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# Digital Simulation of Bioluminescent Bacteria Cells Tweeting via Quorum-Sensing **Molecules**

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In this paper, we propose a computational multiagent model of a bacterial light-producing communication system, termed quorum sensing (QS). Specifically, we propose a bottom-up agent-based approach combined with Ordinary Differential Equations, which abstract the intracellular dynamics, such as a proposed bioluminescence model. Results show that bacteria cells have a metabolism allowing them to grow, reproduce, interact, and cooperate at the population level to exhibit nearoptimal light producing behaviors. The ultimate goal is to develop a self-regulated network in which the rules governing the formation of the network are linked to the internal dynamics of its units without any centralized control. Such a bacterial-inspired networks, can address issues such as mobility, and energy that are key factors for the development of new self-organized network such as mobile Ad-hoc networks.

#### Cellular Dynamics Model

To simulate cell-to-cell communication, we use a generic LuxI/LuxR QS. All LuxI/R systems are mediated by autoinducers, such as acylated homoserine lactone (AHL). Explicitly, the AHL and the receptor LuxR form a dimerized complex (C) that regulates the expression of both LuxI and LuxR genes. The QS model is based on the ODE-models proposed in (Melke et al., 2010). We model bioluminescence or light production as a function of the dimerized complex (C) as follows:

$$\frac{d[L]}{dt} = \frac{L_{max}[C]}{K_L + [C]}$$

where L is the light production rate,  $L_{max}$  is the maximum light production rate and  $K_L$  is the concentration of C at which L is at half-maximum. Bioluminescence is expressed as the accumulation of the green fluorescent protein gfp. The bacterial metabolism (ATP cycle) is implemented as follows:

$$\Delta ATP = (ATP_0 + E.ATP_S + ATP_G + ATP_D + ATP_L + ATP_M)(\Delta t)$$

Where:

- $ATP_0$  is the basal energy.
- ATP<sub>S</sub> represents the energy produced from substrates and is calculated as follows:

$$\frac{d[ATP_S]}{dt} = Y_{ATP/S} \cdot \frac{d[S]}{dt}$$

 $\frac{d[ATP_S]}{dt} = Y_{ATP/S} \cdot \frac{d[S]}{dt}$  Where  $Y_{ATP/S}$  is the energetic substrate yield *i.e.* the amount of ATP produced by per a unit of substrate (S).

- *E* is the substrate metabolism efficiency.
- $ATP_G$  represents consumption of ATP due to the cell growth and is calculated as follows:

$$\frac{d[ATP_G]}{dt} = -\frac{1}{Y_{X/ATP}} \cdot \frac{d[X]}{dt}$$
Where  $Y_{X/ATP}$  is the energetic growth yield coefficient.

- ATP<sub>D</sub> is the division energy.
- $ATP_L$  is the light energy. The consumption of energy due to light production is given by:

$$\frac{d[ATP_L]}{dt} = -\frac{1}{Y_{L/ATP}} \cdot \frac{d[L]}{dt}$$
Where  $Y_{L/ATP}$  is the energetic light yield coefficient.

 $ATP_{M}$  is the maintenance energy.

#### Results

The experiments were conducted using "gro" (Jang et al., 2012). The number of bioluminescent cells represents 73% of the population. This corresponds to the empirical rates found in real populations of bacteria (Anetzberger, Pirch, & Jung, 2009). At the population level, figure 1.d shows a self organized behavior of the microbial population. Figure 2a shows the amount of autoinducer over time. In our simulations the accumulation of AHL exceeds the rate achieved in (Melke et al., 2010) (0.6 against 0.1). This shows that the cells in our model provide self-sustainable behaviors that allow them to exploit metabolism (figure 2b) to grow, survive and produce more AHL molecles, which in turn triggers communication.

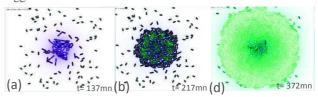
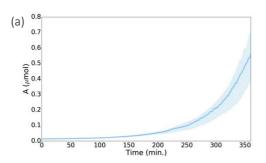


Figure 1: Evolution of bioluminescence. Substrate is shown in purple, AHL in blue, black cells are non-fluorescent. (a) cells grow and divide. In (b) the quorum is being reached, fluorescent cells begin to appear. (c): Bioluminescent cells organize themselves around the colony to be located at the edges of it.



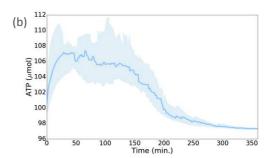


Figure 2: Evolution of the average amount of autoinducer (a) and ATP (b) inside cells over time : median and interquartile range of 20 independent runs of the simulation)

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