



Kinetics Study of Glucose Acetylation in Starch Polymers by Quantitative ^{13}C NMR spectroscopy.

Benoit Laignel, Christophe Bliard, Stéphane Bourg, Jean-Marc Nuzillard, G. Massiot

► To cite this version:

Benoit Laignel, Christophe Bliard, Stéphane Bourg, Jean-Marc Nuzillard, G. Massiot. Kinetics Study of Glucose Acetylation in Starch Polymers by Quantitative ^{13}C NMR spectroscopy.. ISPAC 10, Stephen T. Blake, Aug 1997, Toronto, Canada. pp.36-37. hal-02117657v1

HAL Id: hal-02117657

<https://hal.science/hal-02117657v1>

Submitted on 2 May 2019 (v1), last revised 25 Nov 2019 (v2)

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

10TH

**INTERNATIONAL
SYMPOSIUM ON
POLYMER
ANALYSIS AND
CHARACTERIZATION**

AUGUST 10-13, 1997

**University of Toronto
Toronto, Ontario
Canada**

TENTH INTERNATIONAL SYMPOSIUM ON POLYMER ANALYSIS AND CHARACTERIZATION

August 10-13, 1997

**University of Toronto
Toronto, Ontario
CANADA**

GOVERNING BOARD MEMBERS

Dr. Stephen T. Balke*
Department of Chemical Engineering
University of Toronto
Toronto, Ontario, Canada

Dr. Howard Barth**
Du Pont Experimental Station
Wilmington, Delaware, USA

Dr. Guy C. Berry
Department of Chemistry
Carnegie Mellon University
Pittsburgh, PA, USA

Dr. H.-J. Cantow
University of Freiburg
Freiburg, Germany

Dr. Oscar Chiantore
Department of Chemistry
University of Torino
Torino, Italy

Dr. John V. Dawkins
Chemistry Department
Loughborough University of Technology
Loughborough, Leicestershire
Great Britain

Dr. Nikos Hadjichristidis
Division of Chemistry
University of Athens
Athens, Greece

Dr. Josef Janca
Université de La Rochelle
La Rochelle, France

Dr. Pavel Kratochvil
Academy of Sciences of the Czech Republic
Prague, Czech Republic

Dr. Sadao Mori
Faculty of Engineering
Mie University
Mie, Japan

Dr. Petr Munk
Department of Chemistry
University of Texas at Austin
Austin, Texas, USA

Dr. Marguerite Rinaudo
CERMAV-CNRS
Grenoble, France

* ISPAC-10 Chair, Department of Chemical Engineering and Applied Chemistry,
University of Toronto, Toronto, Ontario, Canada, M5S 3E5, (416) 978-7495

** Program Chairman, Du Pont Co., Experimental Station, Bldg. 228, Wilmington, DE 19898,
(302) 695-4354

- 28 Brito, Z., Torrealba, V., Sánchez, G., Hernández, L., Kinetic Characterization in DGEBA/ETDA with Mono and Bimetallic Fillers from Thermogravimetric Curves**
- 29 Maccone, P., Apostolo, M., Molecular Weight Distribution of Fluorinated Polymers with Long Chain Branching**
- 30 Maccone, P., Dynamic Light Scattering of Concentrated Solutions of Perfluoropolyethers, Polyisocyanate and Apolar Solvent in a Ternary System**
- 31 D'Agnillo, L., Soares, J.B.P., Penlidis, A., A Hierarchical Data Analysis of a Replicate Experiment in Polyethylene Synthesis with High-Temperature Gel Permeation Chromatography**
- 32 Soares, J.B.P., Abbott, R.F., Willis, J.N., Liu, X., New Methodologies for Studying Multiple-Site-Type Catalysts for the Copolymerization of Olefins**
- 33 Ohtani, H., Nonobe, T., Tsuge, S., Kitayama, T., Hatada, K., Stereoregularity of Poly(methyl methacrylate)s Studied by Pyrolysis-Gas Chromatography/Mass Spectroscopy**
- 34 Pagé, J.Y.S.D., Bonin, H.W., Bui, V.T., Bates, P.J., Viscoelastic Analysis of Irradiated Polymer Resins (Polycarbonate, Polysulfone and Polyetheretherketone)**
- 35 Joly, C., Bliard, C., Espuche, E., Escoubès, M., Acetylated Wheat Starch Membranes: Permeability Measurements**
- 36 Laignel, B., Bliard, C., Bourg, S., Nuzillard, J.M., Massiot, G., Kinetics Study of Glucose Acetylation in Starch Polymers by Quantitative ^{13}C NMR Spectroscopy**
- 37 Vancso, G.J., Pearce, R., Real-Time Imaging of Melting and Crystallization in Poly(Ethylene Oxide) by Atomic Force Microscopy with Lamellar Resolution**
- 38 Tanaka, M., Shimbo, K., Yamada, T., Ohfusa, Y., Taguchi, M., Suzuki, H., MacFarlane, J.D., Development of Downsized Columns and System for GPC Analyses**
- 39 Fan, X-D., Deng, Y., Waterhouse, J., Pfromm, P., Synthesis and Characterization of Soy-Based Copolyamides Consisting of Amino Acids or Soy-Protein**
- 40 Wong, W.S., Schmidt, S., Gillespie, D., Application of SEC for the Characterization of Cellulose Materials**

Kinetics Study of Glucose Acetylation in Starch Polymers by Quantitative ^{13}C NMR Spectroscopy

B. Laignel, C. Bliard, S. Bourg, J.M. Nuzillard and G Massiot

Laboratoire de Pharmacognosie, UPRESA 6013 CNRS, Université de Reims Champagne Ardenne, CRPB Bât 18, Moulin de la Housse, BP 1039, 51 097 Reims Cedex 2 - France -

Introduction

Naturally abundant glucose polymers are chemically modified and developed as alternative sources of materials in polymer industries. In the course of a program devoted to the chemical derivatization of starch macromolecules, the acylation kinetics and the rate of modification on each of the monomer positions was investigated. It was found that products having similar acetyl percentages but obtained by different chemical modifications processes had different substitution patterns for intermediate modification (1). In order to determine the relative acetylation rates on positions -2, -3, and -6 of the glucopyranose monomer subunit the acetylation kinetics for each of these positions was analysed.

NMR Study

NMR is the method of choice for investigating the substitution distribution in chemically modified polysaccharides. With a large spectral width, ^{13}C NMR gives a broad spectrum of well differentiated resonances for closely related chemical structures. It is thereby possible to assign and quantify well separated peaks. In order to perform ^{13}C resonance quantification, it is necessary to suppress NOE effects and leave enough time for the magnetization to return to equilibrium with the help of a relaxation agent. Under these conditions, a satisfactory degree of accuracy in the measurement of the area of each peak can be obtained (2).

Quantitative determination of polymaltoside esters

Starch macromolecules are polymaltosides constituted of poly- $\alpha(1\rightarrow 4)$ -glucopyranosyl- subunits on which one primary (-OH -6) and two secondary (-OH -2, -OH -3) hydroxyl groups can be esterified (figure 1)

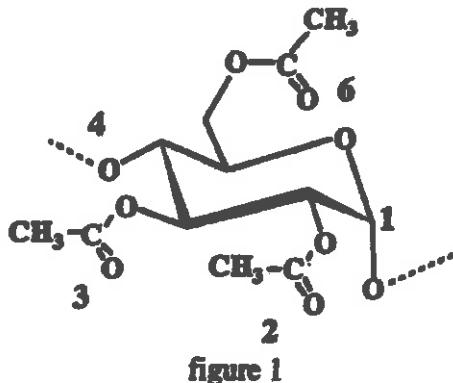


figure 1

The carbonyl ester carbon atoms are separated on the ^{13}C NMR spectra and assigned through long distance ^1H - ^{13}C coupling with each corresponding glucopyranose ring proton. δ 169.3 (C3), 170.2 (C6), 170.6 (C2) ppm for the triacetate and δ 172.8, 173.8 and 174.0 ppm for the amylose tripropionate in deuteriochloroform.

Methods

A chromium salt ($\text{Cr}(\text{Acac})_3$) is added as a relaxation agent to bring the repetition time to ca. 5 s. The spectra were recorded at 75.4 MHz on a AC 300 Bruker spectrometer with a minimum of 1000 scans. A quantitative ^{13}C nmr spectrum (3) is decomposed into a sum of lorentzian peaks with a non-linear adjustment technique. The parameters (width, height and resonance frequencies), obtained after a first analysis of the different peaks, are optimized through a Levenberg-Marquardt (4) type procedure. The areas of the 2, 3, and 6 peaks, the sum of which is proportional to the total substitution degree as determined by titrimetry, can be obtained by the values of these parameters. Partial substitution degrees can thus be determined.

Kinetics study

Acetylation kinetics were investigated on each -2, -3 and -6 position in the course of two different chemical processes. In the first one (method A), native starch was physically destructured by solubilisation in water, lyophilisation followed by dissolution in pyridine and reaction with two -OH equivalents of acetic anhydride for 2, 3, 4 and 5 hours (5). In the second experimental conditions (method B), starch was submitted to acetylation following the procedure of Mehlretter et al. (6). An aliquote was sampled after 1, 2, 3, 4, or 5 hours. The remaining OH groups on each sample were acylated as propionates with propionic anhydride in pyridine for 24 h (7) providing deuteriochloroform soluble triacylated bi-esters. IR spectra were recorded to confirm the absence of OH groups.

Results

The results of the kinetics are shown in figure 2

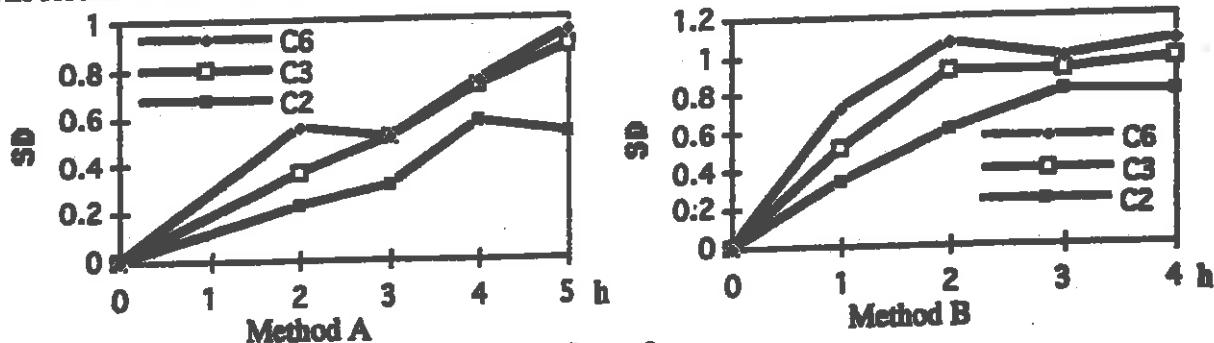


figure 2

Results show that in method A the reactivity of the secondary position -3 is surprisingly close to the primary one in position -6 and even if the primary -OH reacts faster in method B, the difference in reactivity between position -6 and position -3 is the same as that observed between position -3 and position -2.

Conclusion

In this paper ^{13}C NMR spectra of acetylated starch samples were recorded and quantitative ^{13}C NMR allowed the measurement of the relative rates of acetylation on each -OH position during the course of two different chemical processes.

Acknowledgment :

We thank Europol'Agro and Région Champagne Ardenne for financial support.

References

- 1) B. Laignel, C. Bliard, J. M. Nuzillard, G. Massiot, *Carbohydr. Res.*, 298 (1997) 251-260
- 2) Buchanan C. M., Edgar K.J., Wilson A.K.; *Macromol.*, 24 (1991) 3060-3064
- 3) Freeman R., Hill H.D.W.; *J. Magn. Res.* 7, (1972) 327-329
- 4) Marquardt D.W.; *J. of the Soc. for Indust. and Appl. Mathematics* 11 (1963) 431-441
- 5) Laignel B., Ph.D thesis URCA Reims University France 1997
- 6) Mark A.M., Mehlretter C.L.; *Starch* 24 (1972) 73-76-
- 7) Wurtzburg O.B.; *Methods in Carbohydr. Chem.* Ed. Whistler R.L., Acad. Press , 4 (1964) 286-288.