers, zeolites and metal–organic framework (MOF). It is worth noting that only few data are available about ideal CO₂/CH₄ selectivities for a typical natural gas upgrading (CO₂/CH₄, 30:70, v/v); therefore, it appears that the selectivities of the biochars, object of this work, are quite higher than those reported for synthetic carbons (Table 4).

Table 4. CO₂/N₂ and CO₂/CH₄ Selectivities at 25 °C Calculated from IAST Simulations for Different Gas Mixtures ($S_{ij} = C_i/x_j/C_j/x_i$)^a

adsorbent type	CO ₂ /N ₂ (15:85)	CO ₂ /CH ₄ (30:70)	reference
cellulose fibers biochar (CEL-650)	34 (6)	32 (13.7)	this work
cellulose fibers biochar (CEL-700)	119 (21)	14 (6)	this work
<i>P. nigra</i> wood biochar (PN-600)	105 (18.6)	34 (14.7)	this work
synthetic carbon (C125-220)	(4.0) ^b	7.2 ^b	61
synthetic carbon (C200-180)	(3.9) ^b	2.1 ^b	61
synthetic carbon (C _{ReHy12} @600)	(7.1) ^b		62
synthetic carbon (C _{ReHy12} @700)	(5.3) ^b		62
synthetic carbon (C _{ReHy13} @450)	(9.4) ^b		62
synthetic carbon (C _{ReHy13} @500)	(13.0) ^b		62
N-doped carbon (IBN9-NC1)	~32		63
N-doped carbon (IBN9-NC1-A)	~24		63
activated carbon (pomegranate peels)	15.1		64
activated carbon (carrot peels)	8.1		64
activated carbon (fern leaves)	5.6		64
activated carbon (mistletoe branches)	11.4		64
activated carbon (mistletoe leaves)	12.0		64
activated carbon (kiwi fruit peels)	10.6		64
activated carbon (sugar beet pulp)	2.8		64
synthetic carbons (CKHP800-2)	50		65
sulfonate-grafted porous polymer (PPN-6-SO ₃ H)	155 ^c		66
sulfonate-grafted porous polymer (PPN-6-SO ₃ Li)	414 ^c		66
zeolite NaX	110 ^c		66
MOF-505	7.6		67
composite MOF-505@5GO	8.6		67
UiO-66	19.4		68
UiO-66(Zr)–(COOH) ₂	37.9		68
UiO-66(Zr)–(COOLi) ₂	50.8		68
UiO-66(Zr)–(COONa) ₂	58.0		68
UiO-66(Zr)–(COOK) ₂	69.3		68
MIL-101 (Cr)	21		69
MIL-101 (Cr, Mg)	86		69

^aData of carbon materials available in the literature are also compiled for comparison; selectivity values in parentheses have been calculated as $S_{ij} = C_i/C_j$. Unless otherwise stated, experimental conditions were similar for all of the samples. ^bValues calculated at 1.2 bar. ^cValues calculated at 22 °C.

CONCLUSION

The ability of biochars produced by slow steam-assisted pyrolysis to store and separate CO₂ at room temperature was investigated, and some correlations with the biochar textural properties were highlighted. Predictions of the adsorption of binary gaseous mixtures (CO₂/CH₄ and CO₂/N₂) by applying the IAST were also carried out. Moreover, the storage and selectivity properties of selected samples have been compared to available literature data (available data in the literature on CO₂ uptake of biochars is limited).

The collected data revealed that the adsorption capacities (1.5–2.5 mmol/g of CO₂ uptake range) of selected biochars (from cellulose fibers and *P. nigra* wood) were comparable to those reported in the literature for other biomass-derived adsorbents obtained by slow pyrolysis or carbonization but lower than those of activated carbons prepared by physical and/or chemical activation of biomass. The suitability of the selected biochars for CO₂ uptake was also proven by the good correlation obtained between the amount of CO₂ adsorbed and the pore volumes (biochars exhibited a narrow microporosity, whose volume increases with the increasing of the pyrolysis temperature and average pore diameters between 0.55 and 0.6 nm). The samples also exhibited quite good selectivities for CO₂ over both N₂ and CH₄ [between 34 and 119 for a CO₂/N₂ mixture in typical post-combustion conditions (15:85, v/v) and between 14 and 34 for a CO₂/CH₄ mixture typical of natural gas upgrading (30:70, v/v)]. In particular, the CO₂/N₂ selectivity values resulted within the range of those reported for some activated carbons with much higher porosity but lower than those of synthetic polymers, such as zeolites and MOFs.

Overall, the results reported in this work demonstrate that using the biochar as an adsorbent in the gas treatment process could be a sustainable approach and this is even truer when biochar production represents a route for the valorization of a waste material.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.energyfuels.8b01058.

Thermogravimetry profiles of biochar precursors (Figure S1), N_2 isotherms of cellulose-derived biochar (Figure S2), N_2 and CO_2 isotherms of PN-600 NaOH (Figure S3), main textural features of samples PN-600 and PN-600 NaOH (Table S1), isosteric heats of adsorption (Figure S4), correlation graphics (Figures S5–S7 and S9), and IAST-modeled curves (Figure S8) (PDF)

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Notes

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